

The Enzyme List

Class 1 — Oxidoreductases

Nomenclature Committee
of the
International Union of Biochemistry and Molecular Biology
(NC-IUBMB)

L^AT_EX version prepared by Andrew McDonald,
School of Biochemistry and Immunology, Trinity College Dublin, Ireland

Generated from the [ExplorEnz](#) database, May 2023

© 2023 IUBMB

Contents

EC 1.1 Acting on the CH-OH group of donors	4
EC 1.1.1 With NAD ⁺ or NADP ⁺ as acceptor	5
EC 1.1.2 With a cytochrome as acceptor	101
EC 1.1.3 With oxygen as acceptor	103
EC 1.1.4 With a disulfide as acceptor	113
EC 1.1.5 With a quinone or similar compound as acceptor	113
EC 1.1.7 With an iron-sulfur protein as acceptor	117
EC 1.1.9 With a copper protein as acceptor	117
EC 1.1.98 With other, known, physiological acceptors	118
EC 1.1.99 With unknown physiological acceptors	120
EC 1.2 Acting on the aldehyde or oxo group of donors	128
EC 1.2.1 With NAD ⁺ or NADP ⁺ as acceptor	128
EC 1.2.2 With a cytochrome as acceptor	151
EC 1.2.3 With oxygen as acceptor	152
EC 1.2.4 With a disulfide as acceptor	155
EC 1.2.5 With a quinone or similar compound as acceptor	156
EC 1.2.7 With an iron-sulfur protein as acceptor	157
EC 1.2.98 With other, known, physiological acceptors	160
EC 1.2.99 With unknown physiological acceptors	160
EC 1.3 Acting on the CH-CH group of donors	162
EC 1.3.1 With NAD ⁺ or NADP ⁺ as acceptor	162
EC 1.3.2 With a cytochrome as acceptor	189
EC 1.3.3 With oxygen as acceptor	190
EC 1.3.4 With a disulfide as acceptor	194
EC 1.3.5 With a quinone or related compound as acceptor	194
EC 1.3.7 With an iron-sulfur protein as acceptor	196
EC 1.3.8 With a flavin as acceptor	200
EC 1.3.98 With other, known, physiological acceptors	205
EC 1.3.99 With unknown physiological acceptors	206

EC 1.4 Acting on the CH-NH₂ group of donors	214
EC 1.4.1 With NAD ⁺ or NADP ⁺ as acceptor	214
EC 1.4.2 With a cytochrome as acceptor	220
EC 1.4.3 With oxygen as acceptor	221
EC 1.4.4 With a disulfide as acceptor	227
EC 1.4.5 With a quinone or other compound as acceptor	227
EC 1.4.7 With an iron-sulfur protein as acceptor	228
EC 1.4.9 With a copper protein as acceptor	228
EC 1.4.98 With a copper protein as acceptor	229
EC 1.4.99 With unknown physiological acceptors	229
EC 1.5 Acting on the CH-NH group of donors	230
EC 1.5.1 With NAD ⁺ or NADP ⁺ as acceptor	230
EC 1.5.3 With oxygen as acceptor	243
EC 1.5.4 With a disulfide as acceptor	249
EC 1.5.5 With a quinone or similar compound as acceptor	249
EC 1.5.7 With an iron-sulfur protein as acceptor	250
EC 1.5.8 With a flavin or flavoprotein as acceptor	251
EC 1.5.98 With other, known, physiological acceptors	252
EC 1.5.99 With unknown physiological acceptors	253
EC 1.6 Acting on NADH or NADPH	255
EC 1.6.1 With NAD ⁺ or NADP ⁺ as acceptor	255
EC 1.6.2 With a heme protein as acceptor	257
EC 1.6.3 With oxygen as acceptor	258
EC 1.6.5 With a quinone or similar compound as acceptor	260
EC 1.6.6 With a nitrogenous group as acceptor	263
EC 1.6.99 With unknown physiological acceptors	264
EC 1.7 Acting on other nitrogenous compounds as donors	265
EC 1.7.1 With NAD ⁺ or NADP ⁺ as acceptor	265
EC 1.7.2 With a cytochrome as acceptor	269
EC 1.7.3 With oxygen as acceptor	271
EC 1.7.5 With a quinone or similar compound as acceptor	273
EC 1.7.6 With a nitrogenous group as acceptor	273
EC 1.7.7 With an iron-sulfur protein as acceptor	274
EC 1.7.99 With unknown physiological acceptors	274
EC 1.8 Acting on a sulfur group of donors	275
EC 1.8.1 With NAD ⁺ or NADP ⁺ as acceptor	275
EC 1.8.2 With a cytochrome as acceptor	280
EC 1.8.3 With oxygen as acceptor	282
EC 1.8.4 With a disulfide as acceptor	283
EC 1.8.5 With a quinone or similar compound as acceptor	287
EC 1.8.7 With an iron-sulfur protein as acceptor	290
EC 1.8.98 With other, known, physiological acceptors	291
EC 1.8.99 With unknown physiological acceptors	293
EC 1.9 Acting on a heme group of donors	294
EC 1.9.3 With oxygen as acceptor	294
EC 1.9.6 With a nitrogenous group as acceptor	294
EC 1.9.98 With other, known, physiological acceptors	294
EC 1.9.99 With unknown physiological acceptors	295

EC 1.10 Acting on diphenols and related substances as donors	295
EC 1.10.1 With NAD ⁺ or NADP ⁺ as acceptor	295
EC 1.10.2 With a cytochrome as acceptor	295
EC 1.10.3 With oxygen as acceptor	295
EC 1.10.5 With a quinone or related compound as acceptor	298
EC 1.10.9 With a copper protein as acceptor	299
EC 1.10.99 With unknown physiological acceptors	299
EC 1.11 Acting on a peroxide as acceptor	299
EC 1.11.1 Peroxidases	299
EC 1.11.2 Peroxygenases	308
EC 1.12 Acting on hydrogen as donor	310
EC 1.12.1 With NAD ⁺ or NADP ⁺ as acceptor	310
EC 1.12.2 With a cytochrome as acceptor	311
EC 1.12.5 With a quinone or similar compound as acceptor	312
EC 1.12.7 With an iron-sulfur protein as acceptor	312
EC 1.12.98 With other, known, physiological acceptors	312
EC 1.12.99 With unknown physiological acceptors	314
EC 1.13 Acting on single donors with incorporation of molecular oxygen (oxygenases)	314
EC 1.13.1 Acting on single donors with incorporation of molecular oxygen (oxygenases)	314
EC 1.13.11 With incorporation of two atoms of oxygen	315
EC 1.13.12 With incorporation of one atom of oxygen (internal monooxygenases or internal mixed-function oxidases)	337
EC 1.13.99 Miscellaneous	342
EC 1.14 Acting on paired donors, with incorporation or reduction of molecular oxygen	343
EC 1.14.11 With 2-oxoglutarate as one donor, and incorporation of one atom of oxygen into each donor	344
EC 1.14.12 With NADH or NADPH as one donor, and incorporation of two atoms of oxygen into the other donor	364
EC 1.14.13 With NADH or NADPH as one donor, and incorporation of one atom of oxygen into the other donor	369
EC 1.14.14 With reduced flavin or flavoprotein as one donor, and incorporation of one atom of oxygen into the other donor	408
EC 1.14.15 With reduced iron-sulfur protein as one donor, and incorporation of one atom of oxygen into the other donor	462
EC 1.14.16 With reduced pteridine as one donor, and incorporation of one atom of oxygen into the other donor	473
EC 1.14.17 With reduced ascorbate as one donor, and incorporation of one atom of oxygen into the other donor	475
EC 1.14.18 With another compound as one donor, and incorporation of one atom of oxygen into the other donor	476
EC 1.14.19 With oxidation of a pair of donors resulting in the reduction of O ₂ to two molecules of water	481
EC 1.14.20 With 2-oxoglutarate as one donor, and the other dehydrogenated	504
EC 1.14.21 With NADH or NADPH as one donor, and the other dehydrogenated	508
EC 1.14.99 Miscellaneous	509
EC 1.15 Acting on superoxide as acceptor	521
EC 1.15.1 Acting on superoxide as acceptor (only sub-subclass identified to date)	521
EC 1.16 Oxidizing metal ions	521
EC 1.16.1 With NAD ⁺ or NADP ⁺ as acceptor	522
EC 1.16.3 With oxygen as acceptor	524
EC 1.16.5 With a quinone or similar compound as acceptor	525
EC 1.16.8 With a flavin as acceptor	525
EC 1.16.9 With a copper protein as acceptor	525
EC 1.16.98 With other, known, physiological acceptors	526
EC 1.16.99 With unknown physiological acceptors	526

EC 1.17 Acting on CH or CH₂ groups	526
EC 1.17.1 With NAD ⁺ or NADP ⁺ as acceptor	526
EC 1.17.2 With a cytochrome as acceptor	529
EC 1.17.3 With oxygen as acceptor	530
EC 1.17.4 With a disulfide as acceptor	531
EC 1.17.5 With a quinone or similar compound as acceptor	532
EC 1.17.7 With an iron-sulfur protein as acceptor	533
EC 1.17.8 With a flavin as acceptor	535
EC 1.17.9 With a copper protein as acceptor	535
EC 1.17.98 With other, known, physiological acceptors	536
EC 1.17.99 With unknown physiological acceptors	537
EC 1.18 Acting on iron-sulfur proteins as donors	539
EC 1.18.1 With NAD ⁺ or NADP ⁺ as acceptor	539
EC 1.18.6 With dinitrogen as acceptor	542
EC 1.19 Acting on reduced flavodoxin as donor	543
EC 1.19.1 With NAD ⁺ or NADP ⁺ as acceptor	543
EC 1.19.6 With dinitrogen as acceptor	543
EC 1.20 Acting on phosphorus or arsenic in donors	543
EC 1.20.1 With NAD ⁺ or NADP ⁺ as acceptor	543
EC 1.20.2 With a cytochrome as acceptor	544
EC 1.20.4 With disulfide as acceptor	544
EC 1.20.9 With a copper protein as acceptor	545
EC 1.20.98 With other, known, physiological acceptors	545
EC 1.20.99 With unknown physiological acceptors	546
EC 1.21 Catalysing the reaction X-H + Y-H = X-Y	546
EC 1.21.1 With NAD ⁺ or NADP ⁺ as acceptor	546
EC 1.21.3 With oxygen as acceptor	546
EC 1.21.4 With a disulfide as acceptor	549
EC 1.21.98 With other, known, physiological acceptors	550
EC 1.21.99 With unknown physiological acceptors	552
EC 1.22 Acting on halogen in donors	553
EC 1.22.1 With NAD ⁺ or NADP ⁺ as acceptor	553
EC 1.23 Reducing C-O-C group as acceptor	554
EC 1.23.1 With NADH or NADPH as donor	554
EC 1.23.5 With a quinone or similar compound as acceptor	555
EC 1.97 Other oxidoreductases	555
EC 1.97.1 Sole sub-subclass for oxidoreductases that do not belong in the other subclasses	555
EC 1.99.1 Hydroxylases (now covered by EC 1.14)	557
EC 1.99.2 Oxygenases (now covered by EC 1.13)	558
References	559
Index	815

EC 1.1 Acting on the CH-OH group of donors

This subclass contains dehydrogenases that act on primary alcohols, secondary alcohols and hemi-acetals. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.1.1), a cytochrome (EC 1.1.2), oxygen (EC 1.1.3), a disulfide (EC 1.1.4), a quinone or similar compound (EC 1.1.5), or some other acceptor (EC 1.1.99).

EC 1.1.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.1.1.1

- Accepted name:** alcohol dehydrogenase
Reaction: (1) a primary alcohol + NAD⁺ = an aldehyde + NADH + H⁺
(2) a secondary alcohol + NAD⁺ = a ketone + NADH + H⁺
Other name(s): aldehyde reductase; ADH; alcohol dehydrogenase (NAD); aliphatic alcohol dehydrogenase; ethanol dehydrogenase; NAD-dependent alcohol dehydrogenase; NAD-specific aromatic alcohol dehydrogenase; NADH-alcohol dehydrogenase; NADH-aldehyde dehydrogenase; primary alcohol dehydrogenase; yeast alcohol dehydrogenase
Systematic name: alcohol:NAD⁺ oxidoreductase
Comments: A zinc protein. Acts on primary or secondary alcohols or hemi-acetals with very broad specificity; however the enzyme oxidizes methanol much more poorly than ethanol. The animal, but not the yeast, enzyme acts also on cyclic secondary alcohols.
References: [424, 1953, 3035, 4121, 4257]

[EC 1.1.1.1 created 1961, modified 2011]

EC 1.1.1.2

- Accepted name:** alcohol dehydrogenase (NADP⁺)
Reaction: an alcohol + NADP⁺ = an aldehyde + NADPH + H⁺
Other name(s): aldehyde reductase (NADPH₂); NADP-alcohol dehydrogenase; NADP⁺-aldehyde reductase; NADP⁺-dependent aldehyde reductase; NADPH-aldehyde reductase; NADPH-dependent aldehyde reductase; nonspecific succinic semialdehyde reductase; ALR 1; low-*K_m* aldehyde reductase; high-*K_m* aldehyde reductase; alcohol dehydrogenase (NADP)
Systematic name: alcohol:NADP⁺ oxidoreductase
Comments: A zinc protein. Some members of this group oxidize only primary alcohols; others act also on secondary alcohols. May be identical with EC 1.1.1.19 (L-glucuronate reductase), EC 1.1.1.33 [mevaldate reductase (NADPH)] and EC 1.1.1.55 [lactaldehyde reductase (NADPH)]. *Re*-specific with respect to NADPH.
References: [400, 875, 3484, 4157]

[EC 1.1.1.2 created 1961]

EC 1.1.1.3

- Accepted name:** homoserine dehydrogenase
Reaction: L-homoserine + NAD(P)⁺ = L-aspartate 4-semialdehyde + NAD(P)H + H⁺
Other name(s): HSDH; HSD
Systematic name: L-homoserine:NAD(P)⁺ oxidoreductase
Comments: The yeast enzyme acts most rapidly with NAD⁺; the *Neurospora* enzyme with NADP⁺. The enzyme from *Escherichia coli* is a multi-functional protein, which also catalyses the reaction of EC 2.7.2.4 (aspartate kinase).
References: [346, 4007, 4440]

[EC 1.1.1.3 created 1961, modified 1976]

EC 1.1.1.4

- Accepted name:** (*R,R*)-butanediol dehydrogenase
Reaction: (*R,R*)-butane-2,3-diol + NAD⁺ = (*R*)-acetoin + NADH + H⁺

Other name(s): butyleneglycol dehydrogenase; D-butanediol dehydrogenase; D-(–)-butanediol dehydrogenase; butylene glycol dehydrogenase; diacetyl (acetoin) reductase; D-aminopropanol dehydrogenase; 1-amino-2-propanol dehydrogenase; 2,3-butanediol dehydrogenase; D-1-amino-2-propanol dehydrogenase; (R)-diacetyl reductase; (R)-2,3-butanediol dehydrogenase; D-1-amino-2-propanol:NAD⁺ oxidoreductase; 1-amino-2-propanol oxidoreductase; aminopropanol oxidoreductase
Systematic name: (R,R)-butane-2,3-diol:NAD⁺ oxidoreductase
Comments: Also converts diacetyl into acetoin with NADH as reductant.
References: [4059, 4228]

[EC 1.1.1.4 created 1961 (EC 1.1.1.74 created 1972, incorporated 1976)]

[1.1.1.5 *Transferred entry. acetoin dehydrogenase. Now EC 1.1.1.303, diacetyl reductase [(R)-acetoin forming] and EC 1.1.1.304, diacetyl reductase [(S)-acetoin forming]]*

[EC 1.1.1.5 created 1961, modified 1976, deleted 2010]

EC 1.1.1.6

Accepted name: glycerol dehydrogenase
Reaction: glycerol + NAD⁺ = glycerone + NADH + H⁺
Other name(s): glycerin dehydrogenase; NAD-linked glycerol dehydrogenase
Systematic name: glycerol:NAD⁺ 2-oxidoreductase
Comments: Also acts on propane-1,2-diol.
References: [148, 506, 2488]

[EC 1.1.1.6 created 1961]

EC 1.1.1.7

Accepted name: propanediol-phosphate dehydrogenase
Reaction: propane-1,2-diol 1-phosphate + NAD⁺ = hydroxyacetone phosphate + NADH + H⁺
Other name(s): PDP dehydrogenase; 1,2-propanediol-1-phosphate:NAD⁺ oxidoreductase; propanediol phosphate dehydrogenase
Systematic name: propane-1,2-diol-1-phosphate:NAD⁺ oxidoreductase
References: [3796]

[EC 1.1.1.7 created 1961]

EC 1.1.1.8

Accepted name: glycerol-3-phosphate dehydrogenase (NAD⁺)
Reaction: *sn*-glycerol 3-phosphate + NAD⁺ = glycerone phosphate + NADH + H⁺
Other name(s): α-glycerol phosphate dehydrogenase (NAD⁺); α-glycerophosphate dehydrogenase (NAD⁺); glycerol 1-phosphate dehydrogenase; glycerol phosphate dehydrogenase (NAD⁺); glycerophosphate dehydrogenase (NAD⁺); hydroglycerophosphate dehydrogenase; L-α-glycerol phosphate dehydrogenase; L-α-glycerophosphate dehydrogenase; L-glycerol phosphate dehydrogenase; L-glycerophosphate dehydrogenase (ambiguous); NAD⁺-α-glycerophosphate dehydrogenase; NAD⁺-dependent glycerol phosphate dehydrogenase; NAD⁺-dependent glycerol-3-phosphate dehydrogenase; NAD⁺-L-glycerol-3-phosphate dehydrogenase; NAD⁺-linked glycerol 3-phosphate dehydrogenase; NADH-dihydroxyacetone phosphate reductase; glycerol-3-phosphate dehydrogenase (NAD⁺); L-glycerol-3-phosphate dehydrogenase (ambiguous)
Systematic name: *sn*-glycerol-3-phosphate:NAD⁺ 2-oxidoreductase
Comments: Also acts on propane-1,2-diol phosphate and glycerone sulfate (but with a much lower affinity).
References: [214, 451, 3127, 4546, 60, 2188]

[EC 1.1.1.8 created 1961, modified 2005]

EC 1.1.1.9

Accepted name: D-xylulose reductase
Reaction: xylitol + NAD⁺ = D-xylulose + NADH + H⁺
Other name(s): NAD⁺-dependent xylitol dehydrogenase; xylitol dehydrogenase (ambiguous); erythritol dehydrogenase; 2,3-*cis*-polyol(DPN) dehydrogenase (C3-5); pentitol-DPN dehydrogenase (ambiguous); xylitol-2-dehydrogenase
Systematic name: xylitol:NAD⁺ 2-oxidoreductase (D-xylulose-forming)
Comments: Also acts as an L-erythrulose reductase.
References: [649, 1639, 1882]

[EC 1.1.1.9 created 1961]

EC 1.1.1.10

Accepted name: L-xylulose reductase
Reaction: xylitol + NADP⁺ = L-xylulose + NADPH + H⁺
Other name(s): xylitol dehydrogenase (ambiguous)
Systematic name: xylitol:NADP⁺ 4-oxidoreductase (L-xylulose-forming)
References: [955, 1639, 1697, 4316]

[EC 1.1.1.10 created 1961]

EC 1.1.1.11

Accepted name: D-arabinitol 4-dehydrogenase
Reaction: D-arabinitol + NAD⁺ = D-xylulose + NADH + H⁺
Other name(s): D-arabitol dehydrogenase; arabitol dehydrogenase
Systematic name: D-arabinitol:NAD⁺ 4-oxidoreductase
References: [2487, 4666]

[EC 1.1.1.11 created 1961]

EC 1.1.1.12

Accepted name: L-arabinitol 4-dehydrogenase
Reaction: L-arabinitol + NAD⁺ = L-xylulose + NADH + H⁺
Other name(s): pentitol-DPN dehydrogenase (ambiguous); L-arabitol dehydrogenase
Systematic name: L-arabinitol:NAD⁺ 4-oxidoreductase (L-xylulose-forming)
References: [649, 650]

[EC 1.1.1.12 created 1961]

EC 1.1.1.13

Accepted name: L-arabinitol 2-dehydrogenase
Reaction: L-arabinitol + NAD⁺ = L-ribulose + NADH + H⁺
Other name(s): L-arabinitol dehydrogenase (ribulose-forming); L-arabinitol (ribulose-forming) dehydrogenase
Systematic name: L-arabinitol:NAD⁺ 2-oxidoreductase (L-ribulose-forming)
References: [650]

[EC 1.1.1.13 created 1961]

EC 1.1.1.14

Accepted name: L-iditol 2-dehydrogenase
Reaction: L-iditol + NAD⁺ = L-sorbose + NADH + H⁺

Other name(s): polyol dehydrogenase; sorbitol dehydrogenase; L-idoitol:NAD⁺ 5-oxidoreductase; L-idoitol (sorbitol) dehydrogenase; glucitol dehydrogenase; L-idoitol:NAD⁺ oxidoreductase; NAD⁺-dependent sorbitol dehydrogenase; NAD⁺-sorbitol dehydrogenase
Systematic name: L-idoitol:NAD⁺ 2-oxidoreductase
Comments: This enzyme is widely distributed and has been described in archaea, bacteria, yeast, plants and animals. It acts on a number of sugar alcohols, including (but not limited to) L-idoitol, D-glucitol, D-xylitol, and D-galactitol. Enzymes from different organisms or tissues display different substrate specificity. The enzyme is specific to NAD⁺ and can not use NADP⁺.
References: [183, 499, 2413, 3036, 3126, 3056]

[EC 1.1.1.14 created 1961, modified 2011]

EC 1.1.1.15

Accepted name: D-idoitol 2-dehydrogenase
Reaction: D-idoitol + NAD⁺ = D-sorbose + NADH + H⁺
Other name(s): D-sorbitol dehydrogenase
Systematic name: D-idoitol:NAD⁺ 2-oxidoreductase
Comments: Also converts xylitol into L-xylulose and L-glucitol into L-fructose.
References: [3832]

[EC 1.1.1.15 created 1961]

EC 1.1.1.16

Accepted name: galactitol 2-dehydrogenase
Reaction: galactitol + NAD⁺ = D-tagatose + NADH + H⁺
Other name(s): dulcitol dehydrogenase; AtuSorbD (gene name); galactitol:NAD⁺ 2-oxidoreductase
Systematic name: galactitol:NAD⁺ 2-oxidoreductase (D-tagatose-forming)
Comments: Also converts other alditols containing an L-*threo*-configuration adjacent to a primary alcohol group into the corresponding sugars. The enzyme from *Agrobacterium fabrum* C58 is part of D-altritol and galactitol degradation pathways.
References: [3832, 4615]

[EC 1.1.1.16 created 1961]

EC 1.1.1.17

Accepted name: mannitol-1-phosphate 5-dehydrogenase
Reaction: D-mannitol 1-phosphate + NAD⁺ = D-fructose 6-phosphate + NADH + H⁺
Other name(s): hexose reductase; mannitol 1-phosphate dehydrogenase; D-mannitol-1-phosphate dehydrogenase; fructose 6-phosphate reductase
Systematic name: D-mannitol-1-phosphate:NAD⁺ 5-oxidoreductase
References: [2656, 4653, 4654]

[EC 1.1.1.17 created 1961]

EC 1.1.1.18

Accepted name: inositol 2-dehydrogenase
Reaction: *myo*-inositol + NAD⁺ = 2,4,6/3,5-pentahydroxycyclohexanone + NADH + H⁺
Other name(s): *myo*-inositol 2-dehydrogenase; *myo*-inositol:NAD⁺ oxidoreductase; inositol dehydrogenase; *myo*-inositol dehydrogenase
Systematic name: *myo*-inositol:NAD⁺ 2-oxidoreductase
References: [302, 2355, 4448]

[EC 1.1.1.18 created 1961]

EC 1.1.1.19

Accepted name: glucuronate reductase
Reaction: L-gulonate + NADP⁺ = D-glucuronate + NADPH + H⁺
Other name(s): L-hexonate:NADP dehydrogenase; TPN-L-gulonate dehydrogenase; NADP-L-gulonate dehydrogenase; D-glucuronate dehydrogenase; D-glucuronate reductase; L-glucuronate reductase (incorrect)
Systematic name: L-gulonate:NADP⁺ 6-oxidoreductase
Comments: Also reduces D-galacturonate. May be identical with EC 1.1.1.2 [alcohol dehydrogenase (NADP⁺)].
References: [3919, 4470, 4804]

[EC 1.1.1.19 created 1961]

EC 1.1.1.20

Accepted name: glucuronolactone reductase
Reaction: L-gulono-1,4-lactone + NADP⁺ = D-glucurono-3,6-lactone + NADPH + H⁺
Other name(s): GRase; gulonolactone dehydrogenase
Systematic name: L-gulono-1,4-lactone:NADP⁺ 1-oxidoreductase
References: [4138]

[EC 1.1.1.20 created 1961]

EC 1.1.1.21

Accepted name: aldose reductase
Reaction: alditol + NAD(P)⁺ = aldose + NAD(P)H + H⁺
Other name(s): polyol dehydrogenase (NADP⁺); ALR2; alditol:NADP⁺ oxidoreductase; alditol:NADP⁺ 1-oxidoreductase; NADPH-aldopentose reductase; NADPH-aldose reductase; aldehyde reductase (misleading)
Systematic name: alditol:NAD(P)⁺ 1-oxidoreductase
Comments: Has wide specificity.
References: [152, 372, 1630, 3712]

[EC 1.1.1.21 created 1961 (EC 1.1.1.139 created 1972, incorporated 1978), modified 2019]

EC 1.1.1.22

Accepted name: UDP-glucose 6-dehydrogenase
Reaction: UDP- α -D-glucose + 2 NAD⁺ + H₂O = UDP- α -D-glucuronate + 2 NADH + 2 H⁺
Other name(s): UDP-glucose dehydrogenase; uridine diphosphoglucose dehydrogenase; UDPG dehydrogenase; UDPG:NAD oxidoreductase; UDP- α -D-glucose:NAD oxidoreductase; UDP-glucose:NAD⁺ oxidoreductase; uridine diphosphate glucose dehydrogenase; UDP-D-glucose dehydrogenase; uridine diphosphate D-glucose dehydrogenase
Systematic name: UDP- α -D-glucose:NAD⁺ 6-oxidoreductase
Comments: Also acts on UDP- α -D-2-deoxyglucose.
References: [970, 2728, 4072, 4073]

[EC 1.1.1.22 created 1961]

EC 1.1.1.23

Accepted name: histidinol dehydrogenase
Reaction: L-histidinol + 2 NAD⁺ + H₂O = L-histidine + 2 NADH + 3 H⁺
Other name(s): L-histidinol dehydrogenase
Systematic name: L-histidinol:NAD⁺ oxidoreductase
Comments: Also oxidizes L-histidinal. The *Neurospora* enzyme also catalyses the reactions of EC 3.5.4.19 (phosphoribosyl-AMP cyclohydrolase) and EC 3.6.1.31 (phosphoribosyl-ATP diphosphatase).
References: [21, 22, 2538, 4829]

[EC 1.1.1.23 created 1961]

EC 1.1.1.24

Accepted name: quinate/shikimate dehydrogenase (NAD⁺)
Reaction: L-quinatate + NAD⁺ = 3-dehydroquinatate + NADH + H⁺
Other name(s): quinate dehydrogenase (ambiguous); quinic dehydrogenase (ambiguous); quinate:NAD oxidoreductase; quinate 5-dehydrogenase (ambiguous); quinate:NAD⁺ 5-oxidoreductase
Systematic name: L-quinatate:NAD⁺ 3-oxidoreductase
Comments: The enzyme, found mostly in bacteria (mostly, but not exclusively in Gram-positive bacteria), fungi, and plants, participates in the degradation of quinate and shikimate with a strong preference for NAD⁺ as a cofactor. While the enzyme can act on both quinate and shikimate, activity is higher with the former. *cf.* EC 1.1.5.8, quinate/shikimate dehydrogenase (quinone), EC 1.1.1.282, quinate/shikimate dehydrogenase [NAD(P)⁺], and EC 1.1.1.25, shikimate dehydrogenase (NADP⁺).
References: [2833, 1266, 1569, 3910, 4243, 2272, 3280]

[EC 1.1.1.24 created 1961, modified 1976, modified 2004, modified 2021]

EC 1.1.1.25

Accepted name: shikimate dehydrogenase (NADP⁺)
Reaction: shikimate + NADP⁺ = 3-dehydroshikimate + NADPH + H⁺
Other name(s): shikimate dehydrogenase; dehydroshikimic reductase; shikimate oxidoreductase; shikimate:NADP⁺ oxidoreductase; 5-dehydroshikimate reductase; shikimate 5-dehydrogenase; 5-dehydroshikimic reductase; DHS reductase; shikimate:NADP⁺ 5-oxidoreductase; AroE
Systematic name: shikimate:NADP⁺ 3-oxidoreductase
Comments: NAD⁺ cannot replace NADP⁺ [200]. In higher organisms, this enzyme forms part of a multienzyme complex with EC 4.2.1.10, 3-dehydroquinatate dehydratase [617]. *cf.* EC 1.1.1.24, quinate/shikimate dehydrogenase (NAD⁺), EC 1.1.5.8, quinate/shikimate dehydrogenase (quinone), and EC 1.1.1.282, quinate/shikimate dehydrogenase [NAD(P)⁺].
References: [2833, 4776, 200, 617, 110, 4783]

[EC 1.1.1.25 created 1961, modified 1976, modified 2004, modified 2021]

EC 1.1.1.26

Accepted name: glyoxylate reductase
Reaction: glycolate + NAD⁺ = glyoxylate + NADH + H⁺
Other name(s): NADH-glyoxylate reductase; glyoxylic acid reductase; NADH-dependent glyoxylate reductase
Systematic name: glycolate:NAD⁺ oxidoreductase
Comments: Reduces glyoxylate to glycolate or hydroxypyruvate to D-glycerate.
References: [4866, 4867]

[EC 1.1.1.26 created 1961]

EC 1.1.1.27

Accepted name: L-lactate dehydrogenase
Reaction: (S)-lactate + NAD⁺ = pyruvate + NADH + H⁺
Other name(s): lactic acid dehydrogenase; L(+)-nLDH; L-(+)-lactate dehydrogenase; L-lactic dehydrogenase; L-lactic acid dehydrogenase; lactate dehydrogenase; lactate dehydrogenase NAD-dependent; lactic dehydrogenase; NAD-lactate dehydrogenase
Systematic name: (S)-lactate:NAD⁺ oxidoreductase
Comments: Also oxidizes other (S)-2-hydroxymonocarboxylic acids. NADP⁺ also acts, more slowly, with the animal, but not the bacterial, enzyme.
References: [882, 1072, 1689, 3702]

[EC 1.1.1.27 created 1961]

EC 1.1.1.28

Accepted name: D-lactate dehydrogenase
Reaction: (*R*)-lactate + NAD⁺ = pyruvate + NADH + H⁺
Other name(s): lactic acid dehydrogenase; lactic acid dehydrogenase; D-specific lactic dehydrogenase; D-(-)-lactate dehydrogenase (NAD); D-lactic acid dehydrogenase; D-lactic dehydrogenase
Systematic name: (*R*)-lactate:NAD⁺ oxidoreductase
References: [882]

[EC 1.1.1.28 created 1961]

EC 1.1.1.29

Accepted name: glycerate dehydrogenase
Reaction: D-glycerate + NAD⁺ = hydroxypyruvate + NADH + H⁺
Other name(s): D-glycerate dehydrogenase; hydroxypyruvate reductase; (*R*)-glycerate:NAD⁺ oxidoreductase
Systematic name: D-glycerate:NAD⁺ oxidoreductase
References: [1704, 4000]

[EC 1.1.1.29 created 1961]

EC 1.1.1.30

Accepted name: 3-hydroxybutyrate dehydrogenase
Reaction: (*R*)-3-hydroxybutanoate + NAD⁺ = acetoacetate + NADH + H⁺
Other name(s): NAD-β-hydroxybutyrate dehydrogenase; hydroxybutyrate oxidoreductase; β-hydroxybutyrate dehydrogenase; D-β-hydroxybutyrate dehydrogenase; D-3-hydroxybutyrate dehydrogenase; D-(-)-3-hydroxybutyrate dehydrogenase; β-hydroxybutyric acid dehydrogenase; 3-D-hydroxybutyrate dehydrogenase; β-hydroxybutyric dehydrogenase
Systematic name: (*R*)-3-hydroxybutanoate:NAD⁺ oxidoreductase
Comments: Also oxidizes other 3-hydroxymonocarboxylic acids.
References: [299, 871, 2407]

[EC 1.1.1.30 created 1961]

EC 1.1.1.31

Accepted name: 3-hydroxyisobutyrate dehydrogenase
Reaction: 3-hydroxy-2-methylpropanoate + NAD⁺ = 2-methyl-3-oxopropanoate + NADH + H⁺
Other name(s): β-hydroxyisobutyrate dehydrogenase
Systematic name: 3-hydroxy-2-methylpropanoate:NAD⁺ oxidoreductase
References: [3540]

[EC 1.1.1.31 created 1961]

EC 1.1.1.32

Accepted name: mevaldate reductase
Reaction: (*R*)-mevalonate + NAD⁺ = mevaldate + NADH + H⁺
Other name(s): mevalonic dehydrogenase
Systematic name: (*R*)-mevalonate:NAD⁺ oxidoreductase
References: [3723]

[EC 1.1.1.32 created 1961]

EC 1.1.1.33

Accepted name: mevaldate reductase (NADPH)

Reaction: (*R*)-mevalonate + NADP⁺ = mevaldate + NADPH + H⁺
Other name(s): mevaldate (reduced nicotinamide adenine dinucleotide phosphate) reductase; mevaldate reductase (NADPH₂)
Systematic name: (*R*)-mevalonate:NADP⁺ oxidoreductase
Comments: May be identical with EC 1.1.1.2 [alcohol dehydrogenase (NADP⁺)].
References: [726, 4470]

[EC 1.1.1.33 created 1961]

EC 1.1.1.34

Accepted name: hydroxymethylglutaryl-CoA reductase (NADPH)
Reaction: (*R*)-mevalonate + CoA + 2 NADP⁺ = (*S*)-3-hydroxy-3-methylglutaryl-CoA + 2 NADPH + 2 H⁺
Other name(s): hydroxymethylglutaryl coenzyme A reductase (reduced nicotinamide adenine dinucleotide phosphate); 3-hydroxy-3-methylglutaryl-CoA reductase (ambiguous); β-hydroxy-β-methylglutaryl coenzyme A reductase (ambiguous); hydroxymethylglutaryl CoA reductase (NADPH); *S*-3-hydroxy-3-methylglutaryl-CoA reductase (ambiguous); NADPH-hydroxymethylglutaryl-CoA reductase; HMG-CoA reductase-mevalonate:NADP-oxidoreductase (acetylating-CoA); 3-hydroxy-3-methylglutaryl CoA reductase (NADPH); hydroxymethylglutaryl-CoA reductase (NADPH₂)
Systematic name: (*R*)-mevalonate:NADP⁺ oxidoreductase (CoA-acylating)
Comments: The enzyme is inactivated by EC 2.7.11.31 [hydroxymethylglutaryl-CoA reductase (NADPH)] kinase and reactivated by EC 3.1.3.47 [hydroxymethylglutaryl-CoA reductase (NADPH)]-phosphatase.
References: [486, 990, 2040]

[EC 1.1.1.34 created 1961]

EC 1.1.1.35

Accepted name: 3-hydroxyacyl-CoA dehydrogenase
Reaction: (*S*)-3-hydroxyacyl-CoA + NAD⁺ = 3-oxoacyl-CoA + NADH + H⁺
Other name(s): β-hydroxyacyl dehydrogenase; β-keto-reductase; 3-keto reductase; 3-hydroxyacyl coenzyme A dehydrogenase; β-hydroxyacyl-coenzyme A synthetase; β-hydroxyacylcoenzyme A dehydrogenase; β-hydroxybutyrylcoenzyme A dehydrogenase; 3-hydroxyacetyl-coenzyme A dehydrogenase; *L*-3-hydroxyacyl coenzyme A dehydrogenase; *L*-3-hydroxyacyl CoA dehydrogenase; β-hydroxyacyl CoA dehydrogenase; 3β-hydroxyacyl coenzyme A dehydrogenase; 3-hydroxybutyryl-CoA dehydrogenase; β-ketoacyl-CoA reductase; β-hydroxy acid dehydrogenase; 3-*L*-hydroxyacyl-CoA dehydrogenase; 3-hydroxyisobutyryl-CoA dehydrogenase; 1-specific DPN-linked β-hydroxybutyric dehydrogenase
Systematic name: (*S*)-3-hydroxyacyl-CoA:NAD⁺ oxidoreductase
Comments: Also oxidizes *S*-3-hydroxyacyl-*N*-acylthioethanolamine and *S*-3-hydroxyacyl-hydrolipoate. Some enzymes act, more slowly, with NADP⁺. Broad specificity to acyl chain-length (*cf.* EC 1.1.1.211 [long-chain-3-hydroxyacyl-CoA dehydrogenase]).
References: [1656, 2406, 4024, 4492]

[EC 1.1.1.35 created 1961]

EC 1.1.1.36

Accepted name: acetoacetyl-CoA reductase
Reaction: (*R*)-3-hydroxyacyl-CoA + NADP⁺ = 3-oxoacyl-CoA + NADPH + H⁺
Other name(s): acetoacetyl coenzyme A reductase; hydroxyacyl coenzyme-A dehydrogenase; NADP-linked acetoacetyl CoA reductase; NADPH:acetoacetyl-CoA reductase; D(-)-β-hydroxybutyryl CoA-NADP oxidoreductase; short chain β-ketoacetyl(acetoacetyl)-CoA reductase; β-ketoacyl-CoA reductase; D-3-hydroxyacyl-CoA reductase; (*R*)-3-hydroxyacyl-CoA dehydrogenase
Systematic name: (*R*)-3-hydroxyacyl-CoA:NADP⁺ oxidoreductase
References: [4491]

[EC 1.1.1.36 created 1961]

EC 1.1.1.37

- Accepted name:** malate dehydrogenase
Reaction: (S)-malate + NAD⁺ = oxaloacetate + NADH + H⁺
Other name(s): malic dehydrogenase; L-malate dehydrogenase; NAD-L-malate dehydrogenase; malic acid dehydrogenase; NAD-dependent malic dehydrogenase; NAD-malate dehydrogenase; NAD-malic dehydrogenase; malate (NAD) dehydrogenase; NAD-dependent malate dehydrogenase; NAD-specific malate dehydrogenase; NAD-linked malate dehydrogenase; MDH (ambiguous); L-malate-NAD⁺ oxidoreductase
Systematic name: (S)-malate:NAD⁺ oxidoreductase
Comments: There are several forms of malate dehydrogenases that differ by their use of substrate and cofactors. This NAD⁺-dependent enzyme forms oxaloacetate and unlike EC 1.1.1.38, malate dehydrogenase (oxaloacetate-decarboxylating), is unable to convert it to pyruvate. Also oxidizes some other 2-hydroxydicarboxylic acids. *cf.* EC 1.1.1.82, malate dehydrogenase (NADP⁺); EC 1.1.1.299, malate dehydrogenase [NAD(P)⁺]; and EC 1.1.5.4, malate dehydrogenase (quinone).
References: [207, 1439, 2756, 4655]

[EC 1.1.1.37 created 1961]

EC 1.1.1.38

- Accepted name:** malate dehydrogenase (oxaloacetate-decarboxylating)
Reaction: (1) (S)-malate + NAD⁺ = pyruvate + CO₂ + NADH
(2) oxaloacetate = pyruvate + CO₂
Other name(s): 'malic' enzyme (ambiguous); pyruvic-malic carboxylase (ambiguous); NAD⁺-specific malic enzyme; NAD⁺-malic enzyme; NAD⁺-linked malic enzyme
Systematic name: (S)-malate:NAD⁺ oxidoreductase (oxaloacetate-decarboxylating)
Comments: Unlike EC 1.1.1.39, malate dehydrogenase (decarboxylating), this enzyme can also decarboxylate oxaloacetate. *cf.* EC 1.1.1.40, malate dehydrogenase (oxaloacetate-decarboxylating) (NADP⁺).
References: [2037, 4729]

[EC 1.1.1.38 created 1961]

EC 1.1.1.39

- Accepted name:** malate dehydrogenase (decarboxylating)
Reaction: (S)-malate + NAD⁺ = pyruvate + CO₂ + NADH
Other name(s): 'malic' enzyme (ambiguous); pyruvic-malic carboxylase (ambiguous); NAD-specific malic enzyme (ambiguous); NAD-malic enzyme (ambiguous); malate dehydrogenase (decarboxylating) (ambiguous)
Systematic name: (S)-malate:NAD⁺ oxidoreductase (decarboxylating)
Comments: There are several forms of malate dehydrogenases that differ in their use of substrates and cofactors. This particular form is found only in the plant kingdom. Unlike EC 1.1.1.38, which catalyses a similar reaction, this enzyme can not bind oxaloacetate, and thus does not decarboxylate exogenously-added oxaloacetate. *cf.* EC 1.1.1.37, malate dehydrogenase; EC 1.1.1.38, malate dehydrogenase (oxaloacetate-decarboxylating); and EC 1.1.1.83, D-malate dehydrogenase (decarboxylating).
References: [2596, 1432, 4568, 4567]

[EC 1.1.1.39 created 1961]

EC 1.1.1.40

- Accepted name:** malate dehydrogenase (oxaloacetate-decarboxylating) (NADP⁺)
Reaction: (1) (S)-malate + NADP⁺ = pyruvate + CO₂ + NADPH
(2) oxaloacetate = pyruvate + CO₂

Other name(s): 'malic' enzyme (ambiguous); pyruvic-malic carboxylase (ambiguous); malate dehydrogenase (decarboxylating, NADP⁺); NADP⁺-linked decarboxylating malic enzyme; NADP⁺-malic enzyme; NADP⁺-specific malic enzyme; NADP⁺-specific malate dehydrogenase; malate dehydrogenase (NADP⁺, decarboxylating); L-malate:NADP⁺ oxidoreductase

Systematic name: (S)-malate:NADP⁺ oxidoreductase (oxaloacetate-decarboxylating)

Comments: The enzyme catalyses the oxidative decarboxylation of (S)-malate in the presence of NADP⁺ and divalent metal ions, and the decarboxylation of oxaloacetate. *cf.* EC 1.1.1.38, malate dehydrogenase (oxaloacetate-decarboxylating), and EC 1.1.1.39, malate dehydrogenase (decarboxylating).

References: [1529, 3130, 3609, 4028, 4029, 4496]

[EC 1.1.1.40 created 1961, modified 1976]

EC 1.1.1.41

Accepted name: isocitrate dehydrogenase (NAD⁺)

Reaction: isocitrate + NAD⁺ = 2-oxoglutarate + CO₂ + NADH

Other name(s): isocitric dehydrogenase; β-ketoglutaric-isocitric carboxylase; isocitric acid dehydrogenase; NAD dependent isocitrate dehydrogenase; NAD isocitrate dehydrogenase; NAD-linked isocitrate dehydrogenase; NAD-specific isocitrate dehydrogenase; NAD isocitric dehydrogenase; isocitrate dehydrogenase (NAD); IDH (ambiguous); nicotinamide adenine dinucleotide isocitrate dehydrogenase

Systematic name: isocitrate:NAD⁺ oxidoreductase (decarboxylating)

Comments: Requires Mn²⁺ or Mg²⁺ for activity. Unlike EC 1.1.1.42, isocitrate dehydrogenase (NADP⁺), oxalosuccinate cannot be used as a substrate. In eukaryotes, isocitrate dehydrogenase exists in two forms: an NAD⁺-linked enzyme found only in mitochondria and displaying allosteric properties, and a non-allosteric, NADP⁺-linked enzyme that is found in both mitochondria and cytoplasm [532]. The enzyme from some species can also use NADP⁺ but much more slowly [1813].

References: [1560, 2235, 3335, 3336, 3442, 4446, 532, 2114, 1813]

[EC 1.1.1.41 created 1961, modified 2005]

EC 1.1.1.42

Accepted name: isocitrate dehydrogenase (NADP⁺)

Reaction: isocitrate + NADP⁺ = 2-oxoglutarate + CO₂ + NADPH + H⁺ (overall reaction)
(1a) isocitrate + NADP⁺ = oxalosuccinate + NADPH + H⁺
(1b) oxalosuccinate = 2-oxoglutarate + CO₂

Other name(s): oxalosuccinate decarboxylase; oxalsuccinic decarboxylase; isocitrate (NADP) dehydrogenase; isocitrate (nicotinamide adenine dinucleotide phosphate) dehydrogenase; NADP-specific isocitrate dehydrogenase; NADP-linked isocitrate dehydrogenase; NADP-dependent isocitrate dehydrogenase; NADP isocitric dehydrogenase; isocitrate dehydrogenase (NADP-dependent); NADP-dependent isocitric dehydrogenase; triphosphopyridine nucleotide-linked isocitrate dehydrogenase-oxalosuccinate carboxylase; NADP⁺-linked isocitrate dehydrogenase; IDH (ambiguous); dual-cofactor-specific isocitrate dehydrogenase; NADP⁺-ICDH; NADP⁺-IDH; IDP; IDP1; IDP2; IDP3

Systematic name: isocitrate:NADP⁺ oxidoreductase (decarboxylating)

Comments: Requires Mn²⁺ or Mg²⁺ for activity. Unlike EC 1.1.1.41, isocitrate dehydrogenase (NAD⁺), oxalosuccinate can be used as a substrate. In eukaryotes, isocitrate dehydrogenase exists in two forms: an NAD⁺-linked enzyme found only in mitochondria and displaying allosteric properties, and a non-allosteric, NADP⁺-linked enzyme that is found in both mitochondria and cytoplasm [532]. The enzyme from some species can also use NAD⁺ but much more slowly [532, 4010].

References: [37, 2913, 3335, 3899, 4446, 532, 4010, 2196, 581]

[EC 1.1.1.42 created 1961, modified 2005]

EC 1.1.1.43

Accepted name: phosphogluconate 2-dehydrogenase

Reaction: 6-phospho-D-gluconate + NAD(P)⁺ = 6-phospho-2-dehydro-D-gluconate + NAD(P)H + H⁺
Other name(s): 6-phosphogluconic dehydrogenase; phosphogluconate dehydrogenase; gluconate 6-phosphate dehydrogenase; 6-phosphogluconate dehydrogenase (NAD); 2-keto-6-phosphogluconate reductase
Systematic name: 6-phospho-D-gluconate:NAD(P)⁺ 2-oxidoreductase
References: [1160]

[EC 1.1.1.43 created 1961]

EC 1.1.1.44

Accepted name: phosphogluconate dehydrogenase (NADP⁺-dependent, decarboxylating)
Reaction: 6-phospho-D-gluconate + NADP⁺ = D-ribulose 5-phosphate + CO₂ + NADPH + H⁺
Other name(s): phosphogluconic acid dehydrogenase; 6-phosphogluconic dehydrogenase; 6-phosphogluconic carboxylase; 6-phosphogluconate dehydrogenase (decarboxylating); 6-phospho-D-gluconate dehydrogenase
Systematic name: 6-phospho-D-gluconate:NADP⁺ 2-oxidoreductase (decarboxylating)
Comments: The enzyme participates in the oxidative branch of the pentose phosphate pathway, whose main purpose is to produce NADPH and pentose for biosynthetic reactions. Highly specific for NADP⁺. *cf.* EC 1.1.1.343, phosphogluconate dehydrogenase (NAD⁺-dependent, decarboxylating).
References: [903, 3353, 3772, 3773, 440, 4802, 4857]

[EC 1.1.1.44 created 1961, modified 2013]

EC 1.1.1.45

Accepted name: L-gulonate 3-dehydrogenase
Reaction: L-gulonate + NAD⁺ = 3-dehydro-L-gulonate + NADH + H⁺
Other name(s): L-3-aldonate dehydrogenase; L-3-aldonic dehydrogenase; L-gulonic acid dehydrogenase; L-β-hydroxyacid dehydrogenase; L-β-hydroxy-acid-NAD-oxidoreductase; L-3-hydroxyacid dehydrogenase
Systematic name: L-gulonate:NAD⁺ 3-oxidoreductase
Comments: Also oxidizes other L-3-hydroxyacids.
References: [994, 3932]

[EC 1.1.1.45 created 1961]

EC 1.1.1.46

Accepted name: L-arabinose 1-dehydrogenase
Reaction: L-arabinose + NAD⁺ = L-arabinono-1,4-lactone + NADH + H⁺
Systematic name: L-arabinose:NAD⁺ 1-oxidoreductase
References: [4576]

[EC 1.1.1.46 created 1961]

EC 1.1.1.47

Accepted name: glucose 1-dehydrogenase [NAD(P)⁺]
Reaction: D-glucose + NAD(P)⁺ = D-glucono-1,5-lactone + NAD(P)H + H⁺
Other name(s): D-glucose dehydrogenase (NAD(P)⁺); hexose phosphate dehydrogenase; β-D-glucose:NAD(P)⁺ 1-oxidoreductase; glucose 1-dehydrogenase
Systematic name: D-glucose:NAD(P)⁺ 1-oxidoreductase
Comments: This enzyme has similar activity with either NAD⁺ or NADP⁺. *cf.* EC 1.1.1.118, glucose 1-dehydrogenase (NAD⁺) and EC 1.1.1.119, glucose 1-dehydrogenase (NADP⁺).
References: [208, 441, 3266, 4060, 4275, 1215]

[EC 1.1.1.47 created 1961, modified 2013]

EC 1.1.1.48

- Accepted name:** D-galactose 1-dehydrogenase
Reaction: D-galactose + NAD⁺ = D-galactono-1,4-lactone + NADH + H⁺
Other name(s): D-galactose dehydrogenase; β-galactose dehydrogenase (ambiguous); NAD⁺-dependent D-galactose dehydrogenase
Systematic name: D-galactose:NAD⁺ 1-oxidoreductase
Comments: This enzyme is part of the De Ley-Doudoroff pathway, which is used by some bacteria during growth on D-galactose.
References: [2438, 1746]

[EC 1.1.1.48 created 1961, modified 2011]

EC 1.1.1.49

- Accepted name:** glucose-6-phosphate dehydrogenase (NADP⁺)
Reaction: D-glucose 6-phosphate + NADP⁺ = 6-phospho-D-glucono-1,5-lactone + NADPH + H⁺
Other name(s): NADP-glucose-6-phosphate dehydrogenase; Zwischenferment; D-glucose 6-phosphate dehydrogenase; glucose 6-phosphate dehydrogenase (NADP); NADP-dependent glucose 6-phosphate dehydrogenase; 6-phosphoglucose dehydrogenase; Entner-Doudoroff enzyme; glucose-6-phosphate 1-dehydrogenase; G6PDH; GPD; glucose-6-phosphate dehydrogenase
Systematic name: D-glucose-6-phosphate:NADP⁺ 1-oxidoreductase
Comments: The enzyme catalyses a step of the pentose phosphate pathway. The enzyme is specific for NADP⁺. *cf.* EC 1.1.1.363, glucose-6-phosphate dehydrogenase [NAD(P)⁺] and EC 1.1.1.388, glucose-6-phosphate dehydrogenase (NAD⁺).
References: [1047, 1340, 1959, 3098, 2794, 3166, 1513, 1785, 1864, 669]

[EC 1.1.1.49 created 1961, modified 2013, modified 2015]

EC 1.1.1.50

- Accepted name:** 3α-hydroxysteroid 3-dehydrogenase (*Si*-specific)
Reaction: a 3α-hydroxysteroid + NAD(P)⁺ = a 3-oxosteroid + NAD(P)H + H⁺
Other name(s): hydroxyprostaglandin dehydrogenase; 3α-hydroxysteroid oxidoreductase; sterognost 3α; 3α-hydroxysteroid dehydrogenase (*B*-specific); 3α-hydroxysteroid 3-dehydrogenase (*B*-specific); 3α-hydroxysteroid:NAD(P)⁺ 3-oxidoreductase (*B*-specific)
Systematic name: 3α-hydroxysteroid:NAD(P)⁺ 3-oxidoreductase (*Si*-specific)
Comments: The enzyme acts on androsterone and other 3α-hydroxysteroids and on 9-, 11- and 15-hydroxyprostaglandin. *Si*-specific with respect to NAD⁺ or NADP⁺. *cf.* EC 1.1.1.213, 3α-hydroxysteroid 3-dehydrogenase (*Re*-specific).
References: [1892, 2180, 2647, 3287]

[EC 1.1.1.50 created 1961, modified 1986, modified 1990, modified 2012, modified 2013]

EC 1.1.1.51

- Accepted name:** 3(or 17)β-hydroxysteroid dehydrogenase
Reaction: testosterone + NAD(P)⁺ = androstenedione + NAD(P)H + H⁺
Other name(s): β-hydroxy steroid dehydrogenase; 17-ketoreductase; 17β-hydroxy steroid dehydrogenase; 3β-hydroxysteroid dehydrogenase; 3β-hydroxy steroid dehydrogenase
Systematic name: 3(or 17)β-hydroxysteroid:NAD(P)⁺ oxidoreductase
Comments: Also acts on other 3β- or 17β-hydroxysteroids. *cf.* EC 1.1.1.209 3(or 17)α-hydroxysteroid dehydrogenase.
References: [801, 2571, 2647, 3754, 4190]

[EC 1.1.1.51 created 1961]

EC 1.1.1.52

- Accepted name:** 3 α -hydroxycholanate dehydrogenase (NAD⁺)
Reaction: lithocholate + NAD⁺ = 3-oxo-5 β -cholan-24-oate + NADH + H⁺
Other name(s): α -hydroxy-cholanate dehydrogenase; lithocholate:NAD⁺ oxidoreductase; 3 α -hydroxycholanate dehydrogenase
Systematic name: lithocholate:NAD⁺ 3-oxidoreductase
Comments: Also acts on other 3 α -hydroxysteroids with an acidic side-chain. *cf.* EC 1.1.1.392, 3 α -hydroxycholanate dehydrogenase (NADP⁺).
References: [1574]

[EC 1.1.1.52 created 1961, modified 1976, modified 2016]

EC 1.1.1.53

- Accepted name:** 3 α (or 20 β)-hydroxysteroid dehydrogenase
Reaction: androstan-3 α ,17 β -diol + NAD⁺ = 17 β -hydroxyandrostan-3-one + NADH + H⁺
Other name(s): cortisone reductase; (*R*)-20-hydroxysteroid dehydrogenase; 20 β -hydroxy steroid dehydrogenase; Δ^4 -3-ketosteroid hydrogenase; 20 β -hydroxysteroid dehydrogenase; 3 α ,20 β -hydroxysteroid:NAD⁺-oxidoreductase; NADH-20 β -hydroxysteroid dehydrogenase; 20 β -HSD
Systematic name: 3 α (or 20 β)-hydroxysteroid:NAD⁺ oxidoreductase
Comments: The 3 α -hydroxy group or 20 β -hydroxy group of pregnane and androstane steroids can act as donor.
References: [1021, 1760, 1761, 2571, 4064, 4150]

[EC 1.1.1.53 created 1961, modified 1986]

EC 1.1.1.54

- Accepted name:** allyl-alcohol dehydrogenase
Reaction: allyl alcohol + NADP⁺ = acrolein + NADPH + H⁺
Systematic name: allyl-alcohol:NADP⁺ oxidoreductase
Comments: Also acts on saturated primary alcohols.
References: [3206]

[EC 1.1.1.54 created 1965]

EC 1.1.1.55

- Accepted name:** lactaldehyde reductase (NADPH)
Reaction: propane-1,2-diol + NADP⁺ = L-lactaldehyde + NADPH + H⁺
Other name(s): lactaldehyde (reduced nicotinamide adenine dinucleotide phosphate) reductase; NADP-1,2-propanediol dehydrogenase; propanediol dehydrogenase; 1,2-propanediol:NADP⁺ oxidoreductase; lactaldehyde reductase (NADPH₂)
Systematic name: propane-1,2-diol:NADP⁺ oxidoreductase
Comments: May be identical with EC 1.1.1.2 alcohol dehydrogenase (NADP⁺).
References: [1450]

[EC 1.1.1.55 created 1965]

EC 1.1.1.56

- Accepted name:** ribitol 2-dehydrogenase
Reaction: ribitol + NAD⁺ = D-ribulose + NADH + H⁺
Other name(s): adonitol dehydrogenase; ribitol dehydrogenase A (wild type); ribitol dehydrogenase B (mutant enzyme with different properties); ribitol dehydrogenase D (mutant enzyme with different properties)
Systematic name: ribitol:NAD⁺ 2-oxidoreductase
References: [1697, 3104, 4666]

[EC 1.1.1.56 created 1965]

EC 1.1.1.57

Accepted name: fructuronate reductase
Reaction: D-mannonate + NAD⁺ = D-fructuronate + NADH + H⁺
Other name(s): mannonate oxidoreductase; mannonic dehydrogenase; D-mannonate dehydrogenase; D-mannonate:NAD oxidoreductase
Systematic name: D-mannonate:NAD⁺ 5-oxidoreductase
Comments: Also reduces D-tagaturonate.
References: [1640, 2089]

[EC 1.1.1.57 created 1965]

EC 1.1.1.58

Accepted name: tagaturonate reductase
Reaction: D-altronate + NAD⁺ = D-tagaturonate + NADH + H⁺
Other name(s): altronic oxidoreductase; altronate oxidoreductase; TagUAR; altronate dehydrogenase; D-tagaturonate reductase
Systematic name: D-altronate:NAD⁺ 3-oxidoreductase
References: [1640]

[EC 1.1.1.58 created 1965]

EC 1.1.1.59

Accepted name: 3-hydroxypropionate dehydrogenase
Reaction: 3-hydroxypropanoate + NAD⁺ = 3-oxopropanoate + NADH + H⁺
Systematic name: 3-hydroxypropanoate:NAD⁺ oxidoreductase
References: [877]

[EC 1.1.1.59 created 1965]

EC 1.1.1.60

Accepted name: 2-hydroxy-3-oxopropionate reductase
Reaction: D-glycerate + NAD(P)⁺ = 2-hydroxy-3-oxopropanoate + NAD(P)H + H⁺
Other name(s): tartronate semialdehyde reductase; (*R*)-glycerate:NAD(P)⁺ oxidoreductase
Systematic name: D-glycerate:NAD(P)⁺ oxidoreductase
References: [1372]

[EC 1.1.1.60 created 1965]

EC 1.1.1.61

Accepted name: 4-hydroxybutyrate dehydrogenase
Reaction: 4-hydroxybutanoate + NAD⁺ = succinate semialdehyde + NADH + H⁺
Other name(s): γ-hydroxybutyrate dehydrogenase
Systematic name: 4-hydroxybutanoate:NAD⁺ oxidoreductase
References: [3072]

[EC 1.1.1.61 created 1965]

EC 1.1.1.62

Accepted name: 17β-estradiol 17-dehydrogenase
Reaction: 17β-estradiol + NAD(P)⁺ = estrone + NAD(P)H + H⁺
Other name(s): 20α-hydroxysteroid dehydrogenase; 17β,20α-hydroxysteroid dehydrogenase; 17β-estradiol dehydrogenase; estradiol dehydrogenase; estrogen 17-oxidoreductase; 17β-HSD; HSD17B7

Systematic name: 17 β -estradiol:NAD(P)⁺ 17-oxidoreductase
Comments: The enzyme oxidizes or reduces the hydroxy/keto group on C₁₇ of estrogens and androgens in mammals and regulates the biological potency of these steroids. The mammalian enzyme is bifunctional and also catalyses EC 1.1.1.270, 3 β -hydroxysteroid 3-dehydrogenase [2650]. The enzyme also acts on (*S*)-20-hydroxypregn-4-en-3-one and related compounds, oxidizing the (*S*)-20-group, but unlike EC 1.1.1.149, 20 α -hydroxysteroid dehydrogenase, it is *Si*-specific with respect to NAD(P)⁺.
References: [2039, 2347, 2650]

[EC 1.1.1.62 created 1965, modified 1983, modified 1986, modified 2012]

[1.1.1.63 *Transferred entry. testosterone 17 β -dehydrogenase. Now EC 1.1.1.239, 3 α (17 β)-hydroxysteroid dehydrogenase (NAD⁺)*]

[EC 1.1.1.63 created 1965, deleted 2012]

EC 1.1.1.64

Accepted name: testosterone 17 β -dehydrogenase (NADP⁺)
Reaction: testosterone + NADP⁺ = androstenedione + NADPH + H⁺
Other name(s): 17-ketoreductase; NADP-dependent testosterone-17 β -oxidoreductase; testosterone 17 β -dehydrogenase (NADP)
Systematic name: 17 β -hydroxysteroid:NADP⁺ 17-oxidoreductase
Comments: Also oxidizes 3-hydroxyhexobarbital to 3-oxohexobarbital.
References: [1044, 4149, 4452]

[EC 1.1.1.64 created 1965]

EC 1.1.1.65

Accepted name: pyridoxine 4-dehydrogenase
Reaction: pyridoxine + NADP⁺ = pyridoxal + NADPH + H⁺
Other name(s): pyridoxin dehydrogenase; pyridoxol dehydrogenase; pyridoxine dehydrogenase
Systematic name: pyridoxine:NADP⁺ 4-oxidoreductase
Comments: Also oxidizes pyridoxine phosphate.
References: [1705]

[EC 1.1.1.65 created 1965, modified 1976]

EC 1.1.1.66

Accepted name: ω -hydroxydecanoate dehydrogenase
Reaction: 10-hydroxydecanoate + NAD⁺ = 10-oxodecanoate + NADH + H⁺
Systematic name: 10-hydroxydecanoate:NAD⁺ 10-oxidoreductase
Comments: Also acts, more slowly, on 9-hydroxynonanoate and 11-hydroxyundecanoate.
References: [1983, 2836]

[EC 1.1.1.66 created 1965]

EC 1.1.1.67

Accepted name: mannitol 2-dehydrogenase
Reaction: D-mannitol + NAD⁺ = D-fructose + NADH + H⁺
Other name(s): D-mannitol dehydrogenase; mannitol dehydrogenase
Systematic name: D-mannitol:NAD⁺ 2-oxidoreductase
References: [2670]

[EC 1.1.1.67 created 1965]

[1.1.1.68 *Transferred entry. 5,10-methylenetetrahydrofolate reductase. Now EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]*]

[EC 1.1.1.68 created 1965, deleted 1978 [transferred to EC 1.1.99.15, deleted 1980]]

EC 1.1.1.69

Accepted name: gluconate 5-dehydrogenase
Reaction: D-gluconate + NAD(P)⁺ = 5-dehydro-D-gluconate + NAD(P)H + H⁺
Other name(s): 5-keto-D-gluconate 5-reductase; 5-keto-D-gluconate 5-reductase; 5-ketogluconate 5-reductase; 5-ketogluconate reductase; 5-keto-D-gluconate reductase
Systematic name: D-gluconate:NAD(P)⁺ 5-oxidoreductase
References: [77, 2437, 3156]

[EC 1.1.1.69 created 1965, modified 1976]

[1.1.1.70 *Deleted entry. D-glucuronolactone dehydrogenase. Now included with EC 1.2.1.3 aldehyde dehydrogenase (NAD⁺)*]

[EC 1.1.1.70 created 1965, deleted 1978]

EC 1.1.1.71

Accepted name: alcohol dehydrogenase [NAD(P)⁺]
Reaction: an alcohol + NAD(P)⁺ = an aldehyde + NAD(P)H + H⁺
Other name(s): retinal reductase (ambiguous); aldehyde reductase (NADPH/NADH); alcohol dehydrogenase [NAD(P)]
Systematic name: alcohol:NAD(P)⁺ oxidoreductase
Comments: Reduces aliphatic aldehydes of carbon chain length from 2 to 14, with greatest activity on C₄, C₆ and C₈ aldehydes; also reduces retinal to retinol.
References: [1115]

[EC 1.1.1.71 created 1972]

EC 1.1.1.72

Accepted name: glycerol dehydrogenase (NADP⁺)
Reaction: glycerol + NADP⁺ = D-glyceraldehyde + NADPH + H⁺
Other name(s): glycerol dehydrogenase (NADP)
Systematic name: glycerol:NADP⁺ oxidoreductase
References: [2234, 4299]

[EC 1.1.1.72 created 1972]

EC 1.1.1.73

Accepted name: octanol dehydrogenase
Reaction: octan-1-ol + NAD⁺ = octanal + NADH + H⁺
Other name(s): 1-octanol dehydrogenase; octanol:NAD⁺ oxidoreductase
Systematic name: octan-1-ol:NAD⁺ oxidoreductase
Comments: Acts, less rapidly, on other long-chain alcohols.
References: [3542]

[EC 1.1.1.73 created 1972]

[1.1.1.74 *Deleted entry. D-aminopropanol dehydrogenase (reaction due to EC 1.1.1.4 (R,R)-butanediol dehydrogenase)*]

[EC 1.1.1.74 created 1972, deleted 1976]

EC 1.1.1.75

Accepted name: (*R*)-aminopropanol dehydrogenase
Reaction: (*R*)-1-aminopropan-2-ol + NAD⁺ = aminoacetone + NADH + H⁺
Other name(s): L-aminopropanol dehydrogenase; 1-aminopropan-2-ol-NAD⁺ dehydrogenase; L(+)-1-aminopropan-2-ol:NAD⁺ oxidoreductase; 1-aminopropan-2-ol-dehydrogenase; DL-1-aminopropan-2-ol: NAD⁺ dehydrogenase; L(+)-1-aminopropan-2-ol-NAD/NADP oxidoreductase
Systematic name: (*R*)-1-aminopropan-2-ol:NAD⁺ oxidoreductase
Comments: Requires K⁺.
References: [866, 4354, 4355]

[EC 1.1.1.75 created 1972]

EC 1.1.1.76

Accepted name: (*S,S*)-butanediol dehydrogenase
Reaction: (2*S,3S*)-butane-2,3-diol + NAD⁺ = (*S*)-acetoin + NADH + H⁺
Other name(s): L-butanediol dehydrogenase; L-BDH; L(+)-2,3-butanediol dehydrogenase (L-acetoin forming); (*S*)-acetoin reductase [(*S,S*)-butane-2,3-diol forming]
Systematic name: (*S,S*)-butane-2,3-diol:NAD⁺ oxidoreductase
Comments: This enzyme catalyses the reversible reduction of (*S*)-acetoin to (*S,S*)-butane-2,3-diol. It can also catalyse the irreversible reduction of diacetyl to (*S*)-acetoin.
References: [4228, 553, 4189]

[EC 1.1.1.76 created 1972, modified 2010]

EC 1.1.1.77

Accepted name: lactaldehyde reductase
Reaction: (*R*)[or (*S*)]-propane-1,2-diol + NAD⁺ = (*R*)[or (*S*)]-lactaldehyde + NADH + H⁺
Other name(s): propanediol:nicotinamide adenine dinucleotide (NAD) oxidoreductase; L-lactaldehyde:propanediol oxidoreductase
Systematic name: (*R*)[or (*S*)]-propane-1,2-diol:NAD⁺ oxidoreductase
References: [4292]

[EC 1.1.1.77 created 1972]

EC 1.1.1.78

Accepted name: methylglyoxal reductase (NADH)
Reaction: (*R*)-lactaldehyde + NAD⁺ = 2-oxopropanal + NADH + H⁺
Other name(s): methylglyoxal reductase; D-lactaldehyde dehydrogenase; methylglyoxal reductase (NADH-dependent)
Systematic name: (*R*)-lactaldehyde:NAD⁺ oxidoreductase
Comments: This mammalian enzyme differs from the yeast enzyme, EC 1.1.1.283, methylglyoxal reductase (NADPH-dependent), by its coenzyme requirement, reaction direction, and enantiomeric preference.
References: [4291, 3466]

[EC 1.1.1.78 created 1972, modified 2005, modified 2013]

EC 1.1.1.79

Accepted name: glyoxylate reductase (NADP⁺)
Reaction: glycolate + NADP⁺ = glyoxylate + NADPH + H⁺
Other name(s): NADPH-glyoxylate reductase; glyoxylate reductase (NADP)
Systematic name: glycolate:NADP⁺ oxidoreductase
Comments: Also reduces hydroxypyruvate to glycerate; has some affinity for NAD⁺.
References: [567, 2152]

[EC 1.1.1.79 created 1972]

EC 1.1.1.80

Accepted name: isopropanol dehydrogenase (NADP⁺)
Reaction: propan-2-ol + NADP⁺ = acetone + NADPH + H⁺
Other name(s): isopropanol dehydrogenase (NADP)
Systematic name: propan-2-ol:NADP⁺ oxidoreductase
Comments: Also acts on other short-chain secondary alcohols and, slowly, on primary alcohols.
References: [1735, 1736]

[EC 1.1.1.80 created 1972]

EC 1.1.1.81

Accepted name: hydroxypyruvate reductase
Reaction: D-glycerate + NAD(P)⁺ = hydroxypyruvate + NAD(P)H + H⁺
Other name(s): β-hydroxypyruvate reductase; NADH:hydroxypyruvate reductase; D-glycerate dehydrogenase
Systematic name: D-glycerate:NADP⁺ 2-oxidoreductase
References: [2150, 2151, 2201]

[EC 1.1.1.81 created 1972]

EC 1.1.1.82

Accepted name: malate dehydrogenase (NADP⁺)
Reaction: (S)-malate + NADP⁺ = oxaloacetate + NADPH + H⁺
Other name(s): NADP-malic enzyme; NADP-malate dehydrogenase; malic dehydrogenase (nicotinamide adenine dinucleotide phosphate); malate NADP dehydrogenase; NADP malate dehydrogenase; NADP-linked malate dehydrogenase; malate dehydrogenase (NADP)
Systematic name: (S)-malate:NADP⁺ oxidoreductase
Comments: Activated by light.
References: [721, 1932, 1933]

[EC 1.1.1.82 created 1972]

EC 1.1.1.83

Accepted name: D-malate dehydrogenase (decarboxylating)
Reaction: (R)-malate + NAD⁺ = pyruvate + CO₂ + NADH
Other name(s): D-malate dehydrogenase; D-malic enzyme; bifunctional L(+)-tartrate dehydrogenase-D(+)-malate (decarboxylating)
Systematic name: (R)-malate:NAD⁺ oxidoreductase (decarboxylating)
References: [4025]

[EC 1.1.1.83 created 1972]

EC 1.1.1.84

Accepted name: dimethylmalate dehydrogenase
Reaction: (R)-3,3-dimethylmalate + NAD⁺ = 3-methyl-2-oxobutanoate + CO₂ + NADH
Other name(s): β,β-dimethylmalate dehydrogenase
Systematic name: (R)-3,3-dimethylmalate:NAD⁺ oxidoreductase (decarboxylating)
Comments: Requires K⁺ or NH₄⁺ and Mn²⁺ or Co²⁺; also acts on (R)-malate.
References: [2607]

[EC 1.1.1.84 created 1972]

EC 1.1.1.85

- Accepted name:** 3-isopropylmalate dehydrogenase
Reaction: (2*R*,3*S*)-3-isopropylmalate + NAD⁺ = 4-methyl-2-oxopentanoate + CO₂ + NADH + H⁺ (overall reaction)
(1*a*) (2*R*,3*S*)-3-isopropylmalate + NAD⁺ = (2*S*)-2-isopropyl-3-oxosuccinate + NADH + H⁺
(1*b*) (2*S*)-2-isopropyl-3-oxosuccinate = 4-methyl-2-oxopentanoate + CO₂ (spontaneous)
Other name(s): β-isopropylmalic enzyme; β-isopropylmalate dehydrogenase; *threo*-D₅-3-isopropylmalate dehydrogenase; 3-carboxy-2-hydroxy-4-methylpentanoate:NAD⁺ oxidoreductase
Systematic name: (2*R*,3*S*)-3-isopropylmalate:NAD⁺ oxidoreductase
Comments: The product decarboxylates spontaneously to yield 4-methyl-2-oxopentanoate.
References: [501, 3248, 3041, 531]

[EC 1.1.1.85 created 1972, modified 1976]

EC 1.1.1.86

- Accepted name:** ketol-acid reductoisomerase (NADP⁺)
Reaction: (2*R*)-2,3-dihydroxy-3-methylbutanoate + NADP⁺ = (2*S*)-2-hydroxy-2-methyl-3-oxobutanoate + NADPH + H⁺
Other name(s): dihydroxyisovalerate dehydrogenase (isomerizing); acetohydroxy acid isomeroreductase; ketol acid reductoisomerase; α-keto-β-hydroxyacyl reductoisomerase; 2-hydroxy-3-keto acid reductoisomerase; acetohydroxy acid reductoisomerase; acetolactate reductoisomerase; dihydroxyisovalerate (isomerizing) dehydrogenase; isomeroreductase; reductoisomerase; ketol-acid reductoisomerase; (2*R*)-2,3-dihydroxy-3-methylbutanoate:NADP⁺ oxidoreductase (isomerizing)
Systematic name: (2*R*)-2,3-dihydroxy-3-methylbutanoate:NADP⁺ oxidoreductase (isomerizing)
Comments: Also catalyses the reduction of 2-ethyl-2-hydroxy-3-oxobutanoate to 2,3-dihydroxy-3-methylpentanoate. The enzyme, found in many bacteria and archaea, is specific for NADPH (*cf.* EC 1.1.1.382, ketol-acid reductoisomerase (NAD⁺) and EC 1.1.1.383, ketol-acid reductoisomerase [NAD(P)⁺]).
References: [130, 1652, 2126, 3675, 443]

[EC 1.1.1.86 created 1972, modified 1976, modified 1981 (EC 1.1.1.89 created 1972, incorporated 1976), modified 2015]

EC 1.1.1.87

- Accepted name:** homoisocitrate dehydrogenase
Reaction: (1*R*,2*S*)-1-hydroxybutane-1,2,4-tricarboxylate + NAD⁺ = 2-oxoadipate + CO₂ + NADH + H⁺
Other name(s): 2-hydroxy-3-carboxyadipate dehydrogenase; 3-carboxy-2-hydroxyadipate dehydrogenase; homoisocitric dehydrogenase; (-)-1-hydroxy-1,2,4-butanetricarboxylate:NAD⁺ oxidoreductase (decarboxylating); 3-carboxy-2-hydroxyadipate:NAD⁺ oxidoreductase (decarboxylating); HICDH
Systematic name: (1*R*,2*S*)-1-hydroxybutane-1,2,4-tricarboxylate:NAD⁺ oxidoreductase (decarboxylating)
Comments: Forms part of the lysine biosynthesis pathway in fungi [4851].
References: [4054, 3584, 4851]

[EC 1.1.1.87 created 1972 (EC 1.1.1.155 created 1976, incorporated 2004)]

EC 1.1.1.88

- Accepted name:** hydroxymethylglutaryl-CoA reductase
Reaction: (*R*)-mevalonate + CoA + 2 NAD⁺ = 3-hydroxy-3-methylglutaryl-CoA + 2 NADH + 2 H⁺
Other name(s): β-hydroxy-β-methylglutaryl coenzyme A reductase (ambiguous); β-hydroxy-β-methylglutaryl CoA-reductase (ambiguous); 3-hydroxy-3-methylglutaryl coenzyme A reductase (ambiguous); hydroxymethylglutaryl coenzyme A reductase (ambiguous)
Systematic name: (*R*)-mevalonate:NAD⁺ oxidoreductase (CoA-acylating)
References: [1123]

[EC 1.1.1.88 created 1972, modified 2002]

[1.1.1.89 Deleted entry. dihydroxyisovalerate dehydrogenase (isomerizing). Now included with EC 1.1.1.86 ketol-acid reductoisomerase]

[EC 1.1.1.89 created 1972, deleted 1976]

EC 1.1.1.90

Accepted name: aryl-alcohol dehydrogenase
Reaction: an aromatic alcohol + NAD^+ = an aromatic aldehyde + $\text{NADH} + \text{H}^+$
Other name(s): *p*-hydroxybenzyl alcohol dehydrogenase; benzyl alcohol dehydrogenase; coniferyl alcohol dehydrogenase
Systematic name: aryl-alcohol: NAD^+ oxidoreductase
Comments: A group of enzymes with broad specificity towards primary alcohols with an aromatic or cyclohex-1-ene ring, but with low or no activity towards short-chain aliphatic alcohols.
References: [4107, 4745]

[EC 1.1.1.90 created 1972, modified 1989]

EC 1.1.1.91

Accepted name: aryl-alcohol dehydrogenase (NADP^+)
Reaction: an aromatic alcohol + NADP^+ = an aromatic aldehyde + $\text{NADPH} + \text{H}^+$
Other name(s): aryl alcohol dehydrogenase (nicotinamide adenine dinucleotide phosphate); coniferyl alcohol dehydrogenase; NADPH -linked benzaldehyde reductase; aryl-alcohol dehydrogenase (NADP)
Systematic name: aryl-alcohol: NADP^+ oxidoreductase
Comments: Also acts on some aliphatic aldehydes, but cinnamaldehyde was the best substrate found.
References: [1426]

[EC 1.1.1.91 created 1972]

EC 1.1.1.92

Accepted name: oxaloglycolate reductase (decarboxylating)
Reaction: $\text{D-glycerate} + \text{NAD(P)}^+ + \text{CO}_2 = 2\text{-hydroxy-3-oxosuccinate} + \text{NAD(P)H} + 2 \text{H}^+$
Systematic name: $\text{D-glycerate}:\text{NAD(P)}^+$ oxidoreductase (carboxylating)
Comments: Also reduces hydroxypyruvate to D-glycerate and glyoxylate to glycolate.
References: [2200]

[EC 1.1.1.92 created 1972]

EC 1.1.1.93

Accepted name: tartrate dehydrogenase
Reaction: $\text{tartrate} + \text{NAD}^+ = \text{oxaloglycolate} + \text{NADH} + \text{H}^+$
Other name(s): mesotartrate dehydrogenase
Systematic name: tartrate: NAD^+ oxidoreductase
Comments: *meso*-tartrate and (*R,R*)-tartrate act as substrates. Requires Mn^{2+} and a monovalent cation.
References: [2203]

[EC 1.1.1.93 created 1972]

EC 1.1.1.94

Accepted name: glycerol-3-phosphate dehydrogenase [NAD(P)^+]
Reaction: *sn*-glycerol 3-phosphate + $\text{NAD(P)}^+ = \text{glycerone phosphate} + \text{NAD(P)H} + \text{H}^+$

Other name(s): L-glycerol-3-phosphate:NAD(P) oxidoreductase; glycerol phosphate dehydrogenase (nicotinamide adenine dinucleotide (phosphate)); glycerol 3-phosphate dehydrogenase (NADP); glycerol-3-phosphate dehydrogenase [NAD(P)]
Systematic name: *sn*-glycerol-3-phosphate:NAD(P)⁺ 2-oxidoreductase
Comments: The enzyme from *Escherichia coli* shows specificity for the B side of NADPH.
References: [2140, 1015, 1016, 1017]

[EC 1.1.1.94 created 1972, modified 2005]

EC 1.1.1.95

Accepted name: phosphoglycerate dehydrogenase
Reaction: 3-phospho-D-glycerate + NAD⁺ = 3-phosphooxypyruvate + NADH + H⁺
Other name(s): PHGDH (gene name); D-3-phosphoglycerate:NAD⁺ oxidoreductase; α-phosphoglycerate dehydrogenase; 3-phosphoglycerate dehydrogenase; 3-phosphoglyceric acid dehydrogenase; D-3-phosphoglycerate dehydrogenase; glycerate 3-phosphate dehydrogenase; glycerate-1,3-phosphate dehydrogenase; phosphoglycerate oxidoreductase; phosphoglyceric acid dehydrogenase; SerA; 3-phosphoglycerate:NAD⁺ 2-oxidoreductase; SerA 3PG dehydrogenase; 3PHP reductase
Systematic name: 3-phospho-D-glycerate:NAD⁺ 2-oxidoreductase
Comments: This enzyme catalyses the first committed and rate-limiting step in the phosphoserine pathway of serine biosynthesis. The reaction occurs predominantly in the direction of reduction. The enzyme from the bacterium *Escherichia coli* also catalyses the activity of EC 1.1.1.399, 2-oxoglutarate reductase [4900].
References: [3332, 4507, 3926, 4096, 3751, 4900, 9, 897]

[EC 1.1.1.95 created 1972, modified 2006, modified 2016]

EC 1.1.1.96

Accepted name: diiodophenylpyruvate reductase
Reaction: 3-(3,5-diiodo-4-hydroxyphenyl)lactate + NAD⁺ = 3-(3,5-diiodo-4-hydroxyphenyl)pyruvate + NADH + H⁺
Other name(s): aromatic α-keto acid; KAR; 2-oxo acid reductase
Systematic name: 3-(3,5-diiodo-4-hydroxyphenyl)lactate:NAD⁺ oxidoreductase
Comments: Substrates contain an aromatic ring with a pyruvate side chain. The most active substrates are halogenated derivatives. Compounds with hydroxy or amino groups in the 3 or 5 position are inactive.
References: [4860]

[EC 1.1.1.96 created 1972]

EC 1.1.1.97

Accepted name: 3-hydroxybenzyl-alcohol dehydrogenase
Reaction: 3-hydroxybenzyl alcohol + NADP⁺ = 3-hydroxybenzaldehyde + NADPH + H⁺
Other name(s): *m*-hydroxybenzyl alcohol dehydrogenase; *m*-hydroxybenzyl alcohol (NADP) dehydrogenase; *m*-hydroxybenzylalcohol dehydrogenase
Systematic name: 3-hydroxybenzyl-alcohol:NADP⁺ oxidoreductase
References: [1146]

[EC 1.1.1.97 created 1972]

EC 1.1.1.98

Accepted name: (*R*)-2-hydroxy-fatty-acid dehydrogenase
Reaction: (*R*)-2-hydroxystearate + NAD⁺ = 2-oxostearate + NADH + H⁺
Other name(s): D-2-hydroxy fatty acid dehydrogenase; 2-hydroxy fatty acid oxidase
Systematic name: (*R*)-2-hydroxystearate:NAD⁺ oxidoreductase

References: [2430]

[EC 1.1.1.98 created 1972]

EC 1.1.1.99

Accepted name: (*S*)-2-hydroxy-fatty-acid dehydrogenase
Reaction: (*S*)-2-hydroxystearate + NAD⁺ = 2-oxostearate + NADH + H⁺
Other name(s): dehydrogenase, L-2-hydroxy fatty acid; L-2-hydroxy fatty acid dehydrogenase; 2-hydroxy fatty acid oxidase
Systematic name: (*S*)-2-hydroxystearate:NAD⁺ oxidoreductase
References: [2430]

[EC 1.1.1.99 created 1972]

EC 1.1.1.100

Accepted name: 3-oxoacyl-[acyl-carrier-protein] reductase
Reaction: a (3*R*)-3-hydroxyacyl-[acyl-carrier protein] + NADP⁺ = a 3-oxoacyl-[acyl-carrier protein] + NADPH + H⁺
Other name(s): β-ketoacyl-[acyl-carrier protein](ACP) reductase; β-ketoacyl acyl carrier protein (ACP) reductase; β-ketoacyl reductase; β-ketoacyl thioester reductase; β-ketoacyl-ACP reductase; β-ketoacyl-acyl carrier protein reductase; 3-ketoacyl acyl carrier protein reductase; NADPH-specific 3-oxoacyl-[acylcarrier protein]reductase; 3-oxoacyl-[ACP]reductase; (3*R*)-3-hydroxyacyl-[acyl-carrier-protein]:NADP⁺ oxidoreductase
Systematic name: (3*R*)-3-hydroxyacyl-[acyl-carrier protein]:NADP⁺ oxidoreductase
Comments: Exhibits a marked preference for acyl-carrier-protein derivatives over CoA derivatives as substrates.
References: [3375, 3866, 4312]

[EC 1.1.1.100 created 1972, modified 1976]

EC 1.1.1.101

Accepted name: acylglycerone-phosphate reductase
Reaction: 1-palmitoylglycerol 3-phosphate + NADP⁺ = palmitoylglycerone phosphate + NADPH + H⁺
Other name(s): palmitoyldihydroxyacetone-phosphate reductase; palmitoyl dihydroxyacetone phosphate reductase; palmitoyl-dihydroxyacetone-phosphate reductase; acyldihydroxyacetone phosphate reductase; palmitoyl dihydroxyacetone phosphate reductase
Systematic name: 1-palmitoylglycerol-3-phosphate:NADP⁺ oxidoreductase
Comments: Also acts on alkylglycerone 3-phosphate and alkylglycerol 3-phosphate.
References: [2321]

[EC 1.1.1.101 created 1972, modified 1976]

EC 1.1.1.102

Accepted name: 3-dehydrosphinganine reductase
Reaction: sphinganine + NADP⁺ = 3-dehydrosphinganine + NADPH + H⁺
Other name(s): D-3-dehydrosphinganine reductase; D-3-oxosphinganine reductase; DSR; 3-oxosphinganine reductase; 3-oxosphinganine:NADPH oxidoreductase; D-3-oxosphinganine:B-NADPH oxidoreductase
Systematic name: D-*erythro*-dihydrosphingosine:NADP⁺ 3-oxidoreductase
References: [4039, 4040]

[EC 1.1.1.102 created 1972]

EC 1.1.1.103

- Accepted name:** L-threonine 3-dehydrogenase
Reaction: L-threonine + NAD⁺ = L-2-amino-3-oxobutanoate + NADH + H⁺
Other name(s): L-threonine dehydrogenase; threonine 3-dehydrogenase; threonine dehydrogenase; TDH
Systematic name: L-threonine:NAD⁺ oxidoreductase
Comments: This enzyme acts in concert with EC 2.3.1.29, glycine C-acetyltransferase, in the degradation of threonine to glycine. This threonine-degradation pathway is common to prokaryotic and eukaryotic cells and the two enzymes involved form a complex [1542]. In aqueous solution, the product L-2-amino-3-oxobutanoate can spontaneously decarboxylate to form aminoacetone.
References: [1397, 1542, 3054, 1057]

[EC 1.1.1.103 created 1972]

EC 1.1.1.104

- Accepted name:** 4-oxoproline reductase
Reaction: *cis*-4-hydroxy-L-proline + NAD⁺ = 4-oxo-L-proline + NADH + H⁺
Other name(s): *cis*-hydroxy-L-proline oxidase
Systematic name: *cis*-4-hydroxy-L-proline:NAD⁺ oxidoreductase (4-oxo-L-proline forming)
Comments: The enzyme, isolated from animals, is specific for 4-oxo-L-proline and *cis*-4-hydroxy-L-proline. It has no activity with *trans*-4-hydroxy-L-proline.
References: [3946, 2319]

[EC 1.1.1.104 created 1972, modified 2022]

EC 1.1.1.105

- Accepted name:** *all-trans*-retinol dehydrogenase (NAD⁺)
Reaction: *all-trans*-retinol—[cellular-retinol-binding-protein] + NAD⁺ = *all-trans*-retinal—[cellular-retinol-binding-protein] + NADH + H⁺
Other name(s): retinol (vitamin A₁) dehydrogenase; MDR; microsomal retinol dehydrogenase; retinol dehydrogenase (misleading); retinal reductase (ambiguous); retinene reductase; epidermal retinol dehydrogenase 2; SDR16C5 (gene name); RDH16 (gene name)
Systematic name: *all-trans* retinol:NAD⁺ oxidoreductase
Comments: The enzyme recognizes *all-trans*-retinol and *all-trans*-retinal as substrates and exhibits a strong preference for NAD⁺/NADH as cofactors. Recognizes the substrate both in free form and when bound to cellular-retinol-binding-protein (CRBP1), but has higher affinity for the bound form [1375]. No activity with 11-*cis*-retinol or 11-*cis*-retinal (*cf.* EC 1.1.1.315, 11-*cis* retinol dehydrogenase). Also active with 3 α -hydroxysteroids [1375].
References: [2190, 1375, 2721, 2396]

[EC 1.1.1.105 created 1972, modified 2011]

EC 1.1.1.106

- Accepted name:** pantoate 4-dehydrogenase
Reaction: (*R*)-pantoate + NAD⁺ = (*R*)-4-dehydropantoate + NADH + H⁺
Other name(s): pantoate dehydrogenase; pantothenase; D-pantoate:NAD⁺ 4-oxidoreductase
Systematic name: (*R*)-pantoate:NAD⁺ 4-oxidoreductase
References: [1362]

[EC 1.1.1.106 created 1972, modified 1976]

EC 1.1.1.107

- Accepted name:** pyridoxal 4-dehydrogenase
Reaction: pyridoxal + NAD⁺ = 4-pyridoxolactone + NADH + H⁺

Other name(s): pyridoxal dehydrogenase
Systematic name: pyridoxal:NAD⁺ 4-oxidoreductase
Comments: The enzyme acts on the hemiacetal form of the substrate.
References: [495]

[EC 1.1.1.107 created 1972]

EC 1.1.1.108

Accepted name: carnitine 3-dehydrogenase
Reaction: carnitine + NAD⁺ = 3-dehydrocarnitine + NADH + H⁺
Systematic name: carnitine:NAD⁺ 3-oxidoreductase
References: [157, 3745]

[EC 1.1.1.108 created 1972]

[1.1.1.109 Transferred entry. 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase. Now EC 1.3.1.28, 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase]

[EC 1.1.1.109 created 1972, deleted 1976]

EC 1.1.1.110

Accepted name: aromatic 2-oxoacid reductase
Reaction: (1) (*R*)-3-(phenyl)lactate + NAD⁺ = 3-phenylpyruvate + NADH + H⁺
(2) (*R*)-3-(4-hydroxyphenyl)lactate + NAD⁺ = 3-(4-hydroxyphenyl)pyruvate + NADH + H⁺
(3) (*R*)-(indol-3-yl)lactate + NAD⁺ = (indol-3-yl)pyruvate + NADH + H⁺
Other name(s): (*R*)-aromatic lactate dehydrogenase; (*R*)-4-hydroxyphenyllactate dehydrogenase; indolelactate:NAD⁺ oxidoreductase; indolelactate dehydrogenase; *fldH* (gene name); (indol-3-yl)lactate:NAD⁺ oxidoreductase
Systematic name: aromatic 2-oxoacid:NAD⁺ oxidoreductase
Comments: The enzymes from anaerobic bacteria such as *Clostridium sporogenes* participate in the fermentation pathways of L-phenylalanine, L-tyrosine and L-tryptophan. The enzyme from the yeast *Candida maltosa* has similar activity, but, unlike the bacterial enzyme, requires Mn²⁺ and can also use NADPH with lower activity.
References: [1895, 1323, 364, 906, 936]

[EC 1.1.1.110 created 1972 (EC 1.1.1.222 created 2000, incorporated 2018), modified 2018]

EC 1.1.1.111

Accepted name: 3-(imidazol-5-yl)lactate dehydrogenase
Reaction: (*S*)-3-(imidazol-5-yl)lactate + NAD(P)⁺ = 3-(imidazol-5-yl)pyruvate + NAD(P)H + H⁺
Other name(s): imidazol-5-yl lactate dehydrogenase
Systematic name: (*S*)-3-(imidazol-5-yl)lactate:NAD(P)⁺ oxidoreductase
References: [732, 743]

[EC 1.1.1.111 created 1972]

EC 1.1.1.112

Accepted name: indanol dehydrogenase
Reaction: indan-1-ol + NAD(P)⁺ = indanone + NAD(P)H + H⁺
Systematic name: indan-1-ol:NAD(P)⁺ 1-oxidoreductase
Comments: 3(20) α -Hydroxysteroids are also oxidized, more slowly.
References: [334, 1523]

[EC 1.1.1.112 created 1972]

EC 1.1.1.113

Accepted name: L-xylose 1-dehydrogenase
Reaction: L-xylose + NADP⁺ = L-xylono-1,4-lactone + NADPH + H⁺
Other name(s): L-xylose dehydrogenase; NADPH-xylose reductase
Systematic name: L-xylose:NADP⁺ 1-oxidoreductase
Comments: Also oxidizes D-arabinose and D-lyxose.
References: [4367]

[EC 1.1.1.113 created 1972]

EC 1.1.1.114

Accepted name: apiose 1-reductase
Reaction: D-apiitol + NAD⁺ = D-apyiose + NADH + H⁺
Other name(s): D-apyiose reductase; D-apiitol reductase
Systematic name: D-apiitol:NAD⁺ 1-oxidoreductase
References: [1508, 3029]

[EC 1.1.1.114 created 1972]

EC 1.1.1.115

Accepted name: ribose 1-dehydrogenase (NADP⁺)
Reaction: D-ribose + NADP⁺ + H₂O = D-ribonate + NADPH + H⁺
Other name(s): D-ribose dehydrogenase (NADP⁺); NADP-pentose-dehydrogenase; ribose 1-dehydrogenase (NADP)
Systematic name: D-ribose:NADP⁺ 1-oxidoreductase
Comments: Also acts, more slowly, on D-xylose and other pentoses.
References: [3712, 3719]

[EC 1.1.1.115 created 1972]

EC 1.1.1.116

Accepted name: D-arabinose 1-dehydrogenase (NAD⁺)
Reaction: D-arabinose + NAD⁺ = D-arabinono-1,4-lactone + NADH + H⁺
Other name(s): NAD⁺-pentose-dehydrogenase; arabinose(fucose)dehydrogenase
Systematic name: D-arabinose:NAD⁺ 1-oxidoreductase
References: [3224, 3719]

[EC 1.1.1.116 created 1972]

EC 1.1.1.117

Accepted name: D-arabinose 1-dehydrogenase [NAD(P)⁺]
Reaction: D-arabinose + NAD(P)⁺ = D-arabinono-1,4-lactone + NAD(P)H + H⁺
Other name(s): D-arabinose 1-dehydrogenase [NAD(P)]
Systematic name: D-arabinose:NAD(P)⁺ 1-oxidoreductase
Comments: Also acts on L-galactose, 6-deoxy- and 3,6-dideoxy-L-galactose.
References: [706, 704, 705]

[EC 1.1.1.117 created 1972]

EC 1.1.1.118

Accepted name: glucose 1-dehydrogenase (NAD⁺)
Reaction: D-glucose + NAD⁺ = D-glucono-1,5-lactone + NADH + H⁺
Other name(s): D-glucose:NAD oxidoreductase; D-aldohexose dehydrogenase; glucose 1-dehydrogenase (NAD)

Systematic name: D-glucose:NAD⁺ 1-oxidoreductase
References: [1746]

[EC 1.1.1.118 created 1972, modified 1976]

EC 1.1.1.119

Accepted name: glucose 1-dehydrogenase (NADP⁺)
Reaction: D-glucose + NADP⁺ = D-glucono-1,5-lactone + NADPH + H⁺
Other name(s): nicotinamide adenine dinucleotide phosphate-linked aldohexose dehydrogenase; NADP-linked aldohexose dehydrogenase; NADP-dependent glucose dehydrogenase; glucose 1-dehydrogenase (NADP)
Systematic name: D-glucose:NADP⁺ 1-oxidoreductase
Comments: Also oxidizes D-mannose, 2-deoxy-D-glucose and 2-amino-2-deoxy-D-mannose.
References: [13, 158]

[EC 1.1.1.119 created 1972]

EC 1.1.1.120

Accepted name: galactose 1-dehydrogenase (NADP⁺)
Reaction: D-galactose + NADP⁺ = D-galactono-1,5-lactone + NADPH + H⁺
Other name(s): D-galactose dehydrogenase (NADP⁺); galactose 1-dehydrogenase (NADP)
Systematic name: D-galactose:NADP⁺ 1-oxidoreductase
Comments: Also acts on L-arabinose, 6-deoxy- and 2-deoxy-D-galactose.
References: [706, 704, 705, 3718]

[EC 1.1.1.120 created 1972]

EC 1.1.1.121

Accepted name: aldose 1-dehydrogenase (NAD⁺)
Reaction: D-aldose + NAD⁺ = D-aldonolactone + NADH + H⁺
Other name(s): aldose dehydrogenase; D-aldohexose dehydrogenase; aldose 1-dehydrogenase
Systematic name: D-aldose:NAD⁺ 1-oxidoreductase
Comments: Acts on D-glucose, 2-deoxy- and 6-deoxy-D-glucose, D-galactose, 6-deoxy-D-galactose, 2-deoxy-L-arabinose and D-xylose.
References: [706, 704, 705]

[EC 1.1.1.121 created 1972]

EC 1.1.1.122

Accepted name: D-*threo*-aldose 1-dehydrogenase
Reaction: a D-*threo*-aldose + NAD⁺ = a D-*threo*-aldono-1,5-lactone + NADH + H⁺
Other name(s): L-fucose dehydrogenase; (2*S*,3*R*)-aldose dehydrogenase; dehydrogenase, L-fucose; L-fucose (D-arabinose) dehydrogenase
Systematic name: D-*threo*-aldose:NAD⁺ 1-oxidoreductase
Comments: Acts on L-fucose, D-arabinose and L-xylose; the animal enzyme was also shown to act on L-arabinose, and the enzyme from *Pseudomonas caryophylli* on L-glucose.
References: [3666, 3698]

[EC 1.1.1.122 created 1972]

EC 1.1.1.123

Accepted name: sorbose 5-dehydrogenase (NADP⁺)
Reaction: L-sorbose + NADP⁺ = 5-dehydro-D-fructose + NADPH + H⁺

Other name(s): 5-ketofructose reductase; 5-keto-D-fructose reductase; sorbose (nicotinamide adenine dinucleotide phosphate) dehydrogenase; reduced nicotinamide adenine dinucleotide phosphate-linked reductase; sorbose 5-dehydrogenase (NADP⁺)
Systematic name: L-sorbose:NADP⁺ 5-oxidoreductase
References: [1049]

[EC 1.1.1.123 created 1972, modified 1976]

EC 1.1.1.124

Accepted name: fructose 5-dehydrogenase (NADP⁺)
Reaction: D-fructose + NADP⁺ = 5-dehydro-D-fructose + NADPH + H⁺
Other name(s): 5-ketofructose reductase (NADP); 5-keto-D-fructose reductase (NADP⁺); fructose 5-(nicotinamide adenine dinucleotide phosphate) dehydrogenase; D-(-)fructose:(NADP⁺) 5-oxidoreductase; fructose 5-dehydrogenase (NADP)
Systematic name: D-fructose:NADP⁺ 5-oxidoreductase
References: [81, 160]

[EC 1.1.1.124 created 1972, modified 1976]

EC 1.1.1.125

Accepted name: 2-deoxy-D-gluconate 3-dehydrogenase
Reaction: 2-deoxy-D-gluconate + NAD⁺ = 3-dehydro-2-deoxy-D-gluconate + NADH + H⁺
Other name(s): 2-deoxygluconate dehydrogenase
Systematic name: 2-deoxy-D-gluconate:NAD⁺ 3-oxidoreductase
References: [1029]

[EC 1.1.1.125 created 1972]

EC 1.1.1.126

Accepted name: 2-dehydro-3-deoxy-D-gluconate 6-dehydrogenase
Reaction: 2-dehydro-3-deoxy-D-gluconate + NADP⁺ = (4S,5S)-4,5-dihydroxy-2,6-dioxohexanoate + NADPH + H⁺
Other name(s): 2-keto-3-deoxy-D-gluconate dehydrogenase (ambiguous); 2-keto-3-deoxygluconate dehydrogenase (ambiguous)
Systematic name: 2-dehydro-3-deoxy-D-gluconate:NADP⁺ 6-oxidoreductase
References: [3371]

[EC 1.1.1.126 created 1972]

EC 1.1.1.127

Accepted name: 2-dehydro-3-deoxy-D-gluconate 5-dehydrogenase
Reaction: 2-dehydro-3-deoxy-D-gluconate + NAD⁺ = (4S)-4,6-dihydroxy-2,5-dioxohexanoate + NADH + H⁺
Other name(s): 2-keto-3-deoxygluconate 5-dehydrogenase; 2-keto-3-deoxy-D-gluconate dehydrogenase (ambiguous); 2-keto-3-deoxygluconate (nicotinamide adenine dinucleotide (phosphate)) dehydrogenase; 2-keto-3-deoxy-D-gluconate (3-deoxy-D-glycero-2,5-hexodiulosonic acid) dehydrogenase (ambiguous)
Systematic name: 2-dehydro-3-deoxy-D-gluconate:NAD⁺ 5-oxidoreductase
Comments: The enzyme from *Pseudomonas* acts equally well on NAD⁺ or NADP⁺, while that from *Erwinia chrysanthemi* and *Escherichia coli* is more specific for NAD⁺.
References: [719, 3372]

[EC 1.1.1.127 created 1972, modified 1976, modified 1989]

[1.1.1.128 Deleted entry. L-idonate 2-dehydrogenase. The reaction described is covered by EC 1.1.1.264.]

[EC 1.1.1.128 created 1972, modified 1976, deleted 2012]

EC 1.1.1.129

Accepted name: L-threonate 3-dehydrogenase
Reaction: L-threonate + NAD⁺ = 3-dehydro-L-erythronate + NADH + H⁺
Other name(s): threonate dehydrogenase; L-threonic acid dehydrogenase
Systematic name: L-threonate:NAD⁺ 3-oxidoreductase
References: [149]

[EC 1.1.1.129 created 1972]

EC 1.1.1.130

Accepted name: 3-dehydro-L-gulonate 2-dehydrogenase
Reaction: 3-dehydro-L-gulonate + NAD(P)⁺ = (4R,5S)-4,5,6-trihydroxy-2,3-dioxohexanoate + NAD(P)H + H⁺
Other name(s): 3-keto-L-gulonate dehydrogenase; 3-ketogulonate dehydrogenase; 3-keto-L-gulonate dehydrogenase; 3-ketogulonate dehydrogenase
Systematic name: 3-dehydro-L-gulonate:NAD(P)⁺ 2-oxidoreductase
References: [4466]

[EC 1.1.1.130 created 1972]

EC 1.1.1.131

Accepted name: manuronate reductase
Reaction: D-mannonate + NAD(P)⁺ = D-mannuronate + NAD(P)H + H⁺
Other name(s): mannonate dehydrogenase; mannonate (nicotinamide adenine dinucleotide (phosphate))dehydrogenase; mannonate dehydrogenase; manuronate reductase; mannonate dehydrogenase (NAD(P)⁺); D-mannonate:nicotinamide adenine dinucleotide (phosphate oxidoreductase (D-mannuronate-forming))
Systematic name: D-mannonate:NAD(P)⁺ 6-oxidoreductase
References: [1091]

[EC 1.1.1.131 created 1972 (EC 1.2.1.34 created 1972, incorporated 1983; EC 1.1.1.180 created 1983, incorporated 1984)]

EC 1.1.1.132

Accepted name: GDP-mannose 6-dehydrogenase
Reaction: GDP-D-mannose + 2 NAD⁺ + H₂O = GDP-D-mannuronate + 2 NADH + 2 H⁺
Other name(s): guanosine diphosphomannose dehydrogenase; GDP-mannose dehydrogenase; guanosine diphosphomannose dehydrogenase; guanosine diphospho-D-mannose dehydrogenase
Systematic name: GDP-D-mannose:NAD⁺ 6-oxidoreductase
Comments: Also acts on the corresponding deoxynucleoside diphosphate derivative as a substrate.
References: [3370]

[EC 1.1.1.132 created 1972]

EC 1.1.1.133

Accepted name: dTDP-4-dehydrorhamnose reductase
Reaction: dTDP-β-L-rhamnose + NADP⁺ = dTDP-4-dehydro-β-L-rhamnose + NADPH + H⁺
Other name(s): dTDP-4-keto-L-rhamnose reductase; dTDP-4-ketorhamnose reductase; TDP-4-keto-rhamnose reductase; thymidine diphospho-4-ketorhamnose reductase; dTDP-6-deoxy-L-mannose:NADP⁺ 4-oxidoreductase; dTDP-6-deoxy-β-L-mannose:NADP⁺ 4-oxidoreductase
Systematic name: dTDP-β-L-rhamnose:NADP⁺ 4-oxidoreductase

Comments: In the reverse direction, reduction on the 4-position of the hexose moiety takes place only while the substrate is bound to another enzyme that catalyses epimerization at C-3 and C-5; the complex has been referred to as dTDP-L-rhamnose synthase.

References: [2767]

[EC 1.1.1.133 created 1972]

EC 1.1.1.134

Accepted name: dTDP-6-deoxy-L-talose 4-dehydrogenase (NADP⁺)

Reaction: dTDP-6-deoxy-β-L-talose + NADP⁺ = dTDP-4-dehydro-β-L-rhamnose + NADPH + H⁺

Other name(s): thymidine diphospho-6-deoxy-L-talose dehydrogenase; TDP-6-deoxy-L-talose dehydrogenase; dTDP-6-deoxy-L-talose dehydrogenase (4-reductase); dTDP-6-deoxy-L-talose:NADP⁺ 4-oxidoreductase

Systematic name: dTDP-6-deoxy-β-L-talose:NADP⁺ 4-oxidoreductase

Comments: Oxidation on the 4-position of the hexose moiety takes place only while the substrate is bound to another enzyme that catalyses epimerization at C-3 and C-5.

References: [1285]

[EC 1.1.1.134 created 1972]

EC 1.1.1.135

Accepted name: GDP-6-deoxy-D-talose 4-dehydrogenase

Reaction: GDP-6-deoxy-α-D-talose + NAD(P)⁺ = GDP-4-dehydro-α-D-rhamnose + NAD(P)H + H⁺

Other name(s): guanosine diphospho-6-deoxy-D-talose dehydrogenase; GDP-6-deoxy-D-talose:NAD(P)⁺ 4-oxidoreductase

Systematic name: GDP-6-deoxy-α-D-talose:NAD(P)⁺ 4-oxidoreductase

References: [2653]

[EC 1.1.1.135 created 1972, modified 1976]

EC 1.1.1.136

Accepted name: UDP-N-acetylglucosamine 6-dehydrogenase

Reaction: UDP-N-acetyl-α-D-glucosamine + 2 NAD⁺ + H₂O = UDP-2-acetamido-2-deoxy-α-D-glucuronate + 2 NADH + 2 H⁺

Other name(s): uridine diphosphoacetylglucosamine dehydrogenase; UDP-acetylglucosamine dehydrogenase; UDP-2-acetamido-2-deoxy-D-glucose:NAD oxidoreductase; UDP-GlcNAc dehydrogenase; WbpA; WbpO

Systematic name: UDP-N-acetyl-α-D-glucosamine:NAD⁺ 6-oxidoreductase

Comments: This enzyme participates in the biosynthetic pathway for UDP-α-D-ManNAc3NAcA (UDP-2,3-diacetamido-2,3-dideoxy-α-D-mannuronic acid), an important precursor of B-band lipopolysaccharide.

References: [1082, 2815]

[EC 1.1.1.136 created 1972, modified 2012]

EC 1.1.1.137

Accepted name: ribitol-5-phosphate 2-dehydrogenase

Reaction: D-ribitol 5-phosphate + NAD(P)⁺ = D-ribulose 5-phosphate + NAD(P)H + H⁺

Other name(s): ribitol 5-phosphate dehydrogenase

Systematic name: D-ribitol-5-phosphate:NAD(P)⁺ 2-oxidoreductase

Comments: The enzyme, characterized from the bacterium *Lactobacillus plantarum*, can use both NAD⁺ and NADP⁺ as electron acceptor [*cf.* EC 1.1.1.405, ribitol-5-phosphate 2-dehydrogenase (NADP⁺)].

References: [1339]

[EC 1.1.1.137 created 1972, modified 2017]

EC 1.1.1.138

Accepted name: mannitol 2-dehydrogenase (NADP⁺)
Reaction: D-mannitol + NADP⁺ = D-fructose + NADPH + H⁺
Other name(s): mannitol 2-dehydrogenase (NADP)
Systematic name: D-mannitol:NADP⁺ 2-oxidoreductase
References: [1919, 4071]

[EC 1.1.1.138 created 1972]

[1.1.1.139 Deleted entry. polyol dehydrogenase (NADP⁺). Now included with EC 1.1.1.21 aldehyde reductase]

[EC 1.1.1.139 created 1972, deleted 1978]

EC 1.1.1.140

Accepted name: sorbitol-6-phosphate 2-dehydrogenase
Reaction: D-sorbitol 6-phosphate + NAD⁺ = D-fructose 6-phosphate + NADH + H⁺
Other name(s): ketosephosphate reductase; ketosephosphate reductase; D-sorbitol 6-phosphate dehydrogenase; D-sorbitol-6-phosphate dehydrogenase; sorbitol-6-*P*-dehydrogenase; D-glucitol-6-phosphate dehydrogenase
Systematic name: D-sorbitol-6-phosphate:NAD⁺ 2-oxidoreductase
References: [4300, 2506]

[EC 1.1.1.140 created 1972]

EC 1.1.1.141

Accepted name: 15-hydroxyprostaglandin dehydrogenase (NAD⁺)
Reaction: (5*Z*,13*E*,15*S*)-11 α ,15-dihydroxy-9-oxoprost-5,13-dienoate + NAD⁺ = (5*Z*,13*E*)-11 α -hydroxy-9,15-dioxoprost-5,13-dienoate + NADH + H⁺
Other name(s): NAD⁺-dependent 15-hydroxyprostaglandin dehydrogenase (type I); PGDH; 11 α ,15-dihydroxy-9-oxoprost-13-enoate:NAD⁺ 15-oxidoreductase; 15-OH-PGDH; 15-hydroxyprostaglandin dehydrogenase; 15-hydroxyprostanoic dehydrogenase; NAD⁺-specific 15-hydroxyprostaglandin dehydrogenase; prostaglandin dehydrogenase; 15-hydroxyprostaglandin dehydrogenase (NAD⁺); (5*Z*,13*E*)-(15*S*)-11 α ,15-dihydroxy-9-oxoprost-13-enoate:NAD⁺ 15-oxidoreductase
Systematic name: (5*Z*,13*E*,15*S*)-11 α ,15-dihydroxy-9-oxoprost-5,13-dienoate:NAD⁺ 15-oxidoreductase
Comments: Acts on prostaglandin E₂, F_{2 α} and B₁, but not on prostaglandin D₂. *cf.* EC 1.1.1.196 15-hydroxyprostaglandin-D dehydrogenase (NADP⁺) and EC 1.1.1.197 15-hydroxyprostaglandin dehydrogenase (NADP⁺).
References: [104, 422, 2393, 2395]

[EC 1.1.1.141 created 1972]

EC 1.1.1.142

Accepted name: D-pinitol dehydrogenase
Reaction: 1D-3-*O*-methyl-*chiro*-inositol + NADP⁺ = 2D-5-*O*-methyl-2,3,5/4,6-pentahydroxycyclohexanone + NADPH + H⁺
Other name(s): 5D-5-*O*-methyl-*chiro*-inositol:NADP⁺ oxidoreductase
Systematic name: 1D-3-*O*-methyl-*chiro*-inositol:NADP⁺ oxidoreductase
References: [3603]

[EC 1.1.1.142 created 1972]

EC 1.1.1.143

Accepted name: sequoyitol dehydrogenase

Reaction: 5-*O*-methyl-*myo*-inositol + NAD⁺ = 2D-5-*O*-methyl-2,3,5/4,6-pentahydroxycyclohexanone + NADH + H⁺

Other name(s): D-pinitol dehydrogenase

Systematic name: 5-*O*-methyl-*myo*-inositol:NAD⁺ oxidoreductase

References: [3603]

[EC 1.1.1.143 created 1972]

EC 1.1.1.144

Accepted name: perillyl-alcohol dehydrogenase

Reaction: perillyl alcohol + NAD⁺ = perillyl aldehyde + NADH + H⁺

Other name(s): perillyl alcohol dehydrogenase

Systematic name: perillyl-alcohol:NAD⁺ oxidoreductase

Comments: Oxidizes a number of primary alcohols with the alcohol group allylic to an endocyclic double bond and a 6-membered ring, either aromatic or hydroaromatic.

References: [201]

[EC 1.1.1.144 created 1972]

EC 1.1.1.145

Accepted name: 3β-hydroxy-Δ⁵-steroid dehydrogenase

Reaction: a 3β-hydroxy-Δ⁵-steroid + NAD⁺ = a 3-oxo-Δ⁵-steroid + NADH + H⁺

Other name(s): progesterone reductase; Δ⁵-3β-hydroxysteroid dehydrogenase; 3β-hydroxy-5-ene steroid dehydrogenase; 3β-hydroxy steroid dehydrogenase/isomerase; 3β-hydroxy-Δ⁵-C₂₇-steroid dehydrogenase/isomerase; 3β-hydroxy-Δ⁵-C₂₇-steroid oxidoreductase; 3β-hydroxy-5-ene-steroid oxidoreductase; steroid-Δ⁵-3β-ol dehydrogenase; 3β-HSDH; 5-ene-3-β-hydroxysteroid dehydrogenase; 3β-hydroxy-5-ene-steroid dehydrogenase

Systematic name: 3β-hydroxy-Δ⁵-steroid:NAD⁺ 3-oxidoreductase

Comments: This activity is found in several bifunctional enzymes that catalyse the oxidative conversion of Δ⁵-3-hydroxy steroids to a Δ⁴-3-oxo configuration. This conversion is carried out in two separate, sequential reactions; in the first reaction, which requires NAD⁺, the enzyme catalyses the dehydrogenation of the 3β-hydroxy steroid to a 3-oxo intermediate. In the second reaction the reduced coenzyme, which remains attached to the enzyme, activates the isomerization of the Δ⁵ form to a Δ⁴ form (*cf.* EC 5.3.3.1, steroid Δ-isomerase). Substrates include dehydroepiandrosterone (which is converted into androst-5-ene-3,17-dione), pregnenolone (converted to progesterone) and cholest-5-en-3-one, an intermediate of cholesterol degradation.

References: [623, 2233, 3053]

[EC 1.1.1.145 created 1972]

EC 1.1.1.146

Accepted name: 11β-hydroxysteroid dehydrogenase

Reaction: an 11β-hydroxysteroid + NADP⁺ = an 11-oxosteroid + NADPH + H⁺

Other name(s): corticosteroid 11β-dehydrogenase; β-hydroxysteroid dehydrogenase; 11β-hydroxy steroid dehydrogenase; corticosteroid 11-reductase; dehydrogenase, 11β-hydroxy steroid

Systematic name: 11β-hydroxysteroid:NADP⁺ 11-oxidoreductase

References: [32, 509, 2327, 3314]

[EC 1.1.1.146 created 1972]

EC 1.1.1.147

Accepted name: 16α-hydroxysteroid dehydrogenase

Reaction: a 16 α -hydroxysteroid + NAD(P)⁺ = a 16-oxosteroid + NAD(P)H + H⁺
Other name(s): 16 α -hydroxy steroid dehydrogenase
Systematic name: 16 α -hydroxysteroid:NAD(P)⁺ 16-oxidoreductase
References: [2763]

[EC 1.1.1.147 created 1972]

EC 1.1.1.148

Accepted name: estradiol 17 α -dehydrogenase
Reaction: estradiol-17 α + NAD(P)⁺ = estrone + NAD(P)H + H⁺
Other name(s): 17 α -estradiol dehydrogenase; 17 α -hydroxy steroid dehydrogenase; 17 α -hydroxy steroid oxidoreductase; 17 α -hydroxysteroid oxidoreductase; estradiol 17 α -oxidoreductase
Systematic name: 17 α -hydroxysteroid:NAD(P)⁺ 17-oxidoreductase
References: [3501]

[EC 1.1.1.148 created 1972]

EC 1.1.1.149

Accepted name: 20 α -hydroxysteroid dehydrogenase
Reaction: 17 α ,20 α -dihydroxypregn-4-en-3-one + NAD(P)⁺ = 17 α -hydroxypregesterone + NAD(P)H + H⁺
Other name(s): 20 α -hydroxy steroid dehydrogenase; 20 α -HSD; 20 α -HSDH
Systematic name: 20 α -hydroxysteroid:NAD(P)⁺ 20-oxidoreductase
Comments: *Re*-specific with respect to NAD(P)⁺ (*cf.* EC 1.1.1.62 17 β -estradiol 17-dehydrogenase).
References: [3860, 4065]

[EC 1.1.1.149 created 1972, deleted 1983, reinstated 1986]

EC 1.1.1.150

Accepted name: 21-hydroxysteroid dehydrogenase (NAD⁺)
Reaction: pregnan-21-ol + NAD⁺ = pregnan-21-al + NADH + H⁺
Other name(s): 21-hydroxysteroid dehydrogenase (NAD)
Systematic name: 21-hydroxysteroid:NAD⁺ 21-oxidoreductase
Comments: Acts on a number of 21-hydroxycorticosteroids.
References: [2867]

[EC 1.1.1.150 created 1972]

EC 1.1.1.151

Accepted name: 21-hydroxysteroid dehydrogenase (NADP⁺)
Reaction: pregnan-21-ol + NADP⁺ = pregnan-21-al + NADPH + H⁺
Other name(s): 21-hydroxy steroid dehydrogenase; 21-hydroxy steroid (nicotinamide adenine dinucleotide phosphate) dehydrogenase; 21-hydroxy steroid dehydrogenase (nicotinamide adenine dinucleotide phosphate); NADP-21-hydroxysteroid dehydrogenase; 21-hydroxysteroid dehydrogenase (NADP)
Systematic name: 21-hydroxysteroid:NADP⁺ 21-oxidoreductase
Comments: Acts on a number of 21-hydroxycorticosteroids.
References: [2867]

[EC 1.1.1.151 created 1972]

EC 1.1.1.152

Accepted name: 3 α -hydroxy-5 β -androstane-17-one 3 α -dehydrogenase
Reaction: 3 α -hydroxy-5 β -androstane-17-one + NAD⁺ = 5 β -androstane-3,17-dione + NADH + H⁺

Other name(s): etiocholanolone 3 α -dehydrogenase; etiocholanolone 3 α -dehydrogenase; 3 α -hydroxy-5 β -steroid dehydrogenase
Systematic name: 3 α -hydroxy-5 β -steroid:NAD⁺ 3-oxidoreductase
References: [3551]

[EC 1.1.1.152 created 1972]

EC 1.1.1.153

Accepted name: sepiapterin reductase (*L-erythro*-7,8-dihydrobiopterin forming)
Reaction: (1) *L-erythro*-7,8-dihydrobiopterin + NADP⁺ = sepiapterin + NADPH + H⁺
(2) *L-erythro*-tetrahydrobiopterin + 2 NADP⁺ = 6-pyruvoyl-5,6,7,8-tetrahydropterin + 2 NADPH + 2 H⁺
Other name(s): SR
Systematic name: *L-erythro*-7,8-dihydrobiopterin:NADP⁺ oxidoreductase
Comments: This enzyme catalyses the final step in the *de novo* synthesis of tetrahydrobiopterin from GTP. The enzyme, which is found in higher animals and some fungi and bacteria, produces the *erythro* form of tetrahydrobiopterin. *cf.* EC 1.1.1.325, sepiapterin reductase (*L-threo*-7,8-dihydrobiopterin forming).
References: [2025, 2696, 4591, 2111]

[EC 1.1.1.153 created 1972, modified 2012]

EC 1.1.1.154

Accepted name: ureidoglycolate dehydrogenase
Reaction: (*S*)-ureidoglycolate + NAD(P)⁺ = oxalureate + NAD(P)H + H⁺
Systematic name: (*S*)-ureidoglycolate:NAD(P)⁺ oxidoreductase
References: [4402]

[EC 1.1.1.154 created 1976]

[1.1.1.155 Deleted entry. *homoisocitrate dehydrogenase. The enzyme is identical to EC 1.1.1.87, homoisocitrate dehydrogenase*]

[EC 1.1.1.155 created 1976, deleted 2004]

EC 1.1.1.156

Accepted name: glycerol 2-dehydrogenase (NADP⁺)
Reaction: glycerol + NADP⁺ = glycerone + NADPH + H⁺
Other name(s): dihydroxyacetone reductase; dihydroxyacetone (reduced nicotinamide adenine dinucleotide phosphate) reductase; dihydroxyacetone reductase (NADPH); DHA oxidoreductase; glycerol 2-dehydrogenase (NADP)
Systematic name: glycerol:NADP⁺ 2-oxidoreductase (glycerone-forming)
References: [281]

[EC 1.1.1.156 created 1976]

EC 1.1.1.157

Accepted name: 3-hydroxybutyryl-CoA dehydrogenase
Reaction: (*S*)-3-hydroxybutanoyl-CoA + NADP⁺ = 3-acetoacetyl-CoA + NADPH + H⁺
Other name(s): β -hydroxybutyryl coenzyme A dehydrogenase; L(+)-3-hydroxybutyryl-CoA dehydrogenase; BHBD; dehydrogenase, L-3-hydroxybutyryl coenzyme A (nicotinamide adenine dinucleotide phosphate); L-(+)-3-hydroxybutyryl-CoA dehydrogenase; β -hydroxybutyryl-CoA dehydrogenase
Systematic name: (*S*)-3-hydroxybutanoyl-CoA:NADP⁺ oxidoreductase
References: [2598]

[EC 1.1.1.157 created 1976]

[1.1.1.158 Transferred entry. *UDP-N-acetylmuramate dehydrogenase*. Now EC 1.3.1.98, *UDP-N-acetylmuramate dehydrogenase*]

[EC 1.1.1.158 created 1976, modified 1983, modified 2002, deleted 2013]

EC 1.1.1.159

Accepted name: 7 α -hydroxysteroid dehydrogenase
Reaction: cholate + NAD⁺ = 3 α ,12 α -dihydroxy-7-oxo-5 β -cholan-24-oate + NADH + H⁺
Other name(s): 7 α -hydroxy steroid dehydrogenase; 7 α -HSDH
Systematic name: 7 α -hydroxysteroid:NAD⁺ 7-oxidoreductase
Comments: Catalyses the oxidation of the 7 α -hydroxy group of bile acids and alcohols both in their free and conjugated forms. The *Bacteroides fragilis* and *Clostridium* enzymes can also utilize NADP⁺.
References: [1552, 2583, 2585, 2586]

[EC 1.1.1.159 created 1976, modified 1980]

EC 1.1.1.160

Accepted name: dihydrobunolol dehydrogenase
Reaction: (\pm)-5-[(*tert*-butylamino)-2'-hydroxypropoxy]-1,2,3,4-tetrahydro-1-naphthol + NADP⁺ = (\pm)-5-[(*tert*-butylamino)-2'-hydroxypropoxy]-3,4-dihydro-1(2*H*)-naphthalenone + NADPH + H⁺
Other name(s): bunolol reductase
Systematic name: (\pm)-5-[(*tert*-butylamino)-2'-hydroxypropoxy]-1,2,3,4-tetrahydro-1-naphthol:NADP⁺ oxidoreductase
Comments: Also acts, more slowly, with NAD⁺.
References: [2411]

[EC 1.1.1.160 created 1976]

[1.1.1.161 Deleted entry. *cholestanetetraol 26-dehydrogenase*. The activity is part of EC 1.14.13.15, *cholestanetriol 26-monoxygenase*]

[EC 1.1.1.161 created 1976, deleted 2012]

EC 1.1.1.162

Accepted name: erythrulose reductase
Reaction: D-threitol + NADP⁺ = D-erythrulose + NADPH + H⁺
Other name(s): D-erythrulose reductase; erythritol:NADP⁺ oxidoreductase
Systematic name: D-threitol:NADP⁺ oxidoreductase
Comments: NAD⁺ is also utilized, but more slowly.
References: [4368, 4366]

[EC 1.1.1.162 created 1976]

EC 1.1.1.163

Accepted name: cyclopentanol dehydrogenase
Reaction: cyclopentanol + NAD⁺ = cyclopentanone + NADH + H⁺
Systematic name: cyclopentanol:NAD⁺ oxidoreductase
Comments: 4-Methylcyclohexanol and cyclohexanol can also act as substrates.
References: [1408, 1859]

[EC 1.1.1.163 created 1976]

EC 1.1.1.164

- Accepted name:** hexadecanol dehydrogenase
Reaction: hexadecanol + NAD⁺ = hexadecanal + NADH + H⁺
Systematic name: hexadecanol:NAD⁺ oxidoreductase
Comments: The liver enzyme acts on long-chain alcohols from C₈ to C₁₆. The *Euglena* enzyme also oxidizes the corresponding aldehydes to fatty acids.
References: [2211, 4038]

[EC 1.1.1.164 created 1976]

EC 1.1.1.165

- Accepted name:** 2-alkyn-1-ol dehydrogenase
Reaction: 2-butyne-1,4-diol + NAD⁺ = 4-hydroxy-2-butyral + NADH + H⁺
Systematic name: 2-butyne-1,4-diol:NAD⁺ 1-oxidoreductase
Comments: Acts on a variety of 2-alkyn-1-ols, and also on 1,4-butanediol. NADP⁺ also acts as acceptor, but more slowly.
References: [2847]

[EC 1.1.1.165 created 1976]

EC 1.1.1.166

- Accepted name:** hydroxycyclohexanecarboxylate dehydrogenase
Reaction: (1*S*,3*R*,4*S*)-3,4-dihydroxycyclohexane-1-carboxylate + NAD⁺ = (1*S*,4*S*)-4-hydroxy-3-oxocyclohexane-1-carboxylate + NADH + H⁺
Other name(s): dihydroxycyclohexanecarboxylate dehydrogenase; (-)-*t*-3,*t*-4-dihydroxycyclohexane-*c*-1-carboxylate-NAD⁺ oxidoreductase
Systematic name: (1*S*,3*R*,4*S*)-3,4-dihydroxycyclohexane-1-carboxylate:NAD⁺ 3-oxidoreductase
Comments: Acts on hydroxycyclohexanecarboxylates that have an equatorial carboxy group at C-1, an axial hydroxy group at C-3 and an equatorial hydroxy or carbonyl group at C-4, including (-)-quininate and (-)-shikimate.
References: [4608]

[EC 1.1.1.166 created 1976]

EC 1.1.1.167

- Accepted name:** hydroxymalonate dehydrogenase
Reaction: hydroxymalonate + NAD⁺ = oxomalonate + NADH + H⁺
Systematic name: hydroxymalonate:NAD⁺ oxidoreductase
References: [1958]

[EC 1.1.1.167 created 1976]

EC 1.1.1.168

- Accepted name:** 2-dehydropantolactone reductase (*Re*-specific)
Reaction: (*R*)-pantolactone + NADP⁺ = 2-dehydropantolactone + NADPH + H⁺
Other name(s): 2-oxopantoyl lactone reductase; ketopantoyl lactone reductase; 2-ketopantoyl lactone reductase; 2-dehydropantoyl-lactone reductase (*A*-specific); (*R*)-pantolactone:NADP⁺ oxidoreductase (*A*-specific); 2-dehydropantolactone reductase (*A*-specific)
Systematic name: (*R*)-pantolactone:NADP⁺ oxidoreductase (*Re*-specific)
Comments: The yeast enzyme differs from that from *Escherichia coli* [EC 1.1.1.214 2-dehydropantolactone reductase (*Si*-specific)], which is specific for the *Si*-face of NADP⁺, and in receptor requirements from EC 1.1.99.26 3-hydroxycyclohexanone dehydrogenase.
References: [2120, 4627]

[EC 1.1.1.168 created 1976, modified 1986, modified 1999]

EC 1.1.1.169

Accepted name: 2-dehydropantoate 2-reductase
Reaction: (R)-pantoate + NADP⁺ = 2-dehydropantoate + NADPH + H⁺
Other name(s): 2-oxopantoate reductase; 2-ketopantoate reductase; 2-ketopantoic acid reductase; ketopantoate reductase; ketopantoic acid reductase
Systematic name: (R)-pantoate:NADP⁺ 2-oxidoreductase
References: [2120]

[EC 1.1.1.169 created 1976]

EC 1.1.1.170

Accepted name: 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating)
Reaction: a 3 β -hydroxysteroid-4 α -carboxylate + NAD(P)⁺ = a 3-oxosteroid + CO₂ + NAD(P)H
Other name(s): 3 β -hydroxy-4 β -methylcholestene-carboxylate 3-dehydrogenase (decarboxylating); 3 β -hydroxy-4 β -methylcholestenoate dehydrogenase; sterol 4 α -carboxylic decarboxylase; sterol-4 α -carboxylate 3-dehydrogenase (decarboxylating) (ambiguous); ERG26 (gene name); NSDHL (gene name)
Systematic name: 3 β -hydroxysteroid-4 α -carboxylate:NAD(P)⁺ 3-oxidoreductase (decarboxylating)
Comments: The enzyme participates in the biosynthesis of several important sterols such as ergosterol and cholesterol. It is part of a three enzyme system that removes methyl groups from the C-4 position of steroid molecules. The first enzyme, EC 1.14.18.9, 4 α -methylsterol monooxygenase, catalyses three successive oxidations of the methyl group, resulting in a carboxyl group; the second enzyme, EC 1.1.1.170, catalyses an oxidative decarboxylation that results in a reduction of the 3 β -hydroxy group at the C-3 carbon to an oxo group; and the last enzyme, EC 1.1.1.270, 3 β -hydroxysteroid 3-dehydrogenase, reduces the 3-oxo group back to a 3 β -hydroxyl. If a second methyl group remains at the C-4 position, this enzyme also catalyses its epimerization from 4 β to 4 α orientation, so it could serve as a substrate for a second round of demethylation. *cf.* EC 1.1.1.418, plant 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating).
References: [3831, 3430, 420, 1247, 528]

[EC 1.1.1.170 created 1978, modified 2002, modified 2012, modified 2019]

[1.1.1.171 *Transferred entry. methylenetetrahydrofolate reductase (NADPH). Now EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]*]

[EC 1.1.1.171 created 1978, deleted 1984]

EC 1.1.1.172

Accepted name: 2-oxoadipate reductase
Reaction: 2-hydroxyadipate + NAD⁺ = 2-oxoadipate + NADH + H⁺
Other name(s): 2-ketoadipate reductase; α -ketoadipate reductase; 2-ketoadipate reductase
Systematic name: 2-hydroxyadipate:NAD⁺ 2-oxidoreductase
References: [4090]

[EC 1.1.1.172 created 1978]

EC 1.1.1.173

Accepted name: L-rhamnose 1-dehydrogenase
Reaction: L-rhamnofuranose + NAD⁺ = L-rhamno-1,4-lactone + NADH + H⁺
Systematic name: L-rhamnofuranose:NAD⁺ 1-oxidoreductase
References: [3521, 3522]

[EC 1.1.1.173 created 1978]

EC 1.1.1.174

Accepted name: cyclohexane-1,2-diol dehydrogenase
Reaction: *trans*-cyclohexane-1,2-diol + NAD⁺ = 2-hydroxycyclohexan-1-one + NADH + H⁺
Systematic name: *trans*-cyclohexane-1,2-diol:NAD⁺ 1-oxidoreductase
Comments: Also oxidizes, more slowly, the *cis* isomer and 2-hydroxycyclohexanone.
References: [837]

[EC 1.1.1.174 created 1978]

EC 1.1.1.175

Accepted name: D-xylose 1-dehydrogenase
Reaction: D-xylose + NAD⁺ = D-xylonolactone + NADH + H⁺
Other name(s): NAD-D-xylose dehydrogenase; D-xylose dehydrogenase; (NAD)-linked D-xylose dehydrogenase
Systematic name: D-xylose:NAD⁺ 1-oxidoreductase
References: [4744]

[EC 1.1.1.175 created 1978]

EC 1.1.1.176

Accepted name: 12 α -hydroxysteroid dehydrogenase
Reaction: cholate + NADP⁺ = 3 α ,7 α -dihydroxy-12-oxo-5 β -cholan-24-oate + NADPH + H⁺
Other name(s): 12 α -hydroxy steroid dehydrogenase; NAD⁺-dependent 12 α -hydroxysteroid dehydrogenase; NADP⁺-12 α -hydroxysteroid dehydrogenase
Systematic name: 12 α -hydroxysteroid:NADP⁺ 12-oxidoreductase
Comments: Catalyses the oxidation of the 12 α -hydroxy group of bile acids, both in their free and conjugated form. Also acts on bile alcohols.
References: [2582, 2619]

[EC 1.1.1.176 created 1978]

EC 1.1.1.177

Accepted name: glycerol-3-phosphate 1-dehydrogenase (NADP⁺)
Reaction: *sn*-glycerol 3-phosphate + NADP⁺ = D-glyceraldehyde 3-phosphate + NADPH + H⁺
Other name(s): glycerol phosphate (nicotinamide adenine dinucleotide phosphate) dehydrogenase; L-glycerol 3-phosphate:NADP⁺ oxidoreductase; glycerin-3-phosphate dehydrogenase; NADPH-dependent glycerin-3-phosphate dehydrogenase; NADP-specific glycerol 3-phosphate 1-dehydrogenase
Systematic name: *sn*-glycerol-3-phosphate:NADP⁺ 1-oxidoreductase
References: [1346, 4665]

[EC 1.1.1.177 created 1980, modified 1980]

EC 1.1.1.178

Accepted name: 3-hydroxy-2-methylbutyryl-CoA dehydrogenase
Reaction: (2*S*,3*S*)-3-hydroxy-2-methylbutanoyl-CoA + NAD⁺ = 2-methylacetoacetyl-CoA + NADH + H⁺
Other name(s): 2-methyl-3-hydroxybutyryl coenzyme A dehydrogenase; 2-methyl-3-hydroxybutyryl coenzyme A dehydrogenase; 2-methyl-3-hydroxy-butyl CoA dehydrogenase
Systematic name: (2*S*,3*S*)-3-hydroxy-2-methylbutanoyl-CoA:NAD⁺ oxidoreductase
Comments: Also acts, more slowly, on (2*S*,3*S*)-2-hydroxy-3-methylpentanoyl-CoA.
References: [724]

[EC 1.1.1.178 created 1981]

EC 1.1.1.179

- Accepted name:** D-xylose 1-dehydrogenase (NADP⁺, D-xylono-1,5-lactone-forming)
Reaction: D-xylose + NADP⁺ = D-xylono-1,5-lactone + NADPH + H⁺
Other name(s): D-xylose (nicotinamide adenine dinucleotide phosphate) dehydrogenase (ambiguous); D-xylose-NADP dehydrogenase (ambiguous); D-xylose:NADP⁺ oxidoreductase (ambiguous); D-xylose 1-dehydrogenase (NADP) (ambiguous)
Systematic name: D-xylose:NADP⁺ 1-oxidoreductase (D-xylono-1,5-lactone-forming)
Comments: The enzyme, characterized from pig arterial vessels and eye lens, also acts, more slowly, on L-arabinose and D-ribose. *cf.* EC 1.1.1.424, D-xylose 1-dehydrogenase (NADP⁺, D-xylono-1,4-lactone-forming).
References: [4645, 4646]

[EC 1.1.1.179 created 1982, modified 2020]

[1.1.1.180 Deleted entry. mannonate dehydrogenase (NAD(P)⁺). Now included with EC 1.1.1.131 mannuronate reductase]

[EC 1.1.1.180 created 1983, deleted 1984]

EC 1.1.1.181

- Accepted name:** cholest-5-ene-3β,7α-diol 3β-dehydrogenase
Reaction: cholest-5-ene-3β,7α-diol + NAD⁺ = 7α-hydroxycholest-4-en-3-one + NADH + H⁺
Other name(s): 3β-hydroxy-Δ⁵-C₂₇-steroid oxidoreductase (ambiguous)
Systematic name: cholest-5-ene-3β,7α-diol:NAD⁺ 3-oxidoreductase
Comments: Highly specific for 3β,7α-dihydroxy-C₂₇-steroids with Δ⁵-double bond.
References: [4623, 3769]

[EC 1.1.1.181 created 1983]

[1.1.1.182 Deleted entry. fenchol dehydrogenase. Now included with EC 1.1.1.198 (+)-borneol dehydrogenase, EC 1.1.1.227 (-)-borneol dehydrogenase and EC 1.1.1.228 (+)-sabinol dehydrogenase]

[EC 1.1.1.182 created 1983, deleted 1990]

EC 1.1.1.183

- Accepted name:** geraniol dehydrogenase (NADP⁺)
Reaction: geraniol + NADP⁺ = geranial + NADPH + H⁺
Systematic name: geraniol:NADP⁺ oxidoreductase
Comments: Also acts, more slowly on farnesol but not on nerol. The enzyme produces a mixture known as citral, which includes geranial and neral. It is still not known whether neral is produced directly by the enzyme, or by isomerization of geranial.
References: [3360, 3794, 3636]

[EC 1.1.1.183 created 1983]

EC 1.1.1.184

- Accepted name:** carbonyl reductase (NADPH)
Reaction: R-CHOH-R' + NADP⁺ = R-CO-R' + NADPH + H⁺
Other name(s): aldehyde reductase 1; prostaglandin 9-ketoreductase; xenobiotic ketone reductase; NADPH-dependent carbonyl reductase; ALR₃; carbonyl reductase; nonspecific NADPH-dependent carbonyl reductase; carbonyl reductase (NADPH₂)
Systematic name: secondary-alcohol:NADP⁺ oxidoreductase
Comments: Acts on a wide range of carbonyl compounds, including quinones, aromatic aldehydes, ketoaldehydes, daunorubicin and prostaglandins E and F, reducing them to the corresponding alcohol. *Si*-specific with respect to NADPH [*cf.* EC 1.1.1.2 alcohol dehydrogenase (NADP⁺)].
References: [39, 2494, 4589]

[EC 1.1.1.184 created 1983]

EC 1.1.1.185

Accepted name: L-glycol dehydrogenase
Reaction: an L-glycol + NAD(P)⁺ = a 2-hydroxycarbonyl compound + NAD(P)H + H⁺
Other name(s): glycol (nicotinamide adenine dinucleotide (phosphate)) dehydrogenase; L-(+)-glycol:NAD(P) oxidoreductase; L-glycol:NAD(P) dehydrogenase
Systematic name: L-glycol:NAD(P)⁺ oxidoreductase
Comments: The 2-hydroxycarbonyl compound formed can be further oxidized to a vicinal dicarbonyl compound. In the reverse direction, vicinal diketones, glyceraldehyde, glyoxal, methylglyoxal, 2-oxo-hydroxyketones and 2-ketoacid esters can be reduced.
References: [303]

[EC 1.1.1.185 created 1984]

EC 1.1.1.186

Accepted name: dTDP-galactose 6-dehydrogenase
Reaction: dTDP-D-galactose + 2 NADP⁺ + H₂O = dTDP-D-galacturonate + 2 NADPH + 2 H⁺
Other name(s): thymidine-diphosphate-galactose dehydrogenase
Systematic name: dTDP-D-galactose:NADP⁺ 6-oxidoreductase
References: [2009]

[EC 1.1.1.186 created 1984, modified 2002]

EC 1.1.1.187

Accepted name: GDP-4-dehydro-D-rhamnose reductase
Reaction: (1) GDP- α -D-rhamnose + NAD(P)⁺ = GDP-4-dehydro- α -D-rhamnose + NAD(P)H + H⁺
(2) GDP-6-deoxy- α -D-talose + NAD(P)⁺ = GDP-4-dehydro- α -D-rhamnose + NAD(P)H + H⁺
Other name(s): GDP-4-keto-6-deoxy-D-mannose reductase; GDP-4-keto-D-rhamnose reductase; guanosine diphosphate-4-keto-D-rhamnose reductase; GDP-6-deoxy-D-mannose:NAD(P)⁺ 4-oxidoreductase; GDP-6-deoxy- α -D-mannose:NAD(P)⁺ 4-oxidoreductase
Systematic name: GDP-4-dehydro- α -D-rhamnose:NAD(P)⁺ 4-oxidoreductase
Comments: The enzyme, which operates in the opposite direction to that shown, forms a mixture of GDP- α -D-rhamnose and its C-4 epimer, GDP-6-deoxy- α -D-talose. *cf.* EC 1.1.1.281, GDP-4-dehydro-6-deoxy-D-mannose reductase and EC 1.1.1.135, GDP-6-deoxy-D-talose 4-dehydrogenase.
References: [215, 4640]

[EC 1.1.1.187 created 1984]

EC 1.1.1.188

Accepted name: prostaglandin-F synthase
Reaction: (5Z,13E)-(15S)-9 α ,11 α ,15-trihydroxyprosta-5,13-dienoate + NADP⁺ = (5Z,13E)-(15S)-9 α ,15-dihydroxy-11-oxoprosta-5,13-dienoate + NADPH + H⁺
Other name(s): prostaglandin-D₂ 11-reductase; reductase, 15-hydroxy-11-oxoprostaglandin; PGD₂ 11-ketoreductase; PGF_{2 α} synthetase; prostaglandin 11-ketoreductase; prostaglandin D₂-ketoreductase; prostaglandin F synthase; prostaglandin F synthetase; synthetase, prostaglandin F_{2 α} ; PGF synthetase; NADPH-dependent prostaglandin D₂ 11-keto reductase; prostaglandin 11-keto reductase
Systematic name: (5Z,13E)-(15S)-9 α ,11 α ,15-trihydroxyprosta-5,13-dienoate:NADP⁺ 11-oxidoreductase
Comments: Reduces prostaglandin D₂ and prostaglandin H₂ to prostaglandin F₂; prostaglandin D₂ is not an intermediate in the reduction of prostaglandin H₂. Also catalyses the reduction of a number of carbonyl compounds, such as 9,10-phenanthroquinone and 4-nitroacetophenone.
References: [3491, 4553, 4555, 4661, 4662]

[EC 1.1.1.188 created 1984, modified 1989, modified 1990]

EC 1.1.1.189

- Accepted name:** prostaglandin-E₂ 9-reductase
Reaction: (5Z,13E)-(15S)-9 α ,11 α ,15-trihydroxyprosta-5,13-dienoate + NADP⁺ = (5Z,13E)-(15S)-11 α ,15-dihydroxy-9-oxoprosta-5,13-dienoate + NADPH + H⁺
Other name(s): PGE₂-9-OR; reductase, 15-hydroxy-9-oxoprostaglandin; 9-keto-prostaglandin E₂ reductase; 9-ketoprostaglandin reductase; PGE-9-ketoreductase; PGE₂ 9-oxoreductase; PGE₂-9-ketoreductase; prostaglandin 9-ketoreductase; prostaglandin E 9-ketoreductase; prostaglandin E₂-9-oxoreductase
Systematic name: (5Z,13E)-(15S)-9 α ,11 α ,15-trihydroxyprosta-5,13-dienoate:NADP⁺ 9-oxidoreductase
Comments: Reduces prostaglandin E₂ to prostaglandin F₂ α . A number of other 9-oxo- and 15-oxo-prostaglandin derivatives can also be reduced to the corresponding hydroxy compounds. May be identical with EC 1.1.1.197 15-hydroxyprostaglandin dehydrogenase (NADP⁺).
References: [2394, 3721, 4166, 4561]

[EC 1.1.1.189 created 1984, modified 1989]

EC 1.1.1.190

- Accepted name:** indole-3-acetaldehyde reductase (NADH)
Reaction: (indol-3-yl)ethanol + NAD⁺ = (indol-3-yl)acetaldehyde + NADH + H⁺
Other name(s): indoleacetaldehyde reductase; indole-3-acetaldehyde reductase (NADH); indole-3-ethanol:NAD⁺ oxidoreductase
Systematic name: (indol-3-yl)ethanol:NAD⁺ oxidoreductase
References: [460]

[EC 1.1.1.190 created 1984]

EC 1.1.1.191

- Accepted name:** indole-3-acetaldehyde reductase (NADPH)
Reaction: (indol-3-yl)ethanol + NADP⁺ = (indol-3-yl)acetaldehyde + NADPH + H⁺
Other name(s): indoleacetaldehyde (reduced nicotinamide adenine dinucleotide phosphate) reductase; indole-3-acetaldehyde reductase (NADPH); indole-3-ethanol:NADP⁺ oxidoreductase
Systematic name: (indol-3-yl)ethanol:NADP⁺ oxidoreductase
References: [460]

[EC 1.1.1.191 created 1984]

EC 1.1.1.192

- Accepted name:** long-chain-alcohol dehydrogenase
Reaction: a long-chain alcohol + 2 NAD⁺ + H₂O = a long-chain carboxylate + 2 NADH + 2 H⁺
Other name(s): long-chain alcohol dehydrogenase; fatty alcohol oxidoreductase
Systematic name: long-chain-alcohol:NAD⁺ oxidoreductase
Comments: Hexadecanol is a good substrate.
References: [2398]

[EC 1.1.1.192 created 1984]

EC 1.1.1.193

- Accepted name:** 5-amino-6-(5-phosphoribosylamino)uracil reductase
Reaction: 5-amino-6-(5-phospho-D-ribitylamino)uracil + NADP⁺ = 5-amino-6-(5-phospho-D-ribosylamino)uracil + NADPH + H⁺
Other name(s): aminodioxyposphoribosylaminopyrimidine reductase
Systematic name: 5-amino-6-(5-phospho-D-ribitylamino)uracil:NADP⁺ 1'-oxidoreductase
References: [502]

[EC 1.1.1.193 created 1984, modified 2011]

EC 1.1.1.194

- Accepted name:** coniferyl-alcohol dehydrogenase
Reaction: coniferyl alcohol + NADP⁺ = coniferyl aldehyde + NADPH + H⁺
Other name(s): CAD (ambiguous)
Systematic name: coniferyl-alcohol:NADP⁺ oxidoreductase
Comments: Specific for coniferyl alcohol; does not act on cinnamyl alcohol, 4-coumaryl alcohol or sinapyl alcohol.
References: [2640, 4691]

[EC 1.1.1.194 created 1984]

EC 1.1.1.195

- Accepted name:** cinnamyl-alcohol dehydrogenase
Reaction: cinnamyl alcohol + NADP⁺ = cinnamaldehyde + NADPH + H⁺
Other name(s): cinnamyl alcohol dehydrogenase; CAD (ambiguous)
Systematic name: cinnamyl-alcohol:NADP⁺ oxidoreductase
Comments: Acts on coniferyl alcohol, sinapyl alcohol, 4-coumaryl alcohol and cinnamyl alcohol (*cf.* EC 1.1.1.194 coniferyl-alcohol dehydrogenase).
References: [3665, 4691, 4692]

[EC 1.1.1.195 created 1984]

EC 1.1.1.196

- Accepted name:** 15-hydroxyprostaglandin-D dehydrogenase (NADP⁺)
Reaction: (5Z,13E)-(15S)-9 α ,15-dihydroxy-11-oxoprost-5,13-dienoate + NADP⁺ = (5Z,13E)-9 α -hydroxy-11,15-dioxoprost-5,13-dienoate + NADPH + H⁺
Other name(s): prostaglandin-D 15-dehydrogenase (NADP); dehydrogenase, prostaglandin D₂; NADP-PGD₂ dehydrogenase; dehydrogenase, 15-hydroxyprostaglandin (nicotinamide adenine dinucleotide phosphate); 15-hydroxy PGD₂ dehydrogenase; 15-hydroxyprostaglandin dehydrogenase (NADP); NADP-dependent 15-hydroxyprostaglandin dehydrogenase; prostaglandin D₂ dehydrogenase; NADP-linked 15-hydroxyprostaglandin dehydrogenase; NADP-specific 15-hydroxyprostaglandin dehydrogenase; NADP-linked prostaglandin D₂ dehydrogenase; 15-hydroxyprostaglandin-D dehydrogenase (NADP)
Systematic name: (5Z,13E)-(15S)-9 α ,15-dihydroxy-11-oxoprost-5,13-dienoate:NADP⁺ 15-oxidoreductase
Comments: Specific for prostaglandins D [*cf.* EC 1.1.1.141 15-hydroxyprostaglandin dehydrogenase (NAD⁺) and EC 1.1.1.197 15-hydroxyprostaglandin dehydrogenase (NADP⁺)].
References: [4554]

[EC 1.1.1.196 created 1984, modified 1990]

EC 1.1.1.197

- Accepted name:** 15-hydroxyprostaglandin dehydrogenase (NADP⁺)
Reaction: (13E)-(15S)-11 α ,15-dihydroxy-9-oxoprost-13-enoate + NADP⁺ = (13E)-11 α -hydroxy-9,15-dioxoprost-13-enoate + NADPH + H⁺
Other name(s): NADP-dependent 15-hydroxyprostaglandin dehydrogenase; NADP-linked 15-hydroxyprostaglandin dehydrogenase; NADP-specific 15-hydroxyprostaglandin dehydrogenase; type II 15-hydroxyprostaglandin dehydrogenase; 15-hydroxyprostaglandin dehydrogenase (NADP)
Systematic name: (13E)-(15S)-11 α ,15-dihydroxy-9-oxoprost-13-enoate:NADP⁺ 15-oxidoreductase
Comments: Acts on prostaglandins E₂, F_{2 α} and B₁, but not on prostaglandin D₂ [*cf.* EC 1.1.1.141 15-hydroxyprostaglandin dehydrogenase (NAD⁺) and EC 1.1.1.196 15-hydroxyprostaglandin-D dehydrogenase (NADP⁺)]. May be identical with EC 1.1.1.189 prostaglandin-E₂ 9-reductase.
References: [2393, 2395]

[EC 1.1.1.197 created 1984]

EC 1.1.1.198

Accepted name: (+)-borneol dehydrogenase
Reaction: (+)-borneol + NAD⁺ = (+)-camphor + NADH + H⁺
Other name(s): bicyclic monoterpenol dehydrogenase
Systematic name: (+)-borneol:NAD⁺ oxidoreductase
Comments: NADP⁺ can also act, but more slowly.
References: [773, 862]

[EC 1.1.1.198 created 1984, modified 1990 (EC 1.1.1.182 created 1983, part incorporated 1990)]

EC 1.1.1.199

Accepted name: (*S*)-usnate reductase
Reaction: (*6R*)-2-acetyl-6-(3-acetyl-2,4,6-trihydroxy-5-methylphenyl)-3-hydroxy-6-methyl-2,4-cyclohexadien-1-one + NAD⁺ = (*S*)-usnate + NADH + H⁺
Other name(s): L-usnic acid dehydrogenase
Systematic name: reduced-(*S*)-usnate:NAD⁺ oxidoreductase (ether-bond-forming)
References: [1065]

[EC 1.1.1.199 created 1984]

EC 1.1.1.200

Accepted name: aldose-6-phosphate reductase (NADPH)
Reaction: D-sorbitol 6-phosphate + NADP⁺ = D-glucose 6-phosphate + NADPH + H⁺
Other name(s): aldose 6-phosphate reductase; NADP-dependent aldose 6-phosphate reductase; A6PR; aldose-6-*P* reductase; aldose-6-phosphate reductase; alditol 6-phosphate:NADP 1-oxidoreductase; aldose-6-phosphate reductase (NADPH₂)
Systematic name: D-aldose-6-phosphate:NADP⁺ 1-oxidoreductase
Comments: In the reverse reaction, acts also on D-galactose 6-phosphate and, more slowly, on D-mannose 6-phosphate and 2-deoxy-D-glucose 6-phosphate.
References: [3037]

[EC 1.1.1.200 created 1984]

EC 1.1.1.201

Accepted name: 7β-hydroxysteroid dehydrogenase (NADP⁺)
Reaction: a 7β-hydroxysteroid + NADP⁺ = a 7-oxosteroid + NADPH + H⁺
Other name(s): NADP-dependent 7β-hydroxysteroid dehydrogenase; 7β-hydroxysteroid dehydrogenase (NADP)
Systematic name: 7β-hydroxysteroid:NADP⁺ 7-oxidoreductase
Comments: Catalyses the oxidation of the 7β-hydroxy group of bile acids such as ursodeoxycholate.
References: [1667, 2583, 2584]

[EC 1.1.1.201 created 1984]

EC 1.1.1.202

Accepted name: 1,3-propanediol dehydrogenase
Reaction: propane-1,3-diol + NAD⁺ = 3-hydroxypropanal + NADH + H⁺
Other name(s): 3-hydroxypropionaldehyde reductase; 1,3-PD:NAD⁺ oxidoreductase; 1,3-propanediol:NAD⁺ oxidoreductase; 1,3-propanediol dehydrogenase
Systematic name: propane-1,3-diol:NAD⁺ 1-oxidoreductase
References: [2, 1137]

[EC 1.1.1.202 created 1984]

EC 1.1.1.203

- Accepted name:** uronate dehydrogenase
Reaction: (1) β -D-galacturonate + NAD^+ = D-galactaro-1,5-lactone + $\text{NADH} + \text{H}^+$
(2) β -D-glucuronate + NAD^+ = D-glucaro-1,5-lactone + $\text{NADH} + \text{H}^+$
Other name(s): uronate:NAD-oxidoreductase; uronic acid dehydrogenase
Systematic name: uronate:NAD⁺ 1-oxidoreductase
Comments: Requires Mg^{2+} . The enzyme, characterized from the bacterium *Agrobacterium fabrum*, participates in oxidative degradation pathways for galacturonate and glucuronate. The enzyme can only accept the β anomeric form of the substrate [3242]. The 1,5-lactone product is rather stable at cytosolic pH and does not hydrolyse spontaneously at a substantial rate.
References: [2090, 368, 89, 3242]

[EC 1.1.1.203 created 1972 as EC 1.2.1.35, transferred 1984 to EC 1.1.1.203, modified 2014]

[1.1.1.204 Transferred entry. xanthine dehydrogenase. Now EC 1.1.1.4, xanthine dehydrogenase. The enzyme was incorrectly classified as acting on a CH-OH group]

[EC 1.1.1.204 created 1972 as EC 1.2.1.37, transferred 1984 to EC 1.1.1.204, modified 1989, deleted 2004]

EC 1.1.1.205

- Accepted name:** IMP dehydrogenase
Reaction: $\text{IMP} + \text{NAD}^+ + \text{H}_2\text{O} = \text{XMP} + \text{NADH} + \text{H}^+$
Other name(s): inosine-5'-phosphate dehydrogenase; inosinic acid dehydrogenase; inosinate dehydrogenase; inosine 5'-monophosphate dehydrogenase; inosine monophosphate dehydrogenase; IMP oxidoreductase; inosine monophosphate oxidoreductase
Systematic name: IMP:NAD⁺ oxidoreductase
Comments: The enzyme acts on the hydroxy group of the hydrated derivative of the substrate.
References: [2606, 4353]

[EC 1.1.1.205 created 1961 as EC 1.2.1.14, transferred 1984 to EC 1.1.1.205]

EC 1.1.1.206

- Accepted name:** tropinone reductase I
Reaction: $\text{tropine} + \text{NADP}^+ = \text{tropinone} + \text{NADPH} + \text{H}^+$
Other name(s): tropine dehydrogenase; tropinone reductase (ambiguous); TR-I
Systematic name: tropine:NADP⁺ 3 α -oxidoreductase
Comments: Also oxidizes other tropane-3 α -ols, but not the corresponding β -derivatives [2189]. This enzyme along with EC 1.1.1.236, tropinone reductase II, represents a branch point in tropane alkaloid metabolism [961]. Tropine (the product of EC 1.1.1.206) is incorporated into hyoscyamine and scopolamine whereas pseudotropine (the product of EC 1.1.1.236) is the first specific metabolite on the pathway to the calystegines [961]. Both enzymes are always found together in any given tropane-alkaloid-producing species, have a common substrate, tropinone, and are strictly stereospecific [2977].
References: [2189, 749, 2977, 961]

[EC 1.1.1.206 created 1984, modified 2007]

EC 1.1.1.207

- Accepted name:** (-)-menthol dehydrogenase
Reaction: $(-)\text{-menthol} + \text{NADP}^+ = (-)\text{-menthone} + \text{NADPH} + \text{H}^+$
Other name(s): monoterpenoid dehydrogenase
Systematic name: (-)-menthol:NADP⁺ oxidoreductase
Comments: Not identical with EC 1.1.1.208 (+)-neomenthol dehydrogenase. Acts also on a number of other cyclohexanols and cyclohexenols.
References: [2146]

[EC 1.1.1.207 created 1984]

EC 1.1.1.208

Accepted name: (+)-neomenthol dehydrogenase
Reaction: (+)-neomenthol + NADP⁺ = (-)-menthone + NADPH + H⁺
Other name(s): monoterpene dehydrogenase
Systematic name: (+)-neomenthol:NADP⁺ oxidoreductase
Comments: Not identical with EC 1.1.1.207 (-)-menthol dehydrogenase. Acts also on a number of other cyclohexanols and cyclohexenols.
References: [2146]

[EC 1.1.1.208 created 1984]

EC 1.1.1.209

Accepted name: 3(or 17) α -hydroxysteroid dehydrogenase
Reaction: androsterone + NAD(P)⁺ = 5 α -androstane-3,17-dione + NAD(P)H + H⁺
Other name(s): 3(17) α -hydroxysteroid dehydrogenase
Systematic name: 3(or 17) α -hydroxysteroid:NAD(P)⁺ oxidoreductase
Comments: Acts on the 3 α -hydroxy group of androgens of the 5 α -androstane series; and also, more slowly, on the 17 α -hydroxy group of both androgenic and estrogenic substrates (*cf.* EC 1.1.1.51 3(or 17) β -hydroxysteroid dehydrogenase).
References: [2363, 2364]

[EC 1.1.1.209 created 1986]

EC 1.1.1.210

Accepted name: 3 β (or 20 α)-hydroxysteroid dehydrogenase
Reaction: 5 α -androstane-3 β ,17 β -diol + NADP⁺ = 17 β -hydroxy-5 α -androstane-3-one + NADPH + H⁺
Other name(s): progesterone reductase; dehydrogenase, 3 β ,20 α -hydroxy steroid; 3 β ,20 α -hydroxysteroid oxidoreductase
Systematic name: 3 β (or 20 α)-hydroxysteroid:NADP⁺ oxidoreductase
Comments: Also acts on 20 α -hydroxysteroids.
References: [3824]

[EC 1.1.1.210 created 1986]

EC 1.1.1.211

Accepted name: long-chain-3-hydroxyacyl-CoA dehydrogenase
Reaction: a long-chain (*S*)-3-hydroxyacyl-CoA + NAD⁺ = a long-chain 3-oxoacyl-CoA + NADH + H⁺
Other name(s): β -hydroxyacyl-CoA dehydrogenase; long-chain 3-hydroxyacyl coenzyme A dehydrogenase; 3-hydroxyacyl-CoA dehydrogenase; LCHAD
Systematic name: long-chain-(*S*)-3-hydroxyacyl-CoA:NAD⁺ oxidoreductase
Comments: This enzyme was purified from the mitochondrial inner membrane. The enzyme has a preference for long-chain substrates, and activity with a C₁₆ substrate was 6- to 15-fold higher than with a C₄ substrate (*cf.* EC 1.1.1.35 3-hydroxyacyl-CoA dehydrogenase).
References: [1034]

[EC 1.1.1.211 created 1986]

EC 1.1.1.212

Accepted name: 3-oxoacyl-[acyl-carrier-protein] reductase (NADH)

Reaction: a (3*R*)-3-hydroxyacyl-[acyl-carrier protein] + NAD⁺ = a 3-oxoacyl-[acyl-carrier protein] + NADH + H⁺
Other name(s): 3-oxoacyl-[acyl carrier protein] (reduced nicotinamide adenine dinucleotide) reductase; 3-oxoacyl-[acyl-carrier-protein] reductase (NADH); (3*R*)-3-hydroxyacyl-[acyl-carrier-protein]:NAD⁺ oxidoreductase
Systematic name: (3*R*)-3-hydroxyacyl-[acyl-carrier protein]:NAD⁺ oxidoreductase
Comments: Forms part of the fatty acid synthase system in plants. Can be separated from EC 1.1.1.100, 3-oxoacyl-[acyl-carrier-protein] reductase.
References: [575]

[EC 1.1.1.212 created 1986]

EC 1.1.1.213

Accepted name: 3α-hydroxysteroid 3-dehydrogenase (*Re*-specific)
Reaction: a 3α-hydroxysteroid + NAD(P)⁺ = a 3-oxosteroid + NAD(P)H + H⁺
Other name(s): 3α-hydroxysteroid dehydrogenase; 3α-hydroxysteroid:NAD(P)⁺ 3-oxidoreductase (*A*-specific); 3α-hydroxysteroid 3-dehydrogenase (*A*-specific)
Systematic name: 3α-hydroxysteroid:NAD(P)⁺ 3-oxidoreductase (*Re*-specific)
Comments: The enzyme acts on multiple 3α-hydroxysteroids. *Re*-specific with respect to NAD⁺ or NADP⁺ [*cf.* EC 1.1.1.50, 3α-hydroxysteroid 3-dehydrogenase (*Si*-specific)]. Enzymes whose stereo-specificity with respect to NAD⁺ or NADP⁺ is not known are described by EC 1.1.1.357, 3α-hydroxysteroid 3-dehydrogenase.
References: [340, 4304]

[EC 1.1.1.213 created 1986, modified 2012]

EC 1.1.1.214

Accepted name: 2-dehydropantolactone reductase (*Si*-specific)
Reaction: (*R*)-pantolactone + NADP⁺ = 2-dehydropantolactone + NADPH + H⁺
Other name(s): 2-oxopantoyl lactone reductase; 2-ketopantoyl lactone reductase; ketopantoyl lactone reductase; 2-dehydropantoyl-lactone reductase (*B*-specific); (*R*)-pantolactone:NADP⁺ oxidoreductase (*B*-specific); 2-dehydropantolactone reductase (*B*-specific)
Systematic name: (*R*)-pantolactone:NADP⁺ oxidoreductase (*Si*-specific)
Comments: The *Escherichia coli* enzyme differs from that from yeast [EC 1.1.1.168 2-dehydropantolactone reductase (*Re*-specific)], which is specific for the *Re*-face of NADP⁺, and in receptor requirements from EC 1.1.99.26 3-hydroxycyclohexanone dehydrogenase.
References: [4627]

[EC 1.1.1.214 created 1986, modified 1999, modified 2013]

EC 1.1.1.215

Accepted name: gluconate 2-dehydrogenase
Reaction: D-gluconate + NADP⁺ = 2-dehydro-D-gluconate + NADPH + H⁺
Other name(s): 2-keto-D-gluconate reductase; 2-ketogluconate reductase
Systematic name: D-gluconate:NADP⁺ oxidoreductase
Comments: Also acts on L-idonate, D-galactonate and D-xylonate.
References: [14, 661]

[EC 1.1.1.215 created 1989]

EC 1.1.1.216

Accepted name: farnesol dehydrogenase (NADP⁺)
Reaction: (2*E*,6*E*)-farnesol + NADP⁺ = (2*E*,6*E*)-farnesal + NADPH + H⁺

Other name(s): NADP⁺-farnesol dehydrogenase; farnesol (nicotinamide adenine dinucleotide phosphate) dehydrogenase
Systematic name: (2*E*,6*E*)-farnesol:NADP⁺ 1-oxidoreductase
Comments: Also acts, more slowly, on (2*Z*,6*E*)-farnesol, geraniol, citronerol and nerol.
References: [1814]

[EC 1.1.1.216 created 1989]

EC 1.1.1.217

Accepted name: benzyl-2-methyl-hydroxybutyrate dehydrogenase
Reaction: benzyl (2*R*,3*S*)-2-methyl-3-hydroxybutanoate + NADP⁺ = benzyl 2-methyl-3-oxobutanoate + NADPH + H⁺
Other name(s): benzyl 2-methyl-3-hydroxybutyrate dehydrogenase
Systematic name: benzyl-(2*R*,3*S*)-2-methyl-3-hydroxybutanoate:NADP⁺ 3-oxidoreductase
Comments: Also acts on benzyl (2*S*,3*S*)-2-methyl-3-hydroxybutanoate; otherwise highly specific.
References: [1237]

[EC 1.1.1.217 created 1989]

EC 1.1.1.218

Accepted name: morphine 6-dehydrogenase
Reaction: morphine + NAD(P)⁺ = morphinone + NAD(P)H + H⁺
Other name(s): naloxone reductase
Systematic name: morphine:NAD(P)⁺ 6-oxidoreductase
Comments: Also acts on some other alkaloids, including codeine, normorphine and ethylmorphine, but only very slowly on 7,8-saturated derivatives such as dihydromorphine and dihydrocodeine. In the reverse direction, also reduces naloxone to the 6α-hydroxy analogue. Activated by 2-sulfanylethan-1-ol (2-mercaptoethanol).
References: [4750, 4751]

[EC 1.1.1.218 created 1989, modified 1990]

EC 1.1.1.219

Accepted name: dihydroflavonol 4-reductase
Reaction: a (2*R*,3*S*,4*S*)-leucoanthocyanidin + NADP⁺ = a (2*R*,3*R*)-dihydroflavonol + NADPH + H⁺
Other name(s): dihydrokaempferol 4-reductase; dihydromyricetin reductase; NADPH-dihydromyricetin reductase; dihydroquercetin reductase; DFR (gene name); *cis*-3,4-leucopelargonidin:NADP⁺ 4-oxidoreductase; dihydroflavanol 4-reductase (incorrect)
Systematic name: (2*R*,3*S*,4*S*)-leucoanthocyanidin:NADP⁺ 4-oxidoreductase
Comments: This plant enzyme, involved in the biosynthesis of anthocyanidins, is known to act on (+)-dihydrokaempferol, (+)-taxifolin, and (+)-dihydromyricetin, although some enzymes may act only on a subset of these compounds. Each dihydroflavonol is reduced to the corresponding *cis*-flavan-3,4-diol. NAD⁺ can act instead of NADP⁺, but more slowly.
References: [1618, 3999, 1125, 2443]

[EC 1.1.1.219 created 1989, modified 2016]

EC 1.1.1.220

Accepted name: 6-pyruvoyltetrahydropterin 2'-reductase
Reaction: 6-lactoyl-5,6,7,8-tetrahydropterin + NADP⁺ = 6-pyruvoyltetrahydropterin + NADPH + H⁺
Other name(s): 6-pyruvoyltetrahydropterin reductase; 6PPH4(2'-oxo) reductase; 6-pyruvoyl tetrahydropterin (2'-oxo)reductase; 6-pyruvoyl-tetrahydropterin 2'-reductase; pyruvoyl-tetrahydropterin reductase
Systematic name: 6-lactoyl-5,6,7,8-tetrahydropterin:NADP⁺ 2'-oxidoreductase

Comments: Not identical with EC 1.1.1.153 sepiapterin reductase.

References: [2816]

[EC 1.1.1.220 created 1989]

EC 1.1.1.221

Accepted name: vomifoliol dehydrogenase

Reaction: (6*S*,9*R*)-6-hydroxy-3-oxo- α -ionol + NAD⁺ = (6*S*)-6-hydroxy-3-oxo- α -ionone + NADH + H⁺

Other name(s): vomifoliol 4'-dehydrogenase; vomifoliol:NAD⁺ 4'-oxidoreductase

Systematic name: (6*S*,9*R*)-6-hydroxy-3-oxo- α -ionol:NAD⁺ oxidoreductase

Comments: Oxidizes vomifoliol to dehydrovomifoliol; involved in the metabolism of abscisic acid in *Corynebacterium* sp.

References: [1548]

[EC 1.1.1.221 created 1989]

[1.1.1.222 *Transferred entry. (R)-4-hydroxyphenyllactate dehydrogenase. Now included with EC 1.1.1.110, aromatic 2-oxoacid reductase*]

[EC 1.1.1.222 created 1989, deleted 2018]

EC 1.1.1.223

Accepted name: isopiperitenol dehydrogenase

Reaction: (-)-*trans*-isopiperitenol + NAD⁺ = (-)-isopiperitenone + NADH + H⁺

Systematic name: (-)-*trans*-isopiperitenol:NAD⁺ oxidoreductase

Comments: Acts on (-)-*trans*-isopiperitenol, (+)-*trans*-piperitenol and (+)-*trans*-pulegol. Involved in the biosynthesis of menthol and related monoterpenes in peppermint (*Mentha piperita*) leaves.

References: [2147]

[EC 1.1.1.223 created 1989]

EC 1.1.1.224

Accepted name: mannose-6-phosphate 6-reductase

Reaction: D-mannitol 1-phosphate + NADP⁺ = D-mannose 6-phosphate + NADPH + H⁺

Other name(s): NADPH-dependent mannose 6-phosphate reductase; mannose-6-phosphate reductase; 6-phosphomannose reductase; NADP-dependent mannose-6-P:mannitol-1-*P* oxidoreductase; NADPH-dependent M6P reductase; NADPH-mannose-6-*P* reductase

Systematic name: D-mannitol-1-phosphate:NADP⁺ 6-oxidoreductase

Comments: Involved in the biosynthesis of mannitol in celery (*Apium graveolens*) leaves.

References: [3604]

[EC 1.1.1.224 created 1989]

EC 1.1.1.225

Accepted name: chlordecone reductase

Reaction: chlordecone alcohol + NADP⁺ = chlordecone + NADPH + H⁺

Other name(s): CDR

Systematic name: chlordecone-alcohol:NADP⁺ 2-oxidoreductase

Comments: Chlordecone is an organochlorine pesticide.

References: [2865]

[EC 1.1.1.225 created 1989]

EC 1.1.1.226

- Accepted name:** 4-hydroxycyclohexanecarboxylate dehydrogenase
Reaction: *trans*-4-hydroxycyclohexanecarboxylate + NAD⁺ = 4-oxocyclohexanecarboxylate + NADH + H⁺
Other name(s): *trans*-4-hydroxycyclohexanecarboxylate dehydrogenase
Systematic name: *trans*-4-hydroxycyclohexanecarboxylate:NAD⁺ 4-oxidoreductase
Comments: The enzyme from *Corynebacterium cyclohexanicum* is highly specific for the *trans*-4-hydroxy derivative.
References: [3123]

[EC 1.1.1.226 created 1990]

EC 1.1.1.227

- Accepted name:** (-)-borneol dehydrogenase
Reaction: (-)-borneol + NAD⁺ = (-)-camphor + NADH + H⁺
Systematic name: (-)-borneol:NAD⁺ oxidoreductase
Comments: NADP⁺ can also act, but more slowly.
References: [862]

[EC 1.1.1.227 created 1990 (EC 1.1.1.182 created 1983, part incorporated 1990)]

EC 1.1.1.228

- Accepted name:** (+)-sabinol dehydrogenase
Reaction: (+)-*cis*-sabinol + NAD⁺ = (+)-sabinone + NADH + H⁺
Other name(s): (+)-*cis*-sabinol dehydrogenase
Systematic name: (+)-*cis*-sabinol:NAD⁺ oxidoreductase
Comments: NADP⁺ can also act, but more slowly. Involved in the biosynthesis of (+)-3-thujone and (-)-3-isothujone.
References: [862]

[EC 1.1.1.228 created 1990 (EC 1.1.1.182 created 1983, part incorporated 1990)]

EC 1.1.1.229

- Accepted name:** diethyl 2-methyl-3-oxosuccinate reductase
Reaction: diethyl (2*R*,3*R*)-2-methyl-3-hydroxysuccinate + NADP⁺ = diethyl 2-methyl-3-oxosuccinate + NADPH + H⁺
Systematic name: diethyl-(2*R*,3*R*)-2-methyl-3-hydroxysuccinate:NADP⁺ 3-oxidoreductase
Comments: Also acts on diethyl (2*S*,3*R*)-2-methyl-3-hydroxysuccinate; and on the corresponding dimethyl esters.
References: [1238]

[EC 1.1.1.229 created 1990]

EC 1.1.1.230

- Accepted name:** 3 α -hydroxyglycyrrhetinate dehydrogenase
Reaction: 3 α -hydroxyglycyrrhetinate + NADP⁺ = 3-oxoglycyrrhetinate + NADPH + H⁺
Systematic name: 3 α -hydroxyglycyrrhetinate:NADP⁺ 3-oxidoreductase
Comments: Highly specific to 3 α -hydroxy derivatives of glycyrrhetinate and its analogues. Not identical to EC 1.1.1.50 3 α -hydroxysteroid dehydrogenase (*Si*-specific).
References: [47]

[EC 1.1.1.230 created 1990]

EC 1.1.1.231

Accepted name: 15-hydroxyprostaglandin-I dehydrogenase (NADP⁺)
Reaction: (5Z,13E)-(15S)-6,9 α -epoxy-11 α ,15-dihydroxyprosta-5,13-dienoate + NADP⁺ = (5Z,13E)-6,9 α -epoxy-11 α -hydroxy-15-oxoprosta-5,13-dienoate + NADPH + H⁺
Other name(s): prostacyclin dehydrogenase; PG I₂ dehydrogenase; prostacyclin dehydrogenase; NADP-linked 15-hydroxyprostaglandin (prostacyclin) dehydrogenase; NADP⁺-dependent PGI₂-specific 15-hydroxyprostaglandin dehydrogenase; 15-hydroxyprostaglandin-I dehydrogenase (NADP)
Systematic name: (5Z,13E)-(15S)-6,9 α -epoxy-11 α ,15-dihydroxyprosta-5,13-dienoate:NADP⁺ 15-oxidoreductase
Comments: Specific for prostaglandin I₂.
References: [2231]

[EC 1.1.1.231 created 1990]

EC 1.1.1.232

Accepted name: 15-hydroxyicosatetraenoate dehydrogenase
Reaction: (15S)-15-hydroxy-5,8,11-*cis*-13-*trans*-icosatetraenoate + NAD(P)⁺ = 15-oxo-5,8,11-*cis*-13-*trans*-icosatetraenoate + NAD(P)H + H⁺
Other name(s): 15-hydroxyeicosatetraenoate dehydrogenase
Systematic name: (15S)-15-hydroxy-5,8,11-*cis*-13-*trans*-icosatetraenoate:NAD(P)⁺ 15-oxidoreductase
References: [3953]

[EC 1.1.1.232 created 1992]

EC 1.1.1.233

Accepted name: *N*-acylmannosamine 1-dehydrogenase
Reaction: *N*-acyl-D-mannosamine + NAD⁺ = *N*-acyl-D-mannosaminolactone + NADH + H⁺
Other name(s): *N*-acylmannosamine dehydrogenase; *N*-acetyl-D-mannosamine dehydrogenase; *N*-acyl-D-mannosamine dehydrogenase; *N*-acylmannosamine dehydrogenase
Systematic name: *N*-acyl-D-mannosamine:NAD⁺ 1-oxidoreductase
Comments: Acts on acetyl-D-mannosamine and glycolyl-D-mannosamine. Highly specific.
References: [1727]

[EC 1.1.1.233 created 1992]

EC 1.1.1.234

Accepted name: flavanone 4-reductase
Reaction: (2S)-flavan-4-ol + NADP⁺ = (2S)-flavanone + NADPH + H⁺
Systematic name: (2S)-flavan-4-ol:NADP⁺ 4-oxidoreductase
Comments: Involved in the biosynthesis of 3-deoxyanthocyanidins from flavanones such as naringenin or eriodictyol.
References: [4027]

[EC 1.1.1.234 created 1992]

EC 1.1.1.235

Accepted name: 8-oxocoformycin reductase
Reaction: coformycin + NADP⁺ = 8-oxocoformycin + NADPH + H⁺
Other name(s): 8-ketodeoxycoformycin reductase
Systematic name: coformycin:NADP⁺ 8-oxidoreductase
Comments: *Si*-specific with respect to NADPH. Also reduces 8-oxodeoxy-coformycin to the nucleoside antibiotic deoxycoformycin.
References: [1521]

[EC 1.1.1.235 created 1992]

EC 1.1.1.236

- Accepted name:** tropinone reductase II
Reaction: pseudotropine + NADP⁺ = tropinone + NADPH + H⁺
Other name(s): tropinone (ψ -tropine-forming) reductase; pseudotropine forming tropinone reductase; tropinone reductase (ambiguous); TR-II
Systematic name: pseudotropine:NADP⁺ 3-oxidoreductase
Comments: This enzyme along with EC 1.1.1.206, tropine dehydrogenase, represents a branch point in tropane alkaloid metabolism [2977]. Tropine (the product of EC 1.1.1.206) is incorporated into hyoscyamine and scopolamine whereas pseudotropine (the product of EC 1.1.1.236) is the first specific metabolite on the pathway to the calystegines [2977]. Both enzymes are always found together in any given tropane-alkaloid-producing species, have a common substrate, tropinone, and are strictly stereospecific [749].
References: [962, 749, 2977, 961]

[EC 1.1.1.236 created 1992, modified 2007]

EC 1.1.1.237

- Accepted name:** hydroxyphenylpyruvate reductase
Reaction: (1) (*R*)-3-(4-hydroxyphenyl)lactate + NAD(P)⁺ = 3-(4-hydroxyphenyl)pyruvate + NAD(P)H + H⁺
(2) (*R*)-3-(3,4-dihydroxyphenyl)lactate + NAD(P)⁺ = 3-(3,4-dihydroxyphenyl)pyruvate + NAD(P)H + H⁺
Other name(s): HPPR
Systematic name: (*R*)-3-(4-hydroxyphenyl)lactate:NAD(P)⁺ oxidoreductase
Comments: The enzyme participates in the biosynthesis of rosmarinic acid. It belongs to the family of D-isomer-specific 2-hydroxyacid dehydrogenases, and prefers NADPH to NADH.
References: [3300, 2099, 2112, 4514]

[EC 1.1.1.237 created 1992, modified 2018]

EC 1.1.1.238

- Accepted name:** 12 β -hydroxysteroid dehydrogenase
Reaction: 3 α ,7 α ,12 β -trihydroxy-5 β -cholan-24-oate + NADP⁺ = 3 α ,7 α -dihydroxy-12-oxo-5 β -cholan-24-oate + NADPH + H⁺
Other name(s): 12 β -hydroxy steroid (nicotinamide adenine dinucleotide phosphate) dehydrogenase
Systematic name: 12 β -hydroxysteroid:NADP⁺ 12-oxidoreductase
Comments: Acts on a number of bile acids, both in their free and conjugated forms.
References: [1012]

[EC 1.1.1.238 created 1992]

EC 1.1.1.239

- Accepted name:** 3 α (17 β)-hydroxysteroid dehydrogenase (NAD⁺)
Reaction: testosterone + NAD⁺ = androstenedione + NADH + H⁺
Other name(s): 3 α ,17 β -hydroxy steroid dehydrogenase; 3 α (17 β)-HSD; 17-ketoreductase (ambiguous); 17 β -HSD (ambiguous); HSD17B6 (gene name); HSD17B8 (gene name)
Systematic name: 3 α (or 17 β)-hydroxysteroid:NAD⁺ oxidoreductase
Comments: Also acts on other 17 β -hydroxysteroids and on the 3 α -hydroxy group of pregnanes and bile acids. Different from EC 1.1.1.50 3 α -hydroxysteroid dehydrogenase (*Si*-specific) or EC 1.1.1.213 3 α -hydroxysteroid dehydrogenase (*Re*-specific).
References: [4149, 4452, 1044, 3140]

[EC 1.1.1.239 created 1992, modified 2012 (EC 1.1.1.63 created 1965, incorporated 2012)]

EC 1.1.1.240

Accepted name: *N*-acetylhexosamine 1-dehydrogenase
Reaction: *N*-acetyl- α -D-glucosamine + NAD⁺ = *N*-acetyl-D-glucosamine + NADH + H⁺
Other name(s): *N*-acetylhexosamine dehydrogenase; *N*-acetyl-D-hexosamine dehydrogenase
Systematic name: *N*-acetyl-D-hexosamine:NAD⁺ 1-oxidoreductase
Comments: Also acts on *N*-acetylgalactosamine and, more slowly, on *N*-acetylmannosamine. Anomeric specificity was tested with *N*-acetyl-D-glucosamine, and it was shown that the enzyme is specific for the α anomer.
References: [1728]

[EC 1.1.1.240 created 1992]

EC 1.1.1.241

Accepted name: 6-*endo*-hydroxycineole dehydrogenase
Reaction: 6-*endo*-hydroxycineole + NAD⁺ = 6-oxocineole + NADH + H⁺
Systematic name: 6-*endo*-hydroxycineole:NAD⁺ 6-oxidoreductase
References: [4631]

[EC 1.1.1.241 created 1992]

[1.1.1.242] *Transferred entry. zeatin reductase. Now EC 1.3.1.69, zeatin reductase*

[EC 1.1.1.242 created 1992, deleted 2001]

EC 1.1.1.243

Accepted name: carveol dehydrogenase
Reaction: (-)-*trans*-carveol + NADP⁺ = (-)-carvone + NADPH + H⁺
Other name(s): (-)-*trans*-carveol dehydrogenase
Systematic name: (-)-*trans*-carveol:NADP⁺ oxidoreductase
References: [1306]

[EC 1.1.1.243 created 1992]

EC 1.1.1.244

Accepted name: methanol dehydrogenase
Reaction: methanol + NAD⁺ = formaldehyde + NADH + H⁺
Systematic name: methanol:NAD⁺ oxidoreductase
References: [131]

[EC 1.1.1.244 created 1992]

EC 1.1.1.245

Accepted name: cyclohexanol dehydrogenase
Reaction: cyclohexanol + NAD⁺ = cyclohexanone + NADH + H⁺
Systematic name: cyclohexanol:NAD⁺ oxidoreductase
Comments: Also oxidizes some other alicyclic alcohols and diols.
References: [823, 951, 4334]

[EC 1.1.1.245 created 1992]

[1.1.1.246] *Transferred entry. pterocarpin synthase. This activity is now known to be catalysed by two enzymes, vestitone reductase (EC 1.1.1.348) and medicarpin synthase (EC 4.2.1.139).]*

[EC 1.1.1.246 created 1992, deleted 2013]

EC 1.1.1.247

Accepted name: codeinone reductase (NADPH)
Reaction: codeine + NADP⁺ = codeinone + NADPH + H⁺
Systematic name: codeine:NADP⁺ oxidoreductase
Comments: Catalyses the reversible reduction of codeinone to codeine, which is a direct precursor of morphine in the opium poppy plant, *Papaver somniferum*.
References: [2416, 2415]

[EC 1.1.1.247 created 1999, modified 2001]

EC 1.1.1.248

Accepted name: salutaridine reductase (NADPH)
Reaction: salutaridinol + NADP⁺ = salutaridine + NADPH + H⁺
Systematic name: salutaridinol:NADP⁺ 7-oxidoreductase
Comments: Catalyses the reversible reduction of salutaridine to salutaridinol, which is a direct precursor of morphinan alkaloids in the poppy plant.
References: [1301]

[EC 1.1.1.248 created 1999, modified 2001]

[1.1.1.249 Deleted entry. Provisional entry deleted. Revised and reinstated as EC 2.5.1.46 deoxyhypusine synthase]

[EC 1.1.1.249 provisional version created 1999, deleted 1999 (reinstated 2001 as EC 2.5.1.46)]

EC 1.1.1.250

Accepted name: D-arabinitol 2-dehydrogenase
Reaction: D-arabinitol + NAD⁺ = D-ribulose + NADH + H⁺
Other name(s): D-arabinitol 2-dehydrogenase (ribulose-forming)
Systematic name: D-arabinitol:NAD⁺ 2-oxidoreductase (D-ribulose-forming)
References: [4660, 3417]

[EC 1.1.1.250 created 1999]

EC 1.1.1.251

Accepted name: galactitol-1-phosphate 5-dehydrogenase
Reaction: galactitol 1-phosphate + NAD⁺ = D-tagatose 6-phosphate + NADH + H⁺
Other name(s): *gatD* (gene name)
Systematic name: galactitol-1-phosphate:NAD⁺ oxidoreductase
Comments: The enzyme from the bacterium *Escherichia coli* is involved in a galactitol degradation pathway. It contains two zinc atoms per subunit.
References: [4656, 3092, 284]

[EC 1.1.1.251 created 1999]

EC 1.1.1.252

Accepted name: tetrahydroxynaphthalene reductase
Reaction: scytalone + NADP⁺ = 1,3,6,8-tetrahydroxynaphthalene + NADPH + H⁺
Systematic name: scytalone:NADP⁺ Δ⁵-oxidoreductase
Comments: Reduces 1,3,6,8-tetrahydroxynaphthalene to scytalone and also reduces 1,3,8-trihydroxynaphthalene to vermeline. Involved with EC 4.2.1.94 scytalone dehydratase in the biosynthesis of melanin in pathogenic fungi.
References: [4599, 4447, 4273]

[EC 1.1.1.252 created 1992 as EC 1.3.1.50, transferred 1999 to EC 1.1.1.252]

[1.1.1.253 Transferred entry. pteridine reductase. Now EC 1.5.1.33, pteridine reductase]

[EC 1.1.1.253 created 1999, deleted 2003]

EC 1.1.1.254

Accepted name: (*S*)-carnitine 3-dehydrogenase
Reaction: (*S*)-carnitine + NAD⁺ = 3-dehydrocarnitine + NADH + H⁺
Systematic name: (*S*)-carnitine:NAD⁺ oxidoreductase
Comments: Specific for the (*S*)-enantiomer of carnitine, i.e., the enantiomer of the substrate of EC 1.1.1.108 carnitine 3-dehydrogenase
References: [3804]

[EC 1.1.1.254 created 1999]

EC 1.1.1.255

Accepted name: mannitol dehydrogenase
Reaction: D-mannitol + NAD⁺ = D-mannose + NADH + H⁺
Other name(s): MTD; NAD-dependent mannitol dehydrogenase
Systematic name: mannitol:NAD⁺ 1-oxidoreductase
Comments: The enzyme from *Apium graveolens* (celery) oxidizes alditols with a minimum requirement of 2*R* chirality at the carbon adjacent to the primary carbon undergoing the oxidation. The enzyme is specific for NAD⁺ and does not use NADP⁺.
References: [4050, 4051, 4637, 4049]

[EC 1.1.1.255 created 2000]

EC 1.1.1.256

Accepted name: fluoren-9-ol dehydrogenase
Reaction: fluoren-9-ol + NAD(P)⁺ = fluoren-9-one + NAD(P)H + H⁺
Systematic name: fluoren-9-ol:NAD(P)⁺ oxidoreductase
Comments: Involved in the pathway for fluorene metabolism in *Arthrobacter* sp.
References: [570, 1411]

[EC 1.1.1.256 created 2000]

EC 1.1.1.257

Accepted name: 4-(hydroxymethyl)benzenesulfonate dehydrogenase
Reaction: 4-(hydroxymethyl)benzenesulfonate + NAD⁺ = 4-formylbenzenesulfonate + NADH + H⁺
Systematic name: 4-(hydroxymethyl)benzenesulfonate:NAD⁺ oxidoreductase
Comments: Involved in the toluene-4-sulfonate degradation pathway in *Comamonas testosteroni*.
References: [1965]

[EC 1.1.1.257 created 2000]

EC 1.1.1.258

Accepted name: 6-hydroxyhexanoate dehydrogenase
Reaction: 6-hydroxyhexanoate + NAD⁺ = 6-oxohexanoate + NADH + H⁺
Systematic name: 6-hydroxyhexanoate:NAD⁺ oxidoreductase
Comments: Involved in the cyclohexanol degradation pathway in *Acinetobacter* NCIB 9871.
References: [951, 1597]

[EC 1.1.1.258 created 2000]

EC 1.1.1.259

Accepted name: 3-hydroxypimeloyl-CoA dehydrogenase
Reaction: 3-hydroxypimeloyl-CoA + NAD⁺ = 3-oxopimeloyl-CoA + NADH + H⁺
Systematic name: 3-hydroxypimeloyl-CoA:NAD⁺ oxidoreductase
Comments: Involved in the anaerobic pathway of benzoate degradation in bacteria.
References: [1545]

[EC 1.1.1.259 created 2000]

EC 1.1.1.260

Accepted name: sulcatone reductase
Reaction: sulcatol + NAD⁺ = sulcatone + NADH + H⁺
Systematic name: sulcatol:NAD⁺ oxidoreductase
Comments: Studies on the effects of growth-stage and nutrient supply on the stereochemistry of sulcatone reduction in *Clostridia pasteurianum*, *C. tyrobutyricum* and *Lactobacillus brevis* suggest that there may be at least two sulcatone reductases with different stereospecificities.
References: [276, 4286, 4287]

[EC 1.1.1.260 created 2000, modified 2001]

EC 1.1.1.261

Accepted name: *sn*-glycerol-1-phosphate dehydrogenase
Reaction: *sn*-glycerol 1-phosphate + NAD(P)⁺ = glycerone phosphate + NAD(P)H + H⁺
Other name(s): glycerol-1-phosphate dehydrogenase [NAD(P)⁺]; *sn*-glycerol-1-phosphate:NAD⁺ oxidoreductase; G-1-P dehydrogenase; Gro1PDH; AraM
Systematic name: *sn*-glycerol-1-phosphate:NAD(P)⁺ 2-oxidoreductase
Comments: This enzyme is found primarily as a Zn²⁺-dependent form in archaea but a Ni²⁺-dependent form has been found in Gram-positive bacteria [1442]. The Zn²⁺-dependent metalloenzyme is responsible for the formation of archaea-specific *sn*-glycerol-1-phosphate, the first step in the biosynthesis of polar lipids in archaea. It is the enantiomer of *sn*-glycerol 3-phosphate, the form of glycerophosphate found in bacteria and eukaryotes. The other enzymes involved in the biosynthesis of polar lipids in archaea are EC 2.5.1.41 (phosphoglycerol geranylgeranyltransferase) and EC 2.5.1.42 (geranylgeranylglycerol-phosphate geranylgeranyltransferase), which together alkylate the hydroxy groups of glycerol 1-phosphate to give unsaturated archaetidic acid, which is acted upon by EC 2.7.7.67 (CDP-archaeol synthase) to form CDP-unsaturated archaeol. The final step in the pathway involves the addition of L-serine, with concomitant removal of CMP, leading to the production of unsaturated archaetidylserine [2889]. Activity of the enzyme is stimulated by K⁺ [3076].
References: [3075, 3076, 2195, 2889, 1497, 1442]

[EC 1.1.1.261 created 2000, modified 2009]

EC 1.1.1.262

Accepted name: 4-hydroxythreonine-4-phosphate dehydrogenase
Reaction: 4-phosphooxy-L-threonine + NAD⁺ = 3-amino-2-oxopropyl phosphate + CO₂ + NADH + H⁺
Other name(s): NAD⁺-dependent threonine 4-phosphate dehydrogenase; L-threonine 4-phosphate dehydrogenase; 4-(phosphohydroxy)-L-threonine dehydrogenase; PdxA; 4-(phosphonoxy)-L-threonine:NAD⁺ oxidoreductase; 4-phosphooxy-L-threonine:NAD⁺ oxidoreductase
Systematic name: 4-phosphooxy-L-threonine:NAD⁺ 3-oxidoreductase (decarboxylating)
Comments: The enzyme is part of the biosynthesis pathway of the coenzyme pyridoxal 5'-phosphate found in anaerobic bacteria.
References: [544, 2322, 3920, 212]

[EC 1.1.1.262 created 2000, modified 2006]

EC 1.1.1.263

- Accepted name:** 1,5-anhydro-D-fructose reductase
Reaction: 1,5-anhydro-D-glucitol + NADP⁺ = 1,5-anhydro-D-fructose + NADPH + H⁺
Systematic name: 1,5-anhydro-D-glucitol:NADP⁺ oxidoreductase
Comments: Also reduces pyridine-3-aldehyde and 2,3-butanedione. Acetaldehyde, 2-dehydroglucose (glucosone) and glucuronate are poor substrates, but there is no detectable action on glucose, mannose and fructose.
References: [3644]

[EC 1.1.1.263 created 2000]

EC 1.1.1.264

- Accepted name:** L-idonate 5-dehydrogenase
Reaction: L-idonate + NAD(P)⁺ = 5-dehydro-D-gluconate + NAD(P)H + H⁺
Systematic name: L-idonate:NAD(P)⁺ oxidoreductase
Comments: The enzyme from the bacterium *Escherichia coli* is specific for 5-dehydro-D-gluconate. *cf.* EC 1.1.1.366, L-idonate 5-dehydrogenase (NAD⁺).
References: [248]

[EC 1.1.1.264 created 2000, modified 2013]

EC 1.1.1.265

- Accepted name:** 3-methylbutanal reductase
Reaction: 3-methylbutanol + NAD(P)⁺ = 3-methylbutanal + NAD(P)H + H⁺
Systematic name: 3-methylbutanol:NAD(P)⁺ oxidoreductase
Comments: The enzyme purified from *Saccharomyces cerevisiae* catalyses the reduction of a number of straight-chain and branched aldehydes, as well as some aromatic aldehydes.
References: [4410, 3034]

[EC 1.1.1.265 created 2000]

EC 1.1.1.266

- Accepted name:** dTDP-4-dehydro-6-deoxyglucose reductase
Reaction: dTDP- α -D-fucopyranose + NAD(P)⁺ = dTDP-4-dehydro-6-deoxy- α -D-glucose + NAD(P)H + H⁺
Other name(s): dTDP-4-keto-6-deoxyglucose reductase; dTDP-D-fucose:NADP⁺ oxidoreductase; Fcf1; dTDP-6-deoxy-D-xylo-hex-4-ulopyranose reductase
Systematic name: dTDP- α -D-fucopyranose:NAD(P)⁺ oxidoreductase
Comments: The enzymes from the Gram-negative bacteria *Aggregatibacter actinomycetemcomitans* and *Escherichia coli* O52 are involved in activation of fucose for incorporation into capsular polysaccharide O-antigens [4814, 4525]. The enzyme from the Gram-positive bacterium *Anoxybacillus tepidamans* (*Geobacillus tepidamans*) is involved in activation of fucose for incorporation into the organism's S-layer [4863]. The enzyme from *Escherichia coli* O52 has a higher catalytic efficiency with NADH than with NADPH [4525].
References: [4814, 4863, 4525]

[EC 1.1.1.266 created 2001, modified 2013]

EC 1.1.1.267

- Accepted name:** 1-deoxy-D-xylulose-5-phosphate reductoisomerase
Reaction: 2-C-methyl-D-erythritol 4-phosphate + NADP⁺ = 1-deoxy-D-xylulose 5-phosphate + NADPH + H⁺
Other name(s): DXP-reductoisomerase; 1-deoxy-D-xylulose-5-phosphate isomeroreductase; 2-C-methyl-D-erythritol 4-phosphate (MEP) synthase
Systematic name: 2-C-methyl-D-erythritol-4-phosphate:NADP⁺ oxidoreductase (isomerizing)

Comments: The enzyme requires Mn^{2+} , Co^{2+} or Mg^{2+} for activity, with the first being most effective. The enzyme from several eubacteria, including *Escherichia coli*, forms part of an alternative nonmevalonate pathway for terpenoid biosynthesis (for diagram, click here). The mechanism has been shown to be a retroaldol/aldol reaction [2934].

References: [4170, 2934]

[EC 1.1.1.267 created 2001]

EC 1.1.1.268

Accepted name: 2-(*R*)-hydroxypropyl-CoM dehydrogenase

Reaction: 2-(*R*)-hydroxypropyl-CoM + NAD^+ = 2-oxopropyl-CoM + NADH + H^+

Other name(s): 2-(2-(*R*)-hydroxypropylthio)ethanesulfonate dehydrogenase; 2-[2-(*R*)-hydroxypropylthio]ethanesulfonate: NAD^+ oxidoreductase

Systematic name: 2-[(2*R*)-2-hydroxypropyl]sulfanylethane-1-sulfonate: NAD^+ oxidoreductase

Comments: The enzyme is highly specific for (*R*)-2-hydroxyalkyl thioethers of CoM, in contrast to EC 1.1.1.269, 2-(*S*)-hydroxypropyl-CoM dehydrogenase, which is highly specific for the (*S*)-enantiomer. This enzyme forms component III of a four-component enzyme system (comprising EC 4.4.1.23 [2-hydroxypropyl-CoM lyase; component I], EC 1.8.1.5 [2-oxopropyl-CoM reductase (carboxylating); component II], EC 1.1.1.268 [2-(*R*)-hydroxypropyl-CoM dehydrogenase; component III] and EC 1.1.1.269 [2-(*S*)-hydroxypropyl-CoM dehydrogenase; component IV]) that is involved in epoxyalkane carboxylation in *Xanthobacter* sp. strain Py2.

References: [68]

[EC 1.1.1.268 created 2001]

EC 1.1.1.269

Accepted name: 2-(*S*)-hydroxypropyl-CoM dehydrogenase

Reaction: (2*S*)-2-hydroxypropyl-CoM + NAD^+ = 2-oxopropyl-CoM + NADH + H^+

Other name(s): 2-(2-(*S*)-hydroxypropylthio)ethanesulfonate dehydrogenase; 2-[2-(*S*)-hydroxypropylthio]ethanesulfonate: NAD^+ oxidoreductase

Systematic name: 2-[(2*S*)-2-hydroxypropyl]sulfanylethanesulfonate: NAD^+ oxidoreductase

Comments: The enzyme is highly specific for (2*S*)-2-hydroxyalkyl thioethers of CoM, in contrast to EC 1.1.1.268, 2-(*R*)-hydroxypropyl-CoM dehydrogenase, which is highly specific for the (*R*)-enantiomer. This enzyme forms component IV of a four-component enzyme system EC 4.4.1.23 (2-hydroxypropyl-CoM lyase; component I), EC 1.8.1.5 [2-oxopropyl-CoM reductase (carboxylating); component II], EC 1.1.1.268 [2-(*R*)-hydroxypropyl-CoM dehydrogenase; component III] and EC 1.1.1.269 [2-(*S*)-hydroxypropyl-CoM dehydrogenase; component IV].html";click here that is involved in epoxyalkane carboxylation in *Xanthobacter* sp. strain Py2.

References: [68]

[EC 1.1.1.269 created 2001]

EC 1.1.1.270

Accepted name: 3 β -hydroxysteroid 3-dehydrogenase

Reaction: a 3 β -hydroxysteroid + $NADP^+$ = a 3-oxosteroid + NADPH + H^+

Other name(s): 3-keto-steroid reductase; 3-KSR; HSD17B7 (gene name); ERG27 (gene name)

Systematic name: 3 β -hydroxysteroid: $NADP^+$ 3-oxidoreductase

Comments: The enzyme acts on multiple 3 β -hydroxysteroids. Participates in the biosynthesis of zosterol and cholesterol, where it catalyses the reaction in the opposite direction to that shown. The mammalian enzyme is bifunctional and also catalyses EC 1.1.1.62, 17 β -estradiol 17-dehydrogenase [2650].

References: [4151, 333, 1248, 2650]

[EC 1.1.1.270 created 2002, modified 2012]

EC 1.1.1.271

- Accepted name:** GDP-L-fucose synthase
Reaction: $\text{GDP-}\beta\text{-L-fucose} + \text{NADP}^+ = \text{GDP-4-dehydro-}\alpha\text{-D-rhamnose} + \text{NADPH} + \text{H}^+$
Other name(s): GDP-4-keto-6-deoxy-D-mannose-3,5-epimerase-4-reductase; GDP-L-fucose:NADP⁺ 4-oxidoreductase (3,5-epimerizing)
Systematic name: GDP- β -L-fucose:NADP⁺ 4-oxidoreductase (3,5-epimerizing)
Comments: Both human and *Escherichia coli* enzymes can use NADH in place of NADPH to a slight extent.
References: [596, 2724, 2771, 3957]

[EC 1.1.1.271 created 2002, modified 2003]

EC 1.1.1.272

- Accepted name:** D-2-hydroxyacid dehydrogenase (NADP⁺)
Reaction: an (*R*)-2-hydroxycarboxylate + NADP⁺ = a 2-oxocarboxylate + NADPH + H⁺
Other name(s): *ddh* (gene name)
Systematic name: (*R*)-2-hydroxycarboxylate:NADP⁺ oxidoreductase
Comments: This enzyme, characterized from the halophilic archaeon *Haloferax mediterranei* and the mold *Aspergillus oryzae*, catalyses a stereospecific reduction of 2-oxocarboxylic acids into the corresponding D-2-hydroxycarboxylic acids. The enzyme prefers substrates with a main chain of 5 carbons (such as 4-methyl-2-oxopentanoate) to those with a shorter chain, and can use NADH with much lower efficiency. *cf.* EC 1.1.1.345, (*d*)-2-hydroxyacid dehydrogenase (NAD⁺).
References: [941, 3870]

[EC 1.1.1.272 created 2002, modified 2013]

EC 1.1.1.273

- Accepted name:** vellosimine dehydrogenase
Reaction: 10-deoxysarpagine + NADP⁺ = vellosimine + NADPH + H⁺
Systematic name: 10-deoxysarpagine:NADP⁺ oxidoreductase
Comments: Also acts on related alkaloids with an endo-aldehyde group as vellosimine (same stereochemistry at C-16) but only slight activity with exo-aldehydes. Detected in many cell suspension cultures of plants from the family Apocynaceae.
References: [3310]

[EC 1.1.1.273 created 2002]

EC 1.1.1.274

- Accepted name:** 2,5-didehydrogluconate reductase (2-dehydro-D-gluconate-forming)
Reaction: 2-dehydro-D-gluconate + NADP⁺ = 2,5-didehydro-D-gluconate + NADPH + H⁺
Other name(s): 2,5-diketo-D-gluconate reductase (ambiguous)
Systematic name: 2-dehydro-D-gluconate:NADP⁺ 2-oxidoreductase (2-dehydro-D-gluconate-forming)
Comments: The enzyme is involved in the catabolism of 2,5-didehydrogluconate. *cf.* EC 1.1.1.346, 2,5-didehydrogluconate reductase (2-dehydro-L-gulonate-forming).
References: [3965]

[EC 1.1.1.274 created 2002, modified 2013]

EC 1.1.1.275

- Accepted name:** (+)-*trans*-carveol dehydrogenase
Reaction: (+)-*trans*-carveol + NAD⁺ = (+)-(*S*)-carvone + NADH + H⁺
Other name(s): carveol dehydrogenase
Systematic name: (+)-*trans*-carveol:NAD⁺ oxidoreductase

Comments: NADP⁺ cannot replace NAD⁺. Forms part of the monoterpenoid biosynthesis pathway in *Carum carvi* (caraway) seeds.

References: [407]

[EC 1.1.1.275 created 2003]

EC 1.1.1.276

Accepted name: serine 3-dehydrogenase (NADP⁺)

Reaction: L-serine + NADP⁺ = 2-aminoacetaldehyde + CO₂ + NADPH + H⁺ (overall reaction)

(1a) L-serine + NADP⁺ = 2-aminomalonaldehyde + NADPH + H⁺

(1b) 2-aminomalonaldehyde = 2-aminoacetaldehyde + CO₂ (spontaneous)

Other name(s): serine 3-dehydrogenase

Systematic name: L-serine:NADP⁺ 3-oxidoreductase

Comments: NAD⁺ cannot replace NADP⁺ [cf. EC 1.1.1.387, serine 3-dehydrogenase (NAD⁺)].

References: [1209, 681]

[EC 1.1.1.276 created 2003, modified 2015]

EC 1.1.1.277

Accepted name: 3β-hydroxy-5β-steroid dehydrogenase

Reaction: 3β-hydroxy-5β-pregnane-20-one + NADP⁺ = 5β-pregnan-3,20-dione + NADPH + H⁺

Other name(s): 3β-hydroxysteroid 5β-oxidoreductase; 3β-hydroxysteroid 5β-progesterone oxidoreductase

Systematic name: 3β-hydroxy-5β-steroid:NADP⁺ 3-oxidoreductase

References: [4081, 3788, 2496]

[EC 1.1.1.277 created 2003]

EC 1.1.1.278

Accepted name: 3β-hydroxy-5α-steroid dehydrogenase

Reaction: 3β-hydroxy-5α-pregnane-20-one + NADP⁺ = 5α-pregnan-3,20-dione + NADPH + H⁺

Systematic name: 3β-hydroxy-5α-steroid:NADP⁺ 3-oxidoreductase

References: [2496, 4548]

[EC 1.1.1.278 created 2003]

EC 1.1.1.279

Accepted name: (*R*)-3-hydroxyacid-ester dehydrogenase

Reaction: ethyl (*R*)-3-hydroxyhexanoate + NADP⁺ = ethyl 3-oxohexanoate + NADPH + H⁺

Other name(s): 3-oxo ester (*R*)-reductase

Systematic name: ethyl-(*R*)-3-hydroxyhexanoate:NADP⁺ 3-oxidoreductase

Comments: Also acts on ethyl (*R*)-3-oxobutanoate and some other (*R*)-3-hydroxy acid esters. The (*R*)- symbol is allotted on the assumption that no substituents change the order of priority from O-3 > C-2 > C-4. A subunit of yeast fatty acid synthase EC 2.3.1.86, fatty-acyl-CoA synthase system. cf. EC 1.1.1.280, (*S*)-3-hydroxyacid ester dehydrogenase.

References: [1608]

[EC 1.1.1.279 created 1990 as EC 1.2.1.55, transferred 2003 to EC 1.1.1.279, modified 2018]

EC 1.1.1.280

Accepted name: (*S*)-3-hydroxyacid-ester dehydrogenase

Reaction: ethyl (*S*)-3-hydroxyhexanoate + NADP⁺ = ethyl 3-oxohexanoate + NADPH + H⁺

Other name(s): 3-oxo ester (*S*)-reductase

Systematic name: ethyl-(*S*)-3-hydroxyhexanoate:NADP⁺ 3-oxidoreductase
Comments: Also acts on 4-oxo- and 5-oxo-fatty acids and their esters. *cf.* EC 1.1.1.279 (*R*)-3-hydroxyacid-ester dehydrogenase.
References: [1608]

[EC 1.1.1.280 created 1990 as EC 1.2.1.56, transferred 2003 to EC 1.1.1.280]

EC 1.1.1.281

Accepted name: GDP-4-dehydro-6-deoxy-D-mannose reductase
Reaction: GDP- α -D-rhamnose + NAD(P)⁺ = GDP-4-dehydro- α -D-rhamnose + NAD(P)H + H⁺
Other name(s): GDP-4-keto-6-deoxy-D-mannose reductase [ambiguous]; GDP-6-deoxy-D-*lyxo*-4-hexulose reductase; Rmd; GDP-6-deoxy-D-mannose:NAD(P)⁺ 4-oxidoreductase (D-rhamnose-forming); GDP-6-deoxy- α -D-mannose:NAD(P)⁺ 4-oxidoreductase (D-rhamnose-forming)
Systematic name: GDP- α -D-rhamnose:NAD(P)⁺ 4-oxidoreductase
Comments: This enzyme differs from EC 1.1.1.187, GDP-4-dehydro-D-rhamnose reductase, in that the only product formed is GDP- α -D-rhamnose. D-Rhamnose is a constituent of lipopolysaccharides of Gram-negative plant and human pathogenic bacteria.
References: [2165, 2625]

[EC 1.1.1.281 created 2004]

EC 1.1.1.282

Accepted name: quinate/shikimate dehydrogenase [NAD(P)⁺]
Reaction: (1) L-quininate + NAD(P)⁺ = 3-dehydroquininate + NAD(P)H + H⁺
(2) shikimate + NAD(P)⁺ = 3-dehydroshikimate + NAD(P)H + H⁺
Other name(s): YdiB; quinate/shikimate dehydrogenase (ambiguous)
Systematic name: L-quininate:NAD(P)⁺ 3-oxidoreductase
Comments: This is the second shikimate dehydrogenase enzyme found in *Escherichia coli*. It can use both quinate and shikimate as substrates and either NAD⁺ or NADP⁺ as acceptor. The low catalytic efficiency with both quinate and shikimate suggests that neither may be the physiological substrate. *cf.* EC 1.1.1.24, quinate/shikimate dehydrogenase (NAD⁺), EC 1.1.5.8, quinate/shikimate dehydrogenase (quinone), and EC 1.1.1.25, shikimate dehydrogenase (NADP⁺).
References: [2793, 283]

[EC 1.1.1.282 created 2004, modified 2021]

EC 1.1.1.283

Accepted name: methylglyoxal reductase (NADPH)
Reaction: (*S*)-lactaldehyde + NADP⁺ = 2-oxopropanal + NADPH + H⁺
Other name(s): lactaldehyde dehydrogenase (NADP⁺); GRE2 (gene name); methylglyoxal reductase (NADPH-dependent); lactaldehyde:NADP⁺ oxidoreductase
Systematic name: (*S*)-lactaldehyde:NADP⁺ oxidoreductase
Comments: The enzyme from the yeast *Saccharomyces cerevisiae* catalyses the reduction of a keto group in a number of compounds, forming enantiopure products. Among the substrates are methylglyoxal (which is reduced to (*S*)-lactaldehyde) [2943, 627], 3-methylbutanal [1565], hexane-2,5-dione [2924] and 3-chloro-1-phenylpropan-1-one [674]. The enzyme differs from EC 1.1.1.78, methylglyoxal reductase (NADH), which is found in mammals, by its coenzyme requirement, reaction direction, and enantiomeric preference.
References: [2943, 627, 1565, 2924, 674, 433]

[EC 1.1.1.283 created 2005, modified 2013]

EC 1.1.1.284

- Accepted name:** S-(hydroxymethyl)glutathione dehydrogenase
Reaction: S-(hydroxymethyl)glutathione + NAD(P)⁺ = S-formylglutathione + NAD(P)H + H⁺
Other name(s): NAD-linked formaldehyde dehydrogenase (incorrect); formaldehyde dehydrogenase (incorrect); formic dehydrogenase (incorrect); class III alcohol dehydrogenase; ADH3; χ -ADH; FDH (incorrect); formaldehyde dehydrogenase (glutathione) (incorrect); GS-FDH (incorrect); glutathione-dependent formaldehyde dehydrogenase (incorrect); GD-FALDH; NAD- and glutathione-dependent formaldehyde dehydrogenase; NAD-dependent formaldehyde dehydrogenase (incorrect)
Systematic name: S-(hydroxymethyl)glutathione:NAD⁺ oxidoreductase
Comments: The substrate, S-(hydroxymethyl)glutathione, forms spontaneously from glutathione and formaldehyde; its rate of formation is increased in some bacteria by the presence of EC 4.4.1.22, S-(hydroxymethyl)glutathione synthase. This enzyme forms part of the pathway that detoxifies formaldehyde, since the product is hydrolysed by EC 3.1.2.12, S-formylglutathione hydrolase. The human enzyme belongs to the family of zinc-dependent alcohol dehydrogenases. Also specifically reduces S-nitrosylglutathione.
References: [1880, 3573, 2517, 3658, 4412, 3453, 216]

[EC 1.1.1.284 created 2005 (EC 1.2.1.1 created 1961, modified 1982, modified 2002, part transferred 2005 to EC 1.1.1.284)]

EC 1.1.1.285

- Accepted name:** 3''-deamino-3''-oxonicotianamine reductase
Reaction: 2'-deoxymugineic acid + NAD(P)⁺ = 3''-deamino-3''-oxonicotianamine + NAD(P)H + H⁺
Systematic name: 2'-deoxymugineic acid:NAD(P)⁺ 3''-oxidoreductase
References: [3891]

[EC 1.1.1.285 created 2005]

EC 1.1.1.286

- Accepted name:** isocitrate—homoisocitrate dehydrogenase
Reaction: (1) isocitrate + NAD⁺ = 2-oxoglutarate + CO₂ + NADH
(2) (1R,2S)-1-hydroxybutane-1,2,4-tricarboxylate + NAD⁺ = 2-oxoadipate + CO₂ + NADH + H⁺
Other name(s): homoisocitrate—*isocitrate* dehydrogenase; PH1722
Systematic name: isocitrate(homoisocitrate):NAD⁺ oxidoreductase (decarboxylating)
Comments: Requires Mn²⁺ and K⁺ or NH₄⁺ for activity. Unlike EC 1.1.1.41, *isocitrate* dehydrogenase (NAD⁺) and EC 1.1.1.87, homoisocitrate dehydrogenase, this enzyme, from *Pyrococcus horikoshii*, can use both isocitrate and homoisocitrate as substrates. The enzyme may play a role in both the lysine and glutamate biosynthesis pathways.
References: [2842]

[EC 1.1.1.286 created 2005]

EC 1.1.1.287

- Accepted name:** D-arabinitol dehydrogenase (NADP⁺)
Reaction: (1) D-arabinitol + NADP⁺ = D-xylulose + NADPH + H⁺
(2) D-arabinitol + NADP⁺ = D-ribulose + NADPH + H⁺
Other name(s): NADP⁺-dependent D-arabitol dehydrogenase; ARD1p; D-arabitol dehydrogenase 1
Systematic name: D-arabinitol:NADP⁺ oxidoreductase
Comments: The enzyme from the rust fungus *Uromyces fabae* can use D-arabinitol and D-mannitol as substrates in the forward direction and D-xylulose, D-ribulose and, to a lesser extent, D-fructose as substrates in the reverse direction. This enzyme carries out the reactions of both EC 1.1.1.11, D-arabinitol 4-dehydrogenase and EC 1.1.1.250, D-arabinitol 2-dehydrogenase, but unlike them, uses NADP⁺ rather than NAD⁺ as cofactor. D-Arabinitol is capable of quenching reactive oxygen species involved in defense reactions of the host plant.

References: [2504]

[EC 1.1.1.287 created 2005]

EC 1.1.1.288

Accepted name: xanthoxin dehydrogenase
Reaction: xanthoxin + NAD⁺ = abscisic aldehyde + NADH + H⁺
Other name(s): xanthoxin oxidase; ABA2
Systematic name: xanthoxin:NAD⁺ oxidoreductase
Comments: Requires a molybdenum cofactor for activity. NADP⁺ cannot replace NAD⁺ and short-chain alcohols such as ethanol, isopropanol, butanol and cyclohexanol cannot replace xanthoxin as substrate [1361]. Involved in the abscisic-acid biosynthesis pathway in plants, along with EC 1.2.3.14 (abscisic-aldehyde oxidase), EC 1.13.11.51 (9-*cis*-epoxycarotenoid dioxygenase) and EC 1.14.13.93 [(+)-abscisic acid 8'-hydroxylase]. Abscisic acid is a sesquiterpenoid plant hormone that is involved in the control of a wide range of essential physiological processes, including seed development, germination and responses to stress [1361].
References: [3907, 3766, 1361]

[EC 1.1.1.288 created 2005]

EC 1.1.1.289

Accepted name: sorbose reductase
Reaction: D-glucitol + NADP⁺ = L-sorbose + NADPH + H⁺
Other name(s): Sou1p
Systematic name: D-glucitol:NADP⁺ oxidoreductase
Comments: The reaction occurs predominantly in the reverse direction. This enzyme can also convert D-fructose into D-mannitol, but more slowly. Belongs in the short-chain dehydrogenase family.
References: [1399, 1400, 4099, 3885]

[EC 1.1.1.289 created 2006]

EC 1.1.1.290

Accepted name: 4-phosphoerythronate dehydrogenase
Reaction: 4-phospho-D-erythronate + NAD⁺ = (3*R*)-3-hydroxy-2-oxo-4-phosphooxybutanoate + NADH + H⁺
Other name(s): PdxB; PdxB 4PE dehydrogenase; 4-*O*-phosphoerythronate dehydrogenase; 4PE dehydrogenase; erythronate-4-phosphate dehydrogenase
Systematic name: 4-phospho-D-erythronate:NAD⁺ 2-oxidoreductase
Comments: This enzyme catalyses a step in a bacterial pathway for the biosynthesis of pyridoxal 5'-phosphate. The enzyme contains a tightly-bound NAD(H) cofactor that is not re-oxidized by free NAD⁺. In order to re-oxidize the cofactor and restore enzyme activity, the enzyme catalyses the reduction of a 2-oxo acid (such as 2-oxoglutarate, oxaloacetate, or pyruvate) to the respective (*R*)-hydroxy acid [3594]. *cf.* EC 1.1.1.399, 2-oxoglutarate reductase.
References: [2328, 3272, 4900, 1382, 3743, 3594]

[EC 1.1.1.290 created 2006, modified 2016]

EC 1.1.1.291

Accepted name: 2-hydroxymethylglutarate dehydrogenase
Reaction: (*S*)-2-hydroxymethylglutarate + NAD⁺ = 2-formylglutarate + NADH + H⁺
Other name(s): HgD
Systematic name: (*S*)-2-hydroxymethylglutarate:NAD⁺ oxidoreductase

Comments: NADP⁺ cannot replace NAD⁺. Forms part of the nicotinate-fermentation catabolism pathway in *Eubacterium barkeri*. Other enzymes involved in this pathway are EC 1.17.1.5 (nicotinate dehydrogenase), EC 1.3.7.1 (6-hydroxynicotinate reductase), EC 3.5.2.18 (enamidase), EC 5.4.99.4 (2-methyleneglutarate mutase), EC 5.3.3.6 (methylitaconate Δ-isomerase), EC 4.2.1.85 (dimethylmaleate hydratase) and EC 4.1.3.32 (2,3-dimethylmalate lyase).

References: [67]

[EC 1.1.1.291 created 2006]

EC 1.1.1.292

Accepted name: 1,5-anhydro-D-fructose reductase (1,5-anhydro-D-mannitol-forming)
Reaction: 1,5-anhydro-D-mannitol + NADP⁺ = 1,5-anhydro-D-fructose + NADPH + H⁺
Other name(s): 1,5-anhydro-D-fructose reductase (ambiguous); AFR (ambiguous)
Systematic name: 1,5-anhydro-D-mannitol:NADP⁺ oxidoreductase
Comments: This enzyme is present in some but not all *Rhizobium* species and belongs in the GFO/IDH/MocA protein family [814]. This enzyme differs from hepatic 1,5-anhydro-D-fructose reductase, which yields 1,5-anhydro-D-glucitol as the product (see EC 1.1.1.263). In *Sinorhizobium morelense*, the product of the reaction, 1,5-anhydro-D-mannitol, can be further metabolized to D-mannose [2276]. The enzyme also reduces 1,5-anhydro-D-erythro-hexo-2,3-diulose and 2-ketoaldoses (called osones), such as D-glucosone (D-arabino-hexos-2-ulose) and 6-deoxy-D-glucosone. It does not reduce common aldoses and ketoses, or non-sugar aldehydes and ketones [2276].
References: [2276, 814]

[EC 1.1.1.292 created 2007]

[1.1.1.293 Deleted entry. tropinone reductase I. This enzyme was already in the Enzyme List as EC 1.1.1.206, tropine dehydrogenase so EC 1.1.1.293 has been withdrawn at the public-review stage]

[EC 1.1.1.293 created 2007, withdrawn while undergoing public review]

EC 1.1.1.294

Accepted name: chlorophyll(ide) *b* reductase
Reaction: 7¹-hydroxychlorophyllide *a* + NAD(P)⁺ = chlorophyllide *b* + NAD(P)H + H⁺
Other name(s): chlorophyll *b* reductase; Chl *b* reductase
Systematic name: 7¹-hydroxychlorophyllide-*a*:NAD(P)⁺ oxidoreductase
Comments: This enzyme carries out the first step in the conversion of chlorophyll *b* to chlorophyll *a*. It is involved in chlorophyll degradation, which occurs during leaf senescence [1733] and it also forms part of the chlorophyll cycle, which interconverts chlorophyll *a* and *b* in response to changing light conditions [1844, 3592].
References: [3714, 3715, 1733, 1844, 3592]

[EC 1.1.1.294 created 2007]

EC 1.1.1.295

Accepted name: momilactone-A synthase
Reaction: 3β-hydroxy-9β-pimara-7,15-diene-19,6β-olide + NAD(P)⁺ = momilactone A + NAD(P)H + H⁺
Other name(s): momilactone A synthase; OsMAS
Systematic name: 3β-hydroxy-9β-pimara-7,15-diene-19,6β-olide:NAD(P)⁺ oxidoreductase
Comments: The rice phytoalexin momilactone A is a diterpenoid secondary metabolite that is involved in the defense mechanism of the plant. Momilactone A is produced in response to attack by a pathogen through the perception of elicitor signal molecules such as chitin oligosaccharide, or after exposure to UV irradiation. The enzyme, which catalyses the last step in the biosynthesis of momilactone A, can use both NAD⁺ and NADP⁺ but activity is higher with NAD⁺ [151].
References: [151, 3880]

[EC 1.1.1.295 created 2008]

EC 1.1.1.296

- Accepted name:** dihydrocarveol dehydrogenase
Reaction: menth-8-en-2-ol + NAD⁺ = menth-8-en-2-one + NADH + H⁺
Other name(s): carveol dehydrogenase (ambiguous)
Systematic name: menth-8-en-2-ol:NAD⁺ oxidoreductase
Comments: This enzyme from the Gram-positive bacterium *Rhodococcus erythropolis* DCL14 forms part of the carveol and dihydrocarveol degradation pathway. The enzyme accepts all eight stereoisomers of menth-8-en-2-ol as substrate, although some isomers are converted faster than others. The preferred substrates are (+)-neoisodihydrocarveol, (+)-isodihydrocarveol, (+)-dihydrocarveol and (-)-isodihydrocarveol.
References: [4403]

[EC 1.1.1.296 created 2008]

EC 1.1.1.297

- Accepted name:** limonene-1,2-diol dehydrogenase
Reaction: menth-8-ene-1,2-diol + NAD⁺ = 1-hydroxymenth-8-en-2-one + NADH + H⁺ (general reaction)
(1) (1*S*,2*S*,4*R*)-menth-8-ene-1,2-diol + NAD⁺ = (1*S*,4*R*)-1-hydroxymenth-8-en-2-one + NADH + H⁺
(2) (1*R*,2*R*,4*S*)-menth-8-ene-1,2-diol + NAD⁺ = (1*R*,4*S*)-1-hydroxymenth-8-en-2-one + NADH + H⁺
Other name(s): NAD⁺-dependent limonene-1,2-diol dehydrogenase
Systematic name: menth-8-ene-1,2-diol:NAD⁺ oxidoreductase
Comments: While the enzyme from the Gram-positive bacterium *Rhodococcus erythropolis* DCL14 can use both (1*S*,2*S*,4*R*)- and (1*R*,2*R*,4*S*)-menth-8-ene-1,2-diol as substrate, activity is higher with (1*S*,2*S*,4*R*)-menth-8-ene-1,2-diol as substrate.
References: [4404]

[EC 1.1.1.297 created 2008]

EC 1.1.1.298

- Accepted name:** 3-hydroxypropionate dehydrogenase (NADP⁺)
Reaction: 3-hydroxypropanoate + NADP⁺ = malonate semialdehyde + NADPH + H⁺
Other name(s): 3-hydroxypropanoate dehydrogenase (NADP⁺); 3-hydroxypropanoate:NADP⁺ oxidoreductase
Systematic name: 3-hydroxypropanoate:NADP⁺ oxidoreductase
Comments: Catalyses the reduction of malonate semialdehyde to 3-hydroxypropanoate, a key step in the 3-hydroxypropanoate and the 3-hydroxypropanoate/4-hydroxybutanoate cycles, autotrophic CO₂ fixation pathways found in some green non-sulfur phototrophic bacteria and archaea, respectively [4056, 296]. The enzyme from *Chloroflexus aurantiacus* is bifunctional, and also catalyses the upstream reaction in the pathway, EC 1.2.1.75 [1764]. Different from EC 1.1.1.59 [3-hydroxypropanoate dehydrogenase (NAD⁺)] by cofactor preference.
References: [4056, 296, 1764]

[EC 1.1.1.298 created 2009]

EC 1.1.1.299

- Accepted name:** malate dehydrogenase [NAD(P)⁺]
Reaction: (*S*)-malate + NAD(P)⁺ = oxaloacetate + NAD(P)H + H⁺
Other name(s): MdH II, NAD(P)⁺-dependent malate dehydrogenase
Systematic name: (*S*)-malate:NAD(P)⁺ oxidoreductase

Comments: This enzyme, which was characterized from the methanogenic archaeon *Methanobacterium thermoautotrophicum*, catalyses only the reduction of oxaloacetate, and can use NAD⁺ and NADP⁺ with similar specific activity [4271]. Different from EC 1.1.1.37 (malate dehydrogenase (NAD⁺)), EC 1.1.1.82 (malate dehydrogenase (NADP⁺)) and EC 1.1.5.4 (malate dehydrogenase (quinone)).

References: [4271]

[EC 1.1.1.299 created 2009]

EC 1.1.1.300

Accepted name: NADP-retinol dehydrogenase

Reaction: retinol + NADP⁺ = retinal + NADPH + H⁺

Other name(s): *all-trans* retinal reductase (ambiguous); *all-trans*-retinol dehydrogenase; NADP(H)-dependent retinol dehydrogenase/reductase; RDH11; RDH12; RDH13; RDH14; retinol dehydrogenase 12; retinol dehydrogenase 14; retinol dehydrogenase [NADP⁺]; RaIR1; PSDR1

Systematic name: retinol:NADP⁺ oxidoreductase

Comments: Greater catalytic efficiency in the reductive direction. This observation, and the enzyme's localization at the entrance to the mitochondrial matrix, suggest that it may function to protect mitochondria against oxidative stress associated with the highly reactive retinal produced from dietary β-carotene by EC 1.13.11.63 (β-carotene 15,15'-dioxygenase) [279]. *K_m*-values for NADP⁺ and NADPH are at least 800-fold lower than those for NAD⁺ and NADH [280, 2052]. This enzyme differs from EC 1.1.1.105, retinol dehydrogenase, which prefers NAD⁺ and NADH.

References: [280, 279, 1468, 2052]

[EC 1.1.1.300 created 2009]

EC 1.1.1.301

Accepted name: D-arabitol-phosphate dehydrogenase

Reaction: D-arabinitol 1-phosphate + NAD⁺ = D-xylulose 5-phosphate + NADH + H⁺

Other name(s): APDH; D-arabitol 1-phosphate dehydrogenase; D-arabitol 5-phosphate dehydrogenase; D-arabinitol 1-phosphate dehydrogenase; D-arabinitol 5-phosphate dehydrogenase

Systematic name: D-arabinitol-phosphate:NAD⁺ oxidoreductase

Comments: This enzyme participates in arabinitol catabolism. The enzyme also converts D-arabinitol 5-phosphate to D-ribulose 5-phosphate at a lower rate [3363].

References: [3363]

[EC 1.1.1.301 created 2010]

EC 1.1.1.302

Accepted name: 2,5-diamino-6-(ribosylamino)-4(3*H*)-pyrimidinone 5'-phosphate reductase

Reaction: 2,5-diamino-6-(5-phospho-D-ribitylamino)pyrimidin-4(3*H*)-one + NAD(P)⁺ = 2,5-diamino-6-(5-phospho-D-ribosylamino)pyrimidin-4(3*H*)-one + NAD(P)H + H⁺

Other name(s): 2,5-diamino-6-ribosylamino-4(3*H*)-pyrimidinone 5'-phosphate reductase; MjaRED; MJ0671 (gene name)

Systematic name: 2,5-diamino-6-(5-phospho-D-ribosylamino)pyrimidin-4(3*H*)-one:NAD(P)⁺ oxidoreductase

Comments: The reaction proceeds in the opposite direction. A step in riboflavin biosynthesis, NADPH and NADH function equally well as reductant. Differs from EC 1.1.1.193 [5-amino-6-(5-phosphoribosylamino)uracil reductase] since it does not catalyse the reduction of 5-amino-6-ribosylaminopyrimidine-2,4(1*H*,3*H*)-dione 5'-phosphate [1387].

References: [1387, 614]

[EC 1.1.1.302 created 2010, modified 2011]

EC 1.1.1.303

- Accepted name:** diacetyl reductase [(*R*)-acetoin forming]
Reaction: (*R*)-acetoin + NAD⁺ = diacetyl + NADH + H⁺
Other name(s): (*R*)-acetoin dehydrogenase
Systematic name: (*R*)-acetoin:NAD⁺ oxidoreductase
Comments: The reaction is catalysed in the reverse direction. This activity is usually associated with butanediol dehydrogenase activity (EC 1.1.1.4 or EC 1.1.1.76). While the butanediol dehydrogenase activity is reversible, diacetyl reductase activity is irreversible. This enzyme has been reported in the yeast *Saccharomyces cerevisiae* [1609, 1360]. Different from EC 1.1.1.304, diacetyl reductase [(*S*)-acetoin forming].
References: [1609, 1360]

[EC 1.1.1.303 created 2010 (EC 1.1.1.5 created 1961, modified 1976, part incorporated 2010)]

EC 1.1.1.304

- Accepted name:** diacetyl reductase [(*S*)-acetoin forming]
Reaction: (*S*)-acetoin + NAD⁺ = diacetyl + NADH + H⁺
Other name(s): (*S*)-acetoin dehydrogenase
Systematic name: (*S*)-acetoin:NAD⁺ oxidoreductase
Comments: The reaction is catalysed in the reverse direction. This activity is usually associated with butanediol dehydrogenase activity (EC 1.1.1.4 or EC 1.1.1.76). While the butanediol dehydrogenase activity is reversible, diacetyl reductase activity is irreversible. This enzyme has been reported in the bacteria *Geobacillus stearothermophilus*, *Enterobacter aerogenes* and *Klebsiella pneumoniae* [1328, 553, 4371]. Different from EC 1.1.1.303, diacetyl reductase [(*R*)-acetoin forming].
References: [1328, 553, 4371]

[EC 1.1.1.304 created 2010 (EC 1.1.1.5 created 1961, modified 1976, part incorporated 2010)]

EC 1.1.1.305

- Accepted name:** UDP-glucuronic acid dehydrogenase (UDP-4-keto-hexauronic acid decarboxylating)
Reaction: UDP- α -D-glucuronate + NAD⁺ = UDP- β -L-*threo*-pentapyranos-4-ulose + CO₂ + NADH + H⁺
Other name(s): UDP-GlcUA decarboxylase; ArnADH; UDP-glucuronate:NAD⁺ oxidoreductase (decarboxylating)
Systematic name: UDP- α -D-glucuronate:NAD⁺ oxidoreductase (decarboxylating)
Comments: The activity is part of a bifunctional enzyme also performing the reaction of EC 2.1.2.13 (UDP-4-amino-4-deoxy-L-arabinose formyltransferase).
References: [430, 1282, 4633, 1283, 4755]

[EC 1.1.1.305 created 2010]

EC 1.1.1.306

- Accepted name:** *S*-(hydroxymethyl)mycothiol dehydrogenase
Reaction: *S*-(hydroxymethyl)mycothiol + NAD⁺ = *S*-formylmycothiol + NADH + H⁺
Other name(s): NAD/factor-dependent formaldehyde dehydrogenase; mycothiol-dependent formaldehyde dehydrogenase
Systematic name: *S*-(hydroxymethyl)mycothiol:NAD⁺ oxidoreductase
Comments: *S*-hydroxymethylmycothiol is believed to form spontaneously from formaldehyde and mycothiol. This enzyme oxidizes the product of this spontaneous reaction to *S*-formylmycothiol, in a reaction that is analogous to EC 1.1.1.284, *S*-(hydroxymethyl)glutathione dehydrogenase.
References: [2826, 3107, 4459, 3462]

[EC 1.1.1.306 created 2010 as EC 1.2.1.66, transferred 2010 to EC 1.1.1.306]

EC 1.1.1.307

- Accepted name:** D-xylose reductase [NAD(P)H]
Reaction: xylitol + NAD(P)⁺ = D-xylose + NAD(P)H + H⁺
Other name(s): XylR; msXR; dsXR; dual specific xylose reductase; NAD(P)H-dependent xylose reductase; xylose reductase (ambiguous); D-xylose reductase (ambiguous)
Systematic name: xylitol:NAD(P)⁺ oxidoreductase
Comments: Xylose reductases catalyse the reduction of xylose to xylitol, the initial reaction in the fungal D-xylose degradation pathway. Most of the enzymes exhibit a strict requirement for NADPH [*cf.* EC 1.1.1.431, D-xylose reductase (NADPH)]. However, a few D-xylose reductases, such as those from *Neurospora crassa* [4668], *Yamadazyma tenuis* [3045, 1464], *Scheffersomyces stipitis* [4435], and the thermophilic fungus *Chaetomium thermophilum* [1480, 3414], have dual coenzyme specificity, though they still prefer NADPH to NADH. Very rarely the enzyme prefers NADH [*cf.* EC 1.1.1.430, D-xylose reductase (NADH)].
References: [4435, 3045, 1464, 1480, 4668, 1105, 3414]

[EC 1.1.1.307 created 2010, modified 2022]

EC 1.1.1.308

- Accepted name:** sulfopropanediol 3-dehydrogenase
Reaction: (R)-2,3-dihydroxypropane-1-sulfonate + 2 NAD⁺ + H₂O = (R)-3-sulfolactate + 2 NADH + 2 H⁺
Other name(s): DHPS 3-dehydrogenase (sulfolactate forming); 2,3-dihydroxypropane-1-sulfonate 3-dehydrogenase (sulfolactate forming); dihydroxypropanesulfonate 3-dehydrogenase; *hpsN* (gene name)
Systematic name: (R)-2,3-dihydroxypropane-1-sulfonate:NAD⁺ 3-oxidoreductase
Comments: The enzyme is involved in degradation of (R)-2,3-dihydroxypropanesulfonate.
References: [2731]

[EC 1.1.1.308 created 2011]

EC 1.1.1.309

- Accepted name:** phosphonoacetaldehyde reductase (NADH)
Reaction: 2-hydroxyethylphosphonate + NAD⁺ = phosphonoacetaldehyde + NADH + H⁺
Other name(s): PhpC
Systematic name: 2-hydroxyethylphosphonate:NAD⁺ oxidoreductase
Comments: The enzyme from *Streptomyces viridochromogenes* catalyses a step in the biosynthesis of phosphinothricin tripeptide, the reduction of phosphonoacetaldehyde to 2-hydroxyethylphosphonate. The preferred cofactor is NADH, lower activity with NADPH [362].
References: [362]

[EC 1.1.1.309 created 2011]

EC 1.1.1.310

- Accepted name:** (S)-sulfolactate dehydrogenase
Reaction: (2S)-3-sulfolactate + NAD⁺ = 3-sulfo-pyruvate + NADH + H⁺
Other name(s): (2S)-3-sulfolactate dehydrogenase; SlcC
Systematic name: (2S)-sulfolactate:NAD⁺ oxidoreductase
Comments: This enzyme, isolated from the bacterium *Chromohalobacter salexigens* DSM 3043, acts only on the (S)-enantiomer of 3-sulfolactate. Combined with EC 1.1.1.338, (2R)-3-sulfolactate dehydrogenase (NADP⁺), it provides a racemase system that converts (2S)-3-sulfolactate to (2R)-3-sulfolactate, which is degraded further by EC 4.4.1.24, (2R)-sulfolactate sulfo-lyase. The enzyme is specific for NAD⁺.
References: [879]

[EC 1.1.1.310 created 2011, modified 2013]

EC 1.1.1.311

Accepted name: (S)-1-phenylethanol dehydrogenase
Reaction: (S)-1-phenylethanol + NAD⁺ = acetophenone + NADH + H⁺
Other name(s): PED
Systematic name: (S)-1-phenylethanol:NAD⁺ oxidoreductase
Comments: The enzyme is involved in degradation of ethylbenzene.
References: [2168, 1686]

[EC 1.1.1.311 created 2011]

EC 1.1.1.312

Accepted name: 2-hydroxy-4-carboxymuconate semialdehyde hemiacetal dehydrogenase
Reaction: 4-carboxy-2-hydroxymuconate semialdehyde hemiacetal + NADP⁺ = 2-oxo-2H-pyran-4,6-dicarboxylate + NADPH + H⁺
Other name(s): 2-hydroxy-4-carboxymuconate 6-semialdehyde dehydrogenase; 4-carboxy-2-hydroxy-*cis,cis*-muconate-6-semialdehyde:NADP⁺ oxidoreductase; α -hydroxy- γ -carboxymuconic ϵ -semialdehyde dehydrogenase; 4-carboxy-2-hydroxymuconate-6-semialdehyde dehydrogenase; LigC; ProD
Systematic name: 4-carboxy-2-hydroxymuconate semialdehyde hemiacetal:NADP⁺ 2-oxidoreductase
Comments: The enzyme does not act on unsubstituted aliphatic or aromatic aldehydes or glucose; NAD⁺ can replace NADP⁺, but with lower affinity. The enzyme was initially believed to act on 4-carboxy-2-hydroxy-*cis,cis*-muconate 6-semialdehyde and produce 4-carboxy-2-hydroxy-*cis,cis*-muconate [2676]. However, later studies showed that the substrate is the hemiacetal form [2675], and the product is 2-oxo-2H-pyran-4,6-dicarboxylate [2674, 2679].
References: [2676, 2674, 2675, 2679]

[EC 1.1.1.312 created 1978 as EC 1.2.1.45, transferred 2011 to EC 1.1.1.312]

EC 1.1.1.313

Accepted name: sulfoacetaldehyde reductase (NADPH)
Reaction: isethionate + NADP⁺ = 2-sulfoacetaldehyde + NADPH + H⁺
Other name(s): *isfD* (gene name)
Systematic name: isethionate:NADP⁺ oxidoreductase
Comments: Catalyses the reaction only in the opposite direction. Involved in taurine degradation. The bacterium *Chromohalobacter salexigens* strain DSM 3043 possesses two enzymes that catalyse this reaction, a constitutive enzyme (encoded by *isfD2*) and an inducible enzyme (encoded by *isfD*). The latter is induced by taurine, and is responsible for most of the activity observed in taurine-grown cells. *cf.* EC 1.1.1.433, sulfoacetaldehyde reductase (NADH).
References: [2259]

[EC 1.1.1.313 created 2011, modified 2022]

[1.1.1.314 Deleted entry. *germacrene A alcohol dehydrogenase*. Now known to be catalyzed by EC 1.14.14.95, *germacrene A hydroxylase*]

[EC 1.1.1.314 created 2011, deleted 2018]

EC 1.1.1.315

Accepted name: 11-*cis*-retinol dehydrogenase
Reaction: 11-*cis*-retinol—[retinal-binding-protein] + NAD⁺ = 11-*cis*-retinal—[retinol-binding-protein] + NADH + H⁺
Other name(s): RDH5 (gene name)
Systematic name: 11-*cis*-retinol:NAD⁺ oxidoreductase

Comments: This enzyme, abundant in the retinal pigment epithelium, catalyses the reduction of 11-*cis*-retinol to 11-*cis*-retinal [3905] while the substrate is bound to the retinal-binding protein [4686]. This is a crucial step in the regeneration of 11-*cis*-retinal, the chromophore of rhodopsin. The enzyme can also accept other *cis* forms of retinol [4515].

References: [3905, 4515, 2480, 4686]

[EC 1.1.1.315 created 2011]

EC 1.1.1.316

Accepted name: L-galactose 1-dehydrogenase
Reaction: L-galactose + NAD⁺ = L-galactono-1,4-lactone + NADH + H⁺
Other name(s): L-GalDH; L-galactose dehydrogenase
Systematic name: L-galactose:NAD⁺ 1-oxidoreductase
Comments: The enzyme catalyses a step in the ascorbate biosynthesis in higher plants (Smirnoff-Wheeler pathway). The activity with NADP⁺ is less than 10% of the activity with NAD⁺.
References: [2795, 1281, 4598, 3136]

[EC 1.1.1.316 created 2011]

EC 1.1.1.317

Accepted name: perakine reductase
Reaction: raucaffrinoline + NADP⁺ = perakine + NADPH + H⁺
Systematic name: raucaffrinoline:NADP⁺ oxidoreductase
Comments: The biosynthesis of raucaffrinoline from perakine is a side route of the ajmaline biosynthesis pathway. The enzyme is a member of the aldo-keto reductase enzyme superfamily from higher plants.
References: [4118, 3576]

[EC 1.1.1.317 created 2011]

EC 1.1.1.318

Accepted name: eugenol synthase
Reaction: eugenol + a carboxylate + NADP⁺ = a coniferyl ester + NADPH + H⁺
Other name(s): LiCES1; EGS1; EGS2
Systematic name: eugenol:NADP⁺ oxidoreductase (coniferyl ester reducing)
Comments: The enzyme acts in the opposite direction. The enzymes from the plants *Ocimum basilicum* (sweet basil) [2185, 2543], *Clarkia breweri* and *Petunia hybrida* [2186] only accept coniferyl acetate and form eugenol. The enzyme from *Pimpinella anisum* (anise) forms anol (from 4-coumaryl acetate) *in vivo*, although the recombinant enzyme can form eugenol from coniferyl acetate [2184]. The enzyme from *Larrea tridentata* (creosote bush) also forms chavicol from a coumaryl ester and can use NADH [112].
References: [2185, 112, 2543, 2186, 2184]

[EC 1.1.1.318 created 2012]

EC 1.1.1.319

Accepted name: isoeugenol synthase
Reaction: isoeugenol + acetate + NADP⁺ = coniferyl acetate + NADPH + H⁺
Other name(s): IGS1; *t*-anol/isoeugenol synthase 1
Systematic name: eugenol:NADP⁺ oxidoreductase (coniferyl acetate reducing)
Comments: The enzyme acts in the opposite direction. In *Ocimum basilicum* (sweet basil), *Clarkia breweri* and *Petunia hybrida* only isoeugenol is formed [2185, 2186]. However in *Pimpinella anisum* (anise) only anol is formed *in vivo*, although the cloned enzyme does produce isoeugenol [2184].
References: [2185, 2186, 2184]

[EC 1.1.1.319 created 2012]

EC 1.1.1.320

Accepted name: benzil reductase [(*S*)-benzoin forming]
Reaction: (*S*)-benzoin + NADP⁺ = benzil + NADPH + H⁺
Other name(s): YueD
Systematic name: (*S*)-benzoin:NADP⁺ oxidoreductase
Comments: The enzyme also reduces 1-phenylpropane-1,2-dione. The enzyme from *Bacillus cereus* in addition reduces 1,4-naphthoquinone and 1-(4-methylphenyl)-2-phenylethane-1,2-dione with high efficiency [2678].
References: [2677, 2678]

[EC 1.1.1.320 created 2012]

EC 1.1.1.321

Accepted name: benzil reductase [(*R*)-benzoin forming]
Reaction: (*R*)-benzoin + NADP⁺ = benzil + NADPH + H⁺
Systematic name: (*R*)-benzoin:NADP⁺ oxidoreductase
Comments: The enzyme from the bacterium *Xanthomonas oryzae* is able to reduce enantioselectively only one of the two carbonyl groups of benzil to give optically active (*R*)-benzoin.
References: [2223]

[EC 1.1.1.321 created 2012]

EC 1.1.1.322

Accepted name: (–)-*endo*-fenchol dehydrogenase
Reaction: (–)-*endo*-fenchol + NAD(P)⁺ = (+)-fenchone + NAD(P)H + H⁺
Other name(s): *l-endo*-fenchol dehydrogenase; FDH
Systematic name: (–)-*endo*-fenchol:NAD(P)⁺ oxidoreductase
Comments: Isolated from the plant *Foeniculum vulgare* (fennel). NADH is slightly preferred to NADPH.
References: [772]

[EC 1.1.1.322 created 2012]

EC 1.1.1.323

Accepted name: (+)-thujan-3-ol dehydrogenase
Reaction: (+)-thujan-3-ol + NAD(P)⁺ = (+)-thujan-3-one + NAD(P)H + H⁺
Other name(s): *d*-3-thujanol dehydrogenase; TDH
Systematic name: (+)-thujan-3-ol:NAD(P)⁺ oxidoreductase
Comments: Isolated from the plant *Tanacetum vulgare* (tansy). NADH is preferred to NADPH.
References: [772]

[EC 1.1.1.323 created 2012]

EC 1.1.1.324

Accepted name: 8-hydroxygeraniol dehydrogenase
Reaction: (6*E*)-8-hydroxygeraniol + 2 NADP⁺ = (6*E*)-8-oxogeraniol + 2 NADPH + 2 H⁺ (overall reaction)
(1a) (6*E*)-8-hydroxygeraniol + NADP⁺ = (6*E*)-8-hydroxygeranial + NADPH + H⁺
(1b) (6*E*)-8-hydroxygeraniol + NADP⁺ = (6*E*)-8-oxogeraniol + NADPH + H⁺
(1c) (6*E*)-8-hydroxygeranial + NADP⁺ = (6*E*)-8-oxogeraniol + NADPH + H⁺
(1d) (6*E*)-8-oxogeraniol + NADP⁺ = (6*E*)-8-oxogeranial + NADPH + H⁺

Other name(s): 8-hydroxygeraniol oxidoreductase; CYP76B10; G10H; CrG10H; SmG10H; acyclic monoterpene primary alcohol:NADP⁺ oxidoreductase
Systematic name: (6*E*)-8-hydroxygeraniol:NADP⁺ oxidoreductase
Comments: Contains Zn²⁺. The enzyme catalyses the oxidation of (6*E*)-8-hydroxygeraniol to (6*E*)-8-oxogeraniol via either (6*E*)-8-hydroxygeraniol or (6*E*)-8-oxogeraniol. Also acts on geraniol, nerol and citronellol. May be identical to EC 1.1.1.183 geraniol dehydrogenase. The recommended numbering of geraniol gives 8-hydroxygeraniol as the substrate rather than 10-hydroxygeraniol as used by references 1 and 2. See prenol nomenclature Pr-1.
References: [1791, 1483]

[EC 1.1.1.324 created 2012]

EC 1.1.1.325

Accepted name: sepiapterin reductase (*L-threo*-7,8-dihydrobiopterin forming)
Reaction: (1) *L-threo*-7,8-dihydrobiopterin + NADP⁺ = sepiapterin + NADPH + H⁺
(2) *L-threo*-tetrahydrobiopterin + 2 NADP⁺ = 6-pyruvoyl-5,6,7,8-tetrahydropterin + 2 NADPH + 2 H⁺
Systematic name: *L-threo*-7,8-dihydrobiopterin:NADP⁺ oxidoreductase
Comments: This enzyme, isolated from the bacterium *Chlorobium tepidum*, catalyses the final step in the *de novo* synthesis of tetrahydrobiopterin from GTP. *cf.* EC 1.1.1.153, sepiapterin reductase (*L-erythro*-7,8-dihydrobiopterin forming).
References: [668, 4130]

[EC 1.1.1.325 created 2012]

EC 1.1.1.326

Accepted name: zerumbone synthase
Reaction: 10-hydroxy- α -humulene + NAD⁺ = zerumbone + NADH + H⁺
Other name(s): ZSD1
Systematic name: 10-hydroxy- α -humulene:NAD⁺ oxidoreductase
Comments: The enzyme was cloned from shampoo ginger, *Zingiber zerumbet*.
References: [3157]

[EC 1.1.1.326 created 2012]

EC 1.1.1.327

Accepted name: 5-*exo*-hydroxycamphor dehydrogenase
Reaction: 5-*exo*-hydroxycamphor + NAD⁺ = bornane-2,5-dione + NADH + H⁺
Other name(s): F-dehydrogenase; FdeH
Systematic name: 5-*exo*-hydroxycamphor:NAD⁺ oxidoreductase
Comments: Contains Zn²⁺. Isolated from *Pseudomonas putida*, and involved in degradation of (+)-camphor.
References: [3509, 2193, 122]

[EC 1.1.1.327 created 2012]

EC 1.1.1.328

Accepted name: nicotine blue oxidoreductase
Reaction: 3,3'-bipyridine-2,2',5,5',6,6'-hexol + NAD(P)⁺ = (*E*)-2,2',5,5'-tetrahydroxy-6*H*,6'*H*-[3,3'-bipyridinylidene]-6,6'-dione + NAD(P)H + H⁺
Other name(s): *nboR* (gene name)
Systematic name: 3,3'-bipyridine-2,2',5,5',6,6'-hexol:NADP⁺ 11-oxidoreductase

Comments: The enzyme, characterized from the nicotine degrading bacterium *Arthrobacter nicotinovorans*, catalyses the reduction of "nicotine blue" to its hydroquinone form (the opposite direction from that shown). Nicotine blue is the name given to the compound formed by the autocatalytic condensation of two molecules of 2,3,6-trihydroxypyridine, an intermediate in the nicotine degradation pathway. The main role of the enzyme may be to prevent the intracellular formation of nicotine blue semiquinone radicals, which by redox cycling would lead to the formation of toxic reactive oxygen species. The enzyme possesses a slight preference for NADH over NADPH.

References: [2801]

[EC 1.1.1.328 created 2012]

EC 1.1.1.329

Accepted name: 2-deoxy-*scyllo*-inosamine dehydrogenase
Reaction: 2-deoxy-*scyllo*-inosamine + NAD(P)⁺ = 3-amino-2,3-dideoxy-*scyllo*-inosose + NAD(P)H + H⁺
Other name(s): *neoA* (gene name); *kanK* (gene name, ambiguous); *kanE* (gene name, ambiguous)
Systematic name: 2-deoxy-*scyllo*-inosamine:NAD(P)⁺ 1-oxidoreductase
Comments: Requires zinc. Involved in the biosynthetic pathways of several clinically important aminocyclitol antibiotics, including kanamycin, neomycin and ribostamycin. *cf.* EC 1.1.99.38, 2-deoxy-*scyllo*-inosamine dehydrogenase (AdoMet-dependent).
References: [2274, 3042]

[EC 1.1.1.329 created 2012]

EC 1.1.1.330

Accepted name: very-long-chain 3-oxoacyl-CoA reductase
Reaction: a very-long-chain (3*R*)-3-hydroxyacyl-CoA + NADP⁺ = a very-long-chain 3-oxoacyl-CoA + NADPH + H⁺
Other name(s): very-long-chain 3-ketoacyl-CoA reductase; very-long-chain β-ketoacyl-CoA reductase; KCR (gene name); IFA38 (gene name)
Systematic name: (3*R*)-3-hydroxyacyl-CoA:NADP⁺ oxidoreductase
Comments: The second component of the elongase, a microsomal protein complex responsible for extending palmitoyl-CoA and stearoyl-CoA (and modified forms thereof) to very-long-chain acyl CoAs. The enzyme is active with substrates with chain length of C₁₆ to C₃₄, depending on the species. *cf.* EC 2.3.1.199, very-long-chain 3-oxoacyl-CoA synthase, EC 4.2.1.134, very-long-chain (3*R*)-3-hydroxyacyl-[acyl-carrier protein] dehydratase, and EC 1.3.1.93, very-long-chain enoyl-CoA reductase.
References: [257, 1495, 258]

[EC 1.1.1.330 created 2012]

EC 1.1.1.331

Accepted name: secoisolariciresinol dehydrogenase
Reaction: (–)-secoisolariciresinol + 2 NAD⁺ = (–)-matairesinol + 2 NADH + 2 H⁺
Systematic name: (–)-secoisolariciresinol:NAD⁺ oxidoreductase
Comments: Isolated from the plants *Forsythia intermedia* [4693] and *Podophyllum peltatum* [4693, 4827, 2859]. An intermediate lactol is detected *in vitro*.
References: [4693, 4827, 2859]

[EC 1.1.1.331 created 2012]

EC 1.1.1.332

Accepted name: chanoclavine-I dehydrogenase
Reaction: chanoclavine-I + NAD⁺ = chanoclavine-I aldehyde + NADH + H⁺

Other name(s): *easD* (gene name); *fgaDH* (gene name)
Systematic name: chanoclavine-I:NAD⁺ oxidoreductase
Comments: The enzyme catalyses a step in the pathway of ergot alkaloid biosynthesis in certain fungi.
References: [4505, 4504]

[EC 1.1.1.332 created 2012]

EC 1.1.1.333

Accepted name: decaprenylphospho- β -D-erythro-pentofuranosid-2-ulose 2-reductase
Reaction: *trans,octacis*-decaprenylphospho- β -D-arabinofuranose + NAD⁺ = *trans,octacis*-decaprenylphospho- β -D-erythro-pentofuranosid-2-ulose + NADH + H⁺
Other name(s): decaprenylphospho- β -D-ribofuranose 2'-epimerase; Rv3791; DprE2
Systematic name: *trans,octacis*-decaprenylphospho- β -D-arabinofuranose:NAD⁺ 2-oxidoreductase
Comments: The reaction is catalysed in the reverse direction. The enzyme, isolated from the bacterium *Mycobacterium smegmatis*, is involved, along with EC 1.1.98.3, decaprenylphospho- β -D-ribofuranose 2-oxidase, in the epimerization of *trans,octacis*-decaprenylphospho- β -D-ribofuranose to *trans,octacis*-decaprenylphospho- β -D-arabinoofuranose, the arabinosyl donor for the biosynthesis of mycobacterial cell wall arabinan polymers.
References: [4326]

[EC 1.1.1.333 created 2012]

EC 1.1.1.334

Accepted name: methylecgonone reductase
Reaction: ecgonine methyl ester + NADP⁺ = ecgonone methyl ester + NADPH + H⁺
Other name(s): MecgoR (gene name)
Systematic name: ecgonine methyl ester:NADP⁺ oxidoreductase
Comments: The enzyme from the plant *Erythroxylum coca* catalyses the penultimate step in the biosynthesis of cocaine. *In vivo* the reaction proceeds in the opposite direction. With NADH instead of NADPH the reaction rate is reduced to 14%. The enzyme also reduces tropinone, nortropinone and 6-hydroxytropinone but with lower reaction rates.
References: [1918]

[EC 1.1.1.334 created 2012]

EC 1.1.1.335

Accepted name: UDP-*N*-acetyl-2-amino-2-deoxyglucuronate dehydrogenase
Reaction: UDP-*N*-acetyl-2-amino-2-deoxy- α -D-glucuronate + NAD⁺ = UDP-2-acetamido-2-deoxy- α -D-ribohex-3-uluronate + NADH + H⁺
Other name(s): WlbA; WbpB
Systematic name: UDP-*N*-acetyl-2-amino-2-deoxy- α -D-glucuronate:NAD⁺ 3-oxidoreductase
Comments: This enzyme participates in the biosynthetic pathway for UDP- α -D-ManNAc3NAcA (UDP-2,3-diacetamido-2,3-dideoxy- α -D-mannuronic acid), an important precursor of B-band lipopolysaccharide. The enzymes from *Pseudomonas aeruginosa* serotype O5 and *Thermus thermophilus* form a complex with the the enzyme catalysing the next step the pathway (EC 2.6.1.98, UDP-2-acetamido-2-deoxy-ribohexuluronate aminotransferase). The enzyme also possesses an EC 1.1.99.2 (L-2-hydroxyglutarate dehydrogenase) activity, and utilizes the 2-oxoglutarate produced by EC 2.6.1.98 to regenerate the tightly bound NAD⁺. The enzymes from *Bordetella pertussis* and *Chromobacterium violaceum* do not bind NAD⁺ as tightly and do not require 2-oxoglutarate to function.
References: [4596, 2354, 4262, 4263]

[EC 1.1.1.335 created 2012]

EC 1.1.1.336

- Accepted name:** UDP-*N*-acetyl-D-mannosamine dehydrogenase
Reaction: UDP-*N*-acetyl- α -D-mannosamine + 2 NAD⁺ + H₂O = UDP-*N*-acetyl- α -D-mannosaminuronate + 2 NADH + 2 H⁺
Other name(s): UDP-ManNAc 6-dehydrogenase; *wecC* (gene name)
Systematic name: UDP-*N*-acetyl- α -D-mannosamine:NAD⁺ 6-oxidoreductase
Comments: Part of the pathway for acetamido sugar biosynthesis in bacteria and archaea. The enzyme has no activity with NADP⁺.
References: [3006]

[EC 1.1.1.336 created 2012]

EC 1.1.1.337

- Accepted name:** L-2-hydroxycarboxylate dehydrogenase (NAD⁺)
Reaction: a (2*S*)-2-hydroxycarboxylate + NAD⁺ = a 2-oxocarboxylate + NADH + H⁺
Other name(s): (*R*)-sulfolactate:NAD⁺ oxidoreductase; L-sulfolactate dehydrogenase; (*R*)-sulfolactate dehydrogenase; L-2-hydroxyacid dehydrogenase (NAD⁺); ComC
Systematic name: (2*S*)-2-hydroxycarboxylate:NAD⁺ oxidoreductase
Comments: The enzyme from the archaeon *Methanocaldococcus jannaschii* acts on multiple (*S*)-2-hydroxycarboxylates including (2*R*)-3-sulfolactate, (*S*)-malate, (*S*)-lactate, and (*S*)-2-hydroxyglutarate [1380]. Note that (2*R*)-3-sulfolactate has the same stereo configuration as (2*S*)-2-hydroxycarboxylates.
References: [1386, 1385, 1380, 3488]

[EC 1.1.1.337 created 2012]

EC 1.1.1.338

- Accepted name:** (2*R*)-3-sulfolactate dehydrogenase (NADP⁺)
Reaction: (2*R*)-3-sulfolactate + NADP⁺ = 3-sulfofopryuvate + NADPH + H⁺
Other name(s): (*R*)-sulfolactate:NADP⁺ oxidoreductase; L-sulfolactate dehydrogenase; (*R*)-sulfolactate dehydrogenase; ComC
Systematic name: (2*R*)-3-sulfolactate:NADP⁺ oxidoreductase
Comments: The enzyme from the bacterium *Chromohalobacter salexigens* can only utilize NADP⁺. It functions both biosynthetically in coenzyme M biosynthesis and degradatively, in the degradation of sulfolactate. It can not use (*S*)-malate and (*S*)-lactate.
References: [879]

[EC 1.1.1.338 created 2012]

EC 1.1.1.339

- Accepted name:** dTDP-6-deoxy-L-talose 4-dehydrogenase (NAD⁺)
Reaction: dTDP-6-deoxy- β -L-talose + NAD⁺ = dTDP-4-dehydro- β -L-rhamnose + NADH + H⁺
Other name(s): *tll* (gene name)
Systematic name: dTDP-6-deoxy- β -L-talose:NAD⁺ 4-oxidoreductase
Comments: The enzyme has been characterized from the bacterium *Aggregatibacter actinomycetemcomitans*, in which it participates in the biosynthesis of the serotype c-specific polysaccharide antigen. Shows no activity with NADP⁺.
References: [2994]

[EC 1.1.1.339 created 2012]

EC 1.1.1.340

- Accepted name:** 1-deoxy-11 β -hydroxypentalenate dehydrogenase

Reaction: 1-deoxy-11 β -hydroxypentalenate + NAD⁺ = 1-deoxy-11-oxopentalenate + NADH + H⁺
Other name(s): 1-deoxy-11 β -hydroxypentalenic acid dehydrogenase; *ptlF* (gene name); *penF* (gene name)
Systematic name: 1-deoxy-11 β -hydroxypentalenate:NAD⁺ oxidoreductase
Comments: Isolated from the bacterium *Streptomyces avermitilis* and present in many other *Streptomyces* species. Part of the pathway for pentalenolactone biosynthesis.
References: [4823]

[EC 1.1.1.340 created 2012]

EC 1.1.1.341

Accepted name: CDP-abequose synthase
Reaction: CDP- α -D-abequose + NADP⁺ = CDP-4-dehydro-3,6-dideoxy- α -D-glucose + NADPH + H⁺
Other name(s): *rfbJ* (gene name)
Systematic name: CDP- α -D-abequose:NADP⁺ 4-oxidoreductase
Comments: Isolated from *Yersinia pseudotuberculosis* [2074, 4278] and *Salmonella enterica* [2074, 4689].
References: [2074, 4689, 4278]

[EC 1.1.1.341 created 2012]

EC 1.1.1.342

Accepted name: CDP-paratose synthase
Reaction: CDP- α -D-paratose + NADP⁺ = CDP-4-dehydro-3,6-dideoxy- α -D-glucose + NADPH + H⁺
Other name(s): *rfbS* (gene name)
Systematic name: CDP- α -D-paratose:NADP⁺ 4-oxidoreductase
Comments: The enzyme is involved in synthesis of paratose and tyvelose, unusual 3,6-dideoxyhexose sugars that form part of the O-antigen in the lipopolysaccharides of several enteric bacteria. Isolated from *Salmonella enterica* subsp. *enterica* serovar Typhi (*Salmonella typhi*).
References: [4439, 1485]

[EC 1.1.1.342 created 2012]

EC 1.1.1.343

Accepted name: phosphogluconate dehydrogenase (NAD⁺-dependent, decarboxylating)
Reaction: 6-phospho-D-gluconate + NAD⁺ = D-ribulose 5-phosphate + CO₂ + NADH + H⁺
Other name(s): 6-PGDH (ambiguous); *gntZ* (gene name); GNDI
Systematic name: 6-phospho-D-gluconate:NAD⁺ 2-oxidoreductase (decarboxylating)
Comments: Highly specific for NAD⁺. The enzyme catalyses both the oxidation and decarboxylation of 6-phospho-D-gluconate. In the bacterium *Methylobacillus flagellatus* the enzyme participates in a formaldehyde oxidation pathway [658]. *cf.* EC 1.1.1.44, phosphogluconate dehydrogenase (NADP⁺-dependent, decarboxylating).
References: [2127, 3137, 4857, 658]

[EC 1.1.1.343 created 2013]

EC 1.1.1.344

Accepted name: dTDP-6-deoxy-L-talose 4-dehydrogenase [NAD(P)⁺]
Reaction: dTDP-6-deoxy- β -L-talose + NAD(P)⁺ = dTDP-4-dehydro- β -L-rhamnose + NAD(P)H + H⁺
Other name(s): *tal* (gene name)
Systematic name: dTDP-6-deoxy- β -L-talose:NAD(P)⁺ 4-oxidoreductase
Comments: The enzyme works equally well with NAD⁺ and NADP⁺.
References: [1994]

[EC 1.1.1.344 created 2013]

EC 1.1.1.345

- Accepted name:** D-2-hydroxyacid dehydrogenase (NAD⁺)
Reaction: an (*R*)-2-hydroxycarboxylate + NAD⁺ = a 2-oxocarboxylate + NADH + H⁺
Other name(s): LdhA; HdhD; D-2-hydroxyisocaproate dehydrogenase; R-HicDH; D-HicDH; (*R*)-2-hydroxy-4-methylpentanoate:NAD⁺ oxidoreductase; (*R*)-2-hydroxyisocaproate dehydrogenase; D-mandelate dehydrogenase (ambiguous)
Systematic name: (*R*)-2-hydroxycarboxylate:NAD⁺ oxidoreductase
Comments: The enzymes, characterized from bacteria (*Peptoclostridium difficile*, *Enterococcus faecalis* and from lactic acid bacteria) prefer substrates with a main chain of 5 carbons (such as 4-methyl-2-oxopentanoate) to those with a shorter chain. It also utilizes phenylpyruvate. The enzyme from the halophilic archaeon *Haloferax mediterranei* prefers substrates with a main chain of 3-4 carbons (pyruvate and 2-oxobutanoate). *cf.* EC 1.1.1.272, (*d*)-2-hydroxyacid dehydrogenase (NADP⁺).
References: [881, 383, 2094, 4484, 589, 2838]

[EC 1.1.1.345 created 2013]

EC 1.1.1.346

- Accepted name:** 2,5-didehydrogluconate reductase (2-dehydro-L-gulonate-forming)
Reaction: 2-dehydro-L-gulonate + NADP⁺ = 2,5-didehydro-D-gluconate + NADPH + H⁺
Other name(s): 2,5-diketo-D-gluconate-reductase (ambiguous); YqhE reductase; *dkgA* (gene name); *dkgB* (gene name)
Systematic name: 2-dehydro-D-gluconate:NADP⁺ 2-oxidoreductase (2-dehydro-L-gulonate-forming)
Comments: The enzyme is involved in ketogluconate metabolism, and catalyses the reaction *in vivo* in the reverse direction to that shown [3966]. It is used in the commercial microbial production of ascorbate. *cf.* EC 1.1.1.274, 2,5-didehydrogluconate reductase (2-dehydro-D-gluconate-forming).
References: [3966, 2809, 4845, 2648, 2085]

[EC 1.1.1.346 created 2013]

EC 1.1.1.347

- Accepted name:** geraniol dehydrogenase (NAD⁺)
Reaction: geraniol + NAD⁺ = geranial + NADH + H⁺
Other name(s): GeDH; *geoA* (gene name)
Systematic name: geraniol:NAD⁺ oxidoreductase
Comments: The enzyme from the bacterium *Castellaniella defragrans* is most active *in vitro* with perillyl alcohol [2556]. The enzyme from the prune mite *Carpoglyphus lactis* also acts (more slowly) on farnesol but not on nerol [3095].
References: [3095, 2556]

[EC 1.1.1.347 created 2013]

EC 1.1.1.348

- Accepted name:** (3*R*)-2'-hydroxyisoflavanone reductase
Reaction: a (4*R*)-4,2'-dihydroxyisoflavan + NADP⁺ = a (3*R*)-2'-hydroxyisoflavanone + NADPH + H⁺
Other name(s): vestitone reductase; pterocarpin synthase (incorrect); pterocarpin synthase (incorrect)
Systematic name: (3*R*)-2'-hydroxyisoflavanone:NADP⁺ 4-oxidoreductase
Comments: This plant enzyme participates in the biosynthesis of the pterocarpin phytoalexins medicarpin, maackiain, and several forms of glyceollin. The enzyme has a strict stereo specificity for the 3*R*-isoflavanones.
References: [360, 1446, 1447, 1448, 3821]

[EC 1.1.1.348 created 1992 as EC 1.1.1.246, part transferred 2013 to EC 1.1.1.348]

EC 1.1.1.349

Accepted name: norsolorinic acid ketoreductase
Reaction: (1'*S*)-averantin + NADP⁺ = norsolorinic acid + NADPH + H⁺
Other name(s): *aflD* (gene name); nor-1 (gene name)
Systematic name: (1'*S*)-averantin:NADP⁺ oxidoreductase
Comments: Involved in the synthesis of aflatoxins in the fungus *Aspergillus parasiticus*.
References: [4717, 4916]

[EC 1.1.1.349 created 2013]

EC 1.1.1.350

Accepted name: ureidoglycolate dehydrogenase (NAD⁺)
Reaction: (*S*)-ureidoglycolate + NAD⁺ = *N*-carbamoyl-2-oxoglycine + NADH + H⁺
Systematic name: (*S*)-ureidoglycolate:NAD⁺ oxidoreductase
Comments: Involved in catabolism of purines. The enzyme from the bacterium *Escherichia coli* is specific for NAD⁺ [2102]. *cf.* EC 1.1.1.154, ureidoglycolate dehydrogenase [NAD(P)⁺].
References: [789, 2102]

[EC 1.1.1.350 created 2013]

EC 1.1.1.351

Accepted name: phosphogluconate dehydrogenase [NAD(P)⁺-dependent, decarboxylating]
Reaction: 6-phospho-D-gluconate + NAD(P)⁺ = D-ribulose 5-phosphate + CO₂ + NAD(P)H + H⁺
Systematic name: 6-phospho-D-gluconate:NAD(P)⁺ 2-oxidoreductase (decarboxylating)
Comments: The enzyme participates in the oxidative branch of the pentose phosphate pathway, whose main purpose is to produce reducing power and pentose for biosynthetic reactions. Unlike EC 1.1.1.44, phosphogluconate dehydrogenase (NADP⁺-dependent, decarboxylating), it is not specific for NADP⁺ and can accept both cofactors with similar efficiency. *cf.* EC 1.1.1.343, phosphogluconate dehydrogenase [NAD⁺-dependent, decarboxylating].
References: [282, 4052, 2436]

[EC 1.1.1.351 created 2013]

EC 1.1.1.352

Accepted name: 5'-hydroxyaverantin dehydrogenase
Reaction: (1) (1'*S*,5'*S*)-hydroxyaverantin + NAD⁺ = 5'-oxoaverantin + NADH + H⁺
(2) (1'*S*,5'*R*)-hydroxyaverantin + NAD⁺ = 5'-oxoaverantin + NADH + H⁺
Other name(s): HAVN dehydrogenase; *adhA* (gene name)
Systematic name: (1'*S*,5'*S*)-hydroxyaverantin:NAD⁺ oxidoreductase
Comments: Isolated from the aflatoxin-producing mold *Aspergillus parasiticus* [3645]. Involved in aflatoxin biosynthesis. 5'-Oxoaverantin will spontaneously form averufin by intramolecular ketalisation. *cf.* EC 4.2.1.142, 5'-oxoaverantin cyclase.
References: [594, 3645]

[EC 1.1.1.352 created 2013]

EC 1.1.1.353

Accepted name: versiconal hemiacetal acetate reductase
Reaction: (1) versicolorone + NADP⁺ = 1'-hydroxyversicolorone + NADPH + H⁺
(2) versiconol acetate + NADP⁺ = versiconal hemiacetal acetate + NADPH + H⁺
(3) versiconol + NADP⁺ = versiconal + NADPH + H⁺
Other name(s): VHA reductase; VHA reductase I; VHA reductase II; *vrda* (gene name)
Systematic name: versiconol-acetate:NADP⁺ oxidoreductase

Comments: Isolated from the mold *Aspergillus parasiticus*. Involved in a metabolic grid that leads to aflatoxin biosynthesis.

References: [2711, 3862]

[EC 1.1.1.353 created 2013]

EC 1.1.1.354

Accepted name: farnesol dehydrogenase (NAD⁺)

Reaction: (2*E*,6*E*)-farnesol + NAD⁺ = (2*E*,6*E*)-farnesal + NADH + H⁺

Other name(s): NAD⁺-farnesol dehydrogenase

Systematic name: (2*E*,6*E*)-farnesol:NAD⁺ 1-oxidoreductase

Comments: The enzyme from the prune mite *Carpoglyphus lactis* also acts on geraniol with greater activity [*cf.* EC 1.1.1.347, geraniol dehydrogenase (NAD⁺)]. Unlike EC 1.1.1.216, farnesol dehydrogenase (NADP⁺), this enzyme cannot use NADP⁺ as cofactor.

References: [3095]

[EC 1.1.1.354 created 2013]

EC 1.1.1.355

Accepted name: 2'-dehydrokanamycin reductase

Reaction: kanamycin A + NADP⁺ = 2'-dehydrokanamycin A + NADPH + H⁺

Other name(s): *kanK* (gene name, ambiguous)

Systematic name: kanamycin A:NADP⁺ oxidoreductase

Comments: Found in the bacterium *Streptomyces kanamyceticus* where it is involved in the conversion of kanamycin B to kanamycin A.

References: [4089]

[EC 1.1.1.355 created 2013]

EC 1.1.1.356

Accepted name: GDP-L-colitose synthase

Reaction: GDP-β-L-colitose + NAD(P)⁺ = GDP-4-dehydro-3,6-dideoxy-α-D-mannose + NAD(P)H + H⁺

Other name(s): ColC

Systematic name: GDP-β-L-colitose:NAD(P)⁺ 4-oxidoreductase (5-epimerizing)

Comments: The enzyme is involved in biosynthesis of L-colitose, a 3,6-dideoxyhexose found in the O-antigen of Gram-negative lipopolysaccharides, where it catalyses the reaction in the reverse direction. The enzyme also performs the NAD(P)H-dependent epimerisation at C-5 of the sugar. The enzyme from *Yersinia pseudotuberculosis* is *Si*-specific with respect to NAD(P)H [55].

References: [55]

[EC 1.1.1.356 created 2013]

EC 1.1.1.357

Accepted name: 3α-hydroxysteroid 3-dehydrogenase

Reaction: a 3α-hydroxysteroid + NAD(P)⁺ = a 3-oxosteroid + NAD(P)H + H⁺

Other name(s): 3α-hydroxysteroid dehydrogenase; AKR1C4 (gene name); AKR1C2 (gene name); *hSDA* (gene name)

Systematic name: 3α-hydroxysteroid:NAD(P)⁺ 3-oxidoreductase

Comments: The enzyme acts on multiple 3α-hydroxysteroids, such as androsterone and 5 α-dihydrotestosterone. The mammalian enzymes are involved in inactivation of steroid hormones, while the bacterial enzymes are involved in steroid degradation. This entry stands for enzymes whose stereo-specificity with respect to NAD⁺ or NADP⁺ is not known. [*cf.* EC 1.1.1.50, 3α-hydroxysteroid 3-dehydrogenase (*Si*-specific) and EC 1.1.1.213, 3α-hydroxysteroid 3-dehydrogenase (*Re*-specific)].

References: [898, 2080, 3189, 2854, 2966]

[EC 1.1.1.357 created 2013]

EC 1.1.1.358

Accepted name: 2-dehydropantolactone reductase
Reaction: (*R*)-pantolactone + NADP⁺ = 2-dehydropantolactone + NADPH + H⁺
Other name(s): 2-oxopantoyl lactone reductase; 2-ketopantoyl lactone reductase; ketopantoyl lactone reductase; 2-dehydropantoyl-lactone reductase
Systematic name: (*R*)-pantolactone:NADP⁺ oxidoreductase
Comments: The enzyme participates in an alternative pathway for biosynthesis of (*R*)-pantothenate (vitamin B₅). This entry covers enzymes whose stereo specificity for NADP⁺ is not known. *cf.* EC 1.1.1.168 2-dehydropantolactone reductase (*Re*-specific) and EC 1.1.1.214, 2-dehydropantolactone reductase (*Si*-specific).
References: [1556]

[EC 1.1.1.358 created 2013]

EC 1.1.1.359

Accepted name: aldose 1-dehydrogenase [NAD(P)⁺]
Reaction: an aldopyranose + NAD(P)⁺ = an aldono-1,5-lactone + NAD(P)H + H⁺
Systematic name: an aldopyranose:NAD(P)⁺ 1-oxidoreductase
Comments: The enzyme from the archaeon *Sulfolobus solfataricus* shows broad specificity towards aldoses (D-glucose, D-galactose, D-xylose, L-arabinose, 6-deoxy-D-glucose, D-fucose) and can utilize NAD⁺ and NADP⁺ with similar catalytic efficiency. It is involved in aldose catabolism via the branched variant of the Entner-Doudoroff pathway.
References: [1317, 3935, 2334, 4253, 2806, 1469]

[EC 1.1.1.359 created 2013]

EC 1.1.1.360

Accepted name: glucose/galactose 1-dehydrogenase
Reaction: (1) D-glucopyranose + NADP⁺ = D-glucono-1,5-lactone + NADPH + H⁺
(2) D-galactopyranose + NADP⁺ = D-galactono-1,5-lactone + NADPH + H⁺
Other name(s): GdhA; dual-specific glucose/galactose dehydrogenase; glucose (galactose) dehydrogenase; glucose/galactose dehydrogenase
Systematic name: D-glucose/D-galactose 1-dehydrogenase (NADPH)
Comments: A zinc protein. The enzyme from the archaeon *Picrophilus torridus* is involved in glucose and galactose catabolism via the nonphosphorylative variant of the Entner-Doudoroff pathway. It shows 20-fold higher activity with NADP⁺ compared to NAD⁺. The oxidation of D-glucose and D-galactose is catalysed at a comparable rate (*cf.* EC 1.1.1.119, glucose 1-dehydrogenase (NADP⁺) and EC 1.1.1.120, galactose 1-dehydrogenase (NADP⁺)).
References: [103, 2806]

[EC 1.1.1.360 created 2013]

EC 1.1.1.361

Accepted name: glucose-6-phosphate 3-dehydrogenase
Reaction: D-glucose 6-phosphate + NAD⁺ = 3-dehydro-D-glucose 6-phosphate + NADH + H⁺
Other name(s): *ntdC* (gene name)
Systematic name: D-glucose-6-phosphate:NAD⁺ oxidoreductase
Comments: The enzyme, found in the bacterium *Bacillus subtilis*, is involved in a kanosamine biosynthesis pathway.
References: [4441]

[EC 1.1.1.361 created 2013]

EC 1.1.1.362

- Accepted name:** aklaviketone reductase
Reaction: aklavinone + NADP⁺ = aklaviketone + NADPH + H⁺
Other name(s): *dauE* (gene name); *aknU* (gene name)
Systematic name: aklavinone:NADP⁺ oxidoreductase
Comments: The enzyme is involved in the synthesis of the aklavinone aglycone, a common precursor for several anthracycline antibiotics including aclacinomycins, daunorubicin and doxorubicin. The enzyme from the Gram-negative bacterium *Streptomyces* sp. C5 produces daunomycin.
References: [905]

[EC 1.1.1.362 created 2013]

EC 1.1.1.363

- Accepted name:** glucose-6-phosphate dehydrogenase [NAD(P)⁺]
Reaction: D-glucose 6-phosphate + NAD(P)⁺ = 6-phospho-D-glucono-1,5-lactone + NAD(P)H + H⁺
Other name(s): G6PDH; G6PD; Glc6PD
Systematic name: D-glucose-6-phosphate:NAD(P)⁺ 1-oxidoreductase
Comments: The enzyme catalyses a step of the pentose phosphate pathway. The enzyme from the Gram-positive bacterium *Leuconostoc mesenteroides* prefers NADP⁺ while the enzyme from the Gram-negative bacterium *Gluconacetobacter xylinus* prefers NAD⁺. *cf.* EC 1.1.1.49, glucose-6-phosphate dehydrogenase (NADP⁺) and EC 1.1.1.388, glucose-6-phosphate dehydrogenase (NAD⁺).
References: [3168, 2399, 744, 3425]

[EC 1.1.1.363 created 2013, modified 2015]

EC 1.1.1.364

- Accepted name:** dTDP-4-dehydro-6-deoxy- α -D-gulose 4-ketoreductase
Reaction: dTDP-6-deoxy- α -D-allose + NAD(P)⁺ = dTDP-4-dehydro-6-deoxy- α -D-gulose + NAD(P)H + H⁺
Other name(s): dTDP-4-dehydro-6-deoxygulose reductase; *tylD* (gene name); *gerKI* (gene name); *chmD* (gene name); *mydI* (gene name)
Systematic name: dTDP-6-deoxy- α -D-allose:NAD(P)⁺ oxidoreductase
Comments: The enzyme forms an activated deoxy- α -D-allose, which is converted to mycinose after attachment to the aglycones of several macrolide antibiotics, including tylosin, chalcomycin, dihydrochalcomycin, and mycinamicin II.
References: [235, 111, 4280, 2271]

[EC 1.1.1.364 created 2013]

EC 1.1.1.365

- Accepted name:** D-galacturonate reductase
Reaction: L-galactonate + NADP⁺ = D-galacturonate + NADPH + H⁺
Other name(s): GalUR; *gar1* (gene name)
Systematic name: L-galactonate:NADP⁺ oxidoreductase
Comments: The enzyme from plants is involved in ascorbic acid (vitamin C) biosynthesis [1825, 35]. The enzyme from the fungus *Trichoderma reesei* (*Hypocrea jecorina*) is involved in a eukaryotic degradation pathway of D-galacturonate. It is also active with D-galucuronate and glyceraldehyde [2296]. Neither enzyme shows any activity with NADH.
References: [1825, 35, 2296, 2664]

[EC 1.1.1.365 created 2013]

EC 1.1.1.366

- Accepted name:** L-idonate 5-dehydrogenase (NAD⁺)

Reaction: L-idonate + NAD⁺ = 5-dehydro-D-gluconate + NADH + H⁺
Systematic name: L-idonate:NAD⁺ oxidoreductase
Comments: Involved in the catabolism of ascorbate (vitamin C) to tartrate. No activity is observed with NADP⁺ (*cf.* EC 1.1.1.264, L-idonate 5-dehydrogenase).
References: [857]

[EC 1.1.1.366 created 2013]

EC 1.1.1.367

Accepted name: UDP-2-acetamido-2,6-β-L-*arabino*-hexul-4-ose reductase
Reaction: UDP-2-acetamido-2,6-dideoxy-β-L-talose + NAD(P)⁺ = UDP-2-acetamido-2,6-β-L-*arabino*-hexul-4-ose + NAD(P)H + H⁺
Other name(s): WbjC; Cap5F
Systematic name: UDP-2-acetamido-2,6-dideoxy-L-talose:NADP⁺ oxidoreductase
Comments: Part of the biosynthesis of UDP-*N*-acetyl-L-fucosamine. Isolated from the bacteria *Pseudomonas aeruginosa* and *Staphylococcus aureus*.
References: [2166, 2932, 2837]

[EC 1.1.1.367 created 2014]

EC 1.1.1.368

Accepted name: 6-hydroxycyclohex-1-ene-1-carbonyl-CoA dehydrogenase
Reaction: 6-hydroxycyclohex-1-ene-1-carbonyl-CoA + NAD⁺ = 6-oxocyclohex-1-ene-1-carbonyl-CoA + NADH + H⁺
Systematic name: 6-hydroxycyclohex-1-ene-1-carbonyl-CoA:NAD⁺ 6-oxidoreductase
Comments: The enzyme participates in the central benzoyl-CoA degradation pathway of some anaerobic bacteria such as *Thauera aromatica*.
References: [2325]

[EC 1.1.1.368 created 2014]

EC 1.1.1.369

Accepted name: D-*chiro*-inositol 1-dehydrogenase
Reaction: 1D-*chiro*-inositol + NAD⁺ = 2D-2,3,5/4,6-pentahydroxycyclohexanone + NADH + H⁺
Other name(s): DCI 1-dehydrogenase; lolG
Systematic name: 1D-*chiro*-inositol:NAD⁺ 1-oxidoreductase
Comments: The enzyme, found in the bacterium *Bacillus subtilis*, also catalyses the reaction of EC 1.1.1.18, inositol 2-dehydrogenase, and can also use D-glucose and D-xylose. It shows trace activity with D-ribose and D-fructose [3444]. It is part of a *myo*-inositol/D-*chiro*-inositol degradation pathway leading to acetyl-CoA.
References: [3444, 4808]

[EC 1.1.1.369 created 2014]

EC 1.1.1.370

Accepted name: *scyllo*-inositol 2-dehydrogenase (NAD⁺)
Reaction: *scyllo*-inositol + NAD⁺ = 2,4,6/3,5-pentahydroxycyclohexanone + NADH + H⁺
Other name(s): *iolX* (gene name)
Systematic name: *scyllo*-inositol:NAD⁺ 2-oxidoreductase
Comments: The enzyme, found in the bacterium *Bacillus subtilis*, has no activity with NADP⁺ [*cf.* EC 1.1.1.371, *scyllo*-inositol 2-dehydrogenase (NADP⁺)]. It is part of a *scyllo*-inositol degradation pathway leading to acetyl-CoA.
References: [2891]

[EC 1.1.1.370 created 2014]

EC 1.1.1.371

- Accepted name:** *scyllo*-inositol 2-dehydrogenase (NADP⁺)
Reaction: *scyllo*-inositol + NADP⁺ = 2,4,6/3,5-pentahydroxycyclohexanone + NADPH + H⁺
Other name(s): *iolW* (gene name)
Systematic name: *scyllo*-inositol:NADP⁺ 2-oxidoreductase
Comments: The enzyme, found in the bacterium *Bacillus subtilis*, has no activity with NAD⁺ [cf. EC 1.1.1.370, *scyllo*-inositol 2-dehydrogenase (NAD⁺)].
References: [2891]

[EC 1.1.1.371 created 2014]

EC 1.1.1.372

- Accepted name:** D/L-glyceraldehyde reductase
Reaction: (1) glycerol + NADP⁺ = L-glyceraldehyde + NADPH + H⁺
(2) glycerol + NADP⁺ = D-glyceraldehyde + NADPH + H⁺
Other name(s): *gld1* (gene name); *gaaD* (gene name)
Systematic name: glycerol:NADP⁺ oxidoreductase (D/L-glyceraldehyde-forming)
Comments: The enzyme takes part in a D-galacturonate degradation pathway in the fungi *Aspergillus niger* and *Trichoderma reesei* (*Hypocrea jecorina*). It has equal activity with D- and L-glyceraldehyde, and can also reduce glyoxal and methylglyoxal. The reaction is only observed in the direction of glyceraldehyde reduction.
References: [2482, 2664]

[EC 1.1.1.372 created 2014]

EC 1.1.1.373

- Accepted name:** sulfolactaldehyde 3-reductase
Reaction: (2*S*)-2,3-dihydroxypropane-1-sulfonate + NAD⁺ = (2*S*)-3-sulfolactaldehyde + NADH + H⁺
Other name(s): *yihU* (gene name)
Systematic name: (2*S*)-2,3-dihydroxypropane-1-sulfonate:NAD⁺ 3-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Escherichia coli*, is involved in the degradation pathway of sulfoquinovose, the polar headgroup of sulfolipids found in the photosynthetic membranes of all higher plants, mosses, ferns, algae, and most photosynthetic bacteria, as well as the surface layer of some archaea.
References: [880, 3826]

[EC 1.1.1.373 created 2014]

EC 1.1.1.374

- Accepted name:** UDP-*N*-acetylglucosamine 3-dehydrogenase
Reaction: UDP-*N*-acetyl- α -D-glucosamine + NAD⁺ = UDP-2-acetamido-3-dehydro-2-deoxy- α -D-glucopyranose + NADH + H⁺
Systematic name: UDP-*N*-acetyl- α -D-glucosamine:NAD⁺ 3-oxidoreductase
Comments: The enzyme from the archaeon *Methanococcus maripaludis* is activated by KCl (200 mM).
References: [3007]

[EC 1.1.1.374 created 2014]

EC 1.1.1.375

- Accepted name:** L-2-hydroxycarboxylate dehydrogenase [NAD(P)⁺]

Reaction: a (2*S*)-2-hydroxycarboxylate + NAD(P)⁺ = a 2-oxocarboxylate + NAD(P)H + H⁺
Other name(s): MdhII; lactate/malate dehydrogenase
Systematic name: (2*S*)-2-hydroxycarboxylate:NAD(P)⁺ oxidoreductase
Comments: The enzyme from the archaeon *Methanocaldococcus jannaschii* catalyses the reversible oxidation of (2*R*)-3-sulfolactate and (*S*)-malate to 3-sulfofopuvate and oxaloacetate, respectively (note that (2*R*)-3-sulfolactate has the same stereochemical configuration as (2*S*)-2-hydroxycarboxylates) [1386]. The enzyme can use both NADH and NADPH, although activity is higher with NADPH [1386, 2378, 2600]. The oxidation of (2*R*)-3-sulfolactate was observed only in the presence of NADP⁺ [1386]. The same organism also possesses an NAD⁺-specific enzyme with similar activity, cf. EC 1.1.1.337, L-2-hydroxycarboxylate dehydrogenase (NAD⁺).
References: [1386, 2378, 2600]

[EC 1.1.1.375 created 2014]

EC 1.1.1.376

Accepted name: L-arabinose 1-dehydrogenase [NAD(P)⁺]
Reaction: α-L-arabinopyranose + NAD(P)⁺ = L-arabinono-1,4-lactone + NAD(P)H + H⁺
Other name(s): L-*arabino*-aldose dehydrogenase
Systematic name: α-L-arabinopyranose:NAD(P)⁺ 1-oxidoreductase
Comments: The enzymes from the bacterium *Azospirillum brasilense* and the archaeon *Haloferax volcanii* are part of the L-arabinose degradation pathway and prefer NADP⁺ over NAD⁺. *In vitro* the enzyme from *Azospirillum brasilense* shows also high catalytic efficiency with D-galactose. The enzyme is specific for α-L-arabinopyranose [1927, 135].
References: [3111, 4557, 1927, 135]

[EC 1.1.1.376 created 2014, modified 2022]

EC 1.1.1.377

Accepted name: L-rhamnose 1-dehydrogenase (NADP⁺)
Reaction: L-rhamnose + NADP⁺ = L-rhamnono-1,4-lactone + NADPH + H⁺
Systematic name: L-rhamnose:NADP⁺ 1-oxidoreductase
Comments: The enzyme from the archaeon *Thermoplasma acidophilum* is part of the non-phosphorylative degradation pathway for L-rhamnose. The enzyme differs in cofactor specificity from EC 1.1.1.173, L-rhamnose 1-dehydrogenase, which is specific for NAD⁺.
References: [2106]

[EC 1.1.1.377 created 2014]

EC 1.1.1.378

Accepted name: L-rhamnose 1-dehydrogenase [NAD(P)⁺]
Reaction: L-rhamnose + NAD(P)⁺ = L-rhamnono-1,4-lactone + NAD(P)H + H⁺
Systematic name: L-rhamnose:NAD(P)⁺ 1-oxidoreductase
Comments: The enzyme, which occurs in the bacteria *Azotobacter vinelandii* and *Sphingomonas* sp. SKA58, is part of the non-phosphorylative degradation pathway for L-rhamnose. The enzyme differs in cofactor specificity from EC 1.1.1.173, L-rhamnose 1-dehydrogenase, which is specific for NAD⁺ and EC 1.1.1.377, L-rhamnose 1-dehydrogenase (NADP⁺).
References: [4559, 4558]

[EC 1.1.1.378 created 2014]

EC 1.1.1.379

Accepted name: (*R*)-mandelate dehydrogenase
Reaction: (*R*)-mandelate + NAD⁺ = phenylglyoxylate + NADH + H⁺

Other name(s): ManDH₂; D-ManDH₂; D-mandelate dehydrogenase (ambiguous)
Systematic name: (R)-mandelate:NAD⁺ 2-oxidoreductase
Comments: The enzyme, found in bacteria and fungi, can also accept a number of substituted mandelate derivatives, such as 3-hydroxymandelate, 4-hydroxymandelate, 2-methoxymandelate, 4-hydroxy-3-methoxymandelate and 3-hydroxy-4-methoxymandelate. The enzyme has no activity with (S)-mandelate (*cf.* EC 1.1.99.31, (S)-mandelate dehydrogenase) [191, 192]. The enzyme transfers the *pro-R*-hydrogen from NADH [192].
References: [191, 192]

[EC 1.1.1.379 created 2014]

EC 1.1.1.380

Accepted name: L-gulonate 5-dehydrogenase
Reaction: L-gulonate + NAD⁺ = D-fructuronate + NADH + H⁺
Systematic name: L-gulonate:NAD⁺ 5-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Halomonas elongata*, participates in a pathway for L-gulonate degradation.
References: [729, 4614]

[EC 1.1.1.380 created 2014]

EC 1.1.1.381

Accepted name: 3-hydroxy acid dehydrogenase
Reaction: L-*allo*-threonine + NADP⁺ = aminoacetone + CO₂ + NADPH + H⁺ (overall reaction)
(1a) L-*allo*-threonine + NADP⁺ = L-2-amino-3-oxobutanoate + NADPH + H⁺
(1b) L-2-amino-3-oxobutanoate = aminoacetone + CO₂ (spontaneous)
Other name(s): *ydfG* (gene name); YMR226c (gene name)
Systematic name: L-*allo*-threonine:NADP⁺ 3-oxidoreductase
Comments: The enzyme, purified from the bacterium *Escherichia coli* and the yeast *Saccharomyces cerevisiae*, shows activity with a range of 3- and 4-carbon 3-hydroxy acids. The highest activity is seen with L-*allo*-threonine and D-threonine. The enzyme from *Escherichia coli* also shows high activity with L-serine, D-serine, (S)-3-hydroxy-2-methylpropanoate and (R)-3-hydroxy-2-methylpropanoate. The enzyme has no activity with NAD⁺ or L-threonine (*cf.* EC 1.1.1.103, L-threonine 3-dehydrogenase).
References: [1210]

[EC 1.1.1.381 created 2014, modified 2015]

EC 1.1.1.382

Accepted name: ketol-acid reductoisomerase (NAD⁺)
Reaction: (2R)-2,3-dihydroxy-3-methylbutanoate + NAD⁺ = (2S)-2-hydroxy-2-methyl-3-oxobutanoate + NADH + H⁺
Systematic name: (2R)-2,3-dihydroxy-3-methylbutanoate:NAD⁺ oxidoreductase (isomerizing)
Comments: The enzyme, characterized from the bacteria *Thermacetogenium phaeum* and *Desulfococcus oleovorans* and from the archaeon *Archaeoglobus fulgidus*, is specific for NADH [*cf.* EC 1.1.1.86, ketol-acid reductoisomerase (NADP⁺) and EC 1.1.1.383, ketol-acid reductoisomerase [NAD(P)⁺]].
References: [443]

[EC 1.1.1.382 created 2015]

EC 1.1.1.383

Accepted name: ketol-acid reductoisomerase [NAD(P)⁺]
Reaction: (2R)-2,3-dihydroxy-3-methylbutanoate + NAD(P)⁺ = (2S)-2-hydroxy-2-methyl-3-oxobutanoate + NAD(P)H + H⁺

Systematic name: (2R)-2,3-dihydroxy-3-methylbutanoate:NAD(P)⁺ oxidoreductase (isomerizing)
Comments: The enzyme, characterized from the bacteria *Hydrogenobaculum* sp. and *Syntrophomonas wolfei* subsp. *wolfei* and from the archaea *Metallosphaera sedula* and *Ignisphaera aggregans*, can use both NADH and NADPH with similar efficiency [cf. EC 1.1.1.86, ketol-acid reductoisomerase (NADP⁺) and EC 1.1.1.382, ketol-acid reductoisomerase (NAD⁺)].
References: [443]

[EC 1.1.1.383 created 2015]

EC 1.1.1.384

Accepted name: dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose 3-reductase
Reaction: dTDP-4-dehydro-2,6-dideoxy- α -D-glucose + NADP⁺ = dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose + NADPH + H⁺
Other name(s): KijD10; dTDP-4-keto-2,6-dideoxy-D-glucose 3-oxidoreductase; dTDP-4-dehydro-2,6-dideoxy- α -D-glucose 3-oxidoreductase
Systematic name: dTDP-4-dehydro-2,6-dideoxy- α -D-glucose:NADP⁺ 3-oxidoreductase
Comments: The enzyme is involved in the biosynthesis of several deoxysugars, including L-digitoxose, L- and D-olivose, L-oliiose, D-mycarose and forosamine.
References: [38, 4520, 1706, 2270]

[EC 1.1.1.384 created 2015]

EC 1.1.1.385

Accepted name: dihydroantcapsin dehydrogenase
Reaction: L-dihydroantcapsin + NAD⁺ = L-antcapsin + NADH + H⁺
Other name(s): BacC; *ywfD* (gene name)
Systematic name: L-dihydroantcapsin:NAD⁺ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Bacillus subtilis*, is involved in the biosynthesis of the nonribosomally synthesized dipeptide antibiotic bacilysin, composed of L-alanine and L-antcapsin.
References: [3241]

[EC 1.1.1.385 created 2015]

EC 1.1.1.386

Accepted name: ipsdienol dehydrogenase
Reaction: (R)-ipsdienol + NAD(P)⁺ = ipsdienone + NAD(P)H + H⁺
Other name(s): IDOLDH
Systematic name: (R)-ipsdienol:NAD(P)⁺ oxidoreductase
Comments: The enzyme is involved in pheromone production by the pine engraver beetle, *Ips pini*.
References: [1121]

[EC 1.1.1.386 created 2015]

EC 1.1.1.387

Accepted name: L-serine 3-dehydrogenase (NAD⁺)
Reaction: L-serine + NAD⁺ = 2-aminoacetaldehyde + CO₂ + NADH + H⁺ (overall reaction)
(1a) L-serine + NAD⁺ = 2-aminomalonate semialdehyde + NADH + H⁺
(1b) 2-aminomalonate semialdehyde = 2-aminoacetaldehyde + CO₂ (spontaneous)
Other name(s): NAD⁺-dependent L-serine dehydrogenase
Systematic name: L-serine:NAD⁺ 3-oxidoreductase
Comments: The enzyme, purified from the bacterium *Pseudomonas aeruginosa*, also shows activity with L-threonine (cf. EC 1.1.1.103, L-threonine 3-dehydrogenase). The enzyme has only very low activity with NADP⁺ [cf. EC 1.1.1.276, serine 3-dehydrogenase (NADP⁺)].

References: [4232]

[EC 1.1.1.387 created 2015]

EC 1.1.1.388

Accepted name: glucose-6-phosphate dehydrogenase (NAD⁺)
Reaction: D-glucose 6-phosphate + NAD⁺ = 6-phospho-D-glucono-1,5-lactone + NADH + H⁺
Other name(s): Glc6PDH; *azf* (gene name); archaeal zwischenferment
Systematic name: D-glucose-6-phosphate:NAD⁺ 1-oxidoreductase
Comments: The enzyme catalyses a step of the pentose phosphate pathway. The enzyme from the archaeon *Haloferax volcanii* is specific for NAD⁺. *cf.* EC 1.1.1.363, glucose-6-phosphate dehydrogenase [NAD(P)⁺] and EC 1.1.1.49, glucose-6-phosphate dehydrogenase (NADP⁺).
References: [3316]

[EC 1.1.1.388 created 2015]

EC 1.1.1.389

Accepted name: 2-dehydro-3-deoxy-L-galactonate 5-dehydrogenase
Reaction: 2-dehydro-3-deoxy-L-galactonate + NAD⁺ = 3-deoxy-D-*glycero*-2,5-hexodiulosonate + NADH + H⁺
Systematic name: 2-dehydro-3-deoxy-L-galactonate:NAD⁺ 5-oxidoreductase
Comments: The enzyme, characterized from agarose-degrading bacteria, is involved in a degradation pathway for 3,6-anhydro- α -L-galactopyranose, a major component of the polysaccharides of red macroalgae.
References: [2397]

[EC 1.1.1.389 created 2015]

EC 1.1.1.390

Accepted name: sulfoquinovose 1-dehydrogenase
Reaction: sulfoquinovose + NAD⁺ = 6-deoxy-6-sulfo-D-glucono-1,5-lactone + NADH + H⁺
Systematic name: 6-deoxy-6-sulfo-D-glucopyranose:NAD⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Pseudomonas putida* SQ1, participates in a sulfoquinovose degradation pathway. Activity with NADP⁺ is only 4% of that with NAD⁺.
References: [1098]

[EC 1.1.1.390 created 2015]

EC 1.1.1.391

Accepted name: 3 β -hydroxycholanate 3-dehydrogenase (NAD⁺)
Reaction: isolithocholate + NAD⁺ = 3-oxo-5 β -cholan-24-oate + NADH + H⁺
Other name(s): 3 β -hydroxysteroid dehydrogenase
Systematic name: isolithocholate:NAD⁺ 3-oxidoreductase
Comments: This bacterial enzyme is involved, along with EC 1.1.1.52, 3 α -hydroxycholanate dehydrogenase (NAD⁺), or EC 1.1.1.392, 3 α -hydroxycholanate dehydrogenase (NADP⁺), in the modification of secondary bile acids to form 3 β -bile acids (also known as iso-bile acids). The enzyme catalyses the reaction in the reduction direction *in vivo*. Also acts on related 3-oxo bile acids. *cf.* EC 1.1.1.393, 3 β -hydroxycholanate 3-dehydrogenase (NADP⁺).
References: [1013, 1014, 892]

[EC 1.1.1.391 created 2016]

EC 1.1.1.392

Accepted name: 3 α -hydroxycholanate dehydrogenase (NADP⁺)

Reaction: lithocholate + NADP⁺ = 3-oxo-5β-cholan-24-oate + NADPH + H⁺
Other name(s): α-hydroxy-cholanate dehydrogenase (ambiguous)
Systematic name: lithocholate:NADP⁺ 3-oxidoreductase
Comments: This bacterial enzyme is involved in the modification of secondary bile acids to form 3β-bile acids (also known as iso-bile acids) via a 3-oxo intermediate. The enzyme catalyses a reversible reaction *in vitro*. Also acts on related bile acids. *cf.* EC 1.1.1.52, 3α-hydroxychoanate dehydrogenase (NAD⁺).
References: [892]

[EC 1.1.1.392 created 2016]

EC 1.1.1.393

Accepted name: 3β-hydroxychoanate 3-dehydrogenase (NADP⁺)
Reaction: isolithocholate + NADP⁺ = 3-oxo-5β-cholan-24-oate + NADPH + H⁺
Other name(s): 3β-hydroxysteroid dehydrogenase (ambiguous)
Systematic name: isolithocholate:NADP⁺ 3-oxidoreductase
Comments: This bacterial enzyme is involved, along with EC 1.1.1.52, 3α-hydroxychoanate dehydrogenase (NAD⁺), or EC 1.1.1.392, 3α-hydroxychoanate dehydrogenase (NADP⁺), in the modification of secondary bile acids to form 3β-bile acids (also known as iso-bile acids). The enzyme catalyses the reaction in the reduction direction *in vivo*. Also acts on related 3-oxo bile acids. *cf.* EC 1.1.1.391, 3β-hydroxychoanate 3-dehydrogenase (NAD⁺).
References: [46, 892]

[EC 1.1.1.393 created 2016]

EC 1.1.1.394

Accepted name: aurachin B dehydrogenase
Reaction: aurachin B + NAD⁺ + H₂O = 4-[(2*E*,6*E*)-farnesyl]-4-hydroxy-2-methyl-3-oxo-3,4-dihydroquinoline 1-oxide + NADH + H⁺ (overall reaction)
(1a) 4-[(2*E*,6*E*)-farnesyl]-3,4-dihydroxy-2-methyl-3,4-dihydroquinoline 1-oxide + NAD⁺ = 4-[(2*E*,6*E*)-farnesyl]-4-hydroxy-2-methyl-3-oxo-3,4-dihydroquinoline 1-oxide + NADH + H⁺
(1b) aurachin B + H₂O = 4-[(2*E*,6*E*)-farnesyl]-3,4-dihydroxy-2-methyl-3,4-dihydroquinoline 1-oxide (spontaneous)
Other name(s): AuaH
Systematic name: aurachin B:NAD⁺ 3-oxidoreductase
Comments: The enzyme from the bacterium *Stigmatella aurantiaca* catalyses the final step in the conversion of aurachin C to aurachin B. *In vivo* the enzyme catalyses the reduction of 4-[(2*E*,6*E*)-farnesyl]-4-hydroxy-2-methyl-3-oxo-3,4-dihydroquinoline-1-oxide to form 4-[(2*E*,6*E*)-farnesyl]-2-methyl-1-oxo-3,4-dihydroquinoline-3,4-diol (note that the reactions written above proceed from right to left), which then undergoes a spontaneous dehydration to form aurachin B.
References: [2028]

[EC 1.1.1.394 created 2016]

EC 1.1.1.395

Accepted name: 3α-hydroxy bile acid-CoA-ester 3-dehydrogenase
Reaction: a 3α-hydroxy bile acid CoA ester + NAD⁺ = a 3-oxo bile acid CoA ester + NADH + H⁺
Other name(s): *baiA1* (gene name); *baiA2* (gene name); *baiA3* (gene name)
Systematic name: 3α-hydroxy-bile-acid-CoA-ester:NAD⁺ 3-oxidoreductase
Comments: This bacterial enzyme is involved in the 7-dehydroxylation process associated with bile acid degradation. The enzyme has very little activity with unconjugated bile acid substrates. It has similar activity with choloyl-CoA, chenodeoxycholoyl-CoA, deoxycholoyl-CoA, and lithocholoyl-CoA.
References: [2632, 329]

[EC 1.1.1.395 created 2016]

EC 1.1.1.396

- Accepted name:** bacteriochlorophyllide *a* dehydrogenase
- Reaction:** (1) 3-deacetyl-3-(1-hydroxyethyl)bacteriochlorophyllide *a* + NAD⁺ = bacteriochlorophyllide *a* + NADH + H⁺
(2) 3-devinyl-3-(1-hydroxyethyl)chlorophyllide *a* + NAD⁺ = 3-acetyl-3-devinylchlorophyllide *a* + NADH + H⁺
- Other name(s):** *bchC* (gene name)
- Systematic name:** 3-deacetyl-3-(1-hydroxyethyl)bacteriochlorophyllide-*a*:NAD⁺ oxidoreductase (bacteriochlorophyllide *a*-forming)
- Comments:** The enzyme, together with EC 1.3.7.15, chlorophyllide-*a* reductase, and EC 4.2.1.165, chlorophyllide-*a* 3¹-hydratase, is involved in the conversion of chlorophyllide *a* to bacteriochlorophyllide *a*. The enzymes can act in multiple orders, resulting in the formation of different intermediates, but the final product of the cumulative action of the three enzymes is always bacteriochlorophyllide *a*. The enzyme oxidizes a hydroxyl group on ring A, converting it to an oxo group.
- References:** [4581, 2749, 2345]

[EC 1.1.1.396 created 2016]

EC 1.1.1.397

- Accepted name:** β-methylindole-3-pyruvate reductase
- Reaction:** (2*S*,3*R*)-2-hydroxy-3-(indol-3-yl)butanoate + NAD⁺ = (*R*)-3-(indol-3-yl)-2-oxobutanoate + NADH + H⁺
- Other name(s):** ind2 (gene name)
- Systematic name:** (2*S*,3*R*)-2-hydroxy-3-(indol-3-yl)butanoate:NAD⁺ oxidoreductase
- Comments:** The enzyme, characterized from the bacterium *Streptomyces griseus*, participates in the biosynthesis of indolmycin, an antibacterial drug that inhibits the bacterial tryptophan—tRNA ligase (EC 6.1.1.2).
- References:** [973]

[EC 1.1.1.397 created 2016]

EC 1.1.1.398

- Accepted name:** 2-glutathionyl-2-methylbut-3-en-1-ol dehydrogenase
- Reaction:** 2-(glutathion-*S*-yl)-2-methylbut-3-en-1-ol + 2 NAD⁺ + H₂O = 2-(glutathion-*S*-yl)-2-methylbut-3-enoate + 2 NADH + 2 H⁺ (overall reaction)
(1a) 2-(glutathion-*S*-yl)-2-methylbut-3-en-1-ol + NAD⁺ = 2-(glutathion-*S*-yl)-2-methylbut-3-enal + NADH + H⁺
(1b) 2-(glutathion-*S*-yl)-2-methylbut-3-enal + NAD⁺ + H₂O = 2-(glutathion-*S*-yl)-2-methylbut-3-enoate + NADH + H⁺
- Other name(s):** *isoH* (gene name); 4-hydroxy-3-glutathionyl-3-methylbut-1-ene dehydrogenase
- Systematic name:** 2-(glutathion-*S*-yl)-2-methylbut-3-en-1-ol:NAD⁺ oxidoreductase
- Comments:** The enzyme, characterized from the bacterium *Rhodococcus* sp. AD45, is involved in isoprene degradation.
- References:** [4409]

[EC 1.1.1.398 created 2016]

EC 1.1.1.399

- Accepted name:** 2-oxoglutarate reductase
- Reaction:** (*R*)-2-hydroxyglutarate + NAD⁺ = 2-oxoglutarate + NADH + H⁺
- Other name(s):** *serA* (gene name)
- Systematic name:** (*R*)-2-hydroxyglutarate:NAD⁺ 2-oxidoreductase

Comments: The enzyme catalyses a reversible reaction. The enzyme from the bacterium *Peptoniphilus asaccharolyticus* is specific for (*R*)-2-hydroxyglutarate [2420, 1938]. The SerA enzyme from the bacterium *Escherichia coli* can also accept (*S*)-2-hydroxyglutarate with a much higher K_m , and also catalyses the activity of EC 1.1.1.95, phosphoglycerate dehydrogenase [4900].

References: [2420, 1938, 4900]

[EC 1.1.1.399 created 2016]

EC 1.1.1.400

Accepted name: 2-methyl-1,2-propanediol dehydrogenase

Reaction: 2-methylpropane-1,2-diol + NAD^+ = 2-hydroxy-2-methylpropanal + $\text{NADH} + \text{H}^+$

Other name(s): *mpdB* (gene name)

Systematic name: 2-methylpropane-1,2-diol: NAD^+ 1-oxidoreductase

Comments: This bacterial enzyme is involved in the degradation pathways of the alkene 2-methylpropene and the fuel additive *tert*-butyl methyl ether (MTBE), a widely occurring groundwater contaminant.

References: [1110, 2248]

[EC 1.1.1.400 created 2016]

EC 1.1.1.401

Accepted name: 2-dehydro-3-deoxy-L-rhamnonate dehydrogenase (NAD^+)

Reaction: 2-dehydro-3-deoxy-L-rhamnonate + NAD^+ = 2,4-didehydro-3-deoxy-L-rhamnonate + $\text{NADH} + \text{H}^+$

Other name(s): 2-keto-3-deoxy-L-rhamnonate dehydrogenase

Systematic name: 2-dehydro-3-deoxy-L-rhamnonate: NAD^+ 4-oxidoreductase

Comments: The enzyme, characterized from the bacteria *Sphingomonas* sp. SKA58 and *Sulfobacillus thermosulfidooxidans*, is involved in the non-phosphorylative degradation pathway for L-rhamnose. It does not show any detectable activity with NADP^+ or with other aldoses.

References: [4558, 176]

[EC 1.1.1.401 created 2016]

EC 1.1.1.402

Accepted name: D-erythritol 1-phosphate dehydrogenase

Reaction: D-erythritol 1-phosphate + NADP^+ = D-erythrulose 1-phosphate + $\text{NADPH} + \text{H}^+$

Other name(s): *eryB* (gene name)

Systematic name: D-erythritol-1-phosphate 2-oxidoreductase

Comments: The enzyme, characterized from the pathogenic bacterium *Brucella abortus*, which causes brucellosis in livestock, participates in erythritol catabolism.

References: [3984, 3657, 217]

[EC 1.1.1.402 created 2016]

EC 1.1.1.403

Accepted name: D-threitol dehydrogenase (NAD^+)

Reaction: D-threitol + NAD^+ = D-erythrulose + $\text{NADH} + \text{H}^+$

Other name(s): *dthD* (gene name)

Systematic name: D-threitol: NAD^+ oxidoreductase

Comments: The enzyme, characterized from the bacterium *Mycobacterium smegmatis*, participates in the degradation of D-threitol.

References: [1753]

[EC 1.1.1.403 created 2016]

EC 1.1.1.404

Accepted name: tetrachlorobenzoquinone reductase
Reaction: 2,3,5,6-tetrachlorohydroquinone + NAD⁺ = 2,3,5,6-tetrachloro-1,4-benzoquinone + NADH + H⁺
Other name(s): *pcpD* (gene name); TCBQ reductase
Systematic name: 2,3,5,6-tetrachlorohydroquinone:NAD⁺ oxidoreductase
Comments: Contains FMN. The enzyme, characterized from the bacterium *Sphingobium chlorophenolicum*, participates in the degradation of pentachlorophenol.
References: [633, 4718]

[EC 1.1.1.404 created 2017]

EC 1.1.1.405

Accepted name: ribitol-5-phosphate 2-dehydrogenase (NADP⁺)
Reaction: D-ribitol 5-phosphate + NADP⁺ = D-ribulose 5-phosphate + NADPH + H⁺
Other name(s): *acs1* (gene name); *bcs1* (gene name); *tarJ* (gene name); ribulose-5-phosphate reductase; ribulose-5-*P* reductase; D-ribulose 5-phosphate reductase
Systematic name: D-ribitol-5-phosphate:NADP⁺ 2-oxidoreductase
Comments: Requires Zn²⁺. The enzyme, characterized in bacteria, is specific for NADP. It is part of the synthesis pathway of CDP-ribitol. In *Haemophilus influenzae* it is part of a multifunctional enzyme also catalysing EC 2.7.7.40, D-ribitol-5-phosphate cytidylyltransferase. *cf.* EC 1.1.1.137, ribitol-5-phosphate 2-dehydrogenase.
References: [4936, 3288, 3289, 247]

[EC 1.1.1.405 created 2017]

EC 1.1.1.406

Accepted name: galactitol 2-dehydrogenase (L-tagatose-forming)
Reaction: galactitol + NAD⁺ = L-tagatose + NADH + H⁺
Other name(s): GatDH
Systematic name: galactitol:NAD⁺ 2-oxidoreductase (L-tagatose-forming)
Comments: The enzyme, characterized in the bacterium *Rhodobacter sphaeroides*, has a wide substrate specificity. In addition to galactitol, it primarily oxidizes D-threitol and xylitol, and in addition to L-tagatose, it primarily reduces L-erythrulose, D-ribulose and L-glyceraldehyde. It is specific for NAD⁺. The enzyme also shows activity with D-tagatose (*cf.* EC 1.1.1.16, galactitol 2-dehydrogenase).
References: [3736, 556]

[EC 1.1.1.406 created 2017]

EC 1.1.1.407

Accepted name: D-altritol 5-dehydrogenase
Reaction: D-altritol + NAD⁺ = D-tagatose + NADH + H⁺
Systematic name: D-altritol:NAD⁺ 5-oxidoreductase
Comments: The enzyme, characterized in *Agrobacterium fabrum* C58, also has low activity with D-mannitol and D-arabinitol. It is part of a D-altritol degradation pathway.
References: [4615]

[EC 1.1.1.407 created 2017]

EC 1.1.1.408

Accepted name: 4-phospho-D-threonate 3-dehydrogenase
Reaction: 4-phospho-D-threonate + NAD⁺ = glycerone phosphate + CO₂ + NADH + H⁺ (overall reaction)
(1a) 4-phospho-D-threonate + NAD⁺ = 3-dehydro-4-phospho-D-erythronate + NADH + H⁺
(1b) 3-dehydro-4-phospho-D-erythronate = glycerone phosphate + CO₂ (spontaneous)

Other name(s): *pdxA2* (gene name) (ambiguous)
Systematic name: 4-phospho-D-threonate:NAD⁺ 3-oxidoreductase
Comments: The enzyme, characterized from bacteria, is involved in a pathway for D-threonate catabolism.
References: [4887]

[EC 1.1.1.408 created 2017]

EC 1.1.1.409

Accepted name: 4-phospho-D-erythronate 3-dehydrogenase
Reaction: 4-phospho-D-erythronate + NAD⁺ = glycerone phosphate + CO₂ + NADH + H⁺ (overall reaction)
(1a) 4-phospho-D-erythronate + NAD⁺ = 3-dehydro-4-phospho-L-threonate + NADH + H⁺
(1b) 3-dehydro-4-phospho-L-threonate = glycerone phosphate + CO₂ (spontaneous)
Other name(s): *pdxA2* (gene name) (ambiguous)
Systematic name: 4-phospho-D-erythronate:NAD⁺ 3-oxidoreductase
Comments: The enzyme, characterized from bacteria, is involved in a pathway for D-erythronate catabolism.
References: [4887]

[EC 1.1.1.409 created 2017]

EC 1.1.1.410

Accepted name: D-erythronate 2-dehydrogenase
Reaction: D-erythronate + NAD⁺ = 2-dehydro-D-erythronate + NADH + H⁺
Other name(s): *denD* (gene name)
Systematic name: D-erythronate:NAD⁺ 2-oxidoreductase
Comments: The enzyme, characterized from bacteria, is involved in D-erythronate catabolism.
References: [4887]

[EC 1.1.1.410 created 2017]

EC 1.1.1.411

Accepted name: L-threonate 2-dehydrogenase
Reaction: L-threonate + NAD⁺ = 2-dehydro-L-erythronate + NADH + H⁺
Other name(s): *ltnD* (gene name)
Systematic name: L-threonate:NAD⁺ 2-oxidoreductase
Comments: The enzyme, characterized from bacteria, is involved in L-threonate catabolism.
References: [4887]

[EC 1.1.1.411 created 2017]

EC 1.1.1.412

Accepted name: 2-alkyl-3-oxoalkanoate reductase
Reaction: a (2*R*,3*S*)-2-alkyl-3-hydroxyalkanoate + NADP⁺ = an (*R*)-2-alkyl-3-oxoalkanoate + NADPH + H⁺
Other name(s): *oleD* (gene name)
Systematic name: (2*R*,3*S*)-2-alkyl-3-hydroxyalkanoate:NADP⁺ oxidoreductase
Comments: The enzyme, found in certain bacterial species, is part of a pathway for the production of olefins.
References: [386]

[EC 1.1.1.412 created 2017]

EC 1.1.1.413

Accepted name: A-factor type γ -butyrolactone 1'-reductase (1*S*-forming)

Reaction: a (3*R*,4*R*)-3-[(1*S*)-1-hydroxyalkyl]-4-(hydroxymethyl)oxolan-2-one + NADP⁺ = a (3*R*,4*R*)-3-alkanoyl-4-(hydroxymethyl)oxolan-2-one + NADPH + H⁺

Other name(s): *barS1* (gene name)

Systematic name: (3*R*,4*R*)-3-[(1*S*)-1-hydroxyalkyl]-4-(hydroxymethyl)oxolan-2-one:NADP⁺ 1'-oxidoreductase

Comments: The enzyme, which is found in bacteria that produce virginiae-butanolide (VB) type γ -butyrolactone autoregulators, reduces its substrate stereospecifically, forming a hydroxyl group in the (S) configuration.

References: [3861]

[EC 1.1.1.413 created 2017]

EC 1.1.1.414

Accepted name: L-galactonate 5-dehydrogenase

Reaction: L-galactonate + NAD⁺ = D-tagaturonate + NADH + H⁺

Other name(s): *lgoD* (gene name); *lgaC* (gene name)

Systematic name: L-galactonate:NAD⁺ 5-oxidoreductase

Comments: The enzyme, reported from the human gut bacteria *Escherichia coli* and *Bacteroides vulgatus*, participates in an L-galactonate degradation pathway.

References: [728, 2280, 1678]

[EC 1.1.1.414 created 2018]

EC 1.1.1.415

Accepted name: noscapine synthase

Reaction: narcotine hemiacetal + NAD⁺ = noscapine + NADH + H⁺

Other name(s): NOS (gene name)

Systematic name: narcotine hemiacetal:NAD⁺ 1-oxidoreductase

Comments: The enzyme, characterized from the plant *Papaver somniferum* (opium poppy), catalyses the last step in the biosynthesis of the isoquinoline alkaloid noscapine.

References: [637, 2464]

[EC 1.1.1.415 created 2018]

EC 1.1.1.416

Accepted name: isopyridoxal dehydrogenase (5-pyridoxolactone-forming)

Reaction: isopyridoxal + NAD⁺ = 5-pyridoxolactone + NADH + H⁺

Systematic name: isopyridoxal:NAD⁺ oxidoreductase (5-pyridoxolactone-forming)

Comments: The enzyme, characterized from the bacterium *Arthrobacter* sp. Cr-7, participates in the degradation of pyridoxine. The enzyme also catalyses the activity of EC 1.2.1.102, isopyridoxal dehydrogenase (5-pyridoxate-forming).

References: [2401]

[EC 1.1.1.416 created 2018]

EC 1.1.1.417

Accepted name: 3 β -hydroxysteroid-4 β -carboxylate 3-dehydrogenase (decarboxylating)

Reaction: a 3 β -hydroxy-4 α -methylsteroid-4 β -carboxylate + NAD(P)⁺ = a 4 α -methyl-3-oxosteroid + NAD(P)H + CO₂ + H⁺

Other name(s): *sdmB* (gene name)

Systematic name: 3 β -hydroxysteroid-4 β -carboxylate:NAD(P)⁺ 3-oxidoreductase (decarboxylating)

Comments: This bacterial enzyme participates in the biosynthesis of bacterial sterols. Together with EC 1.14.13.246, 4 β -methylsterol monooxygenase (SdmA) it forms an enzyme system that removes one methyl group from the C-4 position of 4,4-dimethylated steroid molecules. SdmA catalyses three successive oxidations of the C-4 β methyl group, turning it into a carboxylate group; SdmB is a bifunctional enzyme that catalyses two different activities. As EC 1.1.1.417 it catalyses an oxidative decarboxylation that results in reduction of the 3 β -hydroxy group at the C-3 carbon to an oxo group. As EC 1.1.1.270, 3 β -hydroxysteroid 3-dehydrogenase, it reduces the 3-oxo group back to a 3 β -hydroxyl. Since the remaining methyl group at C-4 is in an α orientation, it cannot serve as a substrate for a second round of demethylation by this system.

References: [2376]

[EC 1.1.1.417 created 2019]

EC 1.1.1.418

Accepted name: plant 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating)
Reaction: a 3 β -hydroxysteroid-4 α -carboxylate + NAD⁺ = a 3-oxosteroid + CO₂ + NADH
Other name(s): 3 β -HSD/D1 (gene name); 3 β -HSD/D2 (gene name); 3 β -hydroxysteroid dehydrogenases/C-4 decarboxylase (ambiguous)
Systematic name: 3 β -hydroxysteroid-4 α -carboxylate:NAD⁺ 3-oxidoreductase (decarboxylating)
Comments: The enzyme, found in plants, catalyses multiple reactions during plant sterol biosynthesis. Unlike the fungal/animal enzyme EC 1.1.1.170, 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating), the plant enzyme is specific for NAD⁺.
References: [3567, 3427, 3426]

[EC 1.1.1.418 created 2019]

EC 1.1.1.419

Accepted name: nepetalactol dehydrogenase
Reaction: (1) (+)-*cis,cis*-nepetalactol + NAD⁺ = (+)-*cis,cis*-nepetalactone + NADH + H⁺
(2) (+)-*cis,trans*-nepetalactol + NAD⁺ = (+)-*cis,trans*-nepetalactone + NADH + H⁺
Other name(s): NEPS1 (gene name)
Systematic name: nepetalactol:NAD⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the plant *Nepeta mussinii*, binds an NAD⁺ cofactor. It also catalyses the activity of EC 5.5.1.34, (+)-*cis,trans*-nepetalactol synthase.
References: [2476, 2477]

[EC 1.1.1.419 created 2019]

EC 1.1.1.420

Accepted name: D-apiose dehydrogenase
Reaction: D-apiofuranose + NAD⁺ = D-apionolactone + NADH + H⁺
Other name(s): *apsD* (gene name)
Systematic name: D-apiofuranose:NAD⁺ 1-oxidoreductase
Comments: The enzyme, characterized from several bacterial species, is involved in a catabolic pathway for D-apiose.
References: [565]

[EC 1.1.1.420 created 2019]

EC 1.1.1.421

Accepted name: D-apionate oxidoisomerase
Reaction: D-apionate + NAD⁺ = 3-oxoisoapionate + NADH + H⁺
Other name(s): *apnO* (gene name)

Systematic name: D-apionate:NAD⁺ oxidoreductase (isomerizing)
Comments: The enzyme, characterized from several bacterial species, participates in the degradation of D-apionate. The reaction involves migration of a hydroxymethyl group from position 3 to position 2 and oxidation of the 3-hydroxyl group. Stereospecificity of the product, 3-oxoisopionate, has not been determined.
References: [565]

[EC 1.1.1.421 created 2019]

EC 1.1.1.422

Accepted name: pseudoephedrine dehydrogenase
Reaction: (+)-(1*S*,2*S*)-pseudoephedrine + NAD⁺ = (*S*)-2-(methylamino)-1-phenylpropan-1-one + NADH + H⁺
Other name(s): PseDH
Systematic name: (+)-(1*S*,2*S*)-pseudoephedrine:NAD⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Arthrobacter* sp. TS-15, acts on a broad range of different aryl-alkyl ketones, such as haloketones, ketoamines, diketones, and ketoesters. It accepts various types of aryl groups including phenyl-, pyridyl-, thienyl-, and furyl-rings, but the presence of an aromatic ring is essential for the activity. In addition, the presence of a functional group on the alkyl chain, such as an amine, a halogen, or a ketone, is also crucial. The enzyme exhibits a strict anti-Prelog enantioselectivity. When acting on diketones, it catalyses the reduction of only the keto group closest to the ring, with no further reduction to the diol. *cf.* EC 1.1.1.423, ephedrine dehydrogenase.
References: [3819, 3817, 3818]

[EC 1.1.1.422 created 2020]

EC 1.1.1.423

Accepted name: (1*R*,2*S*)-ephedrine 1-dehydrogenase
Reaction: (–)-(1*R*,2*S*)-ephedrine + NAD⁺ = (*S*)-2-(methylamino)-1-phenylpropan-1-one + NADH + H⁺
Other name(s): EDH; ephedrine dehydrogenase
Systematic name: (–)-(1*R*,2*S*)-ephedrine:NAD⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Arthrobacter* sp. TS-15, acts on a broad range of different aryl-alkyl ketones, such as haloketones, ketoamines, diketones, and ketoesters. It exhibits a strict enantioselectivity and accepts various types of aryl groups including phenyl-, pyridyl-, thienyl-, and furyl-rings, but the presence of an aromatic ring is essential for the activity. In addition, the presence of a functional group on the alkyl chain, such as an amine, a halogen, or a ketone, is also crucial. When acting on diketones, it catalyses the reduction of only the keto group closest to the ring, with no further reduction to the diol. *cf.* EC 1.1.1.422, pseudoephedrine dehydrogenase and EC 1.5.1.18, ephedrine dehydrogenase.
References: [3819, 3817]

[EC 1.1.1.423 created 2020, modified 2020]

EC 1.1.1.424

Accepted name: D-xylose 1-dehydrogenase (NADP⁺, D-xylo-1,4-lactone-forming)
Reaction: D-xylose + NADP⁺ = D-xylo-1,4-lactone + NADPH + H⁺
Other name(s): *xacA* (gene name); *xdh* (gene name)
Systematic name: D-xylose:NADP⁺ 1-oxidoreductase (D-xylo-1,4-lactone-forming)
Comments: The enzyme, which participates in the degradation of D-xylose, has been characterized from several halophilic archaeal species. *cf.* EC 1.1.1.179, D-xylose 1-dehydrogenase (NADP⁺, D-xylo-1,5-lactone-forming).
References: [1926, 1925, 4131]

[EC 1.1.1.424 created 2020]

EC 1.1.1.425

- Accepted name:** levoglucosan dehydrogenase
Reaction: levoglucosan + NAD⁺ = 3-dehydrolevoglucosan + NADH + H⁺
Other name(s): 1,6-anhydro-β-D-glucose dehydrogenase
Systematic name: 1,6-anhydro-β-D-glucopyranose:NAD⁺ 3-oxidoreductase
Comments: Levoglucosan is formed from the pyrolysis of carbohydrates such as starch and cellulose and is an important molecular marker for pollution from biomass burning. This enzyme is present only in bacteria, and has been characterized from *Arthrobacter* sp. I-552 and *Pseudarthrobacter phenanthrenivorans*. cf. EC 2.7.1.232, levoglucosan kinase.
References: [2975, 4102]

[EC 1.1.1.425 created 2021]

EC 1.1.1.426

- Accepted name:** UDP-*N*-acetyl-α-D-quinovosamine dehydrogenase
Reaction: UDP-*N*-acetyl-α-D-quinovosamine + NAD(P)⁺ = UDP-2-acetamido-2,6-dideoxy-α-D-xylohex-4-ulose + NAD(P)H + H⁺
Other name(s): *wbpV* (gene name); *wreQ* (gene name)
Systematic name: UDP-*N*-acetyl-α-D-quinovosamine:NAD(P)⁺ 4-dehydrogenase
Comments: The enzyme participates in the biosynthesis of *N*-acetyl-α-D-quinovosamine, a 6-deoxy sugar that is present in the O antigens of many Gram-negative bacteria, including *Pseudomonas aeruginosa* serotypes O6 and O10, *Rhizobium etli*, and *Brucella abortus*.
References: [277, 1147, 2458]

[EC 1.1.1.426 created 2021]

EC 1.1.1.427

- Accepted name:** D-arabinose 1-dehydrogenase (NADP⁺)
Reaction: D-arabinofuranose + NADP⁺ = D-arabinono-1,4-lactone + NADPH + H⁺
Other name(s): AraDH; adh-4 (gene name)
Systematic name: D-arabinose:NADP⁺ 1-oxidoreductase
Comments: The enzyme from the archaeon *Saccharolobus solfataricus* is tetrameric and contains zinc. L-fucose also is a substrate. In contrast to EC 1.1.1.116 (D-arabinose 1-dehydrogenase (NAD⁺)) and EC 1.1.1.117 (D-arabinose 1-dehydrogenase [NAD(P)⁺]), this enzyme is specific for NADP⁺.
References: [455, 454]

[EC 1.1.1.427 created 2022]

EC 1.1.1.428

- Accepted name:** 4-methylthio 2-oxobutanoate reductase (NADH)
Reaction: (2*R*)-2-hydroxy-4-(methylsulfanyl)butanoate + NAD⁺ = 4-(methylsulfanyl)-2-oxobutanoate + NADH + H⁺
Other name(s): CTBP1 (gene name); C-terminal-binding protein 1; MTOB reductase; 4-methylthio 2-oxobutyrate reductase; 4-methylthio 2-oxobutyric acid reductase
Systematic name: (2*R*)-2-hydroxy-4-(methylsulfanyl)butanoate:NAD⁺ 2-oxidoreductase
Comments: The substrate of this enzyme is formed as an intermediate during L-methionine salvage from *S*-methyl-5'-thioadenosine, which is formed during the biosynthesis of polyamines. The human enzyme also functions as a transcriptional co-regulator that downregulates the expression of many tumor-suppressor genes, thus providing a link between gene repression and the methionine salvage pathway. A similar, but NADP-specific, enzyme is involved in dimethylsulfoniopropanoate biosynthesis in algae and phytoplankton.
References: [2284, 7, 1650, 2236]

[EC 1.1.1.428 created 2022]

EC 1.1.1.429

Accepted name: (2S)-[(R)-hydroxy(phenyl)methyl]succinyl-CoA dehydrogenase
Reaction: (2S)-[(R)-hydroxy(phenyl)methyl]succinyl-CoA + NAD⁺ = (S)-2-benzoylsuccinyl-CoA + NADH + H⁺
Other name(s): *bbsCD* (gene name)
Systematic name: (2S)-[(R)-hydroxy(phenyl)methyl]succinyl-CoA:NAD⁺ oxidoreductase
Comments: The enzyme, purified from the bacterium *Thauera aromatica*, is involved in an anaerobic toluene degradation pathway. It is specific for NAD⁺.
References: [4468]

[EC 1.1.1.429 created 2022]

EC 1.1.1.430

Accepted name: D-xylose reductase (NADH)
Reaction: xylitol + NAD⁺ = D-xylose + NADH + H⁺
Other name(s): XYL1 (gene name) (ambiguous)
Systematic name: xylitol:NAD⁺ oxidoreductase
Comments: Xylose reductases catalyse the reduction of xylose to xylitol, the initial reaction in the fungal D-xylose degradation pathway. Most of the enzymes exhibit a strict requirement for NADPH (*cf.* EC 1.1.1.431, D-xylose reductase (NADPH)). Some D-xylose reductases have dual coenzyme specificity, though they still prefer NADPH to NADH (*cf.* EC 1.1.1.307, D-xylose reductase [NAD(P)H]). The enzyme from *Candida parapsilosis* is a rare example of a xylose reductase that significantly prefers NADH, with K_m and V_{max} values for NADH being 10-fold lower and 10-fold higher, respectively, than for NADPH.
References: [2386]

[EC 1.1.1.430 created 2022]

EC 1.1.1.431

Accepted name: D-xylose reductase (NADPH)
Reaction: xylitol + NADP⁺ = D-xylose + NADPH + H⁺
Other name(s): XYL1 (gene name, ambiguous); xyl1 (gene name, ambiguous); *xyrA* (gene name); *xyrB* (gene name)
Systematic name: xylitol:NADP⁺ oxidoreductase
Comments: Xylose reductases catalyse the reduction of xylose to xylitol, the initial reaction in the fungal D-xylose degradation pathway. Most of the enzymes exhibit a strict requirement for NADPH (e.g. the enzymes from *Saccharomyces cerevisiae*, *Aspergillus niger*, *Trichoderma reesei*, *Candida tropicalis*, *Saitozyma flava*, and *Candida intermedia*). Some D-xylose reductases have dual coenzyme specificity, though they still prefer NADPH to NADH (*cf.* EC 1.1.1.307, D-xylose reductase [NAD(P)H]). Very rarely the enzyme prefers NADH (*cf.* EC 1.1.1.430, D-xylose reductase (NADH)).
References: [376, 4144, 3065, 2734, 3797, 1904, 686, 4245]

[EC 1.1.1.431 created 2022]

EC 1.1.1.432

Accepted name: 6-dehydroglucose reductase
Reaction: D-glucose + NADP⁺ = 6-dehydro-D-glucose + NADPH + H⁺
Other name(s): D-glucose 6-dehydrogenase; *smoB* (gene name); *squF* (gene name)
Systematic name: D-glucose:NADP⁺ 6-oxidoreductase
Comments: The enzyme, characterized from alphaproteobacteria, is involved in a D-sulfoquinovose degradation pathway.
References: [3827, 2516]

[EC 1.1.1.432 created 2022]

EC 1.1.1.433

- Accepted name:** sulfoacetaldehyde reductase (NADH)
Reaction: isethionate + NAD^+ = 2-sulfoacetaldehyde + NADH + H^+
Other name(s): *sarD* (gene name); *tauF* (gene name); *sqwF* (gene name); BkTauF
Systematic name: isethionate: NAD^+ oxidoreductase
Comments: The enzymes from the bacteria *Bilophila wadsworthia* and *Clostridium* sp. MSTE9 catalyse the reaction only in the reduction direction. In the bacterium *Bifidobacterium kashiwanohense* the optimal reaction pH for sulfoacetaldehyde reduction is 7.5, while that for isethionate oxidation is 10.0. *cf.* EC 1.1.1.313, sulfoacetaldehyde reductase (NADPH).
References: [3275, 4702, 4918, 2516]

[EC 1.1.1.433 created 2022]

EC 1.1.1.434

- Accepted name:** 2-dehydro-3-deoxy-L-fuconate 4-dehydrogenase
Reaction: 2-dehydro-3-deoxy-L-fuconate + NAD^+ = 2,4-didehydro-3-deoxy-L-fuconate + NADH + H^+
Systematic name: 2-dehydro-3-deoxy-L-fuconate: NAD^+ 4-oxidoreductase
Comments: The enzyme, originally described from the bacterium *Xanthomonas campestris pv. campestris*, participates in an L-fucose degradation pathway. It can also act on 2-dehydro-3-deoxy-L-galactonate and 2-dehydro-3-deoxy-D-pentonate.
References: [4789, 4556]

[EC 1.1.1.434 created 2022]

EC 1.1.1.435

- Accepted name:** L-fucose dehydrogenase
Reaction: β -L-fucopyranose + NADP^+ = L-fucono-1,5-lactone + NADPH + H^+
Systematic name: β -L-fucopyranose: NADP^+ 1-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Burkholderia multivorans*, participates in an L-fucose degradation pathway. The enzyme catalyses the oxidation of β -L-fucopyranose to L-fucono-1,5-lactone, which is unstable and is rapidly converted to L-fucono-1,4-lactone. The α anomer is not recognized. The enzyme can also act on β -L-galactopyranose and D-arabinose with lower activity. NADP^+ is a better cosubstrate than NAD^+ .
References: [1677]

[EC 1.1.1.435 created 2022]

EC 1.1.1.436

- Accepted name:** lactate dehydrogenase (NAD^+ , ferredoxin)
Reaction: lactate + 2 NAD^+ + 2 reduced ferredoxin [iron-sulfur] cluster = pyruvate + 2 NADH + 2 oxidized ferredoxin [iron-sulfur] cluster
Other name(s): electron bifurcating LDH/Etf complex
Systematic name: lactate: NAD^+ , ferredoxin oxidoreductase
Comments: The enzyme, isolated from the bacterium *Acetobacterium woodii*, uses flavin-based electron confurcation to drive endergonic lactate oxidation with NAD^+ as oxidant at the expense of simultaneous exergonic electron flow from reduced ferredoxin to NAD^+ .
References: [4571]

[EC 1.1.1.436 created 2015 as EC 1.3.1.110, transferred 2022 to EC 1.1.1.436]

EC 1.1.1.437

- Accepted name:** 5-dehydrofumagillol 5-reductase
Reaction: fumagillol + NADP^+ = 5-dehydrofumagillol + NADPH + H^+

Other name(s): af490 (gene name); Fma-KR
Systematic name: fumagillol:NADP⁺ 5-oxidoreductase
Comments: The enzyme, characterized from the mold *Aspergillus fumigatus*, participates in the biosynthesis of the meroterpenoid fumagillin. It is a partial polyketide synthase (PKS) consisting of only a dehydratase (DH) and a ketoreductase (KR) domain.
References: [2489]

[EC 1.1.1.437 created 2022]

EC 1.1.2 With a cytochrome as acceptor

[1.1.2.1 Transferred entry. glycerolphosphate dehydrogenase. As the acceptor is now known, the enzyme has been transferred to EC 1.1.5.3, glycerol-3-phosphate dehydrogenase.]

[EC 1.1.2.1 created 1961, deleted 1965]

EC 1.1.2.2

Accepted name: mannitol dehydrogenase (cytochrome)
Reaction: D-mannitol + a ferricytochrome *c* = D-fructose + a ferrocyclochrome *c* + 2 H⁺
Other name(s): polyol dehydrogenase
Systematic name: D-mannitol:cytochrome-*c* 2-oxidoreductase
Comments: The enzyme from the bacterium *Gluconobacter oxydans* acts on polyols with a D-lyxo configuration, such as D-mannitol and D-sorbitol, with preference towards the former.
References: [129, 667]

[EC 1.1.2.2 created 1961]

EC 1.1.2.3

Accepted name: L-lactate dehydrogenase (cytochrome)
Reaction: (S)-lactate + 2 ferricytochrome *c* = pyruvate + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): lactic acid dehydrogenase; cytochrome *b*₂ (flavin-free derivative of flavocytochrome *b*₂); flavocytochrome *b*₂; L-lactate cytochrome *c* reductase; L-(+)-lactate:cytochrome *c* oxidoreductase; dehydrogenase, lactate (cytochrome); L-lactate cytochrome *c* oxidoreductase; lactate dehydrogenase (cytochrome); lactic cytochrome *c* reductase
Systematic name: (S)-lactate:ferricytochrome-*c* 2-oxidoreductase
Comments: Identical with cytochrome *b*₂; a flavohemoprotein (FMN).
References: [120, 119, 171, 3120]

[EC 1.1.2.3 created 1961]

EC 1.1.2.4

Accepted name: D-lactate dehydrogenase (cytochrome)
Reaction: (R)-lactate + 2 ferricytochrome *c* = pyruvate + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): lactic acid dehydrogenase; D-lactate (cytochrome) dehydrogenase; cytochrome-*dependent* D-(-)-lactate dehydrogenase; D-lactate-cytochrome *c* reductase; D-(-)-lactic cytochrome *c* reductase
Systematic name: (R)-lactate:cytochrome-*c* 2-oxidoreductase
Comments: A flavoprotein (FAD).
References: [1403, 1404, 3119, 3120]

[EC 1.1.2.4 created 1961]

EC 1.1.2.5

- Accepted name:** D-lactate dehydrogenase (cytochrome *c*-553)
Reaction: (*R*)-lactate + 2 ferricytochrome *c*-553 = pyruvate + 2 ferrocyclochrome *c*-553 + 2 H⁺
Systematic name: (*R*)-lactate:cytochrome-*c*-553 2-oxidoreductase
Comments: The enzyme from the sulfate-reducing bacterium *Desulfovibrio vulgaris* can also act on (*R*)-2-hydroxybutanoate.
References: [3133]

[EC 1.1.2.5 created 1989]

EC 1.1.2.6

- Accepted name:** polyvinyl alcohol dehydrogenase (cytochrome)
Reaction: polyvinyl alcohol + ferricytochrome *c* = oxidized polyvinyl alcohol + ferrocyclochrome *c* + H⁺
Other name(s): PVA dehydrogenase; PVADH
Systematic name: polyvinyl alcohol:ferricytochrome-*c* oxidoreductase
Comments: A quinoprotein. The enzyme is involved in bacterial polyvinyl alcohol degradation. Some Gram-negative bacteria degrade polyvinyl alcohol by importing it into the periplasmic space, where it is oxidized by polyvinyl alcohol dehydrogenase, an enzyme that is coupled to the respiratory chain via cytochrome *c*. The enzyme contains a pyrroloquinoline quinone cofactor.
References: [3867, 3869, 2634, 1673, 1747, 2042]

[EC 1.1.2.6 created 1989 as EC 1.1.99.23, transferred 2010 to EC 1.1.2.6]

EC 1.1.2.7

- Accepted name:** methanol dehydrogenase (cytochrome *c*)
Reaction: a primary alcohol + 2 ferricytochrome *c*₁ = an aldehyde + 2 ferrocyclochrome *c*₁ + 2 H⁺
Other name(s): methanol dehydrogenase; MDH (ambiguous)
Systematic name: methanol:cytochrome *c* oxidoreductase
Comments: A periplasmic quinoprotein alcohol dehydrogenase that only occurs in methylotrophic bacteria. It uses the novel specific cytochrome *c*₁ as acceptor. Acts on a wide range of primary alcohols, including ethanol, duodecanol, chloroethanol, cinnamyl alcohol, and also formaldehyde. Activity is stimulated by ammonia or methylamine. It is usually assayed with phenazine methosulfate. Like all other quinoprotein alcohol dehydrogenases it has an 8-bladed 'propeller' structure, a calcium ion bound to the PQQ in the active site and an unusual disulfide ring structure in close proximity to the PQQ. It differs from EC 1.1.2.8, alcohol dehydrogenase (cytochrome *c*), in having a high affinity for methanol and in having a second essential small subunit (no known function).
References: [108, 109, 980, 3647, 757, 347, 4694, 30, 107, 4634]

[EC 1.1.2.7 created 1972 as EC 1.1.99.8, modified 1982, part transferred 2010 to EC 1.1.2.7]

EC 1.1.2.8

- Accepted name:** alcohol dehydrogenase (cytochrome *c*)
Reaction: a primary alcohol + 2 ferricytochrome *c* = an aldehyde + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): type I quinoprotein alcohol dehydrogenase; quinoprotein ethanol dehydrogenase
Systematic name: alcohol:cytochrome *c* oxidoreductase
Comments: A periplasmic PQQ-containing quinoprotein. Occurs in *Pseudomonas* and *Rhodospseudomonas*. The enzyme from *Pseudomonas aeruginosa* uses a specific inducible cytochrome *c*₅₅₀ as electron acceptor. Acts on a wide range of primary and secondary alcohols, but not methanol. It has a homodimeric structure [contrasting with the heterotetrameric structure of EC 1.1.2.7, methanol dehydrogenase (cytochrome *c*)]. It is routinely assayed with phenazine methosulfate as electron acceptor. Activity is stimulated by ammonia or amines. Like all other quinoprotein alcohol dehydrogenases it has an 8-bladed 'propeller' structure, a calcium ion bound to the PQQ in the active site and an unusual disulfide ring structure in close proximity to the PQQ.

References: [3605, 4319, 3739, 2058, 2049, 2770]

[EC 1.1.2.8 created 1972 as EC 1.1.99.8, modified 1982, part transferred 2010 to EC 1.1.2.8]

EC 1.1.2.9

Accepted name: 1-butanol dehydrogenase (cytochrome *c*)
Reaction: butan-1-ol + 2 ferricytochrome *c* = butanal + 2 ferrocycytochrome *c* + 2 H⁺
Other name(s): BDH
Systematic name: butan-1-ol:ferricytochrome *c* oxidoreductase
Comments: This periplasmic quinoprotein alcohol dehydrogenase, characterized from the bacterium *Thauera butanivorans*, is involved in butane degradation. It contains both pyrroloquinoline quinone (PQQ) and heme *c* prosthetic groups. *cf.* EC 1.1.5.11, 1-butanol dehydrogenase (quinone).
References: [4421, 4422, 4423]

[EC 1.1.2.9 created 2016]

EC 1.1.2.10

Accepted name: lanthanide-dependent methanol dehydrogenase
Reaction: methanol + 2 oxidized cytochrome *c_I* = formaldehyde + 2 reduced cytochrome *c_I*
Other name(s): XoxF; XoxF-MDH; Ce-MDH; La³⁺-dependent MDH; Ce³⁺-induced methanol dehydrogenase; cerium dependent MDH
Systematic name: methanol:cytochrome *c_I* oxidoreductase
Comments: Isolated from the bacterium *Methylococcus thermophilus* and many *Methylobacterium* species. Requires La³⁺, Ce³⁺, Pr³⁺ or Nd³⁺. The higher lanthanides show decreasing activity with Sm³⁺, Eu³⁺ and Gd³⁺. The lanthanide is coordinated by the enzyme and pyrroloquinoline quinone. Shows little activity with Ca²⁺, the required cofactor of EC 1.1.2.7, methanol dehydrogenase (cytochrome *c*).
References: [1638, 2974, 3344, 370, 3373, 2694]

[EC 1.1.2.10 created 2019]

EC 1.1.2.11

Accepted name: glucoside 3-dehydrogenase (cytochrome *c*)
Reaction: a D-glucoside + a ferric *c*-type cytochrome = a 3-dehydro-D-glucoside + a ferrous *c*-type cytochrome
Other name(s): D-glucoside 3-dehydrogenase (ambiguous); D-aldohexopyranoside dehydrogenase (ambiguous); D-aldohexoside:cytochrome *c* oxidoreductase; hexopyranoside-cytochrome *c* oxidoreductase
Systematic name: a D-glucoside:ferric *c*-type cytochrome 3-oxidoreductase
Comments: This bacterial enzyme acts on D-glucose, D-galactose, D-glucosides and D-galactosides, but the best substrates are disaccharides with a glucose moiety at the non-reducing end. It consists of two subunits, a catalytic subunit that contains an FAD cofactor and an iron-sulfur cluster, and a "hitch-hiker" subunit containing a signal peptide for translocation into the periplasm. A dedicated *c*-type cytochrome protein serves as an electron acceptor, transferring the electrons from the catalytic subunit to the cell's electron transfer chain. *cf.* EC 1.1.99.13, glucoside 3-dehydrogenase (acceptor).
References: [1576, 2979, 4184, 4183, 4340, 2207, 4877, 4876, 2843]

[EC 1.1.2.11 created 2022]

EC 1.1.3 With oxygen as acceptor

[1.1.3.1 Deleted entry. glycolate oxidase. Now included with EC 1.1.3.15 (S)-2-hydroxy-acid oxidase]

[EC 1.1.3.1 created 1961, deleted 1984]

EC 1.1.3.2

Accepted name: L-lactate oxidase
Reaction: (S)-lactate + O₂ = pyruvate + H₂O₂
Other name(s): *lctO* (gene name); LOX
Systematic name: (S)-lactate:oxygen 2-oxidoreductase
Comments: Contains flavin mononucleotide (FMN). The best characterized enzyme is that from the bacterium *Aerococcus viridans*. The enzyme is widely used in biosensors to measure the lactate concentration in blood and other tissues.
References: [983, 2603, 1319, 4376, 1239, 4043]

[EC 1.1.3.2 created 1961, transferred 1972 to EC 1.13.12.4, reinstated 2018]

[1.1.3.3 Deleted entry. malate oxidase. Now classified as EC 1.1.5.4, malate dehydrogenase (quinone).]

[EC 1.1.3.3 created 1961, deleted 2014]

EC 1.1.3.4

Accepted name: glucose oxidase
Reaction: β-D-glucose + O₂ = D-glucono-1,5-lactone + H₂O₂
Other name(s): glucose oxyhydrase; corylophyline; penatin; glucose aerodehydrogenase; microcid; β-D-glucose oxidase; D-glucose oxidase; D-glucose-1-oxidase; β-D-glucose:quinone oxidoreductase; glucose oxyhydrase; deoxin-1; GOD
Systematic name: β-D-glucose:oxygen 1-oxidoreductase
Comments: A flavoprotein (FAD).
References: [291, 750, 2055, 2056]

[EC 1.1.3.4 created 1961]

EC 1.1.3.5

Accepted name: hexose oxidase
Reaction: D-glucose + O₂ = D-glucono-1,5-lactone + H₂O₂
Systematic name: D-hexose:oxygen 1-oxidoreductase
Comments: A copper glycoprotein. Also oxidizes D-galactose, D-mannose, maltose, lactose and cellobiose.
References: [254, 255, 4111]

[EC 1.1.3.5 created 1961, modified 1976]

EC 1.1.3.6

Accepted name: cholesterol oxidase
Reaction: cholesterol + O₂ = cholest-5-en-3-one + H₂O₂
Other name(s): cholesterol- O₂ oxidoreductase; 3β-hydroxy steroid oxidoreductase; 3β-hydroxysteroid:oxygen oxidoreductase
Systematic name: cholesterol:oxygen oxidoreductase
Comments: Contains FAD. Cholesterol oxidases are secreted bacterial bifunctional enzymes that catalyse the first two steps in the degradation of cholesterol. The enzyme catalyses the oxidation of the 3β-hydroxyl group to a keto group, and the isomerization of the double bond in the oxidized steroid ring system from the Δ⁵ position to Δ⁶ position (*cf.* EC 5.3.3.1, steroid Δ-isomerase).
References: [3514, 3997, 2594, 4477]

[EC 1.1.3.6 created 1961, modified 1982, modified 2012]

EC 1.1.3.7

Accepted name: aryl-alcohol oxidase

Reaction: an aromatic primary alcohol + O₂ = an aromatic aldehyde + H₂O₂
Other name(s): aryl alcohol oxidase; veratryl alcohol oxidase; arom. alcohol oxidase
Systematic name: aryl-alcohol:oxygen oxidoreductase
Comments: Oxidizes many primary alcohols containing an aromatic ring; best substrates are (2-naphthyl)methanol and 3-methoxybenzyl alcohol.
References: [1092]

[EC 1.1.3.7 created 1965]

EC 1.1.3.8

Accepted name: L-gulonolactone oxidase
Reaction: L-gulono-1,4-lactone + O₂ = L-ascorbate + H₂O₂ (overall reaction)
(1a) L-gulono-1,4-lactone + O₂ = L-xylo-hex-2-ulono-1,4-lactone + H₂O₂
(1b) L-xylo-hex-2-ulono-1,4-lactone = L-ascorbate (spontaneous)
Other name(s): L-gulono-γ-lactone: O₂ oxidoreductase; L-gulono-γ-lactone oxidase; L-gulono-γ-lactone:oxidoreductase; GLO
Systematic name: L-gulono-1,4-lactone:oxygen 3-oxidoreductase
Comments: A microsomal flavoprotein (FAD). The product spontaneously isomerizes to L-ascorbate. While most higher animals can synthesize ascorbic acid, primates and guinea pigs cannot [3077].
References: [1826, 2141, 3077, 613]

[EC 1.1.3.8 created 1965, modified 2001, modified 2006]

EC 1.1.3.9

Accepted name: galactose oxidase
Reaction: D-galactose + O₂ = D-galacto-hexodialdose + H₂O₂
Other name(s): D-galactose oxidase; β-galactose oxidase
Systematic name: D-galactose:oxygen 6-oxidoreductase
Comments: A copper protein.
References: [159]

[EC 1.1.3.9 created 1965]

EC 1.1.3.10

Accepted name: pyranose oxidase
Reaction: D-glucose + O₂ = 2-dehydro-D-glucose + H₂O₂
Other name(s): glucose 2-oxidase; pyranose-2-oxidase
Systematic name: pyranose:oxygen 2-oxidoreductase
Comments: A flavoprotein (FAD). Also oxidizes D-xylose, L-sorbose and D-glucono-1,5-lactone, which have the same ring conformation and configuration at C-2, C-3 and C-4.
References: [1890, 2592, 3039, 3598]

[EC 1.1.3.10 created 1972]

EC 1.1.3.11

Accepted name: L-sorbose oxidase
Reaction: L-sorbose + O₂ = 5-dehydro-D-fructose + H₂O₂
Systematic name: L-sorbose:oxygen 5-oxidoreductase
Comments: Also acts on D-glucose, D-galactose and D-xylose, but not on D-fructose. 2,6-Dichloroindophenol can act as acceptor.
References: [4728]

[EC 1.1.3.11 created 1972]

EC 1.1.3.12

Accepted name: pyridoxine 4-oxidase
Reaction: pyridoxine + O₂ = pyridoxal + H₂O₂
Other name(s): pyridoxin 4-oxidase; pyridoxol 4-oxidase
Systematic name: pyridoxine:oxygen 4-oxidoreductase
Comments: A flavoprotein. Can also use 2,6-dichloroindophenol as an acceptor.
References: [4122]

[EC 1.1.3.12 created 1972, modified 1976]

EC 1.1.3.13

Accepted name: alcohol oxidase
Reaction: a primary alcohol + O₂ = an aldehyde + H₂O₂
Other name(s): ethanol oxidase; alcohol:oxygen oxidoreductase
Systematic name: alcohol:oxygen oxidoreductase (H₂O₂-forming)
Comments: The enzymes from the fungi *Candida methanosorbosa* and several *Basidiomycetes* species contain an FAD cofactor [1889, 4133]. The enzyme from the phytopathogenic fungi *Colletotrichum graminicola* and *Colletotrichum gloeosporioides* utilize a mononuclear copper-radical mechanism [4792]. The enzyme acts on primary alcohols and unsaturated alcohols, and has much lower activity with branched-chain and secondary alcohols.
References: [1889, 3073, 4133, 4792]

[EC 1.1.3.13 created 1972]

EC 1.1.3.14

Accepted name: catechol oxidase (dimerizing)
Reaction: 4 catechol + 3 O₂ = 2 dibenzo[1,4]dioxin-2,3-dione + 6 H₂O
Systematic name: catechol:oxygen oxidoreductase (dimerizing)
References: [2970]

[EC 1.1.3.14 created 1972]

EC 1.1.3.15

Accepted name: (*S*)-2-hydroxy-acid oxidase
Reaction: an (*S*)-2-hydroxy carboxylate + O₂ = a 2-oxo carboxylate + H₂O₂
Other name(s): hydroxy-acid oxidase A; hydroxy-acid oxidase B; glycolate oxidase; L-2-hydroxy acid oxidase; hydroxyacid oxidase A; L- α -hydroxy acid oxidase
Systematic name: (*S*)-2-hydroxy carboxylate:oxygen 2-oxidoreductase
Comments: A flavoprotein (FMN). Exists as two major isoenzymes; the A form preferentially oxidizes short-chain aliphatic hydroxy acids, and was previously listed as EC 1.1.3.1, glycolate oxidase; the B form preferentially oxidizes long-chain and aromatic hydroxy acids. The rat isoenzyme B also acts as EC 1.4.3.2, L-amino-acid oxidase.
References: [353, 1189, 2287, 2990, 2992, 3315, 3759, 1944]

[EC 1.1.3.15 created 1972 (EC 1.1.3.1 created 1961, incorporated 1984)]

EC 1.1.3.16

Accepted name: ecdysone oxidase
Reaction: ecdysone + O₂ = 3-dehydroecdysone + H₂O₂
Other name(s): β -ecdysone oxidase
Systematic name: ecdysone:oxygen 3-oxidoreductase
Comments: 2,6-Dichloroindophenol can act as an acceptor.

References: [2228]

[EC 1.1.3.16 created 1976]

EC 1.1.3.17

Accepted name: choline oxidase
Reaction: choline + 2 O₂ + H₂O = betaine + 2 H₂O₂ (overall reaction)
(1a) choline + O₂ = betaine aldehyde + H₂O₂
(1b) betaine aldehyde + O₂ + H₂O = betaine + H₂O₂
Systematic name: choline:oxygen 1-oxidoreductase
Comments: A flavoprotein (FAD). In many bacteria, plants and animals, the osmoprotectant betaine is synthesized using different enzymes to catalyse the conversion of (1) choline into betaine aldehyde and (2) betaine aldehyde into betaine. In plants, the first reaction is catalysed by EC 1.14.15.7, choline monooxygenase, whereas in animals and many bacteria, it is catalysed by either membrane-bound choline dehydrogenase (EC 1.1.99.1) or soluble choline oxidase (EC 1.1.3.17) [4485]. The enzyme involved in the second step, EC 1.2.1.8, betaine-aldehyde dehydrogenase, appears to be the same in those plants, animals and bacteria that use two separate enzymes.
References: [1801, 3588, 3448, 1252, 1083, 4485, 1084, 1249]

[EC 1.1.3.17 created 1978, modified 2005, modified 2007]

EC 1.1.3.18

Accepted name: secondary-alcohol oxidase
Reaction: a secondary alcohol + O₂ = a ketone + H₂O₂
Other name(s): polyvinyl alcohol oxidase; secondary alcohol oxidase
Systematic name: secondary-alcohol:oxygen oxidoreductase
Comments: Acts on secondary alcohols with five or more carbons, and polyvinyl alcohols with molecular mass over 300 Da. The *Pseudomonas* enzyme contains one atom of non-heme iron per molecule.
References: [2893, 3637, 4142, 4143]

[EC 1.1.3.18 created 1981]

EC 1.1.3.19

Accepted name: 4-hydroxymandelate oxidase (decarboxylating)
Reaction: (S)-4-hydroxymandelate + O₂ = 4-hydroxybenzaldehyde + CO₂ + H₂O₂
Other name(s): L-4-hydroxymandelate oxidase (decarboxylating); (S)-2-hydroxy-2-(4-hydroxyphenyl)acetate:oxygen 1-oxidoreductase; (S)-4-hydroxymandelate:oxygen 1-oxidoreductase; 4-hydroxymandelate oxidase
Systematic name: (S)-4-hydroxymandelate:oxygen 1-oxidoreductase (decarboxylating)
Comments: A flavoprotein (FAD), requires Mn²⁺. The enzyme from the bacterium *Pseudomonas putida* is involved in the degradation of mandelate.
References: [325]

[EC 1.1.3.19 created 1984, modified 2014]

EC 1.1.3.20

Accepted name: long-chain-alcohol oxidase
Reaction: a long-chain alcohol + O₂ = a long-chain aldehyde + H₂O₂
Other name(s): long-chain fatty alcohol oxidase; fatty alcohol oxidase; fatty alcohol:oxygen oxidoreductase; long-chain fatty acid oxidase
Systematic name: long-chain-alcohol:oxygen oxidoreductase
Comments: Oxidizes long-chain fatty alcohols; best substrate is dodecyl alcohol.
References: [2880, 2881, 644, 4903, 645]

[EC 1.1.3.20 created 1984, modified 2010]

EC 1.1.3.21

Accepted name: glycerol-3-phosphate oxidase
Reaction: *sn*-glycerol 3-phosphate + O₂ = glycerone phosphate + H₂O₂
Other name(s): glycerol phosphate oxidase; glycerol-1-phosphate oxidase; glycerol phosphate oxidase; L- α -glycerophosphate oxidase; α -glycerophosphate oxidase; L- α -glycerol-3-phosphate oxidase
Systematic name: *sn*-glycerol-3-phosphate:oxygen 2-oxidoreductase
Comments: A flavoprotein (FAD).
References: [1268, 2183]

[EC 1.1.3.21 created 1984]

[1.1.3.22 *Transferred entry. xanthine oxidase. Now EC 1.17.3.2, xanthine oxidase. The enzyme was incorrectly classified as acting on a CH-OH group*]

[EC 1.1.3.22 created 1961 as EC 1.2.3.2, transferred 1984 to EC 1.1.3.22, modified 1989, deleted 2004]

EC 1.1.3.23

Accepted name: thiamine oxidase
Reaction: thiamine + 2 O₂ + H₂O = thiamine acetic acid + 2 H₂O₂
Other name(s): thiamin dehydrogenase; thiamine dehydrogenase; thiamin:oxygen 5-oxidoreductase
Systematic name: thiamine:oxygen 5-oxidoreductase
Comments: A flavoprotein (FAD). The product differs from thiamine in replacement of -CH₂.CH₂.OH by -CH₂.COOH; the two-step oxidation proceeds without the release of the intermediate aldehyde from the enzyme.
References: [1018, 1355, 3030]

[EC 1.1.3.23 created 1984]

[1.1.3.24 *Transferred entry. L-galactonolactone oxidase. Now EC 1.3.3.12, L-galactonolactone oxidase. The enzyme had been incorrectly classified as acting upon a CH-OH donor rather than a CH-CH donor*]

[EC 1.1.3.24 created 1984, deleted 2006]

[1.1.3.25 *Transferred entry. cellobiose oxidase. Now included with EC 1.1.99.18, cellobiose dehydrogenase (acceptor)*]

[EC 1.1.3.25 created 1986, deleted 2005]

[1.1.3.26 *Transferred entry. columbamine oxidase. Now EC 1.21.3.2, columbamine oxidase*]

[EC 1.1.3.26 created 1989, deleted 2002]

EC 1.1.3.27

Accepted name: hydroxyphytanate oxidase
Reaction: L-2-hydroxyphytanate + O₂ = 2-oxophytanate + H₂O₂
Other name(s): L-2-hydroxyphytanate:oxygen 2-oxidoreductase
Systematic name: L-2-hydroxyphytanate:oxygen 2-oxidoreductase
References: [4395]

[EC 1.1.3.27 created 1990]

EC 1.1.3.28

Accepted name: nucleoside oxidase
Reaction: inosine + O₂ = 9-riburonosylhypoxanthine + H₂O
(1a) 2 inosine + O₂ = 2 5'-dehydroinosine + 2 H₂O
(1b) 2 5'-dehydroinosine + O₂ = 2 9-riburonosylhypoxanthine
Systematic name: nucleoside:oxygen 5'-oxidoreductase

Comments: Other purine and pyrimidine nucleosides (as well as 2'-deoxyribonucleosides) are substrates, but ribose and nucleotides are not substrates. The overall reaction takes place in two separate steps, with the 5'-dehydro nucleoside being released from the enzyme to serve as substrate for the second reaction. This enzyme differs from EC 1.1.3.39, nucleoside oxidase (H₂O₂-forming), as it produces water rather than hydrogen peroxide.

References: [1839, 1838]

[EC 1.1.3.28 created 1992, modified 2001]

EC 1.1.3.29

Accepted name: *N*-acylhexosamine oxidase

Reaction: (1) *N*-acetyl-D-glucosamine + O₂ + H₂O = *N*-acetyl-D-glucosamine + H₂O₂ (overall reaction)
(1a) *N*-acetyl-D-glucosamine + O₂ = *N*-acetyl-D-glucosamino-1,5-lactone + H₂O₂
(1b) *N*-acetyl-D-glucosamino-1,5-lactone + H₂O = *N*-acetyl-D-glucosamine (spontaneous)
(2) *N*-acetyl-D-galactosamine + O₂ + H₂O = *N*-acetyl-D-galactosamine + H₂O₂ (overall reaction)
(2a) *N*-acetyl-D-galactosamine + O₂ = *N*-acetyl-D-galactosamino-1,5-lactone + H₂O₂
(2b) *N*-acetyl-D-galactosamino-1,5-lactone + H₂O = *N*-acetyl-D-galactosamine (spontaneous)

Other name(s): *N*-acyl-D-hexosamine oxidase; *N*-acyl-β-D-hexosamine:oxygen 1-oxidoreductase

Systematic name: *N*-acyl-D-hexosamine:oxygen 1-oxidoreductase

Comments: The enzyme, found in bacteria, also acts more slowly on *N*-acetyl-D-mannosamine.

References: [1726, 3495]

[EC 1.1.3.29 created 1992, modified 2022]

EC 1.1.3.30

Accepted name: polyvinyl-alcohol oxidase

Reaction: polyvinyl alcohol + O₂ = oxidized polyvinyl alcohol + H₂O₂

Other name(s): dehydrogenase, polyvinyl alcohol; PVA oxidase

Systematic name: polyvinyl-alcohol:oxygen oxidoreductase

References: [3868, 3869]

[EC 1.1.3.30 created 1992]

[1.1.3.31 Deleted entry. methanol oxidase. Cannot be distinguished from EC 1.1.3.13, alcohol oxidase]

[EC 1.1.3.31 created 1992, deleted 2003]

[1.1.3.32 Transferred entry. (*S*)-stylopine synthase. Now EC 1.14.21.1, (*S*)-stylopine synthase]

[EC 1.1.3.32 created 1999, deleted 2002]

[1.1.3.33 Transferred entry. *S*-cheilanthifoline synthase. Now EC 1.14.21.2, (*S*)-cheilanthifoline synthase]

[EC 1.1.3.33 created 1999, deleted 2002]

[1.1.3.34 Transferred entry. berbaminine synthase. Now EC 1.14.21.3, berbaminine synthase]

[EC 1.1.3.34 created 1999, deleted 2002]

[1.1.3.35 Transferred entry. salutaridine synthase. Now EC 1.14.21.4, salutaridine synthase]

[EC 1.1.3.35 created 1999, deleted 2002]

[1.1.3.36 Transferred entry. (*S*)-canadine synthase. Now EC 1.14.21.5, (*S*)-canadine synthase]

[EC 1.1.3.36 created 1999, deleted 2002]

EC 1.1.3.37

- Accepted name:** D-arabinono-1,4-lactone oxidase
Reaction: D-arabinono-1,4-lactone + O₂ = dehydro-D-arabinono-1,4-lactone + H₂O₂
Other name(s): D-arabinono- γ -lactone oxidase; ALO
Systematic name: D-arabinono-1,4-lactone:oxygen oxidoreductase
Comments: A flavoprotein (FAD). L-Galactono-1,4-lactone, L-gulono-1,4-lactone and L-xylono-1,4-lactone can also act as substrates but D-glucono-1,5-lactone, L-arabinono-1,4-lactone, D-galactono-1,4-lactone and D-gulono-1,4-lactone cannot [1767]. With L-galactono-1,4-lactone as substrate, the product is L-ascorbate [2377]. The product dehydro-D-arabinono-1,4-lactone had previously been referred to erroneously as D-erythroascorbate (CAS no.: 5776-48-7; formula: C₆H₈O₆), although it was referred to as a five-carbon compound [1767].
References: [1767, 1768, 2377]

[EC 1.1.3.37 created 1999]

EC 1.1.3.38

- Accepted name:** vanillyl-alcohol oxidase
Reaction: vanillyl alcohol + O₂ = vanillin + H₂O₂
Other name(s): 4-hydroxy-2-methoxybenzyl alcohol oxidase
Systematic name: vanillyl alcohol:oxygen oxidoreductase
Comments: Vanillyl-alcohol oxidase from *Penicillium simplicissimum* contains covalently bound FAD. It converts a wide range of 4-hydroxybenzyl alcohols and 4-hydroxybenzylamines into the corresponding aldehydes. The allyl group of 4-allylphenols is also converted into the -CH=CH-CH₂OH group.
References: [846, 1158]

[EC 1.1.3.38 created 1999]

EC 1.1.3.39

- Accepted name:** nucleoside oxidase (H₂O₂-forming)
Reaction: adenosine + 2 O₂ + H₂O = 9-ribose-5-phosphoribosyladenine + 2 H₂O₂ (overall reaction)
(1a) adenosine + O₂ = 5'-dehydroadenosine + H₂O₂
(1b) 5'-dehydroadenosine + O₂ + H₂O = 9-ribose-5-phosphoribosyladenine + H₂O₂
Systematic name: nucleoside:oxygen 5'-oxidoreductase (H₂O₂-forming)
Comments: A heme-containing flavoprotein (FAD). Other purine and pyrimidine nucleosides (as well as 2'-deoxyribonucleosides and arabinosides) are substrates, but ribose and nucleotides are not substrates. The overall reaction takes place in two separate steps, with the 5'-dehydro nucleoside being released from the enzyme to serve as substrate for the second reaction. This enzyme differs from EC 1.1.3.28, nucleoside oxidase, as it produces hydrogen peroxide rather than water.
References: [2194]

[EC 1.1.3.39 created 2001]

EC 1.1.3.40

- Accepted name:** D-mannitol oxidase
Reaction: D-mannitol + O₂ = D-mannose + H₂O₂
Other name(s): mannitol oxidase; D-arabitol oxidase
Systematic name: mannitol:oxygen oxidoreductase (cyclizing)
Comments: Also catalyses the oxidation of D-arabinitol and, to a lesser extent, D-glucitol (sorbitol), whereas L-arabinitol is not a good substrate. The enzyme from the snails *Helix aspersa* and *Arion ater* is found in a specialised tubular organelle that has been termed the mannosome.
References: [4473, 2630, 2352, 796]

[EC 1.1.3.40 created 2001]

EC 1.1.3.41

- Accepted name:** alditol oxidase
Reaction: an alditol + O₂ = an aldose + H₂O₂
Other name(s): xylitol oxidase; xylitol:oxygen oxidoreductase; AldO
Systematic name: alditol:oxygen oxidoreductase
Comments: The enzyme from *Streptomyces* sp. IKD472 and from *Streptomyces coelicolor* is a monomeric oxidase containing one molecule of FAD per molecule of protein [4752, 1633]. While xylitol (five carbons) and sorbitol (6 carbons) are the preferred substrates, other alditols, including L-threitol (four carbons), D-arabinitol (five carbons), D-galactitol (six carbons) and D-mannitol (six carbons) can also act as substrates, but more slowly [4752, 1633]. Belongs in the vanillyl-alcohol-oxidase family of enzymes [1633].
References: [4752, 1633, 1145]

[EC 1.1.3.41 created 2002, modified 2008]

EC 1.1.3.42

- Accepted name:** prosolanapyrone-II oxidase
Reaction: prosolanapyrone II + O₂ = prosolanapyrone III + H₂O₂
Other name(s): Sol5 (ambiguous); SPS (ambiguous); solanapyrone synthase (bifunctional enzyme: prosolanapyrone II oxidase/prosolanapyrone III cycloisomerase); prosolanapyrone II oxidase
Systematic name: prosolanapyrone-II:oxygen 3'-oxidoreductase
Comments: The enzyme is involved in the biosynthesis of the phytotoxin solanapyrone by some fungi. The bifunctional enzyme catalyses the oxidation of prosolanapyrone II and the subsequent Diels Alder cycloisomerization of the product prosolanapyrone III to (-)-solanapyrone A (*cf.* EC 5.5.1.20, prosolanapyrone III cycloisomerase).
References: [2003, 2013, 2012]

[EC 1.1.3.42 created 2011]

EC 1.1.3.43

- Accepted name:** paromamine 6'-oxidase
Reaction: paromamine + O₂ = 6'-dehydroparomamine + H₂O₂
Other name(s): *btrQ* (gene name); *neoG* (gene name); *kanI* (gene name); *tacB* (gene name); *neoQ* (obsolete gene name)
Systematic name: paromamine:oxygen 6'-oxidoreductase
Comments: Contains FAD. Involved in the biosynthetic pathways of several clinically important aminocyclitol antibiotics, including kanamycin, butirosin, neomycin and ribostamycin. Works in combination with EC 2.6.1.93, neamine transaminase, to replace the 6'-hydroxy group of paromamine with an amino group. The enzyme from the bacterium *Streptomyces fradiae* also catalyses EC 1.1.3.44, 6'''-hydroxyneomycin C oxidase.
References: [1751, 4839, 700]

[EC 1.1.3.43 created 2012]

EC 1.1.3.44

- Accepted name:** 6'''-hydroxyneomycin C oxidase
Reaction: 6'''-deamino-6'''-hydroxyneomycin C + O₂ = 6'''-deamino-6'''-oxoneomycin C + H₂O₂
Other name(s): *neoG* (gene name); *neoQ* (obsolete gene name)
Systematic name: 6'''-deamino-6'''-hydroxyneomycin C:oxygen 6'''-oxidoreductase
Comments: Contains FAD. Involved in the biosynthetic pathway of aminoglycoside antibiotics of the neomycin family. Works in combination with EC 2.6.1.95, neomycin C transaminase, to replace the 6'''-hydroxy group of 6'''-hydroxyneomycin C with an amino group. Also catalyses EC 1.1.3.43, paromamine 6'-oxidase.

References: [1751, 700]

[EC 1.1.3.44 created 2012]

EC 1.1.3.45

Accepted name: aclacinomycin-N oxidase
Reaction: aclacinomycin N + O₂ = aclacinomycin A + H₂O₂
Other name(s): AknOx (ambiguous); aclacinomycin oxidoreductase (ambiguous)
Systematic name: aclacinomycin-N:oxygen oxidoreductase
Comments: A flavoprotein (FAD). This bifunctional enzyme is a secreted flavin-dependent enzyme that is involved in the modification of the terminal sugar residues in the biosynthesis of aclacinomycins. The enzyme utilizes the same active site to catalyse the oxidation of the rhodnose moiety of aclacinomycin N to the cinerulose A moiety of aclacinomycin A and the oxidation of the latter to the L-aculose moiety of aclacinomycin Y (*cf.* EC 1.3.3.14, aclacinomycin A oxidase).
References: [66, 4112]

[EC 1.1.3.45 created 2013]

EC 1.1.3.46

Accepted name: 4-hydroxymandelate oxidase
Reaction: (S)-4-hydroxymandelate + O₂ = 2-(4-hydroxyphenyl)-2-oxoacetate + H₂O₂
Other name(s): 4HmO; HmO
Systematic name: (S)-4-hydroxymandelate:oxygen 1-oxidoreductase
Comments: A flavoprotein (FMN). The enzyme from the bacterium *Amycolatopsis orientalis* is involved in the biosynthesis of L-(4-hydroxyphenyl)glycine and L-(3,5-dihydroxyphenyl)glycine, two non-proteinogenic amino acids occurring in the vancomycin group of antibiotics.
References: [1758, 2459]

[EC 1.1.3.46 created 2014]

EC 1.1.3.47

Accepted name: 5-(hydroxymethyl)furfural oxidase
Reaction: 5-(hydroxymethyl)furfural + 3 O₂ + 2 H₂O = furan-2,5-dicarboxylate + 3 H₂O₂ (overall reaction)
(1a) 5-(hydroxymethyl)furfural + O₂ = furan-2,5-dicarbalddehyde + H₂O₂
(1b) furan-2,5-dicarbalddehyde + H₂O = 5-(dihydroxymethyl)furan-2-carbaldehyde (spontaneous)
(1c) 5-(dihydroxymethyl)furan-2-carbaldehyde + O₂ = 5-formylfuran-2-carboxylate + H₂O₂
(1d) 5-formylfuran-2-carboxylate + H₂O = 5-(dihydroxymethyl)furan-2-carboxylate (spontaneous)
(1e) 5-(dihydroxymethyl)furan-2-carboxylate + O₂ = furan-2,5-dicarboxylate + H₂O₂
Systematic name: 5-(hydroxymethyl)furfural:oxygen oxidoreductase
Comments: The enzyme, characterized from the bacterium *Methylovorus* sp. strain MP688, is involved in the degradation and detoxification of 5-(hydroxymethyl)furfural. The enzyme acts only on alcohol groups and requires the spontaneous hydration of aldehyde groups for their oxidation [914]. The enzyme has a broad substrate range that overlaps with EC 1.1.3.7, aryl-alcohol oxidase.
References: [2229, 913, 914]

[EC 1.1.3.47 created 2014]

EC 1.1.3.48

Accepted name: 3-deoxy- α -D-manno-octulosonate 8-oxidase
Reaction: 3-deoxy- α -D-manno-octulopyranosonate + O₂ = 3,8-dideoxy-8-oxo- α -D-manno-octulosonate + H₂O₂
Other name(s): *kdnB* (gene name)
Systematic name: 3-deoxy- α -D-manno-octulopyranosonate:oxygen 8-oxidoreductase

Comments: The enzyme, characterized from the bacterium *Shewanella oneidensis*, is involved in the formation of 8-amino-3,8-dideoxy- α -D-manno-octulosonate, an aminated form of Kdo found in lipopolysaccharides of members of the *Shewanella* genus. *cf.* EC 2.6.1.109, 8-amino-3,8-dideoxy- α -D-manno-octulosonate transaminase.

References: [1280]

[EC 1.1.3.48 created 2015]

EC 1.1.3.49

Accepted name: (R)-mandelonitrile oxidase

Reaction: (R)-mandelonitrile + O₂ = benzoyl cyanide + H₂O₂

Other name(s): ChuaMOX (gene name)

Systematic name: (R)-mandelonitrile:oxygen oxidoreductase

Comments: Contains FAD. The enzyme, characterized from the millipede *Chamberlinius hualienensis*, is segregated from its substrate, which is contained in special sacs. The sacs are ruptured during defensive behavior, allowing the enzyme and substrate to mix in special reaction chambers leading to production of the defensive chemical benzoyl cyanide.

References: [1829]

[EC 1.1.3.49 created 2016]

EC 1.1.3.50

Accepted name: C-glycoside oxidase

Reaction: carminate + O₂ = 3'-dehydrocarminate + H₂O₂

Other name(s): *carA* (gene name)

Systematic name: carminate:oxygen 3'-oxidoreductase (H₂O₂-forming)

Comments: A flavoprotein (FAD). This bacterial enzyme participates in degradation of certain C-glucosides by catalysing the oxidation of the hydroxyl group at position 3 of the glucose moiety. The enzyme was found active with assorted C-glycosides, such as carminate, mangiferin, and C⁶-glycosylated flavonoids, but not with D-glucose or C⁸-glycosylated flavonoids.

References: [2281]

[EC 1.1.3.50 created 2022]

EC 1.1.4 With a disulfide as acceptor

[1.1.4.1 *Transferred entry. vitamin-K-epoxide reductase (warfarin-sensitive). Now EC 1.17.4.4, vitamin-K-epoxide reductase (warfarin-sensitive)*]

[EC 1.1.4.1 created 1989, deleted 2014]

[1.1.4.2 *Transferred entry. vitamin-K-epoxide reductase (warfarin-insensitive). Now EC 1.17.4.5, vitamin-K-epoxide reductase (warfarin-insensitive)*]

[EC 1.1.4.2 created 1989, deleted 2014]

EC 1.1.5 With a quinone or similar compound as acceptor

[1.1.5.1 *Deleted entry. cellobiose dehydrogenase (quinone). Now known to be proteolytic product of EC 1.1.99.18, cellobiose dehydrogenase (acceptor)*]

[EC 1.1.5.1 created 1983, deleted 2002]

EC 1.1.5.2

- Accepted name:** glucose 1-dehydrogenase (PQQ, quinone)
Reaction: D-glucose + ubiquinone = D-glucono-1,5-lactone + ubiquinol
Other name(s): quinoprotein glucose dehydrogenase; membrane-bound glucose dehydrogenase; mGDH; glucose dehydrogenase (PQQ-dependent); glucose dehydrogenase (pyrroloquinoline-quinone); quinoprotein D-glucose dehydrogenase
- Systematic name:** D-glucose:ubiquinone oxidoreductase
Comments: Integral membrane protein containing PQQ as prosthetic group. It also contains bound ubiquinone and Mg²⁺ or Ca²⁺. Electron acceptor is membrane ubiquinone but usually assayed with phenazine methosulfate. Like in all other quinoprotein alcohol dehydrogenases the catalytic domain has an 8-bladed propeller structure. It occurs in a wide range of bacteria. Catalyses a direct oxidation of the pyranose form of D-glucose to the lactone and thence to D-gluconate in the periplasm. Oxidizes other monosaccharides including the pyranose forms of pentoses.
References: [4726, 893, 981, 80, 760, 762, 1036, 1885, 1035, 2951]

[EC 1.1.5.2 created 1982 as EC 1.1.99.17, transferred 2003 to EC 1.1.5.2, modified 2010]

EC 1.1.5.3

- Accepted name:** glycerol-3-phosphate dehydrogenase
Reaction: *sn*-glycerol 3-phosphate + a quinone = glycerone phosphate + a quinol
Other name(s): α -glycerophosphate dehydrogenase; α -glycerophosphate dehydrogenase (acceptor); anaerobic glycerol-3-phosphate dehydrogenase; DL-glycerol 3-phosphate oxidase (misleading); FAD-dependent glycerol-3-phosphate dehydrogenase; FAD-dependent *sn*-glycerol-3-phosphate dehydrogenase; FAD-GPDH; FAD-linked glycerol 3-phosphate dehydrogenase; FAD-linked L-glycerol-3-phosphate dehydrogenase; flavin-linked glycerol-3-phosphate dehydrogenase; flavoprotein-linked L-glycerol 3-phosphate dehydrogenase; glycerol 3-phosphate cytochrome *c* reductase (misleading); glycerol phosphate dehydrogenase; glycerol phosphate dehydrogenase (acceptor); glycerol phosphate dehydrogenase (FAD); glycerol-3-phosphate CoQ reductase; glycerol-3-phosphate dehydrogenase (flavin-linked); glycerol-3-phosphate:CoQ reductase; glycerophosphate dehydrogenase; L-3-glycerophosphate-ubiquinone oxidoreductase; L-glycerol-3-phosphate dehydrogenase (ambiguous); L-glycerophosphate dehydrogenase; mGPD; mitochondrial glycerol phosphate dehydrogenase; NAD⁺-independent glycerol phosphate dehydrogenase; pyridine nucleotide-independent L-glycerol 3-phosphate dehydrogenase; *sn*-glycerol 3-phosphate oxidase (misleading); *sn*-glycerol-3-phosphate dehydrogenase; *sn*-glycerol-3-phosphate:(acceptor) 2-oxidoreductase; *sn*-glycerol-3-phosphate:acceptor 2-oxidoreductase
- Systematic name:** *sn*-glycerol 3-phosphate:quinone oxidoreductase
Comments: This flavin-dependent dehydrogenase is an essential membrane enzyme, functioning at the central junction of glycolysis, respiration and phospholipid biosynthesis. In bacteria, the enzyme is localized to the cytoplasmic membrane [4509], while in eukaryotes it is tightly bound to the outer surface of the inner mitochondrial membrane [3747]. In eukaryotes, this enzyme, together with the cytosolic enzyme EC 1.1.1.8, glycerol-3-phosphate dehydrogenase (NAD⁺), forms the glycerol-3-phosphate shuttle by which NADH produced in the cytosol, primarily from glycolysis, can be reoxidized to NAD⁺ by the mitochondrial electron-transport chain [2589]. This shuttle plays a critical role in transferring reducing equivalents from cytosolic NADH into the mitochondrial matrix [106, 2359]. Insect flight muscle uses only CoQ₁₀ as the physiological quinone whereas hamster and rat mitochondria use mainly CoQ₉ [3458]. The enzyme is activated by calcium [2589].
References: [3525, 3747, 2589, 3458, 3842, 4509, 106, 2359]

[EC 1.1.5.3 created 1961 as EC 1.1.2.1, transferred 1965 to EC 1.1.99.5, transferred 2009 to EC 1.1.5.3]

EC 1.1.5.4

- Accepted name:** malate dehydrogenase (quinone)
Reaction: (S)-malate + a quinone = oxaloacetate + reduced quinone

Other name(s): FAD-dependent malate-vitamin K reductase; malate-vitamin K reductase; (*S*)-malate:(acceptor) oxidoreductase; L-malate-quinone oxidoreductase; malate:quinone oxidoreductase; malate quinone oxidoreductase; MQO; malate:quinone reductase; malate dehydrogenase (acceptor); FAD-dependent malate dehydrogenase

Systematic name: (*S*)-malate:quinone oxidoreductase

Comments: A flavoprotein (FAD). Vitamin K and several other quinones can act as acceptors. Different from EC 1.1.1.37 (malate dehydrogenase (NAD⁺)), EC 1.1.1.82 (malate dehydrogenase (NADP⁺)) and EC 1.1.1.299 (malate dehydrogenase [NAD(P)⁺]).

References: [1802, 1803, 3475, 2863, 2014]

[EC 1.1.5.4 created 1978 as EC 1.1.99.16, transferred 2009 to EC 1.1.5.4]

EC 1.1.5.5

Accepted name: alcohol dehydrogenase (quinone)

Reaction: ethanol + ubiquinone = acetaldehyde + ubiquinol

Other name(s): type III ADH; membrane associated quinohaemoprotein alcohol dehydrogenase

Systematic name: alcohol:quinone oxidoreductase

Comments: Only described in acetic acid bacteria where it is involved in acetic acid production. Associated with membrane. Electron acceptor is membrane ubiquinone. A model structure suggests that, like all other quinoprotein alcohol dehydrogenases, the catalytic subunit has an 8-bladed 'propeller' structure, a calcium ion bound to the PQQ in the active site and an unusual disulfide ring structure in close proximity to the PQQ; the catalytic subunit also has a heme *c* in the C-terminal domain. The enzyme has two additional subunits, one of which contains three molecules of heme *c*. It does not require amines for activation. It has a restricted substrate specificity, oxidizing a few primary alcohols (C₂ to C₆), but not methanol, secondary alcohols and some aldehydes. It is assayed with phenazine methosulfate or with ferricyanide.

References: [1354, 3884, 654, 1173, 2713, 2719, 2716, 2717, 761]

[EC 1.1.5.5 created 2009, modified 2010]

[1.1.5.6 *Transferred entry. formate dehydrogenase-N. Now EC 1.17.5.3, formate dehydrogenase-N*]

[EC 1.1.5.6 created 2010, deleted 2017]

EC 1.1.5.7

Accepted name: cyclic alcohol dehydrogenase (quinone)

Reaction: a cyclic alcohol + a quinone = a cyclic ketone + a quinol

Other name(s): cyclic alcohol dehydrogenase; MCAD

Systematic name: cyclic alcohol:quinone oxidoreductase

Comments: This enzyme oxidizes a wide variety of cyclic alcohols. Some minor enzyme activity is found with aliphatic secondary alcohols and sugar alcohols, but not primary alcohols. The enzyme is unable to catalyse the reverse reaction of cyclic ketones or aldehydes to cyclic alcohols. This enzyme differs from EC 1.1.5.5, alcohol dehydrogenase (quinone), which shows activity with ethanol [2872].

References: [2872]

[EC 1.1.5.7 created 2010]

EC 1.1.5.8

Accepted name: quinate/shikimate dehydrogenase (quinone)

Reaction: quinate + quinone = 3-dehydroquininate + quinol

Other name(s): NAD(P)⁺-independent quinate dehydrogenase; quinate:pyrroloquinoline-quinone 5-oxidoreductase; quinate dehydrogenase (quinone)

Systematic name: quinate:quinol 3-oxidoreductase

Comments: The enzyme is membrane-bound. Does not use NAD(P)⁺ as acceptor. Contains pyrroloquinoline-quinone. *cf.* EC 1.1.1.24, quinate/shikimate dehydrogenase (NAD⁺), EC 1.1.1.282, quinate/shikimate dehydrogenase [NAD(P)⁺], and EC 1.1.1.25, shikimate dehydrogenase (NADP⁺).

References: [4411, 16, 4424]

[EC 1.1.5.8 created 1992 as EC 1.1.99.25, modified 2004, transferred 2010 to EC 1.1.5.8, modified 2021]

EC 1.1.5.9

Accepted name: glucose 1-dehydrogenase (FAD, quinone)
Reaction: D-glucose + a quinone = D-glucono-1,5-lactone + a quinol
Other name(s): glucose dehydrogenase (*Aspergillus*); FAD-dependent glucose dehydrogenase; D-glucose:(acceptor) 1-oxidoreductase; glucose dehydrogenase (acceptor); *gdh* (gene name)
Systematic name: D-glucose:quinone 1-oxidoreductase
Comments: A glycoprotein containing one mole of FAD per mole of enzyme. 2,6-Dichloroindophenol can act as acceptor. *cf.* EC 1.1.5.2, glucose 1-dehydrogenase (PQQ, quinone).
References: [189, 580, 2546, 1812, 4152, 4153]

[EC 1.1.5.9 created 1972 as EC 1.1.99.10, modified 1976, transferred 2013 to EC 1.1.5.9]

EC 1.1.5.10

Accepted name: D-2-hydroxyacid dehydrogenase (quinone)
Reaction: (R)-2-hydroxyacid + a quinone = 2-oxoacid + a quinol
Other name(s): (R)-2-hydroxy acid dehydrogenase; (R)-2-hydroxy-acid:(acceptor) 2-oxidoreductase; D-lactate dehydrogenase (ambiguous)
Systematic name: (R)-2-hydroxyacid:quinone oxidoreductase
Comments: The enzyme from mammalian kidney contains one mole of FAD per mole of enzyme. (R)-lactate, (R)-malate and *meso*-tartrate are good substrates. Ubiquinone-1 and the dye 2,6-dichloroindophenol can act as acceptors; NAD⁺ and NADP⁺ are not acceptors.
References: [4348, 4349, 536, 537]

[EC 1.1.5.10 created 2014]

EC 1.1.5.11

Accepted name: 1-butanol dehydrogenase (quinone)
Reaction: butan-1-ol + a quinone = butanal + a quinol
Other name(s): BOH
Systematic name: butan-1-ol:quinone oxidoreductase
Comments: This periplasmic quinoprotein alcohol dehydrogenase, characterized from the bacterium *Thauera butanivorans*, is involved in butane degradation. It contains a pyrroloquinoline quinone (PQQ) prosthetic group. *cf.* EC 1.1.2.9, 1-butanol dehydrogenase (cytochrome *c*).
References: [4422, 4423]

[EC 1.1.5.11 created 2016]

EC 1.1.5.12

Accepted name: D-lactate dehydrogenase (quinone)
Reaction: (R)-lactate + a quinone = pyruvate + a quinol
Other name(s): *dld* (gene name)
Systematic name: (R)-lactate:quinone 2-oxidoreductase
Comments: The enzyme is an FAD-dependent peripheral membrane dehydrogenase that participates in respiration. Electrons derived from D-lactate oxidation are transferred to the membrane soluble quinone pool.
References: [2202, 1241, 2712, 3281, 997]

[EC 1.1.5.12 created 2017]

EC 1.1.5.13

Accepted name: (*S*)-2-hydroxyglutarate dehydrogenase
Reaction: (*S*)-2-hydroxyglutarate + a quinone = 2-oxoglutarate + a quinol
Other name(s): L-2-hydroxyglutarate dehydrogenase; *lhgO* (gene name); *ygaF* (gene name)
Systematic name: (*S*)-2-hydroxyglutarate:quinone oxidoreductase
Comments: The enzyme, characterized from the bacterium *Escherichia coli*, contains an FAD cofactor that is not covalently attached. It is bound to the cytoplasmic membrane and is coupled to the membrane quinone pool.
References: [1982, 2172]

[EC 1.1.5.13 created 2019]

EC 1.1.5.14

Accepted name: fructose 5-dehydrogenase
Reaction: D-fructose + a ubiquinone = 5-dehydro-D-fructose + a ubiquinol
Other name(s): fructose 5-dehydrogenase (acceptor); D-fructose dehydrogenase; D-fructose:(acceptor) 5-oxidoreductase
Systematic name: D-fructose:ubiquinone 5-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Gluconobacter japonicus*, is a heterotrimer composed of an FAD-containing large subunit, a small subunit, and a heme *c*-containing subunit, which is responsible for anchoring the complex to the cytoplasmic membrane and for transferring the electrons to ubiquinone.
References: [4727, 79, 2996, 2045]

[EC 1.1.5.14 created 1972 as EC 1.1.99.11, transferred 2021 to EC 1.1.5.14]

EC 1.1.7 With an iron-sulfur protein as acceptor

EC 1.1.7.1

Accepted name: 4-hydroxybenzoyl-CoA reductase
Reaction: benzoyl-CoA + oxidized ferredoxin + H₂O = 4-hydroxybenzoyl-CoA + reduced ferredoxin
Other name(s): 4-hydroxybenzoyl-CoA reductase (dehydroxylating); 4-hydroxybenzoyl-CoA:(acceptor) oxidoreductase; benzoyl-CoA:acceptor oxidoreductase
Systematic name: benzoyl-CoA:oxidized ferredoxin oxidoreductase
Comments: A molybdenum-flavin-iron-sulfur protein that is involved in the anaerobic pathway of phenol metabolism in bacteria. A ferredoxin with two [4Fe-4S] clusters functions as the natural electron donor [432].
References: [1344, 1605, 432, 417, 1606]

[EC 1.1.7.1 created 2000 as EC 1.3.99.20, transferred 2011 to EC 1.3.7.9, transferred 2020 to EC 1.1.7.1]

EC 1.1.9 With a copper protein as acceptor

EC 1.1.9.1

Accepted name: alcohol dehydrogenase (azurin)
Reaction: a primary alcohol + azurin = an aldehyde + reduced azurin
Other name(s): type II quinoprotein alcohol dehydrogenase; quinohaemoprotein ethanol dehydrogenase; QHEDH; ADHIIB
Systematic name: alcohol:azurin oxidoreductase

Comments: A soluble, periplasmic PQQ-containing quinohemoprotein. Also contains a single heme *c*. Occurs in *Comamonas* and *Pseudomonas*. Does not require an amine activator. Oxidizes a wide range of primary and secondary alcohols, and also aldehydes and large substrates such as sterols; methanol is not a substrate. Usually assayed with phenazine methosulfate or ferricyanide. Like all other quinoprotein alcohol dehydrogenases it has an 8-bladed 'propeller' structure, a calcium ion bound to the PQQ in the active site and an unusual disulfide ring structure in close proximity to the PQQ.

References: [1417, 847, 4319, 2720, 641, 3211]

[EC 1.1.9.1 created 2010 as EC 1.1.98.1; transferred 2011 to EC 1.1.9.1]

EC 1.1.98 With other, known, physiological acceptors

[1.1.98.1 Transferred entry. Now EC 1.1.9.1, alcohol dehydrogenase (azurin)]

[EC 1.1.98.1 created 2010, deleted 2011]

EC 1.1.98.2

Accepted name: glucose-6-phosphate dehydrogenase (coenzyme-F₄₂₀)
Reaction: D-glucose 6-phosphate + oxidized coenzyme F₄₂₀ = 6-phospho-D-glucono-1,5-lactone + reduced coenzyme F₄₂₀
Other name(s): coenzyme F₄₂₀-dependent glucose-6-phosphate dehydrogenase; F₄₂₀-dependent glucose-6-phosphate dehydrogenase; FGD1; Rv0407; F₄₂₀-dependent glucose-6-phosphate dehydrogenase 1
Systematic name: D-glucose-6-phosphate:F₄₂₀ 1-oxidoreductase
Comments: The enzyme is very specific for D-glucose 6-phosphate. No activity with NAD⁺, NADP⁺, FAD and FMN [3396].
References: [3396, 231, 3397]

[EC 1.1.98.2 created 2010 as EC 1.1.99.34, transferred 2011 to EC 1.1.98.2]

EC 1.1.98.3

Accepted name: decaprenylphospho-β-D-ribofuranose 2-dehydrogenase
Reaction: *trans*,*octacis*-decaprenylphospho-β-D-ribofuranose + FAD = *trans*,*octacis*-decaprenylphospho-β-D-erythro-pentofuranosid-2-ulose + FADH₂
Other name(s): decaprenylphosphoryl-β-D-ribofuranose 2'-epimerase; Rv3790; DprE1; decaprenylphospho-β-D-ribofuranose 2-oxidase
Systematic name: *trans*,*octacis*-decaprenylphospho-β-D-ribofuranose:FAD 2-oxidoreductase
Comments: The enzyme, isolated from the bacterium *Mycobacterium smegmatis*, is involved, along with EC 1.1.1.333, decaprenylphospho-D-erythro-pentofuranosid-2-ulose 2-reductase, in the epimerization of *trans*,*octacis*-decaprenylphospho-β-D-ribofuranose to *trans*,*octacis*-decaprenylphospho-β-D-arabinofuranose, the arabinosyl donor for the biosynthesis of mycobacterial cell wall arabinan polymers.
References: [3512, 4326]

[EC 1.1.98.3 created 2012, modified 2014]

EC 1.1.98.4

Accepted name: F₄₂₀H₂:quinone oxidoreductase
Reaction: a quinol + oxidized coenzyme F₄₂₀ = a quinone + reduced coenzyme F₄₂₀
Other name(s): FqoF protein
Systematic name: quinol:coenzyme-F₄₂₀ oxidoreductase
Comments: An enzyme complex that contains FAD and iron-sulfur clusters. The enzyme has been described in the archaea *Methanosarcina mazei* and *Archaeoglobus fulgidus*.
References: [467, 2293, 3]

[EC 1.1.98.4 created 2013]

EC 1.1.98.5

- Accepted name:** secondary-alcohol dehydrogenase (coenzyme-F₄₂₀)
Reaction: R-CHOH-R' + oxidized coenzyme F₄₂₀ = R-CO-R' + reduced coenzyme F₄₂₀
Other name(s): F₄₂₀-dependent alcohol dehydrogenase; secondary alcohol:F₄₂₀ oxidoreductase; F₄₂₀-dependent secondary alcohol dehydrogenase
Systematic name: secondary-alcohol:coenzyme F₄₂₀ oxidoreductase
Comments: The enzyme isolated from the methanogenic archaea *Methanogenium liminatans* catalyses the reversible oxidation of various secondary and cyclic alcohols to the corresponding ketones.
References: [359, 154]

[EC 1.1.98.5 created 2013]

EC 1.1.98.6

- Accepted name:** ribonucleoside-triphosphate reductase (formate)
Reaction: ribonucleoside 5'-triphosphate + formate = 2'-deoxyribonucleoside 5'-triphosphate + CO₂ + H₂O
Other name(s): *nrdD* (gene name); class III ribonucleoside-triphosphate reductase; anaerobic ribonucleotide reductase; anaerobic ribonucleoside-triphosphate reductase
Systematic name: ribonucleoside-5'-triphosphate:formate 2'-oxidoreductase
Comments: The enzyme, which is expressed in the bacterium *Escherichia coli* during anaerobic growth, contains an iron sulfur center. The active form of the enzyme contains an oxygen-sensitive glycyI (1-amino-2-oxoethan-1-yl) radical that is generated by the activating enzyme NrdG via chemistry involving *S*-adenosylmethionine (SAM) and a [4Fe-4S] cluster. The glycyI radical is involved in generation of a transient thiyl (sulfanyl) radical on a cysteine residue, which attacks the substrate, forming a ribonucleotide 3'-radical, followed by water loss to form a ketyl (α -oxoalkyl) radical. The ketyl radical gains an electron from a cysteine residue and a proton from formic acid, forming 3'-keto-deoxyribonucleotide and generating a thiosulfuranyl (1 λ^4 -disulfan-1-yl) radical bridge between methionine and cysteine residues. Oxidation of formate by the thiosulfuranyl radical results in the release of CO₂ and regeneration of the thiyl radical. *cf.* EC 1.17.4.1, ribonucleoside-diphosphate reductase and EC 1.17.4.2, ribonucleoside-triphosphate reductase (thioredoxin).
References: [1037, 2930, 2931, 3172, 4574]

[EC 1.1.98.6 created 2017]

EC 1.1.98.7

- Accepted name:** serine-type anaerobic sulfatase-maturating enzyme
Reaction: *S*-adenosyl-L-methionine + a [sulfatase]-L-serine = a [sulfatase]-3-oxo-L-alanine + 5'-deoxyadenosine + L-methionine
Other name(s): *atsB* (gene name)
Systematic name: [sulfatase]-L-serine:*S*-adenosyl-L-methionine oxidoreductase (3-oxo-L-alanine-forming)
Comments: A bacterial radical *S*-adenosyl-L-methionine (AdoMet) enzyme that contains three [4Fe-4S] clusters. The enzyme, found in some bacteria, activates a type I sulfatase enzyme (EC 3.1.6.1) by converting a conserved L-serine residue in the active site to a unique 3-oxo-L-alanine residue that is essential for the sulfatase activity. While the enzyme from *Klebsiella pneumoniae* is specific for L-serine, the enzyme from *Clostridium perfringens* can also act on L-cysteine, see EC 1.8.98.7, cysteine-type anaerobic sulfatase-maturating enzyme.
References: [4155, 1087, 1431]

[EC 1.1.98.7 created 2020]

EC 1.1.99 With unknown physiological acceptors

EC 1.1.99.1

- Accepted name:** choline dehydrogenase
Reaction: choline + acceptor = betaine aldehyde + reduced acceptor
Other name(s): choline oxidase; choline-cytochrome *c* reductase; choline:(acceptor) oxidoreductase; choline:(acceptor) 1-oxidoreductase
Systematic name: choline:acceptor 1-oxidoreductase
Comments: A quinoprotein. In many bacteria, plants and animals, the osmoprotectant betaine is synthesized using different enzymes to catalyse the conversion of (1) choline into betaine aldehyde and (2) betaine aldehyde into betaine. In plants, the first reaction is catalysed by EC 1.14.15.7, choline monooxygenase, whereas in animals and many bacteria, it is catalysed by either membrane-bound choline dehydrogenase (EC 1.1.99.1) or soluble choline oxidase (EC 1.1.3.17) [4485]. The enzyme involved in the second step, EC 1.2.1.8, betaine-aldehyde dehydrogenase, appears to be the same in plants, animals and bacteria.
References: [83, 1010, 1251, 4485]

[EC 1.1.99.1 created 1961, modified 1989, modified 2005]

EC 1.1.99.2

- Accepted name:** L-2-hydroxyglutarate dehydrogenase
Reaction: (S)-2-hydroxyglutarate + acceptor = 2-oxoglutarate + reduced acceptor
Other name(s): α -ketoglutarate reductase; α -hydroxyglutarate dehydrogenase; L- α -hydroxyglutarate dehydrogenase; hydroxyglutaric dehydrogenase; α -hydroxyglutarate oxidoreductase; L- α -hydroxyglutarate:NAD⁺ 2-oxidoreductase; α -hydroxyglutarate dehydrogenase (NAD⁺ specific); (S)-2-hydroxyglutarate:(acceptor) 2-oxidoreductase
Systematic name: (S)-2-hydroxyglutarate:acceptor 2-oxidoreductase
References: [4575]

[EC 1.1.99.2 created 1961, modified 2013]

EC 1.1.99.3

- Accepted name:** gluconate 2-dehydrogenase (acceptor)
Reaction: D-gluconate + acceptor = 2-dehydro-D-gluconate + reduced acceptor
Other name(s): gluconate oxidase; gluconate dehydrogenase; gluconic dehydrogenase; D-gluconate dehydrogenase; gluconic acid dehydrogenase; 2-ketogluconate reductase; D-gluconate dehydrogenase, 2-keto-D-gluconate-yielding; D-gluconate:(acceptor) 2-oxidoreductase
Systematic name: D-gluconate:acceptor 2-oxidoreductase
Comments: A flavoprotein (FAD).
References: [2715, 3443]

[EC 1.1.99.3 created 1961, modified 1976, modified 1989]

EC 1.1.99.4

- Accepted name:** dehydrogluconate dehydrogenase
Reaction: 2-dehydro-D-gluconate + acceptor = 2,5-didehydro-D-gluconate + reduced acceptor
Other name(s): ketogluconate dehydrogenase; α -ketogluconate dehydrogenase; 2-keto-D-gluconate dehydrogenase; 2-oxogluconate dehydrogenase
Systematic name: 2-dehydro-D-gluconate:acceptor 2-oxidoreductase
Comments: A flavoprotein.
References: [831, 3882]

[EC 1.1.99.4 created 1961, modified 1989]

[1.1.99.5 Transferred entry. *glycerol-3-phosphate dehydrogenase*. As the acceptor is now known, the enzyme has been transferred to EC 1.1.5.3, *glycerol-3-phosphate dehydrogenase*.]

[EC 1.1.99.5 created 1961 as EC 1.1.2.1, transferred 1965 to EC 1.1.99.5, deleted 2009]

EC 1.1.99.6

Accepted name: D-lactate dehydrogenase (acceptor)
Reaction: (R)-lactate + acceptor = pyruvate + reduced acceptor
Other name(s): D-2-hydroxy acid dehydrogenase; D-2-hydroxy-acid dehydrogenase; (R)-2-hydroxy-acid:acceptor 2-oxidoreductase
Systematic name: (R)-lactate:acceptor 2-oxidoreductase
Comments: The zinc flavoprotein (FAD) from the archaeon *Archaeoglobus fulgidus* cannot utilize NAD⁺, cytochrome *c*, methylene blue or dimethylnaphthoquinone as acceptors. *In vitro* it is active with artificial electron acceptors such as 2,6-dichlorophenolindophenol, but the physiological acceptor is not yet known.
References: [3477]

[EC 1.1.99.6 created 1965, modified 2013]

EC 1.1.99.7

Accepted name: lactate—malate transhydrogenase
Reaction: (S)-lactate + oxaloacetate = pyruvate + malate
Other name(s): malate-lactate transhydrogenase
Systematic name: (S)-lactate:oxaloacetate oxidoreductase
Comments: Catalyses hydrogen transfer from C₃ or C₄ (S)-2-hydroxy acids to 2-oxo acids. It contains tightly bound nicotinamide nucleotide in its active centre. This prosthetic group cannot be removed without denaturation of the protein.
References: [69, 70]

[EC 1.1.99.7 created 1972]

[1.1.99.8 Transferred entry. *alcohol dehydrogenase (acceptor)*. Now EC 1.1.2.7, *methanol dehydrogenase (cytochrome c)* and EC 1.1.2.8, *alcohol dehydrogenase (cytochrome c)*.]

[EC 1.1.99.8 created 1972, modified 1982, deleted 2010]

EC 1.1.99.9

Accepted name: pyridoxine 5-dehydrogenase
Reaction: pyridoxine + acceptor = isopyridoxal + reduced acceptor
Other name(s): pyridoxal-5-dehydrogenase; pyridoxol 5-dehydrogenase; pyridoxin 5-dehydrogenase; pyridoxine dehydrogenase; pyridoxine 5'-dehydrogenase; pyridoxine:(acceptor) 5-oxidoreductase
Systematic name: pyridoxine:acceptor 5-oxidoreductase
Comments: A flavoprotein (FAD).
References: [4122]

[EC 1.1.99.9 created 1972, modified 1976]

[1.1.99.10 Transferred entry. *glucose dehydrogenase (acceptor)*. Now EC 1.1.5.9, *glucose 1-dehydrogenase (FAD, quinone)*]

[EC 1.1.99.10 created 1972, modified 1976, deleted 2013]

[1.1.99.11 Transferred entry. *fructose 5-dehydrogenase*, now classified as EC 1.1.5.14, *fructose 5-dehydrogenase*.]

[EC 1.1.99.11 created 1972, deleted 2021.]

EC 1.1.99.12

Accepted name: sorbose dehydrogenase
Reaction: L-sorbose + acceptor = 5-dehydro-D-fructose + reduced acceptor
Other name(s): L-sorbose:(acceptor) 5-oxidoreductase
Systematic name: L-sorbose:acceptor 5-oxidoreductase
Comments: 2,6-Dichloroindophenol can act as acceptor.
References: [3668]

[EC 1.1.99.12 created 1972]

EC 1.1.99.13

Accepted name: glucoside 3-dehydrogenase (acceptor)
Reaction: sucrose + acceptor = 3-dehydro- α -D-glucosyl- β -D-fructofuranoside + reduced acceptor
Other name(s): D-glucoside 3-dehydrogenase (ambiguous); D-aldohexopyranoside dehydrogenase (ambiguous); D-aldohexoside:(acceptor) 3-oxidoreductase; *thuA* (gene name); *thuB* (gene name); glucoside 3-dehydrogenase
Systematic name: D-aldohexoside:acceptor 3-oxidoreductase
Comments: The enzymes from members of the *Rhizobiaceae* family (such as *Agrobacterium tumefaciens*) act on disaccharides that contain a glucose moiety at the non-reducing end, such as sucrose, trehalose, leucrose, palatinose, trehalulose, and maltitol, forming the respective 3'-keto derivatives. *cf.* EC 1.1.2.11, glucoside 3-dehydrogenase (cytochrome *c*).
References: [1902, 86, 87]

[EC 1.1.99.13 created 1972, modified 2022]

EC 1.1.99.14

Accepted name: glycolate dehydrogenase
Reaction: glycolate + acceptor = glyoxylate + reduced acceptor
Other name(s): glycolate oxidoreductase; glycolic acid dehydrogenase; glycolate:(acceptor) 2-oxidoreductase
Systematic name: glycolate:acceptor 2-oxidoreductase
Comments: Also acts on (*R*)-lactate. 2,6-Dichloroindophenol and phenazine methosulfate can act as acceptors.
References: [2539]

[EC 1.1.99.14 created 1978]

[1.1.99.15 *Transferred entry. 5,10-methylenetetrahydrofolate reductase (FADH₂). Now EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]*]

[EC 1.1.99.15 created 1978, deleted 1980]

[1.1.99.16 *Transferred entry. malate dehydrogenase (acceptor). As the acceptor is now known, the enzyme has been transferred to EC 1.1.5.4, malate dehydrogenase (quinone).*]

[EC 1.1.99.16 created 1978, deleted 2009]

[1.1.99.17 *Transferred entry. glucose dehydrogenase (pyrroloquinoline-quinone). Now EC 1.1.5.2, quinoprotein glucose dehydrogenase]*

[EC 1.1.99.17 created 1982, deleted 2003]

EC 1.1.99.18

Accepted name: cellobiose dehydrogenase (acceptor)
Reaction: cellobiose + acceptor = cellobiono-1,5-lactone + reduced acceptor

Other name(s): cellobiose dehydrogenase; cellobiose oxidoreductase; *Phanerochaete chrysosporium* cellobiose oxidoreductase; CBOR; cellobiose oxidase; cellobiose:oxygen 1-oxidoreductase; CDH; cellobiose:(acceptor) 1-oxidoreductase

Systematic name: cellobiose:acceptor 1-oxidoreductase

Comments: Also acts, more slowly, on cello-oligosaccharides, lactose and D-glucosyl-1,4-β-D-mannose. The enzyme from the white rot fungus *Phanerochaete chrysosporium* is unusual in having two redoxin domains, one containing a flavin and the other a protoheme group. It transfers reducing equivalents from cellobiose to two types of redox acceptor: two-electron oxidants, including redox dyes, benzoquinones, and molecular oxygen, and one-electron oxidants, including semiquinone species, iron(II) complexes, and the model acceptor cytochrome *c* [2683]. 2,6-Dichloroindophenol can act as acceptor *in vitro*.

References: [748, 867, 868, 1463, 205, 1484, 166, 167, 2683]

[EC 1.1.99.18 created 1983, modified 2002 (EC 1.1.5.1 created 1983, incorporated 2002, EC 1.1.3.25 created 1986, incorporated 2005)]

[1.1.99.19 Transferred entry. uracil dehydrogenase. Now EC 1.17.99.4, uracil/thymine dehydrogenase]

[EC 1.1.99.19 created 1961 as EC 1.2.99.1, transferred 1984 to EC 1.1.99.19, deleted 2006]

EC 1.1.99.20

Accepted name: alkan-1-ol dehydrogenase (acceptor)

Reaction: primary alcohol + acceptor = aldehyde + reduced acceptor

Other name(s): polyethylene glycol dehydrogenase; alkan-1-ol:(acceptor) oxidoreductase

Systematic name: alkan-1-ol:acceptor oxidoreductase

Comments: A quinoprotein. Acts on C₃-C₁₆ linear-chain saturated primary alcohols, C₄-C₇ aldehydes and on non-ionic surfactants containing polyethylene glycol residues, such as Tween 40 and 60, but not on methanol and only very slowly on ethanol. 2,6-Dichloroindophenol can act as acceptor. *cf.* EC 1.1.99.8 alcohol dehydrogenase (acceptor).

References: [2043, 2044]

[EC 1.1.99.20 created 1989]

EC 1.1.99.21

Accepted name: D-sorbitol dehydrogenase (acceptor)

Reaction: D-sorbitol + acceptor = L-sorbose + reduced acceptor

Other name(s): D-sorbitol:(acceptor) 1-oxidoreductase

Systematic name: D-sorbitol:acceptor 1-oxidoreductase

Comments: A flavoprotein (FAD).

References: [3883]

[EC 1.1.99.21 created 1989]

EC 1.1.99.22

Accepted name: glycerol dehydrogenase (acceptor)

Reaction: glycerol + acceptor = glycerone + reduced acceptor

Other name(s): glycerol:(acceptor) 1-oxidoreductase

Systematic name: glycerol:acceptor 1-oxidoreductase

Comments: A quinoprotein. Also acts, more slowly, on a number of other polyols including D-sorbitol, D-arabinitol, *meso*-erythritol, ribitol and propane-1,2-diol.

References: [84]

[EC 1.1.99.22 created 1989]

[1.1.99.23 Transferred entry. polyvinyl-alcohol dehydrogenase (acceptor). Now EC 1.1.2.6, polyvinyl alcohol dehydrogenase (cytochrome)]

[EC 1.1.99.23 created 1989, deleted 2010]

EC 1.1.99.24

Accepted name: hydroxyacid-oxoacid transhydrogenase
Reaction: (*S*)-3-hydroxybutanoate + 2-oxoglutarate = acetoacetate + (*R*)-2-hydroxyglutarate
Other name(s): transhydrogenase, hydroxy acid-oxo acid
Systematic name: (*S*)-3-hydroxybutanoate:2-oxoglutarate oxidoreductase
Comments: 4-Hydroxybutanoate and (*R*)-2-hydroxyglutarate can also act as donors; 4-oxobutanoate can also act as acceptor.
References: [2034]

[EC 1.1.99.24 created 1992]

[1.1.99.25 *Transferred entry. quinate dehydrogenase (pyrroloquinoline-quinone). Now EC 1.1.5.8, quinate dehydrogenase (quinone)*]

[EC 1.1.99.25 created 1992, modified 2004, deleted 2010]

EC 1.1.99.26

Accepted name: 3-hydroxycyclohexanone dehydrogenase
Reaction: 3-hydroxycyclohexanone + acceptor = cyclohexane-1,3-dione + reduced acceptor
Systematic name: 3-hydroxycyclohexanone:acceptor 1-oxidoreductase
Comments: 2,6-Dichloroindophenol and methylene blue can act as acceptors.
References: [823]

[EC 1.1.99.26 created 1992]

EC 1.1.99.27

Accepted name: (*R*)-pantolactone dehydrogenase (flavin)
Reaction: (*R*)-pantolactone + acceptor = 2-dehydropantolactone + reduced acceptor
Other name(s): 2-dehydropantolactone reductase (flavin); 2-dehydropantoyl-lactone reductase (flavin); (*R*)-pantoyllactone dehydrogenase (flavin)
Systematic name: (*R*)-pantolactone:acceptor oxidoreductase (flavin-containing)
Comments: High specificity for (*R*)-pantolactone. Phenazine methosulfate (PMS) can act as acceptor. The enzyme has been studied in the bacterium *Nocardia asteroides* and shown to be membrane-bound and induced by 1,2-propanediol. The FMN cofactor is non-covalently bound.
References: [2011]

[EC 1.1.99.27 created 1999]

EC 1.1.99.28

Accepted name: glucose-fructose oxidoreductase
Reaction: D-glucose + D-fructose = D-gluconolactone + D-glucitol
Systematic name: D-glucose:D-fructose oxidoreductase
Comments: D-mannose, D-xylose, D-galactose, 2-deoxy-D-glucose and L-arabinose will function as aldose substrates, but with low affinities. The ketose substrate must be in the open-chain form. The apparent affinity for fructose is low, because little of the fructose substrate is in the open-chain form. Xylulose and glycerone (dihydroxyacetone) will replace fructose, but they are poor substrates. The enzyme from *Zymomonas mobilis* contains tightly bound NADP⁺.
References: [4852, 1531, 1988]

[EC 1.1.99.28 created 1999]

EC 1.1.99.29

- Accepted name:** pyranose dehydrogenase (acceptor)
Reaction: (1) a pyranose + acceptor = a pyranos-2-ulose (or a pyranos-3-ulose or a pyranos-2,3-diulose) + reduced acceptor
(2) a pyranoside + acceptor = a pyranosid-3-ulose (or a pyranosid-3,4-diulose) + reduced acceptor
Other name(s): pyranose dehydrogenase; pyranose-quinone oxidoreductase; quinone-dependent pyranose dehydrogenase; PDH
Systematic name: pyranose:acceptor oxidoreductase
Comments: Requires FAD. A number of aldoses and ketoses in pyranose form, as well as glycosides, gluco-oligosaccharides, sucrose and lactose can act as a donor. 1,4-Benzoquinone or ferricenium ion (ferrocene oxidized by removal of one electron) can serve as acceptor. Unlike EC 1.1.3.10, pyranose oxidase, this fungal enzyme does not interact with O₂ and exhibits extremely broad substrate tolerance with variable regioselectivity (C-3, C-2 or C-3 + C-2 or C-3 + C-4) for (di)oxidation of different sugars. D-Glucose is exclusively or preferentially oxidized at C-3 (depending on the enzyme source), but can also be oxidized at C-2 + C-3. The enzyme also acts on 1→4- α - and 1→4- β -gluco-oligosaccharides, non-reducing gluco-oligosaccharides and L-arabinose, which are not substrates of EC 1.1.3.10. Sugars are oxidized in their pyranose but not in their furanose form.
References: [4462, 4464, 4465, 4461, 4463]

[EC 1.1.99.29 created 2004]

EC 1.1.99.30

- Accepted name:** 2-oxo-acid reductase
Reaction: a (2*R*)-hydroxy-carboxylate + acceptor = a 2-oxocarboxylate + reduced acceptor
Other name(s): (2*R*)-hydroxycarboxylate-viologen-oxidoreductase; HVOR; 2-oxoacid reductase
Systematic name: (2*R*)-hydroxy-carboxylate:acceptor oxidoreductase
Comments: Contains [4Fe-4S] and a mononucleotide molybdenum (pyranopterin) cofactor. Has broad substrate specificity, with 2-oxo-monocarboxylates and 2-oxo-dicarboxylates acting as substrates. Branching in a substrate at the C-3 position results in loss of activity. The enzyme from *Proteus* sp. is inactivated by oxygen.
References: [4324, 3050]

[EC 1.1.99.30 created 2004]

EC 1.1.99.31

- Accepted name:** (*S*)-mandelate dehydrogenase
Reaction: (*S*)-mandelate + acceptor = phenylglyoxylate + reduced acceptor
Other name(s): MDH (ambiguous)
Systematic name: (*S*)-mandelate:acceptor 2-oxidoreductase
Comments: This enzyme is a member of the FMN-dependent α -hydroxy-acid oxidase/dehydrogenase family [2408]. While all enzymes of this family oxidize the (*S*)-enantiomer of an α -hydroxy acid to an α -oxo acid, the ultimate oxidant (oxygen, intramolecular heme or some other acceptor) depends on the particular enzyme. This enzyme transfers the electron pair from FMNH₂ to a component of the electron transport chain, most probably ubiquinone [2408, 895]. It is part of a metabolic pathway in Pseudomonads that allows these organisms to utilize mandelic acid, derivatized from the common soil metabolite amygdalin, as the sole source of carbon and energy [895]. The enzyme has a large active-site pocket and preferentially binds substrates with longer sidechains, e.g. 2-hydroxyoctanoate rather than 2-hydroxybutyrate [2408]. It also prefers substrates that, like (*S*)-mandelate, have β unsaturation, e.g. (indol-3-yl)glycolate compared with (indol-3-yl)lactate [2408]. Esters of mandelate, such as methyl (*S*)-mandelate, are also substrates [894].
References: [2408, 895, 894]

[EC 1.1.99.31 created 2006]

EC 1.1.99.32

Accepted name: L-sorbose 1-dehydrogenase
Reaction: L-sorbose + acceptor = 1-dehydro-L-sorbose + reduced acceptor
Other name(s): SDH (ambiguous)
Systematic name: L-sorbose:acceptor 1-oxidoreductase
Comments: The product, L-sorbosone, is an intermediate in bacterial 2-keto-L-gulonic-acid formation. The activity of this membrane-bound enzyme is stimulated by Fe(III) or Co²⁺ but is inhibited by Cu²⁺. The enzyme is highly specific for L-sorbose as other sugars, such as glucose, mannitol and sorbitol, are not substrates. Phenazine methosulfate and DCIP can act as artificial acceptors.
References: [4100]

[EC 1.1.99.32 created 2008]

[1.1.99.33 *Transferred entry. formate dehydrogenase (acceptor). Now EC 1.17.99.7, formate dehydrogenase (acceptor)*]

[EC 1.1.99.33 created 2010, deleted 2017]

[1.1.99.34 *Transferred entry. glucose-6-phosphate dehydrogenase (coenzyme-F₄₂₀). As the acceptor is now known, the enzyme has been transferred to EC 1.1.98.2, glucose-6-phosphate dehydrogenase (coenzyme-F₄₂₀)*]

[EC 1.1.99.34 created 2010, deleted 2011]

EC 1.1.99.35

Accepted name: soluble quinoprotein glucose dehydrogenase
Reaction: D-glucose + acceptor = D-glucono-1,5-lactone + reduced acceptor
Other name(s): soluble glucose dehydrogenase; sGDH; glucose dehydrogenase (PQQ-dependent)
Systematic name: D-glucose:acceptor oxidoreductase
Comments: Soluble periplasmic enzyme containing PQQ as prosthetic group, bound to a calcium ion. Electron acceptor is not known. It is assayed with Wurster's Blue or phenazine methosulfate. It has negligible sequence or structure similarity to other quinoproteins. It catalyses an exceptionally high rate of oxidation of a wide range of aldose sugars, including D-glucose, galactose, arabinose and xylose, and also the disaccharides lactose, cellobiose and maltose. It has been described only in *Acinetobacter calcoaceticus*.
References: [1296, 938, 701, 2714, 3210, 2718]

[EC 1.1.99.35 created 2010]

EC 1.1.99.36

Accepted name: alcohol dehydrogenase (nicotinoprotein)
Reaction: ethanol + acceptor = acetaldehyde + reduced acceptor
Other name(s): NDMA-dependent alcohol dehydrogenase; nicotinoprotein alcohol dehydrogenase; np-ADH; ethanol:*N,N*-dimethyl-4-nitrosoaniline oxidoreductase
Systematic name: ethanol:acceptor oxidoreductase
Comments: Contains Zn²⁺. Nicotinoprotein alcohol dehydrogenases are unique medium-chain dehydrogenases/reductases (MDR) alcohol dehydrogenases that have a tightly bound NAD⁺/NADH cofactor that does not dissociate during the catalytic process. Instead, the cofactor is regenerated by a second substrate or electron carrier. While the *in vivo* electron acceptor is not known, *N,N*-dimethyl-4-nitrosoaniline (NDMA), which is reduced to 4-(hydroxylamino)-*N,N*-dimethylaniline, can serve this function *in vitro*. The enzyme from the Gram-positive bacterium *Amycolatopsis methanolica* can accept many primary alcohols as substrates, including benzylalcohol [3188].
References: [3188, 3322, 3710, 3321, 3108]

[EC 1.1.99.36 created 2010]

EC 1.1.99.37

- Accepted name:** methanol dehydrogenase (nicotinoprotein)
Reaction: methanol + acceptor = formaldehyde + reduced acceptor
Other name(s): NDMA-dependent methanol dehydrogenase; nicotinoprotein methanol dehydrogenase; methanol:*N,N*-dimethyl-4-nitrosoaniline oxidoreductase
Systematic name: methanol:acceptor oxidoreductase
Comments: Contains Zn²⁺ and Mg²⁺. Nicotinoprotein methanol dehydrogenases have a tightly bound NADP⁺/NADPH cofactor that does not dissociate during the catalytic process. Instead, the cofactor is regenerated by a second substrate or electron carrier. While the *in vivo* electron acceptor is not known, *N,N*-dimethyl-4-nitrosoaniline (NDMA), which is reduced to 4-(hydroxylamino)-*N,N*-dimethylaniline, can serve this function *in vitro*. The enzyme has been detected in several Gram-positive methylotrophic bacteria, including *Amycolatopsis methanolica*, *Rhodococcus rhodochrous* and *Rhodococcus erythropolis* [4472, 3188, 514]. These enzymes are decameric, and possess a 5-fold symmetry [1616]. Some of the enzymes can also dismutate formaldehyde to methanol and formate [3239].
References: [4472, 3188, 514, 1616, 3239]

[EC 1.1.99.37 created 2010]

EC 1.1.99.38

- Accepted name:** 2-deoxy-*scyllo*-inosamine dehydrogenase (AdoMet-dependent)
Reaction: 2-deoxy-*scyllo*-inosamine + *S*-adenosyl-L-methionine = 3-amino-2,3-dideoxy-*scyllo*-inosose + 5'-deoxyadenosine + L-methionine
Other name(s): *btrN* (gene name); 2-deoxy-*scyllo*-inosamine dehydrogenase (SAM-dependent)
Systematic name: 2-deoxy-*scyllo*-inosamine:*S*-adenosyl-L-methionine 1-oxidoreductase
Comments: Involved in the biosynthetic pathway of the aminoglycoside antibiotics of the butirosin family. The enzyme from *Bacillus circulans* was shown to be a radical *S*-adenosyl-L-methionine (SAM) enzyme. *cf.* EC 1.1.1.329, 2-deoxy-*scyllo*-inosamine dehydrogenase.
References: [4795, 4796]

[EC 1.1.99.38 created 2012, modified 2013]

EC 1.1.99.39

- Accepted name:** D-2-hydroxyglutarate dehydrogenase
Reaction: (*R*)-2-hydroxyglutarate + acceptor = 2-oxoglutarate + reduced acceptor
Other name(s): D2HGDH (gene name)
Systematic name: (*R*)-2-hydroxyglutarate:acceptor 2-oxidoreductase
Comments: Contains FAD. The enzyme has no activity with NAD⁺ or NADP⁺, and was assayed *in vitro* using artificial electron acceptors. It has lower activity with (*R*)-lactate, (*R*)-2-hydroxybutyrate and *meso*-tartrate, and no activity with the (*S*) isomers. The mammalian enzyme is stimulated by Zn²⁺, Co²⁺ and Mn²⁺.
References: [1050, 8]

[EC 1.1.99.39 created 2013]

EC 1.1.99.40

- Accepted name:** (*R*)-2-hydroxyglutarate—pyruvate transhydrogenase
Reaction: (*R*)-2-hydroxyglutarate + pyruvate = 2-oxoglutarate + (*R*)-lactate
Other name(s): DLD3 (gene name)
Systematic name: (*R*)-2-hydroxyglutarate:pyruvate oxidoreductase [(*R*)-lactate-forming]
Comments: The enzyme, characterized in the yeast *Saccharomyces cerevisiae*, also functions as EC 1.1.2.4, D-lactate dehydrogenase (cytochrome), and is active with oxaloacetate as electron acceptor forming (*R*)-malate.

References: [263]

[EC 1.1.99.40 created 2017]

EC 1.1.99.41

Accepted name: 3-hydroxy-1,2-didehydro-2,3-dihydrotabersonine reductase
Reaction: (1) (3*R*)-3-hydroxy-16-methoxy-2,3-dihydrotabersonine + acceptor = (3*R*)-3-hydroxy-16-methoxy-1,2-didehydro-2,3-dihydrotabersonine + reduced acceptor
(2) (3*R*)-3-hydroxy-2,3-dihydrotabersonine + acceptor = (3*R*)-3-hydroxy-1,2-didehydro-2,3-dihydrotabersonine + reduced acceptor
Other name(s): T3R; tabersonine 3-reductase
Systematic name: (3*R*)-3-hydroxy-16-methoxy-2,3-dihydrotabersonine:acceptor oxidoreductase
Comments: This enzyme is involved in the biosynthesis of vindoline and vindorosine in the plant *Catharanthus roseus* (Madagascar periwinkle). *In vivo*, it functions in the direction of reduction. It has no activity with 3-epoxylated compounds, which can form spontaneously from its unstable substrates.
References: [3407]

[EC 1.1.99.41 created 2017]

EC 1.1.99.42

Accepted name: 4-pyridoxic acid dehydrogenase
Reaction: 4-pyridoxate + acceptor = 5-formyl-3-hydroxy-2-methylpyridine-4-carboxylate + reduced acceptor
Other name(s): mlr6792 (locus name)
Systematic name: 4-pyridoxate:acceptor 5-oxidoreductase
Comments: The enzyme, characterized from the bacteria *Pseudomonas* sp. MA-1 and *Mesorhizobium loti*, participates in the degradation of pyridoxine (vitamin B₆). It is membrane bound and contains FAD. The enzyme has been assayed *in vitro* in the presence of the artificial electron acceptor dichloroindophenol (DCPIP).
References: [4721, 1291]

[EC 1.1.99.42 created 2018]

EC 1.2 Acting on the aldehyde or oxo group of donors

This subclass contains enzymes that oxidize aldehydes to the corresponding acids; when this acid is concomitantly phosphorylated or acetylates CoA, this is indicated in parentheses. Oxo groups may be oxidized either with addition of water and cleavage of a carbon-carbon bond or, in the case of ring compounds, by addition of the elements of water and dehydrogenation. Sub-classes are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.2.1), a cytochrome (EC 1.2.2), oxygen (EC 1.2.3), a disulfide (EC 1.2.4), an iron-sulfur protein (EC 1.2.7), or some other acceptor (EC 1.2.99).

EC 1.2.1 With NAD⁺ or NADP⁺ as acceptor

[1.2.1.1 Deleted entry. *glutathione-dependent formaldehyde dehydrogenase*. This enzyme was classified on the basis of an incorrect reaction. It has been replaced by two enzymes, EC 1.1.1.284, *S*-(hydroxymethyl)glutathione dehydrogenase and EC 4.4.1.22, *S*-(hydroxymethyl)glutathione synthase]

[EC 1.2.1.1 created 1961, modified 1982, modified 2002, deleted 2005]

[1.2.1.2 Transferred entry. *formate dehydrogenase*. Now EC 1.17.1.9, *formate dehydrogenase*]

[EC 1.2.1.2 created 1961, deleted 2017]

EC 1.2.1.3

Accepted name: aldehyde dehydrogenase (NAD⁺)
Reaction: an aldehyde + NAD⁺ + H₂O = a carboxylate + NADH + H⁺
Other name(s): CoA-independent aldehyde dehydrogenase; *m*-methylbenzaldehyde dehydrogenase; NAD-aldehyde dehydrogenase; NAD-dependent 4-hydroxynonenal dehydrogenase; NAD-dependent aldehyde dehydrogenase; NAD-linked aldehyde dehydrogenase; propionaldehyde dehydrogenase; aldehyde dehydrogenase (NAD)
Systematic name: aldehyde:NAD⁺ oxidoreductase
Comments: Wide specificity, including oxidation of D-glucuronolactone to D-glucarate.
References: [1880, 3419]

[EC 1.2.1.3 created 1961 (EC 1.1.1.70 created 1965, incorporated 1978)]

EC 1.2.1.4

Accepted name: aldehyde dehydrogenase (NADP⁺)
Reaction: an aldehyde + NADP⁺ + H₂O = a carboxylate + NADPH + H⁺
Other name(s): NADP-acetaldehyde dehydrogenase; NADP-dependent aldehyde dehydrogenase; aldehyde dehydrogenase (NADP)
Systematic name: aldehyde:NADP⁺ oxidoreductase
References: [15, 1880, 2999, 3781]

[EC 1.2.1.4 created 1961]

EC 1.2.1.5

Accepted name: aldehyde dehydrogenase [NAD(P)⁺]
Reaction: an aldehyde + NAD(P)⁺ + H₂O = a carboxylate + NAD(P)H + H⁺
Other name(s): ALDH
Systematic name: aldehyde:NAD(P)⁺ oxidoreductase
References: [343, 1880, 2121, 4020, 4198]

[EC 1.2.1.5 created 1961]

[1.2.1.6 Deleted entry. benzaldehyde dehydrogenase]

[EC 1.2.1.6 created 1961, deleted 1965]

EC 1.2.1.7

Accepted name: benzaldehyde dehydrogenase (NADP⁺)
Reaction: benzaldehyde + NADP⁺ + H₂O = benzoate + NADPH + 2 H⁺
Other name(s): NADP-linked benzaldehyde dehydrogenase; benzaldehyde dehydrogenase (NADP)
Systematic name: benzaldehyde:NADP⁺ oxidoreductase
References: [1443, 3994]

[EC 1.2.1.7 created 1961]

EC 1.2.1.8

Accepted name: betaine-aldehyde dehydrogenase
Reaction: betaine aldehyde + NAD⁺ + H₂O = betaine + NADH + 2 H⁺
Other name(s): betaine aldehyde oxidase; BADH; betaine aldehyde dehydrogenase; BetB
Systematic name: betaine-aldehyde:NAD⁺ oxidoreductase

Comments: In many bacteria, plants and animals, the osmoprotectant betaine is synthesized in two steps: (1) choline to betaine aldehyde and (2) betaine aldehyde to betaine. This enzyme is involved in the second step and appears to be the same in plants, animals and bacteria. In contrast, different enzymes are involved in the first reaction. In plants, this reaction is catalysed by EC 1.14.15.7 (choline monooxygenase), whereas in animals and many bacteria it is catalysed by either membrane-bound EC 1.1.99.1 (choline dehydrogenase) or soluble EC 1.1.3.17 (choline oxidase) [4485]. In some bacteria, betaine is synthesized from glycine through the actions of EC 2.1.1.156 (glycine/sarcosine *N*-methyltransferase) and EC 2.1.1.157 (sarcosine/dimethylglycine *N*-methyltransferase).

References: [3580, 2527, 3113, 1923, 4485]

[EC 1.2.1.8 created 1961, modified 2005, modified 2011]

EC 1.2.1.9

Accepted name: glyceraldehyde-3-phosphate dehydrogenase (NADP⁺)

Reaction: D-glyceraldehyde 3-phosphate + NADP⁺ + H₂O = 3-phospho-D-glycerate + NADPH + 2 H⁺

Other name(s): triosephosphate dehydrogenase (ambiguous); glyceraldehyde phosphate dehydrogenase (nicotinamide adenine dinucleotide phosphate); glyceraldehyde phosphate dehydrogenase (NADP⁺); glyceraldehyde 3-phosphate dehydrogenase (NADP⁺); NADP⁺-glyceraldehyde phosphate dehydrogenase; NADP⁺-glyceraldehyde-3-phosphate dehydrogenase; glyceraldehyde-3-phosphate:NADP⁺ reductase; nonphosphorylating glyceraldehyde-3-phosphate dehydrogenase

Systematic name: D-glyceraldehyde-3-phosphate:NADP⁺ oxidoreductase

References: [3575]

[EC 1.2.1.9 created 1961]

EC 1.2.1.10

Accepted name: acetaldehyde dehydrogenase (acetylating)

Reaction: acetaldehyde + CoA + NAD⁺ = acetyl-CoA + NADH + H⁺

Other name(s): aldehyde dehydrogenase (acylating); ADA; acylating acetaldehyde dehydrogenase; DmpF; BphJ

Systematic name: acetaldehyde:NAD⁺ oxidoreductase (CoA-acetylating)

Comments: Also acts, more slowly, on glycolaldehyde, propanal and butanal. In several bacterial species this enzyme forms a bifunctional complex with EC 4.1.3.39, 4-hydroxy-2-oxovalerate aldolase. The enzymes from the bacteria *Burkholderia xenovorans* and *Thermus thermophilus* also perform the reaction of EC 1.2.1.87, propanal dehydrogenase (propanoylating). Involved in the *meta*-cleavage pathway for the degradation of phenols, methylphenols and catechols. NADP⁺ can replace NAD⁺ but the rate of reaction is much slower [3364].

References: [507, 3936, 3364, 196, 195]

[EC 1.2.1.10 created 1961, modified 2006, modified 2011]

EC 1.2.1.11

Accepted name: aspartate-semialdehyde dehydrogenase

Reaction: L-aspartate 4-semialdehyde + phosphate + NADP⁺ = L-4-aspartyl phosphate + NADPH + H⁺

Other name(s): aspartate semialdehyde dehydrogenase; aspartic semialdehyde dehydrogenase; L-aspartate-β-semialdehyde:NADP⁺ oxidoreductase (phosphorylating); aspartic β-semialdehyde dehydrogenase; ASA dehydrogenase

Systematic name: L-aspartate-4-semialdehyde:NADP⁺ oxidoreductase (phosphorylating)

References: [345, 1880]

[EC 1.2.1.11 created 1961]

EC 1.2.1.12

Accepted name: glyceraldehyde-3-phosphate dehydrogenase (phosphorylating)

Reaction: D-glyceraldehyde 3-phosphate + phosphate + NAD⁺ = 3-phospho-D-glyceroyl phosphate + NADH + H⁺

Other name(s): triosephosphate dehydrogenase (ambiguous); glyceraldehyde phosphate dehydrogenase; phosphoglyceraldehyde dehydrogenase; 3-phosphoglyceraldehyde dehydrogenase; NAD⁺-dependent glyceraldehyde phosphate dehydrogenase; glyceraldehyde phosphate dehydrogenase (NAD⁺); glyceraldehyde-3-phosphate dehydrogenase (NAD⁺); NADH-glyceraldehyde phosphate dehydrogenase; glyceraldehyde-3-*P*-dehydrogenase

Systematic name: D-glyceraldehyde-3-phosphate:NAD⁺ oxidoreductase (phosphorylating)

Comments: Also acts very slowly on D-glyceraldehyde and some other aldehydes; thiols can replace phosphate.

References: [548, 736, 1472, 4429, 4541, 3615]

[EC 1.2.1.12 created 1961]

EC 1.2.1.13

Accepted name: glyceraldehyde-3-phosphate dehydrogenase (NADP⁺) (phosphorylating)

Reaction: D-glyceraldehyde 3-phosphate + phosphate + NADP⁺ = 3-phospho-D-glyceroyl phosphate + NADPH + H⁺

Other name(s): triosephosphate dehydrogenase (NADP⁺); dehydrogenase, glyceraldehyde phosphate (nicotinamide adenine dinucleotide phosphate) (phosphorylating); glyceraldehyde phosphate dehydrogenase (nicotinamide adenine dinucleotide phosphate) (phosphorylating); NADP⁺-glyceraldehyde-3-phosphate dehydrogenase; NADP⁺-glyceraldehyde phosphate dehydrogenase; NADP⁺-dependent glyceraldehyde phosphate dehydrogenase; NADP⁺-triose phosphate dehydrogenase; glyceraldehyde-3-phosphate dehydrogenase (NADP⁺) (phosphorylating); GAPDH

Systematic name: D-glyceraldehyde-3-phosphate:NADP⁺ oxidoreductase (phosphorylating)

References: [438, 1318, 3575]

[EC 1.2.1.13 created 1961]

[1.2.1.14 *Transferred entry. IMP dehydrogenase. Now EC 1.1.1.205, IMP dehydrogenase*]

[EC 1.2.1.14 created 1961, deleted 1984]

EC 1.2.1.15

Accepted name: malonate-semialdehyde dehydrogenase

Reaction: 3-oxopropanoate + NAD(P)⁺ + H₂O = malonate + NAD(P)H + 2 H⁺

Systematic name: 3-oxopropanoate:NAD(P)⁺ oxidoreductase

References: [2978]

[EC 1.2.1.15 created 1965]

EC 1.2.1.16

Accepted name: succinate-semialdehyde dehydrogenase [NAD(P)⁺]

Reaction: succinate semialdehyde + NAD(P)⁺ + H₂O = succinate + NAD(P)H + 2 H⁺

Other name(s): succinate semialdehyde dehydrogenase (nicotinamide adenine dinucleotide (phosphate)); succinate-semialdehyde dehydrogenase [NAD(P)]

Systematic name: succinate-semialdehyde:NAD(P)⁺ oxidoreductase

References: [1880, 1883, 3072]

[EC 1.2.1.16 created 1965]

EC 1.2.1.17

Accepted name: glyoxylate dehydrogenase (acylating)

Reaction: glyoxylate + CoA + NADP⁺ = oxalyl-CoA + NADPH + H⁺

Systematic name: glyoxylate:NADP⁺ oxidoreductase (CoA-oxalylating)
References: [3413]

[EC 1.2.1.17 created 1965]

EC 1.2.1.18

Accepted name: malonate-semialdehyde dehydrogenase (acetylating)
Reaction: 3-oxopropoate + CoA + NAD(P)⁺ = acetyl-CoA + CO₂ + NAD(P)H
Other name(s): malonic semialdehyde oxidative decarboxylase
Systematic name: 3-oxopropoate:NAD(P)⁺ oxidoreductase (decarboxylating, CoA-acetylating)
References: [1573, 1880, 4722]

[EC 1.2.1.18 created 1965]

EC 1.2.1.19

Accepted name: aminobutyraldehyde dehydrogenase
Reaction: 4-aminobutanal + NAD⁺ + H₂O = 4-aminobutanoate + NADH + 2 H⁺
Other name(s): γ-guanidinobutyraldehyde dehydrogenase (ambiguous); ABAL dehydrogenase; 4-aminobutyraldehyde dehydrogenase; 4-aminobutanal dehydrogenase; γ-aminobutyraldehyde dehydrogenase; 1-pyrroline dehydrogenase; ABALDH; YdcW
Systematic name: 4-aminobutanal:NAD⁺ 1-oxidoreductase
Comments: The enzyme from some species exhibits broad substrate specificity and has a marked preference for straight-chain aldehydes (up to 7 carbon atoms) as substrates [1433]. The plant enzyme also acts on 4-guanidinobutanal (*cf.* EC 1.2.1.54 γ-guanidinobutyraldehyde dehydrogenase). As 1-pyrroline and 4-aminobutanal are in equilibrium and can be interconverted spontaneously, 1-pyrroline may act as the starting substrate. The enzyme forms part of the arginine-catabolism pathway [3650] and belongs in the aldehyde dehydrogenase superfamily [1433].
References: [529, 1880, 1881, 2698, 4803, 3383, 3382, 3650, 1433]

[EC 1.2.1.19 created 1965, modified 1989 (EC 1.5.1.35 created 2006, incorporated 2007)]

EC 1.2.1.20

Accepted name: glutarate-semialdehyde dehydrogenase
Reaction: 5-oxopentanoate + NADP⁺ + H₂O = glutarate + NADPH + H⁺
Other name(s): glutarate semialdehyde dehydrogenase; *davD* (gene name)
Systematic name: glutarate-semialdehyde:NADP⁺ oxidoreductase
Comments: The enzyme, characterized from multiple *Pseudomonas* strains, participates in L-lysine degradation. Unlike earlier claims, it prefers NADP⁺ to NAD⁺.
References: [1787, 602, 1152, 603, 4749]

[EC 1.2.1.20 created 1965, modified 2021]

EC 1.2.1.21

Accepted name: glycolaldehyde dehydrogenase
Reaction: glycolaldehyde + NAD⁺ + H₂O = glycolate + NADH + 2 H⁺
Other name(s): glycol aldehyde dehydrogenase
Systematic name: glycolaldehyde:NAD⁺ oxidoreductase
References: [840]

[EC 1.2.1.21 created 1972]

EC 1.2.1.22

- Accepted name:** lactaldehyde dehydrogenase
Reaction: (S)-lactaldehyde + NAD⁺ + H₂O = (S)-lactate + NADH + 2 H⁺
Other name(s): L-lactaldehyde:NAD oxidoreductase; nicotinamide adenine dinucleotide (NAD)-linked dehydrogenase
Systematic name: (S)-lactaldehyde:NAD⁺ oxidoreductase
References: [3496, 3992]

[EC 1.2.1.22 created 1972]

EC 1.2.1.23

- Accepted name:** 2-oxoaldehyde dehydrogenase (NAD⁺)
Reaction: a 2-oxoaldehyde + NAD⁺ + H₂O = a 2-oxo carboxylate + NADH + H⁺
Other name(s): α-ketoaldehyde dehydrogenase; methylglyoxal dehydrogenase; NAD⁺-linked α-ketoaldehyde dehydrogenase; 2-ketoaldehyde dehydrogenase; NAD⁺-dependent α-ketoaldehyde dehydrogenase
Systematic name: 2-oxoaldehyde:NAD⁺ 2-oxidoreductase
Comments: Not identical with EC 1.2.1.49 2-oxoaldehyde dehydrogenase (NADP⁺).
References: [2866, 3465, 3467]

[EC 1.2.1.23 created 1972, modified 1986]

EC 1.2.1.24

- Accepted name:** succinate-semialdehyde dehydrogenase (NAD⁺)
Reaction: succinate semialdehyde + NAD⁺ + H₂O = succinate + NADH + 2 H⁺
Other name(s): succinate semialdehyde dehydrogenase (NAD⁺); succinic semialdehyde dehydrogenase (NAD⁺); succinyl semialdehyde dehydrogenase (NAD⁺); succinate semialdehyde:NAD⁺ oxidoreductase
Systematic name: succinate-semialdehyde:NAD⁺ oxidoreductase
Comments: This enzyme participates in the degradation of glutamate and 4-aminobutyrate. It is similar to EC 1.2.1.79 [succinate-semialdehyde dehydrogenase (NADP⁺)], and EC 1.2.1.16 [succinate-semialdehyde dehydrogenase (NAD(P)⁺)], but is specific for NAD⁺.
References: [58, 3616, 508]

[EC 1.2.1.24 created 1972, modified 2010]

EC 1.2.1.25

- Accepted name:** branched-chain α-keto acid dehydrogenase system
Reaction: 3-methyl-2-oxobutanoate + CoA + NAD⁺ = 2-methylpropanoyl-CoA + CO₂ + NADH
Other name(s): branched-chain α-keto acid dehydrogenase complex; 2-oxoisovalerate dehydrogenase; α-ketoisovalerate dehydrogenase; 2-oxoisovalerate dehydrogenase (acylating)
Systematic name: 3-methyl-2-oxobutanoate:NAD⁺ 2-oxidoreductase (CoA-methylpropanoylating)
Comments: This enzyme system catalyses the oxidative decarboxylation of branched-chain α-keto acids derived from L-leucine, L-isoleucine, and L-valine to branched-chain acyl-CoAs. It belongs to the 2-oxoacid dehydrogenase system family, which also includes EC 1.2.1.104, pyruvate dehydrogenase system, EC 1.2.1.105, 2-oxoglutarate dehydrogenase system, EC 1.4.1.27, glycine cleavage system, and EC 2.3.1.190, acetoin dehydrogenase system. With the exception of the glycine cleavage system, which contains 4 components, the 2-oxoacid dehydrogenase systems share a common structure, consisting of three main components, namely a 2-oxoacid dehydrogenase (E1), a dihydrolipoamide acyltransferase (E2), and dihydrolipoamide dehydrogenase (E3). The reaction catalysed by this system is the sum of three activities: EC 1.2.4.4, 3-methyl-2-oxobutanoate dehydrogenase (2-methylpropanoyl-transferring), EC 2.3.1.168, dihydrolipoyllysine-residue (2-methylpropanoyl)transferase, and EC 1.8.1.4, dihydrolipoyl dehydrogenase. The system also acts on (S)-3-methyl-2-oxopentanoate and 4-methyl-2-oxopentanoate.
References: [3005, 3309, 1538, 1071, 3478]

[EC 1.2.1.25 created 1972, modified 2019, modified 2020]

EC 1.2.1.26

Accepted name: 2,5-dioxovalerate dehydrogenase
Reaction: $2,5\text{-dioxopentanoate} + \text{NADP}^+ + \text{H}_2\text{O} = 2\text{-oxoglutarate} + \text{NADPH} + 2 \text{H}^+$
Other name(s): 2-oxoglutarate semialdehyde dehydrogenase; α -ketoglutaric semialdehyde dehydrogenase
Systematic name: 2,5-dioxopentanoate:NADP⁺ 5-oxidoreductase
References: [25]

[EC 1.2.1.26 created 1972]

EC 1.2.1.27

Accepted name: methylmalonate-semialdehyde dehydrogenase (CoA-acylating)
Reaction: $2\text{-methyl-3-oxopropanoate} + \text{CoA} + \text{H}_2\text{O} + \text{NAD}^+ = \text{propanoyl-CoA} + \text{HCO}_3^- + \text{NADH}$
Other name(s): MSDH; MMSA dehydrogenase; *iolA* (gene name); methylmalonate-semialdehyde dehydrogenase (acylating)
Systematic name: 2-methyl-3-oxopropanoate:NAD⁺ 3-oxidoreductase (CoA-propanoylating)
Comments: Also converts 3-oxopropanoate into acetyl-CoA [4034]. The reaction occurs in two steps with the decarboxylation process preceding CoA-binding [4034]. Bicarbonate rather than CO₂ is released as a final product [4034].
References: [3954, 977, 4034]

[EC 1.2.1.27 created 1972, modified 2014]

EC 1.2.1.28

Accepted name: benzaldehyde dehydrogenase (NAD⁺)
Reaction: $\text{benzaldehyde} + \text{NAD}^+ + \text{H}_2\text{O} = \text{benzoate} + \text{NADH} + 2 \text{H}^+$
Other name(s): benzaldehyde (NAD) dehydrogenase; benzaldehyde dehydrogenase (NAD)
Systematic name: benzaldehyde:NAD⁺ oxidoreductase
References: [1443]

[EC 1.2.1.28 created 1972]

EC 1.2.1.29

Accepted name: aryl-aldehyde dehydrogenase
Reaction: $\text{an aromatic aldehyde} + \text{NAD}^+ + \text{H}_2\text{O} = \text{an aromatic acid} + \text{NADH} + \text{H}^+$
Systematic name: aryl-aldehyde:NAD⁺ oxidoreductase
Comments: Oxidizes a number of aromatic aldehydes, but not aliphatic aldehydes.
References: [3434]

[EC 1.2.1.29 created 1972]

EC 1.2.1.30

Accepted name: carboxylate reductase (NADP⁺)
Reaction: $\text{an aromatic aldehyde} + \text{NADP}^+ + \text{AMP} + \text{diphosphate} = \text{an aromatic acid} + \text{NADPH} + \text{H}^+ + \text{ATP}$
Other name(s): aromatic acid reductase; aryl-aldehyde dehydrogenase (NADP⁺)
Systematic name: aryl-aldehyde:NADP⁺ oxidoreductase (ATP-forming)
Comments: The enzyme contains an adenylation domain, a phosphopantetheinyl binding domain, and a reductase domain, and requires activation by attachment of a phosphopantetheinyl group. The enzyme activates its substrate to an adenylate form, followed by a transfer to the phosphopantetheinyl binding domain. The resulting thioester is subsequently transferred to the reductase domain, where it is reduced to an aldehyde and released.

References: [1425, 1423, 4434, 4046]

[EC 1.2.1.30 created 1972, modified 2019]

EC 1.2.1.31

Accepted name: L-aminoadipate-semialdehyde dehydrogenase
Reaction: (S)-2-amino-6-oxohexanoate + NAD(P)⁺ + H₂O = L-2-aminoadipate + NAD(P)H + H⁺ (overall reaction)
(1a) (S)-2-amino-6-oxohexanoate = (S)-2,3,4,5-tetrahydropyridine-2-carboxylate + H₂O (spontaneous)
(1b) (S)-2,3,4,5-tetrahydropyridine-2-carboxylate + NAD(P)⁺ + 2 H₂O = L-2-aminoadipate + NAD(P)H + H⁺
Other name(s): aminoadipate semialdehyde dehydrogenase; 2-aminoadipate semialdehyde dehydrogenase; α-aminoadipate-semialdehyde dehydrogenase; α-aminoadipate reductase; 2-aminoadipic semialdehyde dehydrogenase; L-α-aminoadipate δ-semialdehyde oxidoreductase; L-α-aminoadipate δ-semialdehyde:NAD⁺ oxidoreductase; L-α-aminoadipate δ-semialdehyde:nicotinamide adenine dinucleotide oxidoreductase; L-2-aminoadipate 6-semialdehyde:NAD(P)⁺ 6-oxidoreductase
Systematic name: (S)-2-amino-6-oxohexanoate:NAD(P)⁺ 6-oxidoreductase
Comments: (S)-2-amino-6-oxohexanoate undergoes a spontaneous dehydration forming the cyclic (S)-2,3,4,5-tetrahydropyridine-2-carboxylate, which serves as a substrate for the hydrogenation reaction.
References: [530, 3550, 849, 1204]

[EC 1.2.1.31 created 1972, modified 2011]

EC 1.2.1.32

Accepted name: aminomuconate-semialdehyde dehydrogenase
Reaction: 2-aminomuconate 6-semialdehyde + NAD⁺ + H₂O = 2-aminomuconate + NADH + 2 H⁺
Other name(s): 2-aminomuconate semialdehyde dehydrogenase; 2-hydroxymuconic acid semialdehyde dehydrogenase; 2-hydroxymuconate semialdehyde dehydrogenase; α-aminomuconic ε-semialdehyde dehydrogenase; α-hydroxymuconic ε-semialdehyde dehydrogenase; 2-hydroxymuconic semialdehyde dehydrogenase
Systematic name: 2-aminomuconate-6-semialdehyde:NAD⁺ 6-oxidoreductase
Comments: Also acts on 2-hydroxymuconate semialdehyde.
References: [1788]

[EC 1.2.1.32 created 1972]

EC 1.2.1.33

Accepted name: (R)-dehydropantoate dehydrogenase
Reaction: (R)-4-dehydropantoate + NAD⁺ + H₂O = (R)-3,3-dimethylmalate + NADH + 2 H⁺
Other name(s): D-aldopantoate dehydrogenase; D-2-hydroxy-3,3-dimethyl-3-formylpropionate:diphosphopyridine nucleotide (DPN⁺) oxidoreductase
Systematic name: (R)-4-dehydropantoate:NAD⁺ 4-oxidoreductase
References: [2607]

[EC 1.2.1.33 created 1972]

[1.2.1.34 Transferred entry. D-mannonate dehydrogenase (NAD(P)⁺). Now EC 1.1.1.131, mannuronate reductase]

[EC 1.2.1.34 created 1972, deleted 1983 [transferred to EC 1.1.1.180, deleted 1984]]

[1.2.1.35 Transferred entry. uronate dehydrogenase. Now EC 1.1.1.203, uronate dehydrogenase]

[EC 1.2.1.35 created 1972, deleted 1984]

EC 1.2.1.36

Accepted name: retinal dehydrogenase
Reaction: retinal + NAD⁺ + H₂O = retinoate + NADH + 2 H⁺
Other name(s): cytosolic retinal dehydrogenase
Systematic name: retinal:NAD⁺ oxidoreductase
Comments: A metalloflavoprotein (FAD). Acts on both the 11-*trans*- and 13-*cis*-forms of retinal.
References: [2856]

[EC 1.2.1.36 created 1972]

[1.2.1.37 Transferred entry. xanthine dehydrogenase. Now EC 1.17.1.4, xanthine dehydrogenase]

[EC 1.2.1.37 created 1972, deleted 1984]

EC 1.2.1.38

Accepted name: *N*-acetyl- γ -glutamyl-phosphate reductase
Reaction: *N*-acetyl-L-glutamate 5-semialdehyde + NADP⁺ + phosphate = *N*-acetyl-L-glutamyl 5-phosphate + NADPH + H⁺
Other name(s): reductase, acetyl- γ -glutamyl phosphate; *N*-acetylglutamate 5-semialdehyde dehydrogenase; *N*-acetylglutamic γ -semialdehyde dehydrogenase; *N*-acetyl-L-glutamate γ -semialdehyde:NADP⁺ oxidoreductase (phosphorylating)
Systematic name: *N*-acetyl-L-glutamate-5-semialdehyde:NADP⁺ 5-oxidoreductase (phosphorylating)
References: [181, 1338]

[EC 1.2.1.38 created 1972]

EC 1.2.1.39

Accepted name: phenylacetaldehyde dehydrogenase
Reaction: phenylacetaldehyde + NAD⁺ + H₂O = phenylacetate + NADH + 2 H⁺
Systematic name: phenylacetaldehyde:NAD⁺ oxidoreductase
References: [1205]

[EC 1.2.1.39 created 1976]

[1.2.1.40 Deleted entry. 3 α ,7 α ,12 α -trihydroxycholestan-26-al 26-oxidoreductase. The activity is part of EC 1.14.13.15, cholestanetriol 26-monoxygenase]

[EC 1.2.1.40 created 1976, deleted 2012]

EC 1.2.1.41

Accepted name: glutamate-5-semialdehyde dehydrogenase
Reaction: L-glutamate 5-semialdehyde + phosphate + NADP⁺ = L-glutamyl 5-phosphate + NADPH + H⁺
Other name(s): β -glutamylphosphate reductase; γ -glutamyl phosphate reductase; β -glutamylphosphate reductase; glutamate semialdehyde dehydrogenase; glutamate- γ -semialdehyde dehydrogenase
Systematic name: L-glutamate-5-semialdehyde:NADP⁺ 5-oxidoreductase (phosphorylating)
References: [180]

[EC 1.2.1.41 created 1976]

EC 1.2.1.42

Accepted name: hexadecanal dehydrogenase (acylating)
Reaction: hexadecanal + CoA + NAD⁺ = hexadecanoyl-CoA + NADH + H⁺
Other name(s): fatty acyl-CoA reductase
Systematic name: hexadecanal:NAD⁺ oxidoreductase (CoA-acylating)

Comments: Also acts, more slowly, on octadecanoyl-CoA.

References: [1937]

[EC 1.2.1.42 created 1978]

[1.2.1.43 *Transferred entry. formate dehydrogenase (NADP⁺). Now EC 1.17.1.10, formate dehydrogenase (NADP⁺)*]

[EC 1.2.1.43 created 1978, deleted 2017]

EC 1.2.1.44

Accepted name: cinnamoyl-CoA reductase

Reaction: cinnamaldehyde + CoA + NADP⁺ = cinnamoyl-CoA + NADPH + H⁺

Other name(s): feruloyl-CoA reductase; cinnamoyl-coenzyme A reductase; ferulyl-CoA reductase; feruloyl coenzyme A reductase; *p*-hydroxycinnamoyl coenzyme A reductase; cinnamoyl-CoA:NADPH reductase

Systematic name: cinnamaldehyde:NADP⁺ oxidoreductase (CoA-cinnamoylating)

Comments: Acts also on a number of substituted cinnamoyl esters of coenzyme A.

References: [1424, 3665, 4587]

[EC 1.2.1.44 created 1978]

[1.2.1.45 *Transferred entry. 4-carboxy-2-hydroxyruconate-6-semialdehyde dehydrogenase. Now EC 1.1.1.312, 2-hydroxy-4-carboxyruconate semialdehyde hemiacetal dehydrogenase.*]

[EC 1.2.1.45 created 1978, deleted 2011]

EC 1.2.1.46

Accepted name: formaldehyde dehydrogenase

Reaction: formaldehyde + NAD⁺ + H₂O = formate + NADH + 2 H⁺

Other name(s): NAD-linked formaldehyde dehydrogenase; NAD-dependent formaldehyde dehydrogenase

Systematic name: formaldehyde:NAD⁺ oxidoreductase

References: [1688]

[EC 1.2.1.46 created 1982]

EC 1.2.1.47

Accepted name: 4-trimethylammoniobutyraldehyde dehydrogenase

Reaction: 4-trimethylammoniobutanal + NAD⁺ + H₂O = 4-trimethylammoniobutanoate + NADH + 2 H⁺

Other name(s): 4-trimethylaminobutyraldehyde dehydrogenase; 4-*N*-trimethylaminobutyraldehyde dehydrogenase

Systematic name: 4-trimethylammoniobutanal:NAD⁺ 1-oxidoreductase

References: [3469]

[EC 1.2.1.47 created 1983]

EC 1.2.1.48

Accepted name: long-chain-aldehyde dehydrogenase

Reaction: a long-chain aldehyde + NAD⁺ + H₂O = a long-chain carboxylate + NADH + 2 H⁺

Other name(s): long-chain aliphatic aldehyde dehydrogenase; long-chain fatty aldehyde dehydrogenase; fatty aldehyde:NAD⁺ oxidoreductase

Systematic name: long-chain-aldehyde:NAD⁺ oxidoreductase

Comments: The best substrate is dodecylaldehyde.

References: [260, 2880, 2881]

[EC 1.2.1.48 created 1984]

EC 1.2.1.49

- Accepted name:** 2-oxoaldehyde dehydrogenase (NADP⁺)
Reaction: a 2-oxoaldehyde + NADP⁺ + H₂O = a 2-oxo carboxylate + NADPH + H⁺
Other name(s): α-ketoaldehyde dehydrogenase; methylglyoxal dehydrogenase; NADP⁺-linked α-ketoaldehyde dehydrogenase; 2-ketoaldehyde dehydrogenase; NADP⁺-dependent α-ketoaldehyde dehydrogenase
Systematic name: 2-oxoaldehyde:NADP⁺ 2-oxidoreductase
Comments: Not identical with EC 1.2.1.23 2-oxoaldehyde dehydrogenase (NAD⁺).
References: [3465, 3467]

[EC 1.2.1.49 created 1986]

EC 1.2.1.50

- Accepted name:** long-chain acyl-protein thioester reductase
Reaction: a long-chain aldehyde + [protein]-L-cysteine + NADP⁺ = a [protein]-S-(long-chain fatty acyl)-L-cysteine + NADPH + H⁺
Other name(s): *luxC* (gene name); acyl-CoA reductase; acyl coenzyme A reductase; long-chain-aldehyde:NADP⁺ oxidoreductase (acyl-CoA-forming); long-chain-fatty-acyl-CoA reductase
Systematic name: long-chain-aldehyde:NADP⁺ oxidoreductase (protein thioester-forming)
Comments: Together with a hydrolase component (EC 3.1.2.2 and EC 3.1.2.14) and a synthetase component (EC 6.2.1.19), this enzyme forms a multienzyme fatty acid reductase complex that produces the long-chain aldehyde substrate of the bacterial luciferase enzyme (EC 1.14.14.3). The enzyme is acylated by receiving an acyl group from EC 6.2.1.19, and catalyses the reduction of the acyl group, releasing the aldehyde product. The enzyme is also able to accept the acyl group from a long-chain acyl-CoA.
References: [3520, 4500, 2491]

[EC 1.2.1.50 created 1986, modified 2016]

EC 1.2.1.51

- Accepted name:** pyruvate dehydrogenase (NADP⁺)
Reaction: pyruvate + CoA + NADP⁺ = acetyl-CoA + CO₂ + NADPH
Systematic name: pyruvate:NADP⁺ 2-oxidoreductase (CoA-acetylating)
Comments: The *Euglena* enzyme can also use FAD or methyl viologen as acceptor, more slowly. The enzyme is inhibited by oxygen.
References: [1820, 1821]

[EC 1.2.1.51 created 1989]

EC 1.2.1.52

- Accepted name:** oxoglutarate dehydrogenase (NADP⁺)
Reaction: 2-oxoglutarate + CoA + NADP⁺ = succinyl-CoA + CO₂ + NADPH
Other name(s): oxoglutarate dehydrogenase (NADP)
Systematic name: 2-oxoglutarate:NADP⁺ 2-oxidoreductase (CoA-succinylating)
Comments: The *Euglena* enzyme can also use NAD⁺ as acceptor, but more slowly.
References: [1820]

[EC 1.2.1.52 created 1989]

EC 1.2.1.53

- Accepted name:** 4-hydroxyphenylacetaldehyde dehydrogenase
Reaction: 4-hydroxyphenylacetaldehyde + NAD⁺ + H₂O = 4-hydroxyphenylacetate + NADH + 2 H⁺
Other name(s): 4-HPAL dehydrogenase
Systematic name: 4-hydroxyphenylacetaldehyde:NAD⁺ oxidoreductase

Comments: With EC 4.2.1.87 octopamine dehydratase, brings about the metabolism of octopamine in *Pseudomonas*.

References: [790]

[EC 1.2.1.53 created 1989]

EC 1.2.1.54

Accepted name: γ -guanidinobutyraldehyde dehydrogenase

Reaction: 4-guanidinobutanal + NAD⁺ + H₂O = 4-guanidinobutanoate + NADH + 2 H⁺

Other name(s): α -guanidinobutyraldehyde dehydrogenase; 4-guanidinobutyraldehyde dehydrogenase; GBAL dehydrogenase

Systematic name: 4-guanidinobutanal:NAD⁺ 1-oxidoreductase

Comments: Involved in the degradation of arginine in *Pseudomonas putida* (cf. EC 1.2.1.19 aminobutyraldehyde dehydrogenase).

References: [4803]

[EC 1.2.1.54 created 1989]

[1.2.1.55 Transferred entry. (*R*)-3-hydroxyacid ester dehydrogenase. Now EC 1.1.1.279, (*R*)-3-hydroxyacid-ester dehydrogenase]

[EC 1.2.1.55 created 1990, deleted 2003]

[1.2.1.56 Transferred entry. (*S*)-3-hydroxyacid ester dehydrogenase. Now EC 1.1.1.280, (*S*)-3-hydroxyacid-ester dehydrogenase]

[EC 1.2.1.56 created 1990, deleted 2003]

EC 1.2.1.57

Accepted name: butanal dehydrogenase

Reaction: butanal + CoA + NAD(P)⁺ = butanoyl-CoA + NAD(P)H + H⁺

Systematic name: butanal:NAD(P)⁺ oxidoreductase (CoA-acylating)

Comments: Also acts on acetaldehyde, but more slowly.

References: [3225]

[EC 1.2.1.57 created 1992]

EC 1.2.1.58

Accepted name: phenylglyoxylate dehydrogenase (acylating)

Reaction: phenylglyoxylate + NAD⁺ + CoA = benzoyl-S-CoA + CO₂ + NADH

Systematic name: phenylglyoxylate:NAD⁺ oxidoreductase

Comments: Requires thiamine diphosphate as cofactor. The enzyme from the denitrifying bacterium *Azoarcus evansii* is specific for phenylglyoxylate. 2-Oxoisovalerate is oxidized at 15% of the rate for phenylglyoxylate. Also reduces viologen dyes. Contains iron-sulfur centres and FAD.

References: [1674]

[EC 1.2.1.58 created 1999]

EC 1.2.1.59

Accepted name: glyceraldehyde-3-phosphate dehydrogenase (NAD(P)⁺) (phosphorylating)

Reaction: D-glyceraldehyde 3-phosphate + phosphate + NAD(P)⁺ = 3-phospho-D-glyceroyl phosphate + NAD(P)H + H⁺

Other name(s): triosephosphate dehydrogenase (NAD(P)); glyceraldehyde-3-phosphate dehydrogenase (NAD(P)) (phosphorylating)

Systematic name: D-glyceraldehyde 3-phosphate:NAD(P)⁺ oxidoreductase (phosphorylating)
Comments: NAD⁺ and NADP⁺ can be used as cofactors with similar efficiency, unlike EC 1.2.1.12 glyceraldehyde-3-phosphate dehydrogenase (phosphorylating) and EC 1.2.1.13 glyceraldehyde-3-phosphate dehydrogenase (NADP⁺) (phosphorylating), which are NAD⁺- and NADP⁺-dependent, respectively.
References: [4393, 4394]

[EC 1.2.1.59 created 1999]

EC 1.2.1.60

Accepted name: 5-carboxymethyl-2-hydroxyumuconic-semialdehyde dehydrogenase
Reaction: 5-carboxymethyl-2-hydroxyumuconate semialdehyde + H₂O + NAD⁺ = 5-carboxymethyl-2-hydroxyumuconate + NADH + 2 H⁺
Other name(s): carboxymethylhydroxyumuconic semialdehyde dehydrogenase
Systematic name: 5-carboxymethyl-2-hydroxyumuconic-semialdehyde:NAD⁺ oxidoreductase
Comments: Involved in the tyrosine degradation pathway in *Arthrobacter* sp.
References: [349, 73, 730, 1277]

[EC 1.2.1.60 created 2000]

EC 1.2.1.61

Accepted name: 4-hydroxyumuconic-semialdehyde dehydrogenase
Reaction: 4-hydroxyumuconic semialdehyde + NAD⁺ + H₂O = maleylacetate + NADH + 2 H⁺
Systematic name: 4-hydroxyumuconic-semialdehyde:NAD⁺ oxidoreductase
Comments: Involved in the 4-nitrophenol degradation pathway.
References: [3971]

[EC 1.2.1.61 created 2000]

EC 1.2.1.62

Accepted name: 4-formylbenzenesulfonate dehydrogenase
Reaction: 4-formylbenzenesulfonate + NAD⁺ + H₂O = 4-sulfobenzoate + NADH + 2 H⁺
Systematic name: 4-formylbenzenesulfonate:NAD⁺ oxidoreductase
Comments: Involved in the toluene-4-sulfonate degradation pathway.
References: [1965, 1963]

[EC 1.2.1.62 created 2000]

EC 1.2.1.63

Accepted name: 6-oxohexanoate dehydrogenase
Reaction: 6-oxohexanoate + NADP⁺ + H₂O = adipate + NADPH + 2 H⁺
Systematic name: 6-oxohexanoate:NADP⁺ oxidoreductase
Comments: Last step in the cyclohexanol degradation pathway in *Acinetobacter* sp.
References: [837, 951]

[EC 1.2.1.63 created 2000]

EC 1.2.1.64

Accepted name: 4-hydroxybenzaldehyde dehydrogenase (NAD⁺)
Reaction: 4-hydroxybenzaldehyde + NAD⁺ + H₂O = 4-hydroxybenzoate + NADH + 2 H⁺
Other name(s): *p*-hydroxybenzaldehyde dehydrogenase (ambiguous); 4-hydroxybenzaldehyde dehydrogenase (ambiguous)

Systematic name: 4-hydroxybenzaldehyde:NAD⁺ oxidoreductase
Comments: The bacterial enzyme (characterized from an unidentified denitrifying bacterium) is involved in an anaerobic toluene degradation pathway. The plant enzyme is involved in formation of 4-hydroxybenzoate, a cell wall-bound phenolic acid that plays a major role in plant defense against pathogens. *cf.* EC 1.2.1.96, 4-hydroxybenzaldehyde dehydrogenase (NADP⁺).
References: [401, 3916]

[EC 1.2.1.64 created 2000, modified 2015]

EC 1.2.1.65

Accepted name: salicylaldehyde dehydrogenase
Reaction: salicylaldehyde + NAD⁺ + H₂O = salicylate + NADH + 2 H⁺
Systematic name: salicylaldehyde:NAD⁺ oxidoreductase
Comments: Involved in the naphthalene degradation pathway in some bacteria.
References: [1006]

[EC 1.2.1.65 created 2000, modified 2011]

[1.2.1.66 *Transferred entry. mycothiol-dependent formaldehyde dehydrogenase. Now EC 1.1.1.306, S-(hydroxymethyl)mycothiol dehydrogenase*]

[EC 1.2.1.66 created 2000, deleted 2010]

EC 1.2.1.67

Accepted name: vanillin dehydrogenase
Reaction: vanillin + NAD⁺ + H₂O = vanillate + NADH + 2 H⁺
Systematic name: vanillin:NAD⁺ oxidoreductase
References: [3351]

[EC 1.2.1.67 created 2000]

EC 1.2.1.68

Accepted name: coniferyl-aldehyde dehydrogenase
Reaction: coniferyl aldehyde + H₂O + NAD(P)⁺ = ferulate + NAD(P)H + 2 H⁺
Systematic name: coniferyl aldehyde:NAD(P)⁺ oxidoreductase
Comments: Also oxidizes other aromatic aldehydes, but not aliphatic aldehydes.
References: [10]

[EC 1.2.1.68 created 2000]

EC 1.2.1.69

Accepted name: fluoroacetaldehyde dehydrogenase
Reaction: fluoroacetaldehyde + NAD⁺ + H₂O = fluoroacetate + NADH + 2 H⁺
Systematic name: fluoroacetaldehyde:NAD⁺ oxidoreductase
Comments: The enzyme from *Streptomyces cattleya* has a high affinity for fluoroacetate and glycolaldehyde but not for acetaldehyde.
References: [2944, 2945]

[EC 1.2.1.69 created 2003]

EC 1.2.1.70

Accepted name: glutamyl-tRNA reductase

Reaction: L-glutamate 1-semialdehyde + NADP⁺ + tRNA^{Glu} = L-glutamyl-tRNA^{Glu} + NADPH + H⁺
Systematic name: L-glutamate-semialdehyde:NADP⁺ oxidoreductase (L-glutamyl-tRNA^{Glu}-forming)
Comments: This enzyme forms part of the pathway for the biosynthesis of 5-aminolevulinic acid from glutamate, known as the C5 pathway. The route shown in the diagram is used in most eubacteria, and in all archaeobacteria, algae and plants. However, in the α -proteobacteria, EC 2.3.1.37, 5-aminolevulinic acid synthase, is used in an alternative route to produce the product 5-aminolevulinic acid from succinyl-CoA and glycine. This route is found in the mitochondria of fungi and animals, organelles that are considered to be derived from an endosymbiotic α -proteobacterium. Although higher plants do not possess EC 2.3.1.37, the protistan *Euglena gracilis* possesses both the C5 pathway and EC 2.3.1.37.
References: [4471, 3352, 3705]

[EC 1.2.1.70 created 2004]

EC 1.2.1.71

Accepted name: succinylglutamate-semialdehyde dehydrogenase
Reaction: N-succinyl-L-glutamate 5-semialdehyde + NAD⁺ + H₂O = N-succinyl-L-glutamate + NADH + 2 H⁺
Other name(s): succinylglutamic semialdehyde dehydrogenase; N-succinylglutamate 5-semialdehyde dehydrogenase; SGSD; AruD; AstD
Systematic name: N-succinyl-L-glutamate 5-semialdehyde:NAD⁺ oxidoreductase
Comments: This is the fourth enzyme in the arginine succinyltransferase (AST) pathway for the catabolism of arginine [4563]. This pathway converts the carbon skeleton of arginine into glutamate, with the concomitant production of ammonia and conversion of succinyl-CoA into succinate and CoA. The five enzymes involved in this pathway are EC 2.3.1.109 (arginine N-succinyltransferase), EC 3.5.3.23 (N-succinylarginine dihydrolase), EC 2.6.1.11 (acetylornithine transaminase), EC 1.2.1.71 (succinylglutamate-semialdehyde dehydrogenase) and EC 3.5.1.96 (succinylglutamate desuccinylase) [4329, 782].
References: [4563, 4564, 4329, 1851, 3732, 782, 783]

[EC 1.2.1.71 created 2006]

EC 1.2.1.72

Accepted name: erythrose-4-phosphate dehydrogenase
Reaction: D-erythrose 4-phosphate + NAD⁺ + H₂O = 4-phosphoerythronate + NADH + 2 H⁺
Other name(s): erythrose 4-phosphate dehydrogenase; E4PDH; GapB; Epd dehydrogenase; E4P dehydrogenase
Systematic name: D-erythrose 4-phosphate:NAD⁺ oxidoreductase
Comments: This enzyme was originally thought to be a glyceraldehyde-3-phosphate dehydrogenase (EC 1.2.1.12), but this has since been disproved, as glyceraldehyde 3-phosphate is not a substrate [4899, 398]. Forms part of the pyridoxal-5'-phosphate coenzyme biosynthesis pathway in *Escherichia coli*, along with EC 1.1.1.290 (4-phosphoerythronate dehydrogenase), EC 2.6.1.52 (phosphoserine transaminase), EC 1.1.1.262 (4-hydroxythreonine-4-phosphate dehydrogenase), EC 2.6.99.2 (pyridoxine 5'-phosphate synthase) and EC 1.4.3.5 (pyridoxamine-phosphate oxidase).
References: [4899, 398, 4771]

[EC 1.2.1.72 created 2006]

EC 1.2.1.73

Accepted name: sulfoacetaldehyde dehydrogenase
Reaction: 2-sulfoacetaldehyde + H₂O + NAD⁺ = sulfoacetate + NADH + 2 H⁺
Other name(s): SafD
Systematic name: 2-sulfoacetaldehyde:NAD⁺ oxidoreductase

Comments: This reaction is part of a bacterial pathway that can utilize the amino group of taurine as a sole source of nitrogen for growth. At physiological concentrations, NAD⁺ cannot be replaced by NADP⁺. The enzyme is specific for sulfoacetaldehyde, as formaldehyde, acetaldehyde, betaine aldehyde, propanal, glyceraldehyde, phosphonoacetaldehyde, glyoxylate, glycolaldehyde and 2-oxobutyrate are not substrates.

References: [2260]

[EC 1.2.1.73 created 2008]

EC 1.2.1.74

Accepted name: abieta-7,13-dien-18-al dehydrogenase

Reaction: abieta-7,13-dien-18-al + H₂O + NAD⁺ = abieta-7,13-dien-18-oate + NADH + H⁺

Other name(s): abietadienal dehydrogenase (ambiguous)

Systematic name: abieta-7,13-dien-18-al:NAD⁺ oxidoreductase

Comments: Abietic acid is the principle component of conifer resin. This enzyme catalyses the last step of the pathway of abietic acid biosynthesis in *Abies grandis* (grand fir). The activity has been demonstrated in cell-free stem extracts of *A. grandis*, was present in the cytoplasm, and required NAD⁺ as cofactor [1230]. The enzyme is expressed constitutively at a high level, and is not inducible by wounding of the plant tissue [1232].

References: [1230, 1232]

[EC 1.2.1.74 created 2009, modified 2012]

EC 1.2.1.75

Accepted name: malonyl-CoA reductase (malonate semialdehyde-forming)

Reaction: malonate semialdehyde + CoA + NADP⁺ = malonyl-CoA + NADPH + H⁺

Other name(s): NADP-dependent malonyl CoA reductase; malonyl CoA reductase (NADP); malonyl CoA reductase (malonate semialdehyde-forming)

Systematic name: malonate semialdehyde:NADP⁺ oxidoreductase (malonate semialdehyde-forming)

Comments: Requires Mg²⁺. Catalyses the reduction of malonyl-CoA to malonate semialdehyde, a key step in the 3-hydroxypropanoate and the 3-hydroxypropanoate/4-hydroxybutanoate cycles, autotrophic CO₂ fixation pathways found in some green non-sulfur phototrophic bacteria and some thermoacidophilic archaea, respectively [4056, 296]. The enzyme from *Sulfolobus tokodaii* has been purified, and found to contain one RNA molecule per two subunits [57]. The enzyme from *Chloroflexus aurantiacus* is bifunctional, and also catalyses the next reaction in the pathway, EC 1.1.1.298 [3-hydroxypropionate dehydrogenase (NADP⁺)] [1764].

References: [4056, 296, 57, 1764]

[EC 1.2.1.75 created 2009]

EC 1.2.1.76

Accepted name: succinate-semialdehyde dehydrogenase (acylating)

Reaction: succinate semialdehyde + CoA + NADP⁺ = succinyl-CoA + NADPH + H⁺

Other name(s): succinyl-coA reductase; coenzyme-A-dependent succinate-semialdehyde dehydrogenase

Systematic name: succinate semialdehyde:NADP⁺ oxidoreductase (CoA-acylating)

Comments: Catalyses the NADPH-dependent reduction of succinyl-CoA to succinate semialdehyde. The enzyme has been described in *Clostridium kluyveri*, where it participates in succinate fermentation [3952], and in *Metallosphaera sedula*, where it participates in the 3-hydroxypropanoate/4-hydroxybutanoate cycle, an autotrophic CO₂ fixation pathway found in some thermoacidophilic archaea [57, 296].

References: [3952, 57, 296]

[EC 1.2.1.76 created 2009]

EC 1.2.1.77

- Accepted name:** 3,4-dehydroadipyl-CoA semialdehyde dehydrogenase (NADP⁺)
Reaction: 3,4-didehydroadipyl-CoA semialdehyde + NADP⁺ + H₂O = 3,4-didehydroadipyl-CoA + NADPH + H⁺
Other name(s): BoxD; 3,4-dehydroadipyl-CoA semialdehyde dehydrogenase
Systematic name: 3,4-didehydroadipyl-CoA semialdehyde:NADP⁺ oxidoreductase
Comments: This enzyme catalyses a step in the aerobic benzoyl-coenzyme A catabolic pathway in *Azoarcus evansii* and *Burkholderia xenovorans*.
References: [1307, 185]

[EC 1.2.1.77 created 2010]

EC 1.2.1.78

- Accepted name:** 2-formylbenzoate dehydrogenase
Reaction: 2-formylbenzoate + NAD⁺ + H₂O = *o*-phthalic acid + NADH + H⁺
Other name(s): 2-carboxybenzaldehyde dehydrogenase; 2CBAL dehydrogenase; PhdK
Systematic name: 2-formylbenzoate:NAD⁺ oxidoreductase
Comments: The enzyme is involved in phenanthrene degradation.
References: [1857, 2145]

[EC 1.2.1.78 created 2010]

EC 1.2.1.79

- Accepted name:** succinate-semialdehyde dehydrogenase (NADP⁺)
Reaction: succinate semialdehyde + NADP⁺ + H₂O = succinate + NADPH + 2 H⁺
Other name(s): succinic semialdehyde dehydrogenase (NADP⁺); succinyl semialdehyde dehydrogenase (NADP⁺); succinate semialdehyde:NADP⁺ oxidoreductase; NADP-dependent succinate-semialdehyde dehydrogenase; GabD
Systematic name: succinate-semialdehyde:NADP⁺ oxidoreductase
Comments: This enzyme participates in the degradation of glutamate and 4-aminobutyrate. It is similar to EC 1.2.1.24 [succinate-semialdehyde dehydrogenase (NAD⁺)], and EC 1.2.1.16 [succinate-semialdehyde dehydrogenase (NAD(P)⁺)], but is specific for NADP⁺. The enzyme from *Escherichia coli* is 20-fold more active with NADP⁺ than NAD⁺ [1875].
References: [229, 1875]

[EC 1.2.1.79 created 2010]

EC 1.2.1.80

- Accepted name:** long-chain acyl-[acyl-carrier-protein] reductase
Reaction: a long-chain aldehyde + an [acyl-carrier protein] + NAD(P)⁺ = a long-chain acyl-[acyl-carrier protein] + NAD(P)H + H⁺
Other name(s): long-chain acyl-[acp] reductase; fatty acyl-[acyl-carrier-protein] reductase; acyl-[acp] reductase
Systematic name: long-chain-aldehyde:NAD(P)⁺ oxidoreductase (acyl-[acyl-carrier protein]-forming)
Comments: Catalyses the reaction in the opposite direction. This enzyme, purified from the cyanobacterium *Synechococcus elongatus* PCC 7942, catalyses the NAD(P)H-dependent reduction of an activated fatty acid (acyl-[acp]) to the corresponding aldehyde. Together with EC 4.1.99.5, octadecanal decarboxylase, it is involved in alkane biosynthesis. The natural substrates of the enzyme are C₁₆ and C₁₈ activated fatty acids. Requires Mg²⁺.
References: [3717]

[EC 1.2.1.80 created 2011]

EC 1.2.1.81

- Accepted name:** sulfoacetaldehyde dehydrogenase (acylating)
Reaction: 2-sulfoacetaldehyde + CoA + NADP⁺ = sulfoacetyl-CoA + NADPH + H⁺
Other name(s): SauS
Systematic name: 2-sulfoacetaldehyde:NADP⁺ oxidoreductase (CoA-acylating)
Comments: The enzyme is involved in degradation of sulfoacetate. In this pathway the reaction is catalysed in the reverse direction. The enzyme is specific for sulfoacetaldehyde and NADP⁺.
References: [4577]

[EC 1.2.1.81 created 2011]

EC 1.2.1.82

- Accepted name:** β-apo-4'-carotenal oxygenase
Reaction: 4'-apo-β,ψ-caroten-4'-al + NAD⁺ + H₂O = neurosporaxanthin + NADH + 2 H⁺
Other name(s): β-apo-4'-carotenal dehydrogenase; YLO-1; *carD* (gene name)
Systematic name: 4'-apo-β,ψ-carotenal:NAD⁺ oxidoreductase
Comments: Neurosporaxanthin is responsible for the orange color of *Neurospora*.
References: [1067, 901]

[EC 1.2.1.82 created 2011]

EC 1.2.1.83

- Accepted name:** 3-succinoylsemialdehyde-pyridine dehydrogenase
Reaction: 4-oxo-4-(pyridin-3-yl)butanal + NADP⁺ + H₂O = 4-oxo-4-(pyridin-3-yl)butanoate + NADPH + H⁺
Systematic name: 4-oxo-4-(pyridin-3-yl)butanal:NADP⁺ oxidoreductase
Comments: The enzyme has been characterized from the soil bacterium *Pseudomonas* sp. HZN6. It participates in the nicotine degradation pathway.
References: [3402]

[EC 1.2.1.83 created 2012]

EC 1.2.1.84

- Accepted name:** alcohol-forming fatty acyl-CoA reductase
Reaction: a long-chain acyl-CoA + 2 NADPH + 2 H⁺ = a long-chain alcohol + 2 NADP⁺ + CoA
Other name(s): FAR (gene name); long-chain acyl-CoA:NADPH reductase
Systematic name: NADPH:long-chain acyl-CoA reductase
Comments: The enzyme has been characterized from the plant *Simmondsia chinensis* (jojoba). The alcohol is formed by a four-electron reduction of fatty acyl-CoA. Although the reaction proceeds through an aldehyde intermediate, a free aldehyde is not released. The recombinant enzyme was shown to accept saturated and mono-unsaturated fatty acyl-CoAs of 16 to 22 carbons.
References: [2782]

[EC 1.2.1.84 created 2012]

EC 1.2.1.85

- Accepted name:** 2-hydroxyomuconate-6-semialdehyde dehydrogenase
Reaction: 2-hydroxyomuconate-6-semialdehyde + NAD⁺ + H₂O = (2Z,4E)-2-hydroxyhexa-2,4-dienedioate + NADH + 2 H⁺
Other name(s): *xyIG* (gene name); *praB* (gene name)
Systematic name: 2-hydroxyomuconate-6-semialdehyde:NAD⁺ oxidoreductase

Comments: This substrate for this enzyme is formed by *meta* ring cleavage of catechol (EC 1.13.11.2, catechol 2,3-dioxygenase), and is an intermediate in the bacterial degradation of several aromatic compounds. Has lower activity with benzaldehyde [1815]. Activity with NAD⁺ is more than 10-fold higher than with NADP⁺ [2005]. *cf.* EC 1.2.1.32, aminomuconate-semialdehyde dehydrogenase.

References: [1815, 3191, 2005]

[EC 1.2.1.85 created 2012]

EC 1.2.1.86

Accepted name: geranial dehydrogenase
Reaction: geranial + H₂O + NAD⁺ = geranate + NADH + H⁺
Other name(s): GaDH; *geoB* (gene name)
Systematic name: geranial:NAD⁺ oxidoreductase
Comments: Does not act on neral.
References: [4658, 2556]

[EC 1.2.1.86 created 2012]

EC 1.2.1.87

Accepted name: propanal dehydrogenase (CoA-propanoylating)
Reaction: propanal + CoA + NAD⁺ = propanoyl-CoA + NADH + H⁺
Other name(s): BphJ
Systematic name: propanal:NAD⁺ oxidoreductase (CoA-propanoylating)
Comments: The enzyme forms a bifunctional complex with EC 4.1.3.43, 4-hydroxy-2-oxohexanoate aldolase, with a tight channel connecting the two subunits [1,2,3]. Also acts, more slowly, on glycolaldehyde and butanal. In *Pseudomonas* species the enzyme forms a bifunctional complex with EC 4.1.3.39, 4-hydroxy-2-oxovalerate aldolase. The enzymes from the bacteria *Burkholderia xenovorans* and *Thermus thermophilus* also perform the reaction of EC 1.2.1.10, acetaldehyde dehydrogenase (acetylating). NADP⁺ can replace NAD⁺ with a much slower rate [195].
References: [196, 555, 195]

[EC 1.2.1.87 created 2013]

EC 1.2.1.88

Accepted name: L-glutamate γ -semialdehyde dehydrogenase
Reaction: L-glutamate 5-semialdehyde + NAD⁺ + H₂O = L-glutamate + NADH + H⁺
Other name(s): 1-pyrroline-5-carboxylate dehydrogenase; Δ^1 -pyrroline-5-carboxylate dehydrogenase; 1-pyrroline dehydrogenase; pyrroline-5-carboxylate dehydrogenase; pyrroline-5-carboxylic acid dehydrogenase; L-pyrroline-5-carboxylate-NAD⁺ oxidoreductase; 1-pyrroline-5-carboxylate:NAD⁺ oxidoreductase; Δ^1 -pyrroline-5-carboxylic acid dehydrogenase
Systematic name: L-glutamate γ -semialdehyde:NAD⁺ oxidoreductase
Comments: This enzyme catalyses the irreversible oxidation of glutamate- γ -semialdehyde to glutamate as part of the proline degradation pathway. (*S*)-1-pyrroline-5-carboxylate, the product of the first enzyme of the pathway (EC 1.5.5.2, proline dehydrogenase) is in spontaneous equilibrium with its tautomer L-glutamate γ -semialdehyde. In many bacterial species, both activities are carried out by a single bifunctional enzyme [1141, 458]. The enzyme can also oxidize other 1-pyrrolines, e.g. 3-hydroxy-1-pyrroline-5-carboxylate is converted into 4-hydroxyglutamate and (*R*)-1-pyrroline-5-carboxylate is converted into D-glutamate. NADP⁺ can also act as acceptor, but with lower activity [1808].
References: [24, 4058, 1141, 458, 1808]

[EC 1.2.1.88 created 1972 as EC 1.5.1.12, modified 2008, transferred 2013 to EC 1.2.1.88]

EC 1.2.1.89

- Accepted name:** D-glyceraldehyde dehydrogenase (NADP⁺)
Reaction: D-glyceraldehyde + NADP⁺ + H₂O = D-glycerate + NADPH + H⁺
Other name(s): glyceraldehyde dehydrogenase; GADH
Systematic name: D-glyceraldehyde:NADP⁺ oxidoreductase
Comments: The enzyme from the archaea *Thermoplasma acidophilum* and *Picrophilus torridus* is involved in the non-phosphorylative Entner-Doudoroff pathway. *cf.* EC 1.2.99.8, glyceraldehyde dehydrogenase (FAD-containing).
References: [1960, 3486]

[EC 1.2.1.89 created 2014]

EC 1.2.1.90

- Accepted name:** glyceraldehyde-3-phosphate dehydrogenase [NAD(P)⁺]
Reaction: D-glyceraldehyde 3-phosphate + NAD(P)⁺ + H₂O = 3-phospho-D-glycerate + NAD(P)H + 2 H⁺
Other name(s): non-phosphorylating glyceraldehyde-3-phosphate dehydrogenase (ambiguous); GAPN
Systematic name: D-glyceraldehyde-3-phosphate:NAD(P)⁺ oxidoreductase
Comments: The enzyme is part of the modified Embden-Meyerhof-Parnas pathway of the archaeon *Thermoproteus tenax*. *cf.* EC 1.2.1.9 [glyceraldehyde-3-phosphate dehydrogenase (NADP⁺)].
References: [476, 477, 3340, 2540]

[EC 1.2.1.90 created 2014]

EC 1.2.1.91

- Accepted name:** 3-oxo-5,6-dehydrosuberil-CoA semialdehyde dehydrogenase
Reaction: 3-oxo-5,6-dehydrosuberil-CoA semialdehyde + NADP⁺ + H₂O = 3-oxo-5,6-dehydrosuberil-CoA + NADPH + H⁺
Other name(s): *paaZ* (gene name)
Systematic name: 3-oxo-5,6-dehydrosuberil-CoA semialdehyde:NADP⁺ oxidoreductase
Comments: The enzyme from *Escherichia coli* is a bifunctional fusion protein that also catalyses EC 3.3.2.12, oxepin-CoA hydrolase. Combined the two activities result in a two-step conversion of oxepin-CoA to 3-oxo-5,6-dehydrosuberil-CoA, part of an aerobic phenylacetate degradation pathway.
References: [1107, 1836, 4250]

[EC 1.2.1.91 created 2011 as EC 1.17.1.7, transferred 2014 to EC 1.2.1.91]

EC 1.2.1.92

- Accepted name:** 3,6-anhydro- α -L-galactose dehydrogenase
Reaction: 3,6-anhydro- α -L-galactopyranose + NAD(P)⁺ + H₂O = 3,6-anhydro-L-galactonate + NAD(P)H + H⁺
Systematic name: 3,6-anhydro- α -L-galactopyranose:NAD(P)⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the marine bacterium *Vibrio* sp. EJY3, is involved in a degradation pathway for 3,6-anhydro- α -L-galactose, a major component of the polysaccharides produced by red macroalgae, such as agarose and porphyran.
References: [4846]

[EC 1.2.1.92 created 2014]

[1.2.1.93 Transferred entry. formate dehydrogenase (NAD⁺, ferredoxin). Now EC 1.17.1.11, formate dehydrogenase (NAD⁺, ferredoxin)]

[EC 1.2.1.93 created 2015, deleted 2017]

EC 1.2.1.94

- Accepted name:** farnesal dehydrogenase
Reaction: (2*E*,6*E*)-farnesal + NAD⁺ + H₂O = (2*E*,6*E*)-farnesoate + NADH + 2 H⁺
Other name(s): AaALDH3
Systematic name: farnesal:NAD⁺ oxidoreductase
Comments: Involved in juvenile hormone production in insects. The enzyme was described from the corpora allata of *Drosophila melanogaster* (fruit fly), *Manduca sexta* (tobacco hornworm) and *Aedes aegypti* (dengue mosquito).
References: [2601, 193, 3530]

[EC 1.2.1.94 created 2015]

EC 1.2.1.95

- Accepted name:** L-2-aminoadipate reductase
Reaction: (S)-2-amino-6-oxohexanoate + NADP⁺ + AMP + diphosphate = L-2-aminoadipate + NADPH + H⁺ + ATP (overall reaction)
(1a) L-2-aminoadipyl-[LYS2 peptidyl-carrier-protein] + AMP + diphosphate = L-2-aminoadipate + holo-[LYS2 peptidyl-carrier-protein] + ATP
(1b) (S)-2-amino-6-oxohexanoate + holo-[LYS2 peptidyl-carrier-protein] + NADP⁺ = L-2-aminoadipyl-[LYS2 peptidyl-carrier-protein] + NADPH + H⁺
Other name(s): LYS2; α-aminoadipate reductase
Systematic name: (S)-2-amino-6-oxohexanoate:NADP⁺ oxidoreductase (ATP-forming)
Comments: This enzyme, characterized from the yeast *Saccharomyces cerevisiae*, catalyses the reduction of L-2-aminoadipate to (S)-2-amino-6-oxohexanoate during L-lysine biosynthesis. An adenylation domain activates the substrate at the expense of ATP hydrolysis, and forms L-2-aminoadipate adenylate, which is attached to a peptidyl-carrier protein (PCP) domain. Binding of NADPH results in reductive cleavage of the acyl-S-enzyme intermediate, releasing (S)-2-amino-6-oxohexanoate. Different from EC 1.2.1.31, L-aminoadipate-semialdehyde dehydrogenase, which catalyses a similar transformation in the opposite direction without ATP hydrolysis.
References: [1025]

[EC 1.2.1.95 created 2015]

EC 1.2.1.96

- Accepted name:** 4-hydroxybenzaldehyde dehydrogenase (NADP⁺)
Reaction: 4-hydroxybenzaldehyde + NADP⁺ + H₂O = 4-hydroxybenzoate + NADPH + 2 H⁺
Other name(s): *p*-hydroxybenzaldehyde dehydrogenase (ambiguous); *pchA* (gene name)
Systematic name: 4-hydroxybenzaldehyde:NADP⁺ oxidoreductase
Comments: Involved in the aerobic pathway for degradation of toluene, 4-methylphenol, and 2,4-xyleneol by several *Pseudomonas* strains. The enzyme is also active with 4-hydroxy-3-methylbenzaldehyde. *cf.* EC 1.2.1.64, 4-hydroxybenzaldehyde dehydrogenase (NAD⁺).
References: [4605, 639]

[EC 1.2.1.96 created 2015]

EC 1.2.1.97

- Accepted name:** 3-sulfolactaldehyde dehydrogenase
Reaction: (2*S*)-3-sulfolactaldehyde + NAD(P)⁺ + H₂O = (2*S*)-3-sulfolactate + NAD(P)H + H⁺
Other name(s): SLA dehydrogenase
Systematic name: (2*S*)-3-sulfolactaldehyde:NAD(P)⁺ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Pseudomonas putida* SQ1, participates in a sulfoquinovose degradation pathway. Also acts on succinate semialdehyde.
References: [1098]

[EC 1.2.1.97 created 2015]

EC 1.2.1.98

Accepted name: 2-hydroxy-2-methylpropanal dehydrogenase
Reaction: 2-hydroxy-2-methylpropanal + NAD⁺ + H₂O = 2-hydroxy-2-methylpropanoate + NADH + H⁺
Other name(s): *mpdC* (gene name)
Systematic name: 2-hydroxy-2-methylpropanal:NAD⁺ oxidoreductase
Comments: This bacterial enzyme is involved in the degradation pathways of the alkene 2-methylpropene and the fuel additive *tert*-butyl methyl ether (MTBE), a widely occurring groundwater contaminant.
References: [1110]

[EC 1.2.1.98 created 2016]

EC 1.2.1.99

Accepted name: 4-(γ -glutamylamino)butanal dehydrogenase
Reaction: 4-(γ -L-glutamylamino)butanal + NAD(P)⁺ + H₂O = 4-(γ -L-glutamylamino)butanoate + NAD(P)H + H⁺
Other name(s): *puuC* (gene name)
Systematic name: 4-(γ -L-glutamylamino)butanal:NAD(P)⁺ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Escherichia coli*, is involved in a putrescine catabolic pathway. It has a broad substrate range, and can also catalyse the activities of EC 1.2.1.19, aminobutyraldehyde dehydrogenase, and EC 1.2.1.24, succinate-semialdehyde dehydrogenase (NAD⁺).
References: [2301, 1920, 3733]

[EC 1.2.1.99 created 2017]

EC 1.2.1.100

Accepted name: 5-formyl-3-hydroxy-2-methylpyridine 4-carboxylic acid 5-dehydrogenase
Reaction: 5-formyl-3-hydroxy-2-methylpyridine-4-carboxylate + NAD⁺ + H₂O = 3-hydroxy-2-methylpyridine-4,5-dicarboxylate + NADH + H⁺
Other name(s): *mlr6793* (locus name)
Systematic name: 5-formyl-3-hydroxy-2-methylpyridine-4-carboxylate:NAD⁺ 5-oxidoreductase
Comments: The enzyme, characterized from the bacteria *Pseudomonas* sp. MA-1 and *Mesorhizobium loti*, participates in the degradation of pyridoxine (vitamin B₆).
References: [2401, 4794, 2916]

[EC 1.2.1.100 created 2018]

EC 1.2.1.101

Accepted name: L-tyrosine reductase
Reaction: L-tyrosinal + NADP⁺ + AMP + diphosphate = L-tyrosine + NADPH + H⁺ + ATP
Other name(s): *InaA* (gene name); *InbA* (gene name)
Systematic name: (2*S*)-2-amino-3-(4-hydroxyphenyl)propanal:NADP⁺ oxidoreductase (ATP-forming)
Comments: The enzyme, characterized from the ascomycete fungus *Aspergillus flavus*, is specific for L-tyrosine. It contains three domains - an adenylation domain, a peptidyl-carrier protein (PCP) domain, and a reductase domain, and requires activation by attachment of a phosphopantetheinyl group. The enzyme activates its substrate to an adenylate form, followed by a transfer to the PCP domain. The resulting thioester is subsequently transferred to the reductase domain, where it is reduced to the aldehyde.
References: [1149]

[EC 1.2.1.101 created 2018]

EC 1.2.1.102

- Accepted name:** isopyridoxal dehydrogenase (5-pyridoxate-forming)
Reaction: isopyridoxal + NAD⁺ + H₂O = 5-pyridoxate + NADH + H⁺
Systematic name: isopyridoxal:NAD⁺ oxidoreductase (5-pyridoxate-forming)
Comments: The enzyme, characterized from the bacterium *Arthrobacter* sp. Cr-7, participates in the degradation of pyridoxine. The enzyme also catalyses the activity of EC 1.1.1.416, isopyridoxal dehydrogenase (5-pyridoxolactone-forming).
References: [2401]

[EC 1.2.1.102 created 2018]

EC 1.2.1.103

- Accepted name:** [amino-group carrier protein]-6-phospho-L-2-aminoadipate reductase
Reaction: an [amino-group carrier protein]-C-terminal-[N-(1-carboxy-5-oxopentyl)-L-glutamine] + phosphate + NADP⁺ = an [amino-group carrier protein]-C-terminal-[N-(1-carboxy-5-phosphooxy-5-oxopentyl)-L-glutamine] + NADPH + H⁺
Other name(s): *lysY* (gene name)
Systematic name: [amino-group carrier protein]-C-terminal-[N-(1-carboxy-5-oxopentyl)-L-glutamine]:NADP⁺ 5-oxidoreductase (phosphorylating)
Comments: The enzyme participates in an L-lysine biosynthesis in certain species of archaea and bacteria.
References: [3074, 1722, 3873]

[EC 1.2.1.103 created 2019]

EC 1.2.1.104

- Accepted name:** pyruvate dehydrogenase system
Reaction: pyruvate + CoA + NAD⁺ = acetyl-CoA + CO₂ + NADH
Other name(s): pyruvate dehydrogenase complex; PDH
Systematic name: pyruvate:NAD⁺ 2-oxidoreductase (CoA-acetylating)
Comments: The pyruvate dehydrogenase system (PDH) is a large enzyme complex that belongs to the 2-oxoacid dehydrogenase system family, which also includes EC 1.2.1.25, branched-chain α-keto acid dehydrogenase system, EC 1.2.1.105, 2-oxoglutarate dehydrogenase system, EC 1.4.1.27, glycine cleavage system, and EC 2.3.1.190, acetoin dehydrogenase system. With the exception of the glycine cleavage system, which contains 4 components, the 2-oxoacid dehydrogenase systems share a common structure, consisting of three main components, namely a 2-oxoacid dehydrogenase (E1), a dihydrolipoamide acyltransferase (E2), and a dihydrolipoamide dehydrogenase (E3). The reaction catalysed by this system is the sum of three activities: EC 1.2.4.1, pyruvate dehydrogenase (acetyltransfering) (E1), EC 2.3.1.12, dihydrolipoyllysine-residue acetyltransferase (E2), and EC 1.8.1.4, dihydrolipoyl dehydrogenase (E3). The mammalian system also includes E3 binding protein, which is involved in the interaction between the E2 and E3 subunits.
References: [3479, 236, 4002, 4764, 3256]

[EC 1.2.1.104 created 2020]

EC 1.2.1.105

- Accepted name:** 2-oxoglutarate dehydrogenase system
Reaction: 2-oxoglutarate + CoA + NAD⁺ = succinyl-CoA + CO₂ + NADH
Other name(s): 2-oxoglutarate dehydrogenase complex
Systematic name: 2-oxoglutarate:NAD⁺ 2-oxidoreductase (CoA-succinylating)

Comments: The 2-oxoglutarate dehydrogenase system is a large enzyme complex that belongs to the 2-oxoacid dehydrogenase system family, which also includes EC 1.2.1.25, branched-chain α -keto acid dehydrogenase system, EC 1.2.1.104, pyruvate dehydrogenase system, EC 1.4.1.27, glycine cleavage system, and EC 2.3.1.190, acetoin dehydrogenase system. With the exception of the glycine cleavage system, which contains 4 components, the 2-oxoacid dehydrogenase systems share a common structure, consisting of three main components, namely a 2-oxoacid dehydrogenase (E1), a dihydrolipoamide acyltransferase (E2), and a dihydrolipoamide dehydrogenase (E3). This enzyme system converts 2-oxoglutarate to succinyl-CoA and produces NADH and CO₂ in a complicated series of irreversible reactions. The reaction catalysed by this system is the sum of three activities: EC 1.2.4.2, oxoglutarate dehydrogenase (succinyl-transferring) (E1), EC 2.3.1.61, dihydrolipoyllysine-residue succinyltransferase (E2) and EC 1.8.1.4, dihydrolipoyl dehydrogenase (E3).

References: [3539, 2164, 3478, 2946, 1165, 494, 795]

[EC 1.2.1.105 created 2020]

EC 1.2.1.106

Accepted name: [amino-group carrier protein]-5-phospho-L-glutamate reductase
Reaction: an [amino-group carrier protein]-C-terminal- γ -(L-glutamate 5-semialdehyde-2-yl)-L-glutamate + phosphate + NADP⁺ = an [amino-group carrier protein]-C-terminal- γ -(5-phospho-L-glutamyl)-L-glutamate + NADPH + H⁺
Other name(s): *lysY* (gene name)
Systematic name: [amino-group carrier protein]-C-terminal- γ -(L-glutamate 5-semialdehyde-2-yl)-L-glutamate:NADP⁺ 5-oxidoreductase (phosphorylating)
Comments: The enzyme participates in an L-arginine biosynthesis pathway in certain species of archaea and bacteria. In some organisms the enzyme is bifunctional and also catalyses the activity of EC 1.2.1.103, [amino-group carrier protein]-6-phospho-L-2-aminoadipate reductase.
References: [3213, 4806]

[EC 1.2.1.106 created 2021]

EC 1.2.1.107

Accepted name: glyceraldehyde-3-phosphate dehydrogenase (arsenate-transferring)
Reaction: D-glyceraldehyde 3-phosphate + arsenate + NAD⁺ = 1-arsono-3-phospho-D-glycerate + NADH + H⁺
Systematic name: D-glyceraldehyde-3-phosphate:NAD⁺ oxidoreductase (arsenate-transferring)
Comments: The enzyme, discovered in bacteria, is very similar to EC 1.2.1.12, glyceraldehyde-3-phosphate dehydrogenase (phosphorylating). However, the gene encoding it is located in arsenic resistance islands in the chromosome, next to a gene (*arsJ*) that encodes a transporter that removes the product, 1-arsono-3-phosphoglycerate, from the cell. Together the two proteins form an arsenic detoxification system.
References: [631, 4677]

[EC 1.2.1.107 created 2021]

EC 1.2.2 With a cytochrome as acceptor

EC 1.2.2.1

Accepted name: formate dehydrogenase (cytochrome)
Reaction: formate + 2 ferricytochrome *b*₁ = CO₂ + 2 ferrocycytochrome *b*₁ + 2 H⁺
Other name(s): formate dehydrogenase; formate:cytochrome *b*₁ oxidoreductase
Systematic name: formate:ferricytochrome-*b*₁ oxidoreductase
References: [1257]

[EC 1.2.2.1 created 1961]

[1.2.2.2 Deleted entry. pyruvate dehydrogenase (cytochrome). Now covered by EC 1.2.5.1, pyruvate dehydrogenase (quinone)]

[EC 1.2.2.2 created 1961, deleted 2010]

[1.2.2.3 Transferred entry. formate dehydrogenase (cytochrome-c-553). Now EC 1.17.2.3, formate dehydrogenase (cytochrome-c-553)]

[EC 1.2.2.3 created 1981, deleted 2017]

[1.2.2.4 Deleted entry. carbon-monoxide dehydrogenase (cytochrome b-561). Now classified as EC 1.2.5.3, aerobic carbon monoxide dehydrogenase]

[EC 1.2.2.4 created 1999 (EC 1.2.3.10 created 1990, incorporated 2003), modified 2003, deleted 2020]

EC 1.2.3 With oxygen as acceptor

EC 1.2.3.1

Accepted name: aldehyde oxidase
Reaction: an aldehyde + H₂O + O₂ = a carboxylate + H₂O₂
Other name(s): quinoline oxidase; retinal oxidase
Systematic name: aldehyde:oxygen oxidoreductase
Comments: Contains molybdenum, [2Fe-2S] centres and FAD. The enzyme from liver exhibits a broad substrate specificity, and is involved in the metabolism of xenobiotics, including the oxidation of *N*-heterocycles and aldehydes and the reduction of *N*-oxides, nitrosamines, hydroxamic acids, azo dyes, nitropolycyclic aromatic hydrocarbons, and sulfoxides [2261, 4815]. The enzyme is also responsible for the oxidation of retinal, an activity that was initially attributed to a distinct enzyme (EC 1.2.3.11, retinal oxidase) [4303, 1750].
References: [1366, 2173, 2615, 2261, 4303, 4815, 1750, 4359]

[EC 1.2.3.1 created 1961, modified 2002, modified 2004, modified 2012]

[1.2.3.2 Transferred entry. xanthine oxidase. Now EC 1.17.3.2, xanthine oxidase]

[EC 1.2.3.2 created 1961, deleted 1984]

EC 1.2.3.3

Accepted name: pyruvate oxidase
Reaction: pyruvate + phosphate + O₂ = acetyl phosphate + CO₂ + H₂O₂
Other name(s): pyruvic oxidase; phosphate-dependent pyruvate oxidase
Systematic name: pyruvate:oxygen 2-oxidoreductase (phosphorylating)
Comments: A flavoprotein (FAD) requiring thiamine diphosphate. Two reducing equivalents are transferred from the resonant carbanion/enamine forms of 2-hydroxyethyl-thiamine-diphosphate to the adjacent flavin cofactor, yielding 2-acetyl-thiamine diphosphate (AcThDP) and reduced flavin. FADH₂ is reoxidized by O₂ to yield H₂O₂ and FAD and AcThDP is cleaved phosphorolytically to acetyl phosphate and thiamine diphosphate [4297].
References: [4632, 4297]

[EC 1.2.3.3 created 1961]

EC 1.2.3.4

Accepted name: oxalate oxidase
Reaction: oxalate + O₂ + 2 H⁺ = 2 CO₂ + H₂O₂
Other name(s): aero-oxalo dehydrogenase; oxalic acid oxidase
Systematic name: oxalate:oxygen oxidoreductase
Comments: Contains Mn²⁺ as a cofactor. The enzyme is not a flavoprotein as had been thought [3502].

References: [832, 2247, 3502]

[EC 1.2.3.4 created 1961]

EC 1.2.3.5

Accepted name: glyoxylate oxidase
Reaction: glyoxylate + H₂O + O₂ = oxalate + H₂O₂
Systematic name: glyoxylate:oxygen oxidoreductase
References: [2006]

[EC 1.2.3.5 created 1972]

EC 1.2.3.6

Accepted name: pyruvate oxidase (CoA-acetylating)
Reaction: pyruvate + CoA + O₂ = acetyl-CoA + CO₂ + H₂O₂
Systematic name: pyruvate:oxygen 2-oxidoreductase (CoA-acetylating)
Comments: A flavoprotein (FAD). May be identical with EC 1.2.7.1 pyruvate synthase.
References: [3485, 4185]

[EC 1.2.3.6 created 1976]

EC 1.2.3.7

Accepted name: indole-3-acetaldehyde oxidase
Reaction: (indol-3-yl)acetaldehyde + H₂O + O₂ = (indol-3-yl)acetate + H₂O₂
Other name(s): indoleacetaldehyde oxidase; IAAld oxidase; AO1; indole-3-acetaldehyde:oxygen oxidoreductase
Systematic name: (indol-3-yl)acetaldehyde:oxygen oxidoreductase
Comments: A hemoprotein. This enzyme is an isoform of aldehyde oxidase (EC 1.2.3.1). It has a preference for aldehydes having an indole-ring structure as substrate [3793, 3798]. It may play a role in plant hormone biosynthesis as its activity is higher in the auxin-overproducing mutant, *super-root1*, than in wild-type *Arabidopsis thaliana* [3798]. While (indol-3-yl)acetaldehyde is the preferred substrate, it also oxidizes indole-3-carbaldehyde and acetaldehyde, but more slowly. The enzyme from maize contains FAD, iron and molybdenum [2238].
References: [411, 2840, 3435, 2238, 2237, 3793, 3798]

[EC 1.2.3.7 created 1984, modified 2004, modified 2006]

EC 1.2.3.8

Accepted name: pyridoxal oxidase
Reaction: pyridoxal + H₂O + O₂ = 4-pyridoxate + (?)
Systematic name: pyridoxal:oxygen 4-oxidoreductase
Comments: A molybdenum protein.
References: [1507, 4549]

[EC 1.2.3.8 created 1984]

EC 1.2.3.9

Accepted name: aryl-aldehyde oxidase
Reaction: an aromatic aldehyde + O₂ + H₂O = an aromatic carboxylate + H₂O₂
Systematic name: aryl-aldehyde:oxygen oxidoreductase
Comments: Acts on benzaldehyde, vanillin and a number of other aromatic aldehydes, but not on aliphatic aldehydes or sugars.
References: [769]

[EC 1.2.3.9 created 1986, modified 2002]

[1.2.3.10 Deleted entry. carbon-monoxide oxidase. Activity due to EC 1.2.2.4 carbon-monoxide dehydrogenase (cytochrome b-561)]

[EC 1.2.3.10 created 1990, deleted 2003]

[1.2.3.11 Deleted entry. retinal oxidase. Now included with EC 1.2.3.1, aldehyde oxidase]

[EC 1.2.3.11 created 1990, modified 2002, deleted 2011]

[1.2.3.12 Transferred entry. vanillate demethylase. Now EC 1.14.13.82, vanillate monooxygenase]

[EC 1.2.3.12 created 2000, deleted 2003]

EC 1.2.3.13

Accepted name: 4-hydroxyphenylpyruvate oxidase
Reaction: $2 \text{ 4-hydroxyphenylpyruvate} + \text{O}_2 = 2 \text{ 4-hydroxyphenylacetate} + 2 \text{ CO}_2$
Systematic name: 4-hydroxyphenylpyruvate:oxygen oxidoreductase (decarboxylating)
Comments: Involved in tyrosine degradation pathway in *Arthrobacter sp.*
References: [349]

[EC 1.2.3.13 created 2000]

EC 1.2.3.14

Accepted name: abscisic-aldehyde oxidase
Reaction: $\text{abscisic aldehyde} + \text{H}_2\text{O} + \text{O}_2 = \text{abscisate} + \text{H}_2\text{O}_2$
Other name(s): abscisic aldehyde oxidase; AAO3; AOD; AO δ
Systematic name: abscisic-aldehyde:oxygen oxidoreductase
Comments: Acts on both (+)- and (-)-abscisic aldehyde. Involved in the abscisic-acid biosynthesis pathway in plants, along with EC 1.1.1.288, (xanthoxin dehydrogenase), EC 1.13.11.51 (9-*cis*-epoxycarotenoid dioxygenase) and EC 1.14.13.93 [(+)-abscisic acid 8'-hydroxylase]. While abscisic aldehyde is the best substrate, the enzyme also acts with indole-3-aldehyde, 1-naphthaldehyde and benzaldehyde as substrates, but more slowly [3799].
References: [3629, 3800, 3799]

[EC 1.2.3.14 created 2005]

EC 1.2.3.15

Accepted name: (methyl)glyoxal oxidase
Reaction: (1) $\text{glyoxal} + \text{H}_2\text{O} + \text{O}_2 = \text{glyoxylate} + \text{H}_2\text{O}_2$
(2) $2 \text{ 2-oxopropanal} + \text{H}_2\text{O} + \text{O}_2 = \text{pyruvate} + \text{H}_2\text{O}_2$
Other name(s): glx1 (gene name); glx2 (gene name)
Systematic name: (methyl)glyoxal:oxygen oxidoreductase
Comments: The enzyme, originally characterized from the white rot fungus *Phanerochaete chrysosporium*, utilizes a free radical-coupled copper complex for catalysis.
References: [2069, 2068, 2071, 4611]

[EC 1.2.3.15 created 2016]

EC 1.2.4 With a disulfide as acceptor

EC 1.2.4.1

- Accepted name:** pyruvate dehydrogenase (acetyl-transferring)
Reaction: pyruvate + [dihydrolipoyllysine-residue acetyltransferase] lipoyllysine = [dihydrolipoyllysine-residue acetyltransferase] *S*-acetyldihydrolipoyllysine + CO₂
Other name(s): pyruvate decarboxylase (ambiguous); pyruvate dehydrogenase (ambiguous); pyruvate dehydrogenase (lipoamide); pyruvate:lipoamide 2-oxidoreductase (decarboxylating and acceptor-acetylation); pyruvic acid dehydrogenase; pyruvic dehydrogenase (ambiguous)
Systematic name: pyruvate:[dihydrolipoyllysine-residue acetyltransferase]-lipoyllysine 2-oxidoreductase (decarboxylating, acceptor-acetylation)
Comments: Contains thiamine diphosphate. It is a component (in multiple copies) of the multienzyme pyruvate dehydrogenase complex, EC 1.2.1.104, in which it is bound to a core of molecules of EC 2.3.1.12, dihydrolipoyllysine-residue acetyltransferase, which also binds multiple copies of EC 1.8.1.4, dihydrolipoyl dehydrogenase. It does not act on free lipoamide or lipoyllysine, but only on the lipoyllysine residue in EC 2.3.1.12.
References: [3129, 3775, 3293]

[EC 1.2.4.1 created 1961, modified 2003]

EC 1.2.4.2

- Accepted name:** oxoglutarate dehydrogenase (succinyl-transferring)
Reaction: 2-oxoglutarate + [dihydrolipoyllysine-residue succinyltransferase] lipoyllysine = [dihydrolipoyllysine-residue succinyltransferase] *S*-succinyldihydrolipoyllysine + CO₂
Other name(s): 2-ketoglutarate dehydrogenase; 2-oxoglutarate dehydrogenase; 2-oxoglutarate:lipoate oxidoreductase; 2-oxoglutarate:lipoamide 2-oxidoreductase (decarboxylating and acceptor-succinylation); α -ketoglutarate dehydrogenase; α -ketoglutaric acid dehydrogenase; α -ketoglutaric dehydrogenase; α -oxoglutarate dehydrogenase; AKGDH; OGDC; ketoglutaric dehydrogenase; oxoglutarate decarboxylase (misleading); oxoglutarate dehydrogenase; oxoglutarate dehydrogenase (lipoamide)
Systematic name: 2-oxoglutarate:[dihydrolipoyllysine-residue succinyltransferase]-lipoyllysine 2-oxidoreductase (decarboxylating, acceptor-succinylation)
Comments: Contains thiamine diphosphate. It is a component of the multienzyme 2-oxoglutarate dehydrogenase complex, EC 1.2.1.105, in which multiple copies of it are bound to a core of molecules of EC 2.3.1.61, dihydrolipoyllysine-residue succinyltransferase, which also binds multiple copies of EC 1.8.1.4, dihydrolipoyl dehydrogenase. It does not act on free lipoamide or lipoyllysine, but only on the lipoyllysine residue in EC 2.3.1.61.
References: [2684, 3129, 3651, 3293]

[EC 1.2.4.2 created 1961, modified 1980, modified 1986, modified 2003]

[1.2.4.3 Deleted entry. 2-oxoisocaproate dehydrogenase. Now included with EC 1.2.4.4, 3-methyl-2-oxobutanoate dehydrogenase (2-methylpropanoyl-transferring)]

[EC 1.2.4.3 created 1972, deleted 1978]

EC 1.2.4.4

- Accepted name:** 3-methyl-2-oxobutanoate dehydrogenase (2-methylpropanoyl-transferring)
Reaction: 3-methyl-2-oxobutanoate + [dihydrolipoyllysine-residue (2-methylpropanoyl)transferase] lipoyllysine = [dihydrolipoyllysine-residue (2-methylpropanoyl)transferase] *S*-(2-methylpropanoyl)dihydrolipoyllysine + CO₂

Other name(s): 2-oxoisocaproate dehydrogenase; 2-oxoisovalerate (lipoate) dehydrogenase; 3-methyl-2-oxobutanoate dehydrogenase (lipoamide); 3-methyl-2-oxobutanoate:lipoamide oxidoreductase (decarboxylating and acceptor-2-methylpropanoylating); α -keto- α -methylvalerate dehydrogenase; α -ketoisocaproate dehydrogenase; α -ketoisocaproic dehydrogenase; α -ketoisocaproic- α -keto- α -methylvaleric dehydrogenase; α -ketoisovalerate dehydrogenase; α -oxoisocaproate dehydrogenase; BCKDH (ambiguous); BCOAD; branched chain keto acid dehydrogenase; branched-chain (-2-oxoacid) dehydrogenase (BCD); branched-chain 2-keto acid dehydrogenase; branched-chain 2-oxo acid dehydrogenase; branched-chain α -keto acid dehydrogenase; branched-chain α -oxo acid dehydrogenase; branched-chain keto acid dehydrogenase; branched-chain ketoacid dehydrogenase; dehydrogenase, 2-oxoisovalerate (lipoate); dehydrogenase, branched chain α -keto acid

Systematic name: 3-methyl-2-oxobutanoate:[dihydrolipoyllysine-residue (2-methylpropanoyl)transferase]-lipoyllysine 2-oxidoreductase (decarboxylating, acceptor-2-methylpropanoylating)

Comments: Contains thiamine diphosphate. It acts not only on 3-methyl-2-oxobutanoate, but also on 4-methyl-2-oxopentanoate and (*S*)-3-methyl-2-oxopentanoate, so that it acts on the 2-oxo acids that derive from the action of transaminases on valine, leucine and isoleucine. It is a component of the multienzyme 3-methyl-2-oxobutanoate dehydrogenase complex in which multiple copies of it are bound to a core of molecules of EC 2.3.1.168, dihydrolipoyllysine-residue (2-methylpropanoyl)transferase, which also binds multiple copies of EC 1.8.1.4, dihydrolipoyl dehydrogenase. It does not act on free lipoamide or lipoyllysine, but only on the lipoyllysine residue in EC 2.3.1.168.

References: [410, 721, 827, 3309, 3293]

[EC 1.2.4.4 created 1972 (EC 1.2.4.3 created 1972, incorporated 1978), modified 2003]

EC 1.2.5 With a quinone or similar compound as acceptor

EC 1.2.5.1

Accepted name: pyruvate dehydrogenase (quinone)

Reaction: pyruvate + ubiquinone + H₂O = acetate + CO₂ + ubiquinol

Other name(s): pyruvate dehydrogenase (ambiguous); pyruvic dehydrogenase (ambiguous); pyruvic (cytochrome *b*₁) dehydrogenase (incorrect); pyruvate:ubiquinone-8-oxidoreductase; pyruvate oxidase (ambiguous); pyruvate dehydrogenase (cytochrome) (incorrect)

Systematic name: pyruvate:ubiquinone oxidoreductase

Comments: Flavoprotein (FAD) [3470]. This bacterial enzyme is located on the inner surface of the cytoplasmic membrane and coupled to the respiratory chain via ubiquinone [785, 2209]. Does not accept menaquinone. Activity is greatly enhanced by lipids [4,5,6]. Requires thiamine diphosphate [3128]. The enzyme can also form acetoin [308].

References: [3470, 785, 2209, 515, 4511, 4848, 3128, 308]

[EC 1.2.5.1 created 2010]

EC 1.2.5.2

Accepted name: aldehyde dehydrogenase (quinone)

Reaction: an aldehyde + a quinone + H₂O = a carboxylate + a quinol

Other name(s): aldehyde dehydrogenase (acceptor)

Systematic name: aldehyde:quinone oxidoreductase

Comments: Wide specificity; acts on straight-chain aldehydes up to C₁₀, aromatic aldehydes, glyoxylate and glyceraldehyde. The enzymes contains a PQQ cofactor and multiple hemes that deliver the electrons to the membrane quinone pool.

References: [78, 82, 3257, 1353]

[EC 1.2.5.2 created 1983 as EC 1.2.99.3, modified 1989, transferred 2015 to EC 1.2.5.2]

EC 1.2.5.3

- Accepted name:** aerobic carbon monoxide dehydrogenase
Reaction: $\text{CO} + \text{a quinone} + \text{H}_2\text{O} = \text{CO}_2 + \text{a quinol}$
Other name(s): MoCu-CODH; *coxSML* (gene names); molybdoenzyme carbon monoxide dehydrogenase
Systematic name: carbon-monoxide,water:quinone oxidoreductase
Comments: This enzyme, found in carboxydophilic bacteria, catalyses the oxidation of CO to CO₂ under aerobic conditions. The enzyme contains a binuclear Mo-Cu cluster in which the copper is ligated to a molybdopterin center via a sulfur bridge. The enzyme also contains two [2Fe-2S] clusters and FAD, and belongs to the xanthine oxidoreductase family. The CO₂ that is produced is assimilated by the Calvin-Benson-Basham cycle, while the electrons are transferred to a quinone via the FAD site, and continue through the electron transfer chain to a dioxygen terminal acceptor [4626]. *cf.* EC 1.2.7.4, anaerobic carbon monoxide dehydrogenase.
References: [1406, 933, 1347, 3503, 4626, 3286, 1655]

[EC 1.2.5.3 created 2016]

EC 1.2.7 With an iron-sulfur protein as acceptor

EC 1.2.7.1

- Accepted name:** pyruvate synthase
Reaction: $\text{pyruvate} + \text{CoA} + 2 \text{ oxidized ferredoxin} = \text{acetyl-CoA} + \text{CO}_2 + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
Other name(s): pyruvate oxidoreductase; pyruvate synthetase; pyruvate:ferredoxin oxidoreductase; pyruvic-ferredoxin oxidoreductase; 2-oxobutyrate synthase; α -ketobutyrate-ferredoxin oxidoreductase; 2-ketobutyrate synthase; α -ketobutyrate synthase; 2-oxobutyrate-ferredoxin oxidoreductase; 2-oxobutanoate:ferredoxin 2-oxidoreductase (CoA-propionylating); 2-oxobutanoate:ferredoxin 2-oxidoreductase (CoA-propanoylating)
Systematic name: pyruvate:ferredoxin 2-oxidoreductase (CoA-acetylating)
Comments: Contains thiamine diphosphate and [4Fe-4S] clusters. The enzyme also decarboxylates 2-oxobutyrate with lower efficiency, but shows no activity with 2-oxoglutarate. This enzyme is a member of the 2-oxoacid oxidoreductases, a family of enzymes that oxidatively decarboxylate different 2-oxoacids to form their CoA derivatives, and are differentiated based on their substrate specificity. For examples of other members of this family, see EC 1.2.7.3, 2-oxoglutarate synthase and EC 1.2.7.7, 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin).
References: [1070, 1295, 4382, 4383, 610]

[EC 1.2.7.1 created 1972, modified 2003, modified 2013]

[1.2.7.2 Deleted entry. 2-oxobutyrate synthase. Now included with EC 1.2.7.1, pyruvate synthase.]

[EC 1.2.7.2 created 1972, deleted 2013]

EC 1.2.7.3

- Accepted name:** 2-oxoglutarate synthase
Reaction: $2\text{-oxoglutarate} + \text{CoA} + 2 \text{ oxidized ferredoxin} = \text{succinyl-CoA} + \text{CO}_2 + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
Other name(s): 2-ketoglutarate ferredoxin oxidoreductase; 2-oxoglutarate:ferredoxin oxidoreductase; KGOR; 2-oxoglutarate ferredoxin oxidoreductase; 2-oxoglutarate:ferredoxin 2-oxidoreductase (CoA-succinylating)
Systematic name: 2-oxoglutarate:ferredoxin oxidoreductase (decarboxylating)
Comments: The enzyme contains thiamine diphosphate and two [4Fe-4S] clusters. Highly specific for 2-oxoglutarate. This enzyme is a member of the 2-oxoacid oxidoreductases, a family of enzymes that oxidatively decarboxylate different 2-oxoacids to form their CoA derivatives, and are differentiated based on their substrate specificity. For examples of other members of this family, see EC 1.2.7.1, pyruvate synthase and EC 1.2.7.7, 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin).
References: [485, 1295, 952, 2620, 3763]

[EC 1.2.7.3 created 1972, modified 2005]

EC 1.2.7.4

- Accepted name:** anaerobic carbon monoxide dehydrogenase
Reaction: $\text{CO} + \text{H}_2\text{O} + 2 \text{ oxidized ferredoxin} = \text{CO}_2 + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
Other name(s): Ni-CODH; carbon-monoxide dehydrogenase (ferredoxin)
Systematic name: carbon-monoxide,water:ferredoxin oxidoreductase
Comments: This prokaryotic enzyme catalyses the reversible reduction of CO_2 to CO. The electrons are transferred to redox proteins such as ferredoxin. In purple sulfur bacteria and methanogenic archaea it catalyses the oxidation of CO to CO_2 , which is incorporated by the Calvin-Benson-Basham cycle or released, respectively. In acetogenic and sulfate-reducing microbes it catalyses the reduction of CO_2 to CO, which is incorporated into acetyl CoA by EC 2.3.1.169, CO-methylating acetyl-CoA synthase, with which the enzyme forms a tight complex in those organisms. The enzyme contains five metal clusters per homodimeric enzyme: two nickel-iron-sulfur clusters called the C-Clusters, one [4Fe-4S] D-cluster; and two [4Fe-4S] B-clusters. In methanogenic archaea additional [4Fe-4S] clusters exist, presumably as part of the electron transfer chain. In purple sulfur bacteria the enzyme forms complexes with the Ni-Fe-S protein EC 1.12.7.2, ferredoxin hydrogenase, which catalyse the overall reaction: $\text{CO} + \text{H}_2\text{O} = \text{CO}_2 + \text{H}_2$. *cf.* EC 1.2.5.3, aerobic carbon monoxide dehydrogenase.
References: [3424, 907, 381, 964, 934, 959, 542]

[EC 1.2.7.4 created 2003 (EC 1.2.99.2 created 1982, modified 1990, modified 2003, incorporated 2015), modified 2016]

EC 1.2.7.5

- Accepted name:** aldehyde ferredoxin oxidoreductase
Reaction: an aldehyde + $\text{H}_2\text{O} + 2 \text{ oxidized ferredoxin} = \text{a carboxylate} + 2 \text{ H}^+ + 2 \text{ reduced ferredoxin}$
Other name(s): AOR
Systematic name: aldehyde:ferredoxin oxidoreductase
Comments: This is an oxygen-sensitive enzyme that contains tungsten-molybdopterin and iron-sulfur clusters. Catalyses the oxidation of aldehydes (including crotonaldehyde, acetaldehyde, formaldehyde and glyceraldehyde) to their corresponding acids. However, it does not oxidize glyceraldehyde 3-phosphate [see EC 1.2.7.6, glyceraldehyde-3-phosphate dehydrogenase (ferredoxin)]. Can use ferredoxin or methyl viologen but not NAD(P)^+ as electron acceptor.
References: [2920, 1934, 593, 3585]

[EC 1.2.7.5 created 2003]

EC 1.2.7.6

- Accepted name:** glyceraldehyde-3-phosphate dehydrogenase (ferredoxin)
Reaction: $\text{D-glyceraldehyde-3-phosphate} + \text{H}_2\text{O} + 2 \text{ oxidized ferredoxin} = 3\text{-phospho-D-glycerate} + 2 \text{ H}^+ + 2 \text{ reduced ferredoxin}$
Other name(s): GAPOR; glyceraldehyde-3-phosphate Fd oxidoreductase; glyceraldehyde-3-phosphate ferredoxin reductase
Systematic name: D-glyceraldehyde-3-phosphate:ferredoxin oxidoreductase
Comments: Contains tungsten-molybdopterin and iron-sulfur clusters. This enzyme is thought to function in place of glyceraldehyde-3-phosphate dehydrogenase and possibly phosphoglycerate kinase in the novel Embden-Meyerhof-type glycolytic pathway found in *Pyrococcus furiosus* [2921]. It is specific for glyceraldehyde-3-phosphate.
References: [2921, 3585]

[EC 1.2.7.6 created 2003]

EC 1.2.7.7

- Accepted name:** 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin)

Reaction: 3-methyl-2-oxobutanoate + CoA + 2 oxidized ferredoxin = S-(2-methylpropanoyl)-CoA + CO₂ + 2 reduced ferredoxin + H⁺

Other name(s): 2-ketoisovalerate ferredoxin reductase; 3-methyl-2-oxobutanoate synthase (ferredoxin); VOR; branched-chain ketoacid ferredoxin reductase; branched-chain oxo acid ferredoxin reductase; ketovaline-ferredoxin oxidoreductase; ketoisovalerate ferredoxin reductase; 2-oxoisovalerate ferredoxin reductase

Systematic name: 3-methyl-2-oxobutanoate:ferredoxin oxidoreductase (decarboxylating; CoA-2-methylpropanoylating)

Comments: The enzyme is CoA-dependent and contains thiamine diphosphate and iron-sulfur clusters. Preferentially utilizes 2-oxo-acid derivatives of branched chain amino acids, e.g. 3-methyl-2-oxopentanoate, 4-methyl-2-oxo-pentanoate, and 2-oxobutanoate. This enzyme is a member of the 2-oxoacid oxidoreductases, a family of enzymes that reversibly catalyse the oxidative decarboxylation of different 2-oxoacids to form their CoA derivatives, and are differentiated based on their substrate specificity. For examples of other members of this family, see EC 1.2.7.1, pyruvate synthase, and EC 1.2.7.3, 2-oxoglutarate synthase.

References: [483, 1607, 4248, 3763]

[EC 1.2.7.7 created 2003]

EC 1.2.7.8

Accepted name: indolepyruvate ferredoxin oxidoreductase

Reaction: (indol-3-yl)pyruvate + CoA + 2 oxidized ferredoxin = S-2-(indol-3-yl)acetyl-CoA + CO₂ + 2 reduced ferredoxin + H⁺

Other name(s): 3-(indol-3-yl)pyruvate synthase (ferredoxin); IOR

Systematic name: 3-(indol-3-yl)pyruvate:ferredoxin oxidoreductase (decarboxylating, CoA-indole-acetyllating)

Comments: Contains thiamine diphosphate and [4Fe-4S] clusters. Preferentially utilizes the transaminated forms of aromatic amino acids and can use phenylpyruvate and *p*-hydroxyphenylpyruvate as substrates. This enzyme, which is found in archaea, is a member of the 2-oxoacid oxidoreductases, a family of enzymes that oxidatively decarboxylate different 2-oxoacids to form their CoA derivatives, and are differentiated based on their substrate specificity. For examples of other members of this family, see EC 1.2.7.3, 2-oxoglutarate synthase and EC 1.2.7.7, 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin).

References: [2621, 3898, 4248, 3763]

[EC 1.2.7.8 created 2003]

[1.2.7.9 Deleted entry. 2-oxoglutarate ferredoxin oxidoreductase. This enzyme is identical to EC 1.2.7.3, 2-oxoglutarate synthase]

[EC 1.2.7.9 created 2003, deleted 2005]

EC 1.2.7.10

Accepted name: oxalate oxidoreductase

Reaction: oxalate + oxidized ferredoxin = 2 CO₂ + reduced ferredoxin

Systematic name: oxalate:ferredoxin oxidoreductase

Comments: Contains thiamine diphosphate and [4Fe-4S] clusters. Acceptors include ferredoxin and the nickel-dependent carbon monoxide dehydrogenase (EC 1.2.7.4)

References: [825, 3319]

[EC 1.2.7.10 created 2011]

EC 1.2.7.11

Accepted name: 2-oxoacid oxidoreductase (ferredoxin)

Reaction: a 2-oxocarboxylate + CoA + 2 oxidized ferredoxin = an acyl-CoA + CO₂ + 2 reduced ferredoxin + 2 H⁺

Other name(s): OFOR
Systematic name: 2-oxocarboxylate:ferredoxin 2-oxidoreductase (decarboxylating, CoA-acylating)
Comments: Contains thiamine diphosphate and [4Fe-4S] clusters [4884]. This enzyme is a member of the 2-oxoacid oxidoreductases, a family of enzymes that oxidatively decarboxylate different 2-oxoacids to form their CoA derivatives, and are differentiated based on their substrate specificity. For example, see EC 1.2.7.3, 2-oxoglutarate synthase and EC 1.2.7.7, 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin).
References: [2066, 4884, 1219, 1220, 3088, 3240]

[EC 1.2.7.11 created 2013]

EC 1.2.7.12

Accepted name: formylmethanofuran dehydrogenase
Reaction: a formylmethanofuran + H₂O + 2 oxidized ferredoxin [iron-sulfur] cluster = CO₂ + a methanofuran + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺
Other name(s): formylmethanofuran:acceptor oxidoreductase
Systematic name: formylmethanofuran:ferredoxin oxidoreductase
Comments: Contains a molybdopterin cofactor and numerous [4Fe-4S] clusters. In some organisms an additional subunit enables the incorporation of tungsten when molybdenum availability is low. The enzyme catalyses a reversible reaction in methanogenic archaea, and is involved in methanogenesis from CO₂ as well as the oxidation of coenzyme M to CO₂. The reaction is endergonic, and is driven by coupling with the soluble CoB-CoM heterodisulfide reductase via electron bifurcation.
References: [1999, 316, 315, 4474, 2783, 2007, 4489]

[EC 1.2.7.12 created 1992 as EC 1.2.99.5, transferred 2017 to EC 1.2.7.12]

EC 1.2.98 With other, known, physiological acceptors

EC 1.2.98.1

Accepted name: formaldehyde dismutase
Reaction: 2 formaldehyde + H₂O = formate + methanol
Other name(s): aldehyde dismutase; cannizzanase; nicotinoprotein aldehyde dismutase
Systematic name: formaldehyde:formaldehyde oxidoreductase
Comments: The enzyme contains a tightly but noncovalently bound NADP(H) cofactor, as well as Zn²⁺ and Mg²⁺. Enzyme-bound NADPH formed by oxidation of formaldehyde to formate is oxidized back to NADP⁺ by reaction with a second formaldehyde, yielding methanol. The enzyme from the bacterium *Mycobacterium* sp. DSM 3803 also catalyses the reactions of EC 1.1.99.36, alcohol dehydrogenase (nicotinoprotein) and EC 1.1.99.37, methanol dehydrogenase (nicotinoprotein) [3239]. Formaldehyde and acetaldehyde can act as donors; formaldehyde, acetaldehyde and propanal can act as acceptors [2018, 2021].
References: [2018, 2021, 3239]

[EC 1.2.98.1 created 1986 as EC 1.2.99.4, modified 2012, transferred 2015 to EC 1.2.98.1]

EC 1.2.99 With unknown physiological acceptors

[1.2.99.1 *Transferred entry. uracil dehydrogenase. Now EC 1.17.99.4, uracil/thymine dehydrogenase*]

[EC 1.2.99.1 created 1961, deleted 1984]

[1.2.99.2 *Transferred entry. carbon-monoxide dehydrogenase (acceptor). Now EC 1.2.7.4, carbon-monoxide dehydrogenase (ferredoxin)]*

[EC 1.2.99.2 created 1982, modified 1990, modified 2003, deleted 2016]

[1.2.99.3 Transferred entry. aldehyde dehydrogenase (pyrroloquinoline-quinone). Now EC 1.2.5.2, aldehyde dehydrogenase (quinone)]

[EC 1.2.99.3 created 1983, modified 1989, deleted 2015]

[1.2.99.4 Transferred entry. formaldehyde dismutase. Now EC 1.2.98.1, formaldehyde dismutase.]

[EC 1.2.99.4 created 1986, modified 2012, deleted 2015]

[1.2.99.5 Transferred entry. formylmethanofuran dehydrogenase. Now EC 1.2.7.12, formylmethanofuran dehydrogenase]

[EC 1.2.99.5 created 1992, deleted 2017]

EC 1.2.99.6

Accepted name: carboxylate reductase
Reaction: an aldehyde + acceptor + H₂O = a carboxylate + reduced acceptor
Other name(s): aldehyde:(acceptor) oxidoreductase
Systematic name: aldehyde:acceptor oxidoreductase
Comments: A tungsten protein. Methyl viologen can act as acceptor. In the reverse direction, non-activated acids are reduced by reduced viologens to aldehydes, but not to the corresponding alcohols.
References: [4603]

[EC 1.2.99.6 created 1992]

EC 1.2.99.7

Accepted name: aldehyde dehydrogenase (FAD-independent)
Reaction: an aldehyde + H₂O + acceptor = a carboxylate + reduced acceptor
Other name(s): aldehyde oxidase; aldehyde oxidoreductase; Mop; AORDd
Systematic name: aldehyde:acceptor oxidoreductase (FAD-independent)
Comments: Belongs to the xanthine oxidase family of enzymes. The enzyme from *Desulfovibrio* sp. contains a molybdenum-molybdopterin-cytosine dinucleotide (MCD) complex and two types of [2Fe-2S] cluster per monomer, but does not contain FAD.
References: [4359, 975, 97, 3565]

[EC 1.2.99.7 created 2004]

EC 1.2.99.8

Accepted name: glyceraldehyde dehydrogenase (FAD-containing)
Reaction: D-glyceraldehyde + H₂O + acceptor = D-glycerate + reduced acceptor
Other name(s): glyceraldehyde oxidoreductase
Systematic name: D-glyceraldehyde:acceptor oxidoreductase (FAD-containing)
Comments: The enzyme from the archaeon *Sulfolobus acidocaldarius* catalyses the oxidation of D-glyceraldehyde in the nonphosphorylative Entner-Doudoroff pathway. With 2,6-dichlorophenolindophenol as artificial electron acceptor, the enzyme shows a broad substrate range, but is most active with D-glyceraldehyde. It is not known which acceptor is utilized *in vivo*. The iron-sulfur protein contains FAD and molybdopterin guanine dinucleotide.
References: [1993]

[EC 1.2.99.8 created 2013]

[1.2.99.9 Transferred entry. formate dehydrogenase (coenzyme F₄₂₀). Now EC 1.17.98.3, formate dehydrogenase (coenzyme F₄₂₀)]

[EC 1.2.99.9 created 2014, deleted 2017]

EC 1.2.99.10

- Accepted name:** 4,4'-diapolycopenoate synthase
- Reaction:** (1) 4,4'-diapolycopene-4-al + H₂O + acceptor = 4,4'-diapolycopene-4-oate + reduced acceptor
(2) 4,4'-diapolycopene-4,4'-dial + 2 H₂O + 2 acceptor = 4,4'-diapolycopene-4,4'-dioate + 2 reduced acceptor
- Other name(s):** *crtNc*; 4,4'-diapolycopenealdehyde oxidase (misleading)
- Systematic name:** 4,4'-diapolycopene-4-al,donor:oxygen oxidoreductase (4,4'-diapolycopene-4-oate-forming)
- Comments:** The enzyme has been described from the bacteria *Methylomonas* sp. 16a and *Bacillus indicus*.
- References:** [4215, 4016]

[EC 1.2.99.10 created 2017]

EC 1.3 Acting on the CH-CH group of donors

This subclass contains enzymes that introduce a double-bond into the substrate by direct dehydrogenation at a carbon-carbon single bond. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.3.1), a cytochrome (EC 1.3.2), oxygen (EC 1.3.3), a quinone or related compound (EC 1.3.5), an iron-sulfur protein (EC 1.3.7), a flavin (EC 1.3.8) or some other acceptor (EC 1.3.99).

EC 1.3.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.3.1.1

- Accepted name:** dihydropyrimidine dehydrogenase (NAD⁺)
- Reaction:** (1) 5,6-dihydrouracil + NAD⁺ = uracil + NADH + H⁺
(2) 5,6-dihydrothymine + NAD⁺ = thymine + NADH + H⁺
- Other name(s):** dihydropyrimidine dehydrogenase; dihydrothymine dehydrogenase; pyrimidine reductase; thymine reductase; uracil reductase; dihydrouracil dehydrogenase (NAD⁺)
- Systematic name:** 5,6-dihydropyrimidine:NAD⁺ oxidoreductase
- Comments:** An iron-sulfur flavoenzyme. The enzyme was originally discovered in the uracil-fermenting bacterium, *Clostridium uracilicum*, which utilizes uracil and thymine as nitrogen and carbon sources for growth [540]. Since then the enzyme was found in additional organisms including *Alcaligenes eutrophus* [3730], *Pseudomonas* strains [2104, 4594] and *Escherichia coli* [4593, 1642].
- References:** [540, 3730, 2104, 4594, 4593, 1642]

[EC 1.3.1.1 created 1961, modified 2011]

EC 1.3.1.2

- Accepted name:** dihydropyrimidine dehydrogenase (NADP⁺)
- Reaction:** 5,6-dihydrouracil + NADP⁺ = uracil + NADPH + H⁺
- Other name(s):** dihydrothymine dehydrogenase; dihydrouracil dehydrogenase (NADP⁺); 4,5-dihydrothymine: oxidoreductase; DPD; DHPDH; dehydrogenase, dihydrouracil (nicotinamide adenine dinucleotide phosphate); DHU dehydrogenase; hydropyrimidine dehydrogenase; dihydropyrimidine dehydrogenase (NADP)
- Systematic name:** 5,6-dihydrouracil:NADP⁺ 5-oxidoreductase
- Comments:** Also acts on dihydrothymine.
- References:** [1192, 3888]

[EC 1.3.1.2 created 1961, modified 1986]

EC 1.3.1.3

- Accepted name:** Δ⁴-3-oxosteroid 5β-reductase

Reaction: a 3-oxo-5 β -steroid + NADP⁺ = a 3-oxo-Delta⁴-steroid + NADPH + H⁺
Other name(s): 3-oxo-Delta⁴-steroid 5 β -reductase; 5 β -reductase; androstenedione 5 β -reductase; cholestenone 5 β -reductase; cortisone 5 β -reductase; cortisone β -reductase; cortisone Δ^4 -5 β -reductase; steroid 5 β -reductase; testosterone 5 β -reductase; Δ^4 -3-ketosteroid 5 β -reductase; Δ^4 -5 β -reductase; Δ^4 -hydrogenase; 4,5 β -dihydrocortisone:NADP⁺ Δ^4 -oxidoreductase; 3-oxo-5 β -steroid:NADP⁺ Δ^4 -oxidoreductase
Systematic name: 3-oxo-5 β -steroid:NADP⁺ 4,5-oxidoreductase
Comments: The enzyme from human efficiently catalyses the reduction of progesterone, androstenedione, 17 α -hydroxyprogesterone and testosterone to 5 β -reduced metabolites; it can also act on aldosterone, corticosterone and cortisol, but to a lesser extent [605]. The bile acid intermediates 7 α ,12 α -dihydroxy-4-cholesten-3-one and 7 α -hydroxy-4-cholesten-3-one can also act as substrates [2219].
References: [1138, 463, 2434, 4305, 4098, 1236, 3163, 605, 2219]

[EC 1.3.1.3 created 1961 (EC 1.3.1.23 created 1972, incorporated 2005), modified 2005]

[1.3.1.4 *Transferred entry. EC 1.3.1.4, cortisone α -reductase, transferred to EC 1.3.1.22, 3-oxo-5 α -steroid 4-dehydrogenase (NADP⁺)*]

[EC 1.3.1.4 created 1965, deleted 2012]

EC 1.3.1.5

Accepted name: cucurbitacin Δ^{23} -reductase
Reaction: 23,24-dihydrocucurbitacin B + NAD(P)⁺ = cucurbitacin B + NAD(P)H + H⁺
Other name(s): NAD(P)H: cucurbitacin B Δ^{23} -oxidoreductase
Systematic name: 23,24-dihydrocucurbitacin:NAD(P)⁺ Δ^{23} -oxidoreductase
Comments: Requires Mn²⁺. Fe²⁺ or Zn²⁺ can replace Mn²⁺ to some extent.
References: [3692, 3694]

[EC 1.3.1.5 created 1965, modified 2011]

EC 1.3.1.6

Accepted name: fumarate reductase (NADH)
Reaction: succinate + NAD⁺ = fumarate + NADH + H⁺
Other name(s): NADH-fumarate reductase; NADH-dependent fumarate reductase; fumarate reductase (NADH₂)
Systematic name: succinate:NAD⁺ oxidoreductase
References: [1712]

[EC 1.3.1.6 created 1972]

EC 1.3.1.7

Accepted name: *meso*-tartrate dehydrogenase
Reaction: *meso*-tartrate + NAD⁺ = dihydroxyfumarate + NADH + H⁺
Systematic name: *meso*-tartrate:NAD⁺ oxidoreductase
References: [2199]

[EC 1.3.1.7 created 1972]

EC 1.3.1.8

Accepted name: acyl-CoA dehydrogenase (NADP⁺)
Reaction: acyl-CoA + NADP⁺ = 2,3-dehydroacyl-CoA + NADPH + H⁺
Other name(s): 2-enoyl-CoA reductase; dehydrogenase, acyl coenzyme A (nicotinamide adenine dinucleotide phosphate); enoyl coenzyme A reductase; crotonyl coenzyme A reductase; crotonyl-CoA reductase; acyl-CoA dehydrogenase (NADP⁺)

Systematic name: acyl-CoA:NADP⁺ 2-oxidoreductase
Comments: The liver enzyme acts on enoyl-CoA derivatives of carbon chain length 4 to 16, with optimum activity on 2-hexenoyl-CoA. In *Escherichia coli*, *cis*-specific and *trans*-specific enzymes exist [EC 1.3.1.37 *cis*-2-enoyl-CoA reductase (NADPH) and EC 1.3.1.38 *trans*-2-enoyl-CoA reductase (NADPH)].
References: [945, 3806]

[EC 1.3.1.8 created 1972, modified 1986]

EC 1.3.1.9

Accepted name: enoyl-[acyl-carrier-protein] reductase (NADH)
Reaction: an acyl-[acyl-carrier protein] + NAD⁺ = a *trans*-2,3-dehydroacyl-[acyl-carrier protein] + NADH + H⁺
Other name(s): enoyl-[acyl carrier protein] reductase; enoyl-ACP reductase; NADH-enoyl acyl carrier protein reductase; NADH-specific enoyl-ACP reductase; acyl-[acyl-carrier-protein]:NAD⁺ oxidoreductase; *fabI* (gene name)
Systematic name: acyl-[acyl-carrier protein]:NAD⁺ oxidoreductase
Comments: The enzyme catalyses an essential step in fatty acid biosynthesis, the reduction of the 2,3-double bond in enoyl-acyl-[acyl-carrier-protein] derivatives of the elongating fatty acid moiety. The enzyme from the bacterium *Escherichia coli* accepts substrates with carbon chain length from 4 to 18 [4838]. The FAS-I enzyme from the bacterium *Mycobacterium tuberculosis* prefers substrates with carbon chain length from 12 to 24 carbons.
References: [3866, 4569, 4838]

[EC 1.3.1.9 created 1972, modified 2013]

EC 1.3.1.10

Accepted name: enoyl-[acyl-carrier-protein] reductase (NADPH, *Si*-specific)
Reaction: an acyl-[acyl-carrier protein] + NADP⁺ = a *trans*-2,3-dehydroacyl-[acyl-carrier protein] + NADPH + H⁺
Other name(s): acyl-ACP dehydrogenase (ambiguous); enoyl-[acyl carrier protein] (reduced nicotinamide adenine dinucleotide phosphate) reductase; NADPH 2-enoyl Co A reductase; enoyl acyl-carrier-protein reductase (ambiguous); enoyl-ACP reductase (ambiguous); acyl-[acyl-carrier-protein]:NADP⁺ oxidoreductase (B-specific); acyl-[acyl-carrier protein]:NADP⁺ oxidoreductase (B-specific); enoyl-[acyl-carrier-protein] reductase (NADPH, B-specific)
Systematic name: acyl-[acyl-carrier protein]:NADP⁺ oxidoreductase (*Si*-specific)
Comments: One of the activities of EC 2.3.1.86, fatty-acyl-CoA synthase system, an enzyme found in yeasts (Ascomycota and Basidiomycota). Catalyses the reduction of enoyl-acyl-[acyl-carrier protein] derivatives of carbon chain length from 4 to 16. The yeast enzyme is *Si*-specific with respect to NADP⁺. *cf.* EC 1.3.1.39, enoyl-[acyl-carrier-protein] reductase (NADPH, *Re*-specific) and EC 1.3.1.104, enoyl-[acyl-carrier-protein] reductase (NADPH), which describes enzymes whose stereo-specificity towards NADPH is not known. See also EC 1.3.1.9, enoyl-[acyl-carrier-protein] reductase (NADH).
References: [3813]

[EC 1.3.1.10 created 1972, modified 1986, modified 2013, modified 2014, modified 2018]

EC 1.3.1.11

Accepted name: 2-coumarate reductase
Reaction: 3-(2-hydroxyphenyl)propanoate + NAD⁺ = 2-coumarate + NADH + H⁺
Other name(s): melilotate dehydrogenase
Systematic name: 3-(2-hydroxyphenyl)propanoate:NAD⁺ oxidoreductase
References: [2433]

[EC 1.3.1.11 created 1972]

EC 1.3.1.12

- Accepted name:** prephenate dehydrogenase
Reaction: prephenate + NAD⁺ = 4-hydroxyphenylpyruvate + CO₂ + NADH
Other name(s): hydroxyphenylpyruvate synthase; chorismate mutase—prephenate dehydrogenase
Systematic name: prephenate:NAD⁺ oxidoreductase (decarboxylating)
Comments: This enzyme in the enteric bacteria also possesses chorismate mutase activity (EC 5.4.99.5 chorismate mutase) and converts chorismate into prephenate.
References: [2179]

[EC 1.3.1.12 created 1972]

EC 1.3.1.13

- Accepted name:** prephenate dehydrogenase (NADP⁺)
Reaction: prephenate + NADP⁺ = 4-hydroxyphenylpyruvate + CO₂ + NADPH
Other name(s): prephenate dehydrogenase (ambiguous); prephenate (nicotinamide adenine dinucleotide phosphate) dehydrogenase; prephenate dehydrogenase (NADP)
Systematic name: prephenate:NADP⁺ oxidoreductase (decarboxylating)
References: [1267]

[EC 1.3.1.13 created 1972]

EC 1.3.1.14

- Accepted name:** dihydroorotate dehydrogenase (NAD⁺)
Reaction: (S)-dihydroorotate + NAD⁺ = orotate + NADH + H⁺
Other name(s): orotate reductase (NADH); orotate reductase (NADH₂); DHODehase (ambiguous); DHOD (ambiguous); DHODase (ambiguous); dihydroorotate oxidase, *pyrD* (gene name)
Systematic name: (S)-dihydroorotate:NAD⁺ oxidoreductase
Comments: Binds FMN, FAD and a [2Fe-2S] cluster. The enzyme consists of two subunits, an FMN binding catalytic subunit and a FAD and iron-sulfur binding electron transfer subunit [3067]. The reaction, which takes place in the cytosol, is the only redox reaction in the *de-novo* biosynthesis of pyrimidine nucleotides. Other class 1 dihydroorotate dehydrogenases use either fumarate (EC 1.3.98.1) or NADP⁺ (EC 1.3.1.15) as electron acceptor. The membrane bound class 2 dihydroorotate dehydrogenase (EC 1.3.5.2) uses quinone as electron acceptor.
References: [1185, 1186, 2481, 3067, 3583, 1976, 2646]

[EC 1.3.1.14 created 1972, modified 2011]

EC 1.3.1.15

- Accepted name:** dihydroorotate dehydrogenase (NADP⁺)
Reaction: (S)-dihydroorotate + NADP⁺ = orotate + NADPH + H⁺
Other name(s): orotate reductase; dihydro-orotic dehydrogenase; L-5,6-dihydro-orotate:NAD⁺ oxidoreductase; orotate reductase (NADPH)
Systematic name: (S)-dihydroorotate:NADP⁺ oxidoreductase
Comments: Binds FMN and FAD [4361]. Other class 1 dihydroorotate dehydrogenases use either fumarate (EC 1.3.98.1) or NAD⁺ (EC 1.3.1.14) as electron acceptor. The membrane bound class 2 dihydroorotate dehydrogenase (EC 1.3.5.2) uses quinone as electron acceptor.
References: [4229, 4361]

[EC 1.3.1.15 created 1972, modified 2011]

EC 1.3.1.16

- Accepted name:** β-nitroacrylate reductase
Reaction: 3-nitropropanoate + NADP⁺ = 3-nitroacrylate + NADPH + H⁺

Systematic name: 3-nitropropanoate:NADP⁺ oxidoreductase
References: [3836]

[EC 1.3.1.16 created 1972]

EC 1.3.1.17

Accepted name: 3-methyleneoxindole reductase
Reaction: 3-methyl-1,3-dihydroindol-2-one + NADP⁺ = 3-methylene-1,3-dihydro-2*H*-indol-2-one + NADPH + H⁺
Other name(s): 3-methyloxindole:NADP⁺ oxidoreductase
Systematic name: 3-methyl-1,3-dihydroindol-2-one:NADP⁺ oxidoreductase
References: [2912]

[EC 1.3.1.17 created 1972]

EC 1.3.1.18

Accepted name: kynurenate-7,8-dihydrodiol dehydrogenase
Reaction: 7,8-dihydro-7,8-dihydroxykynurenate + NAD⁺ = 7,8-dihydroxykynurenate + NADH + H⁺
Other name(s): 7,8-dihydro-7,8-dihydroxykynurenate dehydrogenase; 7,8-dihydroxykynurenic acid 7,8-diol dehydrogenase
Systematic name: 7,8-dihydro-7,8-dihydroxykynurenate:NAD⁺ oxidoreductase
References: [4210]

[EC 1.3.1.18 created 1972]

EC 1.3.1.19

Accepted name: *cis*-1,2-dihydrobenzene-1,2-diol dehydrogenase
Reaction: *cis*-1,2-dihydrobenzene-1,2-diol + NAD⁺ = catechol + NADH + H⁺
Other name(s): *cis*-benzene glycol dehydrogenase; *cis*-1,2-dihydrocyclohexa-3,5-diene (nicotinamide adenine dinucleotide) oxidoreductase;
Systematic name: *cis*-1,2-dihydrobenzene-1,2-diol:NAD⁺ oxidoreductase
References: [163, 1321]

[EC 1.3.1.19 created 1972]

EC 1.3.1.20

Accepted name: *trans*-1,2-dihydrobenzene-1,2-diol dehydrogenase
Reaction: *trans*-1,2-dihydrobenzene-1,2-diol + NADP⁺ = catechol + NADPH + H⁺
Other name(s): dihydrodiol dehydrogenase
Systematic name: *trans*-1,2-dihydrobenzene-1,2-diol:NADP⁺ oxidoreductase
References: [165]

[EC 1.3.1.20 created 1972]

EC 1.3.1.21

Accepted name: 7-dehydrocholesterol reductase
Reaction: cholesterol + NADP⁺ = cholesta-5,7-dien-3β-ol + NADPH + H⁺
Other name(s): DHCR7 (gene name); 7-DHC reductase; 7-dehydrocholesterol dehydrogenase/cholesterol oxidase; Δ⁷-sterol reductase
Systematic name: cholesterol:NADP⁺ Δ⁷-oxidoreductase
Comments: The enzyme is part of the cholesterol biosynthesis pathway.
References: [876, 2855]

[EC 1.3.1.21 created 1972, modified 2013]

EC 1.3.1.22

- Accepted name:** 3-oxo-5 α -steroid 4-dehydrogenase (NADP⁺)
Reaction: a 3-oxo-5 α -steroid + NADP⁺ = a 3-oxo- Δ^4 -steroid + NADPH + H⁺
Other name(s): cholestenone 5 α -reductase; testosterone Δ^4 -5 α -reductase; steroid 5 α -reductase; 3-oxosteroid Δ^4 -dehydrogenase; 5 α -reductase; steroid 5 α -hydrogenase; 3-oxosteroid 5 α -reductase; testosterone Δ^4 -hydrogenase; 4-ene-3-oxosteroid 5 α -reductase; reduced nicotinamide adenine dinucleotide phosphate: Δ^4 -3-ketosteroid 5 α -oxidoreductase; 4-ene-5 α -reductase; Δ^4 -3-ketosteroid 5 α -oxidoreductase; cholest-4-en-3-one 5 α -reductase; testosterone 5 α -reductase; 3-oxo-5 α -steroid 4-dehydrogenase
Systematic name: 3-oxo-5 α -steroid:NADP⁺ Δ^4 -oxidoreductase
Comments: The enzyme catalyses the conversion of assorted 3-oxo- Δ^4 steroids into their corresponding 5 α form. Substrates for the mammalian enzyme include testosterone, progesterone, and corticosterone. Substrates for the plant enzyme are brassinosteroids such as campest-4-en-3-one and (22 α)-hydroxy-campest-4-en-3-one. *cf.* EC 1.3.99.5, 3-oxo-5 α -steroid 4-dehydrogenase (acceptor).
References: [2435, 3837, 646, 3664, 3416, 3345, 2444, 3571]

[EC 1.3.1.22 created 1972, modified 2012]

[1.3.1.23 Deleted entry. cholestenone β -reductase. The enzyme is identical to EC 1.3.1.3, Δ^4 -3-oxosteroid 5 β -reductase]

[EC 1.3.1.23 created 1972, deleted 2005]

EC 1.3.1.24

- Accepted name:** biliverdin reductase
Reaction: bilirubin + NAD(P)⁺ = biliverdin + NAD(P)H + H⁺
Systematic name: bilirubin:NAD(P)⁺ oxidoreductase
References: [3914]

[EC 1.3.1.24 created 1972]

EC 1.3.1.25

- Accepted name:** 1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate dehydrogenase
Reaction: (1R,6S)-1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate + NAD⁺ = catechol + CO₂ + NADH + H⁺
Other name(s): 3,5-cyclohexadiene-1,2-diol-1-carboxylate dehydrogenase; 3,5-cyclohexadiene-1,2-diol-1-carboxylic acid dehydrogenase; dihydrodihydroxybenzoate dehydrogenase; DHBBDH; *cis*-1,2-dihydroxycyclohexa-3,5-diene-1-carboxylate dehydrogenase; 2-hydro-1,2-dihydroxybenzoate dehydrogenase; *cis*-1,2-dihydroxycyclohexa-3,5-diene-1-carboxylate:NAD⁺ oxidoreductase; dihydrodihydroxybenzoate dehydrogenase; (1R,6R)-1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate:NAD⁺ oxidoreductase (decarboxylating)
Systematic name: (1R,6S)-1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate:NAD⁺ oxidoreductase (decarboxylating)
References: [3490, 3038]

[EC 1.3.1.25 created 1976, modified 2004 (EC 1.3.1.55 created 1999, incorporated 2004)]

[1.3.1.26 Transferred entry. dihydrodipicolinate reductase. Now EC 1.17.1.8, 4-hydroxy-tetrahydrodipicolinate reductase.]

[EC 1.3.1.26 created 1976, modified 2011, deleted 2013]

EC 1.3.1.27

- Accepted name:** 2-hexadecenal reductase
Reaction: hexadecanal + NADP⁺ = 2-*trans*-hexadecenal + NADPH + H⁺
Other name(s): 2-alkenal reductase; hexadecanal: NADP⁺ oxidoreductase

Systematic name: hexadecanal:NADP⁺ Δ²-oxidoreductase
Comments: Specific for long chain 2-*trans*- and 2-*cis*-alkenals, with chain length optimum around 14 to 16 carbon atoms.
References: [4037]

[EC 1.3.1.27 created 1976]

EC 1.3.1.28

Accepted name: 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase
Reaction: (2*S*,3*S*)-2,3-dihydro-2,3-dihydroxybenzoate + NAD⁺ = 2,3-dihydroxybenzoate + NADH + H⁺
Other name(s): 2,3-DHB dehydrogenase; 2,3-dihydro-2,3-dihydroxybenzoate:NAD⁺ oxidoreductase
Systematic name: (2*S*,3*S*)-2,3-dihydro-2,3-dihydroxybenzoate:NAD⁺ oxidoreductase
References: [4828]

[EC 1.3.1.28 created 1972 as EC 1.1.1.109, transferred 1976 to EC 1.3.1.28]

EC 1.3.1.29

Accepted name: *cis*-1,2-dihydro-1,2-dihydroxynaphthalene dehydrogenase
Reaction: (1*R*,2*S*)-1,2-dihydronaphthalene-1,2-diol + NAD⁺ = naphthalene-1,2-diol + NADH + H⁺
Other name(s): (+)-*cis*-naphthalene dihydrodiol dehydrogenase; naphthalene dihydrodiol dehydrogenase; *cis*-dihydrodiol naphthalene dehydrogenase; *cis*-1,2-dihydronaphthalene-1,2-diol:NAD⁺ 1,2-oxidoreductase
Systematic name: (1*R*,2*S*)-1,2-dihydronaphthalene-1,2-diol:NAD⁺ 1,2-oxidoreductase
Comments: Also acts, at half the rate, on *cis*-anthracene dihydrodiol and *cis*-phenanthrene dihydrodiol.
References: [3259]

[EC 1.3.1.29 created 1976]

[1.3.1.30 Transferred entry. EC 1.3.1.30, progesterone 5α-reductase, transferred to EC 1.3.1.22, 3-oxo-5α-steroid 4-dehydrogenase (NADP⁺).]

[EC 1.3.1.30 created 1978, deleted 2012]

EC 1.3.1.31

Accepted name: 2-enoate reductase
Reaction: butanoate + NAD⁺ = but-2-enoate + NADH + H⁺
Other name(s): enoate reductase
Systematic name: butanoate:NAD⁺ Δ²-oxidoreductase
Comments: An iron-sulfur-flavoprotein (FAD). Acts (in the reverse direction) on a wide range of alkyl and aryl αβ-unsaturated carboxylate ions; but-2-enoate was the best substrate tested.
References: [4295]

[EC 1.3.1.31 created 1982]

EC 1.3.1.32

Accepted name: maleylacetate reductase
Reaction: 3-oxoadipate + NAD(P)⁺ = 2-maleylacetate + NAD(P)H + H⁺
Other name(s): maleolylacetate reductase
Systematic name: 3-oxoadipate:NAD(P)⁺ oxidoreductase
References: [1242, 1243]

[EC 1.3.1.32 created 1983]

EC 1.3.1.33

Accepted name: protochlorophyllide reductase
Reaction: chlorophyllide *a* + NADP⁺ = protochlorophyllide + NADPH + H⁺
Other name(s): NADPH₂-protochlorophyllide oxidoreductase; NADPH-protochlorophyllide oxidoreductase; NADPH-protochlorophyllide reductase; protochlorophyllide oxidoreductase (ambiguous); protochlorophyllide photooxidoreductase; light-dependent protochlorophyllide reductase
Systematic name: chlorophyllide-*a*:NADP⁺ 7,8-oxidoreductase
Comments: The enzyme catalyses a light-dependent *trans*-reduction of the D-ring of protochlorophyllide; the product has the (7*S*,8*S*)-configuration.
References: [117, 1410]

[EC 1.3.1.33 created 1984]

EC 1.3.1.34

Accepted name: 2,4-dienoyl-CoA reductase [(2*E*)-enoyl-CoA-producing]
Reaction: (1) a (2*E*)-2-enoyl-CoA + NADP⁺ = a (2*E*,4*E*)-2,4-dienoyl-CoA + NADPH + H⁺
(2) a (2*E*)-2-enoyl-CoA + NADP⁺ = a (2*E*,4*Z*)-2,4-dienoyl-CoA + NADPH + H⁺
Other name(s): *fadH* (gene name); 4-enoyl-CoA reductase (NADPH) (ambiguous); 4-enoyl coenzyme A (reduced nicotinamide adenine dinucleotide phosphate) reductase (ambiguous); 4-enoyl-CoA reductase (ambiguous); 2,4-dienoyl-CoA reductase (NADPH) (ambiguous); *trans*-2,3-didehydroacyl-CoA:NADP⁺ 4-oxidoreductase
Systematic name: (2*E*)-2-enoyl-CoA:NADP⁺ 4-oxidoreductase
Comments: This bacterial enzyme catalyses the reduction of either (2*E*,4*E*)-2,4-dienoyl-CoA or (2*E*,4*Z*)-2,4-dienoyl-CoA to (2*E*)-2-enoyl-CoA. The enzyme from *Escherichia coli* contains FAD, FMN, and an [4Fe-4S] iron sulfur cluster. *cf.* EC 1.3.1.124, 2,4-dienoyl-CoA reductase [(3*E*)-enoyl-CoA-producing].
References: [945, 944, 4821, 1592, 2471, 1759, 4347]

[EC 1.3.1.34 created 1984, modified 1986, modified 2020]

[1.3.1.35 Transferred entry. *phosphatidylcholine desaturase*. Now EC 1.14.19.22, *microsomal oleoyl-lipid 12-desaturase*]

[EC 1.3.1.35 created 1984, deleted 2015]

EC 1.3.1.36

Accepted name: geissoschizine dehydrogenase
Reaction: geissoschizine + NADP⁺ = 4,21-didehydrogeissoschizine + NADPH
Systematic name: geissoschizine:NADP⁺ 4,21-oxidoreductase
Comments: Involved in the interconversion of heteroyohimbine alkaloids in *Catharanthus roseus*.
References: [3311]

[EC 1.3.1.36 created 1986]

EC 1.3.1.37

Accepted name: *cis*-2-enoyl-CoA reductase (NADPH)
Reaction: acyl-CoA + NADP⁺ = *cis*-2,3-dehydroacyl-CoA + NADPH + H⁺
Other name(s): NADPH-dependent *cis*-enoyl-CoA reductase; reductase, *cis*-2-enoyl coenzyme A; *cis*-2-enoyl-coenzyme A reductase; *cis*-2-enoyl-CoA reductase (NADPH)
Systematic name: acyl-CoA:NADP⁺ *cis*-2-oxidoreductase
Comments: Not identical with EC 1.3.1.38 *trans*-2-enoyl-CoA reductase (NADPH) [*cf.* EC 1.3.1.8 acyl-CoA dehydrogenase (NADP⁺)].
References: [2848]

[EC 1.3.1.37 created 1986]

EC 1.3.1.38

- Accepted name:** *trans*-2-enoyl-CoA reductase (NADPH)
Reaction: acyl-CoA + NADP⁺ = *trans*-2,3-dehydroacyl-CoA + NADPH + H⁺
Other name(s): NADPH-dependent *trans*-2-enoyl-CoA reductase; reductase, *trans*-enoyl coenzyme A; *trans*-2-enoyl-CoA reductase (NADPH₂)
Systematic name: acyl-CoA:NADP⁺ *trans*-2-oxidoreductase
Comments: Not identical with EC 1.3.1.37 *cis*-2-enoyl-CoA reductase (NADPH) [*cf.* EC 1.3.1.8 acyl-CoA dehydrogenase (NADP⁺)].
References: [2848]

[EC 1.3.1.38 created 1986]

EC 1.3.1.39

- Accepted name:** enoyl-[acyl-carrier-protein] reductase (NADPH, *Re*-specific)
Reaction: an acyl-[acyl-carrier protein] + NADP⁺ = a *trans*-2,3-dehydroacyl-[acyl-carrier protein] + NADPH + H⁺
Other name(s): acyl-ACP dehydrogenase; enoyl-[acyl carrier protein] (reduced nicotinamide adenine dinucleotide phosphate) reductase; NADPH 2-enoyl Co A reductase; enoyl-ACp reductase; enoyl-[acyl-carrier-protein] reductase (NADPH₂, A-specific); acyl-[acyl-carrier-protein]:NADP⁺ oxidoreductase (A-specific); enoyl-[acyl-carrier-protein] reductase (NADPH, A-specific); acyl-[acyl-carrier protein]:NADP⁺ oxidoreductase (A-specific)
Systematic name: acyl-[acyl-carrier protein]:NADP⁺ oxidoreductase (*Re*-specific)
Comments: This enzyme completes each cycle of fatty acid elongation by catalysing the stereospecific reduction of the double bond at position 2 of a growing fatty acid chain, while linked to an acyl-carrier protein. It is one of the activities of EC 2.3.1.85, fatty-acid synthase system. The mammalian enzyme is *Re*-specific with respect to NADP⁺. *cf.* EC 1.3.1.10, enoyl-[acyl-carrier-protein] reductase (NADPH, *Si*-specific) and EC 1.3.1.104, enoyl-[acyl-carrier-protein] reductase (NADPH).
References: [978, 558]

[EC 1.3.1.39 created 1986, modified 2013, modified 2018]

EC 1.3.1.40

- Accepted name:** 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate reductase
Reaction: 2,6-dioxo-6-phenylhexanoate + NADP⁺ = 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate + NADPH + H⁺
Other name(s): 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate (reduced nicotinamide adenine dinucleotide phosphate) reductase
Systematic name: 2,6-dioxo-6-phenylhexanoate:NADP⁺ Δ²-oxidoreductase
Comments: Broad specificity; reduces a number of compounds produced by *Pseudomonas* from aromatic hydrocarbons by ring fission.
References: [3179]

[EC 1.3.1.40 created 1989]

EC 1.3.1.41

- Accepted name:** xanthommatin reductase
Reaction: 5,12-dihydroxanthommatin + NAD⁺ = xanthommatin + NADH + H⁺
Systematic name: 5,12-dihydroxanthommatin:NAD⁺ oxidoreductase
Comments: From *Drosophila melanogaster*.
References: [3661]

[EC 1.3.1.41 created 1989]

EC 1.3.1.42

Accepted name: 12-oxophytodienoate reductase
Reaction: (9*S*,13*S*)-10,11-dihydro-12-oxo-15-phytoenoate + NADP⁺ = (15*Z*)-12-oxophyto-10,15-dienoate + NADPH + H⁺
Other name(s): 12-oxo-phytodienoic acid reductase; 8-[(1*R*,2*R*)-3-oxo-2-(*Z*)-pent-2-enylcyclopentyl]octanoate:NADP⁺ 4-oxidoreductase
Systematic name: (9*S*,13*S*)-10,11-dihydro-12-oxo-15-phytoenoate:NADP⁺ 4-oxidoreductase
Comments: Involved in the conversion of linolenate into jasmonate in *Zea mays*.
References: [4444]

[EC 1.3.1.42 created 1989]

EC 1.3.1.43

Accepted name: aroenate dehydrogenase
Reaction: L-aroenate + NAD⁺ = L-tyrosine + NADH + CO₂
Other name(s): aroenic dehydrogenase (ambiguous); cyclohexadienyl dehydrogenase (ambiguous); pretyrosine dehydrogenase (ambiguous); L-aroenate:NAD⁺ oxidoreductase; aroenate dehydrogenase (NAD⁺)
Systematic name: L-aroenate:NAD⁺ oxidoreductase (decarboxylating)
Comments: Aroenate dehydrogenases may utilize NAD⁺ (EC 1.3.1.43), NADP⁺ (EC 1.3.1.78), or both (EC 1.3.1.79). NAD⁺-specific enzymes have been reported from some bacteria [513] and plants [512]. Some enzymes also possess the activity of EC 1.3.1.12, prephenate dehydrogenase.
References: [4023, 513, 512, 2730, 2503, 4859]

[EC 1.3.1.43 created 1989, modified 2003, modified 2005, modified 2015]

EC 1.3.1.44

Accepted name: *trans*-2-enoyl-CoA reductase (NAD⁺)
Reaction: acyl-CoA + NAD⁺ = *trans*-didehydroacyl-CoA + NADH + H⁺
Other name(s): *trans*-2-enoyl-CoA reductase (NAD)
Systematic name: acyl-CoA:NAD⁺ *trans*-2-oxidoreductase
Comments: The enzyme from *Euglena gracilis* acts on crotonoyl-CoA and, more slowly, on *trans*-hex-2-enoyl-CoA and *trans*-oct-2-enoyl-CoA.
References: [1819]

[EC 1.3.1.44 created 1989]

EC 1.3.1.45

Accepted name: 2'-hydroxyisoflavone reductase
Reaction: vestitone + NADP⁺ = 2'-hydroxyformononetin + NADPH + H⁺
Other name(s): NADPH:2'-hydroxyisoflavone oxidoreductase; isoflavone reductase; 2',7-dihydroxy-4',5'-methylenedioxyisoflavone reductase
Systematic name: vestitone:NADP⁺ oxidoreductase
Comments: In the reverse reaction, a 2'-hydroxyisoflavone is reduced to an isoflavanone; 2'-hydroxypseudobaptigenin also acts. Involved in the biosynthesis of the pterocarpin phytoalexins medicarpin and maackiain.
References: [4288]

[EC 1.3.1.45 created 1990]

EC 1.3.1.46

Accepted name: biochanin-A reductase
Reaction: dihydrobiochanin A + NADP⁺ = biochanin A + NADPH + H⁺
Systematic name: dihydrobiochanin-A:NADP⁺ Δ²-oxidoreductase

Comments: Some other isoflavones are reduced to the corresponding isoflavanones.

References: [4288]

[EC 1.3.1.46 created 1990]

EC 1.3.1.47

Accepted name: α -santonin 1,2-reductase

Reaction: 1,2-dihydrosantonin + NAD(P)⁺ = α -santonin + NAD(P)H + H⁺

Systematic name: 1,2-dihydrosantonin:NAD(P)⁺ 1,2-oxidoreductase

References: [2967]

[EC 1.3.1.47 created 1990]

EC 1.3.1.48

Accepted name: 13,14-dehydro-15-oxoprostaglandin 13-reductase

Reaction: 11 α -hydroxy-9,15-dioxoprostanoate + NAD(P)⁺ = (13E)-11 α -hydroxy-9,15-dioxoprost-13-enoate + NAD(P)H + H⁺

Other name(s): 15-oxo- Δ^{13} -prostaglandin reductase; Δ^{13} -15-ketoprostaglandin reductase; 15-ketoprostaglandin Δ^{13} -reductase; prostaglandin Δ^{13} -reductase; prostaglandin 13-reductase; (5Z)-(15S)-11 α -hydroxy-9,15-dioxoprostanoate:NAD(P)⁺ Δ^{13} -oxidoreductase; (5Z)-11 α -hydroxy-9,15-dioxoprost-5-enoate:NAD(P)⁺ Δ^{13} -oxidoreductase

Systematic name: 11 α -hydroxy-9,15-dioxoprostanoate:NAD(P)⁺ Δ^{13} -oxidoreductase

Comments: Reduces 13,14-dehydro-15-oxoprostaglandins to 13,14-dihydro derivatives. The enzyme from placenta is specific for NAD⁺.

References: [1512, 1891]

[EC 1.3.1.48 created 1990, modified 2014]

EC 1.3.1.49

Accepted name: *cis*-3,4-dihydrophenanthrene-3,4-diol dehydrogenase

Reaction: (+)-*cis*-3,4-dihydrophenanthrene-3,4-diol + NAD⁺ = phenanthrene-3,4-diol + NADH + H⁺

Systematic name: (+)-*cis*-3,4-dihydrophenanthrene-3,4-diol:NAD⁺ 3,4-oxidoreductase

References: [2961]

[EC 1.3.1.49 created 1992]

[1.3.1.50 Deleted entry. tetrahydroxynaphthalene reductase. Now EC 1.1.1.252 tetrahydroxynaphthalene reductase]

[EC 1.3.1.50 created 1992, deleted 1999]

EC 1.3.1.51

Accepted name: 2'-hydroxydaidzein reductase

Reaction: 2'-hydroxy-2,3-dihydrodaidzein + NADP⁺ = 2'-hydroxydaidzein + NADPH + H⁺

Other name(s): NADPH:2'-hydroxydaidzein oxidoreductase; HDR; 2'-hydroxydihydrodaidzein:NADP⁺ 2'-oxidoreductase

Systematic name: 2'-hydroxy-2,3-dihydrodaidzein:NADP⁺ 2'-oxidoreductase

Comments: In the reverse reaction, the 2'-hydroxyisoflavone (2'-hydroxydaidzein) is reduced to an isoflavanone. Also acts on 2'-hydroxyformononetin and to a small extent on 2'-hydroxygenistein. Involved in the biosynthesis of the phytoalexin glyceollin. The isoflavones biochanin A, daidzein and genestein as well as the flavonoids apigenin, kaempferol and quercetin do not act as substrates.

References: [1124]

[EC 1.3.1.51 created 1992, modified 2004]

[1.3.1.52 *Transferred entry. 2-methyl-branched-chain-enoyl-CoA reductase. Now EC 1.3.8.5, 2-methyl-branched-chain-enoyl-CoA reductase*]

[EC 1.3.1.52 created 1992, deleted 2012]

EC 1.3.1.53

Accepted name: (3*S*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1,4-dicarboxylate dehydrogenase
Reaction: (3*S*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1,4-dicarboxylate + NAD⁺ = 3,4-dihydroxybenzoate + CO₂ + NADH
Other name(s): (1*R*,2*S*)-dihydroxy-3,5-cyclohexadiene-1,4-dicarboxylate dehydrogenase; terephthalate 1,2-*cis*-dihydrodiol dehydrogenase; *cis*-4,5-dihydroxycyclohexa-1(6),2-diene-1,4-dicarboxylate:NAD⁺ oxidoreductase (decarboxylating)
Systematic name: (3*S*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1,4-dicarboxylate:NAD⁺ oxidoreductase
Comments: Requires Fe^{II}. Involved in the terephthalate degradation pathway in bacteria [4539].
References: [3648, 4539]

[EC 1.3.1.53 created 1999 (EC 1.3.1.61 created 2000, incorporated 2007)]

EC 1.3.1.54

Accepted name: precorrin-6A reductase
Reaction: precorrin-6B + NADP⁺ = precorrin-6A + NADPH + H⁺
Other name(s): precorrin-6X reductase; precorrin-6Y:NADP⁺ oxidoreductase; CobK
Systematic name: precorrin-6B:NADP⁺ oxidoreductase
Comments: The enzyme, which participates in the aerobic (late cobalt insertion) pathway of adenosylcobalamin biosynthesis, catalyses the reduction of the double bond between C-18 and C-19 of precorrin-6A. See EC 1.3.1.106, cobalt-precorrin-6A reductase, for the corresponding enzyme that participates in the anaerobic cobalamin biosynthesis pathway.
References: [354, 4550]

[EC 1.3.1.54 created 1999, modified 2004]

[1.3.1.55 *Deleted entry. cis-1,2-dihydroxycyclohexa-3,5-diene-1-carboxylate dehydrogenase. Enzyme is identical to EC 1.3.1.25, 1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate dehydrogenase*]

[EC 1.3.1.55 created 1999, deleted 2004]

EC 1.3.1.56

Accepted name: *cis*-2,3-dihydrobiphenyl-2,3-diol dehydrogenase
Reaction: *cis*-3-phenylcyclohexa-3,5-diene-1,2-diol + NAD⁺ = biphenyl-2,3-diol + NADH + H⁺
Other name(s): 2,3-dihydro-2,3-dihydroxybiphenyl dehydrogenase
Systematic name: *cis*-3-phenylcyclohexa-3,5-diene-1,2-diol:NAD⁺ oxidoreductase
Comments: Catalyses the second step in the biphenyl degradation pathway in bacteria.
References: [4154, 1223, 1685]

[EC 1.3.1.56 created 2000]

EC 1.3.1.57

Accepted name: phloroglucinol reductase
Reaction: dihydrophloroglucinol + NADP⁺ = phloroglucinol + NADPH + H⁺
Systematic name: dihydrophloroglucinol:NADP⁺ oxidoreductase
Comments: Involved in the gallate anaerobic degradation pathway in bacteria.
References: [1465]

[EC 1.3.1.57 created 2000]

EC 1.3.1.58

Accepted name: 2,3-dihydroxy-2,3-dihydro-*p*-cumate dehydrogenase
Reaction: *cis*-5,6-dihydroxy-4-isopropylcyclohexa-1,3-dienecarboxylate + NAD⁺ = 2,3-dihydroxy-*p*-cumate + NADH + H⁺
Systematic name: *cis*-2,3-dihydroxy-2,3-dihydro-*p*-cumate:NAD⁺ oxidoreductase
Comments: Involved in the *p*-cymene degradation pathway in *Pseudomonas putida*.
References: [1007]

[EC 1.3.1.58 created 2000]

[1.3.1.59 Deleted entry. 1,2-dihydroxy-3-methyl-1,2-dihydrobenzoate dehydrogenase. No evidence in the paper cited that the enzyme exists]

[EC 1.3.1.59 created 2000, deleted 2006]

EC 1.3.1.60

Accepted name: dibenzothiophene dihydrodiol dehydrogenase
Reaction: *cis*-1,2-dihydroxy-1,2-dihydrodibenzothiophene + NAD⁺ = 1,2-dihydroxydibenzothiophene + NADH + H⁺
Systematic name: *cis*-1,2-dihydroxy-1,2-dihydrodibenzothiophene:NAD⁺ oxidoreductase
Comments: Involved in the dibenzothiophene degradation pathway in bacteria.
References: [2323, 883]

[EC 1.3.1.60 created 2000]

[1.3.1.61 Deleted entry. terephthalate 1,2-*cis*-dihydrodiol dehydrogenase. Enzyme is identical to EC 1.3.1.53, (3*S*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1,4-dicarboxylate dehydrogenase]

[EC 1.3.1.61 created 2000, deleted 2007]

EC 1.3.1.62

Accepted name: pimeloyl-CoA dehydrogenase
Reaction: pimeloyl-CoA + NAD⁺ = 6-carboxyhex-2-enoyl-CoA + NADH + H⁺
Systematic name: pimeloyl-CoA:NAD⁺ oxidoreductase
Comments: Involved in the benzoate degradation (anaerobic) pathway in bacteria.
References: [1262]

[EC 1.3.1.62 created 2000]

[1.3.1.63 Transferred entry. 2,4-dichlorobenzoyl-CoA reductase. Now EC 1.21.1.2, 2,4-dichlorobenzoyl-CoA reductase]

[EC 1.3.1.63 created 2000, modified 2011, deleted 2015]

EC 1.3.1.64

Accepted name: phthalate 4,5-*cis*-dihydrodiol dehydrogenase
Reaction: *cis*-4,5-dihydroxycyclohexa-1(6),2-diene-1,2-dicarboxylate + NAD⁺ = 4,5-dihydroxyphthalate + NADH + H⁺
Systematic name: *cis*-4,5-dihydroxycyclohexa-1(6),2-diene-1,2-dicarboxylate:NAD⁺ oxidoreductase
Comments: Involved in the phthalate degradation pathway in bacteria.
References: [237]

[EC 1.3.1.64 created 2000]

EC 1.3.1.65

- Accepted name:** 5,6-dihydroxy-3-methyl-2-oxo-1,2,5,6-tetrahydroquinoline dehydrogenase
Reaction: 5,6-dihydroxy-3-methyl-2-oxo-1,2,5,6-tetrahydroquinoline + NAD⁺ = 5,6-dihydroxy-3-methyl-2-oxo-1,2-dihydroquinoline + NADH + H⁺
Systematic name: 5,6-dihydroxy-3-methyl-2-oxo-1,2,5,6-tetrahydroquinoline:NAD⁺ oxidoreductase
Comments: Acts in the reverse direction to form part of the 3-methylquinoline degradation pathway in bacteria.
References: [3695]

[EC 1.3.1.65 created 2000]

EC 1.3.1.66

- Accepted name:** *cis*-dihydroethylcatechol dehydrogenase
Reaction: *cis*-1,2-dihydro-3-ethylcatechol + NAD⁺ = 3-ethylcatechol + NADH + H⁺
Systematic name: *cis*-1,2-dihydro-3-ethylcatechol:NAD⁺ oxidoreductase
Comments: Involved in the ethylbenzene degradation pathway in bacteria.
References: [1320]

[EC 1.3.1.66 created 2000]

EC 1.3.1.67

- Accepted name:** *cis*-1,2-dihydroxy-4-methylcyclohexa-3,5-diene-1-carboxylate dehydrogenase
Reaction: *cis*-1,2-dihydroxy-4-methylcyclohexa-3,5-diene-1-carboxylate + NAD(P)⁺ = 4-methylcatechol + NAD(P)H + CO₂
Systematic name: *cis*-1,2-dihydroxy-4-methylcyclohexa-3,5-diene-1-carboxylate:NAD(P)⁺ oxidoreductase (decarboxylating)
Comments: Involved in the *p*-xylene degradation pathway in bacteria.
References: [4607]

[EC 1.3.1.67 created 2000]

EC 1.3.1.68

- Accepted name:** 1,2-dihydroxy-6-methylcyclohexa-3,5-dienecarboxylate dehydrogenase
Reaction: 1,2-dihydroxy-6-methylcyclohexa-3,5-dienecarboxylate + NAD⁺ = 3-methylcatechol + NADH + CO₂
Systematic name: 1,2-dihydroxy-6-methylcyclohexa-3,5-dienecarboxylate:NAD⁺ oxidoreductase (decarboxylating)
Comments: Involved in the *o*-xylene degradation pathway in bacteria.
References: [1647]

[EC 1.3.1.68 created 2000]

EC 1.3.1.69

- Accepted name:** zeatin reductase
Reaction: dihydrozeatin + NADP⁺ = zeatin + NADPH + H⁺
Systematic name: dihydrozeatin:NADP⁺ oxidoreductase
Comments: Previously classified erroneously as EC 1.1.1.242.
References: [2667]

[EC 1.3.1.69 created 1992 as EC 1.1.1.242, transferred 2001 to EC 1.3.1.69]

EC 1.3.1.70

- Accepted name:** Δ¹⁴-sterol reductase

Reaction: 4,4-dimethyl-5 α -cholesta-8,24-dien-3 β -ol + NADP⁺ = 4,4-dimethyl-5 α -cholesta-8,14,24-trien-3 β -ol + NADPH + H⁺
Systematic name: 4,4-dimethyl-5 α -cholesta-8,24-dien-3 β -ol:NADP⁺ Δ^{14} -oxidoreductase
Comments: This enzyme acts on a range of steroids with a 14(15)-double bond.
References: [404, 3222]

[EC 1.3.1.70 created 2001]

EC 1.3.1.71

Accepted name: $\Delta^{24(24^1)}$ -sterol reductase
Reaction: ergosterol + NADP⁺ = ergosta-5,7,22,24(24¹)-tetraen-3 β -ol + NADPH + H⁺
Other name(s): sterol $\Delta^{24(28)}$ -methylene reductase; sterol $\Delta^{24(28)}$ -reductase
Systematic name: ergosterol:NADP⁺ $\Delta^{24(24^1)}$ -oxidoreductase
Comments: Acts on a range of steroids with a 24(24¹)-double bond.
References: [3031, 4945]

[EC 1.3.1.71 created 2001, modified 2002]

EC 1.3.1.72

Accepted name: Δ^{24} -sterol reductase
Reaction: 5 α -cholest-7-en-3 β -ol + NADP⁺ = 5 α -cholesta-7,24-dien-3 β -ol + NADPH + H⁺
Other name(s): lanosterol Δ^{24} -reductase
Systematic name: sterol:NADP⁺ Δ^{24} -oxidoreductase
Comments: Acts on a range of steroids with a 24(25)-double bond, including lanosterol, desmosterol and zymosterol.
References: [177]

[EC 1.3.1.72 created 2001]

EC 1.3.1.73

Accepted name: 1,2-dihydrovomilenine reductase
Reaction: 17-*O*-acetylnorajmaline + NADP⁺ = 1,2-dihydrovomilenine + NADPH + H⁺
Systematic name: 17-*O*-acetylnorajmaline:NADP⁺ oxidoreductase
Comments: Forms part of the ajmaline biosynthesis pathway.
References: [1271]

[EC 1.3.1.73 created 2002]

EC 1.3.1.74

Accepted name: 2-alkenal reductase [NAD(P)⁺]
Reaction: a *n*-alkanal + NAD(P)⁺ = an alk-2-enal + NAD(P)H + H⁺
Other name(s): NAD(P)H-dependent alkenal/one oxidoreductase; NADPH:2-alkenal α,β -hydrogenase; 2-alkenal reductase
Systematic name: *n*-alkanal:NAD(P)⁺ 2-oxidoreductase
Comments: Highly specific for 4-hydroxynon-2-enal and non-2-enal. Alk-2-enals of shorter chain have lower affinities. Exhibits high activities also for alk-2-enones such as but-3-en-2-one and pent-3-en-2-one. Inactive with cyclohex-2-en-1-one and 12-oxophytodienoic acid. Involved in the detoxication of α,β -unsaturated aldehydes and ketones [*cf.* EC 1.3.1.102, 2-alkenal reductase (NADP⁺)].
References: [2636, 902]

[EC 1.3.1.74 created 2003, modified 2014]

EC 1.3.1.75

- Accepted name:** 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (NADPH)
Reaction: protochlorophyllide *a* + NADP⁺ = 3,8-divinyl protochlorophyllide *a* + NADPH + H⁺
Other name(s): DVR (gene name); *bciA* (gene name); [4-vinyl]chlorophyllide *a* reductase; 4VCR; chlorophyllide-*a*:NADP⁺ oxidoreductase; divinyl chlorophyllide *a* 8-vinyl-reductase; plant-type divinyl chlorophyllide *a* 8-vinyl-reductase
Systematic name: protochlorophyllide-*a*:NADP⁺ C-8¹-oxidoreductase
Comments: The enzyme, found in higher plants, green algae, and some phototrophic bacteria, is involved in the production of monovinyl versions of (bacterio)chlorophyll pigments from their divinyl precursors. It can also act on 3,8-divinyl chlorophyllide *a*. *cf.* EC 1.3.7.13, 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (ferredoxin).
References: [4330, 3237, 3238, 2213, 2962, 648]

[EC 1.3.1.75 created 2003, modified 2016]

EC 1.3.1.76

- Accepted name:** precorrin-2 dehydrogenase
Reaction: precorrin-2 + NAD⁺ = sirohydrochlorin + NADH + H⁺
Other name(s): Met8p; SirC; CysG
Systematic name: precorrin-2:NAD⁺ oxidoreductase
Comments: This enzyme catalyses the second of three steps leading to the formation of siroheme from uroporphyrinogen III. The first step involves the donation of two *S*-adenosyl-L-methionine-derived methyl groups to carbons 2 and 7 of uroporphyrinogen III to form precorrin-2 (EC 2.1.1.107, uroporphyrin-III *C*-methyltransferase) and the third step involves the chelation of ferrous iron to sirohydrochlorin to form siroheme (EC 4.99.1.4, sirohydrochlorin ferrochelatase). In *Saccharomyces cerevisiae*, the last two steps are carried out by a single bifunctional enzyme, Met8p. In some bacteria, steps 1-3 are catalysed by a single multifunctional protein called CysG, whereas in *Bacillus megaterium*, three separate enzymes carry out each of the steps, with SirC being responsible for the above reaction.
References: [3748, 4550]

[EC 1.3.1.76 created 2004]

EC 1.3.1.77

- Accepted name:** anthocyanidin reductase [(2*R*,3*R*)-flavan-3-ol-forming]
Reaction: a (2*R*,3*R*)-flavan-3-ol + 2 NAD(P)⁺ = an anthocyanidin with a 3-hydroxy group + 2 NAD(P)H + H⁺
Other name(s): ANR (gene name) (ambiguous); flavan-3-ol:NAD(P)⁺ oxidoreductase; anthocyanidin reductase (ambiguous)
Systematic name: (2*R*,3*R*)-flavan-3-ol:NAD(P)⁺ 3,4-oxidoreductase
Comments: The enzyme participates in the flavonoid biosynthesis pathway found in plants. It catalyses the double reduction of anthocyanidins, producing (2*R*,3*R*)-flavan-3-ol monomers required for the formation of proanthocyanidins. While the enzyme from the legume *Medicago truncatula* (MtANR) can use both NADPH and NADH as reductant, that from the crucifer *Arabidopsis thaliana* (AtANR) uses only NADPH. Also, while the substrate preference of MtANR is cyanidin, pelargonidin, delphinidin, the reverse preference is found with AtANR. *cf.* EC 1.3.1.112, anthocyanidin reductase [(2*S*)-flavan-3-ol-forming].
References: [4697, 4696, 3232]

[EC 1.3.1.77 created 2004, modified 2016]

EC 1.3.1.78

- Accepted name:** arogenate dehydrogenase (NADP⁺)
Reaction: L-arogenate + NADP⁺ = L-tyrosine + NADPH + CO₂

Other name(s): arogenic dehydrogenase (ambiguous); pretyrosine dehydrogenase (ambiguous); TyrAAT1; TyrAAT2; TyrAa
Systematic name: L-arogenate:NADP⁺ oxidoreductase (decarboxylating)
Comments: Arogenate dehydrogenases may utilize NAD⁺ (EC 1.3.1.43), NADP⁺ (EC 1.3.1.78), or both (EC 1.3.1.79). NADP⁺-dependent enzymes usually predominate in higher plants. The enzyme from the cyanobacterium *Synechocystis* sp. PCC 6803 and the TyrAAT1 isoform of the plant *Arabidopsis thaliana* cannot use prephenate as a substrate, while the *Arabidopsis* isoform TyrAAT2 can use it very poorly [3526, 385].
References: [1255, 3526, 385]

[EC 1.3.1.78 created 2005]

EC 1.3.1.79

Accepted name: arogenate dehydrogenase [NAD(P)⁺]
Reaction: L-arogenate + NAD(P)⁺ = L-tyrosine + NAD(P)H + CO₂
Other name(s): arogenic dehydrogenase (ambiguous); cyclohexadienyl dehydrogenase; pretyrosine dehydrogenase (ambiguous)
Systematic name: L-arogenate:NAD(P)⁺ oxidoreductase (decarboxylating)
Comments: Arogenate dehydrogenases may utilize NAD⁺ (EC 1.3.1.43), NADP⁺ (EC 1.3.1.78), or both (EC 1.3.1.79). Enzymes that can utilize both cofactors have been reported from some Proteobacteria, including *Burkholderia caryophylli*, *Burkholderia cepacia*, *Pseudomonas marginata* and *Delftia acidovorans*.
References: [513]

[EC 1.3.1.79 created 2005]

[1.3.1.80 Transferred entry. red chlorophyll catabolite reductase. Now classified as EC 1.3.7.12, red chlorophyll catabolite reductase]

[EC 1.3.1.80 created 2007, deleted 2016]

EC 1.3.1.81

Accepted name: (+)-pulegone reductase
Reaction: (1) (-)-menthone + NADP⁺ = (+)-pulegone + NADPH + H⁺
 (2) (+)-isomenthone + NADP⁺ = (+)-pulegone + NADPH + H⁺
Systematic name: (-)-menthone:NADP⁺ oxidoreductase
Comments: NADH cannot replace NADPH as reductant. The Δ^{8,9}-double bond of (+)-*cis*-isopulegone and the Δ^{1,2}-double bond of (±)-piperitone are not substrates. The enzyme from peppermint (*Mentha × piperita*) converts (+)-pulegone into both (-)-menthone and (+)-isomenthone at a ratio of 70:30 for native enzyme but it does not catalyse the reverse reaction. This enzyme is a member of the medium-chain dehydrogenase/reductase superfamily.
References: [3524]

[EC 1.3.1.81 created 2008]

EC 1.3.1.82

Accepted name: (-)-isopiperitenone reductase
Reaction: (+)-*cis*-isopulegone + NADP⁺ = (-)-isopiperitenone + NADPH + H⁺
Systematic name: (+)-*cis*-isopulegone:NADP⁺ oxidoreductase

Comments: The reaction occurs in the opposite direction to that shown above. The enzyme participates in the menthol-biosynthesis pathway of *Mentha* plants. (+)-Pulegone, (+)-*cis*-isopulegone and (-)-menthone are not substrates. The enzyme has a preference for NADPH as the reductant, with NADH being a poor substitute [3524]. The enzyme is highly regioselective for the reduction of the endocyclic 1,2-double bond, and is stereoselective, producing only the 1*R*-configured product. It is a member of the short-chain dehydrogenase/reductase superfamily.

References: [774, 3524]

[EC 1.3.1.82 created 2008]

EC 1.3.1.83

Accepted name: geranylgeranyl diphosphate reductase
Reaction: phytyl diphosphate + 3 NADP⁺ = geranylgeranyl diphosphate + 3 NADPH + 3 H⁺
Other name(s): geranylgeranyl reductase; CHL P
Systematic name: geranylgeranyl-diphosphate:NADP⁺ oxidoreductase
Comments: This enzyme also acts on geranylgeranyl-chlorophyll *a*. The reaction occurs in three steps. Which order the three double bonds are reduced is not known.
References: [3955, 4197, 2059]

[EC 1.3.1.83 created 2009]

EC 1.3.1.84

Accepted name: acrylyl-CoA reductase (NADPH)
Reaction: propanoyl-CoA + NADP⁺ = acryloyl-CoA + NADPH + H⁺
Systematic name: propanoyl-CoA:NADP⁺ oxidoreductase
Comments: Catalyses a step in the 3-hydroxypropanoate/4-hydroxybutanoate cycle, an autotrophic CO₂ fixation pathway found in some thermoacidophilic archaea [296]. The enzyme from *Sulfolobus tokodaii* does not act on either NADH or crotonyl-CoA [4249]. Different from EC 1.3.1.8, which acts only on enoyl-CoA derivatives of carbon chain length 4 to 16. Contains Zn²⁺.
References: [296, 4249]

[EC 1.3.1.84 created 2009, modified 2014]

EC 1.3.1.85

Accepted name: crotonyl-CoA carboxylase/reductase
Reaction: (2*S*)-ethylmalonyl-CoA + NADP⁺ = (*E*)-but-2-enoyl-CoA + CO₂ + NADPH + H⁺
Other name(s): CCR; crotonyl-CoA reductase (carboxylating)
Systematic name: (2*S*)-ethylmalonyl-CoA:NADP⁺ oxidoreductase (decarboxylating)
Comments: The reaction is catalysed in the reverse direction. This enzyme, isolated from the bacterium *Rhodobacter sphaeroides*, catalyses (*E*)-but-2-enoyl-CoA-dependent oxidation of NADPH in the presence of CO₂. When CO₂ is absent, the enzyme catalyses the reduction of (*E*)-but-2-enoyl-CoA to butanoyl-CoA, but with only 10% of maximal activity (relative to (*E*)-but-2-enoyl-CoA carboxylation).
References: [1059, 1060]

[EC 1.3.1.85 created 2011]

EC 1.3.1.86

Accepted name: crotonyl-CoA reductase
Reaction: butanoyl-CoA + NADP⁺ = (*E*)-but-2-enoyl-CoA + NADPH + H⁺

Other name(s): butyryl-CoA dehydrogenase; butyryl dehydrogenase; unsaturated acyl-CoA reductase; ethylene reductase; enoyl-coenzyme A reductase; unsaturated acyl coenzyme A reductase; butyryl coenzyme A dehydrogenase; short-chain acyl CoA dehydrogenase; short-chain acyl-coenzyme A dehydrogenase; 3-hydroxyacyl CoA reductase; butanoyl-CoA:(acceptor) 2,3-oxidoreductase; CCR

Systematic name: butanoyl-CoA:NADP⁺ 2,3-oxidoreductase

Comments: Catalyses the reaction in the reverse direction. This enzyme from *Streptomyces collinus* is specific for (*E*)-but-2-enoyl-CoA, and is proposed to provide butanoyl-CoA as a starter unit for straight-chain fatty acid biosynthesis.

References: [4501]

[EC 1.3.1.86 created 2011]

EC 1.3.1.87

Accepted name: 3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate dehydrogenase

Reaction:
(1) 3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate + NAD⁺ = 3-(2,3-dihydroxyphenyl)propanoate + NADH + H⁺
(2) (2*E*)-3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)prop-2-enoate + NAD⁺ = (2*E*)-3-(2,3-dihydroxyphenyl)prop-2-enoate + NADH + H⁺

Other name(s): *hcaB* (gene name); *cis*-dihydrodiol dehydrogenase; 2,3-dihydroxy-2,3-dihydro-phenylpropionate dehydrogenase

Systematic name: 3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate:NAD⁺ oxidoreductase

Comments: This enzyme catalyses a step in the pathway of phenylpropanoid compounds degradation.

References: [900]

[EC 1.3.1.87 created 2011]

EC 1.3.1.88

Accepted name: tRNA-dihydrouridine^{16/17} synthase [NAD(P)⁺]

Reaction:
(1) 5,6-dihydrouracil¹⁶ in tRNA + NAD(P)⁺ = uracil¹⁶ in tRNA + NAD(P)H + H⁺
(2) 5,6-dihydrouracil¹⁷ in tRNA + NAD(P)⁺ = uracil¹⁷ in tRNA + NAD(P)H + H⁺

Other name(s): Dus1p; tRNA-dihydrouridine synthase 1

Systematic name: tRNA-5,6-dihydrouracil^{16/17}:NAD(P)⁺ oxidoreductase

Comments: A flavoprotein. The enzyme specifically modifies uracil¹⁶ and uracil¹⁷ in tRNA.

References: [4700, 4701]

[EC 1.3.1.88 created 2011]

EC 1.3.1.89

Accepted name: tRNA-dihydrouridine⁴⁷ synthase [NAD(P)⁺]

Reaction: 5,6-dihydrouracil⁴⁷ in tRNA + NAD(P)⁺ = uracil⁴⁷ in tRNA + NAD(P)H + H⁺

Other name(s): Dus3p; tRNA-dihydrouridine synthase 3

Systematic name: tRNA-5,6-dihydrouracil⁴⁷:NAD(P)⁺ oxidoreductase

Comments: A flavoenzyme. The enzyme specifically modifies uracil⁴⁷ in tRNA.

References: [4700]

[EC 1.3.1.89 created 2011]

EC 1.3.1.90

Accepted name: tRNA-dihydrouridine^{20a/20b} synthase [NAD(P)⁺]

Reaction:
(1) 5,6-dihydrouracil^{20a} in tRNA + NAD(P)⁺ = uracil^{20a} in tRNA + NAD(P)H + H⁺
(2) 5,6-dihydrouracil^{20b} in tRNA + NAD(P)⁺ = uracil^{20b} in tRNA + NAD(P)H + H⁺

Other name(s): Dus4p

Systematic name: tRNA-5,6-dihydrouracil^{20a/20b}:NAD(P)⁺ oxidoreductase
Comments: A flavoenzyme. The enzyme specifically modifies uracil^{20a} and uracil^{20b} in tRNA.
References: [4700]

[EC 1.3.1.90 created 2011]

EC 1.3.1.91

Accepted name: tRNA-dihydrouridine²⁰ synthase [NAD(P)⁺]
Reaction: 5,6-dihydrouracil²⁰ in tRNA + NAD(P)⁺ = uracil²⁰ in tRNA + NAD(P)H + H⁺
Other name(s): Dus2p; tRNA-dihydrouridine synthase 2
Systematic name: tRNA-5,6-dihydrouracil²⁰:NAD(P)⁺ oxidoreductase
Comments: A flavoenzyme [3517]. The enzyme specifically modifies uracil²⁰ in tRNA.
References: [4700, 4701, 3517, 2022]

[EC 1.3.1.91 created 2011]

EC 1.3.1.92

Accepted name: artemisinic aldehyde $\Delta^{11(13)}$ -reductase
Reaction: (11*R*)-dihydroartemisinic aldehyde + NADP⁺ = artemisinic aldehyde + NADPH + H⁺
Other name(s): Dbr2
Systematic name: artemisinic aldehyde:NADP⁺ oxidoreductase
Comments: Cloned from *Artemisia annua*. In addition to the reduction of artemisinic aldehyde it is also able to a lesser extent to reduce artemisinic alcohol and artemisinic acid. Part of the biosynthesis of artemisinin.
References: [309, 4888]

[EC 1.3.1.92 created 2012]

EC 1.3.1.93

Accepted name: very-long-chain enoyl-CoA reductase
Reaction: a very-long-chain acyl-CoA + NADP⁺ = a very-long-chain *trans*-2,3-dehydroacyl-CoA + NADPH + H⁺
Other name(s): TSC13 (gene name); CER10 (gene name)
Systematic name: very-long-chain acyl-CoA:NADP⁺ oxidoreductase
Comments: This is the fourth component of the elongase, a microsomal protein complex responsible for extending palmitoyl-CoA and stearoyl-CoA (and modified forms thereof) to very-long-chain acyl CoAs. *cf.* EC 2.3.1.199, very-long-chain 3-oxoacyl-CoA synthase, EC 1.1.1.330, very-long-chain 3-oxoacyl-CoA reductase, and EC 4.2.1.134, very-long-chain (3*R*)-3-hydroxyacyl-CoA dehydratase.
References: [2198, 1246, 2316, 4909]

[EC 1.3.1.93 created 2012]

EC 1.3.1.94

Accepted name: polyprenol reductase
Reaction: *ditrans, polycis*-dolichol + NADP⁺ = *ditrans, polycis*-polyprenol + NADPH + H⁺
Other name(s): SRD5A3 (gene name); DFG10 (gene name)
Systematic name: *ditrans, polycis*-dolichol:NADP⁺ 2,3-oxidoreductase
Comments: The reaction occurs in the reverse direction with reduction of the terminal double bond next to the alcohol group. Isolated from human fetal brain tissue but present in all eukaryotes. In mammalian cells dolichols are predominantly 18-21 isoprene units in length.
References: [3627, 545]

[EC 1.3.1.94 created 2012]

EC 1.3.1.95

- Accepted name:** acrylyl-CoA reductase (NADH)
Reaction: propanoyl-CoA + NAD⁺ = acryloyl-CoA + NADH + H⁺
Systematic name: propanoyl-CoA:NAD⁺ oxidoreductase
Comments: Contains FAD. The reaction is catalysed in the opposite direction to that shown. The enzyme from the bacterium *Clostridium propionicum* is a complex that includes an electron-transfer flavoprotein (ETF). The ETF is reduced by NADH and transfers the electrons to the active site. Catalyses a step in a pathway for L-alanine fermentation to propanoate [1632]. *cf.* EC 1.3.1.84, acrylyl-CoA reductase (NADPH).
References: [1632, 1989]

[EC 1.3.1.95 created 2012]

EC 1.3.1.96

- Accepted name:** *Botryococcus* squalene synthase
Reaction: squalene + diphosphate + NADP⁺ = presqualene diphosphate + NADPH + H⁺
Other name(s): SSL-2 (gene name)
Systematic name: squalene:NADP⁺ oxidoreductase
Comments: Isolated from the green alga *Botryococcus braunii* BOT22. Acts in the reverse direction. *cf.* EC 2.5.1.21, squalene synthase, where squalene is formed directly from farnesyl diphosphate.
References: [3066]

[EC 1.3.1.96 created 2012]

EC 1.3.1.97

- Accepted name:** botryococcene synthase
Reaction: C₃₀ botryococcene + NADP⁺ + diphosphate = presqualene diphosphate + NADPH + H⁺
Other name(s): SSL-3 (gene name)
Systematic name: C₃₀ botryococcene:NADP⁺ oxidoreductase
Comments: Isolated from the green alga *Botryococcus braunii* BOT22. Acts in the reverse direction. Involved in the production of botryococcenes, which are triterpenoid hydrocarbons of isoprenoid origin produced in large amount by this alga.
References: [3066]

[EC 1.3.1.97 created 2012]

EC 1.3.1.98

- Accepted name:** UDP-*N*-acetylmuramate dehydrogenase
Reaction: UDP-*N*-acetyl- α -D-muramate + NADP⁺ = UDP-*N*-acetyl-3-*O*-(1-carboxyvinyl)- α -D-glucosamine + NADPH + H⁺
Other name(s): MurB reductase; UDP-*N*-acetylenolpyruvoylglucosamine reductase; UDP-*N*-acetylglucosamine-enoylpyruvate reductase; UDP-GlcNAc-enoylpyruvate reductase; uridine diphosphoacetylpyruvoylglucosamine reductase; uridine diphospho-*N*-acetylglucosamine-enolpyruvate reductase; uridine-5'-diphospho-*N*-acetyl-2-amino-2-deoxy-3-*O*-lactylglucose:NADP-oxidoreductase
Systematic name: UDP-*N*-acetyl- α -D-muramate:NADP⁺ oxidoreductase
Comments: A flavoprotein (FAD). NADH can to a lesser extent replace NADPH.
References: [4187, 4188, 4407]

[EC 1.3.1.98 created 1976 as EC 1.1.1.158, modified 1983, modified 2002, transferred 2013 to EC 1.3.1.98]

[1.3.1.99 Transferred entry. *iridoid synthase*. Now known to be catalyzed by two different enzymes, EC 1.3.1.122, (*S*)-8-oxocitronellyl enol synthase, and EC 5.5.1.34, (+)-*cis,trans*-nepetalactol synthase]

[EC 1.3.1.99 created 2013, deleted 2019]

EC 1.3.1.100

- Accepted name:** chanoclavine-I aldehyde reductase
Reaction: dihydrochanoclavine-I aldehyde + NADP⁺ = chanoclavine-I aldehyde + NADPH + H⁺
Other name(s): FgaOx3; *easA* (gene name)
Systematic name: chanoclavine-I aldehyde:NAD⁺ oxidoreductase
Comments: Contains FMN. The enzyme participates in the biosynthesis of fumigaclavine C, an ergot alkaloid produced by some fungi of the *Trichocomaceae* family. The enzyme catalyses the reduction of chanoclavine-I aldehyde to dihydrochanoclavine-I aldehyde. This hydrolyses spontaneously to form 6,8-dimethyl-6,7-didehydroergoline, which is converted to festuclavine by EC 1.5.1.44, festuclavine dehydrogenase.
References: [759, 643, 4506, 4698]

[EC 1.3.1.100 created 2013]

EC 1.3.1.101

- Accepted name:** 2,3-bis-*O*-geranylgeranyl-*sn*-glycerol 1-phosphate reductase [NAD(P)H]
Reaction: 2,3-bis-(*O*-phytanyl)-*sn*-glycerol 1-phosphate + 8 NAD(P)⁺ = 2,3-bis-(*O*-geranylgeranyl)-*sn*-glycerol 1-phosphate + 8 NAD(P)H + 8 H⁺
Other name(s): digeranylgeranyl-glycerophospholipid reductase; Ta0516m (gene name); DGGGPL reductase; 2,3-digeranylgeranyl-glycerophospholipid reductase
Systematic name: 2,3-bis-(*O*-phytanyl)-*sn*-glycerol 1-phosphate:NAD(P)⁺ oxidoreductase
Comments: A flavoprotein (FAD). The enzyme from the archaeon *Thermoplasma acidophilum* is involved in the biosynthesis of membrane lipids. *In vivo* the reaction occurs in the reverse direction with the formation of 2,3-bis-*O*-phytanyl-*sn*-glycerol 1-phosphate. *cf.* EC 1.3.7.11, 2,3-bis-*O*-geranylgeranyl-*sn*-glycero-phospholipid reductase.
References: [3078, 3079, 4707]

[EC 1.3.1.101 created 2013]

EC 1.3.1.102

- Accepted name:** 2-alkenal reductase (NADP⁺)
Reaction: an *n*-alkanal + NADP⁺ = an alk-2-enal + NADPH + H⁺
Other name(s): NADPH-dependent alkenal/one oxidoreductase; NADPH:2-alkenal α,β-hydrogenase
Systematic name: *n*-alkanal:NADP⁺ 2-oxidoreductase
Comments: Shows highest activity with 1-nitrocyclohexene but also has significant activity with 2-methylpentenal and *trans*-cinnamaldehyde [2639]. Involved in the detoxication of α,β-unsaturated aldehydes and ketones. Has very low activity with NAD as reductant (*cf.* EC 1.3.1.74, 2-alkenal reductase [NAD(P)⁺]).
References: [1670, 2710, 2639]

[EC 1.3.1.102 created 2013]

EC 1.3.1.103

- Accepted name:** 2-haloacrylate reductase
Reaction: (*S*)-2-chloropropanoate + NADP⁺ = 2-chloroacrylate + NADPH + H⁺
Other name(s): CAA43 (gene name)
Systematic name: (*S*)-2-chloropropanoate:NADP⁺ oxidoreductase
Comments: The enzyme acts in the degradation pathway of unsaturated organohalogen compounds by the bacterium *Burkholderia* sp. WS.
References: [2298]

[EC 1.3.1.103 created 2013]

EC 1.3.1.104

- Accepted name:** enoyl-[acyl-carrier-protein] reductase (NADPH)
Reaction: an acyl-[acyl-carrier protein] + NADP⁺ = a *trans*-2,3-dehydroacyl-[acyl-carrier protein] + NADPH + H⁺
Other name(s): acyl-ACP dehydrogenase (ambiguous); enoyl-[acyl carrier protein] (reduced nicotinamide adenine dinucleotide phosphate) reductase; NADPH 2-enoyl Co A reductase; enoyl-ACP reductase (ambiguous); *fabL* (gene name)
Systematic name: acyl-[acyl-carrier protein]:NADP⁺ oxidoreductase
Comments: The enzyme completes each cycle of fatty acid elongation by catalysing the stereospecific reduction of the double bond at position 2 of a growing fatty acid chain, while linked to the acyl-carrier protein, in an NADPH-dependent manner. This entry stands for enzymes whose stereo-specificity with respect to NADP⁺ is not known. [*cf.* EC 1.3.1.39 enoyl-[acyl-carrier-protein] reductase (NADPH, *Re*-specific), EC 1.3.1.10, enoyl-[acyl-carrier-protein] reductase (NADPH, *Si*-specific) and EC 1.3.1.9, enoyl-[acyl-carrier-protein] reductase (NADH)].
References: [1595, 2100, 2098]

[EC 1.3.1.104 created 2013]

EC 1.3.1.105

- Accepted name:** 2-methylene-furan-3-one reductase
Reaction: 4-hydroxy-2,5-dimethylfuran-3(2*H*)-one + NADP⁺ = 4-hydroxy-5-methyl-2-methylenefuran-3(2*H*)-one + NADPH + H⁺
Other name(s): FaEO; SIEO; enone oxidoreductase; 4-hydroxy-2,5-dimethylfuran-3(2*H*)-one:NAD(P)⁺ oxidoreductase
Systematic name: 4-hydroxy-2,5-dimethylfuran-3(2*H*)-one:NADP⁺ oxidoreductase
Comments: The enzyme was discovered in strawberry (*Fragaria x ananassa*), where it produces furaneol, one of the major aroma compounds in the fruits. It has also been detected in tomato (*Solanum lycopersicum*) and pineapple (*Ananas comosus*). The enzyme can also act on derivatives substituted at the methylene functional group. The enzyme from the yeast *Saccharomyces cerevisiae* acts on (2*E*)-2-ethylidene-4-hydroxy-5-methylfuran-3(2*H*)-one and produces homofuraneol, an important aroma compound in soy sauce and miso. NADPH is the preferred cofactor.
References: [3418, 2154, 3716, 4369]

[EC 1.3.1.105 created 2013]

EC 1.3.1.106

- Accepted name:** cobalt-precorrin-6A reductase
Reaction: cobalt-precorrin-6B + NAD⁺ = cobalt-precorrin-6A + NADH + H⁺
Other name(s): *cbiJ* (gene name)
Systematic name: cobalt-precorrin-6B:NAD⁺ oxidoreductase
Comments: The enzyme, which participates in the anaerobic (early cobalt insertion) pathway of adenosylcobalamin biosynthesis, catalyses the reduction of the double bond between C-18 and C-19 of cobalt-precorrin-6A. The enzyme from the bacterium *Bacillus megaterium* has no activity with NADPH. See EC 1.3.1.54, precorrin-6A reductase, for the corresponding enzyme that participates in the aerobic cobalamin biosynthesis pathway.
References: [2110, 2877]

[EC 1.3.1.106 created 2014]

EC 1.3.1.107

- Accepted name:** sanguinarine reductase
Reaction: (1) dihydrosanguinarine + NAD(P)⁺ = sanguinarine + NAD(P)H + H⁺
(2) dihydrochelirubine + NAD(P)⁺ = chelirubine + NAD(P)H + H⁺

Systematic name: dihydrosanguinarine:NAD(P)⁺ oxidoreductase
Comments: The enzyme, purified from the California poppy (*Eschscholzia californica*), is involved in detoxifying the phytoalexin sanguinarine produced by poppy itself (cf. EC 1.5.3.12, dihydrobenzophenanthridine oxidase), when it binds to the cell wall of the poppy cell. The reaction with NADPH is up to three times faster than that with NADH at low concentrations (<10 μM) of the dinucleotide. At higher concentrations the reaction with NADPH is inhibited but not that with NADH [4578].
References: [4578, 4457]

[EC 1.3.1.107 created 2014]

EC 1.3.1.108

Accepted name: caffeoyl-CoA reductase
Reaction: 3-(3,4-dihydroxyphenyl)propanoyl-CoA + 2 NAD⁺ + 2 reduced ferredoxin [iron-sulfur] cluster = (2E)-3-(3,4-dihydroxyphenyl)prop-2-enoyl-CoA + 2 NADH + 2 oxidized ferredoxin [iron-sulfur] cluster
Other name(s): electron-bifurcating caffeoyl-CoA reductase; caffeoyl-CoA reductase-Etf complex; hydrocaffeoyl-CoA:NAD⁺, ferredoxin oxidoreductase
Systematic name: 3-(3,4-dihydroxyphenyl)propanoyl-CoA:NAD⁺, ferredoxin oxidoreductase
Comments: The enzyme, characterized from the bacterium *Acetobacterium woodii*, contains two [4Fe-4S] clusters and FAD. The enzyme couples the endergonic ferredoxin reduction with NADH as reductant to the exergonic reduction of caffeoyl-CoA with the same reductant. It uses the mechanism of electron bifurcation to overcome the steep energy barrier in ferredoxin reduction. It also reduces 4-coumaroyl-CoA and feruloyl-CoA.
References: [319]

[EC 1.3.1.108 created 2015]

EC 1.3.1.109

Accepted name: butanoyl-CoA dehydrogenase complex (NAD⁺, ferredoxin)
Reaction: butanoyl-CoA + 2 NAD⁺ + 2 reduced ferredoxin [iron-sulfur] cluster = (E)-but-2-enoyl-CoA + 2 NADH + 2 oxidized ferredoxin [iron-sulfur] cluster
Other name(s): bifurcating butyryl-CoA dehydrogenase; butyryl-CoA dehydrogenase/Etf complex; Etf-Bcd complex; bifurcating butanoyl-CoA dehydrogenase; butanoyl-CoA dehydrogenase/Etf complex; butanoyl-CoA dehydrogenase (NAD⁺, ferredoxin)
Systematic name: butanoyl-CoA:NAD⁺, ferredoxin oxidoreductase
Comments: The enzyme is a complex of a flavin-containing dehydrogenase component (Bcd) and an electron-transfer flavoprotein dimer (EtfAB). The enzyme complex, isolated from the bacteria *Acidaminococcus fermentans* and butanoate-producing *Clostridia* species, couples the exergonic reduction of (E)-but-2-enoyl-CoA to butanoyl-CoA by NADH to the endergonic reduction of ferredoxin by NADH, using electron bifurcation to overcome the steep energy barrier in ferredoxin reduction.
References: [2442, 3328, 683, 682]

[EC 1.3.1.109 created 2015, modified 2021]

[1.3.1.110 Transferred entry. lactate dehydrogenase (NAD⁺, ferredoxin). Now EC 1.1.1.436, lactate dehydrogenase (NAD⁺, ferredoxin)]

[EC 1.3.1.110 created 2015, deleted 2022]

EC 1.3.1.111

Accepted name: geranylgeranyl-bacteriochlorophyllide *a* reductase
Reaction: bacteriochlorophyll *a* + 3 NADP⁺ = geranylgeranyl bacteriochlorophyllide *a* + 3 NADPH + 3 H⁺
Other name(s): geranylgeranyl-bacteriopheophytin reductase; *bchP* (gene name)
Systematic name: bacteriochlorophyll-*a*:NADP⁺ oxidoreductase (geranylgeranyl-reducing)

Comments: The enzyme catalyses the successive reduction of the geranylgeraniol esterifying group to phytol, reducing three out of four double bonds, and transforming geranylgeranyl bacteriochlorophyllide *a* via dihydrogeranylgeranyl bacteriochlorophyllide *a* and tetrahydrogeranylgeranyl bacteriochlorophyllide *a* to bacteriochlorophyll *a*. The enzyme can also accept the pheophytin derivative geranylgeranyl bacteriopheophytin, converting it to bacteriopheophytin *a*.

References: [380, 28, 29, 1525]

[EC 1.3.1.111 created 2016]

EC 1.3.1.112

Accepted name: anthocyanidin reductase [(2*S*)-flavan-3-ol-forming]

Reaction: (1) a (2*S*,3*R*)-flavan-3-ol + 2 NADP⁺ = an anthocyanidin with a 3-hydroxy group + 2 NADPH + H⁺
(2) a (2*S*,3*S*)-flavan-3-ol + 2 NADP⁺ = an anthocyanidin with a 3-hydroxy group + 2 NADPH + H⁺

Systematic name: (2*S*)-flavan-3-ol:NAD(P)⁺ oxidoreductase

Comments: The enzyme, characterized from *Vitis vinifera* (grape), participates in the flavonoid biosynthesis pathway. It catalyses the double reduction of anthocyanidins, producing a mixture of (2*S*,3*S*)- and (2*S*,3*R*)-flavan-3-ols. The enzyme catalyses sequential hydride transfers to C-2 and C-4, respectively. Epimerization at C-3 is achieved by tautomerization that occurs between the two hydride transfers. *cf.* EC 1.3.1.77, anthocyanidin reductase [(2*R*,3*R*)-flavan-3-ol-forming].

References: [1276, 1275]

[EC 1.3.1.112 created 2016]

EC 1.3.1.113

Accepted name: (4-alkanoyl-5-oxo-2,5-dihydrofuran-3-yl)methyl phosphate reductase

Reaction: a [(3*S*,4*R*)-4-alkanoyl-5-oxoxolan-3-yl]methyl phosphate + NADP⁺ = a (4-alkanoyl-5-oxo-2,5-dihydrofuran-3-yl)methyl phosphate + NADPH + H⁺

Other name(s): *bprA* (gene name); *scbC* (gene name)

Systematic name: [(3*S*,4*R*)-4-alkanoyl-5-oxoxolan-3-yl]methyl-phosphate:NADP⁺ oxidoreductase

Comments: The enzyme, characterized from the bacteria *Streptomyces griseus* and *Streptomyces coelicolor*, is involved in the biosynthesis of γ -butyrolactone autoregulators that control secondary metabolism and morphological development in *Streptomyces* bacteria.

References: [2016, 330]

[EC 1.3.1.113 created 2017]

EC 1.3.1.114

Accepted name: 3-dehydro-bile acid $\Delta^{4,6}$ -reductase

Reaction: (1) 3-oxocholan-24-oyl-CoA + NAD⁺ = 3-oxochol-4-en-24-oyl-CoA + NADH + H⁺
(2) 3-oxochol-4-en-24-oyl-CoA + NAD⁺ = 3-oxochol-4,6-dien-24-oyl-CoA + NADH + H⁺
(3) 12 α -hydroxy-3-oxocholan-24-oyl-CoA + NAD⁺ = 12 α -hydroxy-3-oxochol-4-en-24-oyl-CoA + NADH + H⁺
(4) 12 α -hydroxy-3-oxochol-4-en-24-oyl-CoA + NAD⁺ = 12 α -hydroxy-3-oxochol-4,6-dien-24-oyl-CoA + NADH + H⁺

Other name(s): *baiN* (gene name)

Systematic name: 3-oxocholan-24-oyl-CoA $\Delta^{4,6}$ -oxidoreductase

Comments: Contains flavin. The enzyme, characterized from the bacterium *Clostridium scindens*, participates in the bile acid 7 α -dehydroxylation pathway. The enzyme catalyses two subsequent reductions of the double bonds within the bile acid A/B rings, following 7 α -dehydration.

References: [1539]

[EC 1.3.1.114 created 2018]

EC 1.3.1.115

- Accepted name:** 3-oxocholoyl-CoA 4-desaturase
- Reaction:** (1) $7\alpha,12\alpha$ -dihydroxy-3-oxochol-24-oyl-CoA + NAD⁺ = $7\alpha,12\alpha$ -dihydroxy-3-oxochol-4-en-24-oyl-CoA + NADH + H⁺
(2) 7α -hydroxy-3-oxochol-24-oyl-CoA + NAD⁺ = 7α -hydroxy-3-oxochol-4-en-24-oyl-CoA + NADH + H⁺
- Other name(s):** *baiCD* (gene name); 3-oxo-choloyl-CoA dehydrogenase
- Systematic name:** 3-oxocholoyl-CoA Δ^4 -oxidoreductase
- Comments:** Contains flavin. The enzyme, characterized from the bacterium *Clostridium scindens*, participates in the bile acid 7α -dehydroxylation pathway. The enzyme catalyses the stereo-specific oxidation of its substrates and has no activity with the 7β anomers. *cf.* EC 1.3.1.116, 7β -hydroxy-3-oxochol-24-oyl-CoA 4-desaturase.
- References:** [1990]

[EC 1.3.1.115 created 2018]

EC 1.3.1.116

- Accepted name:** 7β -hydroxy-3-oxochol-24-oyl-CoA 4-desaturase
- Reaction:** 7β -hydroxy-3-oxochol-24-oyl-CoA + NAD⁺ = 7β -hydroxy-3-oxochol-4-en-24-oyl-CoA + NADH + H⁺
- Other name(s):** *baiH* (gene name)
- Systematic name:** 7β -hydroxy-3-oxochol-24-oyl-CoA Δ^4 -oxidoreductase
- Comments:** Contains FAD and FMN. The enzyme, characterized from the bacterium *Clostridium scindens*, participates in the bile acid 7α -dehydroxylation pathway. The enzyme catalyses the stereo-specific oxidation of its substrate and has no activity with the 7α anomer. *cf.* EC 1.3.1.115, 3-oxocholoyl-CoA 4-desaturase.
- References:** [223, 1168, 1990]

[EC 1.3.1.116 created 2018]

EC 1.3.1.117

- Accepted name:** hydroxycinnamoyl-CoA reductase
- Reaction:** (1) dihydro-4-coumaroyl-CoA + NADP⁺ = *trans*-4-coumaroyl-CoA + NADPH + H⁺
(2) dihydroferuloyl-CoA + NADP⁺ = *trans*-feruloyl-CoA + NADPH + H⁺
- Other name(s):** MdHCDBR; hydroxycinnamoyl-CoA double bond reductase
- Systematic name:** dihydro-4-coumaroyl-CoA:NADP⁺ 2,3-oxidoreductase
- Comments:** Isolated from *Malus X domestica* (apple). Involved in dihydrochalcone biosynthesis.
- References:** [1784]

[EC 1.3.1.117 created 2018]

EC 1.3.1.118

- Accepted name:** meromycolic acid enoyl-[acyl-carrier-protein] reductase
- Reaction:** a meromycolyl-[acyl-carrier protein] + NAD⁺ = a *trans*- Δ^2 -meromycolyl-[acyl-carrier protein] + NADH + H⁺
- Other name(s):** *inhA* (gene name)
- Systematic name:** meromycolyl-[acyl-carrier protein]:NAD⁺ oxidoreductase
- Comments:** InhA is a component of the fatty acid synthase (FAS) II system of *Mycobacterium tuberculosis*, catalysing an enoyl-[acyl-carrier-protein] reductase step. The enzyme acts on very long and unsaturated fatty acids that form the meromycolic component of mycolic acids. It extends FASI-derived C₂₀ fatty acids to form C₆₀ to C₉₀ mycolic acids. The enzyme, which forms a homotetramer, is the target of the preferred antitubercular drug isoniazid.
- References:** [3415, 3589, 2658, 4451, 1452, 676]

[EC 1.3.1.118 created 2018]

EC 1.3.1.119

- Accepted name:** chlorobenzene dihydrodiol dehydrogenase
Reaction: (1*R*,2*R*)-3-chlorocyclohexa-3,5-diene-1,2-diol + NAD⁺ = 3-chlorocatechol + NADH + H⁺
Other name(s): *tecB* (gene name)
Systematic name: (1*R*,2*R*)-3-chlorocyclohexa-3,5-diene-1,2-diol:NAD⁺ oxidoreductase
Comments: This bacterial enzyme can transform various dihydrodiols of chlorobenzenes into the respective catechols, including the dihydrodiols of mono-, di-, tri-, and tetra-chlorinated benzenes. It also accepts the dihydrodiols of various chlorotoluenes. Substrates for the enzyme are generated by the broad spectrum EC 1.14.12.26, chlorobenzene dioxygenase.
References: [3985, 3347, 3348]

[EC 1.3.1.119 created 2018]

EC 1.3.1.120

- Accepted name:** cyclohexane-1-carbonyl-CoA reductase (NADP⁺)
Reaction: cyclohexane-1-carbonyl-CoA + NADP⁺ = cyclohex-1-ene-1-carbonyl-CoA + NADPH + H⁺
Other name(s): 1-cyclohexenylcarbonyl-CoA reductase (ambiguous); *chcA* (gene name)
Systematic name: cyclohexane-1-carbonyl-CoA:NADP⁺ 1-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Streptomyces collinus*, is involved in a pathway that transforms shikimate to cyclohexane-1-carbonyl-CoA by a series of dehydration and double-bond reduction steps. Most of the steps in this process occur with the carboxylic acid activated as a coenzyme A thioester. The enzyme catalyses three steps in this pathway, also acting on (3*R*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1-carbonyl-CoA and (5*S*)-5-hydroxycyclohex-1-ene-1-carbonyl-CoA.
References: [3506, 4523]

[EC 1.3.1.120 created 2019]

EC 1.3.1.121

- Accepted name:** 4-amino-4-deoxyprephenate dehydrogenase
Reaction: 4-amino-4-deoxyprephenate + NAD⁺ = 3-(4-aminophenyl)pyruvate + CO₂ + NADH + H⁺
Other name(s): *cmlC* (gene name); *papC* (gene name)
Systematic name: 4-amino-4-deoxyprephenate:NAD⁺ oxidoreductase (decarboxylating)
Comments: The enzyme, characterized from the bacteria *Streptomyces venezuelae* and *Streptomyces pristinaespiralis*, participates in the biosynthesis of the antibiotics chloramphenicol and pristinamycin IA, respectively. *cf.* EC 1.3.1.12, prephenate dehydrogenase.
References: [352, 1590]

[EC 1.3.1.121 created 2019]

EC 1.3.1.122

- Accepted name:** (S)-8-oxocitronellyl enol synthase
Reaction: (S)-8-oxocitronellyl enol + NAD(P)⁺ = (6*E*)-8-oxogeranial + NAD(P)H + H⁺
Other name(s): CrISY; 8-oxogeranial:NAD(P)⁺ oxidoreductase (cyclizing, *cis-trans*-nepetalactol forming); iridoid synthase (incorrect)
Systematic name: (S)-8-oxocitronellyl enol:NAD(P)⁺ oxidoreductase
Comments: Isolated from the plants *Catharanthus roseus*, *Olea europaea* (common olive), and several *Nepeta* species. The enzyme reduces 8-oxogeranial, generating an unstable product that is subsequently cyclized into several possible products, either non-enzymically or by dedicated cyclases. The products, known as iridoids, are involved in the biosynthesis of many indole alkaloids. *cf.* EC 1.3.1.123, 7-epi-iridoid synthase.
References: [1312, 1749, 54, 3399, 3846, 2476, 2477]

[EC 1.3.1.122 created 2013 as EC 1.3.1.99, part transferred 2019 to EC 1.3.1.122]

EC 1.3.1.123

- Accepted name:** 8-oxogeranial reductase
Reaction: (R)-8-oxocitronellyl enol + NADP⁺ = (6E)-8-oxogeranial + NADPH + H⁺
Other name(s): *AmISY*
Systematic name: (R)-8-oxocitronellyl enol:NADP⁺ oxidoreductase
Comments: The enzyme, characterized from the plant *Antirrhinum majus* (snapdragon), is involved in biosynthesis of 7-*epi*-iridoids such as antirrhinoside. The enzyme catalyses the stereospecific reduction of 8-oxogeranial, forming an unstable product that in the absence of additional cyclases undergoes spontaneous cyclization to (–)-*cis,trans*-nepetalactol. *cf.* EC 1.3.1.122, (S)-8-oxocitronellyl enol synthase.
References: [2264, 2477]

[EC 1.3.1.123 created 2019]

EC 1.3.1.124

- Accepted name:** 2,4-dienoyl-CoA reductase [(3E)-enoyl-CoA-producing]
Reaction: (1) a (3E)-3-enoyl-CoA + NADP⁺ = a (2E,4E)-2,4-dienoyl-CoA + NADPH + H⁺
(2) a (3E)-3-enoyl-CoA + NADP⁺ = a (2E,4Z)-2,4-dienoyl-CoA + NADPH + H⁺
Other name(s): SPS19 (gene name); DECR1 (gene name); DECR2 (gene name); Δ²,Δ⁴-dienoyl-CoA reductase (ambiguous)
Systematic name: (3E)-3-enoyl-CoA:NADP⁺ 4-oxidoreductase
Comments: This enzyme, characterized from eukaryotic organisms, catalyses the reduction of either (2E,4E)-2,4-dienoyl-CoA or (2E,4Z)-2,4-dienoyl-CoA to (3E)-3-enoyl-CoA. The best substrates for the enzyme from bovine liver have a chain-length of 8 or 10 carbons. Mammals possess both mitochondrial and peroxisomal variants of this enzyme. *cf.* EC 1.3.1.34, 2,4-dienoyl-CoA reductase [(2E)-enoyl-CoA-producing].
References: [2289, 945, 1453, 1297, 3121, 74]

[EC 1.3.1.124 created 2020]

EC 1.3.1.125

- Accepted name:** acrylate reductase
Reaction: propanoate + NAD⁺ = acrylate + NADH + H⁺
Other name(s): *ard* (gene name); NADH:acrylate oxidoreductase
Systematic name: propanoate:NAD⁺ oxidoreductase
Comments: The enzyme, characterized from the marine bacterium *Vibrio harveyi*, enables the organism to utilize acrylate as the terminal electron acceptor for NADH regeneration under anaerobic conditions.
References: [320]

[EC 1.3.1.125 created 2022]

EC 1.3.2 With a cytochrome as acceptor

[1.3.2.1 Transferred entry. butyryl-CoA dehydrogenase. Now EC 1.3.99.2, butyryl-CoA dehydrogenase]

[EC 1.3.2.1 created 1961, deleted 1964]

[1.3.2.2 Transferred entry. acyl-CoA dehydrogenase. Now EC 1.3.99.3, acyl-CoA dehydrogenase]

[EC 1.3.2.2 created 1961, deleted 1964]

EC 1.3.2.3

- Accepted name:** L-galactonolactone dehydrogenase
- Reaction:** L-galactono-1,4-lactone + 4 ferricytochrome *c* = L-dehydroascorbate + 4 ferrocycytochrome *c* + 4 H⁺ (overall reaction)
(1a) L-galactono-1,4-lactone + 2 ferricytochrome *c* = L-ascorbate + 2 ferrocycytochrome *c* + 2 H⁺
(1b) L-ascorbate + 2 ferricytochrome *c* = L-dehydroascorbate + 2 ferrocycytochrome *c* + 2 H⁺ (spontaneous)
- Other name(s):** galactonolactone dehydrogenase; L-galactono- γ -lactone dehydrogenase; L-galactono- γ -lactone:ferricytochrome-*c* oxidoreductase; GLDHase; GLDase
- Systematic name:** L-galactono-1,4-lactone:ferricytochrome-*c* oxidoreductase
- Comments:** This enzyme catalyses the final step in the biosynthesis of L-ascorbic acid in higher plants and in nearly all higher animals with the exception of primates and some birds [3200]. The enzyme is very specific for its substrate L-galactono-1,4-lactone as D-galactono- γ -lactone, D-gulono- γ -lactone, L-gulono- γ -lactone, D-erythronic- γ -lactone, D-xylonic- γ -lactone, L-mannono- γ -lactone, D-galactonate, D-glucuronate and D-gluconate are not substrates [3200]. FAD, NAD⁺, NADP⁺ and O₂ (*cf.* EC 1.3.3.12, L-galactonolactone oxidase) cannot act as electron acceptor [3200].
- References:** [2642, 2643, 1824, 3122, 3200]

[EC 1.3.2.3 created 1961, modified 2006]

EC 1.3.2.4

- Accepted name:** fumarate reductase (cytochrome)
- Reaction:** succinate + 2 ferricytochrome *c* = fumarate + 2 ferrocycytochrome *c*
- Other name(s):** *fccA* (gene name); *fcc3* (gene name); flavocytochrome *c*₃
- Systematic name:** succinate:ferricytochrome-*c* oxidoreductase
- Comments:** Contains a non-covalently bound FAD cofactor and four heme *c* groups. The enzyme, characterized from the bacterium *Shewanella frigidimarina*, is a soluble periplasmic protein that functions as a terminal electron acceptor during anaerobic growth. The direct electron donor is the membrane-bound tetraheme *c*-type cytochrome CymA (EC 7.1.1.8, quinol—cytochrome-*c* reductase), which receives the electrons from the membrane quinol pool.
- References:** [3270, 3271, 1368, 937, 3487, 3764]

[EC 1.3.2.4 created 2022]

EC 1.3.3 With oxygen as acceptor

[1.3.3.1 Transferred entry. dihydroorotate oxidase. Now EC 1.3.98.1 [dihydroorotate dehydrogenase (fumarate)]]

[EC 1.3.3.1 created 1961, deleted 2011]

[1.3.3.2 Transferred entry. now EC 1.14.21.6, lathosterol oxidase. NAD(P)H had not been included previously, so enzyme had to be reclassified]

[EC 1.3.3.2 created 1972, deleted 2005]

EC 1.3.3.3

- Accepted name:** coproporphyrinogen oxidase
- Reaction:** coproporphyrinogen III + O₂ + 2 H⁺ = protoporphyrinogen-IX + 2 CO₂ + 2 H₂O
- Other name(s):** coproporphyrinogen III oxidase; coproporphyrinogenase
- Systematic name:** coproporphyrinogen:oxygen oxidoreductase (decarboxylating)
- References:** [239, 2759, 2205]

[EC 1.3.3.3 created 1972, modified 2003]

EC 1.3.3.4

Accepted name: protoporphyrinogen oxidase
Reaction: protoporphyrinogen IX + 3 O₂ = protoporphyrin IX + 3 H₂O₂
Other name(s): protoporphyrinogen IX oxidase; protoporphyrinogenase; PPO; Protox; HemG; HemY
Systematic name: protoporphyrinogen-IX:oxygen oxidoreductase
Comments: This is the last common enzyme in the biosynthesis of chlorophylls and heme [622]. Two isoenzymes exist in plants: one in plastids and the other in mitochondria. This is the target enzyme of phthalimide-type and diphenylether-type herbicides [622]. The enzyme from oxygen-dependent species contains FAD [809]. Also slowly oxidizes mesoporphyrinogen IX.
References: [3361, 3362, 805, 4518, 739, 1109, 808, 622, 809]

[EC 1.3.3.4 created 1978, modified 2003]

EC 1.3.3.5

Accepted name: bilirubin oxidase
Reaction: 2 bilirubin + O₂ = 2 biliverdin + 2 H₂O
Other name(s): bilirubin oxidase M-1
Systematic name: bilirubin:oxygen oxidoreductase
References: [2942, 4196]

[EC 1.3.3.5 created 1984]

EC 1.3.3.6

Accepted name: acyl-CoA oxidase
Reaction: acyl-CoA + O₂ = *trans*-2,3-dehydroacyl-CoA + H₂O₂
Other name(s): fatty acyl-CoA oxidase; acyl coenzyme A oxidase; fatty acyl-coenzyme A oxidase
Systematic name: acyl-CoA:oxygen 2-oxidoreductase
Comments: A flavoprotein (FAD). Acts on CoA derivatives of fatty acids with chain lengths from 8 to 18.
References: [2041, 3203]

[EC 1.3.3.6 created 1986]

EC 1.3.3.7

Accepted name: dihydrouracil oxidase
Reaction: 5,6-dihhydrouracil + O₂ = uracil + H₂O₂
Systematic name: 5,6-dihhydrouracil:oxygen oxidoreductase
Comments: Also oxidizes dihydrothymine to thymine. A flavoprotein (FMN).
References: [3218]

[EC 1.3.3.7 created 1989]

EC 1.3.3.8

Accepted name: tetrahydroberberine oxidase
Reaction: (*S*)-tetrahydroberberine + 2 O₂ = berberine + 2 H₂O₂
Other name(s): (*S*)-THB oxidase
Systematic name: (*S*)-tetrahydroberberine:oxygen oxidoreductase
Comments: The enzyme from *Berberis* sp. is a flavoprotein; that from *Coptis japonica* is not. (*R*)-Tetrahydroberberines are not oxidized.
References: [76, 3154]

[EC 1.3.3.8 created 1990 (EC 1.5.3.8 created 1989, incorporated 1992)]

[1.3.3.9 Transferred entry. *secologanin synthase*. Now EC 1.14.19.62, *secologanin synthase*]

[EC 1.3.3.9 created 2002, deleted 2018]

EC 1.3.3.10

Accepted name: tryptophan α,β -oxidase
Reaction: L-tryptophan + O₂ = α,β -didehydrotryptophan + H₂O₂
Other name(s): L-tryptophan 2',3'-oxidase; L-tryptophan α,β -dehydrogenase
Systematic name: L-tryptophan:oxygen α,β -oxidoreductase
Comments: Requires heme. The enzyme from *Chromobacterium violaceum* is specific for tryptophan derivatives possessing its carboxyl group free or as an amide or ester, and an unsubstituted indole ring. Also catalyses the α,β dehydrogenation of L-tryptophan side chains in peptides. The product of the reaction can hydrolyse spontaneously to form (indol-3-yl)pyruvate.
References: [1299, 1298]

[EC 1.3.3.10 created 2000 as EC 1.4.3.17, transferred 2003 to EC 1.3.3.10]

EC 1.3.3.11

Accepted name: pyrroloquinoline-quinone synthase
Reaction: 6-(2-amino-2-carboxyethyl)-7,8-dioxo-1,2,3,4,7,8-hexahydroquinoline-2,4-dicarboxylate + 3 O₂ = 4,5-dioxo-4,5-dihydro-1*H*-pyrrolo[2,3-*f*]quinoline-2,7,9-tricarboxylate + 2 H₂O₂ + 2 H₂O
Other name(s): PqqC; 6-(2-amino-2-carboxyethyl)-7,8-dioxo-1,2,3,4,5,6,7,8-octahydroquinoline-2,4-dicarboxylate:oxygen oxidoreductase (cyclizing) [incorrect]
Systematic name: 6-(2-amino-2-carboxyethyl)-7,8-dioxo-1,2,3,4,7,8-hexahydroquinoline-2,4-dicarboxylate:oxygen oxidoreductase (cyclizing)
Comments: So far only a single turnover of the enzyme has been observed, and the pyrroloquinoline quinone remains bound to it. It is not yet known what releases the product in the bacterium.
References: [2610, 2609, 4318, 4320, 3770]

[EC 1.3.3.11 created 2005]

EC 1.3.3.12

Accepted name: L-galactonolactone oxidase
Reaction: L-galactono-1,4-lactone + O₂ = L-ascorbate + H₂O₂
Other name(s): L-galactono-1,4-lactone oxidase
Systematic name: L-galactono-1,4-lactone:oxygen 3-oxidoreductase
Comments: A flavoprotein. Acts on the 1,4-lactones of L-galactonic, D-altronic, L-fuconic, D-arabinic and D-threonic acids; not identical with EC 1.1.3.8 L-gulonolactone oxidase. (*cf.* EC 1.3.2.3 galactonolactone dehydrogenase).
References: [358]

[EC 1.3.3.12 created 1984 as EC 1.1.3.24, transferred 2006 to EC 1.3.3.12]

EC 1.3.3.13

Accepted name: albonoursin synthase
Reaction: cyclo(L-leucyl-L-phenylalanyl) + 2 O₂ = albonoursin + 2 H₂O₂ (overall reaction)
(1a) cyclo(L-leucyl-L-phenylalanyl) + O₂ = cyclo[(*Z*)- α,β -didehydrophenylalanyl-L-leucyl] + H₂O₂
(1b) cyclo[(*Z*)- α,β -didehydrophenylalanyl-L-leucyl] + O₂ = albonoursin + H₂O₂
Other name(s): cyclo(dipeptide):oxygen oxidoreductase; cyclic dipeptide oxidase; AlbA
Systematic name: cyclo(L-leucyl-L-phenylalanyl):oxygen oxidoreductase
Comments: A flavoprotein from the bacterium *Streptomyces noursei*. The enzyme can also oxidize several other cyclo dipeptides, the best being cyclo(L-tryptophyl-L-tryptophyl) and cyclo(L-phenylalanyl-L-phenylalanyl) [1358, 2366].
References: [1358, 2366]

[EC 1.3.3.13 created 2013]

EC 1.3.3.14

- Accepted name:** aclacinomycin-A oxidase
Reaction: aclacinomycin A + O₂ = aclacinomycin Y + H₂O₂
Other name(s): AknOx (ambiguous); aclacinomycin oxidoreductase (ambiguous)
Systematic name: aclacinomycin-A:oxygen oxidoreductase
Comments: A flavoprotein (FAD). This bifunctional enzyme is a secreted flavin-dependent enzyme that is involved in the modification of the terminal sugar residues in the biosynthesis of aclacinomycins. The enzyme utilizes the same active site to catalyse the oxidation of the rhodnose moiety of aclacinomycin N to the cinerulose A moiety of aclacinomycin A (*cf.* EC 1.1.3.45) and the oxidation of the latter to the L-aculose moiety of aclacinomycin Y.
References: [4817, 66, 4112]

[EC 1.3.3.14 created 2013]

EC 1.3.3.15

- Accepted name:** coproporphyrinogen III oxidase (coproporphyrin-forming)
Reaction: coproporphyrinogen III + 3 O₂ = coproporphyrin III + 3 H₂O₂
Other name(s): *hemY* (gene name)
Systematic name: coproporphyrinogen-III:oxygen oxidoreductase (coproporphyrin-forming)
Comments: Contains FAD. The enzyme, present in Gram-positive bacteria, participates in heme biosynthesis. It can also catalyse the reaction of EC 1.3.3.4, protoporphyrinogen oxidase, at a lower level.
References: [1514, 739, 3400, 806]

[EC 1.3.3.15 created 2016]

EC 1.3.3.16

- Accepted name:** oxazoline dehydrogenase
Reaction: (1) a [protein]-(1*S*,4*R*)-2-(C-substituted-aminomethyl)-4-acyl-2-thiazoline + O₂ = a [protein]-(*S*)-2-(C-substituted-aminomethyl)-4-acyl-1,3-thiazole + H₂O₂
(2) a [protein]-(*S*,*S*)-2-(C-substituted-aminomethyl)-4-acyl-2-oxazoline + O₂ = a [protein]-(*S*)-2-(C-substituted-aminomethyl)-4-acyl-1,3-oxazole + H₂O₂
(3) a [protein]-(*S*,*S*)-2-(C-substituted-aminomethyl)-4-acyl-5-methyl-2-oxazoline + O₂ = a [protein]-(*S*)-2-(C-substituted-aminomethyl)-4-acyl-5-methyl-1,3-oxazole + H₂O₂
Other name(s): azoline oxidase; thiazoline oxidase; cyanobactin oxidase; *patG* (gene name); *mcaG* (gene name); *artG* (gene name); *lynG* (gene name); *tenG* (gene name)
Systematic name: a [protein]-2-oxazoline:oxygen oxidoreductase (2-oxazole-forming)
Comments: Contains FMN. This enzyme oxidizes 2-oxazoline, 5-methyl-2-oxazoline, and 2-thiazoline within peptides, which were formed by EC 6.2.2.2, oxazoline synthase, and EC 6.2.2.3, thiazoline synthase, to the respective pyrrole-type rings. The enzyme is found as either a stand-alone protein or as a domain within a multifunctional protein (the G protein) that also functions as a peptidase.
References: [2468, 3725, 290, 1314]

[EC 1.3.3.16 created 2020]

EC 1.3.3.17

- Accepted name:** benzylmalonyl-CoA dehydrogenase
Reaction: benzylmalonyl-CoA + O₂ = (*E*)-cinnamoyl-CoA + CO₂ + H₂O₂
Other name(s): *iaaF* (gene name)
Systematic name: benzylmalonyl-CoA:oxygen oxidoreductase (decarboxylating)
Comments: The enzyme, characterized from the bacterium *Aromatoleum aromaticum*, is involved in degradation of (indol-3-yl)acetate, where it is believed to function on (2-aminobenzyl)malonyl-CoA.
References: [3750]

[EC 1.3.3.17 created 2022]

EC 1.3.4 With a disulfide as acceptor

EC 1.3.4.1

- Accepted name:** fumarate reductase (CoM/CoB)
Reaction: fumarate + CoM + CoB = succinate + CoM-S-S-CoB
Other name(s): thiol:fumarate reductase; Tfr
Systematic name: fumarate CoM:CoB oxidoreductase (succinate-forming)
Comments: The enzyme, isolated from the archaeon *Methanobacterium thermoautotrophicum*, is very oxygen sensitive. It cannot use reduced flavins, reduced coenzyme F₄₂₀, or NAD(P)H as an electron donor. Distinct from EC 1.3.1.6 [fumarate reductase (NADH)], EC 1.3.5.1 [succinate dehydrogenase (ubiquinone)], and EC 1.3.5.4 [fumarate reductase (quinol)].
References: [2078, 1611]

[EC 1.3.4.1 created 2014 as EC 1.3.98.2, transferred 2014 to EC 1.3.4.1]

EC 1.3.5 With a quinone or related compound as acceptor

EC 1.3.5.1

- Accepted name:** succinate dehydrogenase
Reaction: succinate + a quinone = fumarate + a quinol
Other name(s): succinate dehydrogenase (quinone); succinate dehydrogenase (ubiquinone); succinic dehydrogenase; complex II (ambiguous); succinate dehydrogenase complex; SDH (ambiguous); succinate:ubiquinone oxidoreductase; fumarate reductase (quinol); FRD; menaquinol-fumarate oxidoreductase; succinate dehydrogenase (menaquinone); succinate:menaquinone oxidoreductase; fumarate reductase (menaquinone)
Systematic name: succinate:quinone oxidoreductase
Comments: A complex generally comprising an FAD-containing component that also binds the carboxylate substrate (A subunit), a component that contains three different iron-sulfur centers [2Fe-2S], [4Fe-4S], and [3Fe-4S] (B subunit), and a hydrophobic membrane-anchor component (C, or C and D subunits) that is also the site of the interaction with quinones. The enzyme is found in the inner mitochondrial membrane in eukaryotes and the plasma membrane of bacteria and archaea, with the hydrophilic domain extending into the mitochondrial matrix and the cytoplasm, respectively. Under aerobic conditions the enzyme catalyses succinate oxidation, a key step in the citric acid (TCA) cycle, transferring the electrons to quinones in the membrane, thus linking the TCA cycle with the aerobic respiratory chain (where it is known as complex II). Under anaerobic conditions the enzyme functions as a fumarate reductase, transferring electrons from the quinol pool to fumarate, and participating in anaerobic respiration with fumarate as the terminal electron acceptor. The enzyme interacts with the quinone produced by the organism, such as ubiquinone, menaquinone, caldariellaquinone, thermoplasmaquinone, rhodoquinone etc. Some of the enzymes contain two heme subunits in their membrane anchor subunit. These enzymes catalyse an electrogenic reaction and are thus classified as EC 7.1.1.12, succinate dehydrogenase (electrogenic, proton-motive force generating).
References: [2132, 1617, 1855, 583, 1120, 582, 3219, 2303, 1862]

[EC 1.3.5.1 created 1983 (EC 1.3.99.1 created 1961, incorporated 2014, EC 1.3.5.4 created 2010, incorporated 2022), modified 2022]

EC 1.3.5.2

- Accepted name:** dihydroorotate dehydrogenase (quinone)
Reaction: (S)-dihydroorotate + a quinone = orotate + a quinol
Other name(s): dihydroorotate:ubiquinone oxidoreductase; (S)-dihydroorotate:(acceptor) oxidoreductase; (S)-dihydroorotate:acceptor oxidoreductase; DHODhase (ambiguous); DHOD (ambiguous); DHODase (ambiguous); DHODH
Systematic name: (S)-dihydroorotate:quinone oxidoreductase

Comments: This Class 2 dihydroorotate dehydrogenase enzyme contains FMN [1078]. The enzyme is found in eukaryotes in the mitochondrial membrane, in cyanobacteria, and in some Gram-negative and Gram-positive bacteria associated with the cytoplasmic membrane [2,5,6]. The reaction is the only redox reaction in the *de-novo* biosynthesis of pyrimidine nucleotides [1662, 1078]. The best quinone electron acceptors for the enzyme from bovine liver are ubiquinone-6 and ubiquinone-7, although simple quinones, such as benzoquinone, can also act as acceptor at lower rates [1662]. Methyl-, ethyl-, *tert*-butyl and benzyl (*S*)-dihydroorotates are also substrates, but methyl esters of (*S*)-1-methyl and (*S*)-3-methyl and (*S*)-1,3-dimethyldihydroorotates are not [1662]. Class 1 dihydroorotate dehydrogenases use either fumarate (EC 1.3.98.1), NAD⁺ (EC 1.3.1.14) or NADP⁺ (EC 1.3.1.15) as electron acceptor.

References: [1142, 1662, 174, 1078, 341, 3010]

[EC 1.3.5.2 created 1983 as EC 1.3.99.11, transferred 2009 to EC 1.3.5.2, modified 2011]

EC 1.3.5.3

Accepted name: protoporphyrinogen IX dehydrogenase (quinone)
Reaction: protoporphyrinogen IX + 3 quinone = protoporphyrin IX + 3 quinol
Other name(s): HemG; protoporphyrinogen IX dehydrogenase (menaquinone)
Systematic name: protoporphyrinogen IX:quinone oxidoreductase
Comments: Contains FMN. The enzyme participates in heme *b* biosynthesis. In the bacterium *Escherichia coli* it interacts with either ubiquinone or menaquinone, depending on whether the organism grows aerobically or anaerobically.
References: [413, 2853]

[EC 1.3.5.3 created 2010, modified 2020]

[1.3.5.4 Transferred entry. fumarate reductase (quinol), now included in EC 1.3.5.1, succinate dehydrogenase.]

[EC 1.3.5.4 created 2010, modified 2013, deleted 2022]

EC 1.3.5.5

Accepted name: 15-*cis*-phytoene desaturase
Reaction: 15-*cis*-phytoene + 2 plastoquinone = 9,15,9'-*tricus*-ζ-carotene + 2 plastoquinol (overall reaction)
(1a) 15-*cis*-phytoene + plastoquinone = 15,9'-*dicis*-phytofluene + plastoquinol
(1b) 15,9'-*dicis*-phytofluene + plastoquinone = 9,15,9'-*tricus*-ζ-carotene + plastoquinol
Other name(s): phytoene desaturase (ambiguous); PDS; plant-type phytoene desaturase
Systematic name: 15-*cis*-phytoene:plastoquinone oxidoreductase
Comments: This enzyme is involved in carotenoid biosynthesis in plants and cyanobacteria. The enzyme from *Synechococcus* can also use NAD⁺ and NADP⁺ as electron acceptor under anaerobic conditions. The enzyme from *Gentiana lutea* shows no activity with NAD⁺ or NADP⁺ [437].
References: [437, 3734, 1169, 436]

[EC 1.3.5.5 created 2011]

EC 1.3.5.6

Accepted name: 9,9'-*dicis*-ζ-carotene desaturase
Reaction: 9,9'-*dicis*-ζ-carotene + 2 quinone = 7,9,7',9'-*tetracus*-lycopene + 2 quinol (overall reaction)
(1a) 9,9'-*dicis*-ζ-carotene + a quinone = 7,9,9'-*tricus*-neurosporene + a quinol
(1b) 7,9,9'-*tricus*-neurosporene + a quinone = 7,9,7',9'-*tetracus*-lycopene + a quinol
Other name(s): ζ-carotene desaturase; ZDS
Systematic name: 9,9'-*dicis*-ζ-carotene:quinone oxidoreductase
Comments: This enzyme is involved in carotenoid biosynthesis in plants and cyanobacteria.
References: [61, 1954, 434, 436]

[EC 1.3.5.6 created 1999 as EC 1.14.99.30, transferred 2011 to EC 1.3.5.6]

EC 1.3.7 With an iron-sulfur protein as acceptor

EC 1.3.7.1

Accepted name: 6-hydroxynicotinate reductase
Reaction: 6-oxo-1,4,5,6-tetrahydronicotinate + oxidized ferredoxin = 6-hydroxynicotinate + reduced ferredoxin
Other name(s): 6-oxotetrahydronicotinate dehydrogenase; 6-hydroxynicotinic reductase; HNA reductase; 1,4,5,6-tetrahydro-6-oxonicotinate:ferredoxin oxidoreductase
Systematic name: 6-oxo-1,4,5,6-tetrahydronicotinate:ferredoxin oxidoreductase
References: [1691]

[EC 1.3.7.1 created 1972]

EC 1.3.7.2

Accepted name: 15,16-dihydrobiliverdin:ferredoxin oxidoreductase
Reaction: 15,16-dihydrobiliverdin + oxidized ferredoxin = biliverdin IX α + reduced ferredoxin
Other name(s): PebA
Systematic name: 15,16-dihydrobiliverdin:ferredoxin oxidoreductase
Comments: Catalyses the two-electron reduction of biliverdin IX α at the C15 methine bridge. It has been proposed that this enzyme and EC 1.3.7.3, phycoerythrobilin:ferredoxin oxidoreductase, function as a dual enzyme complex in the conversion of biliverdin IX α into phycoerythrobilin.
References: [1167]

[EC 1.3.7.2 created 2002]

EC 1.3.7.3

Accepted name: phycoerythrobilin:ferredoxin oxidoreductase
Reaction: (3Z)-phycoerythrobilin + oxidized ferredoxin = 15,16-dihydrobiliverdin + reduced ferredoxin
Other name(s): PebB
Systematic name: (3Z)-phycoerythrobilin:ferredoxin oxidoreductase
Comments: Catalyses the two-electron reduction of the C2 and C3¹ diene system of 15,16-dihydrobiliverdin. Specific for 15,16-dihydrobiliverdin. It has been proposed that this enzyme and EC 1.3.7.2, 15,16-dihydrobiliverdin:ferredoxin oxidoreductase, function as a dual enzyme complex in the conversion of biliverdin IX α to phycoerythrobilin.
References: [1167]

[EC 1.3.7.3 created 2002]

EC 1.3.7.4

Accepted name: phytochromobilin:ferredoxin oxidoreductase
Reaction: (3Z)-phytochromobilin + 2 oxidized ferredoxin = biliverdin IX α + 2 reduced ferredoxin
Other name(s): HY2; PPhi B synthase; phytochromobilin synthase
Systematic name: (3Z)-phytochromobilin:ferredoxin oxidoreductase
Comments: Catalyses the two-electron reduction of biliverdin IX α . Can use [2Fe-2S] ferredoxins from a number of sources as acceptor but not the [4Fe-4S] ferredoxin from *Clostridium pasteurianum*. The isomerization of (3Z)-phytochromobilin to (3E)-phytochromobilin is thought to occur prior to covalent attachment to apophytochrome in the plant cell cytoplasm. Flavodoxins can be used instead of ferredoxin.
References: [1167, 2747, 4247]

[EC 1.3.7.4 created 2002]

EC 1.3.7.5

- Accepted name:** phycocyanobilin:ferredoxin oxidoreductase
Reaction: (3Z)-phycocyanobilin + 4 oxidized ferredoxin = biliverdin IX α + 4 reduced ferredoxin
Systematic name: (3Z)-phycocyanobilin:ferredoxin oxidoreductase
Comments: Catalyses the four-electron reduction of biliverdin IX α (2-electron reduction at both the A and D rings). Reaction proceeds via an isolatable 2-electron intermediate, 18¹,18²-dihydrobiliverdin. Flavodoxins can be used instead of ferredoxin. The direct conversion of biliverdin IX α (BV) to (3Z)-phycocyanobilin (PCB) in the cyanobacteria *Synechocystis* sp. PCC 6803, *Anabaena* sp. PCC7120 and *Nostoc punctiforme* is in contrast to the proposed pathways of PCB biosynthesis in the red alga *Cyanidium caldarium*, which involves (3Z)-phycoerythrobilin (PEB) as an intermediate [253] and in the green alga *Mesotaenium caldarium*, in which PCB is an isolable intermediate.
References: [1167, 253, 4678]

[EC 1.3.7.5 created 2002, modified 2014]

EC 1.3.7.6

- Accepted name:** phycoerythrobilin synthase
Reaction: (3Z)-phycoerythrobilin + 2 oxidized ferredoxin = biliverdin IX α + 2 reduced ferredoxin
Other name(s): PebS
Systematic name: (3Z)-phycoerythrobilin:ferredoxin oxidoreductase (from biliverdin IX α)
Comments: This enzyme, from a cyanophage infecting oceanic cyanobacteria of the *Prochlorococcus* genus, uses a four-electron reduction to carry out the reactions catalysed by EC 1.3.7.2 (15,16-dihydrobiliverdin:ferredoxin oxidoreductase) and EC 1.3.7.3 (phycoerythrobilin:ferredoxin oxidoreductase). 15,16-Dihydrobiliverdin is formed as a bound intermediate. Free 15,16-dihydrobiliverdin can also act as a substrate to form phycoerythrobilin.
References: [815]

[EC 1.3.7.6 created 2008]

EC 1.3.7.7

- Accepted name:** ferredoxin:protochlorophyllide reductase (ATP-dependent)
Reaction: chlorophyllide *a* + oxidized ferredoxin + 2 ADP + 2 phosphate = protochlorophyllide *a* + reduced ferredoxin + 2 ATP + 2 H₂O
Other name(s): light-independent protochlorophyllide reductase
Systematic name: ATP-dependent ferredoxin:protochlorophyllide-*a* 7,8-oxidoreductase
Comments: Occurs in photosynthetic bacteria, cyanobacteria, green algae and gymnosperms. The enzyme catalyses *trans*-reduction of the D-ring of protochlorophyllide; the product has the (7*S*,8*S*)-configuration. Unlike EC 1.3.1.33 (protochlorophyllide reductase), light is not required. The enzyme contains a [4Fe-4S] cluster, and structurally resembles the Fe protein/MoFe protein complex of nitrogenase (EC 1.18.6.1), which catalyses an ATP-driven reduction.
References: [1214, 3101, 2940]

[EC 1.3.7.7 created 2011, modified 2013]

EC 1.3.7.8

- Accepted name:** benzoyl-CoA reductase
Reaction: cyclohexa-1,5-diene-1-carbonyl-CoA + oxidized ferredoxin + 2 ADP + 2 phosphate = benzoyl-CoA + reduced ferredoxin + 2 ATP + 2 H₂O
Other name(s): benzoyl-CoA reductase (dearomatizing)
Systematic name: cyclohexa-1,5-diene-1-carbonyl-CoA:ferredoxin oxidoreductase (aromatizing, ATP-forming)

Comments: An iron-sulfur protein. Requires Mg²⁺ or Mn²⁺. Inactive towards aromatic acids that are not CoA esters but will also catalyse the reaction: ammonia + acceptor + 2 ADP + 2 phosphate = hydroxylamine + reduced acceptor + 2 ATP + H₂O. In the presence of reduced acceptor, but in the absence of oxidizable substrate, the enzyme catalyses the hydrolysis of ATP to ADP plus phosphate.

References: [378, 2290]

[EC 1.3.7.8 created 1999 as EC 1.3.99.15, transferred 2011 to EC 1.3.7.8, modified 2011]

[1.3.7.9 *Transferred entry. 4-hydroxybenzoyl-CoA reductase. Now classified as EC 1.1.7.1, 4-hydroxybenzoyl-CoA reductase.*]

[EC 1.3.7.9 created 2000 as EC 1.3.99.20, transferred 2011 to EC 1.3.7.9, deleted 2020]

[1.3.7.10 *Transferred entry. pentalenolactone synthase. Now EC 1.14.19.8, pentalenolactone synthase*]

[EC 1.3.7.10 created 2012, deleted 2013]

EC 1.3.7.11

Accepted name: 2,3-bis-*O*-geranylgeranyl-*sn*-glycero-phospholipid reductase
Reaction: a 2,3-bis-(*O*-phytanyl)-*sn*-glycero-phospholipid + **16** oxidized ferredoxin [iron-sulfur] cluster = a 2,3-bis-(*O*-geranylgeranyl)-*sn*-glycero-phospholipid + **16** reduced ferredoxin [iron-sulfur] cluster + **16** H⁺
Other name(s): AF0464 (gene name); 2,3-bis-*O*-geranylgeranyl-*sn*-glycerol 1-phosphate reductase (donor)
Systematic name: 2,3-bis-(*O*-phytanyl)-*sn*-glycero-phospholipid:ferredoxin oxidoreductase
Comments: A flavoprotein (FAD). The enzyme is involved in the biosynthesis of archaeal membrane lipids. It catalyses the reduction of all 8 double bonds in 2,3-bis-*O*-geranylgeranyl-*sn*-glycero-phospholipids and all 4 double bonds in 3-*O*-geranylgeranyl-*sn*-glycerol phospholipids with comparable activity. Unlike EC 1.3.1.101, 2,3-bis-*O*-geranylgeranyl-*sn*-glycerol 1-phosphate reductase [NAD(P)H], this enzyme shows no activity with NADPH, and requires a dedicated ferredoxin [1837].
References: [2939, 3670, 3667, 1837]

[EC 1.3.7.11 created 2013 as EC 1.3.99.34, transferred 2015 to EC 1.3.7.11]

EC 1.3.7.12

Accepted name: red chlorophyll catabolite reductase
Reaction: primary fluorescent chlorophyll catabolite + **2** oxidized ferredoxin [iron-sulfur] cluster = red chlorophyll catabolite + **2** reduced ferredoxin [iron-sulfur] cluster + **2** H⁺
Other name(s): RCCR; RCC reductase; red Chl catabolite reductase
Systematic name: primary fluorescent chlorophyll catabolite:ferredoxin oxidoreductase
Comments: The enzyme participates in chlorophyll degradation, which occurs during leaf senescence and fruit ripening in higher plants. The reaction requires reduced ferredoxin, which is generated from NADPH produced either through the pentose-phosphate pathway or by the action of photosystem I [3545, 4688]. This reaction takes place while red chlorophyll catabolite is still bound to EC 1.14.15.17, pheophorbide *a* oxygenase [3389]. Depending on the plant species used as the source of enzyme, one of two possible C-1 epimers of primary fluorescent chlorophyll catabolite (pFCC), pFCC-1 or pFCC-2, is normally formed, with all genera or species within a family producing the same isomer [3389, 1733]. After modification and export, pFCCs are eventually imported into the vacuole, where the acidic environment causes their non-enzymic conversion into colourless breakdown products called non-fluorescent chlorophyll catabolites (NCCs) [4688].
References: [3545, 4688, 3389, 1733, 3546]

[EC 1.3.7.12 created 2007 as EC 1.3.1.80, transferred 2016 to EC 1.3.7.12]

EC 1.3.7.13

Accepted name: 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (ferredoxin)

Reaction: protochlorophyllide *a* + 2 oxidized ferredoxin [iron-sulfur] cluster = 3,8-divinyl protochlorophyllide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺
Other name(s): *bciB* (gene name); cyano-type divinyl chlorophyllide *a* 8-vinyl-reductase
Systematic name: protochlorophyllide-*a*:ferredoxin C-8¹-oxidoreductase
Comments: The enzyme, found in many phototrophic bacteria, land plants, and some green and red algae, is involved in the production of monovinyl versions of (bacterio)chlorophyll pigments from their divinyl precursors. Binds two [4Fe-4S] clusters and an FAD cofactor. It can also act on 3,8-divinyl chlorophyllide *a*, 3,8-divinyl chlorophyll *a*, and chlorophyll *c*₂. *cf.* EC 1.3.1.75, 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (NADPH).
References: [648, 3676, 1845]

[EC 1.3.7.13 created 2016]

EC 1.3.7.14

Accepted name: 3,8-divinyl chlorophyllide *a* reductase
Reaction: bacteriochlorophyllide *g* + 2 oxidized ferredoxin [iron-sulfur] cluster + ADP + phosphate = 3,8-divinyl chlorophyllide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + ATP + H₂O + 2 H⁺
Systematic name: bacteriochlorophyllide-*g*:ferredoxin C-8¹-oxidoreductase
Comments: The enzyme, found only in bacteriochlorophyll *b*-producing bacteria, catalyses the introduction of a C-8 ethylidene group. The enzyme contains a [4Fe-4S] cluster, and structurally resembles the Fe protein/MoFe protein complex of nitrogenase. It is very similar to EC 1.3.7.15, chlorophyllide *a* reductase, and is composed of three subunits. Two of them form the catalytic component, while the third one functions as an ATP-dependent reductase component that catalyses the electron transfer from ferredoxin to the catalytic component.
References: [4346, 4345]

[EC 1.3.7.14 created 2016]

EC 1.3.7.15

Accepted name: chlorophyllide *a* reductase
Reaction: (1) 3-deacetyl-3-vinylbacteriochlorophyllide *a* + 2 oxidized ferredoxin [iron-sulfur] cluster + ADP + phosphate = chlorophyllide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + ATP + H₂O + 2 H⁺
(2) bacteriochlorophyllide *a* + 2 oxidized ferredoxin [iron-sulfur] cluster + ADP + phosphate = 3-acetyl-3-devinylchlorophyllide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + ATP + H₂O + 2 H⁺
(3) 3-deacetyl-3-(1-hydroxyethyl)bacteriochlorophyllide *a* + 2 oxidized ferredoxin [iron-sulfur] cluster + ADP + phosphate = 3-devinyl-3-(1-hydroxyethyl)chlorophyllide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + ATP + H₂O + 2 H⁺
Other name(s): *bchX* (gene name); *bchY* (gene name); *bchZ* (gene name); COR
Systematic name: bacteriochlorophyllide-*a*:ferredoxin 7,8-oxidoreductase
Comments: The enzyme, together with EC 1.1.1.396, bacteriochlorophyllide-*a* dehydrogenase, and EC 4.2.1.165, chlorophyllide-*a* 3¹-hydratase, is involved in the conversion of chlorophyllide *a* to bacteriochlorophyllide *a*. These enzymes can act in multiple orders, resulting in the formation of different intermediates, but the final product of the cumulative action of the three enzymes is always bacteriochlorophyllide *a*. This enzyme catalyses a *trans*-reduction of the B-ring; the product has the (7*R*,8*R*)-configuration. In addition, the enzyme has a latent activity of EC 1.3.7.13, 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (ferredoxin) [1527]. The enzyme contains a [4Fe-4S] cluster, and structurally resembles the Fe protein/MoFe protein complex of nitrogenase (EC 1.18.6.1), which catalyses an ATP-driven reduction.
References: [3100, 4346, 2345, 1527]

[EC 1.3.7.15 created 1965 as EC 1.3.99.35, modified 2012, transferred 2016 to EC 1.3.7.15]

EC 1.3.8 With a flavin as acceptor

EC 1.3.8.1

- Accepted name:** short-chain acyl-CoA dehydrogenase
Reaction: a short-chain acyl-CoA + electron-transfer flavoprotein = a short-chain *trans*-2,3-dehydroacyl-CoA + reduced electron-transfer flavoprotein
Other name(s): butyryl-CoA dehydrogenase; butanoyl-CoA dehydrogenase; butyryl dehydrogenase; unsaturated acyl-CoA reductase; ethylene reductase; enoyl-coenzyme A reductase; unsaturated acyl coenzyme A reductase; butyryl coenzyme A dehydrogenase; short-chain acyl CoA dehydrogenase; short-chain acyl-coenzyme A dehydrogenase; 3-hydroxyacyl CoA reductase; butanoyl-CoA:(acceptor) 2,3-oxidoreductase; ACADS (gene name).
Systematic name: short-chain acyl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: Contains FAD as prosthetic group. One of several enzymes that catalyse the first step in fatty acids β -oxidation. The enzyme catalyses the oxidation of saturated short-chain acyl-CoA thioesters to give a *trans* 2,3-unsaturated product by removal of the two *pro-R*-hydrogen atoms. The enzyme from beef liver accepts substrates with acyl chain lengths of 3 to 8 carbon atoms. The highest activity was reported with either butanoyl-CoA [1396] or pentanoyl-CoA [3834]. The enzyme from rat has only 10% activity with hexanoyl-CoA (compared to butanoyl-CoA) and no activity with octanoyl-CoA [1795]. *cf.* EC 1.3.8.7, medium-chain acyl-CoA dehydrogenase, EC 1.3.8.8, long-chain acyl-CoA dehydrogenase, and EC 1.3.8.9, very-long-chain acyl-CoA dehydrogenase.
References: [2613, 1396, 275, 3834, 4277, 1795, 2754]

[EC 1.3.8.1 created 1961 as EC 1.3.2.1, transferred 1964 to EC 1.3.99.2, transferred 2011 to EC 1.3.8.1, modified 2012]

EC 1.3.8.2

- Accepted name:** 4,4'-diapophytoene desaturase (4,4'-diapolycopene-forming)
Reaction: 15-*cis*-4,4'-diapophytoene + 4 FAD = *all-trans*-4,4'-diapolycopene + 4 FADH₂ (overall reaction)
(1a) 15-*cis*-4,4'-diapophytoene + FAD = *all-trans*-4,4'-diapophytofluene + FADH₂
(1b) *all-trans*-4,4'-diapophytofluene + FAD = *all-trans*-4,4'-diapo- ζ -carotene + FADH₂
(1c) *all-trans*-4,4'-diapo- ζ -carotene + FAD = *all-trans*-4,4'-diaponeurosporene + FADH₂
(1d) *all-trans*-4,4'-diaponeurosporene + FAD = *all-trans*-4,4'-diapolycopene + FADH₂
Other name(s): dehydrosqualene desaturase (ambiguous); CrtN (ambiguous); 4,4'-diapophytoene:FAD oxidoreductase (ambiguous); 15-*cis*-4,4'-diapophytoene:FAD oxidoreductase; 4,4'-diapophytoene desaturase (ambiguous)
Systematic name: 15-*cis*-4,4'-diapophytoene:FAD oxidoreductase (4,4'-diapolycopene-forming)
Comments: The enzyme catalyses four successive dehydrogenations, resulting in production of 4,4'-diapolycopene. While the enzyme from *Staphylococcus aureus* was only shown to produce 4,4'-diaponeurosporene *in vivo* [4215], it is able to catalyse the last reaction *in vitro* [4807].
References: [4621, 3432, 3433, 4215, 4807]

[EC 1.3.8.2 created 2011, modified 2011]

EC 1.3.8.3

- Accepted name:** (*R*)-benzylsuccinyl-CoA dehydrogenase
Reaction: (*R*)-2-benzylsuccinyl-CoA + electron-transfer flavoprotein = (*E*)-2-benzylidenesuccinyl-CoA + reduced electron-transfer flavoprotein
Other name(s): BbsG; (*R*)-benzylsuccinyl-CoA:(acceptor) oxidoreductase
Systematic name: (*R*)-benzylsuccinyl-CoA:electron transfer flavoprotein oxidoreductase
Comments: Requires FAD as prosthetic group. Unlike other acyl-CoA dehydrogenases, this enzyme exhibits high substrate- and enantiomer specificity; it is highly specific for (*R*)-benzylsuccinyl-CoA and is inhibited by (*S*)-benzylsuccinyl-CoA. Forms the third step in the anaerobic toluene metabolic pathway in *Thauera aromatica*. Ferricenium ion is an effective artificial electron acceptor.
References: [2426, 2427]

[EC 1.3.8.3 created 2003 as EC 1.3.99.21, transferred 2012 to EC 1.3.8.3]

EC 1.3.8.4

Accepted name: isovaleryl-CoA dehydrogenase
Reaction: isovaleryl-CoA + electron-transfer flavoprotein = 3-methylcrotonyl-CoA + reduced electron-transfer flavoprotein
Other name(s): isovaleryl-coenzyme A dehydrogenase; isovaleroyl-coenzyme A dehydrogenase; 3-methylbutanoyl-CoA:(acceptor) oxidoreductase
Systematic name: 3-methylbutanoyl-CoA:electron-transfer flavoprotein oxidoreductase
Comments: Contains FAD as prosthetic group. Pentanoate can act as donor.
References: [172, 1796, 4194]

[EC 1.3.8.4 created 1978 as EC 1.3.99.10, modified 1986, transferred 2012 to EC 1.3.8.4]

EC 1.3.8.5

Accepted name: short-chain 2-methylacyl-CoA dehydrogenase
Reaction: 2-methylbutanoyl-CoA + electron-transfer flavoprotein = (*E*)-2-methylbut-2-enoyl-CoA + reduced electron-transfer flavoprotein + H⁺
Other name(s): ACADSB (gene name); 2-methylacyl-CoA dehydrogenase; branched-chain acyl-CoA dehydrogenase (ambiguous); 2-methyl branched chain acyl-CoA dehydrogenase; 2-methylbutanoyl-CoA:(acceptor) oxidoreductase; 2-methyl-branched-chain-acyl-CoA:electron-transfer flavoprotein 2-oxidoreductase; 2-methyl-branched-chain-enoyl-CoA reductase
Systematic name: short-chain 2-methylacyl-CoA:electron-transfer flavoprotein 2-oxidoreductase
Comments: A flavoprotein (FAD). The mammalian enzyme catalyses an oxidative reaction as a step in the mitochondrial β -oxidation of short-chain 2-methyl fatty acids and participates in isoleucine degradation. The enzyme from the parasitic helminth *Ascaris suum* catalyses a reductive reaction as part of a fermentation pathway, shuttling reducing power from the electron-transport chain to 2-methyl branched-chain enoyl CoA.
References: [1794, 2216, 2217, 4456, 102]

[EC 1.3.8.5 created 1992 as EC 1.3.1.52, transferred 2012 to EC 1.3.8.5 (EC 1.3.99.12, created 1986, incorporated 2020), modified 2020]

EC 1.3.8.6

Accepted name: glutaryl-CoA dehydrogenase (ETF)
Reaction: glutaryl-CoA + electron-transfer flavoprotein = crotonyl-CoA + CO₂ + reduced electron-transfer flavoprotein (overall reaction)
(1a) glutaryl-CoA + electron-transfer flavoprotein = (*E*)-glutaconyl-CoA + reduced electron-transfer flavoprotein
(1b) (*E*)-glutaconyl-CoA = crotonyl-CoA + CO₂
Other name(s): glutaryl coenzyme A dehydrogenase; glutaryl-CoA:(acceptor) 2,3-oxidoreductase (decarboxylating); glutaryl-CoA dehydrogenase
Systematic name: glutaryl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase (decarboxylating)
Comments: Contains FAD. The enzyme catalyses the oxidation of glutaryl-CoA to glutaconyl-CoA (which remains bound to the enzyme), and the decarboxylation of the latter to crotonyl-CoA (*cf.* EC 7.2.4.5, glutaconyl-CoA decarboxylase). FAD is the electron acceptor in the oxidation of the substrate, and its reoxidation by electron-transfer flavoprotein completes the catalytic cycle. The anaerobic, sulfate-reducing bacterium *Desulfococcus multivorans* contains two glutaryl-CoA dehydrogenases: a decarboxylating enzyme (this entry), and a non-decarboxylating enzyme that only catalyses the oxidation to glutaconyl-CoA [EC 1.3.99.32, glutaryl-CoA dehydrogenase (acceptor)].
References: [321, 1541, 995, 3450]

[EC 1.3.8.6 created 1972 as EC 1.3.99.7, transferred 2012 to EC 1.3.8.6, modified 2013, modified 2019]

EC 1.3.8.7

- Accepted name:** medium-chain acyl-CoA dehydrogenase
Reaction: a medium-chain acyl-CoA + electron-transfer flavoprotein = a medium-chain *trans*-2,3-dehydroacyl-CoA + reduced electron-transfer flavoprotein
Other name(s): fatty acyl coenzyme A dehydrogenase (ambiguous); acyl coenzyme A dehydrogenase (ambiguous); acyl dehydrogenase (ambiguous); fatty-acyl-CoA dehydrogenase (ambiguous); acyl CoA dehydrogenase (ambiguous); general acyl CoA dehydrogenase (ambiguous); medium-chain acyl-coenzyme A dehydrogenase; acyl-CoA:(acceptor) 2,3-oxidoreductase (ambiguous); ACADM (gene name).
Systematic name: medium-chain acyl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: Contains FAD as prosthetic group. One of several enzymes that catalyse the first step in fatty acids β -oxidation. The enzyme from pig liver can accept substrates with acyl chain lengths of 4 to 16 carbon atoms, but is most active with C₈ to C₁₂ compounds [768]. The enzyme from rat does not accept C₁₆ at all and is most active with C₆-C₈ compounds [1795]. *cf.* EC 1.3.8.1, short-chain acyl-CoA dehydrogenase, EC 1.3.8.8, long-chain acyl-CoA dehydrogenase, and EC 1.3.8.9, very-long-chain acyl-CoA dehydrogenase.
References: [767, 768, 275, 1795, 4277, 2097, 3303, 4311]

[EC 1.3.8.7 created 1961 as EC 1.3.2.2, transferred 1964 to EC 1.3.99.3, part transferred 2012 to EC 1.3.8.7]

EC 1.3.8.8

- Accepted name:** long-chain acyl-CoA dehydrogenase
Reaction: a long-chain acyl-CoA + electron-transfer flavoprotein = a long-chain *trans*-2,3-dehydroacyl-CoA + reduced electron-transfer flavoprotein
Other name(s): palmitoyl-CoA dehydrogenase; palmitoyl-coenzyme A dehydrogenase; long-chain acyl-coenzyme A dehydrogenase; long-chain-acyl-CoA:(acceptor) 2,3-oxidoreductase; ACADL (gene name).
Systematic name: long-chain acyl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: Contains FAD as prosthetic group. One of several enzymes that catalyse the first step in fatty acids β -oxidation. The enzyme from pig liver can accept substrates with acyl chain lengths of 6 to at least 16 carbon atoms. The highest activity was found with C₁₂, and the rates with C₈ and C₁₆ were 80 and 70%, respectively [1563]. The enzyme from rat can accept substrates with C₈-C₂₂. It is most active with C₁₄ and C₁₆, and has no activity with C₄, C₆ or C₂₄ [1795]. *cf.* EC 1.3.8.1, short-chain acyl-CoA dehydrogenase, EC 1.3.8.8, medium-chain acyl-CoA dehydrogenase, and EC 1.3.8.9, very-long-chain acyl-CoA dehydrogenase.
References: [767, 1563, 1482, 1795, 930]

[EC 1.3.8.8 created 1989 as EC 1.3.99.13, part transferred 2012 to EC 1.3.8.8]

EC 1.3.8.9

- Accepted name:** very-long-chain acyl-CoA dehydrogenase
Reaction: a very-long-chain acyl-CoA + electron-transfer flavoprotein = a very-long-chain *trans*-2,3-dehydroacyl-CoA + reduced electron-transfer flavoprotein
Other name(s): ACADVL (gene name).
Systematic name: very-long-chain acyl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: Contains FAD as prosthetic group. One of several enzymes that catalyse the first step in fatty acids β -oxidation. The enzyme is most active toward long-chain acyl-CoAs such as C₁₄, C₁₆ and C₁₈, but is also active with very-long-chain acyl-CoAs up to 24 carbons. It shows no activity for substrates of less than 12 carbons. Its specific activity towards palmitoyl-CoA is more than 10-fold that of the long-chain acyl-CoA dehydrogenase [1865]. *cf.* EC 1.3.8.1, short-chain acyl-CoA dehydrogenase, EC 1.3.8.7, medium-chain acyl-CoA dehydrogenase, and EC 1.3.8.8, long-chain acyl-CoA dehydrogenase.
References: [1865, 113, 2738]

[EC 1.3.8.9 created 1961 as EC 1.3.2.2, transferred 1964 to EC 1.3.99.3, part transferred 2012 to EC 1.3.8.9]

EC 1.3.8.10

- Accepted name:** cyclohex-1-ene-1-carbonyl-CoA dehydrogenase
Reaction: cyclohex-1-ene-1-carbonyl-CoA + electron-transfer flavoprotein = cyclohex-1,5-diene-1-carbonyl-CoA + reduced electron-transfer flavoprotein
Systematic name: cyclohex-1-ene-1-carbonyl-CoA:electron transfer flavoprotein oxidoreductase
Comments: Contains FAD. The enzyme, characterized from the strict anaerobic bacterium *Syntrophus aciditrophicus*, is involved in production of cyclohexane-1-carboxylate, a byproduct produced by that organism during fermentation of benzoate and crotonate to acetate.
References: [2291]

[EC 1.3.8.10 created 2013]

EC 1.3.8.11

- Accepted name:** cyclohexane-1-carbonyl-CoA dehydrogenase (electron-transfer flavoprotein)
Reaction: cyclohexane-1-carbonyl-CoA + electron-transfer flavoprotein = cyclohex-1-ene-1-carbonyl-CoA + reduced electron-transfer flavoprotein
Other name(s): *aliB* (gene name); cyclohexane-1-carbonyl-CoA dehydrogenase (ambiguous)
Systematic name: cyclohexane-1-carbonyl-CoA:electron transfer flavoprotein oxidoreductase
Comments: Contains FAD. The enzyme, characterized from the strict anaerobic bacterium *Syntrophus aciditrophicus*, is involved in production of cyclohexane-1-carboxylate, a byproduct produced by that organism during fermentation of benzoate and crotonate to acetate.
References: [3283, 2291]

[EC 1.3.8.11 created 2013, modified 2020]

EC 1.3.8.12

- Accepted name:** (2*S*)-methylsuccinyl-CoA dehydrogenase
Reaction: (2*S*)-methylsuccinyl-CoA + electron-transfer flavoprotein = 2-methylfumaryl-CoA + reduced electron-transfer flavoprotein
Other name(s): Mcd
Systematic name: (2*S*)-methylsuccinyl-CoA:electron-transfer flavoprotein oxidoreductase
Comments: The enzyme, characterized from the bacterium *Rhodobacter sphaeroides*, is involved in the ethylmalonyl-CoA pathway for acetyl-CoA assimilation. The enzyme contains FAD.
References: [1061]

[EC 1.3.8.12 created 2015]

EC 1.3.8.13

- Accepted name:** crotonobetainyl-CoA reductase
Reaction: γ -butyrobetainyl-CoA + electron-transfer flavoprotein = crotonobetainyl-CoA + reduced electron-transfer flavoprotein
Other name(s): *caiA* (gene name)
Systematic name: γ -butyrobetainyl-CoA:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: The enzyme has been purified from the bacterium *Escherichia coli* O44 K74, in which it forms a complex with EC 2.8.3.21, L-carnitine CoA-transferase. The electron donor is believed to be an electron-transfer flavoprotein (ETF) encoded by the *fixA* and *fixB* genes.
References: [3578, 3376, 1041, 4508]

[EC 1.3.8.13 created 2017]

EC 1.3.8.14

- Accepted name:** L-prolyl-[peptidyl-carrier protein] dehydrogenase

Reaction: L-prolyl-[peptidyl-carrier protein] + 2 electron-transfer flavoprotein = 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein] + 2 reduced electron-transfer flavoprotein
Other name(s): *pigA* (gene name); *bmp3* (gene name); *pltE* (gene name); *redW* (gene name); (L-prolyl)-[peptidyl-carrier protein]:electron-transfer flavoprotein oxidoreductase
Systematic name: L-prolyl-[peptidyl-carrier protein]:electron-transfer flavoprotein oxidoreductase
Comments: Contains FAD. The enzyme participates in the biosynthesis of several pyrrole-containing compounds, such as undecylprodigiosin, prodigiosin, pyoluteorin, and coumermycin A1. It is believed to catalyse the formation of a Δ^2 -pyrrolin-2-carbonyl-[peptidyl-carrier protein] intermediate, followed by a two-electron oxidation to 1*H*-pyrrol-2-carbonyl-[peptidyl-carrier protein].
References: [4264, 1533]

[EC 1.3.8.14 created 2017]

EC 1.3.8.15

Accepted name: 3-(aryl)acrylate reductase
Reaction: (1) phloretate + electron-transfer flavoprotein = 4-coumarate + reduced electron-transfer flavoprotein
(2) 3-phenylpropanoate + electron-transfer flavoprotein = *trans*-cinnamate + reduced electron-transfer flavoprotein
(3) 3-(1*H*-indol-3-yl)propanoate + electron-transfer flavoprotein = 3-(indol-3-yl)acrylate + reduced electron-transfer flavoprotein
Other name(s): *acdA* (gene name)
Systematic name: 3-(phenyl)propanoate:electron-transfer flavoprotein 2,3-oxidoreductase
Comments: The enzyme, found in some amino acid-fermenting anaerobic bacteria, participates in the fermentation pathways of L-phenylalanine, L-tyrosine, and L-tryptophan. Unlike EC 1.3.1.31, 2-enoate reductase, this enzyme has minimal activity with crotonate.
References: [936]

[EC 1.3.8.15 created 2019]

EC 1.3.8.16

Accepted name: 2-amino-4-deoxychorismate dehydrogenase
Reaction: (2*S*)-2-amino-4-deoxychorismate + FMN = 3-(1-carboxyvinyl)anthranilate + FMNH₂
Other name(s): ADIC dehydrogenase; 2-amino-2-deoxyisochorismate dehydrogenase; SgcG
Systematic name: (2*S*)-2-amino-4-deoxychorismate:FMN oxidoreductase
Comments: The sequential action of EC 2.6.1.86, 2-amino-4-deoxychorismate synthase and this enzyme leads to the formation of the benzoxazolinone moiety of the enediyne antitumour antibiotic C-1027 [2340, 4837].
References: [2340, 4837]

[EC 1.3.8.16 created 2008 as 1.3.99.24, transferred 2020 to EC 1.3.8.16.]

EC 1.3.8.17

Accepted name: dehydro coenzyme F₄₂₀ reductase
Reaction: oxidized coenzyme F₄₂₀-0 + FMN = dehydro coenzyme F₄₂₀-0 + FMNH₂
Other name(s): *fbiB* (gene name)
Systematic name: oxidized coenzyme F₄₂₀-0:FMN oxidoreductase
Comments: This enzyme is involved in the biosynthesis of coenzyme F₄₂₀, a redox-active cofactor found in all methanogenic archaea, as well as some eubacteria. In some eubacteria the enzyme is multifunctional, also catalysing the activities of EC 6.3.2.31, coenzyme F₄₂₀-0:L-glutamate ligase, and EC 6.3.2.34, coenzyme F₄₂₀-1:γ-L-glutamate ligase.
References: [230]

[EC 1.3.8.17 created 2021]

EC 1.3.98 With other, known, physiological acceptors

EC 1.3.98.1

- Accepted name:** dihydroorotate dehydrogenase (fumarate)
Reaction: (S)-dihydroorotate + fumarate = orotate + succinate
Other name(s): DHOdehase (ambiguous); dihydroorotate dehydrogenase (ambiguous); dihydroorotic acid dehydrogenase (ambiguous); DHOD (ambiguous); DHODase (ambiguous); dihydroorotate oxidase; *pyr4* (gene name)
Systematic name: (S)-dihydroorotate:fumarate oxidoreductase
Comments: Binds FMN. The reaction, which takes place in the cytosol, is the only redox reaction in the *de novo* biosynthesis of pyrimidine nucleotides. Molecular oxygen can replace fumarate *in vitro*. Other class 1 dihydroorotate dehydrogenases use either NAD⁺ (EC 1.3.1.14) or NADP⁺ (EC 1.3.1.15) as electron acceptor. The membrane bound class 2 dihydroorotate dehydrogenase (EC 1.3.5.2) uses quinone as electron acceptor.
References: [342, 3582, 3103, 4858, 1809, 626]

[EC 1.3.98.1 created 1961 as EC 1.3.3.1, transferred 2011 to EC 1.3.98.1]

[1.3.98.2 Transferred entry. fumarate reductase (CoM/CoB). Now EC 1.3.4.1, fumarate reductase (CoM/CoB)]

[EC 1.3.98.2 created 2014, deleted 2014]

EC 1.3.98.3

- Accepted name:** coproporphyrinogen dehydrogenase
Reaction: coproporphyrinogen III + 2 *S*-adenosyl-L-methionine = protoporphyrinogen IX + 2 CO₂ + 2 L-methionine + 2 5'-deoxyadenosine
Other name(s): oxygen-independent coproporphyrinogen-III oxidase; HemN; coproporphyrinogen III oxidase
Systematic name: coproporphyrinogen-III:*S*-adenosyl-L-methionine oxidoreductase (decarboxylating)
Comments: This enzyme differs from EC 1.3.3.3, coproporphyrinogen oxidase, by using *S*-adenosyl-L-methionine (AdoMet) instead of oxygen as oxidant. It occurs mainly in bacteria, whereas eukaryotes use the oxygen-dependent oxidase. The reaction starts by using an electron from the reduced form of the enzyme's [4Fe-4S] cluster to split AdoMet into methionine and the radical 5'-deoxyadenosin-5'-yl. This radical initiates attack on the 2-carboxyethyl groups, leading to their conversion into vinyl groups. This conversion, $\text{---CH-CH}_2\text{-COO}^- \rightarrow \text{---CH=CH}_2 + \text{CO}_2 + \text{e}^-$ replaces the electron initially used.
References: [2372, 2371]

[EC 1.3.98.3 created 2004 as EC 1.3.99.22, transferred 2016 to EC 1.3.98.3]

EC 1.3.98.4

- Accepted name:** 5a,11a-dehydrotetracycline reductase
Reaction: tetracycline + oxidized coenzyme F₄₂₀ = 5a,11a-dehydrotetracycline + reduced coenzyme F₄₂₀
Other name(s): *oxyR* (gene name); 12-dehydrotetracycline dehydrogenase; dehydroxytetracycline dehydrogenase; 12-dehydrotetracycline reductase
Systematic name: tetracycline:coenzyme F₄₂₀ dehydrogenase
Comments: The enzyme, characterized from the bacteria *Streptomyces aureofaciens* and *Streptomyces rimosus*, catalyses the last step in the biosynthesis of the tetracycline antibiotics tetracycline and oxytetracycline.
References: [2743, 2811, 2744, 4522]

[EC 1.3.98.4 created 2016]

EC 1.3.98.5

- Accepted name:** hydrogen peroxide-dependent heme synthase

Reaction: Fe-coproporphyrin III + 2 H₂O₂ = protoheme + 2 CO₂ + 4 H₂O (overall reaction)
(1a) Fe-coproporphyrin III + H₂O₂ = harderoheme III + CO₂ + 2 H₂O
(1b) harderoheme III + H₂O₂ = protoheme + CO₂ + 2 H₂O

Other name(s): coproheme III oxidative decarboxylase; *hemQ* (gene name)

Systematic name: Fe-coproporphyrin III:hydrogen peroxide oxidoreductase (decarboxylating)

Comments: The enzyme participates in a heme biosynthesis pathway found in Gram-positive bacteria. The initial decarboxylation step is fast and yields the three-propanoate harderoheme isomer III. The second decarboxylation is much slower. *cf.* EC 1.3.98.6, SAM-dependent heme synthase.

References: [807, 585, 1684, 584]

[EC 1.3.98.5 created 2019]

EC 1.3.98.6

Accepted name: AdoMet-dependent heme synthase

Reaction: Fe-coproporphyrin III + 2 *S*-adenosyl-L-methionine = protoheme + 2 CO₂ + 2 5'-deoxyadenosine + 2 L-methionine

Other name(s): *ahbD* (gene name); SAM-dependent heme synthase

Systematic name: Fe-coproporphyrin III:*S*-adenosyl-L-methionine oxidoreductase (decarboxylating)

Comments: This radical AdoMet enzyme participates in a heme biosynthesis pathway found in archaea and sulfur-reducing bacteria. *cf.* EC 1.3.98.5, hydrogen peroxide-dependent heme synthase.

References: [198, 2278]

[EC 1.3.98.6 created 2019]

EC 1.3.98.7

Accepted name: [mycofactocin precursor peptide]-tyrosine decarboxylase

Reaction: C-terminal [mycofactocin precursor peptide]-glycyl-L-valyl-L-tyrosine + *S*-adenosyl-L-methionine = C-terminal [mycofactocin precursor peptide]-glycyl-L-valyl-4-[2-aminoethenyl]phenol + CO₂ + 5'-deoxyadenosine + L-methionine

Other name(s): *mftC* (gene name)

Systematic name: C-terminal [mycofactocin precursor peptide]-glycyl-L-valyl-L-tyrosine L-tyrosine-carboxylase

Comments: This is a bifunctional radical AdoMet (radical SAM) enzyme that catalyses the first two steps in the biosynthesis of the enzyme cofactor mycofactocin. Activity requires the presence of the MftB chaperone. The other activity of the enzyme is EC 4.1.99.26, 3-amino-5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidin-2-one synthase.

References: [1470, 466, 2077, 168]

[EC 1.3.98.7 created 2021]

EC 1.3.99 With unknown physiological acceptors

[1.3.99.1 Deleted entry. succinate dehydrogenase. The activity is included in EC 1.3.5.1, succinate dehydrogenase (quinone).]

[EC 1.3.99.1 created 1961, deleted 2014]

[1.3.99.2 Transferred entry. butyryl-CoA dehydrogenase. Now EC 1.3.8.1, butyryl-CoA dehydrogenase.]

[EC 1.3.99.2 created 1961 as EC 1.3.2.1, transferred 1964 to EC 1.3.99.2, deleted 2011]

[1.3.99.3 Transferred entry. acyl-CoA dehydrogenase, now EC 1.3.8.7, medium-chain acyl-CoA dehydrogenase, EC 1.3.8.8, long-chain acyl-CoA dehydrogenase and EC 1.3.8.9, very-long-chain acyl-CoA dehydrogenase]

[EC 1.3.99.3 created 1961 as EC 1.3.2.2, transferred 1964 to EC 1.3.99.3, deleted 2012]

EC 1.3.99.4

Accepted name: 3-oxosteroid 1-dehydrogenase
Reaction: a 3-oxosteroid + acceptor = a 3-oxo- Δ^1 -steroid + reduced acceptor
Other name(s): 3-oxosteroid Δ^1 -dehydrogenase; Δ^1 -dehydrogenase; 3-ketosteroid-1-en-dehydrogenase; 3-ketosteroid- Δ^1 -dehydrogenase; 1-ene-dehydrogenase; 3-oxosteroid:(2,6-dichlorphenolindophenol) Δ^1 -oxidoreductase; 4-en-3-oxosteroid:(acceptor)-1-en-oxido-reductase; Δ^1 -steroid reductase; 3-oxosteroid:(acceptor) Δ^1 -oxidoreductase
Systematic name: 3-oxosteroid:acceptor Δ^1 -oxidoreductase
References: [2435]

[EC 1.3.99.4 created 1965]

EC 1.3.99.5

Accepted name: 3-oxo-5 α -steroid 4-dehydrogenase (acceptor)
Reaction: a 3-oxo-5 α -steroid + acceptor = a 3-oxo- Δ^4 -steroid + reduced acceptor
Other name(s): steroid 5 α -reductase; 3-oxosteroid Δ^4 -dehydrogenase; 3-oxo-5 α -steroid Δ^4 -dehydrogenase; steroid Δ^4 -5 α -reductase; Δ^4 -3-keto steroid 5 α -reductase; Δ^4 -3-oxo steroid reductase; Δ^4 -3-ketosteroid5 α -oxidoreductase; Δ^4 -3-oxosteroid-5 α -reductase; 3-keto- Δ^4 -steroid-5 α -reductase; 5 α -reductase; testosterone 5 α -reductase; 4-ene-3-ketosteroid-5 α -oxidoreductase; Δ^4 -5 α -dehydrogenase; 3-oxo-5 α -steroid:(acceptor) Δ^4 -oxidoreductase; *tesI* (gene name)
Systematic name: 3-oxo-5 α -steroid:acceptor Δ^4 -oxidoreductase
Comments: A flavoprotein. This bacterial enzyme, characterized from *Comamonas testosteroni*, is involved in androsterone degradation. cf. EC 1.3.1.22, 3-oxo-5 α -steroid 4-dehydrogenase (NADP⁺).
References: [2435, 1132, 1723]

[EC 1.3.99.5 created 1965, modified 2012]

EC 1.3.99.6

Accepted name: 3-oxo-5 β -steroid 4-dehydrogenase
Reaction: a 3-oxo-5 β -steroid + acceptor = a 3-oxo- Δ^4 -steroid + reduced acceptor
Other name(s): 3-oxo-5 β -steroid:(acceptor) Δ^4 -oxidoreductase
Systematic name: 3-oxo-5 β -steroid:acceptor Δ^4 -oxidoreductase
References: [838]

[EC 1.3.99.6 created 1972]

[1.3.99.7 Transferred entry. glutaryl-CoA dehydrogenase. Now EC 1.3.8.6, glutaryl-CoA dehydrogenase]

[EC 1.3.99.7 created 1972, deleted 2012]

EC 1.3.99.8

Accepted name: 2-furoyl-CoA dehydrogenase
Reaction: 2-furoyl-CoA + H₂O + acceptor = S-(5-hydroxy-2-furoyl)-CoA + reduced acceptor
Other name(s): furoyl-CoA hydroxylase; 2-furoyl coenzyme A hydroxylase; 2-furoyl coenzyme A dehydrogenase; 2-furoyl-CoA:(acceptor) 5-oxidoreductase (hydroxylating)
Systematic name: 2-furoyl-CoA:acceptor 5-oxidoreductase (hydroxylating)
Comments: A copper protein. The oxygen atom of the -OH produced is derived from water, not O₂; the actual oxidative step is probably dehydrogenation of a hydrated form -CHOH-CH₂- to -C(OH)=CH-, which tautomerizes non-enzymically to -CO-CH₂-, giving (5-oxo-4,5-dihydro-2-furoyl)-CoA. Methylene blue, nitro blue, tetrazolium and a membrane fraction from *Pseudomonas putida* can act as acceptors.
References: [2138]

[EC 1.3.99.8 created 1976]

[1.3.99.9 Transferred entry. β -cyclopiazonate dehydrogenase. Now EC 1.21.99.1, β -cyclopiazonate dehydrogenase]

[EC 1.3.99.9 created 1976, deleted 2002]

[1.3.99.10 Transferred entry. *isovaleryl-CoA dehydrogenase*. Now EC 1.3.8.4, *isovaleryl-CoA dehydrogenase*]

[EC 1.3.99.10 created 1978, modified 1986, deleted 2012]

[1.3.99.11 Transferred entry. *dihydroorotate dehydrogenase*. As the acceptor is now known, the enzyme has been transferred to EC 1.3.5.2, *dihydroorotate dehydrogenase*]

[EC 1.3.99.11 created 1983, deleted 2009]

[1.3.99.12 Transferred entry. *2-methylacyl-CoA dehydrogenase*. Now classified as EC 1.3.8.5, *2-methyl-branched-chain-enoyl-CoA reductase*.]

[EC 1.3.99.12 created 1986, deleted 2020]

[1.3.99.13 Transferred entry. *long-chain-acyl-CoA dehydrogenase*. Now EC 1.3.8.8, *long-chain-acyl-CoA dehydrogenase*]

[EC 1.3.99.13 created 1989, deleted 2012]

EC 1.3.99.14

Accepted name: cyclohexanone dehydrogenase
Reaction: cyclohexanone + acceptor = cyclohex-2-enone + reduced acceptor
Other name(s): cyclohexanone:(acceptor) 2-oxidoreductase
Systematic name: cyclohexanone:acceptor 2-oxidoreductase
Comments: 2,6-Dichloroindophenol can act as acceptor. The corresponding ketones of cyclopentane and cycloheptane cannot act as donors.
References: [823]

[EC 1.3.99.14 created 1992]

[1.3.99.15 Transferred entry. *benzoyl-CoA reductase*. Now EC 1.3.7.8.]

[EC 1.3.99.15 created 1999, deleted 2011]

EC 1.3.99.16

Accepted name: isoquinoline 1-oxidoreductase
Reaction: isoquinoline + acceptor + H₂O = isoquinolin-1(2*H*)-one + reduced acceptor
Systematic name: isoquinoline:acceptor 1-oxidoreductase (hydroxylating)
Comments: The enzyme from *Pseudomonas diminuta* is specific towards *N*-containing *N*-heterocyclic substrates, including isoquinoline, isoquinolin-5-ol, phthalazine and quinazoline. Electron acceptors include 1,2-benzoquinone, cytochrome *c*, ferricyanide, iodonitrotetrazolium chloride, nitroblue tetrazolium, Meldola blue and phenazine methosulfate.
References: [2405, 2404]

[EC 1.3.99.16 created 1999]

EC 1.3.99.17

Accepted name: quinoline 2-oxidoreductase
Reaction: quinoline + acceptor + H₂O = quinolin-2(1*H*)-one + reduced acceptor
Systematic name: quinoline:acceptor 2-oxidoreductase (hydroxylating)
Comments: Quinolin-2-ol, quinolin-7-ol, quinolin-8-ol, 3-, 4- and 8-methylquinolines and 8-chloroquinoline are substrates. Iodonitrotetrazolium chloride can act as an electron acceptor.
References: [240, 4337, 3296, 3696]

[EC 1.3.99.17 created 1999]

EC 1.3.99.18

- Accepted name:** quinaldate 4-oxidoreductase
Reaction: quinaldate + acceptor + H₂O = kynurenate + reduced acceptor
Other name(s): quinaldic acid 4-oxidoreductase
Systematic name: quinoline-2-carboxylate:acceptor 4-oxidoreductase (hydroxylating)
Comments: The enzyme from *Pseudomonas* sp. AK2 also acts on quinoline-8-carboxylate, whereas that from *Serratia marcescens* 2CC-1 will oxidize nicotinate; quinaldate is a substrate for both of these enzymes. 2,4,6-Trinitrobenzene sulfonate, 1,4-benzoquinone, 1,2-naphthoquinone, nitroblue tetrazolium, thionine and menadione will serve as an electron acceptor for the former enzyme and ferricyanide for the latter; Meldola blue, iodonitrotetrazolium chloride, phenazine methosulfate, 2,6-dichlorophenolindophenol and cytochrome *c* will act as electron acceptors for both.
References: [3678, 1111]

[EC 1.3.99.18 created 1999]

EC 1.3.99.19

- Accepted name:** quinoline-4-carboxylate 2-oxidoreductase
Reaction: quinoline-4-carboxylate + acceptor + H₂O = 2-oxo-1,2-dihydroquinoline-4-carboxylate + reduced acceptor
Other name(s): quinaldic acid 4-oxidoreductase; quinoline-4-carboxylate:acceptor 2-oxidoreductase (hydroxylating)
Systematic name: quinoline-4-carboxylate:acceptor 2-oxidoreductase (hydroxylating)
Comments: A molybdenum—iron—sulfur flavoprotein with molybdopterin cytosine dinucleotide as the molybdenum cofactor. Quinoline, 4-methylquinoline and 4-chloroquinoline can also serve as substrates for the enzyme from *Agrobacterium* sp. 1B. Iodonitrotetrazolium chloride, thionine, menadione and 2,6-dichlorophenolindophenol can act as electron acceptors.
References: [241]

[EC 1.3.99.19 created 1999, modified 2006]

[1.3.99.20 Transferred entry. EC 1.3.99.20, 4-hydroxybenzoyl-CoA reductase. Now EC 1.3.7.9, 4-hydroxybenzoyl-CoA reductase.]

[EC 1.3.99.20 created 2000, deleted 2011]

[1.3.99.21 Transferred entry. (R)-benzylsuccinyl-CoA dehydrogenase. Now EC 1.3.8.3, (R)-benzylsuccinyl-CoA dehydrogenase]

[EC 1.3.99.21 created 2003 as EC 1.3.99.21, deleted 2012]

[1.3.99.22 Transferred entry. coproporphyrinogen dehydrogenase. Now EC 1.3.98.3, coproporphyrinogen dehydrogenase]

[EC 1.3.99.22 created 2004, deleted 2016]

EC 1.3.99.23

- Accepted name:** all-trans-retinol 13,14-reductase
Reaction: all-trans-13,14-dihydroretinol + acceptor = all-trans-retinol + reduced acceptor
Other name(s): retinol saturase; RetSat; (13,14)-all-trans-retinol saturase; all-trans-retinol:all-trans-13,14-dihydroretinol saturase
Systematic name: all-trans-13,14-dihydroretinol:acceptor 13,14-oxidoreductase
Comments: The reaction is only known to occur in the opposite direction to that given above, with the enzyme being specific for all-trans-retinol as substrate. Neither all-trans-retinoic acid nor 9-cis, 11-cis or 13-cis-retinol isomers are substrates. May play a role in the metabolism of vitamin A.
References: [2862]

[EC 1.3.99.23 created 2005]

[1.3.99.24 Transferred entry. 2-amino-4-deoxychorismate dehydrogenase. Now EC 1.3.8.16, 2-amino-4-deoxychorismate dehydrogenase]

[EC 1.3.99.24 created 2008, deleted 2020]

EC 1.3.99.25

Accepted name: carvone reductase
Reaction: (1) (+)-dihydrocarvone + acceptor = (-)-carvone + reduced acceptor
(2) (-)-isodihydrocarvone + acceptor = (+)-carvone + reduced acceptor
Systematic name: (+)-dihydrocarvone:acceptor 1,6-oxidoreductase
Comments: This enzyme participates in the carveol and dihydrocarveol degradation pathway of the Gram-positive bacterium *Rhodococcus erythropolis* DCL14. The enzyme has not been purified, and requires an unknown cofactor, which is different from NAD⁺, NADP⁺ or a flavin.
References: [4403]

[EC 1.3.99.25 created 2008]

EC 1.3.99.26

Accepted name: *all-trans*-ζ-carotene desaturase
Reaction: *all-trans*-ζ-carotene + 2 acceptor = *all-trans*-lycopene + 2 reduced acceptor (overall reaction)
(1a) *all-trans*-ζ-carotene + acceptor = *all-trans*-neurosporene + reduced acceptor
(1b) *all-trans*-neurosporene + acceptor = *all-trans*-lycopene + reduced acceptor
Other name(s): CrtIb; phytoene desaturase (ambiguous); 2-step phytoene desaturase (ambiguous); two-step phytoene desaturase (ambiguous); CrtI (ambiguous)
Systematic name: *all-trans*-ζ-carotene:acceptor oxidoreductase
Comments: This enzyme is involved in carotenoid biosynthesis.
References: [1811]

[EC 1.3.99.26 created 2011]

EC 1.3.99.27

Accepted name: 1-hydroxycarotenoid 3,4-desaturase
Reaction: 1-hydroxy-1,2-dihydrolycopene + acceptor = 1-hydroxy-3,4-didehydro-1,2-dihydrolycopene + reduced acceptor
Other name(s): CrtD; hydroxyneurosporene desaturase; carotenoid 3,4-dehydrogenase; 1-hydroxy-carotenoid 3,4-dehydrogenase
Systematic name: 1-hydroxy-1,2-dihydrolycopene:acceptor oxidoreductase
Comments: The enzymes from *Rubrivivax gelatinosus* and *Rhodobacter sphaeroides* prefer the acyclic carotenoids (e.g. 1-hydroxy-1,2-dihydroneurosporene, 1-hydroxy-1,2-dihydrolycopene) as substrates. The conversion rate for the 3,4-desaturation of the monocyclic 1'-hydroxy-1',2'-dihydro-γ-carotene is lower [4015, 62]. The enzyme from the marine bacterium strain P99-3 shows high activity with the monocyclic carotenoid 1'-hydroxy-1',2'-dihydro-γ-carotene [4244]. The enzyme from *Rhodobacter sphaeroides* utilizes molecular oxygen as the electron acceptor *in vitro* [62]. However, oxygen is unlikely to be the natural electron acceptor under anaerobic conditions.
References: [4244, 4015, 62]

[EC 1.3.99.27 created 2011]

EC 1.3.99.28

Accepted name: phytoene desaturase (neurosporene-forming)
Reaction: 15-*cis*-phytoene + 3 acceptor = *all-trans*-neurosporene + 3 reduced acceptor (overall reaction)
(1a) 15-*cis*-phytoene + acceptor = *all-trans*-phytofluene + reduced acceptor
(1b) *all-trans*-phytofluene + acceptor = *all-trans*-ζ-carotene + reduced acceptor
(1c) *all-trans*-ζ-carotene + acceptor = *all-trans*-neurosporene + reduced acceptor
Other name(s): 3-step phytoene desaturase; three-step phytoene desaturase; phytoene desaturase (ambiguous); CrtI (ambiguous)

Systematic name: 15-*cis*-phytoene:acceptor oxidoreductase (neurosporene-forming)
Comments: This enzyme is involved in carotenoid biosynthesis and catalyses up to three desaturation steps (*cf.* EC 1.3.99.29 [phytoene desaturase (ζ -carotene-forming)], EC 1.3.99.30 [phytoene desaturase (3,4-didehydrolycopene-forming)], EC 1.3.99.31 [phytoene desaturase (lycopene-forming)]). The enzyme is activated by FAD. NAD⁺, NADP⁺ or ATP show no activating effect [3431].
References: [3431, 4513]

[EC 1.3.99.28 created 2011]

EC 1.3.99.29

Accepted name: phytoene desaturase (ζ -carotene-forming)
Reaction: 15-*cis*-phytoene + 2 acceptor = *all-trans*- ζ -carotene + 2 reduced acceptor (overall reaction)
(1a) 15-*cis*-phytoene + acceptor = *all-trans*-phytofluene + reduced acceptor
(1b) *all-trans*-phytofluene + acceptor = *all-trans*- ζ -carotene + reduced acceptor
Other name(s): CrtIa; 2-step phytoene desaturase (ambiguous); two-step phytoene desaturase (ambiguous)
Systematic name: 15-*cis*-phytoene:acceptor oxidoreductase (ζ -carotene-forming)
Comments: The enzyme is involved in carotenoid biosynthesis and catalyses up to two desaturation steps (*cf.* EC 1.3.99.28 [phytoene desaturase (neurosporene-forming)], EC 1.3.99.30 [phytoene desaturase (3,4-didehydrolycopene-forming)] and EC 1.3.99.31 [phytoene desaturase (lycopene-forming)]).
References: [1811]

[EC 1.3.99.29 created 2011]

EC 1.3.99.30

Accepted name: phytoene desaturase (3,4-didehydrolycopene-forming)
Reaction: 15-*cis*-phytoene + 5 acceptor = *all-trans*-3,4-didehydrolycopene + 5 reduced acceptor (overall reaction)
(1a) 15-*cis*-phytoene + acceptor = *all-trans*-phytofluene + reduced acceptor
(1b) *all-trans*-phytofluene + acceptor = *all-trans*- ζ -carotene + reduced acceptor
(1c) *all-trans*- ζ -carotene + acceptor = *all-trans*-neurosporene + reduced acceptor
(1d) *all-trans*-neurosporene + acceptor = *all-trans*-lycopene + reduced acceptor
(1e) *all-trans*-lycopene + acceptor = *all-trans*-3,4-didehydrolycopene + reduced acceptor
Other name(s): 5-step phytoene desaturase; five-step phytoene desaturase; phytoene desaturase (ambiguous); Al-1
Systematic name: 15-*cis*-phytoene:acceptor oxidoreductase (3,4-didehydrolycopene-forming)
Comments: This enzyme is involved in carotenoid biosynthesis and catalyses up to five desaturation steps (*cf.* EC 1.3.99.28 [phytoene desaturase (neurosporene-forming)], EC 1.3.99.29 [phytoene desaturase (ζ -carotene-forming)] and EC 1.3.99.31 [phytoene desaturase (lycopene-forming)]).
References: [1566, 1066]

[EC 1.3.99.30 created 2011]

EC 1.3.99.31

Accepted name: phytoene desaturase (lycopene-forming)
Reaction: 15-*cis*-phytoene + 4 acceptor = *all-trans*-lycopene + 4 reduced acceptor (overall reaction)
(1a) 15-*cis*-phytoene + acceptor = *all-trans*-phytofluene + reduced acceptor
(1b) *all-trans*-phytofluene + acceptor = *all-trans*- ζ -carotene + reduced acceptor
(1c) *all-trans*- ζ -carotene + acceptor = *all-trans*-neurosporene + reduced acceptor
(1d) *all-trans*-neurosporene + acceptor = *all-trans*-lycopene + reduced acceptor
Other name(s): 4-step phytoene desaturase; four-step phytoene desaturase; phytoene desaturase (ambiguous); CrtI (ambiguous)
Systematic name: 15-*cis*-phytoene:acceptor oxidoreductase (lycopene-forming)

Comments: Requires FAD. The enzyme is involved in carotenoid biosynthesis and catalyses up to four desaturation steps (*cf.* EC 1.3.99.28 [phytoene desaturase (neurosporene-forming)], EC 1.3.99.29 [phytoene desaturase (ζ -carotene-forming)] and EC 1.3.99.30 [phytoene desaturase (3,4-didehydrolycopene-forming)]).

References: [1170]

[EC 1.3.99.31 created 2011]

EC 1.3.99.32

Accepted name: glutaryl-CoA dehydrogenase (acceptor)

Reaction: glutaryl-CoA + acceptor = (*E*)-glutaconyl-CoA + reduced acceptor

Other name(s): GDHDes; nondecarboxylating glutaryl-coenzyme A dehydrogenase; nondecarboxylating glutaconyl-coenzyme A-forming GDH; glutaryl-CoA dehydrogenase (non-decarboxylating)

Systematic name: glutaryl-CoA:acceptor 2,3-oxidoreductase (non-decarboxylating)

Comments: The enzyme contains FAD. The anaerobic, sulfate-reducing bacterium *Desulfococcus multivorans* contains two glutaryl-CoA dehydrogenases: a decarboxylating enzyme (EC 1.3.8.6), and a nondecarboxylating enzyme (this entry). The two enzymes cause different structural changes around the glutaconyl carboxylate group, primarily due to the presence of either a tyrosine or a valine residue, respectively, at the active site.

References: [4643, 4642]

[EC 1.3.99.32 created 2012, modified 2013]

EC 1.3.99.33

Accepted name: urocanate reductase

Reaction: dihydrourocanate + acceptor = urocanate + reduced acceptor

Other name(s): *urdA* (gene name)

Systematic name: dihydrourocanate:acceptor oxidoreductase

Comments: The enzyme from the bacterium *Shewanella oneidensis* MR-1 contains a noncovalently-bound FAD and a covalently-bound FMN. It functions as part of an anaerobic electron transfer chain that utilizes urocanate as the terminal electron acceptor. The activity has been demonstrated with the artificial donor reduced methyl viologen.

References: [369]

[EC 1.3.99.33 created 2013]

[1.3.99.34 *Transferred entry. 2,3-bis-O-geranylgeranyl-sn-glycerol 1-phosphate reductase (donor). Now classified as EC 1.3.7.11, 2,3-bis-O-geranylgeranyl-sn-glycero-phospholipid reductase.*]

[EC 1.3.99.34 created 2013, deleted 2015]

[1.3.99.35 *Transferred entry. chlorophyllide a reductase. Now EC 1.3.7.15, chlorophyllide a reductase*]

[EC 1.3.99.35 created 2014, deleted 2016]

EC 1.3.99.36

Accepted name: cypemycin cysteine dehydrogenase (decarboxylating)

Reaction: cypemycin(1-18)-L-Cys-L-Leu-L-Val-L-Cys + acceptor = C^{3,19},S²¹-cyclocypemycin(1-18)-L-Ala-L-Leu-N-thioethenyl-L-valinamide + CO₂ + H₂S + reduced acceptor

Other name(s): cypemycin decarboxylase; CypD

Systematic name: cypemycin(1-18)-L-Cys-L-Leu-L-Val-L-Cys:acceptor oxidoreductase (decarboxylating, cyclizing)

Comments: Cypemycin, isolated from the bacterium *Streptomyces* sp. OH-4156, is a peptide antibiotic, member of the linaridins, a class of posttranslationally modified ribosomally synthesized peptides. The enzyme decarboxylates and reduces the C-terminal L-cysteine residue, producing a reactive ethenethiol group that reacts with a dethiolated cysteine upstream to form an aminovinyl-methyl-cysteine loop that is important for the antibiotic activity of the mature peptide.

References: [694]

[EC 1.3.99.36 created 2014]

EC 1.3.99.37

Accepted name: 1-hydroxy-2-isopentenylcarotenoid 3,4-desaturase

Reaction: (1) dihydroisopentenyldehydrorhodopin + acceptor = isopentenyldehydrorhodopin + reduced acceptor

(2) dihydrobisanhydrobacterioruberin + acceptor = bisanhydrobacterioruberin + reduced acceptor

Other name(s): *crtD* (gene name)

Systematic name: dihydroisopentenyldehydrorhodopin:acceptor 3,4-oxidoreductase

Comments: The enzyme, isolated from the archaeon *Haloarcula japonica*, is involved in the biosynthesis of the C₅₀ carotenoid bacterioruberin. In this pathway it catalyses the desaturation of the C-3,4 double bond in dihydroisopentenyldehydrorhodopin and the desaturation of the C-3',4' double bond in dihydrobisanhydrobacterioruberin.

References: [4768]

[EC 1.3.99.37 created 2015]

EC 1.3.99.38

Accepted name: menaquinone-9 β -reductase

Reaction: menaquinone-9 + reduced acceptor = β -dihydromenaquinone-9 + acceptor

Other name(s): MenJ

Systematic name: menaquinone-9 oxidoreductase (β -dihydromenaquinone-9-forming)

Comments: The enzyme from the bacterium *Mycobacterium tuberculosis* reduces the β -isoprene unit of menaquinone-9, forming the predominant form of menaquinone found in mycobacteria. Contains FAD.

References: [4377]

[EC 1.3.99.38 created 2017]

EC 1.3.99.39

Accepted name: carotenoid ϕ -ring synthase

Reaction: carotenoid β -end group + 2 acceptor = carotenoid ϕ -end group + 2 reduced acceptor

Other name(s): *crtU* (gene name) (ambiguous)

Systematic name: carotenoid β -ring:acceptor oxidoreductase/methyltransferase (ϕ -ring-forming)

Comments: The enzyme, found in green sulfur bacteria, some cyanobacteria and some actinobacteria, introduces additional double bonds to the carotenoid β -end group, leading to aromatization of the ionone ring. As a result, one of the methyl groups at C-1 is transferred to position C-2. It is involved in the biosynthesis of carotenoids with ϕ -type aromatic end groups such as chlorobactene, β -isorenieratene, and isorenieratene.

References: [2907, 2267, 1188]

[EC 1.3.99.39 created 2018]

EC 1.3.99.40

Accepted name: carotenoid χ -ring synthase

Reaction: carotenoid β -end group + 2 acceptor = carotenoid χ -end group + 2 reduced acceptor

Other name(s): *crtU* (gene name) (ambiguous); *cruE* (gene name)
Systematic name: carotenoid β -ring:acceptor oxidoreductase/methyltransferase (χ -ring-forming)
Comments: The enzyme, found in purple sulfur bacteria (*Chromatiaceae*) and some cyanobacteria, is involved in the biosynthesis of carotenoids that contain χ -type end groups, such as okenone, renierapurpurin, and synechoxanthin.
References: [1381, 4458]

[EC 1.3.99.40 created 2018]

EC 1.3.99.41

Accepted name: 3-(methylsulfanyl)propanoyl-CoA 2-dehydrogenase
Reaction: 3-(methylsulfanyl)propanoyl-CoA + acceptor = 3-(methylsulfanyl)acryloyl-CoA + reduced acceptor
Other name(s): *dmdC* (gene name)
Systematic name: 3-(methylsulfanyl)propanoyl-CoA:acceptor 2-oxidoreductase
Comments: The enzyme, found in marine bacteria, participates in a 3-(methylsulfanyl)propanoate degradation pathway. Based on similar enzymes, the *in vivo* electron acceptor is likely electron-transfer flavoprotein (ETF).
References: [3493, 491, 3822]

[EC 1.3.99.41 created 2022]

EC 1.4 Acting on the CH-NH₂ group of donors

This subclass contains the amino-acid dehydrogenases and the amine oxidases. In most cases, the imine formed is hydrolysed to give an oxo-group and NH₃. This is indicated as "(deaminating)". Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.4.1), a cytochrome (EC 1.4.2), oxygen (EC 1.4.3), a disulfide (EC 1.4.4), an iron-sulfur protein (EC 1.4.7), or some other acceptor (EC 1.4.99).

EC 1.4.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.4.1.1

Accepted name: alanine dehydrogenase
Reaction: L-alanine + H₂O + NAD⁺ = pyruvate + NH₃ + NADH + H⁺
Other name(s): AlaDH; L-alanine dehydrogenase; NAD-linked alanine dehydrogenase; α -alanine dehydrogenase; NAD-dependent alanine dehydrogenase; alanine oxidoreductase; NADH-dependent alanine dehydrogenase
Systematic name: L-alanine:NAD⁺ oxidoreductase (deaminating)
References: [3131, 3317, 4805]

[EC 1.4.1.1 created 1961]

EC 1.4.1.2

Accepted name: glutamate dehydrogenase
Reaction: L-glutamate + H₂O + NAD⁺ = 2-oxoglutarate + NH₃ + NADH + H⁺
Other name(s): glutamic dehydrogenase; glutamate dehydrogenase (NAD); glutamate oxidoreductase; glutamic acid dehydrogenase; L-glutamate dehydrogenase; NAD-dependent glutamate dehydrogenase; NAD-dependent glutamic dehydrogenase; NAD-glutamate dehydrogenase; NAD-linked glutamate dehydrogenase; NAD-linked glutamic dehydrogenase; NAD-specific glutamic dehydrogenase; NAD-specific glutamate dehydrogenase; NAD:glutamate oxidoreductase; NADH-linked glutamate dehydrogenase
Systematic name: L-glutamate:NAD⁺ oxidoreductase (deaminating)
References: [1180, 3089, 3220, 3934]

[EC 1.4.1.2 created 1961]

EC 1.4.1.3

Accepted name: glutamate dehydrogenase [NAD(P)⁺]
Reaction: L-glutamate + H₂O + NAD(P)⁺ = 2-oxoglutarate + NH₃ + NAD(P)H + H⁺
Other name(s): glutamic dehydrogenase; glutamate dehydrogenase [NAD(P)]
Systematic name: L-glutamate:NAD(P)⁺ oxidoreductase (deaminating)
References: [3176, 3934, 4057]

[EC 1.4.1.3 created 1961]

EC 1.4.1.4

Accepted name: glutamate dehydrogenase (NADP⁺)
Reaction: L-glutamate + H₂O + NADP⁺ = 2-oxoglutarate + NH₃ + NADPH + H⁺
Other name(s): glutamic dehydrogenase; dehydrogenase, glutamate (nicotinamide adenine dinucleotide (phosphate)); glutamic acid dehydrogenase; L-glutamate dehydrogenase; L-glutamic acid dehydrogenase; NAD(P)-glutamate dehydrogenase; NAD(P)H-dependent glutamate dehydrogenase; glutamate dehydrogenase (NADP)
Systematic name: L-glutamate:NADP⁺ oxidoreductase (deaminating)
References: [751, 1415, 3859, 3934]

[EC 1.4.1.4 created 1961]

EC 1.4.1.5

Accepted name: L-amino-acid dehydrogenase
Reaction: an L-amino acid + H₂O + NAD⁺ = a 2-oxo carboxylate + NH₃ + NADH + H⁺
Systematic name: L-amino-acid:NAD⁺ oxidoreductase (deaminating)
Comments: Acts on aliphatic amino acids.
References: [3090]

[EC 1.4.1.5 created 1961]

[1.4.1.6 Deleted entry. D-proline reductase. Now included with EC 1.21.4.1, D-proline reductase (dithiol)]

[EC 1.4.1.6 created 1961, deleted 1982]

EC 1.4.1.7

Accepted name: serine 2-dehydrogenase
Reaction: L-serine + H₂O + NAD⁺ = 3-hydroxypyruvate + NH₃ + NADH + H⁺
Other name(s): L-serine:NAD oxidoreductase (deaminating); serine dehydrogenase
Systematic name: L-serine:NAD⁺ 2-oxidoreductase (deaminating)
References: [2262]

[EC 1.4.1.7 created 1972, modified 2003]

EC 1.4.1.8

Accepted name: valine dehydrogenase (NADP⁺)
Reaction: L-valine + H₂O + NADP⁺ = 3-methyl-2-oxobutanoate + NH₃ + NADPH + H⁺
Other name(s): valine dehydrogenase (nicotinamide adenine dinucleotide phosphate); valine dehydrogenase (NADP)
Systematic name: L-valine:NADP⁺ oxidoreductase (deaminating)
References: [1973, 1974, 1975]

[EC 1.4.1.8 created 1972]

EC 1.4.1.9

Accepted name: leucine dehydrogenase
Reaction: L-leucine + H₂O + NAD⁺ = 4-methyl-2-oxopentanoate + NH₃ + NADH + H⁺
Other name(s): L-leucine dehydrogenase; L-leucine:NAD⁺ oxidoreductase, deaminating; LeuDH
Systematic name: L-leucine:NAD⁺ oxidoreductase (deaminating)
Comments: Also acts on isoleucine, valine, norvaline and norleucine.
References: [3662, 4933]

[EC 1.4.1.9 created 1972]

EC 1.4.1.10

Accepted name: glycine dehydrogenase
Reaction: glycine + H₂O + NAD⁺ = glyoxylate + NH₃ + NADH + H⁺
Systematic name: glycine:NAD⁺ oxidoreductase (deaminating)
References: [1351]

[EC 1.4.1.10 created 1972]

EC 1.4.1.11

Accepted name: L-erythro-3,5-diaminohexanoate dehydrogenase
Reaction: L-erythro-3,5-diaminohexanoate + H₂O + NAD⁺ = (S)-5-amino-3-oxohexanoate + NH₃ + NADH + H⁺
Other name(s): L-3,5-diaminohexanoate dehydrogenase
Systematic name: L-erythro-3,5-diaminohexanoate:NAD⁺ oxidoreductase (deaminating)
References: [194]

[EC 1.4.1.11 created 1976]

EC 1.4.1.12

Accepted name: 2,4-diaminopentanoate dehydrogenase
Reaction: (2R,4S)-2,4-diaminopentanoate + H₂O + NAD(P)⁺ = (2R)-2-amino-4-oxopentanoate + NH₃ + NAD(P)H + H⁺
Other name(s): 2,4-diaminopentanoic acid C₄ dehydrogenase
Systematic name: (2R,4S)-2,4-diaminopentanoate:NAD(P)⁺ oxidoreductase (deaminating)
Comments: Also acts, more slowly, on 2,5-diaminohexanoate forming 2-amino-5-oxohexanoate, which then cyclizes non-enzymically to 1-pyrroline-2-methyl-5-carboxylate. It has equal activity with NAD⁺ and NADP⁺ [cf. EC 1.4.1.26, 2,4-diaminopentanoate dehydrogenase (NAD⁺)].
References: [3956, 3996, 4339]

[EC 1.4.1.12 created 1976, modified 2017]

EC 1.4.1.13

Accepted name: glutamate synthase (NADPH)
Reaction: 2 L-glutamate + NADP⁺ = L-glutamine + 2-oxoglutarate + NADPH + H⁺ (overall reaction)
(1a) L-glutamate + NH₃ = L-glutamine + H₂O
(1b) L-glutamate + NADP⁺ + H₂O = NH₃ + 2-oxoglutarate + NADPH + H⁺
Other name(s): glutamate (reduced nicotinamide adenine dinucleotide phosphate) synthase; L-glutamate synthase; L-glutamate synthetase; glutamate synthetase (NADP); NADPH-dependent glutamate synthase; glutamine-ketoglutaric aminotransferase; NADPH-glutamate synthase; NADPH-linked glutamate synthase; glutamine amide-2-oxoglutarate aminotransferase (oxidoreductase, NADP); L-glutamine:2-oxoglutarate aminotransferase, NADPH oxidizing; GOGAT
Systematic name: L-glutamate:NADP⁺ oxidoreductase (transaminating)

Comments: Binds FMN, FAD, 2 [4Fe-4S] clusters and 1 [3Fe-4S] cluster. The reaction takes place in the direction of L-glutamate production. The protein is composed of two subunits, α and β . The α subunit is composed of two domains, one hydrolysing L-glutamine to NH_3 and L-glutamate (*cf.* EC 3.5.1.2, glutaminase), the other combining the produced NH_3 with 2-oxoglutarate to produce a second molecule of L-glutamate (*cf.* EC 1.4.1.4, glutamate dehydrogenase [NADP^+]). The β subunit transfers electrons from the cosubstrate. The NH_3 is channeled within the α subunit through a 31 Å channel. The channelling is very efficient and in the intact α - β complex ammonia is produced only within the complex. In the absence of the β subunit, coupling between the two domains of the α subunit is compromised and some ammonium can leak.

References: [2812, 4239, 4425, 3459]

[EC 1.4.1.13 created 1972 as EC 2.6.1.53, transferred 1976 to EC 1.4.1.13, modified 2001, modified 2012]

EC 1.4.1.14

Accepted name: glutamate synthase (NADH)

Reaction: 2 L-glutamate + NAD^+ = L-glutamine + 2-oxoglutarate + NADH + H^+

(1a) L-glutamate + NH_3 = L-glutamine + H_2O

(1b) L-glutamate + NAD^+ + H_2O = NH_3 + 2-oxoglutarate + NADH + H^+

Other name(s): glutamate (reduced nicotinamide adenine dinucleotide) synthase; NADH: GOGAT; L-glutamate synthase (NADH); L-glutamate synthetase; NADH-glutamate synthase; NADH-dependent glutamate synthase

Systematic name: L-glutamate: NAD^+ oxidoreductase (transaminating)

Comments: A flavoprotein (FMN). The reaction takes place in the direction of L-glutamate production. The protein is composed of two domains, one hydrolysing L-glutamine to NH_3 and L-glutamate (*cf.* EC 3.5.1.2, glutaminase), the other combining the produced NH_3 with 2-oxoglutarate to produce a second molecule of L-glutamate (*cf.* EC 1.4.1.2, glutamate dehydrogenase).

References: [3569, 375, 2692, 95, 355]

[EC 1.4.1.14 created 1978, modified 2019]

EC 1.4.1.15

Accepted name: lysine dehydrogenase

Reaction: L-lysine + NAD^+ = 1,2-didehydropiperidine-2-carboxylate + NH_3 + NADH + H^+

Systematic name: L-lysine: NAD^+ oxidoreductase (deaminating, cyclizing)

References: [496]

[EC 1.4.1.15 created 1978]

EC 1.4.1.16

Accepted name: diaminopimelate dehydrogenase

Reaction: *meso*-2,6-diaminoheptanedioate + H_2O + NADP^+ = L-2-amino-6-oxoheptanedioate + NH_3 + NADPH + H^+

Other name(s): *meso*- α,ϵ -diaminopimelate dehydrogenase; *meso*-diaminopimelate dehydrogenase

Systematic name: *meso*-2,6-diaminoheptanedioate: NADP^+ oxidoreductase (deaminating)

References: [2823, 2824]

[EC 1.4.1.16 created 1981]

EC 1.4.1.17

Accepted name: *N*-methylalanine dehydrogenase

Reaction: *N*-methyl-L-alanine + H_2O + NADP^+ = pyruvate + methylamine + NADPH + H^+

Systematic name: *N*-methyl-L-alanine: NADP^+ oxidoreductase (demethylating, deaminating)

References: [2492]

[EC 1.4.1.17 created 1984]

EC 1.4.1.18

- Accepted name:** lysine 6-dehydrogenase
Reaction: L-lysine + NAD⁺ = (S)-2,3,4,5-tetrahydropyridine-2-carboxylate + NADH + H⁺ + NH₃ (overall reaction)
(1a) L-lysine + NAD⁺ + H₂O = (S)-2-amino-6-oxohexanoate + NADH + H⁺ + NH₃
(1b) (S)-2-amino-6-oxohexanoate = (S)-2,3,4,5-tetrahydropyridine-2-carboxylate + H₂O (spontaneous)
Other name(s): L-lysine ε-dehydrogenase; L-lysine 6-dehydrogenase; LysDH
Systematic name: L-lysine:NAD⁺ 6-oxidoreductase (deaminating)
Comments: The enzyme is highly specific for L-lysine as substrate, although S-(2-aminoethyl)-L-cysteine can act as a substrate, but more slowly. While the enzyme from *Agrobacterium tumefaciens* can use only NAD⁺, that from the thermophilic bacterium *Geobacillus stearothermophilus* can also use NADP⁺, but more slowly [2822, 1635].
References: [2822, 2825, 2821, 1635]

[EC 1.4.1.18 created 1989, modified 2006, modified 2011]

EC 1.4.1.19

- Accepted name:** tryptophan dehydrogenase
Reaction: L-tryptophan + NAD(P)⁺ + H₂O = (indol-3-yl)pyruvate + NH₃ + NAD(P)H + H⁺
Other name(s): NAD(P)⁺-L-tryptophan dehydrogenase; L-tryptophan dehydrogenase; L-Trp-dehydrogenase; TDH
Systematic name: L-tryptophan:NAD(P)⁺ oxidoreductase (deaminating)
Comments: Activated by Ca²⁺.
References: [4386]

[EC 1.4.1.19 created 1989]

EC 1.4.1.20

- Accepted name:** phenylalanine dehydrogenase
Reaction: L-phenylalanine + H₂O + NAD⁺ = phenylpyruvate + NH₃ + NADH + H⁺
Other name(s): L-phenylalanine dehydrogenase; PHD
Systematic name: L-phenylalanine:NAD⁺ oxidoreductase (deaminating)
Comments: The enzymes from *Bacillus badius* and *Sporosarcina ureae* are highly specific for L-phenylalanine; that from *Bacillus sphaericus* also acts on L-tyrosine.
References: [141, 142]

[EC 1.4.1.20 created 1989]

EC 1.4.1.21

- Accepted name:** aspartate dehydrogenase
Reaction: L-aspartate + H₂O + NAD(P)⁺ = oxaloacetate + NH₃ + NAD(P)H + H⁺ (overall reaction)
(1a) L-aspartate + NAD(P)⁺ = 2-iminosuccinate + NAD(P)H + H⁺
(1b) 2-iminosuccinate + H₂O = oxaloacetate + NH₃ (spontaneous)
Other name(s): AspDH; NAD-dependent aspartate dehydrogenase; NADH₂-dependent aspartate dehydrogenase; NADP⁺-dependent aspartate dehydrogenase; *nadX* (gene name); L-aspartate:NAD(P)⁺ oxidoreductase (deaminating)
Systematic name: L-aspartate:NAD(P)⁺ oxidoreductase (2-iminosuccinate-forming)
Comments: The enzyme is strictly specific for L-aspartate as substrate. It produces the unstable compound 2-iminosuccinate, which, in the presence of water, hydrolyses spontaneously to form oxaloacetate. The enzyme from some archaea and thermophilic bacteria is likely to transfer 2-iminosuccinate directly to EC 2.5.1.72, quinolinate synthase, preventing its hydrolysis and enabling the *de novo* biosynthesis of NAD⁺.

References: [2263, 3158, 4773, 4798, 4799, 2463, 2462, 2466]

[EC 1.4.1.21 created 2005, modified 2022]

[1.4.1.22 Deleted entry: ornithine cyclodeaminase. It was pointed out during the public-review process that there is no overall consumption of NAD⁺ during the reaction. As a result, transfer of the enzyme from EC 4.3.1.12 was not necessary and EC 1.4.1.22 was withdrawn before being made official]

[EC 1.4.1.22 created 2006, deleted 2006]

EC 1.4.1.23

Accepted name: valine dehydrogenase (NAD⁺)
Reaction: L-valine + H₂O + NAD⁺ = 3-methyl-2-oxobutanoate + NH₃ + NADH + H⁺
Systematic name: L-valine:NAD⁺ oxidoreductase (deaminating)
Comments: The enzyme from *Streptomyces* spp. has no activity with NADP⁺ [cf. EC 1.4.1.8, valine dehydrogenase (NADP⁺)].
References: [4418, 3025]

[EC 1.4.1.23 created 2012]

EC 1.4.1.24

Accepted name: 3-dehydroquininate synthase II
Reaction: 2-amino-3,7-dideoxy-D-threo-hept-6-ulose-6-phosphate + H₂O + NAD⁺ = 3-dehydroquininate + NH₃ + NADH + H⁺
Other name(s): DHQ synthase II; MJ1249 (gene name); *aroB'* (gene name)
Systematic name: 2-amino-3,7-dideoxy-D-threo-hept-6-ulose-6-phosphate:NAD⁺ oxidoreductase (deaminating)
Comments: The enzyme, which was isolated from the archaeon *Methanocaldococcus jannaschii*, plays a key role in an alternative pathway for the biosynthesis of 3-dehydroquininate (DHQ), an intermediate of the canonical pathway for the biosynthesis of aromatic amino acids. The enzyme catalyses a two-step reaction - an oxidative deamination, followed by cyclization.
References: [4604]

[EC 1.4.1.24 created 2012]

EC 1.4.1.25

Accepted name: L-arginine dehydrogenase
Reaction: L-arginine + H₂O + NAD(P)⁺ = 5-guanidino-2-oxopentanoate + NH₃ + NAD(P)H + H⁺
Other name(s): *dauB* (gene name); anabolic L-arginine dehydrogenase
Systematic name: L-arginine:NAD(P)⁺ oxidoreductase (deaminating)
Comments: The enzyme, which has been isolated from the bacterium *Pseudomonas aeruginosa* PAO1, forms with EC 1.4.99.6, D-arginine dehydrogenase, a two-enzyme complex involved in the racemization of D- and L-arginine.
References: [2440]

[EC 1.4.1.25 created 2017]

EC 1.4.1.26

Accepted name: 2,4-diaminopentanoate dehydrogenase (NAD⁺)
Reaction: (2R,4S)-2,4-diaminopentanoate + H₂O + NAD⁺ = (2R)-2-amino-4-oxopentanoate + NH₃ + NADH + H⁺
Other name(s): DAPDH (ambiguous)
Systematic name: (2R,4S)-2,4-diaminopentanoate:NADP⁺ oxidoreductase (deaminating)

Comments: The enzyme, characterized from an unknown bacterium in an environmental sample, has some activity with (2*R*,4*R*)-2,4-diaminopentanoate. It has very low activity with NADP⁺ (*cf.* EC 1.4.1.12, 2,4-diaminopentanoate dehydrogenase).

References: [1135]

[EC 1.4.1.26 created 2017]

EC 1.4.1.27

Accepted name: glycine cleavage system
Reaction: glycine + tetrahydrofolate + NAD⁺ = 5,10-methylenetetrahydrofolate + NH₃ + CO₂ + NADH
Other name(s): GCV
Systematic name: glycine:NAD⁺ 2-oxidoreductase (tetrahydrofolate-methylene-adding)
Comments: The glycine cleavage (GCV) system is a large multienzyme complex that belongs to the 2-oxoacid dehydrogenase complex family, which also includes EC 1.2.1.25, branched-chain α-keto acid dehydrogenase system, EC 1.2.1.105, 2-oxoglutarate dehydrogenase system, EC 1.2.1.104, pyruvate dehydrogenase system, and EC 2.3.1.190, acetoin dehydrogenase system. The GCV system catalyses the reversible oxidation of glycine, yielding carbon dioxide, ammonia, 5,10-methylenetetrahydrofolate and a reduced pyridine nucleotide. Tetrahydrofolate serves as a recipient for one-carbon units generated during glycine cleavage to form the methylene group. The GCV system consists of four protein components, the P protein (EC 1.4.4.2, glycine dehydrogenase (aminomethyl-transferring)), T protein (EC 2.1.2.10, aminomethyltransferase), L protein (EC 1.8.1.4, dihydrolipoyl dehydrogenase), and the non-enzyme H protein (lipoyl-carrier protein). The P protein catalyses the pyridoxal phosphate-dependent liberation of CO₂ from glycine, leaving a methylamine moiety. The methylamine moiety is transferred to the lipoic acid group of the H protein, which is bound to the P protein prior to decarboxylation of glycine. The T protein catalyses the release of ammonia from the methylamine group and transfers the remaining C₁ unit to tetrahydrofolate, forming 5,10-methylenetetrahydrofolate. The L protein then oxidizes the lipoic acid component of the H protein and transfers the electrons to NAD⁺, forming NADH.
References: [2910, 1665, 3159, 1216, 3160]

[EC 1.4.1.27 created 2020]

EC 1.4.1.28

Accepted name: secondary-alkyl amine dehydrogenase [NAD(P)⁺]
Reaction: a secondary-alkyl amine + H₂O + NAD(P)⁺ = a ketone + NH₃ + NAD(P)H + H⁺
Other name(s): AmDH (ambiguous); amine dehydrogenase (ambiguous)
Systematic name: secondary-alkyl amine:NAD(P)⁺ oxidoreductase (deaminating)
Comments: The enzyme has been shown to react preferentially with short-chain ketones such as cyclohexanone, primary amine groups attached to secondary alkyl groups, or D- and L-amino acids. It also reduces aldehydes to primary amines. Cosubstrate preference depends on the substrate.
References: [1850, 2733, 2732, 2392]

[EC 1.4.1.28 created 2022]

EC 1.4.2 With a cytochrome as acceptor

EC 1.4.2.1

Accepted name: glycine dehydrogenase (cytochrome)
Reaction: glycine + H₂O + 2 ferricytochrome *c* = glyoxylate + NH₃ + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): glycine—cytochrome *c* reductase
Systematic name: glycine:ferricytochrome-*c* oxidoreductase (deaminating)
References: [3655]

[EC 1.4.2.1 created 1976]

EC 1.4.2.2

Accepted name: nicotine dehydrogenase
Reaction: (*S*)-nicotine + 2 ferricytochrome *c* = *N*-methylmyosmine + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): *nicA2* (gene name)
Systematic name: (*S*)-nicotine:cytochrome *c* oxidoreductase (*N*-methylmimosine-forming)
Comments: The enzyme, characterized from the bacterium *Pseudomonas putida* S16, contains an FAD cofactor and belongs to the flavin-containing amine oxidase family. The enzyme from this bacterium is specific for the *c*-type cytochrome CycN. The product undergoes spontaneous hydrolysis to form pseudooxynicotine.
References: [4201, 982]

[EC 1.4.2.2 created 2022]

EC 1.4.2.3

Accepted name: pseudooxynicotine dehydrogenase
Reaction: pseudooxynicotine + H₂O + 2 ferricytochrome *c* = 4-oxo-4-(pyridin-3-yl)butanal + methylamine + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): *pnaO* (gene name)
Systematic name: 4-(methylamino)-1-(pyridin-3-yl)butan-1-one:*c*-type cytochrome oxidoreductase (methylamine releasing)
Comments: Contains one non-covalently bound FAD molecule per dimer. This enzyme, characterized from the soil bacteria *Pseudomonas* sp. HZN6 and *Pseudomonas putida* S16, is involved in nicotine degradation.
References: [3402, 679]

[EC 1.4.2.3 created 2012 as EC 1.4.3.24, transferred 2022 to EC 1.4.2.3]

EC 1.4.3 With oxygen as acceptor

EC 1.4.3.1

Accepted name: D-aspartate oxidase
Reaction: D-aspartate + H₂O + O₂ = oxaloacetate + NH₃ + H₂O₂
Other name(s): aspartic oxidase; D-aspartic oxidase
Systematic name: D-aspartate:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD).
References: [925, 4031, 4032]

[EC 1.4.3.1 created 1961]

EC 1.4.3.2

Accepted name: L-amino-acid oxidase
Reaction: an L-amino acid + H₂O + O₂ = a 2-oxo carboxylate + NH₃ + H₂O₂
Other name(s): ophio-amino-acid oxidase (ambiguous)
Systematic name: L-amino-acid:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD).
References: [2766, 4585]

[EC 1.4.3.2 created 1961]

EC 1.4.3.3

- Accepted name:** D-amino-acid oxidase
Reaction: a D-amino acid + H₂O + O₂ = a 2-oxo carboxylate + NH₃ + H₂O₂
Other name(s): ophio-amino-acid oxidase (ambiguous); L-amino acid:O₂ oxidoreductase; new yellow enzyme
Systematic name: D-amino-acid:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD). Wide specificity for D-amino acids. Also acts on glycine.
References: [926, 928, 927, 2687, 2766]

[EC 1.4.3.3 created 1961]

EC 1.4.3.4

- Accepted name:** monoamine oxidase
Reaction: RCH₂NHR' + H₂O + O₂ = RCHO + R'NH₂ + H₂O₂
Other name(s): adrenalin oxidase; adrenaline oxidase; amine oxidase (ambiguous); amine oxidase (flavin-containing); amine:oxygen oxidoreductase (deaminating) (flavin-containing); epinephrine oxidase; MAO; MAO A; MAO B; MAO-A; MAO-B; monoamine oxidase A; monoamine oxidase B; monoamine:O₂ oxidoreductase (deaminating); polyamine oxidase (ambiguous); serotonin deaminase; spermidine oxidase (ambiguous); spermine oxidase (ambiguous); tyraminase; tyramine oxidase
Systematic name: amine:oxygen oxidoreductase (deaminating)
Comments: A mitochondrial outer-membrane flavoprotein (FAD) that catalyses the oxidative deamination of neurotransmitters and biogenic amines [1019]. Acts on primary amines, and also on some secondary and tertiary amines. It differs from EC 1.4.3.21, primary-amine oxidase as it can oxidize secondary and tertiary amines but not methylamine. This enzyme is inhibited by acetylenic compounds such as chlorgyline, 1-deprenyl and pargyline but, unlike EC 1.4.3.21 and EC 1.4.3.22 (diamine oxidase), it is not inhibited by semicarbazide.
References: [356, 954, 1019, 3858, 4294, 711, 4826, 4825]

[EC 1.4.3.4 created 1961, modified 1983 (EC 1.4.3.9 created 1972, incorporated 1984), modified 2008]

EC 1.4.3.5

- Accepted name:** pyridoxal 5'-phosphate synthase
Reaction: (1) pyridoxamine 5'-phosphate + H₂O + O₂ = pyridoxal 5'-phosphate + NH₃ + H₂O₂
(2) pyridoxine 5'-phosphate + O₂ = pyridoxal 5'-phosphate + H₂O₂
Other name(s): pyridoxamine 5'-phosphate oxidase; pyridoxamine phosphate oxidase; pyridoxine (pyridoxamine)phosphate oxidase; pyridoxine (pyridoxamine) 5'-phosphate oxidase; pyridoxaminephosphate oxidase (EC 1.4.3.5: deaminating); PMP oxidase; pyridoxol-5'-phosphate:oxygen oxidoreductase (deaminating) (incorrect); pyridoxamine-phosphate oxidase; PdxH
Systematic name: pyridoxamine-5'-phosphate:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FMN). In *Escherichia coli*, the coenzyme pyridoxal 5'-phosphate is synthesized *de novo* by a pathway that involves EC 1.2.1.72 (erythrose-4-phosphate dehydrogenase), EC 1.1.1.290 (4-phosphoerythronate dehydrogenase), EC 2.6.1.52 (phosphoserine transaminase), EC 1.1.1.262 (4-hydroxythreonine-4-phosphate dehydrogenase), EC 2.6.99.2 (pyridoxine 5'-phosphate synthase) and EC 1.4.3.5 (with pyridoxine 5'-phosphate as substrate). N^{4'}-Substituted pyridoxamine derivatives are also oxidized in reaction (1) to form pyridoxal 5-phosphate and the corresponding primary amine.
References: [672, 4483, 3109, 2322, 2950, 3626, 4890]

[EC 1.4.3.5 created 1961, modified 2006]

[1.4.3.6 Deleted entry. amine oxidase (copper-containing). This was classified on the basis of cofactor content rather than reaction catalysed and is now known to contain two distinct enzyme activities. It has been replaced by two enzymes, EC 1.4.3.21 (primary-amine oxidase) and EC 1.4.3.22 (diamine oxidase)]

[EC 1.4.3.6 created 1961, modified 1983, modified 1989, deleted 2008]

EC 1.4.3.7

Accepted name: D-glutamate oxidase
Reaction: D-glutamate + H₂O + O₂ = 2-oxoglutarate + NH₃ + H₂O₂
Other name(s): D-glutamic oxidase; D-glutamic acid oxidase
Systematic name: D-glutamate:oxygen oxidoreductase (deaminating)
References: [3541, 4379]

[EC 1.4.3.7 created 1972]

EC 1.4.3.8

Accepted name: ethanolamine oxidase
Reaction: ethanolamine + H₂O + O₂ = glycolaldehyde + NH₃ + H₂O₂
Systematic name: ethanolamine:oxygen oxidoreductase (deaminating)
Comments: A cobamide-protein.
References: [3013]

[EC 1.4.3.8 created 1972]

[1.4.3.9 Deleted entry. tyramine oxidase. Now included with EC 1.4.3.4 amine oxidase (flavin-containing)]

[EC 1.4.3.9 created 1972, deleted 1984]

EC 1.4.3.10

Accepted name: putrescine oxidase
Reaction: putrescine + O₂ + H₂O = 4-aminobutanal + NH₃ + H₂O₂
Systematic name: putrescine:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD). 4-Aminobutanal condenses non-enzymically to 1-pyrroline.
References: [888, 4723]

[EC 1.4.3.10 created 1976]

EC 1.4.3.11

Accepted name: L-glutamate oxidase
Reaction: L-glutamate + O₂ + H₂O = 2-oxoglutarate + NH₃ + H₂O₂
Other name(s): glutamate (acceptor) dehydrogenase; glutamate oxidase; glutamic acid oxidase; glutamic dehydrogenase (acceptor); L-glutamic acid oxidase
Systematic name: L-glutamate:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD). The enzyme from *Azotobacter* previously listed under this number, which did not produce H₂O₂, was a crude cell-free extract that probably contained catalase.
References: [2309]

[EC 1.4.3.11 created 1976, modified 1989]

EC 1.4.3.12

Accepted name: cyclohexylamine oxidase
Reaction: cyclohexylamine + O₂ + H₂O = cyclohexanone + NH₃ + H₂O₂
Systematic name: cyclohexylamine:oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD). Some other cyclic amines can act instead of cyclohexylamine, but not simple aliphatic and aromatic amides.
References: [4301]

[EC 1.4.3.12 created 1978]

EC 1.4.3.13

- Accepted name:** protein-lysine 6-oxidase
Reaction: [protein]-L-lysine + O₂ + H₂O = [protein]-(S)-2-amino-6-oxohexanoate + NH₃ + H₂O₂
Other name(s): lysyl oxidase
Systematic name: protein-L-lysine:oxygen 6-oxidoreductase (deaminating)
Comments: Also acts on protein 5-hydroxylysine. This enzyme catalyses the final known enzymic step required for collagen and elastin cross-linking in the biosynthesis of normal mature extracellular matrices [3223]. These reactions play an important role for the development, elasticity and extensibility of connective tissue. The enzyme is also active on free amines, such as cadaverine or benzylamine [3223, 1972]. Some isoforms can also use [protein]-N(6)-acetyl-L-lysine as substrate deacetamidating the substrate [3548].
References: [1535, 3468, 4008, 3223, 1972, 3548, 2113, 4705, 2580]

[EC 1.4.3.13 created 1980, modified 1983]

EC 1.4.3.14

- Accepted name:** L-lysine oxidase
Reaction: L-lysine + O₂ + H₂O = 6-amino-2-oxohexanoate + NH₃ + H₂O₂
Other name(s): L-lysine α-oxidase; L-lysyl-α-oxidase
Systematic name: L-lysine:oxygen 2-oxidoreductase (deaminating)
Comments: Also acts, more slowly, on L-ornithine, L-phenylalanine, L-arginine and L-histidine.
References: [2307, 2558]

[EC 1.4.3.14 created 1981]

EC 1.4.3.15

- Accepted name:** D-glutamate(D-aspartate) oxidase
Reaction: (1) D-glutamate + H₂O + O₂ = 2-oxoglutarate + NH₃ + H₂O₂
(2) D-aspartate + H₂O + O₂ = oxaloacetate + NH₃ + H₂O₂
Other name(s): D-glutamic-aspartic oxidase; D-monoaminodicarboxylic acid oxidase
Systematic name: D-glutamate(D-aspartate):oxygen oxidoreductase (deaminating)
Comments: A flavoprotein (FAD). D-Glutamate and D-aspartate are oxidized at the same rate. Other D-monoaminodicarboxylates, and other D- and L-amino acids, are not oxidized. *cf.* EC 1.4.3.7, D-glutamate oxidase and EC 1.4.3.1, D-aspartate oxidase.
References: [2849]

[EC 1.4.3.15 created 1983, modified 2012]

EC 1.4.3.16

- Accepted name:** L-aspartate oxidase
Reaction: L-aspartate + O₂ = iminosuccinate + H₂O₂
Other name(s): NadB; Laspo; AO
Systematic name: L-aspartate:oxygen oxidoreductase
Comments: A flavoprotein (FAD). L-Aspartate oxidase catalyses the first step in the *de novo* biosynthesis of NAD⁺ in some bacteria. O₂ can be replaced by fumarate as electron acceptor, yielding succinate [402]. The ability of the enzyme to use both O₂ and fumarate in cofactor reoxidation enables it to function under both aerobic and anaerobic conditions [402]. Iminosuccinate can either be hydrolysed to form oxaloacetate and NH₃ or can be used by EC 2.5.1.72, quinolinate synthase, in the production of quinolinate. The enzyme is a member of the succinate dehydrogenase/fumarate-reductase family of enzymes [402].
References: [3021, 2903, 4236, 2722, 402, 2024]

[EC 1.4.3.16 created 1984, modified 2008]

[1.4.3.17 Transferred entry. tryptophan α,β -oxidase. Now EC 1.3.3.10, tryptophan α,β -oxidase. Enzyme was incorrectly classified as acting on a CH-NH bond rather than a CH-CH bond]

[EC 1.4.3.17 created 2000, deleted 2003]

[1.4.3.18 Deleted entry. cytokinin oxidase. Not approved as the enzyme was shown to be a dehydrogenase and not an oxidase (see EC 1.5.99.12, cytokinin dehydrogenase)]

[EC 1.4.3.18 proposed 2000]

EC 1.4.3.19

Accepted name: glycine oxidase
Reaction: glycine + H₂O + O₂ = glyoxylate + NH₃ + H₂O₂ (overall reaction)
(1a) glycine + O₂ = 2-iminoacetate + H₂O₂
(1b) 2-iminoacetate + H₂O = glyoxylate + NH₃
Systematic name: glycine:oxygen oxidoreductase (deaminating)
Comments: A flavoenzyme containing non-covalently bound FAD. The enzyme from *Bacillus subtilis* is active with glycine, sarcosine, *N*-ethylglycine, D-alanine, D- α -aminobutyrate, D-proline, D-pipecolate and *N*-methyl-D-alanine. It differs from EC 1.4.3.3, D-amino-acid oxidase, due to its activity on sarcosine and D-pipecolate. The intermediate 2-iminoacetate is used directly by EC 2.8.1.10, thiazole synthase.
References: [1921, 3085]

[EC 1.4.3.19 created 2002, modified 2012]

EC 1.4.3.20

Accepted name: L-lysine 6-oxidase
Reaction: L-lysine + O₂ + H₂O = (*S*)-2-amino-6-oxohexanoate + H₂O₂ + NH₃
Other name(s): L-lysine- ϵ -oxidase; Lod; LodA; marinocine
Systematic name: L-lysine:oxygen 6-oxidoreductase (deaminating)
Comments: Differs from EC 1.4.3.13, protein-lysine 6-oxidase, by using free L-lysine rather than the protein-bound form. *N*²-Acetyl-L-lysine is also a substrate, but *N*⁶-acetyl-L-lysine, which has an acetyl group at position 6, is not a substrate. Also acts on L-ornithine, D-lysine and 4-hydroxy-L-lysine, but more slowly. The amines cadaverine and putrescine are not substrates [1352].
References: [2555, 1352]

[EC 1.4.3.20 created 2006, modified 2011]

EC 1.4.3.21

Accepted name: primary-amine oxidase
Reaction: RCH₂NH₂ + H₂O + O₂ = RCHO + NH₃ + H₂O₂
Other name(s): amine oxidase (ambiguous); amine oxidase (copper-containing); amine oxidase (pyridoxal containing) (incorrect); benzylamine oxidase (incorrect); CAO (ambiguous); copper amine oxidase (ambiguous); Cu-amine oxidase (ambiguous); Cu-containing amine oxidase (ambiguous); diamine oxidase (incorrect); diamino oxhydrase (incorrect); histamine deaminase (ambiguous); histamine oxidase (ambiguous); monoamine oxidase (ambiguous); plasma monoamine oxidase (ambiguous); polyamine oxidase (ambiguous); semicarbazide-sensitive amine oxidase (ambiguous); SSAO (ambiguous)
Systematic name: primary-amine:oxygen oxidoreductase (deaminating)
Comments: A group of enzymes that oxidize primary monoamines but have little or no activity towards diamines, such as histamine, or towards secondary and tertiary amines. They are copper quinoproteins (2,4,5-trihydroxyphenylalanine quinone) and, unlike EC 1.4.3.4, monoamine oxidase, are sensitive to inhibition by carbonyl-group reagents, such as semicarbazide. In some mammalian tissues the enzyme also functions as a vascular-adhesion protein (VAP-1).
References: [1584, 4293, 2570, 4625, 2400, 1742, 101, 3690, 3202, 43]

[EC 1.4.3.21 created 2007 (EC 1.4.3.6 created 1961, part-incorporated 2008)]

EC 1.4.3.22

- Accepted name:** diamine oxidase
Reaction: histamine + H₂O + O₂ = (imidazol-4-yl)acetaldehyde + NH₃ + H₂O₂
Other name(s): amine oxidase (ambiguous); amine oxidase (copper-containing) (ambiguous); CAO (ambiguous); Cu-containing amine oxidase (ambiguous); copper amine oxidase (ambiguous); diamine oxidase (ambiguous); diamino oxyhydrase (ambiguous); histaminase; histamine deaminase (incorrect); semicarbazide-sensitive amine oxidase (incorrect); SSAO (incorrect)
Systematic name: histamine:oxygen oxidoreductase (deaminating)
Comments: A group of enzymes that oxidize diamines, such as histamine, and also some primary monoamines but have little or no activity towards secondary and tertiary amines. They are copper quinoproteins (2,4,5-trihydroxyphenylalanine quinone) and, like EC 1.4.3.21 (primary-amine oxidase) but unlike EC 1.4.3.4 (monoamine oxidase), they are sensitive to inhibition by carbonyl-group reagents, such as semicarbazide.
References: [4868, 763, 612, 1742, 1040]

[EC 1.4.3.22 created 2007 (EC 1.4.3.6 created 1961, part-incorporated 2008)]

EC 1.4.3.23

- Accepted name:** 7-chloro-L-tryptophan oxidase
Reaction: 7-chloro-L-tryptophan + O₂ = 2-imino-3-(7-chloroindol-3-yl)propanoate + H₂O₂
Other name(s): RebO
Systematic name: 7-chloro-L-tryptophan:oxygen oxidoreductase
Comments: Contains a noncovalently bound FAD [3086, 1743]. This enzyme catalyses a step in the biosynthesis of rebeccamycin, an indolocarbazole alkaloid produced by the bacterium *Lechevalieria aerocolonigenes*. During catalysis, the bound FAD is reoxidized at the expense of molecular oxygen, producing one molecule of hydrogen peroxide. The enzyme shows significant preference for 7-chloro-L-tryptophan over L-tryptophan [3086].
References: [3086, 1743]

[EC 1.4.3.23 created 2010]

[1.4.3.24 Transferred entry. pseudooxynicotine oxidase, now classified as EC 1.4.2.3, pseudooxynicotine dehydrogenase]

[EC 1.4.3.24 created 2012, deleted 2022]

EC 1.4.3.25

- Accepted name:** L-arginine oxidase
Reaction: L-arginine + H₂O + O₂ = 5-guanidino-2-oxopentanoate + NH₃ + H₂O₂
Systematic name: L-arginine:oxygen oxidoreductase (deaminating)
Comments: Contains FAD. The enzyme from cyanobacteria can also act on other basic amino acids with lower activity. The enzyme from the bacterium *Pseudomonas* sp. TPU 7192 is highly specific.
References: [2808, 3331, 1284, 2699]

[EC 1.4.3.25 created 2017]

EC 1.4.3.26

- Accepted name:** pre-mycofactocin synthase
Reaction: 3-amino-5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidin-2-one + O₂ + H₂O = 5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidine-2,3-dione + NH₃ + H₂O₂ (overall reaction)
(1a) 3-amino-5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidin-2-one + O₂ = 5-[(4-hydroxyphenyl)methyl]-3-imino-4,4-dimethylpyrrolidin-2-one + H₂O₂
(1b) 5-[(4-hydroxyphenyl)methyl]-3-imino-4,4-dimethylpyrrolidin-2-one + H₂O = 5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidine-2,3-dione + NH₃ (spontaneous)
Other name(s): *mftD* (gene name)

Systematic name: 3-amino-5-[(4-hydroxyphenyl)methyl]-4,4-dimethylpyrrolidin-2-one:oxygen oxidoreductase
Comments: A flavoprotein (FMN). The enzyme participates in the biosynthesis of the enzyme cofactor mycofocin. The enzyme uses oxygen as an electron source to oxidize a C-N bond, followed by spontaneous exchange with water to form an α -keto moiety on the resulting molecule.
References: [169]

[EC 1.4.3.26 created 2020]

EC 1.4.3.27

Accepted name: homospermidine oxidase
Reaction: *sym*-homospermidine + 2 O₂ + H₂O = 1-formylpyrrolizidine + 2 H₂O₂ + 2 NH₃ (overall reaction)
(1a) *sym*-homospermidine + O₂ = *N*-(4-aminobutylpyrrolinium) ion + H₂O₂ + NH₃
(1b) *N*-(4-aminobutylpyrrolinium) ion + O₂ + H₂O = *N*-(4-oxobutylpyrrolinium) ion + NH₃ + H₂O₂
(1c) *N*-(4-oxobutylpyrrolinium) ion = 1-formylpyrrolizidine (spontaneous)
Other name(s): HSO
Systematic name: homospermidine:oxygen oxidase (deaminating, cyclizing)
Comments: The copper-containing enzyme has been isolated from the plant *Heliotropium indicum*. It is involved in the biosynthesis of the pyrrolizidine alkaloid (–)-trachelanthamidine which acts as a secondary metabolite for the defense against herbivores. The oxidation of *sym*-homospermidine proceeds in three steps and results in a cyclization.
References: [4854]

[EC 1.4.3.27 created 2022]

EC 1.4.4 With a disulfide as acceptor

[1.4.4.1 Transferred entry. D-proline reductase (dithiol). Now EC 1.21.4.1, D-proline reductase (dithiol)]

[EC 1.4.4.1 created 1972, modified 1982 (EC 1.4.1.6 created 1961, incorporated 1982), deleted 2003]

EC 1.4.4.2

Accepted name: glycine dehydrogenase (aminomethyl-transferring)
Reaction: glycine + [glycine-cleavage complex H protein]-*N*⁶-lipoyl-L-lysine = [glycine-cleavage complex H protein]-*S*-aminomethyl-*N*⁶-dihydrolipoyl-L-lysine + CO₂
Other name(s): P-protein; glycine decarboxylase; glycine-cleavage complex; glycine:lipoylprotein oxidoreductase (decarboxylating and acceptor-aminomethylating); protein P1; glycine dehydrogenase (decarboxylating); glycine cleavage system P-protein; glycine-cleavage complex P-protein
Systematic name: glycine:H-protein-lipoyllysine oxidoreductase (decarboxylating, acceptor-amino-methylating)
Comments: A pyridoxal-phosphate protein. A component of the glycine cleavage system, which is composed of four components that only loosely associate: the P protein (EC 1.4.4.2), the T protein (EC 2.1.2.10, aminomethyltransferase), the L protein (EC 1.8.1.4, dihydrolipoyl dehydrogenase) and the lipoyl-bearing H protein [3043]. Previously known as glycine synthase.
References: [1665, 3293, 3043]

[EC 1.4.4.2 created 1984, modified 2003, modified 2006, modified 2013]

EC 1.4.5 With a quinone or other compound as acceptor

EC 1.4.5.1

Accepted name: D-amino acid dehydrogenase (quinone)
Reaction: a D-amino acid + H₂O + a quinone = a 2-oxo carboxylate + NH₃ + a quinol
Other name(s): DadA

Systematic name: D-amino acid:quinone oxidoreductase (deaminating)
Comments: An iron-sulfur flavoprotein (FAD). The enzyme from the bacterium *Helicobacter pylori* is highly specific for D-proline, while the enzyme from the bacterium *Escherichia coli B* is most active with D-alanine, D-phenylalanine and D-methionine. This enzyme may be the same as EC 1.4.99.6.
References: [3175, 4207]

[EC 1.4.5.1 created 2010]

EC 1.4.7 With an iron-sulfur protein as acceptor

EC 1.4.7.1

Accepted name: glutamate synthase (ferredoxin)
Reaction: $2 \text{ L-glutamate} + 2 \text{ oxidized ferredoxin} = \text{L-glutamine} + 2\text{-oxoglutarate} + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
(overall reaction)
(1a) $\text{L-glutamate} + \text{NH}_3 = \text{L-glutamine} + \text{H}_2\text{O}$
(1b) $\text{L-glutamate} + 2 \text{ oxidized ferredoxin} + \text{H}_2\text{O} = \text{NH}_3 + 2\text{-oxoglutarate} + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
Other name(s): ferredoxin-dependent glutamate synthase; ferredoxin-glutamate synthase; glutamate synthase (ferredoxin-dependent)
Systematic name: L-glutamate:ferredoxin oxidoreductase (transaminating)
Comments: Binds a [3Fe-4S] cluster as well as FAD and FMN. The protein is composed of two domains, one hydrolysing L-glutamine to NH₃ and L-glutamate (*cf.* EC 3.5.1.2, glutaminase), the other combining the produced NH₃ with 2-oxoglutarate to produce a second molecule of L-glutamate. The NH₃ is channeled through a 24 Å channel in the active protein. No hydrolysis of glutamine takes place without ferredoxin and 2-oxoglutarate being bound to the protein [4399, 4400].
References: [1264, 2373, 3460, 3026, 4399, 4400]

[EC 1.4.7.1 created 1976, modified 2012]

EC 1.4.9 With a copper protein as acceptor

EC 1.4.9.1

Accepted name: methylamine dehydrogenase (amicyanin)
Reaction: $\text{methylamine} + \text{H}_2\text{O} + 2 \text{ amicyanin} = \text{formaldehyde} + \text{NH}_3 + 2 \text{ reduced amicyanin}$
Other name(s): amine dehydrogenase; primary-amine dehydrogenase; amine: (acceptor) oxidoreductase (deaminating); primary-amine:(acceptor) oxidoreductase (deaminating)
Systematic name: methylamine:amicyanin oxidoreductase (deaminating)
Comments: Contains tryptophan tryptophylquinone (TTQ) cofactor. The enzyme oxidizes aliphatic monoamines and diamines, histamine and ethanolamine, but not secondary and tertiary amines, quaternary ammonium salts or aromatic amines.
References: [267, 1000, 1002, 577, 2774]

[EC 1.4.9.1 created 1978 as EC 1.4.99.3, modified 1986, transferred 2011 to EC 1.4.98.1, transferred 2011 to EC 1.4.9.1]

EC 1.4.9.2

Accepted name: aralkylamine dehydrogenase (azurin)
Reaction: $\text{ArCH}_2\text{NH}_2 + \text{H}_2\text{O} + 2 \text{ azurin} = \text{ArCHO} + \text{NH}_3 + 2 \text{ reduced azurin}$
Other name(s): aromatic amine dehydrogenase; arylamine dehydrogenase; tyramine dehydrogenase; aralkylamine:(acceptor) oxidoreductase (deaminating)
Systematic name: aralkylamine:azurin oxidoreductase (deaminating)
Comments: Phenazine methosulfate can act as acceptor. Acts on aromatic amines and, more slowly, on some long-chain aliphatic amines, but not on methylamine or ethylamine

References: [1860, 1781, 1782, 839, 4110]

[EC 1.4.9.2 created 1986 as EC 1.4.99.4, transferred 2011 to EC 1.4.9.2]

EC 1.4.98 With a copper protein as acceptor

[1.4.98.1 *Transferred entry. amine dehydrogenase. Now EC 1.4.9.1, methylamine dehydrogenase (amicyanin)]*

[EC 1.4.98.1 created 1978 as EC 1.4.99.3, modified 1986, transferred 2011 to EC 1.4.98.1, deleted 2011]

EC 1.4.99 With unknown physiological acceptors

[1.4.99.1 *Transferred entry. D-amino-acid dehydrogenase. Now listed as EC 1.4.99.6, D-arginine dehydrogenase]*

[EC 1.4.99.1 created 1972, deleted 2015]

EC 1.4.99.2

Accepted name: taurine dehydrogenase
Reaction: taurine + H₂O + acceptor = 2-sulfoacetaldehyde + NH₃ + reduced acceptor
Other name(s): taurine:(acceptor) oxidoreductase (deaminating)
Systematic name: taurine:acceptor oxidoreductase (deaminating)
References: [2218]

[EC 1.4.99.2 created 1976]

[1.4.99.3 *Transferred entry. amine dehydrogenase. Now EC 1.4.9.1, methylamine dehydrogenase (amicyanin)]*

[EC 1.4.99.3 created 1978, modified 1986, deleted 2011]

[1.4.99.4 *Transferred entry. aralkylamine dehydrogenase. Now EC 1.4.9.2, aralkylamine dehydrogenase (azurin)]*

[EC 1.4.99.4 created 1986, deleted 2011]

EC 1.4.99.5

Accepted name: glycine dehydrogenase (cyanide-forming)
Reaction: glycine + 2 acceptor = hydrogen cyanide + CO₂ + 2 reduced acceptor
Other name(s): hydrogen cyanide synthase; HCN synthase
Systematic name: glycine:acceptor oxidoreductase (hydrogen-cyanide-forming)
Comments: The enzyme from *Pseudomonas* sp. contains FAD. The enzyme is membrane-bound, and the 2-electron acceptor is a component of the respiratory chain. The enzyme can act with various artificial electron acceptors, including phenazine methosulfate.
References: [4644, 574, 2367, 363]

[EC 1.4.99.5 created 2002]

EC 1.4.99.6

Accepted name: D-arginine dehydrogenase
Reaction: D-arginine + acceptor + H₂O = 5-guanidino-2-oxopentanoate + NH₃ + reduced acceptor (overall reaction)
(1a) D-arginine + acceptor = iminoarginine + reduced acceptor
(1b) iminoarginine + H₂O = 5-guanidino-2-oxopentanoate + NH₃ (spontaneous)
Other name(s): D-amino-acid:(acceptor) oxidoreductase (deaminating); D-amino-acid dehydrogenase; D-amino-acid:acceptor oxidoreductase (deaminating)
Systematic name: D-arginine:acceptor oxidoreductase (deaminating)

- Comments:** Contains a non-covalent FAD cofactor. The enzyme, which has been isolated from the bacterium *Pseudomonas aeruginosa* PAO1, forms with EC 1.4.1.25, L-arginine dehydrogenase, a two-enzyme complex involved in the racemization of D- and L-arginine. The enzyme has a broad substrate range and can act on most D-amino acids with the exception of D-glutamate and D-aspartate. However, activity is maximal with D-arginine and D-lysine. Not active on glycine.
- References:** [4343, 2440, 1199, 4841, 1200, 4842]

[EC 1.4.99.6 created 1972 as EC 1.4.99.1, transferred 2015 to EC 1.4.99.6, modified 2017]

EC 1.5 Acting on the CH-NH group of donors

This subclass contains enzymes that dehydrogenate secondary amines, introducing a C=N double bond as the primary reaction. In some cases, this is later hydrolysed. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.5.1), oxygen (EC 1.5.3), a disulfide (EC 1.5.4), a quinone or similar compound (EC 1.5.5), an iron-sulfur protein (EC 1.5.7), a flavin (EC 1.5.8), or some other acceptor (EC 1.5.99).

EC 1.5.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.5.1.1

- Accepted name:** 1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase [NAD(P)H]
- Reaction:** (1) L-pipecolate + NAD(P)⁺ = 1-piperideine-2-carboxylate + NAD(P)H + H⁺
(2) L-proline + NAD(P)⁺ = 1-pyrroline-2-carboxylate + NAD(P)H + H⁺
- Other name(s):** Δ¹-pyrroline-2-carboxylate reductase; DELTA1-pyrroline-2-carboxylate reductase; DELTA1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase (ambiguous); AbLhpI; pyrroline-2-carboxylate reductase; L-proline:NAD(P)⁺ 2-oxidoreductase
- Systematic name:** L-pipecolate/L-proline:NAD(P)⁺ 2-oxidoreductase
- Comments:** The enzymes, characterized from the bacterium *Azospirillum brasilense*, is involved in *trans*-3-hydroxy-L-proline metabolism. In contrast to EC 1.5.1.21, 1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase (NADPH), which is specific for NADPH, this enzyme shows similar activity with NADPH and NADH.
- References:** [2765, 4560]

[EC 1.5.1.1 created 1961, modified 2015]

EC 1.5.1.2

- Accepted name:** pyrroline-5-carboxylate reductase
- Reaction:** L-proline + NAD(P)⁺ = 1-pyrroline-5-carboxylate + NAD(P)H + H⁺
- Other name(s):** proline oxidase; L-proline oxidase; 1-pyrroline-5-carboxylate reductase; NADPH-L-Δ¹-pyrroline carboxylic acid reductase; L-proline-NAD(P)⁺ 5-oxidoreductase
- Systematic name:** L-proline:NAD(P)⁺ 5-oxidoreductase
- Comments:** Also reduces 1-pyrroline-3-hydroxy-5-carboxylate to L-hydroxyproline.
- References:** [23, 2765, 3940, 4847]

[EC 1.5.1.2 created 1961]

EC 1.5.1.3

- Accepted name:** dihydrofolate reductase
- Reaction:** 5,6,7,8-tetrahydrofolate + NADP⁺ = 7,8-dihydrofolate + NADPH + H⁺

Other name(s): tetrahydrofolate dehydrogenase; DHFR; pteridine reductase: dihydrofolate reductase; dihydrofolate reductase: thymidylate synthase; thymidylate synthetase-dihydrofolate reductase; folic acid reductase; folic reductase; dihydrofolic acid reductase; dihydrofolic reductase; 7,8-dihydrofolate reductase; NADPH-dihydrofolate reductase

Systematic name: 5,6,7,8-tetrahydrofolate:NADP⁺ oxidoreductase

Comments: The enzyme from animals and some micro-organisms also slowly reduces folate to 5,6,7,8-tetrahydrofolate.

References: [351, 377, 2033, 4828]

[EC 1.5.1.3 created 1961, modified 1976 (EC 1.5.1.4 created 1961, incorporated 1976)]

[1.5.1.4 Deleted entry. dihydrofolate dehydrogenase. Now included with EC 1.5.1.3 dihydrofolate reductase]

[EC 1.5.1.4 created 1961, deleted 1976]

EC 1.5.1.5

Accepted name: methylenetetrahydrofolate dehydrogenase (NADP⁺)

Reaction: 5,10-methylenetetrahydrofolate + NADP⁺ = 5,10-methenyltetrahydrofolate + NADPH + H⁺

Other name(s): N⁵,N¹⁰-methylenetetrahydrofolate dehydrogenase; 5,10-methylenetetrahydrofolate:NADP oxidoreductase; 5,10-methylenetetrahydrofolate dehydrogenase; methylenetetrahydrofolate dehydrogenase; methylenetetrahydrofolate dehydrogenase (NADP)

Systematic name: 5,10-methylenetetrahydrofolate:NADP⁺ oxidoreductase

Comments: In eukaryotes, occurs as a trifunctional enzyme also having methenyltetrahydrofolate cyclohydrolase (EC 3.5.4.9) and formate—tetrahydrofolate ligase (EC 6.3.4.3) activity. In some prokaryotes occurs as a bifunctional enzyme also having methenyltetrahydrofolate cyclohydrolase activity (EC 3.5.4.9).

References: [1559, 3195, 3446, 4788]

[EC 1.5.1.5 created 1961]

EC 1.5.1.6

Accepted name: formyltetrahydrofolate dehydrogenase

Reaction: 10-formyltetrahydrofolate + NADP⁺ + H₂O = tetrahydrofolate + CO₂ + NADPH + H⁺

Other name(s): 10-formyl tetrahydrofolate:NADP oxidoreductase; 10-formyl-H₂PtGlu:NADP oxidoreductase; 10-formyl-H₄folate dehydrogenase; N¹⁰-formyltetrahydrofolate dehydrogenase; 10-formyltetrahydrofolate dehydrogenase

Systematic name: 10-formyltetrahydrofolate:NADP⁺ oxidoreductase

References: [2313]

[EC 1.5.1.6 created 1972]

EC 1.5.1.7

Accepted name: saccharopine dehydrogenase (NAD⁺, L-lysine-forming)

Reaction: N⁶-(L-1,3-dicarboxypropyl)-L-lysine + NAD⁺ + H₂O = L-lysine + 2-oxoglutarate + NADH + H⁺

Other name(s): lysine-2-oxoglutarate reductase; dehydrogenase, saccharopine (nicotinamide adenine dinucleotide, lysine forming); ε-N-(L-glutaryl-2)-L-lysine:NAD oxidoreductase (L-lysine forming); N⁶-(glutar-2-yl)-L-lysine:NAD oxidoreductase (L-lysine-forming); 6-N-(L-1,3-dicarboxypropyl)-L-lysine:NAD⁺ oxidoreductase (L-lysine-forming)

Systematic name: N⁶-(L-1,3-dicarboxypropyl)-L-lysine:NAD⁺ oxidoreductase (L-lysine-forming)

References: [1206, 3677]

[EC 1.5.1.7 created 1972]

EC 1.5.1.8

- Accepted name:** saccharopine dehydrogenase (NADP⁺, L-lysine-forming)
Reaction: N^6 -(L-1,3-dicarboxypropyl)-L-lysine + NADP⁺ + H₂O = L-lysine + 2-oxoglutarate + NADPH + H⁺
Other name(s): lysine-2-oxoglutarate reductase; lysine-ketoglutarate reductase; L-lysine- α -ketoglutarate reductase; lysine: α -ketoglutarate:TPNH oxidoreductase (ϵ -N-[gultaryl-2]-L-lysine forming); saccharopine (nicotinamide adenine dinucleotide phosphate, lysine-forming) dehydrogenase; 6-*N*-(L-1,3-dicarboxypropyl)-L-lysine:NADP⁺ oxidoreductase (L-lysine-forming)
Systematic name: N^6 -(L-1,3-dicarboxypropyl)-L-lysine:NADP⁺ oxidoreductase (L-lysine-forming)
References: [1778, 2654]

[EC 1.5.1.8 created 1972]

EC 1.5.1.9

- Accepted name:** saccharopine dehydrogenase (NAD⁺, L-glutamate-forming)
Reaction: N^6 -(L-1,3-dicarboxypropyl)-L-lysine + NAD⁺ + H₂O = L-glutamate + (*S*)-2-amino-6-oxohexanoate + NADH + H⁺
Other name(s): dehydrogenase, saccharopine (nicotinamide adenine dinucleotide, glutamate-forming); saccharopin dehydrogenase; NAD⁺ oxidoreductase (L-2-aminoadipic- δ -semialdehyde and glutamate forming); aminoadipic semialdehyde synthase; 6-*N*-(L-1,3-dicarboxypropyl)-L-lysine:NAD⁺ oxidoreductase (L-glutamate-forming)
Systematic name: N^6 -(L-1,3-dicarboxypropyl)-L-lysine:NAD⁺ oxidoreductase (L-glutamate-forming)
Comments: The activities of this enzyme along with EC 1.5.1.8, saccharopine dehydrogenase (NADP⁺, L-lysine-forming), occur on a single protein.
References: [1778, 2654]

[EC 1.5.1.9 created 1972, modified 2011]

EC 1.5.1.10

- Accepted name:** saccharopine dehydrogenase (NADP⁺, L-glutamate-forming)
Reaction: N^6 -(L-1,3-dicarboxypropyl)-L-lysine + NADP⁺ + H₂O = L-glutamate + (*S*)-2-amino-6-oxohexanoate + NADPH + H⁺
Other name(s): saccharopine (nicotinamide adenine dinucleotide phosphate, glutamate-forming) dehydrogenase; aminoadipic semialdehyde-glutamic reductase; aminoadipate semialdehyde-glutamate reductase; aminoadipic semialdehyde-glutamate reductase; ϵ -*N*-(L-glutaryl-2)-L-lysine:NAD⁺(P) oxidoreductase (L-2-aminoadipate-semialdehyde forming); saccharopine reductase; 6-*N*-(L-1,3-dicarboxypropyl)-L-lysine:NADP⁺ oxidoreductase (L-glutamate-forming)
Systematic name: N^6 -(L-1,3-dicarboxypropyl)-L-lysine:NADP⁺ oxidoreductase (L-glutamate-forming)
References: [1943]

[EC 1.5.1.10 created 1972, modified 2011]

EC 1.5.1.11

- Accepted name:** D-octopine dehydrogenase
Reaction: N^2 -(D-1-carboxyethyl)-L-arginine + NAD⁺ + H₂O = L-arginine + pyruvate + NADH + H⁺
Other name(s): D-octopine synthase; octopine dehydrogenase; octopine:NAD⁺ oxidoreductase; ODH; 2-*N*-(D-1-carboxyethyl)-L-arginine:NAD⁺ oxidoreductase (L-arginine-forming)
Systematic name: N^2 -(D-1-carboxyethyl)-L-arginine:NAD⁺ oxidoreductase (L-arginine-forming)
Comments: In the reverse direction, acts also on L-ornithine, L-lysine and L-histidine.
References: [2062, 4415]

[EC 1.5.1.11 created 1972]

[1.5.1.12 Transferred entry. 1-pyrroline-5-carboxylate dehydrogenase. Now EC 1.2.1.88, L-glutamate γ -semialdehyde dehydrogenase.]

[EC 1.5.1.12 created 1972, modified 2008, deleted 2013]

[1.5.1.13 *Transferred entry. nicotinate dehydrogenase. Now EC 1.17.1.5, nicotinate dehydrogenase. The enzyme was incorrectly classified as acting on a CH-NH group*]

[EC 1.5.1.13 created 1972, deleted 2004]

[1.5.1.14 *Deleted entry. 1,2-didehydropipecolate reductase. Now included with EC 1.5.1.21 Δ^1 -piperidine-2-carboxylate reductase*]

[EC 1.5.1.14 created 1976, deleted 1989]

EC 1.5.1.15

Accepted name: methylenetetrahydrofolate dehydrogenase (NAD⁺)
Reaction: 5,10-methylenetetrahydrofolate + NAD⁺ = 5,10-methenyltetrahydrofolate + NADH + H⁺
Other name(s): methylenetetrahydrofolate dehydrogenase (NAD⁺)
Systematic name: 5,10-methylenetetrahydrofolate:NAD⁺ oxidoreductase
References: [2876]

[EC 1.5.1.15 created 1978]

EC 1.5.1.16

Accepted name: D-lysopine dehydrogenase
Reaction: N²-(D-1-carboxyethyl)-L-lysine + NADP⁺ + H₂O = L-lysine + pyruvate + NADPH + H⁺
Other name(s): D-lysopine synthase; lysopine dehydrogenase; D(+)-lysopine dehydrogenase; 2-N-(D-1-carboxyethyl)-L-lysine:NADP⁺ oxidoreductase (L-lysine-forming)
Systematic name: N²-(D-1-carboxyethyl)-L-lysine:NADP⁺ oxidoreductase (L-lysine-forming)
Comments: In the reverse reaction, a number of L-amino acids can act instead of L-lysine, and 2-oxobutanoate and, to a lesser extent, glyoxylate can act instead of pyruvate.
References: [3207]

[EC 1.5.1.16 created 1978]

EC 1.5.1.17

Accepted name: alanopine dehydrogenase
Reaction: 2,2'-iminodipropanoate + NAD⁺ + H₂O = L-alanine + pyruvate + NADH + H⁺
Other name(s): ALPDH; alanopine[*meso*-N-(1-carboxyethyl)-alanine]dehydrogenase; *meso*-N-(1-carboxyethyl)-alanine:NAD⁺ oxidoreductase; alanopine: NAD⁺ oxidoreductase; ADH (ambiguous); alanopine:NAD⁺ oxidoreductase
Systematic name: 2,2'-iminodipropanoate:NAD⁺ oxidoreductase (L-alanine-forming)
Comments: In the reverse reaction, L-alanine can be replaced by L-cysteine, L-serine or L-threonine; glycine acts very slowly (*cf.* EC 1.5.1.22 strombine dehydrogenase).
References: [818, 1118, 1119]

[EC 1.5.1.17 created 1983, modified 1986]

EC 1.5.1.18

Accepted name: ephedrine dehydrogenase
Reaction: (-)-ephedrine + NAD⁺ = (R)-2-methylimino-1-phenylpropan-1-ol + NADH + H⁺
Systematic name: (-)-ephedrine:NAD⁺ 2-oxidoreductase
Comments: The product immediately hydrolyses to methylamine and 1-hydroxy-1-phenylpropan-2-one. Acts on a number of related compounds including (-)-sympatol, (+)-pseudoephedrine and (+)-norephedrine.
References: [2148]

[EC 1.5.1.18 created 1984]

EC 1.5.1.19

- Accepted name:** D-nopaline dehydrogenase
Reaction: N^2 -(D-1,3-dicarboxypropyl)-L-arginine + NADP⁺ + H₂O = L-arginine + 2-oxoglutarate + NADPH + H⁺
Other name(s): D-nopaline synthase; nopaline dehydrogenase; nopaline synthase; NOS; 2-*N*-(D-1,3-dicarboxypropyl)-L-arginine:NADP⁺ oxidoreductase (L-arginine-forming)
Systematic name: N^2 -(D-1,3-dicarboxypropyl)-L-arginine:NADP⁺ oxidoreductase (L-arginine-forming)
Comments: In the reverse direction, forms D-nopaline from L-arginine and D-ornaline from L-ornithine.
References: [2063]

[EC 1.5.1.19 created 1984]

EC 1.5.1.20

- Accepted name:** methylenetetrahydrofolate reductase [NAD(P)H]
Reaction: 5-methyltetrahydrofolate + NAD(P)⁺ = 5,10-methylenetetrahydrofolate + NAD(P)H + H⁺
Other name(s): MTHFR (gene name)
Systematic name: 5-methyltetrahydrofolate:NAD(P)⁺ oxidoreductase
Comments: A flavoprotein (FAD). The enzyme catalyses the reversible conversion of 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate, playing an important role in folate metabolism by regulating the distribution of one-carbon moieties between cellular methylation reactions and nucleic acid synthesis. This enzyme, characterized from Protozoan parasites of the genus *Leishmania*, is unique among similar characterized eukaryotic enzymes in that it lacks the C-terminal allosteric regulatory domain (allowing it to catalyse a reversible reaction) and uses NADH and NADPH with equal efficiency under physiological conditions. *cf.* EC 1.5.1.53, methylenetetrahydrofolate reductase (NADPH); EC 1.5.1.54, methylenetetrahydrofolate reductase (NADH); and EC 1.5.7.1, methylenetetrahydrofolate reductase (ferredoxin).
References: [4445]

[EC 1.5.1.20 created 1978 as EC 1.1.1.171, transferred 1984 to EC 1.5.1.20 (EC 1.7.99.5 incorporated 2005), modified 2005., modified 2021]

EC 1.5.1.21

- Accepted name:** 1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase (NADPH)
Reaction: (1) L-pipecolate + NADP⁺ = 1-piperideine-2-carboxylate + NADPH + H⁺
(2) L-proline + NADP⁺ = 1-pyrroline-2-carboxylate + NADPH + H⁺
Other name(s): Pyr2C reductase; 1,2-didehydropipecolate reductase; P₂C reductase; 1,2-didehydropipecolic reductase; DELTA1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase (ambiguous); L-pipecolate:NADP⁺ 2-oxidoreductase; DELTA1-piperideine-2-carboxylate reductase; Δ¹-piperideine-2-carboxylate reductase
Systematic name: L-pipecolate/L-proline:NADP⁺ 2-oxidoreductase
Comments: The enzyme is involved in the catabolism of D-lysine and D-proline in bacteria that belong to the *Pseudomonas* genus. In contrast to EC 1.5.1.1, 1-piperideine-2-carboxylate/1-pyrroline-2-carboxylate reductase [NAD(P)H], which shows similar activity with NADPH and NADH, this enzyme is specific for NADPH.
References: [3269, 2941, 4560]

[EC 1.5.1.21 created 1984 (EC 1.5.1.14 created 1976, incorporated 1989), modified 2015]

EC 1.5.1.22

- Accepted name:** strombine dehydrogenase
Reaction: *N*-(carboxymethyl)-D-alanine + NAD⁺ + H₂O = glycine + pyruvate + NADH + H⁺
Other name(s): strombine[*N*-(carboxymethyl)-D-alanine]dehydrogenase; *N*-(carboxymethyl)-D-alanine: NAD⁺ oxidoreductase
Systematic name: *N*-(carboxymethyl)-D-alanine:NAD⁺ oxidoreductase (glycine-forming)

Comments: Also catalyses the reaction of EC 1.5.1.17 alanopine dehydrogenase, but more slowly. Does not act on L-strombine.

References: [818]

[EC 1.5.1.22 created 1986]

EC 1.5.1.23

Accepted name: tauropine dehydrogenase

Reaction: tauropine + NAD⁺ + H₂O = taurine + pyruvate + NADH + H⁺

Other name(s): 2-*N*-(D-1-carboxyethyl)taurine:NAD⁺ oxidoreductase (taurine-forming)

Systematic name: *N*²-(D-1-carboxyethyl)taurine:NAD⁺ oxidoreductase (taurine-forming)

Comments: In the reverse reaction, alanine can act instead of taurine, but more slowly, and 2-oxobutanoate and 2-oxopentanoate can act instead of pyruvate.

References: [1253]

[EC 1.5.1.23 created 1989]

EC 1.5.1.24

Accepted name: *N*⁵-(carboxyethyl)ornithine synthase

Reaction: *N*⁵-(L-1-carboxyethyl)-L-ornithine + NADP⁺ + H₂O = L-ornithine + pyruvate + NADPH + H⁺

Other name(s): 5-*N*-(L-1-carboxyethyl)-L-ornithine:NADP⁺ oxidoreductase (L-ornithine-forming)

Systematic name: *N*⁵-(L-1-carboxyethyl)-L-ornithine:NADP⁺ oxidoreductase (L-ornithine-forming)

Comments: In the reverse direction, L-lysine can act instead of L-ornithine, but more slowly. Acts on the amino group. *cf.* EC 1.5.1.16, D-lysopine dehydrogenase.

References: [4272]

[EC 1.5.1.24 created 1990]

EC 1.5.1.25

Accepted name: thiomorpholine-carboxylate dehydrogenase

Reaction: thiomorpholine 3-carboxylate + NAD(P)⁺ = 3,4-dehydro-thiomorpholine-3-carboxylate + NAD(P)H + H⁺

Other name(s): ketimine reductase; ketimine-reducing enzyme

Systematic name: thiomorpholine-3-carboxylate:NAD(P)⁺ 5,6-oxidoreductase

Comments: The product is the cyclic imine of the 2-oxoacid corresponding to *S*-(2-aminoethyl)cysteine. In the reverse direction, a number of other cyclic unsaturated compounds can act as substrates, but more slowly.

References: [3011]

[EC 1.5.1.25 created 1990]

EC 1.5.1.26

Accepted name: β-alanopine dehydrogenase

Reaction: β-alanopine + NAD⁺ + H₂O = β-alanine + pyruvate + NADH + H⁺

Systematic name: *N*-(D-1-carboxyethyl)-β-alanine:NAD⁺ oxidoreductase (β-alanine-forming)

References: [3669]

[EC 1.5.1.26 created 1990]

EC 1.5.1.27

Accepted name: 1,2-dehydroreticulium reductase (NADPH)

Reaction: (*R*)-reticuline + NADP⁺ = 1,2-dehydroreticulium + NADPH + H⁺

Other name(s): 1,2-dehydroreticulinium ion reductase
Systematic name: (*R*)-reticuline:NADP⁺ oxidoreductase
Comments: Reduces the 1,2-dehydroreticulinium ion to (*R*)-reticuline, which is a direct precursor of morphinan alkaloids in the poppy plant. The enzyme does not catalyse the reverse reaction to any significant extent under physiological conditions.
References: [845]

[EC 1.5.1.27 created 1999, modified 2004]

EC 1.5.1.28

Accepted name: opine dehydrogenase
Reaction: (2*S*)-2-[1-(*R*)-carboxyethyl]aminopentanoate + NAD⁺ + H₂O = L-2-aminopentanoic acid + pyruvate + NADH + H⁺
Other name(s): (2*S*)-2-[1-(*R*)-carboxyethyl]aminopentanoate dehydrogenase (NAD⁺, L-aminopentanoate-forming)
Systematic name: (2*S*)-2-[1-(*R*)-carboxyethyl]aminopentanoate:NAD⁺ oxidoreductase (L-aminopentanoate-forming)
Comments: In the forward direction, the enzyme from *Arthrobacter* sp. acts also on secondary amine dicarboxylates such as *N*-(1-carboxyethyl)methionine and *N*-(1-carboxyethyl)phenylalanine. Dehydrogenation forms an imine, which dissociates to the amino acid and pyruvate. In the reverse direction, the enzyme acts also on neutral amino acids as an amino donor. They include L-amino acids such as 2-aminopentanoic acid, 2-aminobutyric acid, 2-aminohexanoic acid, 3-chloroalanine, *O*-acetylserine, methionine, isoleucine, valine, phenylalanine, leucine and alanine. The amino acceptors include 2-oxoacids such as pyruvate, oxaloacetate, glyoxylate and 2-oxobutyrate.
References: [143, 812, 2023]

[EC 1.5.1.28 created 1999]

[1.5.1.29 Deleted entry. FMN reductase [NAD(P)H]. Now covered by EC 1.5.1.38 [FMN reductase (NADPH)], EC 1.5.1.39 [FMN reductase [NAD(P)H]] and EC 1.5.1.41 (riboflavin reductase [NAD(P)H])]

[EC 1.5.1.29 created 1981 as EC 1.6.8.1, transferred 2002 to EC 1.5.1.29, modified 2002, deleted 2011]

EC 1.5.1.30

Accepted name: flavin reductase (NADPH)
Reaction: reduced riboflavin + NADP⁺ = riboflavin + NADPH + H⁺
Other name(s): NADPH:flavin oxidoreductase; riboflavin mononucleotide (reduced nicotinamide adenine dinucleotide phosphate) reductase; flavin mononucleotide reductase; flavine mononucleotide reductase; FMN reductase (NADPH); NADPH-dependent FMN reductase; NADPH-flavin reductase; NADPH-FMN reductase; NADPH-specific FMN reductase; riboflavin mononucleotide reductase; riboflavine mononucleotide reductase; NADPH₂ dehydrogenase (flavin); NADPH₂:riboflavin oxidoreductase
Systematic name: reduced-riboflavin:NADP⁺ oxidoreductase
Comments: The enzyme reduces riboflavin, and, less efficiently, FMN and FAD. NADH is oxidized less efficiently than NADPH.
References: [4843, 787]

[EC 1.5.1.30 created 1982 as EC 1.6.8.2, transferred 2002 to EC 1.5.1.30, modified 2011]

EC 1.5.1.31

Accepted name: berberine reductase
Reaction: (*R*)-canadine + 2 NADP⁺ = berberine + 2 NADPH + H⁺
Other name(s): (*R*)-canadine synthase
Systematic name: (*R*)-tetrahydroberberine:NADP⁺ oxidoreductase
Comments: Involved in alkaloid biosynthesis in *Corydalis cava* to give (*R*)-canadine with the opposite configuration to the precursor of berberine (see EC 1.3.3.8 tetrahydroberberine oxidase). Also acts on 7,8-dihydroberberine.

References: [244]

[EC 1.5.1.31 created 2002]

EC 1.5.1.32

Accepted name: vomilenine reductase
Reaction: 1,2-dihydrovomilenine + NADP⁺ = vomilenine + NADPH + H⁺
Systematic name: 1,2-dihydrovomilenine:NADP⁺ oxidoreductase
Comments: Forms part of the ajmaline biosynthesis pathway.
References: [4469]

[EC 1.5.1.32 created 2002]

EC 1.5.1.33

Accepted name: pteridine reductase
Reaction: 5,6,7,8-tetrahydrobiopterin + 2 NADP⁺ = biopterin + 2 NADPH + 2 H⁺
Other name(s): PTR1; pteridine reductase 1
Systematic name: 5,6,7,8-tetrahydrobiopterin:NADP⁺ oxidoreductase
Comments: The enzyme from *Leishmania* (both amastigote and promastigote forms) catalyses the reduction by NADPH of folate and a wide variety of unconjugated pterins, including biopterin, to their tetrahydro forms. It also catalyses the reduction of 7,8-dihydropterins and 7,8-dihydrofolate to their tetrahydro forms. In contrast to EC 1.5.1.3 (dihydrofolate reductase) and EC 1.5.1.34 (6,7-dihydropteridine reductase), pteridine reductase will not catalyse the reduction of the quinonoid form of dihydrobiopterin. The enzyme is specific for NADPH; no activity has been detected with NADH. It also differs from EC 1.5.1.3 (dihydrofolate reductase) in being specific for the *Si*-face of NADPH.
References: [3012, 1378, 1129]

[EC 1.5.1.33 created 1999 as EC 1.1.1.253, transferred 2003 to EC 1.5.1.33]

EC 1.5.1.34

Accepted name: 6,7-dihydropteridine reductase
Reaction: a 5,6,7,8-tetrahydropteridine + NAD(P)⁺ = a 6,7-dihydropteridine + NAD(P)H + H⁺
Other name(s): 6,7-dihydropteridine:NAD(P)H oxidoreductase; DHPR; NAD(P)H:6,7-dihydropteridine oxidoreductase; NADH-dihydropteridine reductase; NADPH-dihydropteridine reductase; NADPH-specific dihydropteridine reductase; dihydropteridine (reduced nicotinamide adenine dinucleotide) reductase; dihydropteridine reductase; dihydropteridine reductase (NADH); 5,6,7,8-tetrahydropteridine:NAD(P)H⁺ oxidoreductase
Systematic name: 5,6,7,8-tetrahydropteridine:NAD(P)⁺ oxidoreductase
Comments: The substrate is the quinonoid form of dihydropteridine. Not identical with EC 1.5.1.3 dihydrofolate reductase.
References: [1528, 1547, 2036, 2495, 2988]

[EC 1.5.1.34 created 1972 as EC 1.6.99.7, modified 1981 (EC 1.6.99.10 created 1978, incorporated 1981), transferred 2003 to EC 1.5.1.34]

[1.5.1.35 Deleted entry. 1-pyrroline dehydrogenase. The enzyme is identical to EC 1.2.1.19, aminobutyraldehyde dehydrogenase, as the substrates 1-pyrroline and 4-aminobutanal are interconvertible]

[EC 1.5.1.35 created 2006, deleted 2007]

EC 1.5.1.36

Accepted name: flavin reductase (NADH)
Reaction: reduced flavin + NAD⁺ = flavin + NADH + H⁺
Other name(s): NADH-dependent flavin reductase; flavin:NADH oxidoreductase

Systematic name: flavin:NAD⁺ oxidoreductase
Comments: The enzyme from *Escherichia coli* W catalyses the reduction of free flavins by NADH. The enzyme has similar affinity to FAD, FMN and riboflavin. Activity with NADPH is more than 2 orders of magnitude lower than activity with NADH.
References: [1256]

[EC 1.5.1.36 created 2011]

EC 1.5.1.37

Accepted name: FAD reductase (NADH)
Reaction: $\text{FADH}_2 + \text{NAD}^+ = \text{FAD} + \text{NADH} + \text{H}^+$
Other name(s): NADH-FAD reductase; NADH-dependent FAD reductase; NADH:FAD oxidoreductase; NADH:flavin adenine dinucleotide oxidoreductase
Systematic name: FADH₂:NAD⁺ oxidoreductase
Comments: The enzyme from *Burkholderia phenoliruptrix* can reduce either FAD or flavin mononucleotide (FMN) but prefers FAD. Unlike EC 1.5.1.36, flavin reductase (NADH), the enzyme can not reduce riboflavin. The enzyme does not use NADPH as acceptor.
References: [1330]

[EC 1.5.1.37 created 2011]

EC 1.5.1.38

Accepted name: FMN reductase (NADPH)
Reaction: $\text{FMNH}_2 + \text{NADP}^+ = \text{FMN} + \text{NADPH} + \text{H}^+$
Other name(s): FRP; flavin reductase P; SsuE
Systematic name: FMNH₂:NADP⁺ oxidoreductase
Comments: The enzymes from bioluminescent bacteria contain FMN [2409], while the enzyme from *Escherichia coli* does not [1028]. The enzyme often forms a two-component system with monooxygenases such as luciferase. Unlike EC 1.5.1.39, this enzyme does not use NADH as acceptor [1305, 1869]. While FMN is the preferred substrate, the enzyme can also use FAD and riboflavin with lower activity [3,6,8].
References: [1305, 1869, 1870, 2409, 4213, 2520, 2410, 1028]

[EC 1.5.1.38 created 2011]

EC 1.5.1.39

Accepted name: FMN reductase [NAD(P)H]
Reaction: $\text{FMNH}_2 + \text{NAD(P)}^+ = \text{FMN} + \text{NAD(P)H} + \text{H}^+$
Other name(s): FRG
Systematic name: FMNH₂:NAD(P)⁺ oxidoreductase
Comments: Contains FMN. The enzyme can utilize NADH and NADPH with similar reaction rates. Different from EC 1.5.1.42, FMN reductase (NADH) and EC 1.5.1.38, FMN reductase (NADPH). The luminescent bacterium *Vibrio harveyi* possesses all three enzymes [4552]. Also reduces riboflavin and FAD, but more slowly.
References: [4552]

[EC 1.5.1.39 created 2011]

EC 1.5.1.40

Accepted name: 8-hydroxy-5-deazaflavin:NADPH oxidoreductase
Reaction: reduced coenzyme F₄₂₀ + NADP⁺ = oxidized coenzyme F₄₂₀ + NADPH + H⁺
Other name(s): 8-OH-5dFl:NADPH oxidoreductase
Systematic name: reduced coenzyme F₄₂₀:NADP⁺ oxidoreductase

Comments: The enzyme has an absolute requirement for both the 5-deazaflavin structure and the presence of an 8-hydroxy group in the substrate [1033].

References: [1033]

[EC 1.5.1.40 created 2011]

EC 1.5.1.41

Accepted name: riboflavin reductase [NAD(P)H]

Reaction: reduced riboflavin + NAD(P)⁺ = riboflavin + NAD(P)H + H⁺

Other name(s): NAD(P)H-FMN reductase (ambiguous); NAD(P)H-dependent FMN reductase (ambiguous); NAD(P)H:FMN oxidoreductase (ambiguous); NAD(P)H:flavin oxidoreductase (ambiguous); NAD(P)H₂ dehydrogenase (FMN) (ambiguous); NAD(P)H₂:FMN oxidoreductase (ambiguous); riboflavin mononucleotide reductase (ambiguous); flavine mononucleotide reductase (ambiguous); riboflavin mononucleotide (reduced nicotinamide adenine dinucleotide (phosphate)) reductase; flavin mononucleotide reductase (ambiguous); riboflavine mononucleotide reductase (ambiguous); Fre

Systematic name: riboflavin:NAD(P)⁺ oxidoreductase

Comments: Catalyses the reduction of soluble flavins by reduced pyridine nucleotides. Highest activity with riboflavin. When NADH is used as acceptor, the enzyme can also utilize FMN and FAD as substrates, with lower activity than riboflavin. When NADPH is used as acceptor, the enzyme has a very low activity with FMN and no activity with FAD [1136].

References: [1136, 3991, 1810]

[EC 1.5.1.41 created 2011]

EC 1.5.1.42

Accepted name: FMN reductase (NADH)

Reaction: FMNH₂ + NAD⁺ = FMN + NADH + H⁺

Other name(s): NADH-FMN reductase; NADH-dependent FMN reductase; NADH:FMN oxidoreductase; NADH:flavin oxidoreductase

Systematic name: FMNH₂:NAD⁺ oxidoreductase

Comments: The enzyme often forms a two-component system with monooxygenases. Unlike EC 1.5.1.38, FMN reductase (NADPH), and EC 1.5.1.39, FMN reductase [NAD(P)H], this enzyme has a strong preference for NADH over NADPH, although some activity with the latter is observed [974, 1305]. While FMN is the preferred substrate, FAD can also be used with much lower activity [974, 4370].

References: [974, 1305, 4370, 1866]

[EC 1.5.1.42 created 2011]

EC 1.5.1.43

Accepted name: carboxynorspermidine synthase

Reaction: (1) carboxynorspermidine + H₂O + NADP⁺ = L-aspartate 4-semialdehyde + propane-1,3-diamine + NADPH + H⁺

(2) carboxyspermidine + H₂O + NADP⁺ = L-aspartate 4-semialdehyde + putrescine + NADPH + H⁺

Other name(s): carboxynorspermidine dehydrogenase; carboxyspermidine dehydrogenase; CASDH; CANSDH; VC1624 (gene name)

Systematic name: carboxynorspermidine:NADP⁺ oxidoreductase

Comments: The reaction takes place in the opposite direction. Part of a bacterial polyamine biosynthesis pathway. L-aspartate 4-semialdehyde and propane-1,3-diamine/putrescine form a Schiff base that is reduced to form carboxynorspermidine/carboxyspermidine, respectively [2995]. The enzyme from the bacterium *Vibrio cholerae* is essential for biofilm formation [2384]. The enzyme from *Campylobacter jejuni* only produces carboxyspermidine *in vivo* even though it also can produce carboxynorspermidine *in vitro* [1504].

References: [2995, 2384, 1504]

[EC 1.5.1.43 created 2012]

EC 1.5.1.44

Accepted name: festuclavine dehydrogenase
Reaction: festuclavine + NAD⁺ = 6,8-dimethyl-6,7-didehydroergoline + NADH + H⁺
Other name(s): FgaFS; festuclavine synthase
Systematic name: festuclavine:NAD⁺ oxidoreductase
Comments: The enzyme participates in the biosynthesis of fumigaclavine C, an ergot alkaloid produced by some fungi of the *Trichocomaceae* family. The reaction proceeds *in vivo* in the opposite direction to the one shown here.
References: [4506]

[EC 1.5.1.44 created 2012]

EC 1.5.1.45

Accepted name: FAD reductase [NAD(P)H]
Reaction: FADH₂ + NAD(P)⁺ = FAD + NAD(P)H + H⁺
Other name(s): GTNG_3158 (gene name)
Systematic name: FADH₂:NAD(P)⁺ oxidoreductase
Comments: This enzyme, isolated from the bacterium *Geobacillus thermodenitrificans*, participates in the pathway of tryptophan degradation. The enzyme is part of a system that also includes a bifunctional riboflavin kinase/FMN adenylyltransferase and EC 1.14.14.8, anthranilate 3-monooxygenase (FAD). It can reduce either FAD or flavin mononucleotide (FMN) but prefers FAD. The enzyme has a slight preference for NADPH as acceptor. *cf.* EC 1.5.1.37, FAD reductase (NADH).
References: [2525]

[EC 1.5.1.45 created 2012]

EC 1.5.1.46

Accepted name: agroclavine dehydrogenase
Reaction: agroclavine + NADP⁺ = 6,8-dimethyl-6,7,8,9-tetrahydroergoline + NADPH + H⁺
Other name(s): *easG* (gene name)
Systematic name: agroclavine:NADP⁺ oxidoreductase
Comments: The enzyme participates in the biosynthesis of ergotamine, an ergot alkaloid produced by some fungi of the Clavicipitaceae family. The reaction is catalysed in the opposite direction to that shown. The substrate for the enzyme is an iminium intermediate that is formed spontaneously from chanoclavine-I aldehyde in the presence of glutathione.
References: [2725]

[EC 1.5.1.46 created 2013]

EC 1.5.1.47

Accepted name: dihydromethanopterin reductase [NAD(P)⁺]
Reaction: 5,6,7,8-tetrahydromethanopterin + NAD(P)⁺ = 7,8-dihydromethanopterin + NAD(P)H + H⁺
Other name(s): DmrA; H₂MPT reductase; 5,6,7,8-tetrahydromethanopterin 5,6-oxidoreductase; dihydromethanopterin reductase
Systematic name: 5,6,7,8-tetrahydromethanopterin:NAD(P)⁺ 5,6-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Methylobacterium extorquens*, is involved in biosynthesis of dephospho-tetrahydromethanopterin. The specific activity with NADH is 15% of that with NADPH at the same concentration [516]. It does not reduce 7,8-dihydrofolate (*cf.* EC 1.5.1.3, dihydrofolate reductase).
References: [516]

[EC 1.5.1.47 created 2013, modified 2014]

EC 1.5.1.48

- Accepted name:** 2-methyl-1-pyrroline reductase
Reaction: (*R*)-2-methylpyrrolidine + NADP⁺ = 2-methyl-1-pyrroline + NADPH + H⁺
Other name(s): (*R*)-imine reductase (ambiguous)
Systematic name: (*R*)-2-methylpyrrolidine:NADP⁺ 2-oxidoreductase
Comments: The enzyme from the bacterium *Streptomyces* sp. GF3587 is highly specific for its substrate and forms only the (*R*) isomer.
References: [2834]

[EC 1.5.1.48 created 2014]

EC 1.5.1.49

- Accepted name:** 1-pyrroline-2-carboxylate reductase [NAD(P)H]
Reaction: L-proline + NAD(P)⁺ = 1-pyrroline-2-carboxylate + NAD(P)H + H⁺
Systematic name: L-proline:NAD(P)⁺ 2-oxidoreductase
Comments: The enzyme from the bacterium *Colwellia psychrerythraea* is involved in *trans*-3-hydroxy-L-proline metabolism. In contrast to EC 1.5.1.1, 1-piperidine-2-carboxylate/1-pyrroline-2-carboxylate reductase [NAD(P)H], which shows similar activity with 1-piperidine-2-carboxylate and 1-pyrroline-2-carboxylate, this enzyme is specific for the latter. While the enzyme is active with both NADH and NADPH, activity is higher with NADPH.
References: [4560]

[EC 1.5.1.49 created 2015]

EC 1.5.1.50

- Accepted name:** dihydromonapterin reductase
Reaction: 5,6,7,8-tetrahydromonapterin + NADP⁺ = 7,8-dihydromonapterin + NADPH + H⁺
Other name(s): FolM; H₂-MPt reductase
Systematic name: 5,6,7,8-tetrahydromonapterin:NADP⁺ oxidoreductase
Comments: The enzyme, found in many Gram negative bacteria, also slowly reduces 7,8-dihydrofolate to 5,6,7,8-tetrahydrofolate (*cf.* EC 1.5.1.3, dihydrofolate reductase). The enzyme has no activity with NADH.
References: [3378]

[EC 1.5.1.50 created 2015]

EC 1.5.1.51

- Accepted name:** *N*-[(2*S*)-2-amino-2-carboxyethyl]-L-glutamate dehydrogenase
Reaction: *N*-[(2*S*)-2-amino-2-carboxyethyl]-L-glutamate + NAD⁺ + H₂O = 2-oxoglutarate + L-2,3-diaminopropanoate + NADH + H⁺
Other name(s): SbnB
Systematic name: *N*-[(2*S*)-2-amino-2-carboxyethyl]-L-glutamate:NAD⁺ dehydrogenase (L-2,3-diaminopropanoate-forming)
Comments: The enzyme, characterized from the bacterium *Staphylococcus aureus*, is involved in the biosynthesis of the siderophore staphyloferrin B.
References: [256, 2178]

[EC 1.5.1.51 created 2017]

EC 1.5.1.52

- Accepted name:** staphylopine dehydrogenase
Reaction: staphylopine + NADP⁺ + H₂O = (2*S*)-2-amino-4-[(1*R*)-1-carboxy-2-(1*H*-imidazol-4-yl)ethyl]aminobutanoate + pyruvate + NADPH + H⁺
Other name(s): *cntM* (gene name); staphylopine synthase

Systematic name: staphylopine:NADP⁺ oxidoreductase [(2*S*)-2-amino-4-[(1*R*)-1-carboxy-2-(1*H*-imidazol-4-yl)ethyl]aminobutanoate]-forming

Comments: The enzyme, characterized from the bacterium *Staphylococcus aureus*, catalyses the last reaction in the biosynthesis of the metallophore staphylopine, which is involved in the acquisition of nickel, copper, and cobalt.

References: [1316, 2748]

[EC 1.5.1.52 created 2018]

EC 1.5.1.53

Accepted name: methylenetetrahydrofolate reductase (NADPH)

Reaction: 5-methyltetrahydrofolate + NADP⁺ = 5,10-methylenetetrahydrofolate + NADPH + H⁺

Other name(s): MTHFR (gene name); methylenetetrahydrofolate (reduced nicotinamide adenine dinucleotide phosphate) reductase; 5,10-methylenetetrahydrofolate reductase (NADPH); 5,10-methylenetetrahydrofolic acid reductase (ambiguous); 5,10-CH₂-H₄folate reductase (ambiguous); methylenetetrahydrofolate reductase (NADPH₂); 5,10-methylenetetrahydrofolate reductase (ambiguous); methylenetetrahydrofolate reductase (ambiguous); *N*⁵,10-methylenetetrahydrofolate reductase (ambiguous); 5,10-methylenetetrahydropteroylglutamate reductase (ambiguous); *N*⁵,*N*¹⁰-methylenetetrahydrofolate reductase (ambiguous); methylenetetrahydrofolic acid reductase (ambiguous); 5-methyltetrahydrofolate:(acceptor) oxidoreductase (incorrect); 5,10-methylenetetrahydrofolate reductase (FADH₂) (ambiguous)

Systematic name: 5-methyltetrahydrofolate:NADP⁺ oxidoreductase

Comments: A flavoprotein (FAD). The enzyme from yeast and mammals catalyses a physiologically irreversible reduction of 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate using NADPH as the electron donor. It plays an important role in folate metabolism by regulating the distribution of one-carbon moieties between cellular methylation reactions and nucleic acid synthesis. The enzyme contains an N-terminal catalytic domain and a C-terminal allosteric regulatory domain that binds *S*-adenosyl-L-methionine, which acts as an inhibitor. *cf.* EC 1.5.1.54, methylenetetrahydrofolate reductase (NADH); EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]; and EC 1.5.7.1, methylenetetrahydrofolate reductase (ferredoxin).

References: [946, 2314, 834, 4912, 3559, 1193]

[EC 1.5.1.53 created 2021]

EC 1.5.1.54

Accepted name: methylenetetrahydrofolate reductase (NADH)

Reaction: 5-methyltetrahydrofolate + NAD⁺ = 5,10-methylenetetrahydrofolate + NADH + H⁺

Other name(s): *metF* (gene name); 5,10-methylenetetrahydrofolic acid reductase (ambiguous); 5,10-CH₂-H₄folate reductase (ambiguous); methylenetetrahydrofolate (reduced riboflavin adenine dinucleotide) reductase; 5,10-methylenetetrahydrofolate reductase (ambiguous); methylenetetrahydrofolate reductase (ambiguous); *N*⁵,10-methylenetetrahydrofolate reductase (ambiguous); 5,10-methylenetetrahydropteroylglutamate reductase (ambiguous); *N*⁵,*N*¹⁰-methylenetetrahydrofolate reductase (ambiguous); methylenetetrahydrofolic acid reductase (ambiguous); 5-methyltetrahydrofolate:(acceptor) oxidoreductase (incorrect); 5,10-methylenetetrahydrofolate reductase (FADH₂) (ambiguous)

Systematic name: 5-methyltetrahydrofolate:NAD⁺ oxidoreductase

Comments: A flavoprotein (FAD). The enzyme, found in plants and some bacteria, catalyses the reversible conversion of 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate using NADH as the electron donor. It play an important role in folate metabolism by regulating the distribution of one-carbon moieties between cellular methylation reactions and nucleic acid synthesis. These proteins either contain a C-terminal domain that does not mediate allosteric regulation (as in plants) or lack this domain entirely (as in *Escherichia coli*). As a result, the plant enzymes are not inhibited by S-adenosyl-L-methionine, unlike other eukaryotic enzymes, and catalyse a reversible reaction. *cf.* EC 1.5.1.53, methylenetetrahydrofolate reductase (NADPH); EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]; and EC 1.5.7.1, methylenetetrahydrofolate reductase (ferredoxin).

References: [4651, 3845, 1438, 3560, 318]

[EC 1.5.1.54 created 2021]

EC 1.5.3 With oxygen as acceptor

EC 1.5.3.1

Accepted name: sarcosine oxidase (formaldehyde-forming)
Reaction: sarcosine + H₂O + O₂ = glycine + formaldehyde + H₂O₂
Other name(s): MSOX; monomeric sarcosine oxidase; sarcosine oxidase (ambiguous)
Systematic name: sarcosine:oxygen oxidoreductase (demethylating)
Comments: The enzyme, reported from bacteria and fungi, catalyses the oxidative demethylation of sarcosine. It contains a FAD cofactor bound to an L-cysteine residue. *cf.* EC 1.5.3.24, sarcosine oxidase (5,10-methylenetetrahydrofolate-forming).
References: [2888, 3083, 3084, 4328, 4488, 4898, 1952, 481]

[EC 1.5.3.1 created 1961, modified 2022]

EC 1.5.3.2

Accepted name: *N*-methyl-L-amino-acid oxidase
Reaction: an *N*-methyl-L-amino acid + H₂O + O₂ = an L-amino acid + formaldehyde + H₂O₂
Other name(s): *N*-methylamino acid oxidase; demethylase
Systematic name: *N*-methyl-L-amino-acid:oxygen oxidoreductase (demethylating)
Comments: A flavoprotein.
References: [2894, 2895, 2896]

[EC 1.5.3.2 created 1961]

[1.5.3.3 Deleted entry. spermine oxidase]

[EC 1.5.3.3 created 1961, deleted 1972]

EC 1.5.3.4

Accepted name: *N*⁶-methyl-lysine oxidase
Reaction: *N*⁶-methyl-L-lysine + H₂O + O₂ = L-lysine + formaldehyde + H₂O₂
Other name(s): ε-alkyl-L-lysine:oxygen oxidoreductase ; *N*⁶-methyllysine oxidase; ε-*N*-methyllysine demethylase; ε-alkyllysine; 6-*N*-methyl-L-lysine:oxygen oxidoreductase (demethylating)
Systematic name: *N*⁶-methyl-L-lysine:oxygen oxidoreductase (demethylating)
References: [2103]

[EC 1.5.3.4 created 1972]

EC 1.5.3.5

- Accepted name:** (S)-6-hydroxynicotine oxidase
- Reaction:** (S)-6-hydroxynicotine + H₂O + O₂ = 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one + H₂O₂ (overall reaction)
(1a) (S)-6-hydroxynicotine + O₂ = 5-(N-methyl-4,5-dihydro-1H-pyrrol-2-yl)pyridin-2-ol + H₂O₂
(1b) 5-(N-methyl-4,5-dihydro-1H-pyrrol-2-yl)pyridin-2-ol + H₂O = 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one (spontaneous)
- Other name(s):** L-6-hydroxynicotine oxidase; 6-hydroxy-L-nicotine oxidase; 6-hydroxy-L-nicotine:oxygen oxidoreductase; *nctB* (gene name)
- Systematic name:** (S)-6-hydroxynicotine:oxygen oxidoreductase
- Comments:** A flavoprotein (FAD). The enzyme, which participates in nicotine degradation, is specific for the (S) isomer of 6-hydroxynicotine. The bacterium *Arthrobacter nicotinovorans*, in which this enzyme was originally discovered, has a different enzyme that catalyses a similar reaction with the less common (R)-isomer (cf. EC 1.5.3.6, (R)-6-hydroxynicotine oxidase).
- References:** [859, 802, 3708, 3403]

[EC 1.5.3.5 created 1972, modified 2015]

EC 1.5.3.6

- Accepted name:** (R)-6-hydroxynicotine oxidase
- Reaction:** (R)-6-hydroxynicotine + H₂O + O₂ = 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one + H₂O₂ (overall reaction)
(1a) (R)-6-hydroxynicotine + O₂ = 5-(N-methyl-4,5-dihydro-1H-pyrrol-2-yl)pyridin-2-ol + H₂O₂
(1b) 5-(N-methyl-4,5-dihydro-1H-pyrrol-2-yl)pyridin-2-ol + H₂O = 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one (spontaneous)
- Other name(s):** D-6-hydroxynicotine oxidase; 6-hydroxy-D-nicotine oxidase
- Systematic name:** (R)-6-hydroxynicotine:oxygen oxidoreductase
- Comments:** A flavoprotein (FAD). The enzyme, which participates in nicotine degradation, is specific for (R) isomer of 6-hydroxynicotine, derived from the uncommon (R)-nicotine. The bacterium *Arthrobacter nicotinovorans*, in which this enzyme was originally discovered, has a different enzyme that catalyses a similar reaction with the (S)-isomer (cf. EC 1.5.3.5, (S)-6-hydroxynicotine oxidase).
- References:** [859, 471, 425, 3708, 2192]

[EC 1.5.3.6 created 1972, modified 2015]

EC 1.5.3.7

- Accepted name:** L-pipecolate oxidase
- Reaction:** L-pipecolate + O₂ = (S)-2,3,4,5-tetrahydropyridine-2-carboxylate + H₂O₂
- Other name(s):** pipecolate oxidase; L-pipecolic acid oxidase
- Systematic name:** L-pipecolate:oxygen 1,6-oxidoreductase
- Comments:** The product reacts with water to form (S)-2-amino-6-oxohexanoate.
- References:** [179, 2124]

[EC 1.5.3.7 created 1986, modified 2011]

[1.5.3.8 Deleted entry. (S)-tetrahydroprotoberberine oxidase. Now included with EC 1.3.3.8, tetrahydroberberine oxidase]

[EC 1.5.3.8 created 1989, deleted 1992]

[1.5.3.9 Transferred entry. reticuline oxidase. Now EC 1.21.3.3, reticuline oxidase]

[EC 1.5.3.9 created 1989, modified 1999, deleted 2002]

EC 1.5.3.10

- Accepted name:** dimethylglycine oxidase

Reaction: N,N -dimethylglycine + 5,6,7,8-tetrahydrofolate + O_2 = sarcosine + 5,10-methylenetetrahydrofolate + H_2O_2

Other name(s): *dmg* (gene name); N,N -dimethylglycine:oxygen oxidoreductase (demethylating)

Systematic name: N,N -dimethylglycine,5,6,7,8-tetrahydrofolate:oxygen oxidoreductase (demethylating,5,10-methylenetetrahydrofolate-forming)

Comments: A flavoprotein (FAD). The enzyme, characterized from the bacterium *Arthrobacter globiformis*, contains two active sites connected by a large "reaction chamber". An imine intermediate is transferred between the sites, eliminating the production of toxic formaldehyde. In the absence of folate the enzyme does form formaldehyde. Does not oxidize sarcosine. *cf.* EC 1.5.8.4, dimethylglycine dehydrogenase.

References: [2887, 2775, 232, 2439, 233, 4322, 568]

[EC 1.5.3.10 created 1992, modified 2022]

[1.5.3.11 Deleted entry. polyamine oxidase. Now included with EC 1.5.3.13 (N^1 -acetylpolyamine oxidase), EC 1.5.3.14 (polyamine oxidase (propane-1,3-diamine-forming)), EC 1.5.3.15 (N^8 -acetylspermidine oxidase (propane-1,3-diamine-forming)), EC 1.5.3.16 (spermine oxidase) and EC 1.5.3.17 (non-specific polyamine oxidase)]

[EC 1.5.3.11 created 1992, deleted 2009]

EC 1.5.3.12

Accepted name: dihydrobenzophenanthridine oxidase

Reaction: (1) dihydrosanguinarine + O_2 = sanguinarine + H_2O_2
(2) dihydrochelirubine + O_2 = chelirubine + H_2O_2
(3) dihydromacarpine + O_2 = macarpine + H_2O_2

Systematic name: dihydrobenzophenanthridine:oxygen oxidoreductase

Comments: A Cu^{II} enzyme found in higher plants that produces oxidized forms of the benzophenanthridine alkaloids

References: [3756, 121]

[EC 1.5.3.12 created 1999]

EC 1.5.3.13

Accepted name: N^1 -acetylpolyamine oxidase

Reaction: (1) N^1 -acetylspermidine + O_2 + H_2O = putrescine + 3-acetamidopropanal + H_2O_2
(2) N^1 -acetylspermine + O_2 + H_2O = spermidine + 3-acetamidopropanal + H_2O_2

Other name(s): hPAO-1; PAO (ambiguous); mPAO; hPAO; polyamine oxidase (ambiguous)

Systematic name: N^1 -acetylpolyamine:oxygen oxidoreductase (3-acetamidopropanal-forming)

Comments: The enzyme also catalyses the reaction: N^1,N^{12} -diacetylspermine + O_2 + H_2O = N^1 -acetylspermidine + 3-acetamidopropanal + H_2O_2 [4480]. No or very weak activity with spermine, or spermidine in absence of aldehydes. In presence of aldehydes the enzyme catalyses the reactions: 1. spermine + O_2 + H_2O = spermidine + 3-aminopropanal + H_2O_2 , and with weak efficiency 2. spermidine + O_2 + H_2O = putrescine + 3-aminopropanal + H_2O_2 [1893]. A flavoprotein (FAD). This enzyme, encoded by the PAOX gene, is found in mammalian peroxisomes and oxidizes N^1 -acetylated polyamines at the exo (three-carbon) side of the secondary amine, forming 3-acetamidopropanal. Since the products of the reactions are deacetylated polyamines, this process is known as polyamine back-conversion. Differs in specificity from EC 1.5.3.14 [polyamine oxidase (propane-1,3-diamine-forming)], EC 1.5.3.15 [N^8 -acetylspermidine oxidase (propane-1,3-diamine-forming)], EC 1.5.3.16 (spermine oxidase) and EC 1.5.3.17 (non-specific polyamine oxidase).

References: [4480, 1893, 4537, 4679]

[EC 1.5.3.13 created 2009]

EC 1.5.3.14

Accepted name: polyamine oxidase (propane-1,3-diamine-forming)
Reaction: spermidine + O₂ + H₂O = propane-1,3-diamine + 4-aminobutanal + H₂O₂
Other name(s): MPAO (ambiguous); maize PAO
Systematic name: spermidine:oxygen oxidoreductase (propane-1,3-diamine-forming)
Comments: As the products of the reaction cannot *be* converted directly to other polyamines, this class of polyamine oxidases is considered to be involved in the terminal catabolism of polyamines [4225]. This enzyme less efficiently catalyses the oxidation of *N*¹-acetylspermine and spermine. A flavoprotein (FAD). Differs in specificity from EC 1.5.3.13 (*N*¹-acetylpolyamine oxidase), EC 1.5.3.15 [*N*⁸-acetylspermidine oxidase (propane-1,3-diamine-forming)], EC 1.5.3.16 (spermine oxidase) and EC 1.5.3.17 (non-specific polyamine oxidase).
References: [4225, 1096]

[EC 1.5.3.14 created 2009]

EC 1.5.3.15

Accepted name: *N*⁸-acetylspermidine oxidase (propane-1,3-diamine-forming)
Reaction: *N*⁸-acetylspermidine + O₂ + H₂O = propane-1,3-diamine + 4-acetamidobutanal + H₂O₂
Systematic name: *N*⁸-acetylspermidine:oxygen oxidoreductase (propane-1,3-diamine-forming)
Comments: Also active with *N*¹-acetylspermine, weak activity with *N*¹,*N*¹²-diacetylspermine. No activity with diaminopropane, putrescine, cadaverine, diaminohexane, norspermidine, spermine and spermidine. Absence of monoamine oxidase (EC 1.4.3.4) activity. Differs in specificity from EC 1.5.3.13 (*N*¹-acetylpolyamine oxidase), EC 1.5.3.14 (polyamine oxidase (propane-1,3-diamine-forming)), EC 1.5.3.16 (spermine oxidase) and EC 1.5.3.17 (non-specific polyamine oxidase).
References: [3896]

[EC 1.5.3.15 created 2009]

EC 1.5.3.16

Accepted name: spermine oxidase
Reaction: spermine + O₂ + H₂O = spermidine + 3-aminopropanal + H₂O₂
Other name(s): PAOh1/SMO; PAOh1 (ambiguous); AtPAO1; AtPAO4; SMO; mSMO; SMO(PAOh1); SMO/PAOh1; SMO5; mSMOmu
Systematic name: spermidine:oxygen oxidoreductase (spermidine-forming)
Comments: The enzyme from *Arabidopsis thaliana* (AtPAO1) oxidizes norspermine to norspermidine with high efficiency [4224]. The mammalian enzyme, encoded by the SMOX gene, is a cytosolic enzyme that catalyses the oxidation of spermine at the exo (three-carbon) side of the tertiary amine. No activity with spermidine. Weak activity with *N*¹-acetylspermine. A flavoprotein (FAD). Differs in specificity from EC 1.5.3.13 (*N*¹-acetylpolyamine oxidase), EC 1.5.3.14 (polyamine oxidase (propane-1,3-diamine-forming)), EC 1.5.3.15 (*N*⁸-acetylspermidine oxidase (propane-1,3-diamine-forming)) and EC 1.5.3.17 (non-specific polyamine oxidase).
References: [2947, 586, 4224, 4538]

[EC 1.5.3.16 created 2009]

EC 1.5.3.17

Accepted name: non-specific polyamine oxidase
Reaction:
(1) spermine + O₂ + H₂O = spermidine + 3-aminopropanal + H₂O₂
(2) spermidine + O₂ + H₂O = putrescine + 3-aminopropanal + H₂O₂
(3) *N*¹-acetylspermine + O₂ + H₂O = spermidine + 3-acetamidopropanal + H₂O₂
(4) *N*¹-acetylspermidine + O₂ + H₂O = putrescine + 3-acetamidopropanal + H₂O₂
Other name(s): polyamine oxidase (ambiguous); Fms1; AtPAO3
Systematic name: polyamine:oxygen oxidoreductase (3-aminopropanal or 3-acetamidopropanal-forming)

Comments: A flavoprotein (FAD). The non-specific polyamine oxidases may differ from each other considerably. The enzyme from *Saccharomyces cerevisiae* shows a rather broad specificity and also oxidizes N^8 -acetylspermidine [2339]. The enzyme from *Ascaris suum* shows high activity with spermine and spermidine, but also oxidizes norspermine [2927]. The enzyme from *Arabidopsis thaliana* shows high activity with spermidine, but also oxidizes other polyamines [2904]. The specific polyamine oxidases are classified as EC 1.5.3.13 (N^1 -acetylpolyamine oxidase), EC 1.5.3.14 (polyamine oxidase (propane-1,3-diamine-forming)), EC 1.5.3.15 (N^8 -acetylspermidine oxidase (propane-1,3-diamine-forming)) and EC 1.5.3.16 (spermine oxidase).

References: [2904, 2927, 2339]

[EC 1.5.3.17 created 2009]

EC 1.5.3.18

Accepted name: L-saccharopine oxidase

Reaction: N^6 -(L-1,3-dicarboxypropyl)-L-lysine + H₂O + O₂ = (S)-2-amino-6-oxohexanoate + L-glutamate + H₂O₂

Other name(s): FAP2

Systematic name: L-saccharopine:oxygen oxidoreductase (L-glutamate-forming)

Comments: The enzyme is involved in pipercolic acid biosynthesis. A flavoprotein (FAD).

References: [4811, 4617]

[EC 1.5.3.18 created 2011]

EC 1.5.3.19

Accepted name: 4-methylaminobutanoate oxidase (formaldehyde-forming)

Reaction: 4-methylaminobutanoate + O₂ + H₂O = 4-aminobutanoate + formaldehyde + H₂O₂

Other name(s): *mabO* (gene name)

Systematic name: 4-methylaminobutanoate:oxygen oxidoreductase (formaldehyde-forming)

Comments: A flavoprotein (FAD). In the enzyme from the soil bacterium *Arthrobacter nicotinovorans* the cofactor is covalently bound. Participates in the nicotine degradation pathway of this organism.

References: [657]

[EC 1.5.3.19 created 2012]

EC 1.5.3.20

Accepted name: *N*-alkylglycine oxidase

Reaction: *N*-alkylglycine + H₂O + O₂ = alkylamine + glyoxalate + H₂O₂

Other name(s): *N*-carboxymethylalkylamine:oxygen oxidoreductase (decarboxymethylating)

Systematic name: *N*-alkylglycine:oxygen oxidoreductase (alkylamine-forming)

Comments: Isolated from the mold *Cladosporium* sp. G-10. Acts on N^6 -(carboxymethyl)lysine, 6-[(carboxymethyl)amino]hexanoic acid, sarcosine and *N*-ethylglycine. It has negligible action on glycine (*cf.* EC 1.4.3.19 glycine oxidase).

References: [1356]

[EC 1.5.3.20 created 2012]

EC 1.5.3.21

Accepted name: 4-methylaminobutanoate oxidase (methylamine-forming)

Reaction: 4-methylaminobutanoate + O₂ + H₂O = succinate semialdehyde + methylamine + H₂O₂

Other name(s): *mao* (gene name, ambiguous)

Systematic name: 4-methylaminobutanoate methylamidohydrolase

Comments: The enzyme participates in the nicotine degradation pathway of the soil bacterium *Arthrobacter nicotinovorans*. Has a very weak monoamine oxidase (EC 1.4.3.4) activity with 4-aminobutanoate [657].

References: [657, 656]

[EC 1.5.3.21 created 2012]

EC 1.5.3.22

Accepted name: coenzyme F₄₂₀H₂ oxidase

Reaction: 2 reduced coenzyme F₄₂₀ + O₂ = 2 oxidized coenzyme F₄₂₀ + 2 H₂O

Other name(s): FprA

Systematic name: reduced coenzyme F₄₂₀:oxygen oxidoreductase

Comments: The enzyme contains FMN and a binuclear iron center. The enzyme from the archaeon *Methanothermobacter marburgensis* is Si-face specific with respect to C-5 of coenzyme F₄₂₀ [3780].

References: [3778, 3780, 3779]

[EC 1.5.3.22 created 2013]

EC 1.5.3.23

Accepted name: glyphosate oxidoreductase

Reaction: 2 glyphosate + O₂ = 2 aminomethylphosphonate + 2 glyoxylate

Other name(s): *gox* (gene name)

Systematic name: glyphosate oxidoreductase (aminomethylphosphonate-forming)

Comments: The enzyme, characterized from the bacterium *Ochrobactrum* sp. G-1, contains an FAD cofactor. The catalytic cycle starts with a reduction of the FAD cofactor by one molecule of glyphosate, yielding reduced FAD and a Schiff base of aminomethylphosphonate with glyoxylate that is hydrolysed to the single components. The reduced FAD is reoxidized by oxygen, generating water and an oxygenated flavin intermediate, which catalyses the oxygenation of a second molecule of glyphosate, forming the second pair of aminomethylphosphonate and glyoxylate.

References: [1076, 4146]

[EC 1.5.3.23 created 2016]

EC 1.5.3.24

Accepted name: sarcosine oxidase (5,10-methylenetetrahydrofolate-forming)

Reaction: sarcosine + 5,6,7,8-tetrahydrofolate + O₂ = glycine + 5,10-methylenetetrahydrofolate + H₂O₂

Other name(s): TSOX; sarcosine oxidase (ambiguous); heterotetrameric sarcosine oxidase

Systematic name: sarcosine, 5,6,7,8-tetrahydrofolate:O₂ oxidoreductase (demethylating,5,10-methylenetetrahydrofolate-forming)

Comments: The enzyme, found in some bacterial species, is composed of four different subunits and two active sites connected by a large "reaction chamber". An imine intermediate is transferred between the sites, eliminating the production of toxic formaldehyde. The enzyme contains three cofactors: noncovalently bound FAD and NAD⁺, and FMN that is covalently bound to a histidine residue. In the absence of folate the enzyme catalyses the reaction of EC 1.5.3.1, sarcosine oxidase (formaldehyde-forming).

References: [1581, 4140, 663, 662, 1063]

[EC 1.5.3.24 created 2022]

EC 1.5.3.25

Accepted name: fructosyl amine oxidase (glucosone-forming)

Reaction: an *N*-(1-deoxy-D-fructos-1-yl)amine + O₂ + H₂O = D-glucosone + an amine + H₂O₂ (overall reaction)

(1a) an *N*-(1-deoxy-D-fructos-1-yl)amine + O₂ = a 2-[(3*S*,4*R*,5*R*)-3,4,5,6-tetrahydroxy-2-oxohexylidene]amine + H₂O₂

(1b) a 2-[(3*S*,4*R*,5*R*)-3,4,5,6-tetrahydroxy-2-oxohexylidene]amine + H₂O = D-glucosone + an amine (spontaneous)

Other name(s): amadoriase
Systematic name: *N*-(1-deoxy-D-fructos-1-yl)amine:oxygen 2-oxidoreductase (glucosone-forming)
Comments: Reducing sugars such as glucose react with amino groups in proteins via the spontaneous Maillard reaction, forming an unstable product that undergoes spontaneous rearrangement to a keto amine compound. These reactions are known as glycation reactions, and the stable products are known as Amadori products. This enzyme, which contains an FAD cofactor, catalyses a deglycation reaction that regenerates the amine reactant. By-products are glucosone and hydrogen peroxide. The enzymes have been reported from fungi and bacteria, but not from higher eukaryotes. Specific enzymes differ in their substrate specificity. *cf.* EC 1.5.3.26, fructosyl amine oxidase (fructosamine-forming).
References: [1729, 4169, 1672, 4681, 3643]

[EC 1.5.3.25 created 2022]

EC 1.5.3.26

Accepted name: fructosyl amine oxidase (fructosamine-forming)
Reaction: an *N*-(1-deoxy-D-fructos-1-yl)amine + O₂ + H₂O = (1-deoxy-D-fructos-1-yl)amine + an aldehyde + H₂O₂
Systematic name: *N*-(1-deoxy-D-fructos-1-yl)amine:oxygen oxidoreductase (fructosamine-forming)
Comments: Reducing sugars such as glucose react with amino groups in proteins via the spontaneous Maillard reaction, forming an unstable product that undergoes spontaneous rearrangement to a keto amine compound. These reactions are known as glycation reactions, and the stable products are known as Amadori products. This enzyme, characterized from a *Pseudomonas* sp. strain, cleaves the Amadori products at the alkylamine bond. All other known fructosyl amine oxidases cleave the ketoamine bond (*cf.* EC 1.5.3.25, fructosyl amine oxidase (glucosone-forming)).
References: [1303, 3687, 4681]

[EC 1.5.3.26 created 2022]

EC 1.5.4 With a disulfide as acceptor

EC 1.5.4.1

Accepted name: pyrimidodiazepine synthase
Reaction: 2-amino-6-acetyl-3,7,8,9-tetrahydro-3*H*-pyrimido[4,5-*b*][1,4]diazepin-4-one + glutathione disulfide + H₂O = 6-pyruvoyltetrahydropterin + 2 glutathione
Other name(s): PDA synthase; pyrimidodiazepine:oxidized-glutathione oxidoreductase (ring-opening, cyclizing); pyrimidodiazepine:glutathione-disulfide oxidoreductase (ring-opening, cyclizing)
Systematic name: 2-amino-6-acetyl-3,7,8,9-tetrahydro-3*H*-pyrimido[4,5-*b*][1,4]diazepin-4-one:glutathione-disulfide oxidoreductase (ring-opening, cyclizing)
Comments: In the reverse direction, the reduction of 6-pyruvoyl-tetrahydropterin is accompanied by the opening of the 6-membered pyrazine ring and the formation of the 7-membered diazepine ring. The pyrimidodiazepine formed is an acetyldihydro derivative. Involved in the formation of the eye pigment drosopterin in *Drosophila melanogaster*.
References: [4620, 2096]

[EC 1.5.4.1 created 1990, modified 2014]

EC 1.5.5 With a quinone or similar compound as acceptor

EC 1.5.5.1

- Accepted name:** electron-transferring-flavoprotein dehydrogenase
Reaction: reduced electron-transferring flavoprotein + ubiquinone = electron-transferring flavoprotein + ubiquinol
Other name(s): ETF-QO; ETF:ubiquinone oxidoreductase; electron transfer flavoprotein dehydrogenase; electron transfer flavoprotein Q oxidoreductase; electron transfer flavoprotein-ubiquinone oxidoreductase; electron transfer flavoprotein reductase
Systematic name: electron-transferring-flavoprotein:ubiquinone oxidoreductase
Comments: An iron-sulfur flavoprotein, forming part of the mitochondrial electron-transfer system.
References: [265, 3610]

[EC 1.5.5.1 created 1986]

EC 1.5.5.2

- Accepted name:** proline dehydrogenase
Reaction: L-proline + a quinone = (S)-1-pyrroline-5-carboxylate + a quinol
Other name(s): L-proline dehydrogenase; L-proline:(acceptor) oxidoreductase
Systematic name: L-proline:quinone oxidoreductase
Comments: A flavoprotein (FAD). The electrons from L-proline are transferred to the FAD cofactor, and from there to a quinone acceptor [2911]. In many organisms, ranging from bacteria to mammals, proline is oxidized to glutamate in a two-step process involving this enzyme and EC 1.2.1.88, L-glutamate γ -semialdehyde dehydrogenase. Both activities are carried out by the same enzyme in enterobacteria.
References: [3691, 458, 2911]

[EC 1.5.5.2 created 1980 as EC 1.5.99.8, transferred 2013 to EC 1.5.5.2]

EC 1.5.5.3

- Accepted name:** hydroxyproline dehydrogenase
Reaction: *trans*-4-hydroxy-L-proline + a quinone = (3*R*,5*S*)-3-hydroxy-1-pyrroline-5-carboxylate + a quinol
Other name(s): HYPDH; OH-POX; hydroxyproline oxidase; PRODH2 (gene name)
Systematic name: *trans*-4-hydroxy-L-proline:quinone oxidoreductase
Comments: A flavoprotein (FAD). The enzyme from human also has low activity with L-proline (*cf.* EC 1.5.5.2, proline dehydrogenase).
References: [731, 4116]

[EC 1.5.5.3 created 2017]

EC 1.5.7 With an iron-sulfur protein as acceptor

EC 1.5.7.1

- Accepted name:** methylenetetrahydrofolate reductase (ferredoxin)
Reaction: 5-methyltetrahydrofolate + 2 oxidized ferredoxin = 5,10-methylenetetrahydrofolate + 2 reduced ferredoxin + 2 H⁺
Other name(s): 5,10-methylenetetrahydrofolate reductase
Systematic name: 5-methyltetrahydrofolate:ferredoxin oxidoreductase
Comments: An iron-sulfur flavoprotein that also contains zinc. The enzyme from *Clostridium formicoaceticum* catalyses the reduction of methylene blue, menadione, benzyl viologen, rubredoxin or FAD with 5-methyltetrahydrofolate and the oxidation of reduced ferredoxin or FADH₂ with 5,10-methylenetetrahydrofolate. However, unlike EC 1.5.1.53, methylenetetrahydrofolate reductase (NADPH); EC 1.5.1.54, methylenetetrahydrofolate reductase (NADH); or EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H], there is no activity with either NADH or NADP⁺.
References: [696]

[EC 1.5.7.1 created 2005, modified 2021]

EC 1.5.7.2

Accepted name: coenzyme F₄₂₀ oxidoreductase (ferredoxin)
Reaction: reduced coenzyme F₄₂₀ + 2 oxidized ferredoxin = oxidized coenzyme F₄₂₀ + 2 reduced ferredoxin + 2 H⁺
Other name(s): Fd:F420 oxidoreductase; FpoF protein; ferredoxin:F420 oxidoreductase
Systematic name: coenzyme F₄₂₀:ferredoxin oxidoreductase
Comments: The enzyme from the archaeon *Methanosarcina mazei* contains iron-sulfur centres and FAD.
References: [4586]

[EC 1.5.7.2 created 2013]

EC 1.5.7.3

Accepted name: *N,N*-dimethylglycine/sarcosine dehydrogenase (ferredoxin)
Reaction: (1) *N,N*-dimethylglycine + 2 oxidized ferredoxin + H₂O = sarcosine + formaldehyde + 2 reduced ferredoxin + 2 H⁺
(2) sarcosine + 2 oxidized ferredoxin + H₂O = glycine + formaldehyde + 2 reduced ferredoxin + 2 H⁺
Other name(s): *ddhC* (gene name); *dgcA* (gene name)
Systematic name: *N,N*-dimethylglycine/sarcosine:ferredoxin oxidoreductase (demethylating)
Comments: This bacterial enzyme is involved in degradation of glycine betaine. The enzyme contains non-covalently bound FAD and NAD(P) cofactors, and catalyses the demethylation of both *N,N*-dimethylglycine and sarcosine, releasing formaldehyde and forming glycine as the final product. The enzyme can utilize both NAD⁺ and NADP⁺, but the catalytic efficiency with NAD⁺ is 5-fold higher. The native electron acceptor of the enzyme is a membrane-bound clostridial-type ferredoxin, which transfers the electrons to an electron-transfer flavoprotein (ETF).
References: [4543, 4765]

[EC 1.5.7.3 created 2022]

EC 1.5.8 With a flavin or flavoprotein as acceptor

EC 1.5.8.1

Accepted name: dimethylamine dehydrogenase
Reaction: dimethylamine + H₂O + electron-transfer flavoprotein = methylamine + formaldehyde + reduced electron-transfer flavoprotein
Systematic name: dimethylamine:electron-transfer flavoprotein oxidoreductase
Comments: Contains FAD and a [4Fe-4S] cluster.
References: [4762]

[EC 1.5.8.1 created 1999 as EC 1.5.99.10, transferred 2002 to EC 1.5.8.1]

EC 1.5.8.2

Accepted name: trimethylamine dehydrogenase
Reaction: trimethylamine + H₂O + electron-transfer flavoprotein = dimethylamine + formaldehyde + reduced electron-transfer flavoprotein
Systematic name: trimethylamine:electron-transfer flavoprotein oxidoreductase (demethylating)
Comments: A number of alkyl-substituted derivatives of trimethylamine can also act as electron donors; phenazine methosulfate and 2,6-dichloroindophenol can act as electron acceptors. Contains FAD and a [4Fe-4S] cluster.
References: [709, 4012, 1755, 1947, 3776]

[EC 1.5.8.2 created 1976 as EC 1.5.99.7, transferred 2002 to EC 1.5.8.2]

EC 1.5.8.3

- Accepted name:** sarcosine dehydrogenase
Reaction: sarcosine + 5,6,7,8-tetrahydrofolate + oxidized [electron-transfer flavoprotein] = glycine + 5,10-methylenetetrahydrofolate + reduced [electron-transfer flavoprotein]
Other name(s): sarcosine *N*-demethylase; monomethylglycine dehydrogenase; sarcosine:(acceptor) oxidoreductase (demethylating); sarcosine:electron-transfer flavoprotein oxidoreductase (demethylating)
Systematic name: sarcosine, 5,6,7,8-tetrahydrofolate:electron-transferflavoprotein oxidoreductase (demethylating,5,10-methylenetetrahydrofolate-forming)
Comments: A flavoprotein (FMN) found in eukaryotes. In the absence of tetrahydrofolate the enzyme produces formaldehyde. *cf.* EC 1.5.3.1, sarcosine oxidase (formaldehyde-forming), and EC 1.5.3.24, sarcosine oxidase (5,10-methylenetetrahydrofolate-forming).
References: [1738, 1190, 4649, 4011]

[EC 1.5.8.3 created 1972 as EC 1.5.99.1, transferred 2012 to EC 1.5.8.3, modified 2022]

EC 1.5.8.4

- Accepted name:** dimethylglycine dehydrogenase
Reaction: *N,N*-dimethylglycine + 5,6,7,8-tetrahydrofolate + electron-transfer flavoprotein = sarcosine + 5,10-methylenetetrahydrofolate + reduced electron-transfer flavoprotein
Other name(s): *N,N*-dimethylglycine oxidase; *N,N*-dimethylglycine:(acceptor) oxidoreductase (demethylating); Me2GlyDH; *N,N*-dimethylglycine:electron-transfer flavoprotein oxidoreductase (demethylating)
Systematic name: *N,N*-dimethylglycine,5,6,7,8-tetrahydrofolate:electron-transferflavoprotein oxidoreductase (demethylating,5,10-methylenetetrahydrofolate-forming)
Comments: A flavoprotein, containing a histidyl(*N*^T)-(8 α)FAD linkage at position 91 in the human protein. An imine intermediate is channeled from the FAD binding site to the 5,6,7,8-tetrahydrofolate binding site through a 40 Å tunnel [5,8,9]. In the absence of 5,6,7,8-tetrahydrofolate the enzyme forms formaldehyde [3356, 155].
References: [1190, 1738, 4650, 4649, 3356, 446, 447, 2557, 155]

[EC 1.5.8.4 created 1972 as EC 1.5.99.2, transferred 2012 to EC 1.5.8.4, modified 2017]

EC 1.5.98 With other, known, physiological acceptors

EC 1.5.98.1

- Accepted name:** methylenetetrahydromethanopterin dehydrogenase
Reaction: 5,10-methylenetetrahydromethanopterin + oxidized coenzyme F₄₂₀ = 5,10-methenyltetrahydromethanopterin + reduced coenzyme F₄₂₀
Other name(s): *N*⁵,*N*¹⁰-methylenetetrahydromethanopterin dehydrogenase; 5,10-methylenetetrahydromethanopterin dehydrogenase
Systematic name: 5,10-methylenetetrahydromethanopterin:coenzyme-F₄₂₀ oxidoreductase
Comments: Coenzyme F₄₂₀ is a 7,8-didemethyl-8-hydroxy-5-deazariboflavin derivative; methanopterin is a pterin analogue. The enzyme is involved in the formation of methane from CO₂ in the methanogen *Methanothermobacter thermoautotrophicus*.
References: [1543, 4233]

[EC 1.5.98.1 created 1989 as EC 1.5.99.9, modified 2004, transferred to EC 1.5.98.1 2014]

EC 1.5.98.2

- Accepted name:** 5,10-methylenetetrahydromethanopterin reductase
Reaction: 5-methyltetrahydromethanopterin + oxidized coenzyme F₄₂₀ = 5,10-methylenetetrahydromethanopterin + reduced coenzyme F₄₂₀

Other name(s): 5,10-methylenetetrahydromethanopterin cyclohydrolase; *N*⁵,*N*¹⁰-methylenetetrahydromethanopterin reductase; methylene-H₄MPT reductase; coenzyme F₄₂₀-dependent *N*⁵,*N*¹⁰-methylenetetrahydromethanopterin reductase; *N*⁵,*N*¹⁰-methylenetetrahydromethanopterin:coenzyme-F₄₂₀ oxidoreductase
Systematic name: 5-methyltetrahydromethanopterin:coenzyme-F₄₂₀ oxidoreductase
Comments: Catalyses an intermediate step in methanogenesis from CO₂ and H₂ in methanogenic archaea.
References: [2576, 4233, 2577, 4235, 4234]

[EC 1.5.98.2 created 2000 as EC 1.5.99.11, modified 2004, transferred to EC 1.5.98.2 2014]

EC 1.5.98.3

Accepted name: coenzyme F₄₂₀:methanophenazine dehydrogenase
Reaction: reduced coenzyme F₄₂₀ + methanophenazine = oxidized coenzyme F₄₂₀ + dihydromethanophenazine
Other name(s): F₄₂₀H₂ dehydrogenase; fpoBCDIF (gene names)
Systematic name: reduced coenzyme F₄₂₀:methanophenazine oxidoreductase
Comments: The enzyme, found in some methanogenic archaea, is responsible for the reoxidation of coenzyme F₄₂₀, which is reduced during methanogenesis, and for the reduction of methanophenazine to dihydromethanophenazine, which is required by EC 1.8.98.1, dihydromethanophenazine:CoB-CoM heterodisulfide reductase. The enzyme is membrane-bound, and is coupled to proton translocation across the cytoplasmic membrane, generating a proton motive force that is used for ATP generation.
References: [449, 246, 884, 1867]

[EC 1.5.98.3 created 2017]

EC 1.5.99 With unknown physiological acceptors

[1.5.99.1 *Transferred entry. sarcosine dehydrogenase. Now EC 1.5.8.3, sarcosine dehydrogenase*]

[EC 1.5.99.1 created 1972, deleted 2012]

[1.5.99.2 *Transferred entry. dimethylglycine dehydrogenase. Now EC 1.5.8.4, dimethylglycine dehydrogenase*]

[EC 1.5.99.2 created 1972, deleted 2012]

EC 1.5.99.3

Accepted name: L-pipecolate dehydrogenase
Reaction: L-pipecolate + acceptor = (*S*)-2,3,4,5-tetrahydropyridine-2-carboxylate + reduced acceptor
Other name(s): L-pipecolate:(acceptor) 1,6-oxidoreductase
Systematic name: L-pipecolate:acceptor 1,6-oxidoreductase
Comments: The product reacts with water to form (*S*)-2-amino-6-oxohexanoate.
References: [179]

[EC 1.5.99.3 created 1972, modified 1986, modified 2011]

EC 1.5.99.4

Accepted name: nicotine dehydrogenase
Reaction: (*S*)-nicotine + acceptor + H₂O = (*S*)-6-hydroxynicotine + reduced acceptor
Other name(s): nicotine oxidase; D-nicotine oxidase; nicotine:(acceptor) 6-oxidoreductase (hydroxylating); L-nicotine oxidase
Systematic name: nicotine:acceptor 6-oxidoreductase (hydroxylating)
Comments: A metalloprotein (FMN). The enzyme can act on both the naturally found (*S*)-enantiomer and the synthetic (*R*)-enantiomer of nicotine, with retention of configuration in both cases [1682].
References: [271, 859, 1681, 1682]

[EC 1.5.99.4 created 1972]

EC 1.5.99.5

Accepted name: methylglutamate dehydrogenase
Reaction: *N*-methyl-L-glutamate + acceptor + H₂O = L-glutamate + formaldehyde + reduced acceptor
Other name(s): *N*-methylglutamate dehydrogenase; *N*-methyl-L-glutamate:(acceptor) oxidoreductase (demethylating)
Systematic name: *N*-methyl-L-glutamate:acceptor oxidoreductase (demethylating)
Comments: A number of *N*-methyl-substituted amino acids can act as donor; 2,6-dichloroindophenol is the best acceptor.
References: [1631]

[EC 1.5.99.5 created 1976]

EC 1.5.99.6

Accepted name: spermidine dehydrogenase
Reaction: spermidine + acceptor + H₂O = propane-1,3-diamine + 4-aminobutanal + reduced acceptor
Other name(s): spermidine:(acceptor) oxidoreductase
Systematic name: spermidine:acceptor oxidoreductase
Comments: A flavohemoprotein (FAD). Ferricyanide, 2,6-dichloroindophenol and cytochrome *c* can act as acceptor. 4-Aminobutanal condenses non-enzymically to 1-pyrroline.
References: [4158, 4159]

[EC 1.5.99.6 created 1976]

[1.5.99.7 *Transferred entry. trimethylamine dehydrogenase. Now EC 1.5.8.2, trimethylamine dehydrogenase*]

[EC 1.5.99.7 created 1976, deleted 2002]

[1.5.99.8 *Transferred entry. proline dehydrogenase. Now EC 1.5.5.2, proline dehydrogenase.]*

[EC 1.5.99.8 created 1980, deleted 2013]

[1.5.99.9 *Transferred entry. methylenetetrahydromethanopterin dehydrogenase. As the acceptor is known the enzyme has been transferred to EC 1.5.98.1, methylenetetrahydromethanopterin dehydrogenase*]

[EC 1.5.99.9 created 1989, modified 2004, deleted 2014]

[1.5.99.10 *Transferred entry. dimethylamine dehydrogenase. Now EC 1.5.8.1, dimethylamine dehydrogenase*]

[EC 1.5.99.10 created 1999, deleted 2002]

[1.5.99.11 *Transferred entry. methylenetetrahydromethanopterin dehydrogenase. As the acceptor is known the enzyme has been transferred to EC 1.5.98.2, 5,10-methylenetetrahydromethanopterin reductase*]

[EC 1.5.99.11 created 2000, modified 2004, deleted 2014]

EC 1.5.99.12

Accepted name: cytokinin dehydrogenase
Reaction: *N*⁶-prenyladenine + acceptor + H₂O = adenine + 3-methylbut-2-enal + reduced acceptor
Other name(s): *N*⁶-dimethylallyladenine:(acceptor) oxidoreductase; 6-*N*-dimethylallyladenine:acceptor oxidoreductase; OsCKX2; CKX; cytokinin oxidase/dehydrogenase; *N*⁶-dimethylallyladenine:acceptor oxidoreductase
Systematic name: *N*⁶-prenyladenine:acceptor oxidoreductase
Comments: A flavoprotein (FAD). Catalyses the oxidation of cytokinins, a family of *N*⁶-substituted adenine derivatives that are plant hormones, where the substituent is a prenyl group. Although this activity was previously thought to be catalysed by a hydrogen-peroxide-forming oxidase, this enzyme does not require oxygen for activity and does not form hydrogen peroxide. 2,6-Dichloroindophenol, methylene blue, nitroblue tetrazolium, phenazine methosulfate and copper(II) in the presence of imidazole can act as acceptors. This enzyme plays a part in regulating rice-grain production, with lower levels of the enzyme resulting in enhanced grain production [144].

References: [1263, 144]

[EC 1.5.99.12 created 2001]

EC 1.5.99.13

Accepted name: D-proline dehydrogenase
Reaction: D-proline + acceptor = 1-pyrroline-2-carboxylate + reduced acceptor
Other name(s): D-Pro DH; D-Pro dehydrogenase; dye-linked D-proline dehydrogenase
Systematic name: D-proline:acceptor oxidoreductase
Comments: A flavoprotein (FAD). The enzyme prefers D-proline and acts on other D-amino acids with lower efficiency.
References: [4206, 3674]

[EC 1.5.99.13 created 2010, modified 2011]

EC 1.5.99.14

Accepted name: 6-hydroxypseudooxynicotine dehydrogenase
Reaction: 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one + acceptor + H₂O = 1-(2,6-dihydroxypyridin-3-yl)-4-(methylamino)butan-1-one + reduced acceptor
Systematic name: 1-(6-hydroxypyridin-3-yl)-4-(methylamino)butan-1-one:acceptor 6-oxidoreductase (hydroxylating)
Comments: Contains a cytidyl molybdenum cofactor [3621]. The enzyme, which participates in the nicotine degradation pathway, has been characterized from the soil bacterium *Arthrobacter nicotinovorans* [1174, 1407].
References: [1174, 1407, 3621]

[EC 1.5.99.14 created 2012]

EC 1.5.99.15

Accepted name: dihydromethanopterin reductase (acceptor)
Reaction: 5,6,7,8-tetrahydromethanopterin + oxidized acceptor = 7,8-dihydromethanopterin + reduced acceptor
Other name(s): DmrX
Systematic name: 5,6,7,8-tetrahydromethanopterin:acceptor 5,6-oxidoreductase
Comments: This archaeal enzyme catalyses the last step in the biosynthesis of tetrahydromethanopterin, a coenzyme used in methanogenesis. The enzyme, characterized from the archaea *Methanosarcina mazei* and *Methanocaldococcus jannaschii*, is an iron-sulfur flavoprotein. *cf.* EC 1.5.1.47, dihydromethanopterin reductase [NAD(P)⁺].
References: [4531]

[EC 1.5.99.15 created 2014]

EC 1.6 Acting on NADH or NADPH

In general, enzymes using NADH or NADPH to reduce a substrate are classified according to the reverse reaction, in which NAD⁺ or NADP⁺ is formally regarded as acceptor. This subclass contains only those enzymes in which some other redox carrier is the acceptor. This can be NAD⁺ or NADP⁺ (EC 1.6.1), a heme protein (EC 1.6.2), oxygen (EC 1.6.3), a quinone or similar compound (EC 1.6.5), a nitrogenous group (EC 1.6.6), or some other acceptor (EC 1.6.99).

EC 1.6.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.6.1.1

- Accepted name:** NAD(P)⁺ transhydrogenase (*Si*-specific)
Reaction: NADPH + NAD⁺ = NADP⁺ + NADH
Other name(s): pyridine nucleotide transhydrogenase; transhydrogenase; NAD(P)⁺ transhydrogenase; nicotinamide adenine dinucleotide (phosphate) transhydrogenase; NAD⁺ transhydrogenase; NADH transhydrogenase; nicotinamide nucleotide transhydrogenase; NADPH-NAD⁺ transhydrogenase; pyridine nucleotide transferase; NADPH-NAD⁺ oxidoreductase; NADH-NADP⁺-transhydrogenase; NADPH:NAD⁺ transhydrogenase; H⁺-Thase; non-energy-linked transhydrogenase; NADPH:NAD⁺ oxidoreductase (B-specific); NAD(P)⁺ transhydrogenase (B-specific)
Systematic name: NADPH:NAD⁺ oxidoreductase (*Si*-specific)
Comments: The enzyme from *Azotobacter vinelandii* is a flavoprotein (FAD). It is *Si*-specific with respect to both NAD⁺ and NADP⁺. Also acts on deamino coenzymes [*cf.* EC 1.6.1.2 NAD(P)⁺ transhydrogenase (*Re/Si*-specific)].
References: [1772, 4820]

[EC 1.6.1.1 created 1961, modified 1986, modified 2013]

EC 1.6.1.2

- Accepted name:** NAD(P)⁺ transhydrogenase (*Re/Si*-specific)
Reaction: NADPH + NAD⁺ = NADP⁺ + NADH
Other name(s): pyridine nucleotide transhydrogenase; transhydrogenase; NAD(P)⁺ transhydrogenase; nicotinamide adenine dinucleotide (phosphate) transhydrogenase; NAD⁺ transhydrogenase; NADH transhydrogenase; nicotinamide nucleotide transhydrogenase; NADPH-NAD⁺ transhydrogenase; pyridine nucleotide transferase; NADPH-NAD⁺ oxidoreductase; NADH-NADP⁺-transhydrogenase; NADPH:NAD⁺ transhydrogenase; H⁺-Thase; energy-linked transhydrogenase; NAD(P) transhydrogenase (AB-specific); NAD(P)⁺ transhydrogenase (AB-specific); NADPH:NAD⁺ oxidoreductase (AB-specific)
Systematic name: NADPH:NAD⁺ oxidoreductase (*Re/Si*-specific)
Comments: The enzyme from heart mitochondria is *Re*-specific with respect to NAD⁺ and *Si*-specific with respect to NADP⁺ [*cf.* EC 1.6.1.1 NAD(P)⁺ transhydrogenase (*Si*-specific)].
References: [1127, 4820]

[EC 1.6.1.2 created 1986, modified 2013]

EC 1.6.1.3

- Accepted name:** NAD(P)⁺ transhydrogenase
Reaction: NADPH + NAD⁺ = NADP⁺ + NADH
Other name(s): pyridine nucleotide transhydrogenase; transhydrogenase (ambiguous); nicotinamide adenine dinucleotide (phosphate) transhydrogenase (ambiguous); NAD⁺ transhydrogenase (ambiguous); NADH transhydrogenase (misleading); nicotinamide nucleotide transhydrogenase (ambiguous); NADPH-NAD⁺ transhydrogenase (ambiguous); pyridine nucleotide transferase (ambiguous); NADPH-NAD⁺ oxidoreductase (ambiguous); NADH-NADP⁺-transhydrogenase (ambiguous); NADPH:NAD⁺ transhydrogenase; H⁺-Thase (ambiguous); non-energy-linked transhydrogenase (ambiguous)
Systematic name: NADPH:NAD⁺ oxidoreductase
Comments: The enzyme catalyses the NADPH-driven reduction of NAD⁺. This entry stands for enzymes whose stereo-specificity with respect to NADPH is not known. [*cf.* EC 1.6.1.1, NAD(P)⁺ transhydrogenase (*Si*-specific) and EC 1.6.1.2 NAD(P)⁺ transhydrogenase (*Re/Si*-specific)].
References: [931]

[EC 1.6.1.3 created 2013]

EC 1.6.1.4

- Accepted name:** NAD(P)⁺ transhydrogenase (ferredoxin)
Reaction: NADH + H⁺ + 2 NADP⁺ + 2 reduced ferredoxin [iron-sulfur] cluster = NAD⁺ + 2 NADPH + 2 oxidized ferredoxin [iron-sulfur] cluster
Other name(s): NADH-dependent reduced ferredoxin:NADP⁺ oxidoreductase; Nfn; *nfnAB* (gene names)
Systematic name: NADH:NADP⁺, ferredoxin oxidoreductase
Comments: The iron-sulfur flavoprotein complex, originally isolated from the bacterium *Clostridium kluyveri*, couples the exergonic reduction of NADP⁺ with reduced ferredoxin and the endergonic reduction of NADP⁺ with NADH.
References: [4530, 874, 2554]

[EC 1.6.1.4 created 2015]

[1.6.1.5 *Transferred entry. proton-translocating NAD(P)⁺ transhydrogenase. Now EC 7.1.1.1, proton-translocating NAD(P)⁺ transhydrogenase*]

[EC 1.6.1.5 created 2015, deleted 2018]

EC 1.6.2 With a heme protein as acceptor

[1.6.2.1 *Transferred entry. NADH₂ cytochrome c reductase. Now EC 1.6.99.3, NADH dehydrogenase*]

[EC 1.6.2.1 created 1961, deleted 1965]

EC 1.6.2.2

- Accepted name:** cytochrome-*b*₅ reductase
Reaction: NADH + 2 ferricytochrome *b*₅ = NAD⁺ + H⁺ + 2 ferrocycytochrome *b*₅
Other name(s): cytochrome *b*₅ reductase; dihydronicotinamide adenine dinucleotide-cytochrome *b*₅ reductase; reduced nicotinamide adeninedinucleotide-cytochrome *b*₅ reductase; NADH-ferricytochrome *b*₅ oxidoreductase; NADH-cytochrome *b*₅ reductase; NADH 5α-reductase ; NADH-cytochrome-*b*₅ reductase
Systematic name: NADH:ferricytochrome-*b*₅ oxidoreductase
Comments: A flavoprotein (FAD).
References: [2616, 4068, 4070]

[EC 1.6.2.2 created 1961]

[1.6.2.3 *Deleted entry. cytochrome reductase (NADPH)*]

[EC 1.6.2.3 created 1972, deleted 1965]

EC 1.6.2.4

- Accepted name:** NADPH—hemoprotein reductase
Reaction: NADPH + H⁺ + *n* oxidized hemoprotein = NADP⁺ + *n* reduced hemoprotein
Other name(s): CPR; FAD-cytochrome *c* reductase; NADP-cytochrome *c* reductase; NADP-cytochrome reductase; NADPH-dependent cytochrome *c* reductase; NADPH:*P*-450 reductase; NADPH:ferrihemoprotein oxidoreductase; NADPH—cytochrome *P*-450 oxidoreductase; NADPH-cytochrome *c* oxidoreductase; NADPH-cytochrome *c* reductase; NADPH—cytochrome *p*-450 reductase; NADPH-ferricytochrome *c* oxidoreductase; NADPH-ferrihemoprotein reductase; TPNH₂ cytochrome *c* reductase; TPNH-cytochrome *c* reductase; aldehyde reductase (NADPH-dependent); cytochrome *P*-450 reductase; cytochrome *c* reductase (reduced nicotinamide adenine dinucleotide phosphate, NADPH, NADPH-dependent); dihydroxynicotinamide adenine dinucleotide phosphate-cytochrome *c* reductase; ferrihemoprotein *P*-450 reductase; reduced nicotinamide adenine dinucleotide phosphate-cytochrome *c* reductase; reductase, cytochrome *c* (reduced nicotinamide adenine dinucleotide phosphate)
Systematic name: NADPH:hemoprotein oxidoreductase

Comments: A flavoprotein containing both FMN and FAD. This enzyme catalyses the transfer of electrons from NADPH, an obligatory two-electron donor, to microsomal *P*-450 monooxygenases (e.g. EC 1.14.14.1, unspecific monooxygenase) by stabilizing the one-electron reduced form of the flavin co-factors FAD and FMN. It also reduces cytochrome *b*₅ and cytochrome *c*. The number *n* in the equation is 1 if the hemoprotein undergoes a 2-electron reduction, and is 2 if it undergoes a 1-electron reduction.

References: [1462, 1720, 2548, 2691, 4630, 2690, 3810, 4521, 2935, 1456]

[EC 1.6.2.4 created 1972, modified 2003]

EC 1.6.2.5

Accepted name: NADPH—cytochrome-*c*₂ reductase
Reaction: $\text{NADPH} + 2 \text{ ferricytochrome } c_2 = \text{NADP}^+ + \text{H}^+ + 2 \text{ ferrocycytochrome } c_2$
Other name(s): cytochrome *c*₂ reductase (reduced nicotinamide adenine dinucleotide phosphate); cytochrome *c*₂ reductase (reduced nicotinamide adenine dinucleotide phosphate, NADPH)
Systematic name: NADPH:ferricytochrome-*c*₂ oxidoreductase
Comments: A flavoprotein (FAD).
References: [3620]

[EC 1.6.2.5 created 1972]

EC 1.6.2.6

Accepted name: leghemoglobin reductase
Reaction: $\text{NAD(P)H} + \text{H}^+ + 2 \text{ ferrileghemoglobin} = \text{NAD(P)}^+ + 2 \text{ ferroleghemoglobin}$
Other name(s): ferric leghemoglobin reductase
Systematic name: NAD(P)H:ferrileghemoglobin oxidoreductase
References: [3619]

[EC 1.6.2.6 created 1989]

EC 1.6.3 With oxygen as acceptor

EC 1.6.3.1

Accepted name: NAD(P)H oxidase (H₂O₂-forming)
Reaction: $\text{NAD(P)H} + \text{H}^+ + \text{O}_2 = \text{NAD(P)}^+ + \text{H}_2\text{O}_2$
Other name(s): THOX2; ThOX; dual oxidase; p138tox; thyroid NADPH oxidase; thyroid oxidase; thyroid oxidase 2; NADPH oxidase; NAD(P)H:oxygen oxidoreductase; NAD(P)H oxidase
Systematic name: NAD(P)H:oxygen oxidoreductase (H₂O₂-forming)
Comments: Requires FAD, heme and calcium. When calcium is present, this transmembrane glycoprotein generates H₂O₂ by transferring electrons from intracellular NAD(P)H to extracellular molecular oxygen. The electron bridge within the enzyme contains one molecule of FAD and probably two heme groups. This flavoprotein is expressed at the apical membrane of thyrocytes, and provides H₂O₂ for the thyrocyte peroxidase-catalysed biosynthesis of thyroid hormones.
References: [2883, 864, 865, 987, 2421, 988]

[EC 1.6.3.1 created 2003, modified 2013]

EC 1.6.3.2

Accepted name: NAD(P)H oxidase (H₂O-forming)
Reaction: $2 \text{ NAD(P)H} + 2 \text{ H}^+ + \text{O}_2 = 2 \text{ NAD(P)}^+ + 2 \text{ H}_2\text{O}$
Systematic name: NAD(P)H:oxygen oxidoreductase (H₂O-forming)

Comments: A flavoprotein (FAD). NADPH is a better substrate than NADH [457, 1912]. By removal of oxygen the enzyme is involved in aerobic tolerance in the thermophilic anaerobic archaeon *Thermococcus profundus* and in *Giardia intestinalis*, a microaerophilic single-celled parasite of the order Diplomonadida.

References: [457, 2450, 1912, 1911]

[EC 1.6.3.2 created 2013]

EC 1.6.3.3

Accepted name: NADH oxidase (H₂O₂-forming)

Reaction: $\text{NADH} + \text{H}^+ + \text{O}_2 = \text{NAD}^+ + \text{H}_2\text{O}_2$

Other name(s): NOX-1; H₂O₂-forming NADH oxidase

Systematic name: NADH:oxygen oxidoreductase (H₂O₂-forming)

Comments: A flavoprotein (FAD). The bacterium *Streptococcus mutans* contains two distinct NADH oxidases, a H₂O₂-forming enzyme and a H₂O-forming enzyme (cf. EC 1.6.3.4, NADH oxidase (H₂O-forming)) [1648]. The enzymes from the anaerobic archaea *Methanocaldococcus jannaschii* [569] and *Pyrococcus furiosus* [2064] also produce low amounts of H₂O. Unlike EC 1.6.3.1 (NAD(P)H oxidase) it has no activity towards NADPH.

References: [1648, 4542, 2064, 4767, 1666, 569]

[EC 1.6.3.3 created 2013]

EC 1.6.3.4

Accepted name: NADH oxidase (H₂O-forming)

Reaction: $2 \text{NADH} + 2 \text{H}^+ + \text{O}_2 = 2 \text{NAD}^+ + 2 \text{H}_2\text{O}$

Other name(s): H₂O-forming NADH oxidase; Nox-2

Systematic name: NADH:oxygen oxidoreductase (H₂O-forming)

Comments: A flavoprotein (FAD). The bacterium *Streptococcus mutans* contains two distinct NADH oxidases, a H₂O-forming enzyme and a H₂O₂-forming enzyme (cf. EC 1.6.3.3, NADH oxidase (H₂O₂-forming)) [3729].

References: [3729, 1648, 2701, 2047, 4889]

[EC 1.6.3.4 created 2013]

EC 1.6.3.5

Accepted name: renalase

Reaction: (1) $1,2\text{-dihydro-}\beta\text{-NAD(P)} + \text{H}^+ + \text{O}_2 = \beta\text{-NAD(P)}^+ + \text{H}_2\text{O}_2$

(2) $1,6\text{-dihydro-}\beta\text{-NAD(P)} + \text{H}^+ + \text{O}_2 = \beta\text{-NAD(P)}^+ + \text{H}_2\text{O}_2$

Other name(s): α NAD(P)H oxidase/anomerase (incorrect); NAD(P)H:oxygen oxidoreductase (H₂O₂-forming, epimerising) (incorrect)

Systematic name: dihydro-NAD(P):oxygen oxidoreductase (H₂O₂-forming)

Comments: Requires FAD. Renalase, previously thought to be a hormone, is a flavoprotein secreted into the blood by the kidney that oxidizes the 1,2-dihydro- and 1,6-dihydro- isomeric forms of β -NAD(P)H back to β -NAD(P)⁺. These isomeric forms, generated by nonspecific reduction of β -NAD(P)⁺ or by tautomerization of β -NAD(P)H, are potent inhibitors of primary metabolism dehydrogenases and pose a threat to normal respiration.

References: [4704, 261]

[EC 1.6.3.5 created 2014, modified 2015]

EC 1.6.4 With a disulfide as acceptor (deleted sub-subclass)

- [1.6.4.1 *Transferred entry. cystine reductase (NADH). Now EC 1.8.1.6, cystine reductase*]
[EC 1.6.4.1 created 1961, deleted 2002]
- [1.6.4.2 *Transferred entry. glutathione reductase (NADPH). Now EC 1.8.1.7, glutathione-disulfide reductase*]
[EC 1.6.4.2 created 1961, modified 1989, deleted 2002]
- [1.6.4.3 *Transferred entry. dihydrolipoamide reductase (NAD⁺). Now EC 1.8.1.4, dihydrolipoyl dehydrogenase*]
[EC 1.6.4.3 created 1961, modified 1976, deleted 1983]
- [1.6.4.4 *Transferred entry. protein-disulfide reductase [NAD(P)H]. Now EC 1.8.1.8, protein-disulfide reductase*]
[EC 1.6.4.4 created 1965, deleted 2002]
- [1.6.4.5 *Transferred entry. thioredoxin reductase (NADPH). Now EC 1.8.1.9, thioredoxin-disulfide reductase*]
[EC 1.6.4.5 created 1972, deleted 2002]
- [1.6.4.6 *Transferred entry. CoA-glutathione reductase (NADPH). Now EC 1.8.1.10, CoA-glutathione reductase*]
[EC 1.6.4.6 created 1972, deleted 2002]
- [1.6.4.7 *Transferred entry. asparaguate reductase (NADH). Now EC 1.8.1.11, asparaguate reductase*]
[EC 1.6.4.7 created 1978, deleted 2002]
- [1.6.4.8 *Transferred entry. trypanothione reductase. Now EC 1.8.1.12, trypanothione-disulfide reductase*]
[EC 1.6.4.8 created 1989, deleted 2002]
- [1.6.4.9 *Transferred entry. bis- γ -glutamylcystine reductase (NADPH). Now EC 1.8.1.13, bis- γ -glutamylcystine reductase*]
[EC 1.6.4.9 created 1992, deleted 2002]
- [1.6.4.10 *Transferred entry. CoA-disulfide reductase (NADH). Now EC 1.8.1.14, CoA-disulfide reductase*]
[EC 1.6.4.10 created 1992, deleted 2002]

EC 1.6.5 With a quinone or similar compound as acceptor

- [1.6.5.1 *Deleted entry. quinone reductase*]
[EC 1.6.5.1 created 1961, deleted 1965]

EC 1.6.5.2

- Accepted name:** NAD(P)H dehydrogenase (quinone)
Reaction: NAD(P)H + H⁺ + a quinone = NAD(P)⁺ + a hydroquinone
Other name(s): menadione reductase; phyloquinone reductase; quinone reductase; dehydrogenase, reduced nicotinamide adenine dinucleotide (phosphate, quinone); DT-diaphorase; flavoprotein NAD(P)H-quinone reductase; menadione oxidoreductase; NAD(P)H dehydrogenase; NAD(P)H menadione reductase; NAD(P)H-quinone dehydrogenase; NAD(P)H-quinone oxidoreductase; NAD(P)H: (quinone-acceptor)oxidoreductase; NAD(P)H: menadione oxidoreductase; NADH-menadione reductase; naphthoquinone reductase; *p*-benzoquinone reductase; reduced NAD(P)H dehydrogenase; viologen accepting pyridine nucleotide oxidoreductase; vitamin K reductase; diaphorase; reduced nicotinamide-adenine dinucleotide (phosphate) dehydrogenase; vitamin-K reductase; NAD(P)H₂ dehydrogenase (quinone); NQO1; QR1; NAD(P)H:(quinone-acceptor) oxidoreductase
Systematic name: NAD(P)H:quinone oxidoreductase

Comments: A flavoprotein. The enzyme catalyses a two-electron reduction and has a preference for short-chain acceptor quinones, such as ubiquinone, benzoquinone, juglone and duroquinone [3972]. The animal, but not the plant, form of the enzyme is inhibited by dicoumarol.

References: [899, 1332, 2651, 2820, 4671, 3972, 427, 1878, 2454]

[EC 1.6.5.2 created 1961, transferred 1965 to EC 1.6.99.2, transferred 2005 to EC 1.6.5.2]

[1.6.5.3 *Transferred entry. NADH:ubiquinone reductase (H⁺-translocating). Now EC 7.1.1.2, NADH:ubiquinone reductase (H⁺-translocating)*]

[EC 1.6.5.3 created 1961, deleted 1965, reinstated 1983, modified 2011, modified 2013, deleted 2018]

EC 1.6.5.4

Accepted name: monodehydroascorbate reductase (NADH)

Reaction: NADH + H⁺ + 2 monodehydroascorbate = NAD⁺ + 2 ascorbate

Other name(s): NADH:semidehydroascorbic acid oxidoreductase; MDHA; semidehydroascorbate reductase; AFR (ambiguous); AFR-reductase; ascorbic free radical reductase; ascorbate free radical reductase; SOR (ambiguous); MDAsA reductase (NADPH); SDA reductase; NADH:ascorbate radical oxidoreductase; NADH-semidehydroascorbate oxidoreductase; ascorbate free-radical reductase; NADH:AFR oxidoreductase; monodehydroascorbate reductase (NADH₂)

Systematic name: NADH:monodehydroascorbate oxidoreductase

References: [3755]

[EC 1.6.5.4 created 1961]

EC 1.6.5.5

Accepted name: NADPH:quinone reductase

Reaction: NADPH + H⁺ + 2 quinone = NADP⁺ + 2 semiquinone

Other name(s): NADPH₂:quinone reductase

Systematic name: NADPH:quinone oxidoreductase

Comments: A zinc enzyme, specific for NADPH. Catalyses the one-electron reduction of certain quinones, with the orthoquinones 1,2-naphthoquinone and 9,10-phenanthrenequinone being the best substrates [3451]. Dicoumarol [*cf.* EC 1.6.5.2 NAD(P)H dehydrogenase (quinone)] and nitrofurantoin are competitive inhibitors with respect to the quinone substrate. The semiquinone free-radical product may be non-enzymically reduced to the hydroquinone or oxidized back to quinone in the presence of O₂ [3451]. In some mammals, the enzyme is abundant in the lens of the eye, where it is identified with the protein ζ-crystallin.

References: [3451, 979, 251, 4199]

[EC 1.6.5.5 created 1999]

EC 1.6.5.6

Accepted name: *p*-benzoquinone reductase (NADPH)

Reaction: NADPH + H⁺ + *p*-benzoquinone = NADP⁺ + hydroquinone

Systematic name: NADPH:*p*-benzoquinone oxidoreductase

Comments: Involved in the 4-nitrophenol degradation pathway in bacteria.

References: [3971]

[EC 1.6.5.6 created 2000]

EC 1.6.5.7

Accepted name: 2-hydroxy-1,4-benzoquinone reductase

Reaction: 2-hydroxy-1,4-benzoquinone + NADH + H⁺ = hydroxyquinol + NAD⁺

Other name(s): hydroxybenzoquinone reductase; 1,2,4-trihydroxybenzene:NAD oxidoreductase
Systematic name: NADH:2-hydroxy-1,4-benzoquinone oxidoreductase
Comments: A flavoprotein (FMN) that differs in substrate specificity from other quinone reductases. The enzyme in *Burkholderia cepacia* is inducible by 2,4,5-trichlorophenoxyacetate.
References: [4850]

[EC 1.6.5.7 created 2000, modified 2004]

[1.6.5.8 *Transferred entry. NADH:ubiquinone reductase (Na⁺-transporting). Now EC 7.2.1.1, NADH:ubiquinone reductase (Na⁺-transporting)*]

[EC 1.6.5.8 created 2011, deleted 2018]

EC 1.6.5.9

Accepted name: NADH:quinone reductase (non-electrogenic)
Reaction: NADH + H⁺ + a quinone = NAD⁺ + a quinol
Other name(s): type II NAD(P)H:quinone oxidoreductase; NDE2 (gene name); *ndh* (gene name); NDH-II; NDH-2; NADH dehydrogenase (quinone) (ambiguous); ubiquinone reductase (ambiguous); coenzyme Q reductase (ambiguous); dihydronicotinamide adenine dinucleotide-coenzyme Q reductase (ambiguous); DPNH-coenzyme Q reductase (ambiguous); DPNH-ubiquinone reductase (ambiguous); NADH-coenzyme Q oxidoreductase (ambiguous); NADH-coenzyme Q reductase (ambiguous); NADH-CoQ oxidoreductase (ambiguous); NADH-CoQ reductase (ambiguous); NADH-ubiquinone reductase (ambiguous); NADH-ubiquinone oxidoreductase (ambiguous); reduced nicotinamide adenine dinucleotide-coenzyme Q reductase (ambiguous); NADH-Q6 oxidoreductase (ambiguous); NADH₂ dehydrogenase (ubiquinone) (ambiguous); NADH:ubiquinone oxidoreductase; NADH:ubiquinone reductase (non-electrogenic)
Systematic name: NADH:quinone oxidoreductase
Comments: A flavoprotein (FAD or FMN). Occurs in mitochondria of yeast and plants, and in aerobic bacteria. Has low activity with NADPH. Unlike EC 7.1.1.2, NADH:ubiquinone reductase (H⁺-translocating), this enzyme does not pump protons or sodium ions across the membrane. It is also not sensitive to rotenone.
References: [300, 2864, 855, 2067, 3454, 2768]

[EC 1.6.5.9 created 2011 (EC 1.6.5.11 created 1972 as EC 1.6.99.5, transferred 2015 to EC 1.6.5.11, incorporated 2019), modified 2019]

EC 1.6.5.10

Accepted name: NADPH dehydrogenase (quinone)
Reaction: NADPH + H⁺ + a quinone = NADP⁺ + a quinol
Other name(s): reduced nicotinamide adenine dinucleotide phosphate (quinone) dehydrogenase; NADPH oxidase; NADPH₂ dehydrogenase (quinone)
Systematic name: NADPH:(quinone-acceptor) oxidoreductase
Comments: A flavoprotein [1, 2]. The enzyme from *Escherichia coli* is specific for NADPH and is most active with quinone derivatives and ferricyanide as electron acceptors [1580]. Menaquinone can act as acceptor. The enzyme from hog liver is inhibited by dicoumarol and folic acid derivatives but not by 2,4-dinitrophenol [2212].
References: [2212, 1579, 1580]

[EC 1.6.5.10 created 1972 as EC 1.6.99.6, transferred 2011 to EC 1.6.5.10]

[1.6.5.11 *Deleted entry. NADH dehydrogenase (quinone). Identical to EC 1.6.5.9, NADH:quinone reductase (non-electrogenic)*]

[EC 1.6.5.11 created 1972 as EC 1.6.99.5, transferred 2015 to EC 1.6.5.11, deleted 2019]

EC 1.6.5.12

Accepted name: demethylphyloquinone reductase

Reaction: demethylphyloquinone + NADPH + H⁺ = demethylphyloquinol + NADP⁺
Other name(s): *ndbB* (gene name); NDC1 (gene name); demethylphyloquinone:NADPH oxidoreductase
Systematic name: NADPH:demethylphyloquinone oxidoreductase
Comments: The enzyme, found in plants and cyanobacteria, is involved in the biosynthesis of phyloquinone (vitamin K₁), an electron carrier associated with photosystem I. The enzyme is a type II NADPH dehydrogenase and requires a flavine adenine dinucleotide cofactor.
References: [1095]

[EC 1.6.5.12 created 2015]

EC 1.6.6 With a nitrogenous group as acceptor

[1.6.6.1 *Transferred entry. nitrate reductase (NADH). Now EC 1.7.1.1, nitrate reductase (NADH)*]

[EC 1.6.6.1 created 1961, deleted 2002]

[1.6.6.2 *Transferred entry. nitrate reductase [NAD(P)H]. Now EC 1.7.1.2, nitrate reductase [NAD(P)H]*]

[EC 1.6.6.2 created 1961, deleted 2002]

[1.6.6.3 *Transferred entry. nitrate reductase (NADPH). Now EC 1.7.1.3, nitrate reductase (NADPH)*]

[EC 1.6.6.3 created 1961, deleted 2002]

[1.6.6.4 *Transferred entry. nitrite reductase [NAD(P)H]. Now EC 1.7.1.4, nitrite reductase [NAD(P)H]*]

[EC 1.6.6.4 created 1961, deleted 2002]

[1.6.6.5 *Transferred entry. now EC 1.7.2.1, nitrite reductase (NO-forming)*]

[EC 1.6.6.5 created 1961, deleted 1964]

[1.6.6.6 *Transferred entry. hyponitrite reductase. Now EC 1.7.1.5, hyponitrite reductase*]

[EC 1.6.6.6 created 1961, deleted 2002]

[1.6.6.7 *Transferred entry. azobenzene reductase. Now EC 1.7.1.6, azobenzene reductase*]

[EC 1.6.6.7 created 1961, deleted 2002]

[1.6.6.8 *Transferred entry. GMP reductase. Now EC 1.7.1.7, GMP reductase*]

[EC 1.6.6.8 created 1965, deleted 2002]

[1.6.6.9 *Deleted entry. The activity is now known to be catalysed by EC 1.7.2.3, trimethylamine-N-oxide reductase.*]

[EC 1.6.6.9 created 1972, deleted 2018]

[1.6.6.10 *Transferred entry. nitroquinoline-N-oxide reductase. Now EC 1.7.1.9, nitroquinoline-N-oxide reductase*]

[EC 1.6.6.10 created 1972, deleted 2002]

[1.6.6.11 *Transferred entry. hydroxylamine reductase (NADH). Now EC 1.7.1.10, hydroxylamine reductase (NADH)*]

[EC 1.6.6.11 created 1972, deleted 2002]

[1.6.6.12 *Transferred entry. 4-(dimethylamino)phenylazoxybenzene reductase. Now EC 1.7.1.11, 4-(dimethylamino)phenylazoxybenzene reductase*]

[EC 1.6.6.12 created 1989, deleted 2002]

[1.6.6.13 *Transferred entry. N-hydroxy-2-acetamidofluorene reductase. Now EC 1.7.1.12, N-hydroxy-2-acetamidofluorene reductase*]

[EC 1.6.6.13 created 1989, deleted 2002]

EC 1.6.7 With an iron-sulfur protein as acceptor (deleted sub-subclass)

[1.6.7.1 Transferred entry. *ferredoxin—NADP⁺ reductase*. Now EC 1.18.1.2, *ferredoxin—NADP⁺ reductase*]

[EC 1.6.7.1 created 1972, deleted 1978]

[1.6.7.2 Transferred entry. *rubredoxin—NAD⁺ reductase*. Now EC 1.18.1.1, *rubredoxin—NAD⁺ reductase*]

[EC 1.6.7.2 created 1972, deleted 1978]

[1.6.7.3 Transferred entry. now EC 1.18.1.3, *ferredoxin—NAD⁺ reductase*]

[EC 1.6.7.3 created 1978, deleted 1978]

EC 1.6.8 With a flavin as acceptor (deleted sub-subclass)

[1.6.8.1 Transferred entry. *NAD(P)H dehydrogenase (FMN)*. Now EC 1.5.1.29, *FMN reductase*]

[EC 1.6.8.1 created 1981, deleted 2002]

[1.6.8.2 Transferred entry. *NADPH dehydrogenase (flavin)*. Now EC 1.5.1.30, *flavin reductase*]

[EC 1.6.8.2 created 1982, deleted 2002]

EC 1.6.99 With unknown physiological acceptors

EC 1.6.99.1

Accepted name: NADPH dehydrogenase

Reaction: NADPH + H⁺ + acceptor = NADP⁺ + reduced acceptor

Other name(s): NADPH₂ diaphorase; NADPH diaphorase; OYE; diaphorase; dihydronicotinamide adenine dinucleotide phosphate dehydrogenase; NADPH-dehydrogenase; NADPH-diaphorase; NADPH₂-dehydrogenase; old yellow enzyme; reduced nicotinamide adenine dinucleotide phosphate dehydrogenase; TPNH dehydrogenase; TPNH-diaphorase; triphosphopyridine diaphorase; triphosphopyridine nucleotide diaphorase; NADPH₂ dehydrogenase; NADPH:(acceptor) oxidoreductase

Systematic name: NADPH:acceptor oxidoreductase

Comments: A flavoprotein (FMN in yeast, FAD in plants).

References: [50, 162, 1876, 4255, 4258]

[EC 1.6.99.1 created 1961, modified 1976]

[1.6.99.2 Transferred entry. *NAD(P)H dehydrogenase (quinone)*. Now EC 1.6.5.2, *NAD(P)H dehydrogenase (quinone)*. The enzyme was erroneously transferred from this sub-subclass in 1965]

[EC 1.6.99.2 created 1961 as EC 1.6.5.2, transferred 1965 to EC 1.6.99.2, deleted 2005]

[1.6.99.3 Deleted entry. *NADH dehydrogenase*. The activity is covered by EC 7.1.1.2, *NADH:ubiquinone reductase (H⁺-translocating)*]

[EC 1.6.99.3 created 1961 as EC 1.6.2.1, transferred 1965 to EC 1.6.99.3, modified 2018, deleted 2020]

[1.6.99.4 Transferred entry. *nitrite reductase*. Now EC 1.18.1.2, *ferredoxin—NADP⁺ reductase*]

[EC 1.6.99.4 created 1965, deleted 1972]

[1.6.99.5 Transferred entry. *NADH dehydrogenase (quinone)*. Transferred to EC 1.6.5.11, *NADH dehydrogenase (quinone)*]

[EC 1.6.99.5 created 1972, deleted 2014]

[1.6.99.6 Transferred entry. *NADPH dehydrogenase (quinone)*. Now EC 1.6.5.10, *NADPH dehydrogenase (quinone)*]

[EC 1.6.99.6 created 1972, deleted 2011]

[1.6.99.7 *Transferred entry. dihydropteridine reductase. Now EC 1.5.1.34, 6,7-dihydropteridine reductase*]

[EC 1.6.99.7 created 1972, modified 1981 (EC 1.6.99.10 created 1978, incorporated 1981), deleted 2003]

[1.6.99.8 *Deleted entry. aquacobalamin reductase. This entry has been deleted since no specific enzyme catalysing this activity has been identified and it has been shown that aquacobalamin is efficiently reduced by free dihydroflavins and by non-specific reduced flavoproteins.*]

[EC 1.6.99.8 created 1972, deleted 2002]

[1.6.99.9 *Transferred entry. cob(II)alamin reductase. Now EC 1.16.1.4, cob(II)alamin reductase*]

[EC 1.6.99.9 created 1972, deleted 2002]

[1.6.99.10 *Deleted entry. dihydropteridine reductase (NADH). Now included with EC 1.5.1.34, 6,7-dihydropteridine reductase*]

[EC 1.6.99.10 created 1978, deleted 1981]

[1.6.99.11 *Deleted entry. aquacobalamin reductase (NADPH). This entry has been deleted since the enzyme the entry was based on was later shown to be EC 1.2.1.51, pyruvate dehydrogenase (NADP⁺). On the other hand, it has been shown that non-enzymatic reduction of cob(III)alamin to cob(II)alamin occurs efficiently in the presence of free dihydroflavins or non-specific reduced flavoproteins.*]

[EC 1.6.99.11 created 1989, deleted 2002]

[1.6.99.12 *Transferred entry. cyanocobalamin reductase (NADPH, cyanide-eliminating). Now EC 1.16.1.6, cyanocobalamin reductase (cyanide-eliminating)*]

[EC 1.6.99.12 created 1989, deleted 2002]

[1.6.99.13 *Transferred entry. ferric-chelate reductase. Now EC 1.16.1.7, ferric-chelate reductase*]

[EC 1.6.99.13 created 1992, deleted 2002]

EC 1.7 Acting on other nitrogenous compounds as donors

This subclass contains a small group of enzymes that oxidize diverse nitrogenous substrates. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.7.1), a cytochrome (EC 1.7.2), oxygen (EC 1.7.3), an iron-sulfur protein (EC 1.7.7), or some other acceptor (EC 1.7.99).

EC 1.7.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.7.1.1

- Accepted name:** nitrate reductase (NADH)
Reaction: nitrite + NAD⁺ + H₂O = nitrate + NADH + H⁺
Other name(s): assimilatory nitrate reductase (ambiguous); NADH-nitrate reductase; NADH-dependent nitrate reductase; assimilatory NADH: nitrate reductase; nitrate reductase (NADH₂); NADH₂:nitrate oxidoreductase
Systematic name: nitrite:NAD⁺ oxidoreductase
Comments: An iron-sulfur molybdenum flavoprotein.
References: [1114, 3019, 3063, 3977, 301]

[EC 1.7.1.1 created 1961 as EC 1.6.6.1, transferred 2002 to EC 1.7.1.1]

EC 1.7.1.2

Accepted name: nitrate reductase [NAD(P)H]
Reaction: nitrite + NAD(P)⁺ + H₂O = nitrate + NAD(P)H + H⁺
Other name(s): assimilatory nitrate reductase (ambiguous); assimilatory NAD(P)H-nitrate reductase; NAD(P)H bispecific nitrate reductase; nitrate reductase (reduced nicotinamide adenine dinucleotide (phosphate)); nitrate reductase NAD(P)H; NAD(P)H-nitrate reductase; nitrate reductase [NAD(P)H₂]; NAD(P)H₂:nitrate oxidoreductase
Systematic name: nitrite:NAD(P)⁺ oxidoreductase
Comments: An iron-sulfur molybdenum flavoprotein.
References: [3019, 3230, 541, 301]

[EC 1.7.1.2 created 1961 as EC 1.6.6.2, transferred 2002 to EC 1.7.1.2]

EC 1.7.1.3

Accepted name: nitrate reductase (NADPH)
Reaction: nitrite + NADP⁺ + H₂O = nitrate + NADPH + H⁺
Other name(s): assimilatory nitrate reductase (ambiguous); assimilatory reduced nicotinamide adenine dinucleotide phosphate-nitrate reductase; NADPH-nitrate reductase; assimilatory NADPH-nitrate reductase; triphosphopyridine nucleotide-nitrate reductase; NADPH:nitrate reductase; nitrate reductase (NADPH₂); NADPH₂:nitrate oxidoreductase
Systematic name: nitrite:NADP⁺ oxidoreductase
Comments: An iron-sulfur molybdenum flavoprotein.
References: [3019, 3020, 3062, 4209, 301]

[EC 1.7.1.3 created 1961 as EC 1.6.6.3, transferred 2002 to EC 1.7.1.3]

EC 1.7.1.4

Accepted name: nitrite reductase [NAD(P)H]
Reaction: NH₃ + 3 NAD(P)⁺ + 2 H₂O = nitrite + 3 NAD(P)H + 5 H⁺
Other name(s): nitrite reductase (reduced nicotinamide adenine dinucleotide (phosphate)); assimilatory nitrite reductase (ambiguous); nitrite reductase [NAD(P)H₂]; NAD(P)H₂:nitrite oxidoreductase; nit-6 (gene name)
Systematic name: ammonia:NAD(P)⁺ oxidoreductase
Comments: An iron-sulfur flavoprotein (FAD) containing siroheme. The enzymes from the fungi *Neurospora crassa* [3061], *Emericella nidulans* [3260] and *Candida nitratophila* [2326] can use either NADPH or NADH as electron donor. cf. EC 1.7.1.15, nitrite reductase (NADH).
References: [3061, 3260, 3529, 2326, 4427, 1398, 3386, 1074, 708]

[EC 1.7.1.4 created 1961 as EC 1.6.6.4, transferred 2002 to EC 1.7.1.4, modified 2013]

EC 1.7.1.5

Accepted name: hyponitrite reductase
Reaction: 2 hydroxylamine + 2 NAD⁺ = hyponitrous acid + 2 NADH + 2 H⁺
Other name(s): NADH₂:hyponitrite oxidoreductase
Systematic name: hydroxylamine:NAD⁺ oxidoreductase
Comments: A metalloprotein.
References: [2758]

[EC 1.7.1.5 created 1961 as EC 1.6.6.6, transferred 2002 to EC 1.7.1.5]

EC 1.7.1.6

Accepted name: azobenzene reductase
Reaction: *N,N*-dimethyl-1,4-phenylenediamine + aniline + 2 NADP⁺ = 4-(dimethylamino)azobenzene + 2 NADPH + 2 H⁺

Other name(s): new coccine (NC)-reductase; NC-reductase; azo-dye reductase; orange II azoreductase; NAD(P)H:1-(4'-sulfophenylazo)-2-naphthol oxidoreductase; orange I azoreductase; azoreductase; azoreductase; nicotinamide adenine dinucleotide (phosphate) azoreductase; NADPH₂-dependent azoreductase; dimethylaminobenzene reductase; *p*-dimethylaminoazobenzene azoreductase; dibromopropylaminophenylazobenzoic azoreductase; *N,N*-dimethyl-4-phenylazoaniline azoreductase; *p*-aminoazobenzene reductase; methyl red azoreductase; NADPH₂:4-(dimethylamino)azobenzene oxidoreductase

Systematic name: *N,N*-dimethyl-1,4-phenylenediamine, aniline:NADP⁺ oxidoreductase

Comments: The reaction occurs in the reverse direction to that shown above. Other azo dyes, such as Methyl Red, Rocceline, Solar Orange and Sumifix Black B can also be reduced [4145].

References: [2914, 4145]

[EC 1.7.1.6 created 1961 as EC 1.6.6.7, transferred 2002 to EC 1.7.1.6]

EC 1.7.1.7

Accepted name: GMP reductase

Reaction: $\text{IMP} + \text{NH}_3 + \text{NADP}^+ = \text{GMP} + \text{NADPH} + \text{H}^+$

Other name(s): guanosine 5'-monophosphate reductase; NADPH:GMP oxidoreductase (deaminating); guanosine monophosphate reductase; guanylate reductase; NADPH₂:guanosine-5'-phosphate oxidoreductase (deaminating); guanosine 5'-phosphate reductase

Systematic name: inosine-5'-phosphate:NADP⁺ oxidoreductase (aminating)

References: [2593, 2608]

[EC 1.7.1.7 created 1965 as EC 1.6.6.8, transferred 2002 to EC 1.7.1.7]

[1.7.1.8 Deleted entry. withdrawn in the light of further information on the acceptor]

[EC 1.7.1.8 created 2002, deleted 2002]

EC 1.7.1.9

Accepted name: nitroquinoline-*N*-oxide reductase

Reaction: $4\text{-(hydroxyamino)quinoline } N\text{-oxide} + 2 \text{ NAD(P)}^+ + \text{H}_2\text{O} = 4\text{-nitroquinoline } N\text{-oxide} + 2 \text{ NAD(P)H} + 2 \text{ H}^+$

Other name(s): 4-nitroquinoline 1-oxide reductase; 4NQO reductase; NAD(P)H₂:4-nitroquinoline-*N*-oxide oxidoreductase

Systematic name: 4-(hydroxyamino)quinoline *N*-oxide:NADP⁺ oxidoreductase

References: [4313, 4003]

[EC 1.7.1.9 created 1972 as EC 1.6.6.10, transferred 2002 to EC 1.7.1.9]

EC 1.7.1.10

Accepted name: hydroxylamine reductase (NADH)

Reaction: $\text{NH}_3 + \text{NAD}^+ + \text{H}_2\text{O} = \text{hydroxylamine} + \text{NADH} + \text{H}^+$

Other name(s): hydroxylamine reductase; ammonium dehydrogenase; NADH-hydroxylamine reductase; *N*-hydroxylamine reductase; hydroxylamine reductase (NADH₂); NADH₂:hydroxylamine oxidoreductase

Systematic name: ammonium:NAD⁺ oxidoreductase

Comments: Also acts on some hydroxamates.

References: [306, 307, 4528]

[EC 1.7.1.10 created 1972 as EC 1.6.6.11, transferred 2002 to EC 1.7.1.10]

EC 1.7.1.11

Accepted name: 4-(dimethylamino)phenylazoxybenzene reductase

Reaction: 4-(dimethylamino)phenylazobenzene + NADP⁺ + H₂O = 4-(dimethylamino)phenylazoxybenzene + NADPH + H⁺
Other name(s): *N,N*-dimethyl-*p*-aminoazobenzene oxide reductase; dimethylaminoazobenzene *N*-oxide reductase; NADPH-dependent DMAB *N*-oxide reductase; NADPH:4-(dimethylamino)phenylazoxybenzene oxidoreductase
Systematic name: 4-(dimethylamino)phenylazobenzene:NADP⁺ oxidoreductase
References: [1936]

[EC 1.7.1.11 created 1989 as EC 1.6.6.12, transferred 2002 to EC 1.7.1.11]

EC 1.7.1.12

Accepted name: *N*-hydroxy-2-acetamidofluorene reductase
Reaction: 2-acetamidofluorene + NAD(P)⁺ + H₂O = *N*-hydroxy-2-acetamidofluorene + NAD(P)H + H⁺
Other name(s): *N*-hydroxy-2-acetylaminofluorene reductase; NAD(P)H₂:*N*-hydroxy-2-acetamidofluorene *N*-oxidoreductase
Systematic name: 2-acetamidofluorene:NAD(P)⁺ oxidoreductase
Comments: Also acts, more slowly, on *N*-hydroxy-4-acetamidobiphenyl.
References: [1458, 2134]

[EC 1.7.1.12 created 1989 as EC 1.6.6.13, transferred 2002 to EC 1.7.1.12]

EC 1.7.1.13

Accepted name: preQ₁ synthase
Reaction: 7-aminomethyl-7-carbaguanine + 2 NADP⁺ = 7-cyano-7-carbaguanine + 2 NADPH + 2 H⁺
Other name(s): YkvM; QueF; preQ₀ reductase; preQ₀ oxidoreductase; 7-cyano-7-deazaguanine reductase; queuine synthase (incorrect as queuine is not the product); queuine:NADP⁺ oxidoreductase (incorrect as queuine is not the product)
Systematic name: 7-aminomethyl-7-carbaguanine:NADP⁺ oxidoreductase
Comments: The reaction occurs in the reverse direction. This enzyme catalyses one of the early steps in the synthesis of queuosine (Q-tRNA), and is followed by the action of EC 2.4.2.29, tRNA-guanosine³⁴ transglycosylase. Queuosine is found in the wobble position of tRNA_{GUN} in Eukarya and Bacteria [4797] and is thought to be involved in translational modulation. The enzyme is not a GTP cyclohydrolase, as was thought previously based on sequence-homology studies.
References: [2341, 4797, 2273, 3153, 3097, 4147]

[EC 1.7.1.13 created 2006]

EC 1.7.1.14

Accepted name: nitric oxide reductase [NAD(P)⁺, nitrous oxide-forming]
Reaction: N₂O + NAD(P)⁺ + H₂O = 2 NO + NAD(P)H + H⁺
Other name(s): fungal nitric oxide reductase; cytochrome P450_{nor}; NOR (ambiguous)
Systematic name: nitrous oxide:NAD(P) oxidoreductase
Comments: A heme-thiolate protein (*P*-450). The enzyme from *Fusarium oxysporum* utilizes only NADH, but the isozyme from *Trichosporon cutaneum* utilizes both NADH and NADPH. The electron transfer from NAD(P)H to heme occurs directly, not requiring flavin or other redox cofactors.
References: [3892, 3889, 4879, 3196]

[EC 1.7.1.14 created 2011]

EC 1.7.1.15

Accepted name: nitrite reductase (NADH)
Reaction: NH₃ + 3 NAD⁺ + 2 H₂O = nitrite + 3 NADH + 5 H⁺

Other name(s): nitrite reductase (reduced nicotinamide adenine dinucleotide); NADH-nitrite oxidoreductase; assimilatory nitrite reductase (ambiguous); *nirB* (gene name); *nirD* (gene name)
Systematic name: ammonia:NAD⁺ oxidoreductase
Comments: An iron-sulfur flavoprotein (FAD) containing siroheme. This prokaryotic enzyme is specific for NADH. In addition to catalysing the 6-electron reduction of nitrite to ammonia, the enzyme from *Escherichia coli* can also catalyse the 2-electron reduction of hydroxylamine to ammonia. *cf.* EC 1.7.1.4, nitrite reductase [NAD(P)H].
References: [4428, 1873, 538, 1530]

[EC 1.7.1.15 created 2013]

EC 1.7.1.16

Accepted name: nitrobenzene nitroreductase
Reaction: *N*-phenylhydroxylamine + 2 NADP⁺ + H₂O = nitrobenzene + 2 NADPH + 2 H⁺ (overall reaction)
(1a) *N*-phenylhydroxylamine + NADP⁺ = nitrosobenzene + NADPH + H⁺
(1b) nitrosobenzene + NADP⁺ + H₂O = nitrobenzene + NADPH + H⁺
Other name(s): *cnbA* (gene name)
Systematic name: *N*-phenylhydroxylamine:NADP⁺ oxidoreductase
Comments: Contains FMN. The enzyme, characterized from *Pseudomonas* species, catalyses two successive reductions of nitrobenzene, via a nitrosobenzene intermediate. It is also active on 1-chloro-4-nitrobenzene.
References: [3958, 4674]

[EC 1.7.1.16 created 2017]

EC 1.7.1.17

Accepted name: FMN-dependent NADH-azoreductase
Reaction: anthranilate + *N,N*-dimethyl-1,4-phenylenediamine + 2 NAD⁺ = 2-(4-dimethylaminophenyl)diazenylbenzoate + 2 NADH + 2 H⁺
Other name(s): *azoR* (gene name); NADH-azoreductase
Systematic name: *N,N*-dimethyl-1,4-phenylenediamine, anthranilate:NAD⁺ oxidoreductase
Comments: Requires FMN. The enzyme catalyses the reductive cleavage of an azo bond in aromatic azo compounds to form the corresponding amines. Does not accept NADPH. *cf.* EC 1.7.1.6, azobenzene reductase.
References: [2987, 1846, 1847, 2773]

[EC 1.7.1.17 created 2018]

EC 1.7.2 With a cytochrome as acceptor

EC 1.7.2.1

Accepted name: nitrite reductase (NO-forming)
Reaction: nitric oxide + H₂O + ferricytochrome *c* = nitrite + ferrocycytochrome *c* + 2 H⁺
Other name(s): cd-cytochrome nitrite reductase; [nitrite reductase (cytochrome)] [misleading, see comments.]; cytochrome *c*-551:O₂, NO₂⁺ oxidoreductase; cytochrome *cd*; cytochrome *cd*₁; hydroxylamine (acceptor) reductase; methyl viologen-nitrite reductase; nitrite reductase (cytochrome; NO-forming)
Systematic name: nitric-oxide:ferricytochrome-*c* oxidoreductase

- Comments:** The reaction is catalysed by two types of enzymes, found in the periplasm of denitrifying bacteria. One type comprises proteins containing multiple copper centres, the other a heme protein, cytochrome *cd*₁. Acceptors include *c*-type cytochromes such as cytochrome *c*-550 or cytochrome *c*-551 from *Paracoccus denitrificans* or *Pseudomonas aeruginosa*, and small blue copper proteins such as azurin and pseudoazurin. Cytochrome *cd*₁ also has oxidase and hydroxylamine reductase activities. May also catalyse the reaction of hydroxylamine reductase (EC 1.7.99.1) since this is a well-known activity of cytochrome *cd*₁.
- References:** [2839, 691, 4499, 3909, 2791, 1349, 4635, 1695, 4941, 1103, 4450]

[EC 1.7.2.1 created 1961, modified 1976, modified 2001, modified 2002 (EC 1.7.99.3 created 1961 as EC 1.6.6.5, transferred 1964 to EC 1.7.99.3, modified 1976, incorporated 2002, EC 1.9.3.2 created 1965, incorporated 2002)]

EC 1.7.2.2

- Accepted name:** nitrite reductase (cytochrome; ammonia-forming)
- Reaction:** $\text{NH}_3 + 2 \text{H}_2\text{O} + 6 \text{ ferricytochrome } c = \text{nitrite} + 6 \text{ ferrocycytochrome } c + 7 \text{ H}^+$
- Other name(s):** cytochrome *c* nitrite reductase; multiheme nitrite reductase
- Systematic name:** ammonia:ferricytochrome-*c* oxidoreductase
- Comments:** Found as a multiheme cytochrome in many bacteria. The enzyme from *Escherichia coli* contains five hemes *c* and requires Ca^{2+} . It also reduces nitric oxide and hydroxylamine to ammonia, and sulfite to sulfide.
- References:** [1030]

[EC 1.7.2.2 created 2001]

EC 1.7.2.3

- Accepted name:** trimethylamine-*N*-oxide reductase
- Reaction:** $\text{trimethylamine} + 2 \text{ (ferricytochrome } c\text{-subunit)} + \text{H}_2\text{O} = \text{trimethylamine } N\text{-oxide} + 2 \text{ (ferrocycytochrome } c\text{-subunit)} + 2 \text{ H}^+$
- Other name(s):** TMAO reductase; TOR; *torA* (gene name); *torZ* (gene name); *bisZ* (gene name); trimethylamine-*N*-oxide reductase (cytochrome *c*)
- Systematic name:** trimethylamine:cytochrome *c* oxidoreductase
- Comments:** Contains bis(molybdopterin guanine dinucleotide)molybdenum cofactor. The reductant is a membrane-bound multiheme cytochrome *c*. Also reduces dimethyl sulfoxide to dimethyl sulfide.
- References:** [125, 2161, 794, 1357, 4881, 4793]

[EC 1.7.2.3 created 2002, modified 2018]

EC 1.7.2.4

- Accepted name:** nitrous-oxide reductase
- Reaction:** $\text{nitrogen} + \text{H}_2\text{O} + 2 \text{ ferricytochrome } c = \text{nitrous oxide} + 2 \text{ ferrocycytochrome } c + 2 \text{ H}^+$
- Other name(s):** nitrous oxide reductase; N_2O reductase; nitrogen:(acceptor) oxidoreductase (N_2O -forming)
- Systematic name:** nitrogen:cytochrome *c* oxidoreductase (N_2O -forming)
- Comments:** The reaction is observed only in the direction of nitrous oxide reduction. Contains the mixed-valent dinuclear CuA species at the electron entry site of the enzyme, and the tetranuclear Cu-Z centre in the active site. In *Paracoccus pantotrophus*, the electron donor is cytochrome *c*₅₅₂.
- References:** [758, 4942, 873]

[EC 1.7.2.4 created 1989 as EC 1.7.99.6, modified 1999, transferred 2011 to EC 1.7.2.4]

EC 1.7.2.5

- Accepted name:** nitric oxide reductase (cytochrome *c*)
- Reaction:** $\text{nitrous oxide} + 2 \text{ ferricytochrome } c + \text{H}_2\text{O} = 2 \text{ nitric oxide} + 2 \text{ ferrocycytochrome } c + 2 \text{ H}^+$
- Systematic name:** nitrous oxide:ferricytochrome-*c* oxidoreductase

Comments: The enzyme from *Pseudomonas aeruginosa* contains a dinuclear centre comprising a non-heme iron centre and heme *b*₃, plus heme *c*, heme *b* and calcium; the acceptor is cytochrome *c*₅₅₁
References: [1626, 1625, 1614, 624, 2286, 1663]

[EC 1.7.2.5 created 1992 as EC 1.7.99.7, transferred 2011 to EC 1.7.2.5]

EC 1.7.2.6

Accepted name: hydroxylamine dehydrogenase
Reaction: hydroxylamine + H₂O + 4 ferricytochrome *c* = nitrite + 4 ferrocycytochrome *c* + 5 H⁺
Other name(s): HAO (ambiguous); hydroxylamine oxidoreductase (ambiguous); hydroxylamine oxidase (misleading)
Systematic name: hydroxylamine:ferricytochrome-*c* oxidoreductase (nitrite-forming)
Comments: The enzymes from the nitrifying bacterium *Nitrosomonas europaea* [3480, 2505] and the methylotrophic bacterium *Methylococcus capsulatus* [3354] are hemoproteins with seven *c*-type hemes and one specialized *P*-460-type heme per subunit. The enzyme converts hydroxylamine to nitrite via an enzyme-bound nitroxyl intermediate [1710]. While nitrite is the main product, the enzyme from *Nitrosomonas europaea* can also produce nitric oxide by catalysing the activity of EC 1.7.2.9, hydroxylamine oxidase [1711].
References: [3480, 1711, 1710, 2505, 3354]

[EC 1.7.2.6 created 1972 as EC 1.7.3.4, part transferred 2012 to EC 1.7.2.6, modified 2021, modified 2021]

EC 1.7.2.7

Accepted name: hydrazine synthase
Reaction: hydrazine + H₂O + 3 ferricytochrome *c* = nitric oxide + ammonium + 3 ferrocycytochrome *c*
Other name(s): HZS
Systematic name: hydrazine:ferricytochrome-*c* oxidoreductase
Comments: The enzyme, characterized from anaerobic ammonia oxidizers (anammox bacteria), is one of only a few enzymes that are known to form an N-N bond (other examples include EC 1.7.1.14, nitric oxide reductase [NAD(P)⁺, nitrous oxide-forming] and EC 4.8.1.1, L-piperazate synthase). The enzyme from the bacterium *Candidatus Kuenenia stuttgartiensis* is a dimer of heterotrimers and contains multiple *c*-type cytochromes.
References: [2002, 910]

[EC 1.7.2.7 created 2016, modified 2021]

EC 1.7.2.8

Accepted name: hydrazine dehydrogenase
Reaction: hydrazine + 4 ferricytochrome *c* = N₂ + 4 ferrocycytochrome *c*
Other name(s): HDH
Systematic name: hydrazine:ferricytochrome *c* oxidoreductase
Comments: The enzyme, which is involved in the pathway of anaerobic ammonium oxidation in anammox bacteria, has been purified from the bacterium *Candidatus Kuenenia stuttgartiensis*. The electrons derived from hydrazine are eventually transferred to the quinone pool.
References: [3701, 1909, 2002, 2001]

[EC 1.7.2.8 created 2003 as EC 1.7.99.8, modified 2010, transferred 2016 to EC 1.7.2.8]

EC 1.7.2.9

Accepted name: hydroxylamine oxidase
Reaction: hydroxylamine + 3 ferricytochrome *c* = nitric oxide + 3 ferrocycytochrome *c* + 3 H⁺
Other name(s): HOX
Systematic name: hydroxylamine:ferricytochrome-*c* oxidoreductase (nitric acid-forming)

Comments: The enzyme, characterized from the anaerobic ammonium-oxidizing (anammox) bacterium *Kueneinia stuttgartiensis*, is very similar to EC 1.7.2.6, hydroxylamine dehydrogenase. Both enzymes are homotrimeric enzymes in which each subunit contains seven *c*-type hemes and one specialized *P460*-type heme that is bound to a tyrosine residue in an adjacent subunit. However, this enzyme catalyses only the 3 electron oxidation of hydroxylamine, forming nitric oxide, and is not capable of performing further oxidation to form nitrite.

References: [2581]

[EC 1.7.2.9 created 2021]

EC 1.7.3 With oxygen as acceptor

EC 1.7.3.1

Accepted name: nitroalkane oxidase
Reaction: a nitroalkane + H₂O + O₂ = an aldehyde or ketone + nitrite + H₂O₂
Other name(s): nitroethane oxidase; NAO; nitroethane:oxygen oxidoreductase
Systematic name: nitroalkane:oxygen oxidoreductase
Comments: Has an absolute requirement for FAD [1130]. While nitroethane may be the physiological substrate [2086], the enzyme also acts on several other nitroalkanes, including 1-nitropropane, 2-nitropropane, 1-nitrobutane, 1-nitropentane, 1-nitrohexane, nitrocyclohexane and some nitroalkanols [1130]. Differs from EC 1.13.12.16, nitronate monooxygenase, in that the preferred substrates are neutral nitroalkanes rather than anionic nitronates [1130].
References: [2509, 2086, 833, 1130, 4392]

[EC 1.7.3.1 created 1961, modified 2006, modified 2009]

EC 1.7.3.2

Accepted name: acetyloxindoxyl oxidase
Reaction: *N*-acetyloxindoxyl + O₂ = *N*-acetylisatin + (?)
Systematic name: *N*-acetyloxindoxyl:oxygen oxidoreductase
References: [269]

[EC 1.7.3.2 created 1961]

EC 1.7.3.3

Accepted name: factor-independent urate hydroxylase
Reaction: urate + O₂ + H₂O = 5-hydroxyisourate + H₂O₂
Other name(s): uric acid oxidase; uricase; uricase II; urate oxidase
Systematic name: urate:oxygen oxidoreductase
Comments: This enzyme was previously thought to be a copper protein, but it is now known that the enzymes from soy bean (*Glycine max*), the mould *Aspergillus flavus* and *Bacillus subtilis* contains no copper nor any other transition-metal ion. The 5-hydroxyisourate formed decomposes spontaneously to form allantoin and CO₂, although there is an enzyme-catalysed pathway in which EC 3.5.2.17, hydroxyisourate hydrolase, catalyses the first step. The enzyme is different from EC 1.14.13.113 (FAD-dependent urate hydroxylase).
References: [2537, 2614, 3535, 1977, 714, 1806]

[EC 1.7.3.3 created 1961, modified 2002, modified 2005, modified 2010]

[1.7.3.4 Transferred entry. hydroxylamine oxidase. Now covered by EC 1.7.2.6, hydroxylamine dehydrogenase, and EC 1.7.3.6, hydroxylamine oxidase (cytochrome)]

[EC 1.7.3.4 created 1972, deleted 2013]

EC 1.7.3.5

- Accepted name:** 3-*aci*-nitropropanoate oxidase
Reaction: 3-*aci*-nitropropanoate + O₂ + H₂O = 3-oxopropanoate + nitrite + H₂O₂
Other name(s): propionate-3-nitronate oxidase
Systematic name: 3-*aci*-nitropropanoate:oxygen oxidoreductase
Comments: A flavoprotein (FMN). The primary products of the enzymic reaction are probably the nitropropanoate free radical and superoxide. Also acts, more slowly, on 4-*aci*-nitrobutanoate.
References: [3357]

[EC 1.7.3.5 created 1990]

EC 1.7.3.6

- Accepted name:** hydroxylamine oxidase (cytochrome)
Reaction: hydroxylamine + O₂ = nitrite + H₂O + H⁺ (overall reaction)
(1a) hydroxylamine + 2 ferricytochrome *c* = nitroxyl + 2 ferrocyclochrome *c* + 2 H⁺
(1b) nitroxyl + 2 ferrocyclochrome *c* + O₂ + H⁺ = nitrite + 2 ferricytochrome *c* + H₂O (spontaneous)
Other name(s): HAO (ambiguous); hydroxylamine oxidoreductase (ambiguous); hydroxylamine oxidase (misleading)
Systematic name: hydroxylamine:oxygen oxidoreductase
Comments: The enzyme from the heterotrophic nitrifying bacterium *Paracoccus denitrificans* contains three to five non-heme, non-iron-sulfur iron atoms and interacts with cytochrome *c*₅₅₆ and pseudoazurin [4573, 2861]. Under anaerobic conditions *in vitro* only nitrous oxide is formed [2861]. Presumably nitroxyl is released and combines with a second nitroxyl to give nitrous oxide and water. When oxygen is present, nitrite is formed.
References: [2302, 4573, 2861, 4572]

[EC 1.7.3.6 created 1972 as EC 1.7.3.4, part transferred 2013 to EC 1.7.3.6, modified 2015]

EC 1.7.5 With a quinone or similar compound as acceptor

EC 1.7.5.1

- Accepted name:** nitrate reductase (quinone)
Reaction: nitrate + a quinol = nitrite + a quinone + H₂O
Other name(s): nitrate reductase A; nitrate reductase Z; quinol/nitrate oxidoreductase; quinol-nitrate oxidoreductase; quinol:nitrate oxidoreductase; NarA; NarZ; NarGHI; dissimilatory nitrate reductase
Systematic name: nitrite:quinone oxidoreductase
Comments: A membrane-bound enzyme which supports anaerobic respiration on nitrate under anaerobic conditions and in the presence of nitrate. Contains the bicyclic form of the molybdo-bis(molybdopterin guanine dinucleotide) cofactor, iron-sulfur clusters and heme *b*. *Escherichia coli* expresses two forms NarA and NarZ, both being comprised of three subunits.
References: [1051, 313, 2337, 312, 384, 1440, 1822]

[EC 1.7.5.1 created 2010]

EC 1.7.5.2

- Accepted name:** nitric oxide reductase (menaquinol)
Reaction: 2 nitric oxide + menaquinol = nitrous oxide + menaquinone + H₂O
Comments: Contains copper.
References: [764, 4109, 4108]

[EC 1.7.5.2 created 2011]

EC 1.7.6 With a nitrogenous group as acceptor

EC 1.7.6.1

- Accepted name:** nitrite dismutase
Reaction: $3 \text{ nitrite} + 2 \text{ H}^+ = 2 \text{ nitric oxide} + \text{ nitrate} + \text{ H}_2\text{O}$
Other name(s): Prolixin S; Nitrophorin 7
Systematic name: nitrite:nitrite oxidoreductase
Comments: Contains ferriheme *b*. The enzyme is one of the nitrophorins from the salivary gland of the blood-feeding insect *Rhodnius prolixus*. Nitric oxide produced induces vasodilation after injection. Nitrophorins 2 and 4 can also catalyze this reaction.
References: [1585, 1586]

[EC 1.7.6.1 created 2011]

EC 1.7.7 With an iron-sulfur protein as acceptor

EC 1.7.7.1

- Accepted name:** ferredoxin—nitrite reductase
Reaction: $\text{NH}_3 + 2 \text{ H}_2\text{O} + 6 \text{ oxidized ferredoxin} = \text{ nitrite} + 6 \text{ reduced ferredoxin} + 7 \text{ H}^+$
Systematic name: ammonia:ferredoxin oxidoreductase
Comments: An iron protein. Contains siroheme and [4Fe-4S] clusters.
References: [1956, 3447, 4944]

[EC 1.7.7.1 created 1972, modified 1999]

EC 1.7.7.2

- Accepted name:** ferredoxin—nitrate reductase
Reaction: $\text{ nitrite} + \text{ H}_2\text{O} + 2 \text{ oxidized ferredoxin} = \text{ nitrate} + 2 \text{ reduced ferredoxin} + 2 \text{ H}^+$
Other name(s): assimilatory nitrate reductase (ambiguous); nitrate (ferredoxin) reductase; assimilatory ferredoxin-nitrate reductase
Systematic name: nitrite:ferredoxin oxidoreductase
Comments: A molybdenum-iron-sulfur protein.
References: [2803]

[EC 1.7.7.2 created 1986]

EC 1.7.99 With unknown physiological acceptors

EC 1.7.99.1

- Accepted name:** hydroxylamine reductase
Reaction: $\text{NH}_3 + \text{H}_2\text{O} + \text{acceptor} = \text{hydroxylamine} + \text{reduced acceptor}$
Other name(s): hydroxylamine (acceptor) reductase; ammonia:(acceptor) oxidoreductase
Systematic name: ammonia:acceptor oxidoreductase
Comments: A flavoprotein. Reduced pyocyanine, methylene blue and flavins act as donors for the reduction of hydroxylamine. May be identical to EC 1.7.2.1, nitrite reductase (NO-forming).
References: [4209, 4498, 3516]

[EC 1.7.99.1 created 1961, modified 1999, modified 2002]

[1.7.99.2 Deleted entry. nitric-oxide reductase. Reaction may have been due to the combined action of EC 1.7.99.6 nitrous-oxide reductase and EC 1.7.99.7 nitric-oxide reductase]

[EC 1.7.99.2 created 1961, modified 1976, deleted 1992]

[1.7.99.3 *Transferred entry. nitrite reductase. Now included with EC 1.7.2.1, nitrite reductase (NO-forming)*]

[EC 1.7.99.3 created 1961 as EC 1.6.6.5, transferred 1964 to EC 1.7.99.3, modified 1976, deleted 2002]

[1.7.99.4 *Transferred entry. nitrate reductase, Now EC 1.7.1.1, nitrate reductase (NADH), EC 1.7.1.2, nitrate reductase [NAD(P)H], EC 1.7.1.3, nitrate reductase (NADPH), EC 1.7.5.1, nitrate reductase (quinone), EC 1.7.7.2, nitrate reductase (ferredoxin) and EC 1.9.6.1, nitrate reductase (cytochrome)*]

[EC 1.7.99.4 created 1972, modified 1976, deleted 2017]

[1.7.99.5 *Deleted entry. 5,10-methylenetetrahydrofolate reductase (FADH₂). Now included with EC 1.5.1.20, methylenetetrahydrofolate reductase [NAD(P)H]. Based on the reference, it had been thought that this was a separate enzyme from EC 1.5.1.20 but the reference upon which the entry was based has since been disproved*]

[EC 1.7.99.5 created 1965 as EC 1.1.1.68, transferred 1978 to EC 1.1.99.15, transferred 1980 to EC 1.7.99.5, deleted 2005]

[1.7.99.6 *Transferred entry. EC 1.7.99.6, nitrous-oxide reductase. Now EC 1.7.2.4.*]

[EC 1.7.99.6 created 1989, modified 1999, deleted 2011]

[1.7.99.7 *Transferred entry. nitric-oxide reductase. Now EC 1.7.2.5 nitric oxide reductase (cytochrome c)*]

[EC 1.7.99.7 created 1992, modified 1999, deleted 2011]

[1.7.99.8 *Transferred entry. hydrazine oxidoreductase. Now classified as EC 1.7.2.8, hydrazine dehydrogenase.*]

[EC 1.7.99.8 created 2003, modified 2010, deleted 2016]

EC 1.8 Acting on a sulfur group of donors

This small subclass contains enzymes that act either on inorganic substrates or organic thiols. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.8.1), a cytochrome (EC 1.8.2), oxygen (EC 1.8.3), a disulfide (EC 1.8.4); a quinone or similar compound (EC 1.8.5), an iron-sulfur protein (EC 1.8.7), other, known, acceptors (EC 1.8.98), or some other acceptor (EC 1.8.99).

EC 1.8.1 With NAD⁺ or NADP⁺ as acceptor

[1.8.1.1 *Deleted entry. cysteamine dehydrogenase*]

[EC 1.8.1.1 created 1961, deleted 1972]

EC 1.8.1.2

Accepted name: assimilatory sulfite reductase (NADPH)

Reaction: hydrogen sulfide + 3 NADP⁺ + 3 H₂O = sulfite + 3 NADPH + 3 H⁺

Other name(s): sulfite reductase (NADPH); sulfite (reduced nicotinamide adenine dinucleotide phosphate) reductase; NADPH-sulfite reductase; NADPH-dependent sulfite reductase; H₂S-NADP oxidoreductase; sulfite reductase (NADPH₂); MET5 (gene name); MET10 (gene name); *cysI* (gene name); *cysJ* (gene name)

Systematic name: hydrogen-sulfide:NADP⁺ oxidoreductase

Comments: Contains siroheme, [4Fe-4S] cluster, FAD and FMN. The enzyme, which catalyses the six-electron reduction of sulfite to sulfide, is involved in sulfate assimilation in bacteria and yeast. Different from EC 1.8.99.5, dissimilatory sulfite reductase, which is involved in prokaryotic sulfur-based energy metabolism. *cf.* EC 1.8.7.1, assimilatory sulfite reductase (ferredoxin).

References: [1660, 4818, 3901, 2175, 3902, 756, 766]

[EC 1.8.1.2 created 1961, modified 2015]

[1.8.1.3 *Deleted entry. hypotaurine dehydrogenase. The reaction is now known to be catalyzed by EC 1.14.13.8, flavin-containing monooxygenase.*]

[EC 1.8.1.3 created 1972, deleted 2022]

EC 1.8.1.4

- Accepted name:** dihydrolipoyl dehydrogenase
Reaction: protein N^6 -(dihydrolipoyl)lysine + NAD^+ = protein N^6 -(lipoyl)lysine + $NADH + H^+$
Other name(s): LDP-Glc; LDP-Val; dehydrolipoate dehydrogenase; diaphorase; dihydrolipoamide dehydrogenase; dihydrolipoamide: NAD^+ oxidoreductase; dihydrolipoic dehydrogenase; dihydrothioctic dehydrogenase; lipoamide dehydrogenase ($NADH$); lipoamide oxidoreductase ($NADH$); lipoamide reductase; lipoamide reductase ($NADH$); lipoate dehydrogenase; lipoic acid dehydrogenase; lipoyl dehydrogenase; protein-6- N -(dihydrolipoyl)lysine: NAD^+ oxidoreductase
Systematic name: protein- N^6 -(dihydrolipoyl)lysine: NAD^+ oxidoreductase
Comments: A flavoprotein (FAD). A component of the multienzyme 2-oxo-acid dehydrogenase complexes. In the pyruvate dehydrogenase complex, it binds to the core of EC 2.3.1.12, dihydrolipoyllysine-residue acetyltransferase, and catalyses oxidation of its dihydrolipoyl groups. It plays a similar role in the oxoglutarate and 3-methyl-2-oxobutanoate dehydrogenase complexes. Another substrate is the dihydrolipoyl group in the H-protein of the glycine-cleavage system (click here for diagram), in which it acts, together with EC 1.4.4.2, glycine dehydrogenase (decarboxylating), and EC 2.1.2.10, aminomethyltransferase, to break down glycine. It can also use free dihydrolipoate, dihydrolipoamide or dihydrolipoyllysine as substrate. This enzyme was first shown to catalyse the oxidation of $NADH$ by methylene blue; this activity was called diaphorase. The glycine cleavage system is composed of four components that only loosely associate: the P protein (EC 1.4.4.2), the T protein (EC 2.1.2.10), the L protein (EC 1.8.1.4) and the lipoyl-bearing H protein [3043].
References: [2685, 2686, 3679, 4055, 3293, 3043]

[EC 1.8.1.4 created 1961 as EC 1.6.4.3, modified 1976, transferred 1983 to EC 1.8.1.4, modified 2003, modified 2006]

EC 1.8.1.5

- Accepted name:** 2-oxopropyl-CoM reductase (carboxylating)
Reaction: CoM + acetoacetate + $NADP^+$ = 2-oxopropyl-CoM + $CO_2 + NADPH$
Other name(s): $NADPH$:2-(2-ketopropylthio)ethanesulfonate oxidoreductase/carboxylase; $NADPH$:2-ketopropyl-coenzyme M oxidoreductase/carboxylase; 2-mercaptoethanesulfonate,acetoacetate: $NADP^+$ oxidoreductase (decarboxylating)
Systematic name: 2-sulfanylethane-1-sulfonate,acetoacetate: $NADP^+$ oxidoreductase (decarboxylating)
Comments: Also acts on thioethers longer in chain length on the oxo side, e.g. 2-oxobutyl-CoM, but this portion must be attached to CoM (2-sulfanylethane-1-sulfonate); no CoM analogs will substitute. This enzyme forms component II of a four-component enzyme system EC 4.4.1.23 (2-hydroxypropyl-CoM lyase; component I), EC 1.8.1.5 [2-oxopropyl-CoM reductase (carboxylating); component II], EC 1.1.1.268 [2-(*R*)-hydroxypropyl-CoM dehydrogenase; component III] and EC 1.1.1.269 [2-(*S*)-hydroxypropyl-CoM dehydrogenase; component IV].html";click here that is involved in epoxyalkane carboxylation in *Xanthobacter* sp. strain Py2.
References: [68, 695]

[EC 1.8.1.5 created 2001]

EC 1.8.1.6

- Accepted name:** cystine reductase
Reaction: 2 L-cysteine + NAD^+ = L-cystine + $NADH + H^+$
Other name(s): cystine reductase ($NADH$); $NADH$ -dependent cystine reductase; cystine reductase ($NADH_2$); $NADH_2$:L-cystine oxidoreductase
Systematic name: L-cysteine: NAD^+ oxidoreductase
References: [3562, 564, 2649]

[EC 1.8.1.6 created 1961 as EC 1.6.4.1, transferred 2002 to EC 1.8.1.6]

EC 1.8.1.7

Accepted name: glutathione-disulfide reductase
Reaction: $2 \text{ glutathione} + \text{NADP}^+ = \text{glutathione disulfide} + \text{NADPH} + \text{H}^+$
Other name(s): glutathione reductase; glutathione reductase (NADPH); NADPH-glutathione reductase; GSH reductase; GSSG reductase; NADPH-GSSG reductase; glutathione *S*-reductase; NADPH:oxidized-glutathione oxidoreductase
Systematic name: glutathione:NADP⁺ oxidoreductase
Comments: A dimeric flavoprotein (FAD); activity is dependent on a redox-active disulfide in each of the active centres.
References: [3221, 3324, 3421, 4408, 4670, 373, 2475]

[EC 1.8.1.7 created 1961 as EC 1.6.4.2, modified 1989, transferred 2002 to EC 1.8.1.7]

EC 1.8.1.8

Accepted name: protein-disulfide reductase
Reaction: $\text{protein-dithiol} + \text{NAD(P)}^+ = \text{protein-disulfide} + \text{NAD(P)H} + \text{H}^+$
Other name(s): protein disulphide reductase; insulin-glutathione transhydrogenase; disulfide reductase; NAD(P)H₂:protein-disulfide oxidoreductase
Systematic name: protein-dithiol:NAD(P)⁺ oxidoreductase
References: [1557]

[EC 1.8.1.8 created 1965 as EC 1.6.4.4, transferred 2002 to EC 1.8.1.8]

EC 1.8.1.9

Accepted name: thioredoxin-disulfide reductase
Reaction: $\text{thioredoxin} + \text{NADP}^+ = \text{thioredoxin disulfide} + \text{NADPH} + \text{H}^+$
Other name(s): NADP-thioredoxin reductase; NADPH-thioredoxin reductase; thioredoxin reductase (NADPH); NADPH₂:oxidized thioredoxin oxidoreductase
Systematic name: thioredoxin:NADP⁺ oxidoreductase
Comments: A flavoprotein (FAD).
References: [2874, 3978, 133]

[EC 1.8.1.9 created 1972 as EC 1.6.4.5, transferred 2002 to EC 1.8.1.9]

EC 1.8.1.10

Accepted name: CoA-glutathione reductase
Reaction: $\text{CoA} + \text{glutathione} + \text{NADP}^+ = \text{CoA-glutathione} + \text{NADPH} + \text{H}^+$
Other name(s): coenzyme A glutathione disulfide reductase; NADPH-dependent coenzyme A-SS-glutathione reductase; coenzyme A disulfide-glutathione reductase; NADPH₂:CoA-glutathione oxidoreductase
Systematic name: glutathione:NADP⁺ oxidoreductase (CoA-acylating)
Comments: A flavoprotein. The substrate is a mixed disulfide. May be identical to EC 1.8.1.9, thioredoxin-disulfide reductase.
References: [3182, 3183, 557]

[EC 1.8.1.10 created 1972 as EC 1.6.4.6, transferred 2002 to EC 1.8.1.10]

EC 1.8.1.11

Accepted name: asparagusate reductase
Reaction: $3\text{-sulfanyl-2-(sulfanylmethyl)propanoate} + \text{NAD}^+ = \text{asparagusate} + \text{NADH} + \text{H}^+$
Other name(s): asparagusate dehydrogenase; asparagusic dehydrogenase; asparagusate reductase (NADH₂); NADH₂:asparagusate oxidoreductase; 3-mercapto-2-mercaptomethylpropanoate:NAD⁺ oxidoreductase
Systematic name: 3-sulfanyl-2-(sulfanylmethyl)propanoate:NAD⁺ oxidoreductase

Comments: Also acts on lipoate.

References: [4759, 4760]

[EC 1.8.1.11 created 1978 as EC 1.6.4.7, transferred 2002 to EC 1.8.1.11]

EC 1.8.1.12

Accepted name: trypanothione-disulfide reductase

Reaction: trypanothione + NADP⁺ = trypanothione disulfide + NADPH + H⁺

Other name(s): trypanothione reductase; NADPH₂:trypanothione oxidoreductase

Systematic name: trypanothione:NADP⁺ oxidoreductase

Comments: Trypanothione disulfide is the oxidized form of *N*¹,*N*⁸-bis(glutathionyl)-spermidine from the insect-parasitic trypanosomatid *Crithidia fasciculata*. The enzyme from *Crithidia fasciculata* is a flavoprotein (FAD), whose activity is dependent on a redox-active cystine at the active centre. (*cf.* EC 1.8.1.7, glutathione-disulfide reductase)

References: [3816, 2660, 786]

[EC 1.8.1.12 created 1989 as EC 1.6.4.8, transferred 2002 to EC 1.8.1.12]

EC 1.8.1.13

Accepted name: bis- γ -glutamylcystine reductase

Reaction: 2 γ -glutamylcystine + NADP⁺ = bis- γ -glutamylcystine + NADPH + H⁺

Other name(s): NADPH₂:bis- γ -glutamylcystine oxidoreductase; GSR

Systematic name: γ -glutamylcystine:NADP⁺ oxidoreductase

Comments: Contains FAD. The enzyme, which is found only in halobacteria, maintains the concentration of γ -glutamylcystine, the major low molecular weight thiol in halobacteria. Not identical with EC 1.8.1.7 (glutathione-disulfide reductase) or EC 1.8.1.14 (CoA-disulfide reductase).

References: [4127, 4128, 2093]

[EC 1.8.1.13 created 1992 as EC 1.6.4.9, transferred 2002 to EC 1.8.1.13, modified 2013]

EC 1.8.1.14

Accepted name: CoA-disulfide reductase

Reaction: 2 CoA + NADP⁺ = CoA-disulfide + NADPH + H⁺

Other name(s): CoA-disulfide reductase (NADH₂); NADH₂:CoA-disulfide oxidoreductase; CoA:NAD⁺ oxidoreductase (misleading); CoADR; coenzyme A disulfide reductase

Systematic name: CoA:NADP⁺ oxidoreductase

Comments: A flavoprotein. Not identical with EC 1.8.1.6 (cystine reductase), EC 1.8.1.7 (glutathione-disulfide reductase) or EC 1.8.1.13 (bis- γ -glutamylcystine reductase). The enzyme from the bacterium *Staphylococcus aureus* has a strong preference for NADPH [2553], while the bacterium *Bacillus megaterium* contains both NADH and NADPH-dependent enzymes [3802].

References: [3802, 872, 2553]

[EC 1.8.1.14 created 1992 as EC 1.6.4.10, transferred 2002 to EC 1.8.1.14, modified 2005, modified 2013]

EC 1.8.1.15

Accepted name: mycothione reductase

Reaction: 2 mycothiol + NAD(P)⁺ = mycothione + NAD(P)H + H⁺

Other name(s): mycothiol-disulfide reductase

Systematic name: mycothiol:NAD(P)⁺ oxidoreductase

Comments: Contains FAD. No activity with glutathione, trypanothione or coenzyme A as substrate.

References: [3254, 3255]

[EC 1.8.1.15 created 2002]

EC 1.8.1.16

Accepted name: glutathione amide reductase
Reaction: $2 \text{ glutathione amide} + \text{NAD}^+ = \text{glutathione amide disulfide} + \text{NADH} + \text{H}^+$
Other name(s): GAR
Systematic name: glutathione amide:NAD⁺ oxidoreductase
Comments: A dimeric flavoprotein (FAD). The enzyme restores glutathione amide disulfide, which is produced during the reduction of peroxide by EC 1.11.1.17 (glutathione amide-dependent peroxidase), back to glutathione amide (it catalyses the reaction in the opposite direction to that shown). The enzyme belongs to the family of flavoprotein disulfide oxidoreductases, but unlike other members of the family, which are specific for NADPH, it prefers NADH [4436].
References: [4436, 4437]

[EC 1.8.1.16 created 2010]

EC 1.8.1.17

Accepted name: dimethylsulfone reductase
Reaction: $\text{dimethyl sulfoxide} + \text{H}_2\text{O} + \text{NAD}^+ = \text{dimethyl sulfone} + \text{NADH} + \text{H}^+$
Comments: A molybdoprotein.
References: [394, 395]

[EC 1.8.1.17 created 2011]

EC 1.8.1.18

Accepted name: NAD(P)H sulfur oxidoreductase (CoA-dependent)
Reaction: $\text{hydrogen sulfide} + \text{NAD(P)}^+ = \text{sulfur} + \text{NAD(P)H} + \text{H}^+$
Other name(s): NADPH NSR; S⁰ reductase; coenzyme A-dependent NADPH sulfur oxidoreductase
Systematic name: hydrogen sulfide:NAD(P)⁺ oxidoreductase (CoA-dependent)
Comments: This FAD-dependent enzyme, characterized from the archaeon *Pyrococcus furiosus*, is responsible for NAD(P)H-linked sulfur reduction. The activity with NADH is about half of that with NADPH. The reaction is dependent on CoA, although the nature of this dependency is not well understood.
References: [3762, 439, 1534]

[EC 1.8.1.18 created 2013]

EC 1.8.1.19

Accepted name: sulfide dehydrogenase
Reaction: $\text{hydrogen sulfide} + (\text{sulfide})_n + \text{NADP}^+ = (\text{sulfide})_{n+1} + \text{NADPH} + \text{H}^+$
Other name(s): SuDH
Systematic name: hydrogen sulfide, polysulfide:NADP⁺ oxidoreductase
Comments: A iron-sulfur flavoprotein. In the archaeon *Pyrococcus furiosus* the enzyme is involved in the oxidation of NADPH which is produced in peptide degradation. The enzyme also catalyses the reduction of sulfur with lower activity.
References: [2573, 1473]

[EC 1.8.1.19 created 2013]

EC 1.8.1.20

Accepted name: 4,4'-dithiodibutanoate disulfide reductase
Reaction: $2 \text{ 4-sulfanylbutanoate} + \text{NAD}^+ = 4,4'\text{-disulfanediyldibutanoate} + \text{NADH} + \text{H}^+$
Systematic name: 4-sulfanylbutanoate:NAD⁺ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Rhodococcus erythropolis* MI2, contains an FMN cofactor.
References: [2075, 2076]

[EC 1.8.1.20 created 2017]

EC 1.8.1.21

Accepted name: dissimilatory dimethyldisulfide reductase
Reaction: $2 \text{ methanethiol} + \text{NAD}^+ = \text{dimethyl disulfide} + \text{NADH} + \text{H}^+$
Systematic name: methanethiol:NAD⁺ oxidoreductase (dimethyl disulfide-forming)
Comments: The enzyme's activity has been demonstrated in the bacterium *Thiobacillus thioparus* E6. The methanethiol formed is eventually oxidized to sulfate and carbon dioxide, and the latter assimilated for autotrophic growth.
References: [3942, 3943]

[EC 1.8.1.21 created 2019]

EC 1.8.2 With a cytochrome as acceptor

EC 1.8.2.1

Accepted name: sulfite dehydrogenase (cytochrome)
Reaction: $\text{sulfite} + 2 \text{ ferricytochrome } c + \text{H}_2\text{O} = \text{sulfate} + 2 \text{ ferrocyclochrome } c + 2 \text{ H}^+$
Other name(s): sulfite cytochrome *c* reductase; sulfite-cytochrome *c* oxidoreductase; sulfite oxidase (ambiguous); sulfite dehydrogenase (ambiguous); *sorAB* (gene names)
Systematic name: sulfite:ferricytochrome-*c* oxidoreductase
Comments: Associated with cytochrome *c*-551. The enzyme from the bacterium *Starkeya novella* contains a molybdopyranopterin cofactor and a smaller monoheme cytochrome *c* subunit. *cf.* EC 1.8.5.6, sulfite dehydrogenase (quinone).
References: [607, 2572, 4747, 2551, 1992]

[EC 1.8.2.1 created 1972, modified 2016]

EC 1.8.2.2

Accepted name: thiosulfate dehydrogenase
Reaction: $2 \text{ thiosulfate} + 2 \text{ ferricytochrome } c = \text{tetrathionate} + 2 \text{ ferrocyclochrome } c$
Other name(s): *tsdA* (gene name); tetrathionate synthase; thiosulfate oxidase; thiosulfate-oxidizing enzyme; thiosulfate-acceptor oxidoreductase
Systematic name: thiosulfate:ferricytochrome-*c* oxidoreductase
Comments: The enzyme catalyses the reversible formation of a sulfur-sulfur bond between the sulfane atoms of two thiosulfate molecules, yielding tetrathionate and releasing two electrons. In many bacterial species the enzyme is a diheme *c*-type cytochrome. In a number of organisms, including *Thiomonas intermedia* and *Sideroxydans lithotrophicus*, a second diheme cytochrome (TsdB) acts as the electron acceptor. However, some organisms, such as *Allochromatium vinosum*, lack TsdB. The electron acceptor in these organisms may be the high-potential iron-sulfur protein (HiPIP).
References: [2552, 1224, 2526, 444, 2305]

[EC 1.8.2.2 created 1990]

EC 1.8.2.3

Accepted name: sulfide-cytochrome-*c* reductase (flavocytochrome *c*)
Reaction: $\text{hydrogen sulfide} + 2 \text{ ferricytochrome } c = \text{sulfur} + 2 \text{ ferrocyclochrome } c + 2 \text{ H}^+$
Systematic name: hydrogen-sulfide:flavocytochrome *c* oxidoreductase
Comments: The enzyme from *Allochromatium vinosum* contains covalently bound FAD and covalently-bound *c*-type hemes.
References: [2306, 1225, 1390, 640, 3970, 2242]

[EC 1.8.2.3 created 2011]

EC 1.8.2.4

- Accepted name:** dimethyl sulfide:cytochrome *c*₂ reductase
Reaction: dimethyl sulfide + 2 ferricytochrome *c*₂ + H₂O = dimethyl sulfoxide + 2 ferrocyclochrome *c*₂ + 2 H⁺
Other name(s): Ddh (gene name)
Systematic name: dimethyl sulfide:cytochrome-*c*₂ oxidoreductase
Comments: The enzyme from the bacterium *Rhodovulum sulfidophilum* binds molybdopterin guanine dinucleotide, heme *b* and [4Fe-4S] clusters.
References: [1506, 2746]

[EC 1.8.2.4 created 2011]

EC 1.8.2.5

- Accepted name:** thiosulfate reductase (cytochrome)
Reaction: sulfite + hydrogen sulfide + 2 ferricytochrome *c*₃ = thiosulfate + 2 ferrocyclochrome *c*₃
Systematic name: sulfite,hydrogen sulfide:ferricytochrome-*c*₃ oxidoreductase (thiosulfate-forming)
Comments: The enzyme is found in sulfate-reducing bacteria. The source of the electrons is molecular hydrogen, via EC 1.12.2.1, cytochrome-*c*₃ hydrogenase. The organisms utilize the sulfite that is produced for energy generation by EC 1.8.99.5, dissimilatory sulfite reductase.
References: [1834, 1833, 2998, 1546, 1558, 51]

[EC 1.8.2.5 created 2017]

EC 1.8.2.6

- Accepted name:** *S*-disulfanyl-L-cysteine oxidoreductase
Reaction: [SoxY protein]-*S*-disulfanyl-L-cysteine + 6 ferricytochrome *c* + 3 H₂O = [SoxY protein]-*S*-sulfosulfanyl-L-cysteine + 6 ferrocyclochrome *c* + 6 H⁺
Other name(s): SoxCD; sulfur dehydrogenase
Systematic name: [SoxY protein]-*S*-disulfanyl-L-cysteine:cytochrome-*c* oxidoreductase
Comments: The enzyme is part of the Sox enzyme system, which participates in a bacterial thiosulfate oxidation pathway that produces sulfate. The enzyme from the bacterium *Paracoccus pantotrophus* contains a molybdoprotein component and a diheme *c*-type cytochrome component. The enzyme successively oxidizes the outer sulfur atom in [SoxY protein]-*S*-disulfanyl-L-cysteine, using three water molecules and forming [SoxY protein]-*S*-sulfosulfanyl-L-cysteine. During the process, six electrons are transferred to the electron chain via cytochrome *c*.
References: [1187, 219, 1379]

[EC 1.8.2.6 created 2018]

EC 1.8.2.7

- Accepted name:** thiocyanate desulfurase
Reaction: thiocyanate + 2 ferricytochrome *c* + H₂O = cyanate + sulfur + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): TcDH; thiocyanate dehydrogenase
Systematic name: thiocyanate:cytochrome *c* oxidoreductase (cyanate and sulfur-forming)
Comments: The enzyme, characterized from the haloalkaliphilic sulfur-oxidizing bacterium *Thioalkalivibrio paradoxus*, contains three copper ions in its active site. It catalyses the direct conversion of thiocyanate into cyanate and elemental sulfur without involvement of molecular oxygen.
References: [4290]

[EC 1.8.2.7 created 2020]

EC 1.8.3 With oxygen as acceptor

EC 1.8.3.1

Accepted name: sulfite oxidase
Reaction: sulfite + O₂ + H₂O = sulfate + H₂O₂
Systematic name: sulfite:oxygen oxidoreductase
Comments: A molybdohemoprotein.
References: [2073, 2595, 4163]

[EC 1.8.3.1 created 1961]

EC 1.8.3.2

Accepted name: thiol oxidase
Reaction: 2 R'C(R)SH + O₂ = R'C(R)S-S(R)CR' + H₂O₂
Other name(s): sulfhydryl oxidase
Systematic name: thiol:oxygen oxidoreductase
Comments: R may be =S or =O, or a variety of other groups. The enzyme is not specific for R'.
References: [156, 3044, 3201, 1709, 1879, 3807, 797, 1093, 1422, 850, 3519]

[EC 1.8.3.2 created 1961, modified 2010, modified 2011]

EC 1.8.3.3

Accepted name: glutathione oxidase
Reaction: 2 glutathione + O₂ = glutathione disulfide + H₂O₂
Systematic name: glutathione:oxygen oxidoreductase
Comments: A flavoprotein (FAD). Also acts, more slowly, on L-cysteine and several other thiols.
References: [2308]

[EC 1.8.3.3 created 1989]

EC 1.8.3.4

Accepted name: methanethiol oxidase
Reaction: methanethiol + O₂ + H₂O = formaldehyde + hydrogen sulfide + H₂O₂
Other name(s): methylmercaptan oxidase; methyl mercaptan oxidase; (MM)-oxidase; MT-oxidase
Systematic name: methanethiol:oxygen oxidoreductase
References: [4134]

[EC 1.8.3.4 created 1990]

EC 1.8.3.5

Accepted name: prenylcysteine oxidase
Reaction: an *S*-prenyl-L-cysteine + O₂ + H₂O = a prenal + L-cysteine + H₂O₂
Other name(s): prenylcysteine lyase
Systematic name: *S*-prenyl-L-cysteine:oxygen oxidoreductase
Comments: A flavoprotein (FAD). Cleaves the thioether bond of *S*-prenyl-L-cysteines, such as *S*-farnesylcysteine and *S*-geranylgeranylgeranyl-L-cysteine. *N*-Acetyl-prenylcysteine and prenylcysteinyll peptides are not substrates. May represent the final step in the degradation of prenylated proteins in mammalian tissues. Originally thought to be a simple lyase so it had been classified as EC 4.4.1.18.
References: [4882, 4336]

[EC 1.8.3.5 created 2000 as EC 4.4.1.18, transferred 2002 to EC 1.8.3.5]

EC 1.8.3.6

- Accepted name:** farnesylcysteine lyase
Reaction: $S\text{-}(2E,6E)\text{-farnesyl-L-cysteine} + O_2 + H_2O = (2E,6E)\text{-farnesal} + L\text{-cysteine} + H_2O_2$
Other name(s): FC lyase; FCLY
Systematic name: $S\text{-}(2E,6E)\text{-farnesyl-L-cysteine oxidase}$
Comments: A flavoprotein (FAD). In contrast to mammalian EC 1.8.3.5 (prenylcysteine oxidase) the farnesylcysteine lyase from *Arabidopsis* is specific for *S*-farnesyl-L-cysteine and shows no activity with *S*-geranylgeranyl-L-cysteine.
References: [1769, 775]

[EC 1.8.3.6 created 2011]

EC 1.8.3.7

- Accepted name:** formylglycine-generating enzyme
Reaction: a [sulfatase]-L-cysteine + $O_2 + 2$ a thiol = a [sulfatase]-3-oxo-L-alanine + hydrogen sulfide + a disulfide + H_2O
Other name(s): sulfatase-modifying factor 1; α -formylglycine-generating enzyme 1; SUMF1 (gene name)
Systematic name: [sulfatase]-L-cysteine:oxygen oxidoreductase (3-oxo-L-alanine-forming)
Comments: Requires a copper cofactor and Ca^{2+} . The enzyme, which is found in both prokaryotes and eukaryotes, catalyses a modification of a conserved L-cysteine residue in the active site of sulfatases, generating a unique 3-oxo-L-alanine residue that is essential for sulfatase activity. The exact nature of the thiol involved is still not clear - dithiothreitol and cysteamine are the most efficiently used thiols *in vitro*. Glutathione also acts *in vitro*, but it is not known whether it is used *in vivo*.
References: [909, 908, 3377, 3554, 559, 1693, 2171, 2170, 2784]

[EC 1.8.3.7 created 2014]

EC 1.8.4 With a disulfide as acceptor

EC 1.8.4.1

- Accepted name:** glutathione—homocystine transhydrogenase
Reaction: 2 glutathione + homocystine = glutathione disulfide + 2 homocystine
Systematic name: glutathione:homocystine oxidoreductase
Comments: The reactions catalysed by this enzyme and by others in this subclass may be similar to those catalysed by EC 2.5.1.18 glutathione transferase.
References: [3420]

[EC 1.8.4.1 created 1961]

EC 1.8.4.2

- Accepted name:** protein-disulfide reductase (glutathione)
Reaction: 2 glutathione + protein-disulfide = glutathione-disulfide + protein-dithiol
Other name(s): glutathione-insulin transhydrogenase; insulin reductase; reductase, protein disulfide (glutathione); protein disulfide transhydrogenase; glutathione-protein disulfide oxidoreductase; protein disulfide reductase (glutathione); GSH-insulin transhydrogenase; protein-disulfide interchange enzyme; protein-disulfide isomerase/oxidoreductase; thiol:protein-disulfide oxidoreductase; thiol-protein disulfide oxidoreductase
Systematic name: glutathione:protein-disulfide oxidoreductase
Comments: Reduces insulin and some other proteins.
References: [2031, 2204]

[EC 1.8.4.2 created 1965]

EC 1.8.4.3

Accepted name: glutathione—CoA-glutathione transhydrogenase
Reaction: CoA + glutathione disulfide = CoA-glutathione + glutathione
Other name(s): glutathione-coenzyme A glutathione disulfide transhydrogenase; glutathione-coenzyme A glutathione disulfide transhydrogenase; glutathione coenzyme A-glutathione transhydrogenase; glutathione:coenzyme A-glutathione transhydrogenase; coenzyme A:oxidized-glutathione oxidoreductase; coenzyme A:glutathione-disulfide oxidoreductase
Systematic name: CoA:glutathione-disulfide oxidoreductase
References: [597]

[EC 1.8.4.3 created 1972]

EC 1.8.4.4

Accepted name: glutathione—cystine transhydrogenase
Reaction: 2 glutathione + cystine = glutathione disulfide + 2 cysteine
Other name(s): GSH-cystine transhydrogenase; NADPH-dependent GSH-cystine transhydrogenase
Systematic name: glutathione:cystine oxidoreductase
References: [2956]

[EC 1.8.4.4 created 1972]

[1.8.4.5 *Transferred entry. methionine-S-oxide reductase. Now EC 1.8.4.13, L-methionine (S)-S-oxide reductase and EC 1.8.4.14, L-methionine (R)-S-oxide reductase*]

[EC 1.8.4.5 created 1984, deleted 2006]

[1.8.4.6 *Transferred entry. protein-methionine-S-oxide reductase. Proved to be due to EC 1.8.4.11, peptide-methionine (S)-S-oxide reductase*]

[EC 1.8.4.6 created 1984, deleted 2006]

EC 1.8.4.7

Accepted name: enzyme-thiol transhydrogenase (glutathione-disulfide)
Reaction: [xanthine dehydrogenase] + glutathione disulfide = [xanthine oxidase] + 2 glutathione
Other name(s): [xanthine-dehydrogenase]:oxidized-glutathione S-oxidoreductase; enzyme-thiol transhydrogenase (oxidized-glutathione); glutathione-dependent thiol:disulfide oxidoreductase; thiol:disulphide oxidoreductase
Systematic name: [xanthine-dehydrogenase]:glutathione-disulfide S-oxidoreductase
Comments: Converts EC 1.17.1.4 xanthine dehydrogenase into EC 1.17.3.2 xanthine oxidase in the presence of glutathione disulfide; also reduces the disulfide bond of ricin. Not inhibited by Cu²⁺ or thiol reagents.
References: [238]

[EC 1.8.4.7 created 1989, modified 2002]

EC 1.8.4.8

Accepted name: phosphoadenylyl-sulfate reductase (thioredoxin)
Reaction: adenosine 3',5'-bisphosphate + sulfite + thioredoxin disulfide = 3'-phosphoadenylyl sulfate + thioredoxin
Other name(s): PAPS reductase, thioredoxin-dependent; PAPS reductase; thioredoxin:adenosine 3'-phosphate 5'-phosphosulfate reductase; 3'-phosphoadenylylsulfate reductase; thioredoxin:3'-phosphoadenylylsulfate reductase; phosphoadenosine-phosphosulfate reductase; adenosine 3',5'-bisphosphate,sulfite:oxidized-thioredoxin oxidoreductase (3'-phosphoadenosine-5'-phosphosulfate-forming)
Systematic name: adenosine 3',5'-bisphosphate,sulfite:thioredoxin-disulfide oxidoreductase (3'-phosphoadenosine-5'-phosphosulfate-forming)

Comments: Specific for PAPS. The enzyme from *Escherichia coli* will use thioredoxins from other species.
References: [293]

[EC 1.8.4.8 created 1999 as EC 1.8.99.4, transferred 2000 to EC 1.8.4.8]

EC 1.8.4.9

Accepted name: adenylyl-sulfate reductase (glutathione)
Reaction: AMP + sulfite + glutathione disulfide = adenylyl sulfate + 2 glutathione
Other name(s): 5'-adenylylsulfate reductase (also used for EC 1.8.99.2); AMP,sulfite:oxidized-glutathione oxidoreductase (adenosine-5'-phosphosulfate-forming); plant-type 5'-adenylylsulfate reductase
Systematic name: AMP,sulfite:glutathione-disulfide oxidoreductase (adenosine-5'-phosphosulfate-forming)
Comments: This enzyme differs from EC 1.8.99.2, adenylyl-sulfate reductase, in using glutathione as the reductant. Glutathione can be replaced by γ -glutamylcysteine or dithiothreitol, but not by thioredoxin, glutaredoxin or 2-sulfanylethan-1-ol (2-mercaptoethanol). The enzyme from the mouse ear cross, *Arabidopsis thaliana*, contains a glutaredoxin-like domain. The enzyme is also found in other photosynthetic eukaryotes, e.g., the Madagascar periwinkle, *Catharanthus roseus* and the hollow green seaweed, *Ulva intestinalis*.
References: [1457, 3803, 331]

[EC 1.8.4.9 created 2000, modified 2002]

EC 1.8.4.10

Accepted name: adenylyl-sulfate reductase (thioredoxin)
Reaction: AMP + sulfite + thioredoxin disulfide = 5'-adenylyl sulfate + thioredoxin
Other name(s): thioredoxin-dependent 5'-adenylylsulfate reductase
Systematic name: AMP,sulfite:thioredoxin-disulfide oxidoreductase (adenosine-5'-phosphosulfate-forming)
Comments: Uses adenylyl sulfate, not phosphoadenylyl sulfate, distinguishing this enzyme from EC 1.8.4.8, phosphoadenylyl-sulfate reductase (thioredoxin). Uses thioredoxin as electron donor, not glutathione or other donors, distinguishing it from EC 1.8.4.9 [adenylyl-sulfate reductase (glutathione)] and EC 1.8.99.2 (adenylyl-sulfate reductase).
References: [332, 5, 4636, 3051]

[EC 1.8.4.10 created 2003]

EC 1.8.4.11

Accepted name: peptide-methionine (S)-S-oxide reductase
Reaction: (1) peptide-L-methionine + thioredoxin disulfide + H₂O = peptide-L-methionine (S)-S-oxide + thioredoxin
(2) L-methionine + thioredoxin disulfide + H₂O = L-methionine (S)-S-oxide + thioredoxin
Other name(s): MsrA; methionine sulfoxide reductase (ambiguous); methionine sulphoxide reductase A; methionine S-oxide reductase (ambiguous); methionine S-oxide reductase (S-form oxidizing); methionine sulfoxide reductase A; peptide methionine sulfoxide reductase
Systematic name: peptide-L-methionine:thioredoxin-disulfide S-oxidoreductase [L-methionine (S)-S-oxide-forming]
Comments: The reaction occurs in the reverse direction to that shown above. The enzyme exhibits high specificity for the reduction of the S-form of L-methionine S-oxide, acting faster on the residue in a peptide than on the free amino acid [3174]. On the free amino acid, it can also reduce D-methionine (S)-S-oxide but more slowly [3174]. The enzyme plays a role in preventing oxidative-stress damage caused by reactive oxygen species by reducing the oxidized form of methionine back to methionine and thereby reactivating peptides that had been damaged. In some species, e.g. *Neisseria meningitidis*, both this enzyme and EC 1.8.4.12, peptide-methionine (R)-S-oxide reductase, are found within the same protein whereas, in other species, they are separate proteins [2908, 399]. The reaction proceeds via a sulfenic-acid intermediate [1075, 452].
References: [2908, 4226, 3913, 399, 1075, 4579, 2032, 4475, 3174, 452]

[EC 1.8.4.11 created 2006]

EC 1.8.4.12

- Accepted name:** peptide-methionine (*R*)-*S*-oxide reductase
Reaction: peptide-L-methionine + thioredoxin disulfide + H₂O = peptide-L-methionine (*R*)-*S*-oxide + thioredoxin
Other name(s): MsrB; methionine sulfoxide reductase (ambiguous); pMSR; methionine *S*-oxide reductase (ambiguous); selenoprotein R; methionine *S*-oxide reductase (*R*-form oxidizing); methionine sulfoxide reductase B; SelR; SelX; PilB; pRMsr
Systematic name: peptide-methionine:thioredoxin-disulfide *S*-oxidoreductase [methionine (*R*)-*S*-oxide-forming]
Comments: The reaction occurs in the reverse direction to that shown above. The enzyme exhibits high specificity for reduction of the *R*-form of methionine *S*-oxide, with higher activity being observed with L-methionine *S*-oxide than with D-methionine *S*-oxide [3174]. While both free and protein-bound methionine (*R*)-*S*-oxide act as substrates, the activity with the peptide-bound form is far greater [3628]. The enzyme plays a role in preventing oxidative-stress damage caused by reactive oxygen species by reducing the oxidized form of methionine back to methionine and thereby reactivating peptides that had been damaged. In some species, e.g. *Neisseria meningitidis*, both this enzyme and EC 1.8.4.11, peptide-methionine (*S*)-*S*-oxide reductase, are found within the same protein whereas in other species, they are separate proteins [3913, 1075]. The reaction proceeds via a sulfenic-acid intermediate [1075, 3628]. For MsrB2 and MsrB3, thioredoxin is a poor reducing agent but thionein works well []. The enzyme from some species contains selenocysteine and Zn²⁺.
References: [2908, 4226, 3913, 399, 1075, 4579, 2032, 4475, 3174, 3628]

[EC 1.8.4.12 created 2006]

EC 1.8.4.13

- Accepted name:** L-methionine (*S*)-*S*-oxide reductase
Reaction: L-methionine + thioredoxin disulfide + H₂O = L-methionine (*S*)-*S*-oxide + thioredoxin
Other name(s): fMSr; methyl sulfoxide reductase I and II; acetylmethionine sulfoxide reductase; methionine sulfoxide reductase; L-methionine:oxidized-thioredoxin *S*-oxidoreductase; methionine-*S*-oxide reductase; free-methionine (*S*)-*S*-oxide reductase
Systematic name: L-methionine:thioredoxin-disulfide *S*-oxidoreductase
Comments: Requires NADPH [1031]. The reaction occurs in the opposite direction to that given above. Dithiothreitol can replace reduced thioredoxin. L-Methionine (*R*)-*S*-oxide is not a substrate [see EC 1.8.4.14, L-methionine (*R*)-*S*-oxide reductase].
References: [344, 1031, 1032, 4579]

[EC 1.8.4.13 created 1984 as EC 1.8.4.5, part transferred 2006 to EC 1.8.4.13]

EC 1.8.4.14

- Accepted name:** L-methionine (*R*)-*S*-oxide reductase
Reaction: L-methionine + thioredoxin disulfide + H₂O = L-methionine (*R*)-*S*-oxide + thioredoxin
Other name(s): fRMsr; FRMsR; free met-R-(o) reductase; free-methionine (*R*)-*S*-oxide reductase
Systematic name: L-methionine:thioredoxin-disulfide *S*-oxidoreductase [L-methionine (*R*)-*S*-oxide-forming]
Comments: Requires NADPH. Unlike EC 1.8.4.12, peptide-methionine (*R*)-*S*-oxide reductase, this enzyme cannot use peptide-bound methionine (*R*)-*S*-oxide as a substrate [1068]. Differs from EC 1.8.4.13, L-methionine (*S*)-*S*-oxide in that L-methionine (*S*)-*S*-oxide is not a substrate.
References: [1068]

[EC 1.8.4.14 created 1984 as EC 1.8.4.5, part transferred 2006 to EC 1.8.4.14]

EC 1.8.4.15

- Accepted name:** protein dithiol oxidoreductase (disulfide-forming)

Reaction: a [DsbA protein] carrying a disulfide bond + a [protein] with reduced L-cysteine residues = a [DsbA protein] with reduced L-cysteine residues + a [protein] carrying a disulfide bond

Other name(s): *dsbA* (gene name)

Systematic name: protein dithiol:[DsbA protein] oxidoreductase (protein disulfide-forming)

Comments: DsbA is a periplasmic thiol:disulfide oxidoreductase found in Gram-negative bacteria that promotes protein disulfide bond formation. DsbA contains a redox active disulfide bond that is catalytically transferred via disulfide exchange to a diverse range of newly translocated protein substrates. The protein is restored to the oxidized state by EC 1.8.5.9, protein dithiol:quinone oxidoreductase DsbB.

References: [220, 52, 4861, 175, 1436, 1968]

[EC 1.8.4.15 created 2019]

EC 1.8.4.16

Accepted name: thioredoxin:protein disulfide reductase

Reaction: a [protein] with reduced L-cysteine residues + thioredoxin disulfide = a [protein] carrying a disulfide bond + thioredoxin (overall reaction)
(1a) a [DsbD protein] with reduced L-cysteine residues + thioredoxin disulfide = a [DsbD protein] carrying a disulfide bond + thioredoxin
(1b) a [DsbD protein] carrying a disulfide bond + a [protein] with reduced L-cysteine residues = a [DsbD protein] with reduced L-cysteine residues + a [protein] carrying a disulfide bond

Other name(s): *dsbD* (gene name); *dipZ* (gene name)

Systematic name: thioredoxin:protein disulfide oxidoreductase (dithiol-forming)

Comments: DsbD is an inner membrane protein found in Gram-negative bacteria that transfers electrons from cytoplasmic thioredoxin to the periplasmic substrate proteins DsbC, DsbG and CcmG, reducing disulfide bonds in the target proteins to dithiols. DsbD consists of three domains: a periplasmic N-terminal domain, a central transmembrane domain and a periplasmic C-terminal domain.

References: [2827, 1367, 2029, 1376, 2030, 3587]

[EC 1.8.4.16 created 2019]

EC 1.8.5 With a quinone or similar compound as acceptor

EC 1.8.5.1

Accepted name: glutathione dehydrogenase (ascorbate)

Reaction: 2 glutathione + dehydroascorbate = glutathione disulfide + ascorbate

Other name(s): dehydroascorbic reductase; dehydroascorbic acid reductase; glutathione dehydroascorbate reductase; DHA reductase ; dehydroascorbate reductase; GDOR; glutathione:dehydroascorbic acid oxidoreductase

Systematic name: glutathione:dehydroascorbate oxidoreductase

References: [771]

[EC 1.8.5.1 created 1961]

EC 1.8.5.2

Accepted name: thiosulfate dehydrogenase (quinone)

Reaction: 2 thiosulfate + 6-decylubiquinone = tetrathionate + 6-decylubiquinol

Other name(s): thiosulfate:quinone oxidoreductase; thiosulphate:quinone oxidoreductase; thiosulfate oxidoreductase, tetrathionate-forming; TQO

Systematic name: thiosulfate:6-decylubiquinone oxidoreductase

Comments: The reaction can also proceed with ferricyanide as the electron acceptor, but more slowly. Unlike EC 1.8.2.2, thiosulfate dehydrogenase, this enzyme cannot utilize cytochrome *c* as an acceptor.

References: [2922]

[EC 1.8.5.2 created 2004]

EC 1.8.5.3

Accepted name: respiratory dimethylsulfoxide reductase
Reaction: dimethylsulfide + menaquinone + H₂O = dimethylsulfoxide + menaquinol
Other name(s): *dmsABC* (gene names); DMSO reductase (ambiguous); dimethylsulfoxide reductase (ambiguous)
Systematic name: dimethyl sulfide:menaquinone oxidoreductase
Comments: The enzyme participates in bacterial electron transfer pathways in which dimethylsulfoxide (DMSO) is the terminal electron acceptor. It is composed of three subunits - DmsA contains a bis(guanylyl molybdopterin) cofactor and a [4Fe-4S] cluster, DmsB is an iron-sulfur protein, and DmsC is a trans-membrane protein that anchors the enzyme and accepts electrons from the quinol pool. The electrons are passed through DmsB to DmsA and on to DMSO. The enzyme can also reduce pyridine-*N*-oxide and trimethylamine *N*-oxide to the corresponding amines with lower activity.
References: [830, 2798, 3903, 3579]

[EC 1.8.5.3 created 2011, modified 2019]

EC 1.8.5.4

Accepted name: bacterial sulfide:quinone reductase
Reaction: $n \text{ HS}^- + n \text{ quinone} = \text{polysulfide} + n \text{ quinol}$
Other name(s): *sqr* (gene name); sulfide:quinone reductase (ambiguous); sulfide:quinone oxidoreductase
Systematic name: sulfide:quinone oxidoreductase (polysulfide-producing)
Comments: Contains FAD. Ubiquinone, plastoquinone or menaquinone can act as acceptor in different species. In some organisms the enzyme catalyses the formation of sulfur globules. It repeats the catalytic cycle without releasing the product, producing a polysulfide of up to 10 sulfur atoms. The reaction stops when the maximum length of the polysulfide that can be accommodated in the sulfide oxidation pocket is achieved. The enzyme also plays an important role in anoxygenic bacterial photosynthesis. *cf.* EC 1.8.5.8, sulfide quinone oxidoreductase.
References: [132, 3489, 3115, 445, 647, 2645, 4699]

[EC 1.8.5.4 created 2011, modified 2017, modified 2019]

EC 1.8.5.5

Accepted name: thiosulfate reductase (quinone)
Reaction: sulfite + hydrogen sulfide + a quinone = thiosulfate + a quinol
Other name(s): *phsABC* (gene names)
Systematic name: sulfite,hydrogen sulfide:quinone oxidoreductase
Comments: The enzyme, characterized from the bacterium *Salmonella enterica*, is similar to EC 1.17.5.3, formate dehydrogenase-N. It contains a molybdopterin-guanine dinucleotide, five [4Fe-4S] clusters and two heme *b* groups. The reaction occurs *in vivo* in the direction of thiosulfate disproportionation, which is highly endergonic. It is driven by the proton motive force that occurs across the cytoplasmic membrane.
References: [2318, 697, 56, 1613, 4042]

[EC 1.8.5.5 created 2016, modified 2017]

EC 1.8.5.6

Accepted name: sulfite dehydrogenase (quinone)
Reaction: sulfite + a quinone + H₂O = sulfate + a quinol
Other name(s): *soeABC* (gene name)
Systematic name: sulfite:quinone oxidoreductase

Comments: This membrane-bound bacterial enzyme catalyses the direct oxidation of sulfite to sulfate in the cytoplasm. The enzyme, characterized from the bacteria *Ruegeria pomeroyi* and *Allochromatium vinosum*, is a complex that consists of a membrane anchor (SoeC) and two cytoplasmic subunits: an iron-sulfur protein (SoeB) and a molybdoprotein that contains a [4Fe-4S] iron-sulfur cluster (SoeA). *cf.* EC 1.8.2.1, sulfite dehydrogenase (cytochrome).

References: [799]

[EC 1.8.5.6 created 2016]

EC 1.8.5.7

Accepted name: glutathionyl-hydroquinone reductase
Reaction: glutathione + 2-(glutathione-*S*-yl)-hydroquinone = glutathione disulfide + hydroquinone
Other name(s): *pcpF* (gene name); *yqiG* (gene name)
Systematic name: 2-(glutathione-*S*-yl)-hydroquinone:glutathione oxidoreductase
Comments: This type of enzymes, which are found in bacteria, halobacteria, fungi, and plants, catalyse the glutathione-dependent reduction of glutathionyl-hydroquinones. The enzyme from the bacterium *Sphingobium chlorophenicum* can act on halogenated substrates such as 2,6-dichloro-3-(glutathione-*S*-yl)-hydroquinone and 2,3,5-trichloro-6-(glutathione-*S*-yl)-hydroquinone. Substrates for these enzymes are often formed spontaneously by interaction of benzoquinones with glutathione.
References: [1756, 4711, 2329, 1395]

[EC 1.8.5.7 created 2017]

EC 1.8.5.8

Accepted name: eukaryotic sulfide quinone oxidoreductase
Reaction: hydrogen sulfide + glutathione + a quinone = *S*-sulfanylglutathione + a quinol
Other name(s): SQR; SQOR; SQRDL (gene name)
Systematic name: sulfide:glutathione,quinone oxidoreductase
Comments: Contains FAD. This eukaryotic enzyme, located at the inner mitochondrial membrane, catalyses the first step in the metabolism of sulfide. While both sulfite and glutathione have been shown to act as sulfane sulfur acceptors *in vitro*, it is thought that the latter acts as the main acceptor *in vivo*. The electrons are transferred via FAD and quinones to the electron transfer chain. Unlike the bacterial homolog (EC 1.8.5.4, bacterial sulfide:quinone reductase), which repeats the catalytic cycle without releasing the product, producing a polysulfide, the eukaryotic enzyme transfers the persulfide to an acceptor at the end of each catalytic cycle.
References: [4570, 1651, 1872, 2474]

[EC 1.8.5.8 created 2017]

EC 1.8.5.9

Accepted name: protein dithiol:quinone oxidoreductase DsbB
Reaction: a [DsbA protein] with reduced L-cysteine residues + a quinone = a [DsbA protein] carrying a disulfide bond + a quinol (overall reaction)
(1a) a [DsbA protein] with reduced L-cysteine residues + a [DsbB protein] carrying a disulfide bond = a [DsbA protein] carrying a disulfide bond + a [DsbB protein] with reduced L-cysteine residues
(1b) a [DsbB protein] with reduced L-cysteine residues + a quinone = a [DsbB protein] carrying a disulfide bond + a quinol
Other name(s): *dsbB* (gene name)
Systematic name: protein dithiol:quinone oxidoreductase (disulfide-forming)
Comments: DsbB is a protein found in Gram-negative bacteria that functions within a pathway for protein disulfide bond formation. The enzyme catalyses the oxidation of the DsbA protein by generating disulfide bonds *de novo* via the reduction of membrane quinones. *cf.* EC 1.8.4.15, protein dithiol oxidoreductase (disulfide-forming).

References: [1441, 2130, 2129, 712, 993, 1807]

[EC 1.8.5.9 created 2019]

EC 1.8.6 With a nitrogenous group as acceptor (deleted sub-subclass)

[1.8.6.1 Deleted entry. Nitrate-ester reductase. Now included with EC 2.5.1.18 glutathione transferase]

[EC 1.8.6.1 created 1961, deleted 1976]

EC 1.8.7 With an iron-sulfur protein as acceptor

EC 1.8.7.1

Accepted name: assimilatory sulfite reductase (ferredoxin)
Reaction: hydrogen sulfide + 6 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O = sulfite + 6 reduced ferredoxin [iron-sulfur] cluster + 6 H⁺
Other name(s): ferredoxin-sulfite reductase; SIR (gene name); sulfite reductase (ferredoxin)
Systematic name: hydrogen-sulfide:ferredoxin oxidoreductase
Comments: An iron protein. The enzyme participates in sulfate assimilation. While it is usually found in cyanobacteria, plants and algae, it has also been reported in bacteria [3051]. Different from EC 1.8.99.5, dissimilatory sulfite reductase, which is involved in prokaryotic sulfur-based energy metabolism. *cf.* EC 1.8.1.2, assimilatory sulfite reductase (NADPH).
References: [3724, 1331, 393, 3051]

[EC 1.8.7.1 created 1972, modified 2015]

EC 1.8.7.2

Accepted name: ferredoxin:thioredoxin reductase
Reaction: 2 reduced ferredoxin + thioredoxin disulfide = 2 oxidized ferredoxin + thioredoxin + 2 H⁺
Systematic name: ferredoxin:thioredoxin disulfide oxidoreductase
Comments: The enzyme contains a [4Fe-4S] cluster and internal disulfide. It forms a mixed disulfide with thioredoxin on one side, and docks ferredoxin on the other side, enabling two one-electron transfers. The reduced thioredoxins generated by the enzyme activate the Calvin cycle enzymes EC 3.1.3.11 (fructose-bisphosphatase), EC 3.1.3.37 (sedoheptulose-bisphosphatase) and EC 2.7.1.19 (phosphoribulokinase) as well as other chloroplast enzymes by disulfide reduction.
References: [484, 680, 4004]

[EC 1.8.7.2 created 2010]

EC 1.8.7.3

Accepted name: ferredoxin:CoB-CoM heterodisulfide reductase
Reaction: 2 oxidized ferredoxin [iron-sulfur] cluster + CoB + CoM = 2 reduced ferredoxin [iron-sulfur] cluster + CoM-S-S-CoB + 2 H⁺
Other name(s): *hdrABC* (gene names); *hdrA1B1C1* (gene names); *hdrA2B2C2* (gene names)
Systematic name: CoB,CoM:ferredoxin oxidoreductase
Comments: HdrABC is an enzyme complex that is found in most methanogens and catalyses the reduction of the CoB-CoM heterodisulfide back to CoB and CoM. HdrA contains a FAD cofactor that acts as the entry point for electrons, which are transferred via HdrC to the HdrB catalytic subunit. One form of the enzyme from *Methanosarcina acetivorans* (HdrA2B2C2) can also catalyse EC 1.8.98.4, coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase. *cf.* EC 1.8.98.5, H₂:CoB-CoM heterodisulfide,ferredoxin reductase, EC 1.8.98.6, formate:CoB-CoM heterodisulfide,ferredoxin reductase, and EC 1.8.98.1, dihydromethanophenazine:CoB-CoM heterodisulfide reductase.

References: [480, 4758]

[EC 1.8.7.3 created 2017]

EC 1.8.98 With other, known, physiological acceptors

EC 1.8.98.1

Accepted name: dihydromethanophenazine:CoB-CoM heterodisulfide reductase
Reaction: CoB + CoM + methanophenazine = CoM-S-S-CoB + dihydromethanophenazine
Other name(s): *hdrDE* (gene names); CoB—CoM heterodisulfide reductase (ambiguous); heterodisulfide reductase (ambiguous); coenzyme B:coenzyme M:methanophenazine oxidoreductase
Systematic name: CoB:CoM:methanophenazine oxidoreductase
Comments: This enzyme, found in methanogenic archaea that belong to the *Methanosarcinales* order, regenerates CoM and CoB after the action of EC 2.8.4.1, coenzyme-B sulfoethylthiotransferase. It is a membrane-bound enzyme that contains (per heterodimeric unit) two distinct *b*-type hemes and two [4Fe-4S] clusters. *cf.* EC 1.8.7.3, ferredoxin:CoB-CoM heterodisulfide reductase, EC 1.8.98.5, H₂:CoB-CoM heterodisulfide,ferredoxin reductase, EC 1.8.98.6, formate:CoB-CoM heterodisulfide,ferredoxin reductase and EC 1.8.98.4, coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase.
References: [1598, 4, 3904, 2936]

[EC 1.8.98.1 created 2003, modified 2017]

EC 1.8.98.2

Accepted name: sulfiredoxin
Reaction: peroxiredoxin-(*S*-hydroxy-*S*-oxocysteine) + ATP + 2 R-SH = peroxiredoxin-(*S*-hydroxycysteine) + ADP + phosphate + R-S-S-R
Other name(s): Srx1; sulphiredoxin; peroxiredoxin-(*S*-hydroxy-*S*-oxocysteine) reductase
Systematic name: peroxiredoxin-(*S*-hydroxy-*S*-oxocysteine):thiol oxidoreductase [ATP-hydrolysing; peroxiredoxin-(*S*-hydroxycysteine)-forming]
Comments: In the course of the reaction of EC 1.11.1.15, peroxiredoxin, its cysteine residue is alternately oxidized to the sulfenic acid, *S*-hydroxycysteine, and reduced back to cysteine. Occasionally the *S*-hydroxycysteine residue is further oxidized to the sulfinic acid *S*-hydroxy-*S*-oxocysteine, thereby inactivating the enzyme. The reductase provides a mechanism for regenerating the active form of peroxiredoxin, i.e. the peroxiredoxin-(*S*-hydroxycysteine) form. Apparently the reductase first catalyses the phosphorylation of the -S(O)-OH group by ATP to give -S(O)-O-P, which is attached to the peroxiredoxin by a cysteine residue, forming an -S(O)-S- link between the two enzymes. Attack by a thiol splits this bond, leaving the peroxiredoxin as the sulfenic acid and the reductase as the thiol.
References: [338, 599, 4663]

[EC 1.8.98.2 created 2005]

EC 1.8.98.3

Accepted name: sulfite reductase (coenzyme F₄₂₀)
Reaction: hydrogen sulfide + 3 oxidized coenzyme F₄₂₀ + 3 H₂O = sulfite + 3 reduced coenzyme F₄₂₀
Other name(s): coenzyme F₄₂₀-dependent sulfite reductase; Fsr
Systematic name: hydrogen sulfide:coenzyme F₄₂₀ oxidoreductase
Comments: The enzyme, isolated from the archaeon *Methanocaldococcus jannaschii*, is involved in sulfite detoxification and assimilation.
References: [1928, 1929]

[EC 1.8.98.3 created 2014]

EC 1.8.98.4

- Accepted name:** coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase
- Reaction:** 2 oxidized coenzyme F₄₂₀ + 2 reduced ferredoxin [iron-sulfur] cluster + CoB + CoM + 2 H⁺ = 2 reduced coenzyme F₄₂₀ + 2 oxidized ferredoxin [iron-sulfur] cluster + CoM-S-S-CoB
- Other name(s):** *hdrA2B2C2* (gene names)
- Systematic name:** CoB,CoM,ferredoxin:coenzyme F₄₂₀ oxidoreductase
- Comments:** The enzyme, characterized from the archaeon *Methanosarcina acetivorans*, catalyses the reduction of CoB-CoM heterodisulfide back to CoB and CoM. The enzyme consists of three components, HdrA, HdrB and HdrC, all of which contain [4Fe-4S] clusters. Electrons enter at HdrA, which also contains FAD, and are transferred via HdrC to the catalytic component, HdrB. During methanogenesis from acetate the enzyme catalyses the activity of EC 1.8.7.3, ferredoxin:CoB-CoM heterodisulfide reductase. However, it can also use electron bifurcation to direct electron pairs from reduced coenzyme F₄₂₀ towards the reduction of both ferredoxin and CoB-CoM heterodisulfide. This activity is proposed to take place during Fe(III)-dependent anaerobic methane oxidation. *cf.* EC 1.8.98.5, H₂:CoB-CoM heterodisulfide,ferredoxin reductase, EC 1.8.98.6, formate:CoB-CoM heterodisulfide,ferredoxin reductase, and EC 1.8.98.1, dihydromethanophenazine:CoB-CoM heterodisulfide reductase.
- References:** [4758]

[EC 1.8.98.4 created 2017]

EC 1.8.98.5

- Accepted name:** H₂:CoB-CoM heterodisulfide,ferredoxin reductase
- Reaction:** 2 reduced ferredoxin [iron-sulfur] cluster + CoB + CoM + 2 H⁺ = 2 H₂ + 2 oxidized ferredoxin [iron-sulfur] cluster + CoM-S-S-CoB
- Systematic name:** CoB,CoM,ferredoxin:H₂ oxidoreductase
- Comments:** This enzyme complex is found in H₂-oxidizing CO₂-reducing methanogenic archaea such as *Methanothermobacter thermautotrophicus*. It consists of a cytoplasmic complex of HdrABC reductase and MvhAGD hydrogenase. Electron pairs donated by the hydrogenase are transferred via its δ subunit to the HdrA subunit of the reductase, where they are bifurcated, reducing both ferredoxin and CoB-CoM heterodisulfide. The reductase can also form a similar complex with formate dehydrogenase, see EC 1.8.98.6, formate:CoB-CoM heterodisulfide,ferredoxin reductase. *cf.* EC 1.8.7.3, ferredoxin:CoB-CoM heterodisulfide reductase, EC 1.8.98.4, coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase, and EC 1.8.98.1, dihydromethanophenazine:CoB-CoM heterodisulfide reductase.
- References:** [3482, 1599, 3805, 4044, 2007, 745]

[EC 1.8.98.5 created 2017]

EC 1.8.98.6

- Accepted name:** formate:CoB-CoM heterodisulfide,ferredoxin reductase
- Reaction:** 2 CO₂ + 2 reduced ferredoxin [iron-sulfur] cluster + CoB + CoM + 2 H⁺ = 2 formate + 2 oxidized ferredoxin [iron-sulfur] cluster + CoM-S-S-CoB
- Systematic name:** coenzyme B,coenzyme M,ferredoxin:formate oxidoreductase
- Comments:** The enzyme is found in formate-oxidizing CO₂-reducing methanogenic archaea such as *Methanococcus maripaludis*. It consists of a cytoplasmic complex of HdrABC reductase and formate dehydrogenase. Electron pairs donated by formate dehydrogenase are transferred to the HdrA subunit of the reductase, where they are bifurcated, reducing both ferredoxin and CoB-CoM heterodisulfide. *cf.* EC 1.8.7.3, ferredoxin:CoB-CoM heterodisulfide reductase, EC 1.8.98.4, coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase, EC 1.8.98.5, H₂:CoB-CoM heterodisulfide,ferredoxin reductase, and EC 1.8.98.1, dihydromethanophenazine:CoB-CoM heterodisulfide reductase.
- References:** [746, 745]

[EC 1.8.98.6 created 2017]

EC 1.8.98.7

- Accepted name:** cysteine-type anaerobic sulfatase-maturing enzyme
Reaction: S -adenosyl-L-methionine + a [sulfatase]-L-cysteine + H_2O = a [sulfatase]-3-oxo-L-alanine + 5'-deoxyadenosine + L-methionine + hydrogen sulfide
Other name(s): anSME; Cys-type anaerobic sulfatase-maturing enzyme; anaerobic sulfatase maturase
Systematic name: [sulfatase]-L-cysteine: S -adenosyl-L-methionine oxidoreductase (3-oxo-L-alanine-forming)
Comments: A radical S -adenosylmethionine (AdoMet) enzyme that contains three [4Fe-4S] clusters. The enzyme, found in some bacteria, activates a type I sulfatase enzyme (EC 3.1.6.1) by converting a conserved L-cysteine residue in the active site to a unique 3-oxo-L-alanine residue that is essential for the sulfatase activity. Some enzymes can also act on L-serine, see EC 1.1.98.7, serine-type anaerobic sulfatase-maturing enzyme and EC 1.8.3.7, formylglycine-generating enzyme.
References: [311, 287, 286, 288, 1430]

[EC 1.8.98.7 created 2020]

EC 1.8.99 With unknown physiological acceptors

[1.8.99.1 Deleted entry. sulfite reductase. Now covered by EC 1.8.1.2, assimilatory sulfite reductase (NADPH) and EC 1.8.7.1, assimilatory sulfite reductase (ferredoxin).]

[EC 1.8.99.1 created 1972, deleted 2015]

EC 1.8.99.2

- Accepted name:** adenylyl-sulfate reductase
Reaction: AMP + sulfite + acceptor = adenylyl sulfate + reduced acceptor
Other name(s): adenosine phosphosulfate reductase; adenosine 5'-phosphosulfate reductase; APS-reductase; APS reductase; AMP, sulfite:(acceptor) oxidoreductase (adenosine-5'-phosphosulfate-forming)
Systematic name: AMP,sulfite:acceptor oxidoreductase (adenosine-5'-phosphosulfate-forming)
Comments: An iron flavoprotein (FAD). Methyl viologen can act as acceptor.
References: [2786]

[EC 1.8.99.2 created 1972]

[1.8.99.3 Deleted entry. hydrogensulfite reductase, now known to be an in vitro artifact of EC 1.8.99.5, dissimilatory sulfite reductase]

[EC 1.8.99.3 created 1986, deleted 2016]

[1.8.99.4 Transferred entry. phosphoadenosine-phosphosulfate reductase. Now EC 1.8.4.8, phosphoadenylyl-sulfate reductase (thioredoxin)]

[EC 1.8.99.4 created 1999, deleted 2000]

EC 1.8.99.5

- Accepted name:** dissimilatory sulfite reductase
Reaction: (1) hydrogen sulfide + a [DsrC protein]-disulfide + 2 acceptor + 3 H_2O = sulfite + a [DsrC protein]-dithiol + 2 reduced acceptor + 2 H^+ (overall reaction)
(1a) hydrogen sulfide + a [DsrC protein]-disulfide = a [DsrC protein]- S -sulfanyl-L-cysteine
(1b) a [DsrC protein]- S -sulfanyl-L-cysteine + 2 acceptor + 3 H_2O = sulfite + a [DsrC protein]-dithiol + 2 reduced acceptor + 2 H^+
(2) a [DsrC protein]- S -sulfanyl-L-cysteine + 3 acceptor + 3 H_2O = sulfite + a [DsrC protein]-disulfide + 3 reduced acceptor + 2 H^+ (overall reaction)
(2a) a [DsrC protein]- S -sulfanyl-L-cysteine + 3 acceptor + 3 H_2O = a [DsrC]- S -sulfo-L-cysteine + 3 reduced acceptor + H^+
(2b) a [DsrC]- S -sulfo-L-cysteine = sulfite + a [DsrC protein]-disulfide

Other name(s): siroheme sulfite reductase; hydrogen-sulfide:(acceptor) oxidoreductase (ambiguous); DsrAB
Systematic name: hydrogen-sulfide:[DsrC sulfur-carrier protein],acceptor oxidoreductase
Comments: Contain siroheme. The enzyme is essential in prokaryotic sulfur-based energy metabolism, including sulfate/sulfite reducing organisms, sulfur-oxidizing bacteria, and organosulfonate reducers. In sulfur reducers it catalyses the reduction of sulfite to sulfide (reaction 1 in the right to left direction), while in sulfur oxidizers it catalyses the opposite reaction (reaction 2 in the left to right direction) [3706]. The reaction involves the small protein DsrC, which is present in all the organisms that contain dissimilatory sulfite reductase. During the process an intramolecular disulfide bond is formed between two L-cysteine residues of DsrC. This disulfide can be reduced by a number of proteins including DsrK and TcmB [4432]. This enzyme is different from EC 1.8.1.2, assimilatory sulfite reductase (NADPH), and EC 1.8.7.1, assimilatory sulfite reductase (ferredoxin), which are involved in sulfate assimilation.
References: [3706, 3792, 3358, 3169, 4432]

[EC 1.8.99.5 created 2015]

EC 1.9 Acting on a heme group of donors

This subclass contains the cytochrome oxidases and nitrate reductases. Sub-subclasses are based on the acceptor: oxygen (EC 1.9.3), a nitrogenous group (EC 1.9.6), or some other acceptor (EC 1.9.99).

EC 1.9.3 With oxygen as acceptor

[1.9.3.1 *Transferred entry. cytochrome-c oxidase. Now EC 7.1.1.9, cytochrome-c oxidase*]

[EC 1.9.3.1 created 1961, modified 2000, deleted 2019]

[1.9.3.2 *Transferred entry. Pseudomonas cytochrome oxidase. Now included with EC 1.7.2.1, nitrite reductase (NO-forming)*]

[EC 1.9.3.2 created 1965, deleted 2002]

EC 1.9.6 With a nitrogenous group as acceptor

EC 1.9.6.1

Accepted name: nitrate reductase (cytochrome)
Reaction: 2 ferrocycytochrome + 2 H⁺ + nitrate = 2 ferricytochrome + nitrite
Other name(s): respiratory nitrate reductase; benzyl viologen-nitrate reductase
Systematic name: ferrocycytochrome:nitrate oxidoreductase
References: [3622]

[EC 1.9.6.1 created 1961]

EC 1.9.98 With other, known, physiological acceptors

EC 1.9.98.1

Accepted name: iron—cytochrome-*c* reductase
Reaction: ferrocycytochrome *c* + Fe³⁺ = ferricytochrome *c* + Fe²⁺
Other name(s): iron-cytochrome *c* reductase
Systematic name: ferrocycytochrome-*c*:Fe³⁺ oxidoreductase
Comments: An iron protein.
References: [4782]

[EC 1.9.98.1 created 1972 as EC 1.9.99.1, transferred 2014 to EC 1.9.98.1]

EC 1.9.99 With unknown physiological acceptors

[1.9.99.1 *Transferred entry. iron—cytochrome-c reductase. Now EC 1.9.98.1, iron—cytochrome-c reductase*]

[EC 1.9.99.1 created 1972, deleted 2014]

EC 1.10 Acting on diphenols and related substances as donors

This subclass contains enzymes that catalyse the oxidation of diphenols or ascorbate. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.10.1), a cytochrome (EC 1.10.2), oxygen (EC 1.10.3), or some other acceptor (EC 1.10.99). Some enzymes that catalyse the oxidation of phenols are oxygenases (EC 1.14.18).

EC 1.10.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.10.1.1

Accepted name: *trans*-acenaphthene-1,2-diol dehydrogenase
Reaction: (±)-*trans*-acenaphthene-1,2-diol + 2 NADP⁺ = acenaphthenequinone + 2 NADPH + 2 H⁺
Other name(s): *trans*-1,2-acenaphthenediol dehydrogenase
Systematic name: (±)-*trans*-acenaphthene-1,2-diol:NADP⁺ oxidoreductase
Comments: Some preparations also utilize NAD⁺.
References: [1713]

[EC 1.10.1.1 created 1976]

EC 1.10.2 With a cytochrome as acceptor

[1.10.2.1 *Deleted entry. L-ascorbate—cytochrome-b₅ reductase. The activity is covered by EC 7.2.1.3, ascorbate ferrireductase (transmembrane)*]

[EC 1.10.2.1 created 1972, modified 2000, deleted 2021]

[1.10.2.2 *Transferred entry. quinol—cytochrome-c reductase. Now EC 7.1.1.8, quinol—cytochrome-c reductase*]

[EC 1.10.2.2 created 1978, modified 2013, deleted 2018]

EC 1.10.3 With oxygen as acceptor

EC 1.10.3.1

Accepted name: catechol oxidase
Reaction: 2 catechol + O₂ = 2 1,2-benzoquinone + 2 H₂O
Other name(s): diphenol oxidase; *o*-diphenolase; polyphenol oxidase; pyrocatechol oxidase; dopa oxidase; catecholase; *o*-diphenol:oxygen oxidoreductase; *o*-diphenol oxidoreductase
Systematic name: 1,2-benzenediol:oxygen oxidoreductase
Comments: A type 3 copper protein that catalyses exclusively the oxidation of catechol (i.e., *o*-diphenol) to the corresponding *o*-quinone. The enzyme also acts on a variety of substituted catechols. It is different from tyrosinase, EC 1.14.18.1, which can catalyse both the monooxygenation of monophenols and the oxidation of catechols.
References: [459, 842, 1405, 2682, 2729, 3261, 3350, 3533, 1302]

[EC 1.10.3.1 created 1961, deleted 1972, reinstated 1978]

EC 1.10.3.2

Accepted name: laccase
Reaction: $4 \text{ benzenediol} + \text{O}_2 = 4 \text{ benzosemiquinone} + 2 \text{ H}_2\text{O}$
Other name(s): urishiol oxidase; urushiol oxidase; *p*-diphenol oxidase
Systematic name: benzenediol:oxygen oxidoreductase
Comments: A group of multi-copper proteins of low specificity acting on both *o*- and *p*-quinols, and often acting also on aminophenols and phenylenediamine. The semiquinone may react further either enzymically or non-enzymically.
References: [842, 2057, 2633, 2729, 2981, 2982, 3282, 3492]

[EC 1.10.3.2 created 1961, deleted 1972, reinstated 1978]

EC 1.10.3.3

Accepted name: L-ascorbate oxidase
Reaction: $4 \text{ L-ascorbate} + \text{O}_2 = 4 \text{ monodehydroascorbate} + 2 \text{ H}_2\text{O}$
Other name(s): ascorbase; ascorbic acid oxidase; ascorbate oxidase; ascorbic oxidase; ascorbate dehydrogenase; L-ascorbic acid oxidase; AAO; L-ascorbate:O₂ oxidoreductase; AA oxidase
Systematic name: L-ascorbate:oxygen oxidoreductase
Comments: A multicopper protein.
References: [4753, 4006, 2779]

[EC 1.10.3.3 created 1961, modified 2011]

EC 1.10.3.4

Accepted name: *o*-aminophenol oxidase
Reaction: $4 \text{ 2-aminophenol} + 3 \text{ O}_2 = 2 \text{ 2-aminophenoxazin-3-one} + 6 \text{ H}_2\text{O}$
Other name(s): isophenoxazine synthase; *o*-aminophenol:O₂ oxidoreductase; 2-aminophenol:O₂ oxidoreductase
Systematic name: 2-aminophenol:oxygen oxidoreductase
Comments: A flavoprotein which catalyses a 6-electron oxidation. The enzyme from the plant *Tecoma stans* requires Mn²⁺ and FAD [2969] whereas the fungus *Pycnoporus coccineus* requires Mn²⁺ and riboflavin 5'-phosphate [2971], the bacteria *Streptomyces antibioticus* requires Cu²⁺ [227] and the plant *Bauhinia monandra* does not require any co-factors [3452].
References: [2969, 2971, 3452, 227]

[EC 1.10.3.4 created 1972, modified 2006]

EC 1.10.3.5

Accepted name: 3-hydroxyanthranilate oxidase
Reaction: $3\text{-hydroxyanthranilate} + \text{O}_2 = 6\text{-imino-5-oxocyclohexa-1,3-dienecarboxylate} + \text{H}_2\text{O}_2$
Other name(s): 3-hydroxyanthranilic acid oxidase
Systematic name: 3-hydroxyanthranilate:oxygen oxidoreductase
References: [2884]

[EC 1.10.3.5 created 1972]

EC 1.10.3.6

Accepted name: rifamycin-B oxidase
Reaction: $\text{rifamycin B} + \text{O}_2 = \text{rifamycin O} + \text{H}_2\text{O}_2$
Other name(s): rifamycin B oxidase
Systematic name: rifamycin-B:oxygen oxidoreductase
Comments: Acts also on benzene-1,4-diol and, more slowly, on some other *p*-quinols. Not identical with EC 1.10.3.1 (catechol oxidase), EC 1.10.3.2 (laccase), EC 1.10.3.4 (*o*-aminophenol oxidase) or EC 1.10.3.5 (3-hydroxyanthranilate oxidase).

References: [1501]

[EC 1.10.3.6 created 1986]

[1.10.3.7 *Transferred entry. sulochrin oxidase [(+)-bisdechlorogeodin-forming]. Now EC 1.21.3.4, sulochrin oxidase [(+)-bisdechlorogeodin-forming]]*

[EC 1.10.3.7 created 1986, deleted 2002]

[1.10.3.8 *Transferred entry. sulochrin oxidase [(+)-bisdechlorogeodin-forming]. Now EC 1.21.3.5, sulochrin oxidase [(-)-bisdechlorogeodin-forming]]*

[EC 1.10.3.8 created 1986, deleted 2002]

EC 1.10.3.9

Accepted name: photosystem II
Reaction: $2 \text{H}_2\text{O} + 2 \text{ plastoquinone} + 4 h\nu = \text{O}_2 + 2 \text{ plastoquinol}$
Systematic name: H₂O:plastoquinone reductase (light-dependent)
Comments: Contains chlorophyll *a*, β-carotene, pheophytin, plastoquinone, a Mn₄Ca cluster, heme and non-heme iron. Four successive photoreactions, resulting in a storage of four positive charges, are required to oxidize two water molecules to one oxygen molecule.
References: [2163, 1454]

[EC 1.10.3.9 created 2011]

[1.10.3.10 *Transferred entry. ubiquinol oxidase (H⁺-transporting). Now EC 7.1.1.3, ubiquinol oxidase (H⁺-transporting)]*

[EC 1.10.3.10 created 2011, modified 2014, deleted 2018]

EC 1.10.3.11

Accepted name: ubiquinol oxidase (non-electrogenic)
Reaction: $2 \text{ ubiquinol} + \text{O}_2 = 2 \text{ ubiquinone} + 2 \text{H}_2\text{O}$
Other name(s): plant alternative oxidase; cyanide-insensitive oxidase; AOX (gene name); ubiquinol oxidase; ubiquinol:O₂ oxidoreductase (non-electrogenic)
Systematic name: ubiquinol:oxygen oxidoreductase (non-electrogenic)
Comments: The enzyme, described from the mitochondria of plants and some fungi and protists, is *an* alternative terminal oxidase that is not sensitive to cyanide inhibition and does not generate a proton motive force. Unlike the electrogenic terminal oxidases that contain hemes (*cf.* EC 1.10.3.10 and EC 1.10.3.14), this enzyme contains a dinuclear non-heme iron complex. The function of this oxidase is believed to be dissipating excess reducing power, minimizing oxidative stress, and optimizing photosynthesis in response to changing conditions.
References: [285, 3900, 314, 4629, 1269]

[EC 1.10.3.11 created 2011, modified 2014]

[1.10.3.12 *Transferred entry. menaquinol oxidase (H⁺-transporting). Now EC 7.1.1.5, menaquinol oxidase (H⁺-transporting)]*

[EC 1.10.3.12 created 2011, deleted 2018]

[1.10.3.13 *Transferred entry. caldariellaquinol oxidase (H⁺-transporting). Now EC 7.1.1.4, caldariellaquinol oxidase (H⁺-transporting)]*

[EC 1.10.3.13 created 2013, deleted 2018]

[1.10.3.14 *Transferred entry. ubiquinol oxidase (electrogenic, non H⁺-transporting). Now EC 7.1.1.7, ubiquinol oxidase (electrogenic, proton-motive force generating)]*

[EC 1.10.3.14 created 2014, modified 2017, deleted 2018]

EC 1.10.3.15

- Accepted name:** grixazone synthase
Reaction: 2 3-amino-4-hydroxybenzoate + *N*-acetyl-L-cysteine + 2 O₂ = grixazone B + 4 H₂O + CO₂
Other name(s): GriF
Systematic name: 3-amino-4-hydroxybenzoate:*N*-acetyl-L-cysteine:oxygen oxidoreductase
Comments: A type 3 multi copper protein. The enzyme, isolated from the bacterium *Streptomyces griseus*, catalyses an 8 electron oxidation. Activation of the enzyme requires a copper chaperone (GriE). It also acts on 3-amino-4-hydroxybenzaldehyde, giving grixazone A. The second aldehyde group is presumably lost as formate. The enzyme also catalyses the reaction of EC 1.10.3.4 *o*-aminophenol oxidase.
References: [4135, 3553]

[EC 1.10.3.15 created 2014]

EC 1.10.3.16

- Accepted name:** dihydrophenazinedicarboxylate synthase
Reaction:
(1) (1*R*,6*R*)-1,4,5,5a,6,9-hexahydrophenazine-1,6-dicarboxylate + O₂ = (1*R*,10a*S*)-1,4,10,10a-tetrahydrophenazine-1,6-dicarboxylate + H₂O₂
(2) (1*R*,10a*S*)-1,4,10,10a-tetrahydrophenazine-1,6-dicarboxylate + O₂ = (5a*S*)-5,5a-dihydrophenazine-1,6-dicarboxylate + H₂O₂
(3) (1*R*,10a*S*)-1,4,10,10a-tetrahydrophenazine-1-carboxylate + O₂ = (10a*S*)-10,10a-dihydrophenazine-1-carboxylate + H₂O₂
(4) (1*R*)-1,4,5,10-tetrahydrophenazine-1-carboxylate + O₂ = (10a*S*)-5,10-dihydrophenazine-1-carboxylate + H₂O₂
Other name(s): *phzG* (gene name)
Systematic name: 1,4,5a,6,9,10a-hexahydrophenazine-1,6-dicarboxylate:oxygen oxidoreductase
Comments: Requires FMN. The enzyme, isolated from the bacteria *Pseudomonas fluorescens* 2-79 and *Burkholderia lata* 383, is involved in biosynthesis of the reduced forms of phenazine, phenazine-1-carboxylate, and phenazine-1,6-dicarboxylate, where it catalyses multiple reactions.
References: [4706]

[EC 1.10.3.16 created 2016]

EC 1.10.3.17

- Accepted name:** superoxide oxidase
Reaction: 2 O₂ + ubiquinol = 2 superoxide + ubiquinone + 2 H⁺
Other name(s): SOO; CybB; cytochrome *b*₅₆₁; superoxide:ubiquinone oxidoreductase
Systematic name: ubiquinol:oxygen oxidoreductase (superoxide-forming)
Comments: This membrane-bound, di-heme containing enzyme, identified in the bacterium *Escherichia coli*, is responsible for the detoxification of superoxide in the periplasm. *In vivo* the reaction proceeds in the opposite direction of that shown and produces oxygen. Superoxide production was only observed when the enzyme was incubated *in vitro* with an excess of ubiquinol.
References: [2937, 2938, 2567]

[EC 1.10.3.17 created 2019]

EC 1.10.5 With a quinone or related compound as acceptor

EC 1.10.5.1

- Accepted name:** ribosyldihydronicotinamide dehydrogenase (quinone)
Reaction: 1-(β-D-ribofuranosyl)-1,4-dihydronicotinamide + a quinone = 1-(β-D-ribofuranosyl)nicotinamide + a quinol

Other name(s): NRH:quinone oxidoreductase 2; NQO2; NAD(P)H:quinone oxidoreductase-2 (misleading); QR2; quinone reductase 2; *N*-ribosyl-dihydronicotinamide dehydrogenase (quinone); NAD(P)H:quinone oxidoreductase2 (misleading)

Systematic name: 1-(β-D-ribofuranosyl)-1,4-dihydronicotinamide:quinone oxidoreductase

Comments: A flavoprotein. Unlike EC 1.6.5.2, NAD(P)H dehydrogenase (quinone), this quinone reductase cannot use NADH or NADPH; instead it uses *N*-ribosyl- and *N*-alkyldihydronicotinamides. Polycyclic aromatic hydrocarbons, such as benz[*a*]anthracene, and the estrogens 17β-estradiol and diethylstilbestrol are potent inhibitors, but dicoumarol is only a very weak inhibitor [4902]. This enzyme can catalyse both 2-electron and 4-electron reductions, but one-electron acceptors, such as potassium ferricyanide, cannot be reduced [4676].

References: [2473, 4902, 4676, 1877]

[EC 1.10.5.1 created 2005 as EC 1.10.99.2, transferred 2015 to EC 1.10.5.1]

EC 1.10.9 With a copper protein as acceptor

[1.10.9.1 *Transferred entry. plastoquinol—plastocyanin reductase. Now EC 7.1.1.6, plastoquinol—plastocyanin reductase*]

[EC 1.10.9.1 created 1984 as EC 1.10.99.1, transferred 2011 to EC 1.10.9.1, deleted 2018]

EC 1.10.99 With unknown physiological acceptors

[1.10.99.1 *Transferred entry. Now EC 1.10.9.1 plastoquinol—plastocyanin reductase*]

[EC 1.10.99.1 created 1984, deleted 2011]

[1.10.99.2 *Transferred entry. ribosyl-dihydronicotinamide dehydrogenase (quinone). Now classified as EC 1.10.5.1, ribosyl-dihydronicotinamide dehydrogenase (quinone).*]

[EC 1.10.99.2 created 2005, deleted 2014]

[1.10.99.3 *Transferred entry. violaxanthin de-epoxidase. Now classified as EC 1.23.5.1, violaxanthin de-epoxidase.*]

[EC 1.10.99.3 created 2005, deleted 2014]

EC 1.11 Acting on a peroxide as acceptor

This subclass contains two sub-subclasses: the peroxidases (EC 1.11.1) and the peroxygenases (EC 1.11.2).

EC 1.11.1 Peroxidases

Acting on a peroxide as acceptor (peroxidases)

EC 1.11.1.1

Accepted name: NADH peroxidase

Reaction: $\text{NADH} + \text{H}^+ + \text{H}_2\text{O}_2 = \text{NAD}^+ + 2 \text{H}_2\text{O}$

Other name(s): DPNH peroxidase; NAD peroxidase; diphosphopyridine nucleotide peroxidase; NADH-peroxidase; nicotinamide adenine dinucleotide peroxidase; NADH₂ peroxidase

Systematic name: NADH:hydrogen-peroxide oxidoreductase

Comments: A flavoprotein (FAD). Ferricyanide, quinones, etc., can replace H₂O₂.

References: [940, 2850, 4497]

[EC 1.11.1.1 created 1961]

EC 1.11.1.2

Accepted name: NADPH peroxidase
Reaction: $\text{NADPH} + \text{H}^+ + \text{H}_2\text{O}_2 = \text{NADP}^+ + 2 \text{H}_2\text{O}$
Other name(s): TPNH peroxidase; NADP peroxidase; nicotinamide adenine dinucleotide phosphate peroxidase; TPN peroxidase; triphosphopyridine nucleotide peroxidase; NADPH₂ peroxidase
Systematic name: NADPH:hydrogen-peroxide oxidoreductase
References: [720]

[EC 1.11.1.2 created 1961]

EC 1.11.1.3

Accepted name: fatty-acid peroxidase
Reaction: $\text{palmitate} + 2 \text{H}_2\text{O}_2 = \text{pentadecanal} + \text{CO}_2 + 3 \text{H}_2\text{O}$
Other name(s): long chain fatty acid peroxidase
Systematic name: hexadecanoate:hydrogen-peroxide oxidoreductase
Comments: Acts on long-chain fatty acids from dodecanoic to octadecanoic acid.
References: [2668]

[EC 1.11.1.3 created 1961]

[1.11.1.4 Transferred entry. now EC 1.13.11.11 tryptophan 2,3-dioxygenase]

[EC 1.11.1.4 created 1961, deleted 1964, reinstated 1965 as EC 1.13.1.12, deleted 1972]

EC 1.11.1.5

Accepted name: cytochrome-*c* peroxidase
Reaction: $2 \text{ferrocycytochrome } c + \text{H}_2\text{O}_2 = 2 \text{ferricytochrome } c + 2 \text{H}_2\text{O}$
Other name(s): cytochrome peroxidase; cytochrome *c*-551 peroxidase; apocytochrome *c* peroxidase; mesocytochrome *c* peroxidase azide; mesocytochrome *c* peroxidase cyanide; mesocytochrome *c* peroxidase cyanate; cytochrome *c*-H₂O oxidoreductase; cytochrome *c* peroxidase
Systematic name: ferrocycytochrome-*c*:hydrogen-peroxide oxidoreductase
Comments: A hemoprotein.
References: [75, 4746, 4800]

[EC 1.11.1.5 created 1961]

EC 1.11.1.6

Accepted name: catalase
Reaction: $2 \text{H}_2\text{O}_2 = \text{O}_2 + 2 \text{H}_2\text{O}$
Other name(s): equilase; caperase; optidase; catalase-peroxidase; CAT
Systematic name: hydrogen-peroxide:hydrogen-peroxide oxidoreductase
Comments: A hemoprotein. A manganese protein containing Mn^{III} in the resting state, which also belongs here, is often called pseudocatalase. The enzymes from some organisms, such as *Penicillium simplicissimum*, can also act as a peroxidase (EC 1.11.1.7) for which several organic substances, especially ethanol, can act as a hydrogen donor. Enzymes that exhibit both catalase and peroxidase activity belong under EC 1.11.1.21, catalase-peroxidase.
References: [1628, 1629, 2054, 2224, 3064, ?]

[EC 1.11.1.6 created 1961, modified 1986, modified 1999, modified 2013]

EC 1.11.1.7

Accepted name: peroxidase
Reaction: $2 \text{phenolic donor} + \text{H}_2\text{O}_2 = 2 \text{phenoxy radical of the donor} + 2 \text{H}_2\text{O}$

Other name(s): lactoperoxidase; guaiacol peroxidase; plant peroxidase; Japanese radish peroxidase; horseradish peroxidase (HRP); soybean peroxidase (SBP); extensin peroxidase; heme peroxidase; oxypoxidase; protoheme peroxidase; pyrocatechol peroxidase; scopoletin peroxidase; *Coprinus cinereus* peroxidase; *Arthromyces ramosus* peroxidase

Systematic name: phenolic donor:hydrogen-peroxide oxidoreductase

Comments: Heme proteins with histidine as proximal ligand. The iron in the resting enzyme is Fe(III). They also peroxidize non-phenolic substrates such as 3,3',5,5'-tetramethylbenzidine (TMB) and 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS). Certain peroxidases (e.g. lactoperoxidase, SBP) oxidize bromide, iodide and thiocyanate.

References: [2065, 2900, 3264, 4162, 4256, 1089, 44, 984, 4314]

[EC 1.11.1.7 created 1961, modified 2011]

EC 1.11.1.8

Accepted name: iodide peroxidase

Reaction: (1) $2 \text{ iodide} + \text{H}_2\text{O}_2 + 2 \text{H}^+ = \text{diiodine} + 2 \text{H}_2\text{O}$
(2) $[\text{thyroglobulin}]\text{-L-tyrosine} + \text{iodide} + \text{H}_2\text{O}_2 = [\text{thyroglobulin}]\text{-3-iodo-L-tyrosine} + 2 \text{H}_2\text{O}$
(3) $[\text{thyroglobulin}]\text{-3-iodo-L-tyrosine} + \text{iodide} + \text{H}_2\text{O}_2 = [\text{thyroglobulin}]\text{-3,5-diiodo-L-tyrosine} + 2 \text{H}_2\text{O}$
(4) $2 [\text{thyroglobulin}]\text{-3,5-diiodo-L-tyrosine} + \text{H}_2\text{O}_2 = [\text{thyroglobulin}]\text{-L-thyroxine} + [\text{thyroglobulin}]\text{-aminoacrylate} + 2 \text{H}_2\text{O}$
(5) $[\text{thyroglobulin}]\text{-3-iodo-L-tyrosine} + [\text{thyroglobulin}]\text{-3,5-diiodo-L-tyrosine} + \text{H}_2\text{O}_2 = [\text{thyroglobulin}]\text{-3,5,3'-triiodo-L-thyronine} + [\text{thyroglobulin}]\text{-aminoacrylate} + 2 \text{H}_2\text{O}$

Other name(s): thyroid peroxidase; iodoperoxidase (heme type); iodide peroxidase-tyrosine iodinase; thyroperoxidase; tyrosine iodinase; TPO; iodinase

Systematic name: iodide:hydrogen-peroxide oxidoreductase

Comments: Thyroid peroxidase catalyses the biosynthesis of the thyroid hormones L-thyroxine and triiodo-L-thyronine. It catalyses both the iodination of tyrosine residues in thyroglobulin (forming mono- and di-iodinated forms) and their coupling to form either L-thyroxine or triiodo-L-thyronine.

References: [784, 1740, 755, 1289, 3149, 2611, 4453, 3464, 4119, 4222, 3599]

[EC 1.11.1.8 created 1961, modified 2012]

EC 1.11.1.9

Accepted name: glutathione peroxidase

Reaction: $2 \text{ glutathione} + \text{H}_2\text{O}_2 = \text{glutathione disulfide} + 2 \text{H}_2\text{O}$

Other name(s): GSH peroxidase; selenium-glutathione peroxidase; reduced glutathione peroxidase

Systematic name: glutathione:hydrogen-peroxide oxidoreductase

Comments: A protein containing a selenocysteine residue. Steroid and lipid hydroperoxides, but not the product of reaction of EC 1.13.11.12 lipoxygenase on phospholipids, can act as acceptor, but more slowly than H_2O_2 (cf. EC 1.11.1.12 phospholipid-hydroperoxide glutathione peroxidase).

References: [618, 1429, 2985]

[EC 1.11.1.9 created 1965, modified 1989]

EC 1.11.1.10

Accepted name: chloride peroxidase

Reaction: $\text{RH} + \text{chloride} + \text{H}_2\text{O}_2 = \text{RCl} + 2 \text{H}_2\text{O}$

Other name(s): chloroperoxidase; CPO; vanadium haloperoxidase

Systematic name: chloride:hydrogen-peroxide oxidoreductase

Comments: Brings about the chlorination of a range of organic molecules, forming stable C-Cl bonds. Also oxidizes bromide and iodide. Enzymes of this type are either heme-thiolate proteins, or contain vanadate. A secreted enzyme produced by the ascomycetous fungus *Caldariomyces fumago* (*Leptoxyphium fumago*) is an example of the heme-thiolate type. It catalyses the production of hypochlorous acid by transferring one oxygen atom from H₂O₂ to chloride. At a separate site it catalyses the chlorination of activated aliphatic and aromatic substrates, via HClO and derived chlorine species. In the absence of halides, it shows peroxidase (e.g. phenol oxidation) and peroxygenase activities. The latter inserts oxygen from H₂O₂ into, for example, styrene (side chain epoxidation) and toluene (benzylic hydroxylation), however, these activities are less pronounced than its activity with halides. Has little activity with non-activated substrates such as aromatic rings, ethers or saturated alkanes. The chlorinating peroxidase produced by ascomycetous fungi (e.g. *Curvularia inaequalis*) is an example of a vanadium chloroperoxidase, and is related to bromide peroxidase (EC 1.11.1.18). It contains vanadate and oxidizes chloride, bromide and iodide into hypohalous acids. In the absence of halides, it peroxygenates organic sulfides and oxidizes ABTS [2,2'-azinobis(3-ethylbenzthiazoline-6-sulfonic acid)] but no phenols.

References: [2899, 1474, 4252, 4123, 4241, 4240, 2637, 2277, 2638]

[EC 1.11.1.10 created 1972, modified 2011]

EC 1.11.1.11

Accepted name: L-ascorbate peroxidase

Reaction: 2 L-ascorbate + H₂O₂ + 2 H⁺ = L-ascorbate + L-dehydroascorbate + 2 H₂O (overall reaction)

(1a) 2 L-ascorbate + H₂O₂ + 2 H⁺ = 2 monodehydroascorbate + 2 H₂O

(1b) 2 monodehydroascorbate = L-ascorbate + L-dehydroascorbate (spontaneous)

Other name(s): L-ascorbic acid peroxidase; L-ascorbic acid-specific peroxidase; ascorbate peroxidase; ascorbic acid peroxidase

Systematic name: L-ascorbate:hydrogen-peroxide oxidoreductase

Comments: A heme protein. Oxidizes ascorbate and low molecular weight aromatic substrates. The monodehydroascorbate radical produced is either directly reduced back to ascorbate by EC 1.6.5.4 [monodehydroascorbate reductase (NADH)] or undergoes non-enzymic disproportionation to ascorbate and dehydroascorbate.

References: [3857, 3856, 2993, 3263, 3830, 2587]

[EC 1.11.1.11 created 1983, modified 2010, modified 2011]

EC 1.11.1.12

Accepted name: phospholipid-hydroperoxide glutathione peroxidase

Reaction: 2 glutathione + a hydroperoxy-fatty-acyl-[lipid] = glutathione disulfide + a hydroxy-fatty-acyl-[lipid] + H₂O

Other name(s): peroxidation-inhibiting protein; PHGPX; peroxidation-inhibiting protein:peroxidase,glutathione (phospholipid hydroperoxide-reducing); phospholipid hydroperoxide glutathione peroxidase; hydroperoxide glutathione peroxidase

Systematic name: glutathione:lipid-hydroperoxide oxidoreductase

Comments: A protein containing a selenocysteine residue. The products of action of EC 1.13.11.12 lipoxygenase on phospholipids can act as acceptors; H₂O₂ can also act, but much more slowly (*cf.* EC 1.11.1.9 glutathione peroxidase).

References: [4381, 3738]

[EC 1.11.1.12 created 1989, modified 2015]

EC 1.11.1.13

Accepted name: manganese peroxidase

Reaction: 2 Mn(II) + 2 H⁺ + H₂O₂ = 2 Mn(III) + 2 H₂O

Other name(s): peroxidase-M2; Mn-dependent (NADH-oxidizing) peroxidase
Systematic name: Mn(II):hydrogen-peroxide oxidoreductase
Comments: A hemoprotein. The enzyme from white rot basidiomycetes is involved in the oxidative degradation of lignin. The enzyme oxidizes a bound Mn²⁺ ion to Mn³⁺ in the presence of hydrogen peroxide. The product, Mn³⁺, is released from the active site in the presence of a chelator (mostly oxalate and malate) that stabilizes it against disproportionation to Mn²⁺ and insoluble Mn⁴⁺ [2269]. The complexed Mn³⁺ ion can diffuse into the lignified cell wall, where it oxidizes phenolic components of lignin and other organic substrates [1343]. It is inactive with veratryl alcohol or nonphenolic substrates.
References: [1343, 3252, 4544, 2269]

[EC 1.11.1.13 created 1992]

EC 1.11.1.14

Accepted name: lignin peroxidase
Reaction: (1) 1-(3,4-dimethoxyphenyl)-2-(2-methoxyphenoxy)propane-1,3-diol + H₂O₂ = 3,4-dimethoxybenzaldehyde + 2-methoxyphenol + glycolaldehyde + H₂O
 (2) 2 (3,4-dimethoxyphenyl)methanol + H₂O₂ = 2 (3,4-dimethoxyphenyl)methanol radical + 2 H₂O
Other name(s): diarylpropane oxygenase; ligninase I; diarylpropane peroxidase; LiP; diarylpropane:oxygen,hydrogen-peroxide oxidoreductase (C-C-bond-cleaving); 1,2-bis(3,4-dimethoxyphenyl)propane-1,3-diol:hydrogen-peroxide oxidoreductase (incorrect); (3,4-dimethoxyphenyl)methanol:hydrogen-peroxide oxidoreductase
Systematic name: 1-(3,4-dimethoxyphenyl)-2-(2-methoxyphenoxy)propane-1,3-diol:hydrogen-peroxide oxidoreductase
Comments: A hemoprotein, involved in the oxidative breakdown of lignin by white-rot basidiomycete fungi. The reaction involves an initial oxidation of the heme iron by hydrogen peroxide, forming compound I (Fe^{IV}=O radical cation) at the active site. A single one-electron reduction of compound I by an electron derived from a substrate molecule yields compound II (Fe^{IV}=O non-radical cation), followed by a second one-electron transfer that returns the enzyme to the ferric oxidation state. The electron transfer events convert the substrate molecule into a transient cation radical intermediate that fragments spontaneously. The enzyme can act on a wide range of aromatic compounds, including methoxybenzenes and nonphenolic β-O-4 linked arylglycerol β-aryl ethers, but cannot act directly on the lignin molecule, which is too large to fit into the active site. However larger lignin molecules can be degraded in the presence of veratryl alcohol. It has been suggested that the free radical that is formed when the enzyme acts on veratryl alcohol can diffuse into the lignified cell wall, where it oxidizes lignin and other organic substrates. In the presence of high concentration of hydrogen peroxide and lack of substrate, the enzyme forms a catalytically inactive form (compound III). This form can be rescued by interaction with two molecules of the free radical products. In the case of veratryl alcohol, such an interaction yields two molecules of veratryl aldehyde.
References: [2070, 3252, 1544, 4545, 526, 2083, 2084, 2082, 960, 3346]

[EC 1.11.1.14 created 1992, modified 2006, modified 2011, modified 2016]

[1.11.1.15 *Transferred entry. peroxiredoxin. Now described by EC 1.11.1.24, thioredoxin-dependent peroxiredoxin; EC 1.11.1.25, glutaredoxin-dependent peroxiredoxin; EC 1.11.1.26, NADH-dependent peroxiredoxin; EC 1.11.1.27, glutathione-dependent peroxiredoxin; EC 1.11.1.28, lipoyl-dependent peroxiredoxin; and EC 1.11.1.29, mycoredoxin-dependent peroxiredoxin.*]

[EC 1.11.1.15 created 2004, deleted 2020]

EC 1.11.1.16

Accepted name: versatile peroxidase
Reaction: (1) 1-(4-hydroxy-3-methoxyphenyl)-2-(2-methoxyphenoxy)propane-1,3-diol + H₂O₂ = 4-hydroxy-3-methoxybenzaldehyde + 2-methoxyphenol + glycolaldehyde + H₂O
 (2) 2 manganese(II) + 2 H⁺ + H₂O₂ = 2 manganese(III) + 2 H₂O
Other name(s): VP; hybrid peroxidase; polyvalent peroxidase; reactive-black-5:hydrogen-peroxide oxidoreductase

Systematic name: 1-(4-hydroxy-3-methoxyphenyl)-2-(2-methoxyphenoxy)propane-1,3-diol:hydrogen-peroxide oxidoreductase

Comments: A hemoprotein. This ligninolytic peroxidase combines the substrate-specificity characteristics of the two other ligninolytic peroxidases, EC 1.11.1.13, manganese peroxidase and EC 1.11.1.14, lignin peroxidase. Unlike these two enzymes, it is also able to oxidize phenols, hydroquinones and both low- and high-redox-potential dyes, due to a hybrid molecular architecture that involves multiple binding sites for substrates [1612, 534].

References: [2671, 1612, 3018, 534, 3017, 533, 3016, 209, 3290, 552]

[EC 1.11.1.16 created 2006, modified 2016]

EC 1.11.1.17

Accepted name: glutathione amide-dependent peroxidase

Reaction: 2 glutathione amide + H₂O₂ = glutathione amide disulfide + 2 H₂O

Systematic name: glutathione amide:hydrogen-peroxide oxidoreductase

Comments: This enzyme, which has been characterized from the proteobacterium *Marichromatium gracile*, is a chimeric protein, containing a peroxiredoxin-like N-terminus and a glutaredoxin-like C terminus. The enzyme has peroxidase activity towards hydrogen peroxide and several small alkyl hydroperoxides, and is thought to represent an early adaptation for fighting oxidative stress [4436]. The glutathione amide disulfide produced by this enzyme can be restored to glutathione amide by EC 1.8.1.16 (glutathione amide reductase).

References: [4436]

[EC 1.11.1.17 created 2010]

EC 1.11.1.18

Accepted name: bromide peroxidase

Reaction: RH + HBr + H₂O₂ = RBr + 2 H₂O

Other name(s): bromoperoxidase; haloperoxidase (ambiguous); eosinophil peroxidase

Systematic name: bromide:hydrogen-peroxide oxidoreductase

Comments: Bromoperoxidases of red and brown marine algae (Rhodophyta and Phaeophyta) contain vanadate. They catalyse the bromination of a range of organic molecules such as sesquiterpenes, forming stable C-Br bonds. Bromoperoxidases also oxidize iodides.

References: [367, 4333, 1840, 566, 3147]

[EC 1.11.1.18 created 2010]

EC 1.11.1.19

Accepted name: dye decolorizing peroxidase

Reaction: Reactive Blue 5 + 2 H₂O₂ = phthalate + 2,2'-disulfonyl azobenzene + 3-[(4-amino-6-chloro-1,3,5-triazin-2-yl)amino]benzenesulfonate + 2 H₂O

Other name(s): DyP; DyP-type peroxidase

Systematic name: Reactive-Blue-5:hydrogen-peroxide oxidoreductase

Comments: Heme proteins with proximal histidine secreted by basidiomycetous fungi and eubacteria. They are similar to EC 1.11.1.16 versatile peroxidase (oxidation of Reactive Black 5, phenols, veratryl alcohol), but differ from the latter in their ability to efficiently oxidize a number of recalcitrant anthraquinone dyes, and inability to oxidize Mn(II). The model substrate Reactive Blue 5 is converted with high efficiency via a so far unique mechanism that combines oxidative and hydrolytic steps and leads to the formation of phthalic acid. Bacterial TfuDyP catalyses sulfoxidation.

References: [2105, 4094, 4938, 4095, 4093, 3135, 4398, 2483, 1687]

[EC 1.11.1.19 created 2011, modified 2015]

EC 1.11.1.20

- Accepted name:** prostamide/prostaglandin F_{2α} synthase
Reaction: thioredoxin + (5Z,9α,11α,13E,15S)-9,11-epidioxy-15-hydroxy-prosta-5,13-dienoate = thioredoxin disulfide + (5Z,9α,11α,13E,15S)-9,11,15-trihydroxyprosta-5,13-dienoate
Other name(s): prostamide/PGF synthase; prostamide F synthase; prostamide/prostaglandin F synthase; tPGF synthase
Systematic name: thioredoxin:(5Z,9α,11α,13E,15S)-9,11-epidioxy-15-hydroxy-prosta-5,13-dienoate oxidoreductase
Comments: The enzyme contains a thioredoxin-type disulfide as a catalytic group. Prostamide H₂ and prostaglandin H₂ are the best substrates; the latter is converted to prostaglandin F_{2α}. The enzyme also reduces *tert*-butyl hydroperoxide, cumene hydroperoxide and H₂O₂, but not prostaglandin D₂ or prostaglandin E₂.
References: [2897, 4816]

[EC 1.11.1.20 created 2011]

EC 1.11.1.21

- Accepted name:** catalase-peroxidase
Reaction: (1) donor + H₂O₂ = oxidized donor + 2 H₂O
(2) 2 H₂O₂ = O₂ + 2 H₂O
Other name(s): *katG* (gene name)
Systematic name: donor:hydrogen-peroxide oxidoreductase
Comments: Differs from EC 1.11.1.7, peroxidase in having a relatively high catalase (EC 1.11.1.6) activity with H₂O₂ as donor, releasing O₂; both activities use the same heme active site. In *Mycobacterium tuberculosis* it is responsible for activation of the commonly used antitubercular drug, isoniazid.
References: [2532, 1680, 1157, 317, 4455]

[EC 1.11.1.21 created 2011]

EC 1.11.1.22

- Accepted name:** hydroperoxy fatty acid reductase
Reaction: a hydroperoxy fatty acid + NADPH + H⁺ = a hydroxy fatty acid + NADP⁺ + H₂O
Other name(s): slr1171 (gene name); slr1992 (gene name); hydroperoxy fatty acid:NADPH oxidoreductase
Systematic name: NADPH:hydroperoxy fatty acid oxidoreductase
Comments: The enzyme, characterized from the cyanobacterium *Synechocystis* PCC 6803, can reduce unsaturated fatty acid hydroperoxides and alkyl hydroperoxides. The enzyme, which utilizes NADPH generated by the photosynthetic electron transfer system, protects the cells from lipid peroxidation.
References: [1244, 1245]

[EC 1.11.1.22 created 2013]

EC 1.11.1.23

- Accepted name:** (S)-2-hydroxypropylphosphonic acid epoxidase
Reaction: (S)-2-hydroxypropylphosphonate + H₂O₂ = (1R,2S)-1,2-epoxypropylphosphonate + 2 H₂O
Other name(s): HPP epoxidase; HppE; 2-hydroxypropylphosphonic acid epoxidase; Fom4; (S)-2-hydroxypropylphosphonate epoxidase
Systematic name: (S)-2-hydroxypropylphosphonate:hydrogen-peroxide epoxidase
Comments: This is the last enzyme in the biosynthetic pathway of fosfomicin, a broad-spectrum antibiotic produced by certain *Streptomyces* species. Contains non heme iron that forms a iron(IV)-oxo (ferryl) complex with hydrogen peroxide, which functions as a proton abstractor from the substrate [4512].
References: [2933, 4756, 1641, 2521, 1645, 535, 4512]

[EC 1.11.1.23 created 2011 as EC 1.14.19.7, transferred 2014 to EC 1.11.1.23]

EC 1.11.1.24

- Accepted name:** thioredoxin-dependent peroxiredoxin
Reaction: thioredoxin + ROOH = thioredoxin disulfide + H₂O + ROH
Other name(s): thioredoxin peroxidase; *bcp* (gene name); *tpx* (gene name); PrxQ
Systematic name: thioredoxin:hydroperoxide oxidoreductase
Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the 'resolving' cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. Thioredoxin-dependent peroxiredoxins are the most common. They have been reported from archaea, bacteria, fungi, plants, and animals.
References: [1991, 2221, 1905, 4667, 1903, 3292]

[EC 1.11.1.24 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.24]

EC 1.11.1.25

- Accepted name:** glutaredoxin-dependent peroxiredoxin
Reaction: glutaredoxin + ROOH = glutaredoxin disulfide + H₂O + ROH
Other name(s): PRXIIB (gene name)
Systematic name: glutaredoxin:hydroperoxide oxidoreductase
Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the 'resolving' cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. To recycle the disulfide, known atypical 2-Cys Prxs appear to use thioredoxin as an electron donor. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. Glutaredoxin-dependent peroxiredoxins have been reported from bacteria, fungi, plants, and animals. These enzymes are often able to use an alternative reductant such as thioredoxin or glutathione.
References: [3581, 4667, 3278, 1509, 2485, 754]

[EC 1.11.1.25 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.25]

EC 1.11.1.26

- Accepted name:** NADH-dependent peroxiredoxin
Reaction: NADH + ROOH + H⁺ = NAD⁺ + H₂O + ROH
Other name(s): *ahpC* (gene name); *ahpF* (gene name); alkyl hydroperoxide reductase
Systematic name: NADH:hydroperoxide oxidoreductase

Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the ‘resolving’ cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. This bacterial peroxiredoxin differs from most other forms by comprising two types of subunits. One subunit (AhpC) is a typical 2-Cys peroxiredoxin. Following the reduction of the substrate, one AhpC subunit forms a disulfide bond with an identical unit. The disulfide bond is reduced by the second type of subunit (AhpF). This second subunit is a flavin-containing protein that uses electrons from NADH to reduce the cysteine residues on the AhpC subunits back to their active state.

References: [4667, 921, 3014]

[EC 1.11.1.26 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.26]

EC 1.11.1.27

Accepted name: glutathione-dependent peroxiredoxin
Reaction: 2 glutathione + ROOH = glutathione disulfide + H₂O + ROH
Other name(s): PRDX6 (gene name); *prx3* (gene name)
Systematic name: glutathione:hydroperoxide oxidoreductase
Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the ‘resolving’ cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. Glutathione-dependent peroxiredoxins have been reported from bacteria and animals, and appear to be 1-Cys enzymes. The mechanism for the mammalian PRDX6 enzyme involves heterodimerization of the enzyme with π -glutathione *S*-transferase, followed by glutathionylation of the oxidized cysteine residue. Subsequent dissociation of the heterodimer yields glutathionylated peroxiredoxin, which is restored to the active form via spontaneous reduction by a second glutathione molecule.

References: [4667, 3267, 2635, 1402, 2485]

[EC 1.11.1.27 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.27]

EC 1.11.1.28

Accepted name: lipoyl-dependent peroxiredoxin
Reaction: a [lipoyl-carrier protein]-N⁶-[(*R*)-dihydrolipoyl]-L-lysine + ROOH = a [lipoyl-carrier protein]-N⁶-[(*R*)-lipoyl]-L-lysine + H₂O + ROH
Other name(s): Ohr; *ahpC* (gene name); *ahpD* (gene name)

Systematic name: [lipoyl-carrier protein]-*N*⁶-[(*R*)-dihydrolipoyl]-L-lysine:hydroperoxide oxidoreductase
Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the ‘resolving’ cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. Two types of lipoyl-dependent peroxiredoxins have been reported from bacteria. One type is the AhpC/AhpD system, originally described from *Mycobacterium tuberculosis*. In that system, AhpC catalyses reduction of the substrate, resulting in an intramolecular disulfide. AhpD then forms an intermolecular disulfide crosslink with AhpC, reducing it back to active state. AhpD is reduced in turn by lipoylated proteins. The second type, which has been characterized in *Xylella fastidiosa*, consists of only one type of subunit, which interacts directly with lipoylated proteins.

References: [1653, 4667, 2240, 2239, 3850, 791]

[EC 1.11.1.28 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.28]

EC 1.11.1.29

Accepted name: mycoredoxin-dependent peroxiredoxin
Reaction: mycoredoxin + ROOH = mycoredoxin disulfide + H₂O + ROH
Other name(s): *ahpE* (gene name)
Systematic name: mycoredoxin:hydroperoxide oxidoreductase
Comments: Peroxiredoxins (Prxs) are a ubiquitous family of antioxidant proteins. They can be divided into three classes: typical 2-Cys, atypical 2-Cys and 1-Cys peroxiredoxins [4667]. The peroxidase reaction comprises two steps centred around a redox-active cysteine called the peroxidatic cysteine. All three peroxiredoxin classes have the first step in common, in which the peroxidatic cysteine attacks the peroxide substrate and is oxidized to *S*-hydroxycysteine (a sulfenic acid) (see mechanism). The second step of the peroxidase reaction, the regeneration of cysteine from *S*-hydroxycysteine, distinguishes the three peroxiredoxin classes. For typical 2-Cys Prxs, in the second step, the peroxidatic *S*-hydroxycysteine from one subunit is attacked by the ‘resolving’ cysteine located in the C-terminus of the second subunit, to form an intersubunit disulfide bond, which is then reduced by one of several cell-specific thiol-containing reductants completing the catalytic cycle. In the atypical 2-Cys Prxs, both the peroxidatic cysteine and its resolving cysteine are in the same polypeptide, so their reaction forms an intrachain disulfide bond. The 1-Cys Prxs conserve only the peroxidatic cysteine, so its regeneration involves direct interaction with a reductant molecule. Mycoredoxin-dependent enzymes are found in Mycobacteria. Following the reduction of the substrate, the sulfenic acid derivative of the peroxidatic cysteine forms a protein mixed disulfide with the N-terminal cysteine of mycoredoxin, which is then reduced by the C-terminal cysteine of mycoredoxin, restoring the peroxiredoxin to active state and resulting in an intra-protein disulfide in mycoredoxin. The disulfide is eventually reduced by mycothiol.

References: [4667, 1766, 1765, 2282, 3279]

[EC 1.11.1.29 created 1983 as EC 1.11.1.15, part transferred 2020 to EC 1.11.1.29]

EC 1.11.2 Peroxygenases

With a peroxide as acceptor, one oxygen atom of which is incorporated into the product

EC 1.11.2.1

- Accepted name:** unspecific peroxygenase
Reaction: $\text{RH} + \text{H}_2\text{O}_2 = \text{ROH} + \text{H}_2\text{O}$
Other name(s): aromatic peroxygenase; mushroom peroxygenase; haloperoxidase-peroxygenase; *Agrocybe aegerita* peroxidase
Systematic name: substrate:hydrogen-peroxide oxidoreductase (RH-hydroxylating or -epoxidising)
Comments: A heme-thiolate protein (*P*-450). Enzymes of this type include glycoproteins secreted by agaric basidiomycetes. They catalyse the insertion of an oxygen atom from H_2O_2 into a wide variety of substrates, including aromatic rings such as naphthalene, toluene, phenanthrene, pyrene and *p*-nitrophenol, recalcitrant heterocycles such as pyridine, dibenzofuran, various ethers (resulting in *O*-dealkylation) and alkanes such as propane, hexane and cyclohexane. Reactions catalysed include hydroxylation, epoxidation, *N*-oxidation, sulfoxidation, *O*- and *N*-dealkylation, bromination and one-electron oxidations. They have little or no activity toward chloride. Mechanistically, the catalytic cycle of unspecific (mono)-peroxygenases combines elements of the "shunt" pathway of cytochrome *P*-450s (a side activity that utilizes a peroxide in place of dioxygen and NAD[P]H) and the classic heme peroxidase cycle.
References: [4375, 4374, 105, 4373, 123, 2122, 2160, 2123, 3276]

[EC 1.11.2.1 created 2011]

EC 1.11.2.2

- Accepted name:** myeloperoxidase
Reaction: $\text{Cl}^- + \text{H}_2\text{O}_2 + \text{H}^+ = \text{HClO} + \text{H}_2\text{O}$
Other name(s): MPO; verdoperoxidase
Systematic name: chloride:hydrogen-peroxide oxidoreductase (hypochlorite-forming)
Comments: Contains calcium and covalently bound heme (proximal ligand histidine). It is present in phagosomes of neutrophils and monocytes, where the hypochlorite produced is strongly bactericidal. It differs from EC 1.11.1.10 chloride peroxidase in its preference for formation of hypochlorite over the chlorination of organic substrates under physiological conditions (pH 5-8). Hypochlorite in turn forms a number of antimicrobial products (Cl_2 , chloramines, hydroxyl radical, singlet oxygen). MPO also oxidizes bromide, iodide and thiocyanate. In the absence of halides, it oxidizes phenols and has a moderate peroxygenase activity toward styrene.
References: [36, 1540, 1235, 4356, 2149, 1116, 1286]

[EC 1.11.2.2 created 2011]

EC 1.11.2.3

- Accepted name:** plant seed peroxygenase
Reaction: $\text{R}^1\text{H} + \text{R}^2\text{OOH} = \text{R}^1\text{OH} + \text{R}^2\text{OH}$
Other name(s): plant peroxygenase; soybean peroxygenase
Systematic name: substrate:hydroperoxide oxidoreductase (RH-hydroxylating or epoxidising)
Comments: A heme protein with calcium binding motif (caleosin-type). Enzymes of this type include membrane-bound proteins found in seeds of different plants. They catalyse the direct transfer of one oxygen atom from an organic hydroperoxide, which is reduced into its corresponding alcohol to a substrate which will be oxidized. Reactions catalysed include hydroxylation, epoxidation and sulfoxidation. Preferred substrate and co-substrate are unsaturated fatty acids and fatty acid hydroperoxides, respectively. Plant seed peroxygenase is involved in the synthesis of cutin.
References: [1832, 357, 1488, 2419, 1503]

[EC 1.11.2.3 created 2011]

EC 1.11.2.4

- Accepted name:** fatty-acid peroxygenase

Reaction: fatty acid + H₂O₂ = 3- or 2-hydroxy fatty acid + H₂O
Other name(s): fatty acid hydroxylase (ambiguous); P450 peroxygenase; CYP152A1; P450BS; P450SP α
Systematic name: fatty acid:hydroperoxide oxidoreductase (RH-hydroxylating)
Comments: A cytosolic heme-thiolate protein with sequence homology to *P*-450 monooxygenases. Unlike the latter, it needs neither NAD(P)H, dioxygen nor specific reductases for function. Enzymes of this type are produced by bacteria (e.g. *Sphingomonas paucimobilis*, *Bacillus subtilis*). Catalytic turnover rates are high compared with those of monooxygenation reactions as well as peroxide shunt reactions catalysed by the common *P*-450s. A model substrate is myristate, but other saturated and unsaturated fatty acids are also hydroxylated. Oxidizes the peroxidase substrate 3,3',5,5'-tetramethylbenzidine (TMB) and peroxygenates aromatic substrates in a fatty-acid-dependent reaction.
References: [2708, 2707, 2705, 1804, 2706, 2382, 2704, 3890]

[EC 1.11.2.4 created 2011]

EC 1.11.2.5

Accepted name: 3-methyl-L-tyrosine peroxygenase
Reaction: 3-methyl-L-tyrosine + H₂O₂ = 3-hydroxy-5-methyl-L-tyrosine + H₂O
Other name(s): SfmD; SacD; 3-methyltyrosine peroxidase; 3-methyl-L-tyrosine peroxidase
Systematic name: 3-methyl-L-tyrosine:hydrogen-peroxide oxidoreductase (3-hydroxy-5-methyl-L-tyrosine-forming)
Comments: The heme-containing peroxygenase from the bacterium *Streptomyces lavendulae* is involved in biosynthesis of saframycin A, a potent antitumor antibiotic that belongs to the tetrahydroisoquinoline family.
References: [4204]

[EC 1.11.2.5 created 2014]

EC 1.11.2.6

Accepted name: L-tyrosine peroxygenase
Reaction: L-tyrosine + H₂O₂ = L-dopa + H₂O
Systematic name: L-tyrosine:hydrogen-peroxide oxidoreductase (L-dopa-forming)
Comments: The enzyme from the bacterium *Streptomyces lincolnensis* participates in the biosynthesis of the antibiotic lincomycin A, while that from *Streptomyces refuineus* is involved in anthramycin biosynthesis. The enzyme, which contains a heme *b* cofactor, is rapidly inactivated in the presence of hydrogen peroxide, but the presence of L-tyrosine protects it. *cf.* EC 1.11.2.5, 3-methyl-L-tyrosine peroxygenase.
References: [3052, 722]

[EC 1.11.2.6 created 2020]

EC 1.12 Acting on hydrogen as donor

This subclass contains hydrogenases other than those that use iron-sulfur compounds as donor (EC 1.18) for the reduction of H⁺ to H₂. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.12.1), a cytochrome (EC 1.12.2), a quinone or similar compound (EC 1.12.5), an iron-sulfur protein (EC 1.12.7), other, known, acceptors (EC 1.12.9), or some other acceptor (EC 1.12.99).

EC 1.12.1 With NAD⁺ or NADP⁺ as acceptor

[1.12.1.1 Transferred entry. peroxidase. Now EC 1.12.7.2, ferredoxin hydrogenase]

[EC 1.12.1.1 created 1965, deleted 1972]

EC 1.12.1.2

Accepted name: hydrogen dehydrogenase
Reaction: $\text{H}_2 + \text{NAD}^+ = \text{H}^+ + \text{NADH}$
Other name(s): H_2 :NAD⁺ oxidoreductase; NAD-linked hydrogenase; bidirectional hydrogenase; hydrogenase
Systematic name: hydrogen:NAD⁺ oxidoreductase
Comments: An iron-sulfur flavoprotein (FMN or FAD). Some forms of this enzyme contain nickel.
References: [382, 3735]

[EC 1.12.1.2 created 1972, modified 2002]

EC 1.12.1.3

Accepted name: hydrogen dehydrogenase (NADP⁺)
Reaction: $\text{H}_2 + \text{NADP}^+ = \text{H}^+ + \text{NADPH}$
Other name(s): NADP⁺-linked hydrogenase; NADP⁺-reducing hydrogenase; hydrogenase (ambiguous); hydrogenase I (ambiguous)
Systematic name: hydrogen:NADP⁺ oxidoreductase
Comments: The protein from the bacterium *Desulfovibrio fructosovorans* is an iron-sulfur protein that exclusively functions as a hydrogen dehydrogenase [852], while the enzyme from the archaeon *Pyrococcus furiosus* is a nickel, iron, iron-sulfur protein, that is part of a heterotetrameric complex where the α and δ subunits function as a hydrogenase while the β and γ subunits function as sulfur reductase (EC 1.12.98.4, sulfhydrogenase). Different from EC 1.12.1.5, hydrogen dehydrogenase [NAD(P)⁺].
References: [852, 478, 2575, 2579, 4406]

[EC 1.12.1.3 created 2002, modified 2013]

EC 1.12.1.4

Accepted name: hydrogenase (NAD⁺, ferredoxin)
Reaction: $2 \text{H}_2 + \text{NAD}^+ + 2 \text{ oxidized ferredoxin} = 5 \text{H}^+ + \text{NADH} + 2 \text{ reduced ferredoxin}$
Other name(s): bifurcating [FeFe] hydrogenase
Systematic name: hydrogen:NAD⁺, ferredoxin oxidoreductase
Comments: The enzyme from *Thermotoga maritima* contains a [FeFe] cluster (*H*-cluster) and iron-sulfur clusters. It works in the direction evolving hydrogen as a means of eliminating excess reducing equivalents.
References: [4438, 3761]

[EC 1.12.1.4 created 2011]

EC 1.12.1.5

Accepted name: hydrogen dehydrogenase [NAD(P)⁺]
Reaction: $\text{H}_2 + \text{NAD(P)}^+ = \text{H}^+ + \text{NAD(P)H}$
Other name(s): hydrogenase II (ambiguous)
Systematic name: hydrogen:NAD(P)⁺ oxidoreductase
Comments: A nickel, iron, iron-sulfur protein. The enzyme from the archaeon *Pyrococcus furiosus* is part of a heterotetrameric complex where the α and δ subunits function as a hydrogenase while the β and γ subunits function as sulfur reductase (EC 1.12.98.4, sulfhydrogenase). Different from EC 1.12.1.3, hydrogen dehydrogenase (NADP⁺).
References: [2578]

[EC 1.12.1.5 created 2013]

EC 1.12.2 With a cytochrome as acceptor

EC 1.12.2.1

- Accepted name:** cytochrome- c_3 hydrogenase
Reaction: $H_2 + 2$ ferricytochrome $c_3 = 2 H^+ + 2$ ferrocycytochrome c_3
Other name(s): H_2 :ferricytochrome c_3 oxidoreductase; cytochrome c_3 reductase; cytochrome hydrogenase; hydrogenase [ambiguous]
Systematic name: hydrogen:ferricytochrome- c_3 oxidoreductase
Comments: An iron-sulfur protein. Some forms of the enzyme contain nickel ([NiFe]-hydrogenases) and, of these, some contain selenocysteine ([NiFeSe]-hydrogenases). Methylene blue and other acceptors can also be reduced.
References: [887, 1649, 3523, 3623, 4460, 1272]

[EC 1.12.2.1 created 1972, modified 2002]

EC 1.12.5 With a quinone or similar compound as acceptor

EC 1.12.5.1

- Accepted name:** hydrogen:quinone oxidoreductase
Reaction: $H_2 +$ menaquinone = menaquinol
Other name(s): hydrogen-ubiquinone oxidoreductase; hydrogen:menaquinone oxidoreductase; membrane-bound hydrogenase; quinone-reactive Ni/Fe-hydrogenase
Systematic name: hydrogen:quinone oxidoreductase
Comments: Contains nickel, iron-sulfur clusters and cytochrome b . Also catalyses the reduction of water-soluble quinones (e.g. 2,3-dimethylnaphthoquinone) or viologen dyes (benzyl viologen or methyl viologen).
References: [968, 969, 1427, 304, 1102, 1830]

[EC 1.12.5.1 created 1999 as EC 1.12.99.3, transferred 2002 to EC 1.12.5.1]

EC 1.12.7 With an iron-sulfur protein as acceptor

[1.12.7.1 *Transferred entry. ferredoxin hydrogenase. Now EC 1.12.7.2, ferredoxin hydrogenase*]

[EC 1.12.7.1 created 1972, deleted 1978]

EC 1.12.7.2

- Accepted name:** ferredoxin hydrogenase
Reaction: $H_2 + 2$ oxidized ferredoxin = 2 reduced ferredoxin + $2 H^+$
Other name(s): H_2 oxidizing hydrogenase; H_2 producing hydrogenase [ambiguous]; bidirectional hydrogenase; hydrogen-lyase [ambiguous]; hydrogenase (ferredoxin); hydrogenase I; hydrogenase II; hydrogenlyase [ambiguous]; uptake hydrogenase [ambiguous]
Systematic name: hydrogen:ferredoxin oxidoreductase
Comments: Contains iron-sulfur clusters. The enzymes from some sources contains nickel. Can use molecular hydrogen for the reduction of a variety of substances.
References: [3895, 4161, 4391, 4943, 26, 3299]

[EC 1.12.7.2 created 1961 as EC 1.98.1.1, transferred 1965 to EC 1.12.1.1, transferred 1972 to EC 1.12.7.1, transferred 1978 to EC 1.18.3.1, transferred 1984 to EC 1.18.99.1, transferred 2002 to EC 1.12.7.2]

EC 1.12.98 With other, known, physiological acceptors

EC 1.12.98.1

- Accepted name:** coenzyme F_{420} hydrogenase

Reaction: $\text{H}_2 + \text{oxidized coenzyme F}_{420} = \text{reduced coenzyme F}_{420}$
Other name(s): 8-hydroxy-5-deazaflavin-reducing hydrogenase; F_{420} -reducing hydrogenase; coenzyme F_{420} -dependent hydrogenase
Systematic name: hydrogen:coenzyme F_{420} oxidoreductase
Comments: An iron-sulfur flavoprotein (FAD) containing nickel. The enzyme from some sources contains selenocysteine. The enzyme also reduces the riboflavin analogue of F_{420} , flavins and methyl viologen, but to a lesser extent. The hydrogen acceptor coenzyme F_{420} is a deazaflavin derivative.
References: [27, 4754, 1156, 2952, 222]

[EC 1.12.98.1 created 1989 as EC 1.12.99.1, transferred 2002 to EC 1.12.98.1]

EC 1.12.98.2

Accepted name: 5,10-methenyltetrahydromethanopterin hydrogenase
Reaction: $\text{H}_2 + 5,10\text{-methenyltetrahydromethanopterin} = \text{H}^+ + 5,10\text{-methylenetetrahydromethanopterin}$
Other name(s): H_2 -forming N^5, N^{10} -methylenetetrahydromethanopterin dehydrogenase; nonmetal hydrogenase; N^5, N^{10} -methenyltetrahydromethanopterin hydrogenase; hydrogen: N^5, N^{10} -methenyltetrahydromethanopterin oxidoreductase
Systematic name: hydrogen:5,10-methenyltetrahydromethanopterin oxidoreductase
Comments: Does not catalyse the reduction of artificial dyes. Does not by itself catalyse a H_2/H^+ exchange reaction. Does not contain nickel or iron-sulfur clusters.
References: [4934, 2153]

[EC 1.12.98.2 created 1999 as EC 1.12.99.4, transferred 2002 to EC 1.12.98.2, modified 2004]

EC 1.12.98.3

Accepted name: *Methanosarcina*-phenazine hydrogenase
Reaction: $\text{H}_2 + 2\text{-}(2,3\text{-dihydropentaprenyloxy})\text{phenazine} = 2\text{-dihydropentaprenyloxyphenazine}$
Other name(s): methanophenazine hydrogenase; methylviologen-reducing hydrogenase
Systematic name: hydrogen:2-(2,3-dihydropentaprenyloxy)phenazine oxidoreductase
Comments: Contains nickel, iron-sulfur clusters and cytochrome *b*. The enzyme from some sources contains selenocysteine.
References: [4, 885, 272]

[EC 1.12.98.3 created 2002]

EC 1.12.98.4

Accepted name: sulfhydrogenase
Reaction: $\text{H}_2 + (\text{sulfide})_n = \text{hydrogen sulfide} + (\text{sulfide})_{n-1}$
Other name(s): sulfur reductase
Systematic name: H_2 :polysulfide oxidoreductase
Comments: An iron-sulfur protein. The enzyme from the hyperthermophilic archaeon *Pyrococcus furiosus* is part of two heterotetrameric complexes where the β and γ subunits function as sulfur reductase and the α and δ subunits function as hydrogenases (EC 1.12.1.3, hydrogen dehydrogenase [NADP^+] and EC 1.12.1.4, hydrogen dehydrogenase [NAD(P)^+], respectively). Sulfur can also be used as substrate, but since it is insoluble in aqueous solution and polysulfide is generated abiotically by the reaction of hydrogen sulfide and sulfur, polysulfide is believed to be the true substrate [2575].
References: [4937, 2575, 2579, 2578]

[EC 1.12.98.4 created 1992 as EC 1.97.1.3, transferred 2013 to EC 1.12.98.4]

EC 1.12.99 With unknown physiological acceptors

[1.12.99.1 *Transferred entry. coenzyme F₄₂₀ hydrogenase. Now EC 1.12.98.1, coenzyme F₄₂₀ hydrogenase*]

[EC 1.12.99.1 created 1989, deleted 2002]

[1.12.99.2 *Deleted entry. coenzyme-M-7-mercaptoheptanoylthreonine-phosphate-heterodisulfide hydrogenase. Now shown to be two enzymes, EC 1.12.98.3, Methanosarcina-phenazine hydrogenase and EC 1.8.98.1, CoB—CoM heterodisulfide reductase*]

[EC 1.12.99.2 created 1992, deleted 2002]

[1.12.99.3 *Transferred entry. hydrogen:quinone oxidoreductase. Now EC 1.12.5.1, hydrogen:quinone oxidoreductase*]

[EC 1.12.99.3 created 1999, deleted 2002]

[1.12.99.4 *Transferred entry. N⁵,N¹⁰-methenyltetrahydromethanopterin hydrogenase. Now EC 1.12.98.2, 5,10-methenyltetrahydromethanopterin hydrogenase*]

[EC 1.12.99.4 created 1999, deleted 2002]

[1.12.99.5 *Deleted entry. 3,4-dihydroxyquinoline 2,4-dioxygenase. Identical to EC 1.13.11.47, 3-hydroxy-4-oxoquinoline 2,4-dioxygenase*]

[EC 1.12.99.5 created 1999, deleted 2001]

EC 1.12.99.6

Accepted name: hydrogenase (acceptor)
Reaction: H₂ + acceptor = reduced acceptor
Other name(s): H₂ producing hydrogenase (ambiguous); hydrogen-lyase (ambiguous); hydrogenlyase (ambiguous); uptake hydrogenase (ambiguous); hydrogen:(acceptor) oxidoreductase
Systematic name: hydrogen:acceptor oxidoreductase
Comments: Uses molecular hydrogen for the reduction of a variety of substances. Contains iron-sulfur clusters. The enzyme from some sources contains nickel.
References: [3895, 27, 4449]

[EC 1.12.99.6 created 2002, modified 2003]

EC 1.13 Acting on single donors with incorporation of molecular oxygen (oxygenases)

This subclass contains oxygenases that incorporate oxygen into the substrate. They differ from those in EC 1.14 in that a second hydrogen donor is not required. Sub-subclasses are based on the number of atoms of oxygen that are incorporated: two atoms of oxygen (EC 1.13.11), one atom of oxygen (EC 1.13.12), or other cases (EC 1.13.99). This classification replaces an earlier version. Common names in this subclass are usually of the form 'monooxygenase' and 'dioxygenase'.

EC 1.13.1 Acting on single donors with incorporation of molecular oxygen (oxygenases)

[1.13.1.1 *Transferred entry. Now EC 1.13.11.1, catechol 1,2-dioxygenase*]

[EC 1.13.1.1 created 1961 as EC 1.99.2.2, transferred 1965 to EC 1.13.1.1, deleted 1972]

[1.13.1.2 *Transferred entry. Now EC 1.13.11.2, catechol 2,3-dioxygenase*]

[EC 1.13.1.2 created 1965, deleted 1972]

[1.13.1.3 *Transferred entry. Now EC 1.13.11.3, protocatechuate 3,4-dioxygenase*]

[EC 1.13.1.3 created 1961 as EC 1.99.2.3, transferred 1965 to EC 1.13.1.3, deleted 1972]

[1.13.1.4 *Transferred entry. Now EC 1.13.11.4, gentisate 1,2-dioxygenase*]

[EC 1.13.1.4 created 1961 as EC 1.99.2.4, transferred 1965 to EC 1.13.1.4, deleted 1972]

[1.13.1.5 *Transferred entry. Now EC 1.13.11.5, homogentisate 1,2-dioxygenase*]

[EC 1.13.1.5 created 1961 as EC 1.99.2.5, transferred 1965 to EC 1.13.1.5, deleted 1972]

[1.13.1.6 *Transferred entry. Now EC 1.13.11.6, 3-hydroxyanthranilate 3,4-dioxygenase*]

[EC 1.13.1.6 created 1965, deleted 1972]

[1.13.1.7 *Deleted entry. 3,4-dihydroxyphenylacetate 3,4-dioxygenase*]

[EC 1.13.1.7 created 1965, transferred 1972 to EC 1.13.11.7, deleted 1980]

[1.13.1.8 *Transferred entry. Now EC 1.13.11.8, protocatechuate 4,5-dioxygenase*]

[EC 1.13.1.8 created 1965, deleted 1972]

[1.13.1.9 *Transferred entry. Now EC 1.13.11.9, 2,5-dihydropyridine 5,6-dioxygenase*]

[EC 1.13.1.9 created 1965, deleted 1972]

[1.13.1.10 *Transferred entry. Now EC 1.13.11.10, 7,8-dihydroxykynurenate 8,8a-dioxygenase*]

[EC 1.13.1.10 created 1965, deleted 1972]

[1.13.1.11 *Transferred entry. Now EC 1.13.99.1, inositol oxygenase*]

[EC 1.13.1.11 created 1961 as EC 1.99.2.6, transferred 1965 to EC 1.13.1.11, deleted 1972]

[1.13.1.12 *Transferred entry. Now EC 1.13.11.11, tryptophan 2,3-dioxygenase*]

[EC 1.13.1.12 created 1961 as EC 1.11.1.4, deleted 1964, reinstated 1965 as EC 1.13.1.12, deleted 1972]

[1.13.1.13 *Transferred entry. Now EC 1.13.11.12, lipoxygenase*]

[EC 1.13.1.13 created 1961 as EC 1.99.2.1, transferred 1965 to EC 1.13.1.13, deleted 1972]

EC 1.13.11 With incorporation of two atoms of oxygen

EC 1.13.11.1

- Accepted name:** catechol 1,2-dioxygenase
Reaction: catechol + O₂ = *cis,cis*-muconate
Other name(s): catechol-oxygen 1,2-oxidoreductase; 1,2-pyrocatechase; catechase; catechol 1,2-oxygenase; catechol dioxygenase; pyrocatechase; pyrocatechol 1,2-dioxygenase; CD I; CD II
Systematic name: catechol:oxygen 1,2-oxidoreductase
Comments: Requires Fe³⁺. Involved in the metabolism of nitro-aromatic compounds by a strain of *Pseudomonas putida*.
References: [1570, 1571, 3918, 4872]

[EC 1.13.11.1 created 1961 as EC 1.99.2.2, transferred 1965 to EC 1.13.1.1, transferred 1972 to EC 1.13.11.1]

EC 1.13.11.2

- Accepted name:** catechol 2,3-dioxygenase
Reaction: catechol + O₂ = 2-hydroxymuconate-6-semialdehyde

Other name(s): 2,3-pyrocatechase; catechol 2,3-oxygenase; catechol oxygenase; metapyrocatechase; pyrocatechol 2,3-dioxygenase; *xylE* (gene name); catechol:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: catechol:oxygen 2,3-oxidoreductase (ring-opening)
Comments: Requires Fe^{II}. The enzyme initiates the *meta*-cleavage pathway of catechol degradation.
References: [1570, 2208, 3114, 2976, 1962, 1964]

[EC 1.13.11.2 created 1965 as EC 1.13.1.2, transferred 1972 to EC 1.13.11.2, modified 1999, modified 2013]

EC 1.13.11.3

Accepted name: protocatechuate 3,4-dioxygenase
Reaction: 3,4-dihydroxybenzoate + O₂ = 3-carboxy-*cis,cis*-muconate
Other name(s): protocatechuate oxygenase; protocatechuic acid oxidase; protocatechuic 3,4-dioxygenase; protocatechuic 3,4-oxygenase; protocatechuate:oxygen 3,4-oxidoreductase (decyclizing)
Systematic name: protocatechuate:oxygen 3,4-oxidoreductase (ring-opening)
Comments: Requires Fe³⁺. The enzyme, which participates in the degradation of aromatic compounds, catalyses the intradiol addition of both oxygen atoms from molecular oxygen, resulting in *ortho*-cleavage of the aromatic ring. The type of cleavage leads to mineralization via the intermediate 3-oxoadipate.
References: [1208, 1428, 4001]

[EC 1.13.11.3 created 1961 as EC 1.99.2.3, transferred 1965 to EC 1.13.1.3, transferred 1972 to EC 1.13.11.3]

EC 1.13.11.4

Accepted name: gentisate 1,2-dioxygenase
Reaction: 2,5-dihydroxybenzoate + O₂ = maleylpyruvate
Other name(s): gentisate oxygenase; 2,5-dihydroxybenzoate dioxygenase; gentisate dioxygenase; gentisic acid oxidase; gentisate:oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: gentisate:oxygen 1,2-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺.
References: [1570, 4104, 4103]

[EC 1.13.11.4 created 1961 as EC 1.99.2.4, transferred 1965 to EC 1.13.1.4, transferred 1972 to EC 1.13.11.4]

EC 1.13.11.5

Accepted name: homogentisate 1,2-dioxygenase
Reaction: homogentisate + O₂ = 4-maleylacetoacetate
Other name(s): homogentisicase; homogentisate oxygenase; homogentisate dioxygenase; homogentisate oxidase; homogentisic acid oxidase; homogentisic acid oxygenase; homogentisic oxygenase; homogentisate:oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: homogentisate:oxygen 1,2-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺.
References: [11, 765, 1570, 2131, 2174, 3461]

[EC 1.13.11.5 created 1961 as EC 1.99.2.5, transferred 1965 to EC 1.13.1.5, transferred 1972 to EC 1.13.11.5]

EC 1.13.11.6

Accepted name: 3-hydroxyanthranilate 3,4-dioxygenase
Reaction: 3-hydroxyanthranilate + O₂ = 2-amino-3-carboxymuconate semialdehyde
Other name(s): 3-hydroxyanthranilate oxygenase; 3-hydroxyanthranilic acid oxygenase; 3-hydroxyanthranilic oxygenase; 3-hydroxyanthranilic acid oxidase; 3HAO; 3-hydroxyanthranilate:oxygen 3,4-oxidoreductase (decyclizing)
Systematic name: 3-hydroxyanthranilate:oxygen 3,4-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺.

References: [860, 1570]

[EC 1.13.11.6 created 1965 as EC 1.13.1.6, transferred 1972 to EC 1.13.11.6]

[1.13.11.7 Deleted entry. 3,4-dihydroxyphenylacetate 3,4-dioxygenase]

[EC 1.13.11.7 created 1965 as EC 1.13.1.7, transferred 1972 to EC 1.13.11.7, deleted 1980]

EC 1.13.11.8

Accepted name: protocatechuate 4,5-dioxygenase
Reaction: 3,4-dihydroxybenzoate + O₂ = 4-carboxy-2-hydroxyruconate semialdehyde
Other name(s): protocatechuate 4,5-oxygenase; protocatechuic 4,5-dioxygenase; protocatechuic 4,5-oxygenase; protocatechuate:oxygen 4,5-oxidoreductase (deacyclizing); protocatechuate:oxygen 4,5-oxidoreductase (ring-opening)
Systematic name: 3,4-dihydroxybenzoate:oxygen 4,5-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺.
References: [4332]

[EC 1.13.11.8 created 1965 as EC 1.13.1.8, transferred 1972 to EC 1.13.11.8]

EC 1.13.11.9

Accepted name: 2,5-dihydropyridine 5,6-dioxygenase
Reaction: 2,5-dihydropyridine + O₂ = *N*-formylmaleamic acid
Other name(s): 2,5-dihydropyridine oxygenase; pyridine-2,5-diol dioxygenase; NicX
Systematic name: 2,5-dihydropyridine:oxygen 5,6-oxidoreductase
Comments: Requires Fe²⁺.
References: [271, 1287, 1288, 1915]

[EC 1.13.11.9 created 1965 as EC 1.13.1.9, transferred 1972 to EC 1.13.11.9, modified 2010]

EC 1.13.11.10

Accepted name: 7,8-dihydroxykynurenate 8,8 α -dioxygenase
Reaction: 7,8-dihydroxykynurenate + O₂ = 5-(3-carboxy-3-oxopropenyl)-4,6-dihydropyridine-2-carboxylate
Other name(s): 7,8-dihydroxykynurenate oxygenase; 7,8-dihydroxykynurenate 8,8 α -dioxygenase; 7,8-dihydroxykynurenate:oxygen 8,8 α -oxidoreductase (deacyclizing)
Systematic name: 7,8-dihydroxykynurenate:oxygen 8,8 α -oxidoreductase (ring-opening)
Comments: Requires Fe²⁺.
References: [2292]

[EC 1.13.11.10 created 1965 as EC 1.13.1.10, transferred 1972 to EC 1.13.11.10]

EC 1.13.11.11

Accepted name: tryptophan 2,3-dioxygenase
Reaction: L-tryptophan + O₂ = *N*-formyl-L-kynurenine
Other name(s): tryptophan pyrrolase (ambiguous); tryptophanase; tryptophan oxygenase; tryptamine 2,3-dioxygenase; tryptophan peroxidase; indoleamine 2,3-dioxygenase (ambiguous); indoleamine 2,3-dioxygenase (ambiguous); L-tryptophan pyrrolase; TDO; L-tryptophan 2,3-dioxygenase; L-tryptophan:oxygen 2,3-oxidoreductase (deacyclizing)
Systematic name: L-tryptophan:oxygen 2,3-oxidoreductase (ring-opening)
Comments: A protohemoprotein. In mammals, the enzyme appears to be located only in the liver. This enzyme, together with EC 1.13.11.52, indoleamine 2,3-dioxygenase, catalyses the first and rate-limiting step in the kynurenine pathway, the major pathway of tryptophan metabolism [2510]. The enzyme is specific for tryptophan as substrate, but is far more active with L-tryptophan than with D-tryptophan [3497].

References: [4360, 3497, 2402, 822, 2510]

[EC 1.13.11.11 created 1961 as EC 1.11.1.4, deleted 1964, reinstated 1965 as EC 1.13.1.12, transferred 1972 to EC 1.13.11.11, modified 1989, modified 2006]

EC 1.13.11.12

Accepted name: linoleate 13*S*-lipoxygenase
Reaction: (1) linoleate + O₂ = (9*Z*,11*E*,13*S*)-13-hydroperoxyoctadeca-9,11-dienoate
(2) α-linolenate + O₂ = (9*Z*,11*E*,13*S*,15*Z*)-13-hydroperoxyoctadeca-9,11,15-trienoate
Other name(s): 13-lipoxidase; carotene oxidase; 13-lipoperoxidase; fat oxidase; 13-lipoxydase; lionoleate:O₂ 13-oxidoreductase
Systematic name: linoleate:oxygen 13-oxidoreductase
Comments: Contains nonheme iron. A common plant lipoxygenase that oxidizes linoleate and α-linolenate, the two most common polyunsaturated fatty acids in plants, by inserting molecular oxygen at the C-13 position with (*S*)-configuration. This enzyme produces precursors for several important compounds, including the plant hormone jasmonic acid. EC 1.13.11.58, linoleate 9*S*-lipoxygenase, catalyses a similar reaction at the second available position of these fatty acids.
References: [685, 4259, 4932, 3586, 173]

[EC 1.13.11.12 created 1961 as EC 1.99.2.1, transferred 1965 to EC 1.13.1.13, transferred 1972 to EC 1.13.11.12, modified 2011, modified 2012]

[1.13.11.13 Deleted entry. ascorbate 2,3-dioxygenase. The activity is the sum of several enzymatic and spontaneous reactions]

[EC 1.13.11.13 created 1972, deleted 2012]

EC 1.13.11.14

Accepted name: 2,3-dihydroxybenzoate 3,4-dioxygenase
Reaction: 2,3-dihydroxybenzoate + O₂ = 3-carboxy-2-hydroxymuconate semialdehyde
Other name(s): *o*-pyrocatechuate oxygenase; 2,3-dihydroxybenzoate 1,2-dioxygenase; 2,3-dihydroxybenzoic oxygenase; 2,3-dihydroxybenzoate oxygenase; 2,3-dihydroxybenzoate:oxygen 3,4-oxidoreductase (decyclizing)
Systematic name: 2,3-dihydroxybenzoate:oxygen 3,4-oxidoreductase (ring-opening)
References: [3511]

[EC 1.13.11.14 created 1972, modified 1976]

EC 1.13.11.15

Accepted name: 3,4-dihydroxyphenylacetate 2,3-dioxygenase
Reaction: 3,4-dihydroxyphenylacetate + O₂ = 2-hydroxy-5-carboxymethylmuconate semialdehyde
Other name(s): 3,4-dihydroxyphenylacetic acid 2,3-dioxygenase; HPC dioxygenase; homoprotocatechuate 2,3-dioxygenase; 3,4-dihydroxyphenylacetate:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: 3,4-dihydroxyphenylacetate:oxygen 2,3-oxidoreductase (ring-opening)
Comments: An iron protein.
References: [12, 218, 2312]

[EC 1.13.11.15 created 1972]

EC 1.13.11.16

Accepted name: 3-carboxyethylcatechol 2,3-dioxygenase
Reaction: (1) 3-(2,3-dihydroxyphenyl)propanoate + O₂ = (2*Z*,4*E*)-2-hydroxy-6-oxonona-2,4-diene-1,9-dioate

(2) (2E)-3-(2,3-dihydroxyphenyl)prop-2-enoate + O₂ = (2Z,4E,7E)-2-hydroxy-6-oxonona-2,4,7-triene-1,9-dioate

- Other name(s):** 2,3-dihydroxy-β-phenylpropionic dioxygenase; 2,3-dihydroxy-β-phenylpropionate oxygenase; 3-(2,3-dihydroxyphenyl)propanoate:oxygen 1,2-oxidoreductase; 3-(2,3-dihydroxyphenyl)propanoate:oxygen 1,2-oxidoreductase (decyclizing)
- Systematic name:** 3-(2,3-dihydroxyphenyl)propanoate:oxygen 1,2-oxidoreductase (ring-opening)
- Comments:** An iron protein. This enzyme catalyses a step in the pathway of phenylpropanoid compounds degradation.
- References:** [798, 2331, 900]

[EC 1.13.11.16 created 1972, modified 2011, modified 2012]

EC 1.13.11.17

- Accepted name:** indole 2,3-dioxygenase
- Reaction:** indole + O₂ = 2-formylaminobenzaldehyde
- Other name(s):** indole oxidase; indoleamine 2,3-dioxygenase (ambiguous); indole:O₂ oxidoreductase; indole-oxygen 2,3-oxidoreductase (decyclizing); IDO (ambiguous); indole:oxygen 2,3-oxidoreductase (decyclizing)
- Systematic name:** indole:oxygen 2,3-oxidoreductase (ring-opening)
- Comments:** Enzymes from the plants *Tecoma stans*, *Jasminum grandiflorum* and *Zea mays* are flavoproteins containing copper. They are part of enzyme systems that form either anthranil (2,1-benzisoxazole) (*Tecoma stans*), anthranilate (*Jasminum grandiflorum*) or both (*Zea mays*) as the final product. A second enzyme from *Tecoma stans* is not a flavoprotein, does not require copper, and is part of a system that forms anthranilate as the final product.
- References:** [2968, 620, 924, 2288]

[EC 1.13.11.17 created 1972, modified 1986]

EC 1.13.11.18

- Accepted name:** persulfide dioxygenase
- Reaction:** S-sulfanylglutathione + O₂ + H₂O = glutathione + sulfite + 2 H⁺ (overall reaction)
(1a) S-sulfanylglutathione + O₂ = S-sulfinatoglutathione + H⁺
(1b) S-sulfinatoglutathione + H₂O = glutathione + sulfite + H⁺ (spontaneous)
- Other name(s):** sulfur oxygenase (incorrect); sulfur:oxygen oxidoreductase (incorrect); sulfur dioxygenase (incorrect)
- Systematic name:** S-sulfanylglutathione:oxygen oxidoreductase
- Comments:** An iron protein. Perthiols, formed spontaneously by interactions between thiols and elemental sulfur or sulfide, are the only acceptable substrate to the enzyme. The sulfite that is formed by the enzyme can be further converted into sulfate, thiosulfate or S-sulfoglutathione (GSSO₃⁻) non-enzymically [3557].
- References:** [4136, 3557, 2514, 1694, 3308]

[EC 1.13.11.18 created 1972, modified 2015]

EC 1.13.11.19

- Accepted name:** cysteamine dioxygenase
- Reaction:** cysteamine + O₂ = hypotaurine
- Other name(s):** ADO (gene name); persulfurase; cysteamine oxygenase; cysteamine:oxygen oxidoreductase
- Systematic name:** 2-aminoethanethiol:oxygen oxidoreductase
- Comments:** A non-heme iron protein that is involved in the biosynthesis of taurine. 3-Aminopropanethiol (homocysteamine) and 2-sulfanylethan-1-ol (2-mercaptoethanol) can also act as substrates, but glutathione, cysteine, and cysteine ethyl- and methyl esters are not good substrates [578, 579].
- References:** [578, 4664, 579, 3513, 943]

[EC 1.13.11.19 created 1972, modified 2006]

EC 1.13.11.20

Accepted name: cysteine dioxygenase
Reaction: L-cysteine + O₂ = 3-sulfinoalanine
Other name(s): cysteine oxidase
Systematic name: L-cysteine:oxygen oxidoreductase
Comments: Requires Fe²⁺ and NAD(P)H.
References: [2536]

[EC 1.13.11.20 created 1972, modified 1976]

[1.13.11.21 Transferred entry. *β-carotene 15,15'-dioxygenase*. Now EC 1.14.99.36, *β-carotene 15,15'-monooxygenase*]

[EC 1.13.11.21 created 1972, deleted 2001]

EC 1.13.11.22

Accepted name: caffeate 3,4-dioxygenase
Reaction: 3,4-dihydroxy-*trans*-cinnamate + O₂ = 3-(2-carboxyethenyl)-*cis,cis*-muconate
Other name(s): 3,4-dihydroxy-*trans*-cinnamate:oxygen 3,4-oxidoreductase (decyclizing)
Systematic name: 3,4-dihydroxy-*trans*-cinnamate:oxygen 3,4-oxidoreductase (ring-opening)
References: [3787]

[EC 1.13.11.22 created 1972]

EC 1.13.11.23

Accepted name: 2,3-dihydroxyindole 2,3-dioxygenase
Reaction: 2,3-dihydroxyindole + O₂ = anthranilate + CO₂
Other name(s): 2,3-dihydroxyindole:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: 2,3-dihydroxyindole:oxygen 2,3-oxidoreductase (ring-opening)
References: [1207]

[EC 1.13.11.23 created 1972]

EC 1.13.11.24

Accepted name: quercetin 2,3-dioxygenase
Reaction: quercetin + O₂ = 2-(3,4-dihydroxybenzoyloxy)-4,6-dihydroxybenzoate + CO + H⁺
Other name(s): quercetinase; flavonol 2,4-oxygenase; quercetin:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: quercetin:oxygen 2,3-oxidoreductase (ring-opening)
Comments: The enzyme from *Aspergillus* sp. is a copper protein whereas that from *Bacillus subtilis* contains iron. Quercetin is a flavonol (5,7,3',4'-tetrahydroxyflavonol).
References: [3151, 4018, 409]

[EC 1.13.11.24 created 1972]

EC 1.13.11.25

Accepted name: 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione 4,5-dioxygenase
Reaction: 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione + O₂ = 3-hydroxy-5,9,17-trioxo-4,5:9,10-disecoandrosta-1(10),2-dien-4-oate
Other name(s): steroid 4,5-dioxygenase; 3-alkylcatechol 2,3-dioxygenase; 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione:oxygen 4,5-oxidoreductase (decyclizing)
Systematic name: 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione:oxygen 4,5-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺. Also acts on 3-isopropylcatechol and 3-*tert*-butyl-5-methylcatechol.
References: [1322]

[EC 1.13.11.25 created 1972]

EC 1.13.11.26

Accepted name: peptide-tryptophan 2,3-dioxygenase
Reaction: [protein]-L-tryptophan + O₂ = [protein]-*N*-formyl-L-kynurenine
Other name(s): pyrroloxygenase; peptidyltryptophan 2,3-dioxygenase; tryptophan pyrroloxygenase; [protein]-L-tryptophan:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: [protein]-L-tryptophan:oxygen 2,3-oxidoreductase (ring-opening)
Comments: Also acts on tryptophan.
References: [1198, 539]

[EC 1.13.11.26 created 1972, modified 2011]

EC 1.13.11.27

Accepted name: 4-hydroxyphenylpyruvate dioxygenase
Reaction: 4-hydroxyphenylpyruvate + O₂ = homogentisate + CO₂
Other name(s): *p*-hydroxyphenylpyruvic hydroxylase; *p*-hydroxyphenylpyruvate hydroxylase; *p*-hydroxyphenylpyruvate oxidase; *p*-hydroxyphenylpyruvic oxidase; *p*-hydroxyphenylpyruvate dioxygenase; *p*-hydroxyphenylpyruvic acid hydroxylase; 4-hydroxyphenylpyruvic acid dioxygenase
Systematic name: 4-hydroxyphenylpyruvate:oxygen oxidoreductase (hydroxylating, decarboxylating)
Comments: The *Pseudomonas* enzyme contains one Fe³⁺ per mole of enzyme; the enzymes from other sources may contain essential iron or copper.
References: [2502, 3543]

[EC 1.13.11.27 created 1961 as EC 1.99.1.14, transferred 1965 to EC 1.14.2.2, transferred 1972 to EC 1.13.11.27]

EC 1.13.11.28

Accepted name: 2,3-dihydroxybenzoate 2,3-dioxygenase
Reaction: 2,3-dihydroxybenzoate + O₂ = 2-carboxy-*cis,cis*-muconate
Other name(s): 2,3-dihydroxybenzoate 2,3-oxygenase; 2,3-dihydroxybenzoate:oxygen 2,3-oxidoreductase (decyclizing)
Systematic name: 2,3-dihydroxybenzoate:oxygen 2,3-oxidoreductase (ring-opening)
Comments: Also acts, more slowly, with 2,3-dihydroxy-4-methylbenzoate and 2,3-dihydroxy-4-isopropylbenzoate.
References: [971, 3825]

[EC 1.13.11.28 created 1978]

EC 1.13.11.29

Accepted name: stizolobate synthase
Reaction: L-dopa + O₂ = 4-(L-alanin-3-yl)-2-hydroxy-*cis,cis*-muconate 6-semialdehyde
Systematic name: 3,4-dihydroxy-L-phenylalanine:oxygen 4,5-oxidoreductase (recyclizing)
Comments: The intermediate product undergoes ring closure and oxidation, with NAD(P)⁺ as acceptor, to stizolobic acid. The enzyme requires Zn²⁺.
References: [3631, 3632]

[EC 1.13.11.29 created 1978]

EC 1.13.11.30

Accepted name: stizolobinate synthase
Reaction: L-dopa + O₂ = 5-(L-alanin-3-yl)-2-hydroxy-*cis,cis*-muconate 6-semialdehyde
Systematic name: 3,4-dihydroxy-L-phenylalanine:oxygen 2,3-oxidoreductase (recyclizing)

Comments: The intermediate product undergoes ring closure and oxidation, with NAD(P)⁺ as acceptor, to stizolobinic acid. The enzyme requires Zn²⁺.

References: [3631, 3632]

[EC 1.13.11.30 created 1978]

EC 1.13.11.31

Accepted name: arachidonate 12-lipoxygenase

Reaction: arachidonate + O₂ = (5Z,8Z,10E,14Z)-(12S)-12-hydroperoxyicosa-5,8,10,14-tetraenoate

Other name(s): Δ¹²-lipoxygenase; 12-lipoxygenase; 12Δ-lipoxygenase; C-12 lipoxygenase; 12S-lipoxygenase; leukotriene A₄ synthase; LTA₄ synthase

Systematic name: arachidonate:oxygen 12-oxidoreductase

Comments: The product is rapidly reduced to the corresponding 12S-hydroxy compound.

References: [1489, 3118, 4502]

[EC 1.13.11.31 created 1983]

[1.13.11.32 *Transferred entry. 2-nitropropane dioxygenase. Now EC 1.13.12.16, nitronate monooxygenase*]

[EC 1.13.11.32 created 1984, modified 2006, deleted 2009]

EC 1.13.11.33

Accepted name: arachidonate 15-lipoxygenase

Reaction: arachidonate + O₂ = (5Z,8Z,11Z,13E)-(15S)-15-hydroperoxyicosa-5,8,11,13-tetraenoate

Other name(s): 15-lipoxygenase; linoleic acid ω⁶-lipoxygenase; ω⁶ lipoxygenase

Systematic name: arachidonate:oxygen 15-oxidoreductase

Comments: The product is rapidly reduced to the corresponding 15S-hydroxy compound.

References: [479, 3015, 3170, 3853]

[EC 1.13.11.33 created 1984]

EC 1.13.11.34

Accepted name: arachidonate 5-lipoxygenase

Reaction: arachidonate + O₂ = leukotriene A₄ + H₂O (overall reaction)

(1a) arachidonate + O₂ = (6E,8Z,11Z,14Z)-(5S)-5-hydroperoxyicosa-6,8,11,14-tetraenoate

(1b) (6E,8Z,11Z,14Z)-(5S)-5-hydroperoxyicosa-6,8,11,14-tetraenoate = leukotriene A₄ + H₂O

Other name(s): leukotriene-A₄ synthase; Δ⁵-lipoxygenase; 5Δ-lipoxygenase; arachidonic 5-lipoxygenase; arachidonic acid 5-lipoxygenase; C-5-lipoxygenase; LTA synthase; leukotriene A₄ synthase

Systematic name: arachidonate:oxygen 5-oxidoreductase

References: [2703, 3138, 3871, 3872]

[EC 1.13.11.34 created 1984, modified 1990]

EC 1.13.11.35

Accepted name: pyrogallol 1,2-oxygenase

Reaction: 1,2,3-trihydroxybenzene + O₂ = (2Z,4E)-2-hydroxyhexa-2,4-dienedioate

Other name(s): pyrogallol 1,2-dioxygenase; 1,2,3-trihydroxybenzene:oxygen 1,2-oxidoreductase (decyclizing)

Systematic name: 1,2,3-trihydroxybenzene:oxygen 1,2-oxidoreductase (ring-opening)

References: [1421]

[EC 1.13.11.35 created 1984, modified 2012]

EC 1.13.11.36

- Accepted name:** chloridazon-catechol dioxygenase
Reaction: 5-amino-4-chloro-2-(2,3-dihydroxyphenyl)-3(2*H*)-pyridazinone + O₂ = 5-amino-4-chloro-2-(2-hydroxymuconoyl)-3(2*H*)-pyridazinone
Other name(s): 5-amino-4-chloro-2-(2,3-dihydroxyphenyl)-3(2*H*)-pyridazinone 1,2-oxidoreductase (decyclizing)
Systematic name: 5-amino-4-chloro-2-(2,3-dihydroxyphenyl)-3(2*H*)-pyridazinone 1,2-oxidoreductase (ring-opening)
Comments: An iron protein, requiring additional Fe²⁺. Not identical with EC 1.13.11.1 (catechol 1,2-dioxygenase), EC 1.13.11.2 (catechol 2,3-dioxygenase) or EC 1.13.11.5 (homogentisate 1,2-dioxygenase). Involved in the breakdown of the herbicide chloridazon.
References: [2925, 2926]

[EC 1.13.11.36 created 1984]

EC 1.13.11.37

- Accepted name:** hydroxyquinol 1,2-dioxygenase
Reaction: hydroxyquinol + O₂ = maleylacetate
Other name(s): hydroxyquinol dioxygenase; benzene-1,2,4-triol: oxygen 1,2-oxidoreductase (decyclizing); benzene-1,2,4-triol: oxygen 1,2-oxidoreductase (ring-opening)
Systematic name: hydroxyquinol: oxygen 1,2-oxidoreductase (ring-opening)
Comments: An iron protein. Highly specific; catechol and pyrogallol are acted on at less than 1% of the rate at which hydroxyquinol is oxidized.
References: [4156, 1108, 1562]

[EC 1.13.11.37 created 1989, modified 2013]

EC 1.13.11.38

- Accepted name:** 1-hydroxy-2-naphthoate 1,2-dioxygenase
Reaction: 1-hydroxy-2-naphthoate + O₂ = (3*Z*)-4-(2-carboxyphenyl)-2-oxobut-3-enoate
Other name(s): 1-hydroxy-2-naphthoate dioxygenase; 1-hydroxy-2-naphthoate-degrading enzyme; 1-hydroxy-2-naphthoic acid dioxygenase; 1-hydroxy-2-naphthoate: oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: 1-hydroxy-2-naphthoate: oxygen 1,2-oxidoreductase (ring-opening)
Comments: Requires Fe²⁺. Involved, with EC 4.1.2.34 4-(2-carboxyphenyl)-2-oxobut-3-enoate aldolase, in the metabolism of phenanthrene in bacteria.
References: [221]

[EC 1.13.11.38 created 1989]

EC 1.13.11.39

- Accepted name:** biphenyl-2,3-diol 1,2-dioxygenase
Reaction: biphenyl-2,3-diol + O₂ = 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate
Other name(s): 2,3-dihydroxybiphenyl dioxygenase; biphenyl-2,3-diol dioxygenase; *bphC* (gene name); biphenyl-2,3-diol: oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: biphenyl-2,3-diol: oxygen 1,2-oxidoreductase (ring-opening)
Comments: Contains Fe²⁺ or Mn²⁺ [1561]. This enzyme participates in the degradation pathway of biphenyl and PCB (poly chlorinated biphenyls), and catalyses the first ring cleavage step by incorporating two oxygen atoms into the catechol ring formed by EC 1.3.1.56, *cis*-2,3-dihydroxybiphenyl-2,3-diol dehydrogenase. The enzyme from the bacterium *Burkholderia xenovorans* LB400 can also process catechol, 3-methylcatechol, and 4-methylcatechol, but less efficiently [1042]. The enzyme from the carbazole-degrader *Pseudomonas resinovorans* strain CA10 also accepts 2'-aminobiphenyl-2,3-diol [1863]. The enzyme from *Ralstonia* sp. SBUG 290 can also accept 1,2-dihydroxydibenzofuran and 1,2-dihydroxynaphthalene [4592]. The enzyme is strongly inhibited by the substrate [1042]. Not identical with EC 1.13.11.2 catechol 2,3-dioxygenase.
References: [1042, 4378, 1561, 4592, 1863]

[EC 1.13.11.39 created 1989]

EC 1.13.11.40

Accepted name: arachidonate 8-lipoxygenase
Reaction: arachidonate + O₂ = (5Z,9E,11Z,14Z)-(8R)-8-hydroperoxyicoso-5,9,11,14-tetraenoate
Other name(s): 8-lipoxygenase; 8(R)-lipoxygenase
Systematic name: arachidonate:oxygen 8-oxidoreductase
Comments: From the coral *Pseudoplexaura porosa*.
References: [493]

[EC 1.13.11.40 created 1989]

EC 1.13.11.41

Accepted name: 2,4'-dihydroxyacetophenone dioxygenase
Reaction: 2,4'-dihydroxyacetophenone + O₂ = 4-hydroxybenzoate + formate
Other name(s): (4-hydroxybenzoyl)methanol oxygenase
Systematic name: 2,4'-dihydroxyacetophenone oxidoreductase (C-C-bond-cleaving)
References: [1715]

[EC 1.13.11.41 created 1989]

[1.13.11.42 Deleted entry. indoleamine-pyrrole 2,3-dioxygenase. The enzyme was identical to EC 1.13.11.11, tryptophan 2,3-dioxygenase]

[EC 1.13.11.42 created 1992, deleted 2006]

EC 1.13.11.43

Accepted name: lignostilbene αβ-dioxygenase
Reaction: 1,2-bis(4-hydroxy-3-methoxyphenyl)ethylene + O₂ = 2 vanillin
Systematic name: 1,2-bis(4-hydroxy-3-methoxyphenyl)ethylene:oxygen oxidoreductase (αβ-bond-cleaving)
Comments: An iron protein. The enzyme catalyses oxidative cleavage of the interphenyl double bond in the synthetic substrate and lignin-derived stilbenes. It is responsible for the degradation of a diarylpropane-type structure in lignin.
References: [1987]

[EC 1.13.11.43 created 1992]

[1.13.11.44 Deleted entry. linoleate diol synthase. Activity is covered by EC 1.13.11.60, linoleate 8R-lipoxygenase and EC 5.4.4.6, 9,12-octadecadienoate 8-hydroperoxide 8S-isomerase.]

[EC 1.13.11.44 created 2000, deleted 2011]

EC 1.13.11.45

Accepted name: linoleate 11-lipoxygenase
Reaction: linoleate + O₂ = (9Z,12Z)-(11S)-11-hydroperoxyoctadeca-9,12-dienoate
Other name(s): linoleate dioxygenase; manganese lipoxygenase
Systematic name: linoleate:oxygen 11S-oxidoreductase
Comments: The product (9Z,12Z)-(11S)-11-hydroperoxyoctadeca-9,12-dienoate, is converted, more slowly, into (9Z,11E)-(13R)-13-hydroperoxyoctadeca-9,11-dienoate. The enzyme from the fungus *Gaeumannomyces graminis* requires Mn²⁺. It also acts on α-linolenate, whereas γ-linolenate is a poor substrate. Oleate and arachidonate are not substrates.
References: [1491, 3171, 4084]

[EC 1.13.11.45 created 2000]

EC 1.13.11.46

Accepted name: 4-hydroxymandelate synthase
Reaction: 4-hydroxyphenylpyruvate + O₂ = (*S*)-4-hydroxymandelate + CO₂
Other name(s): 4-hydroxyphenylpyruvate dioxygenase II
Systematic name: (*S*)-4-hydroxyphenylpyruvate:oxygen oxidoreductase (decarboxylating)
Comments: Requires Fe²⁺. Involved in the biosynthesis of the vancomycin group of glycopeptide antibiotics.
References: [1758, 678]

[EC 1.13.11.46 created 2001]

EC 1.13.11.47

Accepted name: 3-hydroxy-4-oxoquinoline 2,4-dioxygenase
Reaction: 3-hydroxy-1*H*-quinolin-4-one + O₂ = *N*-formylantranilate + CO
Other name(s): (1*H*)-3-hydroxy-4-oxoquinoline 2,4-dioxygenase; 3-hydroxy-4-oxo-1,4-dihydroquinoline 2,4-dioxygenase; 3-hydroxy-4(1*H*)-one, 2,4-dioxygenase; quinoline-3,4-diol 2,4-dioxygenase
Systematic name: 3-hydroxy-1*H*-quinolin-4-one 2,4-dioxygenase (CO-forming)
Comments: Does not contain a metal centre or organic cofactor. Fission of two C-C bonds: 2,4-dioxygenolytic cleavage with concomitant release of carbon monoxide. The enzyme from *Pseudomonas putida* is highly specific for this substrate.
References: [242, 243, 1126]

[EC 1.13.11.47 created 1999 as EC 1.13.99.5, transferred 2001 to EC 1.13.11.47 (EC 1.12.99.5 created 1999 deleted 2001 as identical)]

EC 1.13.11.48

Accepted name: 3-hydroxy-2-methylquinolin-4-one 2,4-dioxygenase
Reaction: 3-hydroxy-2-methyl-1*H*-quinolin-4-one + O₂ = *N*-acetylantranilate + CO
Other name(s): (1*H*)-3-hydroxy-4-oxoquinoline 2,4-dioxygenase
Systematic name: 3-hydroxy-2-methyl-1*H*-quinolin-4-one 2,4-dioxygenase (CO-forming)
Comments: Does not contain a metal centre or organic cofactor. Fission of two C-C bonds: 2,4-dioxygenolytic cleavage with concomitant release of carbon monoxide. The enzyme from *Arthrobacter sp.* can also act on 3-hydroxy-4-oxoquinoline, forming *N*-formylantranilate and CO (*cf.* EC 1.13.11.47, 3-hydroxy-4-oxoquinoline 2,4-dioxygenase), but more slowly.
References: [242, 243, 1126]

[EC 1.13.11.48 created 2001]

EC 1.13.11.49

Accepted name: chlorite O₂-lyase
Reaction: chloride + O₂ = chlorite
Systematic name: chloride:oxygen oxidoreductase
Comments: Reaction occurs in the reverse direction in chlorate- and perchlorate-reducing bacteria. There is no activity when chlorite is replaced by hydrogen peroxide, perchlorate, chlorate or nitrite. The term 'chlorite dismutase' is misleading as the reaction does not involve dismutation/disproportionation. Contains iron and protoheme IX.
References: [4405, 4021]

[EC 1.13.11.49 created 2001]

EC 1.13.11.50

Accepted name: acetylacetone-cleaving enzyme
Reaction: pentane-2,4-dione + O₂ = acetate + 2-oxopropanal
Other name(s): Dke1; acetylacetone dioxygenase; diketone cleaving dioxygenase; diketone cleaving enzyme
Systematic name: acetylacetone:oxygen oxidoreductase

Comments: An iron(II)-dependent enzyme. Forms the first step in the acetylacetone degradation pathway of *Acinetobacter johnsonii*. While acetylacetone is by far the best substrate, heptane-3,5-dione, octane-2,4-dione, 2-acetylcyclohexanone and ethyl acetoacetate can also act as substrates.

References: [4053]

[EC 1.13.11.50 created 2003]

EC 1.13.11.51

Accepted name: 9-*cis*-epoxycarotenoid dioxygenase

Reaction: (1) a 9-*cis*-epoxycarotenoid + O₂ = 2-*cis*,4-*trans*-xanthoxin + a 12'-apo-carotenal
(2) 9-*cis*-violaxanthin + O₂ = 2-*cis*,4-*trans*-xanthoxin + (3*S*,5*R*,6*S*)-5,6-epoxy-3-hydroxy-5,6-dihydro-12'-apo-β-caroten-12'-al
(3) 9'-*cis*-neoxanthin + O₂ = 2-*cis*,4-*trans*-xanthoxin + (3*S*,5*R*,6*R*)-5,6-dihydroxy-6,7-didehydro-5,6-dihydro-12'-apo-β-caroten-12'-al

Other name(s): nine-*cis*-epoxycarotenoid dioxygenase; NCED; AtNCED3; PvNCED1; VP14

Systematic name: 9-*cis*-epoxycarotenoid 11,12-dioxygenase

Comments: Requires iron(II). Acts on 9-*cis*-violaxanthin and 9'-*cis*-neoxanthin but not on the *all-trans* isomers [4192, 3401]. In vitro, it will cleave 9-*cis*-zeaxanthin. Catalyses the first step of abscisic-acid biosynthesis from carotenoids in chloroplasts, in response to water stress. The other enzymes involved in the abscisic-acid biosynthesis pathway are EC 1.1.1.288 (xanthoxin dehydrogenase), EC 1.2.3.14 (abscisic-aldehyde oxidase) and EC 1.14.13.93 [(+)-abscisic acid 8'-hydroxylase].

References: [3768, 4192, 3401, 4269, 1852, 1853]

[EC 1.13.11.51 created 2005]

EC 1.13.11.52

Accepted name: indoleamine 2,3-dioxygenase

Reaction: (1) D-tryptophan + O₂ = *N*-formyl-D-kynurenine
(2) L-tryptophan + O₂ = *N*-formyl-L-kynurenine

Other name(s): IDO (ambiguous); tryptophan pyrrolase (ambiguous); D-tryptophan:oxygen 2,3-oxidoreductase (decyclizing)

Systematic name: D-tryptophan:oxygen 2,3-oxidoreductase (ring-opening)

Comments: A protohemoprotein. Requires ascorbic acid and methylene blue for activity. This enzyme has broader substrate specificity than EC 1.13.11.11, tryptophan 2,3-dioxygenase [4742]. It is induced in response to pathological conditions and host-defense mechanisms and its distribution in mammals is not confined to the liver [4779]. While the enzyme is more active with D-tryptophan than L-tryptophan, its only known function to date is in the metabolism of L-tryptophan [4779, 2510]. Super-oxide radicals can replace O₂ as oxygen donor [1668, 4267].

References: [4742, 4779, 4186, 1668, 822, 2510, 4267, 3964]

[EC 1.13.11.52 created 2006]

EC 1.13.11.53

Accepted name: acireductone dioxygenase (Ni²⁺-requiring)

Reaction: 1,2-dihydroxy-5-(methylsulfanyl)pent-1-en-3-one + O₂ = 3-(methylsulfanyl)propanoate + formate + CO

Other name(s): ARD; 2-hydroxy-3-keto-5-thiomethylpent-1-ene dioxygenase (ambiguous); acireductone dioxygenase (ambiguous); E-2; 1,2-dihydroxy-5-(methylthio)pent-1-en-3-one:oxygen oxidoreductase (formate- and CO-forming)

Systematic name: 1,2-dihydroxy-5-(methylsulfanyl)pent-1-en-3-one:oxygen oxidoreductase (formate- and CO-forming)

Comments: Requires Ni²⁺. If iron(II) is bound instead of Ni²⁺, the reaction catalysed by EC 1.13.11.54, acireductone dioxygenase [iron(II)-requiring], occurs instead [4672]. The enzyme from the bacterium *Klebsiella oxytoca* (formerly *Klebsiella pneumoniae*) ATCC strain 8724 is involved in the methionine salvage pathway.

References: [4672, 4673, 1233, 804, 2852, 803, 53, 3338]

[EC 1.13.11.53 created 2006]

EC 1.13.11.54

Accepted name: acireductone dioxygenase [iron(II)-requiring]

Reaction: 1,2-dihydroxy-5-(methylsulfanyl)pent-1-en-3-one + O₂ = 4-(methylsulfanyl)-2-oxobutanoate + formate

Other name(s): ARD'; 2-hydroxy-3-keto-5-thiomethylpent-1-ene dioxygenase (ambiguous); acireductone dioxygenase (ambiguous); E-2'; E-3 dioxygenase; 1,2-dihydroxy-5-(methylthio)pent-1-en-3-one:oxygen oxidoreductase (formate-forming)

Systematic name: 1,2-dihydroxy-5-(methylsulfanyl)pent-1-en-3-one:oxygen oxidoreductase (formate-forming)

Comments: Requires iron(II). If Ni²⁺ is bound instead of iron(II), the reaction catalysed by EC 1.13.11.53, acireductone dioxygenase (Ni²⁺-requiring), occurs instead. The enzyme from the bacterium *Klebsiella oxytoca* (formerly *Klebsiella pneumoniae*) ATCC strain 8724 is involved in the methionine salvage pathway.

References: [4672, 4673, 1233, 804, 2852, 803, 53, 3338]

[EC 1.13.11.54 created 2006]

EC 1.13.11.55

Accepted name: sulfur oxygenase/reductase

Reaction: 4 sulfur + 4 H₂O + O₂ = 2 hydrogen sulfide + 2 sulfite

Other name(s): SOR; sulfur oxygenase; sulfur oxygenase reductase

Systematic name: sulfur:oxygen oxidoreductase (hydrogen-sulfide- and sulfite-forming)

Comments: This enzyme, which is found in thermophilic microorganisms, contains one mononuclear non-heme iron centre per subunit. Elemental sulfur is both the electron donor and one of the two known acceptors, the other being oxygen. Thiosulfate is also observed as a product, but is likely formed non-enzymically by a reaction between sulfite and sulfur [2155]. This enzyme differs from EC 1.13.11.18, sulfur dioxygenase and EC 1.12.98.4, sulfhydrogenase, in that both activities occur simultaneously.

References: [2155, 2156, 4117, 4380]

[EC 1.13.11.55 created 2006]

EC 1.13.11.56

Accepted name: 1,2-dihydroxynaphthalene dioxygenase

Reaction: naphthalene-1,2-diol + O₂ = 2-hydroxy-2H-chromene-2-carboxylate

Other name(s): 1,2-DHN dioxygenase; DHNDO; 1,2-dihydroxynaphthalene oxygenase; 1,2-dihydroxynaphthalene:oxygen oxidoreductase

Systematic name: naphthalene-1,2-diol:oxygen oxidoreductase

Comments: This enzyme is involved in naphthalene degradation. Requires Fe²⁺.

References: [2275, 2051, 3258]

[EC 1.13.11.56 created 2010, modified 2010]

EC 1.13.11.57

Accepted name: gallate dioxygenase

Reaction: 3,4,5-trihydroxybenzoate + O₂ = (1E)-4-oxobut-1-ene-1,2,4-tricarboxylate

Other name(s): GalA; gallate:oxygen oxidoreductase
Systematic name: 3,4,5-trihydroxybenzoate:oxygen oxidoreductase
Comments: Contains non-heme Fe²⁺. The enzyme is a ring-cleavage dioxygenase that acts specifically on 3,4,5-trihydroxybenzoate to produce the keto-tautomer of 4-oxalomesaconate [3094, 3093].
References: [3094, 3093]

[EC 1.13.11.57 created 2011]

EC 1.13.11.58

Accepted name: linoleate 9*S*-lipoxygenase
Reaction: linoleate + O₂ = (9*S*,10*E*,12*Z*)-9-hydroperoxy-10,12-octadecadienoate
Other name(s): 9-lipoxygenase; 9*S*-lipoxygenase; linoleate 9-lipoxygenase; LOX1 (gene name); 9*S*-LOX
Systematic name: linoleate:oxygen 9*S*-oxidoreductase
Comments: Contains nonheme iron. A common plant lipoxygenase that oxidizes linoleate and α-linolenate, the two most common polyunsaturated fatty acids in plants, by inserting molecular oxygen at the C₉ position with (*S*)-configuration. The enzyme plays a physiological role during the early stages of seedling growth. The enzyme from *Arabidopsis thaliana* shows comparable activity towards linoleate and linolenate [213]. EC 1.13.11.12 (linoleate 13*S*-lipoxygenase) catalyses a similar reaction at another position of these fatty acids.
References: [4430, 366, 99, 213]

[EC 1.13.11.58 created 2011]

EC 1.13.11.59

Accepted name: torulene dioxygenase
Reaction: torulene + O₂ = 4'-apo-β,ψ-caroten-4'-al + 3-methylbut-2-enal
Other name(s): CAO-2; CarT
Systematic name: torulene:oxygen oxidoreductase
Comments: It is assumed that 3-methylbut-2-enal is formed. The enzyme cannot cleave the saturated 3',4'-bond of γ-carotene which implies that a 3',4'-double bond is necessary for this reaction.
References: [3368, 3624, 1066]

[EC 1.13.11.59 created 2011]

EC 1.13.11.60

Accepted name: linoleate 8*R*-lipoxygenase
Reaction: linoleate + O₂ = (8*R*,9*Z*,12*Z*)-8-hydroperoxyoctadeca-9,12-dienoate
Other name(s): linoleic acid 8*R*-dioxygenase; 5,8-LDS (bifunctional enzyme); 7,8-LDS (bifunctional enzyme); 5,8-linoleate diol synthase (bifunctional enzyme); 7,8-linoleate diol synthase (bifunctional enzyme); PpoA
Systematic name: linoleate:oxygen (8*R*)-oxidoreductase
Comments: The enzyme contains heme [450, 4083]. The bifunctional enzyme from *Aspergillus nidulans* uses different heme domains to catalyse two separate reactions. Linoleic acid is oxidized within the N-terminal heme peroxidase domain to (8*R*,9*Z*,12*Z*)-8-hydroperoxyoctadeca-9,12-dienoate, which is subsequently isomerized by the C-terminal *P*-450 heme thiolate domain to (5*S*,8*R*,9*Z*,12*Z*)-5,8-dihydroxyoctadeca-9,12-dienoate (*cf.* EC 5.4.4.5, 9,12-octadecadienoate 8-hydroperoxide 8*R*-isomerase) [450]. The bifunctional enzyme from *Gaeumannomyces graminis* also catalyses the oxidation of linoleic acid to (8*R*,9*Z*,12*Z*)-8-hydroperoxyoctadeca-9,12-dienoate, but its second domain isomerizes it to (7*S*,8*S*,9*Z*,12*Z*)-5,8-dihydroxyoctadeca-9,12-dienoate (*cf.* EC 5.4.4.6, 9,12-octadecadienoate 8-hydroperoxide 8*S*-isomerase) [4083].
References: [450, 1492, 1278, 4083]

[EC 1.13.11.60 created 2011]

EC 1.13.11.61

- Accepted name:** linolenate 9*R*-lipoxygenase
Reaction: α -linolenate + O₂ = (9*R*,10*E*,12*Z*,15*Z*)-9-hydroperoxyoctadeca-10,12,15-trienoate
Other name(s): NspLOX; (9*R*)-LOX; linoleate 9*R*-dioxygenase
Systematic name: α -linolenate:oxygen (9*R*)-oxidoreductase
Comments: In cyanobacteria the enzyme is involved in oxylipin biosynthesis. The enzyme also converts linoleate to (9*R*,10*E*,12*Z*)-9-hydroperoxyoctadeca-10,12-dienoate.
References: [1908, 100, 2342]

[EC 1.13.11.61 created 2011]

EC 1.13.11.62

- Accepted name:** linoleate 10*R*-lipoxygenase
Reaction: linoleate + O₂ = (8*E*,10*R*,12*Z*)-10-hydroperoxy-8,12-octadecadienoate
Other name(s): 10*R*-DOX; (10*R*)-dioxygenase; 10*R*-dioxygenase
Systematic name: linoleate:oxygen (10*R*)-oxidoreductase
Comments: The enzyme is involved in biosynthesis of oxylipins, which affect sporulation, development, and pathogenicity of *Aspergillus* spp.
References: [1279, 1907]

[EC 1.13.11.62 created 2011]

EC 1.13.11.63

- Accepted name:** β -carotene 15,15'-dioxygenase
Reaction: β -carotene + O₂ = 2 *all-trans*-retinal
Other name(s): *blh* (gene name); BCO1 (gene name); BCDO (gene name); carotene dioxygenase; carotene 15,15'-dioxygenase; BCMO1 (misleading); β -carotene 15,15'-monooxygenase (incorrect)
Systematic name: β -carotene:oxygen 15,15'-dioxygenase (bond-cleaving)
Comments: Requires Fe²⁺. The enzyme cleaves β -carotene symmetrically, producing two molecules of *all-trans*-retinal. Both atoms of the oxygen molecule are incorporated into the products [870]. The enzyme can also process β -cryptoxanthin, 8'-apo- β -carotenal, 4'-apo- β -carotenal, α -carotene and γ -carotene in decreasing order. The presence of at least one unsubstituted β -ionone ring in a substrate greater than C₃₀ is mandatory [2116]. A prokaryotic enzyme has been reported from the uncultured marine bacterium 66A03, where it is involved in the proteorhodopsin system, which uses retinal as its chromophore [2115, 2117].
References: [1364, 1363, 4757, 2424, 2116, 2115, 2117, 870]

[EC 1.13.11.63 created 2012 (EC 1.14.99.36 created 1972 as EC 1.13.11.21, transferred 2001 to EC 1.14.99.36, incorporated 2015), modified 2016]

EC 1.13.11.64

- Accepted name:** 5-nitrosalicylate dioxygenase
Reaction: 5-nitrosalicylate + O₂ = 2-oxo-3-(5-oxofuran-2-ylidene)propanoate + nitrite (overall reaction)
(1a) 5-nitrosalicylate + O₂ = 4-nitro-6-oxohepta-2,4-dienedioate
(1b) 4-nitro-6-oxohepta-2,4-dienedioate = 2-oxo-3-(5-oxofuran-2-ylidene)propanoate + nitrite (spontaneous)
Other name(s): *naaB* (gene name); 5-nitrosalicylate:oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: 5-nitrosalicylate:oxygen 1,2-oxidoreductase (ring-opening)
Comments: The enzyme, characterized from the soil bacterium *Bradyrhizobium* sp. JS329, is involved in the pathway of 5-nitroanthranilate degradation. It is unusual in being able to catalyse the ring fission without the requirement for prior removal of the nitro group. The product undergoes spontaneous lactonization, with concurrent elimination of the nitro group.
References: [3408, 3409]

[EC 1.13.11.64 created 2012]

EC 1.13.11.65

- Accepted name:** carotenoid isomeroxygenase
Reaction: zeaxanthin + O₂ = (3*R*)-11-*cis*-3-hydroxyretinal + (3*R*)-*all-trans*-3-hydroxyretinal
Other name(s): *ninaB* (gene name)
Systematic name: zeaxanthin:oxygen 15,15'-oxidoreductase (bond-cleaving, *cis*-isomerizing)
Comments: The enzyme, characterized from the moth *Galleria mellonella* and the fruit fly *Drosophila melanogaster*, is involved in the synthesis of retinal from dietary carotenoids in insects. The enzyme accepts different *all-trans* carotenoids, including β-carotene, α-carotene and lutein, and catalyses the symmetrical cleavage of the carotenoid and the simultaneous isomerization of only one of the products to a *cis* configuration. When the substrate is hydroxylated only in one side (as in cryptoxanthin), the enzyme preferentially isomerizes the hydroxylated part of the molecule.
References: [3124]

[EC 1.13.11.65 created 2012 as EC 1.14.13.164, transferred 2012 to EC 1.13.11.65]

EC 1.13.11.66

- Accepted name:** hydroquinone 1,2-dioxygenase
Reaction: benzene-1,4-diol + O₂ = (2*Z*,4*E*)-4-hydroxy-6-oxohexa-2,4-dienoate
Other name(s): hydroquinone dioxygenase; benzene-1,4-diol:oxygen 1,2-oxidoreductase (decyclizing)
Systematic name: benzene-1,4-diol:oxygen 1,2-oxidoreductase (ring-opening)
Comments: The enzyme is an extradiol-type dioxygenase, and is a member of the nonheme-iron(II)-dependent dioxygenase family. It catalyses the ring cleavage of a wide range of hydroquinone substrates to produce the corresponding 4-hydroxymuconic semialdehydes.
References: [2841, 2871, 3841]

[EC 1.13.11.66 created 2012]

EC 1.13.11.67

- Accepted name:** 8'-apo-β-carotenoid 14',13'-cleaving dioxygenase
Reaction: 8'-apo-β-carotenol + O₂ = 14'-apo-β-carotenal + an uncharacterized product
Other name(s): 8'-apo-β-carotenol:O₂ oxidoreductase (14',13'-cleaving)
Systematic name: 8'-apo-β-carotenol:oxygen oxidoreductase (14',13'-cleaving)
Comments: A thiol-dependent enzyme isolated from rat and rabbit. Unlike EC 1.13.11.63, β-carotene-15,15'-dioxygenase, it is not active towards β-carotene. The secondary product has not been characterized, but may be (3*E*,5*E*)-7-hydroxy-6-methylhepta-3,5-dien-2-one.
References: [932]

[EC 1.13.11.67 created 2000 as EC 1.13.12.12, transferred 2012 to EC 1.13.11.67]

EC 1.13.11.68

- Accepted name:** 9-*cis*-β-carotene 9',10'-cleaving dioxygenase
Reaction: 9-*cis*-β-carotene + O₂ = 9-*cis*-10'-apo-β-carotenal + β-ionone
Other name(s): CCD7 (gene name); MAX3 (gene name); NCED7 (gene name)
Systematic name: 9-*cis*-β-carotene:oxygen oxidoreductase (9',10'-cleaving)
Comments: Requires Fe²⁺. The enzyme participates in a pathway leading to biosynthesis of strigolactones, plant hormones involved in promotion of symbiotic associations known as arbuscular mycorrhiza.
References: [388, 63]

[EC 1.13.11.68 created 2012]

EC 1.13.11.69

- Accepted name:** carlactone synthase
Reaction: 9-*cis*-10'-apo- β -carotenal + 2 O₂ = carlactone + (2*E*,4*E*,6*E*)-7-hydroxy-4-methylhepta-2,4,6-trienal
Other name(s): CCD8 (gene name); MAX4 (gene name); NCED8 (gene name)
Systematic name: 9-*cis*-10'-apo- β -carotenal:oxygen oxidoreductase (14,15-cleaving, carlactone-forming)
Comments: Requires Fe²⁺. The enzyme participates in a pathway leading to biosynthesis of strigolactones, plant hormones involved in promotion of symbiotic associations known as arbuscular mycorrhiza. Also catalyses EC 1.13.11.70, *all-trans*-10'-apo- β -carotenal 13,14-cleaving dioxygenase, but 10-fold slower.
References: [3969, 3767, 63]

[EC 1.13.11.69 created 2012]

EC 1.13.11.70

- Accepted name:** *all-trans*-10'-apo- β -carotenal 13,14-cleaving dioxygenase
Reaction: *all-trans*-10'-apo- β -carotenal + O₂ = 13-apo- β -carotenone + (2*E*,4*E*,6*E*)-4-methylocta-2,4,6-trienedial
Other name(s): CCD8 (gene name); MAX4 (gene name); NCED8 (gene name); *all-trans*-10'-apo- β -carotenal:O₂ oxidoreductase (13,14-cleaving)
Systematic name: *all-trans*-10'-apo- β -carotenal:oxygen oxidoreductase (13,14-cleaving)
Comments: Requires Fe²⁺. The enzyme from the plant *Arabidopsis thaliana* also catalyses EC 1.13.11.69, carlactone synthase, 10-fold faster.
References: [3767]

[EC 1.13.11.70 created 2012]

EC 1.13.11.71

- Accepted name:** carotenoid-9',10'-cleaving dioxygenase
Reaction: *all-trans*- β -carotene + O₂ = *all-trans*-10'-apo- β -carotenal + β -ionone
Other name(s): BCO₂ (gene name); β -carotene 9',10'-monooxygenase (misleading); *all-trans*- β -carotene:O₂ oxidoreductase (9',10'-cleaving)
Systematic name: *all-trans*- β -carotene:oxygen oxidoreductase (9',10'-cleaving)
Comments: Requires Fe²⁺. The enzyme catalyses the asymmetric oxidative cleavage of carotenoids. The mammalian enzyme can also cleave *all-trans*-lycopene.
References: [2087, 2500]

[EC 1.13.11.71 created 2012]

EC 1.13.11.72

- Accepted name:** 2-hydroxyethylphosphonate dioxygenase
Reaction: 2-hydroxyethylphosphonate + O₂ = hydroxymethylphosphonate + formate
Other name(s): HEPD; *phpD* (gene name); 2-hydroxyethylphosphonate:O₂ 1,2-oxidoreductase (hydroxymethylphosphonate forming)
Systematic name: 2-hydroxyethylphosphonate:oxygen 1,2-oxidoreductase (hydroxymethylphosphonate-forming)
Comments: Requires non-heme-iron(II). Isolated from some bacteria including *Streptomyces hygroscopicus* and *Streptomyces viridochromogenes*. The *pro-R* hydrogen at C-2 of the ethyl group is retained by the formate ion. Any stereochemistry at C-1 of the ethyl group is lost. One atom from dioxygen is present in each product. Involved in phosphinothricin biosynthesis.
References: [693, 4612, 3274]

[EC 1.13.11.72 created 2012]

EC 1.13.11.73

Accepted name: methylphosphonate synthase
Reaction: 2-hydroxyethylphosphonate + O₂ = methylphosphonate + HCO₃⁻
Other name(s): *mpnS* (gene name); 2-hydroxyethylphosphonate:O₂ 1,2-oxidoreductase (methylphosphonate forming)
Systematic name: 2-hydroxyethylphosphonate:oxygen 1,2-oxidoreductase (methylphosphonate-forming)
Comments: Isolated from the marine archaeon *Nitrosopumilus maritimus*.
References: [2781]

[EC 1.13.11.73 created 2012]

EC 1.13.11.74

Accepted name: 2-aminophenol 1,6-dioxygenase
Reaction: 2-aminophenol + O₂ = 2-aminomuconate 6-semialdehyde
Other name(s): *amnA* (gene name); *amnB* (gene name); 2-aminophenol:oxygen 1,6-oxidoreductase (decyclizing)
Systematic name: 2-aminophenol:oxygen 1,6-oxidoreductase (ring-opening)
Comments: The enzyme, a member of the nonheme-iron(II)-dependent dioxygenase family, is an extradiol-type dioxygenase that utilizes a non-heme ferrous iron to cleave the aromatic ring at the *meta* position (relative to the hydroxyl substituent). The enzyme also has some activity with 2-amino-5-methylphenol and 2-amino-4-methylphenol [4182]. The enzyme from the bacterium *Comamonas testosteroni* CNB-1 also has the activity of EC 1.13.11.76, 2-amino-5-chlorophenol 1,6-dioxygenase [4675].
References: [4182, 4675, 2441]

[EC 1.13.11.74 created 2013]

EC 1.13.11.75

Accepted name: *all-trans*-8'-apo-β-carotenal 15,15'-oxygenase
Reaction: *all-trans*-8'-apo-β-carotenal + O₂ = *all-trans*-retinal + (2*E*,4*E*,6*E*)-2,6-dimethylocta-2,4,6-trienedial
Other name(s): Diox1; ACO; 8'-apo-β-carotenal 15,15'-oxygenase
Systematic name: *all-trans*-8'-apo-β-carotenal:oxygen 15,15'-oxidoreductase (bond-cleaving)
Comments: Contains an Fe²⁺-4His arrangement. The enzyme is involved in retinal biosynthesis in bacteria [2157].
References: [3591, 2157]

[EC 1.13.11.75 created 2010 as EC 1.14.99.41, transferred 2013 to EC 1.13.11.75]

EC 1.13.11.76

Accepted name: 2-amino-5-chlorophenol 1,6-dioxygenase
Reaction: 2-amino-5-chlorophenol + O₂ = 2-amino-5-chloromuconate 6-semialdehyde
Other name(s): *cnbC* (gene name); 2-amino-5-chlorophenol:oxygen 1,6-oxidoreductase (decyclizing)
Systematic name: 2-amino-5-chlorophenol:oxygen 1,6-oxidoreductase (ring-opening)
Comments: The enzyme, a member of the nonheme-iron(II)-dependent dioxygenase family, is an extradiol-type dioxygenase that utilizes a non-heme ferrous iron to cleave the aromatic ring at the *meta* position (relative to the hydroxyl substituent). The enzyme from the bacterium *Comamonas testosteroni* CNB-1 also has the activity of EC 1.13.11.74, 2-aminophenol 1,6-dioxygenase.
References: [4675]

[EC 1.13.11.76 created 2013]

EC 1.13.11.77

Accepted name: oleate 10*S*-lipoxygenase
Reaction: (1) oleate + O₂ = (8*E*,10*S*)-10-hydroperoxyoctadeca-8-enoate
(2) linoleate + O₂ = (8*E*,10*S*,12*Z*)-10-hydroperoxyoctadeca-8,12-dienoate
(3) α-linolenate + O₂ = (8*E*,10*S*,12*Z*,15*Z*)-10-hydroperoxyoctadeca-8,12,15-trienoate
Other name(s): 10*S*-DOX; (10*S*)-dioxygenase; 10*S*-dioxygenase

Systematic name: oleate:oxygen (10S)-oxidoreductase
Comments: Binds Fe²⁺. The enzyme isolated from the bacterium *Pseudomonas* sp. 42A2 has similar activity with all the three Δ⁹ fatty acids. *cf.* EC 1.13.11.62, linoleate 10R-lipoxygenase.
References: [511]

[EC 1.13.11.77 created 2013]

EC 1.13.11.78

Accepted name: 2-amino-1-hydroxyethylphosphonate dioxygenase (glycine-forming)
Reaction: (2-amino-1-hydroxyethyl)phosphonate + O₂ = glycine + phosphate
Other name(s): *phnZ* (gene name)
Systematic name: 2-amino-1-hydroxyethylphosphonate:oxygen 1-oxidoreductase (glycine-forming)
Comments: Requires Fe²⁺. The enzyme, characterized from a marine bacterium, is involved in a 2-aminoethylphosphonate degradation pathway.
References: [2757, 4669]

[EC 1.13.11.78 created 2014]

EC 1.13.11.79

Accepted name: aerobic 5,6-dimethylbenzimidazole synthase
Reaction: FMNH₂ + O₂ = 5,6-dimethylbenzimidazole + D-erythrose 4-phosphate + other product(s)
Other name(s): BluB; flavin destructase
Systematic name: FMNH₂ oxidoreductase (5,6-dimethylbenzimidazole-forming)
Comments: The enzyme catalyses a complex oxygen-dependent conversion of reduced flavin mononucleotide to form 5,6-dimethylbenzimidazole, the lower ligand of vitamin B₁₂. This conversion involves many sequential steps in two distinct stages, and an alloxan intermediate that acts as a proton donor, a proton acceptor, and a hydride acceptor [4536]. The C-2 of 5,6-dimethylbenzimidazole is derived from C-1' of the ribityl group of FMNH₂ and 2-H from the ribityl 1'-*pro-S* hydrogen. While D-erythrose 4-phosphate has been shown to be one of the byproducts, the nature of the other product(s) has not been verified yet.
References: [1392, 1005, 4160, 4536, 713]

[EC 1.13.11.79 created 2010 as EC 1.14.99.40, transferred 2014 to EC 1.13.11.79, modified 2019]

EC 1.13.11.80

Accepted name: (3,5-dihydroxyphenyl)acetyl-CoA 1,2-dioxygenase
Reaction: (3,5-dihydroxyphenyl)acetyl-CoA + O₂ = 2-(3,5-dihydroxyphenyl)-2-oxoacetate + CoA
Other name(s): DpgC
Systematic name: (3,5-dihydroxyphenyl)acetyl-CoA:oxygen oxidoreductase
Comments: The enzyme, characterized from bacteria *Streptomyces toyocaensis* and *Amycolatopsis orientalis*, is involved in the biosynthesis of (3,5-dihydroxyphenyl)glycine, a component of the glycopeptide antibiotic vancomycin.
References: [630, 4618, 1117]

[EC 1.13.11.80 created 2015]

EC 1.13.11.81

Accepted name: 7,8-dihydroneopterin oxygenase
Reaction: 7,8-dihydroneopterin + O₂ = 7,8-dihydroxanthopterin + formate + glycolaldehyde
Systematic name: 7,8-dihydroneopterin:oxygen oxidoreductase
Comments: The enzyme from the bacterium *Mycobacterium tuberculosis* is multifunctional and also catalyses the epimerisation of the 2'-hydroxy group of 7,8-dihydroneopterin (EC 5.1.99.8, 7,8-dihydroneopterin epimerase) and the reaction of EC 4.1.2.25 (dihydroneopterin aldolase).

References: [793]

[EC 1.13.11.81 created 2015]

EC 1.13.11.82

Accepted name: 8'-apo-carotenoid 13,14-cleaving dioxygenase
Reaction: 8'-apo- β -carotenal + O₂ = 13-apo- β -carotenone + 2,6-dimethyldeca-2,4,6,8-tetraenedial
Other name(s): NACOX1 (gene name)
Systematic name: 8'-apo- β -carotenal:oxygen 13,14-dioxygenase (bond-cleaving)
Comments: Isolated from the bacterium *Novosphingobium aromaticivorans*. It is less active with 4'-apo- β -carotenal and γ -carotene.
References: [2118]

[EC 1.13.11.82 created 2015]

EC 1.13.11.83

Accepted name: 4-hydroxy-3-prenylphenylpyruvate oxygenase
Reaction: 3-(4-hydroxy-3-prenylphenyl)pyruvate + O₂ = 4-hydroxy-3-prenylmandelate + CO₂
Other name(s): CloR
Systematic name: 3-(4-hydroxy-3-prenylphenyl)pyruvate:oxygen 1,2-oxidoreductase (4-hydroxy-3-prenylmandelate-forming)
Comments: Requires non-heme-iron(II). Isolated from the bacterium *Streptomyces roseochromogenes* DS 12976. A bifunctional enzyme involved in chlorobiocin biosynthesis that also catalyses the activity of EC 1.13.12.23, 4-hydroxy-3-prenylbenzoate synthase.
References: [3342]

[EC 1.13.11.83 created 2017]

EC 1.13.11.84

Accepted name: crocetin dialdehyde synthase
Reaction: zeaxanthin + 2 O₂ = crocetin dialdehyde + 2 3 β -hydroxy- β -cyclocitral (overall reaction)
(1a) zeaxanthin + O₂ = 3 β -hydroxy-8'-apo- β -carotenal + 3 β -hydroxy- β -cyclocitral
(1b) 3 β -hydroxy-8'-apo- β -carotenal + O₂ = crocetin dialdehyde + 3 β -hydroxy- β -cyclocitral
Other name(s): CCD2; zeaxanthin 7,8-dioxygenase
Systematic name: zeaxanthin:oxygen 7',8'-oxidoreductase (bond-cleaving)
Comments: The enzyme, characterized from the plant *Crocus sativus* (saffron), acts twice, cleaving 3 β -hydroxy- β -cyclocitral off each 3-hydroxy end group. It is part of the zeaxanthin degradation pathway in that plant, leading to the different compounds that impart the color, flavor and aroma of the saffron spice. The enzyme can similarly cleave the 7-8 double bond of other carotenoids with a 3-hydroxy- β -carotenoid end group.
References: [1197, 41, 40]

[EC 1.13.11.84 created 2011 as EC 1.14.99.42, modified 2014, transferred 2017 to EC 1.13.11.84]

EC 1.13.11.85

Accepted name: exo-cleaving rubber dioxygenase
Reaction: *cis*-1,4-polyisoprene + *n* O₂ = *n* (4*Z*,8*Z*)-4,8-dimethyl-12-oxotrideca-4,8-dienal
Other name(s): *roxA* (gene name); heme-dependent rubber oxygenase (ambiguous)
Systematic name: *cis*-1,4-polyisoprene:oxygen dioxygenase [(4*Z*,8*Z*)-4,8-dimethyl-12-oxotrideca-4,8-dienal-forming]
Comments: The enzyme, studied mainly from the bacterium *Xanthomonas* sp. 35Y, catalyses the cleavage of the double bonds in natural and synthetic rubber (*cis*-1,4-polyisoprene polymers), generating ends that contain ketone and aldehyde groups. The enzyme from *Xanthomonas* sp. 35Y contains two *c*-type cytochromes. It attacks the substrate from its end, producing a single product of 15 carbons.

References: [4338, 1898, 416, 415, 3786, 336]

[EC 1.13.11.85 created 2018]

EC 1.13.11.86

Accepted name: 5-aminosalicylate 1,2-dioxygenase
Reaction: 5-aminosalicylate + O₂ = (2Z,4E)-4-amino-6-oxohepta-2,4-dienedioate
Other name(s): *mabB* (gene name)
Systematic name: 5-aminosalicylate:oxygen 1,2-oxidoreductase (ring-opening)
Comments: Requires iron(II). The enzyme, characterized from different bacteria, is a nonheme iron dioxygenase in the bicupin family.
References: [4047, 4834]

[EC 1.13.11.86 created 2018]

EC 1.13.11.87

Accepted name: endo-cleaving rubber dioxygenase
Reaction: Cleavage of *cis*-1,4-polyisoprene polymers into a mixture of compounds, including a C₂₀ compound ((4Z,8Z,12Z,16Z,20Z,24Z)-4,8,12,16,20,24-hexamethyl-28-oxononacos-4,8,12,16,20,24-hexaenal), a C₂₅ compound ((4Z,8Z,12Z,16Z,20Z)-4,8,12,16,20-pentamethyl-24-oxopentacos-4,8,12,16,20-pentaenal), a C₃₀ compound ((4Z,8Z,12Z,16Z)-4,8,12,16-tetramethyl-20-oxohenicosa-4,8,12,16-tetraenal), and larger isoprenologes such as C₃₅, C₄₀, C₄₅, and higher analogues.
Other name(s): latex clearing protein; *lcp* (gene name); *roxB* (gene name)
Systematic name: *cis*-1,4-polyisoprene:oxygen dioxygenase (endo-cleaving)
Comments: The enzyme catalyses the cleavage of the double bonds in natural and synthetic rubber, producing a mixture of C₂₀, C₂₅, C₃₀, and higher oligo-isoprenoids with ketone and aldehyde groups at their ends. Two unrelated bacterial enzymes are known to possess this activity - the enzyme from *Streptomyces* sp. K30 (*Lcp*) contains a *b*-type cytochrome, while the enzyme from *Xanthomonas* sp. 35Y, (*RoxB*) contains two *c*-type cytochromes. Both enzymes attack the substrate at random locations, and are not able to cleave the C₃₅ or smaller products into shorter fragments.
References: [4338, 1898, 416, 415, 3786, 336, 337]

[EC 1.13.11.87 created 2018]

EC 1.13.11.88

Accepted name: isoeugenol monooxygenase
Reaction: isoeugenol + O₂ = vanillin + acetaldehyde
Other name(s): *iem* (gene name)
Systematic name: isoeugenol:oxygen 7,8-oxidoreductase (bond-cleaving)
Comments: Contains iron(II). The enzyme, characterised from the bacteria *Pseudomonas putida* and *Pseudomonas nitroreducens*, catalyses the epoxidation of the double bond in the side chain of isoeugenol, followed by a second oxygenation and cleavage of the side chain in the form of acetaldehyde.
References: [3879, 4724, 4725, 3614, 3613]

[EC 1.13.11.88 created 2019]

EC 1.13.11.89

Accepted name: (hydroxymethyl)phosphonate dioxygenase
Reaction: (hydroxymethyl)phosphonate + O₂ = formate + phosphate
Other name(s): *phnZ1* (gene name)
Systematic name: (hydroxymethyl)phosphonate:oxygen 1-oxidoreductase (formate-forming)

Comments: Requires iron(II). The enzyme, characterized from the marine bacterium *Gimesia maris*, participates in a methylphosphonate degradation pathway. It also has the activity of EC 1.13.11.78, (2-amino-1-hydroxyethyl)phosphonate dioxygenase (glycine-forming).

References: [1265]

[EC 1.13.11.89 created 2019]

EC 1.13.11.90

Accepted name: [1-hydroxy-2-(trimethylamino)ethyl]phosphonate dioxygenase (glycine-betaine-forming)

Reaction: [(1*R*)-1-hydroxy-2-(trimethylamino)ethyl]phosphonate + O₂ = glycine betaine + phosphate

Other name(s): *tmpB* (gene name)

Systematic name: [(1*R*)-1-hydroxy-2-(trimethylamino)ethyl]phosphonate:oxygen 1*R*-oxidoreductase (glycine-betaine-forming)

Comments: Requires Fe²⁺. This bacterial enzyme is involved in a degradation pathway for [2-(trimethylamino)ethyl]phosphonate.

References: [3437]

[EC 1.13.11.90 created 2020]

EC 1.13.11.91

Accepted name: 3-mercaptopropionate dioxygenase

Reaction: 3-sulfanylpropanoate + O₂ = 3-sulfinopropanoate

Other name(s): *mdo* (gene name); 3-mercaptopropionic acid dioxygenase; 3-sulfanylpropanoate dioxygenase

Systematic name: 3-sulfanylpropanoate:oxygen oxidoreductase

Comments: This bacterial enzyme contains an iron(2+) atom coordinated by three protein-derived histidines and a Ser-His-Tyr motif. It is similar to EC 1.13.11.20, cysteine dioxygenase, and can act on L-cysteine, but has a much higher activity with its native substrate, 3-sulfanylpropanoate.

References: [473, 966, 3318, 4231, 1097, 776, 72, 3663]

[EC 1.13.11.91 created 2020]

EC 1.13.11.92

Accepted name: fatty acid α-dioxygenase

Reaction: a fatty acid + O₂ = a (2*R*)-2-hydroperoxyfatty acid

Other name(s): DOX1 (gene name)

Systematic name: fatty acid:oxygen 2-oxidoreductase [(2*R*)-2-hydroperoxyfatty acid-forming]

Comments: Contains heme. This plant enzyme catalyses the (2*R*)-hydroperoxidation of fatty acids. It differs from lipoxygenases and cyclooxygenases in that the oxygen addition does not target an unsaturated region in the fatty acid. *In vitro* the product undergoes spontaneous decarboxylation, resulting in formation of a chain-shortened aldehyde. *In vivo* the product may be reduced to a (2*R*)-2-hydroxyfatty acid. The enzyme, which is involved in responses to different abiotic and biotic stresses, has a wide substrate range that includes both saturated and unsaturated fatty acids.

References: [45, 1490, 3625, 2187, 2524, 2764]

[EC 1.13.11.92 created 2021]

EC 1.13.11.93

Accepted name: 2-oxoadipate dioxygenase/decarboxylase

Reaction: 2-oxoadipate + O₂ = (*R*)-2-hydroxyglutarate + CO₂

Other name(s): *ycdJ* (gene name)

Systematic name: 2-oxoadipate dioxygenase/carboxy lyase

Comments: The enzyme, characterized from the bacterium *Pseudomonas putida*, is involved in an L-lysine catabolic pathway. Contains Fe(II).

References: [4274]

[EC 1.13.11.93 created 2022]

EC 1.13.12 With incorporation of one atom of oxygen (internal monooxygenases or internal mixed-function oxidases)

EC 1.13.12.1

Accepted name: arginine 2-monooxygenase
Reaction: L-arginine + O₂ = 4-guanidinobutanamide + CO₂ + H₂O
Other name(s): arginine monooxygenase; arginine decarboxylase (incorrect); arginine oxygenase (decarboxylating); arginine decarboxy-oxidase
Systematic name: L-arginine:oxygen 2-oxidoreductase (decarboxylating)
Comments: A flavoprotein. Also acts on canavanine and homoarginine.
References: [3173, 4260, 4261]

[EC 1.13.12.1 created 1972]

EC 1.13.12.2

Accepted name: lysine 2-monooxygenase
Reaction: L-lysine + O₂ = 5-aminopentanamide + CO₂ + H₂O
Other name(s): lysine oxygenase; lysine monooxygenase; L-lysine-2-monooxygenase
Systematic name: L-lysine:oxygen 2-oxidoreductase (decarboxylating)
Comments: A flavoprotein (FAD). Also acts on other diamino acids.
References: [3002, 4177, 4178]

[EC 1.13.12.2 created 1972]

EC 1.13.12.3

Accepted name: tryptophan 2-monooxygenase
Reaction: L-tryptophan + O₂ = (indol-3-yl)acetamide + CO₂ + H₂O
Other name(s): tms1 (gene name); *iaaM* (gene name)
Systematic name: L-tryptophan:oxygen 2-oxidoreductase (decarboxylating)
Comments: The enzyme, studied from phytopathogenic bacteria such as *Pseudomonas savastanoi*, is involved in a pathway for the production of (indol-3-yl)acetate (IAA), the main auxin hormone in plants.
References: [2243, 2295, 1776, 3181, 1043]

[EC 1.13.12.3 created 1972]

EC 1.13.12.4

Accepted name: lactate 2-monooxygenase
Reaction: (S)-lactate + O₂ = acetate + CO₂ + H₂O
Other name(s): lactate oxidative decarboxylase; lactate oxidase; lactic oxygenase; lactate oxygenase; lactic oxidase; L-lactate monooxygenase; lactate monooxygenase; L-lactate-2-monooxygenase
Systematic name: (S)-lactate:oxygen 2-oxidoreductase (decarboxylating)
Comments: A flavoprotein (FMN).
References: [1575, 4132]

[EC 1.13.12.4 created 1961 as EC 1.1.3.2, transferred 1972 to EC 1.13.12.4]

EC 1.13.12.5

- Accepted name:** *Renilla*-type luciferase
- Reaction:** coelenterazine h + O₂ = excited coelenteramide h monoanion + CO₂ (over-all reaction)
(1a) coelenterazine h + O₂ = coelenterazine h dioxetanone
(1b) coelenterazine h dioxetanone = excited coelenteramide h monoanion + CO₂
- Other name(s):** *Renilla*-luciferin 2-monooxygenase; luciferase (*Renilla* luciferin); *Renilla*-luciferin:oxygen 2-oxidoreductase (decarboxylating)
- Systematic name:** coelenterazine h:oxygen 2-oxidoreductase (decarboxylating)
- Comments:** This enzyme has been studied from the soft coral *Renilla reniformis*. Before the reaction occurs the substrate is sequestered by a coelenterazine-binding protein. Elevation in the concentration of calcium ions releases the substrate, which then interacts with the luciferase. Upon binding the substrate, the enzyme catalyses an oxygenation, producing a very short-lived hydroperoxide that cyclizes into a dioxetanone structure, which collapses, releasing a CO₂ molecule. The spontaneous breakdown of the dioxetanone releases the energy (about 50 kcal/mole) that is necessary to generate the excited state of the coelenteramide product, which is the singlet form of the monoanion. *In vivo* the product undergoes the process of nonradiative energy transfer to an accessory protein, a green fluorescent protein (GFP), which results in green bioluminescence. *In vitro*, in the absence of GFP, the product emits blue light.
- References:** [738, 1721, 94, 3876, 606, 2541, 2531]

[EC 1.13.12.5 created 1976, modified 1981, modified 1982, modified 2004, modified 2017]

EC 1.13.12.6

- Accepted name:** *Cypridina*-luciferin 2-monooxygenase
- Reaction:** *Cypridina* luciferin + O₂ = oxidized *Cypridina* luciferin + CO₂ + *hν*
- Other name(s):** *Cypridina*-type luciferase; luciferase (*Cypridina* luciferin); *Cypridina* luciferase
- Systematic name:** *Cypridina*-luciferin:oxygen 2-oxidoreductase (decarboxylating)
- Comments:** *Cypridina* is a bioluminescent crustacea. The luciferins (and presumably the luciferases, since they cross-react) of some luminous fish (e.g. *Apogon*, *Parapriacanthus*, *Porichthys*) are apparently similar. The enzyme may be assayed by measurement of light emission.
- References:** [737, 1997, 2128, 4341]

[EC 1.13.12.6 created 1976, modified 1982]

EC 1.13.12.7

- Accepted name:** firefly luciferase
- Reaction:** D-firefly luciferin + O₂ + ATP = firefly oxyluciferin + CO₂ + AMP + diphosphate + *hν*
- Other name(s):** *Photinus*-luciferin 4-monooxygenase (ATP-hydrolysing); luciferase (firefly luciferin); *Photinus* luciferin 4-monooxygenase (adenosine triphosphate-hydrolysing); firefly luciferin luciferase; *Photinus pyralis* luciferase; *Photinus*-luciferin:oxygen 4-oxidoreductase (decarboxylating, ATP-hydrolysing)
- Systematic name:** D-firefly luciferin:oxygen 4-oxidoreductase (decarboxylating, ATP-hydrolysing)
- Comments:** The enzyme, which is found in fireflies (*Lampyridae*), is responsible for their bioluminescence. The reaction begins with the formation of an acid anhydride between the carboxylic group of D-firefly luciferin and AMP, with the release of diphosphate. An oxygenation follows, with release of the AMP group and formation of a very short-lived peroxide that cyclizes into a dioxetanone structure, which collapses, releasing a CO₂ molecule. The spontaneous breakdown of the dioxetanone (rather than the hydrolysis of the adenylate) releases the energy (about 50 kcal/mole) that is necessary to generate the excited state of oxyluciferin. The excited luciferin then emits a photon, returning to its ground state. The enzyme has a secondary acyl-CoA ligase activity when acting on L-firefly luciferin (see EC 6.2.1.52).
- References:** [1394, 4601, 1714, 4602, 2227, 856, 2980, 4126]

[EC 1.13.12.7 created 1976, modified 1981, modified 1982, modified 2017]

EC 1.13.12.8

Accepted name: *Watasenia*-luciferin 2-monooxygenase
Reaction: *Watasenia* luciferin + O₂ = oxidized *Watasenia* luciferin + CO₂ + *hν*
Other name(s): *Watasenia*-type luciferase
Systematic name: *Watasenia*-luciferin:oxygen 2-oxidoreductase (decarboxylating)
Comments: The enzyme from the luminous squid *Watasenia* may be assayed by measurement of light emission.
References: [1816]

[EC 1.13.12.8 created 1982]

EC 1.13.12.9

Accepted name: phenylalanine 2-monooxygenase
Reaction: L-phenylalanine + O₂ = 2-phenylacetamide + CO₂ + H₂O
Other name(s): L-phenylalanine oxidase (deaminating and decarboxylating); phenylalanine (deaminating, decarboxylating)oxidase
Systematic name: L-phenylalanine:oxygen 2-oxidoreductase (decarboxylating)
Comments: The reaction shown above is about 80% of the reaction catalysed; the remaining 20% is: L-phenylalanine + O₂ + H₂O = 3-phenylpyruvic acid + ammonia + H₂O₂; a reaction similar to that of EC 1.4.3.2, L-amino-acid oxidase.
References: [2250, 2252, 2251, 2253]

[EC 1.13.12.9 created 1986, modified 2003]

[1.13.12.10 Deleted entry. lysine 6-monooxygenase. Reaction covered by EC 1.14.13.59, L-lysine 6-monooxygenase (NADPH)]

[EC 1.13.12.10 created 1989, modified 1999, deleted 2001]

[1.13.12.11 Deleted entry. methylphenyltetrahydropyridine N-monooxygenase. The activity is due to EC 1.14.13.8, flavin-containing monooxygenase]

[EC 1.13.12.11 created 1992, deleted 2006]

[1.13.12.12 Transferred entry. apo-β-carotenoid-14',13'-dioxygenase. The enzyme was misclassified and has been transferred to EC 1.13.11.67, 8-apo-β-carotenoid 14',13'-cleaving dioxygenase]

[EC 1.13.12.12 created 2000, modified 2001, deleted 2012]

EC 1.13.12.13

Accepted name: *Oplophorus*-luciferin 2-monooxygenase
Reaction: *Oplophorus* luciferin + O₂ = oxidized *Oplophorus* luciferin + CO₂ + *hν*
Other name(s): *Oplophorus* luciferase
Systematic name: *Oplophorus*-luciferin:oxygen 2-oxidoreductase (decarboxylating)
Comments: The luciferase from the deep sea shrimp *Oplophorus gracilirostris* is a complex composed of more than one protein. The enzyme's specificity is quite broad, with both coelenterazine and bisdeoxycoelenterazine being good substrates.
References: [3878, 1818]

[EC 1.13.12.13 created 2004]

[1.13.12.14 Transferred entry. chlorophyllide-a oxygenase. Now EC 1.14.13.122, chlorophyllide-a oxygenase]

[EC 1.13.12.14 created 2006, deleted 2011]

EC 1.13.12.15

Accepted name: 3,4-dihydroxyphenylalanine oxidative deaminase
Reaction: 2 L-dopa + O₂ = 2 3,4-dihydroxyphenylpyruvate + 2 NH₃

Other name(s): 3,4-dihydroxy-L-phenylalanine: oxidative deaminase; oxidative deaminase; DOPA oxidative deaminase; DOPAODA

Systematic name: 3,4-dihydroxy-L-phenylalanine:oxygen oxidoreductase (deaminating)

Comments: This enzyme is one of the three enzymes involved in L-dopa (3,4-dihydroxy-L-phenylalanine) catabolism in the non-oxygenic phototrophic bacterium *Rubrivivax benzoatilyticus* OU5 (and not *Rhodobacter sphaeroides* OU5 as had been thought [3449]), the other two being EC 4.3.1.22 (dihydroxyphenylalanine reductive deaminase) and EC 2.6.1.49 (3,4-dihydroxyphenylalanine transaminase). In addition to L-dopa, the enzyme can also use L-tyrosine, L-phenylalanine, L-tryptophan and glutamate as substrate, but more slowly. The enzyme is inhibited by NADH and 2-oxoglutarate.

References: [3449]

[EC 1.13.12.15 created 2008]

EC 1.13.12.16

Accepted name: nitronate monooxygenase

Reaction: ethylnitronate + O₂ = acetaldehyde + nitrite + other products

Other name(s): NMO; 2-nitropropane dioxygenase (incorrect)

Systematic name: nitronate:oxygen 2-oxidoreductase (nitrite-forming)

Comments: Previously classified as 2-nitropropane dioxygenase (EC 1.13.11.32), but it is now recognized that this was the result of the slow ionization of nitroalkanes to their nitronate (anionic) forms. The enzymes from the fungus *Neurospora crassa* and the yeast *Williopsis saturnus* var. *mrakii* (formerly classified as *Hansenula mrakii*) contain non-covalently bound FMN as the cofactor. Neither hydrogen peroxide nor superoxide were detected during enzyme turnover. Active towards linear alkyl nitronates of lengths between 2 and 6 carbon atoms and, with lower activity, towards propyl-2-nitronate. The enzyme from *N. crassa* can also utilize neutral nitroalkanes, but with lower activity.

References: [1164, 1459, 1250, 1163]

[EC 1.13.12.16 created 1984 as EC 1.13.11.32, transferred 2009 to EC 1.13.12.16, modified 2011]

EC 1.13.12.17

Accepted name: dichloroarcyriaflavin A synthase

Reaction: dichlorochromopyrrolate + 4 O₂ + 4 NADH + 4 H⁺ = dichloroarcyriaflavin A + 2 CO₂ + 6 H₂O + 4 NAD⁺

Systematic name: dichlorochromopyrrolate,NADH:oxygen 2,5-oxidoreductase (dichloroarcyriaflavin A-forming)

Comments: The conversion of dichlorochromopyrrolate to dichloroarcyriaflavin A is a complex process that involves two enzyme components. RebP is an NAD-dependent cytochrome *P*-450 oxygenase that performs an aryl-aryl bond formation yielding the six-ring indolocarbazole scaffold [2627]. Along with RebC, a flavin-dependent hydroxylase, it also catalyses the oxidative decarboxylation of both carboxyl groups. The presence of RebC ensures that the only product is the rebeccamycin aglycone dichloroarcyriaflavin A [1744]. The enzymes are similar, but not identical, to StaP and StaC, which are involved in the synthesis of staurosporine [3652].

References: [2627, 1744, 3652]

[EC 1.13.12.17 created 2010]

EC 1.13.12.18

Accepted name: dinoflagellate luciferase

Reaction: dinoflagellate luciferin + O₂ = oxidized dinoflagellate luciferin + H₂O + hν

Other name(s): (dinoflagellate luciferin) luciferase; *Gonyaulax* luciferase

Systematic name: dinoflagellate-luciferin:oxygen 13²-oxidoreductase

Comments: A luciferase from dinoflagellates such as *Gonyaulax polyedra*, *Lingulodinium polyedrum*, *Noctiluca scintillans*, and *Pyrocystis lunula*. It is a single protein with three luciferase domains. The luciferin is strongly bound by a luciferin binding protein above a pH of 7.

References: [985, 2902, 178, 2448, 2901, 3753]

[EC 1.13.12.18 created 2011]

EC 1.13.12.19

Accepted name: 2-oxoglutarate dioxygenase (ethene-forming)
Reaction: 2-oxoglutarate + O₂ = ethene + 3 CO₂ + H₂O
Other name(s): ethylene-forming enzyme; EFE; 2-oxoglutarate dioxygenase (ethylene-forming); 2-oxoglutarate:oxygen oxidoreductase (decarboxylating, ethylene-forming)
Systematic name: 2-oxoglutarate:oxygen oxidoreductase (decarboxylating, ethene-forming)
Comments: This is one of two simultaneous reactions catalysed by the enzyme, which is responsible for ethene production in bacteria of the *Pseudomonas syringae* group. In the other reaction [EC 1.14.20.7, 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming)] the enzyme catalyses the mono-oxygenation of both 2-oxoglutarate and L-arginine, forming succinate, carbon dioxide and L-hydroxyarginine, which is subsequently cleaved into guanidine and (S)-1-pyrroline-5-carboxylate. The enzymes catalyse two cycles of the ethene-forming reaction for each cycle of the succinate-forming reaction, so that the stoichiometry of the products ethene and succinate is 2:1.
References: [2954, 1222, 1221]

[EC 1.13.12.19 created 2011]

EC 1.13.12.20

Accepted name: noranthrone monooxygenase
Reaction: norsolorinic acid anthrone + O₂ = norsolorinic acid + H₂O
Other name(s): norsolorinate anthrone oxidase
Systematic name: norsolorinic acid anthrone:oxygen 9-oxidoreductase (norsolorinic acid-forming)
Comments: Involved in the synthesis of aflatoxins in the fungus *Aspergillus parasiticus*.
References: [1026]

[EC 1.13.12.20 created 2013]

EC 1.13.12.21

Accepted name: tetracenomycin-F1 monooxygenase
Reaction: tetracenomycin F1 + O₂ = tetracenomycin D3 + H₂O
Other name(s): *tcmH* (gene name)
Systematic name: tetracenomycin-F1:oxygen C5-monooxygenase
Comments: The enzyme is involved in biosynthesis of the anthracycline antibiotic tetracenomycin C by the bacterium *Streptomyces glaucescens*.
References: [3838]

[EC 1.13.12.21 created 2013]

EC 1.13.12.22

Accepted name: deoxynogalenate monooxygenase
Reaction: deoxynogalenate + O₂ = nogalenate + H₂O
Other name(s): SnoaB (gene name); 12-deoxynogalonic acid oxidoreductase; [4,5-dihydroxy-10-oxo-3-(3-oxobutanoyl)-9,10-dihydroanthracen-2-yl]acetate oxidase; [4,5-dihydroxy-10-oxo-3-(3-oxobutanoyl)-9,10-dihydroanthracen-2-yl]acetate monooxygenase; deoxynogalenate oxidoreductase
Systematic name: deoxynogalenate:oxygen oxidoreductase
Comments: The enzyme, characterized from the bacterium *Streptomyces nogalater*, is involved in the biosynthesis of the aromatic polyketide nogalamycin.
References: [2241, 1416]

[EC 1.13.12.22 created 2015]

EC 1.13.12.23

- Accepted name:** 4-hydroxy-3-prenylbenzoate synthase
Reaction: 4-hydroxy-3-prenylmandelate + O₂ = 4-hydroxy-3-prenylbenzoate + CO₂ + H₂O
Other name(s): CloR; *novR* (gene name)
Systematic name: 4-hydroxy-3-prenylmandelate:oxygen oxidoreductase (4-hydroxy-3-prenylbenzoate forming)
Comments: Isolated from the bacterium *Streptomyces roseochromogenes* DS 12976. A bifunctional enzyme involved in clorobiocin biosynthesis that also catalyses the activity of EC 1.13.11.83, 4-hydroxy-3-prenylphenylpyruvate oxygenase.
References: [3342]

[EC 1.13.12.23 created 2017]

EC 1.13.12.24

- Accepted name:** calcium-regulated photoprotein
Reaction: [apoaequorin] + coelenterazine + O₂ + 3 Ca²⁺ = [excited state blue fluorescent protein] + CO₂ (overall reaction)
(1a) [apoaequorin] + coelenterazine = [apoaequorin containing coelenterazine]
(1b) [apoaequorin containing coelenterazine] + O₂ = [aequorin]
(1c) [aequorin] + 3 Ca²⁺ = [aequorin] 1,2-dioxetan-3-one
(1d) [aequorin] 1,2-dioxetan-3-one = [excited state blue fluorescent protein] + CO₂
Other name(s): Ca²⁺-regulated photoprotein; calcium-activated photoprotein; aequorin; obelin; halistaurin; mitrocomin; phialidin; clytin; mnemiopsin; berovin
Systematic name: coelenterazine:oxygen 2-oxidoreductase (decarboxylating, calcium-dependent)
Comments: Ca²⁺-regulated photoproteins are found in a variety of bioluminescent marine organisms, mostly coelenterates, and are responsible for their light emission. The best studied enzyme is from the jellyfish *Aequorea victoria*. The enzyme tightly binds the imidazolopyrazinone derivative coelenterazine, which is then peroxidized by oxygen. The hydroperoxide is stably bound until three Ca²⁺ ions bind to the protein, inducing a structural change that results in the formation of a 1,2-dioxetan-3-one ring, followed by decarboxylation and generation of a protein-bound coelenteramide in an excited state. The calcium-bound protein-product complex is known as a blue fluorescent protein. *In vivo* the energy is transferred to a green fluorescent protein (GFP) by Förster resonance energy transfer. *In vitro*, in the absence of GFP, coelenteramide emits a photon of blue light while returning to its ground state.
References: [3874, 2892, 1817, 1594, 878]

[EC 1.13.12.24 created 2018]

EC 1.13.99 Miscellaneous

EC 1.13.99.1

- Accepted name:** inositol oxygenase
Reaction: *myo*-inositol + O₂ = D-glucuronate + H₂O
Other name(s): *meso*-inositol oxygenase; *myo*-inositol oxygenase; MOO
Systematic name: *myo*-inositol:oxygen oxidoreductase
Comments: An iron protein.
References: [604, 3472, 134]

[EC 1.13.99.1 created 1961 as EC 1.99.2.6, transferred 1965 to EC 1.13.1.11, transferred 1972 to EC 1.13.99.1, modified 2002]

[1.13.99.2 Transferred entry. benzoate 1,2-dioxygenase. Now EC 1.14.12.10, benzoate 1,2-dioxygenase]

[EC 1.13.99.2 created 1972, deleted 1992]

EC 1.13.99.3

Accepted name: tryptophan 2'-dioxygenase
Reaction: L-tryptophan + O₂ = (indol-3-yl)glycolaldehyde + CO₂ + NH₃
Other name(s): indole-3-alkane α-hydroxylase; tryptophan side-chain α,β-oxidase; tryptophan side chain oxidase II; tryptophan side-chain oxidase; TSO; indolyl-3-alkan α-hydroxylase; tryptophan side chain oxidase type I; TSO I ; TSO II; tryptophan side chain oxidase
Systematic name: L-tryptophan:oxygen 2'-oxidoreductase (side-chain-cleaving)
Comments: A hemoprotein. Acts on a number of indole-3-alkane derivatives, oxidizing the 3-side-chain in the 2'-position. Best substrates were L-tryptophan and 5-hydroxy-L-tryptophan.
References: [3536, 4173]

[EC 1.13.99.3 created 1984]

[1.13.99.4 *Transferred entry. 4-chlorophenylacetate 3,4-dioxygenase. Now EC 1.14.12.9, 4-chlorophenylacetate 3,4-dioxygenase*]

[EC 1.13.99.4 created 1989, deleted 1992]

[1.13.99.5 *Transferred entry. now EC 1.13.11.47, 3-hydroxy-4-oxoquinoline 2,4-dioxygenase*]

[EC 1.13.99.5 created 1999, deleted 2001]

EC 1.14 Acting on paired donors, with incorporation or reduction of molecular oxygen

This subclass contains enzymes that act on two hydrogen-donors, and oxygen is incorporated into one or both of them. Sub-subclasses are based on the second donor and the number of oxygen atoms that are incorporated into one or both donors: 2-oxoglutarate is one donor and one atom of oxygen is incorporated into each donor (EC 1.14.11), NADH or NADPH is one donor, and two atoms of oxygen are incorporated into the other donor (EC 1.14.12), NADH or NADPH is one donor, but only one atom of oxygen is incorporated into the other donor (EC 1.14.13). In sub-subclasses EC 1.14.14-1.14.18, one atom of oxygen is incorporated into one donor, the other donor being a reduced flavin or flavoprotein (EC 1.14.14), a reduced iron-sulfur protein (EC 1.14.15), a reduced pteridine (EC 1.14.16), reduced ascorbate (EC 1.14.17), or some other compound (EC 1.14.18). Sub-subclass EC 1.14.19 differs from others in subclass EC 1.14 in that hydrogen atoms removed from the two donors are combined with O₂ to form two molecules of water. Sub-subclass EC 1.14.20 has 2-oxoglutarate as one donor, and the other is dehydrogenated. Sub-subclass EC 1.14.21 has NADH or NADPH as one donor, and the other is dehydrogenated. Sub-subclass EC 1.14.99 is for cases where information about the second donor is incomplete.

EC 1.14.1 With NADH or NADPH as one donor (deleted sub-subclass)

[1.14.1.1 *Transferred entry. now EC 1.14.14.1, unspecific monooxygenase*]

[EC 1.14.1.1 created 1961 as EC 1.99.1.1, transferred 1965 to EC 1.14.14.1, deleted 1972]

[1.14.1.2 *Transferred entry. now EC 1.14.13.9, kynurenine 3-monooxygenase*]

[EC 1.14.1.2 created 1965, deleted 1972]

[1.14.1.3 *Deleted entry. squalene hydroxylase. Activity is covered by EC 1.14.99.7, squalene monooxygenase and EC 5.4.99.7, lanosterol synthase*]

[EC 1.14.1.3 created 1961 as EC 1.99.1.13, transferred 1965 to EC 1.14.1.3, deleted 1972]

[1.14.1.4 *Transferred entry. now EC 1.14.99.2, kynurenine 7,8-hydroxylase*]

[EC 1.14.1.4 created 1965, deleted 1972]

[1.14.1.5 *Transferred entry. now EC 1.14.13.5, imidazoleacetate 4-monooxygenase*]

[EC 1.14.1.5 created 1965, deleted 1972]

- [1.14.1.6 *Transferred entry. now EC 1.14.15.4, steroid 11 β -monooxygenase*
[EC 1.14.1.6 created 1961 as EC 1.99.1.7, transferred 1965 to EC 1.14.1.6, deleted 1972]
- [1.14.1.7 *Transferred entry. now EC 1.14.99.9, steroid 17 α -monooxygenase*
[EC 1.14.1.7 created 1965, deleted 1972]
- [1.14.1.8 *Transferred entry. now EC 1.14.99.10, steroid 21-monooxygenase*
[EC 1.14.1.8 created 1965, deleted 1972]
- [1.14.1.9 *Deleted entry. cholesterol 20-hydroxylase*
[EC 1.14.1.9 created 1965, deleted 1972]
- [1.14.1.10 *Transferred entry. now EC 1.14.99.11, estradiol 6 β -monooxygenase*
[EC 1.14.1.10 created 1965, deleted 1972]
- [1.14.1.11 *Deleted entry. oestriol 2-hydroxylase*
[EC 1.14.1.11 created 1965, deleted 1972]

EC 1.14.2 With ascorbate as one donor (deleted sub-subclass)

- [1.14.2.1 *Transferred entry. now EC 1.14.17.1, dopamine β -monooxygenase*
[EC 1.14.2.1 created 1965, deleted 1972]
- [1.14.2.2 *Transferred entry. now EC 1.13.11.27, 4-hydroxyphenylpyruvate dioxygenase*
[EC 1.14.2.2 created 1961 as EC 1.99.1.14, transferred 1965 to EC 1.14.2.2, deleted 1972]

EC 1.14.3 With reduced pteridine as one donor (deleted sub-subclass)

- [1.14.3.1 *Transferred entry. now EC 1.14.16.1, phenylalanine 4-monooxygenase*
[EC 1.14.3.1 created 1961 as EC 1.99.1.2, transferred 1965 to EC 1.14.3.1, deleted 1972]

EC 1.14.11 With 2-oxoglutarate as one donor, and incorporation of one atom of oxygen into each donor

EC 1.14.11.1

- Accepted name:** γ -butyrobetaine dioxygenase
Reaction: 4-trimethylammoniobutanoate + 2-oxoglutarate + O₂ = 3-hydroxy-4-trimethylammoniobutanoate + succinate + CO₂
Other name(s): α -butyrobetaine hydroxylase; γ -butyrobetaine hydroxylase; butyrobetaine hydroxylase
Systematic name: 4-trimethylammoniobutanoate,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate.
References: [2501]

[EC 1.14.11.1 created 1972]

EC 1.14.11.2

- Accepted name:** procollagen-proline 4-dioxygenase
Reaction: procollagen L-proline + 2-oxoglutarate + O₂ = procollagen *trans*-4-hydroxy-L-proline + succinate + CO₂
Other name(s): P4HA (gene name); P4HB (gene name); procollagen hydroxylase; proline hydroxylase; proline,2-oxoglutarate 4-dioxygenase; collagen proline hydroxylase; hydroxylase, collagen proline; peptidyl proline hydroxylase; proline procollagen hydroxylase; proline, 2-oxoglutarate dioxygenase; prolyl hydroxylase; prolylprocollagen dioxygenase; prolylprocollagen hydroxylase; procollagen proline 4-hydroxylase; procollagen proline dioxygenase; procollagen proline hydroxylase; procollagen prolyl hydroxylase; prolyl 4-hydroxylase; prolyl-glycyl-peptide, 2-oxoglutarate:oxygen oxidoreductase, 4-hydroxylating; procollagen-proline 4-dioxygenase (ambiguous)
Systematic name: procollagen-L-proline,2-oxoglutarate:oxygen oxidoreductase (4-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme, which is located within the lumen of the endoplasmic reticulum, catalyses the 4-hydroxylation of prolines in -X-Pro-Gly- sequences. The 4-hydroxyproline residues are essential for the formation of the collagen triple helix. The enzyme forms a complex with protein disulfide isomerase and acts not only on procollagen but also on more than 15 other proteins that have collagen-like domains.
References: [1777, 2144, 2142, 297, 1924, 2333, 2953, 2143]

[EC 1.14.11.2 created 1972, modified 1981, modified 1983, modified 2017]

EC 1.14.11.3

- Accepted name:** pyrimidine-deoxynucleoside 2'-dioxygenase
Reaction: 2'-deoxyuridine + 2-oxoglutarate + O₂ = uridine + succinate + CO₂
Other name(s): deoxyuridine 2'-dioxygenase; deoxyuridine 2'-hydroxylase; pyrimidine deoxyribonucleoside 2'-hydroxylase; thymidine 2'-dioxygenase; thymidine 2'-hydroxylase; thymidine 2-oxoglutarate dioxygenase; thymidine dioxygenase
Systematic name: 2'-deoxyuridine,2-oxoglutarate:oxygen oxidoreductase (2'-hydroxylating)
Comments: Requires iron(II) and ascorbate. Also acts on thymidine. *cf.* EC 1.14.11.10, pyrimidine-deoxynucleoside 1'-dioxygenase.
References: [211, 4076, 4547]

[EC 1.14.11.3 created 1972, modified 1976, modified 1989, modified 2002]

EC 1.14.11.4

- Accepted name:** procollagen-lysine 5-dioxygenase
Reaction: [procollagen]-L-lysine + 2-oxoglutarate + O₂ = [procollagen]-(2*S*,5*R*)-5-hydroxy-L-lysine + succinate + CO₂
Other name(s): lysine hydroxylase; lysine,2-oxoglutarate 5-dioxygenase; procollagen lysine dioxygenase; collagen lysine hydroxylase; lysine-2-oxoglutarate dioxygenase; lysyl hydroxylase; lysylprocollagen dioxygenase; procollagen lysyl hydroxylase; peptidyl-lysine, 2-oxoglutarate: oxygen oxidoreductase; peptidyllysine, 2-oxoglutarate:oxygen 5-oxidoreductase; procollagen lysine hydroxylase; procollagen-L-lysine,2-oxoglutarate:oxygen oxidoreductase (5-hydroxylating); L-lysine-[procollagen],2-oxoglutarate:oxygen oxidoreductase (5-hydroxylating)
Systematic name: [procollagen]-L-lysine,2-oxoglutarate:oxygen oxidoreductase (5-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate.
References: [1567, 3510, 3394, 3395]

[EC 1.14.11.4 created 1972, modified 1983]

[1.14.11.5 Deleted entry. 5-hydroxymethyluracil,2-oxoglutarate dioxygenase. Now included with EC 1.14.11.6 thymine dioxygenase]

[EC 1.14.11.5 created 1972, deleted 1976]

EC 1.14.11.6

- Accepted name:** thymine dioxygenase
Reaction: thymine + 2-oxoglutarate + O₂ = 5-hydroxymethyluracil + succinate + CO₂
Other name(s): thymine 7-hydroxylase; 5-hydroxy-methyluracil dioxygenase; 5-hydroxymethyluracil oxygenase
Systematic name: thymine,2-oxoglutarate:oxygen oxidoreductase (7-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. Also acts on 5-hydroxymethyluracil to oxidize its -CH₂OH group first to -CHO and then to -COOH.
References: [210, 2511, 4547]

[EC 1.14.11.6 created 1972, modified 1976 (EC 1.14.11.5 created 1972, incorporated 1976)]

EC 1.14.11.7

- Accepted name:** procollagen-proline 3-dioxygenase
Reaction: [procollagen]-L-proline + 2-oxoglutarate + O₂ = [procollagen]-*trans*-3-hydroxy-L-proline + succinate + CO₂
Other name(s): proline,2-oxoglutarate 3-dioxygenase; prolyl 3-hydroxylase; procollagen proline 3-hydroxylase; prolyl-4-hydroxyprolyl-glycyl-peptide,2-oxoglutarate:oxygen oxidoreductase, 3-hydroxylating
Systematic name: [procollagen]-L-proline,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme forms a complex with protein disulfide isomerase, and is located in the endoplasmic reticulum. It modifies proline residues within the procollagen peptide of certain collagen types. The modification is essential for proper collagen triple helix formation.
References: [3527, 3528, 4476, 4282]

[EC 1.14.11.7 created 1981, modified 1983, modified 2017]

EC 1.14.11.8

- Accepted name:** trimethyllysine dioxygenase
Reaction: N⁶,N⁶,N⁶-trimethyl-L-lysine + 2-oxoglutarate + O₂ = (3*S*)-3-hydroxy-N⁶,N⁶,N⁶-trimethyl-L-lysine + succinate + CO₂
Other name(s): trimethyllysine α-ketoglutarate dioxygenase; TML-α-ketoglutarate dioxygenase; TML hydroxylase; 6-*N*,6-*N*,6-*N*-trimethyl-L-lysine,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Systematic name: N⁶,N⁶,N⁶-trimethyl-L-lysine,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate.
References: [1771, 4238, 2422, 3476]

[EC 1.14.11.8 created 1983]

EC 1.14.11.9

- Accepted name:** flavanone 3-dioxygenase
Reaction: a (2*S*)-flavan-4-one + 2-oxoglutarate + O₂ = a (2*R*,3*R*)-dihydroflavonol + succinate + CO₂
Other name(s): naringenin 3-hydroxylase; flavanone 3-hydroxylase; flavanone 3β-hydroxylase; flavanone synthase I; (2*S*)-flavanone 3-hydroxylase; naringenin,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating); F₃H; flavanone,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Systematic name: (2*S*)-flavan-4-one,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. This plant enzyme catalyses an early step in the flavonoid biosynthesis pathway, leading to the production of flavanols and anthocyanins. Substrates include (2*S*)-naringenin, (2*S*)-eriodictyol, (2*S*)-dihydrotricetin and (2*S*)-pinocembrin. Some enzymes are bifunctional and also catalyse EC 1.14.20.6, flavonol synthase.
References: [1140, 611, 3284, 4584, 1917, 3840]

[EC 1.14.11.9 created 1983, modified 1989, modified 2004, modified 2016]

EC 1.14.11.10

Accepted name: pyrimidine-deoxynucleoside 1'-dioxygenase
Reaction: 2'-deoxyuridine + 2-oxoglutarate + O₂ = uracil + 2-deoxyribonolactone + succinate + CO₂
Other name(s): deoxyuridine-uridine 1'-dioxygenase
Systematic name: 2'-deoxyuridine,2-oxoglutarate:oxygen oxidoreductase (1'-hydroxylating)
Comments: Requires iron(II) and ascorbate. *cf.* EC 1.14.11.3, pyrimidine-deoxynucleoside 2'-dioxygenase.
References: [4076]

[EC 1.14.11.10 created 1989, modified 2002]

EC 1.14.11.11

Accepted name: hyoscyamine (6*S*)-dioxygenase
Reaction: L-hyoscyamine + 2-oxoglutarate + O₂ = (6*S*)-hydroxyhyoscyamine + succinate + CO₂
Other name(s): hyoscyamine 6β-hydroxylase; hyoscyamine 6β-dioxygenase; hyoscyamine 6-hydroxylase
Systematic name: L-hyoscyamine,2-oxoglutarate:oxygen oxidoreductase [(6*S*)-hydroxylating]
Comments: Requires Fe²⁺ and ascorbate.
References: [1551]

[EC 1.14.11.11 created 1989]

EC 1.14.11.12

Accepted name: gibberellin-44 dioxygenase
Reaction: gibberellin 44 + 2-oxoglutarate + O₂ = gibberellin 19 + succinate + CO₂
Other name(s): oxygenase, gibberellin A44 oxidase; (gibberellin-44), 2-oxoglutarate:oxygen oxidoreductase
Systematic name: (gibberellin-44),2-oxoglutarate:oxygen oxidoreductase
Comments: Requires Fe²⁺.
References: [1327]

[EC 1.14.11.12 created 1990]

EC 1.14.11.13

Accepted name: gibberellin 2β-dioxygenase
Reaction: gibberellin 1 + 2-oxoglutarate + O₂ = 2β-hydroxygibberellin 1 + succinate + CO₂
Other name(s): gibberellin 2β-hydroxylase
Systematic name: (gibberellin-1),2-oxoglutarate:oxygen oxidoreductase (2β-hydroxylating)
Comments: Also acts on a number of other gibberellins.
References: [3947]

[EC 1.14.11.13 created 1990]

[1.14.11.14 *Transferred entry. 6β-hydroxyhyoscyamine epoxidase. Now EC 1.14.20.13, 6β-hydroxyhyoscyamine epoxidase*]

[EC 1.14.11.14 created 1992, deleted 2018]

EC 1.14.11.15

Accepted name: gibberellin 3β-dioxygenase
Reaction: gibberellin 20 + 2-oxoglutarate + O₂ = gibberellin 1 + succinate + CO₂
Other name(s): gibberellin 3β-hydroxylase; (gibberellin-20),2-oxoglutarate: oxygen oxidoreductase (3β-hydroxylating)
Systematic name: (gibberellin-20),2-oxoglutarate:oxygen oxidoreductase (3β-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate.
References: [2317]

[EC 1.14.11.15 created 1992]

EC 1.14.11.16

Accepted name: peptide-aspartate β -dioxygenase
Reaction: peptide-L-aspartate + 2-oxoglutarate + O₂ = peptide-3-hydroxy-L-aspartate + succinate + CO₂
Other name(s): aspartate β -hydroxylase; aspartylpeptide β -dioxygenase
Systematic name: peptide-L-aspartate,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺. Some vitamin K-dependent coagulation factors, as well as synthetic peptides based on the structure of the first epidermal growth factor domain of human coagulation factor IX or X, can act as acceptors.
References: [1419]

[EC 1.14.11.16 created 1992]

EC 1.14.11.17

Accepted name: taurine dioxygenase
Reaction: taurine + 2-oxoglutarate + O₂ = sulfite + aminoacetaldehyde + succinate + CO₂
Other name(s): 2-aminoethanesulfonate dioxygenase; α -ketoglutarate-dependent taurine dioxygenase
Systematic name: taurine, 2-oxoglutarate:oxygen oxidoreductase (sulfite-forming)
Comments: Requires Fe^{II}. The enzyme from *Escherichia coli* also acts on pentanesulfonate, 3-(*N*-morpholino)propanesulfonate and 2-(1,3-dioxoisindolin-2-yl)ethanesulfonate, but at lower rates.
References: [1027]

[EC 1.14.11.17 created 2000]

EC 1.14.11.18

Accepted name: phytanoyl-CoA dioxygenase
Reaction: phytanoyl-CoA + 2-oxoglutarate + O₂ = 2-hydroxyphytanoyl-CoA + succinate + CO₂
Other name(s): phytanoyl-CoA hydroxylase
Systematic name: phytanoyl-CoA, 2-oxoglutarate:oxygen oxidoreductase (2-hydroxylating)
Comments: Part of the peroxisomal phytanic acid α -oxidation pathway. Requires Fe²⁺ and ascorbate.
References: [1886, 1887, 1888, 2800, 2799]

[EC 1.14.11.18 created 2000]

[1.14.11.19] *Transferred entry. anthocyanidin synthase. Now EC 1.14.20.4, anthocyanidin synthase*

[EC 1.14.11.19 created 2001, modified 2017, deleted 2018]

EC 1.14.11.20

Accepted name: deacetoxyvindoline 4-hydroxylase
Reaction: deacetoxyvindoline + 2-oxoglutarate + O₂ = deacetylvindoline + succinate + CO₂
Other name(s): desacetoxyvindoline 4-hydroxylase; desacetoxyvindoline-17-hydroxylase; D17H; desacetoxyvindoline,2-oxoglutarate:oxygen oxidoreductase (4 β -hydroxylating)
Systematic name: deacetoxyvindoline,2-oxoglutarate:oxygen oxidoreductase (4 β -hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. Also acts on 3-hydroxy-16-methoxy-2,3-dihydrotabersonine and to a lesser extent on 16-methoxy-2,3-dihydrotabersonine.
References: [560, 561, 4426]

[EC 1.14.11.20 created 2002, modified 2005]

EC 1.14.11.21

Accepted name: clavamate synthase
Reaction: (1) deoxyamidinoproclavamate + 2-oxoglutarate + O₂ = amidinoproclavamate + succinate + CO₂
(2) proclavamate + 2-oxoglutarate + O₂ = dihydroclavamate + succinate + CO₂ + H₂O

(3) dihydroclavamate + 2-oxoglutarate + O₂ = clavamate + succinate + CO₂ + H₂O

Other name(s): clavamate synthase 2; clavamic acid synthase
Systematic name: deoxyamidinoproclavamate,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Contains nonheme iron. Catalyses three separate oxidative reactions in the pathway for the biosynthesis of the β-lactamase inhibitor clavulanate in *Streptomyces clavuligerus*. The first step (hydroxylation) is separated from the latter two (oxidative cyclization and desaturation) by the action of EC 3.5.3.22, proclavamate amidinohydrolase. The three reactions are all catalysed at the same nonheme iron site.
References: [3649, 4911, 4892, 4913, 4317]

[EC 1.14.11.21 created 2003]

[1.14.11.22 Transferred entry. flavone synthase. Now EC 1.14.20.5, flavone synthase]

[EC 1.14.11.22 created 2004, deleted 2018]

[1.14.11.23 Transferred entry. flavonol synthase. Now EC 1.14.20.6, flavonol synthase]

[EC 1.14.11.23 created 2004, deleted 2018]

EC 1.14.11.24

Accepted name: 2'-deoxymugineic-acid 2'-dioxygenase
Reaction: 2'-deoxymugineic acid + 2-oxoglutarate + O₂ = mugineic acid + succinate + CO₂
Other name(s): IDS3
Systematic name: 2'-deoxymugineic acid,2-oxoglutarate:oxygen oxidoreductase (2-hydroxylating)
Comments: Requires iron(II). It is also likely that this enzyme can catalyse the hydroxylation of 3-epihydroxy-2'-deoxymugineic acid to form 3-epihydroxymugineic acid.
References: [2986, 2177]

[EC 1.14.11.24 created 2005]

EC 1.14.11.25

Accepted name: mugineic-acid 3-dioxygenase
Reaction: (1) mugineic acid + 2-oxoglutarate + O₂ = 3-epihydroxymugineic acid + succinate + CO₂
(2) 2'-deoxymugineic acid + 2-oxoglutarate + O₂ = 3-epihydroxy-2'-deoxymugineic acid + succinate + CO₂
Other name(s): IDS2
Systematic name: mugineic acid,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires iron(II).
References: [2986, 3165]

[EC 1.14.11.25 created 2005]

EC 1.14.11.26

Accepted name: deacetoxycephalosporin-C hydroxylase
Reaction: deacetoxycephalosporin C + 2-oxoglutarate + O₂ = deacetylcephalosporin C + succinate + CO₂
Other name(s): deacetylcephalosporin C synthase; 3'-methylcephem hydroxylase; DACS; DAOC hydroxylase; deacetoxycephalosporin C hydroxylase
Systematic name: deacetoxycephalosporin-C,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires iron(II). The enzyme can also use 3-exomethylenecephalosporin C as a substrate to form deacetoxycephalosporin C, although more slowly [190]. In *Acremonium chrysogenum*, the enzyme forms part of a bifunctional protein along with EC 1.14.20.1, deacetoxycephalosporin-C synthase. It is a separate enzyme in *Streptomyces clavuligerus*.
References: [956, 190, 733, 1313, 2529, 4682, 2665]

[EC 1.14.11.26 created 2005]

EC 1.14.11.27

- Accepted name:** [histone H3]-dimethyl-L-lysine³⁶ demethylase
- Reaction:** a [histone H3]-N⁶,N⁶-dimethyl-L-lysine³⁶ + 2 2-oxoglutarate + 2 O₂ = a [histone H3]-L-lysine³⁶ + 2 succinate + 2 formaldehyde + 2 CO₂ (overall reaction)
(1a) a [histone H3]-N⁶,N⁶-dimethyl-L-lysine³⁶ + 2-oxoglutarate + O₂ = a [histone H3]-N⁶-methyl-L-lysine³⁶ + succinate + formaldehyde + CO₂
(1b) a [histone H3]-N⁶-methyl-L-lysine³⁶ + 2-oxoglutarate + O₂ = a [histone H3]-L-lysine³⁶ + succinate + formaldehyde + CO₂
- Other name(s):** KDM2A (gene name); KDM2B (gene name); JHDM1A (gene name); JHDM1B (gene name); JmjC domain-containing histone demethylase 1A; H3-K36-specific demethylase (ambiguous); histone-lysine (H3-K36) demethylase (ambiguous); histone demethylase (ambiguous); protein-6-N,6-N-dimethyl-L-lysine,2-oxoglutarate:oxygen oxidoreductase; protein-N⁶,N⁶-dimethyl-L-lysine,2-oxoglutarate:oxygen oxidoreductase; [histone-H3]-lysine-36 demethylase
- Systematic name:** [histone H3]-N⁶,N⁶-dimethyl-L-lysine³⁶,2-oxoglutarate:oxygen oxidoreductase
- Comments:** Requires iron(II). Of the seven potential methylation sites in histones H3 (K4, K9, K27, K36, K79) and H4 (K20, R3) from HeLa cells, the enzyme is specific for Lys³⁶. Lysine residues exist in three methylation states (mono-, di- and trimethylated). The enzyme preferentially demethylates the dimethyl form of Lys³⁶ (K36me₂), which is its natural substrate, to form the monomethylated and unmethylated forms of Lys³⁶. It can also demethylate monomethylated (but not the trimethylated) Lys³⁶. *cf.* EC 1.14.11.69, [histone H3]-trimethyl-L-lysine³⁶ demethylase.
- References:** [4344]

[EC 1.14.11.27 created 2006, modified 2019]

EC 1.14.11.28

- Accepted name:** proline 3-hydroxylase
- Reaction:** L-proline + 2-oxoglutarate + O₂ = *cis*-3-hydroxy-L-proline + succinate + CO₂
- Other name(s):** P-3-H
- Systematic name:** L-proline,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
- Comments:** Requires iron(II) for activity. Unlike the proline hydroxylases involved in collagen biosynthesis [EC 1.14.11.2 (procollagen-proline dioxygenase) and EC 1.14.11.7 (procollagen-proline 3-dioxygenase)], this enzyme does not require ascorbate for activity although it does increase the activity of the enzyme [2886]. The enzyme is specific for L-proline as D-proline, *trans*-4-hydroxy-L-proline, *cis*-4-hydroxy-L-proline and 3,4-dehydro-DL-proline are not substrates [2886].
- References:** [2885, 2886, 703]

[EC 1.14.11.28 created 2006]

EC 1.14.11.29

- Accepted name:** hypoxia-inducible factor-proline dioxygenase
- Reaction:** hypoxia-inducible factor-L-proline + 2-oxoglutarate + O₂ = hypoxia-inducible factor-*trans*-4-hydroxy-L-proline + succinate + CO₂
- Other name(s):** HIF hydroxylase
- Systematic name:** hypoxia-inducible factor-L-proline, 2-oxoglutarate:oxygen oxidoreductase (4-hydroxylating)
- Comments:** Contains iron, and requires ascorbate. Specifically hydroxylates a proline residue in HIF- α , the α subunit of the transcriptional regulator HIF (hypoxia-inducible factor), which targets HIF for proteasomal destruction. The requirement of oxygen for the hydroxylation reaction enables animals to respond to hypoxia.
- References:** [1868, 1854, 472, 1058, 3132, 2755]

[EC 1.14.11.29 created 2010]

EC 1.14.11.30

Accepted name: hypoxia-inducible factor-asparagine dioxygenase
Reaction: hypoxia-inducible factor-L-asparagine + 2-oxoglutarate + O₂ = hypoxia-inducible factor-(3S)-3-hydroxy-L-asparagine + succinate + CO₂
Other name(s): HIF hydroxylase
Systematic name: hypoxia-inducible factor-L-asparagine, 2-oxoglutarate:oxygen oxidoreductase (4-hydroxylating)
Comments: Contains iron, and requires ascorbate. Catalyses hydroxylation of an asparagine in the C-terminal transcriptional activation domain of HIF- α , the α subunit of the transcriptional regulator HIF (hypoxia-inducible factor), which reduces its interaction with the transcriptional coactivator protein p300. The requirement of oxygen for the hydroxylation reaction enables animals to respond to hypoxia.
References: [2618, 1634, 826, 2338, 2206, 1038]

[EC 1.14.11.30 created 2010]

EC 1.14.11.31

Accepted name: thebaine 6-*O*-demethylase
Reaction: thebaine + 2-oxoglutarate + O₂ = neopinone + formaldehyde + succinate + CO₂
Other name(s): T6ODM
Systematic name: thebaine,2-oxoglutarate:oxygen oxidoreductase (6-*O*-demethylating)
Comments: Requires Fe²⁺. Catalyses a step in morphine biosynthesis. The product neopinone spontaneously rearranges to the more stable codeinone. The enzyme also catalyses the 6-*O*-demethylation of oripavine to morphinone, with lower efficiency.
References: [1471]

[EC 1.14.11.31 created 2010]

EC 1.14.11.32

Accepted name: codeine 3-*O*-demethylase
Reaction: codeine + 2-oxoglutarate + O₂ = morphine + formaldehyde + succinate + CO₂
Other name(s): codeine *O*-demethylase; CODM
Systematic name: codeine,2-oxoglutarate:oxygen oxidoreductase (3-*O*-demethylating)
Comments: Requires Fe²⁺. Catalyses a step in morphine biosynthesis. The enzyme also catalyses the 3-*O*-demethylation of thebaine to oripavine, with lower efficiency.
References: [1471]

[EC 1.14.11.32 created 2010]

EC 1.14.11.33

Accepted name: DNA oxidative demethylase
Reaction: DNA-base-CH₃ + 2-oxoglutarate + O₂ = DNA-base + formaldehyde + succinate + CO₂
Other name(s): alkylated DNA repair protein; α -ketoglutarate-dependent dioxygenase ABH1; *alkB* (gene name)
Systematic name: methyl DNA-base, 2-oxoglutarate:oxygen oxidoreductase (formaldehyde-forming)
Comments: Contains iron; activity is slightly stimulated by ascorbate. Catalyses oxidative demethylation of the DNA base lesions *N*¹-methyladenine, *N*³-methylcytosine, *N*¹-methylguanine, and *N*³-methylthymine. It works better on single-stranded DNA (ssDNA) and is capable of repairing damaged bases in RNA.
References: [1081, 4791, 4790]

[EC 1.14.11.33 created 2011]

[1.14.11.34 *Transferred entry. 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming). Now EC 1.14.20.7, 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming)*]

[EC 1.14.11.34 created 2011, deleted 2018]

EC 1.14.11.35

Accepted name: 1-deoxypentalenic acid 11 β -hydroxylase
Reaction: 1-deoxypentalenate + 2-oxoglutarate + O₂ = 1-deoxy-11 β -hydroxypentalenate + succinate + CO₂
Other name(s): *ptlH* (gene name); *sav2991* (gene name); *pntH* (gene name)
Systematic name: 1-deoxypentalenic acid,2-oxoglutarate:oxygen oxidoreductase
Comments: The enzyme requires iron(II) and ascorbate. Isolated from the bacterium *Streptomyces avermitilis*. Part of the pathway for pentalenolactone biosynthesis.
References: [4822, 4824]

[EC 1.14.11.35 created 2012]

EC 1.14.11.36

Accepted name: pentalenolactone F synthase
Reaction: pentalenolactone D + 2 2-oxoglutarate + 2 O₂ = pentalenolactone F + 2 succinate + 2 CO₂ + H₂O (overall reaction)
(1a) pentalenolactone D + 2-oxoglutarate + O₂ = pentalenolactone E + succinate + CO₂ + H₂O
(1b) pentalenolactone E + 2-oxoglutarate + O₂ = pentalenolactone F + succinate + CO₂
Other name(s): *penD* (gene name); *pntD* (gene name); *ptlD* (gene name)
Systematic name: pentalenolactone-D,2-oxoglutarate:oxygen oxidoreductase
Comments: Requires iron(II) and ascorbate. Isolated from the bacteria *Streptomyces exfoliatus*, *Streptomyces arenae* and *Streptomyces avermitilis*. Part of the pentalenolactone biosynthesis pathway.
References: [3801]

[EC 1.14.11.36 created 2012]

EC 1.14.11.37

Accepted name: kanamycin B dioxygenase
Reaction: kanamycin B + 2-oxoglutarate + O₂ = 2'-dehydrokanamycin A + succinate + NH₃ + CO₂
Other name(s): *kanJ* (gene name)
Systematic name: kanamycin-B,2-oxoglutarate:oxygen oxidoreductase (deaminating, 2'-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. Found in the bacterium *Streptomyces kanamyceticus* where it is involved in the conversion of the aminoglycoside antibiotic kanamycin B to kanamycin A.
References: [4089]

[EC 1.14.11.37 created 2013, modified 2013]

EC 1.14.11.38

Accepted name: verruculogen synthase
Reaction: fumitremorgin B + 2-oxoglutarate + 2 O₂ + reduced acceptor = verruculogen + succinate + CO₂ + H₂O + acceptor
Other name(s): *fmtF* (gene name); FmtOx1
Systematic name: fumitremorgin B,2-oxoglutarate:oxygen oxidoreductase (verruculogen-forming)
Comments: Requires Fe²⁺ and ascorbate. Found in the fungus *Aspergillus fumigatus*. Both atoms of a dioxygen molecule are incorporated into verruculogen [4013, 2020]. Involved in the biosynthetic pathways of several indole alkaloids such as fumitremorgin A.
References: [4013, 2020]

[EC 1.14.11.38 created 2013]

EC 1.14.11.39

Accepted name: L-asparagine hydroxylase
Reaction: L-asparagine + 2-oxoglutarate + O₂ = (2S,3S)-3-hydroxyasparagine + succinate + CO₂

Other name(s): L-asparagine 3-hydroxylase; AsnO
Systematic name: L-asparagine,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires Fe²⁺. The enzyme is only able to hydroxylate free L-asparagine. It is not active toward D-asparagine. The β-hydroxylated asparagine produced is incorporated at position 9 of the calcium-dependent antibiotic (CDA), an 11-residue non-ribosomally synthesized acidic lipopeptide lactone.
References: [4066]

[EC 1.14.11.39 created 2013]

EC 1.14.11.40

Accepted name: enduracididine β-hydroxylase
Reaction: L-enduracididine + 2-oxoglutarate + O₂ = (3*S*)-3-hydroxy-L-enduracididine + succinate + CO₂
Other name(s): MppO; L-enduracididine,2-oxoglutarate:O₂ oxidoreductase (3-hydroxylating)
Systematic name: L-enduracididine,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Fe²⁺-dependent enzyme. The enzyme is involved in biosynthesis of the nonproteinogenic amino acid β-hydroxyenduracididine, a component of the mannopeptimycins (cyclic glycopeptide antibiotic), produced by *Streptomyces hygroscopicus* NRRL 30439.
References: [1486, 2605]

[EC 1.14.11.40 created 2013]

EC 1.14.11.41

Accepted name: L-arginine hydroxylase
Reaction: L-arginine + 2-oxoglutarate + O₂ = (3*S*)-3-hydroxy-L-arginine + succinate + CO₂
Other name(s): VioC (ambiguous); L-arginine,2-oxoglutarate:O₂ oxidoreductase (3-hydroxylating)
Systematic name: L-arginine,2-oxoglutarate:oxygen oxidoreductase (3-hydroxylating)
Comments: Fe²⁺-dependent enzyme. The enzyme is involved in the biosynthesis of the cyclic pentapeptide antibiotic viomycin. It differs from EC 1.14.20.7, 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming), because it does not form guanidine and (*S*)-1-pyrroline-5-carboxylate from 3-hydroxy-L-arginine.
References: [1957, 1621]

[EC 1.14.11.41 created 2013]

EC 1.14.11.42

Accepted name: tRNA^{Phe} 7-(3-amino-3-carboxypropyl)wyosine³⁷-C²-hydroxylase
Reaction: 7-(3-amino-3-carboxypropyl)wyosine³⁷ in tRNA^{Phe} + 2-oxoglutarate + O₂ = 7-(2-hydroxy-3-amino-3-carboxypropyl)wyosine³⁷ in tRNA^{Phe} + succinate + CO₂
Other name(s): TYW5; tRNA yW-synthesizing enzyme 5
Systematic name: tRNA^{Phe} 7-(3-amino-3-carboxypropyl)wyosine³⁷,2-oxoglutarate:oxygen oxidoreductase (2-hydroxylating)
Comments: Requires Fe²⁺. The enzyme is not active with wybutosine.
References: [3099, 2017]

[EC 1.14.11.42 created 2013]

EC 1.14.11.43

Accepted name: (*S*)-dichlorprop dioxygenase (2-oxoglutarate)
Reaction: (1) (*S*)-2-(4-chloro-2-methylphenoxy)propanoate + 2-oxoglutarate + O₂ = 4-chloro-2-methylphenol + pyruvate + succinate + CO₂
(2) (*S*)-(2,4-dichlorophenoxy)propanoate + 2-oxoglutarate + O₂ = 2,4-dichlorophenol + pyruvate + succinate + CO₂

Other name(s): SdpA; α -ketoglutarate-dependent (*S*)-dichlorprop dioxygenase; (*S*)-phenoxypropionate/ α -ketoglutarate-dioxygenase; 2-oxoglutarate-dependent (*S*)-dichlorprop dioxygenase; (*S*)-mecoprop dioxygenase; 2-oxoglutarate-dependent (*S*)-mecoprop dioxygenase
Systematic name: (*S*)-2-(4-chloro-2-methylphenoxy)propanoate,2-oxoglutarate:oxygen oxidoreductase (pyruvate-forming)
Comments: Fe²⁺-dependent enzyme. The enzymes from the Gram-negative bacteria *Delftia acidovorans* MC1 and *Sphingomonas herbicidovorans* MH are involved in the degradation of the (*S*)-enantiomer of the phenoxyalkanoic acid herbicides mecoprop and dichlorprop [4595, 2928].
References: [4595, 2928, 2929]

[EC 1.14.11.43 created 2013]

EC 1.14.11.44

Accepted name: (*R*)-dichlorprop dioxygenase (2-oxoglutarate)
Reaction: (1) (*R*)-2-(4-chloro-2-methylphenoxy)propanoate + 2-oxoglutarate + O₂ = 4-chloro-2-methylphenol + pyruvate + succinate + CO₂
(2) (*R*)-(2,4-dichlorophenoxy)propanoate + 2-oxoglutarate + O₂ = 2,4-dichlorophenol + pyruvate + succinate + CO₂
Other name(s): RdpA; α -ketoglutarate-dependent (*R*)-dichlorprop dioxygenase; (*R*)-phenoxypropionate/ α -ketoglutarate-dioxygenase; 2-oxoglutarate-dependent (*R*)-dichlorprop dioxygenase; (*R*)-mecoprop dioxygenase; 2-oxoglutarate-dependent (*R*)-mecoprop dioxygenase
Systematic name: (*R*)-2-(4-chloro-2-methylphenoxy)propanoate,2-oxoglutarate:oxygen oxidoreductase (pyruvate-forming)
Comments: Fe²⁺-dependent enzyme. The enzymes from the Gram-negative bacteria *Delftia acidovorans* MC1 and *Sphingomonas herbicidovorans* MH are involved in the degradation of the (*R*)-enantiomer of the phenoxyalkanoic acid herbicides mecoprop and dichlorprop [4595, 2928].
References: [4595, 2928, 2929]

[EC 1.14.11.44 created 2013]

EC 1.14.11.45

Accepted name: L-isoleucine 4-hydroxylase
Reaction: L-isoleucine + 2-oxoglutarate + O₂ = (4*S*)-4-hydroxy-L-isoleucine + succinate + CO₂
Other name(s): *ido* (gene name)
Systematic name: L-isoleucine,2-oxoglutarate:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Bacillus thuringiensis*, can also catalyse the hydroxylation of L-leucine, L-norvaline, L-norleucine, and L-*allo*-isoleucine, as well as the sulfoxidation of L-methionine, L-ethionine, *S*-methyl-L-cysteine, *S*-ethyl-L-cysteine, and *S*-allyl-L-cysteine.
References: [2182, 1636, 1637]

[EC 1.14.11.45 created 2014]

EC 1.14.11.46

Accepted name: 2-aminoethylphosphonate dioxygenase
Reaction: (2-aminoethyl)phosphonate + 2-oxoglutarate + O₂ = (2-amino-1-hydroxyethyl)phosphonate + succinate + CO₂
Other name(s): *phnY* (gene name)
Systematic name: (2-aminoethyl)phosphonate,2-oxoglutarate:oxygen oxidoreductase (1-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme, characterized from an uncultured marine bacterium, is involved in a (2-aminoethyl)phosphonate degradation pathway.
References: [2757]

[EC 1.14.11.46 created 2014]

EC 1.14.11.47

- Accepted name:** [50S ribosomal protein L16]-arginine 3-hydroxylase
Reaction: [50S ribosomal protein L16]-L-Arg⁸¹ + 2-oxoglutarate + O₂ = [50S ribosomal protein L16]-(3R)-3-hydroxy-L-Arg⁸¹ + succinate + CO₂
Other name(s): *ycfD* (gene name)
Systematic name: [50S ribosomal protein L16]-L-Arg⁸¹,2-oxoglutarate:oxygen oxidoreductase (3R-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Escherichia coli*, hydroxylates an arginine residue on the 50S ribosomal protein L16, and is involved in regulation of bacterial ribosome assembly.
References: [1294, 4413]

[EC 1.14.11.47 created 2014]

EC 1.14.11.48

- Accepted name:** xanthine dioxygenase
Reaction: xanthine + 2-oxoglutarate + O₂ = urate + succinate + CO₂
Other name(s): XanA; α -ketoglutarate-dependent xanthine hydroxylase
Systematic name: xanthine,2-oxoglutarate:oxygen oxidoreductase
Comments: Requires Fe²⁺ and L-ascorbate. The enzyme, which was characterized from fungi, is specific for xanthine.
References: [781, 2869, 2451]

[EC 1.14.11.48 created 2015]

EC 1.14.11.49

- Accepted name:** uridine-5'-phosphate dioxygenase
Reaction: UMP + 2-oxoglutarate + O₂ = 5'-dehydrouridine + succinate + CO₂ + phosphate
Other name(s): *lipL* (gene name)
Systematic name: UMP,2-oxoglutarate:oxygen oxidoreductase
Comments: The enzyme catalyses a net dephosphorylation and oxidation of UMP to generate 5'-dehydrouridine, the first intermediate in the biosynthesis of the unusual aminoribosyl moiety found in several C⁷-furanosyl nucleosides such as A-90289s, caprazamycins, liposidomycins, muraymycins and FR-900453. Requires Fe²⁺.
References: [4772, 4774]

[EC 1.14.11.49 created 2015]

[1.14.11.50] *Transferred entry. (-)-deoxypodophyllotoxin synthase. Now EC 1.14.20.8, (-)-deoxypodophyllotoxin synthase*

[EC 1.14.11.50 created 2016, deleted 2018]

EC 1.14.11.51

- Accepted name:** DNA N⁶-methyladenine demethylase
Reaction: N⁶-methyladenine in DNA + 2-oxoglutarate + O₂ = adenine in DNA + formaldehyde + succinate + CO₂
Other name(s): ALKBH1
Systematic name: DNA-N⁶-methyladenosine,2-oxoglutarate:oxygen oxidoreductase (formaldehyde-forming)
Comments: Contains iron(II). Catalyses oxidative demethylation of DNA N⁶-methyladenine, a prevalent modification in LINE-1 transposons, which are specifically enriched on the human X chromosome.
References: [4680]

[EC 1.14.11.51 created 2016]

EC 1.14.11.52

Accepted name: validamycin A dioxygenase
Reaction: validamycin A + 2-oxoglutarate + O₂ = validamycin B + succinate + CO₂
Other name(s): *vldW* (gene name)
Systematic name: validamycin-A,2-oxoglutarate:oxygen oxidoreductase (6'-hydroxylating)
Comments: The enzyme was characterized from the bacterium *Streptomyces hygrosopicus* subsp. *limoneus*. Requires Fe²⁺.
References: [71]

[EC 1.14.11.52 created 2016]

EC 1.14.11.53

Accepted name: mRNA N⁶-methyladenine demethylase
Reaction: N⁶-methyladenine in mRNA + 2-oxoglutarate + O₂ = adenine in mRNA + formaldehyde + succinate + CO₂
Other name(s): ALKBH5; FTO
Systematic name: mRNA-N⁶-methyladenosine,2-oxoglutarate:oxygen oxidoreductase (formaldehyde-forming)
Comments: Contains iron(II). Catalyses oxidative demethylation of mRNA N⁶-methyladenine. The FTO enzyme from human can also demethylate N³-methylthymine from single stranded DNA and N³-methyluridine from single stranded RNA [1914, 1502] with low activity [1913].
References: [1914, 1502, 1913, 4908, 1099, 4703, 42]

[EC 1.14.11.53 created 2016]

EC 1.14.11.54

Accepted name: mRNA N¹-methyladenine demethylase
Reaction: N¹-methyladenine in mRNA + 2-oxoglutarate + O₂ = adenine in mRNA + formaldehyde + succinate + CO₂
Other name(s): ALKBH3
Systematic name: mRNA-N¹-methyladenine,2-oxoglutarate:oxygen oxidoreductase (formaldehyde-forming)
Comments: Contains iron(II). Catalyses oxidative demethylation of mRNA N¹-methyladenine. The enzyme is also involved in alkylation repair in DNA [824].
References: [4125, 824, 2461]

[EC 1.14.11.54 created 2016]

EC 1.14.11.55

Accepted name: ectoine hydroxylase
Reaction: ectoine + 2-oxoglutarate + O₂ = 5-hydroxyectoine + succinate + CO₂
Other name(s): *ectD* (gene name); ectoine dioxygenase
Systematic name: ectoine,2-oxoglutarate:oxygen oxidoreductase (5-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme, found in bacteria, is specific for ectoine.
References: [505, 504, 3505]

[EC 1.14.11.55 created 2017]

EC 1.14.11.56

Accepted name: L-proline *cis*-4-hydroxylase
Reaction: L-proline + 2-oxoglutarate + O₂ = *cis*-4-hydroxy-L-proline + succinate + CO₂
Systematic name: L-proline,2-oxoglutarate:oxygen oxidoreductase (*cis*-4-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme, isolated from *Rhizobium* species, only produces *cis*-4-hydroxy-L-proline (*cf.* EC 1.14.11.57, L-proline *trans*-4-hydroxylase).

References: [1524]

[EC 1.14.11.56 created 2017]

EC 1.14.11.57

Accepted name: L-proline *trans*-4-hydroxylase
Reaction: L-proline + 2-oxoglutarate + O₂ = *trans*-4-hydroxy-L-proline + succinate + CO₂
Systematic name: L-proline,2-oxoglutarate:oxygen oxidoreductase (*trans*-4-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme, isolated from multiple bacterial species, only produces *trans*-4-hydroxy-L-proline (*cf.* EC 1.14.11.56, L-proline *cis*-4-hydroxylase).
References: [2369, 3852]

[EC 1.14.11.57 created 2017]

EC 1.14.11.58

Accepted name: ornithine lipid ester-linked acyl 2-hydroxylase
Reaction: an ornithine lipid + 2-oxoglutarate + O₂ = a 2-hydroxyornithine lipid + succinate + CO₂
Other name(s): *olsC* (gene name)
Systematic name: ornithine lipid,2-oxoglutarate:oxygen oxidoreductase (ester-linked acyl 2-hydroxylase)
Comments: The enzyme, characterized from the bacterium *Rhizobium tropici*, catalyses the hydroxylation of C-2 of the fatty acyl group that is ester-linked to the 3-hydroxy position of the amide-linked fatty acid.
References: [3558, 4431]

[EC 1.14.11.58 created 2018]

EC 1.14.11.59

Accepted name: 2,4-dihydroxy-1,4-benzoxazin-3-one-glucoside dioxygenase
Reaction: (2*R*)-4-hydroxy-3-oxo-3,4-dihydro-2*H*-1,4-benzoxazin-2-yl β-D-glucopyranoside + 2-oxoglutarate + O₂ = (2*R*)-4,7-dihydroxy-3-oxo-3,4-dihydro-2*H*-1,4-benzoxazin-2-yl β-D-glucopyranoside + succinate + CO₂ + H₂O
Other name(s): BX6 (gene name); DIBOA-Glc dioxygenase
Systematic name: (2*R*)-4-hydroxy-3-oxo-3,4-dihydro-2*H*-1,4-benzoxazin-2-yl β-D-glucopyranoside:oxygen oxidoreductase (7-hydroxylating)
Comments: The enzyme is involved in the biosynthesis of protective and allelopathic benzoxazinoids in some plants, most commonly from the family of Poaceae (grasses).
References: [1942]

[EC 1.14.11.59 created 2012 as EC 1.14.20.2, transferred 2018 to EC 1.14.11.59]

EC 1.14.11.60

Accepted name: scopoletin 8-hydroxylase
Reaction: scopoletin + 2-oxoglutarate + O₂ = fraxetin + succinate + CO₂
Other name(s): S8H (gene name)
Systematic name: scopoletin,2-oxoglutarate:oxygen oxidoreductase (8-hydroxylating)
Comments: Requires iron(II) and ascorbate. A protein involved in biosynthesis of iron(III)-chelating coumarins in higher plants.
References: [3921, 3439]

[EC 1.14.11.60 created 2018]

EC 1.14.11.61

Accepted name: feruloyl-CoA 6-hydroxylase

Reaction: *trans*-feruloyl-CoA + 2-oxoglutarate + O₂ = *trans*-6-hydroxyferuloyl-CoA + succinate + CO₂
Systematic name: feruloyl-CoA,2-oxoglutarate:oxygen oxidoreductase (6-hydroxylating)
Comments: Requires iron(II) and ascorbate. The product spontaneously undergoes *trans*-cis isomerization and lactonization to form scopoletin, liberating CoA in the process. The enzymes from the plants *Ruta graveolens* and *Ipomoea batatas* also act on *trans*-4-coumaroyl-CoA. *cf.* EC 1.14.11.62, *trans*-4-coumaroyl-CoA 2-hydroxylase.
References: [1980, 250, 4443, 2702]

[EC 1.14.11.61 created 2019]

EC 1.14.11.62

Accepted name: *trans*-4-coumaroyl-CoA 2-hydroxylase
Reaction: *trans*-4-coumaroyl-CoA + 2-oxoglutarate + O₂ = 2,4-dihydroxycinnamoyl-CoA + succinate + CO₂
Other name(s): Diox4 (gene name); C2'H (gene name)
Systematic name: (2*E*)-3-(4-hydroxyphenyl)prop-2-enoyl-CoA,2-oxoglutarate:oxygen oxidoreductase (2-hydroxylating)
Comments: Requires iron(II) and ascorbate. The product spontaneously undergoes *trans*-cis isomerization followed by lactonization and cyclization, liberating CoA and forming umbelliferone. The enzymes from the plants *Ruta graveolens* and *Ipomoea batatas* also act on *trans*-feruloyl-CoA (*cf.* EC 1.14.11.61, feruloyl-CoA 6-hydroxylase).
References: [4443, 2702]

[EC 1.14.11.62 created 2019]

EC 1.14.11.63

Accepted name: peptidyl-lysine (3*S*)-dioxygenase
Reaction: a [protein]-L-lysine + 2-oxoglutarate + O₂ = a [protein]-(3*S*)-3-hydroxy-L-lysine + succinate + CO₂
Other name(s): JMJD7 (gene name); Jumonji domain-containing protein 7; JmjC domain-containing protein 7
Systematic name: [protein]-L-lysine,2-oxoglutarate:oxygen oxidoreductase (3*S*-hydroxylating)
Comments: Requires iron(II). The enzyme acts on specific lysine residues in its substrates, and is stereo-specific. The enzyme encoded by the human JMJD7 gene acts specifically on two related members of the translation factor family of GTPases, DRG1 and DRG2.
References: [2652]

[EC 1.14.11.63 created 2019]

EC 1.14.11.64

Accepted name: glutarate dioxygenase
Reaction: glutarate + 2-oxoglutarate + O₂ = (*S*)-2-hydroxyglutarate + succinate + CO₂
Other name(s): *csiD* (gene name)
Systematic name: glutarate, 2-oxoglutarate:oxygen oxidoreductase ((*S*)-2-hydroxyglutarate-forming)
Comments: Requires iron(II). The enzyme, characterized from the bacteria *Escherichia coli* and *Pseudomonas putida*, participates in L-lysine degradation in many bacteria. It provides an alternative route for L-glutarate degradation that does not proceed via CoA-activated intermediates.
References: [2172, 4883]

[EC 1.14.11.64 created 2019]

EC 1.14.11.65

Accepted name: [histone H3]-dimethyl-L-lysine⁹ demethylase
Reaction: a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁹ + 2 2-oxoglutarate + 2 O₂ = a [histone H3]-L-lysine⁹ + 2 succinate + 2 formaldehyde + 2 CO₂ (overall reaction)
(1a) a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁹ + 2-oxoglutarate + O₂ = a [histone H3]-*N*⁶-methyl-L-lysine⁹ + succinate + formaldehyde + CO₂

(1b) a [histone H3]-*N*⁶-methyl-L-lysine⁹ + 2-oxoglutarate + O₂ = a [histone H3]-L-lysine⁹ + succinate + formaldehyde + CO₂

Other name(s): KDM3A (gene name); KDM3B (gene name); JMJD1A (gene name); JMJD1B (gene name); JHDM2A (gene name); JHDM2B (gene name); KDM7B (gene name); PHF8 (gene name); HR (gene name)

Systematic name: [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁹,2-oxoglutarate:oxygen oxidoreductase

Comments: Requires iron(II). This entry describes a group of enzymes that demethylate *N*-methylated Lys-9 residues in the tail of the histone protein H3 (H3K9). This lysine residue can exist in three methylation states (mono-, di- and trimethylated), but this group of enzymes only act on the the di- and mono-methylated forms. The enzymes are dioxygenases and act by hydroxylating the methyl group, forming an unstable hemiaminal that leaves as formaldehyde. *cf.* EC 1.14.11.66, [histone H3]-trimethyl-L-lysine⁹ demethylase.

References: [4748, 2533, 1101, 2304, 2518]

[EC 1.14.11.65 created 2019]

EC 1.14.11.66

Accepted name: [histone H3]-trimethyl-L-lysine⁹ demethylase

Reaction: a [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁹ + 2 2-oxoglutarate + 2 O₂ = a [histone H3]-*N*⁶-methyl-L-lysine⁹ + 2 succinate + 2 formaldehyde + 2 CO₂ (overall reaction)

(1a) a [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁹ + 2-oxoglutarate + O₂ = a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁹ + succinate + formaldehyde + CO₂

(1b) a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁹ + 2-oxoglutarate + O₂ = a [histone H3]-*N*⁶-methyl-L-lysine⁹ + succinate + formaldehyde + CO₂

Other name(s): KDM4A (gene name); KDM4B (gene name); KDM4C (gene name); KDM4D (gene name); JHDM3A (gene name); JMJD2 (gene name); JMJD2A (gene name); GASC1 (gene name)

Systematic name: [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁹,2-oxoglutarate:oxygen oxidoreductase

Comments: Requires iron(II). This entry describes a group of enzymes that demethylate *N*-methylated Lys-9 residues in the tail of the histone protein H3 (H3K9). This lysine residue can exist in three methylation states (mono-, di- and trimethylated), but this group of enzymes only act on the the tri- and di-methylated forms. The enzymes are dioxygenases and act by hydroxylating the methyl group, forming an unstable hemiaminal that leaves as formaldehyde. *cf.* EC 1.14.11.65, [histone H3]-dimethyl-L-lysine⁹ demethylase.

References: [707, 1133, 2158, 4600]

[EC 1.14.11.66 created 2019]

EC 1.14.11.67

Accepted name: [histone H3]-trimethyl-L-lysine⁴ demethylase

Reaction: a [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁴ + 3 2-oxoglutarate + 3 O₂ = a [histone H3]-L-lysine⁴ + 3 succinate + 3 formaldehyde + 3 CO₂ (overall reaction)

(1a) a [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁴ + 2-oxoglutarate + O₂ = a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ + succinate + formaldehyde + CO₂

(1b) a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ + 2-oxoglutarate + O₂ = a [histone H3]-*N*⁶-methyl-L-lysine⁴ + succinate + formaldehyde + CO₂

(1c) a [histone H3]-*N*⁶-methyl-L-lysine⁴ + 2-oxoglutarate + O₂ = a [histone H3]-L-lysine⁴ + succinate + formaldehyde + CO₂

Other name(s): KDM5A (gene name); KDM5B (gene name); KDM5C (gene name); KDM5D (gene name); JARID1A (gene name)

Systematic name: [histone H3]-*N*⁶,*N*⁶,*N*⁶-trimethyl-L-lysine⁴,2-oxoglutarate:oxygen oxidoreductase

Comments: Requires iron(II). This entry describes a group of enzymes that demethylate *N*-methylated L-lysine residues at position 4 of histone H3 (H3K4). The enzymes are dioxygenases and act by hydroxylating the methyl group, forming an unstable hemiaminal that leaves as formaldehyde. They can act on tri-, di-, and mono-methylated forms.

References: [3812, 2159, 1861, 684]

[EC 1.14.11.67 created 2019]

EC 1.14.11.68

Accepted name: [histone H3]-trimethyl-L-lysine²⁷ demethylase
Reaction: a [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine²⁷ + 2 2-oxoglutarate + 2 O₂ = a [histone H3]-N⁶-methyl-L-lysine²⁷ + 2 succinate + 2 formaldehyde + 2 CO₂ (overall reaction)
(1a) a [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine²⁷ + 2-oxoglutarate + O₂ = a [histone H3]-N⁶,N⁶-dimethyl-L-lysine²⁷ + succinate + formaldehyde + CO₂
(1b) a [histone H3]-N⁶,N⁶-dimethyl-L-lysine²⁷ + 2-oxoglutarate + O₂ = a [histone H3]-N⁶-methyl-L-lysine²⁷ + succinate + formaldehyde + CO₂
Other name(s): KDM6A (gene name); KDM6C (gene name); UTX (gene name); UTY (gene name); JMJD3 (gene name)
Systematic name: [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine²⁷,2-oxoglutarate:oxygen oxidoreductase
Comments: Requires iron(II). This entry describes a group of enzymes that demethylate *N*-methylated L-lysine residues at position 27 of histone H3 (H3K27). The enzymes are dioxygenases and act by hydroxylating the methyl group, forming an unstable hemiaminal that leaves as formaldehyde. They can act on tri- and di-methylated forms, but have no activity with the mono-methylated form.
References: [3659, 1707, 2336, 2389, 4695]

[EC 1.14.11.68 created 2019]

EC 1.14.11.69

Accepted name: [histone H3]-trimethyl-L-lysine³⁶ demethylase
Reaction: a [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine³⁶ + 2 2-oxoglutarate + 2 O₂ = a [histone H3]-N⁶-methyl-L-lysine³⁶ + 2 succinate + 2 formaldehyde + 2 CO₂ (overall reaction)
(1a) a [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine³⁶ + 2-oxoglutarate + O₂ = a [histone H3]-N⁶,N⁶-dimethyl-L-lysine³⁶ + succinate + formaldehyde + CO₂
(1b) a [histone H3]-N⁶,N⁶-dimethyl-L-lysine³⁶ + 2-oxoglutarate + O₂ = a [histone H3]-N⁶-methyl-L-lysine³⁶ + succinate + formaldehyde + CO₂
Other name(s): KDM4A (gene name); KDM4B (gene name); RPH1 (gene name); JHDM3A (gene name); JHDM3B (gene name); JMJD2A (gene name); JMJD2B (gene name)
Systematic name: [histone H3]-N⁶,N⁶,N⁶-trimethyl-L-lysine³⁶,2-oxoglutarate:oxygen oxidoreductase
Comments: Requires iron(II). This entry describes a group of enzymes that demethylate *N*-methylated Lys³⁶ residues in the tail of the histone protein H3 (H3K36). This lysine residue can exist in three methylation states (mono-, *di*- and trimethylated), but this group of enzymes only act on the tri- and dimethylated forms. The enzymes are dioxygenases and act by hydroxylating the methyl group, forming an unstable hemiaminal that leaves as formaldehyde. Since trimethylation of H3K36 enhances transcription, this enzyme acts as a transcription repressor. The enzymes that possess this activity often also catalyse the activity of EC 1.14.11.66, [histone H3]-trimethyl-L-lysine⁹ demethylase. *cf.* EC 1.14.11.27, [histone H3]-dimethyl-L-lysine³⁶ demethylase.
References: [4600, 2158, 2108, 753, 2486, 716]

[EC 1.14.11.69 created 2019]

EC 1.14.11.70

Accepted name: 7-deoxycylindrospermopsin hydroxylase
Reaction: (1) 7-deoxycylindrospermopsin + 2-oxoglutarate + O₂ = cylindrospermopsin + succinate + CO₂
(2) 7-deoxycylindrospermopsin + 2-oxoglutarate + O₂ = 7-*epi*-cylindrospermopsin + succinate + CO₂
Other name(s): *cyrI* (gene name)
Systematic name: 7-deoxycylindrospermopsin,2-oxoglutarate:oxygen oxidoreductase (7-hydroxylating)

Comments: Requires iron(II). The enzyme, found in some cyanobacterial species, catalyses the last step in the biosynthesis of the toxins cylindrospermopsin and 7-*epi*-cylindrospermopsin. The ratio of the two products differs among different strains.

References: [2735, 2736]

[EC 1.14.11.70 created 2019]

EC 1.14.11.71

Accepted name: methylphosphonate hydroxylase

Reaction: methylphosphonate + 2-oxoglutarate + O₂ = hydroxymethylphosphonate + succinate + CO₂

Other name(s): *phnY** (gene name)

Systematic name: methylphosphonate,2-oxoglutarate:oxygen oxidoreductase (1-hydroxylating)

Comments: Requires iron(II). The enzyme, characterized from the marine bacterium *Gimesia maris*, participates in a methylphosphonate degradation pathway.

References: [1265]

[EC 1.14.11.71 created 2019]

EC 1.14.11.72

Accepted name: [2-(trimethylamino)ethyl]phosphonate dioxygenase

Reaction: [2-(trimethylamino)ethyl]phosphonate + 2-oxoglutarate + O₂ = [(1*R*)-1-hydroxy-2-(trimethylamino)ethyl]phosphonate + succinate + CO₂

Other name(s): *tmpA* (gene name)

Systematic name: [2-(trimethylamino)ethyl]phosphonate,2-oxoglutarate:oxygen oxidoreductase (1*R*-hydroxylating)

Comments: Requires Fe²⁺ and ascorbate. The enzyme, found in bacteria, participates in a degradation pathway for [2-(trimethylamino)ethyl]phosphonate.

References: [3437]

[EC 1.14.11.72 created 2020]

EC 1.14.11.73

Accepted name: [protein]-arginine 3-hydroxylase

Reaction: [protein]-L-arginine + 2-oxoglutarate + O₂ = [protein]-(3*R*)-3-hydroxy-L-arginine + succinate + CO₂

Other name(s): JMJD5 (gene name)

Systematic name: [protein]-L-arginine,2-oxoglutarate:oxygen oxidoreductase (3*R*-hydroxylating)

Comments: The enzyme, characterized from humans, catalyses the stereoselective formation of the (2*S*,3*R*)-hydroxy-L-arginine stereoisomer. So far the enzyme has been shown to act on two substrates - the 40S ribosomal protein S6 (RPS6), which is hydroxylated at R137, and, at a lower activity, RCCD1, a protein involved in chromatin stability, which is hydroxylated at R141. Even though the same stereoisomer is produced by the bacterial EC 1.14.11.47, [50S ribosomal protein L16]-arginine 3-hydroxylase, the two enzymes do not exhibit any cross-reactivity on their respective ribosomal protein substrates.

References: [4628]

[EC 1.14.11.73 created 2020]

EC 1.14.11.74

Accepted name: L-isoleucine 3¹-dioxygenase

Reaction: L-isoleucine + 2-oxoglutarate + O₂ = 3¹-hydroxy-L-isoleucine + succinate + CO₂

Other name(s): *hila* (gene name); L-isoleucine 4'-dioxygenase (incorrect)

Systematic name: L-isoleucine,2-oxoglutarate:oxygen oxidoreductase (3¹-hydroxylating)

Comments: Requires Fe²⁺ and ascorbate. The enzyme has been characterized from the bacterium *Pantoea ananatis*.

References: [3933]

[EC 1.14.11.74 created 2020]

EC 1.14.11.75

Accepted name: 3¹-hydroxy-L-isoleucine 4-dioxygenase
Reaction: 3¹-hydroxy-L-isoleucine + 2-oxoglutarate + O₂ = (4*S*)-3¹,4-dihydroxy-L-isoleucine + succinate + CO₂
Other name(s): *hilB* (gene name); 4'-hydroxy-L-isoleucine 4-dioxygenase (incorrect)
Systematic name: 3¹-hydroxy-L-isoleucine,2-oxoglutarate:oxygen oxidoreductase (4*S*-hydroxylating)
Comments: Requires Fe²⁺ and ascorbate. The enzyme has been characterized from the bacterium *Pantoea ananatis*.
References: [3933]

[EC 1.14.11.75 created 2020]

EC 1.14.11.76

Accepted name: L-glutamate 3(*R*)-hydroxylase
Reaction: L-glutamate + 2-oxoglutarate + O₂ = (3*R*)-3-hydroxy-L-glutamate + succinate + CO₂
Other name(s): *iboH* (gene name)
Systematic name: L-glutamate,2-oxoglutarate:oxygen oxidoreductase (3*R*-hydroxylating)
Comments: Requires Fe²⁺ and L-ascorbate. The enzyme, characterized from the basidiomycete mushroom *Amanita muscaria*, participates in the biosynthesis of the psychoactive compounds ibotenate and muscimol.
References: [3125]

[EC 1.14.11.76 created 2020]

EC 1.14.11.77

Accepted name: alkyl sulfatase
Reaction: a primary alkyl sulfate ester + 2-oxoglutarate + O₂ = an aldehyde + succinate + CO₂ + sulfate
Other name(s): *atsK* (gene name); α-ketoglutarate-dependent sulfate ester dioxygenase; 2-oxoglutarate-dependent sulfate ester dioxygenase; type II alkyl sulfatase
Systematic name: primary alkyl sulfate ester, 2-oxoglutarate:oxygen oxidoreductase (sulfate-hydrolyzing)
Comments: Sulfatase enzymes are classified as type I, in which the key catalytic residue is 3-oxo-L-alanine, type II, which are non-heme iron-dependent dioxygenases, or type III, whose catalytic domain adopts a metallo-β-lactamase fold and binds two zinc ions as cofactors. The type II sulfatasases oxidize the C-H bond of the carbon next to the sulfate ester, using 2-oxoglutarate and oxygen as substrates. The resulting hemiacetal sulfate ester collapses, liberating inorganic sulfate and an alkyl aldehyde along with carbon dioxide and succinate. The enzymes often desulfate a broad spectrum of linear and branched-chain sulfate esters. The enzyme from *Pseudomonas putida* acts on a range of medium-chain alkyl sulfate esters, with chain lengths ranging from C₄ to C₁₂. *cf.* sulfatase EC 3.1.6.1, arylsulfatase (type I), EC 3.1.6.21, linear primary-alkylsulfatase, and EC 3.1.6.22, branched primary-alkylsulfatase.
References: [1979, 2923, 3950]

[EC 1.14.11.77 created 2021]

EC 1.14.11.78

Accepted name: (*R*)-3-[(carboxymethyl)amino]fatty acid dioxygenase/decarboxylase
Reaction: a (3*R*)-3-[(carboxymethyl)amino]fatty acid + 2 2-oxoglutarate + 2 O₂ = a (3*R*)-3-isocyanyl-fatty acid + 2 succinate + 3 CO₂ + 2 H₂O (overall reaction)
(1a) a (3*R*)-3-[(carboxymethyl)amino]fatty acid + 2-oxoglutarate + O₂ = a (3*R*)-3-[carboxy(hydroxy)methyl]aminofatty acid + succinate + CO₂

(1b) a (3*R*)-3-[carboxy(hydroxy)methyl]aminofatty acid + 2-oxoglutarate + O₂ = a (3*R*)-3-isocyanyl-fatty acid + succinate + 2 CO₂ + 2 H₂O

- Other name(s):** *scoE* (gene name); *mmaE* (gene name); Rv0097 (locus name)
Systematic name: (3*R*)-3-[(carboxylmethyl)amino]fatty acid,2-oxoglutarate:oxygen oxidoreductase (isonitrile-forming)
Comments: Requires Fe(II). The enzyme, found in actinobacterial species, participates in the biosynthesis of isonitrile-containing lipopeptides. The reaction comprises two catalytic cycles, each consuming an oxygen molecule and a 2-oxoglutarate molecule. In the first cycle the substrate is hydroxylated, while in the second cycle the enzyme catalyses a decarboxylation/oxidation reaction that produces an isonitrile group.
References: [1537, 1536, 1948]

[EC 1.14.11.78 created 2022]

EC 1.14.11.79

- Accepted name:** protein-L-histidine (3*S*)-3-hydroxylase
Reaction: a [protein]-L-histidine + 2-oxoglutarate + O₂ = a [protein]-(3*S*)-3-hydroxy-L-histidine + succinate + CO₂
Other name(s): RIOX1 (gene name); RIOX2 (gene name); protein histidyl hydroxylase
Systematic name: protein-L-histidine,2-oxoglutarate:oxygen oxidoreductase (3*S*-hydroxylating)
Comments: The human enzymes encoded by the RIOX1 and RIOX2 genes catalyse the hydroxylation of L-histidine residues in the 60S ribosomal proteins Rpl8 and L27a, respectively. Both proteins contain JmjC and winged helix domains, and both also catalyse histone L-lysine demethylation activities.
References: [1294, 492]

[EC 1.14.11.79 created 2022]

EC 1.14.11.80

- Accepted name:** methylcytosine dioxygenase
Reaction:
(1) 5-methylcytosine in DNA + 2-oxoglutarate + O₂ = 5-hydroxymethylcytosine in DNA + succinate + CO₂
(2) 5-hydroxymethylcytosine in DNA + 2-oxoglutarate + O₂ = 5-formylcytosine in DNA + succinate + CO₂ + H₂O
(3) 5-formylcytosine in DNA + 2-oxoglutarate + O₂ = 5-carboxycytosine in DNA + succinate + CO₂
Other name(s): TET1 (gene name); TET2 (gene name); TET3 (gene name)
Systematic name: 5-methylcytosine in DNA,2-oxoglutarate:oxygen oxidoreductase
Comments: The TET proteins mediate iterative oxidation of 5-methylcytosine in DNA (5mc) to 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC), and 5-carboxylcytosine (5caC). 5fC and 5caC are recognized by EC 3.2.2.29, thymine-DNA glycosylase (TDG), which excises them, leaving an apyrimidinic site. Coupled with the base excision repair (BER) pathway, these activities result in a cytosine demethylation pathway.
References: [1848, 1849, 1593, 2623, 4880]

[EC 1.14.11.80 created 2022]

EC 1.14.11.81

- Accepted name:** (–)-cyclopenine synthase
Reaction:
(1) cycloheptene + 2-oxoglutarate + O₂ = dehydrocycloheptene + succinate + CO₂ + H₂O
(2) dehydrocycloheptene + 2-oxoglutarate + O₂ = (–)-cyclopenine + succinate + CO₂
Other name(s): *asqJ* (gene name)
Systematic name: cycloheptene,2-oxoglutarate:oxygen oxidoreductase ((–)-cyclopenine-forming)
Comments: This fungal enzyme is involved in the biosynthesis of quinolone compounds. It catalyses two oxidation reactions: the first reaction results in a desaturation; the second reaction is a monooxygenation of the double bond, forming an epoxide. The enzyme is also active with 4'-methoxycycloheptene.

References: [3110, 1831, 426, 600, 3963, 2472, 2599, 4652, 2447, 4200]

[EC 1.14.11.81 created 2022]

EC 1.14.11.82

Accepted name: 5-dehydro-6-demethoxyfumagillol dioxygenase
Reaction: 5-dehydro-6-demethoxyfumagillol + 2-oxoglutarate + O₂ = 5-dehydro-6-demethoxy-6-hydroxyfumagillol + succinate + CO₂
Other name(s): *fmaF* (gene name); Fma-C6H
Systematic name: 5-dehydro-6-demethoxyfumagillol,2-oxoglutarate:oxygen oxidoreductase (6-hydroxylating)
Comments: Requires iron(II). The enzyme, characterized from the mold *Aspergillus fumigatus*, participates in the biosynthesis of the meroterpenoid fumagillin.
References: [2489]

[EC 1.14.11.82 created 2022]

EC 1.14.12 With NADH or NADPH as one donor, and incorporation of two atoms of oxygen into the other donor

EC 1.14.12.1

Accepted name: anthranilate 1,2-dioxygenase (deaminating, decarboxylating)
Reaction: anthranilate + NAD(P)H + 2 H⁺ + O₂ = catechol + CO₂ + NAD(P)⁺ + NH₃
Other name(s): anthranilate hydroxylase; anthranilic hydroxylase; anthranilic acid hydroxylase
Systematic name: anthranilate,NAD(P)H:oxygen oxidoreductase (1,2-hydroxylating, deaminating, decarboxylating)
Comments: Requires Fe²⁺.
References: [2176, 4208]

[EC 1.14.12.1 created 1972]

[1.14.12.2 Transferred entry. now EC 1.14.13.35 anthranilate 3-monooxygenase (deaminating)]

[EC 1.14.12.2 created 1972, deleted 1990]

EC 1.14.12.3

Accepted name: benzene 1,2-dioxygenase
Reaction: benzene + NADH + H⁺ + O₂ = *cis*-cyclohexa-3,5-diene-1,2-diol + NAD⁺
Other name(s): benzene hydroxylase; benzene dioxygenase
Systematic name: benzene,NADH:oxygen oxidoreductase (1,2-hydroxylating)
Comments: A system, containing a reductase which is an iron-sulfur flavoprotein (FAD), an iron-sulfur oxygenase and ferredoxin. Requires Fe²⁺.
References: [1321]

[EC 1.14.12.3 created 1972]

[1.14.12.4 Transferred entry. 3-hydroxy-2-methylpyridinecarboxylate dioxygenase. Now EC 1.14.13.242, 3-hydroxy-2-methylpyridinecarboxylate monooxygenase]

[EC 1.14.12.4 created 1972, deleted 2018]

[1.14.12.5 Transferred entry. 5-pyridoxate dioxygenase. Now EC 1.14.13.241, 5-pyridoxate monooxygenase]

[EC 1.14.12.5 created 1972, deleted 2018]

[1.14.12.6 Transferred entry. 2-hydroxycyclohexanone 2-monooxygenase. Now EC 1.14.13.66, 2-hydroxycyclohexanone 2-monooxygenase]

[EC 1.14.12.6 created 1978, deleted 1999]

EC 1.14.12.7

Accepted name: phthalate 4,5-dioxygenase
Reaction: phthalate + NADH + H⁺ + O₂ = *cis*-4,5-dihydroxycyclohexa-1(6),2-diene-1,2-dicarboxylate + NAD⁺
Other name(s): PDO ; phthalate dioxygenase
Systematic name: phthalate,NADH:oxygen oxidoreductase (4,5-hydroxylating)
Comments: A system, containing a reductase which is an iron-sulfur flavoprotein (FMN), an iron-sulfur oxygenase, and no independent ferredoxin. Requires Fe²⁺.
References: [237]

[EC 1.14.12.7 created 1990]

EC 1.14.12.8

Accepted name: 4-sulfobenzoate 3,4-dioxygenase
Reaction: 4-sulfobenzoate + NADH + H⁺ + O₂ = 3,4-dihydroxybenzoate + sulfite + NAD⁺
Other name(s): 4-sulfobenzoate dioxygenase; 4-sulfobenzoate 3,4-dioxygenase system
Systematic name: 4-sulfobenzoate,NADH:oxygen oxidoreductase (3,4-hydroxylating, sulfite-forming)
Comments: A system, containing a reductase which is an iron-sulfur flavoprotein (FMN), an iron-sulfur oxygenase, and no independent ferredoxin. Requires Fe²⁺.
References: [2530]

[EC 1.14.12.8 created 1992]

EC 1.14.12.9

Accepted name: 4-chlorophenylacetate 3,4-dioxygenase
Reaction: 4-chlorophenylacetate + NADH + H⁺ + O₂ = 3,4-dihydroxyphenylacetate + chloride + NAD⁺
Systematic name: 4-chlorophenylacetate,NADH:oxygen oxidoreductase (3,4-hydroxylating, dechlorinating)
Comments: A system, containing a reductase and an iron-sulfur oxygenase, and no independent ferredoxin. Requires Fe²⁺. Also acts on 4-bromophenyl acetate.
References: [2655]

[EC 1.14.12.9 created 1989 as EC 1.13.99.4, transferred 1992 to EC 1.14.12.9]

EC 1.14.12.10

Accepted name: benzoate 1,2-dioxygenase
Reaction: benzoate + NADH + H⁺ + O₂ = (1*R*,6*S*)-1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate + NAD⁺
Other name(s): benzoate hydroxylase; benzoate hydroxylase; benzoic hydroxylase; benzoate dioxygenase; benzoate,NADH:oxygen oxidoreductase (1,2-hydroxylating, decarboxylating) [incorrect]
Systematic name: benzoate,NADH:oxygen oxidoreductase (1,2-hydroxylating)
Comments: A system, containing a reductase which is an iron-sulfur flavoprotein (FAD), and an iron-sulfur oxygenase. Requires Fe²⁺.
References: [4730, 4731, 4732]

[EC 1.14.12.10 created 1972 as EC 1.13.99.2, transferred 1992 to EC 1.14.12.10]

EC 1.14.12.11

Accepted name: toluene dioxygenase
Reaction: toluene + NADH + H⁺ + O₂ = (1*S*,2*R*)-3-methylcyclohexa-3,5-diene-1,2-diol + NAD⁺
Other name(s): toluene 2,3-dioxygenase
Systematic name: toluene,NADH:oxygen oxidoreductase (1,2-hydroxylating)

Comments: A system, containing a reductase which is an iron-sulfur flavoprotein (FAD), an iron-sulfur oxygenase, and a ferredoxin. Some other aromatic compounds, including ethylbenzene, 4-xylene and some halogenated toluenes, are converted into the corresponding *cis*-dihydrodiols.

References: [3499, 4087]

[EC 1.14.12.11 created 1992]

EC 1.14.12.12

Accepted name: naphthalene 1,2-dioxygenase
Reaction: naphthalene + NADH + H⁺ + O₂ = (1*R*,2*S*)-1,2-dihydronaphthalene-1,2-diol + NAD⁺
Other name(s): naphthalene dioxygenase; naphthalene oxygenase; NDO
Systematic name: naphthalene,NADH:oxygen oxidoreductase (1,2-hydroxylating)
Comments: This enzyme is a member of the ring-hydroxylating dioxygenase (RHD) family of bacterial enzymes that play a critical role in the degradation of aromatic compounds, such as polycyclic aromatic hydrocarbons [1955]. This enzyme comprises a multicomponent system, containing a reductase that is an iron-sulfur flavoprotein (FAD; EC 1.18.1.3, ferredoxin—NAD⁺ reductase), an iron-sulfur oxygenase, and ferredoxin. Requires Fe²⁺.
References: [1056, 1897, 2038, 3236, 1955]

[EC 1.14.12.12 created 1992]

EC 1.14.12.13

Accepted name: 2-halobenzoate 1,2-dioxygenase
Reaction: a 2-halobenzoate + NADH + H⁺ + O₂ = catechol + a halide anion + NAD⁺ + CO₂
Other name(s): 2-chlorobenzoate 1,2-dioxygenase
Systematic name: 2-halobenzoate,NADH:oxygen oxidoreductase (1,2-hydroxylating, dehalogenating, decarboxylating)
Comments: A multicomponent enzyme system composed of a dioxygenase component and an electron transfer component. The latter contains FAD. The enzyme, characterized from the bacterium *Burkholderia cepacia* 2CBS, has a broad substrate specificity. Substrates include 2-fluorobenzoate, 2-chlorobenzoate, 2-bromobenzoate, and 2-iodobenzoate, which are processed in this order of preference.
References: [1112, 1113, 1460]

[EC 1.14.12.13 created 1992, modified 2012]

EC 1.14.12.14

Accepted name: 2-aminobenzenesulfonate 2,3-dioxygenase
Reaction: 2-aminobenzenesulfonate + NADH + H⁺ + O₂ = 2,3-dihydroxybenzenesulfonate + NH₃ + NAD⁺
Other name(s): 2-aminosulfobenzene 2,3-dioxygenase
Systematic name: 2-aminobenzenesulfonate,NADH:oxygen oxidoreductase (2,3-hydroxylating, ammonia-forming)
References: [1962, 1964]

[EC 1.14.12.14 created 1999]

EC 1.14.12.15

Accepted name: terephthalate 1,2-dioxygenase
Reaction: terephthalate + NADH + H⁺ + O₂ = (1*R*,6*S*)-dihydroxycyclohexa-2,4-diene-1,4-dicarboxylate + NAD⁺
Other name(s): benzene-1,4-dicarboxylate 1,2-dioxygenase; 1,4-dicarboxybenzoate 1,2-dioxygenase
Systematic name: benzene-1,4-dicarboxylate,NADH:oxygen oxidoreductase (1,2-hydroxylating)
Comments: Has been shown to contain a Rieske [2Fe-2S] cluster
References: [3720]

[EC 1.14.12.15 created 1999]

EC 1.14.12.16

- Accepted name:** 2-hydroxyquinoline 5,6-dioxygenase
Reaction: quinolin-2-ol + NADH + H⁺ + O₂ = 2,5,6-trihydroxy-5,6-dihydroquinoline + NAD⁺
Other name(s): 2-oxo-1,2-dihydroquinoline 5,6-dioxygenase; quinolin-2-ol 5,6-dioxygenase; quinolin-2(1*H*)-one 5,6-dioxygenase
Systematic name: quinolin-2-ol,NADH:oxygen oxidoreductase (5,6-hydroxylating)
Comments: 3-Methylquinolin-2-ol, quinolin-8-ol and quinolin-2,8-diol are also substrates. Quinolin-2-ols exist largely as their quinolin-2(1*H*)-one tautomers
References: [3697]

[EC 1.14.12.16 created 1999]

EC 1.14.12.17

- Accepted name:** nitric oxide dioxygenase
Reaction: 2 nitric oxide + 2 O₂ + NAD(P)H = 2 nitrate + NAD(P)⁺ + H⁺
Systematic name: nitric oxide,NAD(P)H:oxygen oxidoreductase
Comments: A flavohemoglobin (FAD). It has been proposed that FAD functions as the electron carrier from NADPH to the ferric heme prosthetic group.
References: [1273, 1274]

[EC 1.14.12.17 created 2000]

EC 1.14.12.18

- Accepted name:** biphenyl 2,3-dioxygenase
Reaction: biphenyl + NADH + H⁺ + O₂ = (1*S*,2*R*)-3-phenylcyclohexa-3,5-diene-1,2-diol + NAD⁺
Other name(s): biphenyl dioxygenase
Systematic name: biphenyl,NADH:oxygen oxidoreductase (2,3-hydroxylating)
Comments: Requires Fe²⁺. The enzyme from *Burkholderia fungorum* LB400 (previously *Pseudomonas* sp.) is part of a multicomponent system composed of an NADH:ferredoxin oxidoreductase (FAD cofactor), a [2Fe-2S] Rieske-type ferredoxin, and a terminal oxygenase that contains a [2Fe-2S] Rieske-type iron-sulfur cluster and a catalytic mononuclear nonheme iron centre. Chlorine-substituted biphenyls can also act as substrates. Similar to the three-component enzyme systems EC 1.14.12.3 (benzene 1,2-dioxygenase) and EC 1.14.12.11 (toluene dioxygenase).
References: [1466, 1467, 448]

[EC 1.14.12.18 created 2001]

EC 1.14.12.19

- Accepted name:** 3-phenylpropanoate dioxygenase
Reaction: (1) 3-phenylpropanoate + NADH + H⁺ + O₂ = 3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate + NAD⁺
(2) (2*E*)-3-phenylprop-2-enoate + NADH + H⁺ + O₂ = (2*E*)-3-(2,3-dihydroxyphenyl)prop-2-enoate + NAD⁺
Other name(s): HcaA1A2CD; Hca dioxygenase; 3-phenylpropionate dioxygenase
Systematic name: 3-phenylpropanoate,NADH:oxygen oxidoreductase (2,3-hydroxylating)
Comments: This enzyme catalyses a step in the pathway of phenylpropanoid compounds degradation. It catalyses the insertion of both atoms of molecular oxygen into positions 2 and 3 of the phenyl ring of 3-phenylpropanoate or (2*E*)-3-phenylprop-2-enoate.
References: [900, 497]

[EC 1.14.12.19 created 2005, modified 2011]

[1.14.12.20 Transferred entry. pheophorbide a oxygenase. Now classified as EC 1.14.15.17, pheophorbide a oxygenase.]

[EC 1.14.12.20 created 2007, deleted 2016]

[1.14.12.21 Transferred entry. benzoyl-CoA 2,3-dioxygenase. Now EC 1.14.13.208, benzoyl-CoA 2,3-epoxidase]

[EC 1.14.12.21 created 2010, deleted 2015]

EC 1.14.12.22

- Accepted name:** carbazole 1,9a-dioxygenase
Reaction: 9*H*-carbazole + NAD(P)H + H⁺ + O₂ = 2'-aminobiphenyl-2,3-diol + NAD(P)⁺
Other name(s): CARDO
Systematic name: 9*H*-carbazole,NAD(P)H:oxygen oxidoreductase (2,3-hydroxylating)
Comments: This enzyme catalyses the first reaction in the pathway of carbazole degradation. The enzyme attacks at the 1 and 9a positions of carbazole, resulting in the formation of a highly unstable hemiaminal intermediate that undergoes a spontaneous cleavage and rearomatization, resulting in 2'-aminobiphenyl-2,3-diol. In most bacteria the enzyme is a complex composed of a terminal oxygenase, a ferredoxin, and a ferredoxin reductase. The terminal oxygenase component contains a nonheme iron centre and a Rieske [2Fe-2S] iron-sulfur cluster.
References: [3004, 1254]

[EC 1.14.12.22 created 2010]

EC 1.14.12.23

- Accepted name:** nitroarene dioxygenase
Reaction: nitrobenzene + NADH + O₂ = catechol + nitrite + NAD⁺
Other name(s): *cnbA* (gene name)
Systematic name: nitrobenzene,NADH:oxygen oxidoreductase (1,2-hydroxylating, nitrite-releasing)
Comments: This enzyme is a member of the naphthalene family of bacterial Rieske non-heme iron dioxygenases. It comprises a multicomponent system, containing a Rieske [2Fe-2S] ferredoxin, an NADH-dependent flavoprotein reductase (EC 1.18.1.3, ferredoxin—NAD⁺ reductase), and an α3β3 oxygenase. The enzyme forms a *cis*-dihydroxylated product that spontaneously rearranges to form a catechol with accompanying release of nitrite. It can typically act on many different nitroaromatic compounds, including chlorinated species. Enzymes found in different strains may have different substrate preferences. Requires Fe²⁺.
References: [3235, 2423, 2513, 3908]

[EC 1.14.12.23 created 2015]

EC 1.14.12.24

- Accepted name:** 2,4-dinitrotoluene dioxygenase
Reaction: 2,4-dinitrotoluene + NADH + O₂ = 4-methyl-5-nitrocatechol + nitrite + NAD⁺
Other name(s): *dntA* (gene name)
Systematic name: 2,4-dinitrotoluene,NADH:oxygen oxidoreductase (4,5-hydroxylating, nitrite-releasing)
Comments: This enzyme, characterized from the bacterium *Burkholderia* sp. strain DNT, is a member of the naphthalene family of bacterial Rieske non-heme iron dioxygenases. It comprises a multicomponent system, containing a Rieske [2Fe-2S] ferredoxin, an NADH-dependent flavoprotein reductase (EC 1.18.1.3, ferredoxin—NAD⁺ reductase), and an α3β3 oxygenase. The enzyme forms a *cis*-dihydroxylated product that spontaneously rearranges to form a catechol with accompanying release of nitrite. It does not act on nitrobenzene. *cf.* EC 1.14.12.23, nitroarene dioxygenase.
References: [4091]

[EC 1.14.12.24 created 2015]

EC 1.14.12.25

- Accepted name:** *p*-cumate 2,3-dioxygenase
Reaction: *p*-cumate + NADH + H⁺ + O₂ = (2*R*,3*S*)-2,3-dihydroxy-2,3-dihydro-*p*-cumate + NAD⁺

Systematic name: 4-isopropylbenzoate:oxygen 2,3-oxidoreductase
Comments: The enzyme, characterized from several *Pseudomonas* strains, is involved in the degradation of *p*-cymene and *p*-cumate. It comprises four components: a ferredoxin, a ferredoxin reductase, and two subunits of a catalytic component. The enzyme can also act on indole, transforming it to the water-insoluble blue dye indigo.
References: [861, 4622, 1009, 1007]

[EC 1.14.12.25 created 2016]

EC 1.14.12.26

Accepted name: chlorobenzene dioxygenase
Reaction: chlorobenzene + NADH + H⁺ + O₂ = (1*R*,2*R*)-3-chlorocyclohexa-3,5-diene-1,2-diol + NAD⁺
Other name(s): TecA
Systematic name: chlorobenzene,NADH:oxygen oxidoreductase (1,2-hydroxylating)
Comments: This bacterial enzyme is a class IIB dioxygenase, comprising three components - a heterodimeric terminal dioxygenase, a ferredoxin protein, and a ferredoxin reductase. The enzyme acts on a range of aromatic compounds, including mono-, di-, tri-, and tetra-chlorinated benzenes and toluenes.
References: [3986, 3959, 273, 274]

[EC 1.14.12.26 created 2018]

EC 1.14.13 With NADH or NADPH as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.13.1

Accepted name: salicylate 1-monooxygenase
Reaction: salicylate + NADH + 2 H⁺ + O₂ = catechol + NAD⁺ + H₂O + CO₂
Other name(s): salicylate hydroxylase; salicylate 1-hydroxylase; salicylate monooxygenase; salicylate hydroxylase (decarboxylating)
Systematic name: salicylate,NADH:oxygen oxidoreductase (1-hydroxylating, decarboxylating)
Comments: A flavoprotein (FAD).
References: [4139, 4180, 4179, 4743]

[EC 1.14.13.1 created 1972]

EC 1.14.13.2

Accepted name: 4-hydroxybenzoate 3-monooxygenase
Reaction: 4-hydroxybenzoate + NADPH + H⁺ + O₂ = 3,4-dihydroxybenzoate + NADP⁺ + H₂O
Other name(s): *p*-hydroxybenzoate hydrolyase; *p*-hydroxybenzoate hydroxylase; 4-hydroxybenzoate 3-hydroxylase; 4-hydroxybenzoate monooxygenase; 4-hydroxybenzoic hydroxylase; *p*-hydroxybenzoate-3-hydroxylase; *p*-hydroxybenzoic acid hydrolase; *p*-hydroxybenzoic acid hydroxylase; *p*-hydroxybenzoic hydroxylase
Systematic name: 4-hydroxybenzoate,NADPH:oxygen oxidoreductase (3-hydroxylating)
Comments: A flavoprotein (FAD). Most enzymes from *Pseudomonas* are highly specific for NADPH (*cf.* EC 1.14.13.33 4-hydroxybenzoate 3-monooxygenase [NAD(P)H]).
References: [1739, 1745, 3976, 3974, 3975, 3785]

[EC 1.14.13.2 created 1972, modified 1999]

[1.14.13.3 Transferred entry. 4-hydroxyphenylacetate 3-monooxygenase. Now EC 1.14.14.9, 4-hydroxyphenylacetate 3-monooxygenase.]

[EC 1.14.13.3 created 1972, deleted 2011]

EC 1.14.13.4

- Accepted name:** melilotate 3-monooxygenase
Reaction: 3-(2-hydroxyphenyl)propanoate + NADH + H⁺ + O₂ = 3-(2,3-dihydroxyphenyl)propanoate + NAD⁺ + H₂O
Other name(s): 2-hydroxyphenylpropionate hydroxylase; melilotate hydroxylase; 2-hydroxyphenylpropionic hydroxylase; melilotic hydroxylase
Systematic name: 3-(2-hydroxyphenyl)propanoate,NADH:oxygen oxidoreductase (3-hydroxylating)
Comments: A flavoprotein (FAD).
References: [2431, 2432, 4063, 4062]

[EC 1.14.13.4 created 1972]

EC 1.14.13.5

- Accepted name:** imidazoleacetate 4-monooxygenase
Reaction: 4-imidazoleacetate + NADH + H⁺ + O₂ = 5-hydroxy-4-imidazoleacetate + NAD⁺ + H₂O
Other name(s): imidazoleacetic hydroxylase; imidazoleacetate hydroxylase; imidazoleacetic monooxygenase
Systematic name: 4-imidazoleacetate,NADH:oxygen oxidoreductase (5-hydroxylating)
Comments: A flavoprotein (FAD).
References: [2626]

[EC 1.14.13.5 created 1965 as EC 1.14.1.5, transferred 1972 to EC 1.14.13.5]

EC 1.14.13.6

- Accepted name:** orcinol 2-monooxygenase
Reaction: orcinol + NADH + H⁺ + O₂ = 2,3,5-trihydroxytoluene + NAD⁺ + H₂O
Other name(s): orcinol hydroxylase
Systematic name: orcinol,NADH:oxygen oxidoreductase (2-hydroxylating)
Comments: A flavoprotein (FAD).
References: [3205]

[EC 1.14.13.6 created 1972]

EC 1.14.13.7

- Accepted name:** phenol 2-monooxygenase (NADPH)
Reaction: phenol + NADPH + H⁺ + O₂ = catechol + NADP⁺ + H₂O
Other name(s): phenol hydroxylase; phenol *o*-hydroxylase
Systematic name: phenol,NADPH:oxygen oxidoreductase (2-hydroxylating)
Comments: A flavoprotein (FAD). The enzyme from the fungus *Trichosporon cutaneum* has a broad substrate specificity, and has been reported to catalyse the hydroxylation of a variety of substituted phenols, such as fluoro-, chloro-, amino- and methyl-phenols and also dihydroxybenzenes. *cf.* EC 1.14.14.20, phenol 2-monooxygenase (FADH₂).
References: [2973, 3046, 3047]

[EC 1.14.13.7 created 1972, modified 2011, modified 2016]

EC 1.14.13.8

- Accepted name:** flavin-containing monooxygenase
Reaction: *N,N*-dimethylaniline + NADPH + H⁺ + O₂ = *N,N*-dimethylaniline *N*-oxide + NADP⁺ + H₂O
Other name(s): dimethylaniline oxidase; dimethylaniline *N*-oxidase; FAD-containing monooxygenase; *N,N*-dimethylaniline monooxygenase; DMA oxidase; flavin mixed function oxidase; Ziegler's enzyme; mixed-function amine oxidase; FMO; FMO-I; FMO-II; FMO1; FMO2; FMO3; FMO4; FMO5; flavin monooxygenase; methylphenyltetrahydropyridine *N*-monooxygenase; 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine:oxygen *N*-oxidoreductase; dimethylaniline monooxygenase (*N*-oxide-forming)

Systematic name: *N,N*-dimethylaniline,NADPH:oxygen oxidoreductase (*N*-oxide-forming)
Comments: A flavoprotein. A broad spectrum monooxygenase that accepts substrates as diverse as hydrazines, phosphines, boron-containing compounds, sulfides, selenides, iodide, as well as primary, secondary and tertiary amines [571, 572]. This enzyme is distinct from other monooxygenases in that the enzyme forms a relatively stable hydroperoxy flavin intermediate [572, 1945]. This microsomal enzyme generally converts nucleophilic heteroatom-containing chemicals and drugs into harmless, readily excreted metabolites. For example, *N*-oxygenation is largely responsible for the detoxification of the dopaminergic neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) [652, 651]
References: [4930, 652, 571, 572, 1945, 651]

[EC 1.14.13.8 created 1972 (EC 1.13.12.11 created 1992, part-incorporated 2006), modified 2006]

EC 1.14.13.9

Accepted name: kynurenine 3-monooxygenase
Reaction: L-kynurenine + NADPH + H⁺ + O₂ = 3-hydroxy-L-kynurenine + NADP⁺ + H₂O
Other name(s): kynurenine 3-hydroxylase; kynurenine hydroxylase; L-kynurenine-3-hydroxylase
Systematic name: L-kynurenine,NADPH:oxygen oxidoreductase (3-hydroxylating)
Comments: A flavoprotein (FAD).
References: [843, 3155, 3635]

[EC 1.14.13.9 created 1961 as EC 1.99.1.5, transferred 1965 to EC 1.14.1.2, transferred 1972 to EC 1.14.13.9]

EC 1.14.13.10

Accepted name: 2,6-dihydroxypyridine 3-monooxygenase
Reaction: 2,6-dihydroxypyridine + NADH + H⁺ + O₂ = 2,3,6-trihydroxypyridine + NAD⁺ + H₂O
Other name(s): 2,6-dihydroxypyridine oxidase
Systematic name: 2,6-dihydroxypyridine,NADH:oxygen oxidoreductase (3-hydroxylating)
Comments: A flavoprotein.
References: [1700, 1701]

[EC 1.14.13.10 created 1976]

[1.14.13.11 *Transferred entry. trans-cinnamate 4-monooxygenase. Now EC 1.14.14.91, trans-cinnamate 4-monooxygenase*]

[EC 1.14.13.11 created 1976, deleted 2018]

[1.14.13.12 *Transferred entry. benzoate 4-monooxygenase. Now EC 1.14.14.92, benzoate 4-monooxygenase*]

[EC 1.14.13.12 created 1976, deleted 2018]

[1.14.13.13 *Transferred entry. calcidiol 1-monooxygenase. Now classified as EC 1.14.15.18, calcidiol 1-monooxygenase*]

[EC 1.14.13.13 created 1976, deleted 2016]

EC 1.14.13.14

Accepted name: *trans*-cinnamate 2-monooxygenase
Reaction: *trans*-cinnamate + NADPH + H⁺ + O₂ = 2-hydroxycinnamate + NADP⁺ + H₂O
Other name(s): cinnamic acid 2-hydroxylase; cinnamate 2-monooxygenase; cinnamic 2-hydroxylase; cinnamate 2-hydroxylase; *trans*-cinnamic acid 2-hydroxylase
Systematic name: *trans*-cinnamate,NADPH:oxygen oxidoreductase (2-hydroxylating)
References: [1309]

[EC 1.14.13.14 created 1976]

[1.14.13.15 *Transferred entry. cholestanetriol 26-monooxygenase. Now EC 1.14.15.15, cholestanetriol 26-monooxygenase.*]

[EC 1.14.13.15 created 1976, modified 2005, modified 2012, deleted 2016]

EC 1.14.13.16

Accepted name: cyclopentanone monooxygenase
Reaction: cyclopentanone + NADPH + H⁺ + O₂ = 5-valerolactone + NADP⁺ + H₂O
Other name(s): cyclopentanone oxygenase
Systematic name: cyclopentanone,NADPH:oxygen oxidoreductase (5-hydroxylating, lactonizing)
References: [1408, 1409]

[EC 1.14.13.16 created 1976]

[1.14.13.17 *Transferred entry. cholesterol 7 α -monooxygenase. Now EC 1.14.14.23, cholesterol 7 α -monooxygenase*]

[EC 1.14.13.17 created 1976, deleted 2016]

EC 1.14.13.18

Accepted name: 4-hydroxyphenylacetate 1-monooxygenase
Reaction: 4-hydroxyphenylacetate + NAD(P)H + H⁺ + O₂ = homogentisate + NAD(P)⁺ + H₂O
Other name(s): 4-hydroxyphenylacetate 1-hydroxylase; 4-hydroxyphenylacetic 1-hydroxylase; 4-HPA 1-hydroxylase
Systematic name: 4-hydroxyphenylacetate,NAD(P)H:oxygen oxidoreductase (1-hydroxylating)
Comments: A flavoprotein (FAD). Also acts on 4-hydroxyhydratropate (forming 2-methylhomogentisate) and on 4-hydroxyphenoxyacetate (forming hydroquinone and glycolate).
References: [1532]

[EC 1.14.13.18 created 1976]

EC 1.14.13.19

Accepted name: taxifolin 8-monooxygenase
Reaction: taxifolin + NAD(P)H + H⁺ + O₂ = 2,3-dihydrogossypetin + NAD(P)⁺ + H₂O
Other name(s): taxifolin hydroxylase
Systematic name: taxifolin,NAD(P)H:oxygen oxidoreductase (8-hydroxylating)
Comments: A flavoprotein, converting a flavanol into a flavanone. Also acts on fustin, but not on catechin, quercetin or mollisacidin.
References: [1896]

[EC 1.14.13.19 created 1976]

EC 1.14.13.20

Accepted name: 2,4-dichlorophenol 6-monooxygenase
Reaction: 2,4-dichlorophenol + NADPH + H⁺ + O₂ = 3,5-dichlorocatechol + NADP⁺ + H₂O
Other name(s): 2,4-dichlorophenol hydroxylase; 2,4-dichlorophenol monooxygenase
Systematic name: 2,4-dichlorophenol,NADPH:oxygen oxidoreductase (6-hydroxylating)
Comments: A flavoprotein (FAD). Also acts, more slowly, on 4-chlorophenol and 4-chloro-2-methylphenol; NADH can act instead of NADPH, but more slowly.
References: [252]

[EC 1.14.13.20 created 1983]

[1.14.13.21 *Transferred entry. flavonoid 3'-monooxygenase. Now EC 1.14.14.82, flavonoid 3'-monooxygenase.*]

[EC 1.14.13.21 created 1983, deleted 2018]

EC 1.14.13.22

Accepted name: cyclohexanone monooxygenase
Reaction: cyclohexanone + NADPH + H⁺ + O₂ = hexano-6-lactone + NADP⁺ + H₂O

Other name(s): cyclohexanone 1,2-monooxygenase; cyclohexanone oxygenase; cyclohexanone:NADPH:oxygen oxidoreductase (6-hydroxylating, 1,2-lactonizing)
Systematic name: cyclohexanone,NADPH:oxygen oxidoreductase (lactone-forming)
Comments: A flavoprotein (FAD). In the catalytic mechanism of this enzyme, the nucleophilic species that attacks the carbonyl group is a peroxyflavin intermediate that is generated by reaction of the enzyme-bound flavin cofactor with NAD(P)H and oxygen [3843]. This enzyme is able to catalyse a wide range of oxidative reactions, including enantioselective Baeyer-Villiger reactions [4026], sulfoxidations [628], amine oxidations [3209] and epoxidations [718].
References: [950, 3843, 4026, 628, 3209, 718]

[EC 1.14.13.22 created 1984, modified 2004]

EC 1.14.13.23

Accepted name: 3-hydroxybenzoate 4-monooxygenase
Reaction: 3-hydroxybenzoate + NADPH + H⁺ + O₂ = 3,4-dihydroxybenzoate + NADP⁺ + H₂O
Other name(s): 3-hydroxybenzoate 4-hydroxylase
Systematic name: 3-hydroxybenzoate,NADPH:oxygen oxidoreductase (4-hydroxylating)
Comments: A flavoprotein (FAD). Acts also on a number of analogues of 3-hydroxybenzoate substituted in the 2, 4, 5 and 6 positions.
References: [2790, 3374]

[EC 1.14.13.23 created 1972 as EC 1.14.99.13, transferred 1984 to EC 1.14.13.23]

EC 1.14.13.24

Accepted name: 3-hydroxybenzoate 6-monooxygenase
Reaction: 3-hydroxybenzoate + NADH + H⁺ + O₂ = 2,5-dihydroxybenzoate + NAD⁺ + H₂O
Other name(s): 3-hydroxybenzoate 6-hydroxylase; *m*-hydroxybenzoate 6-hydroxylase; 3-hydroxybenzoic acid-6-hydroxylase
Systematic name: 3-hydroxybenzoate,NADH:oxygen oxidoreductase (6-hydroxylating)
Comments: A flavoprotein (FAD). Acts also on a number of analogues of 3-hydroxybenzoate substituted in the 2, 4, 5 and 6 positions; NADPH can act instead of NADH, but more slowly.
References: [1420]

[EC 1.14.13.24 created 1984]

EC 1.14.13.25

Accepted name: methane monooxygenase (soluble)
Reaction: methane + NAD(P)H + H⁺ + O₂ = methanol + NAD(P)⁺ + H₂O
Other name(s): methane hydroxylase
Systematic name: methane,NAD(P)H:oxygen oxidoreductase (hydroxylating)
Comments: The enzyme is soluble, in contrast to the particulate enzyme, EC 1.14.18.3. Broad specificity; many alkanes can be hydroxylated, and alkenes are converted into the corresponding epoxides; CO is oxidized to CO₂, ammonia is oxidized to hydroxylamine, and some aromatic compounds and cyclic alkanes can also be hydroxylated, but more slowly.
References: [710, 1780, 4035, 4307]

[EC 1.14.13.25 created 1984, modified 2011]

[1.14.13.26 *Transferred entry. phosphatidylcholine 12-monooxygenase. Now classified as EC 1.14.18.4, phosphatidylcholine 12-monooxygenase.*]

[EC 1.14.13.26 created 1984, deleted 2015]

EC 1.14.13.27

Accepted name: 4-aminobenzoate 1-monooxygenase
Reaction: 4-aminobenzoate + NAD(P)H + 2 H⁺ + O₂ = 4-hydroxyaniline + NAD(P)⁺ + H₂O + CO₂
Other name(s): 4-aminobenzoate hydroxylase; 4-aminobenzoate monooxygenase
Systematic name: 4-aminobenzoate,NAD(P)H:oxygen oxidoreductase (1-hydroxylating, decarboxylating)
Comments: A flavoprotein (FAD). Acts on anthranilate and 4-aminosalicylate but not on salicylate (*cf.* EC 1.14.13.1 salicylate 1-monooxygenase).
References: [4342]

[EC 1.14.13.27 created 1989]

[1.14.13.28 *Transferred entry. 3,9-dihydroxypterocarpan 6a-monooxygenase. Now EC 1.14.14.93, 3,9-dihydroxypterocarpan 6a-monooxygenase*]

[EC 1.14.13.28 created 1989, deleted 2018]

EC 1.14.13.29

Accepted name: 4-nitrophenol 2-monooxygenase
Reaction: 4-nitrophenol + NADH + H⁺ + O₂ = 4-nitrocatechol + NAD⁺ + H₂O
Other name(s): 4-nitrophenol hydroxylase; 4-nitrophenol-2-hydroxylase
Systematic name: 4-nitrophenol,NADH:oxygen oxidoreductase (2-hydroxylating)
Comments: A flavoprotein (FAD).
References: [2832]

[EC 1.14.13.29 created 1989]

[1.14.13.30 *Transferred entry. leukotriene-B₄ 20-monooxygenase. Now EC 1.14.14.94, leukotriene-B₄ 20-monooxygenase*]

[EC 1.14.13.30 created 1989, deleted 2018]

EC 1.14.13.31

Accepted name: 2-nitrophenol 2-monooxygenase
Reaction: 2-nitrophenol + 2 NADPH + 2 H⁺ + O₂ = catechol + nitrite + 2 NADP⁺ + H₂O
Other name(s): 2-nitrophenol oxygenase; nitrophenol oxygenase
Systematic name: 2-nitrophenol,NADPH:oxygen 2-oxidoreductase (2-hydroxylating, nitrite-forming)
Comments: Involved in the metabolism of nitro-aromatic compounds by a strain of *Pseudomonas putida*.
References: [4872]

[EC 1.14.13.31 created 1989]

EC 1.14.13.32

Accepted name: albendazole monooxygenase
Reaction: albendazole + NADPH + H⁺ + O₂ = albendazole S-oxide + NADP⁺ + H₂O
Other name(s): albendazole oxidase (misleading); albendazole sulfoxidase (ambiguous); FMO3 (gene name); albendazole monooxygenase (flavin-containing)
Systematic name: albendazole,NADPH:oxygen oxidoreductase (sulfoxide-forming)
Comments: A microsomal flavin-containing monooxygenase. A similar conversion is also carried out by some microsomal cytochrome *P*-450 enzymes [EC 1.14.14.73, albendazole monooxygenase (sulfoxide-forming)]. It is estimated that cytochrome *P*-450s are responsible for 70% of the activity.
References: [1088, 2898, 3463]

[EC 1.14.13.32 created 1989, modified 2018]

EC 1.14.13.33

- Accepted name:** 4-hydroxybenzoate 3-monooxygenase [NAD(P)H]
Reaction: 4-hydroxybenzoate + NAD(P)H + H⁺ + O₂ = 3,4-dihydroxybenzoate + NAD(P)⁺ + H₂O
Other name(s): 4-hydroxybenzoate 3-monooxygenase (reduced nicotinamide adenine dinucleotide (phosphate)); 4-hydroxybenzoate-3-hydroxylase; 4-hydroxybenzoate 3-hydroxylase
Systematic name: 4-hydroxybenzoate,NAD(P)H:oxygen oxidoreductase (3-hydroxylating)
Comments: A flavoprotein (FAD). The enzyme from *Corynebacterium cyclohexanicum* is highly specific for 4-hydroxybenzoate, but uses NADH and NADPH at approximately equal rates (*cf.* EC 1.14.13.2 4-hydroxybenzoate 3-monooxygenase). It is less specific for NADPH than EC 1.14.13.2.
References: [1203, 3785]

[EC 1.14.13.33 created 1989, modified 1999]

EC 1.14.13.34

- Accepted name:** leukotriene-E₄ 20-monooxygenase
Reaction: (7*E*,9*E*,11*Z*,14*Z*)-(5*S*,6*R*)-6-(cystein-*S*-yl)-5-hydroxyicosa-7,9,11,14-tetraenoate + NADPH + H⁺ + O₂ = 20-hydroxyleukotriene E₄ + NADP⁺ + H₂O
Other name(s): leukotriene-E₄ ω-hydroxylase
Systematic name: (7*E*,9*E*,11*Z*,14*Z*)-(5*S*,6*R*)-6-(cystein-*S*-yl)-5-hydroxyicosa-7,9,11,14-tetraenoate,NADPH:oxygen oxidoreductase (20-hydroxylating)
Comments: Also acts on *N*-acetyl-leukotriene E₄, but more slowly. Not identical with EC 1.14.13.30 leukotriene-B₄ 20-monooxygenase.
References: [3192]

[EC 1.14.13.34 created 1989]

EC 1.14.13.35

- Accepted name:** anthranilate 3-monooxygenase (deaminating)
Reaction: anthranilate + NADPH + H⁺ + O₂ = 2,3-dihydroxybenzoate + NADP⁺ + NH₃
Other name(s): anthranilate hydroxylase; anthranilate 2,3-dioxygenase (deaminating); anthranilate hydroxylase (deaminating); anthranilic hydroxylase; anthranilate 2,3-hydroxylase (deaminating)
Systematic name: anthranilate,NADPH:oxygen oxidoreductase (3-hydroxylating, deaminating)
Comments: The enzyme from *Aspergillus niger* is an iron protein; that from the yeast *Trichosporon cutaneum* is a flavoprotein (FAD).
References: [3367, 4088]

[EC 1.14.13.35 created 1972 as EC 1.14.12.2, transferred 1990 to EC 1.14.13.35]

[1.14.13.36 Transferred entry. 5-*O*-(4-coumaroyl)-*D*-quininate 3'-monooxygenase. Now EC 1.14.14.96, 5-*O*-(4-coumaroyl)-*D*-quininate 3'-monooxygenase]

[EC 1.14.13.36 created 1990, deleted 2018]

[1.14.13.37 Transferred entry. methyltetrahydroprotoberberine 14-monooxygenase. Now EC 1.14.14.97, methyltetrahydroprotoberberine 14-monooxygenase]

[EC 1.14.13.37 created 1990, deleted 2018]

EC 1.14.13.38

- Accepted name:** anhydrotetracycline 6-monooxygenase
Reaction: anhydrotetracycline + NADPH + H⁺ + O₂ = 12-dehydrotetracycline + NADP⁺ + H₂O
Other name(s): ATC oxygenase; anhydrotetracycline oxygenase; *oxyS* (gene name); anhydrotetracycline monooxygenase
Systematic name: anhydrotetracycline,NADPH:oxygen oxidoreductase (6-hydroxylating)

Comments: The enzyme, characterized from the bacterium *Streptomyces rimosus*, participates in the biosynthesis of tetracycline antibiotics. It can also catalyse EC 1.14.13.234, 12-dehydrotetracycline 5-monoxygenase.

References: [270, 335, 4419, 4522]

[EC 1.14.13.38 created 1990, modified 2016]

EC 1.14.13.39

Accepted name: nitric-oxide synthase (NADPH)

Reaction: $2 \text{ L-arginine} + 3 \text{ NADPH} + 3 \text{ H}^+ + 4 \text{ O}_2 = 2 \text{ L-citrulline} + 2 \text{ nitric oxide} + 3 \text{ NADP}^+ + 4 \text{ H}_2\text{O}$ (overall reaction)

(1a) $2 \text{ L-arginine} + 2 \text{ NADPH} + 2 \text{ H}^+ + 2 \text{ O}_2 = 2 \text{ N}^{\omega}\text{-hydroxy-L-arginine} + 2 \text{ NADP}^+ + 2 \text{ H}_2\text{O}$

(1b) $2 \text{ N}^{\omega}\text{-hydroxy-L-arginine} + \text{NADPH} + \text{H}^+ + 2 \text{ O}_2 = 2 \text{ L-citrulline} + 2 \text{ nitric oxide} + \text{NADP}^+ + 2 \text{ H}_2\text{O}$

Other name(s): NOS (gene name); nitric oxide synthetase (ambiguous); endothelium-derived relaxation factor-forming enzyme; endothelium-derived relaxing factor synthase; NO synthase (ambiguous); NADPH-diaphorase (ambiguous)

Systematic name: L-arginine,NADPH:oxygen oxidoreductase (nitric-oxide-forming)

Comments: The enzyme consists of linked oxygenase and reductase domains. The eukaryotic enzyme binds FAD, FMN, heme (iron protoporphyrin IX) and tetrahydrobiopterin, and its two domains are linked via a regulatory calmodulin-binding domain. Upon calcium-induced calmodulin binding, the reductase and oxygenase domains form a complex, allowing electrons to flow from NADPH via FAD and FMN to the active center. The reductase domain of the enzyme from the bacterium *Sorangium cellulosum* utilizes a [2Fe-2S] cluster to transfer the electrons from NADPH to the active center. *cf.* EC 1.14.14.47, nitric-oxide synthase (flavodoxin).

References: [431, 4080, 4079, 31, 1139]

[EC 1.14.13.39 created 1992, modified 2012, modified 2017]

EC 1.14.13.40

Accepted name: anthraniloyl-CoA monoxygenase

Reaction: $\text{anthraniloyl-CoA} + 2 \text{ NAD(P)H} + 2 \text{ H}^+ + \text{O}_2 = 2\text{-amino-5-oxocyclohex-1-enecarboxyl-CoA} + \text{H}_2\text{O} + 2 \text{ NAD(P)}^+$

Other name(s): anthraniloyl coenzyme A reductase; 2-aminobenzoyl-CoA monoxygenase/reductase

Systematic name: anthraniloyl-CoA,NAD(P)H:oxygen oxidoreductase (de-aromatizing)

Comments: A flavoprotein (FAD). The non-aromatic product is unstable and releases CO₂ and NH₃, forming 1,4-cyclohexanedione.

References: [488, 489, 2348]

[EC 1.14.13.40 created 1992]

[1.14.13.41 *Transferred entry. tyrosine N-monoxygenase. Now EC 1.14.14.36, tyrosine N-monoxygenase*]

[EC 1.14.13.41 created 1992, modified 2001, modified 2005, deleted 2016]

[1.14.13.42 *Deleted entry. hydroxyphenylacetonitrile 2-monoxygenase. The activity is covered by EC 1.14.13.68, 4-hydroxyphenylacetaldehyde oxime monoxygenase, that performs the two consecutive reactions in the conversion of (Z)-4-hydroxyphenylacetaldehyde oxime to (S)-4-hydroxymandelonitrile*]

[EC 1.14.13.42 created 1992, deleted 2011]

EC 1.14.13.43

Accepted name: questin monoxygenase

Reaction: $\text{questin} + \text{NADPH} + \text{H}^+ + \text{O}_2 = \text{demethylsulochrin} + \text{NADP}^+$

Other name(s): questin oxygenase

Systematic name: questin,NADPH:oxygen oxidoreductase (hydroxylating, anthraquinone-ring-opening)
Comments: The enzyme cleaves the anthraquinone ring of questin to form a benzophenone. Involved in the biosynthesis of the seco-anthraquinone (+)-geodin.
References: [1202]

[EC 1.14.13.43 created 1992]

EC 1.14.13.44

Accepted name: 2-hydroxybiphenyl 3-monoxygenase
Reaction: 2-hydroxybiphenyl + NADH + H⁺ + O₂ = 2,3-dihydroxybiphenyl + NAD⁺ + H₂O
Systematic name: 2-hydroxybiphenyl,NADH:oxygen oxidoreductase (3-hydroxylating)
Comments: Also converts 2,2'-dihydroxybiphenyl into 2,2',3-trihydroxy-biphenyl.
References: [2197]

[EC 1.14.13.44 created 1992]

[1.14.13.45 *Transferred entry. CMP-N-acetylneuraminate monoxygenase. Now EC 1.14.18.2, CMP-N-acetylneuraminate monoxygenase*]

[EC 1.14.13.45 created 1992, deleted 2003]

EC 1.14.13.46

Accepted name: (-)-menthol monoxygenase
Reaction: (-)-menthol + NADPH + H⁺ + O₂ = *p*-menthane-3,8-diol + NADP⁺ + H₂O
Other name(s): *l*-menthol monoxygenase
Systematic name: (-)-menthol,NADPH:oxygen oxidoreductase (8-hydroxylating)
References: [2602]

[EC 1.14.13.46 created 1992]

[1.14.13.47 *Transferred entry. (S)-limonene 3-monoxygenase. Now EC 1.14.14.99, (S)-limonene 3-monoxygenase*]

[EC 1.14.13.47 created 1992, modified 2003, deleted 2018]

[1.14.13.48 *Transferred entry. (S)-limonene 6-monoxygenase. Now classified as EC 1.14.14.51, (S)-limonene 6-monoxygenase*]

[EC 1.14.13.48 created 1992, modified 2003, deleted 2017]

[1.14.13.49 *Transferred entry. (S)-limonene 7-monoxygenase. Now classified as EC 1.14.14.52, (S)-limonene 7-monoxygenase*]

[EC 1.14.13.49 created 1992, modified 2003, deleted 2017]

EC 1.14.13.50

Accepted name: pentachlorophenol monoxygenase
Reaction: (1) pentachlorophenol + NADPH + H⁺ + O₂ = 2,3,5,6-tetrachloro-1,4-benzoquinone + NADP⁺ + chloride + H₂O
(2) 2,3,5,6-tetrachlorophenol + NADPH + H⁺ + O₂ = 2,3,5,6-tetrachlorohydroquinone + NADP⁺ + H₂O
Other name(s): *pcpB* (gene name); pentachlorophenol dechlorinase; pentachlorophenol dehalogenase; pentachlorophenol 4-monoxygenase; PCP hydroxylase; pentachlorophenol hydroxylase; PCB 4-monoxygenase; PCB4MO
Systematic name: pentachlorophenol,NADPH:oxygen oxidoreductase (hydroxylating, dechlorinating)
Comments: A flavoprotein (FAD). The enzyme displaces a diverse range of substituents from the 4-position of polyhalogenated phenols but requires that a halogen substituent be present at the 2-position [4714]. If C-4 carries a halogen substituent, reaction 1 is catalysed; if C-4 is unsubstituted, reaction 2 is catalysed.

References: [3709, 4714, 4713, 2346, 2984, 633, 1676, 3593]

[EC 1.14.13.50 created 1992, modified 2005, modified 2017]

EC 1.14.13.51

Accepted name: 6-oxocineole dehydrogenase
Reaction: 6-oxocineole + NADPH + H⁺ + O₂ = 1,6,6-trimethyl-2,7-dioxabicyclo[3.2.2]nonan-3-one + NADP⁺ + H₂O
Other name(s): 6-oxocineole oxygenase
Systematic name: 6-oxocineole,NADPH:oxygen oxidoreductase
Comments: The product undergoes non-enzymic cleavage and subsequent ring closure to form the lactone 4,5-dihydro-5,5-dimethyl-4-(3-oxobutyl)furan-2(3*H*)-one.
References: [4631]

[EC 1.14.13.51 created 1992]

[1.14.13.52 *Transferred entry. isoflavone 3'-hydroxylase. Now EC 1.14.14.88, isoflavone 3'-hydroxylase*]

[EC 1.14.13.52 created 1992, deleted 2018]

[1.14.13.53 *Transferred entry. 4'-methoxyisoflavone 2'-hydroxylase. Now EC 1.14.14.89, 4'-methoxyisoflavone 2'-hydroxylase*]

[EC 1.14.13.53 created 1992, modified 2005, deleted 2018]

EC 1.14.13.54

Accepted name: ketosteroid monooxygenase
Reaction: a ketosteroid + NADPH + H⁺ + O₂ = a steroid ester/lactone + NADP⁺ + H₂O (general reaction)
(1) progesterone + NADPH + H⁺ + O₂ = testosterone acetate + NADP⁺ + H₂O
(2) androstenedione + NADPH + H⁺ + O₂ = testololactone + NADP⁺ + H₂O
(3) 17 α -hydroxyprogesterone + NADPH + H⁺ + O₂ = androstenedione + acetate + NADP⁺ + H₂O
Other name(s): steroid-ketone monooxygenase; progesterone, NADPH₂:oxygen oxidoreductase (20-hydroxylating, ester-producing); 17 α -hydroxyprogesterone, NADPH₂:oxygen oxidoreductase (20-hydroxylating, side-chain cleaving); androstenedione, NADPH₂:oxygen oxidoreductase (17-hydroxylating, lactonizing)
Systematic name: ketosteroid,NADPH:oxygen oxidoreductase (20-hydroxylating, ester-producing/20-hydroxylating, side-chain cleaving/17-hydroxylating, lactonizing)
Comments: A single FAD-containing enzyme catalyses three types of monooxygenase (Baeyer-Villiger oxidation) reaction. The oxidative esterification of a number of derivatives of progesterone to produce the corresponding 17 α -hydroxysteroid 17-acetate ester, such as testosterone acetate, is shown in Reaction (1). The oxidative lactonization of a number of derivatives of androstenedione to produce the 13,17-secoandrosten-17,13 α -lactone, such as testololactone, is shown in Reaction (2). The oxidative cleavage of the 17 β -side-chain of 17 α -hydroxyprogesterone to produce androstenedione and acetate is shown in Reaction (3). Reaction (1) is also catalysed by EC 1.14.99.4 (progesterone monooxygenase), and Reactions (2) and (3) correspond to that catalysed by EC 1.14.99.12 (androst-4-ene-3,17-dione monooxygenase). The possibility that a single enzyme is responsible for the reactions ascribed to EC 1.14.99.4 and EC 1.14.99.12 in other tissues cannot be excluded.
References: [2008, 1842, 1843]

[EC 1.14.13.54 created 1999]

[1.14.13.55 *Transferred entry. protopine 6-monooxygenase. Now EC 1.14.14.98, protopine 6-monooxygenase*]

[EC 1.14.13.55 created 1999, deleted 2018]

[1.14.13.56 *Transferred entry. dihydrosanguinarine 10-monooxygenase. Now EC 1.14.14.100, dihydrosanguinarine 10-monooxygenase*]

[EC 1.14.13.56 created 1999, deleted 2018]

[1.14.13.57 Transferred entry. dihydrochelirubine 12-monooxygenase. Now EC 1.14.14.101, dihydrochelirubine 12-monooxygenase]

[EC 1.14.13.57 created 1999, deleted 2018]

EC 1.14.13.58

Accepted name: benzoyl-CoA 3-monooxygenase
Reaction: benzoyl-CoA + NADPH + H⁺ + O₂ = 3-hydroxybenzoyl-CoA + NADP⁺ + H₂O
Other name(s): benzoyl-CoA 3-hydroxylase
Systematic name: benzoyl-CoA,NADPH:oxygen oxidoreductase (3-hydroxylating)
Comments: The enzyme from the denitrifying bacterium *Pseudomonas KB740* catalyses a flavin-requiring reaction (FAD or FMN). Benzoate is not a substrate.
References: [3069]

[EC 1.14.13.58 created 1999]

EC 1.14.13.59

Accepted name: L-lysine N⁶-monooxygenase (NADPH)
Reaction: L-lysine + NADPH + H⁺ + O₂ = N⁶-hydroxy-L-lysine + NADP⁺ + H₂O
Other name(s): lysine N⁶-hydroxylase; L-lysine 6-monooxygenase (NADPH) (ambiguous)
Systematic name: L-lysine,NADPH:oxygen oxidoreductase (6-hydroxylating)
Comments: A flavoprotein (FAD). The enzyme from strain EN 222 of *Escherichia coli* is highly specific for L-lysine; L-ornithine and L-homolysine are, for example, not substrates.
References: [3334, 2591, 4251, 851, 2659, 1350]

[EC 1.14.13.59 created 1999, modified 2001, modified 2012]

[1.14.13.60 Transferred entry. 27-hydroxycholesterol 7 α -monooxygenase. Now classified as EC 1.14.14.29, 25/26-hydroxycholesterol 7 α -hydroxylase]

[EC 1.14.13.60 created 1999, deleted 2013]

EC 1.14.13.61

Accepted name: 2-hydroxyquinoline 8-monooxygenase
Reaction: quinolin-2-ol + NADH + H⁺ + O₂ = quinolin-2,8-diol + NAD⁺ + H₂O
Other name(s): 2-oxo-1,2-dihydroquinoline 8-monooxygenase
Systematic name: quinolin-2(1H)-one,NADH:oxygen oxidoreductase (8-oxygenating)
Comments: Requires iron. Quinolin-2-ol exists largely as the quinolin-2(1H)-one tautomer.
References: [3572]

[EC 1.14.13.61 created 1999]

EC 1.14.13.62

Accepted name: 4-hydroxyquinoline 3-monooxygenase
Reaction: quinolin-4-ol + NADH + H⁺ + O₂ = quinolin-3,4-diol + NAD⁺ + H₂O
Other name(s): quinolin-4(1H)-one 3-monooxygenase
Systematic name: quinolin-4(1H)-one,NADH:oxygen oxidoreductase (3-oxygenating)
Comments: Quinolin-4-ol exists largely as the quinolin-4(1H)-one tautomer.
References: [361]

[EC 1.14.13.62 created 1999]

EC 1.14.13.63

Accepted name: 3-hydroxyphenylacetate 6-hydroxylase
Reaction: 3-hydroxyphenylacetate + NAD(P)H + H⁺ + O₂ = 2,5-dihydroxyphenylacetate + NAD(P)⁺ + H₂O
Other name(s): 3-hydroxyphenylacetate 6-monooxygenase
Systematic name: 3-hydroxyphenylacetate,NAD(P)H:oxygen oxidoreductase (6-hydroxylating)
Comments: 3-hydroxyphenylacetate 6-hydroxylase from *Flavobacterium* sp. is highly specific for 3-hydroxyphenylacetate and uses NADH and NADPH as electron donors with similar efficiency.
References: [4396]

[EC 1.14.13.63 created 1999]

EC 1.14.13.64

Accepted name: 4-hydroxybenzoate 1-hydroxylase
Reaction: 4-hydroxybenzoate + NAD(P)H + 2 H⁺ + O₂ = hydroquinone + NAD(P)⁺ + H₂O + CO₂
Other name(s): 4-hydroxybenzoate 1-monooxygenase
Systematic name: 4-hydroxybenzoate,NAD(P)H:oxygen oxidoreductase (1-hydroxylating, decarboxylating)
Comments: Requires FAD. The enzyme from *Candida parapsilosis* is specific for 4-hydroxybenzoate derivatives and prefers NADH to NADPH as electron donor.
References: [4397]

[EC 1.14.13.64 created 1999]

[1.14.13.65 Deleted entry. 2-hydroxyquinoline 8-monooxygenase]

[EC 1.14.13.65 created 1999, deleted 2006]

EC 1.14.13.66

Accepted name: 2-hydroxycyclohexanone 2-monooxygenase
Reaction: 2-hydroxycyclohexan-1-one + NADPH + H⁺ + O₂ = 6-hydroxyhexan-6-olide + NADP⁺ + H₂O
Systematic name: 2-hydroxycyclohexan-1-one,NADPH:oxygen 2-oxidoreductase (1,2-lactonizing)
Comments: The product decomposes spontaneously to 6-oxohexanoic acid (adipic semialdehyde).
References: [837]

[EC 1.14.13.66 created 1978 as EC 1.14.12.6, transferred 1999 to EC 1.14.13.66]

[1.14.13.67 Transferred entry. quinine 3-monooxygenase. Now EC 1.14.14.55, quinine 3-monooxygenase]

[EC 1.14.13.67 created 2000, deleted 2017]

[1.14.13.68 Transferred entry. 4-hydroxyphenylacetaldehyde oxime monooxygenase. Now EC 1.14.14.37, 4-hydroxyphenylacetaldehyde oxime monooxygenase]

[EC 1.14.13.68 created 2000, modified 2005, deleted 2016]

EC 1.14.13.69

Accepted name: alkene monooxygenase
Reaction: propene + NADH + H⁺ + O₂ = 1,2-epoxypropane + NAD⁺ + H₂O
Other name(s): alkene epoxygenase; etnABCD (gene names); amoABCDE (gene names)
Systematic name: alkene,NADH:oxygen oxidoreductase
Comments: This bacterial binuclear non-heme iron enzyme is a multicomponent enzyme complex comprising an oxygenase, a reductase, and a Rieske-type ferredoxin. The enzyme from the bacterium *Xanthobacter* sp. strain Py2 contains an additional small protein of unknown function that is essential for activity. In general, the enzyme oxygenates C₂ to C₆ aliphatic alkenes, although enzymes from different organisms show different substrate range. With propene as substrate, the stereospecificity of the epoxypropane formed is 95% (R) and 5% (S).

References: [3929, 1258, 4915, 591, 590]

[EC 1.14.13.69 created 2001]

[1.14.13.70] *Transferred entry. sterol 14 α -demethylase. Now EC 1.14.14.154, sterol 14 α -demethylase]*

[EC 1.14.13.70 created 2001, modified 2013, deleted 2018]

[1.14.13.71] *Transferred entry. N-methylcoclaurine 3'-monooxygenase. Now EC 1.14.14.102, N-methylcoclaurine 3'-monooxygenase]*

[EC 1.14.13.71 created 2001, deleted 2018]

[1.14.13.72] *Transferred entry. methylsterol monooxygenase. Now classified as EC 1.14.18.9, methylsterol monooxygenase]*

[EC 1.14.13.72 created 1972 as EC 1.14.99.16, transferred 2002 to EC 1.14.13.72, deleted 2017]

[1.14.13.73] *Transferred entry. tabersonine 16-hydroxylase. Now EC 1.14.14.103, tabersonine 16-hydroxylase]*

[EC 1.14.13.73 created 2002, deleted 2018]

[1.14.13.74] *Transferred entry. 7-deoxyloganin 7-hydroxylase. Now EC 1.14.14.85, 7-deoxyloganin 7-hydroxylase]*

[EC 1.14.13.74 created 2002, deleted 2018]

[1.14.13.75] *Transferred entry. vinorine hydroxylase. Now EC 1.14.14.104, vinorine hydroxylase]*

[EC 1.14.13.75 created 2002, deleted 2018]

[1.14.13.76] *Transferred entry. taxane 10 β -hydroxylase. Now EC 1.14.14.105, taxane 10 β -hydroxylase]*

[EC 1.14.13.76 created 2002, deleted 2018]

[1.14.13.77] *Transferred entry. taxane 13 α -hydroxylase. Now EC 1.14.14.106, taxane 13 α -hydroxylase]*

[EC 1.14.13.77 created 2002, deleted 2018]

[1.14.13.78] *Transferred entry. ent-kaurene oxidase. Now EC 1.14.14.86, ent-kaurene monooxygenase]*

[EC 1.14.13.78 created 2002, deleted 2018]

[1.14.13.79] *Transferred entry. ent-kaurenoic acid oxidase. Now EC 1.14.14.107, ent-kaurenoic acid oxidase]*

[EC 1.14.13.79 created 2002, deleted 2018]

[1.14.13.80] *Transferred entry. (R)-limonene 6-monooxygenase. Now classified as EC 1.14.14.53, (R)-limonene 6-monooxygenase]*

[EC 1.14.13.80 created 2003, deleted 2017]

EC 1.14.13.81

Accepted name: magnesium-protoporphyrin IX monomethyl ester (oxidative) cyclase

Reaction: magnesium-protoporphyrin IX 13-monomethyl ester + 3 NADPH + 3 H⁺ + 3 O₂ = 3,8-divinyl protochlorophyllide *a* + 3 NADP⁺ + 5 H₂O (overall reaction)

(1a) magnesium-protoporphyrin IX 13-monomethyl ester + NADPH + H⁺ + O₂ = 13¹-hydroxy-magnesium-protoporphyrin IX 13-monomethyl ester + NADP⁺ + H₂O

(1b) 13¹-hydroxy-magnesium-protoporphyrin IX 13-monomethyl ester + NADPH + H⁺ + O₂ = 13¹-oxo-magnesium-protoporphyrin IX 13-monomethyl ester + NADP⁺ + 2 H₂O

(1c) 13¹-oxo-magnesium-protoporphyrin IX 13-monomethyl ester + NADPH + H⁺ + O₂ = 3,8-divinyl protochlorophyllide *a* + NADP⁺ + 2 H₂O

Other name(s): Mg-protoporphyrin IX monomethyl ester (oxidative) cyclase

Systematic name: magnesium-protoporphyrin-IX 13-monomethyl ester,NADPH:oxygen oxidoreductase (hydroxylating)

Comments: Requires iron(II) for activity. The enzyme participates in the biosynthesis of chlorophyllide *a* in aerobic organisms. The same transformation is achieved in anaerobic organisms by EC 1.21.98.3, anaerobic magnesium-protoporphyrin IX monomethyl ester cyclase. Some facultative phototrophic bacteria, such as *Rubrivivax gelatinosus*, possess both enzymes.

References: [4495, 379, 3329, 4315]

[EC 1.14.13.81 created 2003, modified 2017]

EC 1.14.13.82

Accepted name: vanillate monooxygenase
Reaction: vanillate + O₂ + NADH + H⁺ = 3,4-dihydroxybenzoate + NAD⁺ + H₂O + formaldehyde
Other name(s): 4-hydroxy-3-methoxybenzoate demethylase; vanillate demethylase
Systematic name: vanillate:oxygen oxidoreductase (demethylating)
Comments: Forms part of the vanillin degradation pathway in *Arthrobacter sp.*
References: [475, 3379]

[EC 1.14.13.82 created 2000 as EC 1.2.3.12, transferred 2003 to EC 1.14.13.82]

EC 1.14.13.83

Accepted name: precorrin-3B synthase
Reaction: precorrin-3A + NADH + H⁺ + O₂ = precorrin-3B + NAD⁺ + H₂O
Other name(s): precorrin-3X synthase; CobG
Systematic name: precorrin-3A,NADH:oxygen oxidoreductase (20-hydroxylating)
Comments: An iron-sulfur protein. An oxygen atom from dioxygen is incorporated into the macrocycle at C-20. In the aerobic cobalamin biosynthesis pathway, four enzymes are involved in the conversion of precorrin-3A to precorrin-6A. The first of the four steps is carried out by EC 1.14.13.83, precorrin-3B synthase (CobG), yielding precorrin-3B as the product. This is followed by three methylation reactions, which introduce a methyl group at C-17 (CobJ; EC 2.1.1.131), C-11 (CobM; EC 2.1.1.133) and C-1 (CobF; EC 2.1.1.152) of the macrocycle, giving rise to precorrin-4, precorrin-5 and precorrin-6A, respectively.
References: [858, 3771, 4550]

[EC 1.14.13.83 created 2004]

EC 1.14.13.84

Accepted name: 4-hydroxyacetophenone monooxygenase
Reaction: (4-hydroxyphenyl)ethan-1-one + NADPH + H⁺ + O₂ = 4-hydroxyphenyl acetate + NADP⁺ + H₂O
Other name(s): HAPMO
Systematic name: (4-hydroxyphenyl)ethan-1-one,NADPH:oxygen oxidoreductase (ester-forming)
Comments: Contains FAD. The enzyme from *Pseudomonas fluorescens* ACB catalyses the conversion of a wide range of acetophenone derivatives. Highest activity occurs with compounds bearing an electron-donating substituent at the para position of the aromatic ring [1984]. In the absence of substrate, the enzyme can act as an NAD(P)H oxidase (EC 1.6.3.1).
References: [1984, 1985]

[EC 1.14.13.84 created 2004]

[1.14.13.85 Transferred entry. glyceollin synthase. Now EC 1.14.14.135, glyceollin synthase]

[EC 1.14.13.85 created 2004, deleted 2018]

[1.14.13.86 Deleted entry. 2-hydroxyisoflavanone synthase. This enzyme was classified on the basis of an incorrect reaction. The activity is covered by EC 1.14.14.87, 2-hydroxyisoflavanone synthase]

[EC 1.14.13.86 created 2004, deleted 2013]

- [1.14.13.87] *Transferred entry. licodione synthase. Now EC 1.14.14.140, licodione synthase*
[EC 1.14.13.87 created 2004, deleted 2018]
- [1.14.13.88] *Transferred entry. flavanoid 3,5-hydroxylase. Now EC 1.14.14.81, flavanoid 3,5-hydroxylase*
[EC 1.14.13.88 created 2004, deleted 2018]
- [1.14.13.89] *Transferred entry. isoflavone 2-hydroxylase. Now EC 1.14.14.90, isoflavone 2-hydroxylase*
[EC 1.14.13.89 created 2005, deleted 2018]
- [1.14.13.90] *Transferred entry. zeaxanthin epoxidase. Now EC 1.14.15.21, zeaxanthin epoxidase*
[EC 1.14.13.90 created 2005, deleted 2016]
- [1.14.13.91] *Transferred entry. deoxysarpagine hydroxylase. Now EC 1.14.14.136, deoxysarpagine hydroxylase*
[EC 1.14.13.91 created 2005, deleted 2018]

EC 1.14.13.92

- Accepted name:** phenylacetone monooxygenase
Reaction: phenylacetone + NADPH + H⁺ + O₂ = benzyl acetate + NADP⁺ + H₂O
Other name(s): PAMO
Systematic name: phenylacetone,NADPH:oxygen oxidoreductase
Comments: A flavoprotein (FAD). NADH cannot replace NADPH as coenzyme. In addition to phenylacetone, which is the best substrate found to date, this Baeyer-Villiger monooxygenase can oxidize other aromatic ketones [1-(4-hydroxyphenyl)propan-2-one, 1-(4-hydroxyphenyl)propan-2-one and 3-phenylbutan-2-one], some alipatic ketones (e.g. dodecan-2-one) and sulfides (e.g. 1-methyl-4-(methylsulfanyl)benzene).
References: [2631, 1159]

[EC 1.14.13.92 created 2005]

- [1.14.13.93] *Transferred entry. (+)-abscisic acid 8-hydroxylase. Now EC 1.14.14.137, (+)-abscisic acid 8-hydroxylase*
[EC 1.14.13.93 created 2005, deleted 2018]
- [1.14.13.94] *Transferred entry. lithocholate 6β-hydroxylase. Now EC 1.14.14.138, lithocholate 6β-hydroxylase*
[EC 1.14.13.94 created 2005, deleted 2018]
- [1.14.13.95] *Transferred entry. 7α-hydroxycholest-4-en-3-one 12α-hydroxylase. Now included with EC 1.14.14.139, 5β-cholestane-3α,7α-diol 12α-hydroxylase*
[EC 1.14.13.95 created 2005, deleted 2015]
- [1.14.13.96] *Transferred entry. 5β-cholestane-3α,7α-diol 12α-hydroxylase. Now EC 1.14.14.139, 5β-cholestane-3α,7α-diol 12α-hydroxylase*
[EC 1.14.13.96 created 2005, deleted 2018]
- [1.14.13.97] *Transferred entry. taurochenodeoxycholate 6α-hydroxylase. Now EC 1.14.14.57, taurochenodeoxycholate 6α-hydroxylase*
[EC 1.14.13.97 created 2005, deleted 2018]
- [1.14.13.98] *Transferred entry. cholesterol 24-hydroxylase. Now EC 1.14.14.25, cholesterol 24-hydroxylase]*
[EC 1.14.13.98 created 2005, deleted 2016]
- [1.14.13.99] *Transferred entry. 24-hydroxycholesterol 7α-hydroxylase. Now EC 1.14.14.26, 24-hydroxycholesterol 7α-hydroxylase*

[EC 1.14.13.99 created 2005, deleted 2016]

[1.14.13.100 Transferred entry. 25/26-hydroxycholesterol 7 α -hydroxylase. Now classified as EC 1.14.14.29, 25/26-hydroxycholesterol 7 α -hydroxylase]

[EC 1.14.13.100 created 2005, modified 2013 (EC 1.14.13.60 created 1999, incorporated 2013), deleted 2016]

EC 1.14.13.101

Accepted name: senecionine *N*-oxygenase
Reaction: senecionine + NADPH + H⁺ + O₂ = senecionine *N*-oxide + NADP⁺ + H₂O
Other name(s): senecionine monooxygenase (*N*-oxide-forming); SNO
Systematic name: senecionine,NADPH:oxygen oxidoreductase (*N*-oxide-forming)
Comments: A flavoprotein. NADH cannot replace NADPH. While pyrrolizidine alkaloids of the senecionine and monocrotaline types are generally good substrates (e.g. senecionine, retrorsine and monocrotaline), the enzyme does not use ester alkaloids lacking an hydroxy group at C-7 (e.g. supinine and phalaenopsine), 1,2-dihydro-alkaloids (e.g. sarracine) or unesterified necine bases (e.g. senkirikine) as substrates [2498]. *Senecionine N*-oxide is used by insects as a chemical defense: senecionine *N*-oxide is non-toxic, but it is bioactivated to a toxic form by the action of cytochrome *P*-450 oxidase when absorbed by insectivores.
References: [2498, 3022]

[EC 1.14.13.101 created 2006]

[1.14.13.102 Transferred entry. psoralen synthase. Now EC 1.14.14.141, psoralen synthase]

[EC 1.14.13.102 created 2007, deleted 2018]

[1.14.13.103 Transferred entry. 8-dimethylallylnaringenin 2-hydroxylase. Now EC 1.14.14.142, 8-dimethylallylnaringenin 2-hydroxylase]

[EC 1.14.13.103 created 2007, deleted 2018]

[1.14.13.104 Transferred entry. (+)-menthofuran synthase. Now EC 1.14.14.143, (+)-menthofuran synthase]

[EC 1.14.13.104 created 2008, deleted 2018]

EC 1.14.13.105

Accepted name: monocyclic monoterpene ketone monooxygenase
Reaction:
(1) (-)-menthone + NADPH + H⁺ + O₂ = (4*R*,7*S*)-7-isopropyl-4-methyloxepan-2-one + NADP⁺ + H₂O
(2) dihydrocarvone + NADPH + H⁺ + O₂ = 4-isopropenyl-7-methyloxepan-2-one + NADP⁺ + H₂O
(3) (iso)-dihydrocarvone + NADPH + H⁺ + O₂ = 6-isopropenyl-3-methyloxepan-2-one + NADP⁺ + H₂O
(4a) 1-hydroxymenth-8-en-2-one + NADPH + H⁺ + O₂ = 7-hydroxy-4-isopropenyl-7-methyloxepan-2-one + NADP⁺ + H₂O
(4b) 7-hydroxy-4-isopropenyl-7-methyloxepan-2-one = 3-isopropenyl-6-oxoheptanoate (spontaneous)
Other name(s): 1-hydroxy-2-oxolimonene 1,2-monooxygenase; dihydrocarvone 1,2-monooxygenase; MMKMO
Systematic name: (-)-menthone,NADPH:oxygen oxidoreductase
Comments: A flavoprotein (FAD). This Baeyer-Villiger monooxygenase enzyme from the Gram-positive bacterium *Rhodococcus erythropolis* DCL14 has wide substrate specificity, catalysing the lactonization of a large number of monocyclic monoterpene ketones and substituted cyclohexanones [4588]. Both (1*R*,4*S*)- and (1*S*,4*R*)-1-hydroxymenth-8-en-2-one are metabolized, with the lactone product spontaneously rearranging to form 3-isopropenyl-6-oxoheptanoate [4404].
References: [4404, 4588, 4403]

[EC 1.14.13.105 created 2008]

[1.14.13.106 Transferred entry. epi-isozizaene 5-monooxygenase, now classified as EC 1.14.15.39, epi-isozizaene 5-monooxygenase.]

[EC 1.14.13.106 created 2008, deleted 2021]

EC 1.14.13.107

Accepted name: limonene 1,2-monooxygenase
Reaction: (1) (*S*)-limonene + NAD(P)H + H⁺ + O₂ = 1,2-epoxymenth-8-ene + NAD(P)⁺ + H₂O
(2) (*R*)-limonene + NAD(P)H + H⁺ + O₂ = 1,2-epoxymenth-8-ene + NAD(P)⁺ + H₂O
Systematic name: limonene,NAD(P)H:oxygen oxidoreductase
Comments: A flavoprotein (FAD). Limonene is the most widespread terpene and is formed by more than 300 plants. *Rhodococcus erythropolis* DCL14, a Gram-positive bacterium, is able to grow on both (*S*)-limonene and (*R*)-limonene as the sole source of carbon and energy. NADPH can act instead of NADH, although more slowly. It has not been established if the product formed is optically pure or a mixture of two enantiomers.
References: [4404]

[EC 1.14.13.107 created 2009]

[1.14.13.108 Transferred entry. *abieta-7,13-diene hydroxylase*. Now EC 1.14.14.144, *abieta-7,13-diene hydroxylase*]

[EC 1.14.13.108 created 2009, modified 2012, deleted 2018]

[1.14.13.109 Transferred entry. *abieta-7,13-dien-18-ol hydroxylase*. Now EC 1.14.14.145, *abieta-7,13-dien-18-ol hydroxylase*]

[EC 1.14.13.109 created 2009, modified 2012, deleted 2018]

[1.14.13.110 Transferred entry. *geranylgeraniol 18-hydroxylase*. Now EC 1.14.14.146, *geranylgeraniol 18-hydroxylase*]

[EC 1.14.13.110 created 2009, deleted 2018]

EC 1.14.13.111

Accepted name: methanesulfonate monooxygenase (NADH)
Reaction: methanesulfonate + NADH + H⁺ + O₂ = formaldehyde + NAD⁺ + sulfite + H₂O
Other name(s): mesylate monooxygenase; mesylate,reduced-FMN:oxygen oxidoreductase; MsmABC; methanesulfonic acid monooxygenase; MSA monooxygenase; MSAMO
Systematic name: methanesulfonate,NADH:oxygen oxidoreductase
Comments: A flavoprotein. Methanesulfonate is the simplest of the sulfonates and is a substrate for the growth of certain methylotrophic microorganisms. Compared with EC 1.14.14.5, alkanesulfonate monooxygenase, this enzyme has a restricted substrate range that includes only the short-chain aliphatic sulfonates (methanesulfonate to butanesulfonate) and excludes all larger molecules, such as arylsulfonates [853]. The enzyme from the bacterium *Methylosulfonomonas methylovora* is a multicomponent system comprising a hydroxylase, a reductase (MsmD) and a ferredoxin (MsmC). The hydroxylase has both large (MsmA) and small (MsmB) subunits, with each large subunit containing a Rieske-type [2Fe-2S] cluster. *cf.* EC 1.14.14.34, methanesulfonate monooxygenase (FMNH₂).
References: [853, 1646]

[EC 1.14.13.111 created 2009 as EC 1.14.14.6, transferred 2010 to EC 1.14.13.111, modified 2016]

[1.14.13.112 Transferred entry. *3-epi-6-deoxocathasterone 23-monooxygenase*. Now EC 1.14.14.147, *3-epi-6-deoxocathasterone 23-monooxygenase*]

[EC 1.14.13.112 created 2010, deleted 2018]

EC 1.14.13.113

Accepted name: FAD-dependent urate hydroxylase
Reaction: urate + NADH + H⁺ + O₂ = 5-hydroxyisourate + NAD⁺ + H₂O
Other name(s): HpxO enzyme; FAD-dependent urate oxidase; urate hydroxylase

Systematic name: urate,NADH:oxygen oxidoreductase (5-hydroxyisourate-forming)
Comments: A flavoprotein. The reaction is part of the purine catabolic pathway in the bacterium *Klebsiella pneumoniae*. The enzyme is different from EC 1.7.3.3, factor-independent urate hydroxylase, found in most plants, which produces hydrogen peroxide. The product of the enzyme is a substrate for EC 3.5.2.17, hydroxyisourate hydrolase.
References: [3167]

[EC 1.14.13.113 created 2010]

EC 1.14.13.114

Accepted name: 6-hydroxynicotinate 3-monoxygenase
Reaction: 6-hydroxynicotinate + NADH + H⁺ + O₂ = 2,5-dihydroxypyridine + NAD⁺ + H₂O + CO₂
Other name(s): NicC; 6HNA monoxygenase; HNA-3-monoxygenase
Systematic name: 6-hydroxynicotinate,NADH:oxygen oxidoreductase (3-hydroxylating, decarboxylating)
Comments: A flavoprotein (FAD) [2989]. The reaction is involved in the aerobic catabolism of nicotinic acid.
References: [2989, 1915]

[EC 1.14.13.114 created 2010]

[1.14.13.115] *Transferred entry. angelicin synthase. Now EC 1.14.14.148, angelicin synthase]*

[EC 1.14.13.115 created 2010, deleted 2018]

[1.14.13.116] *Transferred entry. geranylhydroquinone 3-hydroxylase. Now EC 1.14.14.174, geranylhydroquinone 3-hydroxylase.]*

[EC 1.14.13.116 created 2010, deleted 2020]

[1.14.13.117] *Transferred entry. isoleucine N-monoxygenase, Now EC 1.14.14.39, isoleucine N-monoxygenase]*

[EC 1.14.13.117 created 2010, deleted 2017]

[1.14.13.118] *Transferred entry. valine N-monoxygenase. Now EC 1.14.14.38, valine N-monoxygenase]*

[EC 1.14.13.118 created 2010, deleted 2017]

[1.14.13.119] *Transferred entry. 5-epiaristolochene 1,3-dihydroxylase. Now EC 1.14.14.149, 5-epiaristolochene 1,3-dihydroxylase]*

[EC 1.14.13.119 created 2011, deleted 2018]

[1.14.13.120] *Transferred entry. costunolide synthase. Now EC 1.14.14.150, costunolide synthase]*

[EC 1.14.13.120 created 2011, deleted 2018]

[1.14.13.121] *Transferred entry. premnaspirodiene oxygenase. Now EC 1.14.14.151, premnaspirodiene oxygenase]*

[EC 1.14.13.121 created 2011, deleted 2018]

EC 1.14.13.122

Accepted name: chlorophyllide-*a* oxygenase
Reaction: chlorophyllide *a* + 2 O₂ + 2 NADPH + 2 H⁺ = chlorophyllide *b* + 3 H₂O + 2 NADP⁺ (overall reaction)
(1a) chlorophyllide *a* + O₂ + NADPH + H⁺ = 7¹-hydroxychlorophyllide *a* + H₂O + NADP⁺
(1b) 7¹-hydroxychlorophyllide *a* + O₂ + NADPH + H⁺ = chlorophyllide *b* + 2 H₂O + NADP⁺
Other name(s): chlorophyllide *a* oxygenase; chlorophyll-*b* synthase; CAO
Systematic name: chlorophyllide-*a*:oxygen 7¹-oxidoreductase

Comments: Chlorophyll *b* is required for the assembly of stable light-harvesting complexes (LHCs) in the chloroplast of green algae, cyanobacteria and plants [3199, 1024]. Contains a mononuclear iron centre [1024]. The enzyme catalyses two successive hydroxylations at the 7-methyl group of chlorophyllide *a*. The second step yields the aldehyde hydrate, which loses H₂O spontaneously to form chlorophyllide *b* [3199]. Chlorophyll *a* and protochlorophyllide *a* are not substrates [3199].

References: [1064, 3199, 1024, 3355]

[EC 1.14.13.122 created 2006 as EC 1.13.12.14, transferred 2011 to EC 1.14.13.122, modified 2011]

[1.14.13.123 *Transferred entry. germacrene A hydroxylase. Now EC 1.14.14.95, germacrene A hydroxylase*]

[EC 1.14.13.123 created 2011, deleted 2018]

[1.14.13.124 *Transferred entry. phenylalanine N-monooxygenase, now classified as EC 1.14.14.40, phenylalanine N-monooxygenase*]

[EC 1.14.13.124 created 2011, deleted 2017]

[1.14.13.125 *Transferred entry. tryptophan N-monooxygenase. Now EC 1.14.14.156, tryptophan N-monooxygenase*]

[EC 1.14.13.125 created 2011, deleted 2018]

[1.14.13.126 *Transferred entry. vitamin D₃ 24-hydroxylase. Now EC 1.14.15.16, vitamin D₃ 24-hydroxylase*]

[EC 1.14.13.126 created 2011, deleted 2016]

EC 1.14.13.127

Accepted name: 3-(3-hydroxyphenyl)propanoate hydroxylase

Reaction: (1) 3-(3-hydroxyphenyl)propanoate + NADH + H⁺ + O₂ = 3-(2,3-dihydroxyphenyl)propanoate + H₂O + NAD⁺
(2) (2*E*)-3-(3-hydroxyphenyl)prop-2-enoate + NADH + H⁺ + O₂ = (2*E*)-3-(2,3-dihydroxyphenyl)prop-2-enoate + H₂O + NAD⁺

Other name(s): *mhpA* (gene name)

Systematic name: 3-(3-hydroxyphenyl)propanoate,NADH:oxygen oxidoreductase (2-hydroxylating)

Comments: A flavoprotein (FAD). This enzyme participates in a meta-cleavage pathway employed by the bacterium *Escherichia coli* for the degradation of various phenylpropanoid compounds.

References: [497, 498, 1106, 900]

[EC 1.14.13.127 created 2011]

EC 1.14.13.128

Accepted name: 7-methylxanthine demethylase

Reaction: 7-methylxanthine + O₂ + NAD(P)H + H⁺ = xanthine + NAD(P)⁺ + H₂O + formaldehyde

Other name(s): *ndmC* (gene name)

Systematic name: 7-methylxanthine:oxygen oxidoreductase (demethylating)

Comments: A non-heme iron oxygenase. The enzyme from the bacterium *Pseudomonas putida* prefers NADH over NADPH. The enzyme is specific for 7-methylxanthine [4113]. Forms part of the caffeine degradation pathway.

References: [4114, 4113]

[EC 1.14.13.128 created 2011]

[1.14.13.129 *Transferred entry. β-carotene 3-hydroxylase. Now EC 1.14.15.24, β-carotene 3-hydroxylase.*]

[EC 1.14.13.129 created 2011, deleted 2017]

EC 1.14.13.130

Accepted name: pyrrole-2-carboxylate monooxygenase
Reaction: pyrrole-2-carboxylate + NADH + H⁺ + O₂ = 5-hydroxypyrrole-2-carboxylate + NAD⁺ + H₂O
Other name(s): pyrrole-2-carboxylate oxygenase
Systematic name: pyrrole-2-carboxylate,NADH:oxygen oxidoreductase (5-hydroxylating)
Comments: A flavoprotein (FAD). The enzyme initiates the degradation of pyrrole-2-carboxylate.
References: [1731, 262]

[EC 1.14.13.130 created 2011]

EC 1.14.13.131

Accepted name: dissimilatory dimethyl sulfide monooxygenase
Reaction: dimethyl sulfide + O₂ + NADH + H⁺ = methanethiol + formaldehyde + NAD⁺ + H₂O
Other name(s): *dmoAB* (gene names); dimethyl sulfide C-monooxygenase; dimethylsulfide monooxygenase (ambiguous); dimethyl sulfide monooxygenase (ambiguous)
Systematic name: dimethyl sulfide,NADH:oxygen oxidoreductase
Comments: The enzyme participates exclusively in sulfur dissimilation. It has lower activity with diethyl sulfide and other short-chain alkyl methyl sulfides. Its activity is stimulated by combined addition of FMN, and, after depletion of cations, of Mg²⁺ and Fe²⁺. The enzymes from bacteria of the *Hyphomicrobium* genus are a two component system that includes an FMN-dependent reductase subunit and a monooxygenase subunit.
References: [387, 365]

[EC 1.14.13.131 created 2011]

[1.14.13.132 Transferred entry. squalene monooxygenase. Now EC 1.14.14.17, squalene monooxygenase]

[EC 1.14.13.132 created 1961 as EC 1.99.1.13, transferred 1965 to EC 1.14.1.3, part transferred 1972 to EC 1.14.99.7, transferred 2011 to EC 1.14.13.132, deleted 2015]

[1.14.13.133 Transferred entry. pentalenene oxygenase. Now EC 1.14.15.32, pentalenene oxygenase]

[EC 1.14.13.133 created 2011, deleted 2018]

[1.14.13.134 Transferred entry. β-amyrin 11-oxidase. Now EC 1.14.14.152, β-amyrin 11-oxidase]

[EC 1.14.13.134 created 2011, deleted 2018]

EC 1.14.13.135

Accepted name: 1-hydroxy-2-naphthoate hydroxylase
Reaction: 1-hydroxy-2-naphthoate + NAD(P)H + H⁺ + O₂ = 1,2-dihydroxynaphthalene + NAD(P)⁺ + H₂O + CO₂
Other name(s): 1-hydroxy-2-naphthoic acid hydroxylase
Systematic name: 1-hydroxy-2-naphthoate,NAD(P)H:oxygen oxidoreductase (2-hydroxylating, decarboxylating)
Comments: The enzyme is involved in the catabolic pathway for the degradation of chrysene in some bacteria [3028].
References: [891, 3028]

[EC 1.14.13.135 created 2011]

[1.14.13.136 Transferred entry. 2-hydroxyisoflavanone synthase. Now EC 1.14.14.87, 2-hydroxyisoflavanone synthase]

[EC 1.14.13.136 created 2011, modified 2013, deleted 2018]

[1.14.13.137 Transferred entry. indole-2-monooxygenase. Now EC 1.14.14.153, indole-2-monooxygenase]

[EC 1.14.13.137 created 2012, deleted 2018]

[1.14.13.138 Transferred entry. indolin-2-one monooxygenase. Now EC 1.14.14.157, indolin-2-one monooxygenase]

[EC 1.14.13.138 created 2012, deleted 2018]

[1.14.13.139 Transferred entry. 3-hydroxyindolin-2-one monooxygenase. Now EC 1.14.14.109, 3-hydroxyindolin-2-one monooxygenase]

[EC 1.14.13.139 created 2012, deleted 2018]

[1.14.13.140 Transferred entry. 2-hydroxy-1,4-benzoxazin-3-one monooxygenase. Now EC 1.14.14.110, 2-hydroxy-1,4-benzoxazin-3-one monooxygenase.]

[EC 1.14.13.140 created 2012, deleted 2018]

[1.14.13.141 Transferred entry. cholest-4-en-3-one 26-monooxygenase [(25S)-3-oxocholest-4-en-26-oate forming]. Now EC 1.14.15.29, cholest-4-en-3-one 26-monooxygenase [(25S)-3-oxocholest-4-en-26-oate forming].]

[EC 1.14.13.141 created 2012, modified 2016, deleted 2018]

[1.14.13.142 Transferred entry. 3-ketosteroid 9 α -monooxygenase. Now EC 1.14.15.30, 3-ketosteroid 9 α -monooxygenase]

[EC 1.14.13.142 created 2012, deleted 2018]

[1.14.13.143 Transferred entry. ent-isokaurene C2-hydroxylase. Now EC 1.14.14.76 ent-isokaurene C2/C3-hydroxylase]

[EC 1.14.13.143 created 2012, deleted 2018]

[1.14.13.144 Transferred entry. 9 β -pimara-7,15-diene oxidase. Now EC 1.14.14.111, 9 β -pimara-7,15-diene oxidase.]

[EC 1.14.13.144 created 2012, deleted 2018]

[1.14.13.145 Transferred entry. ent-cassa-12,15-diene 11-hydroxylase. Now EC 1.14.14.112, ent-cassa-12,15-diene 11-hydroxylase.]

[EC 1.14.13.145 created 2012, deleted 2018]

EC 1.14.13.146

Accepted name: taxoid 14 β -hydroxylase
Reaction: 10 β -hydroxytaxa-4(20),11-dien-5 α -yl acetate + O₂ + NADPH + H⁺ = 10 β ,14 β -dihydroxytaxa-4(20),11-dien-5 α -yl acetate + NADP⁺ + H₂O
Systematic name: 10 β -hydroxytaxa-4(20),11-dien-5 α -yl-acetate,NADPH:oxygen 14-oxidoreductase
Comments: Requires cytochrome P450. From the yew *Taxus cuspidata*. Also acts on taxa-4(20),11-dien-5 α -yl acetate.
References: [1899]

[EC 1.14.13.146 created 2012]

[1.14.13.147 Transferred entry. taxoid 7 β -hydroxylase. Now EC 1.14.14.182, taxoid 7 β -hydroxylase]

[EC 1.14.13.147 created 2012, deleted 2022]

EC 1.14.13.148

Accepted name: trimethylamine monooxygenase
Reaction: *N,N,N*-trimethylamine + NADPH + H⁺ + O₂ = *N,N,N*-trimethylamine *N*-oxide + NADP⁺ + H₂O
Other name(s): flavin-containing monooxygenase 3; FMO3; tmm (gene name)
Systematic name: *N,N,N*-trimethylamine,NADPH:oxygen oxidoreductase (*N*-oxide-forming)
Comments: A flavoprotein. The bacterial enzyme enables bacteria to use trimethylamine as the sole source of carbon and energy [2353, 638]. The mammalian enzyme is involved in detoxification of trimethylamine. Mutations in the human enzyme cause the inheritable disease known as trimethylaminuria (fish odor syndrome) [939, 4325].
References: [2353, 939, 4325, 638]

[EC 1.14.13.148 created 2012]

EC 1.14.13.149

Accepted name: phenylacetyl-CoA 1,2-epoxidase
Reaction: phenylacetyl-CoA + NADPH + H⁺ + O₂ = 2-(1,2-epoxy-1,2-dihydrophenyl)acetyl-CoA + NADP⁺ + H₂O
Other name(s): ring 1,2-phenylacetyl-CoA epoxidase; phenylacetyl-CoA monooxygenase; PaaAC; PaaABC(D)E
Systematic name: phenylacetyl-CoA:oxygen oxidoreductase (1,2-epoxidizing)
Comments: Part of the aerobic pathway of phenylacetate catabolism in *Escherichia coli* and *Pseudomonas putida*.
References: [4250, 1414, 1413]

[EC 1.14.13.149 created 2012]

[1.14.13.150 Transferred entry. *α*-humulene 10-hydroxylase. Now EC 1.14.14.113, *α*-humulene 10-hydroxylase.]

[EC 1.14.13.150 created 2012, deleted 2018]

[1.14.13.151 Transferred entry. linalool 8-monooxygenase. Now EC 1.14.14.84, linalool 8-monooxygenase]

[EC 1.14.13.151 created 1989 as EC 1.14.99.28, transferred 2012 to EC 1.14.13.151, deleted 2018]

[1.14.13.152 Transferred entry. geraniol 8-hydroxylase. Now EC 1.14.14.83, geraniol 8-hydroxylase]

[EC 1.14.13.152 created 2012, deleted 2018]

EC 1.14.13.153

Accepted name: (+)-sabinene 3-hydroxylase
Reaction: (+)-sabinene + NADPH + H⁺ + O₂ = (+)-*cis*-sabinol + NADP⁺ + H₂O
Systematic name: (+)-sabinene,NADPH:oxygen oxidoreductase (3-hydroxylating)
Comments: Requires cytochrome *P*-450. The enzyme has been characterized from *Salvia officinalis* (sage).
References: [1995]

[EC 1.14.13.153 created 2012]

EC 1.14.13.154

Accepted name: erythromycin 12-hydroxylase
Reaction: erythromycin D + NADPH + H⁺ + O₂ = erythromycin C + NADP⁺ + H₂O
Other name(s): EryK
Systematic name: erythromycin-D,NADPH:oxygen oxidoreductase (12-hydroxylating)
Comments: The enzyme is responsible for the C-12 hydroxylation of the macrolactone ring, one of the last steps in erythromycin biosynthesis. It shows 1200-1900-fold preference for erythromycin D over the alternative substrate erythromycin B [2332].
References: [2332, 3680, 2868]

[EC 1.14.13.154 created 2012]

EC 1.14.13.155

Accepted name: *α*-pinene monooxygenase
Reaction: (–)-*α*-pinene + NADH + H⁺ + O₂ = *α*-pinene oxide + NAD⁺ + H₂O
Systematic name: (–)-*α*-pinene,NADH:oxygen oxidoreductase
Comments: Involved in the catabolism of *α*-pinene.
References: [717]

[EC 1.14.13.155 created 2012]

[1.14.13.156 Transferred entry. 1,8-cineole 2-endo-monooxygenase. Now EC 1.14.14.133, 1,8-cineole 2-endo-monooxygenase]

[EC 1.14.13.156 created 2012, deleted 2018]

[1.14.13.157 Transferred entry. 1,8-cineole 2-*exo*-monooxygenase. Now EC 1.14.14.56, 1,8-cineole 2-*exo*-monooxygenase]

[EC 1.14.13.157 created 2012, deleted 2017]

[1.14.13.158 Transferred entry. amorpho-4,11-diene 12-monooxygenase. Now EC 1.14.14.114, amorpho-4,11-diene 12-monooxygenase.]

[EC 1.14.13.158 created 2012, deleted 2018]

[1.14.13.159 Transferred entry. vitamin D 25-hydroxylase. Now EC 1.14.14.24, vitamin D 25-hydroxylase]

[EC 1.14.13.159 created 2012, deleted 2016]

EC 1.14.13.160

Accepted name: (2,2,3-trimethyl-5-oxocyclopent-3-enyl)acetyl-CoA 1,5-monooxygenase
Reaction: [(1R)-2,2,3-trimethyl-5-oxocyclopent-3-enyl]acetyl-CoA + O₂ + NADPH + H⁺ = [(2R)-3,3,4-trimethyl-6-oxo-3,6-dihydro-1H-pyran-2-yl]acetyl-CoA + NADP⁺ + H₂O
Other name(s): 2-oxo-Δ³-4,5,5-trimethylcyclopentenylacetyl-CoA monooxygenase; 2-oxo-Δ³-4,5,5-trimethylcyclopentenylacetyl-CoA 1,2-monooxygenase; OTEMO
Systematic name: [(1R)-2,2,3-trimethyl-5-oxocyclopent-3-enyl]acetyl-CoA,NADPH:oxygen oxidoreductase (1,5-lactonizing)
Comments: A FAD dependent enzyme isolated from *Pseudomonas putida*. Forms part of the catabolism pathway of camphor. It acts on the CoA ester in preference to the free acid.
References: [3215, 2412, 1969]

[EC 1.14.13.160 created 2012]

EC 1.14.13.161

Accepted name: (+)-camphor 6-*exo*-hydroxylase
Reaction: (+)-camphor + NADPH + H⁺ + O₂ = (+)-6-*exo*-hydroxycamphor + NADP⁺ + H₂O
Other name(s): (+)-camphor 6-hydroxylase
Systematic name: (+)-camphor,NADPH:oxygen oxidoreductase (6-*exo*-hydroxylating)
Comments: A cytochrome *P*-450 monooxygenase isolated from *Salvia officinalis* (sage). Involved in the catabolism of camphor in senescent tissue.
References: [1231, 1229]

[EC 1.14.13.161 created 2012]

[1.14.13.162 Transferred entry. 2,5-diketocamphane 1,2-monooxygenase. Now EC 1.14.14.108, 2,5-diketocamphane 1,2-monooxygenase]

[EC 1.14.13.162 created 1972 as EC 1.14.15.2, transferred 2012 to EC 1.14.13.162, deleted 2018]

EC 1.14.13.163

Accepted name: 6-hydroxy-3-succinoylpyridine 3-monooxygenase
Reaction: 4-(6-hydroxypyridin-3-yl)-4-oxobutanoate + 2 NADH + 2 H⁺ + O₂ = 2,5-dihydroxypyridine + succinate semialdehyde + 2 NAD⁺ + H₂O
Other name(s): 6-hydroxy-3-succinoylpyridine hydroxylase; *hspA* (gene name); *hspB* (gene name)
Systematic name: 4-(6-hydroxypyridin-3-yl)-4-oxobutanoate,NADH:oxygen oxidoreductase (3-hydroxylating, succinate semialdehyde releasing)
Comments: The enzyme catalyses a reaction in the nicotine degradation pathway of *Pseudomonas* species. One of the enzymes from the soil bacterium *Pseudomonas putida* S16 contains an FAD cofactor [4203].
References: [4202, 4203]

[EC 1.14.13.163 created 2012]

[1.14.13.164 Transferred entry. carotenoid isomeroxygenase. The enzyme was discovered at the public-review stage to have been misclassified and so was withdrawn. See EC 1.13.11.65, carotenoid isomeroxygenase]

[EC 1.14.13.164 created 2012, deleted 2012]

[1.14.13.165 Transferred entry. nitric-oxide synthase [NAD(P)H]. Now classified as EC 1.14.14.47, nitric-oxide synthase (flavodoxin)]

[EC 1.14.13.165 created 2012, deleted 2017]

EC 1.14.13.166

Accepted name: 4-nitrocatechol 4-monooxygenase
Reaction: 4-nitrocatechol + NAD(P)H + H⁺ + O₂ = 2-hydroxy-1,4-benzoquinone + nitrite + NAD(P)⁺ + H₂O
Systematic name: 4-nitrocatechol,NAD(P)H:oxygen 4-oxidoreductase (4-hydroxylating, nitrite-forming)
Comments: Contains FAD. The enzyme catalyses the oxidation of 4-nitrocatechol with the concomitant removal of the nitro group as nitrite. Forms a two-component system with a flavoprotein reductase [1967]. The enzymes from the bacteria *Lysinibacillus sphaericus* JS905 and *Rhodococcus* sp. strain PN1 were shown to also catalyse EC 1.14.13.29, 4-nitrophenol 2-monooxygenase [1967, 2133] while the enzyme from *Pseudomonas* sp. WBC-3 was shown to also catalyse EC 1.14.13.167, 4-nitrophenol 4-monooxygenase [4878].
References: [1967, 2133, 4878]

[EC 1.14.13.166 created 2012]

EC 1.14.13.167

Accepted name: 4-nitrophenol 4-monooxygenase
Reaction: 4-nitrophenol + NADPH + H⁺ + O₂ = 1,4-benzoquinone + nitrite + NADP⁺ + H₂O
Other name(s): *pnpA* (gene name); *pdca* (gene name)
Systematic name: 4-nitrophenol,NAD(P)H:oxygen 4-oxidoreductase (4-hydroxylating, nitrite-forming)
Comments: Contains FAD. The enzyme catalyses the first step in a degradation pathway for 4-nitrophenol, the oxidation of 4-nitrophenol at position 4 with the concomitant removal of the nitro group as nitrite. The enzyme from the bacterium *Pseudomonas* sp. strain WBC-3 also catalyses EC 1.14.13.166, 4-nitrocatechol 4-monooxygenase.
References: [4878]

[EC 1.14.13.167 created 2012]

EC 1.14.13.168

Accepted name: indole-3-pyruvate monooxygenase
Reaction: (indol-3-yl)pyruvate + NADPH + H⁺ + O₂ = (indol-3-yl)acetate + NADP⁺ + H₂O + CO₂
Other name(s): YUC2 (gene name); *spi1* (gene name)
Systematic name: indole-3-pyruvate,NADPH:oxygen oxidoreductase (1-hydroxylating, decarboxylating)
Comments: This plant enzyme, along with EC 2.6.1.99 L-tryptophan—pyruvate aminotransferase, is responsible for the biosynthesis of the plant hormone indole-3-acetate from L-tryptophan.
References: [2681, 4906]

[EC 1.14.13.168 created 2012]

[1.14.13.169 Transferred entry. sphinganine C4-monooxygenase. Now EC 1.14.18.5, sphingolipid C4-monooxygenase]

[EC 1.14.13.169 created 2012, deleted 2015]

EC 1.14.13.170

Accepted name: pentalenolactone D synthase
Reaction: 1-deoxy-11-oxopentalenate + NADPH + H⁺ + O₂ = pentalenolactone D + NADP⁺ + H₂O
Other name(s): *penE* (gene name); *pntE* (gene name)
Systematic name: 1-deoxy-11-oxopentalenate,NADH:oxygen oxidoreductase (pentalenolactone-D-forming)

Comments: A FAD-dependent oxygenase. Isolated from the bacteria *Streptomyces exfoliatus* and *Streptomyces arenae*. The ketone undergoes a biological Baeyer-Villiger reaction. Part of the pathway of pentalenolactone biosynthesis.

References: [3801]

[EC 1.14.13.170 created 2012]

EC 1.14.13.171

Accepted name: neopentalenolactone D synthase

Reaction: 1-deoxy-11-oxopentalenate + NADPH + H⁺ + O₂ = neopentalenolactone D + NADP⁺ + H₂O

Other name(s): *ptlE* (gene name)

Systematic name: 1-deoxy-11-oxopentalenate,NADH:oxygen oxidoreductase (neopentalenolactone-D-forming)

Comments: A FAD-dependent oxygenase. Isolated from the bacterium *Streptomyces avermitilis*. The ketone undergoes a biological Baeyer-Villiger reaction.

References: [3801]

[EC 1.14.13.171 created 2012]

EC 1.14.13.172

Accepted name: salicylate 5-hydroxylase

Reaction: salicylate + NADH + H⁺ + O₂ = 2,5-dihydroxybenzoate + NAD⁺ + H₂O

Other name(s): *nagG* (gene name); *nagH* (gene name)

Systematic name: salicylate,NADH:oxygen oxidoreductase (5-hydroxylating)

Comments: This enzyme, which was characterized from the bacterium *Ralstonia* sp. U2, comprises a multi-component system, containing a reductase that is an iron-sulfur flavoprotein (FAD; EC 1.18.1.7, ferredoxin—NAD(P)⁺ reductase), an iron-sulfur oxygenase, and ferredoxin.

References: [1201]

[EC 1.14.13.172 created 2013]

[1.14.13.173 Transferred entry. 11-oxo-β-amyrin 30-oxidase. Now EC 1.14.14.115, 11-oxo-β-amyrin 30-oxidase.]

[EC 1.14.13.173 created 2013, deleted 2018]

[1.14.13.174 Transferred entry. averantin hydroxylase. Now EC 1.14.14.116, averantin hydroxylase]

[EC 1.14.13.174 created 2013, deleted 2018]

[1.14.13.175 Transferred entry. aflatoxin B synthase. Now EC 1.14.14.117, aflatoxin B synthase]

[EC 1.14.13.175 created 2013, deleted 2018]

[1.14.13.176 Transferred entry. tryprostatin B 6-hydroxylase. Now EC 1.14.14.118, tryprostatin B 6-hydroxylase]

[EC 1.14.13.176 created 2013, deleted 2018]

[1.14.13.177 Transferred entry. fumitremorgin C monooxygenase. Now EC 1.14.14.119, fumitremorgin C monooxygenase]

[EC 1.14.13.177 created 2013, deleted 2018]

EC 1.14.13.178

Accepted name: methylxanthine N¹-demethylase

Reaction: (1) caffeine + O₂ + NAD(P)H + H⁺ = theobromine + NAD(P)⁺ + H₂O + formaldehyde

(2) theophylline + O₂ + NAD(P)H + H⁺ = 3-methylxanthine + NAD(P)⁺ + H₂O + formaldehyde

(3) paraxanthine + O₂ + NAD(P)H + H⁺ = 7-methylxanthine + NAD(P)⁺ + H₂O + formaldehyde

Other name(s): *ndmA* (gene name)

Systematic name: caffeine:oxygen oxidoreductase (N¹-demethylating)

Comments: A non-heme iron oxygenase. The enzyme from the bacterium *Pseudomonas putida* shares an NAD(P)H-FMN reductase subunit with EC 1.14.13.179, methylxanthine *N*³-demethylase, and has a 5-fold higher activity with NADH than with NADPH [4113]. Also demethylate 1-methylxanthine with lower efficiency. Forms part of the degradation pathway of methylxanthines.

References: [4114, 4113]

[EC 1.14.13.178 created 2013]

EC 1.14.13.179

Accepted name: methylxanthine *N*³-demethylase

Reaction: (1) theobromine + O₂ + NAD(P)H + H⁺ = 7-methylxanthine + NAD(P)⁺ + H₂O + formaldehyde
(2) 3-methylxanthine + O₂ + NAD(P)H + H⁺ = xanthine + NAD(P)⁺ + H₂O + formaldehyde

Other name(s): *ndmB* (gene name)

Systematic name: theobromine:oxygen oxidoreductase (*N*³-demethylating)

Comments: A non-heme iron oxygenase. The enzyme from the bacterium *Pseudomonas putida* shares an NAD(P)H-FMN reductase subunit with EC 1.14.13.178, methylxanthine *N*¹-demethylase, and has higher activity with NADH than with NADPH [4114]. Also demethylates caffeine and theophylline with lower efficiency. Forms part of the degradation pathway of methylxanthines.

References: [4114, 4113]

[EC 1.14.13.179 created 2013]

EC 1.14.13.180

Accepted name: aklavinone 12-hydroxylase

Reaction: aklavinone + NADPH + H⁺ + O₂ = ε-rhodomyconone + NADP⁺ + H₂O

Other name(s): DnrF; RdmE; aklavinone 11-hydroxylase (incorrect)

Systematic name: aklavinone,NADPH:oxygen oxidoreductase (12-hydroxylating)

Comments: The enzymes from the Gram-positive bacteria *Streptomyces peucetius* and *Streptomyces purpurascens* participate in the biosynthesis of daunorubicin, doxorubicin and rhodomycins. The enzyme from *Streptomyces purpurascens* is an FAD monooxygenase.

References: [1122, 3070]

[EC 1.14.13.180 created 2013]

EC 1.14.13.181

Accepted name: 13-deoxydaunorubicin hydroxylase

Reaction: (1) 13-deoxydaunorubicin + NADPH + H⁺ + O₂ = 13-dihydrodaunorubicin + NADP⁺ + H₂O
(2) 13-dihydrodaunorubicin + NADPH + H⁺ + O₂ = daunorubicin + NADP⁺ + 2 H₂O

Other name(s): DoxA

Systematic name: 13-deoxydaunorubicin,NADPH:oxygen oxidoreductase (13-hydroxylating)

Comments: The enzymes from the Gram-positive bacteria *Streptomyces* sp. C5 and *Streptomyces peucetius* show broad substrate specificity for structures based on an anthracycline aglycone, but have a strong preference for 4-methoxy anthracycline intermediates (13-deoxydaunorubicin and 13-dihydrodaunorubicin) over their 4-hydroxy analogues (13-deoxycarminomycin and 13-dihydrocarminomycin), as well as a preference for substrates hydroxylated at the C-13 rather than the C-14 position.

References: [4493, 904]

[EC 1.14.13.181 created 2013]

EC 1.14.13.182

Accepted name: 2-heptyl-3-hydroxy-4(1*H*)-quinolone synthase

Reaction: 2-heptyl-4(1*H*)-quinolone + NADH + H⁺ + O₂ = 2-heptyl-3-hydroxy-4(1*H*)-quinolone + NAD⁺ + H₂O

Other name(s): PqsH; 2-heptyl-3,4-dihydroxyquinoline synthase

Systematic name: 2-heptyl-4(1*H*)-quinolone,NADH:oxygen oxidoreductase (3-hydroxylating)

Comments: The enzyme from the bacterium *Pseudomonas aeruginosa* catalyses the terminal step in biosynthesis of the signal molecule 2-heptyl-3,4-dihydroxyquinoline that plays a role in regulation of virulence genes.

References: [3713]

[EC 1.14.13.182 created 2013]

[1.14.13.183 Transferred entry. dammarenediol 12-hydroxylase. Now EC 1.14.14.120, dammarenediol 12-hydroxylase]

[EC 1.14.13.183 created 2013, deleted 2018]

[1.14.13.184 Transferred entry. protopanaxadiol 6-hydroxylase. Now EC 1.14.14.121, protopanaxadiol 6-hydroxylase]

[EC 1.14.13.184 created 2013, deleted 2018]

[1.14.13.185 Transferred entry. pikromycin synthase. Now EC 1.14.15.33, pikromycin synthase]

[EC 1.14.13.185 created 2014, deleted 2018]

[1.14.13.186 Transferred entry. 20-oxo-5-*O*-mycaminosyltylactone 23-monoxygenase. Now EC 1.14.15.34, 20-oxo-5-*O*-mycaminosyltylactone 23-monoxygenase]

[EC 1.14.13.186 created 2014, deleted 2018]

EC 1.14.13.187

Accepted name: L-evernosamine nitrososynthase

Reaction: dTDP-β-L-evernosamine + 2 NADPH + 2 H⁺ + 2 O₂ = dTDP-2,3,6-trideoxy-3-*C*-methyl-4-*O*-methyl-3-nitroso-β-L-*arabino*-hexopyranose + 2 NADP⁺ + 3 H₂O (overall reaction)

(1a) dTDP-β-L-evernosamine + NADPH + H⁺ + O₂ = dTDP-*N*-hydroxy-β-L-evernosamine + NADP⁺ + H₂O

(1b) dTDP-*N*-hydroxy-β-L-evernosamine + NADPH + H⁺ + O₂ = dTDP-2,3,6-trideoxy-3-*C*-methyl-4-*O*-methyl-3-nitroso-β-L-*arabino*-hexopyranose + NADP⁺ + 2 H₂O

Systematic name: dTDP-β-L-evernosamine,NADPH:oxygen oxidoreductase (*N*-hydroxylating)

Comments: Requires FAD. Isolated from the bacterium *Micromonospora carbonacea* var. *africana*. The nitroso group is probably spontaneously oxidized to a nitro group giving dTDP-β-L-evernitrose, which is involved in the biosynthesis of the antibiotic everninomycin. The reaction was studied using dTDP-β-L-4-*epi*-vancosamine (dTDP-4-*O*-desmethyl-β-L-evernitrosamine).

References: [1748, 4442]

[EC 1.14.13.187 created 2014]

[1.14.13.188 Transferred entry. 6-deoxyerythronolide B hydroxylase. Now EC 1.14.15.35, 6-deoxyerythronolide B hydroxylase]

[EC 1.14.13.188 created 2014, deleted 2018]

EC 1.14.13.189

Accepted name: 5-methyl-1-naphthoate 3-hydroxylase

Reaction: 5-methyl-1-naphthoate + NADPH + H⁺ + O₂ = 3-hydroxy-5-methyl-1-naphthoate + NADP⁺ + H₂O

Other name(s): AziB1

Systematic name: 5-methyl-1-naphthoate,NADPH:oxygen oxidoreductase (3-hydroxylating)

Comments: The enzyme from the bacterium *Streptomyces sahachiroi* is involved in the biosynthesis of 3-methoxy-5-methyl-1-naphthoate, a component of the antitumor antibiotic azinomycin B.

References: [919]

[EC 1.14.13.189 created 2014]

[1.14.13.190 Transferred entry. *ferruginol synthase*. Now EC 1.14.14.175, *ferruginol synthase*]

[EC 1.14.13.190 created 2014, modified 2015, deleted 2020]

[1.14.13.191 Transferred entry. *ent-sandaracopimaradiene 3-hydroxylase*. Now EC 1.14.14.70, *ent-sandaracopimaradiene 3-hydroxylase*]

[EC 1.14.13.191 created 2014, deleted 2018]

[1.14.13.192 Transferred entry. *oryzalexin E synthase*. Now EC 1.14.14.122, *oryzalexin E synthase*]

[EC 1.14.13.192 created 2014, deleted 2018]

[1.14.13.193 Transferred entry. *oryzalexin D synthase*. Now EC 1.14.14.123, *oryzalexin D synthase*]

[EC 1.14.13.193 created 2014, deleted 2018]

[1.14.13.194 Transferred entry. *phyloquinone ω -hydroxylase*. Now EC 1.14.14.78, *phyloquinone ω -hydroxylase*]

[EC 1.14.13.194 created 2014, deleted 2018]

EC 1.14.13.195

- Accepted name:** L-ornithine N^5 -monooxygenase (NADPH)
Reaction: L-ornithine + NADPH + H^+ + O_2 = N^5 -hydroxy-L-ornithine + $NADP^+$ + H_2O
Other name(s): CchB; ornithine hydroxylase; EtcB; PvdA; Af-OMO; *dffA* (gene name)
Systematic name: L-ornithine,NADPH:oxygen oxidoreductase (N^5 -hydroxylating)
Comments: A flavoprotein (FAD). The enzyme is involved in biosynthesis of N^5 -hydroxy-L-ornithine, N^5 -formyl- N^5 -hydroxy-L-ornithine or N^5 -acetyl- N^5 -hydroxy-L-ornithine. These nonproteinogenic amino acids are building blocks of siderophores produced by some bacteria (e.g. *Streptomyces coelicolor*, *Saccharopolyspora erythraea* and *Pseudomonas aeruginosa*). The enzyme is specific for NADPH. *cf.* EC 1.14.13.196, L-ornithine N^5 -monooxygenase [NAD(P)H].
References: [1293, 2769, 3341, 3534]

[EC 1.14.13.195 created 2014]

EC 1.14.13.196

- Accepted name:** L-ornithine N^5 -monooxygenase [NAD(P)H]
Reaction: L-ornithine + NAD(P)H + H^+ + O_2 = N^5 -hydroxy-L-ornithine + $NAD(P)^+$ + H_2O
Other name(s): SidA (ambiguous)
Systematic name: L-ornithine,NAD(P)H:oxygen oxidoreductase (N^5 -hydroxylating)
Comments: A flavoprotein (FAD). The enzyme from the pathogenic fungus *Aspergillus fumigatus* catalyses a step in the biosynthesis of the siderophores triacetylfusarinine and desferri-ferricrocin, while the enzyme from the bacterium *Kutzneria* sp. 744 is involved in the biosynthesis of piperazate, a building block of the kutzneride family of antifungal antibiotics. Activity of the fungal enzyme is higher with NADPH, due to the fact that following the reduction of the flavin, $NADP^+$ (but not NAD^+) stabilizes the C4a-hydroperoxyflavin intermediate that oxidizes the substrate [3566]. *cf.* EC 1.14.13.195, L-ornithine N^5 -monooxygenase (NADPH).
References: [670, 1161, 3566, 3049]

[EC 1.14.13.196 created 2014]

[1.14.13.197 Transferred entry. *dihydromonacolin L hydroxylase*. Now EC 1.14.14.124, *dihydromonacolin L hydroxylase*]

[EC 1.14.13.197 created 2014, deleted 2018]

[1.14.13.198 Transferred entry. *monacolin L hydroxylase*. Now EC 1.14.14.125, *monacolin L hydroxylase*]

[EC 1.14.13.198 created 2014, deleted 2018]

[1.14.13.199 Transferred entry. docosahexaenoic acid ω -hydroxylase. Now EC 1.14.14.79, docosahexaenoic acid ω -hydroxylase]

[EC 1.14.13.199 created 2014, deleted 2018]

EC 1.14.13.200

Accepted name: tetracenomycin A2 monooxygenase-dioxygenase
Reaction: tetracenomycin A2 + 2 O₂ + 2 NAD(P)H + 2 H⁺ = tetracenomycin C + 2 NAD(P)⁺ + H₂O
Other name(s): TcmG; ElmG; tetracenomycin A2,NAD(P)H:O₂ oxidoreductase (tetracenomycin C forming)
Systematic name: tetracenomycin A2,NAD(P)H:oxygen oxidoreductase (tetracenomycin-C-forming)
Comments: Isolated from the bacterium *Streptomyces glaucescens*. The enzyme was also isolated from the bacterium *Streptomyces olivaceus*, where it acts on 8-demethyltetracenomycin A2 (tetracenomycin B2) as part of elloramycin biosynthesis. The reaction involves a monooxygenase reaction which is followed by a dioxygenase reaction giving a gem-diol and an epoxide. Water opens the epoxide giving two hydroxy groups. The gem-diol eliminates water to give a ketone which is then reduced to a hydroxy group.
References: [3839, 3423, 324]

[EC 1.14.13.200 created 2014]

[1.14.13.201 Transferred entry. β -amyrin 28-monooxygenase. Now EC 1.14.14.126, β -amyrin 28-monooxygenase]

[EC 1.14.13.201 created 2015, deleted 2018]

[1.14.13.202 Transferred entry. methyl farnesoate epoxidase. Now EC 1.14.14.127, methyl farnesoate epoxidase]

[EC 1.14.13.202 created 2015, deleted 2018]

[1.14.13.203 Transferred entry. farnesoate epoxidase. Now EC 1.14.14.128, farnesoate epoxidase]

[EC 1.14.13.203 created 2015, deleted 2018]

[1.14.13.204 Transferred entry. long-chain acyl-CoA ω -monooxygenase. Now EC 1.14.14.129, long-chain acyl-CoA ω -monooxygenase]

[EC 1.14.13.204 created 2015, deleted 2018]

[1.14.13.205 Transferred entry. long-chain fatty acid ω -monooxygenase. Now EC 1.14.14.80, long-chain fatty acid ω -monooxygenase]

[EC 1.14.13.205 created 2015, deleted 2018]

[1.14.13.206 Transferred entry. laurate 7-monooxygenase. Now EC 1.14.14.130, laurate 7-monooxygenase]

[EC 1.14.13.206 created 2015, deleted 2018]

[1.14.13.207 Transferred entry. ipsdienol synthase. Now EC 1.14.14.31, ipsdienol synthase]

[EC 1.14.13.207 created 2015, deleted 2016]

EC 1.14.13.208

Accepted name: benzoyl-CoA 2,3-epoxidase
Reaction: benzoyl-CoA + NADPH + H⁺ + O₂ = 2,3-epoxy-2,3-dihydrobenzoyl-CoA + NADP⁺ + H₂O
Other name(s): benzoyl-CoA dioxygenase/reductase (incorrect); BoxBA; BoxA/BoxB system; benzoyl-CoA 2,3-dioxygenase (incorrect)
Systematic name: benzoyl-CoA,NADPH:oxygen oxidoreductase (2,3-epoxydizing)

Comments: The enzyme is involved in aerobic benzoate metabolism in *Azoarcus evansii*. BoxB functions as the oxygenase part of benzoyl-CoA oxygenase in conjunction with BoxA, the reductase component, which upon binding of benzoyl-CoA, transfers two electrons to the ring in the course of monooxygenation. BoxA is a homodimeric 46 kDa iron-sulfur-flavoprotein (FAD), BoxB is a monomeric iron-protein [4849].

References: [4849, 1308, 2857, 3455]

[EC 1.14.13.208 created 2010 as EC 1.14.12.21, transferred 2015 to EC 1.14.13.208]

EC 1.14.13.209

Accepted name: salicyloyl-CoA 5-hydroxylase
Reaction: 2-hydroxybenzoyl-CoA + NADH + H⁺ + O₂ = gentisyl-CoA + NAD⁺ + H₂O
Other name(s): *sdgC* (gene name)
Systematic name: salicyloyl-CoA:NADH:oxygen oxidoreductase (5-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Streptomyces* sp. WA46, participates in a pathway for salicylate degradation. *cf.* EC 1.14.13.172, salicylate 5-hydroxylase.
References: [1835]

[EC 1.14.13.209 created 2015]

EC 1.14.13.210

Accepted name: 4-methyl-5-nitrocatechol 5-monooxygenase
Reaction: 4-methyl-5-nitrocatechol + NAD(P)H + H⁺ + O₂ = 2-hydroxy-5-methylquinone + nitrite + NAD(P)⁺ + H₂O
Other name(s): *dntB* (gene name); 4-methyl-5-nitrocatechol oxygenase; MNC monooxygenase
Systematic name: 4-methyl-5-nitrocatechol,NAD(P)H:oxygen 5-oxidoreductase (5-hydroxylating, nitrite-forming)
Comments: Contains FAD. The enzyme, isolated from the bacterium *Burkholderia* sp. DNT, can use both NADH and NADPH, but prefers NADPH. It has a narrow substrate range, but can also act on 4-nitrocatechol.
References: [1478, 2425]

[EC 1.14.13.210 created 2016]

EC 1.14.13.211

Accepted name: rifampicin monooxygenase
Reaction: rifampicin + NAD(P)H + O₂ = 2-hydroxy-2,27-secorifampicin + NAD(P)⁺ + H₂O
Other name(s): RIF-O; ROX; RIFMO; rifampicin:NAD(P)H:oxygen oxidoreductase (2'-N-hydroxyrifampicin-forming) (incorrect)
Systematic name: rifampicin:NAD(P)H:oxygen oxidoreductase (2-hydroxy-2,27-secorifampicin-forming; ring-cleaving)
Comments: The enzyme has been found in a variety of environmental bacteria, notably *Rhodococcus*, *Nocardia*, and *Streptomyces*. It hydroxylates C-2 of rifampicin leading to its macro-ring cleaving.
References: [92, 1737, 2246, 2519]

[EC 1.14.13.211 created 2016, modified 2022]

EC 1.14.13.212

Accepted name: 1,3,7-trimethyluric acid 5-monooxygenase
Reaction: 1,3,7-trimethylurate + NADH + H⁺ + O₂ = 1,3,7-trimethyl-5-hydroxyisourate + NAD⁺ + H₂O
Other name(s): *tmuM* (gene name)
Systematic name: 1,3,7-trimethylurate,NADH:oxygen oxidoreductase (1,3,7-trimethyl-5-hydroxyisourate-forming)
Comments: The enzyme, characterized from the bacterium *Pseudomonas* sp. CBB1, is part of the bacterial C-8 oxidation-based caffeine degradation pathway. The product decomposes spontaneously to a racemic mixture of 3,6,8-trimethylallantoin. The enzyme shows no activity with urate. *cf.* EC 1.14.13.113, FAD-dependent urate hydroxylase.

References: [2858, 4115]

[EC 1.14.13.212 created 2016]

[1.14.13.213 Transferred entry. *bursehernin 5-monoxygenase*. Now EC 1.14.14.131, *bursehernin 5-monoxygenase*]

[EC 1.14.13.213 created 2016, deleted 2018]

[1.14.13.214 Transferred entry. (-)-4'-demethyl-deoxypodophyllotoxin 4-hydroxylase. Now EC 1.14.14.132, (-)-4'-demethyl-deoxypodophyllotoxin 4-hydroxylase]

[EC 1.14.13.214 created 2016, deleted 2018]

EC 1.14.13.215

Accepted name: protoasukamycin 4-monoxygenase
Reaction: protoasukamycin + NADH + H⁺ + O₂ = 4-hydroxyprotoasukamycin + NAD⁺ + H₂O
Systematic name: protoasukamycin,NADH:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Streptomyces nodosus* subsp. *asukaensis*, is involved in the biosynthesis of the antibiotic asukamycin. Requires a flavin cofactor, with no preference among FMN, FAD or riboflavin. When flavin concentration is low, activity is enhanced by the presence of the NADH-dependent flavin-reductase AsuE2.
References: [3602]

[EC 1.14.13.215 created 2016]

EC 1.14.13.216

Accepted name: asperlicin C monoxygenase
Reaction: asperlicin C + NAD(P)H + H⁺ + O₂ = asperlicin E + NAD(P)⁺ + H₂O
Other name(s): AspB
Systematic name: asperlicin C,NAD(P)H:oxygen oxidoreductase
Comments: The enzyme, characterized from the fungus *Aspergillus alliaceus*, contains an FAD cofactor. The enzyme inserts a hydroxyl group, leading to formation of a N-C bond that creates an additional cycle between the bicyclic indole and the tetracyclic core moieties, resulting in the heptacyclic asperlicin E.
References: [1583]

[EC 1.14.13.216 created 2016]

EC 1.14.13.217

Accepted name: protodeoxyviolaceinate monoxygenase
Reaction: protodeoxyviolaceinate + NAD(P)H + O₂ = protoviolaceinate + NAD(P)⁺ + H₂O
Other name(s): *vioD* (gene name); protoviolaceinate synthase
Systematic name: protodeoxyviolaceinate,NAD(P)H:O₂ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Chromobacterium violaceum*, participates in the biosynthesis of the violet pigment violacein. The product, protoviolaceinate, can be acted upon by EC 1.14.13.224, violacein synthase, leading to violacein production. However, it is very labile, and in the presence of oxygen can undergo non-enzymic autooxidation to the shunt product proviolacein.
References: [199, 3887]

[EC 1.14.13.217 created 2016, modified 2016]

EC 1.14.13.218

Accepted name: 5-methylphenazine-1-carboxylate 1-monoxygenase
Reaction: 5-methylphenazine-1-carboxylate + NADH + O₂ = pyocyanin + NAD⁺ + CO₂ + H₂O
Other name(s): *phzS* (gene name)

Systematic name: 5-methylphenazine-1-carboxylate,NADH:oxygen oxidoreductase (1-hydroxylating, decarboxylating)
Comments: The enzyme, characterized from the bacterium *Pseudomonas aeruginosa*, is involved in the biosynthesis of pyocyanin, a toxin produced and secreted by the organism. It can also act on phenazine-1-carboxylate, converting it into phenazin-1-ol.
References: [2727, 3247, 1401]

[EC 1.14.13.218 created 2016]

EC 1.14.13.219

Accepted name: resorcinol 4-hydroxylase (NADPH)
Reaction: resorcinol + NADPH + H⁺ + O₂ = hydroxyquinol + NADP⁺ + H₂O
Systematic name: resorcinol,NADPH:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Corynebacterium glutamicum*, is a single-component hydroxylase. The enzyme has no activity with NADH. *cf.* EC 1.14.13.220, resorcinol 4-hydroxylase (NADH), and EC 1.14.14.27, resorcinol 4-hydroxylase (FADH₂).
References: [1757]

[EC 1.14.13.219 created 2016]

EC 1.14.13.220

Accepted name: resorcinol 4-hydroxylase (NADH)
Reaction: resorcinol + NADH + H⁺ + O₂ = hydroxyquinol + NAD⁺ + H₂O
Other name(s): *tsdB* (gene name)
Systematic name: resorcinol,NADH:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Rhodococcus jostii* RHA1, is a single-component hydroxylase. The enzyme has no activity with NADPH. *cf.* EC 1.14.13.219, resorcinol 4-hydroxylase (NADPH), and EC 1.14.14.27, resorcinol 4-hydroxylase (FADH₂).
References: [2004]

[EC 1.14.13.220 created 2016]

[1.14.13.221 *Transferred entry. cholest-4-en-3-one 26-monooxygenase [(25R)-3-oxocholest-4-en-26-oate forming]. Now EC 1.14.15.28, cholest-4-en-3-one 26-monooxygenase [(25R)-3-oxocholest-4-en-26-oate forming]*]

[EC 1.14.13.221 created 2016, deleted 2018]

EC 1.14.13.222

Accepted name: aurachin C monooxygenase/isomerase
Reaction: aurachin C + NAD(P)H + H⁺ + O₂ = 4-hydroxy-2-methyl-3-oxo-4-[(2E,6E)-farnesyl]-3,4-dihydroquinoline 1-oxide + NAD(P)⁺ + H₂O (overall reaction)
(1a) aurachin C + NAD(P)H + H⁺ + O₂ = 2-hydroxy-1a-methyl-7a-[(2E,6E)-farnesyl]-1a,2-dihydrooxireno[2,3-*b*]quinolin-7(7aH)-one + NAD(P)⁺ + H₂O
(1b) 2-hydroxy-1a-methyl-7a-[(2E,6E)-farnesyl]-1a,2-dihydrooxireno[2,3-*b*]quinolin-7(7aH)-one = 4-hydroxy-2-methyl-3-oxo-4-[(2E,6E)-farnesyl]-3,4-dihydroquinoline 1-oxide
Other name(s): *auaG* (gene name); aurachin C monooxygenase
Systematic name: aurachin C:NAD(P)H:oxygen oxidoreductase (4-hydroxy-2-methyl-3-oxo-4-farnesyl-3,4-dihydroquinoline-1-oxide-forming)
Comments: The aurachin C monooxygenase from the bacterium *Stigmatella aurantiaca* accepts both NADH and NADPH as cofactor, but has a preference for NADH. It catalyses the initial steps in the conversion of aurachin C to aurachin B. The FAD-dependent monooxygenase catalyses the epoxidation of the C₂-C₃ double bond of aurachin C, which is followed by a semipinacol rearrangement, causing migration of the farnesyl group from C₃ to C₄.
References: [2028]

[EC 1.14.13.222 created 2016]

EC 1.14.13.223

- Accepted name:** 3-hydroxy-4-methylanthranilyl-[aryl-carrier protein] 5-monooxygenase
Reaction: 3-hydroxy-4-methylanthranilyl-[aryl-carrier protein] + NADH + H⁺ + O₂ = 3,5-dihydroxy-4-methylanthranilyl-[aryl-carrier protein] + NAD⁺ + H₂O
Other name(s): *sibG* (gene name)
Systematic name: 3-hydroxy-4-methylanthranilyl-[aryl-carrier protein],NADH:oxygen oxidoreductase (5-hydroxylating)
Comments: A flavoprotein (FAD). The enzyme, characterized from the bacterium *Streptosporangium sibiricum*, is involved in the biosynthesis of the antitumor antibiotic sibiromycin. The enzyme is not active with free 3-hydroxy-4-methylanthranilate.
References: [1324]

[EC 1.14.13.223 created 2016]

EC 1.14.13.224

- Accepted name:** violacein synthase
Reaction: (1) protoviolaceinate + NAD(P)H + O₂ = violaceinate + NAD(P)⁺ + H₂O
(2) protodeoxyviolaceinate + NAD(P)H + O₂ = deoxyviolaceinate + NAD(P)⁺ + H₂O
Other name(s): proviolaceinate monooxygenase; *vioC* (gene name)
Systematic name: protoviolaceinate,NAD(P)H:O₂ oxidoreductase
Comments: The enzyme, characterized from the bacterium *Chromobacterium violaceum*, participates in the biosynthesis of the violet pigment violacein. The products, violaceinate and deoxyviolaceinate, undergo non-enzymic autooxidation into violacein and deoxyviolacein, respectively.
References: [199, 3887]

[EC 1.14.13.224 created 2016]

EC 1.14.13.225

- Accepted name:** F-actin monooxygenase
Reaction: [F-actin]-L-methionine + NADPH + O₂ + H⁺ = [F-actin]-L-methionine-(*R*)-*S*-oxide + NADP⁺ + H₂O
Other name(s): MICAL (gene name)
Systematic name: [F-actin]-L-methionine,NADPH:O₂ *S*-oxidoreductase
Comments: The enzyme, characterized from the fruit fly *Drosophila melanogaster*, is a multi-domain oxidoreductase that acts as an F-actin disassembly factor. The enzyme selectively reduces two L-Met residues of F-actin, causing fragmentation of the filaments and preventing repolymerization [1775]. Free methionine is not a substrate [1773]. The reaction is stereospecific and generates the (*R*)-sulfoxide [1774]. In the absence of substrate, the enzyme can act as an NAD(P)H oxidase (EC 1.6.3.1) [4939, 4454].
References: [1775, 1773, 1774, 4939, 4454]

[EC 1.14.13.225 created 2016]

EC 1.14.13.226

- Accepted name:** acetone monooxygenase (methyl acetate-forming)
Reaction: acetone + NADPH + H⁺ + O₂ = methyl acetate + NADP⁺ + H₂O
Other name(s): *acmA* (gene name)
Systematic name: acetone,NADPH:oxygen oxidoreductase (methyl acetate-forming)
Comments: Contains FAD. The enzyme, characterized from the bacterium *Gordonia* sp. TY-5, is a Baeyer-Villiger type monooxygenase and participates in a propane utilization pathway.
References: [2245]

[EC 1.14.13.226 created 2016]

EC 1.14.13.227

Accepted name: propane 2-monooxygenase
Reaction: propane + NADH + H⁺ + O₂ = propan-2-ol + NAD⁺ + H₂O
Other name(s): prmABCD (gene names)
Systematic name: propane,NADH:oxygen oxidoreductase (2-hydroxylating)
Comments: The enzyme, characterized from several bacterial strains, is a multicomponent dinuclear iron monooxygenase that includes a hydroxylase, an NADH-dependent reductase, and a coupling protein. The enzyme has several additional activities, including acetone monooxygenase (acetol-forming) and phenol 4-monooxygenase.
References: [2244, 3829, 1240]

[EC 1.14.13.227 created 2016]

EC 1.14.13.228

Accepted name: jasmonic acid 12-hydroxylase
Reaction: (–)-jasmonate + NADPH + H⁺ + O₂ = *trans*-12-hydroxyjasmonate + NADP⁺ + H₂O
Other name(s): ABM (gene name)
Systematic name: jasmonate,NADPH:oxygen oxidoreductase (12-hydroxylating)
Comments: Although believed to occur in plants, the enzyme has so far been characterized only from the rice blast fungus, *Magnaporthe oryzae*. The fungus strategically deploys the enzyme to hydroxylate and inactivate endogenous jasmonate to evade the jasmonate-based innate immunity in rice plants.
References: [3262]

[EC 1.14.13.228 created 2016]

EC 1.14.13.229

Accepted name: *tert*-butyl alcohol monooxygenase
Reaction: *tert*-butyl alcohol + NADPH + H⁺ + O₂ = 2-methylpropane-1,2-diol + NADP⁺ + H₂O
Other name(s): *mdpJK* (gene names); *tert*-butanol monooxygenase
Systematic name: *tert*-butyl alcohol,NADPH:oxygen oxidoreductase
Comments: The enzyme, characterized from the bacterium *Aquicola tertiaricarbonis*, is a Rieske nonheme mononuclear iron oxygenase. It can also act, with lower efficiency, on propan-2-ol, converting it to propane-1,2-diol. Depending on the substrate, the enzyme also catalyses EC 1.14.19.48, *tert*-amyl alcohol desaturase.
References: [3699, 3760]

[EC 1.14.13.229 created 2016]

EC 1.14.13.230

Accepted name: butane monooxygenase (soluble)
Reaction: butane + NADH + H⁺ + O₂ = butan-1-ol + NAD⁺ + H₂O
Other name(s): sBMO; bmoBCDXYZ (gene names)
Systematic name: butane,NADH:oxygen oxidoreductase
Comments: The enzyme, characterized from the bacterium *Thauera butanivorans*, is similar to EC 1.14.13.25, methane monooxygenase (soluble), but has a very low activity with methane. It comprises three components - a carboxylate-bridged non-heme di-iron center-containing hydroxylase (made of three different subunits), a flavo-iron sulfur-containing NADH-oxidoreductase, and a small regulatory component protein. The enzyme can also act on other C₃-C₆ linear and branched aliphatic alkanes with lower activity.
References: [3928, 976, 958, 725]

[EC 1.14.13.230 created 2016]

EC 1.14.13.231

- Accepted name:** tetracycline 11a-monooxygenase
Reaction: tetracycline + NADPH + H⁺ + O₂ = 11a-hydroxytetracycline + NADP⁺ + H₂O
Other name(s): *tetX* (gene name)
Systematic name: tetracycline,NADPH:oxygen oxidoreductase (11a-hydroxylating)
Comments: A flavoprotein (FAD). This bacterial enzyme confers resistance to all clinically relevant tetracyclines when expressed under aerobic conditions. The hydroxylated products are very unstable and lead to intramolecular cyclization and non-enzymic breakdown to undefined products.
References: [4766, 2875, 4467]

[EC 1.14.13.231 created 2016]

EC 1.14.13.232

- Accepted name:** 6-methylpretetramide 4-monooxygenase
Reaction: 6-methylpretetramide + NADPH + H⁺ + O₂ = 4-hydroxy-6-methylpretetramide + NADP⁺ + H₂O
Systematic name: 6-methylpretetramide,NADPH:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Streptomyces rimosus*, participates in the biosynthesis of tetracycline antibiotics. That bacterium possesses two enzymes that can catalyse the reaction - OxyE is the main isozyme, while OxyL has a lower activity. OxyL is bifunctional, and its main function is EC 1.14.13.233, 4-hydroxy-6-methylpretetramide 12a-monooxygenase. Contains FAD.
References: [4886, 4524]

[EC 1.14.13.232 created 2016]

EC 1.14.13.233

- Accepted name:** 4-hydroxy-6-methylpretetramide 12a-monooxygenase
Reaction: 4-hydroxy-6-methylpretetramide + NADPH + H⁺ + O₂ = 4-de(dimethylamino)-4-oxoanhydrotetracycline + NADP⁺ + H₂O
Other name(s): *oxyL* (gene name)
Systematic name: 4-hydroxy-6-methylpretetramide,NADPH:oxygen oxidoreductase (12a-hydroxylating)
Comments: Contains FAD. The enzyme, characterized from the bacterium *Streptomyces rimosus*, participates in the biosynthesis of tetracycline antibiotics. The enzyme is bifunctional, and can also catalyse EC 1.14.13.232, 6-methylpretetramide 4-monooxygenase.
References: [4886]

[EC 1.14.13.233 created 2016]

EC 1.14.13.234

- Accepted name:** 5a,11a-dehydrotetracycline 5-monooxygenase
Reaction: 5a,11a-dehydrotetracycline + NADPH + H⁺ + O₂ = 5a,11a-dehydroxytetracycline + NADP⁺ + H₂O
Other name(s): *oxyS* (gene name); 12-dehydrotetracycline 5-monooxygenase
Systematic name: 5a,11a-dehydrotetracycline,NADPH:oxygen oxidoreductase (5-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Streptomyces rimosus*, is bifunctional, catalysing two successive monooxygenation reactions. It starts by catalysing the stereospecific hydroxylation of anhydrotetracycline at C-6 (EC 1.14.13.38). If the released product is captured by EC 1.3.98.4, 5a,11a-dehydrotetracycline dehydrogenase (OxyR), it is reduced to tetracycline. However, if the released product is recaptured by OxyS, it performs an additional hydroxylation at C-5, producing 5a,11a-dehydroxytetracycline, which, following the action of OxyR, becomes oxytetracycline.
References: [335, 2810, 4419, 4522]

[EC 1.14.13.234 created 2016]

EC 1.14.13.235

- Accepted name:** indole-3-acetate monooxygenase
Reaction: (indol-3-yl)acetate + NADH + H⁺ + O₂ = (2-hydroxy-1*H*-indol-3-yl)acetate + NAD⁺ + H₂O
Other name(s): *iacA* (gene name)
Systematic name: (indol-3-yl)acetate,NADH:oxygen oxidoreductase (2-hydroxylating)
Comments: The enzyme, characterized from *Pseudomonas putida* strains, catalyses the first step in a pathway for degradation of the plant hormone indole-3-acetate. When acting on indole, the enzyme forms indoxyl, which reacts spontaneously with oxygen to form the blue dye indigo.
References: [2428, 3774]

[EC 1.14.13.235 created 2017]

EC 1.14.13.236

- Accepted name:** toluene 4-monooxygenase
Reaction: toluene + NADH + H⁺ + O₂ = 4-methylphenol + NAD⁺ + H₂O
Other name(s): TMO
Systematic name: toluene,NADH:oxygen oxidoreductase (4-hydroxylating)
Comments: This bacterial enzyme belongs to a family of soluble diiron hydroxylases that includes toluene-, benzene-, xylene- and methane monooxygenases, phenol hydroxylases, and alkene epoxidases. The enzyme comprises a four-component complex that includes a hydroxylase, NADH-ferredoxin oxidoreductase, a Rieske-type [2Fe-2S] ferredoxin, and an effector protein.
References: [4606, 1624, 3765, 184, 1741]

[EC 1.14.13.236 created 2017]

EC 1.14.13.237

- Accepted name:** aliphatic glucosinolate *S*-oxygenase
Reaction: an ω-(methylsulfanyl)alkyl-glucosinolate + NADPH + H⁺ + O₂ = an ω-(methylsulfinyl)alkyl-glucosinolate + NADP⁺ + H₂O
Other name(s): ω-(methylthio)alkylglucosinolate *S*-oxygenase; GS-OX1 (gene name); ω-(methylthio)alkyl-glucosinolate,NADPH:oxygen *S*-oxidoreductase
Systematic name: ω-(methylsulfanyl)alkyl-glucosinolate,NADPH:oxygen *S*-oxidoreductase
Comments: The enzyme is a member of the flavin-dependent monooxygenase (FMO) family (*cf.* EC 1.14.13.8). The plant *Arabidopsis thaliana* contains five isoforms. GS-OX1 through GS-OX4 are able to catalyse the *S*-oxygenation independent of chain length, while GS-OX5 is specific for 8-(methylsulfanyl)octyl glucosinolate.
References: [1510, 2445]

[EC 1.14.13.237 created 2017]

EC 1.14.13.238

- Accepted name:** dimethylamine monooxygenase
Reaction: dimethylamine + NADPH + H⁺ + O₂ = methylamine + formaldehyde + NADP⁺ + H₂O
Other name(s): *dmmABC* (gene names)
Systematic name: dimethylamine,NADPH:oxygen oxidoreductase (formaldehyde-forming)
Comments: The enzyme, characterized from several bacterial species, is involved in a pathway for the degradation of methylated amines. It is composed of three subunits, one of which is a ferredoxin, and contains heme iron and an FMN cofactor.
References: [1001, 999, 59, 2479]

[EC 1.14.13.238 created 2017]

EC 1.14.13.239

- Accepted name:** carnitine monooxygenase
Reaction: L-carnitine + NAD(P)H + H⁺ + O₂ = (3*R*)-3-hydroxy-4-oxobutanoate + trimethylamine + NAD(P)⁺ + H₂O
Other name(s): *cntAB* (gene names); *yeaWX* (gene names)
Systematic name: L-carnitine,NAD(P)H:oxygen oxidoreductase (trimethylamine-forming)
Comments: The bacterial enzyme is a complex consisting of a reductase and an oxygenase components. The reductase subunit contains a flavin and a plant-type ferredoxin [2Fe-2S] cluster, while the oxygenase subunit is a Rieske-type protein in which a [2Fe-2S] cluster is coordinated by two histidine and two cysteine residues.
References: [923, 4928, 2191]

[EC 1.14.13.239 created 2017]

EC 1.14.13.240

- Accepted name:** 2-polyprenylphenol 6-hydroxylase
Reaction: 2-(*all-trans*-polyprenyl)phenol + NADPH + H⁺ + O₂ = 3-(*all-trans*-polyprenyl)benzene-1,2-diol + NADP⁺ + H₂O
Other name(s): *ubiI* (gene name); *ubiM* (gene name)
Systematic name: 2-(*all-trans*-polyprenyl)phenol,NADPH:oxygen oxidoreductase (6-hydroxylating)
Comments: Contains FAD. The enzyme from the bacterium *Escherichia coli* (UbiI) catalyses the first hydroxylation during the aerobic biosynthesis of ubiquinone. The enzyme from the bacterium *Neisseria meningitidis* (UbiM) can also catalyse the two additional hydroxylations that occur in the pathway (*cf.* EC 1.14.99.60, 3-demethoxyubiquinol 3-hydroxylase).
References: [625, 3285]

[EC 1.14.13.240 created 2018]

EC 1.14.13.241

- Accepted name:** 5-pyridoxate monooxygenase
Reaction: 3-hydroxy-4-hydroxymethyl-2-methylpyridine-5-carboxylate + NADPH + H⁺ + O₂ = 2-(acetamidomethylene)-3-(hydroxymethyl)succinate + NADP⁺
Other name(s): 5-pyridoxate,NADPH:oxygen oxidoreductase (decyclizing); 5-pyridoxate oxidase (misleading); 5-pyridoxate dioxygenase (incorrect)
Systematic name: 5-pyridoxate,NADPH:oxygen oxidoreductase (ring-opening)
Comments: Contains FAD. The enzyme, characterized from the bacterium *Arthrobacter* sp. Cr-7, participates in the degradation of pyridoxine (vitamin B₆). Although the enzyme was initially thought to be a dioxygenase, oxygen-tracer experiments have suggested that it is a monooxygenase, incorporating only one oxygen atom from molecular oxygen into the product. The second oxygen atom originates from a water molecule, which is regenerated during the reaction and thus does not show up in the reaction equation.
References: [3973, 3040, 587]

[EC 1.14.13.241 created 2018 (EC 1.14.12.5 created 1972, incorporated 2018)]

EC 1.14.13.242

- Accepted name:** 3-hydroxy-2-methylpyridine-5-carboxylate monooxygenase
Reaction: 3-hydroxy-2-methylpyridine-5-carboxylate + NAD(P)H + H⁺ + O₂ = 2-(acetamidomethylidene)succinate + NAD(P)⁺

Other name(s): MHPCO; 3-hydroxy-2-methylpyridine-5-carboxylate,NAD(P)H:oxygen oxidoreductase (decyclizing); methylhydroxypyridinecarboxylate oxidase (misleading); 2-methyl-3-hydroxypyridine 5-carboxylic acid dioxygenase (incorrect); methylhydroxypyridine carboxylate dioxygenase (incorrect); 3-hydroxy-3-methylpyridinecarboxylate dioxygenase [incorrect]; 3-hydroxy-2-methylpyridinecarboxylate dioxygenase (incorrect)

Systematic name: 3-hydroxy-2-methylpyridine-5-carboxylate,NAD(P)H:oxygen oxidoreductase (ring-opening)

Comments: Contains FAD. The enzyme, characterized from the bacteria *Pseudomonas* sp. MA-1 and *Mesorhizobium loti*, participates in the degradation of pyridoxine (vitamin B₆). Although the enzyme was initially thought to be a dioxygenase, oxygen-tracer experiments have shown that it is a monooxygenase, incorporating only one oxygen atom from molecular oxygen. The second oxygen atom that is incorporated into the product originates from a water molecule, which is regenerated during the reaction and thus does not show up in the reaction equation.

References: [3973, 588, 3187, 4840, 2745, 4284, 4283]

[EC 1.14.13.242 created 2018 (EC 1.14.12.4 created 1972, incorporated 2018)]

EC 1.14.13.243

Accepted name: toluene 2-monooxygenase

Reaction: (1) toluene + NADH + H⁺ + O₂ = 2-methylphenol + NAD⁺ + H₂O
(2) 2-methylphenol + NADH + H⁺ + O₂ = 3-methylcatechol + NAD⁺ + H₂O

Other name(s): *tomA1/2/3/4/5* (gene names); toluene *ortho*-monooxygenase

Systematic name: toluene,NADH:oxygen oxidoreductase (2,3-dihydroxylating)

Comments: The enzyme, characterized from the bacterium *Burkholderia cepacia*, belongs to a class of nonheme, oxygen-dependent diiron enzymes. It contains a hydroxylase component with two binuclear iron centers, an NADH-oxidoreductase component containing FAD and a [2Fe-2S] iron-sulfur cluster, and a third component involved in electron transfer between the hydroxylase and the reductase. The enzyme dihydroxylates its substrate in two sequential hydroxylations, initially forming 2-methylphenol, which is hydroxylated to 3-methylcatechol.

References: [3055, 4784, 543]

[EC 1.14.13.243 created 2019]

EC 1.14.13.244

Accepted name: phenol 2-monooxygenase (NADH)

Reaction: phenol + NADH + H⁺ + O₂ = catechol + NAD⁺ + H₂O

Other name(s): *dmpLMNOP* (gene names)

Systematic name: phenol,NADH:oxygen oxidoreductase (2-hydroxylating)

Comments: The enzyme, characterized from the bacteria *Pseudomonas* sp. CF600 and *Acinetobacter radiore-sistens*, consists of a multisubunit oxygenase component that contains the active site and a dinuclear iron center, a reductase component that contains FAD and one iron-sulfur cluster, and a regulatory component. The reductase component is responsible for transferring electrons from NADH to the dinuclear iron center.

References: [3106, 3366, 3365, 3398, 517]

[EC 1.14.13.244 created 2019]

EC 1.14.13.245

Accepted name: assimilatory dimethylsulfide *S*-monooxygenase

Reaction: (1) dimethyl sulfide + NADH + H⁺ + O₂ = dimethyl sulfoxide + NAD⁺ + H₂O
(2) dimethyl sulfoxide + NADH + H⁺ + O₂ = dimethyl sulfone + NAD⁺ + H₂O

Other name(s): *dsoBCDEF* (gene names)

Systematic name: dimethyl sulfide,NADH:oxygen oxidoreductase (*S*-oxidizing)

Comments: The enzyme, studied from the bacterium *Acinetobacter* sp. strain 20B, is very similar to EC 1.14.13.244, phenol 2-monooxygenase (NADH). It consists of a multisubunit oxygenease component that contains the active site and a dinuclear iron center, a reductase component that contains FAD and one iron-sulfur cluster, and a regulatory component. The three components comprise five different polypeptides. The enzyme catalyses the first two steps of a dimethyl sulfide oxidation pathway in this organism.

References: [1724, 1725]

[EC 1.14.13.245 created 2019]

EC 1.14.13.246

Accepted name: 4 β -methylsterol monooxygenase

Reaction: a 3 β -hydroxy-4,4-dimethylsteroid + 3 NADH + 3 H⁺ + 3 O₂ = a 3 β -hydroxy-4 α -methylsteroid-4 β -carboxylate + 3 NAD⁺ + 4 H₂O (overall reaction)

(1a) a 3 β -hydroxy-4,4-dimethylsteroid + NADH + H⁺ + O₂ = a 3 β -hydroxy-4 β -hydroxymethyl-4 α -methylsteroid + NAD⁺ + H₂O

(1b) a 3 β -hydroxy-4 β -hydroxymethyl-4 α -methylsteroid + NADH + H⁺ + O₂ = a 3 β -hydroxy-4 β -formyl-4 α -methylsteroid + NAD⁺ + 2 H₂O

(1c) a 3 β -hydroxy-4 β -formyl-4 α -methylsteroid + NADH + H⁺ + O₂ = a 3 β -hydroxy-4 α -methylsteroid-4 β -carboxylate + NAD⁺ + H₂O

Other name(s): *sdmA* (gene name)

Systematic name: 3 β -hydroxy-4,4-dimethylsteroid,NADH:oxygen oxidoreductase (C-4m β -hydroxylating)

Comments: Contains a Rieske [2Fe-2S] iron-sulfur cluster. This bacterial enzyme (SdmA) participates in the biosynthesis of bacterial sterols. Together with SdmB it forms an enzyme system that removes one methyl group from the C-4 position of 4,4-dimethylated steroid molecules. SdmA catalyses three successive oxidations of the C-4 β methyl group, turning it into a carboxylate group; the second enzyme, SdmB, is a bifunctional enzyme that catalyses two different activities. As EC 1.1.1.417, 3 β -hydroxysteroid-4 β -carboxylate 3-dehydrogenase (decarboxylating), it catalyses an oxidative decarboxylation that results in reduction of the 3 β -hydroxy group at the C-3 carbon to an oxo group. As EC 1.1.1.270, 3 β -hydroxysteroid 3-dehydrogenase, it reduces the 3-oxo group back to a 3 β -hydroxyl. Unlike the animal/fungal enzyme EC 1.14.18.9, 4 α -methylsterol monooxygenase, and the plant enzymes EC 1.14.18.10, plant 4,4-dimethylsterol C-4 α -methyl-monooxygenase, and EC 1.14.18.11, plant 4 α -monomethylsterol monooxygenase, this enzyme acts preferentially on the 4 β -methyl group. Since no epimerization of the remaining C-4 α methyl group occurs, the enzyme can only remove one methyl group, leaving a 4 α -monomethylated product. Known substrates include 4,4-dimethyl-5 α -cholest-8-en-3 β -ol and 14-demethylsterol.

References: [2376]

[EC 1.14.13.246 created 2019]

EC 1.14.13.247

Accepted name: stachydrine *N*-demethylase

Reaction: L-proline betaine + NAD(P)H + H⁺ + O₂ = *N*-methyl-L-proline + formaldehyde + NAD(P)⁺ + H₂O

Other name(s): L-proline betaine *N*-demethylase; *stc2* (gene name)

Systematic name: L-proline betaine,NAD(P)H:oxygen oxidoreductase (formaldehyde-forming)

Comments: The enzyme, characterized from the bacterium *Sinorhizobium meliloti* 1021, consists of three different types of subunits. The catalytic unit contains a Rieske [2Fe-2S] iron-sulfur cluster, and catalyses the monooxygenation of a methyl group. The resulting *N*-methoxyl group is unstable and decomposes spontaneously to form formaldehyde. The other subunits are involved in the transfer of electrons from NAD(P)H to the catalytic subunit.

References: [835, 2283]

[EC 1.14.13.247 created 2017]

EC 1.14.13.248

- Accepted name:** L-aspartate *N*-monooxygenase (nitrosuccinate-forming)
- Reaction:** L-aspartate + 3 NADPH + 3 H⁺ + 3 O₂ = (2*S*)- 2-nitrobutanedioate + 3 NADP⁺ + 4 H₂O
 (1a) L-aspartate + NADPH + H⁺ + O₂ = *N*-hydroxy-L-aspartate + NADP⁺ + H₂O
 (1b) *N*-hydroxy-L-aspartate + NADPH + H⁺ + O₂ = *N,N*-dihydroxy-L-aspartate + NADP⁺ + H₂O
 (1c) *N,N*-dihydroxy-L-aspartate = (2*S*)-2-nitrosobutanedioate + H₂O (spontaneous)
 (1d) (2*S*)-2-nitrosobutanedioate + NADPH + H⁺ + O₂ = (2*S*)-2-nitrobutanedioate + NADP⁺ + H₂O
- Other name(s):** *creE* (gene name)
- Systematic name:** L-aspartate,NADPH:oxygen oxidoreductase [(2*S*)-2-nitrobutanedioate-forming]
- Comments:** The enzyme, found in some Actinobacteria, is involved in a pathway that forms nitrite, which is subsequently used to generate a diazo group in some secondary metabolites. Requires an FAD cofactor.
- References:** [4092, 1475]

[EC 1.14.13.248 created 2021]

EC 1.14.13.249

- Accepted name:** 3-amino-4-hydroxybenzoate 2-monooxygenase
- Reaction:** 3-amino-4-hydroxybenzoate + NADPH + H⁺ + O₂ = 3-amino-2,4-dihydroxybenzoate + NADP⁺ + H₂O
- Other name(s):** *creL* (gene name); *ptmB3* (gene name); *ptnB3* (gene name)
- Systematic name:** 3-amino-4-hydroxybenzoate,NADPH:oxygen oxidoreductase (2-hydroxylating)
- Comments:** Requires FAD. The CreL enzyme from the bacterium *Streptomyces cremeomycin* participates in the biosynthesis of cremeomycin. The PrmB3 and PtnB3 enzymes from *Streptomyces platensis* are involved in the biosynthesis of platensimycin and platencin, respectively.
- References:** [3931, 4494, 4092, 948]

[EC 1.14.13.249 created 2021]

EC 1.14.13.250

- Accepted name:** nitrosourea synthase
- Reaction:** *N*^ω-methyl-L-arginine + 2 NADH + 2 H⁺ + 3 O₂ = *N*^δ-hydroxy-*N*^ω-methyl-*N*^ω-nitroso-L-citrulline + 2 NAD⁺ + 3 H₂O (overall reaction)
 (1a) *N*^ω-methyl-L-arginine + NADH + H⁺ + O₂ = *N*^δ-hydroxy-*N*^ω-methyl-L-arginine + NAD⁺ + H₂O
 (1b) *N*^δ-hydroxy-*N*^ω-methyl-L-arginine + NADH + H⁺ + O₂ = *N*^δ,*N*^{ω'}-dihydroxy-*N*^ω-methyl-L-arginine + NAD⁺ + H₂O
 (1c) *N*^δ,*N*^{ω'}-dihydroxy-*N*^ω-methyl-L-arginine + O₂ = *N*^δ-hydroxy-*N*^ω-methyl-*N*^ω-nitroso-L-citrulline + H₂O
- Other name(s):** *sznF* (gene name); StzF
- Systematic name:** *N*^ω-methyl-L-arginine,NADH:oxygen oxidoreductase (*N*^δ-hydroxy-*N*^ω-methyl-*N*^ω-nitroso-L-citrulline-forming)
- Comments:** The enzyme, characterized from the bacterium *Streptomyces achromogenes subsp. streptozoticus*, catalyses a complex multi-step reaction during the biosynthesis of the glucosamine-nitrosourea antibiotic streptozotocin. The overall reaction is an oxidative rearrangement of the guanidine group of *N*^ω-methyl-L-arginine, generating an *N*-nitrosourea product. The enzyme hydroxylates its substrate at the *N*^δ position, followed by a second hydroxylation at the *N*^{ω'} position. It then catalyses an oxidative rearrangement to form *N*^δ-hydroxy-*N*^ω-methyl-*N*^ω-nitroso-L-citrulline. This product is unstable, and degrades non-enzymically into nitric oxide and the denitrosated product *N*^δ-hydroxy-*N*^ω-methyl-L-citrulline. The enzyme contains two active sites, each of which utilizes a different iron-containing cofactor.
- References:** [3057, 1588, 2740, 2739, 4517]

[EC 1.14.13.250 created 2021]

EC 1.14.13.251

- Accepted name:** glycine betaine monooxygenase
Reaction: glycine betaine + NADH + H⁺ + O₂ = *N,N*-dimethylglycine + formaldehyde + NAD⁺ + H₂O
Other name(s): glycine betaine dioxygenase (incorrect); *bmoAB* (gene names); *gbcAB* (gene names)
Systematic name: glycine betaine,NADH:oxygen oxidoreductase (demethylating)
Comments: The enzyme, characterized from the bacteria *Pseudomonas aeruginosa* and *Chromohalobacter salexigenis*, is involved in a degradation pathway of glycine betaine. It is composed of two subunits - a ferredoxin reductase component that contains FAD, and a terminal oxygenase component that contains a [2Fe-2S] Rieske-type iron-sulfur cluster and a nonheme iron centre.
References: [4543, 2456, 3823]

[EC 1.14.13.251 created 2022]

EC 1.14.14 With reduced flavin or flavoprotein as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.14.1

- Accepted name:** unspecific monooxygenase
Reaction: RH + [reduced NADPH—hemoprotein reductase] + O₂ = ROH + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): microsomal monooxygenase; xenobiotic monooxygenase; aryl-4-monooxygenase; aryl hydrocarbon hydroxylase; microsomal *P*-450; flavoprotein-linked monooxygenase; flavoprotein monooxygenase; substrate,reduced-flavoprotein:oxygen oxidoreductase (RH-hydroxylating or -epoxidizing)
Systematic name: substrate,NADPH—hemoprotein reductase:oxygen oxidoreductase (RH-hydroxylating or -epoxidizing)
Comments: A group of *P*-450 heme-thiolate proteins, acting on a wide range of substrates including many xenobiotics, steroids, fatty acids, vitamins and prostaglandins; reactions catalysed include hydroxylation, epoxidation, *N*-oxidation, sulfoxidation, *N*-, *S*- and *O*-dealkylations, desulfation, deamination, and reduction of azo, nitro and *N*-oxide groups. Together with EC 1.6.2.4, NADPH—hemoprotein reductase, it forms a system in which two reducing equivalents are supplied by NADPH. Some of the reactions attributed to EC 1.14.15.3, alkane 1-monooxygenase, belong here.
References: [390, 1213, 1564, 1805, 1930, 2297, 2343, 2344, 2417, 2549, 2830, 2831, 3009, 3033, 4106, 4254, 4265]

[EC 1.14.14.1 created 1961 as EC 1.99.1.1, transferred 1965 to EC 1.14.1.1, transferred 1972 to EC 1.14.14.1 (EC 1.14.14.2 created 1972, incorporated 1976, EC 1.14.99.8 created 1972, incorporated 1984), modified 2015]

[1.14.14.2 Deleted entry. benzopyrene 3-monooxygenase. Now included with EC 1.14.14.1 unspecific monooxygenase]

[EC 1.14.14.2 created 1972, deleted 1976]

EC 1.14.14.3

- Accepted name:** bacterial luciferase
Reaction: a long-chain aldehyde + FMNH₂ + O₂ = a long-chain fatty acid + FMN + H₂O + *hν*
Other name(s): aldehyde monooxygenase; luciferase; *Vibrio fischeri* luciferase; alkanal,reduced-FMN:oxygen oxidoreductase (1-hydroxylating, luminescing); alkanal,FMNH₂:oxygen oxidoreductase (1-hydroxylating, luminescing); alkanal monooxygenase (FMN); aldehyde,FMNH₂:oxygen oxidoreductase (1-hydroxylating, luminescing)
Systematic name: long-chain-aldehyde,FMNH₂:oxygen oxidoreductase (1-hydroxylating, luminescing)

Comments: The reaction sequence starts with the incorporation of a molecule of oxygen into reduced FMN bound to the enzyme, forming luciferase peroxyflavin. The peroxyflavin interacts with an aliphatic long-chain aldehyde, producing a highly fluorescent species believed to be luciferase hydroxyflavin. The enzyme is highly specific for reduced FMN and for long-chain aliphatic aldehydes with eight carbons or more. The highest efficiency is achieved with tetradecanal. *cf.* EC 1.13.12.18, dinoflagellate luciferase.

References: [1554, 1553, 1555, 3032, 4137, 2300]

[EC 1.14.14.3 created 1981, modified 2016]

[1.14.14.4 Deleted entry. choline monooxygenase. Identical to EC 1.14.15.7]

[EC 1.14.14.4 created 2000, deleted 2002]

EC 1.14.14.5

Accepted name: alkanesulfonate monooxygenase
Reaction: an alkanesulfonate + FMNH₂ + O₂ = an aldehyde + FMN + sulfite + H₂O
Other name(s): SsuD; sulfate starvation-induced protein 6; alkanesulfonate, reduced-FMN: oxygen oxidoreductase
Systematic name: alkanesulfonate, FMNH₂: oxygen oxidoreductase
Comments: The enzyme from *Escherichia coli* catalyses the desulfonation of a wide range of aliphatic sulfonates (unsubstituted C₁- to C₁₄-sulfonates as well as substituted C₂-sulfonates). Does not desulfonate taurine (2-aminoethanesulfonate) or aromatic sulfonates. Does not use FMN as a bound cofactor. Instead, it uses reduced FMN (i.e., FMNH₂) as a substrate. FMNH₂ is provided by SsuE, the associated FMN reductase (EC 1.5.1.38).
References: [1028]

[EC 1.14.14.5 created 2002]

[1.14.14.6 Transferred entry. methanesulfonate monooxygenase. Now EC 1.14.13.111, methanesulfonate monooxygenase. Formerly thought to involve FMNH₂ but now shown to use NADH.]

[EC 1.14.14.6 created 2009, deleted 2010]

[1.14.14.7 Transferred entry. tryptophan 7-halogenase. As oxygen is completely reduced to H₂O and is not incorporated into the donor chloride, the enzyme has been transferred to EC 1.14.19.9, tryptophan 7-halogenase]

[EC 1.14.14.7 created 2009, deleted 2014]

EC 1.14.14.8

Accepted name: anthranilate 3-monooxygenase (FAD)
Reaction: anthranilate + FADH₂ + O₂ = 3-hydroxyanthranilate + FAD + H₂O
Other name(s): anthranilate 3-hydroxylase; anthranilate hydroxylase
Systematic name: anthranilate, FADH₂: oxygen oxidoreductase (3-hydroxylating)
Comments: This enzyme, isolated from the bacterium *Geobacillus thermodenitrificans*, participates in the pathway of tryptophan degradation. The enzyme is part of a system that also includes a bifunctional riboflavin kinase/FMN adenylyltransferase and an FAD reductase, which ensures ample supply of FAD to the monooxygenase.
References: [2525]

[EC 1.14.14.8 created 2010]

EC 1.14.14.9

Accepted name: 4-hydroxyphenylacetate 3-monooxygenase
Reaction: 4-hydroxyphenylacetate + FADH₂ + O₂ = 3,4-dihydroxyphenylacetate + FAD + H₂O

Other name(s): *p*-hydroxyphenylacetate 3-hydroxylase; 4-hydroxyphenylacetic acid-3-hydroxylase; *p*-hydroxyphenylacetate hydroxylase (FAD); 4 HPA 3-hydroxylase; *p*-hydroxyphenylacetate 3-hydroxylase (FAD); HpaB

Systematic name: 4-hydroxyphenylacetate,FADH₂:oxygen oxidoreductase (3-hydroxylating)

Comments: The enzyme from *Escherichia coli* attacks a broad spectrum of phenolic compounds. The enzyme uses FADH₂ as a substrate rather than a cofactor [4712]. FADH₂ is provided by EC 1.5.1.36, flavin reductase (NADH) [1256, 2545].

References: [12, 3381, 3380, 4712, 1256, 2545]

[EC 1.14.14.9 created 1972 as EC 1.14.13.3, transferred 2011 to EC 1.14.14.9]

EC 1.14.14.10

Accepted name: nitrilotriacetate monooxygenase

Reaction: nitrilotriacetate + FMNH₂ + H⁺ + O₂ = iminodiacetate + glyoxylate + FMN + H₂O

Systematic name: nitrilotriacetate,FMNH₂:oxygen oxidoreductase (glyoxylate-forming)

Comments: Requires Mg²⁺. The enzyme from *Aminobacter aminovorans* (previously *Chelatobacter heintzii*) is part of a two component system that also includes EC 1.5.1.42 (FMN reductase), which provides reduced flavin mononucleotide for this enzyme.

References: [4370, 2169, 4708]

[EC 1.14.14.10 created 2011]

EC 1.14.14.11

Accepted name: styrene monooxygenase

Reaction: styrene + FADH₂ + O₂ = (*S*)-2-phenyloxirane + FAD + H₂O

Other name(s): StyA; SMO; NSMOA

Systematic name: styrene,FADH₂:oxygen oxidoreductase

Comments: The enzyme catalyses the first step in the aerobic styrene degradation pathway. It forms a two-component system with a reductase (StyB) that utilizes NADH to reduce flavin-adenine dinucleotide, which is then transferred to the oxygenase.

References: [3208, 4296]

[EC 1.14.14.11 created 2011]

EC 1.14.14.12

Accepted name: 3-hydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione monooxygenase

Reaction: 3-hydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione + FMNH₂ + O₂ = 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione + FMN + H₂O

Other name(s): HsaA

Systematic name: 3-hydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione,FMNH₂:oxygen oxidoreductase

Comments: This bacterial enzyme participates in the degradation of several steroids, including cholesterol and testosterone. It can use either FADH or FMNH₂ as flavin cofactor. The enzyme forms a two-component system with a reductase (HsaB) that utilizes NADH to reduce the flavin, which is then transferred to the oxygenase subunit.

References: [965]

[EC 1.14.14.12 created 2011]

EC 1.14.14.13

Accepted name: 4-(γ -L-glutamylamino)butanoyl-[BtrI acyl-carrier protein] monooxygenase

Reaction: 4-(γ -L-glutamylamino)butanoyl-[BtrI acyl-carrier protein] + FMNH₂ + O₂ = 4-(γ -L-glutamylamino)-(2*S*)-2-hydroxybutanoyl-[BtrI acyl-carrier protein] + FMN + H₂O

Other name(s): *btrO* (gene name)

Systematic name: 4-(γ -L-glutamylamino)butanoyl-[BtrI acyl-carrier protein],FMNH₂:oxygen oxidoreductase (2-hydroxylating)
Comments: Catalyses a step in the biosynthesis of the side chain of the aminoglycoside antibiotics of the butirosin family. FMNH₂ is used as a free cofactor. Forms a complex with a dedicated NAD(P)H:FMN oxidoreductase. The enzyme is not able to hydroxylate free substrates, activation by the acyl-carrier protein is mandatory. Octanoyl-S-[BtrI acyl-carrier protein] is also accepted.
References: [2465]

[EC 1.14.14.13 created 2012]

EC 1.14.14.14

Accepted name: aromatase
Reaction: (1) testosterone + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = 17 β -estradiol + formate + 4 H₂O + 3 [oxidized NADPH—hemoprotein reductase] (overall reaction)
(1a) testosterone + O₂ + [reduced NADPH—hemoprotein reductase] = 19-hydroxytestosterone + H₂O + [oxidized NADPH—hemoprotein reductase]
(1b) 19-hydroxytestosterone + O₂ + [reduced NADPH—hemoprotein reductase] = 19-oxotestosterone + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
(1c) 19-oxotestosterone + O₂ + [reduced NADPH—hemoprotein reductase] = 17 β -estradiol + formate + H₂O + [oxidized NADPH—hemoprotein reductase]
(2) androst-4-ene-3,17-dione + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = estrone + formate + 4 H₂O + 3 [oxidized NADPH—hemoprotein reductase] (overall reaction)
(2a) androst-4-ene-3,17-dione + O₂ + [reduced NADPH—hemoprotein reductase] = 19-hydroxyandrost-4-ene-3,17-dione + H₂O + [oxidized NADPH—hemoprotein reductase]
(2b) 19-hydroxyandrost-4-ene-3,17-dione + O₂ + [reduced NADPH—hemoprotein reductase] = 19-oxo-androst-4-ene-3,17-dione + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
(2c) 19-oxoandrost-4-ene-3,17-dione + O₂ + [reduced NADPH—hemoprotein reductase] = estrone + formate + H₂O + [oxidized NADPH—hemoprotein reductase]
Other name(s): CYP19A1 (gene name); estrogen synthetase (incorrect)
Systematic name: testosterone1,NADPH—hemoprotein reductase:oxygen oxidoreductase (17 β -estradiol-forming)
Comments: A cytochrome *P*-450. The enzyme catalyses three sequential hydroxylations of the androgens androst-4-ene-3,17-dione and testosterone, resulting in their aromatization and forming the estrogens estrone and 17 β -estradiol, respectively. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [4270, 1128, 2060, 1315]

[EC 1.14.14.14 created 2013]

EC 1.14.14.15

Accepted name: (3*S*)-3-amino-3-(3-chloro-4-hydroxyphenyl)propanoyl-[peptidyl-carrier protein SgcC2] monooxygenase
Reaction: (3*S*)-3-amino-3-(3-chloro-4-hydroxyphenyl)propanoyl-[peptidyl-carrier protein SgcC2] + FADH₂ + O₂ = (3*S*)-3-amino-3-(3-chloro-4,5-dihydroxyphenyl)propanoyl-[peptidyl-carrier protein SgcC2] + FAD + H₂O
Other name(s): SgcC
Systematic name: (3*S*)-3-amino-3-(3-chloro-4-hydroxyphenyl)propanoyl-[peptidyl-carrier protein SgcC2],FADH₂:oxygen oxidoreductase (5-hydroxylating)
Comments: The enzyme from the bacterium *Streptomyces globisporus* is involved in the biosynthesis of the (*S*)-3-chloro-5-hydroxy- β -tyrosine moiety prior to incorporation into the chromoprotein antitumor antibiotic C-1027.
References: [2493]

[EC 1.14.14.15 created 2014]

EC 1.14.14.16

- Accepted name:** steroid 21-monooxygenase
Reaction: a C₂₁ steroid + [reduced NADPH—hemoprotein reductase] + O₂ = a 21-hydroxy-C₂₁-steroid + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): steroid 21-hydroxylase; 21-hydroxylase; P450c21; CYP21A2 (gene name)
Systematic name: steroid,NADPH—hemoprotein reductase:oxygen oxidoreductase (21-hydroxylating)
Comments: A P-450 heme-thiolate protein responsible for the conversion of progesterone and 17 α -hydroxyprogesterone to their respective 21-hydroxylated derivatives, 11-deoxycorticosterone and 11-deoxycortisol. Involved in the biosynthesis of the hormones aldosterone and cortisol. The electron donor is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [1577, 3333, 3611, 2214, 2669, 124]

[EC 1.14.14.16 created 1961 as EC 1.99.1.11, transferred 1965 to EC 1.14.1.8, transferred 1972 to EC 1.14.99.10, modified 2013, transferred 2015 to EC 1.14.14.16]

EC 1.14.14.17

- Accepted name:** squalene monooxygenase
Reaction: squalene + [reduced NADPH—hemoprotein reductase] + O₂ = (3S)-2,3-epoxy-2,3-dihydrosqualene + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): squalene epoxidase; squalene-2,3-epoxide cyclase; squalene 2,3-oxidocyclase; squalene hydroxylase; squalene oxydocyclase; squalene-2,3-epoxidase
Systematic name: squalene,NADPH—hemoprotein:oxygen oxidoreductase (2,3-epoxidizing)
Comments: A flavoprotein (FAD). This enzyme, together with EC 5.4.99.7, lanosterol synthase, was formerly known as squalene oxidocyclase. The electron donor is EC 1.6.2.4, NADPH—hemoprotein reductase [3186, 690].
References: [735, 4230, 4414, 4741, 3186, 3673, 690, 1587]

[EC 1.14.14.17 created 1961 as EC 1.99.1.13, transferred 1965 to EC 1.14.1.3, part transferred 1972 to EC 1.14.99.7, transferred 2011 to EC 1.14.13.132, transferred 2015 to EC 1.14.14.17]

EC 1.14.14.18

- Accepted name:** heme oxygenase (biliverdin-producing)
Reaction: protoheme + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = biliverdin + Fe²⁺ + CO + 3 [oxidized NADPH—hemoprotein reductase] + 3 H₂O
Other name(s): ORP33 proteins; haem oxygenase (ambiguous); heme oxygenase (decyclizing) (ambiguous); heme oxidase (ambiguous); haem oxidase (ambiguous); heme oxygenase (ambiguous); heme,hydrogen-donor:oxygen oxidoreductase (α -methene-oxidizing, hydroxylating)
Systematic name: protoheme,NADPH—hemoprotein reductase:oxygen oxidoreductase (α -methene-oxidizing, hydroxylating)
Comments: This mammalian enzyme participates in the degradation of heme. The terminal oxygen atoms that are incorporated into the carbonyl groups of pyrrole rings A and B of biliverdin are derived from two separate oxygen molecules [3096]. The third oxygen molecule provides the oxygen atom that converts the α -carbon to CO. The enzyme requires NAD(P)H and EC 1.6.2.4, NADPH—hemoprotein reductase. *cf.* EC 1.14.15.20, heme oxygenase (biliverdin-producing, ferredoxin).
References: [2622, 4124, 4812, 3096, 2324]

[EC 1.14.14.18 created 1972 as EC 1.14.99.3, modified 2006, transferred 2015 to EC 1.14.14.18, modified 2016]

EC 1.14.14.19

- Accepted name:** steroid 17 α -monooxygenase
Reaction: a C₂₁-steroid + [reduced NADPH—hemoprotein reductase] + O₂ = a 17 α -hydroxy-C₂₁-steroid + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): steroid 17 α -hydroxylase; cytochrome *P*-450 17 α ; cytochrome *P*-450 (*P*-450 17 α ,lyase); 17 α -hydroxylase-C17,20 lyase; CYP17; CYP17A1 (gene name)

Systematic name: steroid,NADPH—hemoprotein reductase:oxygen oxidoreductase (17 α -hydroxylating)

Comments: Requires NADPH and EC 1.6.2.4, NADPH—hemoprotein reductase. A microsomal hemeprotein that catalyses two independent reactions at the same active site - the 17 α -hydroxylation of pregnenolone and progesterone, which is part of glucocorticoid hormones biosynthesis, and the conversion of the 17 α -hydroxylated products via a 17,20-lyase reaction to form androstenedione and dehydroepiandrosterone, leading to sex hormone biosynthesis (EC 1.14.14.32, 17 α -hydroxyprogesterone deacetylase). The ratio of the 17 α -hydroxylase and 17,20-lyase activities is an important factor in determining the directions of steroid hormone biosynthesis towards biosynthesis of glucocorticoid or sex hormones.

References: [2571, 4809, 1326, 2210, 3273]

[EC 1.14.14.19 created 1961 as EC 1.99.1.9, transferred 1965 to EC 1.14.1.7, transferred 1972 to EC 1.14.99.9, modified 2013, transferred 2015 to EC 1.14.14.19]

EC 1.14.14.20

Accepted name: phenol 2-monoxygenase (FADH₂)

Reaction: phenol + FADH₂ + O₂ = catechol + FAD + H₂O

Other name(s): *pheA1* (gene name)

Systematic name: phenol,FADH₂:oxygen oxidoreductase (2-hydroxylating)

Comments: The enzyme catalyses the *ortho*-hydroxylation of simple phenols into the corresponding catechols. It accepts 4-methylphenol, 4-chlorophenol, and 4-fluorophenol [2125] as well as 4-nitrophenol, 3-nitrophenol, and resorcinol [3618]. The enzyme is part of a two-component system that also includes an NADH-dependent flavin reductase. It is strictly dependent on FADH₂ and does not accept FMNH₂ [2125, 3618]. *cf.* EC 1.14.13.7, phenol 2-monoxygenase (NADPH).

References: [2125, 4401, 3618]

[EC 1.14.14.20 created 2016]

EC 1.14.14.21

Accepted name: dibenzothiophene monoxygenase

Reaction: dibenzothiophene + 2 FMNH₂ + 2 O₂ = dibenzothiophene-5,5-dioxide + 2 FMN + 2 H₂O (overall reaction)
 (1a) dibenzothiophene + FMNH₂ + O₂ = dibenzothiophene-5-oxide + FMN + H₂O
 (1b) dibenzothiophene-5-oxide + FMNH₂ + O₂ = dibenzothiophene-5,5-dioxide + FMN + H₂O

Other name(s): *dszC* (gene name)

Systematic name: dibenzothiophene,FMNH₂:oxygen oxidoreductase

Comments: This bacterial enzyme catalyses the first two steps in the desulfurization pathway of dibenzothiophenes, the oxidation of dibenzothiophene into dibenzothiophene sulfone via dibenzothiophene-5-oxide. The enzyme forms a two-component system with a dedicated NADH-dependent FMN reductase (EC 1.5.1.42) encoded by the *dszD* gene, which also interacts with EC 1.14.14.22, dibenzothiophene sulfone monoxygenase.

References: [1391, 2523, 1435]

[EC 1.14.14.21 created 2016]

EC 1.14.14.22

Accepted name: dibenzothiophene sulfone monoxygenase

Reaction: dibenzothiophene-5,5-dioxide + FMNH₂ + NADH + O₂ = 2'-hydroxybiphenyl-2-sulfinate + H₂O + FMN + NAD⁺ + H⁺ (overall reaction)
 (1a) FMNH₂ + O₂ = FMN-*N*⁵-peroxide
 (1b) dibenzothiophene-5,5-dioxide + FMN-*N*⁵-peroxide = 2'-hydroxybiphenyl-2-sulfinate + FMN-*N*⁵-oxide

(1c) FMN-*N*⁵-oxide + NADH = FMN + H₂O + NAD⁺ + H⁺ (spontaneous)

- Other name(s):** *dszA* (gene name)
Systematic name: dibenzothiophene-5,5-dioxide,FMNH₂:oxygen oxidoreductase
Comments: This bacterial enzyme catalyses a step in the desulfurization pathway of dibenzothiophenes. The enzyme forms a two-component system with a dedicated NADH-dependent FMN reductase (EC 1.5.1.42) encoded by the *dszD* gene, which also interacts with EC 1.14.14.21, dibenzothiophene monooxygenase. The flavin-*N*⁵-oxide that is formed by the enzyme reacts spontaneously with NADH to give oxidized flavin, releasing a water molecule.
References: [1391, 3146, 2222, 3145, 18, 19, 2723]

[EC 1.14.14.22 created 2016, modified 2019]

EC 1.14.14.23

- Accepted name:** cholesterol 7 α -monooxygenase
Reaction: cholesterol + [reduced NADPH—hemoprotein reductase] + O₂ = 7 α -hydroxycholesterol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): cholesterol 7 α -hydroxylase; CYP7A1 (gene name)
Systematic name: cholesterol,NADPH—hemoprotein reductase:oxygen oxidoreductase (7 α -hydroxylating)
Comments: A P-450 heme-thiolate liver protein that catalyses the first step in the biosynthesis of bile acids. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [2835, 412, 3134, 3060, 3059]

[EC 1.14.14.23 created 1976 as EC 1.14.13.17, transferred 2016 to EC 1.14.14.23]

EC 1.14.14.24

- Accepted name:** vitamin D 25-hydroxylase
Reaction: calcidiol + O₂ + [reduced NADPH—hemoprotein reductase] = calcitriol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): vitamin D₂ 25-hydroxylase; vitamin D₃ 25-hydroxylase; CYP2R1
Systematic name: calcidiol,NADPH—hemoprotein reductase:oxygen oxidoreductase (25-hydroxylating)
Comments: A microsomal enzyme isolated from human and mouse liver that bioactivates vitamin D₃. While multiple isoforms (CYP27A1, CYP2J2/3, CYP3A4, CYP2D25 and CYP2C11) are able to catalyse the reaction *in vitro*, only CYP2R1 is thought to catalyse the reaction in humans *in vivo* [4921]. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [642, 3886, 4075, 4921]

[EC 1.14.14.24 created 2012 as EC 1.14.13.159, transferred 2016 to EC 1.14.14.24]

EC 1.14.14.25

- Accepted name:** cholesterol 24-hydroxylase
Reaction: cholesterol + [reduced NADPH—hemoprotein reductase] + O₂ = (24*S*)-cholest-5-ene-3 β ,24-diol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): cholesterol 24-monooxygenase; CYP46; CYP46A1; cholesterol 24*S*-hydroxylase; cytochrome P450 46A1
Systematic name: cholesterol,NADPH—hemoprotein reductase:oxygen oxidoreductase (24-hydroxylating)
Comments: A P-450 heme-thiolate protein. The enzyme can also produce 25-hydroxycholesterol. In addition, it can further hydroxylate the product to 24,25-dihydroxycholesterol and 24,27-dihydroxycholesterol [371]. This reaction is the first step in the enzymic degradation of cholesterol in the brain as hydroxycholesterol can pass the blood—brain barrier whereas cholesterol cannot [2689]. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase [2689].
References: [2562, 371, 2689, 2564, 3607]

[EC 1.14.14.25 created 2005 as EC 1.14.13.98, transferred 2016 to EC 1.14.14.25]

EC 1.14.14.26

- Accepted name:** 24-hydroxycholesterol 7 α -hydroxylase
Reaction: (24*S*)-cholest-5-ene-3 β ,24-diol + [reduced NADPH—hemoprotein reductase] + O₂ = (24*S*)-cholest-5-ene-3 β ,7 α ,24-triol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): 24-hydroxycholesterol 7 α -monooxygenase; CYP39A1; CYP39A1 oxysterol 7 α -hydroxylase
Systematic name: (24*S*)-cholest-5-ene-3 β ,24-diol,NADPH—hemoprotein reductase:oxygen oxidoreductase (7 α -hydroxylating)
Comments: A P-450 heme-thiolate protein that is found in liver microsomes and in ciliary non-pigmented epithelium [1792]. The enzyme is specific for (24*S*)-cholest-5-ene-3 β ,24-diol, which is formed mostly in the brain by EC 1.14.14.25, cholesterol 24-hydroxylase. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [2469, 1792, 3607]

[EC 1.14.14.26 created 2005 as EC 1.14.13.99, transferred 2016 to EC 1.14.14.26]

EC 1.14.14.27

- Accepted name:** resorcinol 4-hydroxylase (FADH₂)
Reaction: resorcinol + FADH₂ + O₂ = hydroxyquinol + FAD + H₂O
Other name(s): *graA* (gene name)
Systematic name: resorcinol,FADH₂:oxygen oxidoreductase (4-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Rhizobium* sp. strain MTP-10005, uses FADH₂ as a substrate rather than a cofactor. FADH₂ is provided by a dedicated EC 1.5.1.36, flavin reductase (NADH). The enzyme participates in the degradation of γ -resorcyate and resorcinol. *cf.* EC 1.14.13.220, resorcinol 4-hydroxylase (NADH), and EC 1.14.13.219, resorcinol 4-hydroxylase (NADPH).
References: [3148, 4810]

[EC 1.14.14.27 created 2016]

EC 1.14.14.28

- Accepted name:** long-chain alkane monooxygenase
Reaction: a long-chain alkane + FMNH₂ + O₂ = a long-chain primary alcohol + FMN + H₂O
Systematic name: long-chain-alkane,FMNH₂:oxygen oxidoreductase
Comments: The enzyme, characterized from the bacterium *Geobacillus thermodenitrificans* NG80-2, is capable of converting alkanes ranging from C₁₅ to C₃₆ into their corresponding primary alcohols [1100, 2449]. The FMNH₂ cofactor is provided by an FMN reductase [949].
References: [1100, 2449, 949]

[EC 1.14.14.28 created 2016]

EC 1.14.14.29

- Accepted name:** 25/26-hydroxycholesterol 7 α -hydroxylase
Reaction: (1) cholest-5-ene-3 β ,25-diol + [reduced NADPH—hemoprotein reductase] + O₂ = cholest-5-ene-3 β ,7 α ,25-triol + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) (25*R*)-cholest-5-ene-3 β ,26-diol + [reduced NADPH—hemoprotein reductase] + O₂ = (25*R*)-cholest-5-ene-3 β ,7 α ,26-triol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): 25-hydroxycholesterol 7 α -monooxygenase; CYP7B1; CYP7B1 oxysterol 7 α -hydroxylase; 27-hydroxycholesterol 7-monooxygenase; 27-hydroxycholesterol 7 α -hydroxylase; cholest-5-ene-3 β ,25-diol,NADPH:oxygen oxidoreductase (7 α -hydroxylating); 25-hydroxycholesterol 7 α -hydroxylase
Systematic name: cholest-5-ene-3 β ,25/26-diol,[NADPH—hemoprotein reductase]:oxygen oxidoreductase (7 α -hydroxylating)

- Comments:** A P-450 (heme-thiolate) protein. Unlike EC 1.14.14.26, 24-hydroxycholesterol 7 α -monooxygenase, which is specific for its oxysterol substrate, this enzyme can also metabolize the oxysterols 24,25-epoxycholesterol, 22-hydroxycholesterol and 24-hydroxycholesterol, but to a lesser extent [4302]. The direct electron donor to the enzyme is EC 1.6.2.4, NADPH—hemoprotein reductase.
- References:** [2285, 4302, 2469, 3498, 3607]

[EC 1.14.14.29 created 2005 as EC 1.14.13.100, modified 2013 (EC 1.14.13.60 created 1999, incorporated 2013), transferred 2016 to EC 1.14.14.29]

EC 1.14.14.30

- Accepted name:** isobutylamine *N*-monooxygenase
- Reaction:** (1) 2-methylpropan-1-amine + FADH₂ + O₂ = *N*-(2-methylpropyl)hydroxylamine + FAD + H₂O
(2) 2-methylpropan-1-amine + FMNH₂ + O₂ = *N*-(2-methylpropyl)hydroxylamine + FMN + H₂O
- Other name(s):** *vlmH* (gene name)
- Systematic name:** 2-methylpropan-1-amine,FADH₂:O₂ *N*-oxidoreductase
- Comments:** The enzyme, characterized from the bacterium *Streptomyces viridifaciens*, is part of a two component system that also includes a flavin reductase, which provides reduced flavin mononucleotide for this enzyme. The enzyme, which is involved in the biosynthesis of the azoxy antibiotic valanimycin, has a similar activity with either FMNH₂ or FADH₂. It exhibits broad specificity, and also accepts propan-1-amine, butan-1-amine, butan-2-amine and benzylamine.
- References:** [3244, 3245, 3243]

[EC 1.14.14.30 created 2016, modified 2017]

EC 1.14.14.31

- Accepted name:** ipsdienol synthase
- Reaction:** myrcene + [reduced NADPH—hemoprotein reductase] + O₂ = (*R*)-ipsdienol + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** myrcene hydroxylase; CYP9T2; CYP9T3
- Systematic name:** myrcene,NADPH—hemoprotein reductase:O₂ oxidoreductase (hydroxylating)
- Comments:** A cytochrome P-450 heme-thiolate protein. Involved in the insect aggregation pheromone production. Isolated from the pine engraver beetle, *Ips pini*. A small amount of (*S*)-ipsdienol is also formed. *In vitro* it also hydroxylated (+)- and (–)- α -pinene, 3-carene, and (+)-limonene, but not α -phellandrene, (–)- β -pinene, γ -terpinene, or terpinolene.
- References:** [3656, 3962]

[EC 1.14.14.31 created 2015 as EC 1.14.13.207, transferred 2016 to EC 1.14.14.31]

EC 1.14.14.32

- Accepted name:** 17 α -hydroxyprogesterone deacetylase
- Reaction:** (1) 17 α -hydroxyprogesterone + [reduced NADPH—hemoprotein reductase] + O₂ = androstenedione + acetate + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) 17 α -hydroxypregnenolone + [reduced NADPH—hemoprotein reductase] + O₂ = 3 β -hydroxyandrost-5-en-17-one + acetate + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** C-17/C-20 lyase; 17 α -hydroxyprogesterone acetaldehyde-lyase; CYP17; CYP17A1 (gene name); 17 α -hydroxyprogesterone 17,20-lyase
- Systematic name:** 17 α -hydroxyprogesterone,NADPH—hemoprotein reductase:oxygen oxidoreductase (17 α -hydroxylating, acetate-releasing)

Comments: A microsomal cytochrome *P*-450 (heme-thiolate) protein that catalyses two independent reactions at the same active site - the 17-hydroxylation of pregnenolone and progesterone, which is part of glucocorticoid hormones biosynthesis (EC 1.14.14.19), and the conversion of the 17-hydroxylated products via a 17,20-lyase reaction to form androstenedione and 3 β -hydroxyandrost-5-en-17-one, leading to sex hormone biosynthesis. The activity of this reaction is dependent on the allosteric interaction of the enzyme with cytochrome *b*₅ without any transfer of electrons from the cytochrome [153, 3906]. The enzymes from different organisms differ in their substrate specificity. While the enzymes from pig, hamster, and rat accept both 17 α -hydroxyprogesterone and 17 α -hydroxypregnenolone, the enzymes from human, bovine, sheep, goat, and bison do not accept the former, and the enzyme from guinea pig does not accept the latter [1326].

References: [1326, 153, 2624, 3906, 328]

[EC 1.14.14.32 created 1976 as EC 4.1.2.30, transferred 2016 to EC 1.14.14.32]

EC 1.14.14.33

Accepted name: ethylenediaminetetraacetate monooxygenase

Reaction: ethylenediaminetetraacetate + 2 FMNH₂ + 2 O₂ = ethylenediamine-*N,N'*-diacetate + 2 glyoxylate + 2 FMN + 2 H₂O (overall reaction)

(1a) ethylenediaminetetraacetate + FMNH₂ + O₂ = ethylenediaminetriacetate + glyoxylate + FMN + H₂O

(1b) ethylenediaminetriacetate + FMNH₂ + O₂ = ethylenediamine-*N,N'*-diacetate + glyoxylate + FMN + H₂O

Systematic name: ethylenediaminetetraacetate,FMNH₂:O₂ oxidoreductase (glyoxylate-forming)

Comments: The enzyme is part of a two component system that also includes EC 1.5.1.42, FMN reductase (NADH), which provides reduced flavin mononucleotide for this enzyme. It acts on EDTA only when it is complexed with divalent cations such as Mg²⁺, Zn²⁺, Mn²⁺, Co²⁺, or Cu²⁺. While the enzyme has a substrate overlap with EC 1.14.14.10, nitrilotriacetate monooxygenase, it has a much wider substrate range, which includes nitrilotriacetate (NTA) and diethylenetriaminepentaacetate (DTPA) in addition to EDTA.

References: [4647, 3268, 374]

[EC 1.14.14.33 created 2016]

EC 1.14.14.34

Accepted name: methanesulfonate monooxygenase (FMNH₂)

Reaction: methanesulfonate + FMNH₂ + O₂ = formaldehyde + FMN + sulfite + H₂O

Other name(s): *msuD* (gene name); *ssuD* (gene name)

Systematic name: methanesulfonate,FMNH₂:oxygen oxidoreductase

Comments: The enzyme, characterized from *Pseudomonas* strains, allows the organisms to utilize methanesulfonate as their sulfur source. It acts in combination with a dedicated NADH-dependent FMN reductase (EC 1.5.1.42), which provides it with reduced FMN. *cf.* EC 1.14.13.111, methanesulfonate monooxygenase (NADH).

References: [2072, 1046]

[EC 1.14.14.34 created 2016]

EC 1.14.14.35

Accepted name: dimethylsulfone monooxygenase

Reaction: dimethyl sulfone + FMNH₂ + O₂ = methanesulfinate + formaldehyde + FMN + H₂O

Other name(s): *sfnG* (gene name)

Systematic name: dimethyl sulfone,FMNH₂:oxygen oxidoreductase

Comments: The enzyme, characterized from *Pseudomonas* spp., is involved in a dimethyl sulfide degradation pathway. It is dependent on NAD(P)H-dependent FMN reductase (EC 1.5.1.38, EC 1.5.1.39, or EC 1.5.1.42), which provides it with reduced FMN. The product, methanesulfinate, is oxidized spontaneously to methanesulfonate in the presence of dioxygen and FMNH₂.

References: [1045, 4616]

[EC 1.14.14.35 created 2016]

EC 1.14.14.36

Accepted name: tyrosine *N*-monooxygenase

Reaction: L-tyrosine + 2 O₂ + 2 [reduced NADPH—hemoprotein reductase] = (*E*)-[4-hydroxyphenylacetaldehyde oxime] + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)
(1a) L-tyrosine + O₂ + [reduced NADPH—hemoprotein reductase] = *N*-hydroxy-L-tyrosine + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) *N*-hydroxy-L-tyrosine + O₂ + [reduced NADPH—hemoprotein reductase] = *N,N*-dihydroxy-L-tyrosine + [oxidized NADPH—hemoprotein reductase] + H₂O
(1c) *N,N*-dihydroxy-L-tyrosine = (*E*)-[4-hydroxyphenylacetaldehyde oxime] + CO₂ + H₂O

Other name(s): tyrosine *N*-hydroxylase; CYP79A1

Systematic name: L-tyrosine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from *Sorghum* is involved in the biosynthesis of the cyanogenic glucoside dhurrin. In *Sinapis alba* (white mustard) the enzyme is involved in the biosynthesis of the glucosinolate sinalbin.

References: [1481, 3897, 289, 1978, 187, 3068, 510, 2265, 699]

[EC 1.14.14.36 created 1992 as EC 1.14.13.41, modified 2001, modified 2005, transferred 2016 to EC 1.14.14.36]

EC 1.14.14.37

Accepted name: 4-hydroxyphenylacetaldehyde oxime monooxygenase

Reaction: (*E*)-4-hydroxyphenylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-4-hydroxymandelonitrile + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) (*E*)-4-hydroxyphenylacetaldehyde oxime = (*Z*)-4-hydroxyphenylacetaldehyde oxime
(1b) (*Z*)-4-hydroxyphenylacetaldehyde oxime = 4-hydroxyphenylacetonitrile + H₂O
(1c) 4-hydroxyphenylacetonitrile + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-4-hydroxymandelonitrile + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): 4-hydroxybenzeneacetaldehyde oxime monooxygenase; cytochrome P450II-dependent monooxygenase; NADPH-cytochrome P450 reductase (CYP71E1); CYP71E1; 4-hydroxyphenylacetaldehyde oxime,NADPH:oxygen oxidoreductase

Systematic name: (*E*)-4-hydroxyphenylacetaldehyde oxime,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase

Comments: This cytochrome *P*-450 (heme thiolate) enzyme is involved in the biosynthesis of the cyanogenic glucoside dhurrin in sorghum. It catalyses three different activities - isomerization of the (*E*) isomer to the (*Z*) isomer, dehydration, and C-hydroxylation.

References: [2590, 3863, 510, 2265, 699]

[EC 1.14.14.37 created 2000 as EC 1.14.13.68, modified 2005, transferred 2016 to EC 1.14.14.37]

EC 1.14.14.38

Accepted name: valine *N*-monooxygenase

Reaction: L-valine + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = (*E*)-2-methylpropanal oxime + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)
(1a) L-valine + [reduced NADPH—hemoprotein reductase] + O₂ = *N*-hydroxy-L-valine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) *N*-hydroxy-L-valine + [reduced NADPH—hemoprotein reductase] + O₂ = *N,N*-dihydroxy-L-valine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1c) *N,N*-dihydroxy-L-valine = (*E*)-2-methylpropanal oxime + CO₂ + H₂O

Other name(s): CYP79D1; CYP79D2

Systematic name: L-valine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. This enzyme catalyses two successive *N*-hydroxylations of L-valine, the committed step in the biosynthesis of the cyanogenic glucoside linamarin in *Manihot esculenta* (cassava). The product of the two hydroxylations, *N,N*-dihydroxy-L-valine, is labile and undergoes dehydration and decarboxylation that produce the (*E*) isomer of the oxime. It is still not known whether the decarboxylation is spontaneous or catalysed by the enzyme. The enzyme can also accept L-isoleucine as substrate, with a lower activity. It is different from EC 1.14.14.39, isoleucine *N*-monooxygenase, which prefers L-isoleucine.

References: [90, 1150]

[EC 1.14.14.38 created 2010 as EC 1.14.13.118, transferred 2017 to EC 1.14.14.38]

EC 1.14.14.39

Accepted name: isoleucine *N*-monooxygenase

Reaction: L-isoleucine + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = (*1E,2S*)-2-methylbutanal oxime + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)

(1a) L-isoleucine + [reduced NADPH—hemoprotein reductase] + O₂ = *N*-hydroxy-L-isoleucine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) *N*-hydroxy-L-isoleucine + [reduced NADPH—hemoprotein reductase] + O₂ = *N,N*-dihydroxy-L-isoleucine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1c) *N,N*-dihydroxy-L-isoleucine = (*1E,2S*)-2-methylbutanal oxime + CO₂ + H₂O (spontaneous)

Other name(s): CYP79D3 (gene name); CYP79D4 (gene name)

Systematic name: L-isoleucine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)

Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, found in plants, catalyses two successive *N*-hydroxylations of L-isoleucine, the committed step in the biosynthesis of the cyanogenic glucoside lotaustralin. The product of the two hydroxylations, *N,N*-dihydroxy-L-isoleucine, is labile and undergoes dehydration followed by decarboxylation, producing the oxime. It is still not known whether the decarboxylation is spontaneous or catalysed by the enzyme. The enzyme can also accept L-valine, but with a lower activity. *cf.* EC 1.14.14.38, valine *N*-monooxygenase.

References: [90, 1150]

[EC 1.14.14.39 created 2010 as EC 1.14.13.117, transferred 2017 to EC 1.14.14.39]

EC 1.14.14.40

Accepted name: phenylalanine *N*-monooxygenase

Reaction: L-phenylalanine + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = (*E*)-phenylacetaldoxime + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)

(1a) L-phenylalanine + [reduced NADPH—hemoprotein reductase] + O₂ = *N*-hydroxy-L-phenylalanine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) *N*-hydroxy-L-phenylalanine + [reduced NADPH—hemoprotein reductase] + O₂ = *N,N*-dihydroxy-L-phenylalanine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1c) *N,N*-dihydroxy-L-phenylalanine = (*E*)-phenylacetaldoxime + CO₂ + H₂O

Other name(s): phenylalanine *N*-hydroxylase; CYP79A2 (gene name); CYP79D16 (gene name)

Systematic name: L-phenylalanine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)

Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, found in plants, catalyses two successive *N*-hydroxylations of L-phenylalanine, a committed step in the biosynthesis of benzylglucosinolate and the cyanogenic glucosides (*R*)-prunasin and (*R*)-amygdalin. The product of the two hydroxylations, *N,N*-dihydroxy-L-phenylalanine, is labile and undergoes dehydration followed by decarboxylation, producing an oxime. It is still not known whether the decarboxylation is spontaneous or catalysed by the enzyme.

References: [4648, 4734]

[EC 1.14.14.40 created 2011 as EC 1.14.13.124, transferred 2017 to EC 1.14.14.40]

EC 1.14.14.41

Accepted name: (*E*)-2-methylbutanal oxime monooxygenase

Reaction: (1) (*E*)-2-methylbutanal oxime + [reduced NADPH—hemoprotein reductase] + O₂ = 2-hydroxy-2-methylbutanenitrile + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)

(1a) (*E*)-2-methylbutanal oxime = (*Z*)-2-methylbutanal oxime

(1b) (*Z*)-2-methylbutanal oxime = 2-methylbutanenitrile + H₂O

(1c) 2-methylbutanenitrile + [reduced NADPH—hemoprotein reductase] + O₂ = 2-hydroxy-2-methylbutanenitrile + [oxidized NADPH—hemoprotein reductase] + H₂O

(2) (*E*)-2-methylpropanal oxime + [reduced NADPH—hemoprotein reductase] + O₂ = 2-hydroxy-2-methylpropanenitrile + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)

(2a) (*E*)-2-methylpropanal oxime = (*Z*)-2-methylpropanal oxime

(2b) (*Z*)-2-methylpropanal oxime = 2-methylpropanenitrile + H₂O

(2c) 2-methylpropanenitrile + [reduced NADPH—hemoprotein reductase] + O₂ = 2-hydroxy-2-methylpropanenitrile + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP71E7 (gene name)

Systematic name: (*E*)-2-methylbutanal oxime,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase

Comments: This cytochrome *P*-450 (heme thiolate) enzyme is involved in the biosynthesis of the cyanogenic glucosides lotaustralin and linamarin. It catalyses three different activities - isomerization of its substrate, the (*E*) isomer, to the (*Z*) isomer, dehydration, and C-hydroxylation.

References: [1949]

[EC 1.14.14.41 created 2017]

EC 1.14.14.42

Accepted name: homomethionine *N*-monooxygenase

Reaction: an L-polyhomomethionine + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = an (*E*)- ω -(methylsulfanyl)alkanal oxime + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)

(1a) an L-polyhomomethionine + [reduced NADPH—hemoprotein reductase] + O₂ = an L-*N*-hydroxypolyhomomethionine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) an L-*N*-hydroxypolyhomomethionine + [reduced NADPH—hemoprotein reductase] + O₂ = an L-*N,N*-dihydroxypolyhomomethionine + [oxidized NADPH—hemoprotein reductase] + H₂O

(1c) an L-*N,N*-dihydroxypolyhomomethionine = an (*E*)- ω -(methylsulfanyl)alkanal oxime + CO₂ + H₂O

Other name(s): CYP79F1 (gene name); CYP79F2 (gene name)

Systematic name: L-polyhomomethionine,[NADPH—hemoprotein reductase]:oxygen oxidoreductase

Comments: This plant cytochrome *P*-450 (heme thiolate) enzyme is involved in methionine-derived aliphatic glucosinolates biosynthesis. It catalyses two successive *N*-hydroxylations, which are followed by dehydration and decarboxylation. CYP79F1 from *Arabidopsis thaliana* can metabolize mono-, di-, tri-, tetra-, penta-, and hexahomomethionine to their corresponding aldoximes, while CYP79F2 from the same plant can only metabolize penta- and hexahomomethionine.

References: [1511, 636]

[EC 1.14.14.42 created 2017]

EC 1.14.14.43

- Accepted name:** (methylsulfanyl)alkanaldoxime *N*-monooxygenase
- Reaction:** an (*E*)- ω -(methylsulfanyl)alkanal oxime + [reduced NADPH—hemoprotein reductase] + glutathione + O₂ = an *S*-[(1*E*)-1-(hydroxyimino)- ω -(methylsulfanyl)alkyl]-L-glutathione + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) an (*E*)- ω -(methylsulfanyl)alkanal oxime + [reduced NADPH—hemoprotein reductase] + O₂ = a 1-(methylsulfanyl)-4-*aci*-nitroalkane + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) a 1-(methylsulfanyl)-4-*aci*-nitroalkane + glutathione = an *S*-[(1*E*)-1-(hydroxyimino)- ω -(methylsulfanyl)alkyl]-L-glutathione + H₂O
- Other name(s):** CYP83A1 (gene name); (methylthio)alkanaldoxime *N*-monooxygenase; (*E*)- ω -(methylthio)alkanaldoxime,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)
- Systematic name:** (*E*)- ω -(methylsulfanyl)alkanaldoxime,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)
- Comments:** This cytochrome *P*-450 (heme thiolate) enzyme is involved in the biosynthesis of glucosinolates in plants. The enzyme catalyses an *N*-hydroxylation of the *E* isomer of ω -(methylsulfanyl)alkanal oximes, forming an *aci*-nitro intermediate that reacts non-enzymically with glutathione to produce an *N*-alkyl-thiohydroximate adduct, the committed precursor of glucosinolates. In the absence of a thiol compound, the enzyme is suicidal, probably due to interaction of the reactive *aci*-nitro intermediate with active site residues.
- References:** [188, 3024, 699]

[EC 1.14.14.43 created 2017]

EC 1.14.14.44

- Accepted name:** phenylacetaldehyde oxime monooxygenase
- Reaction:** (*E*)-phenylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + O₂ = (*R*)-mandelonitrile + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) (*E*)-phenylacetaldehyde oxime = (*Z*)-phenylacetaldehyde oxime
(1b) (*Z*)-phenylacetaldehyde oxime = phenylacetone + H₂O
(1c) phenylacetone + [reduced NADPH—hemoprotein reductase] + O₂ = (*R*)-mandelonitrile + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** CYP71AN24 (gene name)
- Systematic name:** (*E*)-phenylacetaldehyde oxime,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
- Comments:** This cytochrome *P*-450 (heme-thiolate) enzyme is involved in the biosynthesis of the cyanogenic glucosides (*R*)-prunasin and (*R*)-amygdalin. It catalyses three different activities - isomerization of the (*E*) isomer to the (*Z*) isomer, dehydration, and C-hydroxylation.
- References:** [4734]

[EC 1.14.14.44 created 2017]

EC 1.14.14.45

- Accepted name:** aromatic aldoxime *N*-monooxygenase
- Reaction:** (1) (*E*)-indol-3-ylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + glutathione + O₂ = *S*-[(*E*)-*N*-hydroxy(indol-3-yl)acetimidoyl]-L-glutathione + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) (*E*)-indol-3-ylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + O₂ = 1-(1*H*-indol-3-yl)-2-*aci*-nitroethane + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 1-(1*H*-indol-3-yl)-2-*aci*-nitroethane + glutathione = *S*-[(*E*)-*N*-hydroxy(indol-3-yl)acetimidoyl]-L-glutathione + H₂O (spontaneous)
(2) (*E*)-phenylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + glutathione + O₂ = *S*-[(*Z*)-*N*-hydroxy(phenyl)acetimidoyl]-L-glutathione + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)

(2a) (*E*)-phenylacetaldehyde oxime + [reduced NADPH—hemoprotein reductase] + O₂ = 1-*aci*-nitro-2-phenylethane + [oxidized NADPH—hemoprotein reductase] + H₂O
(2b) 1-*aci*-nitro-2-phenylethane + glutathione = *S*-[(*Z*)-*N*-hydroxy(phenyl)acetimidoyl]-L-glutathione + H₂O (spontaneous)

Other name(s): CYP83B1 (gene name)
Systematic name: (*E*)-indol-3-ylacetaldoxime,[reduced NADPH—hemoprotein reductase],glutathione:oxygen oxidoreductase (oxime-hydroxylating)
Comments: This cytochrome *P*-450 (heme thiolate) enzyme is involved in the biosynthesis of glucosinolates in plants. The enzyme catalyses the *N*-hydroxylation of aromatic aldoximes derived from L-tryptophan, L-phenylalanine, and L-tyrosine, forming an *aci*-nitro intermediate that reacts non-enzymically with glutathione to produce an *N*-alkyl-thiohydroximate adduct, the committed precursor of glucosinolates. In the absence of glutathione, the enzyme is suicidal, probably due to interaction of the reactive *aci*-nitro compound with catalytic residues in the active site.
References: [188, 3024, 1311]

[EC 1.14.14.45 created 2017]

EC 1.14.14.46

Accepted name: pimeloyl-[acyl-carrier protein] synthase
Reaction: a long-chain acyl-[acyl-carrier protein] + 2 reduced flavodoxin + 3 O₂ = pimeloyl-[acyl-carrier protein] + an *n*-alkanal + 2 oxidized flavodoxin + 3 H₂O (overall reaction)
(1a) a long-chain acyl-[acyl-carrier protein] + reduced flavodoxin + O₂ = a (*7S*)-7-hydroxy-long-chain-acyl-[acyl-carrier protein] + oxidized flavodoxin + H₂O
(1b) a (*7S*)-7-hydroxy-long-chain-acyl-[acyl-carrier protein] + reduced flavodoxin + O₂ = a (*7R,8R*)-7,8-dihydroxy-long-chain-acyl-[acyl-carrier protein] + oxidized flavodoxin + H₂O
(1c) a (*7R,8R*)-7,8-dihydroxy-long-chain-acyl-[acyl-carrier protein] + reduced flavodoxin + O₂ = a 7-oxoheptanoyl-[acyl-carrier protein] + an *n*-alkanal + oxidized flavodoxin + 2 H₂O
(1d) a 7-oxoheptanoyl-[acyl-carrier protein] + oxidized flavodoxin + H₂O = a pimeloyl-[acyl-carrier protein] + reduced flavodoxin + H⁺
Other name(s): *bioI* (gene name); *P450BioI*; CYP107H1
Systematic name: acyl-[acyl-carrier protein],reduced-flavodoxin:oxygen oxidoreductase (pimeloyl-[acyl-carrier protein]-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme catalyses an oxidative C-C bond cleavage of long-chain acyl-[acyl-carrier protein]s of various lengths to generate pimeloyl-[acyl-carrier protein], an intermediate in the biosynthesis of biotin. The preferred substrate of the enzyme from the bacterium *Bacillus subtilis* is palmitoyl-[acyl-carrier protein] which then gives heptanal as the alkanal. The mechanism is similar to EC 1.14.15.6, cholesterol monooxygenase (side-chain-cleaving), followed by a hydroxylation step, which may occur spontaneously [780].
References: [4045, 780, 779, 777]

[EC 1.14.14.46 created 2013 as EC 1.14.15.12, transferred 2017 to EC 1.14.14.46]

EC 1.14.14.47

Accepted name: nitric-oxide synthase (flavodoxin)
Reaction: 2 L-arginine + 3 reduced flavodoxin + 4 O₂ = 2 L-citrulline + 2 nitric oxide + 3 oxidized flavodoxin + 4 H₂O (overall reaction)
(1a) 2 L-arginine + 2 reduced flavodoxin + 2 O₂ = 2 *N*^ω-hydroxy-L-arginine + 2 oxidized flavodoxin + 2 H₂O
(1b) 2 *N*^ω-hydroxy-L-arginine + reduced flavodoxin + 2 O₂ = 2 L-citrulline + 2 nitric oxide + oxidized flavodoxin + 2 H₂O
Other name(s): nitric oxide synthetase (ambiguous); NO synthase (ambiguous)
Systematic name: L-arginine,reduced-flavodoxin:oxygen oxidoreductase (nitric-oxide-forming)

Comments: Binds heme (iron protoporphyrin IX) and tetrahydrobiopterin. The enzyme, found in bacteria and archaea, consist of only an oxygenase domain and functions together with bacterial ferredoxins or flavodoxins. The orthologous enzymes from plants and animals also contain a reductase domain and use only NADPH as the electron donor (*cf.* EC 1.14.13.39).

References: [3233, 17, 4540, 31, 1692]

[EC 1.14.14.47 created 2012 as EC 1.14.13.165, transferred 2017 to EC 1.14.14.47]

EC 1.14.14.48

Accepted name: jasmonoyl-L-amino acid 12-hydroxylase

Reaction: a jasmonoyl-L-amino acid + [reduced NADPH—hemoprotein reductase] + O₂ = a 12-hydroxyjasmonoyl-L-amino acid + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP94B1 (gene name); CYP94B3 (gene name)

Systematic name: jasmonoyl-L-amino acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12-hydroxylating)

Comments: A cytochrome P450 (heme thiolate) enzyme found in plants. The enzyme acts on jasmonoyl-L-amino acid conjugates, catalysing the hydroxylation of the C-12 position of jasmonic acid. While the best studied substrate is (+)-7-*epi*-jasmonoyl-L-isoleucine, the enzyme was shown to be active with jasmonoyl-L-valine and jasmonoyl-L-phenylalanine, and is likely to be active with other jasmonoyl-amino acid conjugates.

References: [2225, 2136, 1615, 2135, 2226, 4619]

[EC 1.14.14.48 created 2017]

EC 1.14.14.49

Accepted name: 12-hydroxyjasmonoyl-L-amino acid 12-hydroxylase

Reaction: a 12-hydroxyjasmonoyl-L-amino acid + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = a 12-hydroxy-12-oxojasmonoyl-L-amino acid + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)

(1a) a 12-hydroxyjasmonoyl-L-amino acid + [reduced NADPH—hemoprotein reductase] + O₂ = a 12-oxojasmonoyl-L-amino acid + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

(1b) a 12-oxojasmonoyl-L-amino acid + [reduced NADPH—hemoprotein reductase] + O₂ = a 12-hydroxy-12-oxojasmonoyl-L-amino acid + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP94C1 (gene name)

Systematic name: 12-hydroxyjasmonoyl-L-amino acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12-hydroxylating)

Comments: A cytochrome P450 (heme thiolate) enzyme found in plants. The enzyme acts on jasmonoyl-L-amino acid conjugates that have been hydroxylated at the C-12 position of jasmonic acid by EC 1.14.14.48, jasmonoyl-L-amino acid 12-hydroxylase, further oxidizing that position to a carboxylate via an aldehyde intermediate. While the best studied substrate is (+)-7-*epi*-jasmonoyl-L-isoleucine, the enzyme was shown to be active with jasmonoyl-L-phenylalanine, and is likely to be active with other jasmonoyl-amino acid conjugates.

References: [1615, 4619, 465]

[EC 1.14.14.49 created 2017]

EC 1.14.14.50

Accepted name: tabersonine 3-oxygenase

Reaction: (1) 16-methoxytabersonine + [reduced NADPH—hemoprotein reductase] + O₂ = (3*R*)-3-hydroxy-16-methoxy-1,2-didehydro-2,3-dihydrotabersonine + [oxidized NADPH—hemoprotein reductase] + H₂O

(2) tabersonine + [reduced NADPH—hemoprotein reductase] + O₂ = (3*R*)-3-hydroxy-1,2-didehydro-2,3-dihydrotabersonine + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): T3O; CYP71D1V2
Systematic name: 16-methoxytabersonine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3-hydroxylating)
Comments: This cytochrome *P*-450 (heme thiolate) enzyme acts on 16-methoxytabersonine, leading to biosynthesis of vindoline in the plant *Catharanthus roseus* (Madagascar periwinkle). It can also act on tabersonine, resulting in the production of small amounts of vindorosine. The products are unstable and, in the absence of EC 1.1.99.41, 3-hydroxy-1,2-didehydro-2,3-dihydrotabersonine reductase, will convert into 3-epoxylated compounds.
References: [3407]

[EC 1.14.14.50 created 2017]

EC 1.14.14.51

Accepted name: (*S*)-limonene 6-monooxygenase
Reaction: (*S*)-limonene + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-*trans*-carveol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): (–)-limonene 6-hydroxylase; (–)-limonene 6-monooxygenase; (–)-limonene,NADPH:oxygen oxidoreductase (6-hydroxylating)
Systematic name: (*S*)-limonene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6-hydroxylating)
Comments: A cytochrome *P*-450 (heme thiolate) enzyme. The enzyme participates in the biosynthesis of (–)-carvone, which is responsible for the aroma of spearmint.
References: [1996]

[EC 1.14.14.51 created 1992 as EC 1.14.13.48, modified 2003, transferred 2017 to EC 1.14.14.51]

EC 1.14.14.52

Accepted name: (*S*)-limonene 7-monooxygenase
Reaction: (*S*)-limonene + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-perillyl alcohol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): (–)-limonene 7-monooxygenase; (–)-limonene hydroxylase; (–)-limonene monooxygenase; (–)-limonene,NADPH:oxygen oxidoreductase (7-hydroxylating)
Systematic name: (*S*)-limonene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (7-hydroxylating)
Comments: A cytochrome *P*-450 (heme thiolate) enzyme. The enzyme, characterized from the plant *Perilla frutescens*, participates in the biosynthesis of perillyl aldehyde, the major constituent of the essential oil that accumulates in the glandular trichomes of this plant. Some forms of the enzyme also catalyse the oxidation of (–)-perillyl alcohol to (–)-perillyl aldehyde.
References: [1996, 2726, 1217]

[EC 1.14.14.52 created 1992 as EC 1.14.13.49, modified 2003, transferred 2017 to EC 1.14.14.52]

EC 1.14.14.53

Accepted name: (*R*)-limonene 6-monooxygenase
Reaction: (*R*)-limonene + [reduced NADPH—hemoprotein reductase] + O₂ = (+)-*trans*-carveol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): (+)-limonene-6-hydroxylase; (+)-limonene 6-monooxygenase
Systematic name: (*R*)-limonene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6-hydroxylating)
Comments: The reaction is stereospecific with over 95% yield of (+)-*trans*-carveol from (*R*)-limonene. (*S*)-Limonene, the substrate for EC 1.14.14.51, (*S*)-limonene 6-monooxygenase, is not a substrate. Forms part of the carveone biosynthesis pathway in *Carum carvi* (caraway) seeds.
References: [407, 408]

[EC 1.14.14.53 created 2003 as EC 1.14.13.80, transferred 2017 to EC 1.14.14.53]

EC 1.14.14.54

- Accepted name:** phenylacetate 2-hydroxylase
Reaction: phenylacetate + [reduced NADPH—hemoprotein reductase] + O₂ = (2-hydroxyphenyl)acetate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP504; *phaA* (gene name)
Systematic name: phenylacetate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2-hydroxylating)
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, found in *Aspergillus nidulans*, is involved in the degradation of phenylacetate.
References: [2818, 3549]

[EC 1.14.14.54 created 2017]

EC 1.14.14.55

- Accepted name:** quinine 3-monooxygenase
Reaction: quinine + [reduced NADPH—hemoprotein reductase] + O₂ = 3-hydroxyquinine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP3A4 (gene name)
Systematic name: quinine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein.
References: [3494, 4873, 4904, 4905]

[EC 1.14.14.55 created 2000 as EC 1.14.13.67, transferred 2017 to EC 1.14.14.55]

EC 1.14.14.56

- Accepted name:** 1,8-cineole 2-*exo*-monooxygenase
Reaction: 1,8-cineole + [reduced NADPH—hemoprotein reductase] + O₂ = 2-*exo*-hydroxy-1,8-cineole + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP3A4
Systematic name: 1,8-cineole,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2-*exo*-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The mammalian enzyme, expressed in liver microsomes, performs a variety of oxidation reactions of structurally unrelated compounds, including steroids, fatty acids, and xenobiotics. *cf.* EC 1.14.14.55, quinine 3-monooxygenase, EC 1.14.14.57, taurochenodeoxycholate 6-hydroxylase and EC 1.14.14.73, albendazole monooxygenase (sulfoxide-forming).
References: [2845, 2844, 2846]

[EC 1.14.14.56 created 2012 as EC 1.14.13.157, transferred 2017 to EC 1.14.14.56, modified 2018]

EC 1.14.14.57

- Accepted name:** taurochenodeoxycholate 6 α -hydroxylase
Reaction: (1) taurochenodeoxycholate + [reduced NADPH—hemoprotein reductase] + O₂ = taurohyocholate + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) lithocholate + [reduced NADPH—hemoprotein reductase] + O₂ = hyodeoxycholate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP3A4; CYP4A21; taurochenodeoxycholate 6 α -monooxygenase
Systematic name: taurochenodeoxycholate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Requires cytochrome *b*₅ for maximal activity. Acts on taurochenodeoxycholate, taurodeoxycholate and less readily on lithocholate and chenodeoxycholate. In adult pig (*Sus scrofa*), hyocholic acid replaces cholic acid as a primary bile acid [2566].
References: [127, 126, 2257, 2565, 2566, 3607]

[EC 1.14.14.57 created 2005 as EC 1.14.13.97, transferred 2018 to EC 1.14.14.57]

EC 1.14.14.58

- Accepted name:** trimethyltridecatetraene synthase
Reaction: (6*E*,10*E*)-geranylinalool + [reduced NADPH—hemoprotein reductase] + O₂ = (3*E*,7*E*)-4,8,12-trimethyltrideca-1,3,7,11-tetraene + [oxidized NADPH—hemoprotein reductase] + but-3-en-2-one + 2 H₂O
Other name(s): CYP82G1; CYP92C5; CYP92C6; DMNT/TMTT homoterpene synthase
Systematic name: (6*E*,10*E*)-geranylinalool,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plants *Arabidopsis thaliana* (thale cress) and *Zea mays* (maize). It forms this C₁₆ homoterpene in response to herbivore attack. *In vitro* some variants of the enzyme also convert (3*S*,6*E*)-nerolidol to (3*E*)-4,8-dimethylnona-1,3,7-triene (see EC 1.14.14.59, dimethylnonatriene synthase).
References: [2391, 3515]

[EC 1.14.14.58 created 2018]

EC 1.14.14.59

- Accepted name:** dimethylnonatriene synthase
Reaction: (3*S*,6*E*)-nerolidol + [reduced NADPH—hemoprotein reductase] + O₂ = (3*E*)-4,8-dimethylnona-1,3,7-triene + [oxidized NADPH—hemoprotein reductase] + but-3-en-2-one + 2 H₂O
Other name(s): CYP82G1; CYP92C5; DMNT/TMTT homoterpene synthase
Systematic name: (3*S*,6*E*)-nerolidol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plants *Arabidopsis thaliana* (thale cress) and *Zea mays* (maize). It forms this C₁₁ homoterpene in response to herbivore attack. *In vitro* the enzyme also converts (6*E*,10*E*)-geranylinalool to (3*E*,7*E*)-4,8,12-trimethyltrideca-1,3,7,11-tetraene (see EC 1.14.14.58, trimethyltridecatetraene synthase).
References: [2391, 3515]

[EC 1.14.14.59 created 2018]

EC 1.14.14.60

- Accepted name:** ferruginol monooxygenase
Reaction: ferruginol + [reduced NADPH—hemoprotein reductase] + O₂ = 11-hydroxyferruginol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP76AH24; CYP76AH3
Systematic name: ferruginol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (11-hydroxyferruginol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plants *Salvia pomifera* (apple sage) and *Salvia miltiorrhiza* (danshen). 11-Hydroxyferruginol is a precursor of carnosic acid, a potent antioxidant.
References: [1790, 3707, 1444]

[EC 1.14.14.60 created 2018]

EC 1.14.14.61

- Accepted name:** carnosic acid synthase
Reaction: 11-hydroxyferruginol + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = carnosic acid + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O
Other name(s): CYP76AK6; CYP76AK7; CYP76AK8
Systematic name: 11-hydroxyferruginol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plants *Salvia pomifera* (apple sage), *S. miltiorrhiza* (red sage), *S. fruticosa* (Greek sage) and *Rosmarinus officinalis* (Rosemary).
References: [1790, 3707]

[EC 1.14.14.61 created 2018]

EC 1.14.14.62

Accepted name: salviol synthase
Reaction: ferruginol + [reduced NADPH—hemoprotein reductase] + O₂ = salviol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP71BE52
Systematic name: ferruginol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (salviol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Salvia pomifera* (apple sage).
References: [1790]

[EC 1.14.14.62 created 2018]

EC 1.14.14.63

Accepted name: β-amyrin 16β-monooxygenase
Reaction: β-amyrin + [reduced NADPH—hemoprotein reductase] + O₂ = maniladiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP716A141
Systematic name: β-amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (maniladiol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Platycodon grandiflorus* (balloon flower). The enzyme is also able to oxidize oleanolic acid to cochalic acid.
References: [4191]

[EC 1.14.14.63 created 2018]

EC 1.14.14.64

Accepted name: β-amyrin 6β-monooxygenase
Reaction: β-amyrin + [reduced NADPH—hemoprotein reductase] + O₂ = daturadiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP716E26
Systematic name: β-amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (daturadiol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Solanum lycopersicum* (tomato).
References: [4780]

[EC 1.14.14.64 created 2018]

EC 1.14.14.65

Accepted name: sugiol synthase
Reaction: ferruginol + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = sugiol + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O
Other name(s): CYP76AH3
Systematic name: ferruginol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (sugiol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Salvia miltiorrhiza* (danshen). The enzyme also oxidizes 11-hydroxyferruginol to 11-hydroxysugiol. It also oxidizes at C-12 of ferruginol (EC 1.14.14.60 ferruginol monooxygenase).
References: [1444]

[EC 1.14.14.65 created 2018]

EC 1.14.14.66

Accepted name: marmesin synthase
Reaction: demethylsuberosin + [reduced NADPH—hemoprotein reductase] + O₂ = (+)-marmesin + [oxidized NADPH—hemoprotein reductase] + H₂O
Systematic name: demethylsuberosin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A *P*-450 monooxygenase involved in psoralen biosynthesis, see EC 1.14.13.102, psoralen synthase.

References: [1493]

[EC 1.14.14.66 created 2018]

EC 1.14.14.67

Accepted name: 11-hydroxysugiol 20-monooxygenase
Reaction: 11-hydroxysugiol + [reduced NADPH—hemoprotein reductase] + O₂ = 11,20-dihydroxysugiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP76AK1
Systematic name: 11-hydroxysugiol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (11,20-dihydroxysugiol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Salvia miltiorrhiza* (danshen). The enzyme also oxidizes 11-hydroxyferruginol to 11,20-dihydroxyferruginol.
References: [1444]

[EC 1.14.14.67 created 2018]

EC 1.14.14.68

Accepted name: *syn*-pimaradiene 3-monooxygenase
Reaction: 9β-pimara-7,15-diene + [reduced NADPH—hemoprotein reductase] + O₂ = 9β-pimara-7,15-diene-3β-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP701A8
Systematic name: 9β-pimara-7,15-diene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (9β-pimara-7,15-diene-3β-ol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from rice, *Oryza sativa*.
References: [2137]

[EC 1.14.14.68 created 2018]

EC 1.14.14.69

Accepted name: *ent*-cassadiene hydroxylase
Reaction: *ent*-cassa-12,15-diene + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = *ent*-3β-hydroxycassa-12,15-dien-2-one + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) *ent*-cassa-12,15-diene + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-cassa-12,15-dien-2β-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) *ent*-cassa-12,15-dien-2β-ol + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-cassa-12,15-dien-2-one + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1b') *ent*-cassa-12,15-dien-2β-ol + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-cassa-12,15-diene-2β,3β-diol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1c) *ent*-cassa-12,15-dien-2-one + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-3β-hydroxycassa-12,15-dien-2-one + [oxidized NADPH—hemoprotein reductase] + H₂O
(1c') *ent*-cassa-12,15-diene-2β,3β-diol + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-3β-hydroxycassa-12,15-dien-2-one + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): CYP71Z7
Systematic name: *ent*-cassa-12,15-diene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*ent*-3β-hydroxycassa-12,15-dien-2-one-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Oryza sativa* (rice) that is involved in phytocassanes biosynthesis. Depending on the order of activities, the enzyme may form either *ent*-cassa-12,15-dien-2-one or *ent*-cassa-12,15-diene-2β,3β-diol as an intermediate.
References: [2137]

[EC 1.14.14.69 created 2018]

EC 1.14.14.70

- Accepted name:** *ent*-sandaracopimaradiene 3-hydroxylase
Reaction: *ent*-sandaracopimaradiene + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-sandaracopimaradien-3β-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP701A; OsKOL4
Systematic name: *ent*-sandaracopimaradiene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*ent*-sandaracopimaradien-3β-ol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from *Oryza sativa* (rice). Participates in the pathway for the biosynthesis of oryzalexins, a group of related phytoalexins produced by rice. Can also use 9β-pimara-7,15-diene as substrate (*cf.* EC 1.14.14.68, *syn*-pimaradiene 3-monooxygenase).
References: [4527, 4684]

[EC 1.14.14.70 created 2014 as EC 1.14.13.191, transferred 2018 to EC 1.14.14.70]

EC 1.14.14.71

- Accepted name:** cucurbitadienol 11-hydroxylase
Reaction: cucurbitadienol + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = 11-oxocucurbitadienol + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) cucurbitadienol + [reduced NADPH—hemoprotein reductase] + O₂ = 11-hydroxycucurbitadienol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 11-hydroxycucurbitadienol + [reduced NADPH—hemoprotein reductase] + O₂ = 11-oxocucurbitadienol + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): CYP87D18
Systematic name: cucurbitadienol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (11-oxocucurbitadienol-forming)
Comments: Isolated from the plant *Siraitia grosvenorii* (monk fruit).
References: [4875]

[EC 1.14.14.71 created 2018]

EC 1.14.14.72

- Accepted name:** drimenol monooxygenase
Reaction: drimenol + [reduced NADPH—hemoprotein reductase] + O₂ = drimendiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): PhDOX1
Systematic name: drimenol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (drimendiol-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Persicaria hydropiper* (water pepper).
References: [1627]

[EC 1.14.14.72 created 2018]

EC 1.14.14.73

- Accepted name:** albendazole monooxygenase (sulfoxide-forming)
Reaction: (1) albendazole + [reduced NADPH—hemoprotein reductase] + O₂ = albendazole *S*-oxide + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) fenbendazole + [reduced NADPH—hemoprotein reductase] + O₂ = fenbendazole *S*-oxide + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): albendazole sulfoxidase (ambiguous); albendazole hydroxylase (ambiguous); CYP3A4 (gene name); CYP2J2 (gene name); CYP1A2 (gene name)
Systematic name: albendazole,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (sulfoxide-forming)

Comments: This is one of the activities carried out by some microsomal cytochrome *P*-450 monooxygenases. A similar conversion is also carried out by a different microsomal enzyme (EC 1.14.13.32, albendazole monooxygenase (flavin-containing)), but it is estimated that cytochrome *P*-450s are responsible for 70% of the activity.

References: [2898, 3463, 150, 2379, 4685]

[EC 1.14.14.73 created 2018]

EC 1.14.14.74

Accepted name: albendazole monooxygenase (hydroxylating)

Reaction: albendazole + [reduced NADPH—hemoprotein reductase] + O₂ = hydroxyalbendazole + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP2J2 (gene name)

Systematic name: albendazole,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating)

Comments: CYP2J2 is a microsomal cytochrome *P*-450 monooxygenase that catalyses the hydroxylation of the terminal carbon of the propylsulfanyl chain in albendazole, a broad-spectrum anthelmintic used against gastrointestinal nematodes and the larval stages of cestodes. *cf.* EC 1.14.14.73, albendazole monooxygenase (sulfoxide-forming).

References: [4685]

[EC 1.14.14.74 created 2018]

EC 1.14.14.75

Accepted name: fenbendazole monooxygenase (4'-hydroxylating)

Reaction: fenbendazole + [reduced NADPH—hemoprotein reductase] + O₂ = 4'-hydroxyfenbendazole + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP2C19 (gene name)

Systematic name: fenbendazole,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (4'-hydroxylating)

Comments: CYP2C19 is microsomal cytochrome *P*-450 monooxygenase that catalyses the hydroxylation of the benzene ring of fenbendazole, a broad-spectrum anthelmintic used against gastrointestinal nematodes and the larval stages of cestodes. This activity is also carried out by CYP2J2. *cf.* EC 1.14.14.74, albendazole monooxygenase (hydroxylating). CYP2C19 does not act on albendazole.

References: [4685]

[EC 1.14.14.75 created 2018]

EC 1.14.14.76

Accepted name: *ent*-isokaurene C2/C3-hydroxylase

Reaction: *ent*-isokaurene + 2 O₂ + 2 [reduced NADPH—hemoprotein reductase] = *ent*-isokaurene-2β,3β-diol + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)

(1a) *ent*-isokaurene + O₂ + [reduced NADPH—hemoprotein reductase] = *ent*-isokauren-2β-ol + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) *ent*-isokauren-2β-ol + O₂ + [reduced NADPH—hemoprotein reductase] = *ent*-isokaurene-2β,3β-diol + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP71Z6; *ent*-isokaurene C2-hydroxylase

Systematic name: *ent*-isokaurene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*ent*-isokaurene-2β,3β-diol-forming)

Comments: This cytochrome *P*-450 (heme thiolate) enzyme has been characterized from the plant *Oryza sativa* (rice). It may be involved in production of oryzadione.

References: [4683, 2137]

[EC 1.14.14.76 created 2012 as EC 1.14.13.143, transferred 2018 to EC 1.14.14.76]

EC 1.14.14.77

- Accepted name:** phenylacetonitrile α -monooxygenase
Reaction: phenylacetonitrile + [reduced NADPH—hemoprotein reductase] + O₂ = (*R*)-mandelonitrile + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP3201B1 (gene name)
Systematic name: phenylacetonitrile,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase [(*R*)-mandelonitrile-forming]
Comments: The enzyme has been characterized from the cyanogenic millipede *Chamberlinius huaiienensis*. Unlike plant enzymes that can catalyse this reaction (EC 1.14.14.44, phenylacetaldehyde oxime monooxygenase), this enzyme cannot act on phenylacetaldehyde oximes. It can accept (4-hydroxyphenyl)acetonitrile, (2-methylphenyl)acetonitrile, and (3-methylphenyl)acetonitrile as substrates at a lower rate.
References: [4733]

[EC 1.14.14.77 created 2018]

EC 1.14.14.78

- Accepted name:** phylloquinone ω -hydroxylase
Reaction: phylloquinone + [reduced NADPH—hemoprotein reductase] + O₂ = ω -hydroxyphylloquinone + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): vitamin K₁ ω -hydroxylase; CYP4F2; CYP4F11
Systematic name: phylloquinone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (ω -hydroxyphylloquinone-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Isolated from human tissue. The enzyme will also act on menaquinone-4. Prolonged action of CYP4F2, but not CYP4F11, on the ω hydroxyl group oxidizes it to the corresponding carboxylic acid. CYP4F2 also oxidizes leukotriene B₄; see EC 1.14.13.30, leukotriene-B₄ 20-monooxygenase [1916].
References: [1916, 4205, 1020]

[EC 1.14.14.78 created 2014 as EC 1.14.13.194, transferred 2018 to EC 1.14.14.78]

EC 1.14.14.79

- Accepted name:** docosahexaenoic acid ω -hydroxylase
Reaction: docosahexaenoate + [reduced NADPH—hemoprotein reductase] + O₂ = 22-hydroxydocosahexaenoate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP4F3B; CYP4V2; docosahexaenoate,NADPH:O₂ oxidoreductase (22-hydroxydocosahexaenoate forming)
Systematic name: docosahexaenoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (22-hydroxydocosahexaenoate-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from human eye tissue. Defects in the enzyme are associated with Bietti crystalline corneoretinal dystrophy. The enzyme also produces some 21-hydroxydocosahexaenoate. Acts in a similar way on icosapentaenoic acid.
References: [2991]

[EC 1.14.14.79 created 2014 as EC 1.14.13.199, transferred 2018 to EC 1.14.14.79]

EC 1.14.14.80

- Accepted name:** long-chain fatty acid ω -monooxygenase
Reaction: a long-chain fatty acid + [reduced NADPH—hemoprotein reductase] + O₂ = an ω -hydroxy-long-chain fatty acid + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP704B1 (gene name); CYP52M1 (gene name); CYP4A (gene name); CYP86A (gene name)
Systematic name: long-chain fatty acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (ω -hydroxylating)

Comments: A cytochrome *P*-450 (heme thiolate) enzyme. The plant enzyme CYP704B1, which is involved in the synthesis of sporopollenin, a complex polymer found at the outer layer of spores and pollen, acts on palmitate (18:0), stearate (18:0) and oleate (18:1). The plant enzyme CYP86A1 also acts on laurate (12:0). The enzyme from the yeast *Starmerella bombicola* (CYP52M1) acts on C₁₆ to C₂₀ saturated and unsaturated fatty acids and can also hydroxylate the (ω-1) position. The mammalian enzyme CYP4A acts on laurate (12:0), myristate (14:0), palmitate (16:0), oleate (18:1), and arachidonate (20:4).

References: [292, 1679, 935, 1752]

[EC 1.14.14.80 created 2015 as EC 1.14.13.205, transferred 2018 to EC 1.14.14.80]

EC 1.14.14.81

Accepted name: flavanoid 3',5'-hydroxylase
Reaction: a flavanone + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = a 3',5'-dihydroxyflavanone + 2 [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) a flavanone + [reduced NADPH—hemoprotein reductase] + O₂ = a 3'-hydroxyflavanone + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) a 3'-hydroxyflavanone + [reduced NADPH—hemoprotein reductase] + O₂ = a 3',5'-dihydroxyflavanone + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): flavanoid 3',5'-hydroxylase
Systematic name: flavanone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3',5'-dihydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. The 3',5'-dihydroxyflavanone is formed via the 3'-hydroxyflavanone. In *Petunia hybrida* the enzyme acts on naringenin, eriodictyol, dihydroquercetin (taxifolin) and dihydrokaempferol (aromadendrin). The enzyme catalyses the hydroxylation of 5,7,4'-trihydroxyflavanone (naringenin) at either the 3' position to form eriodictyol or at both the 3' and 5' positions to form 5,7,3',4',5'-pentahydroxyflavanone (dihydrotricetin). The enzyme also catalyses the hydroxylation of 3,5,7,3',4'-pentahydroxyflavanone (taxifolin) at the 5' position, forming ampelopsin.

References: [2772, 3865, 854]

[EC 1.14.14.81 created 2004 as EC 1.14.13.88, transferred 2018 to EC 1.14.14.81]

EC 1.14.14.82

Accepted name: flavonoid 3'-monooxygenase
Reaction: a flavonoid + [reduced NADPH—hemoprotein reductase] + O₂ = a 3'-hydroxyflavonoid + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP75B1 (gene name); flavonoid 3'-hydroxylase; flavonoid 3-hydroxylase (incorrect); NADPH:flavonoid-3'-hydroxylase (incorrect); flavonoid 3-monooxygenase (incorrect)

Systematic name: flavonoid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. Acts on a number of flavonoids, including the flavanone naringenin and the flavone apigenin. Does not act on 4-coumarate or 4-coumaroyl-CoA.

References: [1140, 470, 3741]

[EC 1.14.14.82 created 1983 as EC 1.14.13.21, transferred 2018 to EC 1.14.14.82]

EC 1.14.14.83

Accepted name: geraniol 8-hydroxylase
Reaction: geraniol + [reduced NADPH—hemoprotein reductase] + O₂ = (6*E*)-8-hydroxygeraniol + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP76B6 (gene name); G10H (gene name)
Systematic name: geraniol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8-hydroxylating)

Comments: A cytochrome *P*-450 (heme thiolate) protein found in plants. Also hydroxylates nerol and citronellol, *cf.* EC 1.14.14.84, linalool 8-monooxygenase. The recommended numbering of geraniol gives 8-hydroxygeraniol as the product rather than 10-hydroxygeraniol as used by references 1-3. See prenol nomenclature Pr-1. The cloned enzyme also catalysed, but less efficiently, the 3'-hydroxylation of naringenin (*cf.* EC 1.14.14.82, flavonoid 3'-monooxygenase) [4129].

References: [715, 4516, 4129]

[EC 1.14.14.83 created 2012 as EC 1.14.13.152, transferred 2018 to EC 1.14.14.83]

EC 1.14.14.84

Accepted name: linalool 8-monooxygenase

Reaction: linalool + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = (6*E*)-8-oxolinalool + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) linalool + [reduced NADPH—hemoprotein reductase] + O₂ = (6*E*)-8-hydroxylinalool + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) (6*E*)-8-hydroxylinalool + [reduced NADPH—hemoprotein reductase] + O₂ = (6*E*)-8-oxolinalool + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): *P*-450lin; CYP111

Systematic name: linalool,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. The secondary electron donor is a specific [2Fe-2S] ferredoxin from the same bacterial strain.

References: [4372, 3570]

[EC 1.14.14.84 created 1989 as EC 1.14.99.28, transferred 2012 to EC 1.14.13.151, transferred 2018 to EC 1.14.14.84]

EC 1.14.14.85

Accepted name: 7-deoxyloganate 7-hydroxylase

Reaction: 7-deoxyloganate + [reduced NADPH—hemoprotein reductase] + O₂ = loganate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP72A224 (gene name); 7-deoxyloganin 7-hydroxylase (incorrect); 7-deoxyloganin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (7α-hydroxylating) (incorrect)

Systematic name: 7-deoxyloganate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (7α-hydroxylating)

Comments: The enzyme, characterized from the plant *Catharanthus roseus*, is a cytochrome *P*-450 (heme-thiolate) enzyme. It catalyses a reaction in the pathway leading to biosynthesis of monoterpene indole alkaloids.

References: [2010, 2797]

[EC 1.14.14.85 created 2002 as EC 1.14.13.74, transferred 2018 to EC 1.14.14.85, modified 2018]

EC 1.14.14.86

Accepted name: *ent*-kaurene monooxygenase

Reaction: *ent*-kaur-16-ene + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = *ent*-kaur-16-en-19-oate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) *ent*-kaur-16-ene + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-kaur-16-en-19-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) *ent*-kaur-16-en-19-ol + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-kaur-16-en-19-al + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) *ent*-kaur-16-en-19-al + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-kaur-16-en-19-oate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): *ent*-kaurene oxidase (misleading)

Systematic name: *ent*-kaur-16-ene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating)

Comments: A cytochrome *P*-450 (heme thiolate) protein found in plants. Catalyses three successive oxidations of the 4-methyl group of *ent*-kaurene giving kaurenoic acid.

References: [146, 128, 1620]

[EC 1.14.14.86 created 2002 as EC 1.14.13.78, transferred 2018 to EC 1.14.14.86]

EC 1.14.14.87

Accepted name: 2-hydroxyisoflavanone synthase

Reaction: (1) liquiritigenin + O₂ + [reduced NADPH—hemoprotein reductase] = 2,4',7-trihydroxyisoflavanone + H₂O + [oxidized NADPH—hemoprotein reductase]
(2) (2*S*)-naringenin + O₂ + [reduced NADPH—hemoprotein reductase] = 2,4',5,7-tetrahydroxyisoflavanone + H₂O + [oxidized NADPH—hemoprotein reductase]

Other name(s): CYP93C; IFS; isoflavonoid synthase

Systematic name: liquiritigenin, [reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating, aryl migration)

Comments: A cytochrome *P*-450 (heme thiolate) protein found in plants. The reaction involves the migration of the 2-phenyl group of the flavanone to the 3-position of the isoflavanone. The 2-hydroxyl group is derived from the oxygen molecule. EC 4.2.1.105, 2-hydroxyisoflavanone dehydratase, acts on the products with loss of water and formation of genistein and daidzein, respectively.

References: [2181, 1549, 4009, 3685, 3684]

[EC 1.14.14.87 created 2011 as EC 1.14.13.136, modified 2013, transferred 2018 to EC 1.14.14.87]

EC 1.14.14.88

Accepted name: isoflavone 3'-hydroxylase

Reaction: formononetin + [reduced NADPH—hemoprotein reductase] + O₂ = calycosin + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): isoflavone 3'-monooxygenase; CYP81E9

Systematic name: formononetin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3'-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. Also acts on biochanin A and other isoflavones with a 4'-methoxy group. Involved in the biosynthesis of the pterocarpin phytoalexins medicarpin and maackiain.

References: [1661]

[EC 1.14.14.88 created 1992 as EC 1.14.13.52, transferred 2018 to EC 1.14.14.88]

EC 1.14.14.89

Accepted name: 4'-methoxyisoflavone 2'-hydroxylase

Reaction: formononetin + [reduced NADPH—hemoprotein reductase] + O₂ = 2'-hydroxyformononetin + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP81E1 (gene name); CYP81E3 (gene name); CYP81E7 (gene name); isoflavone 2'-monooxygenase (ambiguous); isoflavone 2'-hydroxylase (ambiguous)

Systematic name: formononetin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2'-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. Acts on isoflavones with a 4'-methoxy group, such as formononetin and biochanin A. Involved in the biosynthesis of the pterocarpin phytoalexins medicarpin and maackiain. EC 1.14.14.90, isoflavone 2'-hydroxylase, is less specific and acts on other isoflavones as well as 4'-methoxyisoflavones.

References: [1661, 49, 2512]

[EC 1.14.14.89 created 1992 as EC 1.14.13.53, modified 2005, transferred 2018 to EC 1.14.14.89]

EC 1.14.14.90

Accepted name: isoflavone 2'-hydroxylase

Reaction: an isoflavone + [reduced NADPH—hemoprotein reductase] + O₂ = a 2'-hydroxyisoflavone + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): isoflavone 2'-monooxygenase; CYP81E1; CYP Ge-3
Systematic name: isoflavone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Acts on daidzein, formononetin and genistein. EC 1.14.14.89, 4'-methoxyisoflavone 2'-hydroxylase, has the same reaction but is more specific as it requires a 4'-methoxyisoflavone.
References: [49]

[EC 1.14.14.90 created 2005 as EC 1.14.13.89, transferred 2018 to EC 1.14.14.90]

EC 1.14.14.91

Accepted name: *trans*-cinnamate 4-monooxygenase
Reaction: *trans*-cinnamate + [reduced NADPH—hemoprotein reductase] + O₂ = 4-hydroxycinnamate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): cinnamic acid 4-hydroxylase; CA4H; cytochrome P450 cinnamate 4-hydroxylase; cinnamate 4-hydroxylase; cinnamate 4-monooxygenase; cinnamate hydroxylase; cinnamic 4-hydroxylase; cinnamic acid 4-monooxygenase; cinnamic acid *p*-hydroxylase; *t*-cinnamic acid hydroxylase; *trans*-cinnamate 4-hydroxylase; *trans*-cinnamic acid 4-hydroxylase; CYP73A1 (gene name)
Systematic name: *trans*-cinnamate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (4-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. The enzyme is involved in flavonoid biosynthesis.
References: [3359, 3608, 3320]

[EC 1.14.14.91 created 1976 as EC 1.14.13.11, transferred 2018 to EC 1.14.14.91]

EC 1.14.14.92

Accepted name: benzoate 4-monooxygenase
Reaction: benzoate + [reduced NADPH—hemoprotein reductase] + O₂ = 4-hydroxybenzoate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): benzoic acid 4-hydroxylase; benzoate 4-hydroxylase; benzoic 4-hydroxylase; benzoate-*p*-hydroxylase; *p*-hydroxybenzoate hydroxylase; CYP53A1 (gene name)
Systematic name: benzoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (4-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in *Aspergillus* fungi.
References: [3473, 1077]

[EC 1.14.14.92 created 1976 as EC 1.14.13.12, transferred 2018 to EC 1.14.14.92]

EC 1.14.14.93

Accepted name: 3,9-dihydroxypterocarpan 6 α -monooxygenase
Reaction: (6 α R,11 α R)-3,9-dihydroxypterocarpan + [reduced NADPH—hemoprotein reductase] + O₂ = (6 α S,11 α S)-3,6 α ,9-trihydroxypterocarpan + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): 3,9-dihydroxypterocarpan 6 α -hydroxylase; 3,9-dihydroxypterocarpan 6 α -monooxygenase (erroneous); CYP93A1 (gene name)
Systematic name: (6 α R,11 α R)-3,9-dihydroxypterocarpan,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in soybean. The product of the reaction is the biosynthetic precursor of the glyceollin phytoalexins.
References: [1476, 3744]

[EC 1.14.14.93 created 1989 as EC 1.14.13.28, transferred 2018 to EC 1.14.14.93]

EC 1.14.14.94

- Accepted name:** leukotriene-B₄ 20-monoxygenase
Reaction: (6Z,8E,10E,14Z)-(5S,12R)-5,12-dihydroxyicoso-6,8,10,14-tetraenoate + [reduced NADPH—hemoprotein reductase] + O₂ = (6Z,8E,10E,14Z)-(5S,12R)-5,12,20-trihydroxyicoso-6,8,10,14-tetraenoate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): leukotriene-B₄ 20-hydroxylase; leucotriene-B₄ ω-hydroxylase; LTB₄ 20-hydroxylase; LTB₄ ω-hydroxylase; CYP4F2 (gene name); CYP4F3 (gene name)
Systematic name: (6Z,8E,10E,14Z)-(5S,12R)-5,12-dihydroxyicoso-6,8,10,14-tetraenoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (20-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in mammals.
References: [3563, 3815, 3949]

[EC 1.14.14.94 created 1989 as EC 1.14.13.30, transferred 2018 to EC 1.14.14.94]

EC 1.14.14.95

- Accepted name:** germacrene A hydroxylase
Reaction: (+)-germacrene A + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = germacra-1(10),4,11(13)-trien-12-oate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) (+)-germacrene A + O₂ + [reduced NADPH—hemoprotein reductase] = germacra-1(10),4,11(13)-trien-12-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) germacra-1(10),4,11(13)-trien-12-ol + O₂ + [reduced NADPH—hemoprotein reductase] = germacra-1(10),4,11(13)-trien-12-al + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) germacra-1(10),4,11(13)-trien-12-al + O₂ + [reduced NADPH—hemoprotein reductase] = germacra-1(10),4,11(13)-trien-12-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): GAO (gene name)
Systematic name: (+)-germacrene-A,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. This plant enzyme catalyses three steps in a pathway that leads to the biosynthesis of many sesquiterpenoid lactones.
References: [3058, 2522]

[EC 1.14.14.95 created 2011 as EC 1.14.13.123, transferred 2018 to EC 1.14.14.95]

EC 1.14.14.96

- Accepted name:** 5-*O*-(4-coumaroyl)-D-quininate 3'-monoxygenase
Reaction: *trans*-5-*O*-(4-coumaroyl)-D-quininate + [reduced NADPH—hemoprotein reductase] + O₂ = *trans*-5-*O*-caffeyl-D-quininate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): 5-*O*-(4-coumaroyl)-D-quininate/shikimate 3'-hydroxylase; coumaroylquininate(coumaroylshikimate) 3'-monoxygenase; CYP98A3 (gene name)
Systematic name: *trans*-5-*O*-(4-coumaroyl)-D-quininate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein, found in plants. It also acts on *trans*-5-*O*-(4-coumaroyl)shikimate.
References: [2279, 3740, 1166, 2709]

[EC 1.14.14.96 created 1990 as EC 1.14.13.36, transferred 2018 to EC 1.14.14.96]

EC 1.14.14.97

- Accepted name:** methyltetrahydroprotoberberine 14-monoxygenase
Reaction: (*S*)-*N*-methylcanadine + [reduced NADPH—hemoprotein reductase] + O₂ = allocryptopine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): methyltetrahydroprotoberberine 14-hydroxylase; (*S*)-*cis*-*N*-methyltetrahydroberberine 14-monoxygenase; (*S*)-*cis*-*N*-methyltetrahydroprotoberberine-14-hydroxylase; CYP82N4 (gene name)

Systematic name: (*S*)-*N*-methylcanadine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (14-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants.

References: [3596, 259]

[EC 1.14.14.97 created 1990 as EC 1.14.13.37, transferred 2018 to EC 1.14.14.97]

EC 1.14.14.98

Accepted name: protopine 6-monooxygenase

Reaction: protopine + [reduced NADPH—hemoprotein reductase] + O₂ = 6-hydroxyprotopine + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): protopine 6-hydroxylase; CYP82N2 (gene name)

Systematic name: protopine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in benzophenanthridine alkaloid synthesis in higher plants.

References: [4193, 4181]

[EC 1.14.14.98 created 1999 as EC 1.14.13.55, transferred 2018 to EC 1.14.14.98]

EC 1.14.14.99

Accepted name: (*S*)-limonene 3-monooxygenase

Reaction: (*S*)-limonene + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-*trans*-isopiperitenol + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): (–)-limonene 3-hydroxylase; (–)-limonene 3-monooxygenase; CYP71D15 (gene name)

Systematic name: (*S*)-limonene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein from peppermint (*Mentha piperita*).

References: [1996, 2569, 4687]

[EC 1.14.14.99 created 1992 as EC 1.14.13.47, modified 2003, transferred 2018 1.14.14.99]

EC 1.14.14.100

Accepted name: dihydrosanguinarine 10-monooxygenase

Reaction: dihydrosanguinarine + [reduced NADPH—hemoprotein reductase] + O₂ = 10-hydroxydihydrosanguinarine + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): dihydrosanguinarine 10-hydroxylase

Systematic name: dihydrosanguinarine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (10-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in benzophenanthridine alkaloid synthesis in higher plants.

References: [844]

[EC 1.14.14.100 created 1999 as EC 1.14.13.56, transferred 2018 to EC 1.14.14.100]

EC 1.14.14.101

Accepted name: dihydrochelirubine 12-monooxygenase

Reaction: dihydrochelirubine + [reduced NADPH—hemoprotein reductase] + O₂ = 12-hydroxydihydrochelirubine + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): dihydrochelirubine 12-hydroxylase

Systematic name: dihydrochelirubine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Thalictrum bulgaricum*.

References: [1986]

[EC 1.14.14.101 created 1999 as EC 1.14.13.57, transferred 2018 to EC 1.14.14.101]

EC 1.14.14.102

- Accepted name:** *N*-methylcoclaurine 3'-monooxygenase
Reaction: (*S*)-*N*-methylcoclaurine + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-3'-hydroxy-*N*-methylcoclaurine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): *N*-methylcoclaurine 3'-hydroxylase; CYP80B1 (gene name)
Systematic name: (*S*)-*N*-methylcoclaurine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in benzyloquinoline alkaloid synthesis in higher plants.
References: [3265]

[EC 1.14.14.102 created 2001 as 1.14.13.71, transferred 2018 to EC 1.14.14.102]

EC 1.14.14.103

- Accepted name:** tabersonine 16-hydroxylase
Reaction: tabersonine + [reduced NADPH—hemoprotein reductase] + O₂ = 16-hydroxytabersonine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): tabersonine-11-hydroxylase; T11H; CYP71D12 (gene name)
Systematic name: tabersonine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (16-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant Madagascar periwinkle (*Catharanthus roseus*).
References: [3993, 322]

[EC 1.14.14.103 created 2002 as EC 1.14.13.73, transferred 2018 to EC 1.14.14.103]

EC 1.14.14.104

- Accepted name:** vinorine hydroxylase
Reaction: vinorine + [reduced NADPH—hemoprotein reductase] + O₂ = vomilenine + [oxidized NADPH—hemoprotein reductase] + H₂O
Systematic name: vinorine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (21 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Rauvolfia serpentina*. Forms a stage in the biosynthesis of the indole alkaloid ajmaline.
References: [1080]

[EC 1.14.14.104 created 2002 as EC 1.14.13.75, transferred 2018 to EC 1.14.14.104]

EC 1.14.14.105

- Accepted name:** taxane 10 β -hydroxylase
Reaction: taxa-4(20),11-dien-5 α -yl acetate + [reduced NADPH—hemoprotein reductase] + O₂ = 10 β -hydroxytaxa-4(20),11-dien-5 α -yl acetate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP725A1 (gene name); 5- α -taxadienol-10- β -hydroxylase
Systematic name: taxa-4(20),11-dien-5 α -yl acetate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (10 β -hydroxylating)
Comments: This microsomal cytochrome-*P*-450 (heme-thiolate) enzyme from the plant *Taxus cuspidata* is involved in the biosynthesis of the diterpenoid antineoplastic drug taxol (paclitaxel).
References: [4597, 1900, 3742]

[EC 1.14.14.105 created 2002 as EC 1.14.13.76, transferred 2018 to EC 1.14.14.105]

EC 1.14.14.106

- Accepted name:** taxane 13 α -hydroxylase
Reaction: taxa-4(20),11-dien-5 α -ol + [reduced NADPH—hemoprotein reductase] + O₂ = taxa-4(20),11-dien-5 α ,13 α -diol + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP725A2 (gene name)
Systematic name: taxa-4(20),11-dien-5 α -ol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (13 α -hydroxylating)
Comments: This cytochrome-*P*-450(heme-thiolate) enzyme from the plant *Taxus cuspidata* is involved in the biosynthesis of the diterpenoid antineoplastic drug taxol (paclitaxel).
References: [4597, 1900]

[EC 1.14.14.106 created 2002 as EC 1.14.13.77, transferred 2018 to EC 1.14.14.106]

EC 1.14.14.107

Accepted name: *ent*-kaurenoic acid monooxygenase
Reaction: *ent*-kaur-16-en-19-oate + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = gibberellin A₁₂ + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) *ent*-kaur-16-en-19-oate + [reduced NADPH—hemoprotein reductase] + O₂ = *ent*-7 α -hydroxykaur-16-en-19-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) *ent*-7 α -hydroxykaur-16-en-19-oate + [reduced NADPH—hemoprotein reductase] + O₂ = gibberellin A₁₂ aldehyde + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) gibberellin A₁₂ aldehyde + [reduced NADPH—hemoprotein reductase] + O₂ = gibberellin A₁₂ + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): KAO1 (gene name); CYP88A3 (gene name); *ent*-kaurenoic acid oxidase
Systematic name: *ent*-kaur-16-en-19-oate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from plants. Catalyses three successive oxidations of *ent*-kaurenoic acid. The second step includes a ring-B contraction giving the gibbane skeleton. In pumpkin (*Cucurbita maxima*) *ent*-6 α ,7 α -dihydroxykaur-16-en-19-oate is also formed.
References: [1619]

[EC 1.14.14.107 created 2002 as EC 1.14.13.79, transferred 2018 to EC 1.14.14.107]

EC 1.14.14.108

Accepted name: 2,5-diketocamphane 1,2-monooxygenase
Reaction: (+)-bornane-2,5-dione + FMNH₂ + O₂ = (+)-5-oxo-1,2-campholide + FMN + H₂O
Other name(s): 2,5-diketocamphane lactonizing enzyme; ketolactonase I (ambiguous); 2,5-diketocamphane 1,2-monooxygenase oxygenating component; 2,5-DKCMO; *camP* (gene name); camphor 1,2-monooxygenase; camphor ketolactonase I
Systematic name: (+)-bornane-2,5-dione,FMNH₂:oxygen oxidoreductase (1,2-lactonizing)
Comments: A Baeyer-Villiger monooxygenase isolated from camphor-grown strains of *Pseudomonas putida* and encoded on the cam plasmid. Involved in the degradation of (+)-camphor. Requires a dedicated NADH-FMN reductase [*cf.* EC 1.5.1.42, FMN reductase (NADH)] [723, 4831, 4227]. Can accept several bicyclic ketones including (+)- and (–)-camphor [1970] and adamantanone [3795]. The product spontaneously converts to [(1*R*)-2,2,3-trimethyl-5-oxocyclopent-3-enyl]acetate.
References: [723, 4831, 4227, 3795, 1946, 1970, 1858]

[EC 1.14.14.108 created 1972 as EC 1.14.15.2, transferred 2012 to EC 1.14.13.162, transferred 2018 to EC 1.14.14.108]

EC 1.14.14.109

Accepted name: 3-hydroxyindolin-2-one monooxygenase
Reaction: 3-hydroxyindolin-2-one + [reduced NADPH—hemoprotein reductase] + O₂ = 2-hydroxy-2*H*-1,4-benzoxazin-3(4*H*)-one [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): BX4 (gene name); CYP71C1 (gene name)
Systematic name: 3-hydroxyindolin-2-one,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2-hydroxy-2*H*-1,4-benzoxazin-3(4*H*)-one-forming)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme is involved in the biosynthesis of protective and allelopathic benzoxazinoids in some plants, most commonly from the family of Poaceae (grasses).

References: [1341, 1175, 3987]

[EC 1.14.14.109 created 2012 as EC 1.14.13.139, transferred 2018 to EC 1.14.14.109]

EC 1.14.14.110

Accepted name: 2-hydroxy-1,4-benzoxazin-3-one monooxygenase

Reaction: 2-hydroxy-2*H*-1,4-benzoxazin-3(4*H*)-one + [reduced NADPH—hemoprotein reductase] + O₂ = 2,4-dihydroxy-2*H*-1,4-benzoxazin-3(4*H*)-one + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): BX5 (gene name); CYP71C3 (gene name)

Systematic name: 2-hydroxy-2*H*-1,4-benzoxazin-3(4*H*)-one,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme is involved in the biosynthesis of protective and allelopathic benzoxazinoids in some plants, most commonly from the family of Poaceae (grasses).

References: [182, 1341]

[EC 1.14.14.110 created 2012 as EC 1.14.13.140, transferred 2018 to EC 1.14.14.110]

EC 1.14.14.111

Accepted name: 9β-pimara-7,15-diene oxidase

Reaction: 9β-pimara-7,15-diene + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = 9β-pimara-7,15-dien-19-oate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)

(1a) 9β-pimara-7,15-diene + O₂ + [reduced NADPH—hemoprotein reductase] = 9β-pimara-7,15-dien-19-ol + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) 9β-pimara-7,15-dien-19-ol + O₂ + [reduced NADPH—hemoprotein reductase] = 9β-pimara-7,15-dien-19-al + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

(1c) 9β-pimara-7,15-dien-19-al + O₂ + [reduced NADPH—hemoprotein reductase] = 9β-pimara-7,15-dien-19-oate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP99A3; 9β-pimara-7,15-diene monooxygenase

Systematic name: 9β-pimara-7,15-diene,[reduced NADPH—hemoprotein reductase]:oxygen 19-oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from rice (*Oryza sativa*) is involved in the biosynthesis of the phytoalexin momilactone. It also acts similarly on 9β-stemod-13(17)-ene.

References: [4526]

[EC 1.14.14.111 created 2012 as EC 1.14.13.144, transferred 2018 to EC 1.14.14.111]

EC 1.14.14.112

Accepted name: *ent*-cassa-12,15-diene 11-hydroxylase

Reaction: *ent*-cassa-12,15-diene + O₂ + [reduced NADPH—hemoprotein reductase] = *ent*-11β-hydroxycassa-12,15-diene + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): *ent*-cassadiene C11α-hydroxylase; CYP76M7

Systematic name: *ent*-cassa-12,15-diene,[reduced NADPH—hemoprotein reductase]:oxygen 11-oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from rice (*Oryza sativa*) is involved in the biosynthesis of the antifungal phytocassanes.

References: [4148]

[EC 1.14.14.112 created 2012 as EC 1.14.13.145, transferred 2018 to EC 1.14.14.112]

EC 1.14.14.113

- Accepted name:** α -humulene 10-hydroxylase
Reaction: α -humulene + O₂ + [reduced NADPH—hemoprotein reductase] = 10-hydroxy- α -humulene + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP71BA1
Systematic name: α -humulene,[reduced NADPH—hemoprotein reductase]:oxygen 10-oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The recommended numbering of humulene gives 10-hydroxy- α -humulene as the product rather than 8-hydroxy- α -humulene as used by the reference. See Section F: Natural Product Nomenclature.
References: [4833]

[EC 1.14.14.113 created 2012 as EC 1.14.13.150, transferred 2018 to EC 1.14.14.113]

EC 1.14.14.114

- Accepted name:** amorpha-4,11-diene 12-monooxygenase
Reaction: amorpha-4,11-diene + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = artemisinate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) amorpha-4,11-diene + O₂ + [reduced NADPH—hemoprotein reductase] = artemisinic alcohol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) artemisinic alcohol + O₂ + [reduced NADPH—hemoprotein reductase] = artemisinic aldehyde + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) artemisinic aldehyde + O₂ + [reduced NADPH—hemoprotein reductase] = artemisinate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP71AV1
Systematic name: amorpha-4,11-diene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Cloned from the plant *Artemisia annua* (sweet wormwood). Part of the biosynthetic pathway of artemisinin.
References: [4242]

[EC 1.14.14.114 created 2012 as EC 1.14.13.158, transferred 2018 to EC 1.14.14.114]

EC 1.14.14.115

- Accepted name:** 11-oxo- β -amyrin 30-oxidase
Reaction: 11-oxo- β -amyrin + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = glycyrrhetinate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) 11-oxo- β -amyrin + O₂ + [reduced NADPH—hemoprotein reductase] = 30-hydroxy-11-oxo- β -amyrin + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 30-hydroxy-11-oxo- β -amyrin + O₂ + [reduced NADPH—hemoprotein reductase] = glycyrrhetaldehyde + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) glycyrrhetaldehyde + O₂ + [reduced NADPH—hemoprotein reductase] = glycyrrhetinate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP72A; CYP72A154; 11-oxo- β -amyrin 30-monooxygenase
Systematic name: 11-oxo- β -amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (30-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from the plant *Glycyrrhiza uralensis* (licorice) is involved in the biosynthesis of the triterpenoid saponin glycyrrhizin. The enzyme from the plant *Medicago truncatula* can also hydroxylate β -amyrin.
References: [3790]

[EC 1.14.14.115 created 2013 as EC 1.14.13.173, transferred 2018 to EC 1.14.14.115]

EC 1.14.14.116

Accepted name: averantin hydroxylase
Reaction: (1) (1'*S*)-averantin + [reduced NADPH—hemoprotein reductase] + O₂ = (1'*S*,5'*S*)-5'-hydroxyaverantin + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) (1'*S*)-averantin + [reduced NADPH—hemoprotein reductase] + O₂ = (1'*S*,5'*R*)-5'-hydroxyaverantin + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): AVN hydroxylase; *avnA* (gene name); CYP60A1
Systematic name: (1'*S*)-averantin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (5'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the saprophytic mold *Aspergillus parasiticus*. Involved in aflatoxin biosynthesis. Does not react with (1'*R*)-averantin.
References: [4717, 4835]

[EC 1.14.14.116 created 2013 as EC 1.14.13.174, transferred 2018 to EC 1.14.14.116]

EC 1.14.14.117

Accepted name: aflatoxin B synthase
Reaction: (1) 8-*O*-methylsterigmatocystin + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = aflatoxin B₁ + 2 [oxidized NADPH—hemoprotein reductase] + H₂O + methanol + CO₂
(2) 8-*O*-methylidihydrosterigmatocystin + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = aflatoxin B₂ + 2 [oxidized NADPH—hemoprotein reductase] + H₂O + methanol + CO₂
Other name(s): *ordA* (gene name)
Systematic name: 8-*O*-methylsterigmatocystin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (aflatoxin-B-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Isolated from the mold *Aspergillus parasiticus*.
References: [327, 4836, 4363]

[EC 1.14.14.117 created 2013 as EC 1.14.13.175, transferred 2018 to EC 1.14.14.117]

EC 1.14.14.118

Accepted name: tryprostatin B 6-hydroxylase
Reaction: tryprostatin B + [reduced NADPH—hemoprotein reductase] + O₂ = 6-hydroxytryprostatin B + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): *ftmC* (gene name)
Systematic name: tryprostatin B,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6-hydroxytryprostatin B-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Involved in the biosynthetic pathways of several indole alkaloids such as tryprostatins, fumitremorgins and verruculogen.
References: [2019]

[EC 1.14.14.118 created 2013 as EC 1.14.13.176, transferred 2018 to EC 1.14.14.118]

EC 1.14.14.119

Accepted name: fumitremorgin C monooxygenase
Reaction: fumitremorgin C + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = 12 α ,13 α -dihydroxyfumitremorgin C + 2 [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): *ftmG* (gene name)
Systematic name: fumitremorgin C,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12 α ,13 α -dihydroxyfumitremorgin C-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Involved in the biosynthetic pathway of the indole alkaloid verruculogen.
References: [2019]

[EC 1.14.14.119 created 2013 as EC 1.14.13.177, transferred 2018 to EC 1.14.14.119]

EC 1.14.14.120

Accepted name: dammarenediol 12-hydroxylase
Reaction: dammarenediol-II + [reduced NADPH—hemoprotein reductase] + O₂ = protopanaxadiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): protopanaxadiol synthase; CYP716A47
Systematic name: dammarenediol-II,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12β-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from ginseng (*Panax ginseng*). Involved in the biosynthetic pathway of ginsenosides.
References: [1499]

[EC 1.14.14.120 created 2013 as EC 1.14.13.183, transferred 2018 to EC 1.14.14.120]

EC 1.14.14.121

Accepted name: protopanaxadiol 6-hydroxylase
Reaction: protopanaxadiol + [reduced NADPH—hemoprotein reductase] + O₂ = protopanaxatriol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): protopanaxatriol synthase; P6H; CYP716A53v2
Systematic name: protopanaxadiol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6α-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the rhizomes of ginseng (*Panax ginseng*). Involved in the biosynthetic pathway of ginsenosides.
References: [4844, 1498]

[EC 1.14.14.121 created 2013 as EC 1.14.13.184, transferred 2018 to EC 1.14.14.121]

EC 1.14.14.122

Accepted name: oryzalexin E synthase
Reaction: *ent*-sandaracopimaradien-3β-ol + [reduced NADPH—hemoprotein reductase] + O₂ = oryzalexin E + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP76M6
Systematic name: *ent*-sandaracopimaradien-3β-ol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (oryzalexin-E-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Isolated from *Oryza sativa* (rice). Oryzalexin E is a phytoalexin.
References: [4684]

[EC 1.14.14.122 created 2014 as EC 1.14.13.192, transferred 2018 to EC 1.14.14.122]

EC 1.14.14.123

Accepted name: oryzalexin D synthase
Reaction: *ent*-sandaracopimaradien-3β-ol + [reduced NADPH—hemoprotein reductase] + O₂ = oryzalexin D + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP76M8
Systematic name: *ent*-sandaracopimaradien-3β-ol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (oryzalexin-D-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Isolated from *Oryza sativa* (rice). Oryzalexin D is a phytoalexin.
References: [4684]

[EC 1.14.14.123 created 2014 as EC 1.14.13.193, transferred 2018 to EC 1.14.14.123]

EC 1.14.14.124

- Accepted name:** dihydromonacolin L hydroxylase
- Reaction:** dihydromonacolin L acid + O₂ + [reduced NADPH—hemoprotein reductase] = monacolin L acid + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) dihydromonacolin L acid + O₂ + [reduced NADPH—hemoprotein reductase] = 3 α -hydroxy-3,5-dihydromonacolin L acid + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 3 α -hydroxy-3,5-dihydromonacolin L acid = monacolin L acid + H₂O (spontaneous)
- Other name(s):** LovA (ambiguous)
- Systematic name:** dihydromonacolin L acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein. The dehydration of 3 α -hydroxy-3,5-dihydromonacolin L acid is believed to be spontaneous [4327, 2983]. The enzyme from fungi also catalyses the reaction of EC 1.14.14.125, monacolin L hydroxylase [225].
- References:** [4327, 2983, 225]

[EC 1.14.14.124 created 2014 as EC 1.14.13.197, transferred 2018 to EC 1.14.14.124]

EC 1.14.14.125

- Accepted name:** monacolin L hydroxylase
- Reaction:** monacolin L acid + O₂ + [reduced NADPH—hemoprotein reductase] = monacolin J acid + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** LovA (ambiguous)
- Systematic name:** monacolin L acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from fungi also catalyses the reaction of EC 1.14.14.124, dihydromonacolin L hydroxylase.
- References:** [225]

[EC 1.14.14.125 created 2014 as EC 1.14.13.198, transferred 2018 to EC 1.14.14.125]

EC 1.14.14.126

- Accepted name:** β -amyrin 28-monooxygenase
- Reaction:** β -amyrin + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = oleanolate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) β -amyrin + O₂ + [reduced NADPH—hemoprotein reductase] = erythrodiol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) erythrodiol + O₂ + [reduced NADPH—hemoprotein reductase] = oleanolic aldehyde + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) oleanolic aldehyde + O₂ + [reduced NADPH—hemoprotein reductase] = oleanolate + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** CYP716A52v2; CYP716A12; CYP16A75; β -amyrin 28-oxidase
- Systematic name:** β -amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (28-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein found in plants. The enzyme is involved in the biosynthesis of oleanane-type triterpenoids, such as ginsenoside Ro. The enzyme from *Medicago truncatula* (barrel medic) (CYP716A12) can also convert α -amyrin and lupeol to ursolic acid and betulinic acid, respectively. The enzyme from *Maesa lanceolata* (false assegai) (CYP16A75) does not catalyse the reaction to completion, resulting in accumulation of both intermediates.
- References:** [1226, 1500, 2906]

[EC 1.14.14.126 created 2015 as EC 1.14.13.201, transferred 2018 to EC 1.14.14.126]

EC 1.14.14.127

- Accepted name:** methyl farnesoate epoxidase

Reaction: methyl (2*E*,6*E*)-farnesoate + [reduced NADPH—hemoprotein reductase] + O₂ = juvenile hormone III + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP15A1

Systematic name: methyl (2*E*,6*E*)-farnesoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme, found in insects except for Lepidoptera (moths and butterflies) is specific for methyl farnesoate (*cf.* EC 1.14.14.128, farnesoate epoxidase) [1622, 811].

References: [1622, 811]

[EC 1.14.14.127 created 2015 as EC 1.14.13.202, transferred 2018 to EC 1.14.14.127]

EC 1.14.14.128

Accepted name: farnesoate epoxidase

Reaction: (2*E*,6*E*)-farnesoate + [reduced NADPH—hemoprotein reductase] + O₂ = juvenile-hormone-III carboxylate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP15C1

Systematic name: (2*E*,6*E*)-farnesoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme, found in Lepidoptera (moths and butterflies), is specific for farnesoate (*cf.* EC 1.14.14.127, methyl farnesoate epoxidase) [810, 811]. It is involved in the synthesis of juvenile hormone.

References: [810, 811]

[EC 1.14.14.128 created 2015 as EC 1.14.13.203, transferred 2018 to EC 1.14.14.128]

EC 1.14.14.129

Accepted name: long-chain acyl-CoA ω-monooxygenase

Reaction: (1) oleoyl-CoA + [reduced NADPH—hemoprotein reductase] + O₂ = 18-hydroxyoleoyl-CoA + [oxidized NADPH—hemoprotein reductase] + H₂O

(2) linoleoyl-CoA + [reduced NADPH—hemoprotein reductase] + O₂ = 18-hydroxylinoleoyl-CoA + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): long-chain acyl-CoA ω-hydroxylase; CYP86A₂2 (gene name)

Systematic name: long-chain acyl-CoA,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (ω-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzymes from solanaceous plants are involved in the biosynthesis of stigmatic estolide, a lipid-based polyester that forms a major component of the exudate.

References: [1496]

[EC 1.14.14.129 created 2015 as EC 1.14.13.204, transferred 2018 to EC 1.14.14.129]

EC 1.14.14.130

Accepted name: laurate 7-monooxygenase

Reaction: dodecanoate + [reduced NADPH—hemoprotein reductase] + O₂ = 7-hydroxydodecanoate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): CYP703A2 (gene name)

Systematic name: dodecanoate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (7-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. The enzyme is involved in the synthesis of sporopollenin - a complex polymer found at the outer layer of spores and pollen. It can also act on decanoate (C₁₀), myristate (C₁₄), and palmitate (C₁₆) with lower activity. The enzyme also produces a small amount of products that are hydroxylated at neighboring positions (C-6, C-8 and C-9).

References: [2879]

[EC 1.14.14.130 created 2015 as EC 1.14.13.206, transferred 2018 to EC 1.14.14.130]

EC 1.14.14.131

- Accepted name:** bursehernin 5'-monooxygenase
Reaction: (–)-burshehernin + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-5'-demethylatein + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP71CU1 (gene name); bursehernin 5'-hydroxylase
Systematic name: (–)-burshehernin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (5'-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein characterized from the plant *Sinopodophyllum hexandrum*. The enzyme is involved in the biosynthetic pathway of podophyllotoxin, a non-alkaloid toxin lignan whose derivatives are important anticancer drugs.
References: [2365]

[EC 1.14.14.131 created 2016 as EC 1.14.13.213, transferred 2018 to EC 1.14.14.131]

EC 1.14.14.132

- Accepted name:** (–)-4'-demethyl-deoxypodophyllotoxin 4-hydroxylase
Reaction: (–)-4'-demethyldeoxypodophyllotoxin + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-4'-demethylepipodophyllotoxin + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82D61 (gene name)
Systematic name: (–)-deoxypodophyllotoxin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (4-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein characterized from the plant *Sinopodophyllum hexandrum*. The enzyme produces the direct precursor to etoposide, a potent anticancer drug. It can also act on (–)-deoxypodophyllotoxin with lower efficiency.
References: [2365]

[EC 1.14.14.132 created 2016 as EC 1.14.13.214, transferred 2018 to EC 1.14.14.132]

EC 1.14.14.133

- Accepted name:** 1,8-cineole 2-*endo*-monooxygenase
Reaction: 1,8-cineole + [reduced flavodoxin] + O₂ = 2-*endo*-hydroxy-1,8-cineole + [oxidized flavodoxin] + H₂O
Other name(s): P450_{cin}; CYP176A; CYP176A1
Systematic name: 1,8-cineole,[reduced flavodoxin]:oxygen oxidoreductase (2-*endo*-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein that uses a flavodoxin-like redox partner to reduce the heme iron. Isolated from the bacterium *Citrobacter braakii*, which can use 1,8-cineole as the sole source of carbon.
References: [1568, 2761, 2119, 2762]

[EC 1.14.14.133 created 2012 as EC 1.14.13.156, transferred 2018 to EC 1.14.14.133]

EC 1.14.14.134

- Accepted name:** β-amyrin 24-hydroxylase
Reaction: (1) β-amyrin + [reduced NADPH—hemoprotein reductase] + O₂ = 24-hydroxy-β-amyrin + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) sophoradiol + [reduced NADPH—hemoprotein reductase] + O₂ = 24-hydroxysophoradiol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): sophoradiol 24-hydroxylase; CYP93E1
Systematic name: β-amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (24-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Found in plants and participates in the biosynthesis of soybean saponins.
References: [3854]

[EC 1.14.14.134 created 2011 as EC 1.14.99.43, transferred 2018 to EC 1.14.14.134]

EC 1.14.14.135

- Accepted name:** glyceollin synthase
- Reaction:** (1) (6a*S*,11a*S*)-3,6a,9-trihydroxy-2-prenylpterocarpan + [reduced NADPH—hemoprotein reductase] + O₂ = glyceollin II + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(2) (6a*S*,11a*S*)-3,6a,9-trihydroxy-2-prenylpterocarpan + [reduced NADPH—hemoprotein reductase] + O₂ = glyceollin III + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(3) (6a*S*,11a*S*)-3,6a,9-trihydroxy-4-prenylpterocarpan + [reduced NADPH—hemoprotein reductase] + O₂ = glyceollin I + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
- Other name(s):** dimethylallyl-3,6a,9-trihydroxypterocarpan cyclase
- Systematic name:** (6a*S*,11a*S*)-3,6a,9-trihydroxy-2-prenylpterocarpan,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (cyclizing)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein purified from soybean.
- References:** [4580]

[EC 1.14.14.135 created 2004 as EC 1.14.13.85, transferred 2018 to EC 1.14.14.135]

EC 1.14.14.136

- Accepted name:** deoxysarpagine hydroxylase
- Reaction:** 10-deoxysarpagine + [reduced NADPH—hemoprotein reductase] + O₂ = sarpagine + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** DOSH
- Systematic name:** 10-deoxysarpagine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (10-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Rauvolfia serpentina*.
- References:** [4830]

[EC 1.14.14.136 created 2005 as EC 1.14.13.91, transferred 2018 to EC 1.14.14.136]

EC 1.14.14.137

- Accepted name:** (+)-abscisic acid 8'-hydroxylase
- Reaction:** (+)-abscisate + [reduced NADPH—hemoprotein reductase] + O₂ = 8'-hydroxyabscisate + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** (+)-ABA 8'-hydroxylase; ABA 8'-hydroxylase; CYP707A1 (gene name)
- Systematic name:** abscisate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8'-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein found in plants. Catalyses the first step in the oxidative degradation of abscisic acid and is considered to be the pivotal enzyme in controlling the rate of degradation of this plant hormone [792]. CO inhibits the reaction, but its effects can be reversed by the presence of blue light [792]. The 8'-hydroxyabscisate formed can be converted into (–)-phaseic acid, most probably spontaneously.
- References:** [792, 2266, 3633]

[EC 1.14.14.137 created 2005 as EC 1.14.13.93, transferred 2018 EC 1.14.14.137]

EC 1.14.14.138

- Accepted name:** lithocholate 6β-hydroxylase
- Reaction:** lithocholate + [reduced NADPH—hemoprotein reductase] + O₂ = 6β-hydroxylithocholate + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** lithocholate 6β-monooxygenase; CYP3A10; 6β-hydroxylase; cytochrome P450 3A10; lithocholic acid 6β-hydroxylase
- Systematic name:** lithocholate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6β-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein from *Mesocricetus auratus* (golden hamster). Expression of the gene for this enzyme is 50-fold higher in male compared to female hamsters [4237].
- References:** [4237, 598, 4086, 3607]

[EC 1.14.14.138 created 2005 as EC 1.14.13.94, transferred 2018 to EC 1.14.14.138]

EC 1.14.14.139

- Accepted name:** 5 β -cholestane-3 α ,7 α -diol 12 α -hydroxylase
- Reaction:** (1) 5 β -cholestane-3 α ,7 α -diol + [reduced NADPH—hemoprotein reductase] + O₂ = 5 β -cholestane-3 α ,7 α ,12 α -triol + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) 7 α -hydroxycholest-4-en-3-one + [reduced NADPH—hemoprotein reductase] + O₂ = 7 α ,12 α -dihydroxycholest-4-en-3-one + [oxidized NADPH—hemoprotein reductase] + H₂O
(3) chenodeoxycholate + [reduced NADPH—hemoprotein reductase] + O₂ = cholate + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** 5 β -cholestane-3 α ,7 α -diol 12 α -monooxygenase; sterol 12 α -hydroxylase (ambiguous); CYP8B1; cytochrome *P*450 8B1; 7 α -hydroxycholest-4-en-3-one 12 α -hydroxylase; 7 α -hydroxy-4-cholesten-3-one 12 α -monooxygenase; chenodeoxycholate 12 α monooxygenase
- Systematic name:** 5 β -cholestane-3 α ,7 α -diol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (12 α -hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein found in mammals. This is the key enzyme in the biosynthesis of the bile acid cholate. The enzyme can also hydroxylate 5 β -cholestane-3 α ,7 α -diol at the 25 and 26 position, but to a lesser extent [1516].
- References:** [1515, 1516, 1828, 1023, 2566, 869, 4770, 3607, 1085]

[EC 1.14.14.139 created 2005 as EC 1.14.13.96, transferred 2018 to EC 1.14.14.139 (EC 1.14.18.8 created 2005 as EC 1.14.13.95, transferred 2015 to EC 1.14.18.8, incorporated 2020) , modified 2020]

[1.14.14.140 Transferred entry. licodione synthase. Now included with EC 1.14.14.162, flavanone 2-hydroxylase]

[EC 1.14.14.140 created 2004 as EC 1.14.13.87, transferred 2018 to EC 1.14.14.140, transferred 2018 to EC 1.14.14.162, deleted 2018]

EC 1.14.14.141

- Accepted name:** psoralen synthase
- Reaction:** (+)-marmesin + [reduced NADPH—hemoprotein reductase] + O₂ = psoralen + [oxidized NADPH—hemoprotein reductase] + acetone + 2 H₂O
- Other name(s):** CYP71AJ1
- Systematic name:** (+)-marmesin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
- Comments:** This microsomal cytochrome *P*-450 (heme-thiolate) enzyme is rather specific for (+)-marmesin, although it can also accept 5-hydroxymarmesin to a much lesser extent. Furanocoumarins protect plants from fungal invasion and herbivore attack. (+)-Columbianetin, the angular furanocoumarin analogue of the linear furanocoumarin (+)-marmesin, acts as a competitive inhibitor even though it is not a substrate.
- References:** [2351]

[EC 1.14.14.141 created 2007 as EC 1.14.13.102, transferred 2018 to EC 1.14.14.141]

EC 1.14.14.142

- Accepted name:** 8-dimethylallylnaringenin 2'-hydroxylase
- Reaction:** sophoraflavanone B + [reduced NADPH—hemoprotein reductase] + O₂ = leachianone G + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** 8-DMAN 2'-hydroxylase
- Systematic name:** sophoraflavanone-B,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2'-hydroxylating)
- Comments:** A membrane-bound cytochrome *P*-450 (heme-thiolate) protein that is associated with the endoplasmic reticulum [4738, 4901]. This enzyme is specific for sophoraflavanone B as substrate. Along with EC 2.5.1.70 (naringenin 8-dimethylallyltransferase) and EC 2.5.1.71 (leachianone G 2''-dimethylallyltransferase), this enzyme forms part of the sophoraflavanone G biosynthetic pathway.
- References:** [4738, 4901]

[EC 1.14.14.142 created 2007 as EC 1.14.13.103, transferred 2018 to EC 1.14.14.142]

EC 1.14.14.143

- Accepted name:** (+)-menthofuran synthase
Reaction: (+)-pulegone + [reduced NADPH—hemoprotein reductase] + O₂ = (+)-menthofuran + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): menthofuran synthase; (+)-pulegone 9-hydroxylase; (+)-MFS; cytochrome P450 menthofuran synthase
Systematic name: (+)-pulegone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (9-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The conversion of substrate into product involves the hydroxylation of the *syn*-methyl (C₉), intramolecular cyclization to the hemiketal and dehydration to the furan [310]. This is the second cytochrome *P*-450-mediated step of monoterpene metabolism in peppermint, with the other step being catalysed by EC 1.14.14.99, (*S*)-limonene 3-monooxygenase [310].
References: [310, 2617]

[EC 1.14.14.143 created 2008 as EC 1.14.13.104, transferred 2018 to EC 1.14.14.143]

EC 1.14.14.144

- Accepted name:** abieta-7,13-diene hydroxylase
Reaction: abieta-7,13-diene + [reduced NADPH—hemoprotein reductase] + O₂ = abieta-7,13-dien-18-ol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): abietadiene hydroxylase (ambiguous)
Systematic name: abieta-7,13-diene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (18-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. This enzyme catalyses a step in the pathway of abietic acid biosynthesis. The activity has been demonstrated in cell-free stem extracts of *Abies grandis* (grand fir) and *Pinus contorta* (lodgepole pine). Activity is induced by wounding of the plant tissue [1232].
References: [1230, 1232]

[EC 1.14.14.144 created 2009 as EC 1.14.13.108, modified 2012, transferred 2018 to EC 1.14.14.144]

EC 1.14.14.145

- Accepted name:** abieta-7,13-dien-18-ol hydroxylase
Reaction: abieta-7,13-dien-18-ol + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = abieta-7,13-dien-18-oate + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) abieta-7,13-dien-18-ol + [reduced NADPH—hemoprotein reductase] + O₂ = abieta-7,13-dien-18,18-diol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) abieta-7,13-dien-18,18-diol = abieta-7,13-dien-18-al + H₂O (spontaneous)
(1c) abieta-7,13-dien-18-al + [reduced NADPH—hemoprotein reductase] + O₂ = abieta-7,13-dien-18-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP720B1; PtAO; abietadienol hydroxylase (ambiguous)
Systematic name: abieta-7,13-dien-18-ol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (18-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. This enzyme catalyses a step in the pathway of abietic acid biosynthesis. The activity has been demonstrated in cell-free stem extracts of *Abies grandis* (grand fir) and *Pinus contorta* (lodgepole pine) [1230], and the gene encoding the enzyme has been identified in *Pinus taeda* (loblolly pine) [3531]. The recombinant enzyme catalyses the oxidation of multiple diterpene alcohol and aldehydes, including levopimaradienol, isopimara-7,15-dienol, isopimara-7,15-dienal, dehydroabietadienol and dehydroabietadienal. It is not able to oxidize abietadiene.
References: [1230, 1232, 3531]

[EC 1.14.14.145 created 2009 as EC 1.14.13.109, modified 2012, transferred 2018 to EC 1.14.14.145]

EC 1.14.14.146

Accepted name: geranylgeraniol 18-hydroxylase
Reaction: geranylgeraniol + [reduced NADPH—hemoprotein reductase] + O₂ = 18-hydroxygeranylgeraniol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): GGOH-18-hydroxylase
Systematic name: geranylgeraniol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (18-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Croton sublyratus*.
References: [4214]

[EC 1.14.14.146 created 2009 as EC 1.14.13.110, transferred 2018 to EC 1.14.14.146]

EC 1.14.14.147

Accepted name: 22 α -hydroxysteroid 23-monooxygenase
Reaction: (1) 3-*epi*-6-deoxocathasterone + [reduced NADPH—hemoprotein reductase] + O₂ = 6-deoxytyphasterol + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) (2*S*,24*R*)-22-hydroxy-5 α -ergostan-3-one + [reduced NADPH—hemoprotein reductase] + O₂ = 3-dehydro-6-deoxoteasterone + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): cytochrome *P*450 90C1; CYP90D1; CYP90C1; 3-*epi*-6-deoxocathasterone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*C*-23-hydroxylating); 3-*epi*-6-deoxocathasterone 23-monooxygenase
Systematic name: 22 α -hydroxysteroid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*C*-23-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in brassinosteroid biosynthesis in plants. The enzyme has a relaxed substrate specificity, and *C*-23 hydroxylation can occur at different stages in the pathway. In *Arabidopsis thaliana* two isozymes, encoded by the CYP90C1 and CYP90D1 genes, have redundant activities.
References: [2092, 3142]

[EC 1.14.14.147 created 2010 as EC 1.14.13.112, transferred 2018 to EC 1.14.14.147, modified 2022]

EC 1.14.14.148

Accepted name: angelicin synthase
Reaction: (+)-columbianetin + [reduced NADPH—hemoprotein reductase] + O₂ = angelicin + [oxidized NADPH—hemoprotein reductase] + acetone + 2 H₂O
Other name(s): CYP71AJ4 (gene name)
Systematic name: (+)-columbianetin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme from wild parsnip is involved in the formation of angular furanocoumarins. Attacks its substrate by *syn*-elimination of hydrogen from *C*-3'.
References: [2350]

[EC 1.14.14.148 created 2010 as EC 1.14.13.115, transferred 2018 to EC 1.14.14.148]

EC 1.14.14.149

Accepted name: 5-epiaristolochene 1,3-dihydroxylase
Reaction: 5-epiaristolochene + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = capsidiol + 2 [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): 5-*epi*-aristolochene 1,3-dihydroxylase; EAH; CYP71D20
Systematic name: 5-epiaristolochene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (1- and 3-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. Kinetic studies suggest that 1 β -hydroxyepiaristolochene is mainly formed first followed by hydroxylation at C-3. However the reverse order via 3 α -hydroxyepiaristolochene does occur.

References: [3440, 4172]

[EC 1.14.14.149 created 2011 as EC 1.14.13.119, transferred 2018 to EC 1.14.14.149]

EC 1.14.14.150

Accepted name: costunolide synthase

Reaction: germacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = (+)-costunolide + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) germacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = 6 α -hydroxygermacra-1(10),4,11(13)-trien-12-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 6 α -hydroxygermacra-1(10),4,11(13)-trien-12-oate = (+)-costunolide + H₂O (spontaneous)

Other name(s): CYP71BL2

Systematic name: germacra-1(10),4,11(13)-trien-12-oate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6 α -hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein from chicory plants. The enzyme hydroxylates carbon C-6 of germacra-1(10),4,11(13)-trien-12-oate to give 6 α -hydroxygermacra-1(10),4,11(13)-trien-12-oate, which spontaneously cyclises to form the lactone ring.

References: [848]

[EC 1.14.14.150 created 2011 as EC 1.14.13.120, transferred 2018 to EC 1.14.14.150]

EC 1.14.14.151

Accepted name: premnaspirodiene oxygenase

Reaction: (–)-vetispiradiene + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = solavetivone + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) (–)-vetispiradiene + [reduced NADPH—hemoprotein reductase] + O₂ = solavetivol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) solavetivol + [reduced NADPH—hemoprotein reductase] + O₂ = solavetivone + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): HPO; *Hyoscyamus muticus* premnaspirodiene oxygenase; CYP71D55

Systematic name: (–)-vetispiradiene,[reduced NADPH—hemoprotein reductase]:oxygen 2 α -oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from the plant *Hyoscyamus muticus* also hydroxylates valencene at C-2 to give the α -hydroxy compound, nootkatol, and this is converted into nootkatone. 5-Epiaristolochene and epieremophilene are hydroxylated at C-2 to give a 2 β -hydroxy derivatives that are not oxidized further.

References: [4171]

[EC 1.14.14.151 created 2011 as EC 1.14.13.121, transferred 2018 to EC 1.14.14.151]

EC 1.14.14.152

Accepted name: β -amyryn 11-oxidase

Reaction: β -amyryn + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = 11-oxo- β -amyryn + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) β -amyryn + [reduced NADPH—hemoprotein reductase] + O₂ = 11 α -hydroxy- β -amyryn + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 11 α -hydroxy- β -amyryn + [reduced NADPH—hemoprotein reductase] + O₂ = 11-oxo- β -amyryn + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): CYP88D6

Systematic name: β -amyryn,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Glycyrrhiza uralensis* (Chinese licorice) that participates in the glycyrrhizin biosynthesis pathway. The enzyme is also able to oxidize 30-hydroxy- β -amyrin to 11 α ,30-dihydroxy- β -amyrin but this is not thought to be part of glycyrrhizin biosynthesis.

References: [3789]

[EC 1.14.14.152 created 2011 as EC 1.14.13.134, transferred 2018 to EC 1.14.14.152]

EC 1.14.14.153

Accepted name: indole-2-monooxygenase

Reaction: indole + [reduced NADPH—hemoprotein reductase] + O₂ = indolin-2-one + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): BX2 (gene name); CYP71C4 (gene name)

Systematic name: indole,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme is involved in the biosynthesis of protective and allelopathic benzoxazinoids in some plants, most commonly from the family of Poaceae (grasses).

References: [1175, 1341]

[EC 1.14.14.153 created 2012 as EC 1.14.13.137, transferred 2018 to EC 1.14.14.153]

EC 1.14.14.154

Accepted name: sterol 14 α -demethylase

Reaction: a 14 α -methylsteroid + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = a Δ ¹⁴-steroid + formate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)

(1a) a 14 α -methylsteroid + [reduced NADPH—hemoprotein reductase] + O₂ = a 14 α -hydroxymethylsteroid + [oxidized NADPH—hemoprotein reductase] + H₂O

(1b) a 14 α -hydroxysteroid + [reduced NADPH—hemoprotein reductase] + O₂ = a 14 α -formylsteroid + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

(1c) a 14 α -formylsteroid + [reduced NADPH—hemoprotein reductase] + O₂ = a Δ ¹⁴-steroid + formate + [oxidized NADPH—hemoprotein reductase] + H₂O

Other name(s): obtusifoliol 14-demethylase; lanosterol 14-demethylase; lanosterol 14 α -demethylase; sterol 14-demethylase; CYP51 (gene name); ERG11 (gene name)

Systematic name: sterol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (14-methyl cleaving)

Comments: This cytochrome *P*-450 (heme-thiolate) enzyme acts on a range of steroids with a 14 α -methyl group, such as obtusifoliol and lanosterol. The enzyme catalyses a hydroxylation and a reduction of the 14 α -methyl group, followed by a second hydroxylation, resulting in the elimination of formate and formation of a 14(15) double bond.

References: [65, 4813, 116, 114, 115, 186]

[EC 1.14.14.154 created 2001 as EC 1.14.13.70, modified 2013, transferred 2018 EC 1.14.14.154]

EC 1.14.14.155

Accepted name: 3,6-diketocamphane 1,2-monooxygenase

Reaction: (–)-bornane-2,5-dione + O₂ + FMNH₂ = (–)-5-oxo-1,2-campholide + FMN + H₂O

Other name(s): 3,6-diketocamphane lactonizing enzyme; 3,6-DKCMO

Systematic name: (–)-bornane-2,5-dione,FMNH₂:oxygen oxidoreductase (1,2-lactonizing)

Comments: A Baeyer-Villiger monooxygenase isolated from camphor-grown strains of *Pseudomonas putida* and encoded on the cam plasmid. Involved in the degradation of (–)-camphor. Requires a dedicated NADH—FMN reductase [*cf.* EC 1.5.1.42, FMN reductase (NADH)] [1858, 1841]. The product spontaneously converts to [(1*R*)-2,2,3-trimethyl-5-oxocyclopent-3-enyl]acetate.

References: [1858, 1841]

[EC 1.14.14.155 created 2018]

EC 1.14.14.156

- Accepted name:** tryptophan *N*-monooxygenase
- Reaction:** L-tryptophan + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = (*E*)-indol-3-ylacetaldoxime + 2 [oxidized NADPH—hemoprotein reductase] + CO₂ + 3 H₂O (overall reaction)
(1a) L-tryptophan + [reduced NADPH—hemoprotein reductase] + O₂ = *N*-hydroxy-L-tryptophan + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) *N*-hydroxy-L-tryptophan + [reduced NADPH—hemoprotein reductase] + O₂ = *N,N*-dihydroxy-L-tryptophan + [oxidized NADPH—hemoprotein reductase] + H₂O
(1c) *N,N*-dihydroxy-L-tryptophan = (*E*)-indol-3-ylacetaldoxime + CO₂ + H₂O
- Other name(s):** tryptophan *N*-hydroxylase; CYP79B1; CYP79B2; CYP79B3
- Systematic name:** L-tryptophan,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (*N*-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein from the plant *Arabidopsis thaliana*. This enzyme catalyses two successive *N*-hydroxylations of L-tryptophan, the first steps in the biosynthesis of both auxin and the indole alkaloid phytoalexin camalexin. The product of the two hydroxylations, *N,N*-dihydroxy-L-tryptophan, is extremely labile and dehydrates spontaneously. The dehydrated product is then subject to a decarboxylation that produces an oxime. It is still not known whether the decarboxylation is spontaneous or catalysed by the enzyme.
- References:** [2804, 1770, 4907, 3023]

[EC 1.14.14.156 created 2011 as EC 1.14.13.125, transferred 2018 to EC 1.14.14.156]

EC 1.14.14.157

- Accepted name:** indolin-2-one monooxygenase
- Reaction:** indolin-2-one + [reduced NADPH—hemoprotein reductase] + O₂ = 3-hydroxyindolin-2-one + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** BX3 (gene name); CYP71C2 (gene name)
- Systematic name:** indolin-2-one,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein. The enzyme is involved in the biosynthesis of protective and allelopathic benzoxazinoids in some plants, most commonly from the family of Poaceae (grasses).
- References:** [1175, 1341]

[EC 1.14.14.157 created 2012 as EC 1.14.13.138, transferred 2018 to EC 1.14.14.157]

EC 1.14.14.158

- Accepted name:** carotenoid ϵ hydroxylase
- Reaction:** (1) α -carotene + [reduced NADPH-hemoprotein reductase] + O₂ = α -cryptoxanthin + [oxidized NADPH-hemoprotein reductase] + H₂O
(2) zeinoxanthin + [reduced NADPH-hemoprotein reductase] + O₂ = lutein + [oxidized NADPH-hemoprotein reductase] + H₂O
- Other name(s):** CYP97C1; LUT1; CYP97C; carotene ϵ -monooxygenase
- Systematic name:** α -carotene,[reduced NADPH-hemoprotein reductase]:oxygen oxidoreductase (3-hydroxylating)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein.
- References:** [3339, 4285, 4030, 595, 3474]

[EC 1.14.14.158 created 2011 as EC 1.14.99.45, transferred 2018 to EC 1.14.14.158]

EC 1.14.14.159

- Accepted name:** dolabradiene monooxygenase
- Reaction:** (1) dolabradiene + O₂ + [reduced NADPH—hemoprotein reductase] = 15,16-epoxydolabrene + H₂O + [oxidized NADPH—hemoprotein reductase]

(2) 15,16-epoxydolabrene + O₂ + [reduced NADPH—hemoprotein reductase] = 3β-hydroxy-15,16-epoxydolabrene + H₂O + [oxidized NADPH—hemoprotein reductase]

- Other name(s):** CYP71Z16 (gene name)
Systematic name: dolabradiene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3β-hydroxy-15,16-epoxydolabrene-forming)
Comments: A cytochrome *P*-450 (heme thiolate) enzyme characterized from maize. The enzyme catalyses the epoxidation of dolabradiene at C-16, followed by hydroxylation at C-3.
References: [2604]

[EC 1.14.14.159 created 2018]

EC 1.14.14.160

- Accepted name:** zealexin A1 synthase
Reaction: (*S*)-β-macrocarpene + 3 O₂ + 3 [reduced NADPH—hemoprotein reductase] = zealexin A1 + 4 H₂O + 3 [oxidized NADPH—hemoprotein reductase] (overall reaction)
(1a) (*S*)-β-macrocarpene + O₂ + [reduced NADPH—hemoprotein reductase] = [(4*S*)-4-(5,5-dimethylcyclohex-1-en-1-yl)-cyclohex-1-en-1-yl]methanol + H₂O + [oxidized NADPH—hemoprotein reductase]
(1b) [(4*S*)-4-(5,5-dimethylcyclohex-1-en-1-yl)-cyclohex-1-en-1-yl] methanol + O₂ + [reduced NADPH—hemoprotein reductase] = (4*S*)-4-(5,5-dimethylcyclohex-1-en-1-yl)cyclohex-1-ene-1-carbaldehyde + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
(1c) (4*S*)-4-(5,5-dimethylcyclohex-1-en-1-yl)cyclohex-1-ene-1-carbaldehyde + O₂ + [reduced NADPH—hemoprotein reductase] = zealexin A1 + H₂O + [oxidized NADPH—hemoprotein reductase]
Other name(s): CYP71Z18 (gene name)
Systematic name: (*S*)-β-macrocarpene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (zealexin A1-forming)
Comments: A cytochrome *P*-450 (heme thiolate) enzyme characterized from maize. The enzyme sequentially oxidizes(*S*)-β-macrocarpene via alcohol and aldehyde intermediates to form zealexin A1, a maize phytoalexin that provides biochemical protection against fungal infection.
References: [2641]

[EC 1.14.14.160 created 2018]

EC 1.14.14.161

- Accepted name:** nepetalactol monooxygenase
Reaction: (+)-*cis,trans*-nepetalactol + 3 [reduced NADPH—hemoprotein reductase] + 3 O₂ = 7-deoxyloganetate + 3 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) (+)-*cis,trans*-nepetalactol + [reduced NADPH—hemoprotein reductase] + O₂ = 7-deoxyloganic alcohol + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 7-deoxyloganic alcohol + [reduced NADPH—hemoprotein reductase] + O₂ = iridotrial + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1c) iridotrial + [reduced NADPH—hemoprotein reductase] + O₂ = 7-deoxyloganetate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP76A26 (gene name); iridoid oxidase (misleading)
Systematic name: (+)-*cis,trans*-nepetalactol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (hydroxylating)
Comments: The enzyme, characterized from the plant *Catharanthus roseus*, is a cytochrome *P*-450 (heme thiolate) protein. It catalyses three successive reactions in the pathway leading to biosynthesis of monoterpenoid indole alkaloids.
References: [2797]

[EC 1.14.14.161 created 2018]

EC 1.14.14.162

- Accepted name:** flavanone 2-hydroxylase
Reaction: a flavanone + [reduced NADPH—hemoprotein reductase] + O₂ = a 2-hydroxyflavanone + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP93G2 (gene name); CYP93B1 (gene name); (2*S*)-flavanone 2-hydroxylase; licodione synthase
Systematic name: flavanone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (2-hydroxylating)
Comments: A cytochrome *P*-450 (heme thiolate) plant enzyme that catalyses the 2-hydroxylation of multiple flavanones such as (2*S*)-naringenin, (2*S*)-eriodictyol, (2*S*)-pinocembrin, and (2*S*)-liquiritigenin. The products are *meta*-stable and exist in an equilibrium with open forms such as 1-(4-hydroxyphenyl)-3-(2,4,6-trihydroxyphenyl)propane-1,3-dione.
References: [3204, 48, 972]

[EC 1.14.14.162 created 2018. EC 1.14.14.140 created 2004 as EC 1.14.13.87, transferred 2018 to EC 1.14.14.140, transferred 2018 to EC 1.14.14.162]

EC 1.14.14.163

- Accepted name:** (*S*)-1-hydroxy-*N*-methylcanadine 13-hydroxylase
Reaction: (*S*)-1-hydroxy-*N*-methylcanadine + [reduced NADPH—hemoprotein reductase] + O₂ = (13*S*,14*R*)-1,13-dihydroxy-*N*-methylcanadine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82X2 (gene name)
Systematic name: (*S*)-1-hydroxy-*N*-methylcanadine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (13-hydroxylating)
Comments: The enzyme, characterized from the plant *Papaver somniferum* (opium poppy), participates in the biosynthesis of the isoquinoline alkaloid noscapine.
References: [819, 2467, 2464]

[EC 1.14.14.163 created 2018]

EC 1.14.14.164

- Accepted name:** fraxetin 5-hydroxylase
Reaction: fraxetin + [reduced NADPH—hemoprotein reductase] + O₂ = sideretin (reduced form) + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82C4; fraxetin 5-monooxygenase
Systematic name: fraxetin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (5-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in biosynthesis of iron(III)-chelating coumarins in higher plants.
References: [3439]

[EC 1.14.14.164 created 2018]

EC 1.14.14.165

- Accepted name:** indole-3-carbonyl nitrile 4-hydroxylase
Reaction: indole-3-carbonyl nitrile + [reduced NADPH—hemoprotein reductase] + O₂ = 4-hydroxyindole-3-carbonyl nitrile + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82C2
Systematic name: indole-3-carbonyl nitrile,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (4-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein characterized from the plant *Arabidopsis thaliana*. Involved in biosynthesis of small cyanogenic compounds that take part in pathogen defense. The enzyme also catalyses the 5-hydroxylation of xanthotoxin [2268].
References: [2268, 3438]

[EC 1.14.14.165 created 2018]

EC 1.14.14.166

- Accepted name:** (*S*)-*N*-methylcanadine 1-hydroxylase
Reaction: (*S*)-*N*-methylcanadine + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-1-hydroxy-*N*-methylcanadine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82Y1 (gene name)
Systematic name: (*S*)-*N*-methylcanadine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (1-hydroxylating)
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, characterized from the plant *Papaver somniferum* (opium poppy), participates in the biosynthesis of the isoquinoline alkaloid noscapine.
References: [821, 2464]

[EC 1.14.14.166 created 2018]

EC 1.14.14.167

- Accepted name:** (1*S*,14*R*)-13-*O*-acetyl-1-hydroxy-*N*-methylcanadine 8-hydroxylase
Reaction: (1*S*,14*R*)-13-*O*-acetyl-1-hydroxy-*N*-methylcanadine + [reduced NADPH—hemoprotein reductase] + O₂ = (1*S*,14*R*)-13-*O*-acetyl-1,8-dihydroxy-*N*-methylcanadine + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP82X1 (gene name)
Systematic name: (1*S*,14*R*)-13-*O*-acetyl-1-hydroxy-*N*-methylcanadine 8-hydroxylase,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8-hydroxylating)
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, characterized from the plant *Papaver somniferum* (opium poppy), participates in the biosynthesis of the isoquinoline alkaloid noscapine.
References: [819, 2467, 2464]

[EC 1.14.14.167 created 2018]

EC 1.14.14.168

- Accepted name:** germacrene A acid 8β-hydroxylase
Reaction: germacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = 8β-hydroxygermacra-1(10),4,11(13)-trien-12-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): HaG8H; CYP71BL1; CYP71BL6
Systematic name: germacra-1(10),4,11(13)-trien-12-oate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8β-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Helianthus annuus* (common sunflower). The cyclisation of 8β-hydroxygermacra-1(10),4,11(13)-triene-12-oate to inunolide (12,8β) does not seem to occur spontaneously. The enzyme from *Inula hupehensis* also forms some 8α-hydroxygermacra-1(10),4,11(13)-triene-12-oate, which spontaneously cyclises to 8-*epi*-inunolide (12,8α) (*cf.* EC 1.14.14.170 8-*epi*-inunolide synthase).
References: [1177, 1373]

[EC 1.14.14.168 created 2018]

EC 1.14.14.169

- Accepted name:** eupatolide synthase
Reaction: 8β-hydroxygermacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = eupatolide + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) 8β-hydroxygermacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = 6α,8β-dihydroxygermacra-1(10),4,11(13)-trien-12-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 6α,8β-dihydroxygermacra-1(10),4,11(13)-trien-12-oate = eupatolide + H₂O (spontaneous)
Other name(s): CYP71DD6; HaES

Systematic name: 8 β -hydroxygermacra-1(10),4,11(13)-trien-12-oate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (6 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Helianthus annuus* (common sunflower).
References: [1177]

[EC 1.14.14.169 created 2018]

EC 1.14.14.170

Accepted name: 8-*epi*-inunolide synthase
Reaction: germacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = 8-*epi*-inunolide + [oxidized NADPH—hemoprotein reductase] + 2 H₂O (overall reaction)
(1a) germacra-1(10),4,11(13)-trien-12-oate + [reduced NADPH—hemoprotein reductase] + O₂ = 8 α -hydroxygermacra-1(10),4,11(13)-trien-12-oate + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) 8 α -hydroxygermacra-1(10),4,11(13)-trien-12-oate = 8-*epi*-inunolide + H₂O (spontaneous)
Other name(s): CYP71BL1
Systematic name: germacra-1(10),4,11(13)-trien-12-oate,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (8 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the plant *Inula hupehensis*. The enzyme also produces 8 β -hydroxygermacra-1(10),4,11(13)-triene-12-oate (EC 1.14.14.168, germacrene A acid 8 β -hydroxylase).
References: [1373]

[EC 1.14.14.170 created 2018]

EC 1.14.14.171

Accepted name: β -amyrin 16 α -hydroxylase
Reaction: β -amyrin + [reduced NADPH—hemoprotein reductase] + O₂ = 16 α -hydroxy- β -amyrin + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP87D16
Systematic name: β -amyrin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (16 α -hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the plant *Maesa lanceolata* (false assegai). Involved in the biosynthesis of maesasaponins. It also acts on some derivatives of β -amyrin such as erythrodiol or oleanolic acid.
References: [2905, 2906]

[EC 1.14.14.171 created 2019]

EC 1.14.14.172

Accepted name: 3,5,6-trichloropyridin-2-ol monooxygenase
Reaction: (1) 3,5,6-trichloropyridin-2-ol + FADH₂ + O₂ = 3,6-dichloropyridine-2,5-dione + Cl⁻ + FAD + H₂O
(2) 3,6-dichloropyridine-2,5-diol + FADH₂ + O₂ = 6-chloro-3-hydroxypyridine-2,5-dione + Cl⁻ + FAD + H₂O
(3) 6-chloropyridine-2,3,5-triol + FADH₂ + O₂ = 3,6-dihydroxypyridine-2,5-dione + Cl⁻ + FAD + H₂O
Other name(s): *tcpA* (gene name)
Systematic name: 3,5,6-trichloropyridin-2-ol,FADH₂:oxygen oxidoreductase (dechlorinating)
Comments: The enzyme, characterized from a number of bacterial species, participates in the degradation of 3,5,6-trichloropyridin-2-ol (TCP), a metabolite of the common organophosphorus insecticide chlorpyrifos. The enzyme is a multifunctional flavin-dependent monooxygenase that displaces three chlorine atoms by attacking three different positions in the substrate. Each reaction catalysed by the enzyme displaces a single chlorine and results in formation of a dione, which must be reduced by FADH₂ before the monooxygenase could catalyse the next step. The large amount of FADH₂ that is required is generated by a dedicated flavin reductase (TcpX). *cf.* EC 1.14.14.173, 2,4,6-trichlorophenol monooxygenase.

References: [2446, 1086]

[EC 1.14.14.172 created 2020]

EC 1.14.14.173

Accepted name: 2,4,6-trichlorophenol monooxygenase
Reaction: 2,4,6-trichlorophenol + FADH₂ + O₂ = 6-chloro-2-hydroxy-1,4-benzoquinone + 2 Cl⁻ + FAD (overall reaction)
(1a) 2,4,6-trichlorophenol + FADH₂ + O₂ = 2,6-dichloro-1,4-benzoquinone + Cl⁻ + FAD + H₂O
(1b) 2,6-dichloro-1,4-benzoquinone + H₂O = 6-chloro-2-hydroxy-1,4-benzoquinone + Cl⁻
Other name(s): *tcpA* (gene name)
Systematic name: 2,4,6-trichlorophenol,FADH₂:oxygen oxidoreductase (dechlorinating)
Comments: The enzyme, characterized from *Cupriavidus pinatubonensis*, participates in the degradation of 2,4,6-trichlorophenol, a compound that has been used for decades as a wood preservative. The enzyme is a multifunctional flavin-dependent monooxygenase that catalyses two different reactions to displace two chlorine atoms, a monooxygenase reaction followed by a hydrolysis reaction that takes advantage of the reactivity of the product of the first reaction, 2,6-dichloro-1,4-benzoquinone [4716]. The large amount of FADH₂ that is required is generated by a dedicated flavin reductase (TcbB). *cf.* EC 1.14.14.172, 3,5,6-trichloropyridin-2-ol monooxygenase.
References: [2544, 4716, 1582]

[EC 1.14.14.173 created 2020, modified 2022]

EC 1.14.14.174

Accepted name: geranylhydroquinone 3''-hydroxylase
Reaction: geranylhydroquinone + [reduced NADPH—hemoprotein reductase] + O₂ = 3''-hydroxygeranylhydroquinone + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): GHQ 3''-hydroxylase; CYP76B74 (gene name); geranylhydroquinone,NADPH:oxygen oxidoreductase (3''-hydroxylating)
Systematic name: geranylhydroquinone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (3''-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants, where it is part of the biosynthesis pathway of the red naphthoquinone pigment shikonin.
References: [4735, 4532]

[EC 1.14.14.174 created 2010 as EC 1.14.13.116, transferred 2020 to EC 1.14.14.174]

EC 1.14.14.175

Accepted name: ferruginol synthase
Reaction: abieta-8,11,13-triene + [reduced NADPH—hemoprotein reductase] + O₂ = ferruginol + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): miltiradiene oxidase (incorrect); CYP76AH1; miltiradiene,NADPH:oxygen oxidoreductase (ferruginol forming) (incorrect)
Systematic name: abieta-8,11,13-triene,[reduced NADPH—hemoprotein reductase]:oxygen 12-oxidoreductase (ferruginol-forming)
Comments: A cytochrome *P*-450 (heme thiolate) enzyme found in some members of the *Lamiaceae* (mint family). The enzyme from *Rosmarinus officinalis* (rosemary) is involved in biosynthesis of carnosic acid, while the enzyme from the Chinese medicinal herb *Salvia miltiorrhiza* is involved in the biosynthesis of the tanshinones, abietane-type norditerpenoid naphthoquinones that are the main lipophilic bioactive components found in the plant.
References: [1445, 4929, 414]

[EC 1.14.14.175 created 2014 as EC 1.14.13.190, modified 2015, transferred 2020 to EC 1.14.14.175]

EC 1.14.14.176

- Accepted name:** taxadiene 5 α -hydroxylase
Reaction: taxa-4,11-diene + [reduced NADPH—hemoprotein reductase] + O₂ = taxa-4(20),11-dien-5 α -ol + [oxidized NADPH—hemoprotein reductase] + H₂O
Systematic name: taxa-4,11-diene,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (5 α -hydroxylating)
Comments: This microsomal cytochrome-*P*-450 (heme-thiolate) enzyme is involved in the biosynthesis of the diterpenoid antineoplastic drug taxol (paclitaxel). The reaction includes rearrangement of the 4(5)-double bond to a 4(20)-double bond, possibly through allylic oxidation.
References: [1604]

[EC 1.14.14.176 created 2002 as 1.14.99.37, transferred 2020 to EC 1.14.14.176]

EC 1.14.14.177

- Accepted name:** ultra-long-chain fatty acid ω -hydroxylase
Reaction: an ultra-long-chain fatty acid + [reduced NADPH—hemoprotein reductase] + O₂ = an ultra-long-chain ω -hydroxy fatty acid + [oxidized NADPH—hemoprotein reductase] + H₂O
Other name(s): CYP4F22 (gene name)
Systematic name: ultra-long-chain fatty acid,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (ω -hydroxylating)
Comments: The enzyme, which is expressed in the epidermis of mammals, catalyses the ω -hydroxylation of ultra-long-chain fatty acids (C₂₈ to C₃₆). The products are incorporated into acylceramides, epidermis-specific ceramide species that are very important for skin barrier formation.
References: [3144]

[EC 1.14.14.177 created 2021]

EC 1.14.14.178

- Accepted name:** steroid 22*S*-hydroxylase
Reaction: (1) a C₂₇-steroid + O₂ + [reduced NADPH—hemoprotein reductase] = a (22*S*)-22-hydroxy-C₂₇-steroid + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
(2) a C₂₈-steroid + O₂ + [reduced NADPH—hemoprotein reductase] = a (22*S*)-22-hydroxy-C₂₈-steroid + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
(3) a C₂₉-steroid + O₂ + [reduced NADPH—hemoprotein reductase] = a (22*S*)-22-hydroxy-C₂₉-steroid + 2 H₂O + [oxidized NADPH—hemoprotein reductase]
Other name(s): CYP90B1 (gene name); DWF4 (gene name); steroid C-22 hydroxylase
Systematic name: steroid,NADPH—hemoprotein reductase:oxygen 22*S*-oxidoreductase (hydroxylating)
Comments: This plant cytochrome *P*-450 (heme thiolate) enzyme participates in the biosynthesis of brassinosteroids. While *in vivo* substrates include C₂₈-steroids such as campestanol, campesterol, and 6-oxocampestanol, the enzyme is able to catalyse the C-22 hydroxylation of a variety of C₂₇, C₂₈ and C₂₉ steroids.
References: [139, 671, 140, 1212, 3143]

[EC 1.14.14.178 created 2022]

EC 1.14.14.179

- Accepted name:** brassinosteroid 6-oxygenase
Reaction: 6-deoxocastasterone + 2 O₂ + 2 [reduced NADPH—hemoprotein reductase] = castasterone + 3 H₂O + 2 [oxidized NADPH—hemoprotein reductase] (overall reaction)
(1a) 6-deoxocastasterone + O₂ + [reduced NADPH—hemoprotein reductase] = 6 α -hydroxy-6-deoxocastasterone + H₂O + [oxidized NADPH—hemoprotein reductase]

(1b) 6α -hydroxy-6-deoxocastasterone + O₂ + [reduced NADPH—hemoprotein reductase] = castasterone + 2 H₂O + [oxidized NADPH—hemoprotein reductase]

Other name(s): CYP85A1 (gene name); CYP85A2 (gene name); brassinosteroid 6-oxidase
Systematic name: 6-deoxocastasterone,NADPH—hemoprotein reductase:oxygen 6-oxidoreductase (castasterone-forming)

Comments: This cytochrome *P*-450 (heme thiolate) plant enzyme catalyses the C-6 hydroxylation of several brassinosteroid biosynthesis intermediates, and the further oxidation of the hydroxyl group to an oxo group. Substrates include 6-deoxocastasterone, 6-deoxotyphasterol, 3-dehydro-6-deoxoteasterone, and 6-deoxoteasterone. The CYP85A2 isozyme of *Arabidopsis thaliana* (but not the CYP85A1 isozyme) also catalyses the activity of EC 1.14.14.180, brassinolide synthase.

References: [3864, 3291]

[EC 1.14.14.179 created 2022]

EC 1.14.14.180

Accepted name: brassinolide synthase

Reaction: castasterone + O₂ + [reduced NADPH—hemoprotein reductase] = brassinolide + 2 H₂O + [oxidized NADPH—hemoprotein reductase]

Other name(s): CYP85A2 (gene name); CYP85A3 (gene name)

Systematic name: castasterone,NADPH—hemoprotein reductase:oxygen oxidoreductase (lactonizing, brassinolide-forming)

Comments: This cytochrome *P*-450 (heme thiolate) plant enzyme catalyses the lactonization of several brassinosteroids, including castasterone, teasterone, and typhasterol. The CYP85A2 enzyme of *Arabidopsis thaliana* also catalyses the activity of EC 1.14.14.179, brassinosteroid 6-oxygenase.

References: [3102, 2109, 2027]

[EC 1.14.14.180 created 2022]

EC 1.14.14.181

Accepted name: sulfoquinovose monooxygenase

Reaction: 6-sulfo-D-quinovose + FMNH₂ + O₂ = 6-dehydro-D-glucose + FMN + sulfite + H₂O

Other name(s): 6-deoxy-6-sulfo-D-glucose monooxygenase; *smoC* (gene name); *squD* (gene name)

Systematic name: 6-sulfo-D-quinovose,FMNH₂:oxygen oxidoreductase

Comments: The enzyme, characterized from the bacteria *Agrobacterium fabrum* and *Rhizobium oryzae*, is involved in a D-sulfoquinovose degradation pathway. FMNH₂ is provided by an associated FMN reductase [SmoA, EC 1.5.1.42, FMN reductase (NADH)].

References: [2516, 3827]

[EC 1.14.14.181 created 20022]

EC 1.14.14.182

Accepted name: taxoid 7β-hydroxylase

Reaction: (1) taxusin + [reduced NADPH—hemoprotein reductase] + O₂ = 7β-hydroxytaxusin + [oxidized NADPH—hemoprotein reductase] + H₂O

(2) 2α-hydroxytaxusin + [reduced NADPH—hemoprotein reductase] + O₂ = 2α,7β-dihydroxytaxusin + [oxidized NADPH—hemoprotein reductase] + H₂O

Systematic name: taxusin, [reduced NADPH—hemoprotein reductase]:oxygen 7-oxidoreductase

Comments: A cytochrome *P*-450 (heme-thiolate) protein from the yew tree *Taxus cuspidata*. Does not act on earlier intermediates in taxol biosynthesis.

References: [616, 615]

[EC 1.14.14.182 created 2012 as EC 1.14.13.147, transferred 2022 to EC 1.14.14.182]

EC 1.14.14.183

- Accepted name:** taxoid 2 α -hydroxylase
- Reaction:** (1) taxusin + [reduced NADPH—hemoprotein reductase] + O₂ = 2 α -hydroxytaxusin + [oxidized NADPH—hemoprotein reductase] + H₂O
(2) 7 β -hydroxytaxusin + [reduced NADPH—hemoprotein reductase] + O₂ = 2 α ,7 β -dihydroxytaxusin + [oxidized NADPH—hemoprotein reductase] + H₂O
- Systematic name:** taxusin, [reduced NADPH—hemoprotein reductase]:oxygen 2-oxidoreductase
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein from the yew tree *Taxus cuspidata*. Does not act on earlier intermediates in taxol biosynthesis.
- References:** [616, 615]

[EC 1.14.14.183 created 2022]

EC 1.14.14.184

- Accepted name:** 5-dehydro-6-demethoxyfumagillol synthase
- Reaction:** (+)-*exo*- β -bergamotene + 2 [reduced NADPH—hemoprotein reductase] + 3 O₂ = 5-dehydro-6-demethoxyfumagillol + 2 [oxidized NADPH—hemoprotein reductase] + 3 H₂O (overall reaction)
(1a) (+)-*exo*- β -bergamotene + [reduced NADPH—hemoprotein reductase] + O₂ = (5*R*)-hydroxy-(+)-*exo*- β -bergamotene + [oxidized NADPH—hemoprotein reductase] + H₂O
(1b) (5*R*)-hydroxy-(+)-*exo*- β -bergamotene + O₂ = (3*S*)-3-[2-methyl-3-(3-methylbut-2-en-1-yl)oxiran-2-yl]-4-methylidenecyclohexan-1-one + H₂O
(1c) (3*S*)-3-[2-methyl-3-(3-methylbut-2-en-1-yl)oxiran-2-yl]-4-methylidenecyclohexan-1-one + [reduced NADPH—hemoprotein reductase] + O₂ = 5-dehydro-6-demethoxyfumagillol + [oxidized NADPH—hemoprotein reductase] + H₂O
- Other name(s):** fumagillin multifunctional cytochrome *P*450 monooxygenase; *Fma-P*450; *fmaG* (gene name)
- Systematic name:** (+)-*exo*- β -bergamotene,[reduced NADPH—hemoprotein reductase] oxidoreductase (5-dehydro-6-demethoxyfumagillol-producing)
- Comments:** The enzyme, characterized from the mold *Aspergillus fumigatus*, catalyses a complex transformation comprising hydroxylation, bicyclic ring-opening, and two epoxidations, generating the sesquiterpenoid core skeleton of fumagillin.
- References:** [2489]

[EC 1.14.14.184 created 2022]

EC 1.14.15 With reduced iron-sulfur protein as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.15.1

- Accepted name:** camphor 5-monooxygenase
- Reaction:** (+)-camphor + reduced putidaredoxin + O₂ = (+)-*exo*-5-hydroxycamphor + oxidized putidaredoxin + H₂O
- Other name(s):** camphor 5-*exo*-methylene hydroxylase; 2-bornanone 5-*exo*-hydroxylase; bornanone 5-*exo*-hydroxylase; camphor 5-*exo*-hydroxylase; camphor 5-*exo*hydroxylase; camphor hydroxylase; *d*-camphor monooxygenase; methylene hydroxylase; methylene monooxygenase; *D*-camphor-*exo*-hydroxylase; camphor methylene hydroxylase
- Systematic name:** (+)-camphor, reduced putidaredoxin:oxygen oxidoreductase (5-hydroxylating)
- Comments:** A heme-thiolate protein (*P*-450). Also acts on (-)-camphor and 1,2-campholide, forming 5-*exo*-hydroxy-1,2-campholide.
- References:** [1600, 4358]

[EC 1.14.15.1 created 1972, modified 1986]

[1.14.15.2 Transferred entry. camphor 1,2-monooxygenase. Now EC 1.14.13.162, 2,5-diketocamphane 1,2-monooxygenase.]

[EC 1.14.15.2 created 1972, deleted 2012]

EC 1.14.15.3

- Accepted name:** alkane 1-monooxygenase
Reaction: octane + 2 reduced rubredoxin + O₂ + 2 H⁺ = 1-octanol + 2 oxidized rubredoxin + H₂O
Other name(s): alkane 1-hydroxylase; ω-hydroxylase; fatty acid ω-hydroxylase; alkane monooxygenase; 1-hydroxylase; alkane hydroxylase
Systematic name: alkane, reduced-rubredoxin:oxygen 1-oxidoreductase
Comments: Some enzymes in this group are heme-thiolate proteins (*P*-450). Also hydroxylates fatty acids in the ω-position.
References: [554, 2752, 3301]

[EC 1.14.15.3 created 1972]

EC 1.14.15.4

- Accepted name:** steroid 11β-monooxygenase
Reaction: a steroid + 2 reduced adrenodoxin + O₂ + 2 H⁺ = an 11β-hydroxysteroid + 2 oxidized adrenodoxin + H₂O
Other name(s): steroid 11β-hydroxylase; steroid 11β/18-hydroxylase
Systematic name: steroid, reduced-adrenodoxin:oxygen oxidoreductase (11β-hydroxylating)
Comments: A heme-thiolate protein (*P*-450). Also hydroxylates steroids at the 18-position, and converts 18-hydroxycorticosterone into aldosterone.
References: [1383, 1578, 4306, 4761, 4940]

[EC 1.14.15.4 created 1961 as EC 1.99.1.7, transferred 1965 to EC 1.14.1.6, transferred 1972 to EC 1.14.15.4, modified 1989, modified 2014]

EC 1.14.15.5

- Accepted name:** corticosterone 18-monooxygenase
Reaction: corticosterone + 2 reduced adrenodoxin + O₂ + 2 H⁺ = 18-hydroxycorticosterone + 2 oxidized adrenodoxin + H₂O
Other name(s): corticosterone 18-hydroxylase; corticosterone methyl oxidase
Systematic name: corticosterone, reduced-adrenodoxin:oxygen oxidoreductase (18-hydroxylating)
References: [3445]

[EC 1.14.15.5 created 1972]

EC 1.14.15.6

- Accepted name:** cholesterol monooxygenase (side-chain-cleaving)
Reaction: cholesterol + 6 reduced adrenodoxin + 3 O₂ + 6 H⁺ = pregnenolone + 4-methylpentanal + 6 oxidized adrenodoxin + 4 H₂O (overall reaction)
(1a) cholesterol + 2 reduced adrenodoxin + O₂ + 2 H⁺ = (22*R*)-22-hydroxycholesterol + 2 oxidized adrenodoxin + H₂O
(1b) (22*R*)-22-hydroxycholesterol + 2 reduced adrenodoxin + O₂ + 2 H⁺ = (20*R*,22*R*)-20,22-dihydroxycholesterol + 2 oxidized adrenodoxin + H₂O
(1c) (20*R*,22*R*)-20,22-dihydroxy-cholesterol + 2 reduced adrenodoxin + O₂ + 2 H⁺ = pregnenolone + 4-methylpentanal + 2 oxidized adrenodoxin + 2 H₂O
Other name(s): cholesterol desmolase; cytochrome *P*-450_{sc}; C₂₇-side chain cleavage enzyme; cholesterol 20-22-desmolase; cholesterol C₂₀₋₂₂ desmolase; cholesterol side-chain cleavage enzyme; cholesterol side-chain-cleaving enzyme; steroid 20-22 desmolase; steroid 20-22-lyase; CYP11A1 (gene name)
Systematic name: cholesterol, reduced-adrenodoxin:oxygen oxidoreductase (side-chain-cleaving)

Comments: A heme-thiolate protein (cytochrome *P*-450). The reaction proceeds in three stages, with two hydroxylations at C-22 and C-20 preceding scission of the side-chain between carbons 20 and 22. The initial source of the electrons is NADPH, which transfers the electrons to the adrenodoxin via EC 1.18.1.6, adrenodoxin-NADP⁺ reductase.

References: [503, 1520, 1518, 4074, 2688]

[EC 1.14.15.6 created 1983, modified 2013, modified 2014]

EC 1.14.15.7

Accepted name: choline monooxygenase

Reaction: choline + O₂ + 2 reduced ferredoxin + 2 H⁺ = betaine aldehyde hydrate + H₂O + 2 oxidized ferredoxin

Systematic name: choline, reduced-ferredoxin:oxygen oxidoreductase

Comments: The spinach enzyme, which is located in the chloroplast, contains a Rieske-type [2Fe-2S] cluster, and probably also a mononuclear Fe centre. Requires Mg²⁺. Catalyses the first step of glycine betaine synthesis. In many bacteria, plants and animals, betaine is synthesized in two steps: (1) choline to betaine aldehyde and (2) betaine aldehyde to betaine. Different enzymes are involved in the first reaction. In plants, the reaction is catalysed by this enzyme whereas in animals and many bacteria it is catalysed by either membrane-bound EC 1.1.99.1 (choline dehydrogenase) or soluble EC 1.1.3.17 (choline oxidase) [4485]. The enzyme involved in the second step, EC 1.2.1.8 (betaine-aldehyde dehydrogenase), appears to be the same in plants, animals and bacteria. In some bacteria, betaine is synthesized from glycine through the actions of EC 2.1.1.156 (glycine/sarcosine *N*-methyltransferase) and EC 2.1.1.157 (sarcosine/dimethylglycine *N*-methyltransferase).

References: [456, 500, 3456, 3606, 3116, 3117, 4485]

[EC 1.14.15.7 created 2001, modified 2002 (EC 1.14.14.4 created 2000, incorporated 2002), modified 2005, modified 2011]

EC 1.14.15.8

Accepted name: steroid 15β-monooxygenase

Reaction: progesterone + 2 reduced [2Fe-2S] ferredoxin + O₂ = 15β-hydroxyprogesterone + 2 oxidized [2Fe-2S] ferredoxin + H₂O

Other name(s): cytochrome *P*-450_{meg}; cytochrome P450_{meg}; steroid 15β-hydroxylase; CYP106A2; BmCYP106A2

Systematic name: progesterone, reduced-ferredoxin:oxygen oxidoreductase (15β-hydroxylating)

Comments: The enzyme from the bacterium *Bacillus megaterium* hydroxylates a variety of 3-oxo-Δ⁴-steroids in position 15β. Ring A-reduced, aromatic, and 3β-hydroxy-Δ⁴-steroids do not serve as substrates [294].

References: [295, 294, 2507, 1359, 2508]

[EC 1.14.15.8 created 2010]

EC 1.14.15.9

Accepted name: spheroidene monooxygenase

Reaction: (1) spheroidene + 4 reduced ferredoxin [iron-sulfur] cluster + 2 O₂ + 4 H⁺ = spheroiden-2-one + 4 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O (overall reaction)

(1a) spheroidene + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2-hydroxyspheroidene + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

(1b) 2-hydroxyspheroidene + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2,2-dihydroxyspheroidene + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

(1c) 2,2-dihydroxyspheroidene = spheroiden-2-one + H₂O (spontaneous)

(2) spirilloxanthin + 4 reduced ferredoxin [iron-sulfur] cluster + 2 O₂ + 4 H⁺ = 2-oxospirilloxanthin + 4 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O (overall reaction)

(2a) spirilloxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2-hydroxyspirilloxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

(2b) 2-hydroxyspirilloxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2,2-dihydroxyspirilloxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
 (2c) 2,2-dihydroxyspirilloxanthin = 2-oxospirilloxanthin + H₂O (spontaneous)
 (3) 2-oxospirilloxanthin + 4 reduced ferredoxin [iron-sulfur] cluster + 2 O₂ + 4 H⁺ = 2,2'-dioxospirilloxanthin + 4 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O (overall reaction)
 (3a) 2-oxospirilloxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2'-hydroxy-2-oxospirilloxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
 (3b) 2'-hydroxy-2-oxospirilloxanthin + reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 2',2'-dihydroxy-2-oxospirilloxanthin + oxidized ferredoxin [iron-sulfur] cluster + H₂O
 (3c) 2',2'-dihydroxy-2-oxospirilloxanthin = 2,2'-dioxospirilloxanthin + H₂O (spontaneous)

Other name(s): CrtA; acyclic carotenoid 2-ketolase; spirilloxanthin monooxygenase; 2-oxo-spirilloxanthin monooxygenase
Systematic name: spheroidene, reduced-ferredoxin:oxygen oxidoreductase (spheroidene-2-one-forming)
Comments: The enzyme is involved in spheroidenone biosynthesis and in 2,2'-dioxospirilloxanthin biosynthesis. The enzyme from *Rhodobacter sphaeroides* contains heme at its active site [2390].
References: [2390, 1304]

[EC 1.14.15.9 created 2012, modified 2016]

EC 1.14.15.10

Accepted name: (+)-camphor 6-*endo*-hydroxylase
Reaction: (+)-camphor + reduced putidaredoxin + O₂ = (+)-6-*endo*-hydroxycamphor + oxidized putidaredoxin + H₂O
Other name(s): P450_{camr}
Systematic name: (+)-camphor, reduced putidaredoxin:oxygen oxidoreductase (6-*endo*-hydroxylating)
Comments: A cytochrome *P*-450 monooxygenase from the bacterium *Rhodococcus* sp. NCIMB 9784.
References: [1418]

[EC 1.14.15.10 created 2012]

EC 1.14.15.11

Accepted name: pentalenic acid synthase
Reaction: 1-deoxypentalenate + reduced ferredoxin + O₂ = pentalenate + oxidized ferredoxin + H₂O
Other name(s): CYP105D7; sav7469 (gene name); 1-deoxypentalenate, reduced ferredoxin:O₂ oxidoreductase
Systematic name: 1-deoxypentalenate, reduced ferredoxin:oxygen oxidoreductase
Comments: A heme-thiolate enzyme (*P*-450). Isolated from the bacterium *Streptomyces avermitilis*. The product, pentalenate, is a co-metabolite from pentalenolactone biosynthesis.
References: [4176]

[EC 1.14.15.11 created 2012]

[1.14.15.12 Transferred entry. pimeloyl-[acyl-carrier protein] synthase. Now EC 1.14.14.46, pimeloyl-[acyl-carrier protein] synthase]

[EC 1.14.15.12 created 2013, deleted 2017]

EC 1.14.15.13

Accepted name: pulcherriminic acid synthase
Reaction: cyclo(L-leucyl-L-leucyl) + 6 reduced ferredoxin + 3 O₂ = pulcherriminic acid + 6 oxidized ferredoxin + 4 H₂O
Other name(s): cyclo-L-leucyl-L-leucyl dipeptide oxidase; CYP134A1; CypX (ambiguous)
Systematic name: cyclo(L-leucyl-L-leucyl), reduced-ferredoxin:oxygen oxidoreductase (*N*-hydroxylating, aromatizing)

Comments: A heme-thiolate (*P*-450) enzyme from the bacterium *Bacillus subtilis*. The order of events during the overall reaction is unknown. Pulcherrimic acid spontaneously forms an iron chelate with Fe(3+) to form the red pigment pulcherrimin [778].

References: [2588, 778]

[EC 1.14.15.13 created 2013]

EC 1.14.15.14

Accepted name: methyl-branched lipid ω -hydroxylase

Reaction: a methyl-branched lipid + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = an ω -hydroxy-methyl-branched lipid + H₂O + 2 oxidized ferredoxin [iron-sulfur] cluster

Other name(s): CYP124

Systematic name: methyl-branched lipid, reduced-ferredoxin:oxygen oxidoreductase (ω -hydroxylating)

Comments: The enzyme, found in pathogenic and nonpathogenic mycobacteria species, actinomycetes, and some proteobacteria, hydroxylates the ω -carbon of a number of methyl-branched lipids, including (*2E,6E*)-farnesol, phytanate, geranylgeraniol, 15-methylpalmitate and (*2E,6E*)-farnesyl diphosphate. It is a *P*-450 heme-thiolate enzyme.

References: [1939]

[EC 1.14.15.14 created 2015]

EC 1.14.15.15

Accepted name: cholestanetriol 26-monooxygenase

Reaction: 5 β -cholestane-3 α ,7 α ,12 α -triol + 6 reduced adrenodoxin + 6 H⁺ + 3 O₂ = (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oate + 6 oxidized adrenodoxin + 4 H₂O (overall reaction)
(1a) 5 β -cholestane-3 α ,7 α ,12 α -triol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = (25*R*)-5 β -cholestan-3 α ,7 α ,12 α ,26-tetraol + 2 oxidized adrenodoxin + H₂O
(1b) (25*R*)-5 β -cholestan-3 α ,7 α ,12 α ,26-tetraol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-al + 2 oxidized adrenodoxin + 2 H₂O
(1c) (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-al + 2 reduced adrenodoxin + 2 H⁺ + O₂ = (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oate + 2 oxidized adrenodoxin + H₂O

Other name(s): 5 β -cholestan-3 α ,7 α ,12 α -triol 26-hydroxylase; 5 β -cholestan-3 α ,7 α ,12 α -triol hydroxylase; cholestanetriol 26-hydroxylase; sterol 27-hydroxylase; sterol 26-hydroxylase; cholesterol 27-hydroxylase; CYP27A; CYP27A1; cytochrome P450 27A1'

Systematic name: 5 β -cholestan-3 α ,7 α ,12 α -triol, adrenodoxin:oxygen oxidoreductase (26-hydroxylating)

Comments: This mitochondrial cytochrome *P*-450 enzyme requires adrenodoxin. It catalyses the first three sterol side chain oxidations in bile acid biosynthesis via the neutral (classic) pathway. Can also act on cholesterol, cholest-5-ene-3 β ,7 α -diol, 7 α -hydroxycholest-4-en-3-one, and 5 β -cholestan-3 α ,7 α -diol. The enzyme can also hydroxylate cholesterol at positions 24 and 25. The initial source of the electrons is NADPH, which transfers the electrons to the adrenodoxin via EC 1.18.1.6, adrenodoxin-NADP⁺ reductase.

References: [2695, 3164, 4624, 96, 800, 1698, 3325, 1234, 3326]

[EC 1.14.15.15 created 1976 as EC 1.14.13.15, modified 2005, modified 2012, transferred 2016 to EC 1.14.15.15]

EC 1.14.15.16

Accepted name: vitamin D₃ 24-hydroxylase

Reaction: (1) calcitriol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = calcitretol + 2 oxidized adrenodoxin + H₂O
(2) calcidiol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = secalciferol + 2 oxidized adrenodoxin + H₂O

Other name(s): CYP24A1

Systematic name: calcitriol, adrenodoxin:oxygen oxidoreductase (24-hydroxylating)

Comments: This mitochondrial cytochrome *P*-450 enzyme requires adrenodoxin. The enzyme can perform up to 6 rounds of hydroxylation of the substrate calcitriol leading to calcitroic acid. The human enzyme also shows 23-hydroxylating activity leading to 1,25 dihydroxyvitamin D₃-26,23-lactone as end product while the mouse and rat enzymes do not. The initial source of the electrons is NADPH, which transfers the electrons to the adrenodoxin via EC 1.18.1.6, adrenodoxin-NADP⁺ reductase.

References: [2693, 1487, 3639, 3388, 2310, 3681, 3387]

[EC 1.14.15.16 created 2011 as EC 1.14.13.126, transferred 2016 to EC 1.14.15.16]

EC 1.14.15.17

Accepted name: pheophorbide *a* oxygenase

Reaction: pheophorbide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = red chlorophyll catabolite + 2 oxidized ferredoxin [iron-sulfur] cluster (overall reaction)
(1a) pheophorbide *a* + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = epoxypheophorbide *a* + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(1b) epoxypheophorbide *a* + H₂O = red chlorophyll catabolite (spontaneous)

Other name(s): pheide *a* monooxygenase; pheide *a* oxygenase; PaO; PAO

Systematic name: pheophorbide-*a*,ferredoxin:oxygen oxidoreductase (biladiene-forming)

Comments: This enzyme catalyses a key reaction in chlorophyll degradation, which occurs during leaf senescence and fruit ripening in higher plants. The enzyme from *Arabidopsis* contains a Rieske-type iron-sulfur cluster [3390] and requires reduced ferredoxin, which is generated either by NADPH through the pentose-phosphate pathway or by the action of photosystem I [3545]. While still attached to this enzyme, the product is rapidly converted into primary fluorescent chlorophyll catabolite by the action of EC 1.3.7.12, red chlorophyll catabolite reductase [3390, 3389]. Pheophorbide *b* acts as an inhibitor. In ¹⁸O₂ labelling experiments, only the aldehyde oxygen is labelled, suggesting that the other oxygen atom may originate from H₂O [1734].

References: [1734, 3390, 692, 3545, 1733, 3389]

[EC 1.14.15.17 created 2007 as EC 1.14.12.20, transferred 2016 to EC 1.14.15.17]

EC 1.14.15.18

Accepted name: calcidiol 1-monooxygenase

Reaction: (1) calcidiol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = calcitriol + 2 oxidized adrenodoxin + H₂O
(2) secalciferol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = calcitriol + 2 oxidized adrenodoxin + H₂O

Other name(s): 25-hydroxycholecalciferol 1-hydroxylase; 25-hydroxycholecalciferol 1-monooxygenase; 1-hydroxylase-25-hydroxyvitamin D₃; 25-hydroxy D₃-1α-hydroxylase; 25-hydroxycholecalciferol 1α-hydroxylase; 25-hydroxyvitamin D₃ 1α-hydroxylase

Systematic name: calcidiol,adrenodoxin:oxygen oxidoreductase (1-hydroxylating)

Comments: A *P*-450 (heme-thiolate) enzyme found in mammals.

References: [1393, 3640, 3682]

[EC 1.14.15.18 created 1976 as EC 1.14.13.13, transferred 2016 to EC 1.14.15.18]

EC 1.14.15.19

Accepted name: C-19 steroid 1α-hydroxylase

Reaction: testosterone + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = 1α-hydroxytestosterone + H₂O + 2 oxidized ferredoxin [iron-sulfur] cluster

Other name(s): CYP260A1

Systematic name: testosterone,reduced-ferredoxin:oxygen oxidoreductase (1α-hydroxylating)

Comments: The enzyme, characterized from the bacterium *Sorangium cellulosum*, is a class I cytochrome *P*-450, and uses ferredoxin as its electron donor [1073]. It was shown to act on several C-19 steroid substrates, including testosterone, androstenedione, testosterone-acetate and 11-oxoandrostenedione [2081].

References: [1073, 2081]

[EC 1.14.15.19 created 2016]

EC 1.14.15.20

Accepted name: heme oxygenase (biliverdin-producing, ferredoxin)
Reaction: protoheme + 6 reduced ferredoxin [iron-sulfur] cluster + 3 O₂ + 6 H⁺ = biliverdin + Fe²⁺ + CO + 6 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O
Other name(s): HO1 (gene name); HY1 (gene name); HO3 (gene name); HO4 (gene name); *pbsA1* (gene name)
Systematic name: protoheme, reduced ferredoxin:oxygen oxidoreductase (α-methene-oxidizing, hydroxylating)
Comments: The enzyme, found in plants, algae, and cyanobacteria, participates in the biosynthesis of phytychromobilin and phytobilins. The terminal oxygen atoms that are incorporated into the carbonyl groups of pyrrole rings A and B of biliverdin are derived from two separate oxygen molecules. The third oxygen molecule provides the oxygen atom that converts the α-carbon to CO. Unlike this enzyme, which uses ferredoxin as its electron donor, the electron source for the related mammalian enzyme (EC 1.14.14.18) is EC 1.6.2.4, NADPH—hemoprotein reductase.
References: [2870, 4101, 816]

[EC 1.14.15.20 created 2016]

EC 1.14.15.21

Accepted name: zeaxanthin epoxidase
Reaction: zeaxanthin + 4 reduced ferredoxin [iron-sulfur] cluster + 4 H⁺ + 2 O₂ = violaxanthin + 4 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O (overall reaction)
(1a) zeaxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = antheraxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(1b) antheraxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = violaxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): Zea-epoxidase
Systematic name: zeaxanthin, reduced ferredoxin:oxygen oxidoreductase
Comments: A flavoprotein (FAD) that is active under conditions of low light. Along with EC 1.23.5.1, violaxanthin de-epoxidase, this enzyme forms part of the xanthophyll (or violaxanthin) cycle, which is involved in protecting the plant against damage by excess light. It will also epoxidize lutein in some higher-plant species.
References: [482, 490, 4268, 1643, 1196, 1195, 2697]

[EC 1.14.15.21 created 2005 as EC 1.14.13.90, transferred 2016 to EC 1.14.15.21]

EC 1.14.15.22

Accepted name: vitamin D 1,25-hydroxylase
Reaction: (1) calcidiol + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = calcitriol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(2) calcidiol + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = calcitriol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): CYP105A1; *Streptomyces griseolus* cytochrome P450SU-1
Systematic name: calcidiol, ferredoxin:oxygen oxidoreductase (1,25-hydroxylating)
Comments: A P-450 (heme-thiolate) enzyme found in the bacterium *Streptomyces griseolus*. cf. EC 1.14.14.24, vitamin D 25-hydroxylase and EC 1.14.15.18, calcidiol 1-monooxygenase.
References: [3683, 4097]

[EC 1.14.15.22 created 2016]

EC 1.14.15.23

- Accepted name:** chloroacetanilide *N*-alkylformylase
Reaction: butachlor + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ = 2-chloro-*N*-(2,6-diethylphenyl)acetamide + butyl formate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *cndA* (gene name)
Systematic name: butachlor,ferredoxin:oxygen oxidoreductase (butyl formate-releasing)
Comments: The enzyme, characterized from the bacterium *Sphingomonas* sp. DC-6, initiates the degradation of several chloroacetanilide herbicides, including alachlor, acetochlor, and butachlor. The enzyme is a Rieske non-heme iron oxygenase, and requires a ferredoxin and EC 1.18.1.3, ferredoxin—NAD⁺ reductase, for activity.
References: [635]

[EC 1.14.15.23 created 2017]

EC 1.14.15.24

- Accepted name:** β-carotene 3-hydroxylase
Reaction: β-carotene + 4 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + 2 O₂ = zeaxanthin + 4 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O (overall reaction)
(1a) β-carotene + 2 reduced ferredoxin [iron-sulfur] cluster + H⁺ + O₂ = β-cryptoxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(1b) β-cryptoxanthin + 2 reduced ferredoxin [iron-sulfur] cluster + H⁺ + O₂ = zeaxanthin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): β-carotene 3,3'-monooxygenase; CrtZ
Systematic name: β-carotene,reduced ferredoxin [iron-sulfur] cluster:oxygen 3-oxidoreductase
Comments: Requires ferredoxin and iron(II). Also acts on other carotenoids with a β-end group. In some species canthaxanthin is the preferred substrate.
References: [4120, 1171, 1172, 406, 2497, 4919, 673]

[EC 1.14.15.24 created 2011 as EC 1.14.13.129, transferred 2017 to EC 1.14.15.24]

EC 1.14.15.25

- Accepted name:** *p*-cymene methyl-monooxygenase
Reaction: *p*-cymene + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = 4-isopropylbenzyl alcohol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *cymAa* (gene name); *cymA* (gene name); *p*-cymene methyl hydroxylase
Systematic name: *p*-cymene,ferredoxin:oxygen oxidoreductase (methyl-hydroxylating)
Comments: The enzyme, characterized from several *Pseudomonas* strains, initiates *p*-cymene catabolism through hydroxylation of the methyl group. The enzyme has a distinct preference for substrates containing at least an alkyl or heteroatom substituent at the *para*-position of toluene. The electrons are provided by a reductase (EC 1.18.1.3, ferredoxin—NAD⁺ reductase) that transfers electrons from NADH via FAD and an [2Fe-2S] cluster. In *Pseudomonas chlororaphis* the presence of a third component of unknown function greatly increases the activity. *cf.* EC 1.14.15.26, toluene methyl-monooxygenase.
References: [1008, 992, 3082, 991]

[EC 1.14.15.25 created 2018]

EC 1.14.15.26

- Accepted name:** toluene methyl-monooxygenase
Reaction: (1) toluene + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = benzyl alcohol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(2) *p*-xylene + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = 4-methylbenzyl alcohol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

(3) *m*-xylene + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = 3-methylbenzyl alcohol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

Other name(s): *xylM* (gene names); *ntnM* (gene names)
Systematic name: methylbenzene,ferredoxin:oxygen oxidoreductase (methyl-hydroxylating)
Comments: The enzyme, characterized from several *Pseudomonas* strains, catalyses the first step in the degradation of toluenes and xylenes. It has a broad substrate specificity and is also active with substituted compounds, such as chlorotoluenes. The electrons are provided by a reductase (EC 1.18.1.3, ferredoxin—NAD⁺ reductase) that transfers electrons from NADH via FAD and an [2Fe-2S] cluster. The enzyme can also act on its products, producing gem-diols that spontaneously dehydrate to form aldehydes.

References: [4141, 3833, 442, 1884]

[EC 1.14.15.26 created 2018]

EC 1.14.15.27

Accepted name: β-dihydromenaquinone-9 ω-hydroxylase
Reaction: β-dihydromenaquinone-9 + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ = ω-hydroxy-β-dihydromenaquinone-9 + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *cyp128* (gene name)
Systematic name: β-dihydromenaquinone-9,reduced ferredoxin:oxygen oxidoreductase (ω-hydroxylating)
Comments: The bacterial cytochrome *P*-450 enzyme is involved in the biosynthesis of ω-sulfo-β-dihydromenaquinone-9 by members of the *Mycobacterium tuberculosis* complex.

References: [1703, 3951]

[EC 1.14.15.27 created 2018]

EC 1.14.15.28

Accepted name: cholest-4-en-3-one 26-monooxygenase [(25*R*)-3-oxocholest-4-en-26-oate forming]
Reaction: cholest-4-en-3-one + 6 reduced [2Fe-2S] ferredoxin + 3 O₂ = (25*R*)-3-oxocholest-4-en-26-oate + 6 oxidized [2Fe-2S] ferredoxin + 4 H₂O (overall reaction)
(1a) cholest-4-en-3-one + 2 reduced [2Fe-2S] ferredoxin + O₂ = (25*R*)-26-hydroxycholest-4-en-3-one + 2 oxidized [2Fe-2S] ferredoxin + H₂O
(1b) (25*R*)-26-hydroxycholest-4-en-3-one + 2 reduced [2Fe-2S] ferredoxin + O₂ = (25*R*)-26-oxocholest-4-en-3-one + 2 oxidized [2Fe-2S] ferredoxin + 2 H₂O
(1c) (25*R*)-26-oxocholest-4-en-3-one + 2 reduced [2Fe-2S] ferredoxin + O₂ = (25*R*)-3-oxocholest-4-en-26-oate + 2 oxidized [2Fe-2S] ferredoxin + H₂O
Other name(s): CYP142
Systematic name: cholest-4-en-3-one,reduced [2Fe-2S] ferredoxin:oxygen oxidoreductase [(25*R*)-3-oxocholest-4-en-26-oate-forming]
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, found in several bacterial pathogens, is involved in degradation of the host cholesterol. It catalyses the hydroxylation of the C-26 carbon, followed by oxidation of the alcohol to the carboxylic acid via the aldehyde intermediate, initiating the degradation of the alkyl side-chain of cholesterol. The products are exclusively in the (25*R*) conformation. The enzyme also accepts cholesterol as a substrate. *cf.* EC 1.14.15.29, cholest-4-en-3-one 26-monooxygenase [(25*S*)-3-oxocholest-4-en-26-oate forming]. The enzyme can receive electrons from ferredoxin reductase *in vitro*, its natural electron donor is not known yet.

References: [967, 1940]

[EC 1.14.15.28 created 2016 as EC 1.14.13.221, transferred 2018 to EC 1.14.15.28]

EC 1.14.15.29

Accepted name: cholest-4-en-3-one 26-monooxygenase [(25*S*)-3-oxocholest-4-en-26-oate forming]

Reaction: cholest-4-en-3-one + 6 reduced ferredoxin [iron-sulfur] cluster + 6 H⁺ + 3 O₂ = (25S)-3-oxocholest-4-en-26-oate + 6 oxidized ferredoxin [iron-sulfur] cluster + 4 H₂O (overall reaction)
(1a) cholest-4-en-3-one + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = (25S)-26-hydroxycholest-4-en-3-one + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(1b) (25S)-26-hydroxycholest-4-en-3-one + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = (25S)-26-oxocholest-4-en-3-one + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
(1c) (25S)-26-oxocholest-4-en-3-one + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = (25S)-3-oxocholest-4-en-26-oate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

Other name(s): CYP125; CYP125A1; cholest-4-en-3-one 27-monoxygenase (misleading); cholest-4-en-3-one,NADH:oxygen oxidoreductase (26-hydroxylating); cholest-4-en-3-one 26-monoxygenase (ambiguous)

Systematic name: cholest-4-en-3-one,[reduced ferredoxin]:oxygen oxidoreductase [(25S)-3-oxocholest-4-en-26-oate-forming]

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in several bacterial pathogens. The enzyme is involved in degradation of the host's cholesterol. It catalyses the hydroxylation of the C-26 carbon, followed by oxidation of the alcohol to the carboxylic acid via the aldehyde intermediate, initiating the degradation of the alkyl side-chain of cholesterol [3214]. The products are exclusively in the (25S) configuration. The enzyme is part of a two-component system that also includes a ferredoxin reductase (most likely KshB, which also interacts with EC 1.14.15.30, 3-ketosteroid 9 α -monoxygenase). The enzyme also accepts cholesterol as a substrate. *cf.* EC 1.14.15.28, cholest-4-en-3-one 27-monoxygenase.

References: [3577, 2753, 551, 3214]

[EC 1.14.15.29 created 2012 as EC 1.14.13.141, modified 2016, transferred 2018 to EC 1.14.15.29]

EC 1.14.15.30

Accepted name: 3-ketosteroid 9 α -monoxygenase

Reaction: androsta-1,4-diene-3,17-dione + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = 9 α -hydroxyandrosta-1,4-diene-3,17-dione + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

Other name(s): KshA; 3-ketosteroid 9 α -hydroxylase

Systematic name: androsta-1,4-diene-3,17-dione,[reduced ferredoxin]:oxygen oxidoreductase (9 α -hydroxylating)

Comments: The enzyme is involved in the cholesterol degradation pathway of several bacterial pathogens, such as *Mycobacterium tuberculosis*. It forms a two-component system with a ferredoxin reductase (KshB). The enzyme contains a Rieske-type iron-sulfur center and non-heme iron. The product of the enzyme is unstable, and spontaneously converts to 3-hydroxy-9,10-seconandrost-1,3,5(10)-triene-9,17-dione.

References: [3307, 550, 549]

[EC 1.14.15.30 created 2012 as EC 1.14.13.142, transferred 2018 to EC 1.14.15.30]

EC 1.14.15.31

Accepted name: 2-hydroxy-5-methyl-1-naphthoate 7-hydroxylase

Reaction: 2-hydroxy-5-methyl-1-naphthoate + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = 2,7-dihydroxy-5-methyl-1-naphthoate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O

Other name(s): NcsB3

Systematic name: 2-hydroxy-5-methyl-1-naphthoate,reduced ferredoxin:oxygen oxidoreductase (7-hydroxylating)

Comments: A cytochrome *P*-450 (heme-thiolate) protein involved in the synthesis of neocarzinostatin in the bacterium *Streptomyces carzinostaticus*.

References: [1505]

[EC 1.14.15.31 created 2014 as EC 1.14.99.49, transferred 2018 to EC 1.14.15.31]

EC 1.14.15.32

Accepted name: pentalenene oxygenase
Reaction: pentalenene + 4 reduced ferredoxin [iron-sulfur] cluster + 4 H⁺ + 2 O₂ = pentalen-13-al + 4 oxidized ferredoxin [iron-sulfur] cluster + 3 H₂O (overall reaction)
(1a) pentalenene + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = pentalen-13-ol + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(1b) pentalen-13-ol + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = pentalen-13-al + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): PtlI
Systematic name: pentalenene, reduced ferredoxin:oxygen 13-oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in the bacterium *Streptomyces avermitilis*. The enzyme is involved in the biosynthesis of pentalenolactone and related antibiotics.
References: [3410]

[EC 1.14.15.32 created 2011 as EC 1.14.13.133, transferred 2018 to EC 1.14.15.32]

EC 1.14.15.33

Accepted name: pikromycin synthase
Reaction: (1) narbomycin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = pikromycin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(2) narbomycin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = neopikromycin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(3) narbomycin + 4 reduced ferredoxin [iron-sulfur] cluster + 4 H⁺ + 2 O₂ = novapikromycin + 4 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
(4) 10-deoxymethymycin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = methymycin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(5) 10-deoxymethymycin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = neomethymycin + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
(6) 10-deoxymethymycin + 4 reduced ferredoxin [iron-sulfur] cluster + 4 H⁺ + 2 O₂ = novamethymycin + 4 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): PikC; CYP107L1
Systematic name: narbomycin, reduced ferredoxin:oxygen oxidoreductase (pikromycin-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Involved in the biosynthesis of a number of bacterial macrolide antibiotics containing a desosamine glycoside unit. With narbomycin it hydroxylates at either C-12 to give pikromycin or C-14 to give neopikromycin or both positions to give narvopikromycin. With 10-deoxymethymycin it hydroxylates at either C-10 to give methymycin or C-12 to give neomethymycin or both positions to give novamethymycin.
References: [4710, 3847, 2455]

[EC 1.14.15.33 created 2014 as EC 1.14.13.185, transferred 2018 to EC 1.14.15.33]

EC 1.14.15.34

Accepted name: 20-oxo-5-*O*-mycaminosyltylactone 23-monooxygenase
Reaction: 20-oxo-5-*O*-β-mycaminosyltylactone + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = 5-*O*-β-mycaminosyltylonolide + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *tylH1* (gene name)
Systematic name: 20-oxo-5-*O*-β-mycaminosyltylactone, reduced ferredoxin:oxygen oxidoreductase (23-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Involved in the biosynthetic pathway of the macrolide antibiotic tylosin, which is produced by several species of *Streptomyces* bacteria.
References: [202, 3483]

[EC 1.14.15.34 created 2014 as EC 1.14.13.186, transferred 2018 to EC 1.14.15.34]

EC 1.14.15.35

Accepted name: 6-deoxyerythronolide B hydroxylase
Reaction: 6-deoxyerythronolide B + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = erythronolide B + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): DEB hydroxylase; *eryF* (gene name); P450(*eryF*); CYP107A1
Systematic name: 6-deoxyerythronolide-B,reduced ferredoxin:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the bacterium *Saccharopolyspora erythraea*. The enzyme is involved in the biosynthesis of the antibiotic erythromycin.
References: [4565, 3814, 788, 2960]

[EC 1.14.15.35 created 2014 as EC 1.14.13.188, transferred 2018 to EC 1.14.15.35]

EC 1.14.15.36

Accepted name: sterol 14 α -demethylase (ferredoxin)
Reaction: a 14 α -methylsteroid + 6 reduced ferredoxin [iron-sulfur] cluster + 6 H⁺ + 3 O₂ = a Δ ¹⁴-steroid + formate + 6 oxidized ferredoxin [iron-sulfur] cluster + 4 H₂O (overall reaction)
 (1a) a 14 α -methylsteroid + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = a 14 α -hydroxymethylsteroid + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
 (1b) a 14 α -hydroxymethylsteroid + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = a 14 α -formylsteroid + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
 (1c) a 14 α -formylsteroid + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = a Δ ¹⁴-steroid + formate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *cyp51* (gene name)
Systematic name: sterol,reduced ferredoxin:oxygen oxidoreductase (14-methyl cleaving)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in several bacterial species. The enzyme, which is involved in sterol biosynthesis, catalyses a hydroxylation and a reduction of the 14 α -methyl group, followed by a second hydroxylation, resulting in the elimination of formate and formation of a 14(15) double bond. The enzyme from *Methylococcus capsulatus* is fused to the ferredoxin by an alanine-rich linker. cf. EC 1.14.14.154, sterol 14 α -demethylase.
References: [1871, 3507, 889]

[EC 1.14.15.36 created 2019]

EC 1.14.15.37

Accepted name: luteothin monooxygenase
Reaction: luteothin + 2 O₂ + 4 reduced ferredoxin [iron-sulfur] cluster + 4 H⁺ = aureothin + 3 H₂O + 4 oxidized ferredoxin [iron-sulfur] cluster (overall reaction)
 (1a) luteothin + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = (7*R*)-7-hydroxyluteothin + H₂O + 2 oxidized ferredoxin [iron-sulfur] cluster
 (1b) (7*R*)-7-hydroxyluteothin + O₂ + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ = aureothin + 2 H₂O + 2 oxidized ferredoxin [iron-sulfur] cluster
Other name(s): *aurH* (gene name)
Systematic name: luteothin,ferredoxin:oxygen oxidoreductase (aureothin-forming)
Comments: The enzyme, characterized from the bacterium *Streptomyces thioluteus*, is a bifunctional cytochrome *P*-450 (heme-thiolate) protein that catalyses both the hydroxylation of its substrate and formation of a furan ring, the final step in the biosynthesis of the antibiotic aureothin. In the bacteria *Streptomyces orinoci* and *Streptomyces spectabilis* an orthologous enzyme catalyses a similar reaction that forms spectinabilin.
References: [1591, 4321]

[EC 1.14.15.37 created 2019]

EC 1.14.15.38

Accepted name: *N,N*-dimethyl phenylurea *N*-demethylase
Reaction: an *N,N*-dimethyl-*N'*-phenylurea compound + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = an *N*-methyl-*N'*-phenylurea compound + formaldehyde + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O
Other name(s): *pdmAB* (gene names)
Systematic name: *N,N*-dimethyl-*N'*-phenylurea compound,NAD(P)H:oxygen oxidoreductase (formaldehyde-forming)
Comments: The enzyme, found in members of the *Sphingobium* genus, initiates the degradation of *N,N*-dimethyl-phenylurea herbicides by mono-*N*-demethylation. The catalytic unit contains a Rieske [2Fe-2S] iron-sulfur cluster, and catalyses the monooxygenation of a methyl group. The resulting *N*-methoxyl group is unstable and decomposes spontaneously to form formaldehyde. The enzyme associates with additional proteins (a reductase and a [3Fe-4S] type ferredoxin) that are involved in the transfer of electrons from NAD(P)H to the active site.
References: [1434]

[EC 1.14.15.38 created 2020]

EC 1.14.15.39

Accepted name: *epi*-isozizaene 5-monooxygenase
Reaction: (+)-*epi*-isozizaene + 4 reduced [2Fe-2S] ferredoxin + 4 H⁺ + 2 O₂ = albaflavenone + 4 oxidized [2Fe-2S] ferredoxin + 3 H₂O (overall reaction)
 (1a) (+)-*epi*-isozizaene + 2 reduced [2Fe-2S] ferredoxin + 2 H⁺ + O₂ = (5*S*)-albaflavenol + 2 oxidized [2Fe-2S] ferredoxin + H₂O
 (1b) (5*S*)-albaflavenol + 2 reduced [2Fe-2S] ferredoxin + 2 H⁺ + O₂ = albaflavenone + 2 oxidized [2Fe-2S] ferredoxin + 2 H₂O
 (2a) (+)-*epi*-isozizaene + 2 reduced [2Fe-2S] ferredoxin + 2 H⁺ + O₂ = (5*R*)-albaflavenol + 2 oxidized [2Fe-2S] ferredoxin + H₂O
 (2b) (5*R*)-albaflavenol + 2 reduced [2Fe-2S] ferredoxin + 2 H⁺ + O₂ = albaflavenone + 2 oxidized [2Fe-2S] ferredoxin + 2 H₂O
Other name(s): CYP170A1
Systematic name: (+)-*epi*-isozizaene,reduced-ferredoxin:oxygen oxidoreductase (5-hydroxylating)
Comments: This cytochrome-*P*-450 enzyme, from the soil-dwelling bacterium *Streptomyces coelicolor* A3(2), catalyses two sequential allylic oxidation reactions. The substrate *epi*-isozizaene, which is formed by the action of EC 4.2.3.37, *epi*-isozizaene synthase, is first oxidized to yield the epimeric intermediates (5*R*)-albaflavenol and (5*S*)-albaflavenol, which can be further oxidized to yield the sesquiterpenoid antibiotic albaflavenone.
References: [4897]

[EC 1.14.15.39 created 2008 as EC 1.14.13.106, transferred 2021 to EC 1.14.15.39]

EC 1.14.16 With reduced pteridine as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.16.1

Accepted name: phenylalanine 4-monooxygenase
Reaction: L-phenylalanine + a 5,6,7,8-tetrahydropteridine + O₂ = L-tyrosine + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): phenylalaninase; phenylalanine 4-hydroxylase; phenylalanine hydroxylase
Systematic name: L-phenylalanine,tetrahydropteridine:oxygen oxidoreductase (4-hydroxylating)

Comments: The active centre contains mononuclear iron(II). The reaction involves an arene oxide that rearranges to give the phenolic hydroxy group. This results in the hydrogen at C-4 migrating to C-3 and in part being retained. This process is known as the NIH-shift. The 4a-hydroxytetrahydropteridine formed can dehydrate to 6,7-dihydropteridine, both spontaneously and by the action of EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase. The 6,7-dihydropteridine must be enzymically reduced back to tetrahydropteridine, by EC 1.5.1.34, 6,7-dihydropteridine reductase, before it slowly rearranges into the more stable but inactive compound 7,8-dihydropteridine.

References: [1451, 2035, 2829, 4362, 563, 91, 1062]

[EC 1.14.16.1 created 1961 as EC 1.99.1.2, transferred 1965 to EC 1.14.3.1, transferred 1972 to EC 1.14.16.1, modified 2002, modified 2003, modified 2019]

EC 1.14.16.2

Accepted name: tyrosine 3-monooxygenase
Reaction: L-tyrosine + a 5,6,7,8-tetrahydropteridine + O₂ = L-dopa + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): L-tyrosine hydroxylase; tyrosine 3-hydroxylase; tyrosine hydroxylase
Systematic name: L-tyrosine,tetrahydropteridine:oxygen oxidoreductase (3-hydroxylating)
Comments: The active centre contains mononuclear iron(II). The enzyme is activated by phosphorylation, catalysed by EC 2.7.11.27, [acetyl-CoA carboxylase] kinase. The 4a-hydroxytetrahydropteridine formed can dehydrate to 6,7-dihydropteridine, both spontaneously and by the action of EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase. The 6,7-dihydropteridine must be enzymically reduced back to tetrahydropteridine, by EC 1.5.1.34, 6,7-dihydropteridine reductase, before it slowly rearranges into the more stable but inactive compound 7,8-dihydropteridine.

References: [2780, 1793, 2963, 3323, 1365]

[EC 1.14.16.2 created 1972, modified 2003, modified 2019]

[1.14.16.3 Deleted entry. anthranilate 3-monooxygenase. Withdrawn owing to insufficient evidence.]

[EC 1.14.16.3 created 1972, deleted 2020]

EC 1.14.16.4

Accepted name: tryptophan 5-monooxygenase
Reaction: L-tryptophan + a 5,6,7,8-tetrahydropteridine + O₂ = 5-hydroxy-L-tryptophan + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): L-tryptophan hydroxylase; indoleacetic acid-5-hydroxylase; tryptophan 5-hydroxylase; tryptophan hydroxylase
Systematic name: L-tryptophan,tetrahydropteridine:oxygen oxidoreductase (5-hydroxylating)
Comments: The active centre contains mononuclear iron(II). The enzyme is activated by phosphorylation, catalysed by a Ca²⁺-activated protein kinase. The 4a-hydroxytetrahydropteridine formed can dehydrate to 6,7-dihydropteridine, both spontaneously and by the action of EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase. The 6,7-dihydropteridine must be enzymically reduced back to tetrahydropteridine, by EC 1.5.1.34, 6,7-dihydropteridine reductase, before it slowly rearranges into the more stable but inactive compound 7,8-dihydropteridine.

References: [1183, 1494, 1789, 1906, 4519]

[EC 1.14.16.4 created 1972, modified 2003, modified 2019]

EC 1.14.16.5

Accepted name: alkylglycerol monooxygenase
Reaction: 1-O-alkyl-*sn*-glycerol + a 5,6,7,8-tetrahydropteridine + O₂ = 1-O-(1-hydroxyalkyl)-*sn*-glycerol + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): glyceryl-ether monooxygenase; glyceryl-ether cleaving enzyme; glyceryl ether oxygenase; glyceryl etherase; O-alkylglycerol monooxygenase

Systematic name: 1-alkyl-*sn*-glycerol,tetrahydrobiopteridine:oxygen oxidoreductase
Comments: The enzyme cleaves alkylglycerols, but does not cleave alkenylglycerols (plasmalogens). Requires non-heme iron [4562], reduced glutathione and phospholipids for full activity. The product spontaneously breaks down to form a fatty aldehyde and glycerol. The co-product, 4a-hydroxytetrahydropteridine, is rapidly dehydrated to 6,7-dihydropteridine, either spontaneously or by EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase.
References: [1827, 3312, 3948, 3967, 4289, 4164, 4562, 4590]

[EC 1.14.16.5 created 1972 as EC 1.14.99.17, transferred 1976 to EC 1.14.16.5, modified 2010, modified 2020]

EC 1.14.16.6

Accepted name: mandelate 4-monooxygenase
Reaction: (*S*)-2-hydroxy-2-phenylacetate + a 5,6,7,8-tetrahydropteridine + O₂ = (*S*)-4-hydroxymandelate + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): L-mandelate 4-hydroxylase; mandelic acid 4-hydroxylase
Systematic name: (*S*)-2-hydroxy-2-phenylacetate,tetrahydropteridine:oxygen oxidoreductase (4-hydroxylating)
Comments: Requires Fe²⁺. The enzyme has been characterized from the bacterium *Pseudomonas putida*. The 4a-hydroxytetrahydropteridine formed can dehydrate to 6,7-dihydropteridine, both spontaneously and by the action of EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase. The 6,7-dihydropteridine must be enzymically reduced back to tetrahydropteridine, by EC 1.5.1.34, 6,7-dihydropteridine reductase, before it slowly rearranges into the more stable but inactive compound 7,8-dihydropteridine.
References: [326]

[EC 1.14.16.6 created 1984, modified 2020]

EC 1.14.16.7

Accepted name: phenylalanine 3-monooxygenase
Reaction: L-phenylalanine + a 5,6,7,8-tetrahydropteridine + O₂ = 3-hydroxy-L-phenylalanine + a 4a-hydroxy-5,6,7,8-tetrahydropteridine
Other name(s): PacX; phenylalanine 3-hydroxylase
Systematic name: L-phenylalanine,tetrahydropteridine:oxygen oxidoreductase (3-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Streptomyces coeruleorubidus*, forms 3-hydroxy-L-phenylalanine (i.e. *m*-L-tyrosine), which is one of the building blocks in the biosynthesis of the uridyl peptide antibiotics pacidamycins. The 4a-hydroxytetrahydropteridine formed can dehydrate to 6,7-dihydropteridine, both spontaneously and by the action of EC 4.2.1.96, 4a-hydroxytetrahydrobiopterin dehydratase. The 6,7-dihydropteridine must be enzymically reduced back to tetrahydropteridine, by EC 1.5.1.34, 6,7-dihydropteridine reductase, before it slowly rearranges into the more stable but inactive compound 7,8-dihydropteridine.
References: [4885]

[EC 1.14.16.7 created 2014, modified 2019]

EC 1.14.17 With reduced ascorbate as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.17.1

Accepted name: dopamine β-monooxygenase
Reaction: dopamine + 2 ascorbate + O₂ = noradrenaline + 2 monodehydroascorbate + H₂O

Other name(s): dopamine β -hydroxylase; MDBH (membrane-associated dopamine β -monooxygenase); SDBH (soluble dopamine β -monooxygenase); dopamine-B-hydroxylase; 3,4-dihydroxyphenethylamine β -oxidase; 4-(2-aminoethyl)pyrocatechol β -oxidase; dopa β -hydroxylase; dopamine β -oxidase; dopamine hydroxylase; phenylamine β -hydroxylase; (3,4-dihydroxyphenethylamine) β -monooxygenase; D β M (gene name)

Systematic name: dopamine,ascorbate:oxygen oxidoreductase (β -hydroxylating)

Comments: A copper protein. The enzyme, found in animals, binds two copper ions with distinct roles during catalysis. Stimulated by fumarate.

References: [2429, 1184, 3923, 1069]

[EC 1.14.17.1 created 1965 as EC 1.14.2.1, transferred 1972 to EC 1.14.17.1, modified 2020]

[1.14.17.2 Deleted entry. 4-coumarate 3-monooxygenase. Now included with EC 1.14.18.1 monophenol monooxygenase]

[EC 1.14.17.2 created 1972, deleted 1984]

EC 1.14.17.3

Accepted name: peptidylglycine monooxygenase

Reaction: [peptide]-glycine + 2 ascorbate + O₂ = [peptide]-(2*S*)-2-hydroxyglycine + 2 monodehydroascorbate + H₂O

Other name(s): peptidylglycine 2-hydroxylase; peptidyl α -amidating enzyme; peptide- α -amide synthetase; peptide α -amidating enzyme; peptide α -amide synthase; peptidylglycine α -hydroxylase; peptidylglycine α -amidating monooxygenase; PAM-A; PAM-B; PAM; peptidylglycine,ascorbate:oxygen oxidoreductase (2-hydroxylating)

Systematic name: [peptide]-glycine,ascorbate:oxygen oxidoreductase (2-hydroxylating)

Comments: A copper protein. The enzyme binds two copper ions with distinct roles during catalysis. Peptidylglycines with a neutral amino acid residue in the penultimate position are the best substrates for the enzyme. The product is unstable and dismutates to glyoxylate and the corresponding desglycine peptide amide, a reaction catalysed by EC 4.3.2.5 peptidylamidoglycolate lyase. In mammals, the two activities are part of a bifunctional protein. Involved in the final step of biosynthesis of α -melanotropin and related biologically active peptides.

References: [418, 1342, 2949, 419, 2948, 2026, 3385, 3384, 689, 619]

[EC 1.14.17.3 created 1989, modified 2019]

EC 1.14.17.4

Accepted name: aminocyclopropanecarboxylate oxidase

Reaction: 1-aminocyclopropane-1-carboxylate + ascorbate + O₂ = ethene + cyanide + dehydroascorbate + CO₂ + 2 H₂O

Other name(s): ACC oxidase; ethylene-forming enzyme; 1-aminocyclopropane-1-carboxylate oxygenase (ethylene-forming)

Systematic name: 1-aminocyclopropane-1-carboxylate oxygenase (ethene-forming)

Comments: A nonheme iron enzyme. Requires CO₂ for activity. In the enzyme from plants, the ethene has signalling functions such as stimulation of fruit-ripening.

References: [4893, 4891, 3330, 608, 4279]

[EC 1.14.17.4 created 2003]

EC 1.14.18 With another compound as one donor, and incorporation of one atom of oxygen into the other donor

EC 1.14.18.1

Accepted name: tyrosinase

Reaction: (1) L-tyrosine + O₂ = dopaquinone + H₂O (overall reaction)

(1a) L-tyrosine + $\frac{1}{2}$ O₂ = L-dopa

(1b) L-dopa + $\frac{1}{2}$ O₂ = dopaquinone + H₂O

(2) 2 L-dopa + O₂ = 2 dopaquinone + 2 H₂O

Other name(s): monophenol monooxygenase; phenolase; monophenol oxidase; cresolase; monophenolase; tyrosine-dopa oxidase; monophenol monooxidase; monophenol dihydroxyphenylalanine:oxygen oxidoreductase; *N*-acetyl-6-hydroxytryptophan oxidase; monophenol, dihydroxy-L-phenylalanine oxygen oxidoreductase; *o*-diphenol:O₂ oxidoreductase; phenol oxidase

Systematic name: L-tyrosine,L-dopa:oxygen oxidoreductase

Comments: A type III copper protein found in a broad variety of bacteria, fungi, plants, insects, crustaceans, and mammals, which is involved in the synthesis of betalains and melanin. The enzyme, which is activated upon binding molecular oxygen, can catalyse both a monophenolase reaction cycle (reaction 1) or a diphenolase reaction cycle (reaction 2). During the monophenolase cycle, one of the bound oxygen atoms is transferred to a monophenol (such as L-tyrosine), generating an *o*-diphenol intermediate, which is subsequently oxidized to an *o*-quinone and released, along with a water molecule. The enzyme remains in an inactive deoxy state, and is restored to the active oxy state by the binding of a new oxygen molecule. During the diphenolase cycle the enzyme binds an external diphenol molecule (such as L-dopa) and oxidizes it to an *o*-quinone that is released along with a water molecule, leaving the enzyme in the intermediate met state. The enzyme then binds a second diphenol molecule and repeats the process, ending in a deoxy state [3561]. The second reaction is identical to that catalysed by the related enzyme catechol oxidase (EC 1.10.3.1). However, the latter can not catalyse the hydroxylation or monooxygenation of monophenols.

References: [842, 3261, 3349, 3533, 3653, 4019, 3561]

[EC 1.14.18.1 created 1972, modified 1976, modified 1980 (EC 1.14.17.2 created 1972, incorporated 1984), modified 2012]

EC 1.14.18.2

Accepted name: CMP-*N*-acetylneuraminate monooxygenase

Reaction: CMP-*N*-acetylneuraminate + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = CMP-*N*-glycoloylneuraminate + 2 ferrocyclochrome *b*₅ + H₂O

Other name(s): CMP-*N*-acetylneuraminic acid hydroxylase; CMP-Neu5Ac hydroxylase; cytidine monophosphoacetylneuraminate monooxygenase; *N*-acetylneuraminic monooxygenase; cytidine-5'-monophosphate-*N*-acetylneuraminic acid hydroxylase

Systematic name: CMP-*N*-acetylneuraminate,ferrocyclochrome-*b*₅:oxygen oxidoreductase (*N*-acetyl-hydroxylating)

Comments: This enzyme contains both a Rieske-type [2Fe-2S] cluster and a second iron site. The ferrocyclochrome *b*₅ produced is reduced by NADH and cytochrome-*b*₅ reductase (EC 1.6.2.2). The enzyme can be activated by Fe²⁺ or Fe³⁺.

References: [3835, 2254, 3731, 2046, 3722]

[EC 1.14.18.2 created 1992 as EC 1.14.13.45, transferred 2003 to EC 1.14.18.2]

EC 1.14.18.3

Accepted name: methane monooxygenase (particulate)

Reaction: methane + quinol + O₂ = methanol + quinone + H₂O

Systematic name: methane,quinol:oxygen oxidoreductase

Comments: Contains copper. It is membrane-bound, in contrast to the soluble methane monooxygenase (EC 1.14.13.25).

References: [3855, 234, 2139, 197]

[EC 1.14.18.3 created 2011]

EC 1.14.18.4

Accepted name: phosphatidylcholine 12-monooxygenase

Reaction: a 1-acyl-2-oleoyl-*sn*-glycero-3-phosphocholine + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a 1-acyl-2-[(12*R*)-12-hydroxyoleoyl]-*sn*-glycero-3-phosphocholine + 2 ferricytochrome *b*₅ + H₂O

Other name(s): ricinoleic acid synthase; oleate Δ¹²-hydroxylase; oleate Δ¹²-monooxygenase

Systematic name: 1-acyl-2-oleoyl-*sn*-glycero-3-phosphocholine,ferrocytochrome-*b*₅:oxygen oxidoreductase (12-hydroxylating)

Comments: The enzyme, characterized from the plant *Ricinus communis* (castor bean), is involved in production of the 12-hydroxylated fatty acid ricinoleate. The enzyme, which shares sequence similarity with fatty-acyl desaturases, requires a cytochrome *b*₅ as the electron donor.

References: [1261, 2882, 3939, 2490, 453]

[EC 1.14.18.4 created 1984 as EC 1.14.13.26, transferred 2015 to EC 1.14.18.4]

EC 1.14.18.5

Accepted name: sphingolipid C4-monooxygenase

Reaction: a dihydroceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (4*R*)-4-hydroxysphinganine ceramide + 2 ferricytochrome *b*₅ + H₂O

Other name(s): sphinganine C4-monooxygenase; sphingolipid C4-hydroxylase; SUR2 (gene name); SBH1 (gene name); SBH₂ (gene name); DEGS2 (gene name)

Systematic name: dihydroceramide,ferrocytochrome *b*₅:oxygen oxidoreductase (C4-hydroxylating)

Comments: The enzyme, which belongs to the family of endoplasmic reticular cytochrome *b*₅-dependent enzymes, is involved in the biosynthesis of sphingolipids in eukaryotes. Some enzymes are bifunctional and also catalyse EC 1.14.19.17, sphingolipid 4-desaturase [4246].

References: [1461, 1412, 3982, 4246, 2851]

[EC 1.14.18.5 created 2012 as EC 1.14.13.169, transferred 2015 to EC 1.14.18.5]

EC 1.14.18.6

Accepted name: 4-hydroxysphinganine ceramide fatty acyl 2-hydroxylase

Reaction: a phytoceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (2'*R*)-2'-hydroxyphytoceramide + 2 ferricytochrome *b*₅ + H₂O

Other name(s): FA2H (gene name); SCS7 (gene name)

Systematic name: (4*R*)-4-hydroxysphinganine ceramide,ferrocytochrome-*b*₅:oxygen oxidoreductase (fatty acyl 2-hydroxylating)

Comments: The enzyme, characterized from yeast and mammals, catalyses the hydroxylation of carbon 2 of long- or very-long-chain fatty acids attached to (4*R*)-4-hydroxysphinganine during *de novo* ceramide synthesis. The enzymes from yeast and from mammals contain an N-terminal cytochrome *b*₅ domain that acts as the direct electron donor to the desaturase active site. The newly introduced 2-hydroxyl group has R-configuration. *cf.* EC 1.14.18.7, dihydroceramide fatty acyl 2-hydroxylase.

References: [2828, 986, 64, 1011, 1449]

[EC 1.14.18.6 created 2015]

EC 1.14.18.7

Accepted name: dihydroceramide fatty acyl 2-hydroxylase

Reaction: a dihydroceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (2'*R*)-2'-hydroxydihydroceramide + 2 ferricytochrome *b*₅ + H₂O

Other name(s): FAH1 (gene name); FAH₂ (gene name); plant sphingolipid fatty acid 2-hydroxylase

Systematic name: dihydroceramide,ferrocytochrome-*b*₅:oxygen oxidoreductase (fatty acyl 2-hydroxylating)

Comments: The enzyme, characterized from plants, catalyses the hydroxylation of carbon 2 of long- or very-long-chain fatty acids attached to sphinganine during *de novo* ceramide synthesis. The enzyme requires an external cytochrome *b*₅ as the electron donor. The newly introduced 2-hydroxyl group has R-configuration. *cf.* EC 1.14.18.6, 4-hydroxysphinganine ceramide fatty acyl 2-hydroxylase.

References: [2957, 2958, 2959]

[EC 1.14.18.7 created 2015]

[1.14.18.8 *Transferred entry. 7 α -hydroxycholest-4-en-3-one 12 α -hydroxylase. Now included with EC 1.14.14.139, 5 β -cholestane-3 α ,7 α -diol 12 α -hydroxylase*]

[EC 1.14.18.8 created 2005 as EC 1.14.13.95, transferred 2015 to EC 1.14.18.8, deleted 2020]

EC 1.14.18.9

- Accepted name:** 4 α -methylsterol monooxygenase
- Reaction:** 4,4-dimethyl-5 α -cholest-7-en-3 β -ol + 6 ferrocyclochrome b_5 + 3 O₂ + 6 H⁺ = 3 β -hydroxy-4 β -methyl-5 α -cholest-7-ene-4 α -carboxylate + 6 ferricytochrome b_5 + 4 H₂O (overall reaction)
(1a) 4,4-dimethyl-5 α -cholest-7-en-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 4 α -hydroxymethyl-4 β -methyl-5 α -cholest-7-en-3 β -ol + 2 ferricytochrome b_5 + H₂O
(1b) 4 α -hydroxymethyl-4 β -methyl-5 α -cholest-7-en-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 3 β -hydroxy-4 β -methyl-5 α -cholest-7-ene-4 α -carbaldehyde + 2 ferricytochrome b_5 + 2 H₂O
(1c) 3 β -hydroxy-4 β -methyl-5 α -cholest-7-ene-4 α -carbaldehyde + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 3 β -hydroxy-4 β -methyl-5 α -cholest-7-ene-4 α -carboxylate + 2 ferricytochrome b_5 + H₂O
- Other name(s):** methylsterol hydroxylase (ambiguous); 4-methylsterol oxidase (ambiguous); 4,4-dimethyl-5 α -cholest-7-en-3 β -ol,hydrogen-donor:oxygen oxidoreductase (hydroxylating) (ambiguous); methylsterol monooxygenase (ambiguous); ERG25 (gene name); MSMO1 (gene name); 4,4-dimethyl-5 α -cholest-7-en-3 β -ol,ferrocyclochrome- b_5 :oxygen oxidoreductase (hydroxylating) (ambiguous)
- Systematic name:** 4,4-dimethyl-5 α -cholest-7-en-3 β -ol,ferrocyclochrome- b_5 :oxygen oxidoreductase (C4 α -methyl-hydroxylating)
- Comments:** This enzyme is found in fungi and animals and catalyses a step in the biosynthesis of important sterol molecules such as ergosterol and cholesterol, respectively. The enzyme acts on the 4 α -methyl group. Subsequent decarboxylation by EC 1.1.1.170, 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating), occurs concomitantly with epimerization of the remaining 4 β -methyl into the 4 α position, thus making it a suitable substrate for a second round of catalysis. *cf.* EC 1.14.13.246, 4 β -methylsterol monooxygenase; EC 1.14.18.10, plant 4,4-dimethylsterol C-4 α -methyl-monooxygenase; and EC 1.14.18.11, plant 4 α -monomethylsterol monooxygenase.
- References:** [2814, 1290, 3831, 420, 1227, 2048]

[EC 1.14.18.9 created 1972 as EC 1.14.99.16, transferred 2002 to EC 1.14.13.72, transferred 2017 to EC 1.14.18.9, modified 2019]

EC 1.14.18.10

- Accepted name:** plant 4,4-dimethylsterol C-4 α -methyl-monooxygenase
- Reaction:** 24-methylidenecycloartanol + 6 ferrocyclochrome b_5 + 3 O₂ + 6 H⁺ = 3 β -hydroxy-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-4 α -carboxylate + 6 ferricytochrome b_5 + 4 H₂O (overall reaction)
(1a) 24-methylidenecycloartanol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 4 α -(hydroxymethyl)-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-3 β -ol + 2 ferricytochrome b_5 + H₂O
(1b) 4 α -(hydroxymethyl)-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 4 α -formyl-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-3 β -ol + 2 ferricytochrome b_5 + 2 H₂O
(1c) 4 α -formyl-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 3 β -hydroxy-4 β ,14 α -dimethyl-9 β ,19-cyclo-5 α -ergost-24(24¹)-en-4 α -carboxylate + 2 ferricytochrome b_5 + H₂O
- Other name(s):** SMO1 (gene name)
- Systematic name:** 24-methylidenecycloartanol,ferrocyclochrome- b_5 :oxygen oxidoreductase (C-4 α -methyl-hydroxylating)

Comments: This plant enzyme catalyses a step in the biosynthesis of sterols. It acts on the 4 α -methyl group of the 4,4-dimethylated intermediate 24-methylidenecycloartanol and catalyses three successive oxidations, turning it into a carboxyl group. The carboxylate is subsequently removed by EC 1.1.1.418, plant 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating), which also catalyses the epimerization of the remaining 4 β -methyl into the 4 α position. Unlike the fungal/animal enzyme EC 1.14.18.9, 4 α -methylsterol monooxygenase, this enzyme is not able to remove the methyl group from C-4-monomethylated substrates. That activity is performed in plants by a second enzyme, EC 1.14.18.11, plant 4 α -monomethylsterol monooxygenase.

References: [3250, 3428, 828, 829]

[EC 1.14.18.10 created 2019]

EC 1.14.18.11

Accepted name: plant 4 α -monomethylsterol monooxygenase

Reaction: 24-methylidenelophenol + 6 ferrocyclochrome b_5 + 3 O₂ + 6 H⁺ = 3 β -hydroxyergosta-7,24(24¹)-dien-4 α -carboxylate + 6 ferricytochrome b_5 + 4 H₂O (overall reaction)

(1a) 24-methylidenelophenol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 4 α -(hydroxymethyl)ergosta-7,24(24¹)-dien-3 β -ol + 2 ferricytochrome b_5 + H₂O

(1b) 4 α -(hydroxymethyl)ergosta-7,24(24¹)-dien-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 4 α -formylergosta-7,24(24¹)-dien-3 β -ol + 2 ferricytochrome b_5 + 2 H₂O

(1c) 4 α -formylergosta-7,24(24¹)-dien-3 β -ol + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 3 β -hydroxyergosta-7,24(24¹)-dien-4 α -carboxylate + 2 ferricytochrome b_5 + H₂O

Other name(s): SMO2 (gene name)

Systematic name: 24-ethylidenelophenol,ferrocyclochrome- b_5 :oxygen oxidoreductase (C-4 α -methyl-hydroxylating)

Comments: This plant enzyme catalyses a step in the biosynthesis of sterols. It acts on the methyl group of the 4 α -methylated intermediates 24-ethylidenelophenol and 24-methylidenelophenol and catalyses three successive oxidations, turning it into a carboxyl group. The carboxylate is subsequently removed by EC 1.1.1.418, plant 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating). Unlike the fungal/animal enzyme EC 1.14.18.9, 4 α -methylsterol monooxygenase, this enzyme is not able to act on 4,4-dimethylated substrates. That activity, which occurs earlier in the pathway, is performed in plants by a second enzyme, EC 1.14.18.10, plant 4,4-dimethylsterol C-4 α -methyl-monooxygenase.

References: [3250, 3428, 828, 829]

[EC 1.14.18.11 created 2019]

EC 1.14.18.12

Accepted name: 2-hydroxy fatty acid dioxygenase

Reaction: a (2*R*)-2-hydroxy C_{*n*}-fatty acid + O₂ = a C_{*n*-1}-fatty acid + H₂O + CO₂

Other name(s): MPO1 (gene name)

Systematic name: 2-hydroxyfatty acid:oxygen oxidoreductase (CO₂,H₂O-forming)

Comments: Requires iron(II). The enzyme, characterized from yeast, is involved in phytosphingosine metabolism. The reaction is mediated by iron(IV) peroxide and results in the release of a water molecule and a carbon dioxide molecule, shortening the substrate by a single carbon atom and forming an odd-numbered fatty acid. Both oxygen atoms of the original carboxylate group are released - one as the leaving water molecule, the other as one of the oxygens of the carbon dioxide molecule. The two oxygen atoms in the newly-formed carboxylate originate from the 2-hydroxy group and from molecular oxygen, respectively. The other oxygen atom of the molecular oxygen is incorporated into the leaving CO₂ molecule. The enzyme from the yeast *Saccharomyces cerevisiae* is active at least toward C₁₄ to C₂₆ 2-hydroxyfatty acids, but not against C₈ 2-hydroxyfatty acid.

References: [2220, 3791]

[EC 1.14.18.12 created 2020]

EC 1.14.19 With oxidation of a pair of donors resulting in the reduction of O₂ to two molecules of water

EC 1.14.19.1

- Accepted name:** stearyl-CoA 9-desaturase
Reaction: stearyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = oleoyl-CoA + 2 ferricyclochrome *b*₅ + 2 H₂O
Other name(s): Δ⁹-desaturase; acyl-CoA desaturase; fatty acid desaturase; stearyl-CoA, hydrogen-donor:oxygen oxidoreductase
Systematic name: stearyl-CoA,ferrocyclochrome-*b*₅:oxygen oxidoreductase (9,10-dehydrogenating)
Comments: An iron protein. The rat liver enzyme is an enzyme system involving cytochrome *b*₅ and EC 1.6.2.2, cytochrome-*b*₅ reductase. The ferricyclochrome *b*₅ produced is reduced by NADH and cytochrome-*b*₅ reductase (EC 1.6.2.2).
References: [1228, 3197, 3198, 4069]

[EC 1.14.19.1 created 1972 as EC 1.14.99.5, modified 1986, modified 2000, transferred 2000 to EC 1.14.19.1, modified 2003]

EC 1.14.19.2

- Accepted name:** stearyl-[acyl-carrier-protein] 9-desaturase
Reaction: stearyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = oleoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): stearyl acyl carrier protein desaturase; stearyl-ACP desaturase; acyl-[acyl-carrier-protein] desaturase; acyl-[acyl-carrier protein],hydrogen-donor:oxygen oxidoreductase
Systematic name: stearyl-[acyl-carrier protein],reduced ferredoxin:oxygen oxidoreductase (9,10 *cis*-dehydrogenating)
Comments: The enzyme is found in the lumen of plastids, where *de novo* biosynthesis of fatty acids occurs, and acts on freshly synthesized saturated fatty acids that are still linked to acyl-carrier protein. The enzyme determines the position of the double bond by its distance from the carboxylic acid end of the fatty acid. It also acts on palmitoyl-[acyl-carrier-protein] [521, 547].
References: [1894, 2955, 3820, 521, 547]

[EC 1.14.19.2 created 1972 as EC 1.14.99.6, modified 2000, transferred 2000 to EC 1.14.19.2, modified 2015]

EC 1.14.19.3

- Accepted name:** acyl-CoA 6-desaturase
Reaction: (1) linoleoyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = γ-linolenoyl-CoA + 2 ferricyclochrome *b*₅ + 2 H₂O
(2) α-linolenoyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = stearidonoyl-CoA + 2 ferricyclochrome *b*₅ + 2 H₂O
Other name(s): Δ⁶-desaturase; Δ⁶-fatty acyl-CoA desaturase; Δ⁶-acyl CoA desaturase; fatty acid Δ⁶-desaturase; fatty acid 6-desaturase; linoleate desaturase; linoleic desaturase; linoleic acid desaturase; linoleoyl CoA desaturase; linoleoyl-coenzyme A desaturase; long-chain fatty acid Δ⁶-desaturase; linoleoyl-CoA,hydrogen-donor:oxygen oxidoreductase; linoleoyl-CoA desaturase; FADS2 (gene name)
Systematic name: acyl-CoA,ferrocyclochrome *b*₅:oxygen oxidoreductase (6,7 *cis*-dehydrogenating)
Comments: An iron protein. The enzyme introduces a *cis* double bond at carbon 6 of acyl-CoAs. It is a front-end desaturase, introducing the new double bond between a pre-existing double bond and the carboxyl-end of the fatty acid. The human enzyme has a broad substrate range. It also acts on palmitoyl-CoA, generating sapienoyl-CoA [1292], and on (9Z,12Z,15Z,18Z,21Z)-tetracos-9,12,15,18,21-pentaenoyl-CoA, converting it to (6Z,9Z,12Z,15Z,18Z,21Z)-tetracos-6,9,12,15,18,21-hexaenoyl-CoA as part of a pathway that produces docosahexaenoate [3990]. The enzyme contains a cytochrome *b*₅ domain that is assumed to act *in vivo* as the electron donor to the active site of the desaturase.
References: [3161, 665, 3990, 1292, 942]

[EC 1.14.19.3 created 1986 as EC 1.14.99.25, transferred 2000 to EC 1.14.19.3, modified 2015]

EC 1.14.19.4

- Accepted name:** acyl-lipid (11-3)-desaturase
- Reaction:** (1) an (11Z,14Z)-icosa-11,14-dienoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = an (8Z,11Z,14Z)-icosa-8,11,14-trienoyl-[glycerolipid] + 2 ferrocytochrome b_5 + 2 H₂O
(2) an (11Z,14Z,17Z)-icosa-11,14,17-trienoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = an (8Z,11Z,14Z,17Z)-icosa-8,11,14,17-tetraenoyl-[glycerolipid] + 2 ferrocytochrome b_5 + 2 H₂O
- Other name(s):** acyl-lipid 8-desaturase; Δ^8 fatty acid desaturase; Δ^8 -desaturase; Δ^8 -fatty-acid desaturase; efd1 (gene name); D8Des (gene name); phytosphinganine,hydrogen donor:oxygen Δ^8 -oxidoreductase (incorrect); SLD
- Systematic name:** acyl-lipid,ferrocytochrome b_5 :oxygen oxidoreductase [(11-3),(11-2)-*cis*-dehydrogenating]
- Comments:** The enzyme, characterized from the protist *Euglena gracilis* [4503] and the microalga *Rebecca salina* [4917], introduces a *cis* double bond at the 8-position in 20-carbon fatty acids that are incorporated into a glycerolipid and have an existing Δ^{11} desaturation. The enzyme is a front-end desaturase, introducing the new double bond between the pre-existing double bond and the carboxyl-end of the fatty acid. It contains a cytochrome b_5 domain that acts as the direct electron donor to the active site of the desaturase, and does not require an external cytochrome. Involved in alternative pathways for the biosynthesis of the polyunsaturated fatty acids arachidonate and icosapentaenoate.
- References:** [4503, 4917]

[EC 1.14.19.4 created 2008, modified 2015]

EC 1.14.19.5

- Accepted name:** acyl-CoA 11-(Z)-desaturase
- Reaction:** an acyl-CoA + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = an (11Z)-enoyl-CoA + 2 ferrocytochrome b_5 + 2 H₂O
- Other name(s):** Δ^{11} desaturase; fatty acid Δ^{11} -desaturase; TpDES_N; Cro-PG; Δ^{11} fatty acid desaturase; Z/E11-desaturase; Δ^{11} -palmitoyl-CoA desaturase; acyl-CoA,hydrogen donor:oxygen Δ^{11} -oxidoreductase; Δ^{11} -fatty-acid desaturase
- Systematic name:** acyl-CoA,ferrocytochrome b_5 :oxygen oxidoreductase (11,12 *cis*-dehydrogenating)
- Comments:** The enzyme introduces a *cis* double bond at position C-11 of saturated fatty acyl-CoAs. In moths the enzyme participates in the biosynthesis of their sex pheromones. The enzyme from the marine microalga *Thalassiosira pseudonana* is specific for palmitoyl-CoA (16:0) [4309], that from the leafroller moth *Choristoneura rosaceana* desaturates myristoyl-CoA (14:0) [1522], while that from the moth *Spodoptera littoralis* accepts both substrates [2673]. The enzyme contains three histidine boxes that are conserved in all desaturases [3547]. It is membrane-bound, and contains a cytochrome b_5 -like domain at the N-terminus that serves as the electron donor for the active site of the desaturase.
- References:** [2673, 3547, 3027, 4309, 1522]

[EC 1.14.19.5 created 2008 (EC 1.14.99.32 created 2000, incorporated 2015), modified 2015]

EC 1.14.19.6

- Accepted name:** acyl-CoA (9+3)-desaturase
- Reaction:** (1) oleoyl-CoA + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = linoleoyl-CoA + 2 ferrocytochrome b_5 + 2 H₂O
(2) palmitoleoyl-CoA + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = (9Z,12Z)-hexadeca-9,12-dienoyl-CoA + 2 ferrocytochrome b_5 + 2 H₂O
- Other name(s):** oleoyl-CoA 12-desaturase; Δ^{12} fatty acid desaturase; $\Delta^{12}(\omega^6)$ -desaturase; oleoyl-CoA Δ^{12} desaturase; Δ^{12} desaturase; Δ^{12} -desaturase; Δ^{12} -fatty-acid desaturase; acyl-CoA,hydrogen donor:oxygen Δ^{12} -oxidoreductase
- Systematic name:** acyl-CoA,ferrocytochrome b_5 :oxygen oxidoreductase (12,13 *cis*-dehydrogenating)

Comments: This microsomal enzyme introduces a *cis* double bond at position 12 of fatty-acyl-CoAs that contain a *cis* double bond at position 9. When acting on 19:1 Δ^{10} fatty acyl-CoA the enzyme from the pathogenic protozoan *Trypanosoma brucei* introduces the new double bond at position 13, indicating that the new double bond is introduced three carbons from the existing *cis* double bond, towards the methyl-end of the fatty acid. Requires cytochrome *b*₅ as the electron donor [3306].

References: [391, 2534, 4298, 3306]

[EC 1.14.19.6 created 2008, modified 2015]

[1.14.19.7 *Transferred entry. (S)-2-hydroxypropylphosphonic acid epoxidase. Now EC 1.11.1.23, (S)-2-hydroxypropylphosphonic acid epoxidase.*]

[EC 1.14.19.7 created 2011, deleted 2014]

EC 1.14.19.8

Accepted name: pentalenolactone synthase
Reaction: pentalenolactone F + O₂ + 2 reduced ferredoxin + 2 H⁺ = pentalenolactone + 2 oxidized ferredoxin + 2 H₂O
Other name(s): *penM* (gene name); *pntM* (gene name)
Systematic name: pentalenolactone-reduced-ferredoxin:oxygen oxidoreductase (pentalenolactone-forming)
Comments: A heme-thiolate protein (*P*-450). Isolated from the bacteria *Streptomyces exfoliatus* and *Streptomyces arenae*.
References: [4920]

[EC 1.14.19.8 created 2012 as EC 1.3.7.10, transferred 2013 to EC 1.14.19.8]

EC 1.14.19.9

Accepted name: tryptophan 7-halogenase
Reaction: tryptophan + FADH₂ + chloride + O₂ + H⁺ = 7-chloro-L-tryptophan + FAD + 2 H₂O
Other name(s): *prnA* (gene name); *rebH* (gene name); *ktzQ* (gene name)
Systematic name: L-tryptophan:FADH₂ oxidoreductase (7-halogenating)
Comments: A flavin-dependent halogenase. The enzyme from the bacterium *Lechevalieria aerocolonigenes* catalyses the initial step in the biosynthesis of rebeccamycin [4786]. It utilizes molecular oxygen to oxidize the FADH₂ cofactor, giving C4a-hydroperoxyflavin, which then reacts with chloride to produce a hypochlorite ion. The latter reacts with an active site lysine to generate a chloramine, which chlorinates the substrate. Also acts on bromide ion. *cf.* EC 1.14.19.58, tryptophan 5-halogenase, and EC 1.14.19.59, tryptophan 6-halogenase.
References: [947, 4786, 339, 1603]

[EC 1.14.19.9 created 2009 as EC 1.14.14.7, transferred 2014 to EC 1.14.19.9, modified 2018]

EC 1.14.19.10

Accepted name: icosanoyl-CoA 5-desaturase
Reaction: icosanoyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = (Z)-icos-5-enoyl-CoA + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): acyl-CoA Δ^5 -desaturase (ambiguous)
Systematic name: icosanoyl-CoA,ferrocyclochrome *b*₅:oxygen oxidoreductase (5,6 *cis*-dehydrogenating)
Comments: The enzyme, characterized from the plant *Limnanthes douglasii* (meadowfoam), is involved in the biosynthesis of (5Z)-icos-5-enoate, an unusual monounsaturated fatty acid that makes up to 60% of the total fatty acids in *Limnanthes* sp. seed oil. The enzyme only acts on saturated fatty acids.
References: [522]

[EC 1.14.19.10 created 2015]

EC 1.14.19.11

- Accepted name:** acyl-[acyl-carrier-protein] 4-desaturase
Reaction: palmitoyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = (4Z)-hexadec-4-enoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): Δ⁴-palmitoyl-[acyl carrier protein] desaturase
Systematic name: palmitoyl-[acyl-carrier protein],reduced acceptor:oxygen oxidoreductase (4,5 *cis*-dehydrogenating)
Comments: The enzymes from the plants *Coriandrum sativum* (coriander) and *Hedera helix* (English ivy) are involved in biosynthesis of petroselinic acid [(6Z)-octadec-6-enoate], which is formed by elongation of (4Z)-hexadec-4-enoate. The ivy enzyme can also act on oleoyl-[acyl-carrier protein] and palmitoleoyl-[acyl-carrier protein], generating the corresponding 4,9-diene.
References: [525, 523, 4613]

[EC 1.14.19.11 created 2015]

EC 1.14.19.12

- Accepted name:** acyl-lipid ω-(9-4) desaturase
Reaction: (1) linoleoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = pinolenoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + 2 H₂O
(2) α-linolenoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = coniferonoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + 2 H₂O
Other name(s): acyl-lipid ω-13 desaturase; acyl-lipid 7-desaturase (ambiguous)
Systematic name: acyl-[glycerolipid],ferrocyclochrome *b*₅:oxygen oxidoreductase [ω(9-4),ω(9-5) *cis*-dehydrogenating]
Comments: The enzyme, characterized from the green alga *Chlamydomonas reinhardtii*, is a front-end desaturase that introduces a *cis* double bond in ω⁹ unsaturated C₁₈ or C₂₀ fatty acids incorporated into lipids, at a position 4 carbon atoms from the existing ω⁹ bond, towards the carboxy end of the fatty acid (at the ω¹³ position). When acting on 20:2Δ(11,14) and 20:3Δ(11,14,17) substrates it introduces the new double bond between carbons 7 and 8. The enzyme contains a cytochrome *b*₅ domain that acts as the direct electron donor for the active site of the desaturase.
References: [1981]

[EC 1.14.19.12 created 2015]

EC 1.14.19.13

- Accepted name:** acyl-CoA 15-desaturase
Reaction: (9Z,12Z)-hexadeca-9,12-dienoyl-CoA + reduced acceptor + O₂ = (9Z,12Z,15Z)-hexadeca-9,12,15-trienoyl-CoA + acceptor + 2 H₂O
Other name(s): DES3 (gene name)
Systematic name: acyl-CoA,reduced acceptor:oxygen oxidoreductase (15,16 *cis*-dehydrogenating)
Comments: The enzyme, characterized from the the plant *Sorghum bicolor*, is involved in the biosynthesis of sorgoleone, an allelopathic compound produced in root hair cells. The enzyme inserts a *cis* double bond at carbon 15. When acting on its natural substrate, (9Z,12Z)-hexadeca-9,12-dienoyl-CoA, it produces a product with a terminal double bond.
References: [3229]

[EC 1.14.19.13 created 2015]

EC 1.14.19.14

- Accepted name:** linoleoyl-lipid Δ⁹ conjugase
Reaction: a linoleoyl-[glycerolipid] + reduced acceptor + O₂ = an (8E,10E,12Z)-octadeca-8,10,12-trienoyl-[glycerolipid] + acceptor + 2 H₂O
Systematic name: linoleoyl-lipid,reduced acceptor:oxygen 8,11-allylic oxidase (8E,10E-forming)

Comments: The enzyme, characterized from the plant *Calendula officinalis*, converts a single *cis* double bond at position 9 of fatty acids incorporated into glycerolipids into two conjugated *trans* double bonds at positions 8 and 10.

References: [3406, 524]

[EC 1.14.19.14 created 2015]

EC 1.14.19.15

Accepted name: (11Z)-hexadec-11-enoyl-CoA conjugase

Reaction: (11Z)-hexadec-11-enoyl-CoA + reduced acceptor + O₂ = (10E,12Z)-hexadeca-10,12-dienoyl-CoA + acceptor + 2 H₂O

Other name(s): Bmpgdesat1 (gene name)

Systematic name: (11Z)-hexadec-11-enoyl-CoA, reduced acceptor:oxygen 10,13-allylic oxidase (10E,12E-forming)

Comments: The enzyme, characterized from the silk moth *Bombyx mori*, catalyses a step in the pathway for the biosynthesis of bombykol, a sex pheromone produced by the moth. The enzyme converts a single *cis* double bond at position 11 of (11Z)-hexadec-11-enoyl-CoA into conjugated 10 *trans* and 12 *cis* double bonds. Prior to catalysing this reaction, the enzyme catalyses the introduction of the *cis* bond in position 11 (*cf.* EC 1.14.19.5, acyl-CoA 11-desaturase).

References: [2909]

[EC 1.14.19.15 created 2015]

EC 1.14.19.16

Accepted name: linoleoyl-lipid Δ^{12} conjugase (11E,13Z-forming)

Reaction: a linoleoyl-[glycerolipid] + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (9Z,11E,13Z)-octadeca-9,11,13-trienoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O

Other name(s): Fac (gene name)

Systematic name: linoleoyl-lipid, ferrocytochrome-*b*₅:oxygen 11,14 allylic oxidase (11E,13Z-forming)

Comments: The enzyme, characterized from the plants *Punica granatum* (pomegranate) and *Trichosanthes kirilowii* (Mongolian snake-gourd), converts a single *cis* double bond at position 12 of linoleate incorporated into phosphatidylcholine into conjugated 11-*trans* and 13-*cis* double bonds. *cf.* EC 1.14.19.33, Δ^{12} acyl-lipid conjugase (11E,13E-forming).

References: [1732, 1856]

[EC 1.14.19.16 created 2015]

EC 1.14.19.17

Accepted name: sphingolipid 4-desaturase

Reaction: a dihydroceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (4E)-sphing-4-ene ceramide + 2 ferricytochrome *b*₅ + 2 H₂O

Other name(s): dehydroceramide desaturase

Systematic name: dihydroceramide, ferrocytochrome *b*₅:oxygen oxidoreductase (4,5-dehydrogenating)

Comments: The enzyme, which has been characterized from plants, fungi, and mammals, generates a *trans* double bond at position 4 of sphinganine bases in sphingolipids [4036]. The preferred substrate is dihydroceramide, but the enzyme is also active with dihydroglucosylceramide [2792]. Unlike EC 1.14.19.29, sphingolipid 8-desaturase, this enzyme does not contain an integral cytochrome *b*₅ domain [4246] and requires an external cytochrome *b*₅ [576]. The product serves as an important signalling molecules in mammals and is required for spermatid differentiation [2789].

References: [4036, 2792, 576, 4246, 2789]

[EC 1.14.19.17 created 2015]

EC 1.14.19.18

- Accepted name:** sphingolipid 8-(*E*)-desaturase
Reaction: a (4*E*)-sphing-4-enine ceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (4*E*,8*E*)-sphing-4,8-dienine ceramide + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): 8-sphingolipid desaturase (ambiguous); 8 fatty acid desaturase (ambiguous); DELTA8-sphingolipid desaturase (ambiguous)
Systematic name: (4*E*)-sphing-4-enine ceramide,ferrocytochrome *b*₅:oxygen oxidoreductase (8,9-*trans* dehydrogenating)
Comments: The enzyme, characterized from the yeasts *Kluyveromyces lactis* and *Candida albicans* [4175] and from the diatom *Thalassiosira pseudonana* [4310], introduces a *trans* double bond at the 8-position of sphingoid bases in sphingolipids. The enzyme determines the position of the double bond by its distance from the alcohol end of the sphingoid base, and contains a cytochrome *b*₅ domain that acts as the direct electron donor to the active site of the desaturase [3216]. The homologous enzymes from higher plants, EC 1.14.19.29, sphingolipid 8-(*E/Z*)-desaturase, act on phytosphinganine (4-hydroxysphinganine) and produces a mixture of *trans* and *cis* isomers.
References: [4175, 4310, 3216]

[EC 1.14.19.18 created 2015]

EC 1.14.19.19

- Accepted name:** sphingolipid 10-desaturase
Reaction: a (4*E*,8*E*)-sphinga-4,8-dienine ceramide + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (4*E*,8*E*,10*E*)-sphinga-4,8,10-trienine ceramide + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): *desA* (gene name)
Systematic name: a (4*E*,8*E*)-sphinga-4,8-dienine ceramide,ferrocytochrome *b*₅:oxygen oxidoreductase (10,11 *trans*-dehydrogenating)
Comments: The enzyme, characterized from the marine diatom *Thalassiosira pseudonana*, produces an *all-trans* product. Similar triunsaturated sphingoid bases are found in some marine invertebrates. The enzyme determines the position of the double bond by its distance from the alcohol end of the sphingoid base, and contains a cytochrome *b*₅ domain that acts as the direct electron donor to the active site of the desaturase.
References: [2788]

[EC 1.14.19.19 created 2015]

EC 1.14.19.20

- Accepted name:** Δ⁷-sterol 5(6)-desaturase
Reaction: a Δ⁷-sterol + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a Δ^{5,7}-sterol + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): lathosterol oxidase; Δ⁷-sterol Δ⁵-dehydrogenase; Δ⁷-sterol 5-desaturase; Δ⁷-sterol-C5(6)-desaturase; 5-DES; SC5DL (gene name); ERG3 (gene name)
Systematic name: Δ⁷-sterol,ferrocytochrome *b*₅:oxygen oxidoreductase 5,6-dehydrogenating
Comments: This enzyme, found in eukaryotic organisms, catalyses the introduction of a double bond between the C₅ and C₆ carbons of the B ring of Δ⁷-sterols, to yield the corresponding Δ^{5,7}-sterols. The enzymes from yeast, plants and vertebrates act on avenasterol, episterol, and lathosterol, respectively. The enzyme is located at the endoplasmic reticulum and is membrane bound.
References: [876, 1708, 138, 4218, 3080, 4217, 3343]

[EC 1.14.19.20 created 1972 as EC 1.3.3.2, transferred 2005 to EC 1.14.21.6, transferred 2015 to EC 1.14.19.20]

EC 1.14.19.21

- Accepted name:** cholesterol 7-desaturase
Reaction: cholesterol + O₂ + NAD(P)H + H⁺ = cholesta-5,7-dien-3β-ol + NAD(P)⁺ + 2 H₂O
Other name(s): *nvd* (gene name); *daf-36* (gene name)

Systematic name: cholesterol,NAD(P)H:oxygen oxidoreductase (7,8 dehydrogenating)
Comments: The enzyme, characterized from several organisms including the worm *Caenorhabditis elegans*, the fly *Drosophila melanogaster*, and the ciliate *Tetrahymena thermophila*, is a Rieske oxygenase. In insects it participates in the biosynthesis of ecdysteroid hormones. The electrons are transferred from NAD(P)H via an electron transfer chain likely to include ferredoxin reductase and ferredoxin. The enzyme differs from regular desaturases, such as EC 1.14.19.20, 7-sterol 5(6)-desaturase, which are cytochrome *b*₅-dependent and contain the three His-boxes that are typical to most desaturases.
References: [4819, 4659, 2972, 228]

[EC 1.14.19.21 created 2015]

EC 1.14.19.22

Accepted name: acyl-lipid ω -6 desaturase (cytochrome *b*₅)
Reaction: an oleoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = a linoleoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): oleate desaturase (ambiguous); linoleate synthase (ambiguous); oleoyl-CoA desaturase (incorrect); oleoylphosphatidylcholine desaturase (ambiguous); phosphatidylcholine desaturase (ambiguous); *n*-6 desaturase (ambiguous); FAD2 (gene name)
Systematic name: 1-acyl-2-oleoyl-*sn*-glycero-3-phosphocholine,ferrocyclochrome-*b*₅:oxygen oxidoreductase (12,13 *cis*-dehydrogenating)
Comments: This microsomal enzyme introduces a *cis* double bond in fatty acids attached to lipid molecules at a location 6 carbons away from the methyl end of the fatty acid. The distance from the carboxylic acid end of the molecule does not affect the location of the new double bond. The most common substrates are oleoyl groups attached to either the *sn*-1 or *sn*-2 position of the glycerol backbone in phosphatidylcholine. *cf.* EC 1.14.19.23, acyl-lipid ω -6 desaturase (ferredoxin).
References: [3393, 3925, 4082, 3937, 2050, 2819]

[EC 1.14.19.22 created 1984 as EC 1.3.1.35, transferred 2015 to EC 1.14.19.22]

EC 1.14.19.23

Accepted name: acyl-lipid (*n*+3)-(*Z*)-desaturase (ferredoxin)
Reaction: an oleoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a linoleoyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): acyl-lipid ω ⁶-desaturase (ferredoxin); oleate desaturase (ambiguous); linoleate synthase (ambiguous); oleoyl-CoA desaturase (ambiguous); oleoylphosphatidylcholine desaturase (ambiguous); phosphatidylcholine desaturase (ambiguous); FAD6 (gene name)
Systematic name: oleoyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (12,13 *cis*-dehydrogenating)
Comments: This plastidial enzyme is able to insert a *cis* double bond in monounsaturated fatty acids incorporated into glycerolipids. The enzyme introduces the new bond at a position 3 carbons away from the existing double bond, towards the methyl end of the fatty acid. The native substrates are oleoyl (18:1 Δ^9) and (*Z*)-hexadec-7-enoyl (16:1 Δ^7) groups attached to either position of the glycerol backbone in glycerolipids, resulting in the introduction of the second double bond at positions 12 and 10, respectively. This prompted the suggestion that this is an ω ⁶ desaturase. However, when acting on palmitoleoyl groups (16:1 Δ^9), the enzyme introduces the second double bond at position 12 (ω ⁴), indicating it is an (*n*+3) desaturase [1675]. *cf.* EC 1.14.19.34, acyl-lipid (9+3)-(*E*)-desaturase.
References: [3727, 3728, 1675, 1079, 3726]

[EC 1.14.19.23 created 2015]

EC 1.14.19.24

Accepted name: acyl-CoA 11-(*E*)-desaturase
Reaction: an acyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = an (11*E*)-enoyl-CoA + 2 ferricytochrome *b*₅ + 2 H₂O

Systematic name: acyl-CoA,ferrocytochrome *b*₅:oxygen oxidoreductase (11,12 *trans*-dehydrogenating)
Comments: Involved in sex pheromone synthesis in the Lepidoptera (moths). The enzyme from the moth *Spodoptera littoralis* prefers 13:0 and 14:0 substrates. The product is formed by the stereospecific removal of the *pro-R* H at C-11 and the *pro-S* H at C-12. *cf.* EC 1.14.19.5, acyl-CoA 11-(*Z*)-desaturase.
References: [1151, 2673, 3027, 3327]

[EC 1.14.19.24 created 2000 as EC 1.14.99.31, transferred 2015 to EC 1.14.19.24]

EC 1.14.19.25

Accepted name: acyl-lipid ω -3 desaturase (cytochrome *b*₅)
Reaction: a linoleoyl-[glycerolipid] + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = an α -linolenoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): FAD3
Systematic name: (9*Z*,12*Z*)-octadeca-9,12-dienoyl-[glycerolipid],ferrocytochrome *b*₅:oxygen oxidoreductase (15,16 *cis*-dehydrogenating)
Comments: This microsomal enzyme introduces a *cis* double bond three carbons away from the methyl end of a fatty acid incorporated into a glycerolipid. The distance from the carboxylic acid end of the molecule does not have an effect. The plant enzyme acts on carbon 15 of linoleoyl groups incorporated into both the *sn*-1 and *sn*-2 positions of the glycerol backbone of phosphatidylcholine and other phospholipids, converting them into α -linolenoyl groups. The enzyme from the fungus *Mortierella alpina* acts on γ -linolenoyl and arachidonoyl groups, converting them into stearidonoyl and icosapentaenoyl groups, respectively [3646]. *cf.* EC 1.14.19.35, *sn*-2 acyl-lipid ω -3 desaturase (ferredoxin).
References: [464, 136, 3646]

[EC 1.14.19.25 created 2015]

EC 1.14.19.26

Accepted name: acyl-[acyl-carrier-protein] 6-desaturase
Reaction: palmitoyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = (6*Z*)-hexadec-6-enoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): DELTA6 palmitoyl-ACP desaturase; DELTA6 16:0-ACP desaturase
Systematic name: palmitoyl-[acyl-carrier protein],reduced ferredoxin:oxygen oxidoreductase (6,7 *cis*-dehydrogenating)
Comments: The enzyme, characterized from the endosperm of the plant *Thunbergia alata* (black-eyed Susan vine), introduces a *cis* double bond at carbon 6 of several saturated acyl-[acp]s. It is most active with palmitoyl-[acp] (16:0), but can also act on myristoyl-[acp] (14:0) and stearoyl-[acp] (18:0). The position of the double bond is determined by its distance from the carboxyl end of the fatty acid.
References: [519, 521]

[EC 1.14.19.26 created 2015]

EC 1.14.19.27

Accepted name: *sn*-2 palmitoyl-lipid 9-desaturase
Reaction: a 1-acyl-2-palmitoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1-acyl-2-palmitoleoyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): DesC2
Systematic name: 1-acyl-2-palmitoyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (9,10 *cis*-dehydrogenating)
Comments: The enzyme, characterized from the cyanobacterium *Nostoc* sp. 36, introduces a *cis* double bond at carbon 9 of palmitoyl groups (16:0) attached to the *sn*-2 position of glycerolipids.
References: [655]

[EC 1.14.19.27 created 2015]

EC 1.14.19.28

- Accepted name:** *sn*-1 stearoyl-lipid 9-desaturase
Reaction: a 1-stearoyl-2-acyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1-oleoyl-2-acyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): *desC* (gene name)
Systematic name: 1-stearoyl-2-acyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (9,10 *cis*-dehydrogenating)
Comments: The enzyme, characterized from cyanobacteria, introduces a *cis* double bond at carbon 9 of stearoyl groups (18:0) attached to the *sn*-1 position of glycerolipids. The enzyme is nonspecific with respect to the polar head group of the glycerolipid.
References: [4482, 1644, 3642]

[EC 1.14.19.28 created 2015]

EC 1.14.19.29

- Accepted name:** sphingolipid 8-(*E/Z*)-desaturase
Reaction: (1) a (4*R*)-4-hydroxysphinganine ceramide + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = a (4*R*,8*E*)-4-hydroxysphing-8-enine ceramide + 2 ferricytochrome *b*₅ + 2 H₂O
(2) a (4*R*)-4-hydroxysphinganine ceramide + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = a (4*R*,8*Z*)-4-hydroxysphing-8-enine ceramide + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): 8-sphingolipid desaturase (ambiguous); 8 fatty acid desaturase (ambiguous); DELTA8-sphingolipid desaturase (ambiguous)
Systematic name: (4*R*)-4-hydroxysphinganine ceramide,ferrocyclochrome *b*₅:oxygen oxidoreductase (8,9 *cis/trans*-dehydrogenating)
Comments: The enzymes from higher plants convert sphinganine, 4*E*-sphing-4-enine and phytosphinganine into *E/Z*-mixtures of Δ⁸-desaturated products displaying different proportions of geometrical isomers depending on plant species. The nature of the actual desaturase substrate has not yet been studied experimentally. The enzymes contain an N-terminal cytochrome *b*₅ domain that acts as the direct electron donor to the active site of the desaturase [3983]. The homologous enzymes from some yeasts and diatoms, EC 1.14.19.18, sphingolipid 8-(*E*)-desaturase, act on sphing-4-enine ceramides and produce only the *trans* isomer.
References: [3983, 3979, 3981, 264, 3612, 634]

[EC 1.14.19.29 created 2015]

EC 1.14.19.30

- Accepted name:** acyl-lipid (8-3)-desaturase
Reaction: (1) an (8*Z*,11*Z*,14*Z*)-icosa-8,11,14-trienoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = a (5*Z*,8*Z*,11*Z*,14*Z*)-icosatetra-5,8,11,14-tetraenoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
(2) an (8*Z*,11*Z*,14*Z*,17*Z*)-icosa-8,11,14,17-tetraenoyl-[glycerolipid] + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = a (5*Z*,8*Z*,11*Z*,14*Z*,17*Z*)-icosa-5,8,11,14,17-pentaenoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
Other name(s): acyl-lipid 5-desaturase; Δ⁵-fatty-acid desaturase; DES5 (gene name); D5des (gene name); FADS1
Systematic name: Δ⁸ acyl-lipid,ferrocyclochrome *b*₅:oxygen oxidoreductase (5,6 *cis*-dehydrogenating)
Comments: The enzyme, which has been characterized from multiple organisms including the moss *Physcomitrella patens*, the marine microalga *Rebecca salina*, and the filamentous fungus *Mortierella alpina*, introduces a *cis* double bond at the 5-position in 20-carbon polyunsaturated fatty acids incorporated in a glycerolipid that contain a Δ⁸ double bond. The enzyme contains a cytochrome *b*₅ domain that acts as the direct electron donor to the active site of the desaturase, and does not require an external cytochrome.
References: [2787, 1971, 4917]

[EC 1.14.19.30 created 2015]

EC 1.14.19.31

- Accepted name:** acyl-lipid (7-3)-desaturase
- Reaction:** (1) a (7Z,10Z,13Z,16Z,19Z)-docosa-7,10,13,16,19-pentaenoyl-[glycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = a (4Z,7Z,10Z,13Z,16Z,19Z)-docosa-4,7,10,13,16,19-hexaenoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
(2) a (7Z,10Z,13Z,16Z)-docosa-7,10,13,16-tetraenoyl-[glycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = a (4Z,7Z,10Z,13Z,16Z)-docosa-4,7,10,13,16-pentaenoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
- Other name(s):** D4Des (gene name); des1 (gene name); CrΔ⁴FAD (gene name); acyl-lipid 4-desaturase
- Systematic name:** Δ⁷ acyl-lipid,ferrocyclochrome b_5 :oxygen oxidoreductase (4,5 *cis*-dehydrogenating)
- Comments:** The enzymes from several algae introduce a *cis* double bond at the 4-position in 22-carbon polyunsaturated fatty acids that contain a Δ⁷ double bond. The enzyme from the fresh water alga *Chlamydomonas reinhardtii* acts on the 16 carbon fatty acid (7Z,10Z,13Z)-hexadeca-7,10,13-trienoate [4862]. The enzyme contains an N-terminal cytochrome b_5 domain that acts as the direct electron donor to the active site of the desaturase, and does not require an external cytochrome.
- References:** [3405, 4308, 2785, 4917, 4862]

[EC 1.14.19.31 created 2015]

EC 1.14.19.32

- Accepted name:** palmitoyl-CoA 14-(*E/Z*)-desaturase
- Reaction:** (1) palmitoyl-CoA + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = (14*E*)-hexadec-14-enoyl-CoA + 2 ferricytochrome b_5 + 2 H₂O
(2) palmitoyl-CoA + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = (14*Z*)-hexadec-14-enoyl-CoA + 2 ferricytochrome b_5 + 2 H₂O
- Systematic name:** palmitoyl-CoA,ferrocyclochrome b_5 :oxygen oxidoreductase (14,15 *cis/trans*-dehydrogenating)
- Comments:** The enzyme, found in the moth *Ostrinia furnacalis* (Asian corn borer), produces a mixture of (*E*)- and (*Z*)- isomers. The products are subsequently truncated by partial β-oxidation to a blend of 12(*E/Z*)-tetradec-12-enoyl-CoA, which are converted to the species-specific sex pheromones (*E*)- and (*Z*)-tetradec-12-enoyl acetates.
- References:** [3552, 4709, 3638]

[EC 1.14.19.32 created 2015]

EC 1.14.19.33

- Accepted name:** Δ¹² acyl-lipid conjugase (11*E*,13*E*-forming)
- Reaction:** (1) a linoleoyl-[glycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = an α-eleostearoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
(2) a γ-linolenoyl-[glycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = an α-parinaroyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
- Other name(s):** fatty acid Δ¹²-conjugase (ambiguous); FADX (gene name)
- Systematic name:** Δ¹² acyl-lipid,ferrocyclochrome- b_5 :oxygen 11,14 allylic oxidase (11*E*,13*E*-forming)
- Comments:** The enzyme, characterized from the plants *Impatiens balsamina*, *Momordica charantia* (bitter melon) and *Vernicia fordii* (tung tree), converts a single *cis* double bond at carbon 12 to two conjugated *trans* bonds at positions 11 and 13. The enzyme from *Vernicia fordii* can also act as a 12(*E*) desaturase when acting on the monounsaturated fatty acids oleate and palmitoleate. cf. EC 1.14.19.16, linoleoyl-lipid Δ¹² conjugase (11*E*,13*Z*-forming).
- References:** [518, 996]

[EC 1.14.19.33 created 2015]

EC 1.14.19.34

- Accepted name:** acyl-lipid (9+3)-(*E*)-desaturase

Reaction: (1) an oleoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O_2 + 2 H^+ = a (9Z,12E)-octadeca-9,12-dienoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H_2O
(2) a palmitoleoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O_2 + 2 H^+ = a (9Z,12E)-hexadeca-9,12-dienoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H_2O

Other name(s): acyl-lipid 12-(E)-desaturase; DsFAD2-1; FADX

Systematic name: Δ^9 acyl-lipid, ferrocytochrome b_5 :oxygen oxidoreductase (12,13 *trans*-dehydrogenating)

Comments: The enzymes from the plants *Dimorphotheca sinuata* (African daisy) and *Vernicia fordii* (tung oil tree) insert a *trans* double bond in position C-12 of oleate and palmitoleate incorporated into glycerolipids. The enzyme introduces the new double bond at a position three carbons away from an existing double bond at position 9, towards the methyl end of the fatty acid. The enzyme from tung oil tree also possesses the activity of EC 1.14.19.33, Δ^{12} acyl-lipid conjugase.

References: [996, 520]

[EC 1.14.19.34 created 2015]

EC 1.14.19.35

Accepted name: *sn*-2 acyl-lipid ω -3 desaturase (ferredoxin)

Reaction: (1) a (7Z,10Z)-hexadeca-7,10-dienoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O_2 + 2 H^+ = a (7Z,10Z,13Z)-hexadeca-7,10,13-trienoyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H_2O
(2) a linoleoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O_2 + 2 H^+ = an α -linolenoyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H_2O

Other name(s): FAD7; FAD8

Systematic name: (7Z,10Z)-hexadeca-7,10-dienoyl-[glycerolipid], ferredoxin:oxygen oxidoreductase (13,14 *cis*-dehydrogenating)

Comments: This plastidial enzyme desaturates 16:2 fatty acids attached to the *sn*-2 position of glycerolipids to 16:3 fatty acids, and converts 18:2 to 18:3 in both the *sn*-1 and *sn*-2 positions. It acts on all 16:2- or 18:2-containing chloroplast membrane lipids, including phosphatidylglycerol, monogalactosyldiacylglycerol, digalactosyldiacylglycerol, and sulfoquinovosyldiacylglycerol. The enzyme introduces a *cis* double bond at a location 3 carbons away from the methyl end of the fatty acid. The distance from the carboxylic acid end of the molecule does not affect the location of the new double bond. *cf.* EC 1.14.19.25, acyl-lipid ω -3 desaturase (cytochrome b_5) and EC 1.14.19.36, *sn*-1 acyl-lipid ω -3 desaturase (ferredoxin).

References: [1783, 2742, 4433]

[EC 1.14.19.35 created 2015]

EC 1.14.19.36

Accepted name: *sn*-1 acyl-lipid ω -3 desaturase (ferredoxin)

Reaction: (1) a 1- γ -linolenoyl-2-acyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O_2 + 2 H^+ = a 1-stearidonoyl-2-acyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H_2O
(2) a 1-linoleoyl-2-acyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O_2 + 2 H^+ = a 1- α -linolenoyl-2-acyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H_2O

Other name(s): *desB* (gene name)

Systematic name: 1- γ -linolenoyl-2-acyl-[glycerolipid], ferredoxin:oxygen oxidoreductase (15,16 *cis*-dehydrogenating)

Comments: The enzyme, characterized from cyanobacteria, introduces a *cis* double bond at carbon 15 of linoleoyl and γ -linolenoyl groups attached to the *sn*-1 position of glycerolipids. The enzyme is an ω desaturase, and determines the location of the double bond by counting three carbons from the methyl end of the fatty acid. It is nonspecific with respect to the polar head group of the glycerolipid. *cf.* EC 1.14.19.35, *sn*-2 acyl-lipid ω -3 desaturase (ferredoxin).

References: [3641]

[EC 1.14.19.36 created 2015]

EC 1.14.19.37

- Accepted name:** acyl-CoA 5-desaturase
- Reaction:** (1) (11Z,14Z)-icosa-11,14-dienoyl-CoA + reduced acceptor + O₂ = (5Z,11Z,14Z)-icosa-5,11,14-trienoyl-CoA + acceptor + 2 H₂O
(2) (11Z,14Z,17Z)-icosa-11,14,17-trienoyl-CoA + reduced acceptor + O₂ = (5Z,11Z,14Z,17Z)-icosa-5,11,14,17-tetraenoyl-CoA + acceptor + 2 H₂O
- Other name(s):** acyl-CoA 5-desaturase (non-methylene-interrupted)
- Systematic name:** acyl-CoA,acceptor:oxygen oxidoreductase (5,6 *cis*-dehydrogenating)
- Comments:** The enzyme, characterized from the plant *Anemone leveillei*, introduces a *cis* double bond at carbon 5 of acyl-CoAs that do not contain a double bond at position 8. *In vivo* it forms non-methylene-interrupted polyunsaturated fatty acids such as sciadonate and juniperonate. When expressed in *Arabidopsis thaliana* the enzyme could also act on unsaturated substrates such as palmitoyl-CoA. *cf.* EC 1.14.19.44, acyl-CoA (8-3)-desaturase.
- References:** [3688]

[EC 1.14.19.37 created 2015]

EC 1.14.19.38

- Accepted name:** acyl-lipid Δ^6 -acetylenase
- Reaction:** (1) a γ -linolenoyl-[glycerolipid] + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (9Z,12Z)-octadeca-9,12-dien-6-ynoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
(2) a stearidonoyl-[glycerolipid] + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = a (9Z,12Z,15Z)-octadeca-9,12,15-trien-6-ynoyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
- Systematic name:** Δ^6 acyl-lipid,ferrocytochrome-*b*₅:oxygen oxidoreductase (6,7-dehydrogenating)
- Comments:** The enzyme, characterized from the moss *Ceratodon purpureus*, converts the double bond at position 6 of γ -linolenate and stearidonate into a triple bond. The product of the latter, dicranin, is the main fatty acid found in *C. purpureus*. The enzyme contains a cytochrome *b*₅ domain that acts as the direct electron donor to the desaturase active site. The enzyme also has the activity of EC 1.14.19.47, acyl-lipid (9-3)-desaturase.
- References:** [3980]

[EC 1.14.19.38 created 2015]

EC 1.14.19.39

- Accepted name:** acyl-lipid Δ^{12} -acetylenase
- Reaction:** linoleoyl-[glycerolipid] + 2 ferrocytochrome *b*₅ + O₂ + 2 H⁺ = crepenynyl-[glycerolipid] + 2 ferricytochrome *b*₅ + 2 H₂O
- Systematic name:** Δ^{12} acyl-lipid,ferrocytochrome-*b*₅:oxygen oxidoreductase (12,13-dehydrogenating)
- Comments:** The enzyme, characterized from the plant *Crepis alpina*, converts the double bond at position 12 of linoleate into a triple bond. The product is the main fatty acid found in triacylglycerols in the seed oil of *Crepis alpina*.
- References:** [206, 2388, 3003]

[EC 1.14.19.39 created 2000 as EC 1.14.99.33, transferred 2015 to EC 1.14.19.39]

EC 1.14.19.40

- Accepted name:** hex-5-enoyl-[acyl-carrier protein] acetylenase
- Reaction:** hex-5-enoyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = hex-5-ynoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
- Other name(s):** *jamB* (gene name)
- Systematic name:** hex-5-enoyl-[acyl-carrier protein],reduced ferredoxin:oxygen oxidoreductase (5,6-dehydrogenating)

Comments: The enzyme, characterized from the marine cyanobacterium *Moorea producens*, is involved in production of the ion channel blocker jamaicamide A. It is specific for hexanoate or hex-5-enoate loaded onto a dedicated acyl-carrier protein (JamC), which is encoded by a gene in the same operon.

References: [4926]

[EC 1.14.19.40 created 2015]

EC 1.14.19.41

Accepted name: sterol 22-desaturase

Reaction: ergosta-5,7,24(28)-trien-3 β -ol + NADPH + H⁺ + O₂ = ergosta-5,7,22,24(28)-tetraen-3 β -ol + NADP⁺ + 2 H₂O

Other name(s): ERG5 (gene name); CYP710A (gene name)

Systematic name: ergosta-5,7,24(28)-trien-3 β -ol,NADPH:oxygen oxidoreductase (22,23-dehydrogenating)

Comments: A heme-thiolate protein (*P*-450). The enzyme, found in yeast and plants, catalyses the introduction of a double bond between the C-22 and C-23 carbons of certain sterols. In yeast the enzyme acts on ergosta-5,7,24(28)-trien-3 β -ol, a step in the biosynthesis of ergosterol. The enzyme from the plant *Arabidopsis thaliana* acts on sitosterol and 24-*epi*-campesterol, producing stigmasterol and brassicasterol, respectively.

References: [2061, 3922, 2890]

[EC 1.14.19.41 created 2015]

EC 1.14.19.42

Accepted name: palmitoyl-[glycerolipid] 7-desaturase

Reaction: a 1-acyl-2-palmitoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1-acyl-2-[(7*Z*)-hexadec-7-enoyl]-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O

Other name(s): FAD5

Systematic name: 1-acyl-2-palmitoyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (7,8-*cis*-dehydrogenating)

Comments: The enzyme introduces a *cis* double bond at carbon 7 of a palmitoyl group attached to the *sn*-2 position of glycerolipids. The enzyme from the plant *Arabidopsis thaliana* is specific for palmitate in monogalactosyldiacylglycerol.

References: [2294, 1610]

[EC 1.14.19.42 created 2015]

EC 1.14.19.43

Accepted name: palmitoyl-[glycerolipid] 3-(*E*)-desaturase

Reaction: a 1-acyl-2-palmitoyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1-acyl-2-[(3*E*)-hexadec-3-enoyl]-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O

Other name(s): FAD4

Systematic name: 1-acyl-2-palmitoyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (3,4-*trans* -dehydrogenating)

Comments: The enzyme introduces an unusual *trans* double bond at carbon 3 of a palmitoyl group attached to the *sn*-2 position of glycerolipids. The enzyme from the plant *Arabidopsis thaliana* is specific for palmitate in phosphatidylglycerol. The enzyme from tobacco can also accept oleate and α -linolenate if present at the *sn*-2 position of phosphatidylglycerol [1191].

References: [1191, 1270]

[EC 1.14.19.43 created 2015]

EC 1.14.19.44

Accepted name: acyl-CoA (8-3)-desaturase

Reaction: (1) (8*Z*,11*Z*,14*Z*)-icosa-8,11,14-trienoyl-CoA + 2 ferrocyclochrome *b*₅ + O₂ + 2 H⁺ = arachidonoyl-CoA + 2 ferricytochrome *b*₅ + 2 H₂O

(2) (8Z,11Z,14Z,17Z)-icosa-8,11,14,17-tetraenoyl-CoA + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = (5Z,8Z,11Z,14Z,17Z)-icosa-5,8,11,14,17-pentaenoyl-CoA + 2 ferricytochrome b_5 + 2 H₂O

- Other name(s):** FADS1 (gene name); acyl-CoA 5-desaturase (methylene-interrupted)
Systematic name: Δ^8 -acyl-CoA,ferrocytochrome b_5 :oxygen oxidoreductase (5,6-*cis*-dehydrogenating)
Comments: The enzyme introduces a *cis* double bond at carbon 5 of acyl-CoAs that contain a double bond at position 8. The enzymes from algae, mosses, mammals and the protozoan *Leishmania major* catalyse the desaturation of dihomo- γ -linoleate [(8Z,11Z,14Z)-icosa-8,11,14-trienoate] and (8Z,11Z,14Z,17Z)-icosa-8,11,14,17-tetraenoate to generate arachidonate and (5Z,8Z,11Z,14Z,17Z)-icosa-5,8,11,14,17-pentaenoate, respectively. The enzyme contains a cytochrome b_5 domain that acts as the direct electron donor to the desaturase active site and does not require an external cytochrome. *cf.* EC 1.14.19.37, acyl-CoA 5-desaturase.
References: [664, 2418, 4331, 4223]

[EC 1.14.19.44 created 2015]

EC 1.14.19.45

- Accepted name:** *sn*-1 oleoyl-lipid 12-desaturase
Reaction: a 1-oleoyl-2-acyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1-linoleoyl-2-acyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): *desA* (gene name)
Systematic name: 1-oleoyl-2-acyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (12,13-*cis*-dehydrogenating)
Comments: The enzyme, characterized from cyanobacteria, introduces a *cis* double bond at carbon 12 of oleoyl groups (18:1) attached to the *sn*-1 position of glycerolipids. The enzyme is a methyl-end desaturase, introducing the new double bond between a pre-existing double bond and the methyl-end of the fatty acid. It is nonspecific with respect to the polar head group of the glycerolipid.
References: [4481, 1644, 85]

[EC 1.14.19.45 created 2015]

EC 1.14.19.46

- Accepted name:** *sn*-1 linoleoyl-lipid 6-desaturase
Reaction: a 1-linoleoyl-2-acyl-[glycerolipid] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = a 1- γ -linolenoyl-2-acyl-[glycerolipid] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): *desD* (gene name)
Systematic name: 1-linoleoyl-2-acyl-[glycerolipid],ferredoxin:oxygen oxidoreductase (6,7-*cis*-dehydrogenating)
Comments: The enzyme, characterized from cyanobacteria, introduces a *cis* double bond at carbon 6 of linoleoyl groups (18:2) attached to the *sn*-1 position of glycerolipids. The enzyme is a front-end desaturase, introducing the new double bond between a pre-existing double bond and the carboxyl-end of the fatty acid. It is nonspecific with respect to the polar head group of the glycerolipid.
References: [1644, 3471, 2299]

[EC 1.14.19.46 created 2015]

EC 1.14.19.47

- Accepted name:** acyl-lipid (9-3)-desaturase
Reaction: (1) an α -linolenoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = a stearidonoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
(2) a linoleoyl-[glycerolipid] + 2 ferrocytochrome b_5 + O₂ + 2 H⁺ = a γ -linolenoyl-[glycerolipid] + 2 ferricytochrome b_5 + 2 H₂O
Other name(s): DES6 (gene name); acyl-lipid 6-desaturase; acyl-lipid Δ^6 -desaturase; Δ^6 -desaturase (ambiguous)
Systematic name: Δ^9 acyl-[glycerolipid],ferrocytochrome b_5 :oxygen oxidoreductase (6,7-*cis*-dehydrogenating)

Comments: The enzyme, characterized from the moss *Physcomitrella patens* and the plant *Borago officinalis* (borage), introduces a *cis* double bond at carbon 6 of several acyl-lipids that contain an existing Δ^9 *cis* double bond. The enzyme contains a cytochrome *b*₅ domain that acts as the electron donor for the active site of the desaturase.

References: [3689, 1329]

[EC 1.14.19.47 created 2015]

EC 1.14.19.48

Accepted name: *tert*-amyl alcohol desaturase

Reaction: *tert*-amyl alcohol + NADPH + H⁺ + O₂ = isoprenyl alcohol + NADP⁺ + 2 H₂O

Other name(s): *mdpJK* (gene names)

Systematic name: *tert*-amyl alcohol,NADPH:oxygen oxidoreductase (1,2-dehydrogenating)

Comments: The enzyme, characterized from the bacterium *Aquicola tertiaricarbonis*, is a Rieske nonheme mononuclear iron oxygenase. It can also act, with lower efficiency, on butan-2-ol, converting it to but-1-en-3-ol. Depending on the substrate, the enzyme also catalyses EC 1.14.13.229, *tert*-butanol monooxygenase.

References: [3700, 3760]

[EC 1.14.19.48 created 2016]

EC 1.14.19.49

Accepted name: tetracycline 7-halogenase

Reaction: tetracycline + FADH₂ + chloride + O₂ + H⁺ = 7-chlorotetracycline + FAD + 2 H₂O

Other name(s): *ctcP* (gene name)

Systematic name: tetracycline:FADH₂ oxidoreductase (7-halogenating)

Comments: The enzyme, characterized from the bacterium *Streptomyces aureofaciens*, is a member of the flavin-dependent halogenase family. The enzyme forms a lysine chloramine intermediate on an internal lysine residue before transferring the chlorine to the substrate. It is stereo-selective for the 4*S* (natural) isomer of tetracycline. FADH₂ is provided by a dedicated EC 1.5.1.36, flavin reductase (NADH).

References: [813, 4924]

[EC 1.14.19.49 created 2016]

EC 1.14.19.50

Accepted name: noroxomaritidine synthase

Reaction: (1) 4'-*O*-methylnorbelladine + [reduced NADPH—hemoprotein reductase] + O₂ = (4*aR*,10*bS*)-noroxomaritidine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

(2) 4'-*O*-methylnorbelladine + [reduced NADPH—hemoprotein reductase] + O₂ = (4*aS*,10*bR*)-noroxomaritidine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): CYP96T1 (gene name)

Systematic name: 4'-*O*-methylnorbelladine,NADPH—hemoprotein reductase:oxygen oxidoreductase (noroxomaritidine-forming)

Comments: A P-450 (heme-thiolate) enzyme. The enzyme, characterized from *Narcissus pseudonarcissus* (daffodil), forms the two enantiomers of the *Amaryllidacea* alkaloid noroxomaritidine by catalysing intramolecular oxidative *para-para'* phenol coupling. The oxidation involves molecular oxygen without its incorporation into the product.

References: [2088]

[EC 1.14.19.50 created 2016]

EC 1.14.19.51

Accepted name: (*S*)-corytuberine synthase

Reaction: (S)-reticuline + [reduced NADPH—hemoprotein reductase] + O₂ = (S)-corytuberine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O.
Other name(s): CYP80G2
Systematic name: (S)-reticuline,NADPH:oxygen oxidoreductase (C-C phenol-coupling; (S)-corytuberine-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme is involved in the biosynthesis of the quaternary benzylisoquinoline alkaloid magnoflorine in the plant *Coptis japonica*. It is specific for (S)-reticuline.
References: [1799]

[EC 1.14.19.51 created 2017]

EC 1.14.19.52

Accepted name: camalexin synthase
Reaction: 2-(L-cystein-*S*-yl)-2-(1*H*-indol-3-yl)acetonitrile + 2 [reduced NADPH—hemoprotein reductase] + 2 O₂ = camalexin + hydrogen cyanide + CO₂ + 2 [oxidized NADPH—hemoprotein reductase] + 4 H₂O (overall reaction)
(1a) 2-(L-cystein-*S*-yl)-2-(1*H*-indol-3-yl)acetonitrile + [reduced NADPH—hemoprotein reductase] + O₂ = (*R*)-dihydrocamalexate + hydrogen cyanide + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(1b) (*R*)-dihydrocamalexate + [reduced NADPH—hemoprotein reductase] + O₂ = camalexin + CO₂ + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): CYP71B15 (gene name); bifunctional dihydrocamalexate synthase/camalexin synthase
Systematic name: 2-(cystein-*S*-yl)-2-(1*H*-indol-3-yl)-acetonitrile, [reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (camalexin-forming)
Comments: This cytochrome *P*-450 (heme thiolate) enzyme, which has been characterized from the plant *Arabidopsis thaliana*, catalyses the last two steps in the biosynthesis of camalexin, the main phytoalexin in that plant. The enzyme catalyses two successive oxidation events. During the first oxidation the enzyme introduces a C-N double bond, liberating hydrogen cyanide, and during the second oxidation it catalyses a decarboxylation.
References: [3749, 403]

[EC 1.14.19.52 created 2017]

EC 1.14.19.53

Accepted name: *all-trans*-retinol 3,4-desaturase
Reaction: *all-trans*-retinol + 2 reduced adrenodoxin + 2 H⁺ + O₂ = *all-trans*-3,4-didehydroretinol + 2 oxidized adrenodoxin + 2 H₂O
Other name(s): CYP27C1 (gene name)
Systematic name: *all-trans*-retinol,reduced adrenodoxin:oxygen 3,4-oxidoreductase
Comments: A cytochrome *P*-450 (heme thiolate) enzyme found in vertebrates. The enzyme is also active with retinal and retinoic acid.
References: [1053, 2258]

[EC 1.14.19.53 created 2018]

EC 1.14.19.54

Accepted name: 1,2-dehydroreticuline synthase
Reaction: (S)-reticuline + [reduced NADPH—hemoprotein reductase] + O₂ = 1,2-dehydroreticuline + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): STORR; CYP82Y2 (gene name); DRS (gene name)
Systematic name: (S)-reticuline,[reduced NADPH—hemoprotein reductase]:oxygen 1,2-oxidoreductase

Comments: A P-450 (heme-thiolate) cytochrome. The enzyme from *Papaver rhoeas* (field poppy) is specific for (*S*)-reticuline and does not act on the (*R*)-form. The enzyme from *Papaver somniferum* (opium poppy), which is involved in the biosynthesis of morphine and related alkaloids, forms a fusion protein with EC 1.5.1.27, 1,2-dehydroreticulium reductase (NADPH), which catalyses the reduction of 1,2-dehydroreticuline to (*R*)-reticuline, thus forming an epimerase system that converts (*S*)-reticuline to (*R*)-reticuline.

References: [1669, 4641, 1094]

[EC 1.14.19.54 created 2018]

EC 1.14.19.55

Accepted name: 4-hydroxybenzoate brominase (decarboxylating)

Reaction: (1) 4-hydroxybenzoate + 2 NADPH + 2 bromide + 2 O₂ + 2 H⁺ = 2,4-dibromophenol + 2 NADP⁺ + CO₂ + 4 H₂O (overall reaction)

(1a) 4-hydroxybenzoate + NADPH + bromide + O₂ + H⁺ = 3-bromo-4-hydroxybenzoate + NADP⁺ + 2 H₂O

(1b) 3-bromo-4-hydroxybenzoate + NADPH + bromide + O₂ + H⁺ = 2,4-dibromophenol + NADP⁺ + CO₂ + 2 H₂O

(2) 3,4-dihydroxybenzoate + 2 NADPH + 2 bromide + 2 O₂ + 2 H⁺ = 3,5-dibromobenzene-1,2-diol + 2 NADP⁺ + CO₂ + 4 H₂O (overall reaction)

(2a) 3,4-dihydroxybenzoate + NADPH + bromide + O₂ + H⁺ = 3-bromo-4,5-dihydroxybenzoate + NADP⁺ + 2 H₂O

(2b) 3-bromo-4,5-dihydroxybenzoate + NADPH + bromide + O₂ + H⁺ = 3,5-dibromobenzene-1,2-diol + NADP⁺ + CO₂ + 2 H₂O

Other name(s): bmp5 (gene name)

Systematic name: 4-hydroxybenzoate:NADPH oxidoreductase (brominating, decarboxylating)

Comments: Contains FAD. The enzyme, described from epiphytic marine bacteria of the genera *Pseudoalteromonas* and *Marinomonas*, is an unusual single-component FAD-dependent halogenase that contains a distinct NAD(P)H binding domain and does not require an additional flavin reductase for activity. The enzyme catalyses a bromination of its substrate, followed by a second bromination concurrent with decarboxylation.

References: [33, 34]

[EC 1.14.19.55 created 2018]

EC 1.14.19.56

Accepted name: 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein] chlorinase

Reaction: 1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + 2 FADH₂ + 2 chloride + 2 O₂ = 4,5-dichloro-1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + 2 FAD + 4 H₂O (overall reaction)

(1a) 1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + FADH₂ + chloride + O₂ = 5-chloro-1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + FAD + 2 H₂O

(1b) 5-chloro-1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + FADH₂ + chloride + O₂ = 4,5-dichloro-1*H*-pyrrole-2-carbonyl-[PltL peptidyl-carrier protein] + FAD + H₂O

Other name(s): *pltA* (gene name)

Systematic name: 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein]:FADH₂ oxidoreductase (chlorinating)

Comments: The enzyme, characterized from the bacterium *Pseudomonas protegens* Pf-5, is a flavin-dependent chlorinase that participates in the biosynthesis of the antibacterial and antifungal compound pyoluteorin.

References: [3112, 953, 3231]

[EC 1.14.19.56 created 2018]

EC 1.14.19.57

Accepted name: 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein] brominase
Reaction: 1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + 3 FADH₂ + 3 bromide + 3 O₂ = 3,4,5-tribromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + 3 FAD + 6 H₂O (overall reaction)
(1a) 1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FADH₂ + bromide + O₂ = 5-bromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FAD + 2 H₂O
(1b) 5-bromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FADH₂ + bromide + O₂ = 4,5-dibromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FAD + 2 H₂O
(1c) 4,5-dibromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FADH₂ + bromide + O₂ = 3,4,5-tribromo-1*H*-pyrrole-2-carbonyl-[Bmp1 peptidyl-carrier protein] + FAD + 2 H₂O

Other name(s): bmp2 (gene name)
Systematic name: 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein]:FADH₂ oxidoreductase (brominating)
Comments: The enzyme, characterized from marine bacteria of the *Pseudoalteromonas* genus, belongs to a family of FAD-dependent halogenases that act on acyl-carrier protein-tethered substrates. It catalyses three successive rounds of bromination. While the order has not been verified, it is believed to resemble that of EC 1.14.19.56, *S*-(1*H*-pyrrole-2-carbonyl)-[peptidyl-carrier protein] chlorinase, due to significant sequence homology. Reduced FAD is provided in situ by a dedicated reductase and diffuses into the active site, where it reacts with the oxygen and bromide ion, resulting in formation of a bromoamine intermediate on a catalytic lysine side chain, and the eventual transfer of the bromide to the substrate. The enzyme from *Pseudoalteromonas luteoviolacea* 2ta16 is specific for bromide and does not accept chloride.
References: [33]

[EC 1.14.19.57 created 2018]

EC 1.14.19.58

Accepted name: tryptophan 5-halogenase
Reaction: L-tryptophan + FADH₂ + chloride + O₂ + H⁺ = 5-chloro-L-tryptophan + FAD + 2 H₂O
Other name(s): *pyrH* (gene name)
Systematic name: L-tryptophan:FADH₂ oxidoreductase (5-halogenating)
Comments: A flavin-dependent halogenase. The enzyme from the bacterium *Streptomyces rugosporus* catalyses halogenation of the C-5 position of tryptophan during the biosynthesis of the antibiotic compound pyrroindomycin B. It utilizes molecular oxygen to oxidize the FADH₂ cofactor, giving C4a-hydroperoxyflavin, which then reacts with chloride to produce a hypochlorite ion. The latter reacts with an active site lysine to generate a chloramine, which chlorinates the substrate. *cf.* EC 1.14.19.59, tryptophan 6-halogenase and EC 1.14.19.9, tryptophan 7-halogenase.
References: [4865, 4925]

[EC 1.14.19.58 created 2018]

EC 1.14.19.59

Accepted name: tryptophan 6-halogenase
Reaction: (1) L-tryptophan + FADH₂ + chloride + O₂ + H⁺ = 6-chloro-L-tryptophan + FAD + 2 H₂O
(2) D-tryptophan + FADH₂ + chloride + O₂ + H⁺ = 6-chloro-D-tryptophan + FAD + 2 H₂O
Other name(s): *sttH* (gene name); *thdH* (gene name)
Systematic name: L-tryptophan:FADH₂ oxidoreductase (6-halogenating)
Comments: The enzyme is a flavin-dependent halogenase that has been described from several bacterial species. It utilizes molecular oxygen to oxidize the FADH₂ cofactor, giving C4a-hydroperoxyflavin, which then reacts with chloride to produce a hypochlorite ion. The latter reacts with an active site lysine to generate a chloramine, which chlorinates the substrate. *cf.* EC 1.14.19.58, tryptophan 5-halogenase, and EC 1.14.19.9, tryptophan 7-halogenase.
References: [4869, 2805, 3844]

[EC 1.14.19.59 created 2018]

EC 1.14.19.60

- Accepted name:** 7-chloro-L-tryptophan 6-halogenase
Reaction: 7-chloro-L-tryptophan + FADH₂ + chloride + O₂ + H⁺ = 6,7-dichloro-L-tryptophan + FAD + 2 H₂O
Other name(s): *ktzR* (gene name)
Systematic name: 7-chloro-L-tryptophan:FADH₂ oxidoreductase (6-halogenating)
Comments: An FAD-dependent halogenase. The enzyme, characterized from the bacterium *Kutzneria* sp. 744, works in tandem with EC 1.14.19.9, tryptophan 7-halogenase, (*ktzQ*) to generate 6,7-dichloro-L-tryptophan, which is incorporated as a pyrroloindoline in the kutznerides family of natural products. It has a 120-fold preference for 7-chloro-L-tryptophan over L-tryptophan as substrate.
References: [1603]

[EC 1.14.19.60 created 2018]

EC 1.14.19.61

- Accepted name:** dihydrorhizobitoxine desaturase
Reaction: dihydrorhizobitoxine + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = rhizobitoxine + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): *rtxC* (gene name)
Systematic name: dihydrorhizobitoxine,ferredoxin:oxygen oxidoreductase (3,4 *trans*-dehydrogenating)
Comments: The enzyme, characterized from the bacterium *Bradyrhizobium elkanii*, catalyses the final step in the biosynthesis of the nodulation enhancer compound rhizobitoxine.
References: [4781, 3162]

[EC 1.14.19.61 created 2018]

EC 1.14.19.62

- Accepted name:** secologanin synthase
Reaction: loganin + [reduced NADPH—hemoprotein reductase] + O₂ = secologanin + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Systematic name: loganin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (ring-cleaving)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Secologanin is the precursor of the monoterpene indole alkaloids and ipecac alkaloids.
References: [4737, 4736, 1823]

[EC 1.14.19.62 created 2002 as EC 1.3.3.9, transferred 2018 to EC 1.14.19.62]

EC 1.14.19.63

- Accepted name:** pseudobaptigenin synthase
Reaction: (1) calycosin + [reduced NADPH—hemoprotein reductase] + O₂ = pseudobaptigenin + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
(2) pratensein + [reduced NADPH-hemoprotein reductase] + O₂ = 5-hydroxypseudobaptigenin + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Systematic name: calycosin,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (methylenedioxy-bridge-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) enzyme catalysing an oxidative reaction that does not incorporate oxygen into the product. Catalyses a step in the biosynthesis of (–)-maackiain, the main pterocarpan phytoalexin in chickpea (*Cicer arietinum*).
References: [3617]

[EC 1.14.19.63 created 2011 as EC 1.14.21.8, transferred 2018 to EC 1.14.19.63]

EC 1.14.19.64

- Accepted name:** (*S*)-stylophine synthase

Reaction: (*S*)-cheilanthifoline + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-stylopine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): (*S*)-cheilanthifoline oxidase (methylenedioxy-bridge-forming)

Systematic name: (*S*)-cheilanthifoline,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (methylenedioxy-bridge-forming)

Comments: A cytochrome *P*-450 (heme-thiolate) protein catalysing an oxidative reaction that does not incorporate oxygen into the product. Forms the second methylenedioxy bridge of the protoberberine alkaloid stylopine from oxidative ring closure of adjacent phenolic and methoxy groups of cheilanthifoline.

References: [245]

[EC 1.14.19.64 created 1999 as EC 1.1.3.32, transferred 2002 to EC 1.14.21.1, transferred 2018 to EC 1.14.19.64]

EC 1.14.19.65

Accepted name: (*S*)-cheilanthifoline synthase

Reaction: (*S*)-scoulerine + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-cheilanthifoline + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): CYP719A14 (gene name); (*S*)-scoulerine oxidase (methylenedioxy-bridge-forming) (ambiguous)

Systematic name: (*S*)-scoulerine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase [(*S*)-cheilanthifoline-forming]

Comments: A cytochrome *P*-450 (heme-thiolate) protein catalysing an oxidative reaction that does not incorporate oxygen into the product. Forms the methylenedioxy bridge of the protoberberine alkaloid cheilanthifoline by the oxidative ring closure of adjacent phenolic and methoxy groups of scoulerine. *cf.* EC 1.14.19.73, (*S*)-nandinine synthase, which catalyses a similar reaction at the other side of the (*S*)-scoulerine molecule, forming (*S*)-nandinine.

References: [245, 621]

[EC 1.14.19.65 created 1999 as EC 1.1.3.33, transferred 2002 to EC 1.14.21.2, modified 2016, transferred 2018 to EC 1.14.19.65]

EC 1.14.19.66

Accepted name: berbamunine synthase

Reaction: (*S*)-*N*-methylcoclaurine + (*R*)-*N*-methylcoclaurine + [reduced NADPH—hemoprotein reductase] + O₂ = berbamunine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): (*S*)-*N*-methylcoclaurine oxidase (C-O phenol-coupling)

Systematic name: (*S*)-*N*-methylcoclaurine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (C-O phenol-coupling)

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. Forms the bisbenzylisoquinoline alkaloid berbamunine by phenol oxidation of *N*-methylcoclaurine without the incorporation of oxygen into the product. Reaction of two molecules of (*R*)-*N*-methylcoclaurine gives the dimer guattagaumerine.

References: [3995]

[EC 1.14.19.66 created 1999 as EC 1.1.3.34, transferred 2002 to EC 1.14.21.3, transferred 2018 to EC 1.14.19.66]

EC 1.14.19.67

Accepted name: salutaridine synthase

Reaction: (*R*)-reticuline + [reduced NADPH—hemoprotein reductase] + O₂ = salutaridine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O

Other name(s): (*R*)-reticuline oxidase (C-C phenol-coupling)

Systematic name: (*R*)-reticuline,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (C-C phenol-coupling)

Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. Forms the morphinan alkaloid salutaridine by intramolecular phenol oxidation of reticuline without the incorporation of oxygen into the product.

References: [1300]

[EC 1.14.19.67 created 1999 as EC 1.1.3.35, transferred 2002 to EC 1.14.21.4, transferred 2018 to EC 1.14.19.67]

EC 1.14.19.68

- Accepted name:** (*S*)-canadine synthase
Reaction: (*S*)-tetrahydrocolumbamine + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-canadine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): (*S*)-tetrahydroberberine synthase; (*S*)-tetrahydrocolumbamine oxidase (methylenedioxy-bridge-forming); CYP719A (gene name)
Systematic name: (*S*)-tetrahydrocolumbamine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (methylenedioxy-bridge-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein found in plants. The enzyme catalyses an oxidative reaction that does not incorporate oxygen into the product. Oxidation of the methoxyphenol group of the alkaloid tetrahydrocolumbamine results in the formation of the methylenedioxy bridge of canadine.
References: [3597, 1800, 820]

[EC 1.14.19.68 created 1999 as EC 1.1.3.36, transferred 2002 to EC 1.14.21.5, transferred 2018 to EC 1.14.19.68]

EC 1.14.19.69

- Accepted name:** biflaviolin synthase
Reaction: (1) 2 flaviolin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = 3,3'-biflaviolin + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
(2) 2 flaviolin + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = 3,8'-biflaviolin + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): CYP158A2 (gene name); cytochrome P450 158A2
Systematic name: flaviolin, reduced ferredoxin:oxygen oxidoreductase
Comments: This cytochrome-*P*-450 (heme-thiolate) enzyme, from the soil-dwelling bacterium *Streptomyces coelicolor* A3(2), catalyses a phenol oxidation C-C coupling reaction, which results in the polymerization of flaviolin to form biflaviolin or triflaviolin without the incorporation of oxygen into the product [4894, 4896]. The products are highly conjugated pigments that protect the bacterium from the deleterious effects of UV irradiation [4894].
References: [4894, 4895, 4896]

[EC 1.14.19.69 created 2008 as EC 1.14.21.7, transferred 2018 to EC 1.14.19.69]

EC 1.14.19.70

- Accepted name:** mycocyclusin synthase
Reaction: cyclo(L-tyrosyl-L-tyrosyl) + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺ + O₂ = mycocyclusin + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O
Other name(s): CYP121; rv2276 (locus name)
Systematic name: cyclo(L-tyrosyl-L-tyrosyl), reduced ferredoxin:oxygen oxidoreductase (diarylbridge-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein from the bacterium *Mycobacterium tuberculosis* catalysing an oxidative reaction that does not incorporate oxygen into the product.
References: [278]

[EC 1.14.19.70 created 2013 as EC 1.14.21.9, transferred 2018 to EC 1.14.19.70]

EC 1.14.19.71

- Accepted name:** fumitremorgin C synthase
Reaction: tryprostatin A + [reduced NADPH—hemoprotein reductase] + O₂ = fumitremorgin C + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): *ftmE* (gene name)

Systematic name: tryprostatin A,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The protein from the fungus *Aspergillus fumigatus* also has activity with tryprostatin B forming demethoxyfumitremorgin C. Involved in the biosynthetic pathways of several indole alkaloids such as fumitremorgins and verruculogen.
References: [2019]

[EC 1.14.19.71 created 2013 as EC 1.14.21.10, transferred 2018 to EC 1.14.19.71]

EC 1.14.19.72

Accepted name: (–)-pluviatolide synthase
Reaction: (–)-matairesinol + [reduced NADPH—hemoprotein reductase] + O₂ = (–)-pluviatolide + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): CYP719A23 (gene name)
Systematic name: (–)-matairesinol,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (methylenedioxy-bridge-forming)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. The enzyme from the plants *Sinopodophyllum hexandrum* and *Podophyllum peltatum* catalyses the formation of a methylenedioxy-bridge. It is involved in the biosynthesis of podophyllotoxin, a non-alkaloid toxin lignan whose derivatives are important anticancer drugs.
References: [2657]

[EC 1.14.19.72 created 2016 as EC 1.14.21.11, transferred 2018 to EC 1.14.19.72]

EC 1.14.19.73

Accepted name: (*S*)-nandinine synthase
Reaction: (*S*)-scoulerine + [reduced NADPH—hemoprotein reductase] + O₂ = (*S*)-nandinine + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
Other name(s): CYP719A3
Systematic name: (*S*)-scoulerine,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase [(*S*)-nandinine-forming]
Comments: A cytochrome *P*-450 (heme-thiolate) enzyme found in plants. The enzyme catalyses an oxidative reaction that does not incorporate oxygen into the product. Forms the methylenedioxy bridge of the protoberberine alkaloid (*S*)-nandinine by the oxidative ring closure of adjacent phenolic and methoxy groups of (*S*)-scoulerine. *cf.* EC 1.14.19.65, (*S*)-cheilanthifoline synthase, which catalyses a similar reaction at the other side of the (*S*)-scoulerine molecule, forming (*S*)-cheilanthifoline.
References: [1798, 621]

[EC 1.14.19.73 created 2016 as EC 1.14.21.12, transferred 2018 to EC 1.14.19.73]

EC 1.14.19.74

Accepted name: (+)-piperitol/(+)-sesamin synthase
Reaction: (1) (+)-pinosresinol + [reduced NADPH-hemoprotein reductase] + O₂ = (+)-piperitol + [oxidized NADPH-hemoprotein reductase] + 2 H₂O
(2) (+)-piperitol + [reduced NADPH-hemoprotein reductase] + O₂ = (+)-sesamin + [oxidized NADPH-hemoprotein reductase] + 2 H₂O
Other name(s): CYP81Q1; CYP81Q2; PS; PSS; SS; piperitol synthase; sesamin synthase
Systematic name: (+)-pinosresinol,[reduced NADPH-hemoprotein reductase]:oxygen oxidoreductase (cyclizing)
Comments: A cytochrome *P*-450 (heme-thiolate) protein. Isolated from *Sesamum indicum* (sesame) and *S. radiatum* (black sesame).
References: [3185]

[EC 1.14.19.74 created 2018]

EC 1.14.19.75

- Accepted name:** very-long-chain acyl-lipid ω -9 desaturase
- Reaction:** (1) 1-hexacosanoyl-2-acyl-[phosphoglycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 1-[(17Z)-hexacos-17-enoyl]-2-acyl-[phosphoglycerolipid] + 2 ferricyclochrome b_5 + 2 H₂O
(2) 1-tetracosanoyl-2-acyl-[phosphoglycerolipid] + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = 1-[(15Z)-tetracos-15-enoyl]-2-acyl-[phosphoglycerolipid] + 2 ferricyclochrome b_5 + 2 H₂O
- Other name(s):** ADS2 (gene name)
- Systematic name:** very-long-chain acyl-[glycerolipid],ferrocyclochrome b_5 :oxygen oxidoreductase (ω^9,ω^8 -*cis*-dehydrogenating)
- Comments:** The enzyme, characterized from the plant *Arabidopsis thaliana*, acts on both 24:0 and 26:0 fatty acids, introducing a *cis* double bond at a position 9 carbons from the methyl end. These very-long-chain fatty acids are found as a minor component of seed lipids, but also in the membrane phosphatidylethanolamine and phosphatidylserine, in sphingolipids, as precursors and components of cuticular and epicuticular waxes, and in suberin.
- References:** [1218, 3938]

[EC 1.14.19.75 created 2018]

EC 1.14.19.76

- Accepted name:** flavone synthase II
- Reaction:** a flavanone + [reduced NADPH—hemoprotein reductase] + O₂ = a flavone + [oxidized NADPH—hemoprotein reductase] + 2 H₂O
- Other name(s):** CYP93B16 (gene name); CYP93G1 (gene name); FNS II
- Systematic name:** flavanone,[reduced NADPH—hemoprotein reductase]:oxygen oxidoreductase (flavone-forming)
- Comments:** A cytochrome *P*-450 (heme-thiolate) protein found in plants. The rice enzyme channels flavanones to the biosynthesis of tricin O-linked conjugates. *cf.* EC 1.14.20.5, flavone synthase I.
- References:** [2661, 1131, 2330]

[EC 1.14.19.76 created 2018]

EC 1.14.19.77

- Accepted name:** plasmalyethanolamine desaturase
- Reaction:** a plasmalyethanolamine + 2 ferrocyclochrome b_5 + O₂ + 2 H⁺ = a plasmenylethanolamine + 2 ferricyclochrome b_5 + 2 H₂O
- Other name(s):** TMEM189 (gene name); 2-acyl-1-alkyl-*sn*-glycero-3-phosphoethanolamine desaturase; alkylacyl-glycerophosphoethanolamine desaturase; alkylacylglycero-phosphorylethanolamine dehydrogenase; alkyl-acylglycerophosphorylethanolamine dehydrogenase; 1-*O*-alkyl-2-acyl-*sn*-glycero-3-phosphorylethanolamine desaturase; 1-*O*-alkyl 2-acyl-*sn*-glycero-3-phosphorylethanolamine desaturase
- Systematic name:** plasmalyethanolamine,ferrocyclochrome b_5 :oxygen oxidoreductase (plasmenylethanolamine-forming)
- Comments:** The enzyme catalyses the introduction of a double bond at position 1 of the alkyl group attached by an ether bond at the *sn*-1 position of plasmalyethanolamine, generating a vinyl ether-containing plasmenylethanolamine. The enzyme is found in animals and some bacteria, but not in plants, fungi, or most aerobic bacteria.
- References:** [4041, 3226, 3227, 4690, 1259]

[EC 1.14.19.77 created 1976 as EC 1.14.99.19, transferred 2020 to EC 1.14.19.77]

EC 1.14.19.78

- Accepted name:** decanoyl-[acyl-carrier protein] acetylenase
- Reaction:** decanoyl-[acyl-carrier protein] + 4 reduced ferredoxin [iron-sulfur] cluster + 2 O₂ + 4 H⁺ = dec-9-ynoyl-[acyl-carrier protein] + 4 oxidized ferredoxin [iron-sulfur] cluster + 4 H₂O (overall reaction)

(1a) decanoyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = dec-9-enoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O

(1b) dec-9-enoyl-[acyl-carrier protein] + 2 reduced ferredoxin [iron-sulfur] cluster + O₂ + 2 H⁺ = dec-9-ynoyl-[acyl-carrier protein] + 2 oxidized ferredoxin [iron-sulfur] cluster + 2 H₂O

Other name(s): *ttuB* (gene name) (ambiguous)
Systematic name: decanoyl-[acyl-carrier protein],reduced ferredoxin:oxygen oxidoreductase (9,10-dehydrogenating)
Comments: The enzyme, characterized from the bacterium *Teredinibacter turnerae*, is specific for decanoyl-[acyl-carrier protein]. Activity is maximal when decanoate is loaded onto a dedicated acyl-carrier protein (TtuC), which is encoded by a gene in the same operon.
References: [4927]

[EC 1.14.19.78 created 2021]

EC 1.14.19.79

Accepted name: 3β,22α-dihydroxysteroid 3-dehydrogenase
Reaction: (1) (22*S*)-22-hydroxycampesterol + [reduced NADPH-hemoprotein reductase] + O₂ = (22*S*)-22-hydroxycampest-4-en-3-one + [oxidized NADPH-hemoprotein reductase] + 2 H₂O
(2) 6-deoxoteasterone + [reduced NADPH-hemoprotein reductase] + O₂ = 3-dehydro-6-deoxoteasterone + [oxidized NADPH-hemoprotein reductase] + 2 H₂O
Other name(s): CYP90A1 (gene name)
Systematic name: 3β,22α-dihydroxysteroid,[reduced NADPH-hemoprotein reductase]:oxygen 3-oxidoreductase
Comments: This cytochrome *P*-450 (heme-thiolate) enzyme, characterized from the plant *Arabidopsis thaliana*, catalyses C-3 dehydrogenation of all 3β-hydroxy brassinosteroid synthesis intermediates with 22-hydroxylated or 2²,2³-dihydroxylated side chains.
References: [3141]

[EC 1.14.19.79 created 2022]

EC 1.14.20 With 2-oxoglutarate as one donor, and the other dehydrogenated

EC 1.14.20.1

Accepted name: deacetoxycephalosporin-C synthase
Reaction: penicillin N + 2-oxoglutarate + O₂ = deacetoxycephalosporin C + succinate + CO₂ + H₂O
Other name(s): DAOCS; penicillin N expandase; DAOC synthase
Systematic name: penicillin-N,2-oxoglutarate:oxygen oxidoreductase (ring-expanding)
Comments: Forms part of the penicillin biosynthesis pathway (for pathway, [click here](#)).
References: [546, 2383, 4787, 4390, 957]

[EC 1.14.20.1 created 2002]

[1.14.20.2 *Transferred entry. 2,4-dihydroxy-1,4-benzoxazin-3-one-glucoside dioxygenase. Now EC 1.14.11.59, 2,4-dihydroxy-1,4-benzoxazin-3-one-glucoside dioxygenase*]

[EC 1.14.20.2 created 2012, deleted 2018]

EC 1.14.20.3

Accepted name: (5*R*)-carbapenem-3-carboxylate synthase
Reaction: (3*S*,5*S*)-carbapenam-3-carboxylate + 2-oxoglutarate + O₂ = (5*R*)-carbapen-2-em-3-carboxylate + succinate + CO₂ + H₂O
Other name(s): *carC* (gene name)
Systematic name: (3*S*,5*S*)-carbapenam-3-carboxylate,2-oxoglutarate:oxygen oxidoreductase (dehydrating)

Comments: Requires Fe²⁺. The enzyme is involved in the biosynthesis of the carbapenem β-lactam antibiotic (5*R*)-carbapen-2-em-3-carboxylate in the bacterium *Pectobacterium carotovorum*. It catalyses a stereoinversion at C-5 and introduces a double bond between C-2 and C-3.

References: [702, 4005, 3927]

[EC 1.14.20.3 created 2013]

EC 1.14.20.4

Accepted name: anthocyanidin synthase

Reaction: a (2*R*,3*S*,4*S*)-leucoanthocyanidin + 2-oxoglutarate + O₂ = an anthocyanidin + succinate + CO₂ + 2 H₂O (overall reaction)

(1a) a (2*R*,3*S*,4*S*)-leucoanthocyanidin + 2-oxoglutarate + O₂ = a (4*S*)- 2,3-dehydroflavan-3,4-diol + succinate + CO₂ + H₂O

(1b) a (4*S*)- 2,3-dehydroflavan-3,4-diol = an anthocyanidin + H₂O

Other name(s): leucocyanidin oxygenase; leucocyanidin,2-oxoglutarate:oxygen oxidoreductase; ANS (gene name)

Systematic name: (2*R*,3*S*,4*S*)-leucoanthocyanidin,2-oxoglutarate:oxygen oxidoreductase

Comments: The enzyme requires iron(II) and ascorbate. It is involved in the pathway by which many flowering plants make anthocyanin flower pigments (glycosylated anthocyanidins). The enzyme hydroxylates the C-3 carbon, followed by a *trans* diaxial elimination, forming a C-2,C-3 enol. The product loses a second water molecule to form anthocyanidins. When assayed *in vitro*, non-enzymic epimerization of the product can lead to formation of dihydroflavanols. Thus when the substrate is leucocyanidin, a mixture of (+)-taxifolin and (+)-epitaxifolin are formed. The enzyme can also oxidize the formed (+)-taxifolin to quercetin (*cf.* EC 1.14.20.6, flavonol synthase) [4352, 4639].

References: [3630, 4352, 4639, 4350, 4582]

[EC 1.14.20.4 created 2001 as EC 1.14.11.19, transferred 2018 to EC 1.14.20.4]

EC 1.14.20.5

Accepted name: flavone synthase I

Reaction: a flavanone + 2-oxoglutarate + O₂ = a flavone + succinate + CO₂ + H₂O

Other name(s): FNSI (gene name)

Systematic name: flavanone,2-oxoglutarate:oxygen oxidoreductase (dehydrating)

Comments: The enzyme, which has been found in rice and in members of the Apiaceae (a plant family), is a member of the 2-oxoglutarate-dependent dioxygenases, and requires ascorbate and Fe²⁺ for full activity.

References: [2663, 2559, 2662]

[EC 1.14.20.5 created 2004 as EC 1.14.11.22, transferred 2018 to EC 1.14.20.5]

EC 1.14.20.6

Accepted name: flavonol synthase

Reaction: a dihydroflavonol + 2-oxoglutarate + O₂ = a flavonol + succinate + CO₂ + H₂O

Other name(s): FLS (gene name)

Systematic name: dihydroflavonol,2-oxoglutarate:oxygen oxidoreductase

Comments: In addition to the desaturation of (2*R*,3*R*)-dihydroflavonols to flavonols, the enzyme from *Citrus unshiu* (satsuma mandarin) also has a non-specific activity that *trans*-hydroxylates the flavanones (2*S*)-naringenin and the unnatural (2*R*)-naringenin at C-3 to kaempferol and (2*R*,3*R*)-dihydrokaempferol, respectively [2560]. Requires Fe²⁺.

References: [4583, 2560, 2662, 4351]

[EC 1.14.20.6 created 2004 as EC 1.14.11.23, transferred 2018 to EC 1.14.20.6]

EC 1.14.20.7

- Accepted name:** 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming)
- Reaction:** L-arginine + 2-oxoglutarate + O₂ = succinate + CO₂ + guanidine + (S)-1-pyrroline-5-carboxylate + H₂O (overall reaction)
(1a) L-arginine + 2-oxoglutarate + O₂ = succinate + CO₂ + 5-hydroxy-L-arginine
(1b) 5-hydroxy-L-arginine = guanidine + (S)-1-pyrroline-5-carboxylate + H₂O
- Other name(s):** ethene-forming enzyme; ethylene-forming enzyme; EFE
- Systematic name:** L-arginine,2-oxoglutarate:oxygen oxidoreductase (succinate-forming)
- Comments:** This is one of two simultaneous reactions catalysed by the enzyme, which is responsible for ethylene production in bacteria of the *Pseudomonas syringae* group. In the other reaction [EC 1.13.12.19, 2-oxoglutarate dioxygenase (ethene-forming)] the enzyme catalyses the dioxygenation of 2-oxoglutarate forming ethene and three molecules of carbon dioxide. The enzyme catalyses two cycles of the ethene-forming reaction for each cycle of the succinate-forming reaction, so that the stoichiometry of the products ethene and succinate is 2:1.
- References:** [2954, 1222, 1221, 2672]

[EC 1.14.20.7 created 2011 as EC 1.14.11.34, transferred 2018 to EC 1.14.20.7]

EC 1.14.20.8

- Accepted name:** (–)-deoxypodophyllotoxin synthase
- Reaction:** (–)-yatein + 2-oxoglutarate + O₂ = (–)-deoxypodophyllotoxin + succinate + CO₂ + H₂O
- Other name(s):** 2-ODD (gene name)
- Systematic name:** (–)-yatein,2-oxoglutarate:oxygen oxidoreductase (ring-forming)
- Comments:** The enzyme, characterized from the plant *Sinopodophyllum hexandrum* (mayapple), is involved in the biosynthetic pathway of podophyllotoxin, a non-alkaloid toxin lignan whose derivatives are important anticancer drugs. It catalyses the closure of the central six-membered ring in the aryltetralin scaffold.
- References:** [2365]

[EC 1.14.20.8 created 2016 as EC 1.14.11.50, transferred 2018 to EC 1.14.20.8]

EC 1.14.20.9

- Accepted name:** L-tyrosine isonitrile desaturase
- Reaction:** (2S)-3-(4-hydroxyphenyl)-2-isocyanopropanoate + 2-oxoglutarate + O₂ = (2E)-3-(4-hydroxyphenyl)-2-isocyanoprop-2-enoate + succinate + CO₂ + H₂O
- Other name(s):** *pvcB* (gene name)
- Systematic name:** (2S)-3-(4-hydroxyphenyl)-2-isocyanopropanoate,2-oxoglutarate:oxygen oxidoreductase
- Comments:** The enzyme is a member of the Fe²⁺, 2-oxoglutarate-dependent oxygenases and requires Fe²⁺. It has been characterized from bacteria that form the isonitrile-functionalized compound paerucumarin. *cf.* EC 1.14.20.10, L-tyrosine isonitrile desaturase/decarboxylase.
- References:** [698, 963, 4922]

[EC 1.14.20.9 created 2018]

EC 1.14.20.10

- Accepted name:** L-tyrosine isonitrile desaturase/decarboxylase
- Reaction:** (2S)-3-(4-hydroxyphenyl)-2-isocyanopropanoate + 2-oxoglutarate + O₂ = 4-[(E)-2-isocyanoethenyl]phenol + succinate + 2 CO₂ + H₂O
- Other name(s):** *pvcB* (gene name)
- Systematic name:** (2S)-3-(4-hydroxyphenyl)-2-isocyanopropanoate,2-oxoglutarate:oxygen oxidoreductase (decarboxylating)

Comments: The enzyme, characterized from the bacterium *Xenorhabdus nematophila*, is involved in rhabduscin biosynthesis. The enzyme is a member of the Fe²⁺, 2-oxoglutarate-dependent oxygenases. It is similar to EC 1.14.20.9, L-tyrosine isonitrile desaturase. However, the latter does not catalyse a decarboxylation of the substrate.

References: [770, 4922]

[EC 1.14.20.10 created 2018]

EC 1.14.20.11

Accepted name: 3-[(Z)-2-isocyanoethenyl]-1H-indole synthase

Reaction: (2S)-3-(1H-indol-3-yl)-2-isocyanopropanoate + 2-oxoglutarate + O₂ = 3-[(Z)-2-isocyanoethenyl]-1H-indole + succinate + 2 CO₂ + H₂O

Other name(s): *ambI3* (gene name); *famH3* (gene name); L-tryptophan isonitrile desaturase/decarboxylase (3-[(Z)-2-isocyanoethenyl]-1H-indole-forming)

Systematic name: (2S)-3-(1H-indol-3-yl)-2-isocyanopropanoate,2-oxoglutarate:oxygen oxidoreductase (decarboxylating, 3-[(Z)-2-isocyanoethenyl]-1H-indole-forming)

Comments: The enzyme, characterized from the cyanobacterium *Fischerella ambigua* UTEX 1903, participates in the biosynthesis of hapalindole-type alkaloids. The enzyme catalyses an Fe²⁺, 2-oxoglutarate-dependent monooxygenation at C-3, which is followed by decarboxylation and dehydration, resulting in the generation of a *cis* C-C double bond. *cf.* EC 1.14.20.12, 3-[(E)-2-isocyanoethenyl]-1H-indole synthase.

References: [1659, 601]

[EC 1.14.20.11 created 2018]

EC 1.14.20.12

Accepted name: 3-[(E)-2-isocyanoethenyl]-1H-indole synthase

Reaction: (2S)-3-(1H-indol-3-yl)-2-isocyanopropanoate + 2-oxoglutarate + O₂ = 3-[(E)-2-isocyanoethenyl]-1H-indole + succinate + 2 CO₂ + H₂O

Other name(s): *isnB* (gene name); L-tryptophan isonitrile desaturase/decarboxylase (3-[(E)-2-isocyanoethenyl]-1H-indole-forming)

Systematic name: (2S)-3-(1H-indol-3-yl)-2-isocyanopropanoate,2-oxoglutarate:oxygen oxidoreductase (decarboxylating, 3-[(E)-2-isocyanoethenyl]-1H-indole-forming)

Comments: The enzyme has been characterized from an unidentified soil bacterium. It catalyses an Fe²⁺, 2-oxoglutarate-dependent monooxygenation at C-3, which is followed by decarboxylation and dehydration, resulting in the generation of a *trans* C-C double bond. *cf.* EC 1.14.20.11, 3-[(Z)-2-isocyanoethenyl]-1H-indole synthase.

References: [421, 601]

[EC 1.14.20.12 created 2018]

EC 1.14.20.13

Accepted name: 6β-hydroxyhyoscyamine epoxidase

Reaction: (6S)-6β-hydroxyhyoscyamine + 2-oxoglutarate + O₂ = scopolamine + succinate + CO₂ + H₂O

Other name(s): hydroxyhyoscyamine dioxygenase; (6S)-6-hydroxyhyoscyamine,2-oxoglutarate oxidoreductase (epoxide-forming)

Systematic name: (6S)-6β-hydroxyhyoscyamine,2-oxoglutarate:oxygen oxidoreductase (epoxide-forming)

Comments: Requires Fe²⁺ and ascorbate.

References: [1550]

[EC 1.14.20.13 created 1992 as EC 1.14.11.14, transferred 2018 to EC 1.14.20.13]

EC 1.14.20.14

- Accepted name:** hapalindole-type alkaloid chlorinase
- Reaction:** (1) hapalindole U + 2-oxoglutarate + O₂ + chloride = hapalindole G + succinate + CO₂ + H₂O
(2) 12-*epi*-fischerindole U + 2-oxoglutarate + O₂ + chloride = 12-*epi*-fischerindole G + succinate + CO₂ + H₂O
- Other name(s):** *ambO5* (gene name); *welO5* (gene name)
- Systematic name:** 12-*epi*-fischerindole U,2-oxoglutarate:oxygen oxidoreductase (13-halogenating)
- Comments:** The enzyme, characterized from hapalindole-type alkaloids-producing cyanobacteria, is a specialized iron(II)/2-oxoglutarate-dependent oxygenase that catalyses the chlorination of its substrates in a reaction that requires oxygen, chloride ions, iron(II) and 2-oxoglutarate.
- References:** [1657, 4923, 1658]

[EC 1.14.20.14 created 2018]

EC 1.14.20.15

- Accepted name:** L-threonyl-[L-threonyl-carrier protein] 4-chlorinase
- Reaction:** an L-threonyl-[L-threonyl-carrier protein] + 2-oxoglutarate + O₂ + Cl⁻ = a 4-chloro-L-threonyl-[L-threonyl-carrier protein] + succinate + CO₂ + H₂O
- Other name(s):** *syrB2* (gene name)
- Systematic name:** L-threonyl-[L-threonyl-carrier protein],2-oxoglutarate:oxygen oxidoreductase (4-halogenating)
- Comments:** The enzyme, characterized from the bacterium *Pseudomonas syringae*, participates in syringomycin E biosynthesis. The enzyme is a specialized iron(II)/2-oxoglutarate-dependent oxygenase that catalyses the chlorination of its substrate in a reaction that requires oxygen, chloride ions, ferrous iron and 2-oxoglutarate.
- References:** [4388]

[EC 1.14.20.15 created 2018]

EC 1.14.21 With NADH or NADPH as one donor, and the other dehydrogenated

[1.14.21.1 *Transferred entry. (S)-stylopine synthase. Now EC 1.14.19.64, (S)-stylopine synthase*]

[EC 1.14.21.1 created 2002, deleted 2018]

[1.14.21.2 *Transferred entry. (S)-cheilanthifoline synthase. Now EC 1.14.19.65, (S)-cheilanthifoline synthase*]

[EC 1.14.21.2 created 2002, modified 2016, deleted 2018]

[1.14.21.3 *Transferred entry. berbamunine synthase. Now EC 1.14.19.66, berbamunine synthase*]

[EC 1.14.21.3 created 2002, deleted 2018]

[1.14.21.4 *Transferred entry. salutaridine synthase. Now EC 1.14.19.67, salutaridine synthase*]

[EC 1.14.21.4 created 2002, deleted 2018]

[1.14.21.5 *Transferred entry. (S)-canadine synthase. Now EC 1.14.19.68, (S)-canadine synthase*]

[EC 1.14.21.5 created 2002, deleted 2018]

[1.14.21.6 *Transferred entry. lathosterol oxidase. Now EC 1.14.19.20, Δ⁷-sterol 5(6)-desaturase*]

[EC 1.14.21.6 created 1972 as EC 1.3.3.2, transferred 2005 to EC 1.14.21.6, deleted 2015]

[1.14.21.7 *Transferred entry. biflaviolin synthase. Now EC 1.14.19.69, biflaviolin synthase*]

[EC 1.14.21.7 created 2008, deleted 2018]

[1.14.21.8 *Transferred entry. pseudobaptigenin synthase. Now EC 1.14.19.63, pseudobaptigenin synthase.*]

[EC 1.14.21.8 created 2011, deleted 2018]

[1.14.21.9 Transferred entry. *mycocyclosin synthase*. Now EC 1.14.19.70, *mycocyclosin synthase*]

[EC 1.14.21.9 created 2013, deleted 2018]

[1.14.21.10 Transferred entry. *fumitremorgin C synthase*. Now EC 1.14.19.71, *fumitremorgin C synthase*]

[EC 1.14.21.10 created 2013, deleted 2018]

[1.14.21.11 Transferred entry. *(-)-pluviatolide synthase*. Now EC 1.14.19.72, *(-)-pluviatolide synthase*]

[EC 1.14.21.11 created 2016, deleted 2018]

[1.14.21.12 Transferred entry. *(S)-nandinine synthase*. Now EC 1.14.19.73, *(S)-nandinine synthase*]

[EC 1.14.21.12 created 2016, deleted 2018]

EC 1.14.99 Miscellaneous

EC 1.14.99.1

Accepted name: prostaglandin-endoperoxide synthase
Reaction: arachidonate + reduced acceptor + 2 O₂ = prostaglandin H₂ + acceptor + H₂O
Other name(s): prostaglandin synthase; prostaglandin G/H synthase; (PG)H synthase; PG synthetase; prostaglandin synthetase; fatty acid cyclooxygenase; prostaglandin endoperoxide synthetase
Systematic name: (5Z,8Z,11Z,14Z)-icosa-5,8,11,14-tetraenoate,hydrogen-donor:oxygen oxidoreductase
Comments: This enzyme acts both as a dioxygenase and as a peroxidase.
References: [896, 3139]

[EC 1.14.99.1 created 1972, modified 1990]

EC 1.14.99.2

Accepted name: kynurenine 7,8-hydroxylase
Reaction: kynurenate + reduced acceptor + O₂ = 7,8-dihydro-7,8-dihydroxykynurenate + acceptor
Other name(s): kynurenic acid hydroxylase; kynurenic hydroxylase; kynurenate 7,8-hydroxylase
Systematic name: kynurenate,hydrogen-donor:oxygen oxidoreductase (hydroxylating)
References: [4210]

[EC 1.14.99.2 created 1965 as EC 1.14.1.4, transferred 1972 to EC 1.14.99.2]

[1.14.99.3 Transferred entry. *heme oxygenase (biliverdin-producing)*. Now EC 1.14.14.18, *heme oxygenase (biliverdin-producing)*]

[EC 1.14.99.3 created 1972, modified 2006, deleted 2015]

EC 1.14.99.4

Accepted name: progesterone monooxygenase
Reaction: progesterone + reduced acceptor + O₂ = testosterone acetate + acceptor + H₂O
Other name(s): progesterone hydroxylase
Systematic name: progesterone,hydrogen-donor:oxygen oxidoreductase (hydroxylating)
Comments: Has a wide specificity. A single enzyme from ascomycete the *Neonectria radicola* (EC 1.14.13.54 ketosteroid monooxygenase) catalyses both this reaction and that catalysed by EC 1.14.99.12 androst-4-ene-3,17-dione monooxygenase.
References: [3429]

[EC 1.14.99.4 created 1972, modified 1999]

[1.14.99.5 Transferred entry. *stearoyl-CoA desaturase*. Now EC 1.14.19.1, *stearoyl-CoA 9-desaturase*]

[EC 1.14.99.5 created 1972, modified 1986, modified 2000, deleted 2000]

[1.14.99.6 Transferred entry. *acyl-[acyl-carrier-protein] desaturase*. Now EC 1.14.19.2, *acyl-[acyl-carrier-protein] desaturase*]

[EC 1.14.99.6 created 1972, modified 2000, deleted 2000]

[1.14.99.7 Transferred entry. *squalene monooxygenase*. Transferred to EC 1.14.13.132, *squalene monooxygenase*.]

[EC 1.14.99.7 created 1961 as EC 1.99.1.13, transferred 1965 to EC 1.14.1.3, part transferred 1972 to EC 1.14.99.7 rest to EC 5.4.99.7, deleted 2011]

[1.14.99.8 Deleted entry. *arene monooxygenase (epoxidizing)*. Now included with EC 1.14.14.1 *unspecific monooxygenase*]

[EC 1.14.99.8 created 1972, deleted 1984]

[1.14.99.9 Transferred entry. *steroid 17 α -monooxygenase*, now classified as EC 1.14.14.19, *steroid 17 α -monooxygenase*]

[EC 1.14.99.9 created 1961 as EC 1.99.1.9, transferred 1965 to EC 1.14.1.7, transferred 1972 to EC 1.14.99.9, modified 2013, deleted 2015]

[1.14.99.10 Transferred entry. *steroid 21-monooxygenase*. Now EC 1.14.14.16, *steroid 21-monooxygenase*]

[EC 1.14.99.10 created 1961 as EC 1.99.1.11, transferred 1965 to EC 1.14.1.8, transferred 1972 to EC 1.14.99.10, modified 2013, deleted 2015]

EC 1.14.99.11

Accepted name: estradiol 6 β -monooxygenase
Reaction: estradiol-17 β + reduced acceptor + O₂ = 6 β -hydroxyestradiol-17 β + acceptor + H₂O
Other name(s): estradiol 6 β -hydroxylase
Systematic name: estradiol-17 β ,hydrogen-donor:oxygen oxidoreductase (6 β -hydroxylating)
References: [1479, 2915]

[EC 1.14.99.11 created 1965 as EC 1.14.1.10, transferred 1972 to EC 1.14.99.11]

EC 1.14.99.12

Accepted name: androst-4-ene-3,17-dione monooxygenase
Reaction: androstenedione + reduced acceptor + O₂ = testololactone + acceptor + H₂O
Other name(s): androstene-3,17-dione hydroxylase; androst-4-ene-3,17-dione 17-oxidoreductase; androst-4-ene-3,17-dione hydroxylase; androstenedione monooxygenase; 4-androstene-3,17-dione monooxygenase
Systematic name: androst-4-ene-3,17-dione-hydrogen-donor:oxygen oxidoreductase (13-hydroxylating, lactonizing)
Comments: Has a wide specificity. A single enzyme from the ascomycete *Neonectria radicola* (EC 1.14.13.54, ketosteroid monooxygenase) catalyses both this reaction and that catalysed by EC 1.14.99.4, progesterone monooxygenase.
References: [3369]

[EC 1.14.99.12 created 1972, modified 1999]

[1.14.99.13 Transferred entry. *3-hydroxybenzoate 4-monooxygenase*. Now EC 1.14.13.23, *3-hydroxybenzoate 4-monooxygenase*]

[EC 1.14.99.13 created 1972, deleted 1984]

EC 1.14.99.14

Accepted name: progesterone 11 α -monooxygenase
Reaction: progesterone + reduced acceptor + O₂ = 11 α -hydroxyprogesterone + acceptor + H₂O
Other name(s): progesterone 11 α -hydroxylase
Systematic name: progesterone,hydrogen-donor:oxygen oxidoreductase (11 α -hydroxylating)

References: [3851]

[EC 1.14.99.14 created 1972]

EC 1.14.99.15

Accepted name: 4-methoxybenzoate monooxygenase (*O*-demethylating)
Reaction: 4-methoxybenzoate + reduced acceptor + O₂ = 4-hydroxybenzoate + formaldehyde + acceptor + H₂O
Other name(s): 4-methoxybenzoate 4-monooxygenase (*O*-demethylating); 4-methoxybenzoate *O*-demethylase; *p*-anisic *O*-demethylase; piperonylate-4-*O*-demethylase
Systematic name: 4-methoxybenzoate,hydrogen-donor:oxygen oxidoreductase (*O*-demethylating)
Comments: The bacterial enzyme consists of a ferredoxin-type protein and an iron-sulfur flavoprotein (FMN). Also acts on 4-ethoxybenzoate, *N*-methyl-4-aminobenzoate and toluate. The fungal enzyme acts best on veratrate.
References: [305, 3251, 4357]

[EC 1.14.99.15 created 1972]

[1.14.99.16 Transferred entry. methylsterol monooxygenase. Now EC 1.14.13.72, methylsterol monooxygenase]

[EC 1.14.99.16 created 1972, deleted 2002]

[1.14.99.17 Transferred entry. glyceryl-ether monooxygenase. Now EC 1.14.16.5, glyceryl-ether monooxygenase]

[EC 1.14.99.17 created 1972, deleted 1976]

[1.14.99.18 Deleted entry. CMP-*N*-acetylneuraminate monooxygenase]

[EC 1.14.99.18 created 1976, modified 1999, deleted 2003]

[1.14.99.19 Transferred entry. plasmanylethanolamine desaturase. Now classified as EC 1.14.19.77, plasmanylethanolamine desaturase]

[EC 1.14.99.19 created 1976, deleted 2020]

EC 1.14.99.20

Accepted name: phyloquinone monooxygenase (2,3-epoxidizing)
Reaction: phyloquinone + reduced acceptor + O₂ = 2,3-epoxyphyloquinone + acceptor + H₂O
Other name(s): phyloquinone epoxidase; vitamin K 2,3-epoxidase; vitamin K epoxidase; vitamin K₁ epoxidase
Systematic name: phyloquinone,hydrogen-donor:oxygen oxidoreductase (2,3-epoxidizing)
References: [4638]

[EC 1.14.99.20 created 1976]

EC 1.14.99.21

Accepted name: *Latia*-luciferin monooxygenase (demethylating)
Reaction: *Latia* luciferin + reduced acceptor + 2 O₂ = oxidized *Latia* luciferin + CO₂ + formate + acceptor + H₂O + *hν*
Other name(s): luciferase (*Latia* luciferin); *Latia* luciferin monooxygenase (demethylating)
Systematic name: *Latia*-luciferin,hydrogen-donor:oxygen oxidoreductase (demethylating)
Comments: A flavoprotein. *Latia* is a bioluminescent mollusc. The reaction possibly involves two enzymes, an oxygenase followed by a monooxygenase for the actual light-emitting step.
References: [3875, 3877]

[EC 1.14.99.21 created 1976, modified 1982]

EC 1.14.99.22

Accepted name: ecdysone 20-monooxygenase
Reaction: ecdysone + reduced acceptor + O₂ = 20-hydroxyecdysone + acceptor + H₂O
Other name(s): α-ecdysone C-20 hydroxylase; ecdysone 20-hydroxylase
Systematic name: Ecdysone,hydrogen-donor:oxygen oxidoreductase (20-hydroxylating)
Comments: An enzyme from insect fat body or malpighian tubules involving a heme-thiolate protein (*P*-450). NADPH can act as ultimate hydrogen donor.
References: [1935, 3071, 3944]

[EC 1.14.99.22 created 1978]

EC 1.14.99.23

Accepted name: 3-hydroxybenzoate 2-monooxygenase
Reaction: 3-hydroxybenzoate + reduced acceptor + O₂ = 2,3-dihydroxybenzoate + acceptor + H₂O
Other name(s): 3-hydroxybenzoate 2-hydroxylase; 3-HBA-2-hydroxylase
Systematic name: 3-hydroxybenzoate,hydrogen-donor:oxygen oxidoreductase (2-hydroxylating)
References: [836]

[EC 1.14.99.23 created 1984]

EC 1.14.99.24

Accepted name: steroid 9α-monooxygenase
Reaction: pregna-4,9(11)-diene-3,20-dione + reduced acceptor + O₂ = 9,11α-epoxypregn-4-ene-3,20-dione + acceptor + H₂O
Other name(s): steroid 9α-hydroxylase
Systematic name: steroid,hydrogen-donor:oxygen oxidoreductase (9-epoxidizing)
Comments: An enzyme system involving a flavoprotein (FMN) and two iron-sulfur proteins.
References: [4067]

[EC 1.14.99.24 created 1986]

[1.14.99.25 *Transferred entry. linoleoyl-CoA desaturase. Now EC 1.14.19.3, linoleoyl-CoA desaturase*]

[EC 1.14.99.25 created 1986, deleted 2000]

EC 1.14.99.26

Accepted name: 2-hydroxypyridine 5-monooxygenase
Reaction: 2-hydroxypyridine + reduced acceptor + O₂ = 2,5-dihydroxypyridine + acceptor + H₂O
Other name(s): 2-hydroxypyridine oxygenase
Systematic name: 2-hydroxypyridine,hydrogen-donor:oxygen oxidoreductase (5-hydroxylating)
Comments: Also oxidizes 2,5-dihydroxypyridine, but does not act on 3-hydroxypyridine, 4-hydroxypyridine or 2,6-dihydroxypyridine.
References: [3828]

[EC 1.14.99.26 created 1989]

[1.14.99.27 *Transferred entry. juglone 3-monooxygenase, now classified as EC 1.17.3.4, juglone 3-monooxygenase*]

[EC 1.14.99.27 created 1989, deleted 2016]

[1.14.99.28 *Transferred entry. linalool 8-monooxygenase. Now EC 1.14.14.84, linalool 8-monooxygenase*]

[EC 1.14.99.28 created 1989, deleted 2012]

EC 1.14.99.29

Accepted name: deoxyhypusine monooxygenase
Reaction: [eIF5A]-deoxyhypusine + reduced acceptor + O₂ = [eIF5A]-hypusine + acceptor + H₂O
Other name(s): deoxyhypusine hydroxylase; deoxyhypusine dioxygenase
Systematic name: deoxyhypusine,hydrogen-donor:oxygen oxidoreductase (2-hydroxylating)
Comments: The enzyme catalyses the final step in the formation of the amino acid hypusine in the eukaryotic initiation factor 5A.
References: [1]

[EC 1.14.99.29 created 1989]

[1.14.99.30] *Transferred entry. carotene 7,8-desaturase. Now EC 1.3.5.6, 9,9'-dicis-ζ-carotene desaturase.]*

[EC 1.14.99.30 created 1999, deleted 2011]

[1.14.99.31] *Transferred entry. myristoyl-CoA 11-(E) desaturase. Now classified as EC 1.14.19.24, myristoyl-CoA 11-(E) desaturase]*

[EC 1.14.99.31 created 2000, deleted 2015]

[1.14.99.32] *Transferred entry. myristoyl-CoA 11-(Z) desaturase. Now classified as EC 1.14.19.5, acyl-CoA 11-(Z)-desaturase.]*

[EC 1.14.99.32 created 2000, deleted 2015]

[1.14.99.33] *Transferred entry. Δ¹²-fatty acid dehydrogenase. Now EC 1.14.19.39, acyl-lipid Δ¹²-acetylenase]*

[EC 1.14.99.33 created 2000, deleted 2015]

EC 1.14.99.34

Accepted name: monoprenyl isoflavone epoxidase
Reaction: 7-O-methylfluteone + NADPH + H⁺ + O₂ = dihydrofurano derivatives + NADP⁺ + H₂O
Other name(s): monoprenyl isoflavone monooxygenase; 7-O-methylfluteone:O₂ oxidoreductase; 7-O-methylfluteone,NADPH:O₂ oxidoreductase
Systematic name: 7-O-methylfluteone,NADPH:oxygen oxidoreductase
Comments: A flavoprotein (FAD) with high specificity for monoprenyl isoflavone. The product of the prenyl epoxidation reaction contains an oxygen atom derived from O₂, but not from H₂O. It is slowly and non-enzymically converted into the corresponding dihydrofurano derivative. The enzyme in the fungus *Botrytis cinerea* is induced by the substrate analogue, 6-prenylnaringenin.
References: [4195]

[EC 1.14.99.34 created 2000]

EC 1.14.99.35

Accepted name: thiophene-2-carbonyl-CoA monooxygenase
Reaction: thiophene-2-carbonyl-CoA + reduced acceptor + O₂ = 5-hydroxythiophene-2-carbonyl-CoA + acceptor + H₂O
Other name(s): thiophene-2-carboxyl-CoA dehydrogenase; thiophene-2-carboxyl-CoA hydroxylase; thiophene-2-carboxyl-CoA monooxygenase
Systematic name: thiophene-2-carbonyl-CoA, hydrogen-donor:oxygen oxidoreductase
Comments: A molybdenum enzyme. Highly specific for thiophene-2-carbonyl-CoA. Tetrazolium salts can act as electron acceptors.
References: [203]

[EC 1.14.99.35 created 2000]

[1.14.99.36] *Transferred entry. β-carotene 15,15-monooxygenase. Now classified as EC 1.13.11.63, β-carotene 15,15'-dioxygenase.]*

[EC 1.14.99.36 created 1972 as EC 1.13.11.21, transferred 2001 to EC 1.14.99.36, deleted 2015]

[1.14.99.37 Transferred entry. *taxadiene 5 α -hydroxylase*. Now EC 1.14.14.176, *taxadiene 5 α -hydroxylase*]

[EC 1.14.99.37 created 2002, deleted 2020]

EC 1.14.99.38

Accepted name: cholesterol 25-monooxygenase
Reaction: cholesterol + reduced acceptor + O₂ = 25-hydroxycholesterol + acceptor + H₂O
Other name(s): cholesterol 25-hydroxylase (ambiguous)
Systematic name: cholesterol,hydrogen-donor:oxygen oxidoreductase (25-hydroxylating)
Comments: Unlike most other sterol hydroxylases, this enzyme is not a cytochrome *P*-450. Instead, it uses diiron cofactors to catalyse the hydroxylation of hydrophobic substrates [2563]. The diiron cofactor can be either Fe-O-Fe or Fe-OH-Fe and is bound to the enzyme through interactions with clustered histidine or glutamate residues [1153, 3607]. In cell cultures, this enzyme down-regulates cholesterol synthesis and the processing of sterol regulatory element binding proteins (SREBPs). *cf.* EC 1.17.99.10, cholesterol C-25 hydroxylase.
References: [2563, 632, 2561, 1153, 3607]

[EC 1.14.99.38 created 2005, modified 2020]

EC 1.14.99.39

Accepted name: ammonia monooxygenase
Reaction: NH₃ + a reduced acceptor + O₂ = NH₂OH + an acceptor + H₂O
Other name(s): AMO
Systematic name: ammonia,donor:oxygen oxidoreductase (hydroxylamine-producing)
Comments: The enzyme catalyses the first reaction in the pathway of ammonia oxidation to nitrite. It contains copper [1055], iron [4853] and possibly zinc [1325]. The enzyme requires two electrons, which are derived indirectly from the quinone pool via a membrane-bound donor.
References: [1055, 1779, 298, 1699, 4853, 2860, 4610, 137, 1325]

[EC 1.14.99.39 created 2010]

[1.14.99.40 Transferred entry. *5,6-dimethylbenzimidazole synthase*. Now EC 1.13.11.79, *5,6-dimethylbenzimidazole synthase*]

[EC 1.14.99.40 created 2010, deleted 2014]

[1.14.99.41 Transferred entry. *all-trans-8'-apo- β -carotenal 15,15'-oxygenase*. Now EC 1.13.11.75, *all-trans-8'-apo- β -carotenal 15,15'-oxygenase*]

[EC 1.14.99.41 created 2010, deleted 2013]

[1.14.99.42 Transferred entry. *zeaxanthin 7,8-dioxygenase*. Now EC 1.13.11.84, *crocetin dialdehyde synthase*]

[EC 1.14.99.42 created 2011, modified 2014, deleted 2017]

[1.14.99.43 Transferred entry. *β -amyrin 24-hydroxylase*. Now EC 1.14.14.134, *β -amyrin 24-hydroxylase*]

[EC 1.14.99.43 created 2011, deleted 2018]

EC 1.14.99.44

Accepted name: diapolycopene oxygenase
Reaction: 4,4'-diapolycopene + 4 reduced acceptor + 4 O₂ = 4,4'-diapolycopenedial + 4 acceptor + 6 H₂O
Other name(s): *crtP* (ambiguous)
Systematic name: 4,4'-diapolycopene,AH₂:oxygen oxidoreductase (4,4'-hydroxylating)

Comments: Little activity with neurosporene or lycopene. Involved in the biosynthesis of C₃₀ carotenoids such as staphyloxanthin. The enzyme oxidizes each methyl group to the hydroxymethyl and then a dihydroxymethyl group, followed by the spontaneous loss of water to give an aldehyde group.

References: [2802, 4215]

[EC 1.14.99.44 created 2011]

[1.14.99.45 Transferred entry. carotene ϵ -monoxygenase. Now EC 1.14.14.158, carotene ϵ -monoxygenase]

[EC 1.14.99.45 created 2011, deleted 2018]

EC 1.14.99.46

Accepted name: pyrimidine oxygenase

Reaction: (1) uracil + FMNH₂ + O₂ + NADH = (Z)-3-ureidoacrylate + H₂O + FMN + NAD⁺ + H⁺ (overall reaction)

(1a) FMNH₂ + O₂ = FMN-*N*⁵-peroxide

(1b) uracil + FMN-*N*⁵-peroxide = (Z)-3-ureidoacrylate + FMN-*N*⁵-oxide

(1c) FMN-*N*⁵-oxide + NADH = FMN + H₂O + NAD⁺ + H⁺ (spontaneous)

(2) thymine + FMNH₂ + O₂ + NADH = (Z)-2-methylureidoacrylate + H₂O + FMN + NAD⁺ + H⁺ (overall reaction)

(2a) FMNH₂ + O₂ = FMN-*N*⁵-peroxide

(2b) thymine + FMN-*N*⁵-peroxide = (Z)-2-methylureidoacrylate + FMN-*N*⁵-oxide

(2c) FMN-*N*⁵-oxide + NADH = FMN + H₂O + NAD⁺ + H⁺ (spontaneous)

Other name(s): *rutA* (gene name)

Systematic name: uracil,FMNH₂:oxygen oxidoreductase (uracil hydroxylating, ring-opening)

Comments: The enzyme participates in the Rut pyrimidine catabolic pathway. The flavin-*N*⁵-oxide that is formed by the enzyme reacts spontaneously with NADH to give oxidized flavin, releasing a water molecule.

References: [2918, 2101, 20, 19, 2723]

[EC 1.14.99.46 created 2012, modified 2019]

EC 1.14.99.47

Accepted name: (+)-larreatricin hydroxylase

Reaction: (+)-larreatricin + reduced acceptor + O₂ = (+)-3'-hydroxylarreatricin + acceptor + H₂O

Systematic name: (+)-larreatricin:oxygen 3'-hydroxylase

Comments: Isolated from the plant *Larrea tridentata* (creosote bush). The enzyme has a strong preference for the 3' position of (+)-larreatricin.

References: [666]

[EC 1.14.99.47 created 2012]

EC 1.14.99.48

Accepted name: heme oxygenase (staphylobilin-producing)

Reaction: (1) protoheme + 5 reduced acceptor + 4 O₂ = β -staphylobilin + Fe²⁺ + formaldehyde + 5 acceptor + 4 H₂O

(2) protoheme + 5 reduced acceptor + 4 O₂ = δ -staphylobilin + Fe²⁺ + formaldehyde + 5 acceptor + 4 H₂O

Other name(s): haem oxygenase (ambiguous); heme oxygenase (deacylizing) (ambiguous); heme oxidase (ambiguous); haem oxidase (ambiguous); heme oxygenase (ambiguous); *isdG* (gene name); *isdI* (gene name)

Systematic name: protoheme,hydrogen-donor:oxygen oxidoreductase (δ/β -methene-oxidizing, hydroxylating)

Comments: This enzyme, which is found in some pathogenic bacteria, is involved in an iron acquisition system that catabolizes the host's hemoglobin. The two enzymes from the bacterium *Staphylococcus aureus*, encoded by the *isdG* and *isdI* genes, produce 67.5 % and 56.2 % δ -staphylobilin, respectively.

References: [3500, 2700, 4061]

[EC 1.14.99.48 created 2013]

[1.14.99.49 Transferred entry. 2-hydroxy-5-methyl-1-naphthoate 7-hydroxylase. Now EC 1.14.15.31, 2-hydroxy-5-methyl-1-naphthoate 7-hydroxylase]

[EC 1.14.99.49 created 2014, deleted 2018]

EC 1.14.99.50

Accepted name: γ -glutamyl hercynylcysteine S-oxide synthase
Reaction: hercynine + γ -L-glutamyl-L-cysteine + O₂ = γ -L-glutamyl-S-(hercyn-2-yl)-L-cysteine S-oxide + H₂O
Other name(s): EgtB
Systematic name: hercynine, γ -L-glutamyl-L-cysteine:oxygen oxidoreductase [γ -L-glutamyl-S-(hercyn-2-yl)-L-cysteine S-oxide-forming]
Comments: Requires Fe²⁺ for activity. The enzyme, found in bacteria, is specific for both hercynine and γ -L-glutamyl-L-cysteine. It is part of the biosynthesis pathway of ergothioneine.
References: [3777, 3337]

[EC 1.14.99.50 created 2015]

[1.14.99.51 Transferred entry. hercynylcysteine S-oxide synthase, now listed as EC 1.21.3.10, hercynylcysteine S-oxide synthase.]

[EC 1.14.99.51 created 2015, deleted 2021]

EC 1.14.99.52

Accepted name: L-cysteinyl-L-histidinylsulfoxide synthase
Reaction: L-histidine + L-cysteine + O₂ = S-(L-histidin-5-yl)-L-cysteine S-oxide + H₂O
Other name(s): OvoA
Systematic name: L-histidine,L-cysteine:oxygen [S-(L-histidin-5-yl)-L-cysteine S-oxide-forming]
Comments: Requires Fe²⁺ for activity. The enzyme participates in ovothiol biosynthesis. It also has some activity as EC 1.13.11.20, cysteine dioxygenase, and can perform the reaction of EC 1.14.99.50, γ -glutamyl hercynylcysteine sulfoxide synthase, albeit with low activity [3960].
References: [428, 3961, 2680, 3960]

[EC 1.14.99.52 created 2015]

EC 1.14.99.53

Accepted name: lytic chitin monooxygenase
Reaction: [(1 \rightarrow 4)-N-acetyl- β -D-glucosaminy] (m+n) + reduced acceptor + O₂ = [(1 \rightarrow 4)-N-acetyl- β -D-glucosaminy] (m-1)-(1 \rightarrow 4)-2-(acetylamino)-2-deoxy-D-glucono-1,5-lactone + [(1 \rightarrow 4)-N-acetyl- β -D-glucosaminy] _n + acceptor + H₂O
Other name(s): LPMO (ambiguous); CBP21; chitin oxidohydrolase
Systematic name: chitin, hydrogen-donor:oxygen oxidoreductase (N-acetyl- β -D-glucosaminy C1-hydroxylating/C4-dehydrogenating)
Comments: The enzyme cleaves chitin in an oxidative manner, releasing fragments of chitin with an N-acetylamino-D-glucono-1,5-lactone at the reducing end. The initially formed lactone at the reducing end of the shortened chitin chain quickly hydrolyses spontaneously to the aldonic acid. *In vitro* ascorbate can serve as reducing agent. The enzyme contains copper at the active site.
References: [4385, 4384, 1437, 4874]

[EC 1.14.99.53 created 2017]

EC 1.14.99.54

Accepted name: lytic cellulose monooxygenase (C1-hydroxylating)

Reaction: $[(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_{n+m} + \text{reduced acceptor} + \text{O}_2 = [(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_{m-1}\text{-}(1\rightarrow4)\text{-D-glucono-1,5-lactone} + [(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_n + \text{acceptor} + \text{H}_2\text{O}$

Other name(s): lytic polysaccharide monooxygenase (ambiguous); LPMO (ambiguous); LPMO9A

Systematic name: cellulose, hydrogen-donor:oxygen oxidoreductase (D-glucosyl C1-hydroxylating)

Comments: This copper-containing enzyme, found in fungi and bacteria, cleaves cellulose in an oxidative manner. The cellulose fragments that are formed contain a D-glucono-1,5-lactone residue at the reducing end, which hydrolyses quickly and spontaneously to the aldonic acid. The electrons are provided *in vivo* by the cytochrome *b* domain of EC 1.1.99.18, cellobiose dehydrogenase (acceptor) [3313]. Ascorbate can serve as the electron donor *in vitro*.

References: [3313, 268, 2460, 323, 1194, 3253, 752]

[EC 1.14.99.54 created 2017]

EC 1.14.99.55

Accepted name: lytic starch monooxygenase

Reaction: starch + reduced acceptor + $\text{O}_2 = \text{D-glucono-1,5-lactone-terminated malto-oligosaccharides} + \text{short-chain malto-oligosaccharides} + \text{acceptor} + \text{H}_2\text{O}$

Other name(s): LPMO (ambiguous)

Systematic name: starch, hydrogen-donor:oxygen oxidoreductase (D-glucosyl C1-hydroxylating)

Comments: The enzyme cleaves starch in an oxidative manner. It releases fragments of starch with a D-glucono-1,5-lactone at the reducing end. The initially formed $\alpha\text{-D-glucono-1,5-lactone}$ at the reducing end of the shortend amylose chain quickly hydrolyses spontaneously to the aldonic acid. *In vitro* ascorbate has been found to be able to serve as reducing agent. The enzyme contains copper at the active site.

References: [4479, 1437, 2403]

[EC 1.14.99.55 created 2017]

EC 1.14.99.56

Accepted name: lytic cellulose monooxygenase (C4-dehydrogenating)

Reaction: $[(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_{n+m} + \text{reduced acceptor} + \text{O}_2 = 4\text{-dehydro-}\beta\text{-D-glucosyl-}[(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_{n-1} + [(1\rightarrow4)\text{-}\beta\text{-D-glucosyl}]_m + \text{acceptor} + \text{H}_2\text{O}$

Systematic name: cellulose, hydrogen-donor:oxygen oxidoreductase (D-glucosyl 4-dehydrogenating)

Comments: This copper-containing enzyme, found in fungi and bacteria, cleaves cellulose in an oxidative manner. The cellulose fragments that are formed contain a 4-dehydro-D-glucose residue at the non-reducing end. Some enzymes also oxidize cellulose at the C-1 position of the reducing end forming a D-glucono-1,5-lactone residue [*cf.* EC 1.14.99.54, lytic cellulose monooxygenase (C1-hydroxylating)].

References: [268, 2460, 1148, 392, 3253]

[EC 1.14.99.56 created 2017]

EC 1.14.99.57

Accepted name: heme oxygenase (mycobilin-producing)

Reaction: (1) protoheme + 3 reduced acceptor + 3 $\text{O}_2 = \text{mycobilin a} + \text{Fe}^{2+} + 3 \text{ acceptor} + 3 \text{ H}_2\text{O}$
(2) protoheme + 3 reduced acceptor + 3 $\text{O}_2 = \text{mycobilin b} + \text{Fe}^{2+} + 3 \text{ acceptor} + 3 \text{ H}_2\text{O}$

Other name(s): *mhuD* (gene name)

Systematic name: protoheme,donor:oxygen oxidoreductase (mycobilin-producing)

Comments: The enzyme, characterized from the bacterium *Mycobacterium tuberculosis*, is involved in heme degradation and iron utilization. The enzyme binds two stacked protoheme molecules per monomer. Unlike the canonical heme oxygenases, the enzyme does not release carbon monoxide or formaldehyde. Instead, it forms unique products, named mycobilins, that retain the $\alpha\text{-meso-carbon}$ at the ring cleavage site as an aldehyde group. EC 1.6.2.4, NADPH-hemoprotein reductase, can act as electron donor *in vitro*.

References: [653, 3008, 1388]

[EC 1.14.99.57 created 2017]

EC 1.14.99.58

Accepted name: heme oxygenase (biliverdin-IX- β and δ -forming)
Reaction: (1) protoheme + 3 reduced acceptor + 3 O₂ = biliverdin-IX- δ + CO + Fe²⁺ + 3 acceptor + 3 H₂O
(2) protoheme + 3 reduced acceptor + 3 O₂ = biliverdin-IX- β + CO + Fe²⁺ + 3 acceptor + 3 H₂O
Other name(s): *pigA* (gene name)
Systematic name: protoheme,donor:oxygen oxidoreductase (biliverdin-IX- β and δ -forming)
Comments: The enzyme, characterized from the bacterium *Pseudomonas aeruginosa*, differs from EC 1.14.15.20, heme oxygenase (biliverdin-producing, ferredoxin), in that the heme substrate is rotated by approximately 110 degrees within the active site, resulting in cleavage at a different part of the ring. It forms a mixture of about 70% biliverdin-IX- δ and 30% biliverdin-IX- β .
References: [3457, 527, 1181]

[EC 1.14.99.58 created 2017]

EC 1.14.99.59

Accepted name: tryptamine 4-monooxygenase
Reaction: tryptamine + reduced acceptor + O₂ = 4-hydroxytryptamine + acceptor + H₂O
Other name(s): PsiH
Systematic name: tryptamine,hydrogen-donor:oxygen oxidoreductase (4-hydroxylating)
Comments: A cytochrome *P*-450 (heme-thiolate) protein isolated from the fungus *Psilocybe cubensis*. Involved in the biosynthesis of the psychoactive compound psilocybin.
References: [1179]

[EC 1.14.99.59 created 2017]

EC 1.14.99.60

Accepted name: 3-demethoxyubiquinol 3-hydroxylase
Reaction: 6-methoxy-3-methyl-2-(*all-trans*-polyprenyl)-1,4-benzoquinol + a reduced acceptor + O₂ = 3-demethylubiquinol + acceptor + H₂O
Other name(s): 6-methoxy-3-methyl-2-(*all-trans*-polyprenyl)-1,4-benzoquinol 5-hydroxylase; COQ7 (gene name); clk-1 (gene name); *ubiF* (gene name)
Systematic name: 6-methoxy-3-methyl-2-(*all-trans*-polyprenyl)-1,4-benzoquinol,acceptor:oxygen oxidoreductase (5-hydroxylating)
Comments: The enzyme catalyses the last hydroxylation reaction during the biosynthesis of ubiquinone.
References: [2644, 4389, 2320, 4022, 4323]

[EC 1.14.99.60 created 2018]

EC 1.14.99.61

Accepted name: cyclooctat-9-en-7-ol 5-monooxygenase
Reaction: cyclooctat-9-en-7-ol + reduced acceptor + O₂ = cyclooctat-9-ene-5,7-diol + acceptor + H₂O
Other name(s): CotB3
Systematic name: cyclooctat-9-en-7-ol,hydrogen-donor:oxygen oxidoreductase (5-hydroxylating)
Comments: Isolated from the bacterium *Streptomyces melanosporofaciens* M1614-43f2. Involved in the biosynthesis of cyclooctatin.
References: [2107, 1369]

[EC 1.14.99.61 created 2018]

EC 1.14.99.62

Accepted name: cyclooctatin synthase
Reaction: cyclooctat-9-ene-5,7-diol + reduced acceptor + O₂ = cyclooctatin + acceptor + H₂O
Other name(s): CotB4
Systematic name: cyclooctat-9-ene-5,7-diol,hydrogen-donor:oxygen oxidoreductase (18-hydroxylating)
Comments: Isolated from the bacterium *Streptomyces melanosporofaciens* M1614-43f2.
References: [2107, 1369]

[EC 1.14.99.62 created 2018]

EC 1.14.99.63

Accepted name: β-carotene 4-ketolase
Reaction: (1) β-carotene + 2 reduced acceptor + 2 O₂ = echinenone + 2 acceptor + 3 H₂O
(2) echinenone + 2 reduced acceptor + 2 O₂ = canthaxanthin + 2 acceptor + 3 H₂O
Other name(s): BKT (ambiguous); β-C-4 oxygenase; β-carotene ketolase; *crtS* (gene name); *crtW* (gene name)
Systematic name: β-carotene,donor:oxygen oxidoreductase (echinenone-forming)
Comments: The enzyme, studied from algae, plants, fungi, and bacteria, adds an oxo group at position 4 of a carotenoid β ring. It is involved in the biosynthesis of carotenoids such as astaxanthin and flexixanthin. The enzyme does not act on β rings that are hydroxylated at position 3, such as in zeaxanthin (*cf.* EC 1.14.99.64, zeaxanthin 4-ketolase). The enzyme from the yeast *Xanthophyllomyces dendrorhous* is bifunctional and also catalyses the activity of EC 1.14.15.24, β-carotene 3-hydroxylase.
References: [2542, 435, 4017, 3150, 4216, 2015]

[EC 1.14.99.63 created 2018]

EC 1.14.99.64

Accepted name: zeaxanthin 4-ketolase
Reaction: (1) zeaxanthin + 2 reduced acceptor + 2 O₂ = adonixanthin + 2 acceptor + 3 H₂O
(2) adonixanthin + 2 reduced acceptor + 2 O₂ = (3*S*,3'*S*)-astaxanthin + 2 acceptor + 3 H₂O
Other name(s): BKT (ambiguous); *crtW148* (gene name)
Systematic name: zeaxanthin,donor:oxygen oxidoreductase (adonixanthin-forming)
Comments: The enzyme has a similar activity to that of EC 1.14.99.63, β-carotene 4-ketolase, but unlike that enzyme is able to also act on zeaxanthin.
References: [4910, 1754]

[EC 1.14.99.64 created 2018]

EC 1.14.99.65

Accepted name: 4-amino-L-phenylalanyl-[CmlP-peptidyl-carrier-protein] 3-hydroxylase
Reaction: 4-amino-L-phenylalanyl-[CmlP-peptidyl-carrier-protein] + reduced acceptor + O₂ = 2-(4-aminophenyl)-L-seryl-[CmlP-peptidyl-carrier-protein] + acceptor + H₂O
Other name(s): *cmlA* (gene name)
Systematic name: 4-amino-L-phenylalanyl-[CmlP-peptidyl-carrier-protein],acceptor:oxygen 3-oxidoreductase
Comments: The enzyme, characterized from the bacterium *Streptomyces venezuelae*, participates in the biosynthesis of the antibiotic chloramphenicol. It carries an oxygen-bridged dinuclear iron cluster. The native electron donor remains unknown, and the enzyme was assayed *in vitro* using sodium dithionite. The enzyme only acts on its substrate when it is loaded onto the peptidyl-carrier domain of the CmlP non-ribosomal peptide synthase.
References: [2628]

[EC 1.14.99.65 created 2019]

EC 1.14.99.66

- Accepted name:** [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ FAD-dependent demethylase
- Reaction:** a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ + 2 acceptor + 2 H₂O = a [histone H3]-L-lysine⁴ + 2 formaldehyde + 2 reduced acceptor (overall reaction)
(1a) a [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ + acceptor + H₂O = a [histone H3]-*N*⁶-methyl-L-lysine⁴ + formaldehyde + reduced acceptor
(1b) a [histone H3]-*N*⁶-methyl-L-lysine⁴ + acceptor + H₂O = a [histone H3]-L-lysine⁴ + formaldehyde + reduced acceptor
- Other name(s):** KDM1 (gene name); LSD1 (gene name); lysine-specific histone demethylase 1
- Systematic name:** [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴:acceptor oxidoreductase (demethylating)
- Comments:** The enzyme specifically removes methyl groups from mono- and dimethylated lysine⁴ of histone 3. During the reaction the substrate is oxidized by the FAD cofactor of the enzyme to generate the corresponding imine, which is subsequently hydrolysed in the form of formaldehyde. The enzyme is similar to flavin amine oxidases, and differs from all other known histone lysine demethylases, which are iron(II)- and 2-oxoglutarate-dependent dioxygenases. The physiological electron acceptor is not known with certainty. *In vitro* the enzyme can use oxygen, which is reduced to hydrogen peroxide, but generation of hydrogen peroxide in the chromatin environment is unlikely as it will result in oxidative damage of DNA.
- References:** [1144, 1143]

[EC 1.14.99.66 created 2019]

EC 1.14.99.67

- Accepted name:** α -*N*-dichloroacetyl-*p*-aminophenylserinol *N*-oxygenase
- Reaction:** α -*N*-dichloroacetyl-*p*-aminophenylserinol + reduced acceptor + 2 O₂ = chloramphenicol + acceptor + 2 H₂O
- Other name(s):** *cmlI* (gene name)
- Systematic name:** α -*N*-dichloroacetyl-*p*-aminophenylserinol,acceptor:oxygen oxidoreductase (*N*-hydroxylating)
- Comments:** The enzyme, isolated from the bacterium *Streptomyces venezuelae*, is involved in the biosynthesis of the antibiotic chloramphenicol. It contains a carboxylate-bridged binuclear non-heme iron cluster. The components of the native electron chain have not been identified, although the immediate donor is likely to be an iron-sulfur protein. The reaction mechanism involves formation of an extremely stable peroxo intermediate that catalyses three individual two-electron oxidations via a hydroxylamine and a nitroso intermediates without releasing the intermediates. *cf.* EC 1.14.99.68, 4-aminobenzoate *N*-oxygenase.
- References:** [2550, 2629, 2215]

[EC 1.14.99.67 created 2020]

EC 1.14.99.68

- Accepted name:** 4-aminobenzoate *N*-oxygenase
- Reaction:** 4-aminobenzoate + reduced acceptor + 2 O₂ = 4-nitrobenzoate + acceptor + 2 H₂O
- Other name(s):** *aurF* (gene name)
- Systematic name:** 4-aminobenzoate,acceptor:oxygen oxidoreductase (*N*-hydroxylating)
- Comments:** The enzyme, characterized from the bacterium *Streptomyces thioluteus*, catalyses an early step in the biosynthesis of the antibiotic aureothin. It contains a carboxylate-bridged binuclear non-heme iron cluster. The native electron donor has not been identified, but is likely an iron-sulfur protein. The reaction mechanism involves formation of an extremely stable peroxo intermediate that catalyses three two-electron oxidations via a hydroxylamine and dihydroxylamine intermediates. *cf.* EC 1.14.99.67, *N*-[1-(4-aminophenyl)-1,3-dihydroxypropan-2-yl]-2,2-dichloroacetamide *N*-oxygenase.
- References:** [1589, 2385, 4935, 675, 2230, 2452]

[EC 1.14.99.68 created 2020]

EC 1.14.99.69

- Accepted name:** tRNA 2-(methylsulfanyl)-*N*⁶-isopentenyladenosine³⁷ hydroxylase
- Reaction:** 2-(methylsulfanyl)-*N*⁶-prenyladenosine³⁷ in tRNA + reduced acceptor + O₂ = *N*⁶-[(2*E*)-4-hydroxy-3-methylbut-2-en-1-yl]-2-(methylsulfanyl)adenosine³⁷ in tRNA + acceptor + H₂O
- Other name(s):** *miaE* (gene name); tRNA 2-methylthio-*N*⁶-isopentenyl adenosine(37) hydroxylase; tRNA 2-(methylsulfanyl)-*N*⁶-dimethylallyl-adenosine³⁷ hydroxylase
- Systematic name:** tRNA 2-(methylsulfanyl)-*N*⁶-prenyladenosine³⁷,donor:oxygen 4-oxidoreductase (*trans*-hydroxylating)
- Comments:** The enzyme, found only within a small subset of facultative anaerobic bacteria, belongs to the non-heme diiron family. The enzyme from *Salmonella typhimurium* was shown to catalyse a stereoselective (*E*)-hydroxylation at the terminal C⁴-position of the prenyl group.
- References:** [3294, 3295, 734]

[EC 1.14.99.69 created 2020]

EC 1.15 Acting on superoxide as acceptor

This subclass contains enzymes that act on superoxide as acceptor in a single sub-subclass (EC 1.15.1).

EC 1.15.1 Acting on superoxide as acceptor (only sub-subclass identified to date)

EC 1.15.1.1

- Accepted name:** superoxide dismutase
- Reaction:** 2 superoxide + 2 H⁺ = O₂ + H₂O₂
- Other name(s):** superoxidase dismutase; copper-zinc superoxide dismutase; Cu-Zn superoxide dismutase; ferrisuperoxide dismutase; superoxide dismutase I; superoxide dismutase II; SOD; Cu,Zn-SOD; Mn-SOD; Fe-SOD; SODF; SODS; SOD-1; SOD-2; SOD-3; SOD-4; hemocuprein; erythrocuprein; cytocuprein; cuprein; hepatocuprein
- Systematic name:** superoxide:superoxide oxidoreductase
- Comments:** A metalloprotein; also known as erythrocuprein, hemocuprein or cytocuprein. Enzymes from most eukaryotes contain both copper and zinc; those from mitochondria and most prokaryotes contain manganese or iron.
- References:** [2053, 3686, 4417]

[EC 1.15.1.1 created 1972]

EC 1.15.1.2

- Accepted name:** superoxide reductase
- Reaction:** superoxide + reduced rubredoxin + 2 H⁺ = H₂O₂ + oxidized rubredoxin
- Other name(s):** neelaredoxin; desulfoferrodoxin
- Systematic name:** rubredoxin:superoxide oxidoreductase
- Comments:** The enzyme contains non-heme iron.
- References:** [1901, 4785, 2535, 6]

[EC 1.15.1.2 created 2001 as EC 1.18.96.1, transferred 2001 to EC 1.15.1.2]

EC 1.16 Oxidizing metal ions

This subclass contains enzymes that oxidize metal ions (donors) to a higher valency state. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.16.1), oxygen (EC 1.16.3) and flavin (EC 1.16.8).

EC 1.16.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.16.1.1

Accepted name: mercury(II) reductase
Reaction: $\text{Hg} + \text{NADP}^+ + \text{H}^+ = \text{Hg}^{2+} + \text{NADPH}$
Other name(s): mercuric reductase; mercurate(II) reductase; mercuric ion reductase; mercury reductase; reduced NADP:mercuric ion oxidoreductase; mer A
Systematic name: Hg:NADP⁺ oxidoreductase
Comments: A dithiol enzyme.
References: [1154, 1155]

[EC 1.16.1.1 created 1984]

EC 1.16.1.2

Accepted name: diferric-transferrin reductase
Reaction: $\text{transferrin}[\text{Fe}(\text{II})]_2 + \text{NAD}^+ + \text{H}^+ = \text{transferrin}[\text{Fe}(\text{III})]_2 + \text{NADH}$
Other name(s): diferric transferrin reductase; NADH diferric transferrin reductase; transferrin reductase
Systematic name: transferrin[Fe(II)]₂:NAD⁺ oxidoreductase
References: [2547]

[EC 1.16.1.2 created 1989]

[1.16.1.3 Deleted entry. aquacobalamin reductase. This entry has been deleted since no specific enzyme catalysing this activity has been identified and it has been shown that aquacobalamin is efficiently reduced by free dihydroflavins and by non-specific reduced flavoproteins.]

[EC 1.16.1.3 created 1972 as EC 1.6.99.8, transferred 2002 to EC 1.16.1.3, modified 2020, deleted 2020]

[1.16.1.4 Deleted entry. cob(II)alamin reductase. This entry has been deleted since no specific enzyme catalysing this activity has been identified and it has been shown that cob(II)alamin is efficiently reduced by free dihydroflavins and by non-specific reduced flavoproteins]

[EC 1.16.1.4 created 1972 as EC 1.6.99.9, transferred 2002 to EC 1.16.1.4, deleted 2021]

[1.16.1.5 Deleted entry. aquacobalamin reductase (NADPH). This entry has been deleted since the enzyme the entry was based on was later shown to be EC 1.2.1.51, pyruvate dehydrogenase (NADP⁺). On the other hand, it has been shown that non-enzymatic reduction of cob(III)alamin to cob(II)alamin occurs efficiently in the presence of free dihydroflavins or non-specific reduced flavoproteins.]

[EC 1.16.1.5 created 1989 as EC 1.6.99.11, transferred 2002 to EC 1.16.1.5, modified 2020, deleted 2020]

EC 1.16.1.6

Accepted name: cyanocobalamin reductase
Reaction: $2 \text{cob}(\text{II})\text{alamin} - [\text{cyanocobalamin reductase}] + 2 \text{hydrogen cyanide} + \text{NADP}^+ = 2 \text{cyanocob}(\text{III})\text{alamin} + 2 [\text{cyanocobalamin reductase}] + \text{NADPH} + \text{H}^+$
Other name(s): MMACHC (gene name); CblC; cyanocobalamin reductase (NADPH, cyanide-eliminating); cyanocobalamin reductase (NADPH, CN-eliminating); NADPH:cyanocob(III)alamin oxidoreductase (cyanide-eliminating); cob(I)alamin, cyanide:NADP⁺ oxidoreductase; cyanocobalamin reductase (cyanide-eliminating)
Systematic name: cob(II)alamin, hydrogen cyanide:NADP⁺ oxidoreductase

Comments: The mammalian enzyme, which is cytosolic, can bind internalized cyanocobalamin and process it to cob(II)alamin by removing the upper axial ligand. The product remains bound to the protein, which, together with its interacting partner MMADHC, transfers it directly to downstream enzymes involved in adenosylcobalamin and methylcobalamin biosynthesis. In addition to its decyanase function, the mammalian enzyme also catalyses an entirely different chemical reaction with alkylcobalamins, using the thiolate of glutathione for nucleophilic displacement, generating cob(I)alamin and the corresponding glutathione thioether (*cf.* EC 2.5.1.151, alkylcobalamin dealkylase).

References: [4551, 2095, 2249, 2612]

[EC 1.16.1.6 created 1989 as EC 1.6.99.12, transferred 2002 to EC 1.16.1.6, modified 2018, modified 2021]

EC 1.16.1.7

Accepted name: ferric-chelate reductase (NADH)
Reaction: $2 \text{ Fe(II)-siderophore} + \text{NAD}^+ + \text{H}^+ = 2 \text{ Fe(III)-siderophore} + \text{NADH}$
Other name(s): ferric chelate reductase (ambiguous); iron chelate reductase (ambiguous); NADH:Fe³⁺-EDTA reductase; NADH₂:Fe³⁺ oxidoreductase; *ferB* (gene name); Fe(II):NAD⁺ oxidoreductase
Systematic name: Fe(II)-siderophore:NAD⁺ oxidoreductase
Comments: Contains FAD. The enzyme catalyses the reduction of bound ferric iron in a variety of iron chelators (siderophores), resulting in the release of ferrous iron. The plant enzyme is involved in the transport of iron across plant plasma membranes. The enzyme from the bacterium *Paracoccus denitrificans* can also reduce chromate. *cf.* EC 1.16.1.9, ferric-chelate reductase (NADPH) and EC 1.16.1.10, ferric-chelate reductase [NAD(P)H].
References: [147, 468, 469, 487, 3654, 2737]

[EC 1.16.1.7 created 1992 as EC 1.6.99.13, transferred 2002 to EC 1.16.1.7, modified 2011, modified 2014]

EC 1.16.1.8

Accepted name: [methionine synthase] reductase
Reaction: $2 \text{ [methionine synthase]-methylcob(III)alamin} + 2 \text{ S-adenosyl-L-homocysteine} + \text{NADP}^+ = 2 \text{ [methionine synthase]-cob(II)alamin} + \text{NADPH} + \text{H}^+ + 2 \text{ S-adenosyl-L-methionine}$
Other name(s): methionine synthase cob(II)alamin reductase (methylating); methionine synthase reductase; [methionine synthase]-cobalamin methyltransferase (cob(II)alamin reducing); [methionine synthase]-methylcob(I)alamin,S-adenosylhomocysteine:NADP⁺ oxidoreductase
Systematic name: [methionine synthase]-methylcob(III)alamin,S-adenosyl-L-homocysteine:NADP⁺ oxidoreductase
Comments: In humans, the enzyme is a flavoprotein containing FAD and FMN. The substrate of the enzyme is the inactivated cobalt(II) form of EC 2.1.1.13, methionine synthase. Electrons are transferred from NADPH to FAD to FMN. Defects in this enzyme lead to hereditary hyperhomocysteinemia.
References: [2375, 3177, 3178]

[EC 1.16.1.8 created 1999 as EC 2.1.1.135, transferred 2003 to EC 1.16.1.8, modified 2020]

EC 1.16.1.9

Accepted name: ferric-chelate reductase (NADPH)
Reaction: $2 \text{ Fe(II)-siderophore} + \text{NADP}^+ + \text{H}^+ = 2 \text{ Fe(III)-siderophore} + \text{NADPH}$
Other name(s): ferric chelate reductase (ambiguous); iron chelate reductase (ambiguous); NADPH:Fe³⁺-EDTA reductase; NADPH-dependent ferric reductase; *yqjH* (gene name); Fe(II):NADP⁺ oxidoreductase
Systematic name: Fe(II)-siderophore:NADP⁺ oxidoreductase
Comments: Contains FAD. The enzyme, which is widespread among bacteria, catalyses the reduction of ferric iron bound to a variety of iron chelators (siderophores), including ferric triscatecholates and ferric dicitrate, resulting in the release of ferrous iron. The enzyme from the bacterium *Escherichia coli* has the highest efficiency with the hydrolysed ferric enterobactin complex ferric *N*-(2,3-dihydroxybenzoyl)-L-serine [2796]. *cf.* EC 1.16.1.7, ferric-chelate reductase (NADH) and EC 1.16.1.10, ferric-chelate reductase [NAD(P)H].

References: [204, 4533, 2796]

[EC 1.16.1.9 created 1992 as EC 1.6.99.13, transferred 2002 to EC 1.16.1.7, transferred 2011 to EC 1.16.1.9, modified 2012, modified 2014]

EC 1.16.1.10

Accepted name: ferric-chelate reductase [NAD(P)H]
Reaction: $2 \text{ Fe(II)-siderophore} + \text{NAD(P)}^+ + \text{H}^+ = 2 \text{ Fe(III)-siderophore} + \text{NAD(P)H}$
Other name(s): ferric reductase (ambiguous)
Systematic name: Fe(II)-siderophore:NAD(P)⁺ oxidoreductase
Comments: A flavoprotein. The enzyme catalyses the reduction of bound ferric iron in a variety of iron chelators (siderophores), resulting in the release of ferrous iron. The enzyme from the hyperthermophilic archaeon *Archaeoglobus fulgidus* is not active with uncomplexed Fe(III). *cf.* EC 1.16.1.7, ferric-chelate reductase (NADH) and EC 1.16.1.9, ferric-chelate reductase (NADPH).
References: [4387, 660]

[EC 1.16.1.10 created 2014]

EC 1.16.3 With oxygen as acceptor

EC 1.16.3.1

Accepted name: ferroxidase
Reaction: $4 \text{ Fe(II)} + 4 \text{ H}^+ + \text{O}_2 = 4 \text{ Fe(III)} + 2 \text{ H}_2\text{O}$
Other name(s): ceruloplasmin; caeruloplasmin; ferroxidase I; iron oxidase; iron(II):oxygen oxidoreductase; ferro:O₂ oxidoreductase; iron II:oxygen oxidoreductase; hephaestin; HEPH
Systematic name: Fe(II):oxygen oxidoreductase
Comments: The enzyme in blood plasma (ceruloplasmin) belongs to the family of multicopper oxidases. In humans it accounts for 95% of plasma copper. It oxidizes Fe(II) to Fe(III), which allows the subsequent incorporation of the latter into proteins such as apotransferrin and lactoferrin. An enzyme from iron oxidizing bacterium strain TI-1 contains heme *a*.
References: [3193, 3194, 2499, 4174, 629]

[EC 1.16.3.1 created 1972, modified 2011]

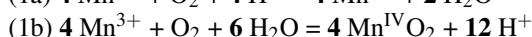
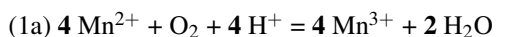
EC 1.16.3.2

Accepted name: bacterial non-heme ferritin
Reaction: $4 \text{ Fe(II)} + \text{O}_2 + 6 \text{ H}_2\text{O} = 4 [\text{FeO(OH)}] + 8 \text{ H}^+$ (overall reaction)
(1a) $2 \text{ Fe(II)} + \text{O}_2 + 4 \text{ H}_2\text{O} = 2 [\text{FeO(OH)}] + 4 \text{ H}^+ + \text{H}_2\text{O}_2$
(1b) $2 \text{ Fe(II)} + \text{H}_2\text{O}_2 + 2 \text{ H}_2\text{O} = 2 [\text{FeO(OH)}] + 4 \text{ H}^+$
Other name(s): FtnA; HuHF
Systematic name: Fe(II):oxygen oxidoreductase ([FeO(OH)]core-producing)
Comments: Ferritins are intracellular iron-storage and detoxification proteins found in all kingdoms of life. They are formed from two subunits that co-assemble in various ratios to form a spherical protein shell. Thousands of mineralized iron atoms are stored within the core of the structure. The product of dioxygen reduction by the bacterial non-heme ferritin is hydrogen peroxide, which is consumed in a subsequent reaction.
References: [1762, 4033, 405]

[EC 1.16.3.2 created 2014]

EC 1.16.3.3

Accepted name: manganese oxidase
Reaction: $4 \text{ Mn}^{2+} + 2 \text{ O}_2 + 4 \text{ H}_2\text{O} = 4 \text{ Mn}^{\text{IV}}\text{O}_2 + 8 \text{ H}^+$ (overall reaction)



Other name(s): *mnxG* (gene name); *mofA* (gene name); *moxA* (gene name); *cotA* (gene name)

Systematic name: manganese(II):oxygen oxidoreductase

Comments: The enzyme, which belongs to the multicopper oxidase family, is found in many bacterial strains. It oxidizes soluble manganese(II) to insoluble manganese(IV) oxides. Since the enzyme is localized to the outer surface of the cell, its activity usually results in encrustation of the cells by the oxides. The physiological function of bacterial manganese(II) oxidation remains unclear.

References: [740, 1162, 3518, 1310, 4085]

[EC 1.16.3.3 created 2017]

EC 1.16.3.4

Accepted name: cuproxidase

Reaction: $4 \text{Cu}^+ + 4 \text{H}^+ + \text{O}_2 = 4 \text{Cu}^{2+} + 2 \text{H}_2\text{O}$

Other name(s): *cueO* (gene name); cuprous oxidase; Cu(I) oxidase; copper efflux oxidase

Systematic name: copper(I):oxygen oxidoreductase

Comments: The enzyme, characterized from the bacterium *Escherichia coli*, is involved in copper tolerance under aerobic conditions. The enzyme contains a substrate binding (type 1) copper site and a trinuclear copper center (consisting of type 2 and type 3 copper sites) in which oxygen binding and reduction takes place. It also contains a methionine rich region that can bind additional copper ions. *In vitro*, if the substrate binding site is occupied by copper(II), the enzyme can function as a laccase-type quinol oxidase (EC 1.10.3.2). However, *in vivo* this site is occupied by a copper(I) ion and the enzyme functions as a cuprous oxidase.

References: [2091, 1384, 3217, 3537, 3538, 3911, 1260, 929, 3912, 742]

[EC 1.16.3.4 created 2021]

EC 1.16.5 With a quinone or similar compound as acceptor

[1.16.5.1 *Transferred entry. ascorbate ferrireductase (transmembrane). Now EC 7.2.1.3, ascorbate ferrireductase (transmembrane)*]

[EC 1.16.5.1 created 2011, deleted 2018]

EC 1.16.8 With a flavin as acceptor

[1.16.8.1 *Deleted entry. cob(II)yrinic acid a,c-diamide reductase. This activity is now known to be catalyzed by EC 2.5.1.17, corrinoid adenosyltransferase*]

[EC 1.16.8.1 created 2004, deleted 2019]

EC 1.16.9 With a copper protein as acceptor

EC 1.16.9.1

Accepted name: iron:rusticyanin reductase

Reaction: $\text{Fe(II)} + \text{rusticyanin} = \text{Fe(III)} + \text{reduced rusticyanin}$

Other name(s): Cyc2 (ambiguous)

Systematic name: Fe(II):rusticyanin oxidoreductase

Comments: Contains *c*-type heme. The enzyme in *Acidithiobacillus ferrooxidans* is a component of an electron transfer chain from Fe(II), comprising this enzyme, the copper protein rusticyanin, cytochrome *c*₄, and cytochrome *c* oxidase (EC 7.1.1.9).

References: [348, 118, 4778, 4777, 4165, 573, 3411]

[EC 1.16.9.1 created 2011 as EC 1.16.98.1, transferred 2011 to EC 1.16.9.1]

EC 1.16.98 With other, known, physiological acceptors

[1.16.98.1 Transferred entry. Now EC 1.16.9.1 iron:rusticyanin reductase]

[EC 1.16.98.1 created 2011, deleted 2011]

EC 1.16.99 With unknown physiological acceptors

EC 1.16.99.1

Accepted name: [Co(II) methylated amine-specific corrinoid protein] reductase

Reaction: (1) ATP + a [Co(II) methylamine-specific corrinoid protein] + reduced acceptor + H₂O = ADP + phosphate + a [Co(I) methylamine-specific corrinoid protein] + acceptor
(2) ATP + a [Co(II) dimethylamine-specific corrinoid protein] + reduced acceptor + H₂O = ADP + phosphate + a [Co(I) dimethylamine-specific corrinoid protein] + acceptor
(3) ATP + a [Co(II) trimethylamine-specific corrinoid protein] + reduced acceptor + H₂O = ADP + phosphate + a [Co(I) trimethylamine-specific corrinoid protein] + acceptor

Systematic name: acceptor:[cobalt(II) methylated amines-specific corrinoid protein] oxidoreductase (ATP-hydrolysing)

Comments: Methyltrophic corrinoid proteins must have the cobalt atom in the active cobalt(I) state to become methylated. Because the cobalt(I)/cobalt(II) transformation has a very low redox potential the corrinoid cofactor is subject to adventitious oxidation to the cobalt(II) state, which renders the proteins inactive. This enzyme, characterized from the methanogenic archaeon *Methanosarcina barkeri*, reduces cobalt(II) back to cobalt(I), restoring activity. The enzyme acts on the corrinoid proteins involved in methanogenesis from methylamine, dimethylamine, and trimethylamine, namely MtmC, MtbC, and MttC, respectively. While *in vitro* the enzyme can use Ti(III)-citrate as the electron donor, the *in vivo* donor is not known. The enzyme from *Methanosarcina barkeri* contains a C-terminal [4Fe-4S] ferredoxin-like domain.

References: [1104, 989]

[EC 1.16.99.1 created 2021]

EC 1.17 Acting on CH or CH₂ groups

This subclass contains enzymes that oxidize the -CH₂- group of donors to -CHOH- (or -CH- to -COH-) and the oxidative cleavage of HC- bonds (as in formate); in the reverse direction, those acting on sugars are involved in the formation of deoxysugars. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.17.1), oxygen (EC 1.17.3), a cytochrome (EC 1.17.2), a disulfide (EC 1.17.4), a quinone or similar compound (EC 1.17.5), another, known, physiological acceptors (EC 1.17.98) or an unknown, physiological acceptor (EC 1.17.99).

EC 1.17.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.17.1.1

Accepted name: CDP-4-dehydro-6-deoxyglucose reductase

Reaction: CDP-4-dehydro-3,6-dideoxy-D-glucose + NAD(P)⁺ + H₂O = CDP-4-dehydro-6-deoxy-D-glucose + NAD(P)H + H⁺

Other name(s): CDP-4-keto-6-deoxyglucose reductase; cytidine diphospho-4-keto-6-deoxy-D-glucose reductase; cytidine diphosphate 4-keto-6-deoxy-D-glucose-3-dehydrogenase; CDP-4-keto-deoxy-glucose reductase; CDP-4-keto-6-deoxy-D-glucose-3-dehydrogenase system; NAD(P)H:CDP-4-keto-6-deoxy-D-glucose oxidoreductase

Systematic name: CDP-4-dehydro-3,6-dideoxy-D-glucose:NAD(P)⁺ 3-oxidoreductase

Comments: The enzyme consists of two proteins. One forms an enzyme-bound adduct of the CDP-4-dehydro-6-deoxyglucose with pyridoxamine phosphate, in which the 3-hydroxy group has been removed. The second catalyses the reduction of this adduct by NAD(P)H and release of the CDP-4-dehydro-3,6-dideoxy-D-glucose and pyridoxamine phosphate.

References: [3234, 3590, 2515]

[EC 1.17.1.1 created 1972, modified 2005]

[1.17.1.2 *Transferred entry. 4-hydroxy-3-methylbut-2-enyl diphosphate reductase, now classified as EC 1.17.7.4, 4-hydroxy-3-methylbut-2-enyl diphosphate reductase.*]

[EC 1.17.1.2 created 2003, modified 2009, deleted 2016]

EC 1.17.1.3

Accepted name: leucoanthocyanidin reductase

Reaction: (2*R*,3*S*)-catechin + NADP⁺ + H₂O = 2,3-*trans*-3,4-*cis*-leucocyanidin + NADPH + H⁺

Other name(s): leucocyanidin reductase

Systematic name: (2*R*,3*S*)-catechin:NADP⁺ 4-oxidoreductase

Comments: The enzyme catalyses the synthesis of catechin, catechin-4β-ol (leucocyanidin) and the related flavan-3-ols afzelechin and gallocatechin, which are initiating monomers in the synthesis of plant polymeric proanthocyanidins or condensed tannins. While 2,3-*trans*-3,4-*cis*-leucocyanidin is the preferred flavan-3,4-diol substrate, 2,3-*trans*-3,4-*cis*-leucodelphinidin and 2,3-*trans*-3,4-*cis*-leucopelargonidin can also act as substrates, but more slowly. NADH can replace NADPH but is oxidized more slowly.

References: [4212, 4211]

[EC 1.17.1.3 created 2003]

EC 1.17.1.4

Accepted name: xanthine dehydrogenase

Reaction: xanthine + NAD⁺ + H₂O = urate + NADH + H⁺

Other name(s): NAD⁺-xanthine dehydrogenase; xanthine-NAD⁺ oxidoreductase; xanthine/NAD⁺ oxidoreductase; xanthine oxidoreductase

Systematic name: xanthine:NAD⁺ oxidoreductase

Comments: Acts on a variety of purines and aldehydes, including hypoxanthine. The mammalian enzyme can also convert *all-trans* retinol to *all-trans*-retinoate, while the substrate is bound to a retinoid-binding protein [4167]. The enzyme from eukaryotes contains [2Fe-2S], FAD and a molybdenum centre. The mammalian enzyme predominantly exists as the NAD-dependent dehydrogenase (EC 1.17.1.4). During purification the enzyme is largely converted to an O₂-dependent form, xanthine oxidase (EC 1.17.3.2). The conversion can be triggered by several mechanisms, including the oxidation of cysteine thiols to form disulfide bonds [2,6,8,15] [which can be catalysed by EC 1.8.4.7, enzyme-thiol transhydrogenase (glutathione-disulfide) in the presence of glutathione disulfide] or limited proteolysis, which results in irreversible conversion. The conversion can also occur *in vivo* [2,7,15].

References: [238, 741, 3249, 3436, 3945, 1797, 1048, 3634, 3246, 1786, 1054, 4335, 1654, 4167, 3081]

[EC 1.17.1.4 created 1972 as EC 1.2.1.37, transferred 1984 to EC 1.1.1.204, modified 1989, transferred 2004 to EC 1.17.1.4, modified 2011]

EC 1.17.1.5

Accepted name: nicotinate dehydrogenase

Reaction: nicotinate + H₂O + NADP⁺ = 6-hydroxynicotinate + NADPH + H⁺

Other name(s): nicotinic acid hydroxylase; nicotinate hydroxylase
Systematic name: nicotinate:NADP⁺ 6-oxidoreductase (hydroxylating)
Comments: A flavoprotein containing non-heme iron. The enzyme is capable of acting on a variety of nicotinate analogues to varying degrees, including pyrazine-2-carboxylate, pyrazine 2,3-dicarboxylate, trigonelline and 6-methylnicotinate. The enzyme from *Clostridium barkeri* also possesses a catalytically essential, labile selenium that can be removed by reaction with cyanide.
References: [1690, 1335, 1334, 916, 915, 2965]

[EC 1.17.1.5 created 1972 as EC 1.5.1.13, transferred 2004 to EC 1.17.1.5]

[1.17.1.6 *Transferred entry. bile-acid 7 α -dehydroxylase. Now EC 1.17.99.5, bile-acid 7 α -dehydroxylase. It is now known that FAD is the acceptor and not NAD⁺ as was thought previously*]

[EC 1.17.1.6 created 2005, deleted 2006]

[1.17.1.7 *Transferred entry. 3-oxo-5,6-dehydrosuberil-CoA semialdehyde dehydrogenase. Now EC 1.2.1.91, 3-oxo-5,6-dehydrosuberil-CoA semialdehyde dehydrogenase*]

[EC 1.17.1.7 created 2011, deleted 2014]

EC 1.17.1.8

Accepted name: 4-hydroxy-tetrahydrodipicolinate reductase
Reaction: (S)-2,3,4,5-tetrahydropyridine-2,6-dicarboxylate + NAD(P)⁺ + H₂O = (2S,4S)-4-hydroxy-2,3,4,5-tetrahydrodipicolinate + NAD(P)H + H⁺
Other name(s): dihydrodipicolinate reductase (incorrect); dihydrodipicolinic acid reductase (incorrect); 2,3,4,5-tetrahydrodipicolinate:NAD(P)⁺ oxidoreductase (incorrect); *dapB* (gene name)
Systematic name: (S)-2,3,4,5-tetrahydropyridine-2,6-dicarboxylate:NAD(P)⁺ 4-oxidoreductase
Comments: The substrate of the enzyme was initially thought to be (S)-2,3-dihydrodipicolinate [1090], and the enzyme was classified accordingly as EC 1.3.1.26, dihydrodipicolinate reductase. Later studies of the enzyme from the bacterium *Escherichia coli* have suggested that the actual substrate of the enzyme is (2S,4S)-4-hydroxy-2,3,4,5-tetrahydrodipicolinate, and that its activity includes a dehydration step [890], and thus the enzyme has been reclassified as 4-hydroxy-tetrahydrodipicolinate reductase. However, the identity of the substrate is still controversial, as more recently it has been suggested that it may be (S)-2,3-dihydrodipicolinate after all [2000].
References: [1090, 890, 2000]

[EC 1.17.1.8 created 1976 as EC 1.3.1.26, transferred 2013 to EC 1.17.1.8, modified 2020]

EC 1.17.1.9

Accepted name: formate dehydrogenase
Reaction: formate + NAD⁺ = CO₂ + NADH
Other name(s): formate-NAD⁺ oxidoreductase; FDH I; FDH II; N-FDH; formic hydrogen-lyase; formate hydrogenlyase; hydrogenlyase; NAD⁺-linked formate dehydrogenase; NAD⁺-dependent formate dehydrogenase; formate dehydrogenase (NAD⁺); NAD⁺-formate dehydrogenase; formate benzyl-viologen oxidoreductase; formic acid dehydrogenase
Systematic name: formate:NAD⁺ oxidoreductase
Comments: The enzyme from most aerobic organisms is devoid of redox-active centres but that from the proteobacterium *Methylosinus trichosporium* contains iron-sulfur centres, flavin and a molybdenum centre [1941]. Together with EC 1.12.1.2 hydrogen dehydrogenase, forms a system previously known as formate hydrogenlyase.
References: [841, 3412, 1941]

[EC 1.17.1.9 created 1961 as EC 1.2.1.2, transferred 2017 to EC 1.17.1.9]

EC 1.17.1.10

- Accepted name:** formate dehydrogenase (NADP⁺)
Reaction: formate + NADP⁺ = CO₂ + NADPH
Other name(s): NADP⁺-dependent formate dehydrogenase
Systematic name: formate:NADP⁺ oxidoreductase
Comments: A tungsten-selenium-iron protein characterized from the bacterium *Moorella thermoacetica*. It is extremely sensitive to oxygen.
References: [98, 4740]

[EC 1.17.1.10 created 1978 as EC 1.2.1.43, transferred 2017 to EC 1.17.1.10]

EC 1.17.1.11

- Accepted name:** formate dehydrogenase (NAD⁺, ferredoxin)
Reaction: 2 formate + NAD⁺ + 2 oxidized ferredoxin [iron-sulfur] cluster = 2 CO₂ + NADH + H⁺ + 2 reduced ferredoxin [iron-sulfur] cluster
Other name(s): electron-bifurcating formate dehydrogenase
Systematic name: formate:NAD⁺, ferredoxin oxidoreductase
Comments: The enzyme complex, isolated from the bacterium *Gottschalkia acidurici*, couples the reduction of NAD⁺ and the reduction of ferredoxin with formate via flavin-based electron bifurcation.
References: [4529]

[EC 1.17.1.11 created 2015 as EC 1.2.1.93, transferred 2017 to EC 1.17.1.11]

EC 1.17.2 With a cytochrome as acceptor

EC 1.17.2.1

- Accepted name:** nicotinate dehydrogenase (cytochrome)
Reaction: nicotinate + a ferricytochrome + H₂O = 6-hydroxynicotinate + a ferrocyclochrome + 2 H⁺
Other name(s): nicotinic acid hydroxylase; nicotinate hydroxylase
Systematic name: nicotinate:cytochrome 6-oxidoreductase (hydroxylating)
Comments: This two-component enzyme from *Pseudomonas* belongs to the family of xanthine dehydrogenases, but differs from most other members of this family. While most members contain an FAD cofactor, the large subunit of this enzyme contains three *c*-type cytochromes, enabling it to interact with the electron transfer chain, probably by delivering the electrons to a cytochrome oxidase. The small subunit contains a typical molybdopterine cytosine dinucleotide (MCD) cofactor and two [2Fe-2S] clusters [1915].
References: [1915, 4769]

[EC 1.17.2.1 created 2010]

EC 1.17.2.2

- Accepted name:** lupanine 17-hydroxylase (cytochrome *c*)
Reaction: lupanine + 2 ferricytochrome *c* + H₂O = 17-hydroxylupanine + 2 ferrocyclochrome *c* + 2 H⁺
Other name(s): lupanine dehydrogenase (cytochrome *c*)
Systematic name: lupanine:cytochrome *c*-oxidoreductase (17-hydroxylating)
Comments: The enzyme isolated from *Pseudomonas putida* contains heme *c* and requires pyrroloquinoline quinone (PQQ) for activity
References: [1718, 1717]

[EC 1.17.2.2 created 2012]

EC 1.17.2.3

- Accepted name:** formate dehydrogenase (cytochrome-*c*-553)
Reaction: formate + 2 ferricytochrome *c*-553 = CO₂ + 2 ferrocyclochrome *c*-553 + H⁺
Systematic name: formate:ferricytochrome-*c*-553 oxidoreductase
Comments: The enzyme has been characterized from the bacterium *Desulfovibrio vulgaris*. *In vitro*, yeast cytochrome *c*, ferricyanide and phenazine methosulfate can act as acceptors.
References: [4719, 4720]

[EC 1.17.2.3 created 1981 as EC 1.2.2.3, transferred 2017 to EC 1.17.2.3]

EC 1.17.3 With oxygen as acceptor

EC 1.17.3.1

- Accepted name:** pteridine oxidase
Reaction: 2-amino-4-hydroxypteridine + O₂ = 2-amino-4,7-dihydroxypteridine + (?)
Systematic name: 2-amino-4-hydroxypteridine:oxygen oxidoreductase (7-hydroxylating)
Comments: Different from EC 1.17.3.2 xanthine oxidase; does not act on hypoxanthine.
References: [4801]

[EC 1.17.3.1 created 1983]

EC 1.17.3.2

- Accepted name:** xanthine oxidase
Reaction: xanthine + H₂O + O₂ = urate + H₂O₂
Other name(s): hypoxanthine oxidase; hypoxanthine:oxygen oxidoreductase; Schardinger enzyme; xanthine oxidoreductase; hypoxanthine-xanthine oxidase; xanthine:O₂ oxidoreductase; xanthine:xanthine oxidase
Systematic name: xanthine:oxygen oxidoreductase
Comments: An iron-molybdenum flavoprotein (FAD) containing [2Fe-2S] centres. Also oxidizes hypoxanthine, some other purines and pterins, and aldehydes, but is distinct from EC 1.2.3.1, aldehyde oxidase. Under some conditions the product is mainly superoxide rather than peroxide: RH + H₂O + 2 O₂ = ROH + 2 O₂^{·-} + 2 H⁺. The mammalian enzyme predominantly exists as an NAD-dependent dehydrogenase (EC 1.17.1.4, xanthine dehydrogenase). During purification the enzyme is largely converted to the O₂-dependent xanthine oxidase form (EC 1.17.3.2). The conversion can be triggered by several mechanisms, including the oxidation of cysteine thiols to form disulfide bonds [4,5,7,10] [which can be catalysed by EC 1.8.4.7, enzyme-thiol transhydrogenase (glutathione-disulfide) in the presence of glutathione disulfide] or limited proteolysis, which results in irreversible conversion. The conversion can also occur *in vivo* [4,6,10].
References: [161, 238, 429, 741, 1797, 1048, 3634, 562, 1022, 3081]

[EC 1.17.3.2 created 1961 as EC 1.2.3.2, transferred 1984 to EC 1.1.3.22, modified 1989, transferred 2004 to EC 1.17.3.2, modified 2011]

EC 1.17.3.3

- Accepted name:** 6-hydroxynicotinate dehydrogenase
Reaction: 6-hydroxynicotinate + H₂O + O₂ = 2,6-dihydroxynicotinate + H₂O₂
Other name(s): 6-hydroxynicotinic acid hydroxylase; 6-hydroxynicotinic acid dehydrogenase; 6-hydroxynicotinate hydroxylase; 6-hydroxynicotinate:O₂ oxidoreductase
Systematic name: 6-hydroxynicotinate:oxygen oxidoreductase
Comments: Contains [2Fe-2S] iron-sulfur centres, FAD and molybdenum. It also has a catalytically essential, labile selenium that can be removed by reaction with cyanide. In *Bacillus niacini*, this enzyme is required for growth on nicotinic acid.
References: [2964, 2965]

[EC 1.17.3.3 created 2004]

EC 1.17.3.4

- Accepted name:** juglone 3-hydroxylase
Reaction: $2 \text{ juglone} + \text{O}_2 = 2 \text{ 3,5-dihydroxy-1,4-naphthoquinone}$ (overall reaction)
(1a) $2 \text{ juglone} + 2 \text{ H}_2\text{O} = 2 \text{ naphthalene-1,2,4,8-tetrol}$
(1b) $2 \text{ naphthalene-1,2,4,8-tetrol} + \text{O}_2 = 2 \text{ 3,5-dihydroxy-1,4-naphthoquinone} + 2 \text{ H}_2\text{O}$
Other name(s): juglone hydroxylase; naphthoquinone hydroxylase; naphthoquinone-hydroxylase
Systematic name: 5-hydroxy-1,4-naphthoquinone,water:oxygen oxidoreductase (3-hydroxylating)
Comments: Even though oxygen is consumed, molecular oxygen is not incorporated into the product. Catalysis starts by incorporation of an oxygen atom from a water molecule into the substrate. The naphthalene-1,2,4,8-tetrol intermediate is then oxidized by molecular oxygen, which is reduced to water. Also acts on 1,4-naphthoquinone, naphthazarin and 2-chloro-1,4-naphthoquinone.
References: [3504]

[EC 1.17.3.4 created 1989 as EC 1.14.99.27, transferred 2016 to EC 1.17.3.4]

EC 1.17.4 With a disulfide as acceptor

EC 1.17.4.1

- Accepted name:** ribonucleoside-diphosphate reductase
Reaction: $2' \text{-deoxyribonucleoside } 5' \text{-diphosphate} + \text{thioredoxin disulfide} + \text{H}_2\text{O} = \text{ribonucleoside } 5' \text{-diphosphate} + \text{thioredoxin}$
Other name(s): ribonucleotide reductase (ambiguous); CDP reductase; ribonucleoside diphosphate reductase; UDP reductase; ADP reductase; nucleoside diphosphate reductase; ribonucleoside $5'$ -diphosphate reductase; ribonucleotide diphosphate reductase; $2'$ -deoxyribonucleoside-diphosphate:oxidized-thioredoxin $2'$ -oxidoreductase; RR; *nrdB* (gene name); *nrdF* (gene name); *nrdJ* (gene name)
Systematic name: $2'$ -deoxyribonucleoside- $5'$ -diphosphate:thioredoxin-disulfide $2'$ -oxidoreductase
Comments: This enzyme is responsible for the *de novo* conversion of ribonucleoside diphosphates into deoxyribonucleoside diphosphates, which are essential for DNA synthesis and repair. There are three types of this enzyme differing in their cofactors. Class Ia enzymes contain a diiron(III)-tyrosyl radical, class Ib enzymes contain a dimanganese-tyrosyl radical, and class II enzymes contain adenosylcobalamin. In all cases the cofactors are involved in generation of a transient thiyl (sulfanyl) radical on a cysteine residue, which attacks the substrate, forming a ribonucleotide $3'$ -radical, followed by water loss to form a ketyl (α -oxoalkyl) radical. The ketyl radical is reduced to $3'$ -keto-deoxynucleotide concomitant with formation of a disulfide anion radical between two cysteine residues. A proton-coupled electron-transfer from the disulfide radical to the substrate generates a $3'$ -deoxynucleotide radical, and the final product is formed when the hydrogen atom that was initially removed from the $3'$ -position of the nucleotide by the thiyl radical is returned to the same position. The disulfide bridge is reduced by the action of thioredoxin. *cf.* EC 1.1.98.6, ribonucleoside-triphosphate reductase (formate) and EC 1.17.4.2, ribonucleoside-triphosphate reductase (thioredoxin).
References: [2357, 2358, 2873, 2356, 2335, 4078, 2414, 2368, 3404]

[EC 1.17.4.1 created 1972, modified 2017]

EC 1.17.4.2

- Accepted name:** ribonucleoside-triphosphate reductase (thioredoxin)
Reaction: $2' \text{-deoxyribonucleoside } 5' \text{-triphosphate} + \text{thioredoxin disulfide} + \text{H}_2\text{O} = \text{ribonucleoside } 5' \text{-triphosphate} + \text{thioredoxin}$
Other name(s): ribonucleotide reductase (ambiguous); $2'$ -deoxyribonucleoside-triphosphate:oxidized-thioredoxin $2'$ -oxidoreductase
Systematic name: $2'$ -deoxyribonucleoside- $5'$ -triphosphate:thioredoxin-disulfide $2'$ -oxidoreductase

Comments: The enzyme, characterized from the bacterium *Lactobacillus leichmannii*, is similar to class II ribonucleoside-diphosphate reductase (*cf.* EC 1.17.4.1). However, it is specific for the triphosphate versions of its substrates. The enzyme contains an adenosylcobalamin cofactor that is involved in generation of a transient thiyl (sulfanyl) radical on a cysteine residue. This radical attacks the substrate, forming a ribonucleotide 3'-radical, followed by water loss to form a ketyl (α -oxoalkyl) radical. The ketyl radical is reduced to 3'-keto-deoxynucleotide concomitant with formation of a disulfide anion radical between two cysteine residues. A proton-coupled electron-transfer from the disulfide radical to the substrate generates a 3'-deoxynucleotide radical, and the final product is formed when the hydrogen atom that was initially removed from the 3'-position of the nucleotide by the thiyl radical is returned to the same position. The disulfide bridge is reduced by the action of thioredoxin. *cf.* EC 1.1.98.6, ribonucleoside-triphosphate reductase (formate).

References: [350, 1377, 4077, 145, 2370, 2478]

[EC 1.17.4.2 created 1972, modified 2017]

[1.17.4.3 *Transferred entry. 4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase. As ferredoxin and not protein-disulfide is now known to take part in the reaction, the enzyme has been transferred to EC 1.17.7.1, (E)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase.*]

[EC 1.17.4.3 created 2003, deleted 2009]

EC 1.17.4.4

Accepted name: vitamin-K-epoxide reductase (warfarin-sensitive)
Reaction: (1) phyloquinone + a protein with a disulfide bond + H₂O = 2,3-epoxyphyloquinone + a protein with reduced L-cysteine residues
(2) phyloquinol + a protein with a disulfide bond = phyloquinone + a protein with reduced L-cysteine residues
Other name(s): VKORC1 (gene name); VKORC1L1 (gene name)
Systematic name: phyloquinone:disulfide oxidoreductase
Comments: The enzyme catalyses the reduction of vitamin K 2,3-epoxide, which is formed by the activity of EC 4.1.1.90, peptidyl-glutamate 4-carboxylase, back to its phyloquinol active form. The enzyme forms a tight complex with EC 5.3.4.1, protein disulfide-isomerase, which transfers the required electrons from newly-synthesized proteins by catalysing the formation of disulfide bridges. The enzyme acts on the epoxide forms of both phyloquinone (vitamin K₁) and menaquinone (vitamin K₂). Inhibited strongly by (*S*)-warfarin and ferulenol.
References: [4609, 2387, 2917, 2457, 4490, 3988, 3752]

[EC 1.17.4.4 created 1989 as EC 1.1.4.1, transferred 2014 to EC 1.17.4.4, modified 2018]

EC 1.17.4.5

Accepted name: vitamin-K-epoxide reductase (warfarin-insensitive)
Reaction: 3-hydroxy-2-methyl-3-phytyl-2,3-dihydro-1,4-naphthoquinone + oxidized dithiothreitol = 2,3-epoxy-2-methyl-3-phytyl-2,3-dihydro-1,4-naphthoquinone + 1,4-dithiothreitol
Systematic name: 3-hydroxy-2-methyl-3-phytyl-2,3-dihydronaphthoquinone:oxidized-dithiothreitol oxidoreductase
Comments: Vitamin K 2,3-epoxide is reduced to 3-hydroxy- (and 2-hydroxy-) vitamin K by 1,4-dithiothreitol, which is oxidized to a disulfide. Not inhibited by warfarin [*cf.* EC 1.17.4.4, vitamin-K-epoxide reductase (warfarin-sensitive)].
References: [2917]

[EC 1.17.4.5 created 1989 as EC 1.1.4.2, transferred 2014 to EC 1.17.4.5]

EC 1.17.5 With a quinone or similar compound as acceptor

EC 1.17.5.1

- Accepted name:** phenylacetyl-CoA dehydrogenase
Reaction: phenylacetyl-CoA + H₂O + 2 quinone = phenylglyoxylyl-CoA + 2 quinol
Other name(s): phenylacetyl-CoA:acceptor oxidoreductase
Systematic name: phenylacetyl-CoA:quinone oxidoreductase
Comments: The enzyme from *Thauera aromatica* is a membrane-bound molybdenum—iron—sulfur protein. The enzyme is specific for phenylacetyl-CoA as substrate. Phenylacetate, acetyl-CoA, benzoyl-CoA, propanoyl-CoA, crotonyl-CoA, succinyl-CoA and 3-hydroxybenzoyl-CoA cannot act as substrates. The oxygen atom introduced into the product, phenylglyoxylyl-CoA, is derived from water and not molecular oxygen. Duroquinone, menaquinone and 2,6-dichlorophenolindophenol (DCPIP) can act as acceptor, but the likely physiological acceptor is ubiquinone [3508]. A second enzyme, EC 3.1.2.25, phenylacetyl-CoA hydrolase, converts the phenylglyoxylyl-CoA formed into phenylglyoxylate.
References: [3508, 3737]

[EC 1.17.5.1 created 2004]

EC 1.17.5.2

- Accepted name:** caffeine dehydrogenase
Reaction: caffeine + ubiquinone + H₂O = 1,3,7-trimethylurate + ubiquinol
Systematic name: caffeine:ubiquinone oxidoreductase
Comments: This enzyme, characterized from the soil bacterium *Pseudomonas* sp. CBB1, catalyses the incorporation of an oxygen atom originating from a water molecule into position C-8 of caffeine. The enzyme utilizes short-tail ubiquinones as the preferred electron acceptor.
References: [4832]

[EC 1.17.5.2 created 2010]

EC 1.17.5.3

- Accepted name:** formate dehydrogenase-N
Reaction: formate + a quinone = CO₂ + a quinol
Other name(s): Fdh-N; FdnGHI; nitrate-inducible formate dehydrogenase; formate dehydrogenase N; FDH-N; nitrate inducible Fdn; nitrate inducible formate dehydrogenase
Systematic name: formate:quinone oxidoreductase
Comments: The enzyme contains molybdopterin-guanine dinucleotides, five [4Fe-4S] clusters and two heme *b* groups. Formate dehydrogenase-N oxidizes formate in the periplasm, transferring electrons via the menaquinone pool in the cytoplasmic membrane to a dissimilatory nitrate reductase (EC 1.7.5.1), which transfers electrons to nitrate in the cytoplasm. The system generates proton motive force under anaerobic conditions [1950].
References: [1052, 1951, 1950]

[EC 1.17.5.3 created 2010 as EC 1.1.5.6, transferred 2017 to EC 1.17.5.3]

EC 1.17.7 With an iron-sulfur protein as acceptor

EC 1.17.7.1

- Accepted name:** (*E*)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase (ferredoxin)
Reaction: (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate + H₂O + 2 oxidized ferredoxin = 2-*C*-methyl-D-erythritol 2,4-cyclodiphosphate + 2 reduced ferredoxin
Other name(s): 4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase (ambiguous); (*E*)-4-hydroxy-3-methylbut-2-en-1-yl-diphosphate:protein-disulfide oxidoreductase (hydrating) (incorrect); (*E*)-4-hydroxy-3-methylbut-2-enyl diphosphate synthase (ambiguous); *gcpE* (gene name); ISPG (gene name); (*E*)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase

Systematic name: (*E*)-4-hydroxy-3-methylbut-2-en-1-yl-diphosphate:oxidized ferredoxin oxidoreductase
Comments: An iron-sulfur protein found in plant chloroplasts and cyanobacteria that contains a [4Fe-4S] cluster [3152]. Forms part of an alternative non-mevalonate pathway for isoprenoid biosynthesis. Bacteria have a similar enzyme that uses flavodoxin rather than ferredoxin (*cf.* EC 1.17.7.3). The enzyme from the plant *Arabidopsis thaliana* is active with photoreduced 5-deazaflavin but not with flavodoxin [3152].
References: [3152, 3784, 3783, 3782]

[EC 1.17.7.1 created 2003 as EC 1.17.4.3, transferred 2009 to EC 1.17.7.1, modified 2014]

EC 1.17.7.2

Accepted name: 7-hydroxymethyl chlorophyll *a* reductase
Reaction: chlorophyll *a* + H₂O + 2 oxidized ferredoxin = 7¹-hydroxychlorophyll *a* + 2 reduced ferredoxin + 2 H⁺
Other name(s): HCAR; 7¹-hydroxychlorophyll-*a*:ferredoxin oxidoreductase
Systematic name: chlorophyll-*a*:ferredoxin oxidoreductase
Comments: Contains FAD and an iron-sulfur center. This enzyme, which is present in plant chloroplasts, carries out the second step in the conversion of chlorophyll *b* to chlorophyll *a*. It similarly reduces chlorophyllide *a*.
References: [2760]

[EC 1.17.7.2 created 2011]

EC 1.17.7.3

Accepted name: (*E*)-4-hydroxy-3-methylbut-2-en-1-yl-diphosphate synthase (flavodoxin)
Reaction: (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate + H₂O + oxidized flavodoxin = 2-*C*-methyl-D-erythritol 2,4-cyclodiphosphate + reduced flavodoxin
Other name(s): 4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase (ambiguous); (*E*)-4-hydroxy-3-methylbut-2-en-1-yl-diphosphate:protein-disulfide oxidoreductase (hydrating) (incorrect); (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate synthase (ambiguous); *ispG* (gene name)
Systematic name: (*E*)-4-hydroxy-3-methylbut-2-en-1-yl-diphosphate:oxidized flavodoxin oxidoreductase
Comments: A bacterial iron-sulfur protein that contains a [4Fe-4S] cluster. Forms part of an alternative non-mevalonate pathway for isoprenoid biosynthesis that is found in most bacteria [4871]. Plants and cyanobacteria have a similar enzyme that utilizes ferredoxin rather than flavodoxin (*cf.* EC 1.17.7.1).
References: [1596, 4871, 3391]

[EC 1.17.7.3 created 2014]

EC 1.17.7.4

Accepted name: 4-hydroxy-3-methylbut-2-en-1-yl diphosphate reductase
Reaction: (1) 3-methylbut-3-en-1-yl diphosphate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O = (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺
(2) prenyl diphosphate + 2 oxidized ferredoxin [iron-sulfur] cluster + H₂O = (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate + 2 reduced ferredoxin [iron-sulfur] cluster + 2 H⁺
Other name(s): isopentenyl-diphosphate:NADP⁺ oxidoreductase; LytB; (*E*)-4-hydroxy-3-methylbut-2-en-1-yl diphosphate reductase; HMBPP reductase; IspH; LytB/IspH; isopentenyl-diphosphate:ferredoxin oxidoreductase
Systematic name: 3-methylbut-3-en-1-yl-diphosphate:ferredoxin oxidoreductase
Comments: An iron-sulfur protein that contains either a [3Fe-4S] [1389] or a [4Fe-4S] [4657] cluster. This enzyme forms a system with a ferredoxin or a flavodoxin and an NAD(P)H-dependent reductase. This is the last enzyme in the non-mevalonate pathway for isoprenoid biosynthesis. This pathway, also known as the 1-deoxy-D-xylulose 5-phosphate (DOXP) or as the 2-*C*-methyl-D-erythritol-4-phosphate (MEP) pathway, is found in most bacteria and in plant chloroplasts. The enzyme acts in the reverse direction, producing a 5:1 mixture of 3-methylbut-3-en-1-yl diphosphate and prenyl diphosphate.

References: [3555, 1664, 609, 3556, 4657, 1389]

[EC 1.17.7.4 created 2003 as EC 1.17.1.2, modified 2009, transferred 2016 to EC 1.17.7.4]

EC 1.17.8 With a flavin as acceptor

EC 1.17.8.1

Accepted name: hydroxysqualene dehydroxylase
Reaction: squalene + FAD + H₂O = hydroxysqualene + FADH₂
Other name(s): *hpnE* (gene name)
Systematic name: squalene:FAD oxidoreductase (hydroxylating)
Comments: This enzyme, isolated from the bacteria *Rhodopseudomonas palustris* and *Zymomonas mobilis*, participates, along with EC 2.5.1.103, presqualene diphosphate synthase, and EC 4.2.3.156, hydroxysqualene synthase, in the conversion of *all-trans*-farnesyl diphosphate to squalene. Eukaryotes achieve the same goal in a single step, catalysed by EC 2.5.1.21, squalene synthase.
References: [3228]

[EC 1.17.8.1 created 2016]

EC 1.17.9 With a copper protein as acceptor

EC 1.17.9.1

Accepted name: 4-methylphenol dehydrogenase (hydroxylating)
Reaction: 4-methylphenol + 4 oxidized azurin + H₂O = 4-hydroxybenzaldehyde + 4 reduced azurin + 4 H⁺ (overall reaction)
(1a) 4-methylphenol + 2 oxidized azurin + H₂O = 4-hydroxybenzyl alcohol + 2 reduced azurin + 2 H⁺
(1b) 4-hydroxybenzyl alcohol + 2 oxidized azurin = 4-hydroxybenzaldehyde + 2 reduced azurin + 2 H⁺
Other name(s): *pchCF* (gene names); *p*-cresol-(acceptor) oxidoreductase (hydroxylating); *p*-cresol methylhydroxylase; 4-cresol dehydrogenase (hydroxylating)
Systematic name: 4-methylphenol:oxidized azurin oxidoreductase (methyl-hydroxylating)
Comments: This bacterial enzyme contains a flavin (FAD) subunit and a cytochrome *c* subunit. The flavin subunit abstracts two hydrogen atoms from the substrate, forming a quinone methide intermediate, then hydrates the latter at the benzylic carbon with a hydroxyl group derived from water. The protons are lost to the bulk solvent, while the electrons are passed to the heme on the cytochrome subunit, and from there to azurin, a small copper-binding protein that is co-localized with the enzyme in the periplasm. The first hydroxylation forms 4-hydroxybenzyl alcohol; a second hydroxylation converts this into 4-hydroxybenzaldehyde.
References: [1719, 2750, 1716, 401, 3481, 3298, 1922]

[EC 1.17.9.1 created 1983 as EC 1.17.99.1, modified 2001, modified 2011, modified 2015, transferred 2018 to EC 1.17.9.1]

EC 1.17.9.2

Accepted name: (+)-pinoselin hydroxylase
Reaction: (+)-pinoselin + 2 oxidized azurin + H₂O = (+)-6-hydroxypinoselin + 2 reduced azurin + 2 H⁺
Other name(s): pinoselin α -hydroxylase; *pinAB* (gene names)
Systematic name: (+)-pinoselin:azurin oxidoreductase
Comments: Contains FAD. This enzyme, characterized from the bacterium *Pseudomonas* sp. SG-MS2, catalyses the incorporation of an oxygen atom originating from a water molecule into position C-6 of the lignan (+)-pinoselin. The enzyme consists of a flavoprotein subunit and a *c*-type cytochrome subunit. Electrons that originate in the substrate are transferred via the FAD cofactor and the cytochrome subunit to the blue-copper protein azurin.

References: [3848]

[EC 1.17.9.2 created 2020]

EC 1.17.98 With other, known, physiological acceptors

[1.17.98.1 Deleted entry. bile-acid 7 α -dehydroxylase. Now known to be catalyzed by multiple enzymes.]

[EC 1.17.98.1 created 2005 as EC 1.17.1.6, transferred 2006 to EC 1.17.99.5, transferred 2014 to EC 1.17.98.1, deleted 2016]

EC 1.17.98.2

Accepted name: bacteriochlorophyllide *c* C-7¹-hydroxylase
Reaction: 2 *S*-adenosyl-L-methionine + a bacteriochlorophyllide *c* + H₂O = a bacteriochlorophyllide *e* + 2 5'-deoxyadenosine + 2 L-methionine (overall reaction)
(1a) *S*-adenosyl-L-methionine + a bacteriochlorophyllide *c* + H₂O = a 7-(hydroxymethyl)bacteriochlorophyllide *c* + 5'-deoxyadenosine + L-methionine
(1b) *S*-adenosyl-L-methionine + a 7-(hydroxymethyl)bacteriochlorophyllide *c* + H₂O = a 7-(dihydroxymethyl)bacteriochlorophyllide *c* + 5'-deoxyadenosine + L-methionine
(1c) a 7-(dihydroxymethyl)bacteriochlorophyllide *c* = a bacteriochlorophyllide *e* + H₂O (spontaneous)
Other name(s): *bciD* (gene name)
Systematic name: bacteriochlorophyllide-*c*:*S*-adenosyl-L-methionine oxidoreductase (C-7¹-hydroxylating)
Comments: The enzyme, found in green sulfur bacteria (Chlorobiaceae), is a radical *S*-adenosyl-L-methionine (AdoMet) enzyme and contains a [4Fe-4S] cluster. It catalyses two consecutive hydroxylation reactions of the C-7 methyl group of bacteriochlorophyllide *c* to form a geminal diol intermediate that spontaneously dehydrates to produce the formyl group of bacteriochlorophyllide *e*.
References: [1526, 4281]

[EC 1.17.98.2 created 2016, modified 2017]

EC 1.17.98.3

Accepted name: formate dehydrogenase (coenzyme F₄₂₀)
Reaction: formate + oxidized coenzyme F₄₂₀ = CO₂ + reduced coenzyme F₄₂₀
Other name(s): coenzyme F₄₂₀ reducing formate dehydrogenase; coenzyme F₄₂₀-dependent formate dehydrogenase
Systematic name: formate:coenzyme-F₄₂₀ oxidoreductase
Comments: The enzyme, characterized from methanogenic archaea, is involved in formate-dependent H₂ production. It contains noncovalently bound FAD [3703].
References: [3703, 3704, 2568]

[EC 1.17.98.3 created 2014 as EC 1.2.99.9, transferred 2017 to EC 1.17.98.3]

EC 1.17.98.4

Accepted name: formate dehydrogenase (hydrogenase)
Reaction: formate + an [oxidized hydrogenase] = CO₂ + a [reduced hydrogenase]
Other name(s): FDHH; FDH-H; FDH-O; formate dehydrogenase H; formate dehydrogenase O
Systematic name: formate:[oxidized hydrogenase] oxidoreductase
Comments: Formate dehydrogenase H is a cytoplasmic enzyme that oxidizes formate without oxygen transfer, transferring electrons to a hydrogenase. The two enzymes form the formate-hydrogen lyase complex [164]. The enzyme contains an [4Fe-4S] cluster, a selenocysteine residue and a molybdopterin cofactor [164].
References: [164, 1333, 2079]

[EC 1.17.98.4 created 2010 as EC 1.1.99.33, transferred 2018 to EC 1.17.99.7, transferred 2020 to 1.17.98.4.]

EC 1.17.99 With unknown physiological acceptors

[1.17.99.1 Transferred entry. 4-methylphenol dehydrogenase (hydroxylating). Now EC 1.17.9.1, 4-methylphenol dehydrogenase (hydroxylating)]

[EC 1.17.99.1 created 1983, modified 2001, modified 2011, modified 2015, deleted 2018]

EC 1.17.99.2

Accepted name: ethylbenzene hydroxylase
Reaction: ethylbenzene + H₂O + acceptor = (S)-1-phenylethanol + reduced acceptor
Other name(s): ethylbenzene dehydrogenase; ethylbenzene:(acceptor) oxidoreductase
Systematic name: ethylbenzene:acceptor oxidoreductase
Comments: Involved in the anaerobic catabolism of ethylbenzene by denitrifying bacteria. Ethylbenzene is the preferred substrate; the enzyme from some strains oxidizes propylbenzene, 1-ethyl-4-fluorobenzene, 3-methylpent-2-ene and ethylidenecyclohexane. Toluene is not oxidized. *p*-Benzoquinone or ferrocenium can act as electron acceptor. Contains molybdopterin, [4Fe-4S] clusters and heme *b*.
References: [2167, 1931]

[EC 1.17.99.2 created 2001]

EC 1.17.99.3

Accepted name: 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanoyl-CoA 24-hydroxylase
Reaction: (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oyl-CoA + H₂O + acceptor = (24*R*,25*R*)-3 α ,7 α ,12 α ,24-tetrahydroxy-5 β -cholestan-26-oyl-CoA + reduced acceptor
Other name(s): trihydroxycoprostanoyl-CoA oxidase; THC-CoA oxidase; THCA-CoA oxidase; 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanoyl-CoA oxidase; 3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oate 24-hydroxylase
Systematic name: (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oyl-CoA:acceptor 24-oxidoreductase (24*R*-hydroxylating)
Comments: Requires ATP. The reaction in mammals possibly involves dehydrogenation to give a 24(25)-double bond followed by hydration [1455]. However, in amphibians such as the Oriental fire-bellied toad (*Bombina orientalis*), it is probable that the product is formed via direct hydroxylation of the saturated side chain of (25*R*)-3 α ,7 α ,12 α -trihydroxy-5 β -cholestan-26-oate and not via hydration of a 24(25) double bond [3277]. In microsomes, the free acid is preferred to the coenzyme A ester, whereas in mitochondria, the coenzyme A ester is preferred to the free-acid form of the substrate [1455].
References: [1455, 3711, 911, 912, 3277, 3607]

[EC 1.17.99.3 created 2005]

EC 1.17.99.4

Accepted name: uracil/thymine dehydrogenase
Reaction: (1) uracil + H₂O + acceptor = barbiturate + reduced acceptor
(2) thymine + H₂O + acceptor = 5-methylbarbiturate + reduced acceptor
Other name(s): uracil oxidase; uracil-thymine oxidase; uracil dehydrogenase
Systematic name: uracil:acceptor oxidoreductase
Comments: Forms part of the oxidative pyrimidine-degrading pathway in some microorganisms, along with EC 3.5.2.1 (barbiturase) and EC 3.5.1.95 (*N*-malonylurea hydrolase). Mammals, plants and other microorganisms utilize the reductive pathway, comprising EC 1.3.1.1 [dihydrouracil dehydrogenase (NAD⁺)] or EC 1.3.1.2 [dihydropyrimidine dehydrogenase (NADP⁺)], EC 3.5.2.2 (dihydropyrimidinase) and EC 3.5.1.6 (β -ureidopropionase), with the ultimate degradation products being an L-amino acid, NH₃ and CO₂ [3968].
References: [1572, 4534, 4535, 2349, 3968]

[EC 1.17.99.4 created 1961 as EC 1.2.99.1, transferred 1984 to EC 1.1.99.19, transferred 2006 to EC 1.17.99.4]

[1.17.99.5 Transferred entry. bile-acid 7 α -dehydroxylase. Now classified as EC 1.17.98.1, bile-acid 7 α -dehydroxylase.]

[EC 1.17.99.5 created 2005 as EC 1.17.1.6, transferred 2006 to EC 1.17.99.5, deleted 2014]

EC 1.17.99.6

Accepted name: epoxyqueuosine reductase
Reaction: queuosine³⁴ in tRNA + acceptor + H₂O = epoxyqueuosine³⁴ in tRNA + reduced acceptor
Other name(s): oQ reductase; *queG* (gene name); *queH* (gene name)
Systematic name: queuosine³⁴ in tRNA:acceptor oxidoreductase
Comments: This enzyme catalyses the last step in the bacterial biosynthetic pathway to queuosine, the modified guanosine base in the wobble position in tRNAs specific for Tyr, His, Asp or Asn. Two types of enzymes are known to catalyse this activity. QueG enzymes are cobalamin-dependent, while QueH enzymes contain a [4Fe-4S] metallocluster along with an adjacent coordinated iron metal.
References: [2807, 4856, 2453]

[EC 1.17.99.6 created 2014]

[1.17.99.7 Transferred entry. formate dehydrogenase (acceptor). Now classified as EC 1.17.98.4, formate dehydrogenase (hydrogenase).]

[EC 1.17.99.7 created 2010 as EC 1.1.99.33, transferred 2017 to EC 1.17.99.7, deleted 2020]

EC 1.17.99.8

Accepted name: limonene dehydrogenase
Reaction: (1) (*S*)-limonene + H₂O + acceptor = (–)-perillyl alcohol + reduced acceptor
(2) (*R*)-limonene + H₂O + acceptor = (+)-perillyl alcohol + reduced acceptor
Other name(s): *ctmAB* (gene names)
Systematic name: limonene:acceptor oxidoreductase (7-hydroxylating)
Comments: Contains FAD. The enzyme, characterized from the bacterium *Castellaniella defragrans* 65Phen, hydroxylates the *R*- and *S*-enantiomers at a similar rate. The *in vivo* electron acceptor may be a heterodimeric electron transfer flavoprotein (ETF).
References: [3297, 3392]

[EC 1.17.99.8 created 2020]

EC 1.17.99.9

Accepted name: heme *a* synthase
Reaction: ferroheme *o* + H₂O + 2 acceptor = ferroheme *a* + 2 reduced acceptor (overall reaction)
(1a) ferroheme *o* + H₂O + acceptor = ferroheme *i* + reduced acceptor
(1b) ferroheme *i* + H₂O + acceptor = hydroxyferroheme *i* + reduced acceptor
(1c) hydroxyferroheme *i* = ferroheme *a* + H₂O (spontaneous)
Other name(s): COX15 (gene name); *ctaA* (gene name)
Systematic name: ferroheme *o*:acceptor C-8¹-oxidoreductase (heme *a*-forming)
Comments: Contains a heme *b* cofactor. The enzyme catalyses the conversion of heme *o* to heme *a* by two successive hydroxylations of the methyl group at C-8, using water as the oxygen source. The first hydroxylation forms heme *i*, the second hydroxylation results in an unstable dihydroxymethyl group, which spontaneously dehydrates, resulting in the formyl group of heme *a* [461, 1602]. The electrons produced by the reaction are transferred to a heme *b* cofactor [3091]. However, the electron acceptor that is used to restore the heme *b* cofactor to its oxidized state is still not known (both a thioredoxin-like protein and menaquinol have been proposed).
References: [226, 461, 462, 1602, 1601, 3091]

[EC 1.17.99.9 created 2020]

EC 1.17.99.10

- Accepted name:** steroid C-25 hydroxylase
Reaction: cholest-4-en-3-one + acceptor + H₂O = 25-hydroxycholest-4-en-3-one + reduced acceptor
Other name(s): s25dA1 (gene name); s25dA1B3 (gene name); s25dA1C3 (gene name); cholesterol C-25 dehydrogenase; steroid C-25 dehydrogenase
Systematic name: cholest-4-en-3-one:acceptor oxidoreductase (25-hydroxylating)
Comments: The enzyme, characterized from the bacterium *Sterolibacterium denitrificans*, participates in the anaerobic degradation of cholesterol. The enzyme can accept several substrates including vitamin D₃. The enzyme contains a bis(guanylyl molybdopterin) cofactor, five [Fe-S] clusters, and one heme *b*. *cf.* EC 1.14.99.38, cholesterol 25-monooxygenase, an oxygen-requiring eukaryotic enzyme that catalyses a similar transformation.
References: [886, 3600, 3601, 1874]

[EC 1.17.99.10 created 2020]

EC 1.17.99.11

- Accepted name:** 3-oxo- Δ^1 -steroid hydratase/dehydrogenase
Reaction: a 3-oxo- Δ^1 -steroid + H₂O + acceptor = a steroid 1,3-dione + reduced acceptor (overall reaction)
(1a) a 3-oxo- Δ^1 -steroid + H₂O = a 1-hydroxy-3-oxo-steroid
(1b) a 1-hydroxy-3-oxo-steroid + acceptor = a steroid 1,3-dione + reduced acceptor
Other name(s): *atcABC* (gene names)
Systematic name: 3-oxo- Δ^1 -steroid:acceptor 1-oxidoreductase
Comments: A molybdenum enzyme. The enzyme, characterized from the bacterium *Steroidobacter denitrificans*, is involved in the anaerobic degradation of steroids. It is specific to 3-oxo- Δ^1 -steroids such as androsta-1-ene-3,17-dione and Δ^1 -dihydrotestosterone and does not act on 3-oxo- Δ^4 -steroids.
References: [4763]

[EC 1.17.99.11 created 2020]

EC 1.18 Acting on iron-sulfur proteins as donors

This subclass contains enzymes that act on iron-sulfur proteins as donors. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.18.1) and dinitrogen (EC 1.18.6).

EC 1.18.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.18.1.1

- Accepted name:** rubredoxin—NAD⁺ reductase
Reaction: 2 reduced rubredoxin + NAD⁺ + H⁺ = 2 oxidized rubredoxin + NADH
Other name(s): rubredoxin reductase; rubredoxin-nicotinamide adenine dinucleotide reductase; dihydronicotinamide adenine dinucleotide-rubredoxin reductase; reduced nicotinamide adenine dinucleotide-rubredoxin reductase; NADH-rubredoxin reductase; rubredoxin-NAD reductase; NADH: rubredoxin oxidoreductase; DPNH-rubredoxin reductase; NADH-rubredoxin oxidoreductase
Systematic name: rubredoxin:NAD⁺ oxidoreductase
Comments: Requires FAD. The enzyme from *Clostridium acetobutylicum* reduces rubredoxin, ferricyanide and dichlorophenolindophenol, but not ferredoxin or flavodoxin. The reaction does not occur when NADPH is substituted for NADH. Contains iron at the redox centre.
References: [3301, 4364, 4365, 3305]

[EC 1.18.1.1 created 1972 as EC 1.6.7.2, transferred 1978 to EC 1.18.1.1, modified 2001]

EC 1.18.1.2

- Accepted name:** ferredoxin—NADP⁺ reductase
Reaction: 2 reduced ferredoxin + NADP⁺ + H⁺ = 2 oxidized ferredoxin + NADPH
Other name(s): ferredoxin-nicotinamide adenine dinucleotide phosphate reductase; ferredoxin-NADP⁺ reductase; TPNH-ferredoxin reductase; ferredoxin-NADP⁺ oxidoreductase; NADP⁺:ferredoxin oxidoreductase; ferredoxin-TPN reductase; ferredoxin-NADP⁺-oxidoreductase; NADPH:ferredoxin oxidoreductase; ferredoxin-nicotinamide-adenine dinucleotide phosphate (oxidized) reductase
Systematic name: ferredoxin:NADP⁺ oxidoreductase
Comments: A flavoprotein (FAD). In chloroplasts and cyanobacteria the enzyme acts on plant-type [2Fe-2S] ferredoxins, but in other bacteria it can also reduce bacterial [4Fe-4S] ferredoxins and flavodoxin.
References: [3881, 2162, 1998, 2878]

[EC 1.18.1.2 created 1965 as EC 1.6.99.4, transferred 1972 as EC 1.6.7.1, transferred 1978 to EC 1.18.1.2, part transferred 2012 to EC 1.18.1.6, modified 2012]

EC 1.18.1.3

- Accepted name:** ferredoxin—NAD⁺ reductase
Reaction: (1) 2 reduced [2Fe-2S] ferredoxin + NAD⁺ + H⁺ = 2 oxidized [2Fe-2S] ferredoxin + NADH
(2) reduced 2[4Fe-4S] ferredoxin + NAD⁺ + H⁺ = oxidized 2[4Fe-4S] ferredoxin + NADH
Other name(s): ferredoxin-nicotinamide adenine dinucleotide reductase; ferredoxin reductase (ambiguous); NAD⁺-ferredoxin reductase; NADH-ferredoxin oxidoreductase; reductase, reduced nicotinamide adenine dinucleotide-ferredoxin; ferredoxin-NAD⁺ reductase; NADH-ferredoxin reductase; NADH₂-ferredoxin oxidoreductase; NADH flavodoxin oxidoreductase; NADH-ferredoxin NAP reductase (component of naphthalene dioxygenase multicomponent enzyme system); ferredoxin-linked NAD⁺ reductase; NADH-ferredoxin TOL reductase (component of toluene dioxygenase); ferredoxin—NAD reductase
Systematic name: ferredoxin:NAD⁺ oxidoreductase
Comments: Contains FAD. Reaction (1) is written for a [2Fe-2S] ferredoxin, which is characteristic of some mono- and dioxygenase systems. The alternative reaction (2) is written for a 2[4Fe-4S] ferredoxin, which transfers two electrons, and occurs in metabolism of anaerobic bacteria.
References: [1961, 1477, 3441, 3833]

[EC 1.18.1.3 created 1976 as EC 1.6.7.3, transferred 1978 to EC 1.18.1.3, modified 2011]

EC 1.18.1.4

- Accepted name:** rubredoxin—NAD(P)⁺ reductase
Reaction: 2 reduced rubredoxin + NAD(P)⁺ + H⁺ = 2 oxidized rubredoxin + NAD(P)H
Other name(s): rubredoxin-nicotinamide adenine dinucleotide (phosphate) reductase; rubredoxin-nicotinamide adenine; dinucleotide phosphate reductase; NAD(P)⁺-rubredoxin oxidoreductase; NAD(P)H-rubredoxin oxidoreductase
Systematic name: rubredoxin:NAD(P)⁺ oxidoreductase
Comments: The enzyme from *Pyrococcus furiosus* requires FAD. It reduces a number of electron carriers, including benzyl viologen, menadione and 2,6-dichloroindophenol, but rubredoxin is the most efficient. Ferredoxin is not utilized.
References: [3304, 2574]

[EC 1.18.1.4 created 1984, modified 2001, modified 2011]

EC 1.18.1.5

- Accepted name:** putidaredoxin—NAD⁺ reductase
Reaction: reduced putidaredoxin + NAD⁺ = oxidized putidaredoxin + NADH + H⁺
Other name(s): putidaredoxin reductase; *camA* (gene name)
Systematic name: putidaredoxin:NAD⁺ oxidoreductase

Comments: Requires FAD. The enzyme from *Pseudomonas putida* reduces putidaredoxin. It contains a [2Fe-2S] cluster. Involved in the camphor monooxygenase system (see EC 1.14.15.1, camphor 5-monooxygenase).

References: [3568, 2193, 3302, 3811, 3808, 3809, 3941]

[EC 1.18.1.5 created 2012]

EC 1.18.1.6

Accepted name: adrenodoxin-NADP⁺ reductase

Reaction: 2 reduced adrenodoxin + NADP⁺ + H⁺ = 2 oxidized adrenodoxin + NADPH

Other name(s): adrenodoxin reductase; nicotinamide adenine dinucleotide phosphate-adrenodoxin reductase; AdR; NADPH:adrenal ferredoxin oxidoreductase; NADPH-adrenodoxin reductase

Systematic name: reduced adrenodoxin:NADP⁺ oxidoreductase

Comments: A flavoprotein (FAD). The enzyme, which transfers electrons from NADPH to adrenodoxin molecules, is the first component of the mitochondrial cytochrome *P*-450 electron transfer systems, and is involved in the biosynthesis of all steroid hormones.

References: [3180, 688, 4105, 1519, 1518, 1517, 4931]

[EC 1.18.1.6 created 1965 as EC 1.6.99.4, transferred 1972 as EC 1.6.7.1, transferred 1978 to EC 1.18.1.2, part transferred 2012 to EC 1.18.1.6, modified 2016]

EC 1.18.1.7

Accepted name: ferredoxin—NAD(P)⁺ reductase (naphthalene dioxygenase ferredoxin-specific)

Reaction: 2 reduced [2Fe-2S] ferredoxin + NAD(P)⁺ + H⁺ = 2 oxidized [2Fe-2S] ferredoxin + NAD(P)H

Other name(s): NADH-ferredoxin(NAP) reductase

Systematic name: ferredoxin:NAD(P)⁺ oxidoreductase

Comments: The enzyme from the aerobic bacterium *Ralstonia* sp. U2 donates electrons to both EC 1.14.12.12, naphthalene 1,2-dioxygenase and EC 1.14.13.172, salicylate 5-hydroxylase [4914]. The enzyme from *Pseudomonas* NCIB 9816 is specific for the ferredoxin associated with naphthalene dioxygenase; it contains FAD and a [2Fe-2S] cluster.

References: [4914, 1477]

[EC 1.18.1.7 created 2013]

[1.18.1.8 *Transferred entry. ferredoxin-NAD⁺ oxidoreductase (Na⁺-transporting). Now EC 7.2.1.2, ferredoxin—NAD⁺ oxidoreductase (Na⁺-transporting)*]

[EC 1.18.1.8 created 2015, deleted 2018]

EC 1.18.2 With dinitrogen as acceptor (deleted sub-subclass)

[1.18.2.1 *Transferred entry. now EC 1.18.6.1, nitrogenase*]

[EC 1.18.2.1 created 1978, deleted 1984]

EC 1.18.3 With H⁺ as acceptor (deleted sub-subclass)

[1.18.3.1 *Transferred entry. hydrogenase. Now EC 1.12.7.2, ferredoxin hydrogenase*]

[EC 1.18.3.1 created 1978, deleted 1984]

EC 1.18.6 With dinitrogen as acceptor

EC 1.18.6.1

- Accepted name:** nitrogenase
- Reaction:** 8 reduced ferredoxin + 8 H⁺ + N₂ + 16 ATP + 16 H₂O = 8 oxidized ferredoxin + H₂ + 2 NH₃ + 16 ADP + 16 phosphate
- Other name(s):** reduced ferredoxin:dinitrogen oxidoreductase (ATP-hydrolysing)
- Systematic name:** ferredoxin:dinitrogen oxidoreductase (ATP-hydrolysing, molybdenum-dependent)
- Comments:** Requires Mg²⁺. The enzyme is a complex of two components (namely dinitrogen reductase and dinitrogenase). Dinitrogen reductase is a [4Fe-4S] protein, which, in the presence of two molecules of ATP, transfers an electron from ferredoxin to the dinitrogenase component. Dinitrogenase is a molybdenum-iron protein that reduces dinitrogen to two molecules of ammonia in three successive two-electron reductions via diazene and hydrazine. The reduction is initiated by formation of hydrogen in stoichiometric amounts [2470]. Acetylene is reduced to ethylene (but only very slowly to ethane), azide to nitrogen and ammonia, and cyanide to methane and ammonia. In the absence of a suitable substrate, hydrogen is slowly formed. Ferredoxin may be replaced by flavodoxin [see EC 1.19.6.1 nitrogenase (flavodoxin)]. The enzyme does not reduce CO (*cf.* EC 1.18.6.2, vanadium-dependent nitrogenase).
- References:** [4944, 2470, 817, 592]

[EC 1.18.6.1 created 1978 as EC 1.18.2.1, transferred 1984 to EC 1.18.6.1, modified 2005, modified 2018]

EC 1.18.6.2

- Accepted name:** vanadium-dependent nitrogenase
- Reaction:** 12 reduced ferredoxin + 12 H⁺ + N₂ + 40 ATP + 40 H₂O = 12 oxidized ferredoxin + 3 H₂ + 2 NH₃ + 40 ADP + 40 phosphate
- Other name(s):** *vnfD* (gene name); *vnfG* (gene name); *vnfK* (gene name)
- Systematic name:** ferredoxin:dinitrogen oxidoreductase (ATP-hydrolysing, vanadium-dependent)
- Comments:** Requires Mg²⁺. This enzyme, originally isolated from the bacterium *Azotobacter vinelandii*, is a complex of two components (namely dinitrogen reductase and dinitrogenase). Dinitrogen reductase is a [4Fe-4S] protein, which, in the presence of ATP, transfers an electron from ferredoxin to the dinitrogenase component. Dinitrogenase is a vanadium-iron protein that reduces dinitrogen to two molecules of ammonia in three successive two-electron reductions via diazine and hydrazine. Compared with molybdenum-dependent nitrogenase (EC 1.18.6.1), this enzyme produces more dihydrogen and consumes more ATP per dinitrogen molecule being reduced. Unlike EC 1.18.6.1, this enzyme can also use CO as substrate, producing ethene, ethane and propane [2380, 3915].
- References:** [1003, 2813, 4276, 917, 918, 998, 2380, 2381, 3915]

[EC 1.18.6.2 created 2018]

EC 1.18.96 With other, known, acceptors (deleted sub-subclass)

[1.18.96.1 Transferred entry. superoxide reductase. Now EC 1.15.1.2, superoxide reductase]

[EC 1.18.96.1 created 2001, deleted 2001]

EC 1.18.99 With H⁺ as acceptor (deleted sub-subclass)

[1.18.99.1 Transferred entry. hydrogenase. Now EC 1.12.7.2, ferredoxin hydrogenase]

[EC 1.18.99.1 created 1961 as EC 1.98.1.1, transferred 1965 to EC 1.12.1.1, transferred 1972 to EC 1.12.7.1, transferred 1978 to EC 1.18.3.1, transferred 1984 to EC 1.18.99.1, deleted 2002]

EC 1.19 Acting on reduced flavodoxin as donor

This subclass contains enzymes that act on reduced flavodoxin as donors. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.19.1) and dinitrogen (EC 1.19.6).

EC 1.19.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.19.1.1

- Accepted name:** flavodoxin—NADP⁺ reductase
Reaction: reduced flavodoxin + NADP⁺ = oxidized flavodoxin + NADPH + H⁺
Other name(s): FPR
Systematic name: flavodoxin:NADP⁺ oxidoreductase
Comments: A flavoprotein (FAD). This activity occurs in some prokaryotes and algae that possess flavodoxin, and provides low-potential electrons for a variety of reactions such as nitrogen fixation, sulfur assimilation and amino acid biosynthesis. In photosynthetic organisms it is involved in the photosynthetic electron transport chain. The enzyme also catalyses EC 1.18.1.2, ferredoxin—NADP⁺ reductase.
References: [2751, 2374, 4510, 396, 397, 3924]

[EC 1.19.1.1 created 2016]

EC 1.19.6 With dinitrogen as acceptor

EC 1.19.6.1

- Accepted name:** nitrogenase (flavodoxin)
Reaction: 4 reduced flavodoxin + N₂ + 16 ATP + 16 H₂O = 4 oxidized flavodoxin + H₂ + 2 NH₃ + 16 ADP + 16 phosphate
Systematic name: reduced flavodoxin:dinitrogen oxidoreductase (ATP-hydrolysing)
Comments: Requires Mg²⁺. It is composed of two components, dinitrogen reductase and dinitrogenase, that can be separated but are both required for nitrogenase activity. Dinitrogen reductase is a [4Fe-4S] protein, which, at the expense of ATP, transfers electrons from a dedicated flavodoxin to dinitrogenase. Dinitrogenase is a protein complex that contains either a molybdenum-iron cofactor, a vanadium-iron cofactor, or an iron-iron cofactor, that reduces dinitrogen in three successive two-electron reductions from nitrogen to diimine to hydrazine to two molecules of ammonia. The reduction is initiated by formation of hydrogen. The enzyme can also reduce acetylene to ethylene (but only very slowly to ethane), azide to nitrogen and ammonia, and cyanide to methane and ammonia. In the absence of a suitable substrate, hydrogen is slowly formed. Some enzymes utilize ferredoxin rather than flavodoxin as the electron donor (see EC 1.18.6.1, nitrogenase).
References: [4943, 1004, 863]

[EC 1.19.6.1 created 1984, modified 2014]

EC 1.20 Acting on phosphorus or arsenic in donors

This subclass contains enzymes that act on phosphorus or arsenic in donors. Sub-subclasses are based on the acceptor: NAD⁺ or NADP⁺ (EC 1.20.1), disulfide (EC 1.20.4), other, known, acceptors (EC 1.20.98), or some other acceptor (EC 1.20.99).

EC 1.20.1 With NAD⁺ or NADP⁺ as acceptor

EC 1.20.1.1

- Accepted name:** phosphonate dehydrogenase
Reaction: phosphonate + NAD⁺ + H₂O = phosphate + NADH + H⁺
Other name(s): NAD:phosphite oxidoreductase; phosphite dehydrogenase
Systematic name: phosphonate:NAD⁺ oxidoreductase
Comments: NADP⁺ is a poor substitute for NAD⁺ in the enzyme from *Pseudomonas stutzeri* WM88.
References: [747, 4478]

[EC 1.20.1.1 created 2001]

EC 1.20.2 With a cytochrome as acceptor

EC 1.20.2.1

- Accepted name:** arsenate reductase (cytochrome *c*)
Reaction: arsenite + H₂O + 2 oxidized cytochrome *c* = arsenate + 2 reduced cytochrome *c* + 2 H⁺
Other name(s): arsenite oxidase (ambiguous)
Systematic name: arsenite:cytochrome *c* oxidoreductase
Comments: A molybdoprotein containing iron-sulfur clusters. Isolated from α -proteobacteria. Unlike EC 1.20.9.1, arsenate reductase (azurin), it does not use azurin as acceptor.
References: [4420, 3660, 423, 2484]

[EC 1.20.2.1 created 2011]

EC 1.20.4 With disulfide as acceptor

EC 1.20.4.1

- Accepted name:** arsenate reductase (glutathione/glutaredoxin)
Reaction: arsenate + glutathione + glutaredoxin = arsenite + a glutaredoxin-glutathione disulfide + H₂O
Other name(s): ArsC (ambiguous); arsenate:glutaredoxin oxidoreductase; arsenate reductase (glutaredoxin)
Systematic name: arsenate:glutathione/glutaredoxin oxidoreductase
Comments: The enzyme is part of a system for detoxifying arsenate. The substrate binds to a catalytic cysteine residue, forming a covalent thiolate—As(V) intermediate. A tertiary intermediate is then formed between the arsenic, the enzyme's cysteine, and a glutathione cysteine. This intermediate is reduced by glutaredoxin, which forms a dithiol with the glutathione, leading to the dissociation of arsenite. Thus reduction of As(V) is mediated by three cysteine residues: one in ArsC, one in glutathione, and one in glutaredoxin. Although the arsenite formed is more toxic than arsenate, it can be extruded from some bacteria by EC 7.3.2.7, arsenite-transporting ATPase; in other organisms, arsenite can be methylated by EC 2.1.1.137, arsenite methyltransferase, in a pathway that produces non-toxic organoarsenical compounds. *cf.* EC 1.20.4.4, arsenate reductase (thioredoxin).
References: [1336, 1337, 1702, 2256, 2666, 3422, 3671, 3849, 2919, 2778]

[EC 1.20.4.1 created 2000 as EC 1.97.1.5, transferred 2001 to EC 1.20.4.1, modified 2015, modified 2019, modified 2020]

EC 1.20.4.2

- Accepted name:** methylarsonate reductase
Reaction: methylarsonate + 2 glutathione = methylarsonite + glutathione disulfide + H₂O
Other name(s): MMA(V) reductase
Systematic name: methylarsonate:glutathione oxidoreductase
Comments: The product, methylarsonite, is biologically methylated by EC 2.1.1.137, arsenite methyltransferase, to form cacodylic acid.
References: [4855]

[EC 1.20.4.2 created 2000 as EC 1.97.1.7, transferred 2001 to EC 1.20.4.2, modified 2003]

EC 1.20.4.3

Accepted name: mycoredoxin
Reaction: arseno-mycothiol + mycoredoxin = arsenite + mycothiol-mycoredoxin disulfide
Other name(s): Mrx1; MrxI
Systematic name: arseno-mycothiol:mycoredoxin oxidoreductase
Comments: Reduction of arsenate is part of a defense mechanism of the cell against toxic arsenate. The substrate arseno-mycothiol is formed by EC 2.8.4.2 (arsenate:mycothiol transferase). A second mycothiol recycles mycoredoxin and forms mycothione.
References: [3190]

[EC 1.20.4.3 created 2010]

EC 1.20.4.4

Accepted name: arsenate reductase (thioredoxin)
Reaction: arsenate + thioredoxin = arsenite + thioredoxin disulfide + H₂O
Other name(s): ArsC (ambiguous)
Systematic name: arsenate:thioredoxin oxidoreductase
Comments: The enzyme, characterized in bacteria of the Firmicutes phylum, is specific for thioredoxin [1910]. It has no activity with glutaredoxin [*cf.* EC 1.20.4.1, arsenate reductase (glutaredoxin)]. Although the arsenite formed is more toxic than arsenate, it can be extruded from some bacteria by EC 7.3.2.7, arsenite-transporting ATPase; in other organisms, arsenite can be methylated by EC 2.1.1.137, arsenite methyltransferase, in a pathway that produces non-toxic organoarsenical compounds. The enzyme also has the activity of EC 3.1.3.48, protein-tyrosine-phosphatase [4864].
References: [1910, 2776, 4864, 2777]

[EC 1.20.4.4 created 2015, modified 2019]

EC 1.20.9 With a copper protein as acceptor

EC 1.20.9.1

Accepted name: arsenate reductase (azurin)
Reaction: arsenite + H₂O + 2 oxidized azurin = arsenate + 2 reduced azurin + 2 H⁺
Other name(s): arsenite oxidase (ambiguous)
Systematic name: arsenite:azurin oxidoreductase
Comments: Contains a molybdopterin centre comprising two molybdopterin guanosine dinucleotide cofactors bound to molybdenum, a [3Fe-4S] cluster and a Rieske-type [2Fe-2S] cluster. Isolated from β -proteobacteria. Also uses a *c*-type cytochrome or O₂ as acceptors.
References: [93, 1039]

[EC 1.20.9.1 created 2001 as EC 1.20.98.1, transferred 2011 to EC 1.20.9.1]

EC 1.20.98 With other, known, physiological acceptors

[1.20.98.1 Transferred entry. arsenate reductase (azurin). Now EC 1.20.9.1, arsenate reductase (azurin)]

[EC 1.20.98.1 created 2001, deleted 2011]

EC 1.20.99 With unknown physiological acceptors

EC 1.20.99.1

- Accepted name:** arsenate reductase (donor)
Reaction: arsenite + acceptor = arsenate + reduced acceptor
Other name(s): arsenate:(acceptor) oxidoreductase
Systematic name: arsenate:acceptor oxidoreductase
Comments: Benzyl viologen can act as an acceptor. Unlike EC 1.20.4.1, arsenate reductase (glutaredoxin), reduced glutaredoxin cannot serve as a reductant.
References: [2256, 3422]

[EC 1.20.99.1 created 2000 as EC 1.97.1.6, transferred 2001 to EC 1.20.99.1]

EC 1.21 Catalysing the reaction $X-H + Y-H = X-Y$

This subclass contains enzymes that catalyse the reaction $X-H + Y-H = X-Y$, forming or breaking an X-Y bond. Sub-subclasses are based on the acceptor: oxygen (EC 1.21.3), a disulfide (EC 1.21.4), or some other unidentified acceptor (EC 1.21.99).

EC 1.21.1 With NAD^+ or $NADP^+$ as acceptor

EC 1.21.1.1

- Accepted name:** iodotyrosine deiodinase
Reaction: L-tyrosine + 2 $NADP^+$ + 2 iodide = 3,5-diiodo-L-tyrosine + 2 $NADPH$ + 2 H^+ (overall reaction)
(1a) L-tyrosine + $NADP^+$ + iodide = 3-iodo-L-tyrosine + $NADPH$ + H^+
(1b) 3-iodo-L-tyrosine + $NADP^+$ + iodide = 3,5-diiodo-L-tyrosine + $NADPH$ + H^+
Other name(s): iodotyrosine dehalogenase 1; DEHAL1
Systematic name: L-tyrosine,iodide: $NADP^+$ oxidoreductase (iodinating)
Comments: The enzyme activity has only been demonstrated in the direction of 3-deiodination. Present in a trans-membrane flavoprotein. Requires FMN.
References: [3574, 1348, 1182, 4266]

[EC 1.21.1.1 created 2010 as EC 1.22.1.1, transferred 2015 to EC 1.21.1.1]

EC 1.21.1.2

- Accepted name:** 2,4-dichlorobenzoyl-CoA reductase
Reaction: 4-chlorobenzoyl-CoA + $NADP^+$ + chloride = 2,4-dichlorobenzoyl-CoA + $NADPH$ + H^+
Systematic name: 4-chlorobenzoyl-CoA: $NADP^+$ oxidoreductase (halogenating)
Comments: The enzyme, characterized from *Corynebacterium* strains able to grow on 2,4-dichlorobenzoate, forms part of the 2,4-dichlorobenzoate degradation pathway.
References: [3564]

[EC 1.21.1.2 created 2000 as EC 1.3.1.63, modified 2011, transferred 2015 to EC 1.21.1.2]

EC 1.21.3 With oxygen as acceptor

EC 1.21.3.1

- Accepted name:** isopenicillin-N synthase
Reaction: *N*-[(5*S*)-5-amino-5-carboxypentanoyl]-L-cysteinyl-D-valine + O_2 = isopenicillin N + 2 H_2O
Other name(s): isopenicillin N synthetase

Systematic name: *N*-[(5*S*)-5-amino-5-carboxypentanoyl]-L-cysteinyl-D-valine:oxygen oxidoreductase (cyclizing)
Comments: Forms part of the penicillin biosynthesis pathway (for pathway, [click here](#)).
References: [1763, 3532]

[EC 1.21.3.1 created 2002]

EC 1.21.3.2

Accepted name: columbamine oxidase
Reaction: 2 columbamine + O₂ = 2 berberine + 2 H₂O
Other name(s): berberine synthase
Systematic name: columbamine:oxygen oxidoreductase (cyclizing)
Comments: An iron protein. Oxidation of the *O*-methoxyphenol structure forms the methylenedioxy group of berberine.
References: [3595]

[EC 1.21.3.2 created 1989 as EC 1.1.3.26, transferred 2002 to EC 1.21.3.2]

EC 1.21.3.3

Accepted name: reticuline oxidase
Reaction: (*S*)-reticuline + O₂ = (*S*)-scoulerine + H₂O₂
Other name(s): BBE; berberine bridge enzyme; berberine-bridge-forming enzyme; tetrahydroprotoberberine synthase
Systematic name: (*S*)-reticuline:oxygen oxidoreductase (methylene-bridge-forming)
Comments: Contains FAD. The enzyme from the plant *Eschscholtzia californica* binds the cofactor covalently [2311]. Acts on (*S*)-reticuline and related compounds, converting the *N*-methyl group into the methylene bridge ('berberine bridge') of (*S*)-tetrahydroprotoberberines. The product of the reaction, (*S*)-scoulerine, is a precursor of protopine, protoberberine and benzophenanthridine alkaloid biosynthesis in plants.
References: [4014, 922, 2311]

[EC 1.21.3.3 created 1989 as EC 1.5.3.9, transferred 2002 to EC 1.21.3.3]

EC 1.21.3.4

Accepted name: sulochrin oxidase [(+)-bisdechlorogeodin-forming]
Reaction: 2 sulochrin + O₂ = 2 (+)-bisdechlorogeodin + 2 H₂O
Other name(s): sulochrin oxidase
Systematic name: sulochrin:oxygen oxidoreductase (cyclizing, (+)-specific)
Comments: Also acts on several diphenols and phenylenediamines, but has low affinity for these substrates. Involved in the biosynthesis of mould metabolites related to the antibiotic griseofulvin.
References: [3105]

[EC 1.21.3.4 created 1986 as EC 1.10.3.7, transferred 2002 to EC 1.21.3.4]

EC 1.21.3.5

Accepted name: sulochrin oxidase [(-)-bisdechlorogeodin-forming]
Reaction: 2 sulochrin + O₂ = 2 (-)-bisdechlorogeodin + 2 H₂O
Other name(s): sulochrin oxidase
Systematic name: sulochrin:oxygen oxidoreductase (cyclizing, (-)-specific)
Comments: Also acts on several diphenols and phenylenediamines, but has low affinity for these substrates. Involved in the biosynthesis of mould metabolites related to the antibiotic griseofulvin.
References: [3105]

[EC 1.21.3.5 created 1986 as EC 1.10.3.8, transferred 2002 to EC 1.21.3.5]

EC 1.21.3.6

- Accepted name:** aureusidin synthase
- Reaction:** (1) 2',4,4',6'-tetrahydroxychalcone 4'-O-β-D-glucoside + O₂ = aureusidin 6-O-β-D-glucoside + H₂O
(2) 2',3,4,4',6'-pentahydroxychalcone 4'-O-β-D-glucoside + $\frac{1}{2}$ O₂ = aureusidin 6-O-β-D-glucoside + H₂O
(3) 2',3,4,4',6'-pentahydroxychalcone 4'-O-β-D-glucoside + O₂ = bracteatin 6-O-β-D-glucoside + H₂O
- Other name(s):** AmAS1
- Systematic name:** 2',4,4',6'-tetrahydroxychalcone 4'-O-β-D-glucoside:oxygen oxidoreductase
- Comments:** A copper-containing glycoprotein that plays a key role in the yellow coloration of flowers such as *Antirrhinum majus* (snapdragon). The enzyme is a homologue of plant polyphenol oxidase [3001] and catalyses two separate chemical transformations, i.e. 3-hydroxylation and oxidative cyclization (2',-dehydrogenation). H₂O₂ activates reaction (1) but inhibits reaction (2). Originally considered to act on the phenol but now thought to act mainly on the 4'-O-β-D-glucoside *in vivo* [3184].
- References:** [3001, 3000, 3672, 3184]

[EC 1.21.3.6 created 2003, modified 2012]

EC 1.21.3.7

- Accepted name:** tetrahydrocannabinolic acid synthase
- Reaction:** cannabigerolate + O₂ = Δ⁹-tetrahydrocannabinolate + H₂O₂
- Other name(s):** THCA synthase; Δ¹-tetrahydrocannabinolic acid synthase
- Systematic name:** cannabigerolate:oxygen oxidoreductase (cyclizing, Δ⁹-tetrahydrocannabinolate-forming)
- Comments:** A flavoprotein (FAD). The cofactor is covalently bound. Part of the cannabinoids biosynthetic pathway in the plant *Cannabis sativa*. The enzyme can also convert cannabinerolate (the (Z)-isomer of cannabigerolate) to Δ⁹-THCA with lower efficiency. Whereas the product was originally called Δ¹-tetrahydrocannabinolate, the recommended name according to systematic peripheral numbering is Δ⁹-tetrahydrocannabinolate.
- References:** [4220, 3917, 3893, 3894]

[EC 1.21.3.7 created 2012]

EC 1.21.3.8

- Accepted name:** cannabidiolic acid synthase
- Reaction:** cannabigerolate + O₂ = cannabidiolate + H₂O₂
- Other name(s):** CBDA synthase
- Systematic name:** cannabigerolate:oxygen oxidoreductase (cyclizing, cannabidiolate-forming)
- Comments:** Binds FAD covalently. Part of the cannabinoids biosynthetic pathway of the plant *Cannabis sativa*. The enzyme can also convert cannabinerolate to cannabidiolate with lower efficiency.
- References:** [4219, 4221]

[EC 1.21.3.8 created 2012]

[1.21.3.9 Transferred entry. dichlorochromopyrrolate synthase, now classified as EC 1.21.98.2, dichlorochromopyrrolate synthase]

[EC 1.21.3.9 created 2010 as EC 4.3.1.26, transferred 2013 to EC 1.21.3.9, deleted 2016]

EC 1.21.3.10

- Accepted name:** hercynylcysteine S-oxide synthase
- Reaction:** hercynine + L-cysteine + O₂ = S-(hercyn-2-yl)-L-cysteine S-oxide + H₂O
- Other name(s):** Egt1; Egt-1
- Systematic name:** hercynine,L-cysteine:oxygen [S-(hercyn-2-yl)-L-cysteine S-oxide-forming]

Comments: Requires Fe²⁺ for activity. The enzyme, found in fungal species, is part of a fusion protein that also has the activity of EC 2.1.1.44, L-histidine N^α-methyltransferase. It is part of the biosynthesis pathway of ergothioneine. The enzyme can also use L-selenocysteine to produce hercynylselenocysteine, which can be converted to selenoneine.

References: [3337]

[EC 1.21.3.10 created 2015 as 1.14.99.51, transferred 2022 to EC 1.21.3.10]

EC 1.21.4 With a disulfide as acceptor

EC 1.21.4.1

Accepted name: D-proline reductase

Reaction: 5-aminopentanoate + a [PrdC protein with a selenide-sulfide bridge] = D-proline + a [PrdC protein with thiol/selenol residues]

Other name(s): *prdAB* (gene names); D-proline reductase (dithiol)

Systematic name: 5-aminopentanoate:[PrdC protein] oxidoreductase (cyclizing)

Comments: A pyruvoyl- and L-selenocysteine-containing enzyme found in a number of Clostridial species. The pyruvoyl group, located on the PrdA subunit, binds the substrate, while the selenocysteine residue, located on the PrdB subunit, attacks the α-C-atom of D-proline, leading to a reductive cleavage of the C-N-bond of the pyrrolidine ring and formation of a selenoether. The selenoether is cleaved by a cysteine residue of PrdB, resulting in a mixed selenide-sulfide bridge, which is restored to its reduced state by another selenocysteine protein, PrdC. 5-aminopentanoate is released from PrdA by hydrolysis, regenerating the pyruvoyl moiety. The resulting mixed selenide-sulfide bridge in PrdC is reduced by NADH.

References: [3998, 1683, 1966, 266, 1134]

[EC 1.21.4.1 created 1972 as EC 1.4.4.1, modified 1982 (EC 1.4.1.6 created 1961, incorporated 1982), transferred 2003 to EC 1.21.4.1, modified 2018]

EC 1.21.4.2

Accepted name: glycine reductase

Reaction: acetyl phosphate + NH₃ + thioredoxin disulfide + H₂O = glycine + phosphate + thioredoxin

Systematic name: acetyl-phosphate ammonia:thioredoxin disulfide oxidoreductase (glycine-forming)

Comments: The reaction is observed only in the direction of glycine reduction. The enzyme from *Eubacterium acidaminophilum* consists of subunits A, B and C. Subunit B contains selenocysteine and a pyruvoyl group, and is responsible for glycine binding and ammonia release. Subunit A, which also contains selenocysteine, is reduced by thioredoxin, and is needed to convert the carboxymethyl group into a ketene equivalent, in turn used by subunit C to produce acetyl phosphate. Only subunit B distinguishes this enzyme from EC 1.21.4.3 (sarcosine reductase) and EC 1.21.4.4 (betaine reductase).

References: [4487, 266]

[EC 1.21.4.2 created 2003]

EC 1.21.4.3

Accepted name: sarcosine reductase

Reaction: acetyl phosphate + methylamine + thioredoxin disulfide + H₂O = N-methylglycine + phosphate + thioredoxin

Systematic name: acetyl-phosphate methylamine:thioredoxin disulfide oxidoreductase (N-methylglycine-forming)

Comments: The reaction is observed only in the direction of sarcosine reduction. The enzyme from *Eubacterium acidaminophilum* consists of subunits A, B and C. Subunit B contains selenocysteine and a pyruvoyl group, and is responsible for sarcosine binding and methylamine release. Subunit A, which also contains selenocysteine, is reduced by thioredoxin, and is needed to convert the carboxymethyl group into a ketene equivalent, in turn used by subunit C to produce acetyl phosphate. Only subunit B distinguishes this enzyme from EC 1.21.4.2 (glycine reductase) and EC 1.21.4.4 (betaine reductase).

References: [4487, 1730]

[EC 1.21.4.3 created 2003]

EC 1.21.4.4

Accepted name: betaine reductase

Reaction: acetyl phosphate + trimethylamine + thioredoxin disulfide + H₂O = betaine + phosphate + thioredoxin

Other name(s): acetyl-phosphate trimethylamine:thioredoxin disulfide oxidoreductase (*N,N,N*-trimethylglycine-forming)

Systematic name: acetyl-phosphate trimethylamine:thioredoxin disulfide oxidoreductase (betaine-forming)

Comments: The reaction is observed only in the direction of betaine reduction. The enzyme from *Eubacterium acidaminophilum* consists of subunits A, B and C. Subunit B contains selenocysteine and a pyruvoyl group, and is responsible for betaine binding and trimethylamine release. Subunit A, which also contains selenocysteine, is reduced by thioredoxin, and is needed to convert the carboxymethyl group into a ketene equivalent, in turn used by subunit C to produce acetyl phosphate. Only subunit B distinguishes this enzyme from EC 1.21.4.2 (glycine reductase) and EC 1.21.4.3 (sarcosine reductase).

References: [4487, 266]

[EC 1.21.4.4 created 2003, modified 2010]

EC 1.21.4.5

Accepted name: tetrachlorohydroquinone reductive dehalogenase

Reaction: (1) 2,6-dichlorohydroquinone + Cl⁻ + glutathione disulfide = 2,3,6-trichlorohydroquinone + 2 glutathione

(2) 2,3,6-trichlorohydroquinone + Cl⁻ + glutathione disulfide = 2,3,5,6-tetrachlorohydroquinone + 2 glutathione

Other name(s): *pcpC* (gene name)

Systematic name: glutathione disulfide:2,6-dichlorohydroquinone (chlorinating)

Comments: The enzyme, characterized from the bacterium *Sphingobium chlorophenicum*, converts tetrachlorohydroquinone to 2,6-dichlorohydroquinone in two steps, via 2,3,6-trichlorohydroquinone, using glutathione as the reducing agent. The enzyme is sensitive to oxidation - when an internal L-cysteine residue is oxidized, the enzyme produces 2,3,5-trichloro-6-(glutathion-S-yl)-hydroquinone and 2,6-dichloro-3-(glutathion-S-yl)-hydroquinone instead of its normal products.

References: [4715, 2741]

[EC 1.21.4.5 created 2018]

EC 1.21.98 With other, known, physiological acceptors

EC 1.21.98.1

Accepted name: cyclic dehypoxanthinyl futasosine synthase

Reaction: dehypoxanthine futasosine + *S*-adenosyl-L-methionine = cyclic dehypoxanthinyl futasosine + 5'-deoxyadenosine + L-methionine

Other name(s): MqnC; dehypoxanthinyl futasosine cyclase

Systematic name: dehypoxanthine futasosine:*S*-adenosyl-L-methionine oxidoreductase (cyclizing)

Comments: This enzyme is a member of the 'AdoMet radical' (radical SAM) family. The enzyme, found in several bacterial species, is part of the futasol pathway for menaquinone biosynthesis.

References: [1671, 727]

[EC 1.21.98.1 created 2014 as EC 1.21.99.2, transferred 2014 to EC 1.21.98.1]

EC 1.21.98.2

Accepted name: dichlorochromopyrrolate synthase

Reaction: 2 3-(7-chloroindol-3-yl)-2-imino-propionate + H₂O₂ = dichlorochromopyrrolate + NH₃ + 2 H₂O

Other name(s): RebD; chromopyrrolic acid synthase; chromopyrrolate synthase

Systematic name: 3-(7-chloroindol-3-yl)-2-imino-propionate ammonia-lyase (dichlorochromopyrrolate-forming)

Comments: This enzyme catalyses a step in the biosynthesis of rebeccamycin, an indolocarbazole alkaloid produced by the bacterium *Lechevalieria aerocolonigenes*. The enzyme is a dimeric heme-protein oxidase that catalyses the oxidative dimerization of two L-tryptophan-derived molecules to form dichlorochromopyrrolic acid, the precursor for the fused six-ring indolocarbazole scaffold of rebeccamycin [3087]. Contains one molecule of heme *b* per monomer, as well as non-heme iron that is not part of an iron-sulfur center [1743]. *In vivo* the enzyme uses hydrogen peroxide, formed by the enzyme upstream in the biosynthetic pathway (EC 1.4.3.23, 7-chloro-L-tryptophan oxidase) as the electron acceptor. However, the enzyme is also able to catalyse the reaction using molecular oxygen [3989].

References: [3087, 1743, 3989]

[EC 1.21.98.2 created 2010 as EC 4.3.1.26, transferred 2013 to EC 1.21.3.9, transferred 2016 to EC 1.21.98.2]

EC 1.21.98.3

Accepted name: anaerobic magnesium-protoporphyrin IX monomethyl ester cyclase

Reaction: magnesium-protoporphyrin IX 13-monomethyl ester + 3 *S*-adenosyl-L-methionine + H₂O = 3,8-divinyl protochlorophyllide *a* + 3 5'-deoxyadenosine + 3 L-methionine (overall reaction)
(1a) magnesium-protoporphyrin IX 13-monomethyl ester + *S*-adenosyl-L-methionine + H₂O = 13¹-hydroxy-magnesium-protoporphyrin IX 13-monomethyl ester + 5'-deoxyadenosine + L-methionine
(1b) 13¹-hydroxy-magnesium-protoporphyrin IX 13-monomethyl ester + *S*-adenosyl-L-methionine = 13¹-oxo-magnesium-protoporphyrin IX 13-monomethyl ester + 5'-deoxyadenosine + L-methionine
(1c) 13¹-oxo-magnesium-protoporphyrin IX 13-monomethyl ester + *S*-adenosyl-L-methionine = 3,8-divinyl protochlorophyllide *a* + 5'-deoxyadenosine + L-methionine

Other name(s): *bchE* (gene name); MPE cyclase (ambiguous)

Systematic name: magnesium-protoporphyrin-IX 13-monomethyl ester,*S*-adenosyl-L-methionine:H₂O oxidoreductase (hydroxylating)

Comments: This radical AdoMet enzyme participates in the biosynthesis of chlorophyllide *a* in anaerobic bacteria, catalysing the formation of an isocyclic ring. Contains a [4Fe-4S] cluster and a cobalamin cofactor. The same transformation is achieved in aerobic organisms by the oxygen-dependent EC 1.14.13.81, magnesium-protoporphyrin IX monomethyl ester (oxidative) cyclase. Some facultative phototrophic bacteria, such as *Rubrivivax gelatinosus*, possess both enzymes.

References: [4775, 1374, 3212, 389]

[EC 1.21.98.3 created 2016]

EC 1.21.98.4

Accepted name: PqqA peptide cyclase

Reaction: a PqqA peptide + *S*-adenosyl-L-methionine = a PqqA peptide with linked Glu-Tyr residues + 5'-deoxyadenosine + L-methionine

Other name(s): *pqqE* (gene name)

Systematic name: PqqA peptide:*S*-adenosyl-L-methionine oxidoreductase (cyclizing)

Comments: This bacterial enzyme, which is a member of the radical SAM protein family, catalyses the formation of a C-C bond between C-4 of glutamate and C-3 of tyrosine residues of the PqqA protein (which are separated by three amino acid residues). This is the first enzymic step in the biosynthesis of the bacterial enzyme cofactor pyrroloquinoline quinone (PQQ). The reaction is dependent on the presence of a reductant (flavodoxin) and the accessory protein PqqD.

References: [4566, 2360, 224]

[EC 1.21.98.4 created 2018]

EC 1.21.98.5

Accepted name: tetraether lipid synthase

Reaction: (1) 2 a 2,3-bis-*O*-phytanyl-*sn*-glycero-phospholipid + 4 *S*-adenosyl-L-methionine + 2 reduced acceptor = a glycerol dibiphytanyl glycerol tetraether phospholipid + 4 L-methionine + 4 5'-deoxyadenosine + 2 acceptor

(2) a 2,3-bis-*O*-phytanyl-*sn*-glycero-phospholipid + 2 *S*-adenosyl-L-methionine + reduced acceptor = a macrocyclic archaeol phospholipid + 2 L-methionine + 2 5'-deoxyadenosine + acceptor

Other name(s): GDGT/MA synthase; GDGT/MAS; tetraether synthase; Tes; Mj0619 (locus name)

Systematic name: a 2,3-bis-*O*-phytanyl-*sn*-glycero-phospholipid:*S*-adenosyl-L-methionine,acceptor oxidoreductase (cyclizing)

Comments: This archaeal enzyme catalyses a C-C bond formation during the biosynthesis of tetraether lipids. The bond is formed between the termini of two lipid tails from two glycerophospholipids to generate the macrocycle glycerol dibiphytanyl glycerol tetraether (GDGT). The enzyme does not distinguish whether the two lipids are connected in antiparallel or parallel geometry, resulting in formation of two forms of the product, which are known as caldarchaeol and isocaldarchaeol, respectively. The enzyme can also form macrocyclic archaeol phospholipids by joining the two lipid tails of a single substrate molecule. Even though the reaction shown here describes phospholipid substrates, the enzyme can also act on glycolipids or lipids that contains mixed types of polar head groups. The enzyme is a radical SAM enzyme that contains 3 [4Fe-4S] clusters and one mononuclear rubredoxin-like iron ion, each found in a separate domain. The enzyme uses the 5'-deoxyadenosyl radical to initiate the reaction, which involves the formation of an intermediate bond between the substrate carbon and a sulfur of one of the [4Fe-4S] clusters. Two radicals are needed per C-C bond formed. The source of the required additional electrons is not known.

References: [4870, 2528]

[EC 1.21.98.5 created 2022]

EC 1.21.99 With unknown physiological acceptors

EC 1.21.99.1

Accepted name: β -cyclopiazonate dehydrogenase

Reaction: β -cyclopiazonate + acceptor = α -cyclopiazonate + reduced acceptor

Other name(s): β -cyclopiazonate oxidocyclase; β -cyclopiazonic oxidocyclase; β -cyclopiazonate:(acceptor) oxidoreductase (cyclizing)

Systematic name: β -cyclopiazonate:acceptor oxidoreductase (cyclizing)

Comments: A flavoprotein (FAD). Cytochrome *c* and various dyes can act as acceptor. Cyclopiazonate is a microbial toxin.

References: [1018, 3693]

[EC 1.21.99.1 created 1976 as EC 1.3.99.9, transferred 2002 to EC 1.21.99.1]

[1.21.99.2 *Transferred entry. EC 1.21.99.2, cyclic dehydropoxanthinyl futasosine synthase. Now classified as EC 1.21.98.1, cyclic dehydropoxanthinyl futasosine synthase.*]

[EC 1.21.99.2 created 2014, deleted 2014]

EC 1.21.99.3

- Accepted name:** thyroxine 5-deiodinase
Reaction: 3,3',5'-triiodo-L-thyronine + iodide + acceptor + H⁺ = L-thyroxine + reduced acceptor
Other name(s): diiodothyronine 5'-deiodinase (ambiguous); iodothyronine 5-deiodinase; iodothyronine inner ring monodeiodinase; type III iodothyronine deiodinase
Systematic name: 3,3',5'-triiodo-L-thyronine,iodide:acceptor oxidoreductase (iodinating)
Comments: The enzyme activity has only been demonstrated in the direction of 5-deiodination. This removal of the 5-iodine, i.e. from the inner ring, largely inactivates the hormone thyroxine.
References: [677, 2232]

[EC 1.21.99.3 created 2003 as EC 1.97.1.11, transferred 2015 to EC 1.21.99.3]

EC 1.21.99.4

- Accepted name:** thyroxine 5'-deiodinase
Reaction: 3,3',5-triiodo-L-thyronine + iodide + acceptor + H⁺ = L-thyroxine + reduced acceptor
Other name(s): diiodothyronine 5'-deiodinase [ambiguous]; iodothyronine 5'-deiodinase; iodothyronine outer ring monodeiodinase; type I iodothyronine deiodinase; type II iodothyronine deiodinase; thyroxine 5-deiodinase [misleading]; L-thyroxine iodohydrolase (reducing)
Systematic name: 3,3',5-triiodo-L-thyronine,iodide:acceptor oxidoreductase (iodinating)
Comments: The enzyme activity has only been demonstrated in the direction of 5'-deiodination, which renders the thyroid hormone more active. The enzyme consists of type I and type II enzymes, both containing selenocysteine, but with different kinetics. For the type I enzyme the first reaction is a reductive deiodination converting the -Se-H group of the enzyme into an -Se-I group; the reductant then reconverts this into -Se-H, releasing iodide.
References: [677, 1371, 3930, 2232]

[EC 1.21.99.4 created 1984 as EC 3.8.1.4, transferred 2003 to EC 1.97.1.10, transferred 2015 to EC 1.21.99.4]

EC 1.21.99.5

- Accepted name:** tetrachloroethene reductive dehalogenase
Reaction: trichloroethene + chloride + acceptor = tetrachloroethene + reduced acceptor
Other name(s): tetrachloroethene reductase
Systematic name: acceptor:trichloroethene oxidoreductase (chlorinating)
Comments: This enzyme allows the common pollutant tetrachloroethene to support bacterial growth and is responsible for disposal of a number of chlorinated hydrocarbons. The reaction occurs in the reverse direction. The enzyme also reduces trichloroethene to dichloroethene. Although the physiological reductant is unknown, the supply of reductant in some organisms involves menaquinol, which is reduced by molecular hydrogen via the action of EC 1.12.5.1, hydrogen:quinone oxidoreductase. The enzyme contains a corrinoid and two iron-sulfur clusters. Methyl viologen can act as electron donor *in vitro*.
References: [1696, 1345, 3048, 3758, 3757]

[EC 1.21.99.5 created 2001 as EC 1.97.1.8, transferred 2017 to EC 1.21.99.5]

EC 1.22 Acting on halogen in donors

EC 1.22.1 With NAD⁺ or NADP⁺ as acceptor

[1.22.1.1 Transferred entry. iodothyrosine deiodinase. Now EC 1.21.1.1, iodothyrosine deiodinase]

[EC 1.22.1.1 created 2010, deleted 2015]

EC 1.23 Reducing C-O-C group as acceptor

EC 1.23.1 With NADH or NADPH as donor

EC 1.23.1.1

- Accepted name:** (+)-pinoresinol reductase
Reaction: (+)-lariciresinol + NADP⁺ = (+)-pinoresinol + NADPH + H⁺
Other name(s): pinoresinol/lariciresinol reductase; pinoresinol-lariciresinol reductases; (+)-pinoresinol/(+)-lariciresinol; (+)-pinoresinol-(+)-lariciresinol reductase; PLR
Systematic name: (+)-lariciresinol:NADP⁺ oxidoreductase
Comments: The reaction is catalysed *in vivo* in the opposite direction to that shown. A multifunctional enzyme that further reduces the product to the lignan (–)-secoisolariciresinol [EC 1.23.1.2, (+)-lariciresinol reductase]. Isolated from the plants *Forsythia intermedia* [687, 920], *Thuja plicata* (western red cedar) [1211], *Linum perenne* (perennial flax) [1623] and *Linum corymbulosum* [249]. The 4-*pro-R* hydrogen of NADH is transferred to the 7-*pro-R* position of lariciresinol [687].
References: [687, 920, 1211, 2817, 1623, 249]

[EC 1.23.1.1 created 2013]

EC 1.23.1.2

- Accepted name:** (+)-lariciresinol reductase
Reaction: (–)-secoisolariciresinol + NADP⁺ = (+)-lariciresinol + NADPH + H⁺
Other name(s): pinoresinol/lariciresinol reductase; pinoresinol-lariciresinol reductases; (+)-pinoresinol/(+)-lariciresinol; (+)-pinoresinol-(+)-lariciresinol reductase; PLR
Systematic name: (–)-secoisolariciresinol:NADP⁺ oxidoreductase
Comments: The reaction is catalysed *in vivo* in the opposite direction to that shown. A multifunctional enzyme that also reduces (+)-pinoresinol [EC 1.23.1.1, (+)-pinoresinol reductase]. Isolated from the plants *Forsythia intermedia* [687, 920], *Thuja plicata* (western red cedar) [1211], *Linum perenne* (perennial flax) [1623] and *Linum corymbulosum* [249].
References: [687, 920, 1211, 2817, 1623, 249]

[EC 1.23.1.2 created 2013]

EC 1.23.1.3

- Accepted name:** (–)-pinoresinol reductase
Reaction: (–)-lariciresinol + NADP⁺ = (–)-pinoresinol + NADPH + H⁺
Other name(s): pinoresinol/lariciresinol reductase; pinoresinol-lariciresinol reductases; (–)-pinoresinol-(–)-lariciresinol reductase; PLR
Systematic name: (–)-lariciresinol:NADP⁺ oxidoreductase
Comments: The reaction is catalysed *in vivo* in the opposite direction to that shown. A multifunctional enzyme that usually further reduces the product to (+)-secoisolariciresinol [EC 1.23.1.4, (–)-lariciresinol reductase]. Isolated from the plants *Thuja plicata* (western red cedar) [1211], *Linum perenne* (perennial flax) [1623] and *Arabidopsis thaliana* (thale cress) [2997].
References: [1211, 1623, 2997]

[EC 1.23.1.3 created 2013]

EC 1.23.1.4

- Accepted name:** (–)-lariciresinol reductase
Reaction: (+)-secoisolariciresinol + NADP⁺ = (–)-lariciresinol + NADPH + H⁺

Other name(s): pinoresinol/lariciresinol reductase; pinoresinol-lariciresinol reductases; (-)-pinoresinol-(-)-lariciresinol reductase; PLR
Systematic name: (+)-secoisolariciresinol:NADP⁺ oxidoreductase
Comments: The reaction is catalysed *in vivo* in the opposite direction to that shown. A multifunctional enzyme that also reduces (-)-pinoresinol [EC 1.23.1.3, (-)-pinoresinol reductase]. Isolated from the plants *Thuja plicata* (western red cedar) [1211] and *Linum corymbulosum* [1623].
References: [1211, 1623]

[EC 1.23.1.4 created 2013]

EC 1.23.5 With a quinone or similar compound as acceptor

EC 1.23.5.1

Accepted name: violaxanthin de-epoxidase
Reaction: violaxanthin + 2 L-ascorbate = zeaxanthin + 2 L-dehydroascorbate + 2 H₂O (overall reaction)
(1a) violaxanthin + L-ascorbate = antheraxanthin + L-dehydroascorbate + H₂O
(1b) antheraxanthin + L-ascorbate = zeaxanthin + L-dehydroascorbate + H₂O
Other name(s): VDE
Systematic name: violaxanthin:ascorbate oxidoreductase
Comments: Along with EC 1.14.15.21, zeaxanthin epoxidase, this enzyme forms part of the xanthophyll (or violaxanthin) cycle for controlling the concentration of zeaxanthin in chloroplasts. It is activated by a low pH of the thylakoid lumen (produced by high light intensity). Zeaxanthin induces the dissipation of excitation energy in the chlorophyll of the light-harvesting protein complex of photosystem II. In higher plants the enzyme reacts with *all-trans*-diepoxides, such as violaxanthin, and *all-trans*-monoepoxides, but in the alga *Mantoniella squamata*, only the diepoxides are good substrates.
References: [4739, 3544, 490, 2315, 2362, 1370, 2361]

[EC 1.23.5.1 created 2005 as EC 1.10.99.3, transferred 2015 to EC 1.23.5.1]

EC 1.97 Other oxidoreductases

This subclass contains a single sub-subclass (EC 1.97.1) and is reserved for oxidoreductases not included in the previous categories.

EC 1.97.1 Sole sub-subclass for oxidoreductases that do not belong in the other subclasses

EC 1.97.1.1

Accepted name: chlorate reductase
Reaction: reduced acceptor + chlorate = acceptor + H₂O + chlorite
Other name(s): chlorate reductase C
Systematic name: chlorite:acceptor oxidoreductase
Comments: Flavins or benzylviologen can act as acceptor.
References: [170]

[EC 1.97.1.1 created 1978]

EC 1.97.1.2

Accepted name: pyrogallol hydroxytransferase
Reaction: 1,2,3,5-tetrahydroxybenzene + 1,2,3-trihydroxybenzene = 1,3,5-trihydroxybenzene + 1,2,3,5-tetrahydroxybenzene

Other name(s): 1,2,3,5-tetrahydroxybenzene hydroxyltransferase; 1,2,3,5-tetrahydroxybenzene:pyrogallol transhydroxylase; 1,2,3,5-tetrahydroxybenzene-pyrogallol hydroxyltransferase (transhydroxylase); pyrogallol hydroxyltransferase; 1,2,3,5-tetrahydroxybenzene:1,2,3-trihydroxybenzene hydroxyltransferase
Systematic name: 1,2,3,5-tetrahydroxybenzene:1,2,3-trihydroxybenzene hydroxytransferase
Comments: 1,2,3,5-Tetrahydroxybenzene acts as a co-substrate for the conversion of pyrogallol into phloroglucinol, and for a number of similar isomerizations. The enzyme is provisionally listed here, but might be considered as the basis for a new class in the transferases, analogous to the aminotransferases.
References: [474]

[EC 1.97.1.2 created 1992]

[1.97.1.3 *Transferred entry. sulfur reductase. Now EC 1.12.98.4, sulfhydrogenase, since hydrogen is known to be the electron donor.*]

[EC 1.97.1.3 created 1992, deleted 2013]

EC 1.97.1.4

Accepted name: [formate-C-acetyltransferase]-activating enzyme
Reaction: *S*-adenosyl-L-methionine + dihydroflavodoxin + [formate *C*-acetyltransferase]-glycine = 5'-deoxyadenosine + L-methionine + flavodoxin semiquinone + [formate *C*-acetyltransferase]-glycine-2-yl radical
Other name(s): PFL activase; PFL-glycine:*S*-adenosyl-L-methionine H transferase (flavodoxin-oxidizing, *S*-adenosyl-L-methionine-cleaving); formate acetyltransferase activating enzyme; formate acetyltransferase-glycine dihydroflavodoxin:*S*-adenosyl-L-methionine oxidoreductase (*S*-adenosyl-L-methionine cleaving); pyruvate formate-lyase activating enzyme; pyruvate formate-lyase 1 activating enzyme
Systematic name: [formate *C*-acetyltransferase]-glycine dihydroflavodoxin:*S*-adenosyl-L-methionine oxidoreductase (*S*-adenosyl-L-methionine cleaving)
Comments: An iron-sulfur protein. A single glycine residue in EC 2.3.1.54, formate *C*-acetyltransferase, is oxidized to the corresponding radical by transfer of H from its CH₂ to AdoMet with concomitant cleavage of the latter. The reaction requires Fe²⁺. The first stage is reduction of the AdoMet to give methionine and the 5'-deoxyadenosin-5'-yl radical, which then abstracts a hydrogen radical from the glycine residue.
References: [1176, 4486, 1178]

[EC 1.97.1.4 created 1999, modified 2004]

[1.97.1.5 *Transferred entry. arsenate reductase (glutaredoxin). Now EC 1.20.4.1, arsenate reductase (glutaredoxin)*]

[EC 1.97.1.5 created 2000 deleted 2001]

[1.97.1.6 *Transferred entry. arsenate reductase (donor). Now EC 1.20.99.1, arsenate reductase (donor)*]

[EC 1.97.1.6 created 2000 deleted 2001]

[1.97.1.7 *Transferred entry. methylarsonate reductase. Now EC 1.20.4.2, methylarsonate reductase*]

[EC 1.97.1.7 created 2000, deleted 2001]

[1.97.1.8 *Transferred entry. tetrachloroethene reductive dehalogenase. Now EC 1.21.99.5, tetrachloroethene reductive dehalogenase*]

[EC 1.97.1.8 created 2001, deleted 2017]

EC 1.97.1.9

Accepted name: selenate reductase
Reaction: selenite + H₂O + acceptor = selenate + reduced acceptor
Systematic name: selenite:reduced acceptor oxidoreductase

Comments: The periplasmic enzyme from *Thauera selenatis* is a complex comprising three heterologous subunits (α , β and γ) that contains molybdenum, iron, acid-labile sulfide and heme b as cofactor constituents. Nitrate, nitrite, chlorate and sulfate are not substrates. A number of compounds, including acetate, lactate, pyruvate, and certain sugars, amino acids, fatty acids, di- and tricarboxylic acids, and benzoate can serve as electron donors.

References: [3746, 2597, 2255, 4048]

[EC 1.97.1.9 created 2003]

[1.97.1.10 *Transferred entry. thyroxine 5'-deiodinase. Now EC 1.21.99.4 thyroxine 5'-deiodinase*]

[EC 1.97.1.10 created 1984 as EC 3.8.1.4, transferred 2003 to EC 1.97.1.10, deleted 2015]

[1.97.1.11 *Transferred entry. thyroxine 5-deiodinase. Now EC 1.21.99.3 thyroxine 5-deiodinase.*]

[EC 1.97.1.11 created 2003, deleted 2015]

EC 1.97.1.12

Accepted name: photosystem I
Reaction: reduced plastocyanin + oxidized ferredoxin + $h\nu$ = oxidized plastocyanin + reduced ferredoxin
Systematic name: plastocyanin:ferredoxin oxidoreductase (light-dependent)
Comments: Contains chlorophyll, phylloquinones, carotenoids and [4Fe-4S] clusters. Cytochrome c_6 can act as an alternative electron donor, and flavodoxin as an alternative acceptor in some species.
References: [4168, 4416, 659, 88]

[EC 1.97.1.12 created 2011]

EC 1.98 Enzymes using H_2 as reductant (deleted subclass)

EC 1.98.1 Enzymes using H_2 as reductant (deleted subclass)

[1.98.1.1 *Transferred entry. hydrogenase. Now EC 1.12.7.2, ferredoxin hydrogenase*]

[EC 1.98.1.1 created 1961, deleted 1965]

EC 1.99 Other enzymes using O_2 as oxidant (deleted subclass)

EC 1.99.1 Hydroxylases (now covered by EC 1.14)

[1.99.1.1 *Transferred entry. Now EC 1.12.7.2, ferredoxin hydrogenase*]

[EC 1.99.1.1 created 1961, deleted 1965]

[1.99.1.2 *Transferred entry. Now EC 1.14.16.1, phenylalanine 4-monooxygenase*]

[EC 1.99.1.2 created 1961, deleted 1965]

[1.99.1.3 *Deleted entry. nicotinate 6-hydroxylase*]

[EC 1.99.1.3 created 1961, deleted 1965]

[1.99.1.4 *Deleted entry. tryptophan 5-hydroxylase*]

[EC 1.99.1.4 created 1961, deleted 1965]

[1.99.1.5 *Transferred entry. Now EC 1.14.13.9, kynurenine 3-monooxygenase*]

[EC 1.99.1.5 created 1961, deleted 1965]

[1.99.1.6 Deleted entry. steroid 11 α -hydroxylase]

[EC 1.99.1.6 created 1961, deleted 1965]

[1.99.1.7 Transferred entry. Now EC 1.14.15.4, steroid 11 β -monooxygenase]

[EC 1.99.1.7 created 1961, deleted 1965]

[1.99.1.8 Deleted entry. steroid 6 β -hydroxylase]

[EC 1.99.1.8 created 1961, deleted 1965]

[1.99.1.9 Transferred entry. Now EC 1.14.99.9, steroid 17 α -monooxygenase]

[EC 1.99.1.9 created 1961, deleted 1965]

[1.99.1.10 Deleted entry. steroid 19-hydroxylase]

[EC 1.99.1.10 created 1961, deleted 1965]

[1.99.1.11 Transferred entry. Now EC 1.14.99.10, steroid 21-monooxygenase]

[EC 1.99.1.11 created 1961, deleted 1965]

[1.99.1.12 Deleted entry. alkoxyaryl hydroxylase]

[EC 1.99.1.12 created 1961, deleted 1965]

[1.99.1.13 Deleted entry. squalene cyclohydroxylase, covered by EC 1.14.99.7 (squalene monooxygenase) and by EC 5.4.99.7 (lanosterol synthase)]

[EC 1.99.1.13 created 1961, deleted 1965]

[1.99.1.14 Transferred entry. Now EC 1.13.11.27, 4-hydroxyphenylpyruvate dioxygenase]

[EC 1.99.1.14 created 1961, deleted 1965]

EC 1.99.2 Oxygenases (now covered by EC 1.13)

[1.99.2.1 Transferred entry. Now EC 1.13.11.12, lipoxygenase]

[EC 1.99.2.1 created 1961, deleted 1965]

[1.99.2.2 Transferred entry. Now EC 1.13.11.1, catechol 1,2-dioxygenase]

[EC 1.99.2.2 created 1961, deleted 1965]

[1.99.2.3 Transferred entry. Now EC 1.13.11.3, protocatechuate 3,4-dioxygenase]

[EC 1.99.2.3 created 1961, deleted 1965]

[1.99.2.4 Transferred entry. Now EC 1.13.11.4, gentisate 1,2-dioxygenase]

[EC 1.99.2.4 created 1961, deleted 1965]

[1.99.2.5 Transferred entry. Now EC 1.13.11.5, homogentisate 1,2-dioxygenase]

[EC 1.99.2.5 created 1961, deleted 1965]

[1.99.2.6 Transferred entry. Now EC 1.13.99.1, inositol oxygenase]

[EC 1.99.2.6 created 1961, deleted 1965]

References

- [1] A. Abbruzzese, M.H. Park, and J.E. Folk. Deoxyhypusine hydroxylase from rat testis. Partial purification and characterization. *J. Biol. Chem.*, 261:3085–3089, 1986.
- [2] R.H. Abeles, A.M. Brownstein, and C.H. Randles. α -Hydroxypropionaldehyde, an intermediate in the formation of 1,3-propanediol by *Aerobacter melanogaster*. *Biochim. Biophys. Acta*, 41:530–530, 1960.
- [3] H.-J. Abken and U. Deppenmeier. Purification and properties of an $F_{420}H_2$ dehydrogenase from *Methanosarcina mazei* Gö1. *FEMS Microbiol. Lett.*, 154:231–237, 1997.
- [4] H.J. Abken, M. Tietze, J. Brodersen, S. Bäumer, U. Beifuss, and U. Deppenmeier. Isolation and characterization of methanophenazine and function of phenazines in membrane-bound electron transport of *Methanosarcina mazei* gol. *J. Bacteriol.*, 180:2027–2032, 1998.
- [5] A.P. Abola, M.G. Willits, R.C. Wang, and S.R. Long. Reduction of adenosine-5'-phosphosulfate instead of 3'-phosphoadenosine-5'-phosphosulfate in cysteine biosynthesis by *Rhizobium meliloti* and other members of the family Rhizobiaceae. *J. Bacteriol.*, 181:5280–5287, 1999.
- [6] I.A. Abreu, L.M. Saraiva, J. Carita, H. Huber, K.O. Stetter, D. Cabelli, and M. Teixeira. Oxygen detoxification in the strict anaerobic archaeon *Archaeoglobus fulgidus*: superoxide scavenging by neelaredoxin. *Mol. Microbiol.*, 38:322–334, 2000.
- [7] Y. Achouri, G. Noel, and E. Van Schaftingen. 2-Keto-4-methylthiobutyrate, an intermediate in the methionine salvage pathway, is a good substrate for CtBP1. *Biochem. Biophys. Res. Commun.*, 352:903–906, 2007.
- [8] Y. Achouri, G. Noel, D. Vertommen, M.H. Rider, M. Veiga-Da-Cunha, and E. Van Schaftingen. Identification of a dehydrogenase acting on D-2-hydroxyglutarate. *Biochem. J.*, 381:35–42, 2004.
- [9] Y. Achouri, M.H. Rider, E.V. Schaftingen, and M. Robbi. Cloning, sequencing and expression of rat liver 3-phosphoglycerate dehydrogenase. *Biochem. J.*, 323:365–370, 1997.
- [10] S. Achterholt, H. Priefert, and A. Steinbuchel. Purification and characterization of the coniferyl 2-hydroxy-1,4-benzoquinonealdehyde dehydrogenase from *Pseudomonas* sp. Strain HR199 and molecular characterization of the gene. *J. Bacteriol.*, 180:4387–4391, 1998.
- [11] K. Adachi, Y. Iwayama, H. Tanioka, and Y. Takeda. Purification and properties of homogentisate oxygenase from *Pseudomonas fluorescens*. *Biochim. Biophys. Acta*, 118:88–97, 1966.
- [12] K. Adachi, Y. Takeda, S. Senoh, and H. Kita. Metabolism of *p*-hydroxyphenylacetic acid in *Pseudomonas ovalis*. *Biochim. Biophys. Acta*, 93:483–493, 1964.
- [13] O. Adachi and M. Ameyama. D-Glucose dehydrogenase from *Gluconobacter suboxydans*. *Methods Enzymol.*, 89:159–163, 1982.
- [14] O. Adachi, T. Chiyonobu, E. Shinagawa, K. Matsushita, and M. Ameyama. Crystalline 2-ketogluconate reductase from *Acetobacter ascendens*, the second instance of crystalline enzyme in genus *Acetobacter*. *Agric. Biol. Chem.*, 42:2057–2057, 1978.
- [15] O. Adachi, K. Matsushita, E. Shinagawa, and M. Ameyama. Crystallization and properties of NADP-dependent aldehyde dehydrogenase from *Gluconobacter melanogenus*. *Agric. Biol. Chem.*, 44:155–164, 1980.
- [16] O. Adachi, S. Tanasupawat, N. Yoshihara, H. Toyama, and K. Matsushita. 3-Dehydroquinone production by oxidative fermentation and further conversion of 3-dehydroquinone to the intermediates in the shikimate pathway. *Biosci. Biotechnol. Biochem.*, 67:2124–2131, 2003.
- [17] S. Adak, K.S. Aulak, and D.J. Stuehr. Direct evidence for nitric oxide production by a nitric-oxide synthase-like protein from *Bacillus subtilis*. *J. Biol. Chem.*, 277:16167–16171, 2002.
- [18] S. Adak and T.P. Begley. Dibenzothiophene catabolism proceeds via a flavin- N^5 -oxide intermediate. *J. Am. Chem. Soc.*, 138:6424–6426, 2016.

- [19] S. Adak and T.P. Begley. Flavin- N^5 -oxide: A new, catalytic motif in flavoenzymology. *Arch. Biochem. Biophys.*, 632:4–10, 2017.
- [20] S. Adak and T.P. Begley. RutA-catalyzed oxidative cleavage of the uracil amide involves formation of a flavin- N^5 -oxide. *Biochemistry*, 56:3708–3709, 2017.
- [21] E. Adams. Enzymatic synthesis of histidine from histidinol. *J. Biol. Chem.*, 209:829–846, 1954.
- [22] E. Adams. L-Histidinal, a biosynthetic precursor of histidine. *J. Biol. Chem.*, 217:325–344, 1955.
- [23] E. Adams and A. Goldstone. Hydroxyproline metabolism. III. Enzymatic synthesis of hydroxyproline from Δ^1 -pyrroline-3-hydroxy-5-carboxylate. *J. Biol. Chem.*, 235:3499–3503, 1960.
- [24] E. Adams and A. Goldstone. Hydroxyproline metabolism. IV. Enzymatic synthesis of γ -hydroxyglutamate from Δ^1 -pyrroline-3-hydroxy-5-carboxylate. *J. Biol. Chem.*, 235:3504–3512, 1960.
- [25] E. Adams and G. Rosso. α -Ketoglutaric semialdehyde dehydrogenase of *Pseudomonas*. Properties of the purified enzyme induced by hydroxyproline and of the glucarate-induced and constitutive enzymes. *J. Biol. Chem.*, 242:1803–1814, 1967.
- [26] M.W.W. Adams. The structure and mechanism of iron-hydrogenases. *Biochim. Biophys. Acta*, 1020:115–145, 1990.
- [27] M.W.W. Adams, L.E. Mortenson, and J.-S. Chen. Hydrogenase. *Biochim. Biophys. Acta*, 594:105–176, 1981.
- [28] H.A. Addlesee and C.N. Hunter. Physical mapping and functional assignment of the geranylgeranyl-bacteriochlorophyll reductase gene, *bchP*, of *Rhodobacter sphaeroides*. *J. Bacteriol.*, 181:7248–7255, 1999.
- [29] H.A. Addlesee and C.N. Hunter. *Rhodospirillum rubrum* possesses a variant of the *bchP* gene, encoding geranylgeranyl-bacteriopheophytin reductase. *J. Bacteriol.*, 184:1578–1586, 2002.
- [30] P.R. Afolabi, F. Mohammed, K. Amaratunga, O. Majekodunmi, S.L. Dales, R. Gill, D. Thompson, J.B. Cooper, S.P. Wood, P.M. Goodwin, and C. Anthony. Site-directed mutagenesis and X-ray crystallography of the PQQ-containing quinoprotein methanol dehydrogenase and its electron acceptor, cytochrome *c₁*. *Biochemistry*, 40:9799–9809, 2001.
- [31] T. Agapie, S. Suseno, J.J. Woodward, S. Stoll, R.D. Britt, and M.A. Marletta. NO formation by a catalytically self-sufficient bacterial nitric oxide synthase from *Sorangium cellulosum*. *Proc. Natl. Acad. Sci. USA*, 106:16221–16226, 2009.
- [32] A.K. Agarwal, C. Monder, B. Eckstein, and P.C. White. Cloning and expression of rat cDNA encoding corticosteroid 11 β -dehydrogenase. *J. Biol. Chem.*, 264:18939–18943, 1989.
- [33] V. Agarwal, A.A. El Gamal, K. Yamanaka, D. Poth, R.D. Kersten, M. Schorn, E.E. Allen, and B.S. Moore. Biosynthesis of polybrominated aromatic organic compounds by marine bacteria. *Nat. Chem. Biol.*, 10:640–647, 2014.
- [34] V. Agarwal and B.S. Moore. Enzymatic synthesis of polybrominated dioxins from the marine environment. *ACS Chem. Biol.*, 9:1980–1984, 2014.
- [35] F. Agius, R. Gonzalez-Lamothe, J.L. Caballero, J. Munoz-Blanco, M.A. Botella, and V. Valpuesta. Engineering increased vitamin C levels in plants by overexpression of a D-galacturonic acid reductase. *Nat. Biotechnol.*, 21:177–181, 2003.
- [36] K. Agner. Myeloperoxidase. *Adv. Enzymol.*, 3:137–148, 1943.
- [37] M.U. Agosin and E.C. Weinbach. Partial purification and characterization of the isocitric dehydrogenase from *Trypanosoma cruzi*. *Biochim. Biophys. Acta*, 21:117–126, 1956.
- [38] I. Aguirrezabalaga, C. Olano, N. Allende, L. Rodriguez, A.F. Brana, C. Mendez, and J.A. Salas. Identification and expression of genes involved in biosynthesis of L-oleandrose and its intermediate L-olivose in the oleandomycin producer *Streptomyces antibioticus*. *Antimicrob. Agents Chemother.*, 44:1266–1275, 2000.
- [39] N.K. Ahmed, R.L. Felsted, and N.R. Bachur. Heterogeneity of anthracycline antibiotic carbonyl reductases in mammalian livers. *Biochem. Pharmacol.*, 27:2713–2719, 1978.
- [40] O. Ahrazem, G. Diretto, J. Argandona, A. Rubio-Moraga, J.M. Julve, D. Orzaez, A. Granell, and L. Gomez-Gomez. Evolutionarily distinct carotenoid cleavage dioxygenases are responsible for crocetin production in *Buddleja davidii*. *J. Exp. Bot.*, 68:4663–4677, 2017.

- [41] O. Ahrazem, A. Rubio-Moraga, J. Berman, T. Capell, P. Christou, C. Zhu, and L. Gomez-Gomez. The carotenoid cleavage dioxygenase CCD2 catalysing the synthesis of crocetin in spring crocuses and saffron is a plastidial enzyme. *New Phytol.*, 209:650–663, 2016.
- [42] W. Aik, J.S. Scotti, H. Choi, L. Gong, M. Demetriades, C.J. Schofield, and M.A. McDonough. Structure of human RNA N^6 -methyladenine demethylase ALKBH5 provides insights into its mechanisms of nucleic acid recognition and demethylation. *Nucleic Acids Res.*, 42:4741–4754, 2014.
- [43] T.T. Airene, Y. Nymalm, H. Kidron, D.J. Smith, M. Pihlavisto, M. Salmi, S. Jalkanen, M.S. Johnson, and T.A. Salminen. Crystal structure of the human vascular adhesion protein-1: unique structural features with functional implications. *Protein Sci.*, 14:1964–1974, 2005.
- [44] M.D. Aitken and P.E. Heck. Turnover capacity of coprinus cinereus peroxidase for phenol and monosubstituted phenol. *Biotechnol. Prog.*, 14:487–492, 1998.
- [45] Y. Akakabe, K. Matsui, , and T. Enantioselective α -hydroperoxylation of long-chain fatty acids with crude enzyme of marine green alga *Ulva pertusa*. *Tetrahedron Lett.*, 40:1137–1140, 1999.
- [46] T. Akao, T. Akao, M. Hattori, T. Namba, and K. Kobashi. 3 β -Hydroxysteroid dehydrogenase of *Ruminococcus* sp. from human intestinal bacteria. *J. Biochem.*, 99:1425–1431, 1986.
- [47] T. Akao, T. Akao, M. Hattori, T. Namba, and K. Kobashi. Purification and properties of 3 α -hydroxyglycyrrhetinate dehydrogenase of *Clostridium innocuum* from human intestine. *J. Biochem. (Tokyo)*, 103:504–507, 1988.
- [48] T. Akashi, T. Aoki, and S. Ayabe. Identification of a cytochrome P_{450} cDNA encoding (2S)-flavanone 2-hydroxylase of licorice (*Glycyrrhiza echinata* L.; Fabaceae) which represents licodione synthase and flavone synthase II. *FEBS Lett.*, 431:287–290, 1998.
- [49] T. Akashi, T. Aoki, and S.-I. Ayabe. CYP81E1, a cytochrome P_{450} cDNA of licorice (*Glycyrrhiza echinata* L.), encodes isoflavone 2'-hydroxylase. *Biochem. Biophys. Res. Commun.*, 251:67–70, 1998.
- [50] Å. Åkesson, A. Ehrenberg, and H. Theorell. Old yellow enzyme. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 477–494. Academic Press, New York, 2nd edition, 1963.
- [51] J. Aketagawa, K. Kobayashi, and M. Ishimoto. Purification and properties of thiosulfate reductase from *Desulfovibrio vulgaris*, Miyazaki F. *J. Biochem.*, 97:1025–1032, 1985.
- [52] Y. Akiyama, S. Kamitani, N. Kusukawa, and K. Ito. *In vitro* catalysis of oxidative folding of disulfide-bonded proteins by the *Escherichia coli dsbA (ppfA)* gene product. *J. Biol. Chem.*, 267:22440–22445, 1992.
- [53] F. Al-Mjeni, T. Ju, T.C. Pochapsky, and M.J. Maroney. XAS investigation of the structure and function of Ni in acireductone dioxygenase. *Biochemistry*, 41:6761–6769, 2002.
- [54] F. Alagna, F. Geu-Flores, H. Kries, F. Panara, L. Baldoni, S.E. O'Connor, and A. Osbourn. Identification and characterization of the iridoid synthase involved in oleuropein biosynthesis in olive (*Olea europaea*) fruits. *J. Biol. Chem.*, 291:5542–5554, 2016.
- [55] J. Alam, N. Beyer, and H.W. Liu. Biosynthesis of colitose: expression, purification, and mechanistic characterization of GDP-4-keto-6-deoxy-D-mannose-3-dehydrase (CoLD) and GDP-L-colitose synthase (CoLC). *Biochemistry*, 43:16450–16460, 2004.
- [56] N. Alami and P.C. Hallenbeck. Cloning and characterization of a gene cluster, phsBCDEF, necessary for the production of hydrogen sulfide from thiosulfate by *Salmonella typhimurium*. *Gene*, 156:53–57, 1995.
- [57] B. Alber, M. Olinger, A. Rieder, D. Kockelkorn, B. Jobst, M. Hugler, and G. Fuchs. Malonyl-coenzyme A reductase in the modified 3-hydroxypropionate cycle for autotrophic carbon fixation in archaeal *Metallosphaera* and *Sulfolobus* spp. *J. Bacteriol.*, 188:8551–8559, 2006.
- [58] R.W. Albers and G.J. Koval. Succinic semialdehyde dehydrogenase : purification and properties of the enzyme from monkey brain. *Biochim. Biophys. Acta*, 52:29–35, 1961.
- [59] J.A. Alberta and J.H. Dawson. Purification to homogeneity and initial physical characterization of secondary amine monooxygenase. *J. Biol. Chem.*, 262:11857–11863, 1987.

- [60] J. Albertyn, A. van Tonder, and B.A. Prior. Purification and characterization of glycerol-3-phosphate dehydrogenase of *Saccharomyces cerevisiae*. *FEBS Lett.*, 308:130–132, 1992.
- [61] M. Albrecht, H. Linden, and G. Sandmann. Biochemical characterization of purified ζ -carotene desaturase from *Anabaena* PCC 7120 after expression in *E. coli*. *Eur. J. Biochem.*, 236:115–120, 1996.
- [62] M. Albrecht, A. Ruther, and G. Sandmann. Purification and biochemical characterization of a hydroxyneurosporene desaturase involved in the biosynthetic pathway of the carotenoid spheroidene in *Rhodobacter sphaeroides*. *J. Bacteriol.*, 179:7462–7467, 1997.
- [63] A. Alder, M. Jamil, M. Marzorati, M. Bruno, M. Vermathen, P. Bigler, S. Ghisla, H. Bouwmeester, P. Beyer, and S. Al-Babili. The path from β -carotene to carlactone, a strigolactone-like plant hormone. *Science*, 335:1348–1351, 2012.
- [64] N.L. Alderson, B.M. Rembiesa, M.D. Walla, A. Bielawska, J. Bielawski, and H. Hama. The human FA2H gene encodes a fatty acid 2-hydroxylase. *J. Biol. Chem.*, 279:48562–48568, 2004.
- [65] K. Alexander, M. Akhtar, R.B. Boar, J.F. McGhie, and D.H.R. Barton. The removal of the 32-carbon atom as formic acid in cholesterol biosynthesis. *J. Chem. Soc. Chem. Commun.*, pages 383–385, 1972.
- [66] I. Alexeev, A. Sultana, P. Mantsala, J. Niemi, and G. Schneider. Aclacinomycin oxidoreductase (AknOx) from the biosynthetic pathway of the antibiotic aclacinomycin is an unusual flavoenzyme with a dual active site. *Proc. Natl. Acad. Sci. USA*, 104:6170–6175, 2007.
- [67] A. Alhapel, D.J. Darley, N. Wagener, E. Eckel, N. Elsner, and A.J. Pierik. Molecular and functional analysis of nicotinate catabolism in *Eubacterium barkeri*. *Proc. Natl. Acad. Sci. USA*, 103:12341–12346, 2006.
- [68] J.R. Allen, D.D. Clark, J.G. Krum, and S.A. Ensign. A role for coenzyme M (2-mercaptoethanesulfonic acid) in a bacterial pathway of aliphatic epoxide carboxylation. *Proc. Natl. Acad. Sci. USA*, 96:8432–8437, 1999.
- [69] S.H.G. Allen. The isolation and characterization of malate-lactate transhydrogenase from *Micrococcus lactilyticus*. *J. Biol. Chem.*, 241:5266–5275, 1966.
- [70] S.H.G. Allen and J.R. Patil. Studies on the structure and mechanism of action of the malate-lactate transhydrogenase. *J. Biol. Chem.*, 247:909–916, 1972.
- [71] K.H. Almabruk, S. Asamizu, A. Chang, S.G. Varghese, and T. Mahmud. The α -ketoglutarate/Fe(II)-dependent dioxygenase VldW is responsible for the formation of validamycin B. *ChemBioChem*, 13:2209–2211, 2012.
- [72] S. Aloi, C.G. Davies, P.A. Karplus, S.M. Wilbanks, and G.N.L. Jameson. Substrate specificity in thiol dioxygenases. *Biochemistry*, 58:2398–2407, 2019.
- [73] J.M. Alonso and A. Garrido-Pertierra. Carboxymethylhydroxybutyric semialdehyde dehydrogenase in the 4-hydroxyphenylacetate catabolic pathway of *Escherichia coli*. *Biochim. Biophys. Acta*, 719:165–167, 1982.
- [74] M.S. Alphey, W. Yu, E. Byres, D. Li, and W.N. Hunter. Structure and reactivity of human mitochondrial 2,4-dienoyl-CoA reductase: enzyme-ligand interactions in a distinctive short-chain reductase active site. *J. Biol. Chem.*, 280:3068–3077, 2005.
- [75] A.M. Altschul, R. Abrams, and T.R. Hogness. Cytochrome c peroxidase. *J. Biol. Chem.*, 136:777–794, 1940.
- [76] M. Amann, N. Nagakura, and M.H. Zenk. (S)-Tetrahydroprotoberberine oxidase the final enzyme in protoberberine biosynthesis. *Tetrahedron Lett.*, 25:953–954, 1984.
- [77] M. Ameyama and O. Adachi. 5-Keto-D-gluconate reductase from *Gluconobacter suboxydans*. *Methods Enzymol.*, 89:198–202, 1982.
- [78] M. Ameyama and O. Adachi. Aldehyde dehydrogenase from acetic acid bacteria, membrane-bound. *Methods Enzymol.*, 89:491–497, 1982.
- [79] M. Ameyama and O. Adachi. D-Fructose dehydrogenase from *Gluconobacter industrius*, membrane-bound. *Methods Enzymol.*, 89:154–159, 1982.

- [80] M. Ameyama, K. Matsushita, Y. Ohno, E. Shinagawa, and O. Adachi. Existence of a novel prosthetic group, PQQ, in membrane-bound, electron transport chain-linked, primary dehydrogenases of oxidative bacteria. *FEBS Lett.*, 130:179–183, 1981.
- [81] M. Ameyama, K. Matsushita, E. Shinagawa, and O. Adachi. 5-keto-D-Fructose reductase of *Gluconobacter industrius*. Purification, crystallization and properties. *Agric. Biol. Chem.*, 45:863–869, 1981.
- [82] M. Ameyama, K. Osada, E. Shinagawa, K. Matsushita, and O. Adachi. Purification and characterization of aldehyde dehydrogenase of *Acetobacter aceti*. *Agric. Biol. Chem.*, 45:1189–1890, 1981.
- [83] M. Ameyama, E. Shinagawa, K. Matsushita, K. Takimoto, K. Nakashima, and O. Adachi. Mammalian choline dehydrogenase is a quinoprotein. *Agric. Biol. Chem.*, 49:3623–3626, 1985.
- [84] M. Ameyama, E. Shinagawa, K. Matsushita, and O. Adachi. Solubilization, purification and properties of membrane-bound glycerol dehydrogenase from *Gluconobacter industrius*. *Agric. Biol. Chem.*, 49:1001–1010, 1985.
- [85] R.M. Amiri, N.O. Yur'eva, K.R. Shimshilashvili, I.V. Goldenkova-Pavlova, V.P. Pchelkin, E.I. Kuznitsova, V.D. Tsydendambaev, T.I. Trunova, D.A. Los, G.S. Jouzani, and A.M. Nosov. Expression of acyl-lipid Δ^{12} -desaturase gene in prokaryotic and eukaryotic cells and its effect on cold stress tolerance of potato. *J. Integr. Plant Biol.*, 52:289–297, 2010.
- [86] O.Y. Ampomah, A. Avetisyan, E. Hansen, J. Svenson, T. Huser, J.B. Jensen, and T.V. Bhuvaneshwari. The *thuEFGKAB* operon of Rhizobia and *Agrobacterium tumefaciens* codes for transport of trehalose, maltitol, and isomers of sucrose and their assimilation through the formation of their 3-keto derivatives. *J. Bacteriol.*, 195:3797–3807, 2013.
- [87] O.Y. Ampomah and J.B. Jensen. The trehalose utilization gene *thuA* ortholog in *Mesorhizobium loti* does not influence competitiveness for nodulation on *Lotus* spp. *World J. Microbiol. Biotechnol.*, 30:1129–1134, 2014.
- [88] A. Amunts, H. Toporik, A. Borovikova, and N. Nelson. Structure determination and improved model of plant photosystem I. *J. Biol. Chem.*, 285:3478–3486, 2010.
- [89] M. Andberg, H. Maaheimo, H. Boer, M. Penttila, A. Koivula, and P. Richard. Characterization of a novel *Agrobacterium tumefaciens* galactarolactone cycloisomerase enzyme for direct conversion of D-galactarolactone to 3-deoxy-2-keto-L-threo-hexarate. *J. Biol. Chem.*, 287:17662–17671, 2012.
- [90] M.D. Andersen, P.K. Busk, I. Svendsen, and B.L. Møller. Cytochromes *P*-450 from cassava (*Manihot esculenta* Crantz) catalyzing the first steps in the biosynthesis of the cyanogenic glucosides linamarin and lotaustralin. Cloning, functional expression in *Pichia pastoris*, and substrate specificity of the isolated recombinant enzymes. *J. Biol. Chem.*, 275:1966–1975, 2000.
- [91] O.A. Andersen, T. Flatmark, and E. Hough. High resolution crystal structures of the catalytic domain of human phenylalanine hydroxylase in its catalytically active Fe(II) form and binary complex with tetrahydrobiopterin. *J. Mol. Biol.*, 314:266–278, 2001.
- [92] S.J. Andersen, S. Quan, B. Gowan, and E.R. Dabbs. Monooxygenase-like sequence of a *Rhodococcus equi* gene conferring increased resistance to rifampin by inactivating this antibiotic. *Antimicrob. Agents Chemother.*, 41:218–221, 1997.
- [93] G.L. Anderson, J. Williams, and R. Hille. The purification and characterization of arsenite oxidase from *Alcaligenes faecalis*, a molybdenum-containing hydroxylase. *J. Biol. Chem.*, 267:23674–23682, 1992.
- [94] J.M. Anderson, H. Charbonneau, and M.J. Cormier. Mechanism of calcium induction of *Renilla* bioluminescence. Involvement of a calcium-triggered luciferin binding protein. *Biochemistry*, 13:1195–1200, 1974.
- [95] M.P. Anderson, C.P. Vance, G.H. Heichel, and S.S. Miller. Purification and characterization of NADH-glutamate synthase from alfalfa root nodules. *Plant Physiol.*, 90:351–358, 1989.
- [96] S. Andersson, D.L. Davis, H. Dahlbäck, H. Jörnvall, and D.W. Russell. Cloning, structure, and expression of the mitochondrial cytochrome *P*-450 sterol 26-hydroxylase, a bile acid biosynthetic enzyme. *J. Biol. Chem.*, 264:8222–8229, 1989.
- [97] S.L. Andrade, C.D. Brondino, M.J. Feio, I. Moura, and J.J. Moura. Aldehyde oxidoreductase activity in *Desulfovibrio alaskensis* NCIMB 13491. EPR assignment of the proximal [2Fe-2S] cluster to the Mo site. *Eur. J. Biochem.*, 267:2054–2061, 2000.

- [98] J.R. Andreesen and L.G. Ljungdahl. Nicotinamide adenine dinucleotide phosphate-dependent formate dehydrogenase from *Clostridium thermoaceticum*: purification and properties. *J. Bacteriol.*, 120:6–14, 1974.
- [99] A.Z. Andreou, E. Hornung, S. Kunze, S. Rosahl, and I. Feussner. On the substrate binding of linoleate 9-lipoxygenases. *Lipids*, 44:207–215, 2009.
- [100] A.Z. Andreou, M. Vanko, L. Bezakova, and I. Feussner. Properties of a mini 9R-lipoxygenase from *Nostoc* sp. PCC 7120 and its mutant forms. *Phytochemistry*, 69:1832–1837, 2008.
- [101] N. Andrés, J.M. Lizcano, M.J. Rodríguez, M. Romera, M. Unzeta, and N. Mahy. Tissue activity and cellular localization of human semicarbazide-sensitive amine oxidase. *J. Histochem. Cytochem.*, 49:209–217, 2001.
- [102] B.S. Andresen, E. Christensen, T.J. Corydon, P. Bross, B. Pilgaard, R.J. Wanders, J.P. Ruiten, H. Simonsen, V. Winter, I. Knudsen, L.D. Schroeder, N. Gregersen, and F. Skovby. Isolated 2-methylbutyryl-glycinuria caused by short/branched-chain acyl-CoA dehydrogenase deficiency: identification of a new enzyme defect, resolution of its molecular basis, and evidence for distinct acyl-CoA dehydrogenases in isoleucine and valine metabolism. *Am. J. Hum. Genet.*, 67:1095–1103, 2000.
- [103] A. Angelov, O. Futterer, O. Valerius, G.H. Braus, and W. Liebl. Properties of the recombinant glucose/galactose dehydrogenase from the extreme thermoacidophile, *Picrophilus torridus*. *FEBS J.*, 272:1054–1062, 2005.
- [104] E. Änggård and B. Samuelsson. Purification and properties of a 15-hydroxyprostaglandin dehydrogenase from swine lung. *Prostaglandins*, 25:293–300, 1996.
- [105] D.H. Anh, R. Ullrich, D. Benndorf, A. Svatos, A. Muck, and M. Hofrichter. The coprophilous mushroom *Coprinus radians* secretes a haloperoxidase that catalyzes aromatic peroxygenation. *Appl. Environ. Microbiol.*, 73:5477–5485, 2007.
- [106] R. Ansell, K. Granath, S. Hohmann, J.M. Thevelein, and L. Adler. The two isoenzymes for yeast NAD⁺-dependent glycerol 3-phosphate dehydrogenase encoded by GPD1 and GPD2 have distinct roles in osmoadaptation and redox regulation. *EMBO J.*, 16:2179–2187, 1997.
- [107] C. Anthony and P. Williams. The structure and mechanism of methanol dehydrogenase. *Biochim. Biophys. Acta*, 1647:18–23, 2003.
- [108] C. Anthony and L.J. Zatman. The microbial oxidation of methanol. 2. The methanol-oxidizing enzyme of *Pseudomonas* sp. M 27. *Biochem. J.*, 92:614–621, 1964.
- [109] C. Anthony and L.J. Zatman. The microbial oxidation of methanol. The prosthetic group of the alcohol dehydrogenase of *Pseudomonas* sp. M27: a new oxidoreductase prosthetic group. *Biochem. J.*, 104:960–969, 1967.
- [110] I.A. Anton and J.R. Coggins. Sequencing and overexpression of the *Escherichia coli* *aroE* gene encoding shikimate dehydrogenase. *Biochem. J.*, 249:319–326, 1988.
- [111] Y. Anzai, N. Saito, M. Tanaka, K. Kinoshita, Y. Koyama, and F. Kato. Organization of the biosynthetic gene cluster for the polyketide macrolide mycinamicin in *Micromonospora griseorubida*. *FEMS Microbiol. Lett.*, 218:135–141, 2003.
- [112] D.G. Vass ao, S.J. Kim, J.K. Milhollan, D. Eichinger, L.B. Davin, and N.G. Lewis. A pinoresinol-lariciresinol reductase homologue from the creosote bush (*Larrea tridentata*) catalyzes the efficient *in vitro* conversion of *p*-coumaryl/coniferyl alcohol esters into the allylphenols chavicol/eugenol, but not the propenylphenols *p*-anol/isoegenol. *Arch. Biochem. Biophys.*, 465:209–218, 2007.
- [113] T. Aoyama, M. Souri, S. Ushikubo, T. Kamijo, S. Yamaguchi, R.I. Kelley, W.J. Rhead, K. Uetake, K. Tanaka, and T. Hashimoto. Purification of human very-long-chain acyl-coenzyme A dehydrogenase and characterization of its deficiency in seven patients. *J. Clin. Invest.*, 95:2465–2473, 1995.
- [114] Y. Aoyama and Y. Yoshida. Different substrate specificities of lanosterol 14 α -demethylase (*P*-450_{14DM}) of *Saccharomyces cerevisiae* and rat liver of 24-methylene-24,25-dihydrolanosterol and 24,25-dihydrolanosterol. *Biochem. Biophys. Res. Commun.*, 178:1064–1071, 1991.
- [115] Y. Aoyama and Y. Yoshida. The 4 β -methyl group of substrate does not affect the activity of lanosterol 14 α -demethylase (*P*₄₅₀14DM) of yeast: differences between the substrate recognition by yeast and plant sterol 14 α -demethylases. *Biochem. Biophys. Res. Commun.*, 183:1266–1272, 1992.

- [116] Y. Aoyama, Y. Yoshida, and R. Sato. Yeast cytochrome *P*-450 catalyzing lanosterol 14 α -demethylation. II. Lanosterol metabolism by purified *P*-450¹⁴DM and by intact microsomes. *J. Biol. Chem.*, 259:1661–1666, 1984.
- [117] K. Apel, H.-J. Santel, T.E. Redlinger, and H. Falk. The protochlorophyllide holochrome of barley (*Hordeum vulgare* L.). Isolation and characterization of the NADPH:protochlorophyllide oxidoreductase. *Eur. J. Biochem.*, 111:251–258, 1980.
- [118] C. Appia-Ayme, A. Bengrine, C. Cavazza, M.T. Giudici-Ortoni, M. Bruschi, M. Chippaux, and V. Bonnefoy. Characterization and expression of the co-transcribed *cyc1* and *cyc2* genes encoding the cytochrome *c*₄ (*c*₅₅₂) and a high-molecular-mass cytochrome *c* from *Thiobacillus ferrooxidans* ATCC 33020. *FEMS Microbiol. Lett.*, 167:171–177, 1998.
- [119] C.A. Appleby and R.K. Morton. Lactic dehydrogenase and cytochrome *b*₂ of baker's yeast. Enzymic and chemical properties of the crystalline enzyme. *Biochem. J.*, 73:539–550, 1959.
- [120] C.A. Appleby and R.K. Morton. Lactic dehydrogenase and cytochrome *b*₂ of baker's yeast. Purification and crystallization. *Biochem. J.*, 71:492–499, 1959.
- [121] H. Arakawa, , and W.G. , Psenak. M. and Coscia, C.J. Purification and characterization of dihydrobenzophenanthridine oxidase from elicited *Sanguinaria canadensis* cell cultures. *Arch. Biochem. Biophys.*, 299:1–7, 1992.
- [122] H. Aramaki, H. Koga, Y. Sagara, M. Hosoi, and T. Horiuchi. Complete nucleotide sequence of the 5-*exo*-hydroxycamphor dehydrogenase gene on the CAM plasmid of *Pseudomonas putida* (ATCC 17453). *Biochim. Biophys. Acta*, 1174:91–94, 1993.
- [123] E. Aranda, M. Kinne, M. Kluge, R. Ullrich, and M. Hofrichter. Conversion of dibenzothiophene by the mushrooms *Agrocybe aegerita* and *Coprinellus radians* and their extracellular peroxygenases. *Appl. Microbiol. Biotechnol.*, 82:1057–1066, 2009.
- [124] M. Arase, M.R. Waterman, and N. Kagawa. Purification and characterization of bovine steroid 21-hydroxylase (P450c21) efficiently expressed in *Escherichia coli*. *Biochem. Biophys. Res. Commun.*, 344:400–405, 2006.
- [125] H. Arata, M. Shimizu, and K. Takamiya. Purification and properties of trimethylamine *N*-oxide reductase from aerobic photosynthetic bacterium *Roseobacter denitrificans*. *J. Biochem. (Tokyo)*, 112:470–475, 1992.
- [126] Z. Araya, U. Hellman, and R. Hansson. Characterisation of taurochenodeoxycholic acid 6 α -hydroxylase from pig liver microsomes. *Eur. J. Biochem.*, 231:855–861, 1995.
- [127] Z. Araya and K. Wikvall. 6 α -Hydroxylation of taurochenodeoxycholic acid and lithocholic acid by CYP3A4 in human liver microsomes. *Biochim. Biophys. Acta*, 1438:47–54, 1999.
- [128] C. Archer, P.J. Ashman, P. Hedden, J.R. Bowyer, and P.M. Bramley. Purification of *ent*-kaurene oxidase from *Gibberella fujikuroi* and *Cucurbita maxima*. *Biochem. Soc. Trans.*, 20:218–218, 1992.
- [129] A.C. Arcus and N.L. Edson. Polyol dehydrogenases. 2. The polyol dehydrogenases of *Acetobacter suboxydans* and *Candida utilis*. *Biochem. J.*, 64:385–394, 1956.
- [130] S.M. Arfin and H.E. Umbarger. Purification and properties of the acetohydroxy acid isomeroreductase of *Salmonella typhimurium*. *J. Biol. Chem.*, 244:1118–1127, 1969.
- [131] N. Arfman, E.M. Watling, W. Clement, R.J. van Oosterwijk, G.E. de Vries, W. Harder, M.M. Attwood, and L. Dijkhuizen. Methanol metabolism in thermotolerant methylotrophic *Bacillus* strains involving a novel catabolic NAD-dependent methanol dehydrogenase as a key enzyme. *Arch. Microbiol.*, 152:280–288, 1989.
- [132] B. Arieli, Y. Shahak, D. Taglicht, G. Hauska, and E. Padan. Purification and characterization of sulfide-quinone reductase, a novel enzyme driving anoxygenic photosynthesis in *Oscillatoria limnetica*. *J. Biol. Chem.*, 269:5705–5711, 1994.
- [133] E.S. Arner and A. Holmgren. Physiological functions of thioredoxin and thioredoxin reductase. *Eur. J. Biochem.*, 267:6102–6109, 2000.
- [134] R.J. Arner, K.S. Prabhu, J.T. Thompson, G.R. Hildenbrandt, A.D. Liken, and C.C. Reddy. *myo*-Inositol oxygenase: molecular cloning and expression of a unique enzyme that oxidizes *myo*-inositol and *D-chiro*-inositol. *Biochem. J.*, 360:313–320, 2001.

- [135] N. Aro-Karkkainen, M. Toivari, H. Maaheimo, M. Ylilauri, O.T. Pentikainen, M. Andberg, M. Oja, M. Penttila, M.G. Wiebe, L. Ruohonen, and A. Koivula. L-arabinose/D-galactose 1-dehydrogenase of *Rhizobium leguminosarum* bv. trifolii characterised and applied for bioconversion of L-arabinose to L-arabonate with *Saccharomyces cerevisiae*. *Appl. Microbiol. Biotechnol.*, 98:9653–9665, 2014.
- [136] V. Arondel, B. Lemieux, I. Hwang, S. Gibson, H.M. Goodman, and C.R. Somerville. Map-based cloning of a gene controlling ω -3 fatty acid desaturation in *Arabidopsis*. *Science*, 258:1353–1355, 1992.
- [137] D.J. Arp, L.A. Sayavedra-Soto, and N.G. Hommes. Molecular biology and biochemistry of ammonia oxidation by *Nitrosomonas europaea*. *Arch. Microbiol.*, 178:250–255, 2002.
- [138] B.A. Arthington, L.G. Bennett, P.L. Skatrud, C.J. Guynn, R.J. Barbuch, C.E. Ulbright, and M. Bard. Cloning, disruption and sequence of the gene encoding yeast C-5 sterol desaturase. *Gene*, 102:39–44, 1991.
- [139] T. Asami, M. Mizutani, S. Fujioka, H. Goda, Y.K. Min, Y. Shimada, T. Nakano, S. Takatsuto, T. Matsuyama, N. Nagata, K. Sakata, and S. Yoshida. Selective interaction of triazole derivatives with DWF4, a cytochrome P450 monooxygenase of the brassinosteroid biosynthetic pathway, correlates with brassinosteroid deficiency in planta. *J. Biol. Chem.*, 276:25687–25691, 2001.
- [140] T. Asami, M. Mizutani, Y. Shimada, H. Goda, N. Kitahata, K. Sekimata, S.Y. Han, S. Fujioka, S. Takatsuto, K. Sakata, and S. Yoshida. Triadimefon, a fungicidal triazole-type P450 inhibitor, induces brassinosteroid deficiency-like phenotypes in plants and binds to DWF4 protein in the brassinosteroid biosynthesis pathway. *Biochem. J.*, 369:71–76, 2003.
- [141] Y. Asano, A. Nakazawa, and K. Endo. Novel phenylalanine dehydrogenases from *Sporosarcina ureae* and *Bacillus sphaericus*. Purification and characterization. *J. Biol. Chem.*, 262:10346–10354, 1987.
- [142] Y. Asano, A. Nakazawa, K. Endo, Y. Hibino, M. Ohmori, N. Numao, and K. Kondo. Phenylalanine dehydrogenase of *Bacillus badius*. Purification, characterization and gene cloning. *Eur. J. Biochem.*, 168:153–159, 1987.
- [143] Y. Asano, K. Yamaguchi, and K. Kondo. A new NAD⁺-dependent opine dehydrogenase from *Arthrobacter* sp. strain 1C. *J. Bacteriol.*, 171:4466–4471, 1989.
- [144] M. Ashikari, H. Sakakibara, S. Lin, T. Yamamoto, T. Takashi, A. Nishimura, E.R. Angeles, Q. Qian, H. Kitano, and M. Matsuoka. Cytokinin oxidase regulates rice grain production. *Science*, 309:741–745, 2005.
- [145] G.W. Ashley, G. Harris, and J. Stubbe. The mechanism of *Lactobacillus leichmannii* ribonucleotide reductase. Evidence for 3' carbon-hydrogen bond cleavage and a unique role for coenzyme B₁₂. *J. Biol. Chem.*, 261:3958–3964, 1986.
- [146] P.J. Ashman, A. Mackenzie, and P.M. Bramley. Characterization of ent-kaurene oxidase activity from *Gibberella fujikuroi*. *Biochim. Biophys. Acta*, 1036:151–157, 1990.
- [147] P. Askerlund, C. Larrson, and S. Widell. Localization of donor and acceptor sites of NADH dehydrogenase activities using inside-out and right-side-out plasma membrane vesicles from plants. *FEBS Lett.*, 239:23–28, 1988.
- [148] R.E. Asnis and A.F. Brodie. A glycerol dehydrogenase from *Escherichia coli*. *J. Biol. Chem.*, 203:153–159, 1953.
- [149] A.J. Aspen and W.B. Jakoby. L-Threonic acid dehydrogenase: purification and properties. *J. Biol. Chem.*, 239:710–713, 1964.
- [150] J. Asteinza, R. Camacho-Carranza, R.E. Reyes-Reyes, V. Dorado-Gonzalez, Espinosa-Aguirre V., and J.J. Induction of cytochrome P450 enzymes by albendazole treatment in the rat. *Environ Toxicol Pharmacol*, 9:31–37, 2000.
- [151] A. Atawong, M. Hasegawa, and O. Kodama. Biosynthesis of rice phytoalexin: enzymatic conversion of 3 β -hydroxy-9 β -pimara-7,15-dien-19,6 β -olide to momilactone A. *Biosci. Biotechnol. Biochem.*, 66:566–570, 2002.
- [152] M.A. Attwood and C.C. Doughty. Purification and properties of calf liver aldose reductase. *Biochim. Biophys. Acta*, 370:358–368, 1974.
- [153] R.J. Auchus, T.C. Lee, and W.L. Miller. Cytochrome b₅ augments the 17,20-lyase activity of human P450c17 without direct electron transfer. *J. Biol. Chem.*, 273:3158–3165, 1998.
- [154] S.W. Aufhammer, E. Warkentin, H. Berk, S. Shima, R.K. Thauer, and U. Ermler. Coenzyme binding in F₄₂₀-dependent secondary alcohol dehydrogenase, a member of the bacterial luciferase family. *Structure*, 12:361–370, 2004.

- [155] P. Augustin, A. Hromic, T. Pavkov-Keller, K. Gruber, and P. Macheroux. Structure and biochemical properties of recombinant human dimethylglycine dehydrogenase and comparison to the disease-related H109R variant. *FEBS J.*, 283:3587–3603, 2016.
- [156] G.D. Aurbach and W.B. Jakoby. The multiple functions of thiooxidase. *J. Biol. Chem.*, 237:565–568, 1962.
- [157] H. Aurich, H.-P. Kleber, H. Sorger, and H. Tauchert. Reinigung und Eigenschaften der Carnitindehydrogenase aus *Pseudomonas aeruginosa*. *Eur. J. Biochem.*, 6:196–201, 1968.
- [158] G. Avigad, Y. Alroy, and S. England. Purification and properties of a nicotinamide adenine dinucleotide phosphate-linked aldohexose dehydrogenase from *Gluconobacter cerinus*. *J. Biol. Chem.*, 243:1936–1941, 1968.
- [159] G. Avigad, D. Amaral, C. Asensio, and B.L. Horecker. The D-galactose oxidase of *Polyporus circinatus*. *J. Biol. Chem.*, 237:2736–2743, 1962.
- [160] G. Avigad, S. England, and S. Pifco. 5-keto-D-Fructose. IV. A specific reduced nicotinamide adenine dinucleotide phosphate-linked reductase from *Gluconobacter cerinus*. *J. Biol. Chem.*, 241:373–378, 1966.
- [161] P.G. Avis, F. Bergel, and R.C. Bray. Cellular constituents. The chemistry of xanthine oxidase. Part I. The preparation of a crystalline xanthine oxidase from cow's milk. *J. Chem. Soc. (Lond.)*, pages 1100–1105, 1955.
- [162] M. Avron and A.T. Jagendorf. Some further investigations on chloroplast TPNH diaphorase. *Arch. Biochem. Biophys.*, 72:17–24, 1957.
- [163] B.C. Axcell and P.J. Geary. The metabolism of benzene by bacteria. Purification and some properties of the enzyme *cis*-1,2-dihydroxycyclohexa-3,5-diene (nicotinamide adenine dinucleotide) oxidoreductase (*cis*-benzene glycol dehydrogenase). *Biochem. J.*, 136:927–934, 1973.
- [164] M.J. Axley, D.A. Grahame, and T.C. Stadtman. *Escherichia coli* formate-hydrogen lyase. Purification and properties of the selenium-dependent formate dehydrogenase component. *J. Biol. Chem.*, 265:18213–18218, 1990.
- [165] P.K. Ayengar, O. Hayaishi, M. Nakajima, and I. Tomida. Enzymic aromatization of 3,5-cyclohexadiene-1,2-diol. *Biochim. Biophys. Acta*, 33:111–119, 1959.
- [166] A.R. Ayers, S.B. Ayers, and K.-E. Eriksson. Cellobiose oxidase, purification and partial characterization of a hemoprotein from *Sporotrichum pulverulentum*. *Eur. J. Biochem.*, 90:171–181, 1978.
- [167] A.R. Ayers and K.-E. Eriksson. Cellobiose oxidase from *Sporotrichum pulverulentum*. *Methods Enzymol.*, 89:129–135, 1982.
- [168] R. Ayikpoe, T. Ngendahimana, M. Langton, S. Bonitatibus, L.M. Walker, S.S. Eaton, G.R. Eaton, M.E. Pandelia, S.J. Elliott, and J.A. Latham. Spectroscopic and electrochemical characterization of the mycofactocin biosynthetic protein, MftC, provides insight into its redox flipping mechanism. *Biochemistry*, 58:940–950, 2019.
- [169] R.S. Ayikpoe and J.A. Latham. MftD catalyzes the formation of a biologically active redox center in the biosynthesis of the ribosomally synthesized and post-translationally modified redox cofactor mycofactocin. *J. Am. Chem. Soc.*, 141:13582–13591, 2019.
- [170] E. Azoulay, S. Mutaftshiev, and M.L. de Sousa. Étude des mutants chlorate-résistants chez *Escherichia coli* K12. III. Mise en évidence et étude de l'activité chlorate-réductase c des mutants chl. C-. *Biochim. Biophys. Acta*, 237:579–590, 1971.
- [171] S.G. Bach, M. Dixon, and L.G. Zervas. Yeast lactic dehydrogenase and cytochrome *b*₂. *Biochem. J.*, 40:229–239, 1946.
- [172] B.K. Bachhawat, W.G. Robinson, and M.J. Coon. Enzymatic carboxylation of β -hydroxyisovaleryl coenzyme A. *J. Biol. Chem.*, 219:539–550, 1956.
- [173] A. Bachmann, B. Hause, H. Maucher, E. Garbe, K. Voros, H. Weichert, C. Wasternack, and I. Feussner. Jasmonate-induced lipid peroxidation in barley leaves initiated by distinct 13-LOX forms of chloroplasts. *Biol. Chem.*, 383:1645–1657, 2002.
- [174] B. Bader, W. Knecht, M. Fries, and M. Löffler. Expression, purification, and characterization of histidine-tagged rat and human flavoenzyme dihydroorotate dehydrogenase. *Protein Expr. Purif.*, 13:414–422, 1998.

- [175] M. Bader, W. Muse, T. Zander, and J. Bardwell. Reconstitution of a protein disulfide catalytic system. *J. Biol. Chem.*, 273:10302–10307, 1998.
- [176] J. Bae, S.M. Kim, and S.B. Lee. Identification and characterization of 2-keto-3-deoxy-L-rhamnonate dehydrogenase belonging to the MDR superfamily from the thermoacidophilic bacterium *Sulfobacillus thermosulfidooxidans*: implications to L-rhamnose metabolism in archaea. *Extremophiles*, 19:469–478, 2015.
- [177] S.H. Bae and Y.K. Paik. Cholesterol biosynthesis from lanosterol: development of a novel assay method and characterization of rat liver microsomal lanosterol Δ^{24} -reductase. *Biochem. J.*, 326:609–616, 1997.
- [178] Y.M. Bae and J.W. Hastings. Cloning, sequencing and expression of dinoflagellate luciferase DNA from a marine alga, *Gonyaulax polyedra*. *Biochim. Biophys. Acta*, 1219:449–456, 1994.
- [179] B.L. Baginsky and V.W. Rodwell. Metabolism of pipercolic acid in a *Pseudomonas* species. V. Pipercolate oxidase and dehydrogenase. *J. Bacteriol.*, 94:1034–1039, 1967.
- [180] A. Baich. The biosynthesis of proline in *Escherichia coli*: phosphate-dependent glutamate-semialdehyde dehydrogenase (NADP), the second enzyme in the pathway. *Biochim. Biophys. Acta*, 244:129–134, 1971.
- [181] A. Baich and H.J. Vogel. *N*-Acetyl- γ -glutamokinase and *N*-acetylglutamic γ -semialdehyde dehydrogenase: repressible enzymes of arginine synthesis in *Escherichia coli*. *Biochem. Biophys. Res. Commun.*, 7:491–496, 1962.
- [182] B.A. Bailey and R.L. Larson. Maize microsomal benzoxazinone *N*-monooxygenase. *Plant Physiol.*, 95:792–796, 1991.
- [183] J.P. Bailey, C. Renz, and E.T. McGuinness. Sorbitol dehydrogenase from horse liver: purification, characterization and comparative properties. *Comp. Biochem. Physiol.*, 69B:909–914, 1981.
- [184] L.J. Bailey, J.F. Acheson, J.G. McCoy, N.L. Elsen, G.N. Phillips, Fox Jr., and B.G. Crystallographic analysis of active site contributions to regioselectivity in the diiron enzyme toluene 4-monooxygenase. *Biochemistry*, 51:1101–1113, 2012.
- [185] J. Bains and M.J. Boulanger. Structural and biochemical characterization of a novel aldehyde dehydrogenase encoded by the benzoate oxidation pathway in *Burkholderia xenovorans* LB400. *J. Mol. Biol.*, 379:597–608, 2008.
- [186] S. Bak, R.A. Kahn, C.E. Olsen, and B.A. Halkier. Cloning and expression in *Escherichia coli* of the obtusifolios 14 α -demethylase of *Sorghum bicolor* (L.) Moench, a cytochrome *P*₄₅₀ orthologous to the sterol 14 α -demethylases (CYP51) from fungi and mammals. *Plant J.*, 11:191–201, 1997.
- [187] S. Bak, C.E. Olsen, B.A. Halkier, and B.L. Møller. Transgenic tobacco and *Arabidopsis* plants expressing the two multifunctional sorghum cytochrome P450 enzymes, CYP79A1 and CYP71E1, are cyanogenic and accumulate metabolites derived from intermediates in Dhurrin biosynthesis. *Plant Physiol.*, 123:1437–1448, 2000.
- [188] S. Bak, F.E. Tax, K.A. Feldmann, D.W. Galbraith, and R. Feyereisen. CYP83B1, a cytochrome P450 at the metabolic branch point in auxin and indole glucosinolate biosynthesis in *Arabidopsis*. *Plant Cell*, 13:101–111, 2001.
- [189] T.-G. Bak. Studies on glucose dehydrogenase of *Aspergillus oryzae*. II. Purification and physical and chemical properties. *Biochim. Biophys. Acta*, 139:277–293, 1967.
- [190] B.J. Baker, J.E. Dotzlar, and W.K. Yeh. Deacetoxycephalosporin C hydroxylase of *Streptomyces clavuligerus*. Purification, characterization, bifunctionality, and evolutionary implication. *J. Biol. Chem.*, 266:5087–5093, 1991.
- [191] D.P. Baker and C.A. Fewson. Purification and characterization of D(–)-mandelate dehydrogenase from *Rhodotorula graminis*. *Microbiology*, 135:2035–2044, 1989.
- [192] D.P. Baker, C. Kleanthous, J.N. Keen, E. Weinhold, and C.A. Fewson. Mechanistic and active-site studies on D(–)-mandelate dehydrogenase from *Rhodotorula graminis*. *Biochem. J.*, 281:211–218, 1992.
- [193] F.C. Baker, B. Mauchamp, L.W. Tsai, and D.A. Schooley. Farnesol and farnesal dehydrogenase(s) in corpora allata of the tobacco hornworm moth, *Manduca sexta*. *J. Lipid Res.*, 24:1586–1594, 1983.
- [194] J.J. Baker, I. Jeng, and H.A. Barker. Purification and properties of L-erythro-3,5-diaminohexanoate dehydrogenase from a lysine-fermenting *Clostridium*. *J. Biol. Chem.*, 247:7724–7734, 1972.
- [195] P. Baker, C. Hillis, J. Carere, and S.Y.K. Seah. Protein-protein interactions and substrate channeling in orthologous and chimeric aldolase-dehydrogenase complexes. *Biochemistry*, 51:1942–1952, 2012.

- [196] P. Baker, D. Pan, J. Carere, A. Rossi, W. Wang, and S.Y.K. Seah. Characterization of an aldolase-dehydrogenase complex that exhibits substrate channeling in the polychlorinated biphenyls degradation pathway. *Biochemistry*, 48:6551–6558, 2009.
- [197] R. Balasubramanian and A.C. Rosenzweig. Structural and mechanistic insights into methane oxidation by particulate methane monooxygenase. *Acc. Chem. Res.*, 40:573–580, 2007.
- [198] S. Bali, A.D. Lawrence, S.A. Lobo, L.M. Saraiva, B.T. Golding, D.J. Palmer, M.J. Howard, S.J. Ferguson, and M.J. Warren. Molecular hijacking of siroheme for the synthesis of heme and d1 heme. *Proc. Natl. Acad. Sci. USA*, 108:18260–18265, 2011.
- [199] C.J. Balibar and C.T. Walsh. *In vitro* biosynthesis of violacein from L-tryptophan by the enzymes VioA-E from *Chromobacterium violaceum*. *Biochemistry*, 45:15444–15457, 2006.
- [200] D. Balinsky and D.D. Davies. Aromatic biosynthesis in higher plants. 1. Preparation and properties of dehydroshikimic reductase. *Biochem. J.*, 80:292–296, 1961.
- [201] N.R. Ballal, P.K. Bhattacharyya, and P.N. Rangachari. Perillyl alcohol dehydrogenase from a soil pseudomonad. *Biochem. Biophys. Res. Commun.*, 23:473–478, 1966.
- [202] R.H. Baltz and E.T. Seno. Properties of *Streptomyces fradiae* mutants blocked in biosynthesis of the macrolide antibiotic tylosin. *Antimicrob. Agents Chemother.*, 20:214–225, 1981.
- [203] A. Bambauer, F.A. Rainey, E. Stackebrandt, and J. Winter. Characterization of *Aquamicrobium defluvii* gen. nov. sp. nov., a thiophene-2-carboxylate-metabolizing bacterium from activated sludge. *Arch. Microbiol.*, 169:293–302, 1998.
- [204] V.A. Bamford, M. Armour, S.A. Mitchell, M. Cartron, S.C. Andrews, and K.A. Watson. Preliminary X-ray diffraction analysis of YqjH from *Escherichia coli*: a putative cytoplasmic ferri-siderophore reductase. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 64:792–796, 2008.
- [205] U. Baminger, S.S. Subramaniam, V. Renganathan, and D. Haltrich. Purification and characterization of cellobiose dehydrogenase from the plant pathogen *Sclerotium (Athelia) rolfsii*. *Appl. Environ. Microbiol.*, 67:1766–1774, 2001.
- [206] A. Banas, M. Bafor, E. Wiberg, M. Lenman, U. Staahl, and S. Stymne. Biosynthesis of an acetylenic fatty acid in microsomal preparations from developing seeds *Crepis alpina*. *Physiol. Biochem. Mol. Biol. Plant. [Proc. Int. Symp. Plant Lipids]*, 12th:57–59, 1997.
- [207] L.J. Banaszak and R.A. Bradshaw. Malate dehydrogenase. In P.D. Boyer, editor, *The Enzymes*, volume 11, pages 369–396. Academic Press, New York, 3rd edition, 1975.
- [208] D. Banauch, W. Brummer, W. Ebeling, H. Metz, H. Rindfrey, H. Lang, K. Leybold, and W. Rick. A glucose dehydrogenase for the determination of glucose concentrations in body fluids. *Z. Klin. Chem. Klin. Biochem.*, 13:101–107, 1975.
- [209] L. Banci, S. Camarero, A.T. Martínez, M.J. Martínez, M. Pérez-Boada, R. Pierattelli, and F.J. Ruiz-Dueñas. NMR study of manganese(II) binding by a new versatile peroxidase from the white-rot fungus *Pleurotus eryngii*. *J. Biol. Inorg. Chem.*, 8:751–760, 2003.
- [210] L. Bankel, E. Holme, G. Lindstedt, and S. Lindstedt. Oxygenases involved in thymine and thymidine metabolism in *Neurospora crassa*. *FEBS Lett.*, 21:135–138, 1972.
- [211] L. Bankel, G. Lindstedt, and S. Lindstedt. Thymidine 2'-hydroxylation in *Neurospora crassa*. *J. Biol. Chem.*, 247:6128–6134, 1972.
- [212] J. Banks and D.E. Cane. Biosynthesis of vitamin B₆: direct identification of the product of the PdxA-catalyzed oxidation of 4-hydroxy-L-threonine-4-phosphate using electrospray ionization mass spectrometry. *Bioorg. Med. Chem. Lett.*, 14:1633–1636, 2004.
- [213] G. Bannenberg, M. Martinez, M. Hamberg, and C. Castresana. Diversity of the enzymatic activity in the lipoxygenase gene family of *Arabidopsis thaliana*. *Lipids*, 44:85–95, 2009.
- [214] T. Baranowski. α -Glycerophosphate dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 85–96. Academic Press, New York, 2nd edition, 1963.

- [215] G.A. Barber. The synthesis of guanosine 5'-diphosphate D-rhamnose by enzymes of a higher plant. *Biochim. Biophys. Acta*, 165:68–75, 1968.
- [216] R.D. Barber, M.A. Rott, and T.J. Donohue. Characterization of a glutathione-dependent formaldehyde dehydrogenase from *Rhodobacter sphaeroides*. *J. Bacteriol.*, 178:1386–1393, 1996.
- [217] T. Barbier, F. Collard, A. Zuniga-Ripa, I. Moriyon, T. Godard, J. Becker, C. Wittmann, E. Van Schaftingen, and J.J. Letesson. Erythritol feeds the pentose phosphate pathway via three new isomerases leading to D-erythrose-4-phosphate in *Brucella*. *Proc. Natl. Acad. Sci. USA*, 111:17815–17820, 2014.
- [218] M.G. Barbour and R.C. Bayly. Control of meta-cleavage degradation of 4-hydroxyphenylacetate in *Pseudomonas putida*. *J. Bacteriol.*, 147:844–850, 1981.
- [219] F. Bardischewsky, A. Quentmeier, D. Rother, P. Hellwig, S. Kostka, and C.G. Friedrich. Sulfur dehydrogenase of *Paracoccus pantotrophus*: the heme-2 domain of the molybdoprotein cytochrome *c* complex is dispensable for catalytic activity. *Biochemistry*, 44:7024–7034, 2005.
- [220] J.C. Bardwell, K. McGovern, and J. Beckwith. Identification of a protein required for disulfide bond formation *in vivo*. *Cell*, 67:581–589, 1991.
- [221] E.A. Barnsley. Phthalate pathway of phenanthrene metabolism: formation of 2'-carboxybenzalpyruvate. *J. Bacteriol.*, 154:113–117, 1983.
- [222] S.F. Baron and J.G. Ferry. Purification and properties of the membrane-associated coenzyme F₄₂₀-reducing hydrogenase from *Methanobacterium formicicum*. *J. Bacteriol.*, 171:3846–3853, 1989.
- [223] S.F. Baron and P.B. Hylemon. Expression of the bile acid-inducible NADH:flavin oxidoreductase gene of *Eubacterium* sp. VPI 12708 in *Escherichia coli*. *Biochim. Biophys. Acta*, 1249:145–154, 1995.
- [224] I. Barr, J.A. Latham, A.T. Iavarone, T. Chantarojsiri, J.D. Hwang, and J.P. Klinman. Demonstration that the radical S-adenosylmethionine (SAM) enzyme PqqE catalyzes *de novo* carbon-carbon cross-linking within a peptide substrate PqqA in the presence of the peptide chaperone PqqD. *J. Biol. Chem.*, 291:8877–8884, 2016.
- [225] J. Barriuso, D.T. Nguyen, J.W. Li, J.N. Roberts, G. MacNevin, J.L. Chaytor, S.L. Marcus, J.C. Vederas, and D.K. Ro. Double oxidation of the cyclic nonaketide dihydromonacolin L to monacolin J by a single cytochrome P450 monooxygenase, LovA. *J. Am. Chem. Soc.*, 133:8078–8081, 2011.
- [226] M.H. Barros, C.G. Carlson, D.M. Glerum, and A. Tzagoloff. Involvement of mitochondrial ferredoxin and Cox15p in hydroxylation of heme O. *FEBS Lett.*, 492:133–138, 2001.
- [227] C.E. Barry, Nayar 3rd, Begley P.G., and T.P. Phenoxazinone synthase: mechanism for the formation of the phenoxazinone chromophore of actinomycin. *Biochemistry*, 28:6323–6333, 1989.
- [228] S.M. Barry and G.L. Challis. Mechanism and catalytic diversity of Rieske non-heme iron-dependent oxygenases. *ACS Catal.*, 3:2362–2370, 2013.
- [229] K. Bartsch, A. von Johnn-Marteville, and A. Schulz. Molecular analysis of two genes of the *Escherichia coli* gab cluster: nucleotide sequence of the glutamate:succinic semialdehyde transaminase gene (*gabT*) and characterization of the succinic semialdehyde dehydrogenase gene (*gabD*). *J. Bacteriol.*, 172:7035–7042, 1990.
- [230] G. Bashiri, J. Antoney, E.N.M. Jirgis, M.V. Shah, B. Ney, J. Copp, S.M. Stuteley, S. Sreebhavan, B. Palmer, M. Middleditch, N. Tokuriki, C. Greening, C. Scott, E.N. Baker, and C.J. Jackson. A revised biosynthetic pathway for the cofactor F₄₂₀ in prokaryotes. *Nat. Commun.*, 10:1558–1558, 2019.
- [231] G. Bashiri, C.J. Squire, E.N. Baker, and N.J. Moreland. Expression, purification and crystallization of native and selenomethionine labeled *Mycobacterium tuberculosis* FGD1 (Rv0407) using a *Mycobacterium smegmatis* expression system. *Protein Expr. Purif.*, 54:38–44, 2007.
- [232] J. Basran, N. Bhanji, A. Basran, D. Nietlispach, S. Mistry, R. Meskys, and N.S. Scrutton. Mechanistic aspects of the covalent flavoprotein dimethylglycine oxidase of *Arthrobacter globiformis* studied by stopped-flow spectrophotometry. *Biochemistry*, 41:4733–4743, 2002.

- [233] J. Basran, S. Fullerton, D. Leys, and N.S. Scrutton. Mechanism of FAD reduction and role of active site residues His-225 and Tyr-259 in *Arthrobacter globiformis* dimethylglycine oxidase: analysis of mutant structure and catalytic function. *Biochemistry*, 45:11151–11161, 2006.
- [234] P. Basu, B. Katterle, K.K. Andersson, and H. Dalton. The membrane-associated form of methane mono-oxygenase from *Methylococcus capsulatus* (Bath) is a copper/iron protein. *Biochem. J.*, 369:417–427, 2003.
- [235] N. Bate and E. Cundliffe. The mycinose-biosynthetic genes of *Streptomyces fradiae*, producer of tylosin. *J Ind Microbiol Biotechnol*, 23:118–122, 1999.
- [236] D.L. Bates, M.J. Danson, G. Hale, E.A. Hooper, and R.N. Perham. Self-assembly and catalytic activity of the pyruvate dehydrogenase multienzyme complex of *Escherichia coli*. *Nature*, 268:313–316, 1977.
- [237] C.J. Batie, E. LaHaie, and D.P. Ballou. Purification and characterization of phthalate oxygenase and phthalate oxygenase reductase from *Pseudomonas cepacia*. *J. Biol. Chem.*, 262:1510–1518, 1987.
- [238] M.G. Battelli and E. Lorenzoni. Purification and properties of a new glutathione-dependent thiol:disulphide oxidoreductase from rat liver. *Biochem. J.*, 207:133–138, 1982.
- [239] A.M. Battle, A. Benson, and C. Rimington. Purification and properties of coproporphyrinogenase. *Biochem. J.*, 97:731–740, 1965.
- [240] R. Bauder, B. Tshisuaka, and F. Lingens. Microbial metabolism of quinoline and related compounds. VII. Quinoline oxidoreductase from *Pseudomonas putida*: a molybdenum-containing enzyme. *Biol. Chem. Hoppe-Seyler*, 371:1137–1144, 1990.
- [241] G. Bauer and F. Lingens. Microbial metabolism of quinoline and related compounds. XV. Quinoline-4-carboxylic acid oxidoreductase from *Agrobacterium* spec.1B: a molybdenum-containing enzyme. *Biol. Chem. Hoppe-Seyler*, 373:699–705, 1992.
- [242] I. Bauer, A. De Beyer, B. Tshisuaka, S. Fetzner, and F. Lingens. A novel type of oxygenolytic ring cleavage: 2,4-Oxygenation and decarbonylation of 1*H*-3-hydroxy-4-oxoquinoline and 1*H*-3-hydroxy-4-oxoquinoline. *FEMS Microbiol. Lett.*, 117:299–304, 1994.
- [243] I. Bauer, N. Max, S. Fetzner, and F. Lingens. 2,4-Dioxygenases catalyzing *N*-heterocyclic-ring cleavage and formation of carbon monoxide. Purification and some properties of 1*H*-3-hydroxy-4-oxoquinoline 2,4-dioxygenase from *Arthrobacter* sp. Ru61a and comparison with 1*H*-3-hydroxy-4-oxoquinoline 2,4-dioxygenase from *Pseudomonas putida* 33/1. *Eur. J. Biochem.*, 240:576–583, 1996.
- [244] W. Bauer and M.H. Zenk. Formation of (*R*)-configured tetrahydroprotoberberine alkaloids in vivo and in vitro. *Tetrahedron Lett.*, 32:487–490, 1991.
- [245] W. Bauer and M.H. Zenk. Two methylenedioxy bridge-forming cytochrome *P*-450 dependent enzymes are involved in (*S*)-stylophine biosynthesis. *Phytochemistry*, 30:2953–2961, 1991.
- [246] S. Baumer, T. Ide, C. Jacobi, A. Johann, G. Gottschalk, and U. Deppenmeier. The F₄₂₀H₂ dehydrogenase from *Methanosarcina mazei* is a Redox-driven proton pump closely related to NADH dehydrogenases. *J. Biol. Chem.*, 275:17968–17973, 2000.
- [247] S. Baur, J. Marles-Wright, S. Buckenmaier, R.J. Lewis, and W. Vollmer. Synthesis of CDP-activated ribitol for teichoic acid precursors in *Streptococcus pneumoniae*. *J. Bacteriol.*, 191:1200–1210, 2009.
- [248] C. Bausch, N. Peekhaus, C. Utz, T. Blais, E. Murray, T. Lowary, and T. Conway. Sequence analysis of the GntII (subsidiary) system for gluconate metabolism reveals a novel pathway for L-idonic acid catabolism in *Escherichia coli*. *J. Bacteriol.*, 180:3704–3710, 1998.
- [249] Ü. Bayindir, A.W. Alfermann, and E. Fuss. Hinokinin biosynthesis in *Linum corymbulosum* Reichenb. *Plant J.*, 55:810–820, 2008.
- [250] S.A. Bayoumi, M.G. Rowan, I.S. Blagbrough, and J.R. Beeching. Biosynthesis of scopoletin and scopolin in cassava roots during post-harvest physiological deterioration: the E-Z-isomerisation stage. *Phytochemistry*, 69:2928–2936, 2008.

- [251] M.D. Bazzi. Interaction of camel lens ζ -crystallin with quinones: portrait of a substrate by fluorescence spectroscopy. *Arch. Biochem. Biophys.*, 395:185–190, 2001.
- [252] C.A. Beadle and A.R.W. Smith. The purification and properties of 2,4-dichlorophenol hydroxylase from a strain of *Acinetobacter species*. *Eur. J. Biochem.*, 123:323–332, 1982.
- [253] S.I. Beale. Biosynthesis of phycobilins. *Chem. Rev.*, 93:785–802, 1993.
- [254] R.C. Bean and W.Z. Hassid. Carbohydrate oxidase from a red alga *Iridophycus flaccidum*. *J. Biol. Chem.*, 218:425–436, 1956.
- [255] R.C. Bean, G.G. Porter, and B.M. Steinberg. Carbohydrate metabolism of citrus fruit. II. Oxidation of sugars by an aereodehydrogenase from young orange fruit. *J. Biol. Chem.*, 236:1235–1240, 1961.
- [256] F.C. Beasley, J. Cheung, and D.E. Heinrichs. Mutation of L-2,3-diaminopropionic acid synthase genes blocks staphyloferrin B synthesis in *Staphylococcus aureus*. *BMC Microbiol.*, 11:199–199, 2011.
- [257] F. Beaudoin, K. Gable, O. Sayanova, T. Dunn, and J.A. Napier. A *Saccharomyces cerevisiae* gene required for heterologous fatty acid elongase activity encodes a microsomal β -keto-reductase. *J. Biol. Chem.*, 277:11481–11488, 2002.
- [258] F. Beaudoin, X. Wu, F. Li, R.P. Haslam, J.E. Markham, H. Zheng, J.A. Napier, and L. Kunst. Functional characterization of the *Arabidopsis* β -ketoacyl-coenzyme A reductase candidates of the fatty acid elongase. *Plant Physiol.*, 150:1174–1191, 2009.
- [259] G.A. Beaudoin and P.J. Facchini. Isolation and characterization of a cDNA encoding (*S*)-*cis*-*N*-methylstylopine 14-hydroxylase from opium poppy, a key enzyme in sanguinarine biosynthesis. *Biochem. Biophys. Res. Commun.*, 431:597–603, 2013.
- [260] J.M. Le Beault, B. Roche, Z. Duvnjak, and E. Azoulay. Alcool- et aldéhyde-déshydrogénases particulières de *Candida tropicalis* cultivé sur hydrocarbures. *Biochim. Biophys. Acta*, 220:373–385, 1970.
- [261] B.A. Beaupre, M.R. Hoag, J. Roman, F.H. Forsterling, and G.R. Moran. Metabolic function for human renalase: oxidation of isomeric forms of β -NAD(P)H that are inhibitory to primary metabolism. *Biochemistry*, 54:795–806, 2015.
- [262] D. Becker, T. Schrader, and J.R. Andreesen. Two-component flavin-dependent pyrrole-2-carboxylate monooxygenase from *Rhodococcus* sp. *Eur. J. Biochem.*, 249:739–747, 1997.
- [263] J. Becker-Ketterm, N. Paczia, J.F. Conrotte, D.P. Kay, C. Guignard, P.P. Jung, and C.L. Linster. *Saccharomyces cerevisiae* forms D-2-hydroxyglutarate and couples its degradation to D-lactate formation via a cytosolic transhydrogenase. *J. Biol. Chem.*, 291:6036–6058, 2016.
- [264] C. Beckmann, J. Rattke, N.J. Oldham, P. Sperling, E. Heinz, and W. Boland. Characterization of a Δ^8 -sphingolipid desaturase from higher plants: a stereochemical and mechanistic study on the origin of *E,Z* isomers. *Angew. Chem. Int. Ed. Engl.*, 41:2298–2300, 2002.
- [265] J.D. Beckmann and F.E. Frerman. Electron-transfer flavoprotein-ubiquinone oxidoreductase from pig liver: purification and molecular, redox, and catalytic properties. *Biochemistry*, 24:3913–3921, 1985.
- [266] B. Bednarski, J.R. Andreesen, and A. Pich. In vitro processing of the proproteins GrdE of protein B of glycine reductase and PrdA of D-proline reductase from *Clostridium sticklandii*: formation of a pyruvoyl group from a cysteine residue. *Eur. J. Biochem.*, 268:3538–3544, 2001.
- [267] R. De Beer, J.A. Duine, J. Frank, Large Jr., and P.J. The prosthetic group of methylamine dehydrogenase from *Pseudomonas* AM1: evidence for a quinone structure. *Biochim. Biophys. Acta*, 622:370–374, 1980.
- [268] W.T. Beeson, C.M. Phillips, J.H. Cate, and M.A. Marletta. Oxidative cleavage of cellulose by fungal copper-dependent polysaccharide monooxygenases. *J. Am. Chem. Soc.*, 134:890–892, 2012.
- [269] H. Beevers and R.C. French. Oxidation of *N*-acetylindoxyl by an enzyme from plants. *Arch. Biochem. Biophys.*, 50:427–439, 1954.
- [270] V. Behal, Z. Hostalek, and Z. Vanek. Anhydrotetracycline oxygenase activity and biosynthesis of tetracyclines in *Streptomyces aureofaciens*. *Biotechnol. Lett.*, 1:177–182, 1979.

- [271] E.J. Behrman and R.Y. Stanier. The bacterial oxidation of nicotinic acid. *J. Biol. Chem.*, 228:923–945, 1957.
- [272] U. Beifuss, M. Tietze, S. Baumer, and U. Deppenmeier. Methanophenazine: structure, total synthesis, and function of a new cofactor from methanogenic *Archaea*. *Angew. Chem. Int. Ed. Engl.*, 39:2470–2472, 2000.
- [273] S. Beil, B. Happe, K.N. Timmis, and D.H. Pieper. Genetic and biochemical characterization of the broad spectrum chlorobenzene dioxygenase from *Burkholderia* sp. strain PS12 - dechlorination of 1,2,4,5-tetrachlorobenzene. *Eur. J. Biochem.*, 247:190–199, 1997.
- [274] S. Beil, J.R. Mason, K.N. Timmis, and D.H. Pieper. Identification of chlorobenzene dioxygenase sequence elements involved in dechlorination of 1,2,4,5-tetrachlorobenzene. *J. Bacteriol.*, 180:5520–5528, 1998.
- [275] H. Beinert. Acyl coenzyme A dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 447–466. Academic Press, New York, 2nd edition, 1963.
- [276] A. Belan, J. Botle, A. Fauve, J.G. Gourcy, and H. Veschambre. Use of biological systems for the preparation of chiral molecules. 3. An application in pheromone synthesis: Preparation of sulcatol enantiomers. *J. Org. Chem.*, 52:256–260, 1987.
- [277] M. Belanger, L.L. Burrows, and J.S. Lam. Functional analysis of genes responsible for the synthesis of the B-band O antigen of *Pseudomonas aeruginosa* serotype O6 lipopolysaccharide. *Microbiology (Reading)*, 145:3505–3521, 1999.
- [278] P. Belin, M.H. Le Du, A. Fielding, O. Lequin, M. Jacquet, J.B. Charbonnier, A. Lecoq, R. Thai, M. Courcon, C. Masson, C. Dugave, R. Genet, J.L. Pernodet, and M. Gondry. Identification and structural basis of the reaction catalyzed by CYP121, an essential cytochrome P450 in *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. USA*, 106:7426–7431, 2009.
- [279] O.V. Belyaeva, O.V. Korkina, A.V. Stetsenko, and N.Y. Kedishvili. Human retinol dehydrogenase 13 (RDH13) is a mitochondrial short-chain dehydrogenase/reductase with a retinaldehyde reductase activity. *FEBS J.*, 275:138–147, 2008.
- [280] O.V. Belyaeva, O.V. Korkina, A.V. Stetsenko, T. Kim, P.S. Nelson, and N.Y. Kedishvili. Biochemical properties of purified human retinol dehydrogenase 12 (RDH12): catalytic efficiency toward retinoids and C₉ aldehydes and effects of cellular retinol-binding protein type I (CRBPI) and cellular retinaldehyde-binding protein (CRALBP) on the oxidation and reduction of retinoids. *Biochemistry*, 44:7035–7047, 2005.
- [281] A. Ben-Amotz and M. Avron. NADP specific dihydroxyacetone reductase from *Dunaliella parva*. *FEBS Lett.*, 29:153–155, 1973.
- [282] A. Ben-Bassat and I. Goldberg. Purification and properties of glucose-6-phosphate dehydrogenase (NADP⁺/NAD⁺) and 6-phosphogluconate dehydrogenase (NADP⁺/NAD⁺) from methanol-grown *Pseudomonas C*. *Biochim. Biophys. Acta*, 611:1–10, 1980.
- [283] J. Benach, I. Lee, W. Edstrom, A.P. Kuzin, Y. Chiang, T.B. Acton, G.T. Montelione, and J.F. Hunt. The 2.3-Å crystal structure of the shikimate 5-dehydrogenase orthologue YdiB from *Escherichia coli* suggests a novel catalytic environment for an NAD-dependent dehydrogenase. *J. Biol. Chem.*, 278:19176–19182, 2003.
- [284] R. Benavente, M. Esteban-Torres, G.W. Kohring, A. Cortes-Cabrera, P.A. Sanchez-Murcia, F. Gago, I. Acebron, B. de las Rivas, R. Munoz, and J.M. Mancheno. Enantioselective oxidation of galactitol 1-phosphate by galactitol-1-phosphate 5-dehydrogenase from *Escherichia coli*. *Acta Crystallogr. D Biol. Crystallogr.*, 71:1540–1554, 2015.
- [285] D.S. Bendall and W.D. Bonner. Cyanide-insensitive respiration in plant mitochondria. *Plant Physiol.*, 47:236–245, 1971.
- [286] A. Benjdia, J. Leprince, C. Sandstrom, H. Vaudry, and O. Berteau. Mechanistic investigations of anaerobic sulfatase-maturating enzyme: direct Cβ H-atom abstraction catalyzed by a radical AdoMet enzyme. *J. Am. Chem. Soc.*, 131:8348–8349, 2009.
- [287] A. Benjdia, S. Subramanian, J. Leprince, H. Vaudry, M.K. Johnson, and O. Berteau. Anaerobic sulfatase-maturating enzymes, first dual substrate radical S-adenosylmethionine enzymes. *J. Biol. Chem.*, 283:17815–17826, 2008.
- [288] A. Benjdia, S. Subramanian, J. Leprince, H. Vaudry, M.K. Johnson, and O. Berteau. Anaerobic sulfatase-maturating enzyme⁻-a mechanistic link with glycyl radical-activating enzymes. *FEBS J.*, 277:1906–1920, 2010.

- [289] R.N. Bennett, G. Kiddle, and R.M. Wallsgrove. Involvement of cytochrome P450 in glucosinolate biosynthesis in white mustard (a biochemical anomaly). *Plant Physiol.*, 114:1283–1291, 1997.
- [290] A.F. Bent, G. Mann, W.E. Houssen, V. Mykhaylyk, R. Duman, L. Thomas, M. Jaspars, A. Wagner, and J.H. Naismith. Structure of the cyanobactin oxidase ThcOx from *Cyanothece* sp. PCC 7425, the first structure to be solved at Diamond Light Source beamline I23 by means of *S*-SAD. *Acta Crystallogr D Struct Biol*, 72:1174–1180, 2016.
- [291] R. Bentley. Glucose oxidase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 567–586. Academic Press, New York, 2nd edition, 1963.
- [292] I. Benveniste, N. Tijet, F. Adas, G. Philipps, J.P. Salaun, and F. Durst. CYP86A1 from *Arabidopsis thaliana* encodes a cytochrome P450-dependent fatty acid ω -hydroxylase. *Biochem. Biophys. Res. Commun.*, 243:688–693, 1998.
- [293] U. Berendt, T. Haverkamp, A. Prior, , and J.D. Reaction mechanism of thioredoxin: 3'-phospho-adenylylsulfate reductase investigated by site-directed mutagenesis. *Eur. J. Biochem.*, 233:347–356, 1995.
- [294] A. Berg, J.A. Gustafsson, and M. Ingelman-Sundberg. Characterization of a cytochrome *P*-450-dependent steroid hydroxylase system present in *Bacillus megaterium*. *J. Biol. Chem.*, 251:2831–2838, 1976.
- [295] A. Berg, M. Ingelman-Sundberg, and J.A. Gustafsson. Purification and characterization of cytochrome *P*-450meg. *J. Biol. Chem.*, 254:5264–5271, 1979.
- [296] I.A. Berg, D. Kockelkorn, W. Buckel, and G. Fuchs. A 3-hydroxypropionate/4-hydroxybutyrate autotrophic carbon dioxide assimilation pathway in *Archaea*. *Science*, 318:1782–1786, 2007.
- [297] R.A. Berg and D.J. Prockop. Affinity column purification of procollagen proline hydroxylase from chick embryos and further characterization of the enzyme. *J. Biol. Chem.*, 248:1175–1182, 1973.
- [298] D.J. Bergmann and A.B. Hooper. Sequence of the gene, *amoB*, for the 43-kDa polypeptide of ammonia monooxygenase of *Nitrosomonas europaea*. *Biochem. Biophys. Res. Commun.*, 204:759–762, 1994.
- [299] H.-U. Bergmeyer, K. Gawehn, H. Klotzsch, H.A. Krebs, and D.H. Williamson. Purification and properties of crystalline 3-hydroxybutyrate dehydrogenase from *Rhodospseudomonas spheroides*. *Biochem. J.*, 102:423–431, 1967.
- [300] J. Bergsma, R. Strijker, J.Y. Alkema, H.G. Seijen, and W.N. Konings. NADH dehydrogenase and NADH oxidation in membrane vesicle from *Bacillus subtilis*. *Eur. J. Biochem.*, 120:599–606, 1981.
- [301] B.C. Berks, S.J. Ferguson, J.W. Moir, and D.J. Richardson. Enzymes and associated electron transport systems that catalyse the respiratory reduction of nitrogen oxides and oxyanions. *Biochim. Biophys. Acta*, 1232:97–173, 1995.
- [302] T. Berman and B. Magasanik. The pathway of *myo*-inositol degradation in *Aerobacter aerogenes*. Dehydrogenation and dehydration. *J. Biol. Chem.*, 241:800–806, 1966.
- [303] A. Bernardo, J. Burgos, and R. Martin. Purification and some properties of L-glycol dehydrogenase from hen's muscle. *Biochim. Biophys. Acta*, 659:189–198, 1981.
- [304] M. Bernhard, B. Benelli, A. Hochkoeppler, D. Zannoni, and B. Friedrich. Functional and structural role of the cytochrome *b* subunit of the membrane-bound hydrogenase complex of *Alcaligenes eutrophus* H16. *Eur. J. Biochem.*, 248:179–186, 1997.
- [305] F.-H. Bernhardt, W. Nastainczyk, and V. Seydewitz. Kinetic studies on a 4-methoxybenzoate *O*-demethylase from *Pseudomonas putida*. *Eur. J. Biochem.*, 72:107–115, 1977.
- [306] M.L.C. Bernheim. The hydroxylamine reductase of mitochondria. *Arch. Biochem. Biophys.*, 134:408–413, 1969.
- [307] M.L.C. Bernheim and P. Hochstein. Reduction of hydroxylamine by rat liver mitochondria. *Arch. Biochem. Biophys.*, 124:436–442, 1968.
- [308] B.L. Bertagnolli and L.P. Hager. Role of flavin in acetoin production by two bacterial pyruvate oxidases. *Arch. Biochem. Biophys.*, 300:364–371, 1993.
- [309] C.M. Berteza, J.R. Freije, H. van der Woude, F.W. Verstappen, L. Perk, V. Marquez, J.W. De Kraker, M.A. Posthumus, B.J. Jansen, A. de Groot, M.C. Franssen, and H.J. Bouwmeester. Identification of intermediates and enzymes involved in the early steps of artemisinin biosynthesis in *Artemisia annua*. *Planta Med.*, 71:40–47, 2005.

- [310] C.M. Berteau, M. Schalk, F. Karp, M. Maffei, and R. Croteau. Demonstration that menthofuran synthase of mint (*Mentha*) is a cytochrome P_{450} monooxygenase: cloning, functional expression, and characterization of the responsible gene. *Arch. Biochem. Biophys.*, 390:279–286, 2001.
- [311] O. Berteau, A. Guillot, A. Benjdia, and S. Rabot. A new type of bacterial sulfatase reveals a novel maturation pathway in prokaryotes. *J. Biol. Chem.*, 281:22464–22470, 2006.
- [312] M.G. Bertero, R.A. Rothery, N. Boroumand, M. Palak, F. Blasco, N. Ginet, J.H. Weiner, and N.C. Strynadka. Structural and biochemical characterization of a quinol binding site of *Escherichia coli* nitrate reductase A. *J. Biol. Chem.*, 280:14836–14843, 2005.
- [313] M.G. Bertero, R.A. Rothery, M. Palak, C. Hou, D. Lim, F. Blasco, J.H. Weiner, and N.C. Strynadka. Insights into the respiratory electron transfer pathway from the structure of nitrate reductase A. *Nat. Struct. Biol.*, 10:681–687, 2003.
- [314] D.A. Berthold, M.E. Andersson, and P. Nordlund. New insight into the structure and function of the alternative oxidase. *Biochim. Biophys. Acta*, 1460:241–254, 2000.
- [315] P.A. Bertram, M. Karrasch, R.A. Schmitz, R. Bocher, S.P. Albracht, and R.K. Thauer. Formylmethanofuran dehydrogenases from methanogenic Archaea. Substrate specificity, EPR properties and reversible inactivation by cyanide of the molybdenum or tungsten iron-sulfur proteins. *Eur. J. Biochem.*, 220:477–484, 1994.
- [316] P.A. Bertram, R.A. Schmitz, D. Linder, and R.K. Thauer. Tungstate can substitute for molybdate in sustaining growth of *Methanobacterium thermoautotrophicum*. Identification and characterization of a tungsten isoenzyme of formylmethanofuran dehydrogenase. *Arch. Microbiol.*, 161:220–228, 1994.
- [317] T. Bertrand, N.A. Eady, J.N. Jones, Nagy Jesmin, Jamart-Gregoire J.M., Raven B., Brown E.L., and K.A. Crystal structure of *Mycobacterium tuberculosis* catalase-peroxidase. *J. Biol. Chem.*, 279:38991–38999, 2004.
- [318] J. Bertsch, C. Oppinger, V. Hess, J.D. Langer, and V. Muller. Heterotrimeric NADH-oxidizing methylenetetrahydrofolate reductase from the acetogenic bacterium *Acetobacterium woodii*. *J. Bacteriol.*, 197:1681–1689, 2015.
- [319] J. Bertsch, A. Parthasarathy, W. Buckel, and V. Muller. An electron-bifurcating caffeoyl-CoA reductase. *J. Biol. Chem.*, 288:11304–11311, 2013.
- [320] Y.V. Bertsova, M.V. Serebryakova, A.A. Baykov, and A.V. Bogachev. A novel, NADH-dependent acrylate reductase in *Vibrio harveyi*. *Appl. Environ. Microbiol.*, 88:e0051922–e0051922, 2022.
- [321] A. Besrat, C.E. Polan, and L.M. Henderson. Mammalian metabolism of glutaric acid. *J. Biol. Chem.*, 244:1461–1467, 1969.
- [322] S. Besseau, F. Kellner, A. Lanoue, A.M. Thamm, V. Salim, B. Schneider, F. Geu-Flores, R. Hofer, G. Guirimand, A. Guihur, A. Oudin, G. Glevarec, E. Foureau, N. Papon, M. Clastre, N. Giglioli-Guivarc’h, B. St-Pierre, D. Werck-Reichhart, V. Burlat, V. De Luca, S.E. O’Connor, and V. Courdavault. A pair of tabersonine 16-hydroxylases initiates the synthesis of vindoline in an organ-dependent manner in *Catharanthus roseus*. *Plant Physiol.*, 163:1792–1803, 2013.
- [323] M. Bey, S. Zhou, L. Poidevin, B. Henrissat, P.M. Coutinho, J.G. Berrin, and J.C. Sigoillot. Cello-oligosaccharide oxidation reveals differences between two lytic polysaccharide monooxygenases (family GH61) from *Podospira anserina*. *Appl. Environ. Microbiol.*, 79:488–496, 2013.
- [324] J. Beynon, E.R. Rafanan, Shen Jr., Fisher B., and A.J. Crystallization and preliminary X-ray analysis of tetracenomycin A2 oxygenase: a flavoprotein hydroxylase involved in polyketide biosynthesis. *Acta Crystallogr. D Biol. Crystallogr.*, 56:1647–1651, 2000.
- [325] S.G. Bhat and C.S. Vaidyanathan. Purification and properties of L-4-hydroxymandelate oxidase from *Pseudomonas convexa*. *Eur. J. Biochem.*, 68:323–331, 1976.
- [326] S.G. Bhat and C.S. Vaidyanathan. Purifications and properties of L-mandelate-4-hydroxylase from *Pseudomonas convexa*. *Arch. Biochem. Biophys.*, 176:314–323, 1976.
- [327] D. Bhatnagar, T.E. Cleveland, and D.G. Kingston. Enzymological evidence for separate pathways for aflatoxin B₁ and B₂ biosynthesis. *Biochemistry*, 30:4343–4350, 1991.

- [328] M.R. Bhatt, Y. Khatri, R.J. Rodgers, and L.L. Martin. Role of cytochrome b_5 in the modulation of the enzymatic activities of cytochrome P450 17 α -hydroxylase/17,20-lyase (P450 17A1). *J. Steroid Biochem. Mol. Biol.*, 2016.
- [329] S. Bhowmik, D.H. Jones, H.P. Chiu, I.H. Park, H.J. Chiu, H.L. Axelrod, C.L. Farr, H.J. Tien, S. Agarwalla, and S.A. Lesley. Structural and functional characterization of BaiA, an enzyme involved in secondary bile acid synthesis in human gut microbe. *Proteins*, 82:216–229, 2014.
- [330] M. Biarnes-Carrera, C.K. Lee, T. Nihira, R. Breitling, and E. Takano. Orthogonal regulatory circuits for *Escherichia coli* based on the γ -butyrolactone system of *Streptomyces coelicolor*. *ACS Synth. Biol.*, 7:1043–1055, 2018.
- [331] J.A. Bick, F. Aslund, Y. Cen, and T. Leustek. Glutaredoxin function for the carboxyl-terminal domain of the plant-type 5'-adenylylsulfate reductase. *Proc. Natl. Acad. Sci. USA*, 95:8404–8409, 1998.
- [332] J.A. Bick, J.J. Dennis, G.J. Zylstra, J. Nowack, and T. Leustek. Identification of a new class of 5-adenylylsulfate (APS) reductase from sulfate-assimilating bacteria. *J. Bacteriol.*, 182:135–142, 2000.
- [333] J.T. Billheimer, M. Alcorn, and J.L. Gaylor. Solubilization and partial purification of a microsomal 3-ketosteroid reductase of cholesterol biosynthesis. Purification and properties of 3 β -hydroxysteroid dehydrogenase and Δ^5 -3-ketosteroid isomerase from bovine corpora lutea. *Arch. Biochem. Biophys.*, 211:430–438, 1981.
- [334] R.E. Billings, H.R. Sullivan, and R.E. McMahon. The dehydrogenation of 1-indanol by a soluble oxidoreductase from bovine liver. *J. Biol. Chem.*, 246:3512–3517, 1971.
- [335] C. Binnie, M. Warren, and M.J. Butler. Cloning and heterologous expression in *Streptomyces lividans* of *Streptomyces rimosus* genes involved in oxytetracycline biosynthesis. *J. Bacteriol.*, 171:887–895, 1989.
- [336] J. Birke and D. Jendrossek. Rubber oxygenase and latex clearing protein cleave rubber to different products and use different cleavage mechanisms. *Appl. Environ. Microbiol.*, 80:5012–5020, 2014.
- [337] J. Birke, W. Röther, and D. Jendrossek. RoxB is a novel type of rubber oxygenase that combines properties of rubber oxygenase RoxA and latex clearing protein (Lcp). *Appl. Environ. Microbiol.*, 83:e00721–17–, 2017.
- [338] B. Biteau, J. Labarre, and M.B. Toledano. ATP-dependent reduction of cysteine-sulphinic acid by *S. cerevisiae* sulphiredoxin. *Nature*, 425:980–984, 2003.
- [339] E. Bitto, Y. Huang, C.A. Bingman, S. Singh, J.S. Thorson, and G.N. Phillips Jr. The structure of flavin-dependent tryptophan 7-halogenase RebH. *Proteins Struct. Funct. Genet.*, 70:289–293, 2008.
- [340] I. Björkhem and H. Danielsson. Stereochemistry of hydrogen transfer from pyridine nucleotides catalyzed by Δ^4 -3-oxosteroid 5- β -reductase and 3- α -hydroxysteroid dehydrogenase from rat liver. *Eur. J. Biochem.*, 12:80–84, 1970.
- [341] O. Björnberg, A.C. Grüner, P. Roepstorff, and K.F. Jensen. The activity of *Escherichia coli* dihydroorotate dehydrogenase is dependent on a conserved loop identified by sequence homology, mutagenesis, and limited proteolysis. *Biochemistry*, 38:2899–2908, 1999.
- [342] O. Björnberg, P. Rowland, S. Larsen, and K.F. Jensen. Active site of dihydroorotate dehydrogenase A from *Lactococcus lactis* investigated by chemical modification and mutagenesis. *Biochemistry*, 36:16197–16205, 1997.
- [343] S. Black. Yeast aldehyde dehydrogenase. *Arch. Biochem. Biophys.*, 34:86–97, 1951.
- [344] S. Black, E.M. Harte, B. Hudson, and L. Wartofsky. A specific enzymatic reduction of L(-)-methionine sulfoxide and a related nonspecific reduction of diulfides. *J. Biol. Chem.*, 235:2910–2916, 1960.
- [345] S. Black and N.G. Wright. Aspartic β -semialdehyde dehydrogenase and aspartic β -semialdehyde. *J. Biol. Chem.*, 213:39–50, 1955.
- [346] S. Black and N.G. Wright. Homoserine dehydrogenase. *J. Biol. Chem.*, 213:51–60, 1955.
- [347] C.C. Blake, M. Ghosh, K. Harlos, A. Avezoux, and C. Anthony. The active site of methanol dehydrogenase contains a disulphide bridge between adjacent cysteine residues. *Nat. Struct. Biol.*, 1:102–105, 1994.
- [348] R.C. Blake, Shute 2nd, and E.A. Respiratory enzymes of *Thiobacillus ferrooxidans*. Kinetic properties of an acid-stable iron:rusticyanin oxidoreductase. *Biochemistry*, 33:9220–9228, 1994.

- [349] E.R. Blakley. The catabolism of L-tyrosine by an *Arthrobacter sp.* *Can. J. Microbiol.*, 23:1128–1139, 1977.
- [350] R.L. Blakley. Cobamides and ribonucleotide reduction. I. Cobamide stimulation of ribonucleotide reduction in extracts of *Lactobacillus leichmannii*. *J. Biol. Chem.*, 240:2173–2180, 1965.
- [351] R.L. Blakley and B.M. MacDougall. Dihydrofolic reductase from *Streptococcus faecalis* R. *J. Biol. Chem.*, 236:1163–1163, 1961.
- [352] V. Blanc, P. Gil, N. Bamas-Jacques, S. Lorenzon, M. Zagorec, J. Schleuniger, D. Bisch, F. Blanche, L. Debussche, J. Crouzet, and D. Thibaut. Identification and analysis of genes from *Streptomyces pristinaespiralis* encoding enzymes involved in the biosynthesis of the 4-dimethylamino-L-phenylalanine precursor of pristinamycin I. *Mol. Microbiol.*, 23:191–202, 1997.
- [353] M. Blanchard, D.E. Green, V. Nocito-Carroll, and S. Ratner. *l*-Hydroxy acid oxidase. *J. Biol. Chem.*, 163:137–144, 1946.
- [354] F. Blanche, D. Thibaut, A. Famechon, L. Debussche, B. Cameron, and J. Crouzet. Precorrin-6X reductase from *Pseudomonas denitrificans*: purification and characterization of the enzyme and identification of the structural gene. *J. Bacteriol.*, 174:1036–1042, 1992.
- [355] L. Blanco, P.M. Reddy, S. Silvente, B. Bucciarelli, S. Khandual, X. Alvarado-Affantranger, F. Sanchez, S. Miller, C. Vance, and M. Lara-Flores. Molecular cloning, characterization and regulation of two different NADH-glutamate synthase cDNAs in bean nodules. *Plant Cell Environ.*, 31:454–472, 2008.
- [356] H. Blaschko. Amine oxidase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 8, pages 337–351. Academic Press, New York, 2nd edition, 1963.
- [357] E. Blee, A.L. Wilcox, L.J. Marnett, and F. Schuber. Mechanism of reaction of fatty acid hydroperoxides with soybean peroxygenase. *J. Biol. Chem.*, 268:1708–1715, 1993.
- [358] H.S. Bleeg and F. Christensen. Biosynthesis of ascorbate in yeast. Purification of L-galactono-1,4-lactone oxidase with properties different from mammalian L-gulonolactone oxidase. *Eur. J. Biochem.*, 127:391–396, 1982.
- [359] K. Bleicher and J. Winter. Purification and properties of F₄₂₀- and NADP⁺-dependent alcohol dehydrogenases of *Methanogenium liminatans* and *Methanobacterium palustre*, specific for secondary alcohols. *Eur. J. Biochem.*, 200:43–51, 1991.
- [360] W. Bless and W. Barz. Isolation of pterocarpan synthase, the terminal enzyme of pterocarpan phytoalexin biosynthesis in cell-suspension cultures of *Cicer arietinum*. *FEBS Lett.*, 235:47–50, 1988.
- [361] D.W. Block and F. Lingens. Microbial metabolism of quinoline and related compounds. XIV. Purification and properties of 1H-3-hydroxy-4-oxoquinoline oxygenase, a new estradiol cleavage enzyme from *Pseudomonas putida* strain 33/1. *Biol. Chem. Hoppe Seyler*, 373:249–254, 1992.
- [362] J.A. Blodgett, P.M. Thomas, G. Li, J.E. Velasquez, W.A. van der Donk, N.L. Kelleher, and W.W. Metcalf. Unusual transformations in the biosynthesis of the antibiotic phosphinothricin tripeptide. *Nat. Chem. Biol.*, 3:480–485, 2007.
- [363] C. Blumer and D. Haas. Mechanism, regulation, and ecological role of bacterial cyanide biosynthesis. *Arch. Microbiol.*, 173:170–177, 2000.
- [364] R. Bode, A. Lippoldt, and D. Birnbaum. Purification and properties of D-aromatic lactate dehydrogenase an enzyme involved in the catabolism of the aromatic amino acids of *Candida maltosa*. *Biochem. Physiol. Pflanzen*, 181:189–198, 1986.
- [365] R. Boden, E. Borodina, A.P. Wood, D.P. Kelly, J.C. Murrell, and H. Schafer. Purification and characterization of dimethylsulfide monooxygenase from *Hyphomicrobium sulfonivorans*. *J. Bacteriol.*, 193:1250–1258, 2011.
- [366] W.E. Boeglin, A. Itoh, Y. Zheng, G. Coffa, G.A. Howe, and A.R. Brash. Investigation of substrate binding and product stereochemistry issues in two linoleate 9-lipoxygenases. *Lipids*, 43:979–987, 2008.
- [367] E. De Boer, M.G.M. Tromp, H. Plat, G.E. Krenn, and R. Wever. Vanadium(v) as an essential element for haloperoxidase activity in marine brown-algae - purification and characterization of a vanadium(V)-containing bromoperoxidase from *Laminaria saccharina*. *Biochim. Biophys. Acta*, 872:104–115, 1986.

- [368] H. Boer, H. Maaheimo, A. Koivula, M. Penttila, and P. Richard. Identification in *Agrobacterium tumefaciens* of the D-galacturonic acid dehydrogenase gene. *Appl. Microbiol. Biotechnol.*, 86:901–909, 2010.
- [369] A.V. Bogachev, Y.V. Bertsova, D.A. Bloch, and M.I. Verkhovsky. Urocanate reductase: identification of a novel anaerobic respiratory pathway in *Shewanella oneidensis* MR-1. *Mol. Microbiol.*, 86:1452–1463, 2012.
- [370] J.A. Bogart, A.J. Lewis, and E.J. Schelter. DFT study of the active site of the XoxF-type natural, cerium-dependent methanol dehydrogenase enzyme. *Chemistry Eur. J.*, 21:1743–1748, 2015.
- [371] N. Bogdanovic, L. Bretillon, E.G. Lund, U. Diczfalusy, L. Lannfelt, B. Winblad, D.W. Russell, and I. Björkhem. On the turnover of brain cholesterol in patients with Alzheimer’s disease. Abnormal induction of the cholesterol-catabolic enzyme CYP46 in glial cells. *Neurosci. Lett.*, 314:45–48, 2001.
- [372] R.A. Boghosian and E.T. McGuinness. Affinity purification and properties of porcine brain aldose reductase. *Biochim. Biophys. Acta*, 567:278–286, 1979.
- [373] C.C. Böhmé, L.D. Arscott, K. Becker, R.H., Williams Schirmer, , and Jr. Kinetic characterization of glutathione reductase from the malarial parasite *Plasmodium falciparum*. Comparison with the human enzyme. *J. Biol. Chem.*, 275:37317–37323, 2000.
- [374] J. Bohuslavek, J.W. Payne, Y. Liu, H. Bolton, Xun Jr., and L. Cloning, sequencing, and characterization of a gene cluster involved in EDTA degradation from the bacterium BNC1. *Appl. Environ. Microbiol.*, 67:688–695, 2001.
- [375] M.J. Boland and A.G. Benny. Enzymes of nitrogen metabolism in legume nodules. Purification and properties of NADH-dependent glutamate synthase from lupin nodules. *Eur. J. Biochem.*, 79:355–362, 1977.
- [376] P.L. Bolen and R.W. Detroy. Induction of NADPH-linked D-xylose reductase and NAD-linked xylitol dehydrogenase activities in *Pachysolen tannophilus* by D-xylose, L-arabinose, or D-galactose. *Biotechnol. Bioeng.*, 27:302–307, 1985.
- [377] J.T. Bolin, D.J. Filman, D.A. Matthews, and J. Kraut. Crystal structures of *Escherichia coli* and *Lactobacillus casei* dihydrofolate reductase refined at 1.7 Å resolution. I. General features and binding of methotrexate. *J. Biol. Chem.*, 257:13650–13662, 1982.
- [378] M. Boll and G. Fuchs. Benzoyl-coenzyme A reductase (dearomatizing), a key enzyme of anaerobic aromatic metabolism. ATP dependence of the reaction, purification and some properties of the enzyme from *Thauera aromatica* strain K172. *Eur. J. Biochem.*, 234:921–933, 1995.
- [379] D.W. Bollivar and S.I. Beale. The chlorophyll biosynthetic enzyme Mg-protoporphyrin IX monomethyl ester (oxidative) cyclase (characterization and partial purification from *Chlamydomonas reinhardtii* and *Synechocystis* sp. PCC 6803). *Plant Physiol.*, 112:105–114, 1996.
- [380] D.W. Bollivar, S. Wang, J.P. Allen, and C.E. Bauer. Molecular genetic analysis of terminal steps in bacteriochlorophyll *a* biosynthesis: characterization of a *Rhodobacter capsulatus* strain that synthesizes geranylgeraniol-esterified bacteriochlorophyll *a*. *Biochemistry*, 33:12763–12768, 1994.
- [381] D. Bonam and P.W. Ludden. Purification and characterization of carbon monoxide dehydrogenase, a nickel, zinc, iron-sulfur protein, from *Rhodospirillum rubrum*. *J. Biol. Chem.*, 262:2980–2987, 1987.
- [382] D.H. Bone, S. Bernstein, and W. Vishniac. Purification and some properties of different forms of hydrogen dehydrogenase. *Biochim. Biophys. Acta*, 67:581–588, 1963.
- [383] M.J. Bonete, J. Ferrer, C. Pire, M. Penades, and J.L. Ruiz. 2-Hydroxyacid dehydrogenase from *Haloferax mediterranei*, a D-isomer-specific member of the 2-hydroxyacid dehydrogenase family. *Biochimie*, 82:1143–1150, 2000.
- [384] V. Bonnefoy and J.A. Demoss. Nitrate reductases in *Escherichia coli*. *Antonie Van Leeuwenhoek*, 66:47–56, 1994.
- [385] C.A. Bonner, R.A. Jensen, J.E. Gander, and N.O. Keyhani. A core catalytic domain of the TyrA protein family: arogenate dehydrogenase from *Synechocystis*. *Biochem. J.*, 382:279–291, 2004.
- [386] S.A. Bonnett, K. Papireddy, S. Higgins, S. del Cardayre, and K.A. Reynolds. Functional characterization of an NADPH dependent 2-alkyl-3-ketoalkanoic acid reductase involved in olefin biosynthesis in *Stenotrophomonas maltophilia*. *Biochemistry*, 50:9633–9640, 2011.

- [387] J.A.M. De Bont, J.P. Van Dijken, and W. Harder. Dimethyl sulphoxide and dimethyl sulphide as a carbon, sulphur and energy source for growth of *Hyphomicrobium* S. *J. Gen. Microbiol.*, 127:315–323, 1981.
- [388] J. Booker, M. Auldridge, S. Wills, D. McCarty, H. Klee, and O. Leyser. MAX3/CCD7 is a carotenoid cleavage dioxygenase required for the synthesis of a novel plant signaling molecule. *Curr. Biol.*, 14:1232–1238, 2004.
- [389] S.J. Booker. Anaerobic functionalization of unactivated C-H bonds. *Curr. Opin. Chem. Biol.*, 13:58–73, 2009.
- [390] J. Booth and E. Boyland. The biochemistry of aromatic amines. 3. Enzymic hydroxylation by rat-liver microsomes. *Biochem. J.*, 66:73–78, 1957.
- [391] C.E. Borgeson, M. de Renobales, and G.J. Blomquist. Characterization of the Δ^{12} desaturase in the American cockroach, *Periplaneta americana*: the nature of the substrate. *Biochim. Biophys. Acta*, 1047:135–140, 1990.
- [392] A.S. Borisova, T. Isaksen, M. Dimarogona, A.A. Kognole, G. Mathiesen, A. Varnai, A.K. Rohr, C.M. Payne, M. Sorlie, M. Sandgren, and V.G. Eijsink. Structural and functional characterization of a lytic polysaccharide monooxygenase with broad substrate specificity. *J. Biol. Chem.*, 290:22955–22969, 2015.
- [393] C. Bork, J.D. Schwenn, and R. Hell. Isolation and characterization of a gene for assimilatory sulfite reductase from *Arabidopsis thaliana*. *Gene*, 212:147–153, 1998.
- [394] E. Borodina, D.P. Kelly, F.A. Rainey, N.L. Ward-Rainey, and A.P. Wood. Dimethylsulfone as a growth substrate for novel methylotrophic species of *Hyphomicrobium* and *Arthrobacter*. *Arch. Microbiol.*, 173:425–437, 2000.
- [395] E. Borodina, D.P. Kelly, P. Schumann, F.A. Rainey, N.L. Ward-Rainey, and A.P. Wood. Enzymes of dimethylsulfone metabolism and the phylogenetic characterization of the facultative methylotrophs *Arthrobacter sulfonivorans* sp. nov., *Arthrobacter methylotrophus* sp. nov., and *Hyphomicrobium sulfonivorans* sp. nov. *Arch. Microbiol.*, 177:173–183, 2002.
- [396] A. Bortolotti, I. Perez-Dorado, G. Goni, M. Medina, J.A. Hermoso, N. Carrillo, and N. Cortez. Coenzyme binding and hydride transfer in *Rhodobacter capsulatus* ferredoxin/ flavodoxin NADP(H) oxidoreductase. *Biochim. Biophys. Acta*, 1794:199–210, 2009.
- [397] A. Bortolotti, A. Sanchez-Azqueta, C.M. Maya, A. Velazquez-Campoy, J.A. Hermoso, M. Medina, and N. Cortez. The C-terminal extension of bacterial flavodoxin-reductases: involvement in the hydride transfer mechanism from the coenzyme. *Biochim. Biophys. Acta*, 1837:33–43, 2014.
- [398] S. Boschi-Muller, S. Azza, D. Pollastro, C. Corbier, and G. Branlant. Comparative enzymatic properties of GapB-encoded erythrose-4-phosphate dehydrogenase of *Escherichia coli* and phosphorylating glyceraldehyde-3-phosphate dehydrogenase. *J. Biol. Chem.*, 272:15106–15112, 1997.
- [399] S. Boschi-Muller, A. Olry, M. Antoine, and G. Branlant. The enzymology and biochemistry of methionine sulfoxide reductases. *Biochim. Biophys. Acta*, 1703:231–238, 2005.
- [400] W.F. Bosron and R.L. Prairie. Triphosphopyridine nucleotide-linked aldehyde reductase. I. Purification and properties of the enzyme from pig kidney cortex. *J. Biol. Chem.*, 247:4480–4485, 1972.
- [401] I.D. Bossert, G. Whited, D.T. Gibson, and L.Y. Young. Anaerobic oxidation of *p*-cresol mediated by a partially purified methylhydroxylase from a denitrifying bacterium. *J. Bacteriol.*, 171:2956–2962, 1989.
- [402] R.T. Bossi, A. Negri, G. Tedeschi, and A. Mattevi. Structure of FAD-bound L-aspartate oxidase: insight into substrate specificity and catalysis. *Biochemistry*, 41:3018–3024, 2002.
- [403] C. Böttcher, L. Westphal, C. Schmotz, E. Prade, D. Scheel, and E. Glawischnig. The multifunctional enzyme CYP71B15 (PHYTOALEXIN DEFICIENT3) converts cysteine-indole-3-acetonitrile to camalexin in the indole-3-acetonitrile metabolic network of *Arabidopsis thaliana*. *Plant Cell*, 21:1830–1845, 2009.
- [404] C.K. Bottema and L.W. Parks. Δ^{14} -Sterol reductase in *Saccharomyces cerevisiae*. *Biochim. Biophys. Acta*, 531:301–307, 1978.
- [405] F. Bou-Abdallah, H. Yang, A. Awomolo, B. Cooper, M.R. Woodhall, S.C. Andrews, and N.D. Chasteen. Functionality of the three-site ferroxidase center of *Escherichia coli* bacterial ferritin (EcFtnA). *Biochemistry*, 53:483–495, 2014.

- [406] F. Bouvier, Y. Keller, A. d'Harlingue, and B. Camara. Xanthophyll biosynthesis: molecular and functional characterization of carotenoid hydroxylases from pepper fruits (*Capsicum annuum* L.). *Biochim. Biophys. Acta*, 1391:320–328, 1998.
- [407] H.J. Bouwmeester, J. Gershenzon, M.C.J.M. Konings, and R. Croteau. Biosynthesis of the monoterpenes limonene and carvone in the fruit of caraway. I. Demonstration of enzyme activities and their changes with development. *Plant Physiol.*, 117:901–912, 1998.
- [408] H.J. Bouwmeester, M.C.J.M. Konings, J. Gershenzon, F. Karp, and R. Croteau. Cytochrome *P*-450 dependent (+)-limonene-6-hydroxylation in fruits of caraway (*Carum carvi*). *Phytochemistry*, 50:243–248, 1999.
- [409] L. Bowater, S.A. Fairhurst, V.J. Just, and S. Bornemann. *Bacillus subtilis* YxaG is a novel Fe-containing quercetin 2,3-dioxygenase. *FEBS Lett.*, 557:45–48, 2004.
- [410] J.A. Bowden and J.L. Connelly. Branched chain α -keto acid metabolism. II. Evidence for the common identity of α -ketoisocaproic acid and α -keto- β -methyl-valeric acid dehydrogenases. *J. Biol. Chem.*, 243:3526–3531, 1968.
- [411] P.J. Bower, H.M. Brown, and W.K. Purves. Cucumber seedling indoleacetaldehyde oxidase. *Plant Physiol.*, 61:107–110, 1978.
- [412] G.S. Boyd, A.M. Grimwade, and M.E. Lawson. Studies on rat-liver microsomal cholesterol 7 α -hydroxylase. *Eur. J. Biochem.*, 37:334–340, 1973.
- [413] T.O. Boynton, L.E. Daugherty, T.A. Dailey, and H.A. Dailey. Identification of *Escherichia coli* HemG as a novel, menadione-dependent flavodoxin with protoporphyrinogen oxidase activity. *Biochemistry*, 48:6705–6711, 2009.
- [414] D. Bozic, D. Papaefthimiou, K. Bruckner, R.C. de Vos, C.A. Tsoleridis, D. Katsarou, A. Papanikolaou, I. Pateraki, F.M. Chatzopoulou, E. Dimitriadou, S. Kostas, D. Manzano, U. Scheler, A. Ferrer, A. Tissier, A.M. Makris, S.C. Kampranis, and A.K. Kanellis. Towards elucidating carnosic acid biosynthesis in *Lamiaceae*: functional characterization of the three first steps of the pathway in *Salvia fruticosa* and *Rosmarinus officinalis*. *PLoS One*, 10:e0124106–e0124106, 2015.
- [415] R. Braaz, W. Armbruster, and D. Jendrossek. Heme-dependent rubber oxygenase RoxA of *Xanthomonas* sp. cleaves the carbon backbone of poly(*cis*-1,4-Isoprene) by a dioxygenase mechanism. *Appl. Environ. Microbiol.*, 71:2473–2478, 2005.
- [416] R. Braaz, P. Fischer, and D. Jendrossek. Novel type of heme-dependent oxygenase catalyzes oxidative cleavage of rubber (*poly-cis*-1,4-isoprene). *Appl. Environ. Microbiol.*, 70:7388–7395, 2004.
- [417] R. Brackmann and G. Fuchs. Enzymes of anaerobic metabolism of phenolic compounds. 4-Hydroxybenzoyl-CoA reductase (dehydroxylating) from a denitrifying *Pseudomonas* species. *Eur. J. Biochem.*, 213:563–571, 1993.
- [418] A.F. Bradbury, M.D.A. Finnie, and D.G. Smyth. Mechanism of C-terminal amide formation by pituitary enzymes. *Nature (Lond.)*, 298:686–688, 1982.
- [419] A.F. Bradbury and D.G. Smyth. Enzyme-catalysed peptide amidation. Isolation of a stable intermediate formed by reaction of the amidating enzyme with an imino acid. *Eur. J. Biochem.*, 169:579–584, 1987.
- [420] D.R. Brady, R.D. Crowder, and W.J. Hayes. Mixed function oxidases in sterol metabolism. Source of reducing equivalents. *J. Biol. Chem.*, 255:10624–10629, 1980.
- [421] S.F. Brady and J. Clardy. Cloning and heterologous expression of isocyanide biosynthetic genes from environmental DNA. *Angew. Chem. Int. Ed. Engl.*, 44:7063–7065, 2005.
- [422] S.S. Braithwaite and J. Jarabak. Studies on a 15-hydroxyprostaglandin dehydrogenase from human placenta. Purification and partial characterization. *J. Biol. Chem.*, 250:2315–2318, 1975.
- [423] R. Branco, R. Francisco, A.P. Chung, and P.V. Morais. Identification of an aox system that requires cytochrome *c* in the highly arsenic-resistant bacterium *Ochrobactrum tritici* SCII24. *Appl. Environ. Microbiol.*, 75:5141–5147, 2009.
- [424] G.-I. Brändén, H. Jörnvall, H. Eklund, and B. Furugren. Alcohol dehydrogenase. In P.D. Boyer, editor, *The Enzymes*, volume 11, pages 103–190. Academic Press, New York, 3rd edition, 1975.

- [425] R. Brandsch, A.E. Hinkkanen, L. Mauch, H. Nagursky, and K. Decker. 6-Hydroxy-D-nicotine oxidase of *Arthrobacter oxidans*. Gene structure of the flavoenzyme and its relationship to 6-hydroxy-L-nicotine oxidase. *Eur. J. Biochem.*, 167:315–320, 1987.
- [426] A. Brauer, P. Beck, L. Hintermann, and M. Groll. Structure of the dioxygenase AsqJ: Mechanistic insights into a one-pot multistep quinolone antibiotic biosynthesis. *Angew. Chem. Int. Ed. Engl.*, 55:422–426, 2016.
- [427] M. Braun, S. Bungert, and T. Friedrich. Characterization of the overproduced NADH dehydrogenase fragment of the NADH:ubiquinone oxidoreductase (complex I) from *Escherichia coli*. *Biochemistry*, 37:1861–1867, 1998.
- [428] A. Braunshausen and F.P. Seebeck. Identification and characterization of the first ovothioliol biosynthetic enzyme. *J. Am. Chem. Soc.*, 133:1757–1759, 2011.
- [429] R.C. Bray. Xanthine oxidase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 533–556. Academic Press, New York, 2nd edition, 1963.
- [430] S.D. Breazeale, A.A. Ribeiro, A.L. McClerren, and C.R.H. Raetz. A formyltransferase required for polymyxin resistance in *Escherichia coli* and the modification of lipid A with 4-amino-4-deoxy-L-arabinose. Identification and function of UDP-4-deoxy-4-formamido-L-arabinose. *J. Biol. Chem.*, 280:14154–14167, 2005.
- [431] D.S. Bredt and S.H. Snyder. Isolation of nitric oxide synthetase, a calmodulin-requiring enzyme. *Proc. Natl. Acad. Sci. USA*, 87:682–685, 1990.
- [432] K. Breese and G. Fuchs. 4-Hydroxybenzoyl-CoA reductase (dehydroxylating) from the denitrifying bacterium *Thauera aromatica* - prosthetic groups, electron donor, and genes of a member of the molybdenum-flavin-iron-sulfur proteins. *Eur. J. Biochem.*, 251:916–923, 1998.
- [433] K. Breicha, M. Muller, W. Hummel, and K. Niefind. Crystallization and preliminary crystallographic analysis of Gre2p, an NADP⁺-dependent alcohol dehydrogenase from *Saccharomyces cerevisiae*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 66:838–841, 2010.
- [434] J. Breitenbach, M. Kuntz, S. Takaichi, and G. Sandmann. Catalytic properties of an expressed and purified higher plant type ζ -carotene desaturase from *Capsicum annuum*. *Eur. J. Biochem.*, 265:376–383, 1999.
- [435] J. Breitenbach, N. Misawa, S. Kajiwara, and G. Sandmann. Expression in *Escherichia coli* and properties of the carotene ketolase from *Haematococcus pluvialis*. *FEMS Microbiol. Lett.*, 140:241–246, 1996.
- [436] J. Breitenbach and G. Sandmann. ζ -Carotene *cis* isomers as products and substrates in the plant *poly-cis* carotenoid biosynthetic pathway to lycopene. *Planta*, 220:785–793, 2005.
- [437] J. Breitenbach, C. Zhu, and G. Sandmann. Bleaching herbicide norflurazon inhibits phytoene desaturase by competition with the cofactors. *J. Agric. Food Chem.*, 49:5270–5272, 2001.
- [438] F.N. Brenneman and W.A. Volk. Glyceraldehyde phosphate dehydrogenase activity with triphosphopyridine nucleotide and with diphosphopyridine nucleotide. *J. Biol. Chem.*, 234:2443–2447, 1959.
- [439] S.L. Bridger, S.M. Clarkson, K. Stirrett, M.B. DeBarry, G.L. Lipscomb, G.J. Schut, J. Westpheling, R.A. Scott, and M.W. Adams. Deletion strains reveal metabolic roles for key elemental sulfur-responsive proteins in *Pyrococcus furiosus*. *J. Bacteriol.*, 193:6498–6504, 2011.
- [440] R.B. Bridges, M.P. Palumbo, and C.L. Wittenberger. Purification and properties of an NADP-specific 6-phosphogluconate dehydrogenase from *Streptococcus faecalis*. *J. Biol. Chem.*, 250:6093–6100, 1975.
- [441] N.G. Brink. Beef liver glucose dehydrogenase. 1. Purification and properties. *Acta Chem. Scand.*, 7:1081–1089, 1953.
- [442] U. Brinkmann and W. Reineke. Degradation of chlorotoluenes by *in vivo* constructed hybrid strains: problems of enzyme specificity, induction and prevention of *meta*-pathway. *FEMS Microbiol. Lett.*, 75:81–87, 1992.
- [443] S. Brinkmann-Chen, J.K. Cahn, and F.H. Arnold. Uncovering rare NADH-preferring ketol-acid reductoisomerases. *Metab. Eng.*, 26C:17–22, 2014.
- [444] J.A. Brito, K. Denkmann, I.A. Pereira, M. Archer, and C. Dahl. Thiosulfate dehydrogenase (TsdA) from *Allochromatium vinosum*: structural and functional insights into thiosulfate oxidation. *J. Biol. Chem.*, 290:9222–9238, 2015.

- [445] J.A. Brito, F.L. Sousa, M. Stelter, T.M. Bandejas, C. Vonrhein, M. Teixeira, M.M. Pereira, and M. Archer. Structural and functional insights into sulfide:quinone oxidoreductase. *Biochemistry*, 48:5613–5622, 2009.
- [446] C. Brizio, R. Brandsch, D. Bufano, L. Pochini, C. Indiveri, and M. Barile. Over-expression in *Escherichia coli*, functional characterization and refolding of rat dimethylglycine dehydrogenase. *Protein Expr. Purif.*, 37:434–442, 2004.
- [447] C. Brizio, R. Brandsch, M. Douka, R. Wait, and M. Barile. The purified recombinant precursor of rat mitochondrial dimethylglycine dehydrogenase binds FAD via an autocatalytic reaction. *Int. J. Biol. Macromol.*, 42:455–462, 2008.
- [448] R.M. Broadus and J.D. Haddock. Purification and characterization of the NADH:ferredoxinBPH oxidoreductase component of biphenyl 2,3-dioxygenase from *Pseudomonas* sp. strain LB400. *Arch. Microbiol.*, 170:106–112, 1998.
- [449] J. Brodersen, G. Gottschalk, and U. Deppenmeier. Membrane-bound F₄₂₀H₂-dependent heterodisulfide reduction in *Methanococcus volta*. *Arch. Microbiol.*, 171:115–121, 1999.
- [450] F. Brodhun, C. Gobel, E. Hornung, and I. Feussner. Identification of PpoA from *Aspergillus nidulans* as a fusion protein of a fatty acid heme dioxygenase/peroxidase and a cytochrome P₄₅₀. *J. Biol. Chem.*, 284:11792–11805, 2009.
- [451] R.W. Brosemer and R.W. Kuhn. Comparative structural properties of honeybee and rabbit α -glycerophosphate dehydrogenases. *Biochemistry*, 8:2095–2105, 1969.
- [452] N. Brot, L. Weissbach, J. Werth, and H. Weissbach. Enzymatic reduction of protein-bound methionine sulfoxide. *Proc. Natl. Acad. Sci. USA*, 78:2155–2158, 1981.
- [453] P. Broun and C. Somerville. Accumulation of ricinoleic, lesquerolic, and densipolic acids in seeds of transgenic *Arabidopsis* plants that express a fatty acyl hydroxylase cDNA from castor bean. *Plant Physiol.*, 113:933–942, 1997.
- [454] S.J. Brouns, A.P. Turnbull, H.L. Willemen, J. Akerboom, and J. van der Oost. Crystal structure and biochemical properties of the D-arabinose dehydrogenase from *Sulfolobus solfataricus*. *J. Mol. Biol.*, 371:1249–1260, 2007.
- [455] S.J. Brouns, J. Walther, A.P. Snijders, H.J. van de Werken, H.L. Willemen, P. Worm, M.G. de Vos, A. Andersson, M. Lundgren, H.F. Mazon, R.H. van den Heuvel, P. Nilsson, L. Salmon, W.M. de Vos, P.C. Wright, R. Bernander, and J. van der Oost. Identification of the missing links in prokaryotic pentose oxidation pathways: evidence for enzyme recruitment. *J. Biol. Chem.*, 281:27378–27388, 2006.
- [456] R. Brouquisse, P. Weigel, D. Rhodes, C.F. Yocum, and A.D. Hanson. Evidence for a ferredoxin-dependent choline monooxygenase from spinach chloroplast stroma. *Plant Physiol.*, 90:322–329, 1989.
- [457] D.M. Brown, J.A. Upcroft, and P. Upcroft. A H₂O-producing NADH oxidase from the protozoan parasite *Giardia duodenalis*. *Eur. J. Biochem.*, 241:155–161, 1996.
- [458] E.D. Brown and J.M. Wood. Redesigned purification yields a fully functional PutA protein dimer from *Escherichia coli*. *J. Biol. Chem.*, 267:13086–13092, 1992.
- [459] F.C. Brown and D.N. Ward. Preparation of a soluble mammalian tyrosinase. *J. Am. Chem. Soc.*, 79:2647–2648, 1957.
- [460] H.M. Brown and W.K. Purves. Isolation and characterization of indole-3-acetaldehyde reductases from *Cucumis sativus*. *J. Biol. Chem.*, 251:907–913, 1976.
- [461] K.R. Brown, B.M. Allan, P. Do, and E.L. Hegg. Identification of novel hemes generated by heme A synthase: evidence for two successive monooxygenase reactions. *Biochemistry*, 41:10906–10913, 2002.
- [462] K.R. Brown, B.M. Brown, E. Hoagland, C.L. Mayne, and E.L. Hegg. Heme A synthase does not incorporate molecular oxygen into the formyl group of heme A. *Biochemistry*, 43:8616–8624, 2004.
- [463] K. Brown-Grant, E. Forchielli, and R.I. Dorfman. The Δ^4 -hydrogenases of guinea pig adrenal gland. *J. Biol. Chem.*, 235:1317–1320, 1960.
- [464] J. Browse, M. McConn, D. James, Miquel Jr., and M. Mutants of *Arabidopsis* deficient in the synthesis of α -linolenate. Biochemical and genetic characterization of the endoplasmic reticulum linoleoyl desaturase. *J. Biol. Chem.*, 268:16345–16351, 1993.
- [465] V. Bruckhoff, S. Haroth, K. Feussner, S. König, F. Brodhun, and I. Feussner. Functional characterization of CYP94-genes and identification of a novel jasmonate catabolite in flowers. *PLoS One*, 11:e0159875–e0159875, 2016.

- [466] N.A. Bruender and V. Bandarian. The radical *S*-adenosyl-L-methionine enzyme MftC catalyzes an oxidative decarboxylation of the C-terminus of the MftA peptide. *Biochemistry*, 55:2813–2816, 2016.
- [467] H. Brüggemann, F. Falinski, and U. Deppenmeier. Structure of the F₄₂₀H₂:quinone oxidoreductase of *Archaeoglobus fulgidus* identification and overproduction of the F₄₂₀H₂-oxidizing subunit. *Eur. J. Biochem.*, 267:5810–5814, 2000.
- [468] W. Brüggemann and P.R. Moog. NADH-dependent Fe³⁺ EDTA and oxygen reduction by plasma membrane vesicles from barley roots. *Physiol. Plant.*, 75:245–254, 1989.
- [469] W. Brüggemann, P.R. Moog, H. Nakagawa, P. Janiesch, and P.J.C. Kuiper. Plasma membrane-bound NADH:Fe³⁺-EDTA reductase and iron deficiency in tomato (*Lycopersicon esculentum*). Is there a Turbo reductase ? *Physiol. Plant.*, 79:339–346, 1990.
- [470] F. Brugliera, G. Barri-Rewell, T.A. Holton, and J.G. Mason. Isolation and characterization of a flavonoid 3'-hydroxylase cDNA clone corresponding to the Ht1 locus of *Petunia hybrida*. *Plant J.*, 19:441–451, 1999.
- [471] M. Brühmüller, H.K. Möhler, and K. Decker. Covalently bound flavin in D-6-hydroxynicotine oxidase from *Arthrobacter oxidans*. Purification and properties of D-6-hydroxynicotine oxidase. *Eur. J. Biochem.*, 29:143–151, 1972.
- [472] R.K. Bruick and S.L. McKnight. A conserved family of prolyl-4-hydroxylases that modify HIF. *Science*, 294:1337–1340, 2001.
- [473] N. Bruland, J.H. Wubbeler, and A. Steinbuchel. 3-Mercaptopropionate dioxygenase, a cysteine dioxygenase homologue, catalyzes the initial step of 3-mercaptopropionate catabolism in the 3,3-thiodipropionic acid-degrading bacterium *Variovorax paradoxus*. *J. Biol. Chem.*, 284:660–672, 2009.
- [474] A. Brune and B. Schink. Pyrogallol-to-phloroglucinol conversion and other hydroxyl-transfer reactions catalyzed by cell extracts of *Pelobacter acidigallici*. *J. Bacteriol.*, 172:1070–1076, 1990.
- [475] F. Brunel and J. Davison. Cloning and sequencing of *Pseudomonas* genes encoding vanillate demethylase. *J. Bacteriol.*, 170:4924–4930, 1988.
- [476] N.A. Brunner, H. Brinkmann, B. Siebers, and R. Hensel. NAD⁺-dependent glyceraldehyde-3-phosphate dehydrogenase from *Thermoproteus tenax*. The first identified archaeal member of the aldehyde dehydrogenase superfamily is a glycolytic enzyme with unusual regulatory properties. *J. Biol. Chem.*, 273:6149–6156, 1998.
- [477] N.A. Brunner, B. Siebers, and R. Hensel. Role of two different glyceraldehyde-3-phosphate dehydrogenases in controlling the reversible Embden-Meyerhof-Parnas pathway in *Thermoproteus tenax*: regulation on protein and transcript level. *Extremophiles*, 5:101–109, 2001.
- [478] F.O. Bryant and M.W. Adams. Characterization of hydrogenase from the hyperthermophilic archaeobacterium, *Pyrococcus furiosus*. *J. Biol. Chem.*, 264:5070–5079, 1989.
- [479] R.W. Bryant, J.M. Bailey, T. Schewq, and S.M. Rapoport. Positional specificity of a reticulocyte lipoxygenase. Conversion of arachidonic acid to 15-*S*-hydroperoxy-ecosatetraenoic acid. *J. Biol. Chem.*, 257:6050–6055, 1982.
- [480] N.R. Buan and W.W. Metcalf. Methanogenesis by *Methanosarcina acetivorans* involves two structurally and functionally distinct classes of heterodisulfide reductase. *Mol. Microbiol.*, 75:843–853, 2010.
- [481] A. Bucci, T.Q. Yu, E. Vanden-Eijnden, and C.F. Abrams. Kinetics of O₂ entry and exit in monomeric sarcosine oxidase via Markovian milestone molecular dynamics. *J Chem Theory Comput*, 12:2964–2972, 2016.
- [482] K. Buch, H. Stransky, and A. Hager. FAD is a further essential cofactor of the NAD(P)H and O₂-dependent zeaxanthin-epoxidase. *FEBS Lett.*, 376:45–48, 1995.
- [483] B.B. Buchanan. Role of ferredoxin in the synthesis of α -ketobutyrate from propionyl coenzyme A and carbon dioxide by enzymes from photosynthetic and nonphotosynthetic bacteria. *J. Biol. Chem.*, 244:4218–4223, 1969.
- [484] B.B. Buchanan. Regulation of CO₂ assimilation in oxygenic photosynthesis: the ferredoxin/thioredoxin system. Perspective on its discovery, present status, and future development. *Arch. Biochem. Biophys.*, 288:1–9, 1991.
- [485] B.B. Buchanan and M.C.W. Evans. The synthesis of α -ketoglutarate from succinate and carbon dioxide by a subcellular preparation of a photosynthetic bacterium. *Proc. Natl. Acad. Sci. USA*, 54:1212–1218, 1965.

- [486] N.L.R. Bucher, P. Overath, and F. Lynen. β -Hydroxy- β -methylglutaryl coenzyme A reductase, cleavage and condensing enzymes in relation to cholesterol formation in rat liver. *Biochim. Biophys. Acta*, 40:491–501, 1960.
- [487] T.J. Buckhout and T.C. Hrubec. Pyridine nucleotide-dependent ferricyanide reduction associated with isolated plasma membranes of maize (*Zea mays* L.) roots. *Protoplasma*, 135:144–154, 1986.
- [488] R. Buder and G. Fuchs. 2-Aminobenzoyl-CoA monooxygenase/reductase, a novel type of flavoenzyme. Purification and some properties of the enzyme. *Eur. J. Biochem.*, 185:629–635, 1989.
- [489] R. Buder, K. Ziegler, G. Fuchs, B. Langkau, and S. Ghisla. 2-Aminobenzoyl-CoA monooxygenase/reductase, a novel type of flavoenzyme. Studies on the stoichiometry and the course of the reaction. *Eur. J. Biochem.*, 185:637–643, 1989.
- [490] R.C. Bugos, A.D. Hieber, and H.Y. Yamamoto. Xanthophyll cycle enzymes are members of the lipocalin family, the first identified from plants. *J. Biol. Chem.*, 273:15321–15324, 1998.
- [491] H.A. Bullock, H. Luo, and W.B. Whitman. Evolution of dimethylsulfoniopropionate metabolism in marine phytoplankton and bacteria. *Front. Microbiol.*, 8:637–637, 2017.
- [492] J.R. Bundred, E. Hendrix, and M.L. Coleman. The emerging roles of ribosomal histidyl hydroxylases in cell biology, physiology and disease. *Cell. Mol. Life Sci.*, 75:4093–4105, 2018.
- [493] G.L. Bundy, E.G. Nidy, D.E. Epps, S.A. Mizsak, and R.J. Wnuk. Discovery of an arachidonic acid C-8 lipoxygenase in the gorgonian coral *Pseudoplexaura porosa*. *J. Biol. Chem.*, 261:747–751, 1986.
- [494] V.I. Bunik and D. Degtyarev. Structure-function relationships in the 2-oxo acid dehydrogenase family: substrate-specific signatures and functional predictions for the 2-oxoglutarate dehydrogenase-like proteins. *Proteins*, 71:874–890, 2008.
- [495] R.W. Burg and E.E. Snell. The bacterial oxidation of vitamin B₆. VI. Pyridoxal dehydrogenase and 4-pyridoxolactonase. *J. Biol. Chem.*, 244:2585–2589, 1969.
- [496] W. Burgi, R. Richterich, and J.P. Colombo. L-Lysine dehydrogenase deficiency in a patient with congenital lysine intolerance. *Nature (Lond.)*, 211:854–855, 1966.
- [497] R. Burlingame and P.J. Chapman. Catabolism of phenylpropionic acid and its 3-hydroxy derivative by *Escherichia coli*. *J. Bacteriol.*, 155:113–121, 1983.
- [498] R.P. Burlingame, L. Wyman, and P.J. Chapman. Isolation and characterization of *Escherichia coli* mutants defective for phenylpropionate degradation. *J. Bacteriol.*, 168:55–64, 1986.
- [499] J.N. Burnell and R.S. Holmes. Purification and properties of sorbitol dehydrogenase from mouse liver. *Int. J. Biochem.*, 15:507–511, 1983.
- [500] M. Burnet, P.J. Lafontaine, and A.D. Hanson. Assay, purification, and partial characterization of choline monooxygenase from spinach. *Plant Physiol.*, 108:581–588, 1995.
- [501] R.O. Burns, H.E. Umbarger, and S.R. Gross. The biosynthesis of leucine. III. The conversion of α -hydroxy- β -carboxyisocaproate to α -ketoisocaproate. *Biochemistry*, 2:1053–1053, 1963.
- [502] R.B. Burrows and G.M. Brown. Presence of *Escherichia coli* of a deaminase and a reductase involved in biosynthesis of riboflavin. *J. Bacteriol.*, 136:657–667, 1978.
- [503] S. Burstein, B.S. Middleditch, and M. Gut. Mass spectrometric study of the enzymatic conversion of cholesterol to (22R)-22-hydroxycholesterol, (20R,22R)-20,22-dihydroxycholesterol, and pregnenolone, and of (22R)-22-hydroxycholesterol to the Igcol and pregnenolone in bovine adrenocortical preparations. Mode of oxygen incorporation. *J. Biol. Chem.*, 250:9028–9037, 1975.
- [504] J. Bursy, A.U. Kuhlmann, M. Pittelkow, H. Hartmann, M. Jebbar, A.J. Pierik, and E. Bremer. Synthesis and uptake of the compatible solutes ectoine and 5-hydroxyectoine by *Streptomyces coelicolor* A3(2) in response to salt and heat stresses. *Appl. Environ. Microbiol.*, 74:7286–7296, 2008.
- [505] J. Bursy, A.J. Pierik, N. Pica, and E. Bremer. Osmotically induced synthesis of the compatible solute hydroxyectoine is mediated by an evolutionarily conserved ectoine hydroxylase. *J. Biol. Chem.*, 282:31147–31155, 2007.

- [506] R.M. Burton and N.O. Kaplan. A DPN specific glycerol dehydrogenase from *Aerobacter aerogenes*. *J. Am. Chem. Soc.*, 75:1005–1007, 1953.
- [507] R.M. Burton and E.R. Stadtman. The oxidation of acetaldehyde to acetyl coenzyme A. *J. Biol. Chem.*, 202:873–890, 1953.
- [508] K.B. Busch and H. Fromm. Plant succinic semialdehyde dehydrogenase. Cloning, purification, localization in mitochondria, and regulation by adenine nucleotides. *Plant Physiol.*, 121:589–597, 1999.
- [509] I.E. Bush, S.A. Hunter, and R.A. Meigs. Metabolism of 11-oxygenated steroids. Metabolism in vitro by preparations of liver. *Biochem. J.*, 107:239–258, 1968.
- [510] P.K. Busk and B.L. Møller. Dhurrin synthesis in sorghum is regulated at the transcriptional level and induced by nitrogen fertilization in older plants. *Plant Physiol.*, 129:1222–1231, 2002.
- [511] M. Busquets, V. Deroncelle, J. Vidal-Mas, E. Rodriguez, A. Guerrero, and A. Manresa. Isolation and characterization of a lipoxygenase from *Pseudomonas* 42A2 responsible for the biotransformation of oleic acid into (S)-(E)-10-hydroxy-8-octadecenoic acid. *Antonie Van Leeuwenhoek*, 85:129–139, 2004.
- [512] G. Byng, R. Whitaker, C. Flick, and R.A. Jensen. Enzymology of L-tyrosine biosynthesis in corn (*Zea mays*). *Phytochemistry*, 20:1289–1292, 1981.
- [513] G.S. Byng, R.J. Whitaker, R.L. Gherna, and R.A. Jensen. Variable enzymological patterning in tyrosine biosynthesis as a means of determining natural relatedness among the Pseudomonadaceae. *J. Bacteriol.*, 144:247–257, 1980.
- [514] L.V. Bystrykh, J. Vonck, E.F. van Bruggen, J. van Beeumen, B. Samyn, N.I. Govorukhina, N. Arfman, J.A. Duine, and L. Dijkhuizen. Electron microscopic analysis and structural characterization of novel NADP(H)-containing methanol: N,N'-dimethyl-4-nitrosoaniline oxidoreductases from the gram-positive methylotrophic bacteria *Amycolatopsis methanolica* and *Mycobacterium gastri* MB19. *J. Bacteriol.*, 175:1814–1822, 1993.
- [515] C., Cronan Grabau, , and Jr. *In vivo* function of *Escherichia coli* pyruvate oxidase specifically requires a functional lipid binding site. *Biochemistry*, 25:3748–3751, 1986.
- [516] M.A. Caccamo, C.S. Malone, and M.E. Rasche. Biochemical characterization of a dihydromethanopterin reductase involved in tetrahydromethanopterin biosynthesis in *Methylobacterium extorquens* AM1. *J. Bacteriol.*, 186:2068–2073, 2004.
- [517] E. Cadieux, V. Vrajmasu, C. Achim, J. Powlowski, and E. Munck. Biochemical, Mossbauer, and EPR studies of the diiron cluster of phenol hydroxylase from *Pseudomonas* sp. strain CF 600. *Biochemistry*, 41:10680–10691, 2002.
- [518] E.B. Cahoon, T.J. Carlson, K.G. Ripp, B.J. Schweiger, G.A. Cook, S.E. Hall, and A.J. Kinney. Biosynthetic origin of conjugated double bonds: production of fatty acid components of high-value drying oils in transgenic soybean embryos. *Proc. Natl. Acad. Sci. USA*, 96:12935–12940, 1999.
- [519] E.B. Cahoon, A.M. Cranmer, J. Shanklin, and J.B. Ohlrogge. Δ^6 Hexadecenoic acid is synthesized by the activity of a soluble Δ^6 palmitoyl-acyl carrier protein desaturase in *Thunbergia alata* endosperm. *J. Biol. Chem.*, 269:27519–27526, 1994.
- [520] E.B. Cahoon and A.J. Kinney. Dimorphecolic acid is synthesized by the coordinate activities of two divergent Δ^{12} -oleic acid desaturases. *J. Biol. Chem.*, 279:12495–12502, 2004.
- [521] E.B. Cahoon, Y. Lindqvist, G. Schneider, and J. Shanklin. Redesign of soluble fatty acid desaturases from plants for altered substrate specificity and double bond position. *Proc. Natl. Acad. Sci. USA*, 94:4872–4877, 1997.
- [522] E.B. Cahoon, E.F. Marillia, K.L. Stecca, S.E. Hall, D.C. Taylor, and A.J. Kinney. Production of fatty acid components of meadowfoam oil in somatic soybean embryos. *Plant Physiol.*, 124:243–251, 2000.
- [523] E.B. Cahoon and J.B. Ohlrogge. Metabolic evidence for the involvement of a Δ^4 -palmitoyl-acyl carrier protein desaturase in petroselinic acid synthesis in coriander endosperm and transgenic tobacco cells. *Plant Physiol.*, 104:827–837, 1994.
- [524] E.B. Cahoon, K.G. Ripp, S.E. Hall, and A.J. Kinney. Formation of conjugated Δ^8, Δ^{10} -double bonds by Δ^{12} -oleic-acid desaturase-related enzymes: biosynthetic origin of calendic acid. *J. Biol. Chem.*, 276:2637–2643, 2001.

- [525] E.B. Cahoon, J. Shanklin, and J.B. Ohlrogge. Expression of a coriander desaturase results in petroselinic acid production in transgenic tobacco. *Proc. Natl. Acad. Sci. USA*, 89:11184–11188, 1992.
- [526] D.Y. Cai and M. Tien. Characterization of the oxycomplex of lignin peroxidases from *Phanerochaete chrysosporium*: equilibrium and kinetics studies. *Biochemistry*, 29:2085–2091, 1990.
- [527] G.A. Caignan, R. Deshmukh, A. Wilks, Y. Zeng, H.W. Huang, P. Moenne-Loccoz, R.A. Bunce, M.A. Eastman, and M. Rivera. Oxidation of heme to β - and δ -biliverdin by *Pseudomonas aeruginosa* heme oxygenase as a consequence of an unusual seating of the heme. *J. Am. Chem. Soc.*, 124:14879–14892, 2002.
- [528] H. Caldas and G.E. Herman. NSDHL, an enzyme involved in cholesterol biosynthesis, traffics through the Golgi and accumulates on ER membranes and on the surface of lipid droplets. *Hum. Mol. Genet.*, 12:2981–2991, 2003.
- [529] D.M. Callewaert, M.S. Roseblatt, and T.T. Tchen. Purification and properties of 4-aminobutanal dehydrogenase from a *Pseudomonas* species. *J. Biol. Chem.*, 249:1737–1741, 1974.
- [530] A.F. Calvert and V.W. Rodwell. Metabolism of pipercolic acid in a *Pseudomonas* species. 3. L- α -Aminoadipate δ -semialdehyde:nicotinamide adenine dinucleotide oxidoreductase. *J. Biol. Chem.*, 241:409–414, 1966.
- [531] J. Calvo. M., Stevens, C. M., Kalyanpur, M. G., and Umbarger, H. E. The absolute configuration of α -hydroxy- β -carboxyisocaproic acid (3-isopropylmalic acid), an intermediate in leucine biosynthesis. *Biochemistry*, 3:2024–2027, 1964.
- [532] M.L. Camacho, R.A. Brown, M.J. Bonete, M.J. Danson, and D.W. Hough. Isocitrate dehydrogenases from *Haloflex volcanii* and *Sulfolobus solfataricus*: enzyme purification, characterisation and N-terminal sequence. *FEMS Microbiol. Lett.*, 134:85–90, 1995.
- [533] S. Camarero, F.J. Ruiz-Due nas, S. Sarkar, M.J. Martínez, and A.T. Martínez. The cloning of a new peroxidase found in lignocellulose cultures of *Pleurotus eryngii* and sequence comparison with other fungal peroxidases. *FEMS Microbiol. Lett.*, 191:37–43, 2000.
- [534] S. Camarero, S. Sarkar, F.J. Ruiz-Due nas, M.J. Martínez, and A.T. Martínez. Description of a versatile peroxidase involved in the natural degradation of lignin that has both manganese peroxidase and lignin peroxidase substrate interaction sites. *J. Biol. Chem.*, 274:10324–10330, 1999.
- [535] S. Cameron, K. McLuskey, R. Chamberlayne, I. Hallyburton, and W.N. Hunter. Initiating a crystallographic analysis of recombinant (S)-2-hydroxypropylphosphonic acid epoxidase from *Streptomyces wedmorensis*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:534–536, 2005.
- [536] R. Cammack. Assay, purification and properties of mammalian D-2-hydroxy acid dehydrogenase. *Biochem. J.*, 115:55–64, 1969.
- [537] R. Cammack. D-2-hydroxy acid dehydrogenase from animal tissue. *Methods Enzymol.*, 41:323–329, 1975.
- [538] R. Cammack, R.H. Jackson, A. Cornish-Bowden, and J.A. Cole. Electron-spin-resonance studies of the NADH-dependent nitrite reductase from *Escherichia coli* K12. *Biochem. J.*, 207:333–339, 1982.
- [539] B. Camoretti-Mercado and R.B. Frydman. Separation of tryptophan pyrroloxygenase into three molecular forms. A study of their substrate specificities using tryptophyl-containing peptides and proteins. *Eur. J. Biochem.*, 156:317–325, 1986.
- [540] L.L. Campbell. Reductive degradation of pyrimidines. III. Purification and properties of dihydrouracil dehydrogenase. *J. Biol. Chem.*, 227:693–700, 1957.
- [541] W.H. Campbell. Structure and function of eukaryotic NAD(P)H:nitrate reductase. *Cell. Mol. Life Sci.*, 58:194–204, 2001.
- [542] M. Can, F.A. Armstrong, and S.W. Ragsdale. Structure, function, and mechanism of the nickel metalloenzymes, CO dehydrogenase, and acetyl-CoA synthase. *Chem. Rev.*, 114:4149–4174, 2014.
- [543] K.A. Canada, S. Iwashita, H. Shim, and T.K. Wood. Directed evolution of toluene *ortho*-monooxygenase for enhanced 1-naphthol synthesis and chlorinated ethene degradation. *J. Bacteriol.*, 184:344–349, 2002.

- [544] D.E. Cane, Y. Hsiung, J.A. Cornish, J.K. Robinson, and I.D. Spenser. Biosynthesis of vitamin B₆: The oxidation of L-threonine 4-phosphate by PdxA. *J. Am. Chem. Soc.*, 120:1936–1937, 1998.
- [545] V. Cantagrel, D.J. Lefeber, B.G. Ng, Z. Guan, J.L. Silhavy, S.L. Bielas, L. Lehle, H. Hombauer, M. Adamowicz, E. Swiezewska, A.P. De Brouwer, P. Blumel, J. Sykut-Cegielska, S. Houliston, D. Swistun, B.R. Ali, W.B. Dobyns, D. Babovic-Vuksanovic, H. van Bokhoven, R.A. Wevers, C.R. Raetz, H.H. Freeze, E. Morava, L. Al-Gazali, and J.G. Gleeson. SRD5A3 is required for converting polyprenol to dolichol and is mutated in a congenital glycosylation disorder. *Cell*, 142:203–217, 2010.
- [546] C. Cantwell, R. Beckmann, P. Whiteman, S.W. Queener, and E.P. Abraham. Isolation of deacetoxycephalosporin-c from fermentation broths of *Penicillium chrysogenum* transformants - construction of a new fungal biosynthetic-pathway. *Proc. R. Soc. Lond. B Biol. Sci.*, 248:283–289, 1992.
- [547] Y. Cao, M. Xian, J. Yang, X. Xu, W. Liu, and L. Li. Heterologous expression of stearoyl-acyl carrier protein desaturase (S-ACP-DES) from *Arabidopsis thaliana* in *Escherichia coli*. *Protein Expr. Purif.*, 69:209–214, 2010.
- [548] R. Caputto and M. Dixon. Crystallization and identity of the triose and triosephosphate dehydrogenase of muscle. *Nature (Lond.)*, 156:630–631, 1945.
- [549] J.K. Capyk, I. Casabon, R. Gruninger, N.C. Strynadka, and L.D. Eltis. Activity of 3-ketosteroid 9 α -hydroxylase (KshAB) indicates cholesterol side chain and ring degradation occur simultaneously in *Mycobacterium tuberculosis*. *J. Biol. Chem.*, 286:40717–40724, 2011.
- [550] J.K. Capyk, I. D'Angelo, N.C. Strynadka, and L.D. Eltis. Characterization of 3-ketosteroid 9 α -hydroxylase, a Rieske oxygenase in the cholesterol degradation pathway of *Mycobacterium tuberculosis*. *J. Biol. Chem.*, 284:9937–9946, 2009.
- [551] J.K. Capyk, R. Kalscheuer, G.R. Stewart, J. Liu, H. Kwon, R. Zhao, S. Okamoto, W.R. Jacobs, Eltis Jr., Mohn L.D., and W.W. Mycobacterial cytochrome P450 125 (Cyp125) catalyzes the terminal hydroxylation of C27 steroids. *J. Biol. Chem.*, 284:35534–35542, 2009.
- [552] L. Caramelo, M.J. Martínez, and A.T. Martínez. A search for ligninolytic peroxidases in the fungus *Pleurotus eryngii* involving α -keto- γ -thiomethylbutyric acid and lignin model dimer. *Appl. Environ. Microbiol.*, 65:916–922, 1999.
- [553] J. Carballo, R. Martin, A. Bernardo, and J. Gonzalez. Purification, characterization and some properties of diacetyl(acetoin) reductase from *Enterobacter aerogenes*. *Eur. J. Biochem.*, 198:327–332, 1991.
- [554] G. Cardini and P. Jurtshuk. The enzymatic hydroxylation of *n*-octane by *Corynebacterium* sp. strain 7E1C. *J. Biol. Chem.*, 245:2789–2796, 1970.
- [555] J. Carere, P. Baker, and S.Y.K. Seah. Investigating the molecular determinants for substrate channeling in BphI-BphJ, an aldolase-dehydrogenase complex from the polychlorinated biphenyls degradation pathway. *Biochemistry*, 50:8407–8416, 2011.
- [556] Y. Carius, H. Christian, A. Faust, U. Zander, B.U. Klink, P. Kornberger, G.W. Kohring, F. Giffhorn, and A.J. Scheidig. Structural insight into substrate differentiation of the sugar-metabolizing enzyme galactitol dehydrogenase from *Rhodobacter sphaeroides* D. *J. Biol. Chem.*, 285:20006–20014, 2010.
- [557] I. Carlberg and B. Mannervik. Purification by affinity chromatography of yeast glutathione reductase, the enzyme responsible for the NADPH-dependent reduction of the mixed disulfide of coenzyme A and glutathione. *Biochim. Biophys. Acta*, 484:268–274, 1977.
- [558] L. Carlisle-Moore, C.R. Gordon, C.A. Machutta, W.T. Miller, and P.J. Tonge. Substrate recognition by the human fatty-acid synthase. *J. Biol. Chem.*, 280:42612–42618, 2005.
- [559] B.L. Carlson, E.R. Ballister, E. Skordalakes, D.S. King, M.A. Breidenbach, S.A. Gilmore, J.M. Berger, and C.R. Bertozzi. Function and structure of a prokaryotic formylglycine-generating enzyme. *J. Biol. Chem.*, 283:20117–20125, 2008.
- [560] E. De Carolis, F. Chan, J. Balsevich, and V. De Luca. Isolation and characterization of a 2-oxoglutarate dependent dioxygenase involved in the 2nd-to-last step in vindoline biosynthesis. *Plant Physiol.*, 94:1323–1329, 1990.
- [561] E. De Carolis and V. De Luca. Purification, characterization, and kinetic analysis of a 2-oxoglutarate-dependent dioxygenase involved in vindoline biosynthesis from *Catharanthus roseus*. *J. Biol. Chem.*, 268:5504–5511, 1993.

- [562] G. Carpani, M. Racchi, P. Ghezzi, M. Terao, and E. Garattini. Purification and characterization of mouse liver xanthine oxidase. *Arch. Biochem. Biophys.*, 279:237–241, 1990.
- [563] R.T. Carr, S. Balasubramanian, P.C. Hawkins, and S.J. Benkovic. Mechanism of metal-independent hydroxylation by *Chromobacterium violaceum* phenylalanine hydroxylase. *Biochemistry*, 34:7525–7532, 1995.
- [564] J.E. Carroll, G.W. Kosicki, and R.J. Thibert. α -Substituted cystines as possible substrates for cystine reductase and L-amino acid oxidase. *Biochim. Biophys. Acta*, 198:601–603, 1970.
- [565] M.S. Carter, X. Zhang, H. Huang, J.T. Bouvier, B.S. Francisco, M.W. Vetting, N. Al-Obaidi, J.B. Bonanno, A. Ghosh, R.G. Zallot, H.M. Andersen, S.C. Almo, and J.A. Gerlt. Functional assignment of multiple catabolic pathways for D-apiose. *Nat. Chem. Biol.*, 14:696–705, 2018.
- [566] J.N. Carter-Franklin and A. Butler. Vanadium bromoperoxidase-catalyzed biosynthesis of halogenated marine natural products. *J. Am. Chem. Soc.*, 126:15060–15066, 2004.
- [567] L.N. Cartwright and R.P. Hullin. Purification and properties of two glyoxylate reductases from a species of *Pseudomonas*. *Biochem. J.*, 101:781–791, 1966.
- [568] V. Casaite, S. Poviloniene, R. Meskiene, R. Rutkiene, and R. Meskys. Studies of dimethylglycine oxidase isoenzymes in *Arthrobacter globiformis* cells. *Curr. Microbiol.*, 62:1267–1273, 2011.
- [569] C.L. Case, J.R. Rodriguez, and B. Mukhopadhyay. Characterization of an NADH oxidase of the flavin-dependent disulfide reductase family from *Methanocaldococcus jannaschii*. *Microbiology*, 155:69–79, 2009.
- [570] M. Casellas, M. Grifoll, J.M. Bayona, and A.M. Solanas. New metabolites in the degradation of fluorene by *Arthrobacter* sp. strain F101. *Appl. Environ. Microbiol.*, 63:819–826, 1997.
- [571] J.R. Cashman. Structural and catalytic properties of the mammalian flavin-containing monooxygenase. *Chem. Res. Toxicol.*, 8:165–181, 1995.
- [572] J.R. Cashman and J. Zhang. Human flavin-containing monooxygenases. *Annu. Rev. Pharmacol. Toxicol.*, 46:65–100, 2006.
- [573] C. Castelle, M. Guiral, G. Malarte, F. Ledgham, G. Leroy, M. Brugna, and M.T. Giudici-Ortoni. A new iron-oxidizing/O₂-reducing supercomplex spanning both inner and outer membranes, isolated from the extreme acidophile *Acidithiobacillus ferrooxidans*. *J. Biol. Chem.*, 283:25803–25811, 2008.
- [574] P.A. Castric. Glycine metabolism by *Pseudomonas aeruginosa*: Hydrogen cyanide biosynthesis. *J. Bacteriol.*, 130:826–831, 1977.
- [575] I. Caughey and R.G.O. Kekwick. The characteristics of some components of the fatty acid synthetase system in the plastids from the mesocarp of avocado (*Persea americana*) fruit. *Eur. J. Biochem.*, 123:553–561, 1982.
- [576] C. Causeret, L. Geeraert, G. Van der Hoeven, G.P. Mannaerts, and P.P. Van Veldhoven. Further characterization of rat dihydroceramide desaturase: tissue distribution, subcellular localization, and substrate specificity. *Lipids*, 35:1117–1125, 2000.
- [577] C. Cavaliere, N. Biermann, M.D. Vlasie, O. Einsle, A. Merli, D. Ferrari, G.L. Rossi, and M. Ubbink. Structural comparison of crystal and solution states of the 138 kDa complex of methylamine dehydrogenase and amicyanin from *Paracoccus versutus*. *Biochemistry*, 47:6560–6570, 2008.
- [578] D. Cavallini, C. de Marco, R. Scandurra, S. Duprè, and M.T. Graziani. The enzymatic oxidation of cysteamine to hypotaurine. Purification and properties of the enzyme. *J. Biol. Chem.*, 241:3189–3196, 1966.
- [579] D. Cavallini, G. Federici, G. Ricci, S. Duprè, and A. Antonucci. The specificity of cysteamine oxygenase. *FEBS Lett.*, 56:348–351, 1975.
- [580] D.R. Cavener and R.J. MacIntyre. Biphasic expression and function of glucose dehydrogenase in *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. USA*, 80:6286–6288, 1983.

- [581] C. Ceccarelli, N.B. Grodsky, N. Ariyaratne, R.F. Colman, and B.J. Bahnson. Crystal structure of porcine mitochondrial NADP⁺-dependent isocitrate dehydrogenase complexed with Mn²⁺ and isocitrate. Insights into the enzyme mechanism. *J. Biol. Chem.*, 277:43454–43462, 2002.
- [582] G. Cecchini. Function and structure of complex II of the respiratory chain. *Annu. Rev. Biochem.*, 72:77–109, 2003.
- [583] G. Cecchini, I. Schroder, R.P. Gunsalus, and E. Maklashina. Succinate dehydrogenase and fumarate reductase from *Escherichia coli*. *Biochim. Biophys. Acta*, 1553:140–157, 2002.
- [584] A.I. Celis, G.H. Gauss, B.R. Streit, K. Shisler, G.C. Moraski, K.R. Rodgers, G.S. Lukat-Rodgers, J.W. Peters, and J.L. DuBois. Structure-based mechanism for oxidative decarboxylation reactions mediated by amino acids and heme propionates in coproheme decarboxylase (HemQ). *J. Am. Chem. Soc.*, 139:1900–1911, 2017.
- [585] A.I. Celis, B.R. Streit, G.C. Moraski, R. Kant, T.D. Lash, G.S. Lukat-Rodgers, K.R. Rodgers, and J.L. DuBois. Unusual peroxide-dependent, heme-transforming reaction catalyzed by HemQ. *Biochemistry*, 54:4022–4032, 2015.
- [586] M. Cervelli, F. Polticelli, R. Federico, and P. Mariottini. Heterologous expression and characterization of mouse spermine oxidase. *J. Biol. Chem.*, 278:5271–5276, 2003.
- [587] P. Chaiyen. Flavoenzymes catalyzing oxidative aromatic ring-cleavage reactions. *Arch. Biochem. Biophys.*, 493:62–70, 2010.
- [588] P. Chaiyen, D.P. Ballou, and V. Massey. Gene cloning, sequence analysis, and expression of 2-methyl-3-hydroxypyridine-5-carboxylic acid oxygenase. *Proc. Natl. Acad. Sci. USA*, 94:7233–7238, 1997.
- [589] E. Chambellon, L. Rijnen, F. Lorquet, C. Gitton, J.E. van Hylckama Vlieg, J.A. Wouters, and M. Yvon. The D-2-hydroxyacid dehydrogenase incorrectly annotated PanE is the sole reduction system for branched-chain 2-keto acids in *Lactococcus lactis*. *J. Bacteriol.*, 191:873–881, 2009.
- [590] V. Champreda, Y.J. Choi, N.Y. Zhou, and D.J. Leak. Alteration of the stereo- and regioselectivity of alkene monooxygenase based on coupling protein interactions. *Appl. Microbiol. Biotechnol.*, 71:840–847, 2006.
- [591] V. Champreda, N.Y. Zhou, and D.J. Leak. Heterologous expression of alkene monooxygenase components from *Xanthobacter autotrophicus* Py2 and reconstitution of the active complex. *FEMS Microbiol. Lett.*, 239:309–318, 2004.
- [592] J.M. Chan, W. Wu, D.R. Dean, and L.C. Seefeldt. Construction and characterization of a heterodimeric iron protein: defining roles for adenosine triphosphate in nitrogenase catalysis. *Biochemistry*, 39:7221–7228, 2000.
- [593] M.K. Chan, S. Mukund, A. Kletzin, M.W.W. Adams, and D.C. Rees. Structure of a hyperthermophilic tungstopterin enzyme, aldehyde ferredoxin oxidoreductase. *Science*, 267:1463–1469, 1995.
- [594] P.K. Chang, J. Yu, K.C. Ehrlich, S.M. Boue, B.G. Montalbano, D. Bhatnagar, and T.E. Cleveland. *adhA* in *Aspergillus parasiticus* is involved in conversion of 5'-hydroxyaverantin to averufin. *Appl. Environ. Microbiol.*, 66:4715–4719, 2000.
- [595] S. Chang, J. Berman, Y. Sheng, Y. Wang, T. Capell, L. Shi, X. Ni, G. Sandmann, P. Christou, and C. Zhu. Cloning and functional characterization of the maize (*Zea mays* L.) carotenoid ϵ hydroxylase gene. *PLoS One*, 10:e0128758–e0128758, 2015.
- [596] S. Chang, B. Duerr, and G. Serif. An epimerase-reductase in L-fucose synthesis. *J. Biol. Chem.*, 263:1693–1697, 1988.
- [597] S.H. Chang and D.R. Wilken. Participation of the unsymmetrical disulfide of coenzyme A and glutathione in an enzymatic sulfhydryl-disulfide interchange. I. Partial purification and properties of the bovine kidney enzyme. *J. Biol. Chem.*, 241:4251–4260, 1966.
- [598] T.K. Chang, J. Teixeira, G., Waxman Gil, , and CYP. 3A10, is an active catalyst of steroid-hormone 6 β -hydroxylation. *Biochem. J.*, 291:429–433, 1993.
- [599] T.S. Chang, W. Jeong, H.A. Woo, S.M. Lee, S. Park, and S.G. Rhee. Characterization of mammalian sulfiredoxin and its reactivation of hyperoxidized peroxiredoxin through reduction of cysteine sulfinic acid in the active site to cysteine. *J. Biol. Chem.*, 279:50994–51001, 2004.
- [600] W.C. Chang, J. Li, J.L. Lee, A.A. Cronican, and Y. Guo. Mechanistic investigation of a non-heme iron enzyme catalyzed epoxidation in (–)-4'-methoxycyclophenin biosynthesis. *J. Am. Chem. Soc.*, 138:10390–10393, 2016.

- [601] W.C. Chang, D. Sanyal, J.L. Huang, K. Ittiarnornkul, Q. Zhu, and X. Liu. *In vitro* stepwise reconstitution of amino acid derived vinyl isocyanide biosynthesis: detection of an elusive intermediate. *Org. Lett.*, 19:1208–1211, 2017.
- [602] Y. F. Chang and E. Adams. Glutaric semialdehyde dehydrogenase (*Pseudomonas putida*). *Methods Enzymol.*, 17B:166–171, 1971.
- [603] Y.F. Chang and E. Adams. Glutarate semialdehyde dehydrogenase of *Pseudomonas*. Purification, properties, and relation to L-lysine catabolism. *J. Biol. Chem.*, 252:7979–7986, 1977.
- [604] F.C. Charalampous. Biochemical studies on inositol. V. Purification and properties of the enzyme that cleaves inositol to D-glucuronic acid. *J. Biol. Chem.*, 234:220–227, 1959.
- [605] A. Charbonneau and V.L. The. Genomic organization of a human 5 β -reductase and its pseudogene and substrate selectivity of the expressed enzyme. *Biochim. Biophys. Acta*, 1517:228–235, 2001.
- [606] H. Charbonneau and M.J. Cormier. Ca²⁺-induced bioluminescence in *Renilla reniformis*. Purification and characterization of a calcium-triggered luciferin-binding protein. *J. Biol. Chem.*, 254:769–780, 1979.
- [607] A.M. Charles and I. Suzuki. Purification and properties of sulfite:cytochrome *c* oxidoreductase from *Thiobacillus novellus*. *Biochim. Biophys. Acta*, 128:522–534, 1966.
- [608] Y. Charng, S.J. Chou, W.T. Jiaang, S.T. Chen, and S.F. Yang. The catalytic mechanism of 1-aminocyclopropane-1-carboxylic acid oxidase. *Arch. Biochem. Biophys.*, 385:179–185, 2001.
- [609] L. Charon, C. Pale-Grosdemange, and M. Rohmer. On the reduction steps in the mevalonate independent 2-C-methyl-D-erythritol 4-phosphate (MEP) pathway for isoprenoid biosynthesis in the bacterium *Zymomonas mobilis*. *Tetrahedron Lett.*, 40:7231–7234, 1999.
- [610] M.-H. Charon, A. Volbeda, E. Chabriere, L. Pieulle, and J.C. Fontecilla-Camps. Structure and electron transfer mechanism of pyruvate:ferredoxin oxidoreductase. *Curr. Opin. Struct. Biol.*, 9:663–669, 1999.
- [611] B. Charrier, C. Coronado, A. Kondorosi, and P. Ratet. Molecular characterization and expression of alfalfa (*Medicago sativa* L.) flavanone-3-hydroxylase and dihydroflavonol-4-reductase encoding genes. *Plant Mol. Biol.*, 29:773–786, 1995.
- [612] O. Chassande, S. Renard, P. Barbry, and M. Lazdunski. The human gene for diamine oxidase, an amiloride binding protein. Molecular cloning, sequencing, and characterization of the promoter. *J. Biol. Chem.*, 269:14484–14489, 1994.
- [613] I.B. Chatterjee, G.C. Chatterjee, N.C. Ghosh, and B.C. Guha. Identification of 2-keto-L-gulonolactone as an intermediate in the biosynthesis of L-ascorbic acid. *Naturwissenschaften*, 46:475–475, 1959.
- [614] L. Chatwell, T. Krojer, A. Fidler, W. Romisch, W. Eisenreich, A. Bacher, R. Huber, and M. Fischer. Biosynthesis of riboflavin: structure and properties of 2,5-diamino-6-ribosylamino-4(3*H*)-pyrimidinone 5'-phosphate reductase of *Methanocaldococcus jannaschii*. *J. Mol. Biol.*, 359:1334–1351, 2006.
- [615] M. Chau and R. Croteau. Molecular cloning and characterization of a cytochrome P450 taxoid 2 α -hydroxylase involved in taxol biosynthesis. *Arch. Biochem. Biophys.*, 427:48–57, 2004.
- [616] M. Chau, S. Jennewein, K. Walker, and R. Croteau. Taxol biosynthesis: Molecular cloning and characterization of a cytochrome P450 taxoid 7 β -hydroxylase. *Chem. Biol.*, 11:663–672, 2004.
- [617] S. Chaudhuri and J.R. Coggins. The purification of shikimate dehydrogenase from *Escherichia coli*. *Biochem. J.*, 226:217–223, 1985.
- [618] J. Chaudiere and A.L. Tappel. Purification and characterization of selenium-glutathione peroxidase from hamster liver. *Arch. Biochem. Biophys.*, 226:448–457, 1983.
- [619] S. Chauhan, P. Hosseinzadeh, Y. Lu, and N.J. Blackburn. Stopped-flow studies of the reduction of the copper centers suggest a bifurcated electron transfer pathway in peptidylglycine monooxygenase. *Biochemistry*, 55:2008–2021, 2016.
- [620] Y.S. Chauhan, V.S. Rathore, G.K. Garg, and A. Bhargava. Detection of an indole oxidizing system in maize leaves. *Biochem. Biophys. Res. Commun.*, 83:1237–1245, 1978.

- [621] M.L. Diaz Chavez, M. Rolf, A. Gesell, and T.M. Kutchan. Characterization of two methylenedioxy bridge-forming cytochrome P450-dependent enzymes of alkaloid formation in the Mexican prickly poppy *Argemone mexicana*. *Arch. Biochem. Biophys.*, 507:186–193, 2011.
- [622] F.S. Che, N. Watanabe, M. Iwano, H. Inokuchi, S. Takayama, S. Yoshida, and A. Isogai. Molecular characterization and subcellular localization of protoporphyrinogen oxidase in spinach chloroplasts. *Plant Physiol.*, 124:59–70, 2000.
- [623] S.G. Cheatum and J.C. Warren. Purification and properties of 3- β -hydroxysteroid dehydrogenase and Δ -5-3-ketosteroid isomerase from bovine corpora lutea. *Biochim. Biophys. Acta*, 122:1–13, 1966.
- [624] M.R. Cheesman, W.G. Zumft, and A.J. Thomson. The MCD and EPR of the heme centers of nitric oxide reductase from *Pseudomonas stutzeri*: evidence that the enzyme is structurally related to the heme-copper oxidases. *Biochemistry*, 37:3994–4000, 1998.
- [625] M. Hajj Chehade, L. Loiseau, M. Lombard, L. Pecqueur, A. Ismail, M. Smadja, B. Golinelli-Pimpaneau, C. Mellot-Draznieks, O. Hamelin, L. Aussel, S. Kieffer-Jaquinod, N. Labessan, F. Barras, M. Fontecave, and F. Pierrel. *ubiI*, a new gene in *Escherichia coli* coenzyme Q biosynthesis, is involved in aerobic C5-hydroxylation. *J. Biol. Chem.*, 288:20085–20092, 2013.
- [626] J. Chelieski, H.J. Wiggers, A.P. Citadini, A.J. da Costa Filho, M.C. Nonato, and C.A. Montanari. Kinetic mechanism and catalysis of *Trypanosoma cruzi* dihydroorotate dehydrogenase enzyme evaluated by isothermal titration calorimetry. *Anal. Biochem.*, 399:13–22, 2010.
- [627] C.N. Chen, L. Porubleva, G. Shearer, M. Svrakic, L.G. Holden, J.L. Dover, M. Johnston, P.R. Chitnis, and D.H. Kohl. Associating protein activities with their genes: rapid identification of a gene encoding a methylglyoxal reductase in the yeast *Saccharomyces cerevisiae*. *Yeast*, 20:545–554, 2003.
- [628] G. Chen, M.M. Kayser, M.D. Milhovilovic, M.E. Mrstik, C.A. Martinez, and J.D. Stewart. Asymmetric oxidations at sulfur catalyzed by engineered strains that overexpress cyclohexanone monooxygenase. *New J. Chem.*, 23:827–832, 1999.
- [629] H. Chen, Z.K. Attieh, T. Su, B.A. Syed, H. Gao, R.M. Alaeddine, T.C. Fox, J. Usta, C.E. Naylor, R.W. Evans, A.T. McKie, G.J. Anderson, and C.D. Vulpe. Hephaestin is a ferroxidase that maintains partial activity in sex-linked anemia mice. *Blood*, 103:3933–3939, 2004.
- [630] H. Chen, C.C. Tseng, B.K. Hubbard, and C.T. Walsh. Glycopeptide antibiotic biosynthesis: enzymatic assembly of the dedicated amino acid monomer (*S*)-3,5-dihydroxyphenylglycine. *Proc. Natl. Acad. Sci. USA*, 98:14901–14906, 2001.
- [631] J. Chen, M. Yoshinaga, L.D. Garbinski, and B.P. Rosen. Synergistic interaction of glyceraldehydes-3-phosphate dehydrogenase and ArsJ, a novel organoarsenical efflux permease, confers arsenate resistance. *Mol. Microbiol.*, 100:945–953, 2016.
- [632] J.J. Chen, Y. Lukyanenko, and J.C. Hutson. 25-Hydroxycholesterol is produced by testicular macrophages during the early postnatal period and influences differentiation of Leydig cells in vitro. *Biol. Reprod.*, 66:1336–1341, 2002.
- [633] L. Chen and J. Yang. Biochemical characterization of the tetrachlorobenzoquinone reductase involved in the biodegradation of pentachlorophenol. *Int. J. Mol. Sci.*, 9:198–212, 2008.
- [634] M. Chen, J.E. Markham, and E.B. Cahoon. Sphingolipid Δ^8 unsaturation is important for glucosylceramide biosynthesis and low-temperature performance in *Arabidopsis*. *Plant J.*, 69:769–781, 2012.
- [635] Q. Chen, C.H. Wang, S.K. Deng, Y.D. Wu, Y. Li, L. Yao, J.D. Jiang, X. Yan, J. He, and S.P. Li. Novel three-component Rieske non-heme iron oxygenase system catalyzing the *N*-dealkylation of chloroacetanilide herbicides in sphingomonads DC-6 and DC-2. *Appl. Environ. Microbiol.*, 80:5078–5085, 2014.
- [636] S. Chen, E. Glawischnig, K. Jørgensen, P. Naur, B. Jørgensen, C.E. Olsen, C.H. Hansen, H. Rasmussen, J.A. Pickett, and B.A. Halkier. CYP79F1 and CYP79F2 have distinct functions in the biosynthesis of aliphatic glucosinolates in *Arabidopsis*. *Plant J.*, 33:923–937, 2003.
- [637] X. Chen and P.J. Facchini. Short-chain dehydrogenase/reductase catalyzing the final step of noscapine biosynthesis is localized to laticifers in opium poppy. *Plant J.*, 77:173–184, 2014.

- [638] Y. Chen, N.A. Patel, A. Crombie, J.H. Scrivens, and J.C. Murrell. Bacterial flavin-containing monooxygenase is trimethylamine monooxygenase. *Proc. Natl. Acad. Sci. USA*, 108:17791–17796, 2011.
- [639] Y.F. Chen, H. Chao, and N.Y. Zhou. The catabolism of 2,4-xyleneol and *p*-cresol share the enzymes for the oxidation of *para*-methyl group in *Pseudomonas putida* NCIMB 9866. *Appl. Microbiol. Biotechnol.*, 98:1349–1356, 2014.
- [640] Z.W. Chen, M. Koh, G. Van Driessche, J.J. Van Beeumen, R.G. Bartsch, T.E. Meyer, M.A. Cusanovich, and F.S. Mathews. The structure of flavocytochrome *c* sulfide dehydrogenase from a purple phototrophic bacterium. *Science*, 266:430–432, 1994.
- [641] Z.W. Chen, K. Matsushita, T. Yamashita, T.A. Fujii, H. Toyama, O. Adachi, H.D. Bellamy, and F.S. Mathews. Structure at 1.9 Å resolution of a quinoxinoprotein alcohol dehydrogenase from *Pseudomonas putida* HK5. *Structure*, 10:837–849, 2002.
- [642] J.B. Cheng, D.L. Motola, D.J. Mangelsdorf, and D.W. Russell. De-orphanization of cytochrome P450 2R1: a microsomal vitamin D 25-hydroxylase. *J. Biol. Chem.*, 278:38084–38093, 2003.
- [643] J.Z. Cheng, C.M. Coyle, D.G. Panaccione, and S.E. O'Connor. A role for Old Yellow Enzyme in ergot alkaloid biosynthesis. *J. Am. Chem. Soc.*, 132:1776–1777, 2010.
- [644] Q. Cheng, H.T. Liu, P. Bombelli, A. Smith, and A.R. Slabas. Functional identification of AtFao3, a membrane bound long chain alcohol oxidase in *Arabidopsis thaliana*. *FEBS Lett.*, 574:62–68, 2004.
- [645] Q. Cheng, D. Sanglard, S. Vanhanen, H.T. Liu, P. Bombelli, A. Smith, and A.R. Slabas. *Candida* yeast long chain fatty alcohol oxidase is a c-type haemoprotein and plays an important role in long chain fatty acid metabolism. *Biochim. Biophys. Acta*, 1735:192–203, 2005.
- [646] Y.-J. Cheng and H.J. Karavolas. Properties and subcellular distribution of Δ^4 -steroid (progesterone) 5 α -reductase in rat anterior pituitary. *Steroids*, 26:57–71, 1975.
- [647] M.M. Cherney, Y. Zhang, M. Solomonson, J.H. Weiner, and M.N. James. Crystal structure of sulfide:quinone oxidoreductase from *Acidithiobacillus ferrooxidans*: insights into sulfidotrophic respiration and detoxification. *J. Mol. Biol.*, 398:292–305, 2010.
- [648] A.G. Chew and D.A. Bryant. Characterization of a plant-like protochlorophyllide *a* divinyl reductase in green sulfur bacteria. *J. Biol. Chem.*, 282:2967–2975, 2007.
- [649] C. Chiang and S.G. Knight. A new pathway of pentose metabolism. *Biochem. Biophys. Res. Commun.*, 3:554–559, 1960.
- [650] C. Chiang and S.G. Knight. L-Arabinose metabolism by cell-free extracts of *Penicillium chrysogenum*. *Biochim. Biophys. Acta*, 46:271–278, 1961.
- [651] K. Chiba, K. Kobayashi, K. Itoh, S. Itoh, T. Chiba, T. Ishizaki, and T. Kamataki. *N*-Oxygenation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine by the rat liver flavin-containing monooxygenase expressed in yeast cells. *Eur. J. Pharmacol.*, 293:97–100, 1995.
- [652] K. Chiba, E. Kubota, T. Miyakawa, Y. Kato, and T. Ishizaki. Characterization of hepatic microsomal metabolism as an *in vivo* detoxication pathway of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine in mice. *J. Pharmacol. Exp. Ther.*, 246:1108–1115, 1988.
- [653] N. Chim, A. Iniguez, T.Q. Nguyen, and C.W. Goulding. Unusual diheme conformation of the heme-degrading protein from *Mycobacterium tuberculosis*. *J. Mol. Biol.*, 395:595–608, 2010.
- [654] P. Chinnawirotpisan, G. Theeragool, S. Limtong, H. Toyama, O.O. Adachi, and K. Matsushita. Quinoxinoprotein alcohol dehydrogenase is involved in catabolic acetate production, while NAD-dependent alcohol dehydrogenase in ethanol assimilation in *Acetobacter pasteurianus* SKU1108. *J. Biosci. Bioeng.*, 96:564–571, 2003.
- [655] S. Chintalapati, J.S. Prakash, P. Gupta, S. Ohtani, I. Suzuki, T. Sakamoto, N. Murata, and S. Shivaji. A novel Δ^9 acyl-lipid desaturase, DesC₂, from cyanobacteria acts on fatty acids esterified to the *sn*-2 position of glycerolipids. *Biochem. J.*, 398:207–214, 2006.
- [656] C.B. Chiribau, M. Mihasan, P. Ganas, G.L. Igloi, V. Artenie, and R. Brandsch. Final steps in the catabolism of nicotine. *FEBS J.*, 273:1528–1536, 2006.

- [657] C.B. Chiribau, C. Sandu, M. Fraaije, E. Schiltz, and R. Brandsch. A novel γ -*N*-methylaminobutyrate demethylating oxidase involved in catabolism of the tobacco alkaloid nicotine by *Arthrobacter nicotinovorans* pAO1. *Eur. J. Biochem.*, 271:4677–4684, 2004.
- [658] L. Chistoserdova, L. Gomelsky, J.A. Vorholt, M. Gomelsky, Y.D. Tsygankov, and M.E. Lidstrom. Analysis of two formaldehyde oxidation pathways in *Methylobacillus flagellatus* KT, a ribulose monophosphate cycle methylotroph. *Microbiology*, 146:233–238, 2000.
- [659] P.R. Chitnis. Photosystem I: function and physiology. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 52:593–626, 2001.
- [660] H.J. Chiu, E. Johnson, I. Schroder, and D.C. Rees. Crystal structures of a novel ferric reductase from the hyperthermophilic archaeon *Archaeoglobus fulgidus* and its complex with NADP⁺. *Structure*, 9:311–319, 2001.
- [661] T. Chiyonobu, E. Shinagawa, O. Adachi, and M. Ameyama. Purification, crystallization and properties of 2-ketogluconate reductase from *Acetobacter rancens*. *Agric. Biol. Chem.*, 40:175–184, 1976.
- [662] L.J. Chlumsky, A.W. Sturgess, E. Nieves, and M.S. Jorns. Identification of the covalent flavin attachment site in sarcosine oxidase. *Biochemistry*, 37:2089–2095, 1998.
- [663] L.J. Chlumsky, L. Zhang, A.J. Ramsey, and M.S. Jorns. Preparation and properties of recombinant corynebacterial sarcosine oxidase: evidence for posttranslational modification during turnover with sarcosine. *Biochemistry*, 32:11132–11142, 1993.
- [664] H.P. Cho, M. Nakamura, and S.D. Clarke. Cloning, expression, and fatty acid regulation of the human Δ^5 desaturase. *J. Biol. Chem.*, 274:37335–37339, 1999.
- [665] H.P. Cho, M.T. Nakamura, and S.D. Clarke. Cloning, expression, and nutritional regulation of the mammalian Δ -6 desaturase. *J. Biol. Chem.*, 274:471–477, 1999.
- [666] M.H. Cho, S.G. Moinuddin, G.L. Helms, S. Hishiyama, D. Eichinger, L.B. Davin, and N.G. Lewis. (+)-Larreatricin hydroxylase, an enantio-specific polyphenol oxidase from the creosote bush (*Larrea tridentata*). *Proc. Natl. Acad. Sci. USA*, 100:10641–10646, 2003.
- [667] N.C. Cho, K. Kim, and D.Y. Jhon. Purification and characterization of polyol dehydrogenase from *Gluconobacter melanogenus*. *Han'guk Saenghwa Hakhaochi*, 23:172–178, 1990.
- [668] S.H. Cho, J.U. Na, H. Youn, C.S. Hwang, C.H. Lee, and S.O. Kang. Sepiapterin reductase producing L-threo-dihydrobiopterin from *Chlorobium tepidum*. *Biochem. J.*, 340:497–503, 1999.
- [669] S.W. Cho and J.G. Joshi. Characterization of glucose-6-phosphate dehydrogenase isozymes from human and pig brain. *Neuroscience*, 38:819–828, 1990.
- [670] S.W. Chocklett and P. Sobrado. *Aspergillus fumigatus* SidA is a highly specific ornithine hydroxylase with bound flavin cofactor. *Biochemistry*, 49:6777–6783, 2010.
- [671] S. Choe, S. Fujioka, T. Noguchi, S. Takatsuto, S. Yoshida, and K.A. Feldmann. Overexpression of DWARF4 in the brassinosteroid biosynthetic pathway results in increased vegetative growth and seed yield in *Arabidopsis*. *Plant J.*, 26:573–582, 2001.
- [672] J.-D. Choi, D.M. Bowers-Komro, M.D. Davis, D.E. Edmondson, and D.B. McCormick. Kinetic properties of pyridoxamine (pyridoxine)-5'-phosphate oxidase from rabbit liver. *J. Biol. Chem.*, 258:840–845, 1983.
- [673] S.K. Choi, S. Matsuda, T. Hoshino, X. Peng, and N. Misawa. Characterization of bacterial β -carotene 3,3'-hydroxylases, CrtZ, and P₄₅₀ in astaxanthin biosynthetic pathway and adonirubin production by gene combination in *Escherichia coli*. *Appl. Microbiol. Biotechnol.*, 72:1238–1246, 2006.
- [674] Y.H. Choi, H.J. Choi, D. Kim, K.N. Uhm, and H.K. Kim. Asymmetric synthesis of (*S*)-3-chloro-1-phenyl-1-propanol using *Saccharomyces cerevisiae* reductase with high enantioselectivity. *Appl. Microbiol. Biotechnol.*, 87:185–193, 2010.
- [675] Y.S. Choi, H. Zhang, J.S. Brunzelle, S.K. Nair, and H. Zhao. *In vitro* reconstitution and crystal structure of *p*-aminobenzoate *N*-oxygenase (AurF) involved in aureothin biosynthesis. *Proc. Natl. Acad. Sci. USA*, 105:6858–6863, 2008.

- [676] A. Chollet, L. Mourey, C. Lherbet, A. Delbot, S. Julien, M. Baltas, J. Bernadou, G. Pratviel, L. Maveyraud, and V. Bernardes-Genisson. Crystal structure of the enoyl-ACP reductase of *Mycobacterium tuberculosis* (InhA) in the apo-form and in complex with the active metabolite of isoniazid pre-formed by a biomimetic approach. *J. Struct. Biol.*, 190:328–337, 2015.
- [677] I.J. Chopra and G.N.C. Teco. Characteristics of inner ring (3 or 5) monodeiodination of 3,5-diiodothyronine in rat liver: evidence suggesting marked similarities of inner and outer ring deiodinases for iodothyronines. *Endocrinology*, 110:89–97, 1982.
- [678] O.W. Choroba, D.H. Williams, and J.B. Spencer. Biosynthesis of the vancomycin group of antibiotics: involvement of an unusual dioxygenase in the pathway to (S)-4-hydroxyphenylglycine. *J. Am. Chem. Soc.*, 122:5389–5390, 2000.
- [679] V. Choudhary, K. Wu, Z. Zhang, M. Dulchavsky, T. Barkman, J.C.A. Bardwell, and F. Stull. The enzyme pseudooxynicotine amine oxidase from *Pseudomonas putida* S16 is not an oxidase, but a dehydrogenase. *J. Biol. Chem.*, 298:102251–102251, 2022.
- [680] L.P. Chow, H. Iwadate, K. Yano, M. Kamo, A. Tsugita, L. Gardet-Salvi, A.L. Stritt-Etter, and P. Schurmann. Amino acid sequence of spinach ferredoxin:thioredoxin reductase catalytic subunit and identification of thiol groups constituting a redox-active disulfide and a [4Fe-4S] cluster. *Eur. J. Biochem.*, 231:149–156, 1995.
- [681] E.K. Chowdhury, K. Higuchi, S. Nagata, and H. Misono. A novel NADP⁺ dependent serine dehydrogenase from *Agrobacterium tumefaciens*. *Biosci. Biotechnol. Biochem.*, 61:152–157, 1997.
- [682] N.P. Chowdhury, J. Kahnt, and W. Buckel. Reduction of ferredoxin or oxygen by flavin-based electron bifurcation in *Megasphaera elsdenii*. *FEBS J.*, 282:3149–3160, 2015.
- [683] N.P. Chowdhury, A.M. Mowafy, J.K. Demmer, V. Upadhyay, S. Koelzer, E. Jayamani, J. Kahnt, M. Hornung, U. Demmer, U. Ermler, and W. Buckel. Studies on the mechanism of electron bifurcation catalyzed by electron transferring flavoprotein (Etf) and butyryl-CoA dehydrogenase (Bcd) of *Acidaminococcus fermentans*. *J. Biol. Chem.*, 289:5145–5157, 2014.
- [684] J. Christensen, K. Agger, P.A. Cloos, D. Pasini, S. Rose, L. Sennels, J. Rappsilber, K.H. Hansen, A.E. Salcini, and K. Helin. RBP2 belongs to a family of demethylases, specific for tri- and dimethylated lysine 4 on histone 3. *Cell*, 128:1063–1076, 2007.
- [685] J. Christopher, E. Pistorius, and B. Axelrod. Isolation of an enzyme of soybean lipoxidase. *Biochim. Biophys. Acta*, 198:12–19, 1970.
- [686] T. Chroumpi, M. Peng, M.V. Aguilar-Pontes, A. Muller, M. Wang, J. Yan, A. Lipzen, V. Ng, I.V. Grigoriev, M.R. Makela, and R.P. de Vries. Revisiting a ‘simple’ fungal metabolic pathway reveals redundancy, complexity and diversity. *Microb. Biotechnol.*, 14:2525–2537, 2021.
- [687] A. Chu, A. Dinkova, L.B. Davin, D.L. Bedgar, and N.G. Lewis. Stereospecificity of (+)-pinosresinol and (+)-lariciresinol reductases from *Forsythia intermedia*. *J. Biol. Chem.*, 268:27026–27033, 1993.
- [688] J.W. Chu and T. Kimura. Studies on adrenal steroid hydroxylases. Molecular and catalytic properties of adrenodoxin reductase (a flavoprotein). *J. Biol. Chem.*, 248:2089–2094, 1973.
- [689] E.E. Chufan, S.T. Prigge, X. Siebert, B.A. Eipper, R.E. Mains, and L.M. Amzel. Differential reactivity between two copper sites in peptidylglycine α -hydroxylating monooxygenase. *J. Am. Chem. Soc.*, 132:15565–15572, 2010.
- [690] A. Chugh, A. Ray, and J.B. Gupta. Squalene epoxidase as hypocholesterolemic drug target revisited. *Prog. Lipid Res.*, 42:37–50, 2003.
- [691] C.W. Chung and V.A. Najjar. Cofactor requirements for enzymatic denitrification. I. Nitrite reductase. *J. Biol. Chem.*, 218:617–625, 1956.
- [692] D.W. Chung, A. Pružinská, S. Hörtensteiner, and D.R. Ort. The role of pheophorbide *a* oxygenase expression and activity in the canola green seed problem. *Plant Physiol.*, 142:88–97, 2006.
- [693] R.M. Cicchillo, H. Zhang, J.A. Blodgett, J.T. Whitteck, G. Li, S.K. Nair, W.A. van der Donk, and W.W. Metcalf. An unusual carbon-carbon bond cleavage reaction during phosphinothricin biosynthesis. *Nature*, 459:871–874, 2009.

- [694] J. Claesen and M. Bibb. Genome mining and genetic analysis of cypemycin biosynthesis reveal an unusual class of posttranslationally modified peptides. *Proc. Natl. Acad. Sci. USA*, 107:16297–16302, 2010.
- [695] D.D. Clark, J.R. Allen, and S.A. Ensign. Characterization of five catalytic activities associated with the NADPH:2-ketopropyl-coenzyme M [2-(2-ketopropylthio)ethanesulfonate] oxidoreductase/carboxylase of the *Xanthobacter* strain Py2 epoxide carboxylase system. *Biochemistry*, 39:1294–1304, 2000.
- [696] J.E. Clark and L.G. Ljungdahl. Purification and properties of 5,10-methylenetetrahydrofolate reductase, an iron-sulfur flavoprotein from *Clostridium formicoaceticum*. *J. Biol. Chem.*, 259:10845–10849, 1984.
- [697] M.A. Clark and E.L. Barrett. The pbs gene and hydrogen sulfide production by *Salmonella typhimurium*. *J. Bacteriol.*, 169:2391–2397, 1987.
- [698] M.F. Clarke-Pearson and S.F. Brady. Paerucumarin, a new metabolite produced by the pvc gene cluster from *Pseudomonas aeruginosa*. *J. Bacteriol.*, 190:6927–6930, 2008.
- [699] M. Clausen, R.M. Kannangara, C.E. Olsen, C.K. Blomstedt, R.M. Gleadow, K. Jørgensen, S. Bak, M.S. Motawie, and B.L. Møller. The bifurcation of the cyanogenic glucoside and glucosinolate biosynthetic pathways. *Plant J.*, 84:558–573, 2015.
- [700] D. Clausnitzer, W. Piepersberg, and U.F. Wehmeier. The oxidoreductases LivQ and NeoQ are responsible for the different 6'-modifications in the aminoglycosides lividomycin and neomycin. *J. Appl. Microbiol.*, 111:642–651, 2011.
- [701] A.M. Cleton-Jansen, N. Goosen, T.J. Wenzel, and P. van de Putte. Cloning of the gene encoding quinoprotein glucose dehydrogenase from *Acinetobacter calcoaceticus*: evidence for the presence of a second enzyme. *J. Bacteriol.*, 170:2121–2125, 1988.
- [702] I.J. Clifton, L.X. Doan, M.C. Sleeman, M. Topf, H. Suzuki, R.C. Wilmoth, and C.J. Schofield. Crystal structure of carbapenem synthase (CarC). *J. Biol. Chem.*, 278:20843–20850, 2003.
- [703] I.J. Clifton, L.C. Hsueh, J.E. Baldwin, K. Harlos, and C.J. Schofield. Structure of proline 3-hydroxylase. Evolution of the family of 2-oxoglutarate dependent oxygenases. *Eur. J. Biochem.*, 268:6625–6636, 2001.
- [704] A.L. Cline and A.S.L. Hu. Enzymatic characterization and comparison of three sugar dehydrogenases from a pseudomonad. *J. Biol. Chem.*, 240:4493–4497, 1965.
- [705] A.L. Cline and A.S.L. Hu. Some physical properties of three sugar dehydrogenases from a pseudomonad. *J. Biol. Chem.*, 240:4498–4502, 1965.
- [706] A.L. Cline and A.S.L. Hu. The isolation of three sugar dehydrogenases from a pseudomonad. *J. Biol. Chem.*, 240:4488–4492, 1965.
- [707] P.A. Cloos, J. Christensen, K. Agger, A. Maiolica, J. Rappsilber, T. Antal, K.H. Hansen, and K. Helin. The putative oncogene GASC1 demethylates tri- and dimethylated lysine 9 on histone H3. *Nature*, 442:307–311, 2006.
- [708] J.D. Colandene and R.H. Garrett. Functional dissection and site-directed mutagenesis of the structural gene for NAD(P)H-nitrite reductase in *Neurospora crassa*. *J. Biol. Chem.*, 271:24096–24104, 1996.
- [709] J. Colby and L.J. Zatman. The purification and properties of a bacterial trimethylamine dehydrogenase. *Biochem. J.*, 121:9P–10P, 1971.
- [710] J. Stirling Colby, Dalton D.I., and H. The soluble methane mono-oxygenase of *Methylococcus capsulatus* (Bath). Its ability to oxygenate *n*-alkanes, *n*-alkenes, ethers, and alicyclic, aromatic and heterocyclic compounds. *Biochem. J.*, 165:395–402, 1977.
- [711] L. De Colibus, M. Li, C. Binda, A. Lustig, D.E. Edmondson, and A. Mattevi. Three-dimensional structure of human monoamine oxidase A (MAO A): relation to the structures of rat MAO A and human MAO B. *Proc. Natl. Acad. Sci. USA*, 102:12684–12689, 2005.
- [712] J.F. Collet and J.C. Bardwell. Oxidative protein folding in bacteria. *Mol. Microbiol.*, 44:1–8, 2002.

- [713] H.F. Collins, R. Biedendieck, H.K. Leech, M. Gray, J.C. Escalante-Semerena, K.J. McLean, A.W. Munro, S.E. Rigby, M.J. Warren, and A.D. Lawrence. *Bacillus megaterium* has both a functional BluB protein required for DMB synthesis and a related flavoprotein that forms a stable radical species. *PLoS One*, 8:e55708–e55708, 2013.
- [714] N. Colloc'h, M. el Hajji, B. Bachet, G. L'Hermite, M. Schiltz, T. Prange, B. Castro, and J.-P. Mornon. Crystal structure of the protein drug urate oxidase-inhibitor complex at 2.05 Å resolution. *Nat. Struct. Biol.*, 4:947–952, 1997.
- [715] G. Collu, N. Unver, A.M. Peltenburg-Looman, R. van der Heijden, R. Verpoorte, and J. Memelink. Geraniol 10-hydroxylase, a cytochrome P450 enzyme involved in terpenoid indole alkaloid biosynthesis. *FEBS Lett.*, 508:215–220, 2001.
- [716] S.U. Colmenares, J.M. Swenson, S.A. Langley, C. Kennedy, S.V. Costes, and G.H. Karpen. *Drosophila* histone demethylase KDM4A has enzymatic and non-enzymatic roles in controlling heterochromatin integrity. *Dev Cell*, 42:156–169.e5, 2017.
- [717] A. Saqib Colocousi, Leak K.M., and D.J. Mutants of *Pseudomonas fluorescens* NCIMB 11671 defective in the catabolism of α -pinene. *Appl. Microbiol. Biotechnol.*, 45:822–830, 1996.
- [718] S. Colonna, N. Gaggero, G. Carrea, G. Ottolina, P. Pasta, and F. Zambianchi. First asymmetric epoxidation catalysed by cyclohexanone monooxygenase. *Tetrahedron Lett.*, 43:1797–1799, 2002.
- [719] G. Condemine, N. Hugouvieux-Cotte-Pattat, and J. Robert-Baudouy. An enzyme in the pectolytic pathway of *Erwinia chrysanthemi*: 3-keto-3-deoxygluconate oxidoreductase. *J. Gen. Microbiol.*, 130:2839–2844, 1984.
- [720] E.E. Conn, L.M. Kraemer, P.N. Liu, and B. Vennesland. The aerobic oxidation of reduced triphosphopyridine nucleotide by a wheat germ enzyme system. *J. Biol. Chem.*, 194:143–151, 1952.
- [721] J.L. Connelly, D.J. Danner, and J.A. Bowden. Branched chain α -keto acid metabolism. I. Isolation, purification, and partial characterization of bovine liver α -ketoisocaproic: α -keto- β -methylvaleric acid dehydrogenase. *J. Biol. Chem.*, 243:1198–1203, 1968.
- [722] K.L. Connor, K.L. Colabroy, and B. Gerratana. A heme peroxidase with a functional role as an L-tyrosine hydroxylase in the biosynthesis of anthramycin. *Biochemistry*, 50:8926–8936, 2011.
- [723] H.E. Conrad, R. DuBus, M.J. Namtvedt, and I.C. Gunsalus. Mixed function oxidation. II. Separation and properties of the enzymes catalyzing camphor lactonization. *J. Biol. Chem.*, 240:495–503, 1965.
- [724] R.S. Conrad, L.K. Massey, and J.R. Sokatch. D- and L-isoleucine metabolism and regulation of their pathways in *Pseudomonas putida*. *J. Bacteriol.*, 118:103–111, 1974.
- [725] R.B. Cooley, B.L. Dubbels, L.A. Sayavedra-Soto, P.J. Bottomley, and D.J. Arp. Kinetic characterization of the soluble butane monooxygenase from *Thaueria butanivorans*, formerly '*Pseudomonas butanovora*'. *Microbiology*, 155:2086–2096, 2009.
- [726] M.J. Coon, F.P. Kupiecki, E.E. Dekker, M.J. Schlesinger, and A. del Campillo. The enzymic synthesis of branched-chain acids. In G.E.W. Wolstenholme and M. O'Connor, editors, *CIBA Symposium on the Biosynthesis of Terpenes and Sterols*, pages 62–74. CIBA Symposium on the Biosynthesis of Terpenes and Sterols, London, 1959.
- [727] L.E. Cooper, D. Fedoseyenko, S.H. Abdelwahed, S.H. Kim, T. Dairi, and T.P. Begley. *In vitro* reconstitution of the radical S-adenosylmethionine enzyme MqnC involved in the biosynthesis of futasoline-derived menaquinone. *Biochemistry*, 52:4592–4594, 2013.
- [728] R.A. Cooper. The pathway for L-galactonate catabolism in *Escherichia coli* K-12. *FEBS Lett.*, 103:216–220, 1979.
- [729] R.A. Cooper. The pathway for L-gulonate catabolism in *Escherichia coli* K-12 and *Salmonella typhimurium* LT-2. *FEBS Lett.*, 115:63–67, 1980.
- [730] R.A. Cooper and M.A. Skinner. Catabolism of 3- and 4-hydroxyphenylacetate by the 3,4-dihydroxyphenylacetate pathway in *Escherichia coli*. *J. Bacteriol.*, 143:302–306, 1980.
- [731] S.K. Cooper, J. Pandhare, S.P. Donald, and J.M. Phang. A novel function for hydroxyproline oxidase in apoptosis through generation of reactive oxygen species. *J. Biol. Chem.*, 283:10485–10492, 2008.

- [732] J.G. Coote and H. Hassall. The role of imidazol-5-yl-lactate-nicotinamide-adenine dinucleotide phosphate oxidoreductase and histidine-2-oxoglutarate aminotransferase in the degradation of imidazol-5-yl-lactate by *Pseudomonas acidovorans*. *Biochem. J.*, 111:237–239, 1969.
- [733] J.J. Coque, F.J. Enguita, R.E. Cardoza, J.F. Martin, and P. Liras. Characterization of the *cefF* gene of *Nocardia lactamdurans* encoding a 3'-methylcephem hydroxylase different from the 7-cephem hydroxylase. *Appl. Microbiol. Biotechnol.*, 44:605–609, 1996.
- [734] A.L. Corder, B.P. Subedi, S. Zhang, A.M. Dark, F.W. Foss, Pierce Jr., and B.S. Peroxide-shunt substrate-specificity for the *Salmonella typhimurium* O₂-dependent tRNA modifying monooxygenase (MiaE). *Biochemistry*, 52:6182–6196, 2013.
- [735] E.J. Corey, W.E. Russey, and P.R. Ortiz de Montellano. 2,3-Oxidosqualene, an intermediate in the biological synthesis of sterols from squalene. *J. Am. Chem. Soc.*, 88:4750–4751, 1966.
- [736] G.T. Cori, M.W. Slein, and C.F. Cori. Crystalline D-glyceraldehyde-3-phosphate dehydrogenase from rabbit muscle. *J. Biol. Chem.*, 173:605–618, 1948.
- [737] M.J. Cormier, J.M. Crane, Nakano Jr., and Y. Evidence for the identity of the luminescent systems of *Porichthys porosissimus* (fish) and *Cypridina hilgendorffii* (crustacean). *Biochem. Biophys. Res. Commun.*, 29:747–752, 1967.
- [738] M.J. Cormier, K. Hori, and J.M. Anderson. Bioluminescence in coelenterates. *Biochim. Biophys. Acta*, 346:137–164, 1974.
- [739] A.V. Corrigall, K.B. Siziba, M.H. Maneli, E.G. Shephard, M. Ziman, T.A. Dailey, H.A. Dailey, R.E. Kirsch, and P.N. Meissner. Purification of and kinetic studies on a cloned protoporphyrinogen oxidase from the aerobic bacterium *Bacillus subtilis*. *Arch. Biochem. Biophys.*, 358:251–256, 1998.
- [740] P.L.A.M. Corstjens, J.P.M. de Vrind, T. Goosen, and E.W. de Vrind-de Jong. Identification and molecular analysis of the *Leptothrix discophora* SS-1 *mofA* gene, a gene putatively encoding a manganese-oxidizing protein with copper domains. *Geomicrobiol. J.*, 14:91–108, 1997.
- [741] E. Della Corte and F. Stirpe. The regulation of rat liver xanthine oxidase. Involvement of thiol groups in the conversion of the enzyme activity from dehydrogenase (type D) into oxidase (type O) and purification of the enzyme. *Biochem. J.*, 126:739–745, 1972.
- [742] L. Cortes, A.G. Wedd, and Z. Xiao. The functional roles of the three copper sites associated with the methionine-rich insert in the multicopper oxidase CueO from *E. coli*. *Metallomics*, 7:776–785, 2015.
- [743] R. Cortese, J. Brevet, J. Hedegaard, and J. Roche. [Identification and purification of an α -ketoacid aromatic reductase of *Escherichia coli* B]. *C.R. Seances Soc. Biol. Fil.*, 162:390–395, 1968.
- [744] M.S. Cosgrove, C. Naylor, S. Paludan, M.J. Adams, and H.R. Levy. On the mechanism of the reaction catalyzed by glucose 6-phosphate dehydrogenase. *Biochemistry*, 37:2759–2767, 1998.
- [745] K.C. Costa, T.J. Lie, Q. Xia, and J.A. Leigh. VhuD facilitates electron flow from H₂ or formate to heterodisulfide reductase in *Methanococcus marisaludis*. *J. Bacteriol.*, 195:5160–5165, 2013.
- [746] K.C. Costa, P.M. Wong, T. Wang, T.J. Lie, J.A. Dodsworth, I. Swanson, J.A. Burn, M. Hackett, and J.A. Leigh. Protein complexing in a methanogen suggests electron bifurcation and electron delivery from formate to heterodisulfide reductase. *Proc. Natl. Acad. Sci. USA*, 107:11050–11055, 2010.
- [747] A.M.G. Costas, A.K. White, and W.W. Metcalf. Purification and characterization of a novel phosphorus-oxidizing enzyme from *Pseudomonas stutzeri* WM88. *J. Biol. Chem.*, 276:17429–17436, 2001.
- [748] M.-R. Coudray, G. Canebascini, and H. Meier. Characterization of a cellobiose dehydrogenase in the cellulolytic fungus *Protrichum* (*Chrysosporium*) thermophile. *Biochem. J.*, 203:277–284, 1982.
- [749] M.M. Couladis, J.B. Friesen, M.E. Landgrebe, and E. Leete. Enzymes catalysing the reduction of tropinone to tropine and ψ -tropine isolated from the roots of *Datura innoxia*. *Pytochemistry*, 30:801–805, 1991.

- [750] C.E. Coulthard, R. Michaelis, W.F. Short, G. Sykes, G.E.H. Skrimshire, A.F.B. Standfast, J.H. Birkinshaw, and H. Raistick. Notatin: an anti-bacterial glucose-aerodehydrogenase from *Penicillium notatum* Westling and *Penicillium resticulosum* sp. nov. *Biochem. J.*, 39:24–36, 1945.
- [751] J.W. Coulton and M. Kapoor. Purification and some properties of the glutamate dehydrogenase of *Salmonella typhimurium*. *Can. J. Microbiol.*, 19:427–438, 1973.
- [752] G. Courtade, R. Wimmer, A.K. Rohr, M. Preims, A.K. Felice, M. Dimarogona, G. Vaaje-Kolstad, M. Sorlie, M. Sandgren, R. Ludwig, V.G. Eijsink, and F.L. Aachmann. Interactions of a fungal lytic polysaccharide monooxygenase with β -glucan substrates and cellobiose dehydrogenase. *Proc. Natl. Acad. Sci. USA*, 113:5922–5927, 2016.
- [753] J.F. Couture, E. Collazo, P.A. Ortiz-Tello, J.S. Brunzelle, and R.C. Trievel. Specificity and mechanism of JMJD2A, a trimethyllysine-specific histone demethylase. *Nat. Struct. Mol. Biol.*, 14:689–695, 2007.
- [754] J. Couturier, P. Prosper, A.M. Winger, A. Hecker, M. Hirasawa, D.B. Knaff, P. Gans, J.P. Jacquot, A. Navaza, A. Haouz, and N. Rouhier. In the absence of thioredoxins, what are the reductants for peroxiredoxins in *Thermotoga maritima*. *Antioxid Redox Signal*, 18:1613–1622, 2013.
- [755] M.L. Coval and A. Taurog. Purification and iodinating activity of hog thyroid peroxidase. *J. Biol. Chem.*, 242:5510–5523, 1967.
- [756] J. Coves, M. Zeghouf, D. Macherel, B. Guigliarelli, M. Asso, and M. Fontecave. Flavin mononucleotide-binding domain of the flavoprotein component of the sulfite reductase from *Escherichia coli*. *Biochemistry*, 36:5921–5928, 1997.
- [757] J.M. Cox, D.J. Day, and C. Anthony. The interaction of methanol dehydrogenase and its electron acceptor, cytochrome c_1 in methylotrophic bacteria. *Biochim. Biophys. Acta*, 1119:97–106, 1992.
- [758] C.L. Coyle, W.G. Zumft, P.M.H. Kroneck, H. Körner, and W. Jakob. Nitrous oxide reductase from denitrifying *Pseudomonas perfectomarina*. Purification and properties of a novel multicopper enzyme. *Eur. J. Biochem.*, 153:459–467, 1985.
- [759] C.M. Coyle, J.Z. Cheng, S.E. O’Connor, and D.G. Panaccione. An old yellow enzyme gene controls the branch point between *Aspergillus fumigatus* and *Claviceps purpurea* ergot alkaloid pathways. *Appl. Environ. Microbiol.*, 76:3898–3903, 2010.
- [760] G.E. Cozier and C. Anthony. Structure of the quinoprotein glucose dehydrogenase of *Escherichia coli* modelled on that of methanol dehydrogenase from *Methylobacterium extorquens*. *Biochem. J.*, 312:679–685, 1995.
- [761] G.E. Cozier, I.G. Giles, and C. Anthony. The structure of the quinoprotein alcohol dehydrogenase of *Acetobacter aceti* modelled on that of methanol dehydrogenase from *Methylobacterium extorquens*. *Biochem. J.*, 308:375–379, 1995.
- [762] G.E. Cozier, R.A. Salleh, and C. Anthony. Characterization of the membrane quinoprotein glucose dehydrogenase from *Escherichia coli* and characterization of a site-directed mutant in which histidine-262 has been changed to tyrosine. *Biochem. J.*, 340:639–647, 1999.
- [763] M.J. Crabbe, R.D. Waight, W.G. Bardsley, R.W. Barker, I.D. Kelly, and P.F. Knowles. Human placental diamine oxidase. Improved purification and characterization of a copper- and manganese-containing amine oxidase with novel substrate specificity. *Biochem. J.*, 155:679–687, 1976.
- [764] R. Cramm, A. Pohlmann, and B. Friedrich. Purification and characterization of the single-component nitric oxide reductase from *Ralstonia eutropha* H16. *FEBS Lett.*, 460:6–10, 1999.
- [765] D.I. Crandall and D.N. Halikis. Homogentisic acid oxidase. I. Distribution in animal tissues and relation to tyrosine metabolism in rat kidney. *J. Biol. Chem.*, 208:629–638, 1954.
- [766] B.R. Crane, L.M. Siegel, and E.D. Getzoff. Structures of the siroheme- and Fe₄S₄-containing active center of sulfite reductase in different states of oxidation: heme activation via reduction-gated exogenous ligand exchange. *Biochemistry*, 36:12101–12119, 1997.
- [767] F.L. Crane, J.G. Hauge, and H. Beinert. Flavoproteins involved in the first oxidative step of the fatty acid cycle. *Biochim. Biophys. Acta*, 17:292–294, 1955.

- [768] F.L. Crane, S. Mii, J.G. Hauge, D.E. Green, and H. Beinert. On the mechanism of dehydrogenation of fatty acyl derivatives of coenzyme A. I. The general fatty acyl coenzyme A dehydrogenase. *J. Biol. Chem.*, 218:701–716, 1956.
- [769] D.L. Crawford, J.B. Sutherland, A.L. Pometto, Miller III, and J.M. Production of an aromatic aldehyde oxidase by *Streptomyces viridosporus*. *Arch. Microbiol.*, 131:351–355, 1982.
- [770] J.M. Crawford, C. Portmann, X. Zhang, M.B. Roeffaers, and J. Clardy. Small molecule perimeter defense in entomopathogenic bacteria. *Proc. Natl. Acad. Sci. USA*, 109:10821–10826, 2012.
- [771] E.M. Crook. The system dehydroascorbic acid-glutathione. *Biochem. J.*, 35:226–236, 1941.
- [772] R. Croteau and N.M. Felton. Substrate specificity of monoterpenol dehydrogenases from *Foeniculum vulgare* and *Tanacetum vulgare*. *Phytochemistry*, 19:1343–1347, 1980.
- [773] R. Croteau, C.L. Hooper, and M. Felton. Biosynthesis of monoterpenes. Partial purification and characterization of a bicyclic monoterpenol dehydrogenase from sage (*Salvia officinalis*). *Arch. Biochem. Biophys.*, 188:182–193, 1978.
- [774] R. Croteau and K.V. Venkatachalam. Metabolism of monoterpenes: demonstration that (+)-*cis*-isopulegone, not piperitenone, is the key intermediate in the conversion of (-)-isopiperitenone to (+)-pulegone in peppermint (*Mentha piperita*). *Arch. Biochem. Biophys.*, 249:306–315, 1986.
- [775] D.N. Crowell, D.H. Huizinga, A.K. Deem, C. Trobaugh, R. Denton, and S.E. Sen. *Arabidopsis thaliana* plants possess a specific farnesylcysteine lyase that is involved in detoxification and recycling of farnesylcysteine. *Plant J.*, 50:839–847, 2007.
- [776] J.K. Crowell, S. Sardar, M.S. Hossain, F.W. Foss, Pierce Jr., and B.S. Non-chemical proton-dependent steps prior to O₂-activation limit *Azotobacter vinelandii* 3-mercaptopropionic acid dioxygenase (MDO) catalysis. *Arch. Biochem. Biophys.*, 604:86–94, 2016.
- [777] M.J. Cryle. Selectivity in a barren landscape: the P450(BioI)-ACP complex. *Biochem. Soc. Trans.*, 38:934–939, 2010.
- [778] M.J. Cryle, S.G. Bell, and I. Schlichting. Structural and biochemical characterization of the cytochrome P450 CypX (CYP134A1) from *Bacillus subtilis*: a cyclo-L-leucyl-L-leucyl dipeptide oxidase. *Biochemistry*, 49:7282–7296, 2010.
- [779] M.J. Cryle and I. Schlichting. Structural insights from a P450 Carrier Protein complex reveal how specificity is achieved in the P450(BioI) ACP complex. *Proc. Natl. Acad. Sci. USA*, 105:15696–15701, 2008.
- [780] M.J. Cryle and J.J. De Voss. Carbon-carbon bond cleavage by cytochrome *p*₄₅₀(BioI)(CYP107H1). *Chem. Commun. (Camb.)*, pages 86–87, 2004.
- [781] A. Cultrone, C. Scazzocchio, M. Rochet, G. Montero-Moran, C. Drevet, and R. Fernandez-Martin. Convergent evolution of hydroxylation mechanisms in the fungal kingdom: molybdenum cofactor-independent hydroxylation of xanthine via α -ketoglutarate-dependent dioxygenases. *Mol. Microbiol.*, 57:276–290, 2005.
- [782] R. Cunin, N. Glansdorff, A. Pierard, and V. Stalon. Biosynthesis and metabolism of arginine in bacteria. *Microbiol. Rev.*, 50:314–352, 1986.
- [783] R. Cunin, N. Glansdorff, A. Pierard, and V. Stalon. Erratum report: Biosynthesis and metabolism of arginine in bacteria. *Microbiol. Rev.*, 51:178–178, 1987.
- [784] B.A. Cunningham and S. Kirkwood. Enzyme systems concerned with the synthesis of monoiodotyrosine. III. Ion requirements of the soluble system. *J. Biol. Chem.*, 236:485–489, 1961.
- [785] C.C. Cunningham and L.P. Hager. Reactivation of the lipid-depleted pyruvate oxidase system from *Escherichia coli* with cell envelope neutral lipids. *J. Biol. Chem.*, 250:7139–7146, 1975.
- [786] M.L. Cunningham and A.H. Fairlamb. Trypanothione reductase from *Leishmania donovani*. Purification, characterisation and inhibition by trivalent antimonials. *Eur. J. Biochem.*, 230:460–468, 1995.
- [787] O. Cunningham, M.G. Gore, and T.J. Mantle. Initial-rate kinetics of the flavin reductase reaction catalysed by human biliverdin-IX β reductase (BVR-B). *Biochem. J.*, 345:393–399, 2000.
- [788] J.R. Cupp-Vickery, H. Li, and T.L. Poulos. Preliminary crystallographic analysis of an enzyme involved in erythromycin biosynthesis: cytochrome P450*eryF*. *Proteins*, 20:197–201, 1994.

- [789] E. Cusa, N. Obradors, L. Baldoma, J. Badia, and J. Aguilar. Genetic analysis of a chromosomal region containing genes required for assimilation of allantoin nitrogen and linked glyoxylate metabolism in *Escherichia coli*. *J. Bacteriol.*, 181:7479–7484, 1999.
- [790] S.M. Cuskey, V. Peccoraro, and R.H. Olsen. Initial catabolism of aromatic biogenic amines by *Pseudomonas aeruginosa* PAO: pathway description, mapping of mutations, and cloning of essential genes. *J. Bacteriol.*, 169:2398–2404, 1987.
- [791] J.R. Cussiol, T.G. Alegria, L.I. Szweda, and L.E. Netto. Ohr (organic hydroperoxide resistance protein) possesses a previously undescribed activity, lipoyl-dependent peroxidase. *J. Biol. Chem.*, 285:21943–21950, 2010.
- [792] A.J. Cutler, T.M. Squires, M.K. Loewen, and J.J. Balsevich. Induction of (+)-abscisic acid 8' hydroxylase by (+)-abscisic acid in cultured maize cells. *J. Exp. Bot.*, 48:1787–1795, 1997.
- [793] C.M. Czekster and J.S. Blanchard. One substrate, five products: reactions catalyzed by the dihydroneopterin aldolase from *Mycobacterium tuberculosis*. *J. Am. Chem. Soc.*, 134:19758–19771, 2012.
- [794] M. Czjzek, J.P. Dos Santos, J. Pommier, G. Giordano, V. Méjean, and R. Haser. Crystal structure of oxidized trimethylamine *N*-oxide reductase from *Shewanella massilia* at 2.5 Å resolution. *J. Mol. Biol.*, 284:435–447, 1998.
- [795] J. Shim da, N.S. Nemeria, A. Balakrishnan, H. Patel, J. Song, J. Wang, F. Jordan, and E.T. Farinas. Assignment of function to histidines 260 and 298 by engineering the E1 component of the *Escherichia coli* 2-oxoglutarate dehydrogenase complex; substitutions that lead to acceptance of substrates lacking the 5-carboxyl group. *Biochemistry*, 50:7705–7709, 2011.
- [796] A. Lobo da Cunha, D. Amaral de Carvalho, E. Oliveira, A. Alves, V. Costa, and G. Calado. Mannitol oxidase and polyol dehydrogenases in the digestive gland of gastropods: Correlations with phylogeny and diet. *PLoS One*, 13:e0193078–e0193078, 2018.
- [797] D.V. Dabir, E.P. Leverich, S.K. Kim, F.D. Tsai, M. Hirasawa, D.B. Knaff, and C.M. Koehler. A role for cytochrome *c* and cytochrome *c* peroxidase in electron shuttling from Erv1. *EMBO J.*, 26:4801–4811, 2007.
- [798] S. Dagle, P.J. Chapman, and D.T. Gibson. The metabolism of β -phenylpropionic acid by an *Achromobacter*. *Biochem. J.*, 97:643–650, 1965.
- [799] C. Dahl, B. Franz, D. Hensen, A. Kesselheim, and R. Zigann. Sulfite oxidation in the purple sulfur bacterium *Allochro-matium vinosum*: identification of SoeABC as a major player and relevance of SoxYZ in the process. *Microbiology*, 159:2626–2638, 2013.
- [800] H. Dahlback and I. Holmberg. Oxidation of 5 β -cholestane-3 α ,7 α ,12 α -triol into 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanoic acid by cytochrome *P*-450₂₆ from rabbit liver mitochondria. *Biochem. Biophys. Res. Commun.*, 167:391–395, 1990.
- [801] K. Dahm and H. Breuer. Anreicherung einer 17 β -hydroxysteroid:NAD(P)-oxydoreduktase aus der Nebenniere der Ratte. *Hoppe-Seyler's Z. Physiol. Chem.*, 336:63–68, 1964.
- [802] V.D. Dai, K. Decker, and H. Sund. Purification and properties of L-6-hydroxynicotine oxidase. *Eur. J. Biochem.*, 4:95–102, 1968.
- [803] Y. Dai, T.C. Pochapsky, and R.H. Abeles. Mechanistic studies of two dioxygenases in the methionine salvage pathway of *Klebsiella pneumoniae*. *Biochemistry*, 40:6379–6387, 2001.
- [804] Y. Dai, P.C. Wensink, and R.H. Abeles. One protein, two enzymes. *J. Biol. Chem.*, 274:1193–1195, 1999.
- [805] H.A. Dailey and T.A. Dailey. Protoporphyrinogen oxidase of *Myxococcus xanthus*. Expression, purification, and characterization of the cloned enzyme. *J. Biol. Chem.*, 271:8714–8718, 1996.
- [806] H.A. Dailey, S. Gerdes, T.A. Dailey, J.S. Burch, and J.D. Phillips. Noncanonical coproporphyrin-dependent bacterial heme biosynthesis pathway that does not use protoporphyrin. *Proc. Natl. Acad. Sci. USA*, 112:2210–2215, 2015.
- [807] T.A. Dailey, T.O. Boynton, A.N. Albetel, S. Gerdes, M.K. Johnson, and H.A. Dailey. Discovery and characterization of HemQ: an essential heme biosynthetic pathway component. *J. Biol. Chem.*, 285:25978–25986, 2010.
- [808] T.A. Dailey and H.A. Dailey. Human protoporphyrinogen oxidase: expression, purification, and characterization of the cloned enzyme. *Protein Sci.*, 5:98–105, 1996.

- [809] T.A. Dailey and H.A. Dailey. Identification of an FAD superfamily containing protoporphyrinogen oxidases, monoamine oxidases, and phytoene desaturase. Expression and characterization of phytoene desaturase of *Myxococcus xanthus*. *J. Biol. Chem.*, 273:13658–13662, 1998.
- [810] T. Daimon, T. Kozaki, R. Niwa, I. Kobayashi, K. Furuta, T. Namiki, K. Uchino, Y. Banno, S. Katsuma, T. Tamura, K. Mita, H. Sezutsu, M. Nakayama, K. Itoyama, T. Shimada, and T. Shinoda. Precocious metamorphosis in the juvenile hormone-deficient mutant of the silkworm, *Bombyx mori*. *PLoS Genet.*, 8:e1002486–e1002486, 2012.
- [811] T. Daimon and T. Shinoda. Function, diversity, and application of insect juvenile hormone epoxidases (CYP15). *Biotechnol. Appl. Biochem.*, 60:82–91, 2013.
- [812] T. Dairi and Y. Asano. Cloning, nucleotide sequencing, and expression of an opine dehydrogenase gene from *Arthrobacter sp.* strain 1C. *Appl. Environ. Microbiol.*, 61:3169–3171, 1995.
- [813] T. Dairi, T. Nakano, K. Aisaka, R. Katsumata, and M. Hasegawa. Cloning and nucleotide sequence of the gene responsible for chlorination of tetracycline. *Biosci. Biotechnol. Biochem.*, 59:1099–1106, 1995.
- [814] T.R. Dambe, A.M. Kühn, T. Brossette, F. Giffhorn, and A.J. Scheidig. Crystal structure of NADP(H)-dependent 1,5-anhydro-D-fructose reductase from *Sinorhizobium morelense* at 2.2 Å resolution: construction of a NADH-accepting mutant and its application in rare sugar synthesis. *Biochemistry*, 45:10030–10042, 2006.
- [815] T. Dammeyer, S.C. Bagby, M.B. Sullivan, S.W. Chisholm, and N. Frankenberg-Dinkel. Efficient phage-mediated pigment biosynthesis in oceanic cyanobacteria. *Curr. Biol.*, 18:442–448, 2008.
- [816] T. Dammeyer and N. Frankenberg-Dinkel. Function and distribution of bilin biosynthesis enzymes in photosynthetic organisms. *Photochem Photobiol Sci*, 7:1121–1130, 2008.
- [817] I. Dance. The mechanism of nitrogenase. Computed details of the site and geometry of binding of alkyne and alkene substrates and intermediates. *J. Am. Chem. Soc.*, 126:11852–11863, 2004.
- [818] P.R. Dando. Strombine [*N*-(carboxymethyl)-D-alanine] dehydrogenase and alanopine [*meso-N*-(1-carboxyethyl)-alanine dehydrogenase from the mussel *Mytilus edulis* L. *Biochem. Soc. Trans.*, 9:297–298, 1981.
- [819] T.T. Dang, X. Chen, and P.J. Facchini. Acetylation serves as a protective group in noscapine biosynthesis in opium poppy. *Nat. Chem. Biol.*, 11:104–106, 2015.
- [820] T.T. Dang and P.J. Facchini. Cloning and characterization of canadine synthase involved in noscapine biosynthesis in opium poppy. *FEBS Lett.*, 588:198–204, 2014.
- [821] T.T. Dang and P.J. Facchini. CYP82Y1 is *N*-methylcanadine 1-hydroxylase, a key noscapine biosynthetic enzyme in opium poppy. *J. Biol. Chem.*, 289:2013–2026, 2014.
- [822] Y. Dang, W.E. Dale, and O.R. Brown. Comparative effects of oxygen on indoleamine 2,3-dioxygenase and tryptophan 2,3-dioxygenase of the kynurenine pathway. *Free Radic. Biol. Med.*, 28:615–624, 2000.
- [823] W. Dangel, A. Tschech, and G. Fuchs. Enzyme-reactions involved in anaerobic cyclohexanol metabolism by a denitrifying *Pseudomonas* species. *Arch. Microbiol.*, 152:273–279, 1989.
- [824] S. Dango, N. Mosammaparast, M.E. Sowa, L.J. Xiong, F. Wu, K. Park, M. Rubin, S. Gygi, J.W. Harper, and Y. Shi. DNA unwinding by ASCC3 helicase is coupled to ALKBH3-dependent DNA alkylation repair and cancer cell proliferation. *Mol. Cell*, 44:373–384, 2011.
- [825] S.L. Daniel, C. Pils, and H.L. Drake. Oxalate metabolism by the acetogenic bacterium *Moorella thermoacetica*. *FEMS Microbiol. Lett.*, 231:39–43, 2004.
- [826] C.E. Dann, Bruick 3rd, Deisenhofer R.K., and J. Structure of factor-inhibiting hypoxia-inducible factor 1: An asparaginyl hydroxylase involved in the hypoxic response pathway. *Proc. Natl. Acad. Sci. USA*, 99:15351–15356, 2002.
- [827] D.J. Danner, S.K. Lemmon, J.C., Elsas Beharse, , and II. Purification and characterization of branched chain α -ketoacid dehydrogenase from bovine liver mitochondria. *J. Biol. Chem.*, 254:5522–5526, 1979.
- [828] S. Darnet, M. Bard, and A. Rahier. Functional identification of sterol-4 α -methyl oxidase cDNAs from *Arabidopsis thaliana* by complementation of a yeast *erg25* mutant lacking sterol-4 α -methyl oxidation. *FEBS Lett.*, 508:39–43, 2001.

- [829] S. Darnet and A. Rahier. Plant sterol biosynthesis: identification of two distinct families of sterol 4 α -methyl oxidases. *Biochem. J.*, 378:889–898, 2004.
- [830] R. Daruwala and R. Meganathan. Dimethyl sulfoxide reductase is not required for trimethylamine *N*-oxide reduction in *Escherichia coli*. *FEMS Microbiol. Lett.*, 67:255–259, 1991.
- [831] A.G. Datta and H. Katznelson. The oxidation of 2-ketogluconate by a partially purified enzyme from *Acetobacter melanogenum*. *Arch. Biochem. Biophys.*, 65:576–578, 1956.
- [832] P.K. Datta, B.J.D. Meeuse, V. Engstrom-Heg, and S.H. Hilal. Moss oxalic acid oxidase - a flavoprotein. *Biochim. Biophys. Acta*, 17:602–603, 1955.
- [833] S.C. Daubner, G. Gadda, M.P. Valley, and P.F. Fitzpatrick. Cloning of nitroalkane oxidase from *Fusarium oxysporum* identifies a new member of the acyl-CoA dehydrogenase superfamily. *Proc. Natl. Acad. Sci. USA*, 99:2702–2707, 2002.
- [834] S.C. Daubner and R.T. Matthews. Purification and properties of methylenetetrahydrofolate reductase from pig liver. *J. Biol. Chem.*, 257:140–145, 1982.
- [835] K.D. Daughtry, Y. Xiao, D. Stoner-Ma, E. Cho, A.M. Orville, P. Liu, and K.N. Allen. Quaternary ammonium oxidative demethylation: X-ray crystallographic, resonance Raman, and UV-visible spectroscopic analysis of a Rieske-type demethylase. *J. Am. Chem. Soc.*, 134:2823–2834, 2012.
- [836] G.O. Daumy and A.S. McColl. Induction of 3-hydroxybenzoate 2-hydroxylase in a *Pseudomonas testosteroni* mutant. *J. Bacteriol.*, 149:384–385, 1982.
- [837] J.F. Davey and P.W. Trudgill. The metabolism of *trans*-cyclohexan-1,2-diol by an *Acinetobacter species*. *Eur. J. Biochem.*, 74:115–127, 1977.
- [838] S.J. Davidson and P. Talalay. Purification and mechanism of action of a steroid Δ^4 -5 β -dehydrogenase. *J. Biol. Chem.*, 241:906–915, 1966.
- [839] V.L. Davidson. Electron transfer in quinoproteins. *Arch. Biochem. Biophys.*, 428:32–40, 2004.
- [840] D.D. Davies. The purification and properties of glycolaldehyde dehydrogenase. *J. Exp. Bot.*, 11:289–295, 1960.
- [841] D.C. Davison. Studies on plant formic dehydrogenase. *Biochem. J.*, 49:520–526, 1951.
- [842] C.R. Dawson and W.B. Tarpley. The copper oxidases. In J.B. Sumner and K. Myrbäck, editors, *The Enzymes*, volume 2, pages 454–498. Academic Press, New York, 1st edition, 1951.
- [843] F.T. de Castro, J.M. Price, and R.R. Brown. Reduced triphosphopyridinenucleotide requirement for the enzymatic formation of 3-hydroxykynurenine from L-kynurenine. *J. Am. Chem. Soc.*, 78:2904–2905, 1956.
- [844] W. De-Eknamkul, T. Tanahashi, and M.H. Zenk. Enzymic 10-hydroxylation and 10-*O*-methylation of dihydrosanguinarine in dihydrochelirubine formation by *Eschscholtzia*. *Phytochemistry*, 31:2713–2717, 1992.
- [845] W. De-Eknamkul and M.H. Zenk. Purification and properties of 1,2-dehydroreticulium reductase from *Papaver somniferum* seedlings. *Phytochemistry*, 31:813–821, 1992.
- [846] E. de Jong, W.J.H. van Berkel, R.P. van der Zwan, and J.A.M. de Bont. Purification and characterization of vanillyl-alcohol oxidase from *Penicillium simplicissimum*, a novel aromatic alcohol oxidase containing covalently bound FAD. *Eur. J. Biochem.*, 208:651–657, 1992.
- [847] G.A. de Jong, J. Caldeira, J. Sun, J.A. Jongejan, S. de Vries, T.M. Loehr, I. Moura, J.J. Moura, and J.A. Duine. Characterization of the interaction between PQQ and heme c in the quinohemoprotein ethanol dehydrogenase from *Comamonas testosteroni*. *Biochemistry*, 34:9451–9458, 1995.
- [848] J.W. de Kraker, M.C. Franssen, M. Joerink, A. de Groot, and H.J. Bouwmeester. Biosynthesis of costunolide, dihydrocostunolide, and leucodin. Demonstration of cytochrome *p*₄₅₀-catalyzed formation of the lactone ring present in sesquiterpene lactones of chicory. *Plant Physiol.*, 129:257–268, 2002.
- [849] J.L. de La Fuente, A. Rumbero, J.F. Martin, and P. Liras. Δ -1-piperideine-6-carboxylate dehydrogenase, a new enzyme that forms α -amino adipate in *Streptomyces clavuligerus* and other cephamycin C-producing actinomycetes. *Biochem. J.*, 327:59–64, 1997.

- [850] R.S. de la Motte and F.W. Wagner. *Aspergillus niger* sulfhydryl oxidase. *Biochemistry*, 26:7363–7371, 1987.
- [851] V. de Lorenzo, A. Bindereif, B.H. Paw, and J.B. Neilands. Aerobactin biosynthesis and transport genes of plasmid ColV-K30 in *Escherichia coli* K-12. *J. Bacteriol.*, 165:570–578, 1986.
- [852] G. de Luca, P. de Philip, M. Rousset, J.P. Belaich, and Z. Dermoun. The NADP-reducing hydrogenase of *Desulfovibrio fructosovorans*: Evidence for a native complex with hydrogen-dependent methyl-viologen-reducing activity. *Biochem. Biophys. Res. Commun.*, 248:591–596, 1998.
- [853] P. de Marco, P. Moradas-Ferreira, T.P. Higgins, I. McDonald, E.M. Kenna, and J.C. Murrell. Molecular analysis of a novel methanesulfonic acid monooxygenase from the methylotroph *Methylosulfonomonas methylovora*. *J. Bacteriol.*, 181:2244–2251, 1999.
- [854] N. de Vetten, J. ter Horst, H.P. van Schaik, A. de Boer, J. Mol, and R. Koes. A cytochrome b_5 is required for full activity of flavonoid 3', 5'-hydroxylase, a cytochrome P450 involved in the formation of blue flower colors. *Proc. Natl. Acad. Sci. USA*, 96:778–783, 1999.
- [855] S. de Vries and L.A. Grivell. Purification and characterization of a rotenone-insensitive NADH:Q6 oxidoreductase from mitochondria of *Saccharomyces cerevisiae*. *Eur. J. Biochem.*, 176:377–384, 1988.
- [856] J.R. de Wet, K.V. Wood, D.R. Helinski, and M. DeLuca. Cloning of firefly luciferase cDNA and the expression of active luciferase in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA*, 82:7870–7873, 1985.
- [857] S. DeBolt, D.R. Cook, and C.M. Ford. L-Tartaric acid synthesis from vitamin C in higher plants. *Proc. Natl. Acad. Sci. USA*, 103:5608–5613, 2006.
- [858] L. Debussche, D. Thibaut, B. Cameron, J. Crouzet, and F. Blanche. Biosynthesis of the corrin macrocycle of coenzyme B₁₂ in *Pseudomonas denitrificans*. *J. Bacteriol.*, 175:7430–7440, 1993.
- [859] K. Decker and H. Bleege. Induction and purification of stereospecific nicotine oxidizing enzymes from *Arthrobacter oxidans*. *Biochim. Biophys. Acta*, 105:313–324, 1965.
- [860] R.H. Decker, H.H. Kang, F.R. Leach, and L.M. Henderson. Purification and properties of 3-hydroxyanthranilic acid oxidase. *J. Biol. Chem.*, 236:3076–3082, 1961.
- [861] J.J. DeFrank and D.W. Ribbons. *p*-cymene pathway in *Pseudomonas putida*: initial reactions. *J. Bacteriol.*, 129:1356–1364, 1977.
- [862] S.S. Dehal and R. Croteau. Metabolism of monoterpenes: specificity of the dehydrogenases responsible for the biosynthesis of camphor, 3-thujone, and 3-isothujone. *Arch. Biochem. Biophys.*, 258:287–291, 1987.
- [863] J. Deistung, F.C. Cannon, M.C. Cannon, S. Hill, and R.N. Thorneley. Electron transfer to nitrogenase in *Klebsiella pneumoniae*. *nifF* gene cloned and the gene product, a flavodoxin, purified. *Biochem. J.*, 231:743–753, 1985.
- [864] X. De Deken, D. Wang, J.E. Dumont, and F. Miot. Characterization of ThOX proteins as components of the thyroid H₂O₂-generating system. *Exp. Cell*, 273:187–196, 2002.
- [865] X. De Deken, D. Wang, M.C. Many, S. Costagliola, F. Libert, G. Vassart, J.E. Dumont, and F. Miot. Cloning of two human thyroid cDNAs encoding new members of the NADPH oxidase family. *J. Biol. Chem.*, 275:23227–23233, 2000.
- [866] E.E. Dekker and R.R. Swain. Formation of D_g-1-amino-2-propanol by a highly purified enzyme from *Escherichia coli*. *Biochim. Biophys. Acta*, 158:306–307, 1968.
- [867] R.F.H. Dekker. Induction and characterization of a cellobiose dehydrogenase produced by a species of *Monilia*. *J. Gen. Microbiol.*, 120:309–316, 1980.
- [868] R.F.H. Dekker. Cellobiose dehydrogenase produced by *Monilia* sp. *Methods Enzymol.*, 160:454–463, 1988.
- [869] A. del Castillo-Olivares and G. Gil. α 1-Fetoprotein transcription factor is required for the expression of sterol 12 α -hydroxylase, the specific enzyme for cholic acid synthesis. Potential role in the bile acid-mediated regulation of gene transcription. *J. Biol. Chem.*, 275:17793–17799, 2000.
- [870] C. dela Seña, K.M. Riedl, S. Narayanasamy, R.W. Curley, Schwartz Jr., Harrison S.J., and E.H. The human enzyme that converts dietary provitamin A carotenoids to vitamin A is a dioxygenase. *J. Biol. Chem.*, 289:13661–13666, 2014.

- [871] F.P. Delafield, K.E. Cooksey, and M. Doudoroff. β -Hydroxybutyric dehydrogenase and dimer hydrolase of *Pseudomonas lemoignei*. *J. Biol. Chem.*, 240:4023–4028, 1965.
- [872] S.B. delCardayré, K.P. Stock, G.L. Newton, R.C. Fahey, and J.E. Davies. Coenzyme A disulfide reductase, the primary low molecular weight disulfide reductase from *Staphylococcus aureus*. Purification and characterization of the native enzyme. *J. Biol. Chem.*, 273:5744–5751, 1998.
- [873] S. Dell'Acqua, S.R. Pauleta, P.M. Paes de Sousa, E. Monzani, L. Casella, J.J. Moura, and I. Moura. A new CuZ active form in the catalytic reduction of N₂O by nitrous oxide reductase from *Pseudomonas nautica*. *J. Biol. Inorg. Chem.*, 15:967–976, 2010.
- [874] J.K. Demmer, H. Huang, S. Wang, U. Demmer, R.K. Thauer, and U. Ermler. Insights into flavin-based electron bifurcation via the NADH-dependent reduced ferredoxin:NADP oxidoreductase Structure. *J. Biol. Chem.*, 290:21985–21995, 2015.
- [875] R. DeMoss. Triphosphopyridine nucleotide-specific ethanol dehydrogenase from *Leuconostoc mesenteroides*. *Bacteriol. Proc.*, pages 81–81, 1953.
- [876] M.E. Dempsey, J.D. Seaton, G.J. Schroepfer, and R.W. Trockman. The intermediary role of $\Delta^{5,7}$ -cholestadien-3 β -ol in cholesterol biosynthesis. *J. Biol. Chem.*, 239:1381–1387, 1964.
- [877] H. Den, W.G. Robinson, and M.J. Coon. Enzymatic conversion of β -hydroxypropionate to malonic semialdehyde. *J. Biol. Chem.*, 234:1666–1671, 1959.
- [878] L. Deng, E.S. Vysotski, S.V. Markova, Z.J. Liu, J. Lee, J. Rose, and B.C. Wang. All three Ca²⁺-binding loops of photoproteins bind calcium ions: the crystal structures of calcium-loaded apo-aequorin and apo-obelin. *Protein Sci.*, 14:663–675, 2005.
- [879] K. Denger and A.M. Cook. Racemase activity effected by two dehydrogenases in sulfolactate degradation by *Chromohalobacter salexigens*: purification of (S)-sulfolactate dehydrogenase. *Microbiology*, 156:967–974, 2010.
- [880] K. Denger, M. Weiss, A.K. Felux, A. Schneider, C. Mayer, D. Spittler, T. Huhn, A.M. Cook, and D. Schleheck. Sulphoglycolysis in *Escherichia coli* K-12 closes a gap in the biogeochemical sulphur cycle. *Nature*, 507:114–117, 2014.
- [881] U. Dengler, K. Niefind, M. Kiess, and D. Schomburg. Crystal structure of a ternary complex of D-2-hydroxyisocaproate dehydrogenase from *Lactobacillus casei*, NAD⁺ and 2-oxoisocaproate at 1.9 Å resolution. *J. Mol. Biol.*, 267:640–660, 1997.
- [882] D. Dennis and N.O. Kaplan. D and L-lactic acid dehydrogenase in *Lactobacillus plantarum*. *J. Biol. Chem.*, 235:810–818, 1960.
- [883] S.A. Denome, D.C. Stanley, E.S. Olson, and K.D. Young. Metabolism of dibenzothiophene and naphthalene in *Pseudomonas* strains: complete DNA sequence of an upper naphthalene catabolic pathway. *J. Bacteriol.*, 175:6890–6901, 1993.
- [884] U. Deppenmeier. The membrane-bound electron transport system of *Methanosarcina* species. *J. Bioenerg. Biomembr.*, 36:55–64, 2004.
- [885] U. Deppenmeier, T. Lienard, and G. Gottschalk. Novel reactions involved in energy conservation by methanogenic archaea. *FEBS Lett.*, 457:291–297, 1999.
- [886] J. Dermer and G. Fuchs. Molybdoenzyme that catalyzes the anaerobic hydroxylation of a tertiary carbon atom in the side chain of cholesterol. *J. Biol. Chem.*, 287:36905–36916, 2012.
- [887] D.V. DerVartanian and J. Le Gall. A monomolecular electron transfer chain: structure and function of cytochrome c₃. *Biochim. Biophys. Acta*, 346:79–99, 1974.
- [888] R.J. DeSa. Putrescine oxidase from *Micrococcus rubens*. Purification and properties of the enzyme. *J. Biol. Chem.*, 247:5527–5534, 1972.
- [889] E. Desmond and S. Gribaldo. Phylogenomics of sterol synthesis: insights into the origin, evolution, and diversity of a key eukaryotic feature. *Genome Biol Evol*, 1:364–381, 2009.

- [890] S.R. Devenish, J.W. Blunt, and J.A. Gerrard. NMR studies uncover alternate substrates for dihydrodipicolinate synthase and suggest that dihydrodipicolinate reductase is also a dehydratase. *J. Med. Chem.*, 53:4808–4812, 2010.
- [891] J. Deveryshetty and P.S. Phale. Biodegradation of phenanthrene by *Alcaligenes* sp. strain PPH: partial purification and characterization of 1-hydroxy-2-naphthoic acid hydroxylase. *FEMS Microbiol. Lett.*, 311:93–101, 2010.
- [892] A.S. Devlin and M.A. Fischbach. A biosynthetic pathway for a prominent class of microbiota-derived bile acids. *Nat. Chem. Biol.*, 11:685–690, 2015.
- [893] A.R. Dewanti and J.A. Duine. Reconstitution of membrane-integrated quinoprotein glucose dehydrogenase apoenzyme with PQQ and the holoenzyme's mechanism of action. *Biochemistry*, 37:6810–6818, 1998.
- [894] A.R. Dewanti, Y. Xu, and B. Mitra. Esters of mandelic acid as substrates for (*S*)-mandelate dehydrogenase from *Pseudomonas putida*: implications for the reaction mechanism. *Biochemistry*, 43:1883–1890, 2004.
- [895] A.R. Dewanti, Y. Xu, and B. Mitra. Role of glycine 81 in (*S*)-mandelate dehydrogenase from *Pseudomonas putida* in substrate specificity and oxidase activity. *Biochemistry*, 43:10692–10700, 2004.
- [896] D.L. DeWitt and W.L. Smith. Primary structure of prostaglandin G/H synthase from sheep vesicular gland determined from the complementary DNA sequence. *Proc. Natl. Acad. Sci. USA*, 85:1412–1416, 1988.
- [897] S. Dey, G.A. Grant, and J.C. Sacchettini. Crystal structure of *Mycobacterium tuberculosis* D-3-phosphoglycerate dehydrogenase: extreme asymmetry in a tetramer of identical subunits. *J. Biol. Chem.*, 280:14892–14899, 2005.
- [898] Y. Deyashiki, A. Ogasawara, T. Nakayama, M. Nakanishi, Y. Miyabe, K. Sato, and A. Hara. Molecular cloning of two human liver 3 α -hydroxysteroid/dihydrodiol dehydrogenase isoenzymes that are identical with chlordecone reductase and bile-acid binder. *Biochem. J.*, 299:545–552, 1994.
- [899] G. di Prisco, L. Casola, and A. Giuditta. Purification and properties of a soluble reduced nicotinamide-adenine dinucleotide (phosphate) dehydrogenase from the hepatopancreas of *Octopus vulgaris*. *Biochem. J.*, 105:455–460, 1967.
- [900] E. Díaz, A. Ferrández, and J.L. García. Characterization of the *hca* cluster encoding the dioxygenolytic pathway for initial catabolism of 3-phenylpropionic acid in *Escherichia coli* K-12. *J. Bacteriol.*, 180:2915–2923, 1998.
- [901] V. Diaz-Sanchez, A.F. Estrada, D. Trautmann, S. Al-Babili, and J. Avalos. The gene *carD* encodes the aldehyde dehydrogenase responsible for neurosporaxanthin biosynthesis in *Fusarium fujikuroi*. *FEBS J.*, 278:3164–3176, 2011.
- [902] R.A. Dick, M.K. Kwak, T.R. Sutter, and T.W. Kensler. Antioxidative function and substrate specificity of NAD(P)H-dependent alkenal/one oxidoreductase. A new role for leukotriene B₄ 12-hydroxydehydrogenase/15-oxoprostaglandin 13-reductase. *J. Biol. Chem.*, 276:40803–40810, 2001.
- [903] F. Dickens and G.E. Glock. Direct oxidation of glucose-6-phosphate, 6-phosphogluconate and pentose-5-phosphate by enzymes of animal origin. *Biochem. J.*, 50:81–95, 1951.
- [904] M.L. Dickens, N.D. Priestley, and W.R. Strohl. *In vivo* and *in vitro* bioconversion of ϵ -rhodomycinone glycoside to doxorubicin: functions of DauP, DauK, and DoxA. *J. Bacteriol.*, 179:2641–2650, 1997.
- [905] M.L. Dickens, J. Ye, and W.R. Strohl. Cloning, sequencing, and analysis of aklaviketone reductase from *Streptomyces* sp. strain C5. *J. Bacteriol.*, 178:3384–3388, 1996.
- [906] S. Dickert, A.J. Pierik, D. Linder, and W. Buckel. The involvement of coenzyme A esters in the dehydration of (*R*)-phenyllactate to (*E*)-cinnamate by *Clostridium sporogenes*. *Eur. J. Biochem.*, 267:3874–3884, 2000.
- [907] G. Diekert and M. Ritter. Purification of the nickel protein carbon monoxide dehydrogenase of *Clostridium thermoaceticum*. *FEBS Lett.*, 151:41–44, 1983.
- [908] T. Dierks, C. Miech, J. Hummerjohann, B. Schmidt, M.A. Kertesz, and K. von Figura. Posttranslational formation of formylglycine in prokaryotic sulfatases by modification of either cysteine or serine. *J. Biol. Chem.*, 273:25560–25564, 1998.
- [909] T. Dierks, B. Schmidt, and K. von Figura. Conversion of cysteine to formylglycine: a protein modification in the endoplasmic reticulum. *Proc. Natl. Acad. Sci. USA*, 94:11963–11968, 1997.

- [910] A. Dietl, C. Ferousi, W.J. Maalcke, A. Menzel, S. de Vries, J.T. Keltjens, M.S. Jetten, B. Kartal, and T.R. Barends. The inner workings of the hydrazine synthase multiprotein complex. *Nature*, 527:394–397, 2015.
- [911] M. Dieuaide-Noubhani, D. Novikov, E. Baumgart, J.C. Vanhooren, M. Fransen, M. Goethals, J. Vandekerckhove, P.P. Van Veldhoven, and G.P. Mannaerts. Further characterization of the peroxisomal 3-hydroxyacyl-CoA dehydrogenases from rat liver. Relationship between the different dehydrogenases and evidence that fatty acids and the C₂₇ bile acids di- and tri-hydroxycoprostanic acids are metabolized by separate multifunctional proteins. *Eur. J. Biochem.*, 240:660–666, 1996.
- [912] M. Dieuaide-Noubhani, D. Novikov, E. Baumgart, J.C. Vanhooren, M. Fransen, M. Goethals, J. Vandekerckhove, P.P. Van Veldhoven, and G.P. Mannaerts. Erratum report. Further characterization of the peroxisomal 3-hydroxyacyl-CoA dehydrogenases from rat liver. Relationship between the different dehydrogenases and evidence that fatty acids and the C₂₇ bile acids di- and tri-hydroxycoprostanic acids are metabolized by separate multifunctional proteins. *Eur. J. Biochem.*, 243:537–537, 1997.
- [913] W.P. Dijkman and M.W. Fraaije. Discovery and characterization of a 5-hydroxymethylfurfural oxidase from *Methylovorus* sp. strain MP688. *Appl. Environ. Microbiol.*, 80:1082–1090, 2014.
- [914] W.P. Dijkman, D.E. Groothuis, and M.W. Fraaije. Enzyme-catalyzed oxidation of 5-hydroxymethylfurfural to furan-2,5-dicarboxylic acid. *Angew. Chem. Int. Ed. Engl.*, 53:6515–6518, 2014.
- [915] G.L. Dilworth. Properties of the selenium-containing moiety of nicotinic acid hydroxylase from *Clostridium barkeri*. *Arch. Biochem. Biophys.*, 219:30–38, 1982.
- [916] G.L. Dilworth. Occurrence of molybdenum in the nicotinic acid hydroxylase from *Clostridium barkeri*. *Arch. Biochem. Biophys.*, 221:565–569, 1983.
- [917] M.J. Dilworth, M.E. Eldridge, and R.R. Eady. Correction for creatine interference with the direct indophenol measurement of NH₃ in steady-state nitrogenase assays. *Anal. Biochem.*, 207:6–10, 1992.
- [918] M.J. Dilworth, M.E. Eldridge, and R.R. Eady. The molybdenum and vanadium nitrogenases of *Azotobacter chroococcum*: effect of elevated temperature on N₂ reduction. *Biochem. J.*, 289:395–400, 1993.
- [919] W. Ding, W. Deng, M. Tang, Q. Zhang, G. Tang, Y. Bi, and W. Liu. Biosynthesis of 3-methoxy-5-methyl naphthoic acid and its incorporation into the antitumor antibiotic azinomycin B. *Mol. Biosyst.*, 6:1071–1081, 2010.
- [920] A.T. Dinkova-Kostova, D.R. Gang, L.B. Davin, D.L. Bedgar, A. Chu, and N.G. Lewis. (+)-Pinoresinol/(+)-lariciresinol reductase from *Forsythia intermedia*. Protein purification, cDNA cloning, heterologous expression and comparison to isoflavone reductase. *J. Biol. Chem.*, 271:29473–29482, 1996.
- [921] P.V. Dip, N. Kamariah, M.S. Subramanian Manimekhalai, W. Nartey, A.M. Balakrishna, F. Eisenhaber, B. Eisenhaber, and G. Gruber. Structure, mechanism and ensemble formation of the alkylhydroperoxide reductase subunits AhpC and AhpF from *Escherichia coli*. *Acta Crystallogr. D Biol. Crystallogr.*, 70:2848–2862, 2014.
- [922] H. Dittrich and T.M. Kutchan. Molecular cloning, expression and induction of the berberine bridge enzyme, an enzyme essential to the formation of benzophenanthridine alkaloids in the response of plants to pathogenic attack. *Proc. Natl. Acad. Sci. USA*, 88:9969–9973, 1991.
- [923] D. Ditullio, D. Anderson, C.S. Chen, and C.J. Sih. L-Carnitine via enzyme-catalyzed oxidative kinetic resolution. *Bioorg. Med. Chem.*, 2:415–420, 1994.
- [924] N.G. Divakar, V. Subramanian, M. Sugumaran, and C.S. Vaidyanathan. Indole oxygenase from the leaves of *Jasminum grandiflorum*. *Plant Sci. Lett.*, 15:177–181, 1979.
- [925] M. Dixon and P. Kenworthy. D-Aspartate oxidase of kidney. *Biochim. Biophys. Acta*, 146:54–76, 1967.
- [926] M. Dixon and K. Kleppe. D-Amino acid oxidase. I. Dissociation and recombination of the haloenzyme. *Biochim. Biophys. Acta*, 96:357–367, 1965.
- [927] M. Dixon and K. Kleppe. D-Amino acid oxidase. III. Effect of pH. *Biochim. Biophys. Acta*, 96:383–389, 1965.
- [928] M. Dixon and K. Kleppe. D-Amino acid oxidase. II. Specificity, competitive inhibition and reaction sequence. *Biochim. Biophys. Acta*, 96:368–382, 1965.

- [929] K.Y. Djoko, L.X. Chong, A.G. Wedd, and Z. Xiao. Reaction mechanisms of the multicopper oxidase CueO from *Escherichia coli* support its functional role as a cuprous oxidase. *J. Am. Chem. Soc.*, 132:2005–2015, 2010.
- [930] S. Djordjevic, Y. Dong, R. Paschke, F.E. Frerman, A.W. Strauss, and J.J. Kim. Identification of the catalytic base in long chain acyl-CoA dehydrogenase. *Biochemistry*, 33:4258–4264, 1994.
- [931] Keister D.L. and Stolzenbach F.E. Pyridine nucleotide transhydrogenase from spinach. I. Purification and properties. *J. Biol. Chem.*, 235:2989–2996, 1960.
- [932] A.A. Dmitrovskii, N.N. Gessler, S.B. Gomboeva, Yu.V. Ershov, and V.Ya. Bykhovskiy. Enzymatic oxidation of β -apo-8'-carotenol to β -apo-14'-carotenal by an enzyme different from β -carotene-15,15'-dioxygenase. *Biochemistry (Mosc.)*, 62:787–792, 1997.
- [933] H. Dobbek, L. Gremer, R. Kiefersauer, R. Huber, and O. Meyer. Catalysis at a dinuclear [CuSMo(=O)OH] cluster in a CO dehydrogenase resolved at 1.1-Å resolution. *Proc. Natl. Acad. Sci. USA*, 99:15971–15976, 2002.
- [934] H. Dobbek, V. Svetlitchnyi, L. Gremer, R. Huber, and O. Meyer. Crystal structure of a carbon monoxide dehydrogenase reveals a [Ni-4Fe-5S] cluster. *Science*, 293:1281–1285, 2001.
- [935] A.A. Dobritsa, J. Shrestha, M. Morant, F. Pinot, M. Matsuno, R. Swanson, B.L. Møller, and D. Preuss. CYP704B1 is a long-chain fatty acid ω -hydroxylase essential for sporopollenin synthesis in pollen of *Arabidopsis*. *Plant Physiol.*, 151:574–589, 2009.
- [936] D. Dodd, M.H. Spitzer, W. Van Treuren, B.D. Merrill, A.J. Hryckowian, S.K. Higginbottom, A. Le, T.M. Cowan, G.P. Nolan, M.A. Fischbach, and J.L. Sonnenburg. A gut bacterial pathway metabolizes aromatic amino acids into nine circulating metabolites. *Nature*, 551:648–652, 2017.
- [937] M.K. Doherty, S.L. Pealing, C.S. Miles, R. Moysey, P. Taylor, M.D. Walkinshaw, G.A. Reid, and S.K. Chapman. Identification of the active site acid/base catalyst in a bacterial fumarate reductase: a kinetic and crystallographic study. *Biochemistry*, 39:10695–10701, 2000.
- [938] P. Dokter, J. Frank, and J.A. Duine. Purification and characterization of quinoprotein glucose dehydrogenase from *Acinetobacter calcoaceticus* L.M.D. 79.41. *Biochem. J.*, 239:163–167, 1986.
- [939] C.T. Dolphin, J.H. Riley, R.L. Smith, E.A. Shephard, and I.R. Phillips. Structural organization of the human flavin-containing monooxygenase 3 gene (FMO3), the favored candidate for fish-odor syndrome, determined directly from genomic DNA. *Genomics*, 46:260–267, 1997.
- [940] G.F. Domagk and B.L. Horecker. Fructose and erythrose metabolism in *Alcaligenes faecalis*. *Arch. Biochem. Biophys.*, 109:342–349, 1965.
- [941] J. Domenech and J. Ferrer. A new D-2-hydroxyacid dehydrogenase with dual coenzyme-specificity from *Haloferax mediterranei*, sequence analysis and heterologous overexpression. *Biochim. Biophys. Acta*, 1760:1667–1674, 2006.
- [942] F. Domergue, A. Abbadi, U. Zähringer, H. Moreau, and E. Heinz. *In vivo* characterization of the first acyl-CoA Δ^6 -desaturase from a member of the plant kingdom, the microalga *Ostreococcus tauri*. *Biochem. J.*, 389:483–490, 2005.
- [943] J.E. Dominy, Simmons Jr., Hirschberger C.R., Hwang L.L., Coloso J., Stipanuk R.M., and M.H. Discovery and characterization of a second mammalian thiol dioxygenase, cysteamine dioxygenase. *J. Biol. Chem.*, 282:25189–25198, 2007.
- [944] V. Dommès and W.H. Kunau. 2,4-Dienoyl coenzyme A reductases from bovine liver and *Escherichia coli*. Comparison of properties. *J. Biol. Chem.*, 259:1781–1788, 1984.
- [945] V. Dommès, W. Luster, M. Cvetanovic, and W.-H. Kunau. Purification by affinity chromatography of 2,4-dienoyl-CoA reductases from bovine liver and *Escherichia coli*. *Eur. J. Biochem.*, 125:335–341, 1982.
- [946] K.O. Donaldson and J.C. Keresztesy. Naturally occurring forms of folic acid. I. *J. Biol. Chem.*, 234:3235–3240, 1959.
- [947] C. Dong, A. Kotzsch, M. Dorward, K.H. van Pee, and J.H. Naismith. Crystallization and X-ray diffraction of a halogenating enzyme, tryptophan 7-halogenase, from *Pseudomonas fluorescens*. *Acta Crystallogr. D Biol. Crystallogr.*, 60:1438–1440, 2004.

- [948] L.B. Dong, J.D. Rudolf, D. Kang, N. Wang, C.Q. He, Y. Deng, Y. Huang, K.N. Houk, Y. Duan, and B. Shen. Biosynthesis of thiocarboxylic acid-containing natural products. *Nat. Commun.*, 9:2362–2362, 2018.
- [949] Y. Dong, J. Yan, H. Du, M. Chen, T. Ma, and L. Feng. Engineering of LadA for enhanced hexadecane oxidation using random- and site-directed mutagenesis. *Appl. Microbiol. Biotechnol.*, 94:1019–1029, 2012.
- [950] N.A. Donoghue, D.B. Morris, and P.W. Trudgill. The purification and properties of cyclohexanone oxygenase from *Nocardia globerula* CL1 and *Acinetobacter* NCIB 9871. *Eur. J. Biochem.*, 63:175–192, 1976.
- [951] N.A. Donoghue and P.W. Trudgill. The metabolism of cyclohexanol by *Acinetobacter* NCIB 9871. *Eur. J. Biochem.*, 60:1–7, 1975.
- [952] E. Dorner and M. Boll. Properties of 2-oxoglutarate:ferredoxin oxidoreductase from *Thauera aromatica* and its role in enzymatic reduction of the aromatic ring. *J. Bacteriol.*, 184:3975–3983, 2002.
- [953] P.C. Dorrestein, E. Yeh, S. Garneau-Tsodikova, N.L. Kelleher, and C.T. Walsh. Dichlorination of a pyrrolyl-S-carrier protein by FADH₂-dependent halogenase PltA during pyoluteorin biosynthesis. *Proc. Natl. Acad. Sci. USA*, 102:13843–13848, 2005.
- [954] P.L. Dostert, M. Strolin Benedetti, and K.F. Tipton. Interactions of monoamine oxidase with substrates and inhibitors. *Med. Res. Rev.*, 9:45–89, 1989.
- [955] R.C. Doten and R.P. Mortlock. Characterization of xylitol-utilizing mutants of *Erwinia uredovora*. *J. Bacteriol.*, 161:529–533, 1985.
- [956] J.E. Dotzlafl and W.K. Yeh. Copurification and characterization of deacetoxycephalosporin C synthetase/hydroxylase from *Cephalosporium acremonium*. *J. Bacteriol.*, 169:1611–1618, 1987.
- [957] J.E. Dotzlafl and W.K. Yeh. Purification and properties of deacetoxycephalosporin C synthase from recombinant *Escherichia coli* and its comparison with the native enzyme purified from *Streptomyces clavuligerus*. *J. Biol. Chem.*, 264:10219–10227, 1989.
- [958] D.M. Doughty, E.G. Kurth, L.A. Sayavedra-Soto, D.J. Arp, and P.J. Bottomley. Evidence for involvement of copper ions and redox state in regulation of butane monooxygenase in *Pseudomonas butanovora*. *J. Bacteriol.*, 190:2933–2938, 2008.
- [959] T.I. Doukov, T. Iverson, J. Seravalli, S.W. Ragsdale, and C.L. Drennan. A Ni-Fe-Cu center in a bifunctional carbon monoxide dehydrogenase/acetyl-CoA synthase. *Science*, 298:567–572, 2002.
- [960] W.A. Doyle, W. Blodig, N.C. Veitch, K. Piontek, and A.T. Smith. Two substrate interaction sites in lignin peroxidase revealed by site-directed mutagenesis. *Biochemistry*, 37:15097–15105, 1998.
- [961] B. Dräger. Tropinone reductases, enzymes at the branch point of tropane alkaloid metabolism. *Phytochemistry*, 67:327–337, 2006.
- [962] B. Dräger, T. Hashimoto, and Y. Yamada. Purification and characterization of pseudotropine forming tropinone reductase from *Hyoscyamus niger* root cultures. *Agric. Biol. Chem.*, 52:2663–2667, 1988.
- [963] E.J. Drake and A.M. Gulick. Three-dimensional structures of *Pseudomonas aeruginosa* PvcA and PvcB, two proteins involved in the synthesis of 2-isocyano-6,7-dihydroxycoumarin. *J. Mol. Biol.*, 384:193–205, 2008.
- [964] C.L. Drennan, J. Heo, M.D. Sintchak, E. Schreiter, and P.W. Ludden. Life on carbon monoxide: X-ray structure of *Rhodospirillum rubrum* Ni-Fe-S carbon monoxide dehydrogenase. *Proc. Natl. Acad. Sci. USA*, 98:11973–11978, 2001.
- [965] C. Dresen, L.Y. Lin, I. D’Angelo, E.I. Tocheva, N. Strynadka, and L.D. Eltis. A flavin-dependent monooxygenase from *Mycobacterium tuberculosis* involved in cholesterol catabolism. *J. Biol. Chem.*, 285:22264–22275, 2010.
- [966] C.M. Driggers, S.J. Hartman, and P.A. Karplus. Structures of Arg- and Gln-type bacterial cysteine dioxygenase homologs. *Protein Sci.*, 24:154–161, 2015.
- [967] M.D. Driscoll, K.J. McLean, C. Levy, N. Mast, I.A. Pikuleva, P. Lafite, S.E. Rigby, D. Leys, and A.W. Munro. Structural and biochemical characterization of *Mycobacterium tuberculosis* CYP142: evidence for multiple cholesterol 27-hydroxylase activities in a human pathogen. *J. Biol. Chem.*, 285:38270–38282, 2010.

- [968] F. Dross, V. Geisler, R. Lenger, F. Theis, T. Krafft, F. Fahrenholz, , and E. , Duchêne, A., Tripier, D., Juvenal, K. and Kröger, A. The quinone-reactive Ni/Fe-hydrogenase of *Wolinella succinogenes*. *Eur. J. Biochem.*, 206:93–102, 1992.
- [969] F. Dross, V. Geisler, R. Lenger, F. Theis, T. Krafft, F. Fahrenholz, E. Kojro, A. Duchene, D. Tripier, and K. Juvenal. Erratum to "The quinone-reactive Ni/Fe-hydrogenase of *Wolinella succinogenes*". *Eur. J. Biochem.*, 214:949–950, 1993.
- [970] T.N. Druzhinina, Y.Y. Kusov, V.N. Shibaev, N.K. Kochetkov, P. Biely, S. Kucar, and S. Bauer. Uridine diphosphate 2-deoxyglucose. Chemical synthesis, enzymic oxidation and epimerization. *Biochim. Biophys. Acta*, 381:301–307, 1975.
- [971] B.N. La Du and V.G. Zannoni. The tyrosine oxidation system of liver. III. Further studies on the oxidation of *p*-hydroxyphenylpyruvic acid. *J. Biol. Chem.*, 219:273–281, 1956.
- [972] Y. Du, H. Chu, I.K. Chu, and C. Lo. CYP93G2 is a flavanone 2-hydroxylase required for C-glycosylflavone biosynthesis in rice. *Plant Physiol.*, 154:324–333, 2010.
- [973] Y.L. Du, L.M. Alkhalaf, and K.S. Ryan. *In vitro* reconstitution of indolmycin biosynthesis reveals the molecular basis of oxazolinone assembly. *Proc. Natl. Acad. Sci. USA*, 112:2717–2722, 2015.
- [974] W. Duane and J.W. Hastings. Flavin mononucleotide reductase of luminous bacteria. *Mol. Cell. Biochem.*, 6:53–64, 1975.
- [975] R.O. Duarte, M. Archer, J.M. Dias, S. Bursakov, R. Huber, I. Moura, M.J. Romao, and J.J. Moura. Biochemical/spectroscopic characterization and preliminary X-ray analysis of a new aldehyde oxidoreductase isolated from *Desulfovibrio desulfuricans* ATCC 27774. *Biochem. Biophys. Res. Commun.*, 268:745–749, 2000.
- [976] B.L. Dubbels, L.A. Sayavedra-Soto, and D.J. Arp. Butane monooxygenase of '*Pseudomonas butanovora*': purification and biochemical characterization of a terminal-alkane hydroxylating diiron monooxygenase. *Microbiology*, 153:1808–1816, 2007.
- [977] H. Dubourg, C. Stines-Chaumeil, C. Didierjean, F. Talfournier, S. Rahuel-Clermont, G. Branlant, and A. Aubry. Expression, purification, crystallization and preliminary X-ray diffraction data of methylmalonate-semialdehyde dehydrogenase from *Bacillus subtilis*. *Acta Crystallogr. D Biol. Crystallogr.*, 60:1435–1437, 2004.
- [978] R.E. Dugan, L.L. Slakey, and L.W. Porter. Stereospecificity of the transfer of hydrogen from reduced nicotinamide adenine dinucleotide phosphate to the acyl chain in the dehydrogenase-catalyzed reactions of fatty acid synthesis. *J. Biol. Chem.*, 245:6312–6316, 1970.
- [979] A.S. Duhaiman. Kinetic properties of camel lens ζ -crystallin. *Int. J. Biochem. Cell Biol.*, 28:1163–1168, 1996.
- [980] J.A. Duine, J. Frank, and P.E.J. Verweil. Structure and activity of the prosthetic group of methanol dehydrogenase. *Eur. J. Biochem.*, 108:187–192, 1980.
- [981] J.A. Duine, J. Frank, and J.K. Van Zeeland. Glucose dehydrogenase from *Acinetobacter calcoaceticus*: a 'quinoprotein'. *FEBS Lett.*, 108:443–446, 1979.
- [982] M. Dulchavsky, C.T. Clark, J.C.A. Bardwell, and F. Stull. A cytochrome *c* is the natural electron acceptor for nicotine oxidoreductase. *Nat. Chem. Biol.*, 17:344–350, 2021.
- [983] J.D. Duncan, J.O. Wallis, and M.R. Azari. Purification and properties of *Aerococcus viridans* lactate oxidase. *Biochem. Biophys. Res. Commun.*, 164:919–926, 1989.
- [984] H.B. Dunford. In *Heme peroxidases*, pages 33–218. Wiley-VCH, New York, 1999.
- [985] J.C. Dunlap and J.W. Hastings. The biological clock in *Gonyaulax* controls luciferase activity by regulating turnover. *J. Biol. Chem.*, 256:10509–10518, 1981.
- [986] T.M. Dunn, D. Haak, E. Monaghan, and T.J. Beeler. Synthesis of monohydroxylated inositolphosphorylceramide (IPC-C) in *Saccharomyces cerevisiae* requires Scs7p, a protein with both a cytochrome *b₅*-like domain and a hydroxylase/desaturase domain. *Yeast*, 14:311–321, 1998.
- [987] C. Dupuy, R. Ohayon, A. Valent, M.S. Noel-Hudson, D. Deme, and A. Virion. Purification of a novel flavoprotein involved in the thyroid NADPH oxidase. Cloning of the porcine and human cDNAs. *J. Biol. Chem.*, 274:37265–37269, 1999.

- [988] C. Dupuy, A. Virion, R. Ohayon, J. Kaniewski, D. Deme, and J. Pommier. Mechanism of hydrogen peroxide formation catalyzed by NADPH oxidase in thyroid plasma membrane. *J. Biol. Chem.*, 266:3739–3743, 1991.
- [989] H. Durichen, G. Diekert, and S. Studenik. Redox potential changes during ATP-dependent corrinoid reduction determined by redox titrations with europium(II)-DTPA. *Protein Sci.*, 28:1902–1908, 2019.
- [990] I.F. Durr and H. Rudney. The reduction of β -hydroxy- β -methylglutaryl coenzyme A to mevalonic acid. *J. Biol. Chem.*, 235:2572–2578, 1960.
- [991] T.K. Dutta, J. Chakraborty, M. Roy, D. Ghosal, P. Khara, and I.C. Gunsalus. Cloning and characterization of a *p*-cymene monooxygenase from *Pseudomonas chlororaphis* subsp. *aureofaciens*. *Res. Microbiol.*, 161:876–882, 2010.
- [992] T.K. Dutta and I.C. Gunsalus. Reductase gene sequences and protein structures: *p*-cymene methyl hydroxylase. *Biochem. Biophys. Res. Commun.*, 233:502–506, 1997.
- [993] R.J. Dutton, D. Boyd, M. Berkmen, and J. Beckwith. Bacterial species exhibit diversity in their mechanisms and capacity for protein disulfide bond formation. *Proc. Natl. Acad. Sci. USA*, 105:11933–11938, 2008.
- [994] P. Dworsky and O. Hoffmann-Ostenhof. L-3-Aldonic acid dehydrogenase from *Schwanniomyces occidentalis*. *Acta Biochim. Pol.*, 11:269–277, 1964.
- [995] T.M. Dwyer, L. Zhang, M. Muller, F. Marrugo, and F. Frerman. The functions of the flavin contact residues, α Arg²⁴⁹ and β Tyr¹⁶, in human electron transfer flavoprotein. *Biochim. Biophys. Acta*, 1433:139–152, 1999.
- [996] J.M. Dyer, D.C. Chapital, J.C. Kuan, R.T. Mullen, C. Turner, T.A. McKeon, and A.B. Pepperman. Molecular analysis of a bifunctional fatty acid conjugase/desaturase from tung. Implications for the evolution of plant fatty acid diversity. *Plant Physiol.*, 130:2027–2038, 2002.
- [997] O. Dym, E.A. Pratt, C. Ho, and D. Eisenberg. The crystal structure of D-lactate dehydrogenase, a peripheral membrane respiratory enzyme. *Proc. Natl. Acad. Sci. USA*, 97:9413–9418, 2000.
- [998] R.R. Eady. Current status of structure function relationships of vanadium nitrogenase. *Coordinat. Chem. Rev.*, 237:23–30, 2003.
- [999] R.R. Eady, T.R. Jarman, and P.J. Large. Microbial oxidation of amines. Partial purification of a mixed-function secondary-amine oxidase system from *Pseudomonas aminovorans* that contains an enzymically active cytochrome-*P*-420-type haemoprotein. *Biochem. J.*, 125:449–459, 1971.
- [1000] R.R. Eady and P.J. Large. Purification and properties of an amine dehydrogenase from *Pseudomonas* AM1 and its role in growth on methylamine. *Biochem. J.*, 106:245–255, 1968.
- [1001] R.R. Eady and P.J. Large. Bacterial oxidation of dimethylamine, a new mono-oxygenase reaction. *Biochem. J.*, 111:37P–38P, 1969.
- [1002] R.R. Eady and P.J. Large. Microbial oxidation of amines. Spectral and kinetic properties of the primary amine dehydrogenase of *Pseudomonas* AM1. *Biochem. J.*, 123:757–771, 1971.
- [1003] R.R. Eady, T.H. Richardson, R.W. Miller, M. Hawkins, and D.J. Lowe. The vanadium nitrogenase of *Azotobacter chroococcum*. Purification and properties of the Fe protein. *Biochem. J.*, 256:189–196, 1988.
- [1004] R.R. Eady, B.E. Smith, K.A. Cook, and J.R. Postgate. Nitrogenase of *Klebsiella pneumoniae*. Purification and properties of the component proteins. *Biochem. J.*, 128:655–675, 1972.
- [1005] S.E. Ealick and T.P. Begley. Biochemistry: molecular cannibalism. *Nature*, 446:387–388, 2007.
- [1006] R. Eaton and P.J. Chapman. Bacterial metabolism of naphthalene: construction and use of recombinant bacteria to study ring cleavage of 1,2-dihydroxynaphthalene and subsequent reactions. *J. Bacteriol.*, 174:7542–7554, 1992.
- [1007] R.W. Eaton. *p*-Cumate catabolic pathway in *Pseudomonas putida* F1: cloning and characterization of DNA carrying the *cmt* operon. *J. Bacteriol.*, 178:1351–1362, 1996.
- [1008] R.W. Eaton. *p*-Cymene catabolic pathway in *Pseudomonas putida* F1: cloning and characterization of DNA encoding conversion of *p*-cymene to *p*-cumate. *J. Bacteriol.*, 179:3171–3180, 1997.

- [1009] R.W. Eaton and P.J. Chapman. Formation of indigo and related compounds from indolecarboxylic acids by aromatic acid-degrading bacteria: chromogenic reactions for cloning genes encoding dioxygenases that act on aromatic acids. *J. Bacteriol.*, 177:6983–6988, 1995.
- [1010] K. Ebisuzaki and J.N. Williams. Preparation and partial purification of soluble choline dehydrogenase from liver mitochondria. *Biochem. J.*, 60:644–646, 1955.
- [1011] M. Eckhardt, A. Yaghootfam, S.N. Fewou, I. Zoller, and V. Gieselmann. A mammalian fatty acid hydroxylase responsible for the formation of α -hydroxylated galactosylceramide in myelin. *Biochem. J.*, 388:245–254, 2005.
- [1012] R. Edenharder and A. Pfützner. Characterization of NADP-dependent 12 β -hydroxysteroid dehydrogenase from *Clostridium paraputrificum*. *Biochim. Biophys. Acta*, 962:362–370, 1988.
- [1013] R. Edenharder, A. Pfützner, and R. Hammann. Characterization of NAD-dependent 3 α - and 3 β -hydroxysteroid dehydrogenase and of NADP-dependent 7 β -hydroxysteroid dehydrogenase from *Peptostreptococcus productus*. *Biochim. Biophys. Acta*, 1004:230–238, 1989.
- [1014] R. Edenharder and M. Pfützner. Partial purification and characterization of an NAD-dependent 3 β -hydroxysteroid dehydrogenase from *Clostridium innocuum*. *Appl. Environ. Microbiol.*, 55:1656–1659, 1989.
- [1015] J.R. Edgar and R.M. Bell. Biosynthesis in *Escherichia coli* of *sn*-glycerol 3-phosphate, a precursor of phospholipid. *J. Biol. Chem.*, 253:6348–6353, 1978.
- [1016] J.R. Edgar and R.M. Bell. Biosynthesis in *Escherichia coli* of *sn*-glycerol 3-phosphate, a precursor of phospholipid. Kinetic characterization of wild type and feedback-resistant forms of the biosynthetic *sn*-glycerol-3-phosphate dehydrogenase. *J. Biol. Chem.*, 253:6354–6363, 1978.
- [1017] J.R. Edgar and R.M. Bell. Biosynthesis in *Escherichia coli* of *sn*-glycerol-3-phosphate, a precursor of phospholipid. Further kinetic characterization of wild type and feedback-resistant forms of the biosynthetic *sn*-glycerol-3-phosphate dehydrogenase. *J. Biol. Chem.*, 255:3492–3497, 1980.
- [1018] D.E. Edmondson, W.C. Kenney, and T.P. Singer. Structural elucidation and properties of 8 α -(N^1 -histidyl)riboflavin: the flavin component of thiamine dehydrogenase and β -cyclopiasonate oxidocyclase. *Biochemistry*, 15:2937–2945, 1976.
- [1019] D.E. Edmondson, A. Mattevi, C. Binda, M. Li, and F. Hubálek. Structure and mechanism of monoamine oxidase. *Curr. Med. Chem.*, 11:1983–1993, 2004.
- [1020] K.Z. Edson, B. Prasad, J.D. Unadkat, Y. Suhara, T. Okano, F.P. Guengerich, and A.E. Rettie. Cytochrome P450-dependent catabolism of vitamin K: ω -hydroxylation catalyzed by human CYP4F2 and CYP4F11. *Biochemistry*, 52:8276–8285, 2013.
- [1021] C.A.F. Edwards and J.C. Orr. Comparison of the 3 α - and 20 β -hydroxysteroid dehydrogenase activities of the cortisone reductase of *Streptomyces hydrogenans*. *Biochemistry*, 17:4370–4376, 1978.
- [1022] B.T. Eger, K. Okamoto, C. Enroth, M. Sato, T. Nishino, E.F. Pai, and T. Nishino. Purification, crystallization and preliminary X-ray diffraction studies of xanthine dehydrogenase and xanthine oxidase isolated from bovine milk. *Acta Crystallogr. D Biol. Crystallogr.*, 56:1656–1658, 2000.
- [1023] G. Eggertsen, M. Olin, U. Andersson, H. Ishida, S. Kubota, U. Hellman, K.I. Okuda, and I. Björkhem. Molecular cloning and expression of rabbit sterol 12 α -hydroxylase. *J. Biol. Chem.*, 271:32269–32275, 1996.
- [1024] L.L. Eggink, R. LoBrutto, D.C. Brune, J. Brusslan, A. Yamasato, A. Tanaka, and J.K. Hooper. Synthesis of chlorophyll *b*: localization of chlorophyllide *a* oxygenase and discovery of a stable radical in the catalytic subunit. *BMC Plant Biol.*, 4:5–5, 2004.
- [1025] D.E. Ehmman, A.M. Gehring, and C.T. Walsh. Lysine biosynthesis in *Saccharomyces cerevisiae*: mechanism of α -aminoadipate reductase (Lys²) involves posttranslational phosphopantetheinylation by Lys⁵. *Biochemistry*, 38:6171–6177, 1999.
- [1026] K.C. Ehrlich, P. Li, L. Scharfenstein, and P.K. Chang. HypC, the anthrone oxidase involved in aflatoxin biosynthesis. *Appl. Environ. Microbiol.*, 76:3374–3377, 2010.

- [1027] E. Eichhorn, J.R. Van Der Poeg, M.A. Kertesz, and T. Leisinger. Characterization of α -ketoglutarate-dependent taurine dioxygenase from *Escherichia coli*. *J. Biol. Chem.*, 272:23031–23036, 1997.
- [1028] E. Eichhorn, J.R. van der Ploeg, and T. Leisinger. Characterization of a two-component alkanesulfonate monooxygenase from *Escherichia coli*. *J. Biol. Chem.*, 274:26639–26646, 1999.
- [1029] M.M. Eichhorn and M.A. Cynkin. Microbial metabolism of 2-deoxyglucose; 2-deoxyglucose acid dehydrogenase. *Biochemistry*, 4:159–165, 1965.
- [1030] O. Einsle, A. Messerschmidt, P. Bourenkov Stach, Bartunik G.P., Huber H.D., Kroneck R., and P.M.H. Structure of cytochrome *c* nitrite reductase. *Nature*, 400:476–480, 1999.
- [1031] S.-I. Ejiri, H. Weissbach, and N. Brot. Reduction of methionine sulfoxide to methionine by *Escherichia coli*. *J. Bacteriol.*, 139:161–164, 1979.
- [1032] S.-I. Ejiri, H. Weissbach, and N. Brot. The purification of methionine sulfoxide reductase from *Escherichia coli*. *Anal. Biochem.*, 102:393–398, 1980.
- [1033] A.P. Eker, J.K. Hessels, and R. Meerwaldt. Characterization of an 8-hydroxy-5-deazaflavin:NADPH oxidoreductase from *Streptomyces griseus*. *Biochim. Biophys. Acta*, 990:80–86, 1989.
- [1034] M. El-Fakhri and B. Middleton. The existence of an inner-membrane-bound, long acyl-chain-specific 3-hydroxyacyl-CoA dehydrogenase in mammalian mitochondria. *Biochim. Biophys. Acta*, 713:270–279, 1982.
- [1035] M.D. Elias, S. Nakamura, C.T. Migita, H. Miyoshi, H. Toyama, K. Matsushita, O. Adachi, and M. Yamada. Occurrence of a bound ubiquinone and its function in *Escherichia coli* membrane-bound quinoprotein glucose dehydrogenase. *J. Biol. Chem.*, 279:3078–3083, 2004.
- [1036] M.D. Elias, M. Tanaka, M. Sakai, H. Toyama, K. Matsushita, O. Adachi, and M. Yamada. C-terminal periplasmic domain of *Escherichia coli* quinoprotein glucose dehydrogenase transfers electrons to ubiquinone. *J. Biol. Chem.*, 276:48356–48361, 2001.
- [1037] R. Eliasson, E. Pontis, M. Fontecave, C. Gerez, J. Harder, H. Jornvall, M. Krook, and P. Reichard. Characterization of components of the anaerobic ribonucleotide reductase system from *Escherichia coli*. *J. Biol. Chem.*, 267:25541–25547, 1992.
- [1038] J.M. Elkins, K.S. Hewitson, L.A. McNeill, J.F. Seibel, I. Schlemminger, C.W. Pugh, P.J. Ratcliffe, and C.J. Schofield. Structure of factor-inhibiting hypoxia-inducible factor (HIF) reveals mechanism of oxidative modification of HIF-1 α . *J. Biol. Chem.*, 278:1802–1806, 2003.
- [1039] P.J. Ellis, T. Conrads, R. Hille, and P. Kuhn. Crystal structure of the 100 kDa arsenite oxidase from *Alcaligenes faecalis* in two crystal forms at 1.64 Å and 2.03 Å. *Structure*, 9:125–132, 2001.
- [1040] B.O. Elmore, J.A. Bollinger, and D.M. Dooley. Human kidney diamine oxidase: heterologous expression, purification, and characterization. *J. Biol. Inorg. Chem.*, 7:565–579, 2002.
- [1041] T. Elssner, L. Hennig, H. Frauendorf, D. Haferburg, and H.P. Kleber. Isolation, identification, and synthesis of γ -butyrobetainyl-CoA and crotonobetainyl-CoA, compounds involved in carnitine metabolism of *E. coli*. *Biochemistry*, 39:10761–10769, 2000.
- [1042] L.D. Eltis, B. Hofmann, H.J. Hecht, H. Lunsdorf, and K.N. Timmis. Purification and crystallization of 2,3-dihydroxybiphenyl 1,2-dioxygenase. *J. Biol. Chem.*, 268:2727–2732, 1993.
- [1043] J.J. Emanuele and P.F. Fitzpatrick. Mechanistic studies of the flavoprotein tryptophan 2-monooxygenase. 1. Kinetic mechanism. *Biochemistry*, 34:3710–3715, 1995.
- [1044] G.L. Endahl, C.D. Kochakia, and D. Hamm. Separation of a triphosphopyridine nucleotide-specific from a diphosphopyridine-specific 17 β -hydroxy (testosterone) dehydrogenase of guinea pig liver. *J. Biol. Chem.*, 235:2792–2796, 1960.
- [1045] T. Endoh, H. Habe, H. Nojiri, H. Yamane, and T. Omori. The σ^{54} -dependent transcriptional activator SfnR regulates the expression of the *Pseudomonas putida* *sfnFG* operon responsible for dimethyl sulphone utilization. *Mol. Microbiol.*, 55:897–911, 2005.

- [1046] T. Endoh, K. Kasuga, M. Horinouchi, T. Yoshida, H. Habe, H. Nojiri, and T. Omori. Characterization and identification of genes essential for dimethyl sulfide utilization in *Pseudomonas putida* strain DS1. *Appl. Microbiol. Biotechnol.*, 62:83–91, 2003.
- [1047] H.J. Engel, W. Domschke, M. Alberti, and G.F. Domagk. Protein structure and enzymatic activity. II. Purification and properties of a crystalline glucose-6-phosphate dehydrogenase from *Candida utilis*. *Biochim. Biophys. Acta*, 191:509–516, 1969.
- [1048] T.D. Engerson, T.G. McKelvey, D.B. Rhyne, E.B. Boggio, S.J. Snyder, and H.P. Jones. Conversion of xanthine dehydrogenase to oxidase in ischemic rat tissues. *J. Clin. Invest.*, 79:1564–1570, 1987.
- [1049] S. Englard, G. Kaysen, and G. Avigad. 5-keto-D-Fructose. VI. A specific reduced nicotinamide adenine dinucleotide phosphate-linked reductase from yeast. *J. Biol. Chem.*, 245:1311–1318, 1970.
- [1050] M. Engqvist, M.F. Drincovich, U.I. Flugge, and V.G. Maurino. Two D-2-hydroxy-acid dehydrogenases in *Arabidopsis thaliana* with catalytic capacities to participate in the last reactions of the methylglyoxal and β -oxidation pathways. *J. Biol. Chem.*, 284:25026–25037, 2009.
- [1051] H.G. Enoch and R.L. Lester. The role of a novel cytochrome *b*-containing nitrate reductase and quinone in the in vitro reconstruction of formate-nitrate reductase activity of *E. coli*. *Biochem. Biophys. Res. Commun.*, 61:1234–1241, 1974.
- [1052] H.G. Enoch and R.L. Lester. The purification and properties of formate dehydrogenase and nitrate reductase from *Escherichia coli*. *J. Biol. Chem.*, 250:6693–6705, 1975.
- [1053] J.M. Enright, M.B. Toomey, S.Y. Sato, S.E. Temple, J.R. Allen, R. Fujiwara, V.M. Kramlinger, L.D. Nagy, K.M. Johnson, Y. Xiao, M.J. How, S.L. Johnson, N.W. Roberts, V.J. Kefalov, F.P. Guengerich, and J.C. Corbo. Cyp27c1 red-shifts the spectral sensitivity of photoreceptors by converting vitamin A₁ into A₂. *Curr. Biol.*, 25:3048–3057, 2015.
- [1054] C. Enroth, B.T. Eger, K. Okamoto, T. Nishino, T. Nishino, and E.F. Pai. Crystal structures of bovine milk xanthine dehydrogenase and xanthine oxidase: structure-based mechanism of conversion. *Proc. Natl. Acad. Sci. USA*, 97:10723–10728, 2000.
- [1055] S.A. Ensign, M.R. Hyman, and D.J. Arp. *In vitro* activation of ammonia monooxygenase from *Nitrosomonas europaea* by copper. *J. Bacteriol.*, 175:1971–1980, 1993.
- [1056] B.D. Ensley and D.T. Gibson. Naphthalene dioxygenase: purification and properties of a terminal oxygenase component. *J. Bacteriol.*, 155:505–511, 1983.
- [1057] B.R. Epperly and E.E. Dekker. L-Threonine dehydrogenase from *Escherichia coli*. Identification of an active site cysteine residue and metal ion studies. *J. Biol. Chem.*, 266:6086–6092, 1991.
- [1058] A.C. Epstein, J.M. Gleadle, L.A. McNeill, K.S. Hewitson, J. O'Rourke, D.R. Mole, M. Mukherji, E. Metzen, M.I. Wilson, A. Dhanda, Y.M. Tian, N. Masson, D.L. Hamilton, P. Jaakkola, R. Barstead, J. Hodgkin, P.H. Maxwell, C.W. Pugh, C.J. Schofield, and P.J. Ratcliffe. *C. elegans* EGL-9 and mammalian homologs define a family of dioxygenases that regulate HIF by prolyl hydroxylation. *Cell*, 107:43–54, 2001.
- [1059] T.J. Erb, I.A. Berg, V. Brecht, M. Muller, G. Fuchs, and B.E. Alber. Synthesis of C₅-dicarboxylic acids from C₂-units involving crotonyl-CoA carboxylase/reductase: the ethylmalonyl-CoA pathway. *Proc. Natl. Acad. Sci. USA*, 104:10631–10636, 2007.
- [1060] T.J. Erb, V. Brecht, G. Fuchs, M. Muller, and B.E. Alber. Carboxylation mechanism and stereochemistry of crotonyl-CoA carboxylase/reductase, a carboxylating enoyl-thioester reductase. *Proc. Natl. Acad. Sci. USA*, 106:8871–8876, 2009.
- [1061] T.J. Erb, G. Fuchs, and B.E. Alber. (2*S*)-Methylsuccinyl-CoA dehydrogenase closes the ethylmalonyl-CoA pathway for acetyl-CoA assimilation. *Mol. Microbiol.*, 73:992–1008, 2009.
- [1062] H. Erlandsen, J.Y. Kim, M.G. Patch, A. Han, A. Volner, M.M. Abu-Omar, and R.C. Stevens. Structural comparison of bacterial and human iron-dependent phenylalanine hydroxylases: similar fold, different stability and reaction rates. *J. Mol. Biol.*, 320:645–661, 2002.
- [1063] M. Eschenbrenner, L.J. Chlumsky, P. Khanna, F. Strasser, and M.S. Jorns. Organization of the multiple coenzymes and subunits and role of the covalent flavin link in the complex heterotetrameric sarcosine oxidase. *Biochemistry*, 40:5352–5367, 2001.

- [1064] C.E. Espineda, A.S. Linford, D. Devine, and J.A. Brusslan. The *AtCAO* gene, encoding chlorophyll *a* oxygenase, is required for chlorophyll *b* synthesis in *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. USA*, 96:10507–10511, 1999.
- [1065] M.P. Estevéz, E. Legaz, L. Olmeda, F.J. Pérez, and C. Vincente. Purification and properties of a new enzyme from *Evernia prunastri*, which reduces L-uscnic acid. *Z. Naturforsch. C: Biosci.*, 36:35–39, 1981.
- [1066] A.F. Estrada, D. Maier, D. Scherzinger, J. Avalos, and S. Al-Babili. Novel apocarotenoid intermediates in *Neurospora crassa* mutants imply a new biosynthetic reaction sequence leading to neurosporaxanthin formation. *Fungal Genet. Biol.*, 45:1497–1505, 2008.
- [1067] A.F. Estrada, L. Youssar, D. Scherzinger, S. Al-Babili, and J. Avalos. The *ylo-1* gene encodes an aldehyde dehydrogenase responsible for the last reaction in the *Neurospora* carotenoid pathway. *Mol. Microbiol.*, 69:1207–1220, 2008.
- [1068] F. Etienne, D. Spector, N. Brot, and H. Weissbach. A methionine sulfoxide reductase in *Escherichia coli* that reduces the R enantiomer of methionine sulfoxide. *Biochem. Biophys. Res. Commun.*, 300:378–382, 2003.
- [1069] J.P. Evans, K. Ahn, and J.P. Klinman. Evidence that dioxygen and substrate activation are tightly coupled in dopamine β -monooxygenase. Implications for the reactive oxygen species. *J. Biol. Chem.*, 278:49691–49698, 2003.
- [1070] M.C.W. Evans and B.B. Buchanan. Photoreduction of ferredoxin and its use in carbon dioxide fixation by a subcellular system from a photosynthetic bacterium. *Proc. Natl. Acad. Sci. USA*, 53:1420–1425, 1965.
- [1071] A. Evarsson, J.L. Chuang, R.M. Wynn, S. Turley, D.T. Chuang, and W.G. Hol. Crystal structure of human branched-chain α -ketoacid dehydrogenase and the molecular basis of multienzyme complex deficiency in maple syrup urine disease. *Structure*, 8:277–291, 2000.
- [1072] J. Everse and N.O. Kaplan. Lactate dehydrogenases: structure and function. *Adv. Enzymol. Relat. Subj. Biochem.*, 37:61–133, 1973.
- [1073] K.M. Ewen, F. Hannemann, Y. Khatri, O. Perlova, R. Kappl, D. Krug, J. Huttermann, R. Muller, and R. Bernhardt. Genome mining in *Sorangium cellulosum* So ce56: identification and characterization of the homologous electron transfer proteins of a myxobacterial cytochrome P450. *J. Biol. Chem.*, 284:28590–28598, 2009.
- [1074] G.E. Exley, J.D. Colandene, and R.H. Garrett. Molecular cloning, characterization, and nucleotide sequence of nit-6, the structural gene for nitrite reductase in *Neurospora crassa*. *J. Bacteriol.*, 175:2379–2392, 1993.
- [1075] B. Ezraty, L. Aussel, and F. Barras. Methionine sulfoxide reductases in prokaryotes. *Biochim. Biophys. Acta*, 1703:221–229, 2005.
- [1076] G. F. and Kishore. G. M. Barry. Glyphosate tolerant plants, 1995.
- [1077] B.W. Faber, R.F. van Gorcom, and J.A. Duine. Purification and characterization of benzoate-para-hydroxylase, a cytochrome P450 (CYP53A1), from *Aspergillus niger*. *Arch. Biochem. Biophys.*, 394:245–254, 2001.
- [1078] R.L. Fagan, M.N. Nelson, P.M. Pagano, and B.A. Palfey. Mechanism of flavin reduction in Class 2 dihydroorotate dehydrogenases. *Biochemistry*, 45:14926–14932, 2006.
- [1079] D.L. Falcone, S. Gibson, B. Lemieux, and C. Somerville. Identification of a gene that complements an *Arabidopsis* mutant deficient in chloroplast ω 6 desaturase activity. *Plant Physiol.*, 106:1453–1459, 1994.
- [1080] H. Falkenhagen and J. Stöckigt. Enzymatic biosynthesis of vomilenine, a key intermediate of the ajmaline pathway, catalysed by a novel cytochrome *P*-450-dependent enzyme from plant cell cultures of *Rauwolfia serpentina*. *Z. Naturforsch. C: Biosci.*, 50:45–53, 1995.
- [1081] P.O. Falnes, R.F. Johansen, and E. Seeberg. AlkB-mediated oxidative demethylation reverses DNA damage in *Escherichia coli*. *Nature*, 419:178–182, 2002.
- [1082] D.-F. Fan, C.E. John, J. Zalitis, and D.S. Feingold. UDPacetylglucosamine dehydrogenase from *Achromobacter georgopolitanum*. *Arch. Biochem. Biophys.*, 135:45–49, 1969.
- [1083] F. Fan and G. Gadda. On the catalytic mechanism of choline oxidase. *J. Am. Chem. Soc.*, 127:2067–2074, 2005.
- [1084] F. Fan, M. Ghanem, and G. Gadda. Cloning, sequence analysis, and purification of choline oxidase from *Arthrobacter globiformis*: a bacterial enzyme involved in osmotic stress tolerance. *Arch. Biochem. Biophys.*, 421:149–158, 2004.

- [1085] L. Fan, J.F. Joseph, P. Durairaj, M.K. Parr, and M. Bureik. Conversion of chenodeoxycholic acid to cholic acid by human CYP8B1. *Biol. Chem.*, 400:625–628, 2019.
- [1086] L. Fang, T. Shi, Y. Chen, X. Wu, C. Zhang, X. Tang, Q.X. Li, and R. Hua. Kinetics and catabolic pathways of the insecticide chlorpyrifos, annotation of the degradation genes, and characterization of enzymes TcpA and Fre in *Cupriavidus nantongensis* X1(T). *J. Agric. Food Chem.*, 67:2245–2254, 2019.
- [1087] Q. Fang, J. Peng, and T. Dierks. Post-translational formylglycine modification of bacterial sulfatases by the radical S-adenosylmethionine protein AtsB. *J. Biol. Chem.*, 279:14570–14578, 2004.
- [1088] X. Fargetton, P. Galtier, and P. Delatour. Sulfoxidation of albendazole by a cytochrome *P*₄₅₀-independent monooxygenase from rat liver microsomes. *Vet. Res. Commun.*, 10:317–324, 1986.
- [1089] Z.S. Farhangrazi, B.R. Copeland, T. Nakayama, T. Amachi, I. Yamazaki, and L.S. Powers. Oxidation-reduction properties of compounds I and II of *Arthromyces ramosus* peroxidase. *Biochemistry*, 33:5647–5652, 1994.
- [1090] W. Farkas and C. Gilvarg. The reduction step in diaminopimelic acid biosynthesis. *J. Biol. Chem.*, 240:4717–4722, 1965.
- [1091] J.J. Farmer, Eagon III, and R.G. Aldoheuxuronic acid catabolism by a soil *Aeromonas*. *J. Bacteriol.*, 97:97–106, 1969.
- [1092] V.C. Farmer, M.E.K. Henderson, and J.D. Russell. Aromatic-alcohol-oxidase activity in the growth medium of *Polystictus versicolor*. *Biochem. J.*, 74:257–262, 1960.
- [1093] S.R. Farrell and C. Thorpe. Augmenter of liver regeneration: a flavin-dependent sulfhydryl oxidase with cytochrome *c* reductase activity. *Biochemistry*, 44:1532–1541, 2005.
- [1094] S.C. Farrow, J.M. Hagel, G.A. Beaudoin, D.C. Burns, and P.J. Facchini. Stereochemical inversion of (*S*)-reticuline by a cytochrome P450 fusion in opium poppy. *Nat. Chem. Biol.*, 11:728–732, 2015.
- [1095] A. Fatihi, S. Latimer, S. Schmollinger, A. Block, P.H. Dussault, W.F. Vermaas, S.S. Merchant, and G.J. Basset. A dedicated type II NADPH dehydrogenase performs the penultimate step in the biosynthesis of vitamin K₁ in *Synechocystis* and *Arabidopsis*. *Plant Cell*, 27:1730–1741, 2015.
- [1096] R. Federico, L. Ercolini, M. Laurenzi, , and R. Oxidation of acetylpolyamines by maize polyamine oxidase. *Phytochemistry*, 43:339–341, 1996.
- [1097] M. Fellner, S. Aloï, E.P. Tchesnokov, S.M. Wilbanks, and G.N. Jameson. Substrate and pH-dependent kinetic profile of 3-mercaptopyruvate dioxygenase from *Pseudomonas aeruginosa*. *Biochemistry*, 55:1362–1371, 2016.
- [1098] A.K. Felux, D. Spiteller, J. Klebensberger, and D. Schleheck. Entner-Doudoroff pathway for sulfoquinovose degradation in *Pseudomonas putida* SQ1. *Proc. Natl. Acad. Sci. USA*, 112:E4298–E4305, 2015.
- [1099] C. Feng, Y. Liu, G. Wang, Z. Deng, Q. Zhang, W. Wu, Y. Tong, C. Cheng, and Z. Chen. Crystal structures of the human RNA demethylase Alkbh5 reveal basis for substrate recognition. *J. Biol. Chem.*, 289:11571–11583, 2014.
- [1100] L. Feng, W. Wang, J. Cheng, Y. Ren, G. Zhao, C. Gao, Y. Tang, X. Liu, W. Han, X. Peng, R. Liu, and L. Wang. Genome and proteome of long-chain alkane degrading *Geobacillus thermodenitrificans* NG80-2 isolated from a deep-subsurface oil reservoir. *Proc. Natl. Acad. Sci. USA*, 104:5602–5607, 2007.
- [1101] W. Feng, M. Yonezawa, J. Ye, T. Jenuwein, and I. Grummt. PHF8 activates transcription of rRNA genes through H₃K4me3 binding and H₃K9me1/2 demethylation. *Nat. Struct. Mol. Biol.*, 17:445–450, 2010.
- [1102] D.M. Ferber and R.J. Maier. Hydrogen-ubiquinone oxidoreductase activity by the *Bradyrhizobium japonicum* membrane-bound hydrogenase. *FEMS Microbiol. Lett.*, 110:257–264, 1993.
- [1103] S.J. Ferguson. Nitrogen cycle enzymology. *Curr. Opin. Chem. Biol.*, 2:182–193, 1998.
- [1104] T. Ferguson, J.A. Soares, T. Lienard, G. Gottschalk, and J.A. Krzycki. RamA, a protein required for reductive activation of corrinoid-dependent methylamine methyltransferase reactions in methanogenic archaea. *J. Biol. Chem.*, 284:2285–2295, 2009.
- [1105] S. Fernandes, M.G. Tuohy, and P.G. Murray. Xylose reductase from the thermophilic fungus *Talaromyces emersonii*: cloning and heterologous expression of the native gene (Texr) and a double mutant (TexrK271R + N273D) with altered coenzyme specificity. *J. Biosci.*, 34:881–890, 2009.

- [1106] A. Ferrández, J.L. García, and E. Díaz. Genetic characterization and expression in heterologous hosts of the 3-(3-hydroxyphenyl)propionate catabolic pathway of *Escherichia coli* K-12. *J. Bacteriol.*, 179:2573–2581, 1997.
- [1107] A. Ferrandez, B. Minambres, B. Garcia, E.R. Olivera, J.M. Luengo, J.L. Garcia, and E. Diaz. Catabolism of phenylacetic acid in *Escherichia coli*. Characterization of a new aerobic hybrid pathway. *J. Biol. Chem.*, 273:25974–25986, 1998.
- [1108] M. Ferraroni, J. Seifert, V.M. Travkin, M. Thiel, S. Kaschabek, A. Scozzafava, L. Golovleva, M. Schlomann, and F. Briganti. Crystal structure of the hydroxyquinol 1,2-dioxygenase from *Nocardioides simplex* 3E, a key enzyme involved in polychlorinated aromatics biodegradation. *J. Biol. Chem.*, 280:21144–21154, 2005.
- [1109] G.C. Ferreira and H.A. Dailey. Mouse protoporphyrinogen oxidase. Kinetic parameters and demonstration of inhibition by bilirubin. *Biochem. J.*, 250:597–603, 1988.
- [1110] N. Lopes Ferreira, D. Labbe, F. Monot, F. Fayolle-Guichard, and C.W. Greer. Genes involved in the methyl *tert*-butyl ether (MTBE) metabolic pathway of *Mycobacterium austroafricanum* IFP 2012. *Microbiology*, 152:1361–1374, 2006.
- [1111] S. Fetzner and F. Lingens. Microbial metabolism of quinoline and related compounds. XVIII. Purification and some properties of the molybdenum- and iron-containing quinaldic acid 4-oxidoreductase from *Serratia marcescens* 2CC-1. *Biol. Chem. Hoppe-Seyler*, 374:363–376, 1993.
- [1112] S. Fetzner, R. Mueller, and F. Lingens. Degradation of 2-chlorobenzoate by *Pseudomonas cepacia* 2CBS. *Biol. Chem. Hoppe-Seyler*, 370:1173–1182, 1989.
- [1113] S. Fetzner, R. Muller, and F. Lingens. Purification and some properties of 2-halobenzoate 1,2-dioxygenase, a two-component enzyme system from *Pseudomonas cepacia* 2CBS. *J. Bacteriol.*, 174:279–290, 1992.
- [1114] C.A. Fewson and D.J.D. Nicholas. Nitrate reductase from *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta*, 49:335–349, 1961.
- [1115] N.H. Fidge and D.S. Goodman. The enzymatic reduction of retinal to retinol in rat intestine. *J. Biol. Chem.*, 243:4372–4379, 1968.
- [1116] T.J. Fiedler, C.A. Davey, and R.E. Fenna. X-ray crystal structure and characterization of halide-binding sites of human myeloperoxidase at 1.8 Å resolution. *J. Biol. Chem.*, 275:11964–11971, 2000.
- [1117] E.N. Fielding, P.F. Widboom, and S.D. Bruner. Substrate recognition and catalysis by the cofactor-independent dioxygenase DpgC. *Biochemistry*, 46:13994–14000, 2007.
- [1118] J.H.A. Fields, A.K. Eng, W.D. Ramsden, P.W. Hochachka, and B. Weinstein. Alanopine and strombine are novel imino acids produced by a dehydrogenase found in the adductor muscle of the oyster, *Crassostrea gigas*. *Arch. Biochem. Biophys.*, 201:110–114, 1980.
- [1119] J.H.A. Fields and P.W. Hochachka. Purification and properties of alanopine dehydrogenase from the adductor muscle of the oyster, *Crassostrea gigas* (Mollusca, Bivalvia). *Eur. J. Biochem.*, 114:615–621, 1981.
- [1120] P. Figueroa, G. Leon, A. Elorza, L. Holuigue, A. Araya, and X. Jordana. The four subunits of mitochondrial respiratory complex II are encoded by multiple nuclear genes and targeted to mitochondria in *Arabidopsis thaliana*. *Plant Mol. Biol.*, 50:725–734, 2002.
- [1121] R. Figueroa-Teran, W.H. Welch, G.J. Blomquist, and C. Tittiger. Ipsdienol dehydrogenase (IDOLDH): a novel oxidoreductase important for *Ips pini* pheromone production. *Insect Biochem. Mol. Biol.*, 42:81–90, 2012.
- [1122] S. Filippini, M.M. Solinas, U. Breme, M.B. Schluter, D. Gabellini, G. Biamonti, A.L. Colombo, and L. Garofano. *Streptomyces peucetius* daunorubicin biosynthesis gene, *dnrF*: sequence and heterologous expression. *Microbiology*, 141:1007–1016, 1995.
- [1123] G.M. Fimognari and V.W. Rodwell. Substrate-competitive inhibition of bacterial mevalonate:nicotinamide-adenine dinucleotide oxidoreductase (acylating CoA). *Biochemistry*, 4:2086–2090, 1965.
- [1124] D. Fischer, C. Ebenau-Jehle, and H. Grisebach. Phytoalexin synthesis in soybean: purification and characterization of NADPH:2'-hydroxydaidzein oxidoreductase from elicitor-challenged soybean cell cultures. *Arch. Biochem. Biophys.*, 276:390–395, 1990.

- [1125] D. Fischer, K. Stich, L. Britsch, and H. Grisebach. Purification and characterization of (+)dihydroflavonol (3-hydroxyflavanone) 4-reductase from flowers of *Dahlia variabilis*. *Arch. Biochem. Biophys.*, 264:40–47, 1988.
- [1126] F. Fischer, S. Kunne, and S. Fetzner. Bacterial 2,4-dioxygenases: new members of the hydrolase-fold superfamily of enzymes functionally related to serine hydrolases. *J. Bacteriol.*, 181:5725–5733, 1999.
- [1127] R.R. Fisher and S.R. Earle. Membrane-bound pyridine dinucleotide transhydrogenases. In J. Everse, B. Anderson, and K. You, editors, *The Pyridine Nucleotide Coenzymes*, pages 279–324. The Pyridine Nucleotide Coenzymes, New York, 1982.
- [1128] J. Fishman and J. Goto. Mechanism of estrogen biosynthesis. Participation of multiple enzyme sites in placental aromatase hydroxylations. *J. Biol. Chem.*, 256:4466–4471, 1981.
- [1129] P.F. Fitzpatrick. The aromatic amino acid hydroxylases. *Adv. Enzymol. Relat. Areas Mol. Biol.*, 74:235–294, 2000.
- [1130] P.F. Fitzpatrick, A.M. Orville, A. Nagpal, and M.P. Valley. Nitroalkane oxidase, a carbanion-forming flavoprotein homologous to acyl-CoA dehydrogenase. *Arch. Biochem. Biophys.*, 433:157–165, 2005.
- [1131] J. Fliegmann, K. Furtwangler, G. Malterer, C. Cantarello, G. Schuler, J. Ebel, and A. Mithofer. Flavone synthase II (CYP93B16) from soybean (*Glycine max* L.). *Phytochemistry*, 71:508–514, 2010.
- [1132] C. Florin, T. Kohler, M. Grandguillot, and P. Plesiat. *Comamonas testosteroni* 3-ketosteroid- $\Delta^4(5\alpha)$ -dehydrogenase: gene and protein characterization. *J. Bacteriol.*, 178:3322–3330, 1996.
- [1133] B.D. Fodor, S. Kubicek, M. Yonezawa, R.J. O’Sullivan, R. Sengupta, L. Perez-Burgos, S. Opravil, K. Mechtler, G. Schotta, and T. Jenuwein. Jmjd2b antagonizes H₃K9 trimethylation at pericentric heterochromatin in mammalian cells. *Genes Dev.*, 20:1557–1562, 2006.
- [1134] N. Fonknechten, S. Chaussonnerie, S. Tricot, A. Lajus, J.R. Andreesen, N. Perchat, E. Pelletier, M. Gouyvenoux, V. Barbe, M. Salanoubat, D. Le Paslier, J. Weissenbach, G.N. Cohen, and A. Kreimeyer. *Clostridium sticklandii*, a specialist in amino acid degradation: revisiting its metabolism through its genome sequence. *BMC Genomics*, 11:555–555, 2010.
- [1135] N. Fonknechten, A. Perret, N. Perchat, S. Tricot, C. Lechaplais, D. Vallenet, C. Vergne, A. Zaparucha, D. Le Paslier, J. Weissenbach, and M. Salanoubat. A conserved gene cluster rules anaerobic oxidative degradation of L-ornithine. *J. Bacteriol.*, 191:3162–3167, 2009.
- [1136] M. Fontecave, R. Eliasson, and P. Reichard. NAD(P)H:flavin oxidoreductase of *Escherichia coli*. A ferric iron reductase participating in the generation of the free radical of ribonucleotide reductase. *J. Biol. Chem.*, 262:12325–12331, 1987.
- [1137] R.G. Forage and M.A. Foster. Glycerol fermentation in *Klebsiella pneumoniae*: functions of the coenzyme B₁₂-dependent glycerol and diol dehydratases. *J. Bacteriol.*, 149:413–419, 1982.
- [1138] E. Forchielli and R.I. Dorfman. Separation of Δ^4 -5 α - and Δ^4 -5 β -hydrogenases from rat liver homogenates. *J. Biol. Chem.*, 223:443–448, 1956.
- [1139] N. Foresi, N. Correa-Aragunde, G. Parisi, G. Calo, G. Salerno, and L. Lamattina. Characterization of a nitric oxide synthase from the plant kingdom: NO generation from the green alga *Ostreococcus tauri* is light irradiance and growth phase dependent. *Plant Cell*, 22:3816–3830, 2010.
- [1140] G. Forkmann, W. Heller, and H. Grisebach. Anthocyanin biosynthesis in flowers of *Matthiola incana* flavanone 3- and flavonoid 3'-hydroxylases. *Z. Naturforsch. C: Biosci.*, 35:691–695, 1980.
- [1141] G. Forlani, D. Scainelli, and E. Nielsen. Δ^1 -Pyrroline-5-carboxylate dehydrogenase from cultured cells of potato (purification and properties). *Plant Physiol.*, 113:1413–1418, 1997.
- [1142] H.J. Forman and J. Kennedy. Mammalian dihydroorotate dehydrogenase: physical and catalytic properties of the primary enzyme. *Arch. Biochem. Biophys.*, 191:23–31, 1978.
- [1143] F. Forneris, E. Battaglioli, A. Mattevi, and C. Binda. New roles of flavoproteins in molecular cell biology: histone demethylase LSD1 and chromatin. *FEBS J.*, 276:4304–4312, 2009.

- [1144] F. Forneris, C. Binda, M.A. Vanoni, A. Mattevi, and E. Battaglioli. Histone demethylation catalysed by LSD1 is a flavin-dependent oxidative process. *FEBS Lett.*, 579:2203–2207, 2005.
- [1145] F. Forneris, D.P. Heuts, M. Delvecchio, S. Rovida, M.W. Fraaije, and A. Mattevi. Structural analysis of the catalytic mechanism and stereoselectivity in *Streptomyces coelicolor* alditol oxidase. *Biochemistry*, 47:978–985, 2008.
- [1146] P.I. Forrester and G.M. Gaucher. m-Hydroxybenzyl alcohol dehydrogenase from *Penicillium urticae*. *Biochemistry*, 11:1108–1114, 1972.
- [1147] L.S. Forsberg, K.D. Noel, J. Box, and R.W. Carlson. Genetic locus and structural characterization of the biochemical defect in the O-antigenic polysaccharide of the symbiotically deficient *Rhizobium etli* mutant, CE166. Replacement of N-acetylquinovosamine with its hexosyl-4-ulose precursor. *J. Biol. Chem.*, 278:51347–51359, 2003.
- [1148] Z. Forsberg, A.K. Mackenzie, M. Sorlie, A.K. Rohr, R. Helland, A.S. Arvai, G. Vaaje-Kolstad, and V.G. Eijsink. Structural and functional characterization of a conserved pair of bacterial cellulose-oxidizing lytic polysaccharide monoxygenases. *Proc. Natl. Acad. Sci. USA*, 111:8446–8451, 2014.
- [1149] R.R. Forseth, S. Amaike, D. Schwenk, K.J. Affeldt, D. Hoffmeister, F.C. Schroeder, and N.P. Keller. Homologous NRPS-like gene clusters mediate redundant small-molecule biosynthesis in *Aspergillus flavus*. *Angew. Chem. Int. Ed. Engl.*, 52:1590–1594, 2013.
- [1150] K. Forslund, M. Morant, B. Jørgensen, C.E. Olsen, E. Asamizu, S. Sato, S. Tabata, and S. Bak. Biosynthesis of the nitrile glucosides rhodiocyanoside A and D and the cyanogenic glucosides lotaustralin and linamarin in *Lotus japonicus*. *Plant Physiol.*, 135:71–84, 2004.
- [1151] S. P. Foster and W. L. Roelofs. Biosynthesis of a monoene and a conjugated diene sex pheromone component of the lightbrown apple moth by 11 desaturation. *Experientia*, 46:269–273, 1990.
- [1152] J.C. Fothergill and J.R. Guest. Catabolism of L-lysine by *Pseudomonas aeruginosa*. *J. Gen. Microbiol.*, 99:139–155, 1977.
- [1153] B.G. Fox, J. Shanklin, J. Ai, T.M. Loehr, and J. Sanders-Loehr. Resonance Raman evidence for an Fe-O-Fe center in stearyl-ACP desaturase. Primary sequence identity with other diiron-oxo proteins. *Biochemistry*, 33:12776–12786, 1994.
- [1154] B.S. Fox and C.T. Walsh. Mercuric reductase. Purification and characterization of a transposon-encoded flavoprotein containing an oxidation-reduction-active disulfide. *J. Biol. Chem.*, 257:2498–2503, 1982.
- [1155] B.S. Fox and C.T. Walsh. Mercuric reductase - homology to glutathione-reductase and lipoamide dehydrogenase - iodoacetamide alkylation and sequence of the active-site peptide. *Biochemistry*, 22:4082–4088, 1983.
- [1156] J.A. Fox, D.J. Livingston, W.H. Orme-Johnson, and C.T. Walsh. 8-Hydroxy-5-deazaflavin-reducing hydrogenase from *Methanobacterium thermoautotrophicum*: 1. Purification and characterization. *Biochemistry*, 26:4219–4228, 1987.
- [1157] M.W. Fraaije, H.P. Roubroeks, , and W.H.J. Purification and characterization of an intracellular catalase-peroxidase from *Penicillium simplicissimum*. *Eur. J. Biochem.*, 235:192–198, 1996.
- [1158] M.W. Fraaije, C. Veeger, and W.J.H. van Berkel. Substrate specificity of flavin-dependent vanillyl-alcohol oxidase from *Penicillium simplicissimum*. Evidence for the production of 4-hydroxycinnamyl alcohols from 4-allylphenols. *Eur. J. Biochem.*, 234:271–277, 1995.
- [1159] M.W. Fraaije, J. Wu, D.P. Heuts, E.W. van Hellemond, J.H. Spelberg, and D.B. Janssen. Discovery of a thermostable Baeyer-Villiger monoxygenase by genome mining. *Appl. Microbiol. Biotechnol.*, 66:393–400, 2005.
- [1160] E.W. Frampton and W.A. Wood. Carbohydrate oxidation by *Pseudomonas fluorescens*. VI. Conversion of 2-keto-6-phosphogluconate to pyruvate. *J. Biol. Chem.*, 236:2571–2577, 1961.
- [1161] S. Franceschini, M. Fedkenheuer, N.J. Vogelaar, H.H. Robinson, P. Sobrado, and A. Mattevi. Structural insight into the mechanism of oxygen activation and substrate selectivity of flavin-dependent N-hydroxylating monoxygenases. *Biochemistry*, 51:7043–7045, 2012.
- [1162] C.A. Francis, K.L. Casciotti, and B.M. Tebo. Localization of Mn(II)-oxidizing activity and the putative multicopper oxidase, MnxG, to the exosporium of the marine *Bacillus* sp. strain SG-1. *Arch. Microbiol.*, 178:450–456, 2002.

- [1163] K. Francis and G. Gadda. Kinetic evidence for an anion binding pocket in the active site of nitronate monooxygenase. *Bioorg. Chem.*, 37:167–172, 2009.
- [1164] K. Francis, B. Russell, and G. Gadda. Involvement of a flavosemiquinone in the enzymatic oxidation of nitroalkanes catalyzed by 2-nitropropane dioxygenase. *J. Biol. Chem.*, 280:5195–5204, 2005.
- [1165] R.A. Frank, A.J. Price, F.D. Northrop, R.N. Perham, and B.F. Luisi. Crystal structure of the E1 component of the *Escherichia coli* 2-oxoglutarate dehydrogenase multienzyme complex. *J. Mol. Biol.*, 368:639–651, 2007.
- [1166] R. Franke, J.M. Humphreys, M.R. Hemm, J.W. Denault, M.O. Ruegger, J.C. Cusumano, and C. Chapple. The *Arabidopsis* REF8 gene encodes the 3-hydroxylase of phenylpropanoid metabolism. *Plant J.*, 30:33–45, 2002.
- [1167] N. Frankenberg, K. Mukougawa, T. Kohchi, and J.C. Lagarias. Functional genomic analysis of the HY2 family of ferredoxin-dependent bilin reductases from oxygenic photosynthetic organisms. *Plant Cell*, 13:965–978, 2001.
- [1168] C.V. Franklund, S.F. Baron, and P.B. Hylemon. Characterization of the *baiH* gene encoding a bile acid-inducible NADH:flavin oxidoreductase from *Eubacterium* sp. strain VPI 12708. *J. Bacteriol.*, 175:3002–3012, 1993.
- [1169] P.D. Fraser, H. Linden, and G. Sandmann. Purification and reactivation of recombinant *Synechococcus* phytoene desaturase from an overexpressing strain of *Escherichia coli*. *Biochem. J.*, 291:687–692, 1993.
- [1170] P.D. Fraser, N. Misawa, H. Linden, S. Yamano, K. Kobayashi, and G. Sandmann. Expression in *Escherichia coli*, purification, and reactivation of the recombinant *Erwinia uredovora* phytoene desaturase. *J. Biol. Chem.*, 267:19891–19895, 1992.
- [1171] P.D. Fraser, Y. Miura, and N. Misawa. *In vitro* characterization of astaxanthin biosynthetic enzymes. *J. Biol. Chem.*, 272:6128–6135, 1997.
- [1172] P.D. Fraser, H. Shimada, and N. Misawa. Enzymic confirmation of reactions involved in routes to astaxanthin formation, elucidated using a direct substrate *in vitro* assay. *Eur. J. Biochem.*, 252:229–236, 1998.
- [1173] J. Frebortova, K. Matsushita, H. Arata, and O. Adachi. Intramolecular electron transport in quinoprotein alcohol dehydrogenase of *Acetobacter methanolicus*: a redox-titration stud. *Biochim. Biophys. Acta*, 1363:24–34, 1998.
- [1174] W. Freudenberg, K. Konig, and J. R. Andreesen. Nicotine dehydrogenase from *Arthrobacter oxidans*: A molybdenum-containing hydroxylase. *FEMS Microbiology Letters*, 52:13–18, 1988.
- [1175] M. Frey, P. Chomet, E. Glawischnig, C. Stettner, S. Grün, A. Winklmaier, W. Eisenreich, A. Bacher, R.B. Meeley, S.P. Briggs, K. Simcox, and A. Gierl. Analysis of a chemical plant defense mechanism in grasses. *Science*, 277:696–699, 1997.
- [1176] M. Frey, M. Rothe, A.F.V. Wagner, and J. Knappe. Adenosylmethionine-dependent synthesis of the glycyl radical in pyruvate formate-lyase by abstraction of the glycine C-2 *pro-S* hydrogen atom. *J. Biol. Chem.*, 269:12432–12437, 1994.
- [1177] M. Frey, K. Schmauder, I. Pateraki, and O. Spring. Biosynthesis of eupatolide-A metabolic route for sesquiterpene lactone formation involving the P450 enzyme CYP71DD6. *ACS Chem. Biol.*, 13:1536–1543, 2018.
- [1178] P.A. Frey. Radical mechanisms in enzymatic catalysis. *Annu. Rev. Biochem.*, 70:121–148, 2001.
- [1179] J. Fricke, F. Blei, and D. Hoffmeister. Enzymatic synthesis of psilocybin. *Angew. Chem. Int. Ed. Engl.*, 56:12352–12355, 2017.
- [1180] C. Frieden. L-Glutamate dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 3–24. Academic Press, New York, 2nd edition, 1963.
- [1181] J. Friedman, L. Lad, H. Li, A. Wilks, and T.L. Poulos. Structural basis for novel δ -regioselective heme oxygenation in the opportunistic pathogen *Pseudomonas aeruginosa*. *Biochemistry*, 43:5239–5245, 2004.
- [1182] J.E. Friedman, J.A. Watson, Lam Jr., Rokita D.W., and S.E. Iodotyrosine deiodinase is the first mammalian member of the NADH oxidase/flavin reductase superfamily. *J. Biol. Chem.*, 281:2812–2819, 2006.
- [1183] P.A. Friedman, A.H. Kappelman, and S. Kaufman. Partial purification and characterization of tryptophan hydroxylase from rabbit hindbrain. *J. Biol. Chem.*, 247:4165–4173, 1972.

- [1184] S. Friedman and S. Kaufman. 3,4-Dihydroxyphenylethylamine β -hydroxylase. Physical properties, copper content, and role of copper in the catalytic activity. *J. Biol. Chem.*, 240:4763–4773, 1965.
- [1185] H.C. Friedmann and B. Vennesland. Purification and properties of dihydroorotic acid dehydrogenase. *J. Biol. Chem.*, 233:1398–1406, 1958.
- [1186] H.C. Friedmann and B. Vennesland. Crystalline dihydroorotic dehydrogenase. *J. Biol. Chem.*, 235:1526–1532, 1960.
- [1187] C.G. Friedrich, D. Rother, F. Bardischewsky, A. Quentmeier, and J. Fischer. Oxidation of reduced inorganic sulfur compounds by bacteria: emergence of a common mechanism. *Appl. Environ. Microbiol.*, 67:2873–2882, 2001.
- [1188] N.U. Frigaard, J.A. Maresca, C.E. Yunker, A.D. Jones, and D.A. Bryant. Genetic manipulation of carotenoid biosynthesis in the green sulfur bacterium *Chlorobium tepidum*. *J. Bacteriol.*, 186:5210–5220, 2004.
- [1189] N.A. Frigerio and H.A. Harbury. Preparation and some properties of crystalline glycolic acid oxidase of spinach. *J. Biol. Chem.*, 231:135–157, 1958.
- [1190] W.R. Frisell and C.G. MacKenzie. Separation and purification of sarcosine dehydrogenase and dimethylglycine dehydrogenase. *J. Biol. Chem.*, 237:94–98, 1962.
- [1191] M. Fritz, H. Lokstein, D. Hackenberg, R. Welti, M. Roth, U. Zähringer, M. Fulda, W. Hellmeyer, C. Ott, F.P. Wolter, and E. Heinz. Channeling of eukaryotic diacylglycerol into the biosynthesis of plastidial phosphatidylglycerol. *J. Biol. Chem.*, 282:4613–4625, 2007.
- [1192] P. Fritzson. Properties and assay of dihydrouracil dehydrogenase of rat liver. *J. Biol. Chem.*, 235:719–725, 1960.
- [1193] D.S. Froese, J. Kopec, E. Rembeza, G.A. Bezerra, A.E. Oberholzer, T. Suormala, S. Lutz, R. Chalk, O. Borkowska, M.R. Baumgartner, and W.W. Yue. Structural basis for the regulation of human 5,10-methylenetetrahydrofolate reductase by phosphorylation and *S*-adenosylmethionine inhibition. *Nat. Commun.*, 9:2261–2261, 2018.
- [1194] M. Frommhagen, S. Sforza, A.H. Westphal, J. Visser, S.W. Hinz, M.J. Koetsier, W.J. van Berkel, H. Gruppen, and M.A. Kabel. Discovery of the combined oxidative cleavage of plant xylan and cellulose by a new fungal polysaccharide monooxygenase. *Biotechnol. Biofuels*, 8:101–101, 2015.
- [1195] R. Frommolt, R. Goss, and C. Wilhelm. Erratum Report. The de-epoxidase and epoxidase reactions of *Mantoniella squamata* (Prasinophyceae) exhibit different substrate-specific reaction kinetics compared to spinach. *Planta*, 213:492–492, 2001.
- [1196] R. Frommolt, R. Goss, and C. Wilhelm. The de-epoxidase and epoxidase reactions of *Mantoniella squamata* (Prasinophyceae) exhibit different substrate-specific reaction kinetics compared to spinach. *Planta*, 213:446–456, 2001.
- [1197] S. Frusciante, G. Diretto, M. Bruno, P. Ferrante, M. Pietrella, A. Prado-Cabrero, A. Rubio-Moraga, P. Beyer, L. Gomez-Gomez, S. Al-Babili, and G. Giuliano. Novel carotenoid cleavage dioxygenase catalyzes the first dedicated step in saffron crocin biosynthesis. *Proc. Natl. Acad. Sci. USA*, 111:12246–12251, 2014.
- [1198] R.B. Frydman, M.L. Tomaro, and B. Frydman. Pyrroloxygenase: its action on tryptophan-containing enzymes and peptides. *Biochim. Biophys. Acta*, 284:80–89, 1972.
- [1199] G. Fu, H. Yuan, C. Li, C.D. Lu, G. Gadda, and I.T. Weber. Conformational changes and substrate recognition in *Pseudomonas aeruginosa* D-arginine dehydrogenase. *Biochemistry*, 49:8535–8545, 2010.
- [1200] G. Fu, H. Yuan, S. Wang, G. Gadda, and I.T. Weber. Atomic-resolution structure of an N5 flavin adduct in D-arginine dehydrogenase. *Biochemistry*, 50:6292–6294, 2011.
- [1201] S.L. Fuenmayor, M. Wild, A.L. Boyes, and P.A. Williams. A gene cluster encoding steps in conversion of naphthalene to gentisate in *Pseudomonas* sp. strain U2. *J. Bacteriol.*, 180:2522–2530, 1998.
- [1202] I. Fujii, Y. Ebizuka, and U. Sankawa. A novel anthraquinone ring cleavage enzyme from *Aspergillus terreus*. *J. Biochem. (Tokyo)*, 103:878–883, 1988.
- [1203] T. Fujii and T. Kaneda. Purification and properties of NADH/NADPH-dependent *p*-hydroxybenzoate hydroxylase from *Corynebacterium cyclohexanicum*. *Eur. J. Biochem.*, 147:97–104, 1985.

- [1204] T. Fujii, T. Narita, H. Agematu, N. Agata, and K. Isshiki. Cloning and characterization of *pcd* encoding Δ^1 -piperidine-6-carboxylate dehydrogenase from *Flavobacterium lutescens* IFO3084. *J. Biochem.*, 128:975–982, 2000.
- [1205] M. Fujioka, Y. Morino, and H. Wada. Metabolism of phenylalanine (*Achromobacter eurydice*). III. Phenylacetaldehyde dehydrogenase. *Methods Enzymol.*, 17A:593–596, 1970.
- [1206] M. Fujioka and Y. Nakatani. Saccharopine dehydrogenase. Interaction with substrate analogues. *Eur. J. Biochem.*, 25:301–307, 1972.
- [1207] M. Fujioka and H. Wada. The bacterial oxidation of indole. *Biochim. Biophys. Acta*, 158:70–78, 1968.
- [1208] H. Fujisawa and O. Hayaishi. Protocatechuate 3,4-dioxygenase. I. Crystallization and characterization. *J. Biol. Chem.*, 243:2673–2681, 1968.
- [1209] H. Fujisawa, S. Nagata, E.K. Chowdhury, M. Matsumoto, and H. Misono. Cloning and sequencing of the serine dehydrogenase gene from *Agrobacterium tumefaciens*. *Biosci. Biotechnol. Biochem.*, 66:1137–1139, 2002.
- [1210] H. Fujisawa, S. Nagata, and H. Misono. Characterization of short-chain dehydrogenase/reductase homologues of *Escherichia coli* (YdfG) and *Saccharomyces cerevisiae* (YMR226C). *Biochim. Biophys. Acta*, 1645:89–94, 2003.
- [1211] M. Fujita, D.R. Gang, L.B. Davin, and N.G. Lewis. Recombinant pinoresinol-lariciresinol reductases from western red cedar (*Thuja plicata*) catalyze opposite enantiospecific conversions. *J. Biol. Chem.*, 274:618–627, 1999.
- [1212] S. Fujita, T. Ohnishi, B. Watanabe, T. Yokota, S. Takatsuto, S. Fujioka, S. Yoshida, K. Sakata, and M. Mizutani. *Arabidopsis* CYP90B1 catalyses the early C-22 hydroxylation of C₂₇, C₂₈ and C₂₉ sterols. *Plant J.*, 45:765–774, 2006.
- [1213] T. Fujita and G.J. Mannering. Differences in soluble P-450 hemoproteins from livers of rats treated with phenobarbital and 3-methylcholanthrene. *Chem. Biol. Interact.*, 3:264–265, 1971.
- [1214] Y. Fujita, H. Matsumoto, Y. Takahashi, and H. Matsubara. Identification of a *nifDK*-like gene (ORF467) involved in the biosynthesis of chlorophyll in the cyanobacterium *Plectonema boryanum*. *Plant Cell Physiol.*, 34:305–314, 1993.
- [1215] Y. Fujita, R. Ramaley, and E. Freese. Location and properties of glucose dehydrogenase in sporulating cells and spores of *Bacillus subtilis*. *J. Bacteriol.*, 132:282–293, 1977.
- [1216] K. Fujiwara, K. Okamura-Ikeda, and Y. Motokawa. Mechanism of the glycine cleavage reaction. Further characterization of the intermediate attached to H-protein and of the reaction catalyzed by T-protein. *J. Biol. Chem.*, 259:10664–10668, 1984.
- [1217] Y. Fujiwara and M. Ito. Molecular cloning and characterization of a *Perilla frutescens* cytochrome P450 enzyme that catalyzes the later steps of perillaldehyde biosynthesis. *Phytochemistry*, 134:26–37, 2017.
- [1218] M. Fukuchi-Mizutani, Y. Tasaka, Y. Tanaka, T. Ashikari, T. Kusumi, and N. Murata. Characterization of δ A9 acyl-lipid desaturase homologues from *Arabidopsis thaliana*. *Plant Cell Physiol.*, 39:247–253, 1998.
- [1219] E. Fukuda, H. Kino, H. Matsuzawa, and T. Wakagi. Role of a highly conserved YPITP motif in 2-oxoacid:ferredoxin oxidoreductase: heterologous expression of the gene from *Sulfolobus* sp. strain 7, and characterization of the recombinant and variant enzymes. *Eur. J. Biochem.*, 268:5639–5646, 2001.
- [1220] E. Fukuda and T. Wakagi. Substrate recognition by 2-oxoacid:ferredoxin oxidoreductase from *Sulfolobus* sp. strain 7. *Biochim. Biophys. Acta*, 1597:74–80, 2002.
- [1221] H. Fukuda, T. Ogawa, K. Ishihara, T. Fujii, K. Nagahama, T. Omata, Y. Inoue, S. Tanase, and Y. Morino. Molecular cloning in *Escherichia coli*, expression, and nucleotide sequence of the gene for the ethylene-forming enzyme of *Pseudomonas syringae* pv. *phaseolicola* PK2. *Biochem. Biophys. Res. Commun.*, 188:826–832, 1992.
- [1222] H. Fukuda, T. Ogawa, M. Tazaki, K. Nagahama, T. Fujii, S. Tanase, and Y. Morino. Two reactions are simultaneously catalyzed by a single enzyme: the arginine-dependent simultaneous formation of two products, ethylene and succinate, from 2-oxoglutarate by an enzyme from *Pseudomonas syringae*. *Biochem. Biophys. Res. Commun.*, 188:483–489, 1992.
- [1223] M. Fukuda, Y. Yasukochi, Y. Kikuchi, Y. Nagata, K. Kimbara, H. Horiuchi, M. Takagi, and K. Yano. Identification of the *bphA* and *bphB* genes of *Pseudomonas* sp. strains KKS102 involved in degradation of biphenyl and polychlorinated biphenyls. *Biochem. Biophys. Res. Commun.*, 202:850–856, 1994.

- [1224] Y. Fukumori and T. Yamanaka. A high-potential nonheme iron protein (HiPIP)-linked, thiosulfate-oxidizing enzyme derived from *Chromatium vinosum*. *Curr. Microbiol.*, 3:117–120, 1979.
- [1225] Y. Fukumori and T. Yamanaka. Flavocytochrome *c* of *Chromatium vinosum*. Some enzymatic properties and subunit structure. *J. Biochem.*, 85:1405–1414, 1979.
- [1226] E.O. Fukushima, H. Seki, K. Ohyama, E. Ono, N. Umemoto, M. Mizutani, K. Saito, and T. Muranaka. CYP716A subfamily members are multifunctional oxidases in triterpenoid biosynthesis. *Plant Cell Physiol.*, 52:2050–2061, 2011.
- [1227] H. Fukushima, G.F. Grinstead, and J.L. Gaylor. Total enzymic synthesis of cholesterol from lanosterol. Cytochrome *b*₅-dependence of 4-methyl sterol oxidase. *J. Biol. Chem.*, 256:4822–4826, 1981.
- [1228] A.J. Fulco and K. Bloch. Cofactor requirements for the formation of Δ^9 -unsaturated fatty acids in *Mycobacterium phlei*. *J. Biol. Chem.*, 239:993–997, 1964.
- [1229] C. Funk and R. Croteau. Induction and characterization of a cytochrome *P*-450-dependent camphor hydroxylase in tissue cultures of common sage (*Salvia officinalis*). *Plant Physiol.*, 101:1231–1237, 1993.
- [1230] C. Funk and R. Croteau. Diterpenoid resin acid biosynthesis in conifers: characterization of two cytochrome P450-dependent monooxygenases and an aldehyde dehydrogenase involved in abietic acid biosynthesis. *Arch. Biochem. Biophys.*, 308:258–266, 1994.
- [1231] C. Funk, A.E. Koepp, and R. Croteau. Catabolism of camphor in tissue cultures and leaf disks of common sage (*Salvia officinalis*). *Arch. Biochem. Biophys.*, 294:306–313, 1992.
- [1232] C. Funk, E. Lewinsohn, B.S. Vogel, C.L. Steele, and R. Croteau. Regulation of oleoresinosis in grand fir (*Abies grandis*) (coordinate induction of monoterpene and diterpene cyclases and two cytochrome P450-dependent diterpenoid hydroxylases by stem wounding). *Plant Physiol.*, 106:999–1005, 1994.
- [1233] E.S. Furfine and R.H. Abeles. Intermediates in the conversion of 5'-*S*-methylthioadenosine to methionine in *Klebsiella pneumoniae*. *J. Biol. Chem.*, 263:9598–9606, 1988.
- [1234] C. Furster, T. Bergman, and K. Wikvall. Biochemical characterization of a truncated form of CYP27A purified from rabbit liver mitochondria. *Biochem. Biophys. Res. Commun.*, 263:663–666, 1999.
- [1235] P.G. Furtmuller, U. Burner, and C. Obinger. Reaction of myeloperoxidase compound I with chloride, bromide, iodide, and thiocyanate. *Biochemistry*, 37:17923–17930, 1998.
- [1236] M. Furuebisu, S. Deguchi, and K. Okuda. Identification of cortisone 5 β -reductase as Δ^4 -3-ketosteroid 5 β -reductase. *Biochim. Biophys. Acta*, 912:110–114, 1987.
- [1237] A. Furuichi, H. Akita, H. Matsukura, T. Oishi, and K. Horikoshi. Purification and properties of an asymmetric reduction enzyme of 2-methyl-3-oxobutyrate in baker's yeast. *Agric. Biol. Chem.*, 49:2563–2570, 1985.
- [1238] A. Furuichi, H. Akita, H. Matsukura, T. Oishi, and K. Horikoshi. Purification and properties of an asymmetric reduction of diethyl 2-methyl-3-oxosuccinate in *Saccharomyces fermentati*. *Agric. Biol. Chem.*, 51:293–299, 1987.
- [1239] M. Furuichi, N. Suzuki, B. Dhakshnamoorthy, H. Minagawa, R. Yamagishi, Y. Watanabe, Y. Goto, H. Kaneko, Y. Yoshida, H. Yagi, I. Waga, P.K. Kumar, and H. Mizuno. X-ray structures of *Aerococcus viridans* lactate oxidase and its complex with D-lactate at pH 4.5 show an α -hydroxyacid oxidation mechanism. *J. Mol. Biol.*, 378:436–446, 2008.
- [1240] T. Furuya, S. Hirose, H. Osanai, H. Semba, and K. Kino. Identification of the monooxygenase gene clusters responsible for the regioselective oxidation of phenol to hydroquinone in mycobacteria. *Appl. Environ. Microbiol.*, 77:1214–1220, 2011.
- [1241] M. Futai. Membrane D-lactate dehydrogenase from *Escherichia coli*. Purification and properties. *Biochemistry*, 12:2468–2474, 1973.
- [1242] A.B. Gaal and H.Y. Neujahr. Maleylacetate reductase from *Trichosporon cutaneum*. *Biochem. J.*, 185:783–786, 1980.
- [1243] A.B. Gaal and H.Y. Neujahr. Induction of phenol-metabolizing enzymes in *Trichosporon cutaneum*. *Arch. Microbiol.*, 130:54–58, 1981.

- [1244] A. Gaber, M. Tamoi, T. Takeda, Y. Nakano, and S. Shigeoka. NADPH-dependent glutathione peroxidase-like proteins (Gpx-1, Gpx-2) reduce unsaturated fatty acid hydroperoxides in *Synechocystis* PCC 6803. *FEBS Lett.*, 499:32–36, 2001.
- [1245] A. Gaber, K. Yoshimura, M. Tamoi, T. Takeda, Y. Nakano, and S. Shigeoka. Induction and functional analysis of two reduced nicotinamide adenine dinucleotide phosphate-dependent glutathione peroxidase-like proteins in *Synechocystis* PCC 6803 during the progression of oxidative stress. *Plant Physiol.*, 136:2855–2861, 2004.
- [1246] K. Gable, S. Garton, J.A. Napier, and T.M. Dunn. Functional characterization of the *Arabidopsis thaliana* orthologue of Tsc13p, the enoyl reductase of the yeast microsomal fatty acid elongating system. *J. Exp. Bot.*, 55:543–545, 2004.
- [1247] D. Gachotte, R. Barbuch, J. Gaylor, E. Nickel, and M. Bard. Characterization of the *Saccharomyces cerevisiae* ERG26 gene encoding the C-3 sterol dehydrogenase (C-4 decarboxylase) involved in sterol biosynthesis. *Proc. Natl. Acad. Sci. USA*, 95:13794–13799, 1998.
- [1248] D. Gachotte, S.E. Sen, J. Eckstein, R. Barbuch, M. Krieger, B.D. Ray, and M. Bard. Characterization of the *Saccharomyces cerevisiae* ERG27 gene encoding the 3-keto reductase involved in C-4 sterol demethylation. *Proc. Natl. Acad. Sci. USA*, 96:12655–12660, 1999.
- [1249] G. Gadda. Kinetic mechanism of choline oxidase from *Arthrobacter globiformis*. *Biochim. Biophys. Acta*, 1646:112–118, 2003.
- [1250] G. Gadda and K. Francis. Nitronate monooxygenase, a model for anionic flavin semiquinone intermediates in oxidative catalysis. *Arch. Biochem. Biophys.*, 493:53–61, 2010.
- [1251] G. Gadda and E.E. McAllister-Wilkins. Cloning, expression, and purification of choline dehydrogenase from the moderate halophile *Halomonas elongata*. *Appl. Environ. Microbiol.*, 69:2126–2132, 2003.
- [1252] G. Gadda, N.L. Powell, and P. Menon. The trimethylammonium headgroup of choline is a major determinant for substrate binding and specificity in choline oxidase. *Arch. Biochem. Biophys.*, 430:264–273, 2004.
- [1253] G. Gäde. Purification and properties of taupine dehydrogenase from the shell adductor muscle of the ormer, *Haliotis lamellosa*. *Eur. J. Biochem.*, 160:311–318, 1986.
- [1254] Z. Gai, X. Wang, X. Liu, C. Tai, H. Tang, X. He, G. Wu, Z. Deng, and P. Xu. The genes coding for the conversion of carbazole to catechol are flanked by IS6100 elements in *Sphingomonas* sp. strain XLDN2-5. *PLoS One*, 5:e10018–e10018, 2010.
- [1255] C.G. Gaines, G.S. Byng, R.J. Whitaker, and R.A. Jensen. L-Tyrosine regulation and biosynthesis via aroenate dehydrogenase in suspension-cultured cells of *Nicotiana glauca* L. *Planta*, 156:233–240, 1982.
- [1256] B. Galan, E. Diaz, M.A. Prieto, and J.L. Garcia. Functional analysis of the small component of the 4-hydroxyphenylacetate 3-monooxygenase of *Escherichia coli* W: a prototype of a new Flavin:NAD(P)H reductase subfamily. *J. Bacteriol.*, 182:627–636, 2000.
- [1257] E.F. Gale. Formic dehydrogenase of *Bacterium coli*: its inactivation by oxygen and its protection in the bacterial cell. *Biochem. J.*, 33:1012–1027, 1939.
- [1258] S.C. Gallagher, R. Cammack, and H. Dalton. Alkene monooxygenase from *Nocardia corallina* B-276 is a member of the class of dinuclear iron proteins capable of stereospecific epoxyoxygenation reactions. *Eur. J. Biochem.*, 247:635–641, 1997.
- [1259] A. Gallego-Garcia, A.J. Monera-Girona, E. Pajares-Martinez, E. Bastida-Martinez, R. Perez-Castano, A.A. Iniesta, M. Fontes, S. Padmanabhan, and M. Elias-Arnanz. A bacterial light response reveals an orphan desaturase for human plasmalogen synthesis. *Science*, 366:128–132, 2019.
- [1260] I. Galli, G. Musci, and M.C. Bonaccorsi di Patti. Sequential reconstitution of copper sites in the multicopper oxidase CueO. *J. Biol. Inorg. Chem.*, 9:90–95, 2004.
- [1261] T. Galliard and P.K. Stumpf. Fat metabolism in higher plants. 30. Enzymatic synthesis of ricinoleic acid by a microsomal preparation from developing *Ricinus communis* seeds. *J. Biol. Chem.*, 241:5806–5812, 1966.
- [1262] C. Gallus and B. Schink. Anaerobic degradation of pimelate by newly isolated denitrifying bacteria. *Microbiology*, 140:409–416, 1994.

- [1263] P. Galuszka, I. Frebort, M. Sebel, S. Jacobsen, and P. Pec. Cytokinin oxidase or dehydrogenase? Mechanism of cytokinin degradation in plants. *Eur. J. Biochem.*, 268:450–461, 2001.
- [1264] F. Galván, A.J. Márquez, and J.M. Vega. Purification and molecular properties of ferredoxin-glutamate synthase from *Chlamydomonas reinhardtii*. *Planta*, 162:180–187, 1984.
- [1265] S.R. Gama, M. Vogt, T. Kalina, K. Hupp, F. Hammerschmidt, K. Pallitsch, and D.L. Zechel. An oxidative pathway for microbial utilization of methylphosphonic acid as a phosphate source. *ACS Chem. Biol.*, 14:735–741, 2019.
- [1266] O.L. Gamborg. Aromatic metabolism in plants. III. Quinate dehydrogenase from mung bean cell suspension cultures. *Biochim. Biophys. Acta*, 128:483–491, 1966.
- [1267] O.L. Gamborg and F.W. Keeley. Aromatic metabolism in plants. I. A study of the prephenate dehydrogenase from bean plants. *Biochim. Biophys. Acta*, 115:65–72, 1966.
- [1268] C. Gancedo, J.M. Gancedo, and A. Sols. Glycerol metabolism in yeasts. Pathways of utilization and production. *Eur. J. Biochem.*, 5:165–172, 1968.
- [1269] A. Gandin, C. Duffes, D.A. Day, and A.B. Cousins. The absence of alternative oxidase AOX1A results in altered response of photosynthetic carbon assimilation to increasing CO₂ in *Arabidopsis thaliana*. *Plant Cell Physiol.*, 53:1627–1637, 2012.
- [1270] J. Gao, I. Ajjawi, A. Manoli, A. Sawin, C. Xu, J.E. Froehlich, R.L. Last, and C. Benning. FATTY ACID DESATURASE4 of *Arabidopsis* encodes a protein distinct from characterized fatty acid desaturases. *Plant J.*, 60:832–839, 2009.
- [1271] S. Gao, G. von Schumann, and J. Stöckigt. A newly-detected reductase from *Rauvolfia* closes a gap in the biosynthesis of the antiarrhythmic alkaloid ajmaline. *Planta Med.*, 68:906–911, 2002.
- [1272] E. Garcin, X. Vernede, E.C. Hatchikian, A. Volbeda, M. Frey, and J.C. Fontecilla-Camps. The crystal structure of a reduced [NiFeSe] hydrogenase provides an image of the activated catalytic center. *Structure*, 7:557–566, 1999.
- [1273] P.R. Gardner, G. Costantino, and A.L. Salzman. Constitutive and adaptive detoxification of nitric oxide in *Escherichia coli*. Role of nitric-oxide dioxygenase in the protection of aconitase. *J. Biol. Chem.*, 273:26528–26533, 1998.
- [1274] P.R. Gardner, A.M. Gardner, L.A. Martin, and A.L. Salzman. Nitric oxide dioxygenase: an enzymic function for flavo-hemoglobin. *Proc. Natl. Acad. Sci. USA*, 95:10378–10383, 1998.
- [1275] M. Gargouri, J. Chaudiere, C. Manigand, C. Mauge, K. Bathany, J.M. Schmitter, and B. Gallois. The epimerase activity of anthocyanidin reductase from *Vitis vinifera* and its regiospecific hydride transfers. *Biol. Chem.*, 391:219–227, 2010.
- [1276] M. Gargouri, C. Manigand, C. Mauge, T. Granier, B. Langlois d'Estaintot, O. Cala, I. Pianet, K. Bathany, J. Chaudiere, and B. Gallois. Structure and epimerase activity of anthocyanidin reductase from *Vitis vinifera*. *Acta Crystallogr. D Biol. Crystallogr.*, 65:989–1000, 2009.
- [1277] A. Garrido-Pertierra and R.A. Cooper. Identification and purification of distinct isomerase and decarboxylase enzymes involved in the 4-hydroxyphenylacetate pathway of *Escherichia coli*. *Eur. J. Biochem.*, 117:581–584, 1981.
- [1278] U. Garscha and E. Oliw. *Pichia* expression and mutagenesis of 7,8-linoleate diol synthase change the dioxygenase and hydroperoxide isomerase. *Biochem. Biophys. Res. Commun.*, 373:579–583, 2008.
- [1279] U. Garscha and E.H. Oliw. Leucine/valine residues direct oxygenation of linoleic acid by (10*R*)- and (8*R*)-dioxygenases: expression and site-directed mutagenesis of (10*R*)-dioxygenase with epoxyalcohol synthase activity. *J. Biol. Chem.*, 284:13755–13765, 2009.
- [1280] S.G. Gattis, H.S. Chung, M.S. Trent, and C.R. Raetz. The origin of 8-amino-3,8-dideoxy-D-manno-octulosonic acid (Kdo8N) in the lipopolysaccharide of *Shewanella oneidensis*. *J. Biol. Chem.*, 288:9216–9225, 2013.
- [1281] S. Gatzek, G.L. Wheeler, and N. Smirnov. Antisense suppression of L-galactose dehydrogenase in *Arabidopsis thaliana* provides evidence for its role in ascorbate synthesis and reveals light modulated L-galactose synthesis. *Plant J.*, 30:541–553, 2002.

- [1282] P.Z. Gatzeva-Topalova, A.P. May, and M.C. Sousa. Crystal structure of *Escherichia coli* ArnA (PmrI) decarboxylase domain. A key enzyme for lipid A modification with 4-amino-4-deoxy-L-arabinose and polymyxin resistance. *Biochemistry*, 43:13370–13379, 2004.
- [1283] P.Z. Gatzeva-Topalova, A.P. May, and M.C. Sousa. Structure and mechanism of ArnA: conformational change implies ordered dehydrogenase mechanism in key enzyme for polymyxin resistance. *Structure*, 13:929–942, 2005.
- [1284] A.E. Gau, A. Heindl, A. Nodop, U. Kahmann, and E.K. Pistorius. L-Amino acid oxidases with specificity for basic L-amino acids in cyanobacteria. *Z. Naturforsch. C*, 62:273–284, 2007.
- [1285] R.W. Gaugler and O. Gabriel. Biological mechanisms involved in the formation of deoxy sugars. VII. Biosynthesis of 6-deoxy-L-talose. *J. Biol. Chem.*, 248:6041–6049, 1973.
- [1286] J.P. Gaut, G.C. Yeh, H.D. Tran, J. Byun, J.P. Henderson, G.M. Richter, M.L. Brennan, A.J. Lulis, A. Belaouaj, R.S. Hotchkiss, and J.W. Heinecke. Neutrophils employ the myeloperoxidase system to generate antimicrobial brominating and chlorinating oxidants during sepsis. *Proc. Natl. Acad. Sci. USA*, 98:11961–11966, 2001.
- [1287] J.J. Gauthier and S.C. Rittenberg. The metabolism of nicotinic acid. I. Purification and properties of 2,5-dihydropyridine oxygenase from *Pseudomonas putida* N-9. *J. Biol. Chem.*, 246:3737–3742, 1971.
- [1288] J.J. Gauthier and S.C. Rittenberg. The metabolism of nicotinic acid. II. 2,5-Dihydropyridine oxidation, product formation, and oxygen 18 incorporation. *J. Biol. Chem.*, 246:3743–3748, 1971.
- [1289] J.M. Gavaret, H.J. Cahnmann, and J. Nunez. Thyroid hormone synthesis in thyroglobulin. The mechanism of the coupling reaction. *J. Biol. Chem.*, 256:9167–9173, 1981.
- [1290] J.L. Gaylor and H.S. Mason. Investigation of the component reactions of oxidative sterol demethylation. Evidence against participation of cytochrome P-450. *J. Biol. Chem.*, 243:4966–4972, 1968.
- [1291] F. Ge, N. Yokochi, Y. Yoshikane, K. Ohnishi, and T. Yagi. Gene identification and characterization of the pyridoxine degradative enzyme 4-pyridoxic acid dehydrogenase from the nitrogen-fixing symbiotic bacterium *Mesorhizobium loti* MAFF303099. *J. Biochem.*, 143:603–609, 2008.
- [1292] L. Ge, J.S. Gordon, C. Hsuan, K. Stenn, and S.M. Prouty. Identification of the Δ -6 desaturase of human sebaceous glands: expression and enzyme activity. *J. Invest. Dermatol.*, 120:707–714, 2003.
- [1293] L. Ge and S.Y. Seah. Heterologous expression, purification, and characterization of an *l*-ornithine N⁵-hydroxylase involved in pyoverdine siderophore biosynthesis in *Pseudomonas aeruginosa*. *J. Bacteriol.*, 188:7205–7210, 2006.
- [1294] W. Ge, A. Wolf, T. Feng, C.H. Ho, R. Sekirnik, A. Zayer, N. Granatino, M.E. Cockman, C. Loenarz, N.D. Loik, A.P. Hardy, T.D.W. Claridge, R.B. Hamed, R. Chowdhury, L. Gong, C.V. Robinson, D.C. Trudgian, M. Jiang, M.M. Mackeen, J.S. McCullagh, Y. Gordiyenko, A. Thalhammer, A. Yamamoto, M. Yang, P. Liu-Yi, Z. Zhang, M. Schmidt-Zachmann, B.M. Kessler, P.J. Ratcliffe, G.M. Preston, M.L. Coleman, and C.J. Schofield. Oxygenase-catalyzed ribosome hydroxylation occurs in prokaryotes and humans. *Nat. Chem. Biol.*, 8:960–962, 2012.
- [1295] U. Gehring and D.I. Arnon. Purification and properties of α -ketoglutarate synthase from a photosynthetic bacterium. *J. Biol. Chem.*, 247:6963–6969, 1972.
- [1296] O. Geiger and H. Gorisch. Crystalline quinoprotein glucose dehydrogenase from *Acinetobacter calcoaceticus*. *Biochemistry*, 25:6043–6048, 1986.
- [1297] B.V. Geisbrecht, X. Liang, J.C. Morrell, H. Schulz, and S.J. Gould. The mouse gene PDCR encodes a peroxisomal δ^2 , δ^4 -dienoyl-CoA reductase. *J. Biol. Chem.*, 274:25814–25820, 1999.
- [1298] R. Genet, P.H. Benetti, A. Hammadi, and A. Menez. L-Tryptophan 2',3'-oxidase from *Chromobacterium violaceum*. Substrate specificity and mechanistic implications. *J. Biol. Chem.*, 270:23540–23545, 1995.
- [1299] R. Genet, C. Denoyelle, and A. Menez. Purification and partial characterization of an amino acid α , β -dehydrogenase, L-tryptophan 2',3'-oxidase from *Chromobacterium violaceum*. *J. Biol. Chem.*, 269:18177–18184, 1994.
- [1300] R. Gerady and M.H. Zenk. Formation of salutaridine from (*R*)-reticuline by a membrane-bound cytochrome P-450 enzyme from *Papaver somniferum*. *Phytochemistry*, 32:79–86, 1993.

- [1301] R. Gerady and M.H. Zenk. Purification and characterization of salutaridine:NADPH 7-oxidoreductase from *Papaver somniferum*. *Phytochemistry*, 34:125–132, 1993.
- [1302] C. Gerdemann, C. Eicken, and B. Krebs. The crystal structure of catechol oxidase: new insight into the function of type-3 copper proteins. *Acc. Chem. Res.*, 35:183–191, 2002.
- [1303] C. Gerhardinger, M.S. Marion, A. Rovner, M. Glomb, and V.M. Monnier. Novel degradation pathway of glycated amino acids into free fructosamine by a *Pseudomonas* sp. soil strain extract. *J. Biol. Chem.*, 270:218–224, 1995.
- [1304] T. Gerjets, S. Steiger, and G. Sandmann. Catalytic properties of the expressed acyclic carotenoid 2-ketolases from *Rhodobacter capsulatus* and *Rubrivivax gelatinosus*. *Biochim. Biophys. Acta*, 1791:125–131, 2009.
- [1305] E. Gerlo and J. Charlier. Identification of NADH-specific and NADPH-specific FMN reductases in *Beneckea harveyi*. *Eur. J. Biochem.*, 57:461–467, 1975.
- [1306] J. Gershenzon, M. Maffei, and R. Croteau. Biochemical and histochemical-localization of monoterpene biosynthesis in the glandular trichomes of spearmint (*Mentha spicata*). *Plant Physiol.*, 89:1351–1357, 1989.
- [1307] J. Gescher, W. Ismail, E. Olgeschlager, W. Eisenreich, J. Worth, and G. Fuchs. Aerobic benzoyl-coenzyme A (CoA) catabolic pathway in *Azoarcus evansii*: conversion of ring cleavage product by 3,4-dehydroadipyl-CoA semialdehyde dehydrogenase. *J. Bacteriol.*, 188:2919–2927, 2006.
- [1308] J. Gescher, A. Zaar, M. Mohamed, H. Schagger, and G. Fuchs. Genes coding for a new pathway of aerobic benzoate metabolism in *Azoarcus evansii*. *J. Bacteriol.*, 184:6301–6315, 2002.
- [1309] B. Gestetner and E.E. Conn. The 2-hydroxylation of *trans*-cinnamic acid by chloroplasts from *Melilotus alba* Desr. *Arch. Biochem. Biophys.*, 163:617–624, 1974.
- [1310] K. Geszvain, J.K. McCarthy, and B.M. Tebo. Elimination of manganese(II,III) oxidation in *Pseudomonas putida* GB-1 by a double knockout of two putative multicopper oxidase genes. *Appl. Environ. Microbiol.*, 79:357–366, 2013.
- [1311] F. Geu-Flores, M.E. Møldrup, C. Böttcher, C.E. Olsen, D. Scheel, and B.A. Halkier. Cytosolic γ -glutamyl peptidases process glutathione conjugates in the biosynthesis of glucosinolates and camalexin in *Arabidopsis*. *Plant Cell*, 23:2456–2469, 2011.
- [1312] F. Geu-Flores, N.H. Sherden, V. Courdavault, V. Burlat, W.S. Glenn, C. Wu, E. Nims, Y. Cui, and S.E. O'Connor. An alternative route to cyclic terpenes by reductive cyclization in iridoid biosynthesis. *Nature*, 492:138–142, 2012.
- [1313] S.K. Ghag, D.N. Brems, T.C. Hassell, and W.K. Yeh. Refolding and purification of *Cephalosporium acremonium* deacetoxycephalosporin C synthetase/hydroxylase from granules of recombinant *Escherichia coli*. *Biotechnol. Appl. Biochem.*, 24:109–119, 1996.
- [1314] D. Ghilarov, C.E.M. Stevenson, D.Y. Travin, J. Piskunova, M. Serebryakova, A. Maxwell, D.M. Lawson, and K. Severinov. Architecture of microcin B17 synthetase: an octameric protein complex converting a ribosomally synthesized peptide into a DNA gyrase poison. *Mol. Cell*, 73:749–762.e5, 2019.
- [1315] D. Ghosh, J. Griswold, M. Erman, and W. Pangborn. Structural basis for androgen specificity and oestrogen synthesis in human aromatase. *Nature*, 457:219–223, 2009.
- [1316] G. Ghssein, C. Brutesco, L. Ouerdane, C. Fojcik, A. Izaute, S. Wang, C. Hajjar, R. Lobinski, D. Lemaire, P. Richaud, R. Voulhoux, A. Espaillat, F. Cava, D. Pignol, E. Borezee-Durant, and P. Arnoux. Biosynthesis of a broad-spectrum nicotianamine-like metallophore in *Staphylococcus aureus*. *Science*, 352:1105–1109, 2016.
- [1317] P. Giardina, M.G. de Biasi, M. de Rosa, A. Gambacorta, and V. Buonocore. Glucose dehydrogenase from the thermoacidophilic archaeobacterium *Sulfolobus solfataricus*. *Biochem. J.*, 239:517–522, 1986.
- [1318] M. Gibbs. TPN triosephosphate dehydrogenase from plant tissue. *Methods Enzymol.*, 1:411–415, 1955.
- [1319] A. Gibello, M.D. Collins, L. Dominguez, J.F. Fernandez-Garayzabal, and P.T. Richardson. Cloning and analysis of the L-lactate utilization genes from *Streptococcus iniae*. *Appl. Environ. Microbiol.*, 65:4346–4350, 1999.
- [1320] D.T. Gibson, B. Gschwendt, W.K. Yeh, and V.M. Kobal. Initial reactions in the oxidation of ethylbenzene by *Pseudomonas putida*. *Biochemistry*, 12:1520–1528, 1973.

- [1321] D.T. Gibson, J.R. Koch, and R.E. Kallio. Oxidative degradation of aromatic hydrocarbons by microorganisms. I. Enzymatic formation of catechol from benzene. *Biochemistry*, 7:2653–2662, 1968.
- [1322] D.T. Gibson, K.C. Wang, C.J. Sih, and J.H. Whitlock. Mechanisms of steroid oxidation by microorganisms. IX. On the mechanism of ring A cleavage in the degradation of 9,10-seco steroids by microorganisms. *J. Biol. Chem.*, 241:551–559, 1966.
- [1323] H. Giesel and H. Simon. On the occurrence of enoate reductase and 2-oxo-carboxylate reductase in clostridia and some observations on the amino acid fermentation by *Peptostreptococcus anaerobius*. *Arch. Microbiol.*, 135:51–57, 1983.
- [1324] T.W. Giessen, F.I. Kraas, and M.A. Marahiel. A four-enzyme pathway for 3,5-dihydroxy-4-methylanthranilic acid formation and incorporation into the antitumor antibiotic sibiromycin. *Biochemistry*, 50:5680–5692, 2011.
- [1325] S. Gilch, O. Meyer, and I. Schmidt. A soluble form of ammonia monooxygenase in *Nitrosomonas europaea*. *Biol. Chem.*, 390:863–873, 2009.
- [1326] A.A. Gilep, R.W. Estabrook, and S.A. Usanov. Molecular cloning and heterologous expression in *E. coli* of cytochrome P45017 α . Comparison of structural and functional properties of substrate-specific cytochromes P450 from different species. *Biochemistry (Mosc.)*, 68:86–98, 2003.
- [1327] S.J. Gilmour, A.B. Bleecker, and J.A.D. Zeevaart. Partial-purification of gibberellin oxidases from spinach leaves. *Plant Physiol.*, 85:87–90, 1987.
- [1328] P.P. Giovannini, A. Medici, C.M. Bergamini, and M. Rippa. Properties of diacetyl (acetoin) reductase from *Bacillus stearothermophilus*. *Bioorg. Med. Chem.*, 4:1197–1201, 1996.
- [1329] T. Girke, H. Schmidt, U. Zähringer, R. Reski, and E. Heinz. Identification of a novel Δ^6 -acyl-group desaturase by targeted gene disruption in *Physcomitrella patens*. *Plant J.*, 15:39–48, 1998.
- [1330] M.R. Gisi and L. Xun. Characterization of chlorophenol 4-monooxygenase (TftD) and NADH:flavin adenine dinucleotide oxidoreductase (TftC) of *Burkholderia cepacia* AC1100. *J. Bacteriol.*, 185:2786–2792, 2003.
- [1331] G. Gisselmann, P. Klausmeier, and J.D. Schwenn. The ferredoxin:sulphite reductase gene from *Synechococcus* PCC7942. *Biochim. Biophys. Acta*, 1144:102–106, 1993.
- [1332] A. Giuditta and H.J. Strecker. Purification and some properties of a brain diaphorase. *Biochim. Biophys. Acta*, 48:10–19, 1961.
- [1333] V.N. Gladyshev, J.C. Boyington, S.V. Khangulov, D.A. Grahame, T.C. Stadtman, and P.D. Sun. Characterization of crystalline formate dehydrogenase H from *Escherichia coli*. Stabilization, EPR spectroscopy, and preliminary crystallographic analysis. *J. Biol. Chem.*, 271:8095–8100, 1996.
- [1334] V.N. Gladyshev, S.V. Khangulov, and T.C. Stadtman. Nicotinic acid hydroxylase from *Clostridium barkeri*: electron paramagnetic resonance studies show that selenium is coordinated with molybdenum in the catalytically active selenium-dependent enzyme. *Proc. Natl. Acad. Sci. USA*, 91:232–236, 1994.
- [1335] V.N. Gladyshev, S.V. Khangulov, and T.C. Stadtman. Properties of the selenium- and molybdenum-containing nicotinic acid hydroxylase from *Clostridium barkeri*. *Biochemistry*, 35:212–223, 1996.
- [1336] T. Gladysheva, J.Y. Liu, and B.P. Rosen. His-8 lowers the pKa of the essential Cys-12 residue of the ArsC arsenate reductase of plasmid R773. *J. Biol. Chem.*, 271:33256–33260, 1996.
- [1337] T.B. Gladysheva, K.L. Oden, and B.P. Rosen. Properties of the arsenate reductase of plasmid R773. *Biochemistry*, 33:7288–7293, 1994.
- [1338] N. Glansdorff and G. Sand. Coordination of enzyme synthesis in the arginine pathway of *Escherichia coli* K-12. *Biochim. Biophys. Acta*, 108:308–311, 1965.
- [1339] L. Glaser. Ribitol-5-phosphate dehydrogenase from *Lactobacillus plantarum*. *Biochim. Biophys. Acta*, 67:525–530, 1963.
- [1340] L. Glaser and D.H. Brown. Purification and properties of D-glucose-6-phosphate dehydrogenase. *J. Biol. Chem.*, 216:67–79, 1955.

- [1341] E. Glawischnig, S. Grun, M. Frey, and A. Gierl. Cytochrome P450 monooxygenases of DIBOA biosynthesis: specificity and conservation among grasses. *Phytochemistry*, 50:925–930, 1999.
- [1342] C.G. Glembofski. Further characterization of the peptidyl α -amidating enzyme in rat anterior pituitary secretory granules. *Arch. Biochem. Biophys.*, 241:673–683, 1985.
- [1343] J.K. Glenn, L. Akileswaran, and M.H. Gold. Mn(II) oxidation is the principal function of the extracellular Mn-peroxidase from *Phanerochaete chrysosporium*. *Arch. Biochem. Biophys.*, 251:688–696, 1986.
- [1344] R. Glockler, A. Tschech, and G. Fuchs. Reductive dehydroxylation of 4-hydroxybenzoyl-CoA to benzoyl-CoA in a denitrifying, phenol-degrading *Pseudomonas* species. *FEBS Lett.*, 251:237–240, 1989.
- [1345] G. Glod, W. Angst, C. Holliger, and R.P. Schwarzenbach. Corrinoid-mediated reduction of tetrachloroethene, trichloroethene, and trichlorofluoroethene in homogeneous aqueous solution: Reaction kinetics and reaction mechanisms. *Environ. Sci. Technol.*, 31:253–260, 1997.
- [1346] P.E. Glushankov, V.E. Epifanova, and A.I. Kolotilova. Pentose phosphate pathway of carbohydrate metabolism and NADP-dependent glycerol 3-phosphate dehydrogenase activity in some white rat tissues. *Biokhimiya*, 41:1788–1790, 1976.
- [1347] M. Gnida, R. Ferner, L. Gremer, O. Meyer, and W. Meyer-Klaucke. A novel binuclear [CuSMo] cluster at the active site of carbon monoxide dehydrogenase: characterization by X-ray absorption spectroscopy. *Biochemistry*, 42:222–230, 2003.
- [1348] S. Gnidehou, B. Caillou, M. Talbot, R. Ohayon, J. Kaniewski, M.S. Noel-Hudson, S. Morand, D. Agnangji, A. Sezan, F. Courtin, A. Virion, and C. Dupuy. Iodotyrosine dehalogenase 1 (DEHAL1) is a transmembrane protein involved in the recycling of iodide close to the thyroglobulin iodination site. *FASEB J.*, 18:1574–1576, 2004.
- [1349] J.W. Godden, S. Turley, D.C. Teller, E.T. Adman, M.Y. Liu, W.J. Payne, and J. Legall. The 2.3 angstrom X-ray structure of nitrite reductase from *Achromobacter cycloclastes*. *Science*, 253:438–442, 1991.
- [1350] C.J. Goh, E.W. Szczepan, N. Menhart, and T. Viswanatha. Studies on lysine: N^6 -hydroxylation by cell-free system of *Aerobacter aerogenes* 62-1. *Biochim. Biophys. Acta*, 990:240–245, 1989.
- [1351] D.S. Goldman and M.J. Wagner. Enzyme systems in the mycobacteria. XIII. Glycine dehydrogenase and the glyoxylic acid cycle. *Biochim. Biophys. Acta*, 65:297–306, 1962.
- [1352] D. Gómez, P. Lucas-Elío, A. Sanchez-Amat, and F. Solano. A novel type of lysine oxidase: L-lysine- ϵ -oxidase. *Biochim. Biophys. Acta*, 1764:1577–1585, 2006.
- [1353] S. Gomez-Manzo, J.L. Chavez-Pacheco, M. Contreras-Zentella, M.E. Sosa-Torres, R. Arreguin-Espinosa, M. Perez de la Mora, J. Membrillo-Hernandez, and J.E. Escamilla. Molecular and catalytic properties of the aldehyde dehydrogenase of *Gluconacetobacter diazotrophicus*, a quinoxaline protein containing pyrroloquinoline quinone, cytochrome *b*, and cytochrome *c*. *J. Bacteriol.*, 192:5718–5724, 2010.
- [1354] S. Gomez-Manzo, M. Contreras-Zentella, A. Gonzalez-Valdez, M. Sosa-Torres, R. Arreguin-Espinoza, and E. Escamilla-Marvan. The PQQ-alcohol dehydrogenase of *Gluconacetobacter diazotrophicus*. *Int. J. Food Microbiol.*, 125:71–78, 2008.
- [1355] C. Gomez-Moreno and D.E. Edmondson. Evidence for an aldehyde intermediate in the catalytic mechanism of thiamine oxidase. *Arch. Biochem. Biophys.*, 239:46–52, 1985.
- [1356] K. Gomi and T. Horiuchi. Purification and characterization of a new enzyme, *N*-alkylglycine oxidase from *Cladosporium* sp. G-10. *Biochim. Biophys. Acta*, 1429:439–445, 1999.
- [1357] S. Gon, M.T. Giudici-Orticoni, V. Mejean, and C. Iobbi-Nivol. Electron transfer and binding of the *c*-type cytochrome TorC to the trimethylamine *N*-oxide reductase in *Escherichia coli*. *J. Biol. Chem.*, 276:11545–11551, 2001.
- [1358] M. Gondry, S. Lautru, G. Fusai, G. Meunier, A. Menez, and R. Genet. Cyclic dipeptide oxidase from *Streptomyces noursei*. Isolation, purification and partial characterization of a novel, amino acyl α,β -dehydrogenase. *Eur. J. Biochem.*, 268:1712–1721, 2001.

- [1359] G. Goni, A. Zollner, M. Lisurek, A. Velazquez-Campoy, S. Pinto, C. Gomez-Moreno, F. Hannemann, R. Bernhardt, and M. Medina. Cyanobacterial electron carrier proteins as electron donors to CYP106A2 from *Bacillus megaterium* ATCC 13368. *Biochim. Biophys. Acta*, 1794:1635–1642, 2009.
- [1360] E. Gonzalez, M.R. Fernandez, C. Larroy, L. Sola, M.A. Pericas, X. Pares, and J.A. Biosca. Characterization of a (2*R*,3*R*)-2,3-butanediol dehydrogenase as the *Saccharomyces cerevisiae* YAL060W gene product. Disruption and induction of the gene. *J. Biol. Chem.*, 275:35876–35885, 2000.
- [1361] M. González-Guzmán, N. Apostolova, J.M. Bellés, J.M. Barrero, P. Piqueras, M.R. Ponce, J.L. Micol, R. Serrano, and P.L. Rodríguez. The short-chain alcohol dehydrogenase ABA2 catalyzes the conversion of xanthoxin to abscisic aldehyde. *Plant Cell*, 14:1833–1846, 2002.
- [1362] C.T. Goodhue and E.E. Snell. The bacterial degradation of pantothenic acid. 3. Enzymatic formation of aldopantoic acid. *Biochemistry*, 5:403–408, 1966.
- [1363] D.S. Goodman, H.S. Huang, M. Kanai, and T. Shiratori. The enzymatic conversion of *all-trans* β -carotene into retinal. *J. Biol. Chem.*, 242:3543–3554, 1967.
- [1364] D.S. Goodman, H.S. Huang, and T. Shiratori. Mechanism of the biosynthesis of vitamin A from β -carotene. *J. Biol. Chem.*, 241:1929–1932, 1966.
- [1365] K.E. Goodwill, C. Sabatier, C. Marks, R. Raag, P.F. Fitzpatrick, and R.C. Stevens. Crystal structure of tyrosine hydroxylase at 2.3 Å and its implications for inherited neurodegenerative diseases. *Nat. Struct. Biol.*, 4:578–585, 1997.
- [1366] A.H. Gordon, D.E. Green, and V. Subrahmanyam. Liver aldehyde oxidase. *Biochem. J.*, 34:764–774, 1940.
- [1367] E.H. Gordon, M.D. Page, A.C. Willis, and S.J. Ferguson. *Escherichia coli* DipZ: anatomy of a transmembrane protein disulphide reductase in which three pairs of cysteine residues, one in each of three domains, contribute differentially to function. *Mol. Microbiol.*, 35:1360–1374, 2000.
- [1368] E.H.J. Gordon, S.L. Pealing, S.K. Chapman, F.B. Ward, and G.A. Reid. Physiological function and regulation of flavocytochrome *c*₃, the soluble fumarate reductase from *Shewanella putrefaciens* NCIMB 400. *Microbiology (Reading)*, 144:937–945, 1998.
- [1369] C. Gorner, P. Schrepfer, V. Redai, F. Wallrapp, B. Loll, W. Eisenreich, M. Haslbeck, and T. Bruck. Identification, characterization and molecular adaptation of class I redox systems for the production of hydroxylated diterpenoids. *Microb. Cell Fact.*, 15:86–86, 2016.
- [1370] R. Goss. Substrate specificity of the violaxanthin de-epoxidase of the primitive green alga *Mantoniella squamata* (Prasinophyceae). *Planta*, 217:801–812, 2003.
- [1371] A. Goswami, J.L. Leonard, and I.N. Rosenberg. Inhibition by coumadin anticoagulants of enzymatic outer ring monoiodination of iodothyronines. *Biochem. Biophys. Res. Commun.*, 104:1231–1238, 1982.
- [1372] A.M. Gotto and H.L. Kornberg. The metabolism of C2 compounds in micro-organisms. 7. Preparation and properties of crystalline tartronic semialdehyde reductase. *Biochem. J.*, 81:273–284, 1961.
- [1373] J. Gou, F. Hao, C. Huang, M. Kwon, F. Chen, C. Li, C. Liu, D.K. Ro, H. Tang, and Y. Zhang. Discovery of a non-stereoselective cytochrome P450 catalyzing either 8 α - or 8 β -hydroxylation of germacrene A acid from the Chinese medicinal plant, *Inula hupehensis*. *Plant J.*, 93:92–106, 2018.
- [1374] S.P. Gough, B.O. Petersen, and J.O. Duus. Anaerobic chlorophyll isocyclic ring formation in *Rhodobacter capsulatus* requires a cobalamin cofactor. *Proc. Natl. Acad. Sci. USA*, 97:6908–6913, 2000.
- [1375] W.H. Gough, S. VanOoteghem, T. Sint, and N.Y. Kedishvili. cDNA cloning and characterization of a new human microsomal NAD⁺-dependent dehydrogenase that oxidizes *all-trans*-retinol and 3 α -hydroxysteroids. *J. Biol. Chem.*, 273:19778–19785, 1998.
- [1376] C.W. Goulding, M.R. Sawaya, A. Parseghian, V. Lim, D. Eisenberg, and D. Missiakas. Thiol-disulfide exchange in an immunoglobulin-like fold: structure of the N-terminal domain of DsbD. *Biochemistry*, 41:6920–6927, 2002.
- [1377] M. Goulian and W.S. Beck. Purification and properties of cobamide-dependent ribonucleotide reductase from *Lactobacillus leichmannii*. *J. Biol. Chem.*, 241:4233–4242, 1966.

- [1378] D.G. Gourley, A.W. Schüttelkopf, G.A. Leonard, J. Luba, L.W. Hardy, S.M. Beverley, and W.N. Hunter. Pteridine reductase mechanism correlates pterin metabolism with drug resistance in trypanosomatid parasites. *Nat. Struct. Biol.*, 8:521–525, 2001.
- [1379] D.B. Grabarczyk and B.C. Berks. Intermediates in the Sox sulfur oxidation pathway are bound to a sulfane conjugate of the carrier protein SoxYZ. *PLoS One*, 12:e0173395–e0173395, 2017.
- [1380] D.E. Graham and R.H. White. Elucidation of methanogenic coenzyme biosyntheses: from spectroscopy to genomics. *Nat. Prod. Rep.*, 19:133–147, 2002.
- [1381] J.E. Graham and D.A. Bryant. The Biosynthetic pathway for synechoxanthin, an aromatic carotenoid synthesized by the euryhaline, unicellular cyanobacterium *Synechococcus* sp. strain PCC 7002. *J. Bacteriol.*, 190:7966–7974, 2008.
- [1382] G.A. Grant. A new family of 2-hydroxyacid dehydrogenases. *Biochem. Biophys. Res. Commun.*, 165:1371–1374, 1989.
- [1383] J.K. Grant and A.C. Brownie. The role of fumarate and TPN in steroid enzymic 11 β -hydroxylation. *Biochim. Biophys. Acta*, 18:433–434, 1955.
- [1384] G. Grass and C. Rensing. CueO is a multi-copper oxidase that confers copper tolerance in *Escherichia coli*. *Biochem. Biophys. Res. Commun.*, 286:902–908, 2001.
- [1385] M. Graupner and R.H. White. The first examples of (*S*)-2-hydroxyacid dehydrogenases catalyzing the transfer of the *pro*-4*S* hydrogen of NADH are found in the archaea. *Biochim. Biophys. Acta*, 1548:169–173, 2001.
- [1386] M. Graupner, H. Xu, and R.H. White. Identification of an archaeal 2-hydroxy acid dehydrogenase catalyzing reactions involved in coenzyme biosynthesis in methanoarchaea. *J. Bacteriol.*, 182:3688–3692, 2000.
- [1387] M. Graupner, H. Xu, and R.H. White. The pyrimidine nucleotide reductase step in riboflavin and F₄₂₀ biosynthesis in archaea proceeds by the eukaryotic route to riboflavin. *J. Bacteriol.*, 184:1952–1957, 2002.
- [1388] A.B. Graves, R.P. Morse, A. Chao, A. Iniguez, C.W. Goulding, and M.D. Liptak. Crystallographic and spectroscopic insights into heme degradation by *Mycobacterium tuberculosis* MhuD. *Inorg. Chem.*, 53:5931–5940, 2014.
- [1389] T. Gräwert, J. Kaiser, F. Zepeck, R. Laupitz, S. Hecht, S. Amslinger, N. Schramek, E. Schleicher, S. Weber, M. Haslbeck, J. Buchner, C. Rieder, D. Arigoni, A. Bacher, W. Eisenreich, and F. Rohdich. IspH protein of *Escherichia coli*: studies on iron-sulfur cluster implementation and catalysis. *J. Am. Chem. Soc.*, 126:12847–12855, 2004.
- [1390] G.O. Gray, D.F. Gaul, and D.B. Knaff. Partial purification and characterization of two soluble *c*-type cytochromes from *Chromatium vinosum*. *Arch. Biochem. Biophys.*, 222:78–86, 1983.
- [1391] K.A. Gray, O.S. Pogrebinsky, G.T. Mrachko, L. Xi, D.J. Monticello, and C.H. Squires. Molecular mechanisms of biocatalytic desulfurization of fossil fuels. *Nat. Biotechnol.*, 14:1705–1709, 1996.
- [1392] M.J. Gray and J.C. Escalante-Semerena. Single-enzyme conversion of FMNH₂ to 5,6-dimethylbenzimidazole, the lower ligand of B₁₂. *Proc. Natl. Acad. Sci. USA*, 104:2921–2926, 2007.
- [1393] R.W. Gray, J.L. Omdahl, J.G. Ghazarian, and H.F. De Luca. 25-Hydroxycholecalciferol-1-hydroxylase. Subcellular location and properties. *J. Biol. Chem.*, 247:7528–7532, 1972.
- [1394] A. A. Green and W. D. McElroy. Crystalline firefly luciferase. *Biochim. Biophys. Acta*, 20:170–176, 1956.
- [1395] A.R. Green, R.P. Hayes, L. Xun, and C. Kang. Structural understanding of the glutathione-dependent reduction mechanism of glutathionyl-hydroquinone reductases. *J. Biol. Chem.*, 287:35838–35848, 2012.
- [1396] D.E. Green, S. Mii, H.R. Mahler, and R.M. Bock. Studies on the fatty acid oxidizing system of animal tissue. III. Butyryl coenzyme A dehydrogenase. *J. Biol. Chem.*, 206:1–12, 1954.
- [1397] M.L. Green and W.H. Elliott. The enzymic formation of aminoacetone from threonine and its further metabolism. *Biochem. J.*, 92:537–549, 1964.
- [1398] P. Greenbaum, K.N. Prodouz, and R.H. Garrett. Preparation and some properties of homogeneous *Neurospora crassa* assimilatory NADPH-nitrite reductase. *Biochim. Biophys. Acta*, 526:52–64, 1978.

- [1399] J.R. Greenberg, N.P. Price, R.P. Oliver, F. Sherman, and E. Rustchenko. *Candida albicans* *SOU1* encodes a sorbose reductase required for L-sorbose utilization. *Yeast*, 22:957–969, 2005.
- [1400] J.R. Greenberg, N.P. Price, R.P. Oliver, F. Sherman, and E. Rustchenko. Erratum report: *Candida albicans* *SOU1* encodes a sorbose reductase required for L-sorbose utilization. *Yeast*, 22:1171–1171, 2005.
- [1401] B.T. Greenhagen, K. Shi, H. Robinson, S. Gamage, A.K. Bera, J.E. Ladner, and J.F. Parsons. Crystal structure of the pyocyanin biosynthetic protein PhzS. *Biochemistry*, 47:5281–5289, 2008.
- [1402] D. Greetham and C.M. Grant. Antioxidant activity of the yeast mitochondrial one-Cys peroxiredoxin is dependent on thioredoxin reductase and glutathione *in vivo*. *Mol. Cell Biol.*, 29:3229–3240, 2009.
- [1403] C. Gregolin and T.P. Singer. The lactate dehydrogenase of yeast. III. D(-)-Lactate cytochrome *c* reductase, a zinc-flavoprotein from aerobic yeast. *Biochim. Biophys. Acta*, 67:201–218, 1963.
- [1404] C. Gregolin, T.P. Singer, E.B. Kearney, and E. Boeri. The formation and enzymatic properties of the various lactic dehydrogenases of yeast. *Ann. N.Y. Acad. Sci.*, 94:780–797, 1961.
- [1405] R.P.F. Gregory and D.S. Bendall. The purification and some properties of the polyphenol oxidase from tea (*Camellia sinensis* L.). *Biochem. J.*, 101:569–581, 1966.
- [1406] L. Gremer, S. Kellner, H. Dobbek, R. Huber, and O. Meyer. Binding of flavin adenine dinucleotide to molybdenum-containing carbon monoxide dehydrogenase from *Oligotropha carboxidovorans*. Structural and functional analysis of a carbon monoxide dehydrogenase species in which the native flavoprotein has been replaced by its recombinant counterpart produced in *Escherichia coli*. *J. Biol. Chem.*, 275:1864–1872, 2000.
- [1407] S. Grether-Beck, G.L. Igloi, S. Pust, E. Schilz, K. Decker, and R. Brandsch. Structural analysis and molybdenum-dependent expression of the pAO1-encoded nicotine dehydrogenase genes of *Arthrobacter nicotinovorans*. *Mol. Microbiol.*, 13:929–936, 1994.
- [1408] M. Griffin and P.W. Trudgill. The metabolism of cyclopentanol by *Pseudomonas* N.C.I.B. 9872. *Biochem. J.*, 129:595–603, 1972.
- [1409] M. Griffin and P.W. Trudgill. Purification and properties of cyclopentanone oxygenase of *Pseudomonas* NCIB 9872. *Eur. J. Biochem.*, 63:199–209, 1976.
- [1410] W.T. Griffiths. Reconstitution of chlorophyllide formation by isolated etioplast membranes. *Biochem. J.*, 174:681–692, 1978.
- [1411] M. Grifoll, M. Casellas, J.M. Bayona, and A.M. Solanas. Isolation and characterization of a fluorene-degrading bacterium: identification of ring oxidation and ring fission products. *Appl. Environ. Microbiol.*, 58:2910–2917, 1992.
- [1412] M.M. Grilley, S.D. Stock, R.C. Dickson, R.L. Lester, and J.Y. Takemoto. Syringomycin action gene *SYR2* is essential for sphingolipid 4-hydroxylation in *Saccharomyces cerevisiae*. *J. Biol. Chem.*, 273:11062–11068, 1998.
- [1413] A.M. Grishin, E. Ajamian, L. Tao, L. Zhang, R. Menard, and M. Cygler. Structural and functional studies of the *Escherichia coli* phenylacetyl-CoA monooxygenase complex. *J. Biol. Chem.*, 286:10735–10743, 2011.
- [1414] A.M. Grishin, E. Ajamian, L. Zhang, and M. Cygler. Crystallization and preliminary X-ray analysis of PaaAC, the main component of the hydroxylase of the *Escherichia coli* phenylacetyl-coenzyme A oxygenase complex. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 66:1045–1049, 2010.
- [1415] S. Grisolia, C.L. Quijada, and M. Fernandez. Glutamate dehydrogenase from yeast and from animal tissues. *Biochim. Biophys. Acta*, 81:61–70, 1964.
- [1416] T. Grocholski, H. Koskiniemi, Y. Lindqvist, P. Mantsala, J. Niemi, and G. Schneider. Crystal structure of the cofactor-independent monooxygenase SnoaB from *Streptomyces nogalater*: implications for the reaction mechanism. *Biochemistry*, 49:934–944, 2010.
- [1417] B.W. Groen, M.A. van Kleef, and J.A. Duine. Quinohaemoprotein alcohol dehydrogenase apoenzyme from *Pseudomonas testosteroni*. *Biochem. J.*, 234:611–615, 1986.

- [1418] G. Grogan, G.A. Roberts, S. Parsons, N.J. Turner, and S.L. Flitsch. P450_{camr}, a cytochrome P450 catalysing the stereospecific 6-*endo*-hydroxylation of (1R)-(+)-camphor. *Appl. Microbiol. Biotechnol.*, 59:449–454, 2002.
- [1419] R.S. Gronke, D.J. Welsch, W.J. VanDusen, V.M. Garsky, M.K. Sardana, A.M. Stern, and P.A. Friedman. Partial purification and characterization of bovine liver aspartyl β -hydroxylase. *J. Biol. Chem.*, 265:8558–8565, 1990.
- [1420] E.E. Groseclose and D.W. Ribbons. 3-Hydroxybenzoate 6-hydroxylase from *Pseudomonas aeruginosa*. *Biochem. Biophys. Res. Commun.*, 55:897–903, 1973.
- [1421] E.E. Groseclose and D.W. Ribbons. Metabolism of resorcinolic compounds by bacteria: new pathway for resorcinol catabolism in *Azotobacter vinelandii*. *J. Bacteriol.*, 146:460–466, 1981.
- [1422] E. Gross, C.S. Sevier, N. Heldman, E. Vitu, M. Bentzur, C.A. Kaiser, C. Thorpe, and D. Fass. Generating disulfides enzymatically: reaction products and electron acceptors of the endoplasmic reticulum thiol oxidase Ero1p. *Proc. Natl. Acad. Sci. USA*, 103:299–304, 2006.
- [1423] G.G. Gross. Formation and reduction of intermediate acyladenylate by aryl-aldehyde. NADP oxidoreductase from *Neurospora crassa*. *Eur. J. Biochem.*, 31:585–592, 1972.
- [1424] G.G. Gross and W. Kreiten. Reduction of coenzyme A thioesters of cinnamic acids with an enzyme preparation from lignifying tissue of *Forsythia*. *FEBS Lett.*, 54:259–262, 1975.
- [1425] G.G. Gross and M.H. Zenk. Reduktion aromatischer Säuren zu Aldehyden und Alkoholen im zellfreien System. 1. Reinigung und Eigenschaften von Aryl-Aldehyd:NADP-Oxidoreduktase aus *Neurospora crassa*. *Eur. J. Biochem.*, 8:413–419, 1969.
- [1426] G.G. Gross and M.H. Zenk. Reduktion aromatischer Säuren zu Aldehyden und Alkoholen im zellfreien System. 2. Reinigung und Eigenschaften von Aryl Alkohol:NADP-Oxidoreduktase aus *Neurospora crassa*. *Eur. J. Biochem.*, 8:420–425, 1969.
- [1427] R. Gross, J. Simon, C.R.D. Lancaster, and A. Kroger. Identification of histidine residues in *Wolinella succinogenes* hydrogenase that are essential for menaquinone reduction by H-2. *Mol. Microbiol.*, 30:639–646, 1998.
- [1428] S.R. Gross, R.D. Gafford, and E.L. Tatum. The metabolism of protocatechuic acid by *Neurospora*. *J. Biol. Chem.*, 219:781–796, 1956.
- [1429] A. Grossmann and A. Wendel. Non-reactivity of the selenoenzyme glutathione peroxidase with enzymatically hydroperoxidized phospholipids. *Eur. J. Biochem.*, 135:549–552, 1983.
- [1430] T.L. Grove, J.H. Ahlum, R.M. Qin, N.D. Lanz, M.I. Radle, C. Krebs, and S.J. Booker. Further characterization of Cys-type and Ser-type anaerobic sulfatase maturing enzymes suggests a commonality in the mechanism of catalysis. *Biochemistry*, 52:2874–2887, 2013.
- [1431] T.L. Grove, K.H. Lee, J. St Clair, C. Krebs, and S.J. Booker. *In vitro* characterization of AtsB, a radical SAM formylglycine-generating enzyme that contains three [4Fe-4S] clusters. *Biochemistry*, 47:7523–7538, 2008.
- [1432] S.D. Grover, P.F. Canellas, and R.T. Wedding. Purification of NAD malic enzyme from potato and investigation of some physical and kinetic properties. *Arch. Biochem. Biophys.*, 209:396–407, 1981.
- [1433] A. Gruez, V. Roig-Zamboni, S. Grisel, A. Salomoni, C. Valencia, V. Campanacci, M. Tegoni, and C. Cambillau. Crystal structure and kinetics identify *Escherichia coli* YdcW gene product as a medium-chain aldehyde dehydrogenase. *J. Mol. Biol.*, 343:29–41, 2004.
- [1434] T. Gu, C. Zhou, S.R. Sorensen, J. Zhang, J. He, P. Yu, X. Yan, and S. Li. The novel bacterial *N*-demethylase PdmAB is responsible for the initial step of *N,N*-dimethyl-substituted phenylurea herbicide degradation. *Appl. Environ. Microbiol.*, 79:7846–7856, 2013.
- [1435] L.J. Guan, W.C. Lee, S. Wang, T. Ohshiro, Y. Izumi, J. Ohtsuka, and M. Tanokura. Crystal structures of apo-DszC and FMN-bound DszC from *Rhodococcus erythropolis* D-1. *FEBS J.*, 282:3126–3135, 2015.
- [1436] L.W. Guddat, J.C. Bardwell, and J.L. Martin. Crystal structures of reduced and oxidized DsbA: investigation of domain motion and thiolate stabilization. *Structure*, 6:757–767, 1998.

- [1437] M. Gudmundsson, S. Kim, M. Wu, T. Ishida, M.H. Momeni, G. Vaaje-Kolstad, D. Lundberg, A. Royant, J. Stahlberg, V.G. Eijsink, G.T. Beckham, and M. Sandgren. Structural and electronic snapshots during the transition from a Cu(II) to Cu(I) metal center of a lytic polysaccharide monooxygenase by X-ray photoreduction. *J. Biol. Chem.*, 289:18782–18792, 2014.
- [1438] B.D. Guenther, C.A. Sheppard, P. Tran, R. Rozen, R.G. Matthews, and M.L. Ludwig. The structure and properties of methylenetetrahydrofolate reductase from *Escherichia coli* suggest how folate ameliorates human hyperhomocysteinemia. *Nat. Struct. Biol.*, 6:359–365, 1999.
- [1439] A. Guha, S. Englard, and I. Listowsky. Beef heart malic dehydrogenases. VII. Reactivity of sulfhydryl groups and conformation of the supernatant enzyme. *J. Biol. Chem.*, 243:609–615, 1968.
- [1440] B. Guigliarelli, M. Asso, C. More, V. Augier, F. Blasco, J. Pommier, G. Giordano, and P. Bertrand. EPR and redox characterization of iron-sulfur centers in nitrate reductases A and Z from *Escherichia coli*. Evidence for a high-potential and a low-potential class and their relevance in the electron-transfer mechanism. *Eur. J. Biochem.*, 207:61–68, 1992.
- [1441] C. Guilhot, G. Jander, N.L. Martin, and J. Beckwith. Evidence that the pathway of disulfide bond formation in *Escherichia coli* involves interactions between the cysteines of DsbB and DsbA. *Proc. Natl. Acad. Sci. USA*, 92:9895–9899, 1995.
- [1442] H. Guldan, R. Sterner, and P. Babinger. Identification and characterization of a bacterial glycerol-1-phosphate dehydrogenase: Ni(2+)-dependent AraM from *Bacillus subtilis*. *Biochemistry*, 47:7376–7384, 2008.
- [1443] C.F. Gunsalus, R.Y. Stanier, and I.C. Gunsalus. The enzymatic conversion of mandelic acid to benzoic acid. III. Fractionation and properties of the soluble enzymes. *J. Bacteriol.*, 66:548–553, 1953.
- [1444] J. Guo, X. Ma, Y. Cai, Y. Ma, Z. Zhan, Y.J. Zhou, W. Liu, M. Guan, J. Yang, G. Cui, L. Kang, L. Yang, Y. Shen, J. Tang, H. Lin, X. Ma, B. Jin, Z. Liu, R.J. Peters, Z.K. Zhao, and L. Huang. Cytochrome P450 promiscuity leads to a bifurcating biosynthetic pathway for tanshinones. *New Phytol.*, 210:525–534, 2016.
- [1445] J. Guo, Y.J. Zhou, M.L. Hillwig, Y. Shen, L. Yang, Y. Wang, X. Zhang, W. Liu, R.J. Peters, X. Chen, Z.K. Zhao, and L. Huang. CYP76AH1 catalyzes turnover of miltiradiene in tanshinones biosynthesis and enables heterologous production of ferruginol in yeasts. *Proc. Natl. Acad. Sci. USA*, 110:12108–12113, 2013.
- [1446] L. Guo, R.A. Dixon, and N.L. Paiva. Conversion of vestitone to medicarpin in alfalfa (*Medicago sativa* L.) is catalyzed by two independent enzymes. Identification, purification, and characterization of vestitone reductase and 7,2'-dihydroxy-4'-methoxyisoflavanol dehydratase. *J. Biol. Chem.*, 269:22372–22378, 1994.
- [1447] L. Guo, R.A. Dixon, and N.L. Paiva. The 'pterocarpan synthase' of alfalfa: association and co-induction of vestitone reductase and 7,2'-dihydroxy-4'-methoxy-isoflavanol (DMI) dehydratase, the two final enzymes in medicarpin biosynthesis. *FEBS Lett.*, 356:221–225, 1994.
- [1448] L. Guo and N.L. Paiva. Molecular cloning and expression of alfalfa (*Medicago sativa* L.) vestitone reductase, the penultimate enzyme in medicarpin biosynthesis. *Arch. Biochem. Biophys.*, 320:353–360, 1995.
- [1449] L. Guo, X. Zhang, D. Zhou, A.L. Okunade, and X. Su. Stereospecificity of fatty acid 2-hydroxylase and differential functions of 2-hydroxy fatty acid enantiomers. *J. Lipid Res.*, 53:1327–1335, 2012.
- [1450] N.K. Gupta and W.G. Robinson. The enzymatic conversion of lactaldehyde to propanediol. *J. Biol. Chem.*, 235:1609–1612, 1960.
- [1451] G. Guroff and C.A. Rhoads. Phenylalanine hydroxylation by *Pseudomonas* species (ATCC 11299a). Nature of the cofactor. *J. Biol. Chem.*, 244:142–146, 1969.
- [1452] A. Gurvitz, J.K. Hiltunen, and A.J. Kastaniotis. Function of heterologous *Mycobacterium tuberculosis* InhA, a type 2 fatty acid synthase enzyme involved in extending C₂₀ fatty acids to C⁶⁰-to-C₉₀ mycolic acids, during *de novo* lipoic acid synthesis in *Saccharomyces cerevisiae*. *Appl. Environ. Microbiol.*, 74:5078–5085, 2008.
- [1453] A. Gurvitz, H. Rottensteiner, S.H. Kilpelainen, A. Hartig, J.K. Hiltunen, M. Binder, I.W. Dawes, and B. Hamilton. The *Saccharomyces cerevisiae* peroxisomal 2,4-dienoyl-CoA reductase is encoded by the oleate-inducible gene SPS19. *J. Biol. Chem.*, 272:22140–22147, 1997.

- [1454] A. Guskov, J. Kern, A. Gabdulkhakov, M. Broser, A. Zouni, and W. Saenger. Cyanobacterial photosystem II at 2.9-Å resolution and the role of quinones, lipids, channels and chloride. *Nat. Struct. Mol. Biol.*, 16:334–342, 2009.
- [1455] J. Gustafsson. Biosynthesis of cholic acid in rat liver. 24-Hydroxylation of 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanic acid. *J. Biol. Chem.*, 250:8243–8247, 1975.
- [1456] A. Gutierrez, A. Grunau, M. Paine, A.W. Munro, C.R. Wolf, G.C. Roberts, and N.S. Scrutton. Electron transfer in human cytochrome P450 reductase. *Biochem. Soc. Trans.*, 31:497–501, 2003.
- [1457] J.F. Gutierrez-Marcos, M.A. Roberts, E.I. Campbell, and J.L. Wray. Three members of a novel small gene-family from *Arabidopsis thaliana* able to complement functionally an *Escherichia coli* mutant defective in PAPS reductase activity encode proteins with a thioredoxin-like domain and ‘APS reductase’ activity. *Proc. Natl. Acad. Sci. USA*, 93:13377–13382, 1996.
- [1458] H.R. Gutmann and R.R. Erickson. The conversion of the carcinogen *N*-hydroxy-2-fluorenylacamide to *o*-amidophenols by rat liver in vitro. An inducible enzymatic reaction. *J. Biol. Chem.*, 244:1729–1740, 1969.
- [1459] J.Y. Ha, J.Y. Min, S.K. Lee, H.S. Kim, J. Kim do, K.H. Kim, H.H. Lee, H.K. Kim, H.J. Yoon, and S.W. Suh. Crystal structure of 2-nitropropane dioxygenase complexed with FMN and substrate. Identification of the catalytic base. *J. Biol. Chem.*, 281:18660–18667, 2006.
- [1460] B. Haak, S. Fetzner, and F. Lingens. Cloning, nucleotide sequence, and expression of the plasmid-encoded genes for the two-component 2-halobenzoate 1,2-dioxygenase from *Pseudomonas cepacia* 2CBS. *J. Bacteriol.*, 177:667–675, 1995.
- [1461] D. Haak, K. Gable, T. Beeler, and T. Dunn. Hydroxylation of *Saccharomyces cerevisiae* ceramides requires Sur2p and Scs7p. *J. Biol. Chem.*, 272:29704–29710, 1997.
- [1462] E. Haas, B.L. Horecker, and T.R. Hogness. The enzymatic reduction of cytochrome *c*, cytochrome *c* reductase. *J. Biol. Chem.*, 136:747–774, 1940.
- [1463] N. Habu, M. Samejima, J.F. Dean, and K.E. Eriksson. Release of the FAD domain from cellobiose oxidase by proteases from cellulolytic cultures of *Phanerochaete chrysosporium*. *FEBS Lett.*, 327:161–164, 1993.
- [1464] B. Hacker, A. Habenicht, M. Kiess, and R. Mattes. Xylose utilisation: cloning and characterisation of the xylose reductase from *Candida tenuis*. *Biol. Chem.*, 380:1395–1403, 1999.
- [1465] J.D. Haddock and J.G. Ferry. Purification and properties of phloroglucinol reductase from *Eubacterium oxidoreducens* G-41. *J. Biol. Chem.*, 264:4423–4427, 1989.
- [1466] J.D. Haddock and D.T. Gibson. Purification and characterization of the oxygenase component of biphenyl 2,3-dioxygenase from *Pseudomonas* sp. strain LB400. *J. Bacteriol.*, 177:5834–5839, 1995.
- [1467] J.D. Haddock, D.A. Pelletier, and D.T. Gibson. Purification and properties of ferredoxinBPH, a component of biphenyl 2,3-dioxygenase of *Pseudomonas* sp. strain LB400. *J. Indust. Microbiol. Biotechnol.*, 19:355–359, 1997.
- [1468] F. Haeseleer, J. Huang, L. Lebioda, J.C. Saari, and K. Palczewski. Molecular characterization of a novel short-chain dehydrogenase/reductase that reduces *all-trans*-retinal. *J. Biol. Chem.*, 273:21790–21799, 1998.
- [1469] P. Haferkamp, S. Kutschki, J. Treichel, H. Hemeda, K. Sewczyk, D. Hoffmann, M. Zaparty, and B. Siebers. An additional glucose dehydrogenase from *Sulfolobus solfataricus*: fine-tuning of sugar degradation. *Biochem. Soc. Trans.*, 39:77–81, 2011.
- [1470] D.H. Haft. Bioinformatic evidence for a widely distributed, ribosomally produced electron carrier precursor, its maturation proteins, and its nicotinoprotein redox partners. *BMC Genomics*, 12:21–21, 2011.
- [1471] J.M. Hagel and P.J. Facchini. Dioxygenases catalyze the *O*-demethylation steps of morphine biosynthesis in opium poppy. *Nat. Chem. Biol.*, 6:273–275, 2010.
- [1472] R.H. Hageman and D.I. Arnon. The isolation of triosephosphate dehydrogenase from pea seeds. *Arch. Biochem. Biophys.*, 55:162–168, 1955.

- [1473] W.R. Hagen, P.J. Silva, M.A. Amorim, P.L. Hagedoorn, H. Wassink, H. Haaker, and F.T. Robb. Novel structure and redox chemistry of the prosthetic groups of the iron-sulfur flavoprotein sulfide dehydrogenase from *Pyrococcus furiosus*; evidence for a [2Fe-2S] cluster with Asp(Cys)₃ ligands. *J. Biol. Inorg. Chem.*, 5:527–534, 2000.
- [1474] L.P. Hager, P.F. Hollenberg, T. Rand-Meir, R. Chiang, and D.L. Doubek. Chemistry of peroxidase intermediates. *Ann. N.Y. Acad. Sci.*, 244:80–93, 1975.
- [1475] R. Hagihara, Y. Katsuyama, Y. Sugai, H. Onaka, and Y. Ohnishi. Novel desferrioxamine derivatives synthesized using the secondary metabolism-specific nitrous acid biosynthetic pathway in *Streptomyces davawensis*. *J. Antibiot. (Tokyo)*, 71:911–919, 2018.
- [1476] M.-L. Hagmann, W. Heller, and H. Grisebach. Induction of phytoalexin synthesis in soybean. Stereospecific 3,9-dihydroxypterocarpan 6 α -hydroxylase from elicitor-induced soybean cell cultures. *Eur. J. Biochem.*, 142:127–131, 1984.
- [1477] B.E. Haigler and D.T. Gibson. Purification and properties of NADH-ferredoxinNAP reductase, a component of naphthalene dioxygenase from *Pseudomonas* sp. strain NCIB 9816. *J. Bacteriol.*, 172:457–464, 1990.
- [1478] B.E. Haigler, W.C. Suen, and J.C. Spain. Purification and sequence analysis of 4-methyl-5-nitrocatechol oxygenase from *Burkholderia* sp. strain DNT. *J. Bacteriol.*, 178:6019–6024, 1996.
- [1479] W.J. Haines. The biosynthesis of adrenal cortex hormones. *Recent Progr. Hormone Res.*, 7:255–305, 1952.
- [1480] N. Hakulinen, O. Turunen, J. Janis, M. Leisola, and J. Rouvinen. Three-dimensional structures of thermophilic β -1,4-xylanases from *Chaetomium thermophilum* and *Nonomuraea flexuosa*. Comparison of twelve xylanases in relation to their thermal stability. *Eur. J. Biochem.*, 270:1399–1412, 2003.
- [1481] B.A. Halkier and B.L. Møller. The biosynthesis of cyanogenic glucosides in higher plants. Identification of three hydroxylation steps in the biosynthesis of dhurrin in *Sorghum bicolor* (L.) Moench and the involvement of 1-ACI-nitro-2-(*p*-hydroxyphenyl)ethane as an intermediate. *J. Biol. Chem.*, 265:21114–21121, 1990.
- [1482] C.L. Hall, L. Heijkenkjold, T. Bartfai, L. Ernster, and H. Kamin. Acyl coenzyme A dehydrogenases and electron-transferring flavoprotein from beef heart mitochondria. *Arch. Biochem. Biophys.*, 177:402–414, 1976.
- [1483] D.L. Hallahan, J.M. West, R.M. Wallsgrove, D.W. Smiley, G.W. Dawson, J.A. Pickett, and J.G. Hamilton. Purification and characterization of an acyclic monoterpene primary alcohol:NADP⁺ oxidoreductase from catmint (*Nepeta racemosa*). *Arch. Biochem. Biophys.*, 318:105–112, 1995.
- [1484] B.M. Hallberg, G. Henriksson, G. Pettersson, and C. Divne. Crystal structure of the flavoprotein domain of the extracellular flavocytochrome cellobiose dehydrogenase. *J. Mol. Biol.*, 315:421–434, 2002.
- [1485] T.M. Hallis, Y. Lei, N.L. Que, and H. Liu. Mechanistic studies of the biosynthesis of paratose: purification and characterization of CDP-paratose synthase. *Biochemistry*, 37:4935–4945, 1998.
- [1486] B. Haltli, Y. Tan, N.A. Magarvey, M. Wagenaar, X. Yin, M. Greenstein, J.A. Hucul, and T.M. Zabriskie. Investigating β -hydroxyenduracididine formation in the biosynthesis of the mannopeptimycins. *Chem. Biol.*, 12:1163–1168, 2005.
- [1487] H. Hamamoto, T. Kusudo, N. Urushino, H. Masuno, K. Yamamoto, S. Yamada, M. Kamakura, M. Ohta, K. Inouye, and T. Sakaki. Structure-function analysis of vitamin D 24-hydroxylase (CYP24A1) by site-directed mutagenesis: amino acid residues responsible for species-based difference of CYP24A1 between humans and rats. *Mol. Pharmacol.*, 70:120–128, 2006.
- [1488] M. Hamberg and G. Hamberg. Peroxygenase-catalyzed fatty acid epoxidation in cereal seeds (sequential oxidation of linoleic acid into 9(*S*),12(*S*),13(*S*)-trihydroxy-10(*E*)-octadecenoic acid). *Plant Physiol.*, 110:807–815, 1996.
- [1489] M. Hamberg and B. Samuelsson. Prostaglandin endoperoxides. Novel transformations of arachidonic acid in human platelets. *Proc. Natl. Acad. Sci. USA*, 71:3400–3404, 1974.
- [1490] M. Hamberg, A. Sanz, and C. Castresana. α -oxidation of fatty acids in higher plants. Identification of a pathogen-inducible oxygenase (piox) as an α -dioxygenase and biosynthesis of 2-hydroperoxylinolenic acid. *J. Biol. Chem.*, 274:24503–24513, 1999.
- [1491] M. Hamberg, C. Su, and E.H. Oliw. Manganese lipoxygenase: Discovery of bis-allylic hydroperoxide as product and intermediate in a lipoxygenase reaction. *J. Biol. Chem.*, 273:13080–13088, 1998.

- [1492] M. Hamberg, L.-Y. Zhang, I.D. Brodowsky, and E.H. Oliw. Sequential oxygenation of linoleic acid in the fungus *Gaeumannomyces graminis*: stereochemistry of dioxygenase and hydroperoxide isomerase reactions. *Arch. Biochem. Biophys.*, 309:77–80, 1994.
- [1493] D. Hamerski and U. Matern. Elicitor-induced biosynthesis of psoralens in *Ammi majus* L. suspension cultures. Microsomal conversion of demethylsuberosin into (+)marmesin and psoralen. *Eur. J. Biochem.*, 171:369–375, 1988.
- [1494] M. Hamon, S. Bourgoïn, F. Artaud, and J. Glowinski. The role of intraneuronal 5-HT and of tryptophan hydroxylase activation in the control of 5-HT synthesis in rat brain slices incubated in K⁺-enriched medium. *J. Neurochem.*, 33:1031–1042, 1979.
- [1495] G. Han, K. Gable, S.D. Kohlwein, F. Beaudoin, J.A. Napier, and T.M. Dunn. The *Saccharomyces cerevisiae* YBR159w gene encodes the 3-ketoreductase of the microsomal fatty acid elongase. *J. Biol. Chem.*, 277:35440–35449, 2002.
- [1496] J. Han, J.M. Clement, J. Li, A. King, S. Ng, and J.G. Jaworski. The cytochrome P450 CYP86A22 is a fatty acyl-CoA ω -hydroxylase essential for estolide synthesis in the stigma of *Petunia hybrida*. *J. Biol. Chem.*, 285:3986–3996, 2010.
- [1497] J.S. Han and K. Ishikawa. Active site of Zn²⁺-dependent *sn*-glycerol-1-phosphate dehydrogenase from *Aeropyrum pernix* K1. *Archaea*, 1:311–317, 2005.
- [1498] J.Y. Han, H.S. Hwang, S.W. Choi, H.J. Kim, and Y.E. Choi. Cytochrome P450 CYP716A53v2 catalyzes the formation of protopanaxatriol from protopanaxadiol during ginsenoside biosynthesis in *Panax ginseng*. *Plant Cell Physiol.*, 53:1535–1545, 2012.
- [1499] J.Y. Han, H.J. Kim, Y.S. Kwon, and Y.E. Choi. The Cyt P450 enzyme CYP716A47 catalyzes the formation of protopanaxadiol from dammarenediol-II during ginsenoside biosynthesis in *Panax ginseng*. *Plant Cell Physiol.*, 52:2062–2073, 2011.
- [1500] J.Y. Han, M.J. Kim, Y.W. Ban, H.S. Hwang, and Y.E. Choi. The involvement of β -amyrin 28-oxidase (CYP716A52v2) in oleanane-type ginsenoside biosynthesis in *Panax ginseng*. *Plant Cell Physiol.*, 54:2034–2046, 2013.
- [1501] M.H. Han, B.-L. Seong, H.-J. Son, and T.-I. Mheen. Rifamycin B oxidase from *Monocillium* spp., a new type of diphenol oxidase. *FEBS Lett.*, 151:36–40, 1983.
- [1502] Z. Han, T. Niu, J. Chang, X. Lei, M. Zhao, Q. Wang, W. Cheng, J. Wang, Y. Feng, and J. Chai. Crystal structure of the FTO protein reveals basis for its substrate specificity. *Nature*, 464:1205–1209, 2010.
- [1503] A. Hanano, M. Burcklen, M. Flenet, A. Ivancich, M. Louwagie, J. Garin, and E. Blee. Plant seed peroxygenase is an original heme-oxygenase with an EF-hand calcium binding motif. *J. Biol. Chem.*, 281:33140–33151, 2006.
- [1504] C.C. Hanfrey, B.M. Pearson, S. Hazeldine, J. Lee, D.J. Gaskin, P.M. Woster, M.A. Phillips, and A.J. Michael. Alternative spermidine biosynthetic route is critical for growth of *Campylobacter jejuni* and is the dominant polyamine pathway in human gut microbiota. *J. Biol. Chem.*, 286:43301–43312, 2011.
- [1505] V.T.T. Hang, T.J. Oh, T. Yamaguchi, and J.K. Sohng. *In vivo* characterization of NcsB3 to establish the complete biosynthesis of the naphthoic acid moiety of the neocarzinostatin chromophore. *FEMS Microbiol. Lett.*, 311:119–125, 2010.
- [1506] S.P. Hanlon, T.H. Toh, P.S. Solomon, R.A. Holt, and A.G. McEwan. Dimethylsulfide:acceptor oxidoreductase from *Rhodobacter sulfidophilus*. The purified enzyme contains b-type haem and a pterin molybdenum cofactor. *Eur. J. Biochem.*, 239:391–396, 1996.
- [1507] E.W. Hanly. Preliminary characterization and physical properties of pyridoxal oxidase activity from *Drosophila melanogaster*. *Mol. Gen. Genet.*, 180:455–462, 1980.
- [1508] R. Hanna, M. Picken, and J. Mendicino. Purification of a specific D-apiitol dehydrogenase from a *Micrococcus* isolated from the surface of germinating parsley seeds. *Biochim. Biophys. Acta*, 315:259–271, 1973.
- [1509] E.M. Hanschmann, M.E. Lonn, L.D. Schutte, M. Funke, J.R. Godoy, S. Eitner, C. Hudemann, and C.H. Lillig. Both thioredoxin 2 and glutaredoxin 2 contribute to the reduction of the mitochondrial 2-Cys peroxiredoxin Prx3. *J. Biol. Chem.*, 285:40699–40705, 2010.

- [1510] B.G. Hansen, D.J. Kliebenstein, and B.A. Halkier. Identification of a flavin-monoxygenase as the S-oxygenating enzyme in aliphatic glucosinolate biosynthesis in *Arabidopsis*. *Plant J.*, 50:902–910, 2007.
- [1511] C.H. Hansen, U. Wittstock, C.E. Olsen, A.J. Hick, J.A. Pickett, and B.A. Halkier. Cytochrome *p*₄₅₀ CYP79F1 from *Arabidopsis* catalyzes the conversion of dihomomethionine and trihomomethionine to the corresponding aldoximes in the biosynthesis of aliphatic glucosinolates. *J. Biol. Chem.*, 276:11078–11085, 2001.
- [1512] H.S. Hansen. Purification and assay of 15-ketoprostaglandin Δ^{13} -reductase from bovine lung. *Methods Enzymol.*, 86:156–163, 1982.
- [1513] T. Hansen, B. Schlichting, and P. Schonheit. Glucose-6-phosphate dehydrogenase from the hyperthermophilic bacterium *Thermotoga maritima*: expression of the *g6pd* gene and characterization of an extremely thermophilic enzyme. *FEMS Microbiol. Lett.*, 216:249–253, 2002.
- [1514] M. Hansson and L. Hederstedt. *Bacillus subtilis* HemY is a peripheral membrane protein essential for protoheme IX synthesis which can oxidize coproporphyrinogen III and protoporphyrinogen IX. *J. Bacteriol.*, 176:5962–5970, 1994.
- [1515] R. Hansson and K. Wikvall. Hydroxylations in biosynthesis and metabolism of bile acids. Catalytic properties of different forms of cytochrome P-450. *J. Biol. Chem.*, 255:1643–1649, 1980.
- [1516] R. Hansson and K. Wikvall. Hydroxylations in biosynthesis of bile acids. Cytochrome P-450 LM4 and 12 α -hydroxylation of 5 β -cholestane-3 α ,7 α -diol. *Eur. J. Biochem.*, 125:423–429, 1982.
- [1517] I. Hanukoglu and T. Gutfinger. cDNA sequence of adrenodoxin reductase. Identification of NADP-binding sites in oxidoreductases. *Eur. J. Biochem.*, 180:479–484, 1989.
- [1518] I. Hanukoglu and Z. Hanukoglu. Stoichiometry of mitochondrial cytochromes P-450, adrenodoxin and adrenodoxin reductase in adrenal cortex and corpus luteum. Implications for membrane organization and gene regulation. *Eur. J. Biochem.*, 157:27–31, 1986.
- [1519] I. Hanukoglu and C.R. Jefcoate. Mitochondrial cytochrome P-450_{sc}. Mechanism of electron transport by adrenodoxin. *J. Biol. Chem.*, 255:3057–3061, 1980.
- [1520] I. Hanukoglu, V. Spitsberg, J.A. Bumpus, K.M. Dus, and C.R. Jefcoate. Adrenal mitochondrial cytochrome P-450_{sc}. Cholesterol and adrenodoxin interactions at equilibrium and during turnover. *J. Biol. Chem.*, 256:4321–4328, 1981.
- [1521] J.C. Hanvey, E.S. Hawkins, D.C. Baker, and R.J. Suhadolnick. 8-Ketodeoxycoformycin and 8-ketocoformycin as intermediates in the biosynthesis of 2'-deoxycoformycin and coformycin. *Biochemistry*, 27:5790–5795, 1988.
- [1522] G. Hao, M. O'Connor, W. Liu, and W.L. Roelofs. Characterization of Z/E11- and Z9-desaturases from the obliquebanded leafroller moth, *Choristoneura rosaceana*. *J. Insect Sci.*, 2:26:1–7, 2002.
- [1523] A. Hara, M. Nakagawa, H. Taniguchi, and H. Sawada. 3(20) α -Hydroxysteroid dehydrogenase activity of monkey liver indanol dehydrogenase. *J. Biochem. (Tokyo)*, 106:900–903, 1989.
- [1524] R. Hara and K. Kino. Characterization of novel 2-oxoglutarate dependent dioxygenases converting L-proline to *cis*-4-hydroxy-L-proline. *Biochem. Biophys. Res. Commun.*, 379:882–886, 2009.
- [1525] J. Harada, S. Miyago, T. Mizoguchi, C. Azai, K. Inoue, H. Tamiaki, and H. Oh-oka. Accumulation of chlorophyllous pigments esterified with the geranylgeranyl group and photosynthetic competence in the CT2256-deleted mutant of the green sulfur bacterium *Chlorobium tepidum*. *Photochem Photobiol Sci*, 7:1179–1187, 2008.
- [1526] J. Harada, T. Mizoguchi, S. Satoh, Y. Tsukatani, M. Yokono, M. Noguchi, A. Tanaka, and H. Tamiaki. Specific gene *bciD* for C7-methyl oxidation in bacteriochlorophyll *e* biosynthesis of brown-colored green sulfur bacteria. *PLoS One*, 8:e60026–e60026, 2013.
- [1527] J. Harada, T. Mizoguchi, Y. Tsukatani, M. Yokono, A. Tanaka, and H. Tamiaki. Chlorophyllide *a* oxidoreductase works as one of the divinyl reductases specifically involved in bacteriochlorophyll *a* biosynthesis. *J. Biol. Chem.*, 289:12716–12726, 2014.
- [1528] T. Harano. New diaphorases from Bombyx silkworm eggs. NADH/NADPH cytochrome *c* reductase activity mediated with 6,7-dimethyltetrahydropterin. *Insect Biochem.*, 2:385–399, 1972.

- [1529] I. Harary, S.R. Korey, and S. Ochoa. Biosynthesis of dicarboxylic acids by carbon dioxide fixation. VII. Equilibrium of "malic" enzyme reaction. *J. Biol. Chem.*, 203:595–604, 1953.
- [1530] N.R. Harborne, L. Griffiths, S.J. Busby, and J.A. Cole. Transcriptional control, translation and function of the products of the five open reading frames of the *Escherichia coli nir* operon. *Mol. Microbiol.*, 6:2805–2813, 1992.
- [1531] M.J. Hardman and R.K. Scopes. The kinetics of glucose-fructose oxidoreductase from *Zymomonas mobilis*. *Eur. J. Biochem.*, 173:203–209, 1988.
- [1532] W.A. Hareland, R.L. Crawford, P.J. Chapman, and S. Dagley. Metabolic function and properties of 4-hydroxyphenylacetic acid 1-hydroxylase from *Pseudomonas acidovorans*. *J. Bacteriol.*, 121:272–285, 1975.
- [1533] A.K. Harris, N.R. Williamson, H. Slater, A. Cox, S. Abbasi, I. Foulds, H.T. Simonsen, F.J. Leeper, and G.P. Salmond. The *Serratia* gene cluster encoding biosynthesis of the red antibiotic, prodigiosin, shows species- and strain-dependent genome context variation. *Microbiology*, 150:3547–3560, 2004.
- [1534] D.R. Harris, D.E. Ward, J.M. Feasel, K.M. Lancaster, R.D. Murphy, T.C., Crane Mallet, , and 3rd. Discovery and characterization of a coenzyme A disulfide reductase from *Pyrococcus horikoshii*. Implications for this disulfide metabolism of anaerobic hyperthermophiles. *FEBS J.*, 272:1189–1200, 2005.
- [1535] E.D. Harris, W.A. Gonnerman, J.E. Savage, and B.L. O'Dell. Connective tissue amine oxidase. II. Purification and partial characterization of lysyl oxidase from chick aorta. *Biochim. Biophys. Acta*, 341:332–344, 1974.
- [1536] N.C. Harris, D.A. Born, W. Cai, Y. Huang, J. Martin, R. Khalaf, C.L. Drennan, and W. Zhang. Isonitrile formation by a non-heme iron(II)-dependent oxidase/decarboxylase. *Angew. Chem. Int. Ed. Engl.*, 57:9707–9710, 2018.
- [1537] N.C. Harris, M. Sato, N.A. Herman, F. Twigg, W. Cai, J. Liu, X. Zhu, J. Downey, R. Khalaf, J. Martin, H. Koshino, and W. Zhang. Biosynthesis of isonitrile lipopeptides by conserved nonribosomal peptide synthetase gene clusters in Actinobacteria. *Proc. Natl. Acad. Sci. USA*, 114:7025–7030, 2017.
- [1538] R.A. Harris, J.W. Hawes, K.M. Popov, Y. Zhao, Y. Shimomura, J. Sato, J. Jaskiewicz, and T.D. Hurley. Studies on the regulation of the mitochondrial α -ketoacid dehydrogenase complexes and their kinases. *Adv. Enzyme Regul.*, 37:271–293, 1997.
- [1539] S.C. Harris, S. Devendran, J.M.P. Alves, S.M. Mythen, P.B. Hylemon, and J.M. Ridlon. Identification of a gene encoding a flavoprotein involved in bile acid metabolism by the human gut bacterium *Clostridium scindens* ATCC 35704. *Biochim. Biophys. Acta*, 1863:276–283, 2018.
- [1540] J.E. Harrison and J. Schultz. Studies on the chlorinating activity of myeloperoxidase. *J. Biol. Chem.*, 251:1371–1374, 1976.
- [1541] U. Hartel, E. Eckel, J. Koch, G. Fuchs, D. Linder, and W. Buckel. Purification of glutaryl-CoA dehydrogenase from *Pseudomonas* sp., an enzyme involved in the anaerobic degradation of benzoate. *Arch. Microbiol.*, 159:174–181, 1993.
- [1542] D. Hartshorne and D.M. Greenberg. Studies on liver threonine dehydrogenase. *Arch. Biochem. Biophys.*, 105:173–178, 1964.
- [1543] P.L. Hartzell, G. Zvilius, J.C. Escalante-Semerena, and M.I. Donnelly. Coenzyme F₄₂₀ dependence of the methylenetetrahydromethanopterin dehydrogenase of *Methanobacterium thermoautotrophicum*. *Biochem. Biophys. Res. Commun.*, 133:884–890, 1985.
- [1544] P.J. Harvey, H.E. Schoemaker, and J.M. Palmer. Veratryl alcohol as a mediator and the role of radical cations in lignin biodegradation by *Phanerochaete chrysosporium*. *FEBS Lett.*, 195:242–246, 1986.
- [1545] C.S. Harwood and J. Gibson. Shedding light on anaerobic benzene ring degradation: a process unique to prokaryotes? *J. Bacteriol.*, 179:301–309, 1997.
- [1546] R.H. Haschke and L.L. Campbell. Thiosulfate reductase of *Desulfovibrio vulgaris*. *J. Bacteriol.*, 106:603–607, 1971.
- [1547] H. Hasegawa. Dihydropteridine reductase from bovine liver. Purification, crystallization, and isolation of a binary complex with NADH. *J. Biochem. (Tokyo)*, 81:169–177, 1977.

- [1548] S. Hasegawa, S.M. Poling, V.P. Maier, and R.D. Bennett. Metabolism of abscisic-acid bacterial conversion to dehydrovomifoliol and vomifoliol dehydrogenase-activity. *Phytochemistry*, 23:2769–2771, 1984.
- [1549] M.F. Hashim, T. Hakamatsuka, Y. Ebizuka, and U. Sankawa. Reaction mechanism of oxidative rearrangement of flavanone in isoflavone biosynthesis. *FEBS Lett.*, 271:219–222, 1990.
- [1550] T. Hashimoto, J. Kohno, and Y. Yamada. 6 β -Hydroxyhyoscyamine epoxidase from cultured roots of *Hyoscyamus niger*. *Phytochemistry*, 28:1077–1082, 1989.
- [1551] T. Hashimoto and Y. Yamada. Hyoscyamine 6 β -hydroxylase, a 2-oxoglutarate-dependent dioxygenase, in alkaloid-producing root cultures. *Plant Physiol.*, 81:619–625, 1986.
- [1552] E.S. Haslewood and G.A.D. Haslewood. The specificity of a 7 α -hydroxy steroid dehydrogenase from *Escherichia coli*. *Biochem. J.*, 157:207–210, 1976.
- [1553] J.W. Hastings. Bacterial bioluminescence light emission in the mixed function oxidation of reduced flavin and fatty aldehyde. *Crit. Rev. Biochem.*, 5:163–184, 1978.
- [1554] J.W. Hastings and K.H. Nealson. Bacterial bioluminescence. *Annu. Rev. Microbiol.*, 31:549–595, 1977.
- [1555] J.W. Hastings and R.P. Presswood. Bacterial luciferase: FMNH₂-aldehyde oxidase. *Methods Enzymol.*, 53:558–570, 1978.
- [1556] H. Hata, S. Shimizu, S. Hattori, and H. Yamada. Ketopantoyl-lactone reductase from *Candida parapsilosis*: purification and characterization as a conjugated polyketone reductase. *Biochim. Biophys. Acta*, 990:175–181, 1989.
- [1557] M.D. Hatch and J.F. Turner. A protein disulphide reductase from pea seeds. *Biochem. J.*, 76:556–562, 1960.
- [1558] E.C. Hatchikian. Purification and properties of thiosulfate reductase from *Desulfovibrio gigas*. *Arch. Microbiol.*, 105:249–256, 1975.
- [1559] Y. Hatefi, M.J. Osborn, L.D. Kay, and F.M. Huennekens. Hydroxymethyl tetrahydrofolic dehydrogenase. *J. Biol. Chem.*, 227:637–647, 1957.
- [1560] J.A. Hathaway and D.E. Atkinson. The effect of adenylic acid on yeast nicotinamide adenine dinucleotide isocitrate dehydrogenase, a possible metabolic control mechanism. *J. Biol. Chem.*, 238:2875–2881, 1963.
- [1561] T. Hatta, G. Mukerjee-Dhar, J. Damborsky, H. Kiyohara, and K. Kimbara. Characterization of a novel thermostable Mn(II)-dependent 2,3-dihydroxybiphenyl 1,2-dioxygenase from a polychlorinated biphenyl- and naphthalene-degrading *Bacillus* sp. JF8. *J. Biol. Chem.*, 278:21483–21492, 2003.
- [1562] T. Hatta, O. Nakano, N. Imai, N. Takizawa, and H. Kiyohara. Cloning and sequence analysis of hydroxyquinol 1,2-dioxygenase gene in 2,4,6-trichlorophenol-degrading *Ralstonia pickettii* DTP0602 and characterization of its product. *J. Biosci. Bioeng.*, 87:267–272, 1999.
- [1563] J.G. Hauge, F.L. Crane, and H. Beinert. On the mechanism of dehydrogenation of fatty acyl derivatives of coenzyme A. III. Palmityl CoA dehydrogenase. *J. Biol. Chem.*, 219:727–733, 1956.
- [1564] D.A. Haugen and M.J. Coon. Properties of electrophoretically homogeneous phenobarbital-inducible and β -naphthoflavone-inducible forms of liver microsomal cytochrome P-450. *J. Biol. Chem.*, 251:7929–7939, 1976.
- [1565] M. Hauser, P. Horn, H. Tournu, N.C. Hauser, J.D. Hoheisel, A.J. Brown, and J.R. Dickinson. A transcriptome analysis of isoamyl alcohol-induced filamentation in yeast reveals a novel role for Gre2p as isovaleraldehyde reductase. *FEMS Yeast Res.*, 7:84–92, 2007.
- [1566] A. Hausmann and G. Sandmann. A single five-step desaturase is involved in the carotenoid biosynthesis pathway to β -carotene and torulene in *Neurospora crassa*. *Fungal Genet. Biol.*, 30:147–153, 2000.
- [1567] E. Hausmann. Cofactor requirements for the enzymatic hydroxylation of lysine in a polypeptide precursor of collagen. *Biochim. Biophys. Acta*, 133:591–598, 1967.
- [1568] D.B. Hawkes, G.W. Adams, A.L. Burlingame, P.R. Ortiz de Montellano, and J.J. De Voss. Cytochrome P450_{cin} (CYP176A), isolation, expression, and characterization. *J. Biol. Chem.*, 277:27725–27732, 2002.

- [1569] A.R. Hawkins, N.H. Giles, and J.R. Kinghorn. Genetical and biochemical aspects of quinate breakdown in the filamentous fungus *Aspergillus nidulans*. *Biochem. Genet.*, 20:271–286, 1982.
- [1570] O. Hayaishi. Direct oxygenation by O₂, oxygenases. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 8, pages 353–371. Academic Press, New York, 2nd edition, 1963.
- [1571] O. Hayaishi, M. Katagiri, and S. Rothberg. Studies on oxygenases: pyrocatechase. . *J. Biol. Chem.*, 229:905–920, 1957.
- [1572] O. Hayaishi and A. Kornberg. Metabolism of cytosine, thymine, uracil, and barbituric acid by bacterial enzymes. *J. Biol. Chem.*, 197:717–723, 1952.
- [1573] O. Hayaishi, Y. Nishizuka, M. Tatibana, M. Takeshita, and S. Kuno. Enzymatic studies on the metabolism of β-alanine. *J. Biol. Chem.*, 236:781–790, 1961.
- [1574] O. Hayaishi, Y. Saito, W.B. Jakoby, and E.F. Stohman. Reversible enzymatic oxidation of bile acids. *Arch. Biochem. Biophys.*, 56:554–555, 1955.
- [1575] O. Hayaishi and W.B. Sutton. Enzymatic oxygen fixation into acetate concomitant with the enzymatic decarboxylation of L-lactate. *J. Am. Chem. Soc.*, 79:4809–4810, 1957.
- [1576] K. Hayano and S. Fukui. Purification and properties of 3-ketosucrose-forming enzyme from the cells of *Agrobacterium tumefaciens*. *J. Biol. Chem.*, 242:3665–3672, 1967.
- [1577] M. Hayano and R.I. Dorfman. The action of adrenal homogenates on progesterone, 17-hydroxyprogesterone and 21-desoxycortisone. *Arch. Biochem. Biophys.*, 36:237–239, 1952.
- [1578] M. Hayano and R.I. Dorfman. On the mechanism of the C-11β-hydroxylation of steroids. *J. Biol. Chem.*, 211:227–235, 1954.
- [1579] M. Hayashi, K. Hasegawa, Y. Oguni, and T. Unemoto. Characterization of FMN-dependent NADH-quinone reductase induced by menadione in *Escherichia coli*. *Biochim. Biophys. Acta*, 1035:230–236, 1990.
- [1580] M. Hayashi, H. Ohzeki, H. Shimada, and T. Unemoto. NADPH-specific quinone reductase is induced by 2-methylene-4-butyrolactone in *Escherichia coli*. *Biochim. Biophys. Acta*, 1273:165–170, 1996.
- [1581] S. Hayashi, S. Nakamura, and M. Suzuki. *Corynebacterium* sarcosine oxidase: a unique enzyme having covalently-bound and noncovalently-bound flavins. *Biochem. Biophys. Res. Commun.*, 96:924–930, 1980.
- [1582] R.P. Hayes, B.N. Webb, A.K. Subramanian, M. Nissen, A. Popchok, L. Xun, and C. Kang. Structural and catalytic differences between two FADH₂-dependent monooxygenases: 2,4,5-TCP 4-monooxygenase (TftD) from *Burkholderia cepacia* AC1100 and 2,4,6-TCP 4-monooxygenase (TcpA) from *Cupriavidus necator* JMP134. *Int. J. Mol. Sci.*, 13:9769–9784, 2012.
- [1583] S.W. Haynes, X. Gao, Y. Tang, and C.T. Walsh. Assembly of asperlicin peptidyl alkaloids from anthranilate and tryptophan: a two-enzyme pathway generates heptacyclic scaffold complexity in asperlicin E. *J. Am. Chem. Soc.*, 134:17444–17447, 2012.
- [1584] G.W. Haywood and P.J. Large. Microbial oxidation of amines. Distribution, purification and properties of two primary-amine oxidases from the yeast *Candida boidinii* grown on amines as sole nitrogen source. *Biochem. J.*, 199:187–201, 1981.
- [1585] C. He and M. Knipp. Formation of nitric oxide from nitrite by the ferriheme *b* protein nitrophorin 7. *J. Am. Chem. Soc.*, 131:12042–12043, 2009.
- [1586] C. He, H. Ogata, and M. Knipp. Formation of the complex of nitrite with the ferriheme *b* β-barrel proteins nitrophorin 4 and nitrophorin 7. *Biochemistry*, 49:5841–5851, 2010.
- [1587] F. He, Y. Zhu, M. He, and Y. Zhang. Molecular cloning and characterization of the gene encoding squalene epoxidase in *Panax notoginseng*. *DNA Seq*, 19:270–273, 2008.
- [1588] H.Y. He, A.C. Henderson, Y.L. Du, and K.S. Ryan. Two-enzyme pathway links *l*-arginine to nitric oxide in *N*-nitroso biosynthesis. *J. Am. Chem. Soc.*, 141:4026–4033, 2019.

- [1589] J. He and C. Hertweck. Biosynthetic origin of the rare nitroaryl moiety of the polyketide antibiotic aureothin: involvement of an unprecedented *N*-oxygenase. *J. Am. Chem. Soc.*, 126:3694–3695, 2004.
- [1590] J. He, N. Magarvey, M. Pirae, and L.C. Vining. The gene cluster for chloramphenicol biosynthesis in *Streptomyces venezuelae* ISP5230 includes novel shikimate pathway homologues and a monomodular non-ribosomal peptide synthetase gene. *Microbiology*, 147:2817–2829, 2001.
- [1591] J. He, M. Muller, and C. Hertweck. Formation of the aureothin tetrahydrofuran ring by a bifunctional cytochrome P450 monooxygenase. *J. Am. Chem. Soc.*, 126:16742–16743, 2004.
- [1592] X.Y. He, S.Y. Yang, and H. Schulz. Cloning and expression of the *fadH* gene and characterization of the gene product 2,4-dienoyl coenzyme A reductase from *Escherichia coli*. *Eur. J. Biochem.*, 248:516–520, 1997.
- [1593] Y.F. He, B.Z. Li, Z. Li, P. Liu, Y. Wang, Q. Tang, J. Ding, Y. Jia, Z. Chen, L. Li, Y. Sun, X. Li, Q. Dai, C.X. Song, K. Zhang, C. He, and G.L. Xu. Tet-mediated formation of 5-carboxylcytosine and its excision by TDG in mammalian DNA. *Science*, 333:1303–1307, 2011.
- [1594] J.F. Head, S. Inouye, K. Teranishi, and O. Shimomura. The crystal structure of the photoprotein aequorin at 2.3 Å resolution. *Nature*, 405:372–376, 2000.
- [1595] R.J. Heath, N. Su, C.K. Murphy, and C.O. Rock. The enoyl-[acyl-carrier-protein] reductases FabI and FabL from *Bacillus subtilis*. *J. Biol. Chem.*, 275:40128–40133, 2000.
- [1596] S. Hecht, W. Eisenreich, P. Adam, S. Amslinger, K. Kis, A. Bacher, D. Arigoni, and F. Rohdich. Studies on the non-mevalonate pathway to terpenes: the role of the GcpE (IspG) protein. *Proc. Natl. Acad. Sci. USA*, 98:14837–14842, 2001.
- [1597] L.I. Hecker, Y. Tondeur, and J.G. Farrelly. Formation of ϵ -hydroxycaproate and ϵ -aminocaproate from *N*-nitrosohexamethyleneimine: evidence that microsomal α -hydroxylation of cyclic nitrosamines may not always involve the insertion of molecular oxygen into the substrate. *Chem. Biol. Interact.*, 49:235–248, 1984.
- [1598] R. Hedderich, A. Berkessel, and R.K. Thauer. Purification and properties of heterodisulfide reductase from *Methanobacterium thermoautotrophicum* (strain Marburg). *Eur. J. Biochem.*, 193:255–261, 1990.
- [1599] R. Hedderich, J. Koch, D. Linder, and R.K. Thauer. The heterodisulfide reductase from *Methanobacterium thermoautotrophicum* contains sequence motifs characteristic of pyridine-nucleotide-dependent thioredoxin reductases. *Eur. J. Biochem.*, 225:253–261, 1994.
- [1600] J. Hedegaard and I.C. Gunsalus. Mixed function oxidation. IV. An induced methylene hydroxylase in camphor oxidation. *J. Biol. Chem.*, 240:4038–4043, 1965.
- [1601] L. Hederstedt. Heme A biosynthesis. *Biochim. Biophys. Acta*, 1817:920–927, 2012.
- [1602] L. Hederstedt, A. Lewin, and M. Throne-Holst. Heme A synthase enzyme functions dissected by mutagenesis of *Bacillus subtilis* CtaA. *J. Bacteriol.*, 187:8361–8369, 2005.
- [1603] J.R. Heemstra, Walsh Jr., and C.T. Tandem action of the O₂⁻ and FADH₂-dependent halogenases KtzQ and KtzR produce 6,7-dichlorotryptophan for kutzneride assembly. *J. Am. Chem. Soc.*, 130:14024–14025, 2008.
- [1604] J. Hefner, S.M. Rubenstein, R.E. Ketchum, D.M. Gibson, R.M. Williams, and R. Croteau. Cytochrome P₄₅₀-catalyzed hydroxylation of taxa-4(5),11(12)-diene to taxa-4(20),11(12)-dien-5 α -ol: the first oxygenation step in taxol biosynthesis. *Chem. Biol.*, 3:479–489, 1996.
- [1605] J. Heider, M. Boll, K. Breese, S. Breinig, C. Ebenau-Jehle, U. Feil, N. Gad'on, D. Laempe, B. Leuthner, M.E. Mohamed, S. Schneider, G. Burchhardt, and G. Fuchs. Differential induction of enzymes involved in anaerobic metabolism of aromatic compounds in the denitrifying bacterium *Thauera aromatica*. *Arch. Microbiol.*, 170:120–131, 1998.
- [1606] J. Heider and G. Fuchs. Anaerobic metabolism of aromatic compounds. *Eur. J. Biochem.*, 243:577–596, 1997.
- [1607] J. Heider, X.H. Mai, and M.W.W. Adams. Characterization of 2-ketoisovalerate ferredoxin oxidoreductase, a new and reversible coenzyme A-dependent enzyme involved in peptide fermentation by hyperthermophilic archaea. *J. Bacteriol.*, 178:780–787, 1996.

- [1608] J. Heidlas, K.-H. Engel, and R. Tressl. Purification and characterization of two oxidoreductases involved in the enantioselective reduction of 3-oxo, 4-oxo and 5-oxo esters in baker's yeast. *Eur. J. Biochem.*, 172:633–639, 1988.
- [1609] J. Heidlas and R. Tressl. Purification and characterization of a (*R*)-2,3-butanediol dehydrogenase from *Saccharomyces cerevisiae*. *Arch. Microbiol.*, 154:267–273, 1990.
- [1610] I. Heilmann, S. Mekhedov, B. King, J. Browse, and J. Shanklin. Identification of the *Arabidopsis* palmitoyl-monogalactosyldiacylglycerol Δ^7 -desaturase gene FAD5, and effects of plastidial retargeting of *Arabidopsis* desaturases on the fad5 mutant phenotype. *Plant Physiol.*, 136:4237–4245, 2004.
- [1611] S. Heim, A. Kunkel, R.K. Thauer, and R. Hedderich. Thiol:fumarate reductase (Tfr) from *Methanobacterium thermoautotrophicum*. Identification of the catalytic sites for fumarate reduction and thiol oxidation. *Eur. J. Biochem.*, 253:292–299, 1998.
- [1612] A. Heinfling, F.J. Ruiz-Dueñas, M.J. Martínez, M. Bergbauer, U. Szewzyk, and A.T. Martínez. A study on reducing substrates of manganese-oxidizing peroxidases from *Pleurotus eryngii* and *Bjerkandera adusta*. *FEBS Lett.*, 428:141–146, 1998.
- [1613] N.K. Heinzinger, S.Y. Fujimoto, M.A. Clark, M.S. Moreno, and E.L. Barrett. Sequence analysis of the *phs* operon in *Salmonella typhimurium* and the contribution of thiosulfate reduction to anaerobic energy metabolism. *J. Bacteriol.*, 177:2813–2820, 1995.
- [1614] B. Heiss, K. Frunzke, and W.G. Zumpf. Formation of the N-N bond from nitric oxide by a membrane-bound cytochrome *bc* complex of nitrate-respiring (denitrifying) *Pseudomonas stutzeri*. *J. Bacteriol.*, 171:3288–3297, 1989.
- [1615] T. Heitz, E. Widemann, R. Lugan, L. Miesch, P. Ullmann, L. Desaubry, E. Holder, B. Grausem, S. Kandel, M. Miesch, D. Werck-Reichhart, and F. Pinot. Cytochromes P450 CYP94C1 and CYP94B3 catalyze two successive oxidation steps of plant hormone jasmonoyl-isoleucine for catabolic turnover. *J. Biol. Chem.*, 287:6296–6306, 2012.
- [1616] H.J. Hektor, H. Kloosterman, and L. Dijkhuizen. Identification of a magnesium-dependent NAD(P)(H)-binding domain in the nicotinoprotein methanol dehydrogenase from *Bacillus methanolicus*. *J. Biol. Chem.*, 277:46966–46973, 2002.
- [1617] J.J. Van Hellemond and A.G. Tielens. Expression and functional properties of fumarate reductase. *Biochem. J.*, 304:321–331, 1994.
- [1618] W. Heller, G. Forkmann, L. Britsch, and H. Grisebach. Enzymatic reduction of (+)-dihydroflavonols to flavan-3,4-*cis*-diols with flower extracts from *Matthiola incana* and its role in anthocyanin biosynthesis. *Planta*, 165:284–287, 1985.
- [1619] C.A. Helliwell, P.M. Chandler, A. Poole, E.S. Dennis, and W.J. Peacock. The CYP88A cytochrome P450, *ent*-kaurenoic acid oxidase, catalyzes three steps of the gibberellin biosynthesis pathway. *Proc. Natl. Acad. Sci. USA*, 98:2065–2070, 2001.
- [1620] C.A. Helliwell, A. Poole, W.J. Peacock, and E.S. Dennis. *Arabidopsis ent*-kaurene oxidase catalyzes three steps of gibberellin biosynthesis. *Plant Physiol.*, 119:507–510, 1999.
- [1621] V. Helmetag, S.A. Samel, M.G. Thomas, M.A. Marahiel, and L.O. Essen. Structural basis for the *erythro*-stereospecificity of the L-arginine oxygenase VioC in viomycin biosynthesis. *FEBS J.*, 276:3669–3682, 2009.
- [1622] C. Helvig, J.F. Koener, G.C. Unnithan, and R. Feyereisen. CYP15A1, the cytochrome P450 that catalyzes epoxidation of methyl farnesoate to juvenile hormone III in cockroach corpora allata. *Proc. Natl. Acad. Sci. USA*, 101:4024–4029, 2004.
- [1623] S. Hemmati, T.J. Schmidt, and E. Fuss. (+)-Pinoresinol/(−)-lariciresinol reductase from *Linum perenne* Himmelszelt involved in the biosynthesis of justicidin B. *FEBS Lett.*, 581:603–610, 2007.
- [1624] H. Hemmi, J.M. Studts, Y.K. Chae, J. Song, J.L. Markley, and B.G. Fox. Solution structure of the toluene 4-monooxygenase effector protein (T4moD). *Biochemistry*, 40:3512–3524, 2001.
- [1625] J. Hendriks, U. Gohlke, and M. Saraste. From NO to O₂: nitric oxide and dioxygen in bacterial respiration. *J. Bioenerg. Biomembr.*, 30:15–24, 1998.
- [1626] J. Hendriks, A. Warne, U. Gohlke, T. Haltia, C. Ludovici, M. Lubben, and M. Saraste. The active site of the bacterial nitric oxide reductase is a dinuclear iron center. *Biochemistry*, 37:13102–13109, 1998.

- [1627] M.G.L. Henquet, N. Prota, J.J.J. van der Hooft, M. Varbanova-Herde, R.J.M. Hulzink, M. de Vos, M. Prins, M.T.J. de Both, M.C.R. Franssen, H. Bouwmeester, and M. Jongsma. Identification of a drimenol synthase and drimenol oxidase from *Persicaria hydropiper*, involved in the biosynthesis of insect deterrent drimanes. *Plant J.*, 90:1052–1063, 2017.
- [1628] D. Herbert and J. Pinsent. Crystalline bacterial catalase. *Biochem. J.*, 43:193–202, 1948.
- [1629] D. Herbert and J. Pinsent. Crystalline human erythrocyte catalase. *Biochem. J.*, 43:203–205, 1948.
- [1630] H.G. Hers. L' Aldose-réductase. *Biochim. Biophys. Acta*, 37:120–126, 1960.
- [1631] L.B. Hersh, M.J. Stark, S. Worthen, and M.K. Fiero. N-Methylglutamate dehydrogenase: kinetic studies on the solubilized enzyme. *Arch. Biochem. Biophys.*, 150:219–226, 1972.
- [1632] M. Hetzel, M. Brock, T. Selmer, A.J. Pierik, B.T. Golding, and W. Buckel. Acryloyl-CoA reductase from *Clostridium propionicum*. An enzyme complex of propionyl-CoA dehydrogenase and electron-transferring flavoprotein. *Eur. J. Biochem.*, 270:902–910, 2003.
- [1633] D.P. Heuts, E.W. van Hellemond, D.B. Janssen, and M.W. Fraaije. Discovery, characterization, and kinetic analysis of an alditol oxidase from *Streptomyces coelicolor*. *J. Biol. Chem.*, 282:20283–20291, 2007.
- [1634] K.S. Hewitson, L.A. McNeill, M.V. Riordan, Y.M. Tian, A.N. Bullock, R.W. Welford, J.M. Elkins, N.J. Oldham, S. Bhattacharya, J.M. Gleadle, P.J. Ratcliffe, C.W. Pugh, and C.J. Schofield. Hypoxia-inducible factor (HIF) asparagine hydroxylase is identical to factor inhibiting HIF (FIH) and is related to the cupin structural family. *J. Biol. Chem.*, 277:26351–26355, 2002.
- [1635] M. Heydari, T. Ohshima, N. Nunoura-Kominato, and H. Sakuraba. Highly stable L-lysine 6-dehydrogenase from the thermophile *Geobacillus stearothermophilus* isolated from a Japanese hot spring: characterization, gene cloning and sequencing, and expression. *Appl. Environ. Microbiol.*, 70:937–942, 2004.
- [1636] M. Hibi, T. Kawashima, T. Kodera, S.V. Smirnov, P.M. Sokolov, M. Sugiyama, S. Shimizu, K. Yokozeki, and J. Ogawa. Characterization of *Bacillus thuringiensis* L-isoleucine dioxygenase for production of useful amino acids. *Appl. Environ. Microbiol.*, 77:6926–6930, 2011.
- [1637] M. Hibi, T. Kawashima, H. Yajima, S.V. Smirnov, T. Kodera, M. Sugiyama, S. Shimizu, K. Yokozeki, , and J. Enzymatic synthesis of chiral amino acid sulfoxides by Fe(II)/ α ketoglutarate-dependent dioxygenase. *Tetrahedron Asym.*, 24:990–994, 2013.
- [1638] Y. Hibi, K. Asai, H. Arafuka, M. Hamajima, T. Iwama, and K. Kawai. Molecular structure of La³⁺-induced methanol dehydrogenase-like protein in *Methylobacterium radiotolerans*. *J. Biosci. Bioeng.*, 111:547–549, 2011.
- [1639] J. Hickman and G. Ashwell. A sensitive and stereospecific enzymatic assay for xylulose. *J. Biol. Chem.*, 234:758–761, 1959.
- [1640] J. Hickman and G. Ashwell. Uronic acid metabolism in bacteria. II. Purification and properties of D-altronic acid and D-mannonic acid dehydrogenases in *Escherichia coli*. *J. Biol. Chem.*, 235:1566–1570, 1960.
- [1641] T. Hidaka, M. Goda, T. Kuzuyama, N. Takei, M. Hidaka, and H. Seto. Cloning and nucleotide sequence of fosfomycin biosynthetic genes of *Streptomyces wedmorensis*. *Mol. Gen. Genet.*, 249:274–280, 1995.
- [1642] R. Hidese, H. Mihara, T. Kurihara, and N. Esaki. *Escherichia coli* dihydropyrimidine dehydrogenase is a novel NAD-dependent heterotetramer essential for the production of 5,6-dihydrouracil. *J. Bacteriol.*, 193:989–993, 2011.
- [1643] A.D. Hieber, R.C. Bugos, and H.Y. Yamamoto. Plant lipocalins: violaxanthin de-epoxidase and zeaxanthin epoxidase. *Biochim. Biophys. Acta*, 1482:84–91, 2000.
- [1644] S. Higashi and N. Murata. An *in vivo* study of substrate specificities of acyl-lipid desaturases and acyltransferases in lipid synthesis in *Synechocystis* PCC6803. *Plant Physiol.*, 102:1275–1278, 1993.
- [1645] L.J. Higgins, F. Yan, P. Liu, H.W. Liu, and C.L. Drennan. Structural insight into antibiotic fosfomycin biosynthesis by a mononuclear iron enzyme. *Nature*, 437:838–844, 2005.
- [1646] T.P. Higgins, M. Davey, J. Trickett, D.P. Kelly, and J.C. Murrell. Metabolism of methanesulfonic acid involves a multi-component monooxygenase enzyme. *Microbiology*, 142:251–260, 1996.

- [1647] F.K. Higson and D.D. Focht. Degradation of 2-methylbenzoic acid by *Pseudomonas cepacia* MB2. *Appl. Environ. Microbiol.*, 58:194–200, 1992.
- [1648] M. Higuchi, M. Shimada, Y. Yamamoto, T. Hayashi, T. Koga, and Y. Kamio. Identification of two distinct NADH oxidases corresponding to H₂O₂-forming oxidase and H₂O-forming oxidase induced in *Streptococcus mutans*. *J. Gen. Microbiol.*, 139:2343–2351, 1993.
- [1649] Y. Higuchi, N. Yasuoka, M. Kakudo, Y. Katsube, T. Yagi, and H. Inokuchi. Single crystals of hydrogenase from *Desulfovibrio vulgaris* Miyazaki F. *J. Biol. Chem.*, 262:2823–2825, 1987.
- [1650] B.J. Hilbert, S.R. Grossman, C.A., Royer Schiffer, , and Jr. Crystal structures of human CtBP in complex with substrate MTOB reveal active site features useful for inhibitor design. *FEBS Lett.*, 588:1743–1748, 2014.
- [1651] T.M. Hildebrandt and M.K. Grieshaber. Three enzymatic activities catalyze the oxidation of sulfide to thiosulfate in mammalian and invertebrate mitochondria. *FEBS J.*, 275:3352–3361, 2008.
- [1652] R.K. Hill, S. Sawada, and S.M. Arfin. Stereochemistry of valine and isoleucine biosynthesis. IV. Synthesis, configuration, and enzymatic specificity of α -acetolactate and α -aceto- α -hydroxybutyrate. *Bioorg. Chem.*, 8:175–189, 1979.
- [1653] P.J. Hillas, F.S. del Alba, J. Oyarzabal, A. Wilks, and P.R. Ortiz De Montellano. The AhpC and AhpD antioxidant defense system of *Mycobacterium tuberculosis*. *J. Biol. Chem.*, 275:18801–18809, 2000.
- [1654] R. Hille. The mononuclear molybdenum enzymes. *Chem. Rev.*, 96:2757–2816, 1996.
- [1655] R. Hille, S. Dingwall, and J. Wilcoxon. The aerobic CO dehydrogenase from *Oligotropha carboxidovorans*. *J. Biol. Inorg. Chem.*, 20:243–251, 2015.
- [1656] P. Hillmer and G. Gottschalk. Solubilization and partial characterisation of particulate dehydrogenases from *Clostridium kluyveri*. *Biochim. Biophys. Acta*, 334:12–23, 1974.
- [1657] M.L. Hillwig and X. Liu. A new family of iron-dependent halogenases acts on freestanding substrates. *Nat. Chem. Biol.*, 10:921–923, 2014.
- [1658] M.L. Hillwig, Q. Zhu, K. Ittiamornkul, and X. Liu. Discovery of a promiscuous non-heme iron halogenase in ambiguine alkaloid biogenesis: implication for an evolvable enzyme family for late-stage halogenation of aliphatic carbons in small molecules. *Angew. Chem. Int. Ed. Engl.*, 55:5780–5784, 2016.
- [1659] M.L. Hillwig, Q. Zhu, and X. Liu. Biosynthesis of ambiguine indole alkaloids in cyanobacterium *Fischerella ambigua*. *ACS Chem. Biol.*, 9:372–377, 2014.
- [1660] H. Hilz, M. Kittler, and G. Knappe. Die Reduktion von Sulfate in der Hefe. *Biochem. Z.*, 332:151–166, 1959.
- [1661] W. Hinderer, U. Flentje, and W. Barz. Microsomal isoflavone 2'-hydroxylases and 3'-hydroxylases from chickpea (*Cicer arietinum* L) cell-suspensions induced for pterocarpan phytoalexin formation. *FEBS Lett.*, 214:101–106, 1987.
- [1662] V. Hines, L.D. Keys, Johnston III, and M. Purification and properties of the bovine liver mitochondrial dihydroorotate dehydrogenase. *J. Biol. Chem.*, 261:11386–11392, 1986.
- [1663] T. Hino, Y. Matsumoto, S. Nagano, H. Sugimoto, Y. Fukumori, T. Murata, S. Iwata, and Y. Shiro. Structural basis of biological N₂O generation by bacterial nitric oxide reductase. *Science*, 330:1666–1670, 2010.
- [1664] M. Hintz, A. Reichenberg, B. Altincicek, U. Bahr, R.M. Gschwind, A.-K. Kollas, E. Beck, J. Wiesner, M. Eberl, and H. Jomaa. Identification of (*E*)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human T cells in *Escherichia coli*. *FEBS Lett.*, 509:317–322, 2001.
- [1665] K. Hiraga and G. Kikuchi. The mitochondrial glycine cleavage system. Functional association of glycine decarboxylase and aminomethyl carrier protein. *J. Biol. Chem.*, 255:11671–11676, 1980.
- [1666] J. Hirano, K. Miyamoto, and H. Ohta. Purification and characterization of thermostable H₂O₂-forming NADH oxidase from 2-phenylethanol-assimilating *Brevibacterium* sp. KU1309. *Appl. Microbiol. Biotechnol.*, 80:71–78, 2008.
- [1667] S. Hirano and N. Masuda. Characterization of NADP-dependent 7 β -hydroxysteroid dehydrogenases from *Peptostreptococcus productus* and *Eubacterium aerofaciens*. *Appl. Environ. Microbiol.*, 43:1057–1063, 1982.

- [1668] F. Hirata, T. Ohnishi, and O. Hayaishi. Indoleamine 2,3-dioxygenase. Characterization and properties of enzyme. O₂⁻ complex. *J. Biol. Chem.*, 252:4637–4642, 1977.
- [1669] K. Hirata, C. Poeaknapo, J. Schmidt, and M.H. Zenk. 1,2-Dehydroreticuline synthase, the branch point enzyme opening the morphinan biosynthetic pathway. *Phytochemistry*, 65:1039–1046, 2004.
- [1670] T. Hirata, Y. Tamura, N. Yokobatake, K. Shimoda, and Y. Ashida. A 38 kDa allylic alcohol dehydrogenase from the cultured cells of *Nicotiana tabacum*. *Phytochemistry*, 55:297–303, 2000.
- [1671] T. Hiratsuka, K. Furihata, J. Ishikawa, H. Yamashita, N. Itoh, H. Seto, and T. Dairi. An alternative menaquinone biosynthetic pathway operating in microorganisms. *Science*, 321:1670–1673, 2008.
- [1672] K. Hirokawa and N. Kajiyama. Recombinant agrobacterium AgaE-like protein with fructosyl amino acid oxidase activity. *Biosci. Biotechnol. Biochem.*, 66:2323–2329, 2002.
- [1673] R. Hirota-Mamoto, R. Nagai, S. Tachibana, M. Yasuda, A. Tani, K. Kimbara, and F. Kawai. Cloning and expression of the gene for periplasmic poly(vinyl alcohol) dehydrogenase from *Sphingomonas* sp. strain 113P3, a novel-type quino-haemoprotein alcohol dehydrogenase. *Microbiology*, 152:1941–1949, 2006.
- [1674] W. Hirsch, H. Schägger, and G. Fuchs. Phenylglyoxylate:NAD⁺ oxidoreductase (CoA benzoylating), a new enzyme of anaerobic phenylalanine metabolism in the denitrifying bacterium *Axoarcus Evansii*. *Eur. J. Biochem.*, 251:907–915, 1998.
- [1675] W.D. Hitz, T.J. Carlson, J.R. Booth, Kinney Jr., Stecca A.J., Yadav K.L., and N.S. Cloning of a higher-plant plastid ω-6 fatty acid desaturase cDNA and its expression in a cyanobacterium. *Plant Physiol.*, 105:635–641, 1994.
- [1676] K. Hlouchova, J. Rudolph, J.M. Pietari, L.S. Behlen, and S.D. Copley. Pentachlorophenol hydroxylase, a poorly functioning enzyme required for degradation of pentachlorophenol by *Sphingobium chlorophenolicum*. *Biochemistry*, 51:3848–3860, 2012.
- [1677] M.E. Hobbs, M. Vetting, H.J. Williams, T. Narindoshvili, D.M. Kebodeaux, B. Hillerich, R.D. Seidel, S.C. Almo, and F.M. Raushel. Discovery of an L-fuco-1,5-lactonase from cog3618 of the amidohydrolase superfamily. *Biochemistry*, 52:239–253, 2013.
- [1678] M.E. Hobbs, H.J. Williams, B. Hillerich, S.C. Almo, and F.M. Raushel. L-Galactose metabolism in *Bacteroides vulgatus* from the human gut microbiota. *Biochemistry*, 53:4661–4670, 2014.
- [1679] U. Hoch, Z. Zhang, D.L. Kroetz, and P.R. Ortiz de Montellano. Structural determination of the substrate specificities and regioselectivities of the rat and human fatty acid ω-hydroxylases. *Arch. Biochem. Biophys.*, 373:63–71, 2000.
- [1680] A. Hochman and I. Goldberg. Purification and characterization of a catalase-peroxidase and a typical catalase from the bacterium *Klebsiella pneumoniae*. *Biochim. Biophys. Acta*, 1077:299–307, 1991.
- [1681] L.I. Hochstein and B.P. Dalton. The purification and properties of nicotine oxidase. *Biochim. Biophys. Acta*, 139:56–68, 1967.
- [1682] L.I. Hochstein and S.C. Rittenberg. The bacterial oxidation of nicotine. II. The isolation of the first oxidative product and its identification as (1)-6-hydroxynicotine. *J. Biol. Chem.*, 234:156–160, 1959.
- [1683] D.S. Hodgins and R.H. Abeles. Studies of the mechanism of action of D-proline reductase: the presence on covalently bound pyruvate and its role in the catalytic process. *Arch. Biochem. Biophys.*, 130:274–285, 1969.
- [1684] S. Hofbauer, G. Mlynek, L. Milazzo, D. Pühringer, D. Maresch, I. Schaffner, P.G. Furtmüller, G. Smulevich, K. Djinovic-Carugo, and C. Obinger. Hydrogen peroxide-mediated conversion of coproheme to heme *b* by HemQ—lessons from the first crystal structure and kinetic studies. *FEBS J.*, 283:4386–4401, 2016.
- [1685] B. Hofer, L.D. Eltis, D.N. Dowling, and K.N. Timmis. Genetic analysis of a *Pseudomonas* locus encoding a pathway for biphenyl/polychlorinated biphenyl degradation. *Gene*, 130:47–55, 1993.
- [1686] H.W. Hoffken, M. Duong, T. Friedrich, M. Breuer, B. Hauer, R. Reinhardt, R. Rabus, and J. Heider. Crystal structure and enzyme kinetics of the (*S*)-specific 1-phenylethanol dehydrogenase of the denitrifying bacterium strain EbN1. *Biochemistry*, 45:82–93, 2006.

- [1687] M. Hofrichter, R. Ullrich, M.J. Pecyna, C. Liers, and T. Lundell. New and classic families of secreted fungal heme peroxidases. *Appl. Microbiol. Biotechnol.*, 87:871–897, 2010.
- [1688] W. Hohnloser, B. Osswald, and F. Lingens. Enzymological aspects of caffeine demethylation and formaldehyde oxidation by *Pseudomonas putida* C1. *Hoppe-Seyler's Z. Physiol. Chem.*, 361:1763–1766, 1980.
- [1689] J.J. Holbrook, A. Liljas, S.J. Steindel, and M.G. Rossmann. Lactate dehydrogenase. In P.D. Boyer, editor, *The Enzymes*, volume 11, pages 191–292. Academic Press, New York, 3rd edition, 1975.
- [1690] J.S. Holcenberg and E.R. Stadtman. Nicotinic acid metabolism. 3. Purification and properties of a nicotinic acid hydroxylase. *J. Biol. Chem.*, 244:1194–1203, 1969.
- [1691] J.S. Holcenberg and L. Tsai. Nicotinic acid metabolism. IV. Ferredoxin-dependent reduction of 6-hydroxynicotinic acid to 6-oxo-1,4,5,6-tetrahydronicotinic acid. *J. Biol. Chem.*, 244:1204–1211, 1969.
- [1692] J.K. Holden, N. Lim, and T.L. Poulos. Identification of redox partners and development of a novel chimeric bacterial nitric oxide synthase for structure activity analyses. *J. Biol. Chem.*, 289:29437–29445, 2014.
- [1693] P.G. Holder, L.C. Jones, P.M. Drake, R.M. Barfield, S. Banas, G.W. de Hart, J. Baker, and D. Rabuka. Reconstitution of formylglycine-generating enzyme with copper(II) for aldehyde tag conversion. *J. Biol. Chem.*, 290:15730–15745, 2015.
- [1694] M.M. Holdorf, H.A. Owen, S.R. Lieber, L. Yuan, N. Adams, C. Dabney-Smith, and C.A. Makaroff. *Arabidopsis* ETHE1 encodes a sulfur dioxygenase that is essential for embryo and endosperm development. *Plant Physiol.*, 160:226–236, 2012.
- [1695] U.H. Hole, K.U. Vollack, W.G. Zumft, E. Eisenmann, R.A. Siddiqui, B. Friedrich, and P.M.H. Kroneck. Characterization of the membranous denitrification enzymes nitrite reductase (cytochrome cd1) and copper-containing nitrous oxide reductase from *Thiobacillus denitrificans*. *Arch. Microbiol.*, 165:55–61, 1996.
- [1696] C. Holliger, G. Wohlfarth, and G. Diekert. Reductive dechlorination in the energy metabolism of anaerobic bacteria. *FEMS Microbiol. Rev.*, 22:383–398, 1998.
- [1697] S. Hollmann and O. Touster. The L-xylulose-xylitol enzyme and other polyol dehydrogenases of guinea pig liver mitochondria. *J. Biol. Chem.*, 225:87–102, 1957.
- [1698] I. Holmberg-Betsholtz, E. Lund, I. Björkhem, and K. Wikvall. Sterol 27-hydroxylase in bile acid biosynthesis. Mechanism of oxidation of 5 β -cholestane-3 α ,7 α ,12 α ,27-tetrol into 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanic acid. *J. Biol. Chem.*, 268:11079–11085, 1993.
- [1699] A.J. Holmes, A. Costello, M.E. Lidstrom, and J.C. Murrell. Evidence that particulate methane monooxygenase and ammonia monooxygenase may be evolutionarily related. *FEMS Microbiol. Lett.*, 132:203–208, 1995.
- [1700] P.E. Holmes and S.C. Rittenberg. The bacterial oxidation of nicotine. VII. Partial purification and properties of 2,6-dihydroxypyridine oxidase. *J. Biol. Chem.*, 247:7622–7627, 1972.
- [1701] P.E. Holmes, S.C. Rittenberg, and H.J. Knackmuss. The bacterial oxidation of nicotine. 8. Synthesis of 2,3,6-trihydroxypyridine and accumulation and partial characterization of the product of 2,6-dihydroxypyridine oxidation. *J. Biol. Chem.*, 247:7628–7633, 1972.
- [1702] A. Holmgren and F. Aslund. Glutaredoxin. *Methods Enzymol.*, 252:283–292, 1995.
- [1703] C.M. Holsclaw, K.M. Sogi, S.A. Gilmore, M.W. Schelle, M.D. Leavell, C.R. Bertozzi, and J.A. Leary. Structural characterization of a novel sulfated menaquinone produced by *stf3* from *Mycobacterium tuberculosis*. *ACS Chem. Biol.*, 3:619–624, 2008.
- [1704] H. Holzer and A. Holldorf. Isolation of a D-glycerate dehydrogenase, its properties, and its use for the optical determination of hydroxypyruvate in the presence of pyruvate. *Biochem. Z.*, 329:292–312, 1957.
- [1705] H. Holzer and S. Schneider. Reinigung und charakterisierung einer TPN-abhängigen Pyridoxol-dehydrogenase aus bierhefe. *Biochim. Biophys. Acta*, 48:71–76, 1961.

- [1706] L. Hong, Z. Zhao, C.E. Melancon, Zhang 3rd, Liu H., and H.W. *In vitro* characterization of the enzymes involved in TDP-D-forosamine biosynthesis in the spinosyn pathway of *Saccharopolyspora spinosa*. *J. Am. Chem. Soc.*, 130:4954–4967, 2008.
- [1707] S. Hong, Y.W. Cho, L.R. Yu, H. Yu, T.D. Veenstra, and K. Ge. Identification of JmjC domain-containing UTX and JMJD3 as histone H3 lysine 27 demethylases. *Proc. Natl. Acad. Sci. USA*, 104:18439–18444, 2007.
- [1708] K. Honjo, T. Ishibashi, and Y. Imai. Partial purification and characterization of lathosterol 5-desaturase from rat liver microsomes. *J. Biochem.*, 97:955–959, 1985.
- [1709] K.L. Hooper, B. Joneja, H.B. White, Thorpe 3rd, and C. A. A sulfhydryl oxidase from chicken egg white. *J. Biol. Chem.*, 271:30510–30516, 1996.
- [1710] A.B. Hooper and C. Balny. Reaction of oxygen with hydroxylamine oxidoreductase of *Nitrosomonas*: fast kinetics. *FEBS Lett.*, 144:299–303, 1982.
- [1711] A.B. Hooper and K.R. Terry. Hydroxylamine oxidoreductase of *Nitrosomonas*. Production of nitric oxide from hydroxylamine. *Biochim. Biophys. Acta*, 571:12–20, 1979.
- [1712] M.F. Hopgood and D.J. Walker. Succinic acid production by rumen bacteria. III. Enzymic studies on the formation of succinate by *Ruminococcus flavefaciens*. *Aust. J. Biol. Sci.*, 22:1413–1424, 1969.
- [1713] R.P. Hopkins, E.C. Drummond, and P. Callaghan. Dehydrogenation of *trans*-acenaphthene-1,2-diol by liver cytosol preparations. *Biochem. Soc. Trans.*, 1:989–991, 1973.
- [1714] T.A. Hopkins, H.H. Seliger, E.H. White, and M.W. Cass. The chemiluminescence of firefly luciferin. A model for the bioluminescent reaction and identification of the product excited state. *J. Am. Chem. Soc.*, 89:7148–7150, 1967.
- [1715] D.J. Hopper. Oxygenase properties of the (4-hydroxybenzoyl)methanol-cleavage enzyme from an *Alcaligenes* sp. *Biochem. J.*, 239:469–472, 1986.
- [1716] D.J. Hopper, M.R. Jones, and M.J. Causer. Periplasmic location of *p*-cresol methylhydroxylase in *Pseudomonas putida*. *FEBS Lett.*, 182:485–488, 1985.
- [1717] D.J. Hopper and M.A. Kaderbhai. The quinohaemoprotein lupanine hydroxylase from *Pseudomonas putida*. *Biochim. Biophys. Acta*, 1647:110–115, 2003.
- [1718] D.J. Hopper, J. Rogozinski, and M. Toczko. Lupanine hydroxylase, a quinocytochrome *c* from an alkaloid-degrading *Pseudomonas* sp. *Biochem. J.*, 279:105–109, 1991.
- [1719] D.J. Hopper and D.G. Taylor. The purification and properties of *p*-cresol-(acceptor) oxidoreductase (hydroxylating), a flavocytochrome from *Pseudomonas putida*. *Biochem. J.*, 167:155–162, 1977.
- [1720] B.L. Horecker. Triphosphopyridine nucleotide-cytochrome *c* reductase in liver. *J. Biol. Chem.*, 183:593–605, 1950.
- [1721] K. Hori, J.M. Anderson, W.W. Ward, and M.J. Cormier. *Renilla* luciferin as the substrate for calcium induced photo-protein bioluminescence. Assignment of luciferin tautomers in aequorin and mnemiopsin. *Biochemistry*, 14:2371–2376, 1975.
- [1722] A. Horie, T. Tomita, A. Saiki, H. Kono, H. Taka, R. Mineki, T. Fujimura, C. Nishiyama, T. Kuzuyama, and M. Nishiyama. Discovery of proteinaceous *N*-modification in lysine biosynthesis of *Thermus thermophilus*. *Nat. Chem. Biol.*, 5:673–679, 2009.
- [1723] M. Horinouchi, T. Hayashi, T. Yamamoto, and T. Kudo. A new bacterial steroid degradation gene cluster in *Comamonas testosteroni* TA441 which consists of aromatic-compound degradation genes for seco-steroids and 3-ketosteroid dehydrogenase genes. *Appl. Environ. Microbiol.*, 69:4421–4430, 2003.
- [1724] M. Horinouchi, K. Kasuga, H. Nojiri, H. Yamane, and T. Omori. Cloning and characterization of genes encoding an enzyme which oxidizes dimethyl sulfide in *Acinetobacter* sp. strain 20B. *FEMS Microbiol. Lett.*, 155:99–105, 1997.
- [1725] M. Horinouchi, T. Yoshida, H. Nojiri, H. Yamane, and T. Omori. Polypeptide requirement of multicomponent monooxygenase DsoABCDEF for dimethyl sulfide oxidizing activity. *Biosci. Biotechnol. Biochem.*, 63:1765–1771, 1999.

- [1726] T. Horiuchi. Purification and properties of *N*-acyl-D-hexosamine oxidase from *Pseudomonas* sp. 15-1. *Agric. Biol. Chem.*, 53:361–368, 1989.
- [1727] T. Horiuchi and T. Kurokawa. Purification and properties of *N*-acyl-D-mannosamine dehydrogenase from *Flavobacterium* sp. 141-8. *J. Biochem. (Tokyo)*, 104:466–471, 1988.
- [1728] T. Horiuchi and T. Kurokawa. Purification and characterization of *N*-acetyl-D-hexosamine dehydrogenase from *Pseudomonas* sp no 53. *Agric. Biol. Chem.*, 53:1919–1925, 1989.
- [1729] T. Horiuchi, T. Kurokawa, and N. Saito. Purification and properties of fructosyl-amino acid oxidase from *Corynebacterium* sp. 2-4-1. *Agr Biol Chem Tokyo*, 53:103–110, 1989.
- [1730] K. Hormann and J.R. Andreesen. Reductive cleavage of sarcosine and betaine by *Eubacterium acidaminophilum* via enzyme systems different from glycine reductase. *Arch. Microbiol.*, 153:50–59, 1989.
- [1731] K. Hormann and J.R. Andreesen. Purification and characterization of a pyrrole-2-carboxylate oxygenase from *Arthrobacter* strain Py1. *Biol. Chem. Hoppe-Seyler*, 375:211–218, 1994.
- [1732] E. Hornung, C. Pernstich, and I. Feussner. Formation of conjugated $\Delta^{11}\Delta^{13}$ -double bonds by Δ^{12} -linoleic acid (1,4)-acyl-lipid-desaturase in pomegranate seeds. *Eur. J. Biochem.*, 269:4852–4859, 2002.
- [1733] S. Hörtensteiner. Chlorophyll degradation during senescence. *Annu. Rev. Plant Biol.*, 57:55–77, 2006.
- [1734] S. Hörtensteiner, K.L. Wüthrich, P. Matile, K.H. Ongania, and B. Kräutler. The key step in chlorophyll breakdown in higher plants. Cleavage of pheophorbide *a* macrocycle by a monooxygenase. *J. Biol. Chem.*, 273:15335–15339, 1998.
- [1735] K. Hoshino. Organism producing isopropanol from acetone. V. Enzymological [studies] on the oxidation-reduction of *Lactobacillus brevis* var. *hofuensis*. [in Japanese]. *Nippon Nogei Kagaku Kaishi*, 34:608–615, 1960.
- [1736] K. Hoshino and K. Udagawa. Organism producing isopropanol from acetone. VI. Isopropanol dehydrogenase and alcohol dehydrogenase of *Lactobacillus brevis* var. *hofuensis*. [in Japanese]. *Nippon Nogei Kagaku Kaishi*, 34:616–619, 1960.
- [1737] Y. Hoshino, S. Fujii, H. Shinonaga, K. Arai, F. Saito, T. Fukai, H. Satoh, Y. Miyazaki, and J. Ishikawa. Monooxygenation of rifampicin catalyzed by the *rox* gene product of *Nocardia farcinica*: structure elucidation, gene identification and role in drug resistance. *J. Antibiot. (Tokyo)*, 63:23–28, 2010.
- [1738] D.D. Hoskins and C.G. MacKenzie. Solubilization and electron transfer flavoprotein requirement of mitochondrial sarcosine dehydrogenase and dimethylglycine dehydrogenase. *J. Biol. Chem.*, 236:177–183, 1961.
- [1739] K. Hosokawa and R.Y. Stanier. Crystallization and properties of *p*-hydroxybenzoate hydroxylase from *Pseudomonas putida*. *J. Biol. Chem.*, 241:2453–2460, 1966.
- [1740] T. Hosoya, Y. Kondo, and N. Ui. Peroxidase activity in thyroid gland and partial purification of the enzyme. *J. Biochem. (Tokyo)*, 52:180–189, 1962.
- [1741] A. Hosseini, M. Brouk, M.F. Lucas, F. Glaser, A. Fishman, and V. Guallar. Atomic picture of ligand migration in toluene 4-monooxygenase. *J. Phys. Chem. B*, 119:671–678, 2015.
- [1742] G. Houen. Mammalian Cu-containing amine oxidases (CAOs): new methods of analysis, structural relationships, and possible functions. *APMIS Suppl.*, 96:1–46, 1999.
- [1743] A.R. Howard-Jones and C.T. Walsh. Enzymatic generation of the chromopyrrolic acid scaffold of rebeccamycin by the tandem action of RebO and RebD. *Biochemistry*, 44:15652–15663, 2005.
- [1744] A.R. Howard-Jones and C.T. Walsh. Staurosporine and rebeccamycin aglycones are assembled by the oxidative action of StaP, StaC, and RebC on chromopyrrolic acid. *J. Am. Chem. Soc.*, 128:12289–12298, 2006.
- [1745] L.G. Howell, T. Spector, and V. Massey. Purification and properties of *p*-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*. *J. Biol. Chem.*, 247:4340–4350, 1972.
- [1746] A.S.L. Hu and A.L. Cline. The regulation of some sugar dehydrogenases in a pseudomonad. *Biochim. Biophys. Acta*, 93:237–245, 1964.

- [1747] X. Hu, R. Mamoto, Y. Fujioka, A. Tani, K. Kimbara, and F. Kawai. The pva operon is located on the megaplasmid of *Sphingopyxis* sp. strain 113P3 and is constitutively expressed, although expression is enhanced by PVA. *Appl. Microbiol. Biotechnol.*, 78:685–693, 2008.
- [1748] Y. Hu, A. Al-Mestarihi, C.L. Grimes, D. Kahne, and B.O. Bachmann. A unifying nitrososynthase involved in nitrosugar biosynthesis. *J. Am. Chem. Soc.*, 130:15756–15757, 2008.
- [1749] Y. Hu, W. Liu, S.R. Malwal, Y. Zheng, X. Feng, T.P. Ko, C.C. Chen, Z. Xu, M. Liu, X. Han, J. Gao, E. Oldfield, and R.T. Guo. Structures of iridoid synthase from *Catharanthus roseus* with bound NAD(+) , NADPH, or NAD(+)/10-oxogeranial: Reaction mechanisms. *Angew. Chem. Int. Ed. Engl.*, 54:15478–15482, 2015.
- [1750] D.-Y. Huang, A. Furukawa, and Y. Ichikawa. Molecular cloning of retinal oxidase/aldehyde oxidase cDNAs from rabbit and mouse livers and functional expression of recombinant mouse retinal oxidase cDNA in *Escherichia coli*. *Arch. Biochem. Biophys.*, 364:264–272, 1999.
- [1751] F. Huang, D. Spiteller, N.A. Koorbanally, Y. Li, N.M. Llewellyn, and J.B. Spencer. Elaboration of neosamine rings in the biosynthesis of neomycin and butirosin. *ChemBioChem*, 8:283–288, 2007.
- [1752] F.C. Huang, A. Peter, and W. Schwab. Expression and characterization of CYP52 genes involved in the biosynthesis of sophorolipid and alkane metabolism from *Starmerella bombicola*. *Appl. Environ. Microbiol.*, 80:766–776, 2014.
- [1753] H. Huang, M.S. Carter, M.W. Vetting, N. Al-Obaidi, Y. Patskovsky, S.C. Almo, and J.A. Gerlt. A general strategy for the discovery of metabolic pathways: D-threitol, L-threitol, and erythritol utilization in *Mycobacterium smegmatis*. *J. Am. Chem. Soc.*, 137:14570–14573, 2015.
- [1754] J. Huang, Y. Zhong, G. Sandmann, J. Liu, and F. Chen. Cloning and selection of carotenoid ketolase genes for the engineering of high-yield astaxanthin in plants. *Planta*, 236:691–699, 2012.
- [1755] L.X. Huang, R.J. Rohlf, and R. Hille. The reaction of trimethylamine dehydrogenase with electron transferring flavo-protein. *J. Biol. Chem.*, 270:23958–23965, 1995.
- [1756] Y. Huang, R. Xun, G. Chen, and L. Xun. Maintenance role of a glutathionyl-hydroquinone lyase (PcpF) in pentachlorophenol degradation by *Sphingobium chlorophenolicum* ATCC 39723. *J. Bacteriol.*, 190:7595–7600, 2008.
- [1757] Y. Huang, K.X. Zhao, X.H. Shen, M.T. Chaudhry, C.Y. Jiang, and S.J. Liu. Genetic characterization of the resorcinol catabolic pathway in *Corynebacterium glutamicum*. *Appl. Environ. Microbiol.*, 72:7238–7245, 2006.
- [1758] B.K. Hubbard, M.G. Thomas, and C.T. Walsh. Biosynthesis of L-p-hydroxyphenylglycine, a non-proteinogenic amino acid constituent of peptide antibiotics. *Chem. Biol.*, 7:931–942, 2000.
- [1759] P.A. Hubbard, X. Liang, H. Schulz, and J.J. Kim. The crystal structure and reaction mechanism of *Escherichia coli* 2,4-dienoyl-CoA reductase. *J. Biol. Chem.*, 278:37553–37560, 2003.
- [1760] H.J. Hübener and F.G. Sahrholz. 20 β -hydroxy-steroid-dehydrogenase. II. Darstellung und Kristallisation. *Biochem. Z.*, 333:95–105, 1960.
- [1761] H.J. Hübener, F.G. Sahrholz, J. Schmidt-Thomé, G. Neseemann, and R. Junk. 20 β -Hydroxy-Steroid-Dehydrogenase, ein neues kristallines Enzym. *Biochim. Biophys. Acta*, 35:270–272, 1959.
- [1762] A.J. Hudson, S.C. Andrews, C. Hawkins, J.M. Williams, M. Izuhara, F.C. Meldrum, S. Mann, P.M. Harrison, and J.R. Guest. Overproduction, purification and characterization of the *Escherichia coli* ferritin. *Eur. J. Biochem.*, 218:985–995, 1993.
- [1763] G.W. Huffman, P.D. Gesellchen, J.R. Turner, R.B. Rothenberger, H.E. Osborne, F.D. Miller, J.L. Chapman, and S.W. Queener. Substrate specificity of isopenicillin N synthase. *J. Med. Chem.*, 35:1897–1914, 1992.
- [1764] M. Hugler, C. Menendez, H. Schagger, and G. Fuchs. Malonyl-coenzyme A reductase from *Chloroflexus aurantiacus*, a key enzyme of the 3-hydroxypropionate cycle for autotrophic CO₂ fixation. *J. Bacteriol.*, 184:2404–2410, 2002.
- [1765] M. Hugo, K. Van Laer, A.M. Reyes, D. Vertommen, J. Messens, R. Radi, and M. Trujillo. Mycothiol/mycoeredoxin 1-dependent reduction of the peroxiredoxin AhpE from *Mycobacterium tuberculosis*. *J. Biol. Chem.*, 289:5228–5239, 2014.

- [1766] M. Hugo, L. Turell, B. Manta, H. Botti, G. Monteiro, L.E. Netto, B. Alvarez, R. Radi, and M. Trujillo. Thiol and sulfenic acid oxidation of AhpE, the one-cysteine peroxiredoxin from *Mycobacterium tuberculosis*: kinetics, acidity constants, and conformational dynamics. *Biochemistry*, 48:9416–9426, 2009.
- [1767] W.K. Huh, S.T. Kim, K.S. Yang, Y.J. Seok, Y.C. Hah, and S.O. Kang. Characterisation of D-arabinono-1,4-lactone oxidase from *Candida albicans* ATCC 10231. *Eur. J. Biochem.*, 225:1073–1079, 1994.
- [1768] W.K. Huh, B.H. Lee, S.T. Kim, Y.R. Kim, G.E. Rhie, Y.W. Baek, C.S. Hwang, J.S. Lee, and S.O. Kang. D-Erythroascorbic acid is an important antioxidant molecule in *Saccharomyces cerevisiae*. *Mol. Microbiol.*, 30:895–903, 1998.
- [1769] D.H. Huizinga, R. Denton, K.G. Koehler, A. Tomasello, L. Wood, S.E. Sen, and D.N. Crowell. Farnesylcysteine lyase is involved in negative regulation of abscisic acid signaling in *Arabidopsis*. *Mol Plant*, 3:143–155, 2010.
- [1770] A.K. Hull, R. Vij, and J.L. Celenza. *Arabidopsis* cytochrome P_{450} s that catalyze the first step of tryptophan-dependent indole-3-acetic acid biosynthesis. *Proc. Natl. Acad. Sci. USA*, 97:2379–2384, 2000.
- [1771] J.D. Hulse, S.R. Ellis, and L.M. Henderson. Carnitine biosynthesis. β -Hydroxylation of trimethyllysine by an α -ketoglutarate-dependent mitochondrial dioxygenase. *J. Biol. Chem.*, 253:1654–1659, 1978.
- [1772] G.F. Humphrey. The distribution and properties of transhydrogenase from animal tissues. *Biochem. J.*, 65:546–550, 1957.
- [1773] R.J. Hung, C.W. Pak, and J.R. Terman. Direct redox regulation of F-actin assembly and disassembly by Mical. *Science*, 334:1710–1713, 2011.
- [1774] R.J. Hung, C.S. Spaeth, H.G. Yesilyurt, and J.R. Terman. SelR reverses Mical-mediated oxidation of actin to regulate F-actin dynamics. *Nat. Cell Biol.*, 15:1445–1454, 2013.
- [1775] R.J. Hung, U. Yazdani, J. Yoon, H. Wu, T. Yang, N. Gupta, Z. Huang, W.J. van Berkel, and J.R. Terman. Mical links semaphorins to F-actin disassembly. *Nature*, 463:823–827, 2010.
- [1776] S.W. Hutcheson and T. Kosuge. Regulation of 3-indoleacetic acid production in *Pseudomonas syringae* pv. savastanoi. Purification and properties of tryptophan 2-monooxygenase. *J. Biol. Chem.*, 260:6281–6287, 1985.
- [1777] J.J. Hutton, Tappel Jr., Udenfriend A.L., and S. Cofactor and substrate requirements of collagen proline hydroxylase. *Arch. Biochem. Biophys.*, 118:231–240, 1967.
- [1778] J. Hutzler and J. Dancis. Conversion of lysine to saccharopine by human tissues. *Biochim. Biophys. Acta*, 158:62–69, 1968.
- [1779] M.R. Hyman, C.L. Page, and D.J. Arp. Oxidation of methyl fluoride and dimethyl ether by ammonia monooxygenase in *Nitrosomonas europaea*. *Appl. Environ. Microbiol.*, 60:3033–3035, 1994.
- [1780] M.R. Hyman and P.M. Wood. Methane oxidation by *Nitrosomonas europaea*. *Biochem. J.*, 212:31–37, 1983.
- [1781] Y.L. Hyun and V.L. Davidson. Electron transfer reactions between aromatic amine dehydrogenase and azurin. *Biochemistry*, 34:12249–12254, 1995.
- [1782] Y.L. Hyun, Z. Zhu, and V.L. Davidson. Gated and ungated electron transfer reactions from aromatic amine dehydrogenase to azurin. *J. Biol. Chem.*, 274:29081–29086, 1999.
- [1783] K. Iba, S. Gibson, T. Nishiuchi, T. Fuse, M. Nishimura, V. Arondel, S. Hugly, and C. Somerville. A gene encoding a chloroplast ω -3 fatty acid desaturase complements alterations in fatty acid desaturation and chloroplast copy number of the fad7 mutant of *Arabidopsis thaliana*. *J. Biol. Chem.*, 268:24099–24105, 1993.
- [1784] M. Ibdah, A. Berim, S. Martens, A.L.H. Valderrama, L. Palmieri, E. Lewinsohn, and D.R. Gang. Identification and cloning of an NADPH-dependant hydroxycinnamoyl-CoA double bond reductase involved in dihydrochalcone formation in *Malus X domestica* Borkh. *Phytochemistry*, 107:24–31–, 2014.
- [1785] O. Ibraheem, I.O. Adewale, and A. Afolayan. Purification and properties of glucose 6-phosphate dehydrogenase from *Aspergillus aculeatus*. *J. Biochem. Mol. Biol.*, 38:584–590, 2005.

- [1786] K. Ichida, Y. Amaya, K. Noda, S. Minoshima, T. Hosoya, O. Sakai, N. Shimizu, and T. Nishino. Cloning of the cDNA encoding human xanthine dehydrogenase (oxidase): structural analysis of the protein and chromosomal location of the gene. *Gene*, 133:279–284, 1993.
- [1787] A. Ichihara and E.A. Ichihara. Metabolism of L-lysine by bacterial enzymes. V. Glutaric semialdehyde dehydrogenase. *J. Biochem. (Tokyo)*, 49:154–157, 1961.
- [1788] A. Ichiyama, S. Nakamura, H. Kawai, T. Honjo, Y. Nishizuka, O. Hayaishi, and S. Senoh. Studies on the metabolism of the benzene ring of tryptophan in mammalian tissues. II. Enzymic formation of α -aminomuconic acid from 3-hydroxyanthranilic acid. *J. Biol. Chem.*, 240:740–749, 1965.
- [1789] A. Ichiyama, S. Nakamura, Y. Nishizuka, and O. Hayaishi. Enzymic studies on the biosynthesis of serotonin in mammalian brain. *J. Biol. Chem.*, 245:1699–1709, 1970.
- [1790] C. Ignea, A. Athanasakoglou, E. Ioannou, P. Georgantea, F.A. Trika, S. Loupassaki, V. Roussis, A.M. Makris, and S.C. Kampranis. Carnosic acid biosynthesis elucidated by a synthetic biology platform. *Proc. Natl. Acad. Sci. USA*, 113:3681–3686, 2016.
- [1791] H. Ikeda, N. Esaki, S. Nakai, K. Hashimoto, S. Uesato, K. Soda, and T. Fujita. Acyclic monoterpene primary alcohol:NADP⁺ oxidoreductase of *Rauwolfia serpentina* cells: the key enzyme in biosynthesis of monoterpene alcohols. *J. Biochem.*, 109:341–347, 1991.
- [1792] H. Ikeda, M. Ueda, M. Ikeda, H. Kobayashi, and Y. Honda. Oxysterol 7 α -hydroxylase (CYP39A1) in the ciliary nonpigmented epithelium of bovine eye. *Lab. Invest.*, 83:349–355, 2003.
- [1793] M. Ikeda, M. Levitt, and S. Udenfriend. Phenylalanine as substrate and inhibitor of tyrosine hydroxylase. *Arch. Biochem. Biophys.*, 120:420–427, 1967.
- [1794] Y. Ikeda, C. Dabrowski, and K. Tanaka. Separation and properties of five distinct acyl-CoA dehydrogenases from rat liver mitochondria. Identification of a new 2-methyl branched chain acyl-CoA dehydrogenase. *J. Biol. Chem.*, 258:1066–1076, 1983.
- [1795] Y. Ikeda, K.O. Ikeda, and K. Tanaka. Purification and characterization of short-chain, medium-chain, and long-chain acyl-CoA dehydrogenases from rat liver mitochondria. Isolation of the holo- and apoenzymes and conversion of the apoenzyme to the holoenzyme. *J. Biol. Chem.*, 260:1311–1325, 1985.
- [1796] Y. Ikeda and K. Tanaka. Purification and characterization of isovaleryl coenzyme A dehydrogenase from rat liver mitochondria. *J. Biol. Chem.*, 258:1077–1085, 1983.
- [1797] T. Ikegami and T. Nishino. The presence of desulfo xanthine dehydrogenase in purified and crude enzyme preparations from rat liver. *Arch. Biochem. Biophys.*, 247:254–260, 1986.
- [1798] N. Ikezawa, K. Iwasa, and F. Sato. Molecular cloning and characterization of methylenedioxy bridge-forming enzymes involved in stylopine biosynthesis in *Eschscholzia californica*. *FEBS J.*, 274:1019–1035, 2007.
- [1799] N. Ikezawa, K. Iwasa, and F. Sato. Molecular cloning and characterization of CYP80G2, a cytochrome P450 that catalyzes an intramolecular C-C phenol coupling of (*S*)-reticuline in magnoflorine biosynthesis, from cultured *Coptis japonica* cells. *J. Biol. Chem.*, 283:8810–8821, 2008.
- [1800] N. Ikezawa, M. Tanaka, M. Nagayoshi, R. Shinkyo, T. Sakaki, K. Inouye, and F. Sato. Molecular cloning and characterization of CYP719, a methylenedioxy bridge-forming enzyme that belongs to a novel P450 family, from cultured *Coptis japonica* cells. *J. Biol. Chem.*, 278:38557–38565, 2003.
- [1801] S. Ikuta, S. Imamura, H. Misaki, and Y. Horiuti. Purification and characterization of choline oxidase from *Arthrobacter globiformis*. *J. Biochem. (Tokyo)*, 82:1741–1749, 1977.
- [1802] D. Imai and A.F. Brodie. A phospholipid-requiring enzyme, malate-vitamin K reductase. *J. Biol. Chem.*, 248:7487–7494, 1973.
- [1803] T. Imai. FAD-dependent malate dehydrogenase, a phospholipid-requiring enzyme from *Mycobacterium* sp. strain Takeo. Purification and some properties. *Biochim. Biophys. Acta*, 523:37–46, 1978.

- [1804] Y. Imai, I. Matsunaga, E. Kusunose, and K. Ichihara. Unique heme environment at the putative distal region of hydrogen peroxide-dependent fatty acid α -hydroxylase from *Sphingomonas paucimobilis* (peroxygenase $P_{450}SP\alpha$). *J. Biochem.*, 128:189–194, 2000.
- [1805] S. Imaoka, K. Inoue, and Y. Funae. Aminopyrine metabolism by multiple forms of cytochrome *P*-450 from rat liver microsomes: simultaneous quantitation of four aminopyrine metabolites by high-performance liquid chromatography. *Arch. Biochem. Biophys.*, 265:159–170, 1988.
- [1806] R.D. Imhoff, N.P. Power, M.J. Borrok, and P.A. Tipton. General base catalysis in the urate oxidase reaction: evidence for a novel Thr-Lys catalytic diad. *Biochemistry*, 42:4094–4100, 2003.
- [1807] K. Inaba. Disulfide bond formation system in *Escherichia coli*. *J. Biochem.*, 146:591–597, 2009.
- [1808] E. Inagaki, N. Ohshima, K. Sakamoto, N.D. Babayeva, H. Kato, S. Yokoyama, and T.H. Tahirov. New insights into the binding mode of coenzymes: structure of *Thermus thermophilus* Δ^1 -pyrroline-5-carboxylate dehydrogenase complexed with NADP⁺. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 63:462–465, 2007.
- [1809] D.K. Inaoka, K. Sakamoto, H. Shimizu, T. Shiba, G. Kurisu, T. Nara, T. Aoki, K. Kita, and S. Harada. Structures of *Trypanosoma cruzi* dihydroorotate dehydrogenase complexed with substrates and products: atomic resolution insights into mechanisms of dihydroorotate oxidation and fumarate reduction. *Biochemistry*, 47:10881–10891, 2008.
- [1810] M. Ingelman, S. Ramaswamy, V. Nivière, M. Fontecave, and H. Eklund. Crystal structure of NAD(P)H:flavin oxidoreductase from *Escherichia coli*. *Biochemistry*, 38:7040–7049, 1999.
- [1811] A.A. Iniesta, M. Cervantes, and F.J. Murillo. Cooperation of two carotene desaturases in the production of lycopene in *Myxococcus xanthus*. *FEBS J.*, 274:4306–4314, 2007.
- [1812] K. Inose, M. Fujikawa, T. Yamazaki, K. Kojima, and K. Sode. Cloning and expression of the gene encoding catalytic subunit of thermostable glucose dehydrogenase from *Burkholderia cepacia* in *Escherichia coli*. *Biochim. Biophys. Acta*, 1645:133–138, 2003.
- [1813] H. Inoue, T. Tamura, N. Ehara, A. Nishito, Y. Nakayama, M. Maekawa, K. Imada, H. Tanaka, and K. Inagaki. Biochemical and molecular characterization of the NAD⁺-dependent isocitrate dehydrogenase from the chemolithotroph *Acidithiobacillus thiooxidans*. *FEMS Microbiol. Lett.*, 214:127–132, 2002.
- [1814] H. Inoue, H. Tsuji, and I. Uritani. Characterization and activity change of farnesol dehydrogenase in black rot fungus-infected sweet-potato. *Agric. Biol. Chem.*, 48:733–738, 1984.
- [1815] J. Inoue, J.P. Shaw, M. Rekik, and S. Harayama. Overlapping substrate specificities of benzaldehyde dehydrogenase (the *xyIC* gene product) and 2-hydroxyruconic semialdehyde dehydrogenase (the *xyIG* gene product) encoded by TOL plasmid pWW0 of *Pseudomonas putida*. *J. Bacteriol.*, 177:1196–1201, 1995.
- [1816] S. Inoue, H. Kakoi, and T. Goto. Squid bioluminescence. III. Isolation and structure of *Watasenia* luciferin. *Tetrahedron Lett.*, pages 2971–2974, 1976.
- [1817] S. Inouye, M. Noguchi, Y. Sakaki, Y. Takagi, T. Miyata, S. Iwanaga, T. Miyata, and F.I. Tsuji. Cloning and sequence analysis of cDNA for the luminescent protein aequorin. *Proc. Natl. Acad. Sci. USA*, 82:3154–3158, 1985.
- [1818] S. Inouye, K. Watanabe, H. Nakamura, , and O. Secretional luciferase of the luminous shrimp *Oplophorus gracilirostris*: cDNA cloning of a novel imidazopyrazinone luciferase. *FEBS Lett.*, 481:19–25, 2000.
- [1819] H. Inui, K. Miyatake, Y. Nakano, and S. Kitaoka. Purification and some properties of short chain-length specific *trans*-2-enoyl-CoA reductase in mitochondria of *Euglena gracilis*. *J. Biochem. (Tokyo)*, 100:995–1000, 1986.
- [1820] H. Inui, K. Miyatake, Y., Kitaoka Nakano, , and NADP. +-dependent pyruvate dehydrogenase in mitochondria of *Euglena gracilis*. *J. Biochem. (Tokyo)*, 96:931–934, 1984.
- [1821] H. Inui, K. Ono, K. Miyatake, Y. Nakano, and S. Kitaoka. Purification and characterization of pyruvate:NADP⁺ oxidoreductase in *Euglena gracilis*. *J. Biol. Chem.*, 262:9130–9135, 1987.
- [1822] C. Iobbi-Nivol, C.L. Santini, F. Blasco, and G. Giordano. Purification and further characterization of the second nitrate reductase of *Escherichia coli* K12. *Eur. J. Biochem.*, 188:679–687, 1990.

- [1823] S. Irmeler, G. Schroder, B. St-Pierre, N.P. Crouch, M. Hotze, J. Schmidt, D. Strack, U. Matern, and J. Schroder. Indole alkaloid biosynthesis in *Catharanthus roseus*: new enzyme activities and identification of cytochrome P-450 CYP72A1 as secologanin synthase. *Plant J.*, 24:797–804, 2000.
- [1824] F.A. Isherwood, Y.T. Chen, and L.W. Mapson. Synthesis of L-ascorbic acid in plants and animals. *Biochem. J.*, 56:1–15, 1954.
- [1825] F.A. Isherwood and L.W. Mapson. Biological synthesis of ascorbic acid: the conversion of derivatives of D-galacturonic acid into L-ascorbic acid by plant extracts. *Biochem. J.*, 64:13–22, 1956.
- [1826] F.A. Isherwood, L.W. Mapson, and Y.T. Chen. Synthesis of L-ascorbic acid in rat liver homogenates. Conversion of L-gulono- and L-galactono- γ -lactone and the respective acids into L-ascorbic acid. *Biochem. J.*, 76:157–171, 1960.
- [1827] T. Ishibashi and Y. Imai. Solubilization and partial characterization of alkylglycerol monooxygenase from rat liver microsomes. *Eur. J. Biochem.*, 132:23–27, 1983.
- [1828] H. Ishida, M. Noshiro, K. Okuda, and M.J. Coon. Purification and characterization of 7 α -hydroxy-4-cholesten-3-one 12 α -hydroxylase. *J. Biol. Chem.*, 267:21319–21323, 1992.
- [1829] Y. Ishida, Y. Kuwahara, M. Dadashpour, A. Ina, T. Yamaguchi, M. Morita, Y. Ichiki, and Y. Asano. A sacrificial millipede altruistically protects its swarm using a drone blood enzyme, mandelonitrile oxidase. *Sci. Rep.*, 6:26998–26998, 2016.
- [1830] M. Ishii, T. Omori, Y. Igarashi, O. Adachi, M. Ameyama, and T. Kodama. Methionaquinone is a direct natural electron-acceptor for the membrane-bound hydrogenase in *Hydrogenobacter thermophilus* strain TK-6. *Agric. Biol. Chem.*, 55:3011–3016, 1991.
- [1831] N. Ishikawa, H. Tanaka, F. Koyama, H. Noguchi, C.C. Wang, K. Hotta, and K. Watanabe. Non-heme dioxygenase catalyzes atypical oxidations of 6,7-bicyclic systems to form the 6,6-quinolone core of viridicatin-type fungal alkaloids. *Angew. Chem. Int. Ed. Engl.*, 53:12880–12884, 2014.
- [1832] A. Ishimaru. Purification and characterization of solubilized peroxygenase from microsomes of pea seeds. *J. Biol. Chem.*, 254:8427–8433, 1979.
- [1833] M. Ishimoto, , and J. Biochemical studies on sulfate reducing bacteria. VI. Separation of hydrogenase and thiosulfate reductase and partial purification of cytochrome and green pigment. *J. Biochem. (Tokyo)*, 44:233–242, 1957.
- [1834] M. Ishimoto and J. Koyama. On the role of a cytochrome in the thiosulfate reduction by sulfate-reducing bacterium. *B. Chem. Soc. Jpn.*, 28:231b–232, 1955.
- [1835] D. Ishiyama, D. Vujaklija, and J. Davies. Novel pathway of salicylate degradation by *Streptomyces* sp. strain WA46. *Appl. Environ. Microbiol.*, 70:1297–1306, 2004.
- [1836] W. Ismail, M. El-Said Mohamed, B.L. Wanner, K.A. Datsenko, W. Eisenreich, F. Rohdich, A. Bacher, and G. Fuchs. Functional genomics by NMR spectroscopy. Phenylacetate catabolism in *Escherichia coli*. *Eur. J. Biochem.*, 270:3047–3054, 2003.
- [1837] K. Isobe, T. Ogawa, K. Hirose, T. Yokoi, T. Yoshimura, and H. Hemmi. Geranylgeranyl reductase and ferredoxin from *Methanosarcina acetivorans* are required for the synthesis of fully reduced archaeal membrane lipid in *Escherichia coli* cells. *J. Bacteriol.*, 196:417–423, 2014.
- [1838] Y. Isono, T. Sudo, and M. Hoshino. Properties of a new enzyme, nucleoside oxidase, from *Pseudomonas maltophilia* LB-86. *Agric. Biol. Chem.*, 53:1671–1677, 1989.
- [1839] Y. Isono, T. Sudo, and M. Hoshino. Purification and reaction of a new enzyme, nucleoside oxidase. *Agric. Biol. Chem.*, 53:1663–1669, 1989.
- [1840] M.N. Isupov, A.R. Dalby, A.A. Brindley, Y. Izumi, T. Tanabe, G.N. Murshudov, and J.A. Littlechild. Crystal structure of dodecameric vanadium-dependent bromoperoxidase from the red algae *Corallina officinalis*. *J. Mol. Biol.*, 299:1035–1049, 2000.

- [1841] M.N. Isupov, E. Schroder, R.P. Gibson, J. Beecher, G. Donadio, V. Saneei, S.A. Dcunha, E.J. McGhie, C. Sayer, C.F. Davenport, P.C. Lau, Y. Hasegawa, H. Iwaki, M. Kadow, K. Balke, U.T. Bornscheuer, G. Bourenkov, and J.A. Littlechild. The oxygenating constituent of 3,6-diketocamphane monooxygenase from the CAM plasmid of *Pseudomonas putida*: the first crystal structure of a type II Baeyer-Villiger monooxygenase. *Acta Crystallogr. D Biol. Crystallogr.*, 71:2344–2353, 2015.
- [1842] E. Itagaki. Studies on a steroid monooxygenase from *Cylindrocarpon radicum* ATCC 11011. Purification and characterization. *J. Biochem. (Tokyo)*, 99:815–824, 1986.
- [1843] E. Itagaki. Studies on a steroid monooxygenase from *Cylindrocarpon radicum* ATCC11011. Oxygenative lactonization of androstenedione to testololactone. *J. Biochem. (Tokyo)*, 99:825–832, 1986.
- [1844] H. Ito, T. Ohtsuka, and A. Tanaka. Conversion of chlorophyll *b* to chlorophyll *a* via 7-hydroxymethyl chlorophyll. *J. Biol. Chem.*, 271:1475–1479, 1996.
- [1845] H. Ito and A. Tanaka. Evolution of a new chlorophyll metabolic pathway driven by the dynamic changes in enzyme promiscuous activity. *Plant Cell Physiol.*, 55:593–603, 2014.
- [1846] K. Ito, M. Nakanishi, W.C. Lee, H. Sasaki, S. Zenno, K. Saigo, Y. Kitade, and M. Tanokura. Crystallization and preliminary X-ray analysis of AzoR (azoreductase) from *Escherichia coli*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:399–402, 2005.
- [1847] K. Ito, M. Nakanishi, W.C. Lee, Y. Zhi, H. Sasaki, S. Zenno, K. Saigo, Y. Kitade, and M. Tanokura. Expansion of substrate specificity and catalytic mechanism of azoreductase by X-ray crystallography and site-directed mutagenesis. *J. Biol. Chem.*, 283:13889–13896, 2008.
- [1848] S. Ito, A.C. D’Alessio, O.V. Taranova, K. Hong, L.C. Sowers, and Y. Zhang. Role of Tet proteins in 5mC to 5hmC conversion, ES-cell self-renewal and inner cell mass specification. *Nature*, 466:1129–1133, 2010.
- [1849] S. Ito, L. Shen, Q. Dai, S.C. Wu, L.B. Collins, J.A. Swenberg, C. He, and Y. Zhang. Tet proteins can convert 5-methylcytosine to 5-formylcytosine and 5-carboxylcytosine. *Science*, 333:1300–1303, 2011.
- [1850] N. Itoh, C. Yachi, and T. Kudome. Determining a novel NAD⁺-dependent amine dehydrogenase with a broad substrate range from *Streptomyces virginiae* IFO 12827: purification and characterization. *Journal of Molecular Catalysis B: Enzymatic*, 10:281–290, 2000.
- [1851] Y. Itoh. Cloning and characterization of the *aru* genes encoding enzymes of the catabolic arginine succinyltransferase pathway in *Pseudomonas aeruginosa*. *J. Bacteriol.*, 179:7280–7290, 1997.
- [1852] S. Iuchi, M. Kobayashi, T. Taji, M. Naramoto, M. Seki, T. Kato, S. Tabata, Y. Kakubari, K. Yamaguchi-Shinozaki, and K. Shinozaki. Regulation of drought tolerance by gene manipulation of 9-*cis*-epoxycarotenoid dioxygenase, a key enzyme in abscisic acid biosynthesis in *Arabidopsis*. *Plant J.*, 27:325–333, 2001.
- [1853] S. Iuchi, M. Kobayashi, T. Taji, M. Naramoto, M. Seki, T. Kato, S. Tabata, Y. Kakubari, K. Yamaguchi-Shinozaki, and K. Shinozaki. Regulation of drought tolerance by gene manipulation of 9-*cis*-epoxycarotenoid dioxygenase, a key enzyme in abscisic acid biosynthesis in *Arabidopsis*. *Plant J.*, 30:611–611, 2002.
- [1854] M. Ivan, K. Kondo, H. Yang, W. Kim, J. Valiando, M. Ohh, A. Salic, J.M. Asara, W.S., Kaelin Lane, , and Jr. HIF α targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ sensing. *Science*, 292:464–468, 2001.
- [1855] T.M. Iverson, C. Luna-Chavez, G. Cecchini, and D.C. Rees. Structure of the *Escherichia coli* fumarate reductase respiratory complex. *Science*, 284:1961–1966, 1999.
- [1856] M. Iwabuchi, J. Kohno-Murase, and J. Imamura. Δ^{12} -oleate desaturase-related enzymes associated with formation of conjugated *trans*- Δ^{11} , *cis*- Δ^{13} double bonds. *J. Biol. Chem.*, 278:4603–4610, 2003.
- [1857] T. Iwabuchi and S. Harayama. Biochemical and genetic characterization of 2-carboxybenzaldehyde dehydrogenase, an enzyme involved in phenanthrene degradation by *Nocardioides sp.* strain KP7. *J. Bacteriol.*, 179:6488–6494, 1997.
- [1858] H. Iwaki, S. Grosse, H. Bergeron, H. Leisch, K. Morley, Y. Hasegawa, and P.C. Lau. Camphor pathway redux: functional recombinant expression of 2,5- and 3,6-diketocamphane monooxygenases of *Pseudomonas putida* ATCC 17453 with their cognate flavin reductase catalyzing Baeyer-Villiger reactions. *Appl. Environ. Microbiol.*, 79:3282–3293, 2013.

- [1859] H. Iwaki, Y. Hasegawa, S. Wang, M.M. Kayser, and P.C. Lau. Cloning and characterization of a gene cluster involved in cyclopentanol metabolism in *Comamonas* sp. strain NCIMB 9872 and biotransformations effected by *Escherichia coli*-expressed cyclopentanone 1,2-monooxygenase. *Appl. Environ. Microbiol.*, 68:5671–5684, 2002.
- [1860] M. Iwaki, T. Yagi, K. Horiike, Y. Saeki, T. Ushijima, and M. Nozaki. Crystallization and properties of aromatic amine dehydrogenase from *Pseudomonas* sp. *Arch. Biochem. Biophys.*, 220:253–262, 1983.
- [1861] S. Iwase, F. Lan, P. Bayliss, L. de la Torre-Ubieta, M. Huarte, H.H. Qi, J.R. Whetstone, A. Bonni, T.M. Roberts, and Y. Shi. The X-linked mental retardation gene SMCX/JARID1C defines a family of histone H3 lysine 4 demethylases. *Cell*, 128:1077–1088, 2007.
- [1862] F. Iwata, N. Shinjyo, H. Amino, K. Sakamoto, M.K. Islam, N. Tsuji, and K. Kita. Change of subunit composition of mitochondrial complex II (succinate-ubiquinone reductase/quinol-fumarate reductase) in *Ascaris suum* during the migration in the experimental host. *Parasitol Int*, 57:54–61, 2008.
- [1863] K. Iwata, H. Nojiri, K. Shimizu, T. Yoshida, H. Habe, and T. Omori. Expression, purification, and characterization of 2'-aminobiphenyl-2,3-diol 1,2-dioxygenase from carbazole-degrader *Pseudomonas resinovorans* strain CA10. *Biosci. Biotechnol. Biochem.*, 67:300–307, 2003.
- [1864] R.B. Iyer, J. Wang, and L.G. Bachas. Cloning, expression, and characterization of the *gsdA* gene encoding thermophilic glucose-6-phosphate dehydrogenase from *Aquifex aeolicus*. *Extremophiles*, 6:283–289, 2002.
- [1865] K. Izai, Y. Uchida, T. Orii, S. Yamamoto, and T. Hashimoto. Novel fatty acid β -oxidation enzymes in rat liver mitochondria. I. Purification and properties of very-long-chain acyl-coenzyme A dehydrogenase. *J. Biol. Chem.*, 267:1027–1033, 1992.
- [1866] Y. Izumoto, T. Mori, and K. Yamamoto. Cloning and nucleotide sequence of the gene for NADH:FMN oxidoreductase from *Vibrio harveyi*. *Biochim. Biophys. Acta*, 1185:243–246, 1994.
- [1867] U. Abken H. J. and Deppenmeier. Purification and properties of an $F_{420}H_2$ dehydrogenase from *Methanosarcina mazei* Gö1. *FEMS Microbiol. Lett.*, 154:231–237, 2006.
- [1868] P. Jaakkola, D.R. Mole, Y.M. Tian, M.I. Wilson, J. Gielbert, S.J. Gaskell, Hebestreit Kriegsheim Av, Mukherji H.F., Schofield M., Maxwell C.J., Pugh P.H., Ratcliffe C.W., and P.J. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O_2 -regulated prolyl hydroxylation. *Science*, 292:468–472, 2001.
- [1869] E. Jablonski and M. DeLuca. Purification and properties of the NADH and NADPH specific FMN oxidoreductases from *Beneckea harveyi*. *Biochemistry*, 16:2932–2936, 1977.
- [1870] E. Jablonski and M. DeLuca. Studies of the control of luminescence in *Beneckea harveyi*: properties of the NADH and NADPH:FMN oxidoreductases. *Biochemistry*, 17:672–678, 1978.
- [1871] C.J. Jackson, D.C. Lamb, T.H. Marczylo, A.G. Warrilow, N.J. Manning, D.J. Lowe, D.E. Kelly, and S.L. Kelly. A novel sterol 14 α -demethylase/ferredoxin fusion protein (MCCYP51FX) from *Methylococcus capsulatus* represents a new class of the cytochrome P450 superfamily. *J. Biol. Chem.*, 277:46959–46965, 2002.
- [1872] M.R. Jackson, S.L. Melideo, and M.S. Jorns. Human sulfide:quinone oxidoreductase catalyzes the first step in hydrogen sulfide metabolism and produces a sulfane sulfur metabolite. *Biochemistry*, 51:6804–6815, 2012.
- [1873] R.H. Jackson, A. Cornish-Bowden, and J.A. Cole. Prosthetic groups of the NADH-dependent nitrite reductase from *Escherichia coli* K12. *Biochem. J.*, 193:861–867, 1981.
- [1874] C. Jacoby, J. Eipper, M. Warnke, O. Tiedt, M. Mergelsberg, H.J. Stark, B. Daus, Z. Martin-Moldes, M.T. Zamarro, E. Diaz, and M. Boll. Four molybdenum-dependent steroid C-25 hydroxylases: heterologous overproduction, role in steroid degradation, and application for 25-hydroxyvitamin D₃ synthesis. *mBio*, 9:e00694–18–, 2018.
- [1875] M. Jaeger, B. Rothacker, and T. Ilg. Saturation transfer difference NMR studies on substrates and inhibitors of succinic semialdehyde dehydrogenases. *Biochem. Biophys. Res. Commun.*, 372:400–406, 2008.
- [1876] A.T. Jagendorf. Chloroplast TPNH diaphorase. *Methods Enzymol.*, 6:430–434, 1963.
- [1877] A.K. Jaiswal. Human NAD(P)H:quinone oxidoreductase2. Gene structure, activity, and tissue-specific expression. *J. Biol. Chem.*, 269:14502–14508, 1994.

- [1878] A.K. Jaiswal. Characterization and partial purification of microsomal NAD(P)H:quinone oxidoreductases. *Arch. Biochem. Biophys.*, 375:62–68, 2000.
- [1879] J. Jaje, H.N. Wolcott, O. Fadugba, D. Cripps, A.J. Yang, I.H. Mather, and C. Thorpe. A flavin-dependent sulfhydryl oxidase in bovine milk. *Biochemistry*, 46:13031–13040, 2007.
- [1880] W.B. Jakoby. Aldehyde dehydrogenases. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 203–221. Academic Press, New York, 2nd edition, 1963.
- [1881] W.B. Jakoby and J. Fredericks. Pyrrolidine and putrescine metabolism: γ -aminobutyraldehyde dehydrogenase. *J. Biol. Chem.*, 234:2145–2150, 1959.
- [1882] W.B. Jakoby and J. Fredericks. Erythritol dehydrogenase from *Aerobacter aerogenes*. *Biochim. Biophys. Acta*, 48:26–32, 1961.
- [1883] W.B. Jakoby and E.M. Scott. Aldehyde oxidation. III. Succinic semialdehyde dehydrogenase. *J. Biol. Chem.*, 234:937–940, 1959.
- [1884] K.D. James and P.A. Williams. ntn genes determining the early steps in the divergent catabolism of 4-nitrotoluene and toluene in *Pseudomonas* sp. strain TW3. *J. Bacteriol.*, 180:2043–2049, 1998.
- [1885] P.L. James and C. Anthony. The metal ion in the active site of the membrane glucose dehydrogenase of *Escherichia coli*. *Biochim. Biophys. Acta*, 1647:200–205, 2003.
- [1886] G.A. Jansen, S.J. Mihalik, P.A. Watkins, C. Jakobs, H.W. Moser, and R.J.A. Wanders. Characterization of phytanoyl-CoA hydroxylase in human liver and activity measurements in patients with peroxisomal disorders. *Clin. Chim. Acta*, 271:203–211, 1998.
- [1887] G.A. Jansen, S.J. Mihalik, P.A. Watkins, H.W. Moser, C. Jakobs, S. Denis, and R.J.A. Wanders. Phytanoyl-CoA hydroxylase is present in human liver, located in peroxisomes, and deficient in Zellweger syndrome: direct, unequivocal evidence for the new, revised pathway of phytanic acid α -oxidation in humans. *Biochem. Biophys. Res. Commun.*, 229:205–210, 1996.
- [1888] G.A. Jansen, R. Ofman, S. Ferdinandusse, L. Ijlst, A.O. Muijsers, O.H. Skjeldal, O. Stokke, C. Jakobs, G.T.N. Besley, J.E. Wraith, and R.J.A. Wanders. Refsum disease is caused by mutations in the phytanoyl-CoA hydroxylase gene. *Nat. Genet.*, 17:190–193, 1997.
- [1889] F.W. Janssen and H.W. Ruelius. Alcohol oxidase, a flavoprotein from several *Basidiomycetes* species. Crystallization by fractional precipitation with polyethylene glycol. *Biochim. Biophys. Acta*, 151:330–342, 1968.
- [1890] F.W. Janssen and H.W. Ruelius. Carbohydrate oxidase, a novel enzyme from *Polyporus obtusus*. II. Specificity and characterization of reaction products. *Biochim. Biophys. Acta*, 167:501–510, 1968.
- [1891] J. Jarabak. Isolation and properties of a 15-ketoprostaglandin Δ^{13} -reductase from human placenta. *Methods Enzymol.*, 86:163–167, 1982.
- [1892] J. Jarabak and P. Talalay. Stereospecificity of hydrogen transfer by pyridine nucleotide-linked hydroxysteroid hydrogenase. *J. Biol. Chem.*, 235:2147–2151, 1960.
- [1893] A. Jarvinen, N. Grigorenko, A.R. Khomutov, M.T. Hyvonen, A. Uimari, J. Vepsalainen, R. Sinervirta, T.A. Keinanen, S. Vujcic, L. Alhonen, C.W. Porter, and J. Janne. Metabolic stability of α -methylated polyamine derivatives and their use as substitutes for the natural polyamines. *J. Biol. Chem.*, 280:6595–6601, 2005.
- [1894] J.G. Jaworski and P.K. Stumpf. Fat metabolism in higher plants. Properties of a soluble stearyl-acyl carrier protein desaturase from maturing *Carthamus tinctorius*. *Arch. Biochem. Biophys.*, 162:158–165, 1974.
- [1895] M. Jean and R.D. DeMoss. Indolelactate dehydrogenase from *Clostridium sporogenes*. *Can. J. Microbiol.*, 14:429–435, 1968.
- [1896] A.M. Jeffrey, M. Knight, and W.C. Evans. The bacterial degradation of flavonoids. Hydroxylation of the A-ring of taxifolin by a soil pseudomonad. *Biochem. J.*, 130:373–381, 1972.

- [1897] A.M. Jeffrey, H.J.C. Yeh, D.M. Jerina, T.R. Patel, J.F. Davey, and D.T. Gibson. Initial reactions in the oxidation of naphthalene by *Pseudomonas putida*. *Biochemistry*, 14:575–584, 1975.
- [1898] D. Jendrossek and S. Reinhardt. Sequence analysis of a gene product synthesized by *Xanthomonas* sp. during growth on natural rubber latex. *FEMS Microbiol. Lett.*, 224:61–65, 2003.
- [1899] S. Jennewein, C.D. Rithner, R.M. Williams, and R. Croteau. Taxoid metabolism: Taxoid 14 β -hydroxylase is a cytochrome P450-dependent monooxygenase. *Arch. Biochem. Biophys.*, 413:262–270, 2003.
- [1900] S. Jennewein, C.D. Rithner, R.M. Williams, and R.B. Croteau. Taxol biosynthesis: taxane 13 α -hydroxylase is a cytochrome P₄₅₀-dependent monooxygenase. *Proc. Natl. Acad. Sci. USA*, 98:13595–13560, 2001.
- [1901] F.E. Jenney, Verhagen Jr., Cui M.F.J.M., Adams X., and M.W.W. Anaerobic microbes: Oxygen detoxification without superoxide dismutase. *Science*, 286:306–309, 1999.
- [1902] J.B. Jensen, O.Y. Ampomah, R. Darrah, N.K. Peters, and T.V. Bhuvanewari. Role of trehalose transport and utilization in *Sinorhizobium meliloti*-alfalfa interactions. *Mol. Plant Microbe Interact.*, 18:694–702, 2005.
- [1903] S.J. Jeon and K. Ishikawa. Characterization of novel hexadecameric thioredoxin peroxidase from *Aeropyrum pernix* K1. *J. Biol. Chem.*, 278:24174–24180, 2003.
- [1904] E.Y. Jeong, C. Sopher, I.S. Kim, and H. Lee. Mutational study of the role of tyrosine-49 in the *Saccharomyces cerevisiae* xylose reductase. *Yeast*, 18:1081–1089, 2001.
- [1905] W. Jeong, M.K. Cha, and I.H. Kim. Thioredoxin-dependent hydroperoxide peroxidase activity of bacterioferritin comigratory protein (BCP) as a new member of the thiol-specific antioxidant protein (TSA)/alkyl hydroperoxide peroxidase C (AhpC) family. *J. Biol. Chem.*, 275:2924–2930, 2000.
- [1906] E. Jequier, B.S. Robinson, W. Lovenberg, and A. Sjoerdsma. Further studies on tryptophan hydroxylase in rat brainstem and beef pineal. *Biochem. Pharmacol.*, 18:1071–1081, 1969.
- [1907] F. Jernerren, U. Garscha, I. Hoffmann, M. Hamberg, and E.H. Oliw. Reaction mechanism of 5,8-linoleate diol synthase, 10R-dioxygenase, and 8,11-hydroperoxide isomerase of *Aspergillus clavatus*. *Biochim. Biophys. Acta*, 1801:503–507, 2010.
- [1908] F. Jernerren, I. Hoffmann, and E.H. Oliw. Linoleate 9R-dioxygenase and allene oxide synthase activities of *Aspergillus terreus*. *Arch. Biochem. Biophys.*, 495:67–73, 2010.
- [1909] M.S. Jetten, M. Wagner, J. Fuerst, M. van Loosdrecht, G. Kuenen, and M. Strous. Microbiology and application of the anaerobic ammonium oxidation ('anammox') process. *Curr. Opin. Biotechnol.*, 12:283–288, 2001.
- [1910] G. Ji, E.A. Garber, L.G. Armes, C.M. Chen, J.A. Fuchs, and S. Silver. Arsenate reductase of *Staphylococcus aureus* plasmid pI258. *Biochemistry*, 33:7294–7299, 1994.
- [1911] B. Jia, S. Lee, B.P. Pham, Y.S. Cho, J.K. Yang, H.S. Byeon, J.C. Kim, and G.W. Cheong. An archaeal NADH oxidase causes damage to both proteins and nucleic acids under oxidative stress. *Mol. Cells*, 29:363–371, 2010.
- [1912] B. Jia, S.C. Park, S. Lee, B.P. Pham, R. Yu, T.L. Le, S.W. Han, J.K. Yang, M.S. Choi, W. Baumeister, and G.W. Cheong. Hexameric ring structure of a thermophilic archaeon NADH oxidase that produces predominantly H₂O. *FEBS J.*, 275:5355–5366, 2008.
- [1913] G. Jia, Y. Fu, X. Zhao, Q. Dai, G. Zheng, Y. Yang, C. Yi, T. Lindahl, T. Pan, Y.G. Yang, and C. He. N⁶-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. *Nat. Chem. Biol.*, 7:885–887, 2011.
- [1914] G. Jia, C.G. Yang, S. Yang, X. Jian, C. Yi, Z. Zhou, and C. He. Oxidative demethylation of 3-methylthymine and 3-methyluracil in single-stranded DNA and RNA by mouse and human FTO. *FEBS Lett.*, 582:3313–3319, 2008.
- [1915] J.I. Jimenez, A. Canales, J. Jimenez-Barbero, K. Ginalska, L. Rychlewski, J.L. Garcia, and E. Diaz. Deciphering the genetic determinants for aerobic nicotinic acid degradation: the *nic* cluster from *Pseudomonas putida* KT2440. *Proc. Natl. Acad. Sci. USA*, 105:11329–11334, 2008.
- [1916] R. Jin, D.R. Koop, J.L. Raucy, and J.M. Lasker. Role of human CYP4F2 in hepatic catabolism of the proinflammatory agent leukotriene B₄. *Arch. Biochem. Biophys.*, 359:89–98, 1998.

- [1917] Z. Jin, E. Grotewold, W. Qu, G. Fu, and D. Zhao. Cloning and characterization of a flavanone 3-hydroxylase gene from *Saussurea medusa*. *DNA Seq*, 16:121–129, 2005.
- [1918] J. Jirschitzka, G.W. Schmidt, M. Reichelt, B. Schneider, J. Gershenzon, and J.C. D’Auria. Plant tropane alkaloid biosynthesis evolved independently in the Solanaceae and Erythroxylaceae. *Proc. Natl. Acad. Sci. USA*, 109:10304–10309, 2012.
- [1919] J.M., Wriston Edmundowicz, , and Jr. Mannitol dehydrogenase from *Agaricus campestris*. *J. Biol. Chem.*, 238:3539–3541, 1963.
- [1920] J.E. Jo, S. Mohan Raj, C. Rathnasingh, E. Selvakumar, W.C. Jung, and S. Park. Cloning, expression, and characterization of an aldehyde dehydrogenase from *Escherichia coli* K-12 that utilizes 3-hydroxypropionaldehyde as a substrate. *Appl. Microbiol. Biotechnol.*, 81:51–60, 2008.
- [1921] V. Job, G.L. Marcone, M.S. Pilone, and L. Pollegioni. Glycine oxidase from *Bacillus subtilis*. Characterization of a new flavoprotein. *J. Biol. Chem.*, 277:6985–6993, 2002.
- [1922] J. Johannes, A. Bluschke, N. Jehmlich, M. von Bergen, and M. Boll. Purification and characterization of active-site components of the putative *p*-cresol methylhydroxylase membrane complex from *Geobacter metallireducens*. *J. Bacteriol.*, 190:6493–6500, 2008.
- [1923] K. Johansson, M. El-Ahmad, S. Ramaswamy, L. Hjelmqvist, H. Jornvall, and H. Eklund. Structure of betaine aldehyde dehydrogenase at 2.1 Å resolution. *Protein Sci.*, 7:2106–2117, 1998.
- [1924] D.C. John and N.J. Bulleid. Prolyl 4-hydroxylase: defective assembly of α -subunit mutants indicates that assembled α -subunits are intramolecularly disulfide bonded. *Biochemistry*, 33:14018–14025, 1994.
- [1925] U. Johnsen, M. Dambeck, H. Zaiss, T. Fuhrer, J. Soppa, U. Sauer, and P. Schönheit. D-Xylose degradation pathway in the halophilic archaeon *Haloferax volcanii*. *J. Biol. Chem.*, 284:27290–27303, 2009.
- [1926] U. Johnsen and P. Schönheit. Novel xylose dehydrogenase in the halophilic archaeon *Haloarcula marismortui*. *J. Bacteriol.*, 186:6198–6207, 2004.
- [1927] U. Johnsen, J.M. Sutter, H. Zaiss, and P. Schönheit. L-Arabinose degradation pathway in the haloarchaeon *Haloferax volcanii* involves a novel type of L-arabinose dehydrogenase. *Extremophiles*, 17:897–909, 2013.
- [1928] E.F. Johnson and B. Mukhopadhyay. A new type of sulfite reductase, a novel coenzyme F₄₂₀-dependent enzyme, from the methanarchaeon *Methanocaldococcus jannaschii*. *J. Biol. Chem.*, 280:38776–38786, 2005.
- [1929] E.F. Johnson and B. Mukhopadhyay. Coenzyme F₄₂₀-dependent sulfite reductase-enabled sulfite detoxification and use of sulfite as a sole sulfur source by *Methanococcus maripaludis*. *Appl. Environ. Microbiol.*, 74:3591–3595, 2008.
- [1930] E.F. Johnson, M. Zounes, and U. Müller-Eberhard. Characterization of three forms of rabbit microsomal cytochrome P-450 by peptide mapping utilizing limited proteolysis in sodium dodecyl sulfate and analysis by gel electrophoresis. *Arch. Biochem. Biophys.*, 192:282–289, 1979.
- [1931] H.A. Johnson, D.A. Pelletier, and A.M. Spormann. Isolation and characterisation of anaerobic ethylbenzene dehydrogenase, a novel Mo-Fe-S enzyme. *J. Bacteriol.*, 183:4536–4542, 2001.
- [1932] H.S. Johnson. NADP-malate dehydrogenase: photoactivation in leaves of plants with Calvin cycle photosynthesis. *Biochem. Biophys. Res. Commun.*, 43:703–709, 1971.
- [1933] H.S. Johnson and M.D. Hatch. Properties and regulation of leaf nicotinamide-adenine dinucleotide phosphate-malate dehydrogenase and ‘malic’ enzyme in plants with the C₄-dicarboxylic acid pathway of photosynthesis. *Biochem. J.*, 119:273–280, 1970.
- [1934] J.L. Johnson, K.V. Rajagopalan, S. Mukund, and M.W.W. Adams. Identification of molybdopterin as the organic component of the tungsten cofactor in four enzymes from hyperthermophilic archaea. *J. Biol. Chem.*, 268:4848–4852, 1993.
- [1935] P. Johnson and H.H. Rees. The mechanism of C-20 hydroxylation of α -ecdysone in the desert locust, *Schistocerca gregaria*. *Biochem. J.*, 168:513–520, 1977.

- [1936] P.R. Lashmet Johnson and D.M. Ziegler. Properties of an *N,N*-dimethyl-*p*-aminoazobenzene oxide reductase purified from rat liver cytosol. *J. Biochem. Toxicol.*, 1:15–27, 1986.
- [1937] R.C. Johnson and J.R. Gilbertson. Isolation, characterization, and partial purification of a fatty acyl coenzyme A reductase from bovine cardiac muscle. *J. Biol. Chem.*, 247:6991–6998, 1972.
- [1938] W.M. Johnson and D.W. Westlake. Purification and characterization of glutamic acid dehydrogenase and α -ketoglutaric acid reductase from *Peptococcus aerogenes*. *Can. J. Microbiol.*, 18:881–892, 1972.
- [1939] J.B. Johnston, P.M. Kells, L.M. Podust, and P.R. Ortiz de Montellano. Biochemical and structural characterization of CYP124: a methyl-branched lipid ω -hydroxylase from *Mycobacterium tuberculosis*. *Proc. Natl. Acad. Sci. USA*, 106:20687–20692, 2009.
- [1940] J.B. Johnston, H. Ouellet, and P.R. Ortiz de Montellano. Functional redundancy of steroid C₂₆-monooxygenase activity in *Mycobacterium tuberculosis* revealed by biochemical and genetic analyses. *J. Biol. Chem.*, 285:36352–36360, 2010.
- [1941] D.R. Jollie and J.D. Lipscomb. Formate dehydrogenase from *Methylosinus trichosporium* OB3b. Purification and spectroscopic characterization of the cofactors. *J. Biol. Chem.*, 266:21853–21863, 1991.
- [1942] R. Jonczyk, H. Schmidt, A. Osterrieder, A. Fiesselmann, K. Schullehner, M. Haslbeck, D. Sicker, D. Hofmann, N. Yalpani, C. Simmons, M. Frey, and A. Gierl. Elucidation of the final reactions of DIMBOA-glucoside biosynthesis in maize: characterization of Bx6 and Bx7. *Plant Physiol.*, 146:1053–1063, 2008.
- [1943] E.E. Jones and H.P. Broquist. Saccharopine, an intermediate of the amino adipic acid pathway of lysine biosynthesis. 3. Amino adipic semialdehyde-glutamate reductase. *J. Biol. Chem.*, 241:3430–3434, 1966.
- [1944] J.M. Jones, J.C. Morrell, and S.J. Gould. Identification and characterization of HAOX1, HAOX2, and HAOX3, three human peroxisomal 2-hydroxy acid oxidases. *J. Biol. Chem.*, 275:12590–12597, 2000.
- [1945] K.C. Jones and D.P. Ballou. Reactions of the 4a-hydroperoxide of liver microsomal flavin-containing monooxygenase with nucleophilic and electrophilic substrates. *J. Biol. Chem.*, 261:2553–2559, 1986.
- [1946] K.H. Jones, R.T. Smith, and P.W. Trudgill. Diketocamphane enantiomer-specific ‘Baeyer-Villiger’ monooxygenases from camphor-grown *Pseudomonas putida* ATCC 17453. *J. Gen. Microbiol.*, 139:797–805, 1993.
- [1947] M. Jones, F. Talfournier, A. Bobrov, J.G. Grossmann, N. Vekshin, M.J. Sutcliffe, and N.S. Scrutton. Electron transfer and conformational change in complexes of trimethylamine dehydrogenase and electron transferring flavoprotein. *J. Biol. Chem.*, 277:8457–8465, 2002.
- [1948] R. Jonnalagadda, A. Del Rio Flores, W. Cai, R. Mehmood, M. Narayanamoorthy, C. Ren, J.P.T. Zaragoza, H.J. Kulik, W. Zhang, and C.L. Drennan. Biochemical and crystallographic investigations into isonitrile formation by a nonheme iron-dependent oxidase/decarboxylase. *J. Biol. Chem.*, 296:100231–100231, 2021.
- [1949] K. Jørgensen, A.V. Morant, M. Morant, N.B. Jensen, C.E. Olsen, R. Kannangara, M.S. Motawia, B.L. Møller, and S. Bak. Biosynthesis of the cyanogenic glucosides linamarin and lotaustralin in cassava: isolation, biochemical characterization, and expression pattern of CYP71E7, the oxime-metabolizing cytochrome P450 enzyme. *Plant Physiol.*, 155:282–292, 2011.
- [1950] M. Jormakka, S. Tornroth, J. Abramson, B. Byrne, and S. Iwata. Purification and crystallization of the respiratory complex formate dehydrogenase-N from *Escherichia coli*. *Acta Crystallogr. D Biol. Crystallogr.*, 58:160–162, 2002.
- [1951] M. Jormakka, S. Tornroth, B. Byrne, and S. Iwata. Molecular basis of proton motive force generation: structure of formate dehydrogenase-N. *Science*, 295:1863–1868, 2002.
- [1952] M.S. Jorns, Z.W. Chen, and F.S. Mathews. Structural characterization of mutations at the oxygen activation site in monomeric sarcosine oxidase. *Biochemistry*, 49:3631–3639, 2010.
- [1953] H. Jörnvall. Differences between alcohol dehydrogenases. Structural properties and evolutionary aspects. *Eur. J. Biochem.*, 72:443–452, 1977.
- [1954] E.M. Josse, A.J. Simkin, J. Gaffe, A.M. Laboure, M. Kuntz, and P. Carol. A plastid terminal oxidase associated with carotenoid desaturation during chromoplast differentiation. *Plant Physiol.*, 123:1427–1436, 2000.

- [1955] Y. Jouanneau, C. Meyer, J. Jakoncic, V. Stojanoff, and J. Gaillard. Characterization of a naphthalene dioxygenase endowed with an exceptionally broad substrate specificity toward polycyclic aromatic hydrocarbons. *Biochemistry*, 45:12380–12391, 2006.
- [1956] K.W. Joy and R.H. Hageman. The purification and properties of nitrite reductase from higher plants, and its dependence on ferredoxin. *Biochem. J.*, 100:263–273, 1966.
- [1957] J. Ju, S.G. Ozanick, B. Shen, and M.G. Thomas. Conversion of (2*S*)-arginine to (2*S*,3*R*)-capreomycin by VioC and VioD from the viomycin biosynthetic pathway of *Streptomyces* sp. strain ATCC11861. *ChemBioChem*, 5:1281–1285, 2004.
- [1958] N.I. Jukova, S.M. Klunova, and Y.B. Philippovich. Biochemistry of Insects, issue 17. pages 56–. V.I. Lenin State Pedagogical Institute, Moscow, 1971.
- [1959] G.R. Julian, R.G. Wolfe, and F.J. Reithel. The enzymes of mammary gland. II. The preparation of glucose 6-phosphate dehydrogenase. *J. Biol. Chem.*, 236:754–758, 1961.
- [1960] J.H. Jung and S.B. Lee. Identification and characterization of *Thermoplasma acidophilum* glyceraldehyde dehydrogenase: a new class of NADP⁺-specific aldehyde dehydrogenase. *Biochem. J.*, 397:131–138, 2006.
- [1961] K. Jungerman, R.F. Thauer, G. Leimenstoll, and K. Decker. Function of reduced pyridine nucleotide-ferredoxin oxidoreductases in saccharolytic *Clostridia*. *Biochim. Biophys. Acta*, 305:268–280, 1973.
- [1962] F. Junker, J.A. Field, F. Bangerter, K. Ramsteiner, H.-P. Kohler, C.L. Joannou, J.R. Mason, T. Leisinger, and A.M. Cook. Oxygenation and spontaneous deamination of 2-aminobenzenesulphonic acid in *Alcaligenes* sp. strain O-1 with subsequent *meta* ring cleavage and spontaneous desulphonation to 2-hydroxymuconic acid. *Biochem. J.*, 300:429–436, 1994.
- [1963] F. Junker, R. Kiewitz, and A.M. Cook. Characterization of the *p*-toluenesulfonate operon *tsaMBCD* and *tsaR* in *Comamonas testosteroni* T-2. *J. Bacteriol.*, 179:919–927, 1997.
- [1964] F. Junker, T. Leisinger, and A.M. Cook. 3-Sulphocatechol 2,3-dioxygenase and other dioxygenases (EC 1.13.11.2 and EC 1.14.12.-) in the degradative pathways of 2-aminobenzenesulphonic, benzenesulphonic and 4-toluenesulphonic acids in *Alcaligenes* sp. strain O-1. *Microbiology*, 140:1713–1722, 1994.
- [1965] F. Junker, E. Saller, H.R. Schläfli Oppenberg, P.M. Kroneck, T. Leisinger, and A.M. Cook. Degradative pathways for *p*-toluenecarboxylate and *p*-toluenesulfonate and their multicomponent oxygenases in *Comamonas testosteroni* strains PSB-4 and T-2. *Microbiology*, 142:2419–2427, 1996.
- [1966] U.C. Kabisch, A. Gräntzdörffer, A. Schierhorn, K.P. Rücknagel, J.R. Andreesen, and A. Pich. Identification of D-proline reductase from *Clostridium sticklandii* as a selenoenzyme and indications for a catalytically active pyruvoyl group derived from a cysteine residue by cleavage of a proprotein. *J. Biol. Chem.*, 274:8445–8454, 1999.
- [1967] V. Kadiyala and J.C. Spain. A two-component monooxygenase catalyzes both the hydroxylation of *p*-nitrophenol and the oxidative release of nitrite from 4-nitrocatechol in *Bacillus sphaericus* JS905. *Appl. Environ. Microbiol.*, 64:2479–2484, 1998.
- [1968] H. Kadokura, H. Tian, T. Zander, J.C. Bardwell, and J. Beckwith. Snapshots of DsbA in action: detection of proteins in the process of oxidative folding. *Science*, 303:534–537, 2004.
- [1969] M. Kadow, K. Loschinski, S. Sass, M. Schmidt, and U.T. Bornscheuer. Completing the series of BVMOs involved in camphor metabolism of *Pseudomonas putida* NCIMB 10007 by identification of the two missing genes, their functional expression in *E. coli*, and biochemical characterization. *Appl. Microbiol. Biotechnol.*, 96:419–429, 2012.
- [1970] M. Kadow, S. Sass, M. Schmidt, and U.T. Bornscheuer. Recombinant expression and purification of the 2,5-diketocamphane 1,2-monooxygenase from the camphor metabolizing *Pseudomonas putida* strain NCIMB 10007. *AMB Express*, 1:13–13, 2011.
- [1971] S. Kaewsuwan, E.B. Cahoon, P.F. Perroud, C. Wiwat, N. Panvisavas, R.S. Quatrano, D.J. Cove, and N. Bunyapraphatsara. Identification and functional characterization of the moss *Physcomitrella patens* Δ^5 -desaturase gene involved in arachidonic and eicosapentaenoic acid biosynthesis. *J. Biol. Chem.*, 281:21988–21997, 2006.

- [1972] H.M. Kagan, M.A. Williams, P.R. Williamson, and J.M. Anderson. Influence of sequence and charge on the specificity of lysyl oxidase toward protein and synthetic peptide substrates. *J. Biol. Chem.*, 259:11203–11207, 1984.
- [1973] Z.S. Kagan, V.L. Kretovich, and V.A. Polyakov. Biosynthesis of valine by reductive amination of its keto analogue in plants. *Enzymologia*, 30:343–366, 1966.
- [1974] Z.S. Kagan, V.A. Polyakov, and V.L. Kretovich. Soluble valine dehydrogenase from roots of plant seedlings. *Biochemistry (Mosc.)*, 33:74–84, 1968.
- [1975] Z.S. Kagan, V.A. Polyakov, and V.L. Kretovich. Purification and properties of valine dehydrogenase. *Biochemistry (Mosc.)*, 34:47–51, 1969.
- [1976] A.E. Kahler, F.S. Nielsen, and R.L. Switzer. Biochemical characterization of the heteromeric *Bacillus subtilis* dihydroorotate dehydrogenase and its isolated subunits. *Arch. Biochem. Biophys.*, 371:191–201, 1999.
- [1977] K. Kahn and P.A. Tipton. Spectroscopic characterization of intermediates in the urate oxidase reaction. *Biochemistry*, 37:11651–11659, 1998.
- [1978] R.A. Kahn, T. Fahrendorf, B.A. Halkier, and B.L. Møller. Substrate specificity of the cytochrome P450 enzymes CYP79A1 and CYP71E1 involved in the biosynthesis of the cyanogenic glucoside dhurrin in *Sorghum bicolor* (L.) Moench. *Arch. Biochem. Biophys.*, 363:9–18, 1999.
- [1979] A. Kahnert and M.A. Kertesz. Characterization of a sulfur-regulated oxygenative alkylsulfatase from *Pseudomonas putida* S-313. *J. Biol. Chem.*, 275:31661–31667, 2000.
- [1980] K. Kai, M. Mizutani, N. Kawamura, R. Yamamoto, M. Tamai, H. Yamaguchi, K. Sakata, and B. Shimizu. Scopoletin is biosynthesized via *ortho*-hydroxylation of feruloyl CoA by a 2-oxoglutarate-dependent dioxygenase in *Arabidopsis thaliana*. *Plant J.*, 55:989–999, 2008.
- [1981] M. Kajikawa, K.T. Yamato, Y. Kohzu, S. Shoji, K. Matsui, Y. Tanaka, Y. Sakai, and H. Fukuzawa. A front-end desaturase from *Chlamydomonas reinhardtii* produces pinolenic and coniferonic acids by ω^{13} desaturation in methylotrophic yeast and tobacco. *Plant Cell Physiol.*, 47:64–73, 2006.
- [1982] E. Kalliri, S.B. Mulrooney, and R.P. Hausinger. Identification of *Escherichia coli* YgaF as an L-2-hydroxyglutarate oxidase. *J. Bacteriol.*, 190:3793–3798, 2008.
- [1983] S. Kamei, K. Wakabayashi, and M. Shimazono. ω -Oxidation of fatty acids in vitro. II. ω -Hydroxy fatty acid-NAD oxidoreductase. *J. Biochem. (Tokyo)*, 56:72–76, 1964.
- [1984] N.M. Kamerbeek, M.J. Moonen, J.G. van der Ven, W.J.H. van Berkel, M.W. Fraaije, and D.B. Janssen. 4-Hydroxyacetophenone monooxygenase from *Pseudomonas fluorescens* ACB: a novel flavoprotein catalyzing Baeyer-Villiger oxidation of aromatic compounds. *Eur. J. Biochem.*, 268:2547–2557, 2001.
- [1985] N.M. Kamerbeek, A.J.J. Olsthoorn, M.W. Fraaije, and D.B. Janssen. Substrate specificity of a novel Baeyer-Villiger monooxygenase, 4-hydroxyacetophenone monooxygenase. *Appl. Environ. Microbiol.*, 69:419–426, 2003.
- [1986] L. Kammerer, W. De-Eknamkul, and M.H. Zenk. Enzymic 12-hydroxylation and 12-*O*-methylation of dihydrochelirubine in dihydromacarpine formation by *Thalictrum bulgaricum*. *Phytochemistry*, 36:1409–1416, 1994.
- [1987] S. Kamoda, N. Habu, M. Samejima, and T. Yoshimoto. Purification and some properties of lignostilbene- $\alpha\beta$ -dioxygenase responsible for the C α -C β cleavage of a diarylpropane type lignin model-compound from *Pseudomonas* sp TMY1009. *Agric. Biol. Chem.*, 53:2757–2761, 1989.
- [1988] V. Kanagasundaram and R.K. Scopes. Cloning, sequence analysis and expression of the structural gene encoding glucose-fructose oxidoreductase. *J. Bacteriol.*, 174:1439–1447, 1992.
- [1989] V. Kandasamy, H. Vaidyanathan, I. Djurdjevic, E. Jayamani, K.B. Ramachandran, W. Buckel, G. Jayaraman, and S. Ramalingam. Engineering *Escherichia coli* with acrylate pathway genes for propionic acid synthesis and its impact on mixed-acid fermentation. *Appl. Microbiol. Biotechnol.*, 97:1191–1200, 2013.
- [1990] D.J. Kang, J.M. Ridlon, D.R. Moore, Barnes 2nd, Hylemon S., and P.B. *Clostridium scindens* baiCD and baiH genes encode stereo-specific 7 α /7 β -hydroxy-3-oxo- Δ^4 -cholenoic acid oxidoreductases. *Biochim. Biophys. Acta*, 1781:16–25, 2008.

- [1991] S.W. Kang, H.Z. Chae, M.S. Seo, K. Kim, I.C. Baines, and S.G. Rhee. Mammalian peroxiredoxin isoforms can reduce hydrogen peroxide generated in response to growth factors and tumor necrosis factor- α . *J. Biol. Chem.*, 273:6297–6302, 1998.
- [1992] U. Kappler, B. Bennett, J. Rethmeier, G. Schwarz, R. Deutzmann, A.G. McEwan, and C. Dahl. Sulfite:Cytochrome *c* oxidoreductase from *Thiobacillus novellus*. Purification, characterization, and molecular biology of a heterodimeric member of the sulfite oxidase family. *J. Biol. Chem.*, 275:13202–13212, 2000.
- [1993] S. Kardinahl, C.L. Schmidt, T. Hansen, S. Anemuller, A. Petersen, and G. Schafer. The strict molybdate-dependence of glucose-degradation by the thermoacidophile *Sulfolobus acidocaldarius* reveals the first crenarchaeotic molybdenum containing enzyme—an aldehyde oxidoreductase. *Eur. J. Biochem.*, 260:540–548, 1999.
- [1994] S. Karki, H.G. Yoo, S.Y. Kwon, J.W. Suh, and H.J. Kwon. Cloning and *in vitro* characterization of dTDP-6-deoxy-L-talose biosynthetic genes from *Kitasatospora kifunensis* featuring the dTDP-6-deoxy-L-lyxo-4-hexulose reductase that synthesizes dTDP-6-deoxy-L-talose. *Carbohydr. Res.*, 345:1958–1962, 2010.
- [1995] F. Karp, J.L. Harris, and R. Croteau. Metabolism of monoterpenes: demonstration of the hydroxylation of (+)-sabinene to (+)-*cis*-sabinol by an enzyme preparation from sage (*Salvia officinalis*) leaves. *Arch. Biochem. Biophys.*, 256:179–193, 1987.
- [1996] F. Karp, C.A. Mihaliak, J.L. Harris, and R. Croteau. Monoterpene biosynthesis: specificity of the hydroxylations of (-)-limonene by enzyme preparations from peppermint (*Mentha piperita*), spearmint (*Mentha spicata*), and perilla (*Perilla frutescens*) leaves. *Arch. Biochem. Biophys.*, 276:219–226, 1990.
- [1997] T.P. Karpetsky and E.H. White. The synthesis of *Cypridina* etioluciferamine and the proof of the structure of *Cypridina* luciferin. *Tetrahedron*, 29:3761–3773, 1973.
- [1998] P.A. Karplus, M.J. Daniels, and J.R. Herriott. Atomic structure of ferredoxin-NADP⁺ reductase: prototype for a structurally novel flavoenzyme family. *Science*, 251:60–66, 1991.
- [1999] M. Karrasch, G. Börner, M. Enssle, and R.K. Thauer. The molybdoenzyme formylmethanofuran dehydrogenase from *Methanosarcina barkeri* contains a pterin cofactor. *Eur. J. Biochem.*, 194:367–372, 1990.
- [2000] W.E. Karsten, S.A. Nimmo, J. Liu, and L. Chooback. Identification of 2,3-dihydrodipicolinate as the product of the dihydrodipicolinate synthase reaction from *Escherichia coli*. *Arch. Biochem. Biophys.*, 653:50–62, 2018.
- [2001] B. Kartal, N.M. de Almeida, W.J. Maalcke, H.J. Op den Camp, M.S. Jetten, and J.T. Keltjens. How to make a living from anaerobic ammonium oxidation. *FEMS Microbiol. Rev.*, 37:428–461, 2013.
- [2002] B. Kartal, W.J. Maalcke, N.M. de Almeida, I. Cirpus, J. Gloerich, W. Geerts, H.J. Op den Camp, H.R. Harhangi, E.M. Janssen-Megens, K.J. Francoijs, H.G. Stunnenberg, J.T. Keltjens, M.S. Jetten, and M. Strous. Molecular mechanism of anaerobic ammonium oxidation. *Nature*, 479:127–130, 2011.
- [2003] K. Kasahara, T. Miyamoto, T. Fujimoto, H. Oguri, T. Tokiwano, H. Oikawa, Y. Ebizuka, and I. Fujii. Solanapyrone synthase, a possible Diels-Alderase and iterative type I polyketide synthase encoded in a biosynthetic gene cluster from *Alternaria solani*. *ChemBioChem*, 11:1245–1252, 2010.
- [2004] D. Kasai, N. Araki, K. Motoi, S. Yoshikawa, T. Iino, S. Imai, E. Masai, and M. Fukuda. γ -Resorcyrate catabolic-pathway genes in the soil actinomycete *Rhodococcus jostii* RHA1. *Appl. Environ. Microbiol.*, 81:7656–7665, 2015.
- [2005] D. Kasai, T. Fujinami, T. Abe, K. Mase, Y. Katayama, M. Fukuda, and E. Masai. Uncovering the protocatechuate 2,3-cleavage pathway genes. *J. Bacteriol.*, 191:6758–6768, 2009.
- [2006] T. Kasai, I. Suzuki, and T. Asai. [Glyoxylic oxidase system in *Acetobacter*.]. *Koso Kagaku Shimpoijumu*, 17:77–81, 1962.
- [2007] A.K. Kaster, J. Moll, K. Parey, and R.K. Thauer. Coupling of ferredoxin and heterodisulfide reduction via electron bifurcation in hydrogenotrophic methanogenic archaea. *Proc. Natl. Acad. Sci. USA*, 108:2981–2986, 2011.
- [2008] M. Katagiri and E. Itagaki. A steroid ketone monooxygenase from *Cylindrocarpum radicum*. In F. Müller, editor, *Chemistry and Biochemistry of Flavoenzymes*, pages 102–108. CRC Press, Florida, 1991.

- [2009] R. Katan and G. Avigad. NADP dependent oxidation of TDP-glucose by an enzyme system from sugar beets. *Biochem. Biophys. Res. Commun.*, 24:18–24, 1966.
- [2010] N. Katano, H. Yamamoto, R. Iio, and K. Inoue. 7-Deoxyloganin 7-hydroxylase in *Lonicera japonica* cell cultures. *Phytochemistry*, 58:53–58, 2001.
- [2011] M. Kataoka, S. Shimizu, and H. Yamada. Purification and characterization of a novel FMN-dependent enzyme. Membrane-bound L-(+)-pantoyl lactone dehydrogenase from *Nocardia asteroides*. *Eur. J. Biochem.*, 204:799–806, 1992.
- [2012] K. Katayama, T. Kobayashi, M. Chijimatsu, A. Ichihara, and H. Oikawa. Purification and N-terminal amino acid sequence of solanapyrone synthase, a natural Diels-Alderase from *Alternaria solani*. *Biosci. Biotechnol. Biochem.*, 72:604–607, 2008.
- [2013] K. Katayama, T. Kobayashi, H. Oikawa, M. Honma, and A. Ichihara. Enzymatic activity and partial purification of solanapyrone synthase: first enzyme catalyzing Diels-Alder reaction. *Biochim. Biophys. Acta*, 1384:387–395, 1998.
- [2014] B. Kather, K. Stingl, M.E. van der Rest, K. Altendorf, and D. Molenaar. Another unusual type of citric acid cycle enzyme in *Helicobacter pylori*: the malate:quinone oxidoreductase. *J. Bacteriol.*, 182:3204–3209, 2000.
- [2015] S. Kathiresan, A. Chandrashekar, G.A. Ravishankar, and R. Sarada. Regulation of astaxanthin and its intermediates through cloning and genetic transformation of β -carotene ketolase in *Haematococcus pluvialis*. *J. Biotechnol.*, 196-197:33–41, 2015.
- [2016] J.Y. Kato, N. Funai, H. Watanabe, Y. Ohnishi, and S. Horinouchi. Biosynthesis of γ -butyrolactone autoregulators that switch on secondary metabolism and morphological development in *Streptomyces*. *Proc. Natl. Acad. Sci. USA*, 104:2378–2383, 2007.
- [2017] M. Kato, Y. Arais, A. Noma, A. Nagao, T. Suzuki, R. Ishitani, and O. Nureki. Crystal structure of a novel JmjC-domain-containing protein, TYW5, involved in tRNA modification. *Nucleic Acids Res.*, 39:1576–1585, 2011.
- [2018] N. Kato, K. Shirakawa, H. Kobayashi, and C. Sakazawa. The dismutation of aldehydes by a bacterial enzyme. *Agric. Biol. Chem.*, 47:39–46, 1983.
- [2019] N. Kato, H. Suzuki, H. Takagi, Y. Asami, H. Takeya, M. Uramoto, T. Usui, S. Takahashi, Y. Sugimoto, and H. Osada. Identification of cytochrome P450s required for fumitremorgin biosynthesis in *Aspergillus fumigatus*. *ChemBioChem*, 10:920–928, 2009.
- [2020] N. Kato, H. Suzuki, H. Takagi, M. Uramoto, S. Takahashi, and H. Osada. Gene disruption and biochemical characterization of verruculogen synthase of *Aspergillus fumigatus*. *ChemBioChem*, 12:711–714, 2011.
- [2021] N. Kato, T. Yamagami, M. Shima, and C. Sakazawa. Formaldehyde dismutase, a novel NAD-binding oxidoreductase from *Pseudomonas putida* F61. *Eur. J. Biochem.*, 156:59–64, 1986.
- [2022] T. Kato, Y. Daigo, S. Hayama, N. Ishikawa, T. Yamabuki, T. Ito, M. Miyamoto, S. Kondo, and Y. Nakamura. A novel human tRNA-dihydrouridine synthase involved in pulmonary carcinogenesis. *Cancer Res.*, 65:5638–5646, 2005.
- [2023] Y. Kato, H. Yamada, and Y. Asano. Stereoselective synthesis of opine-type secondary amine carboxylic acids by a new enzyme opine dehydrogenase. Use of recombinant enzymes. *J. Mol. Catal., B Enzym.*, 1:151–160, 1996.
- [2024] A. Katoh, K. Uenohara, M. Akita, and T. Hashimoto. Early steps in the biosynthesis of NAD in *Arabidopsis* start with aspartate and occur in the plastid. *Plant Physiol.*, 141:851–857, 2006.
- [2025] S. Katoh. Sepiapterin reductase from horse liver: purification and properties of the enzyme. *Arch. Biochem. Biophys.*, 146:202–214, 1971.
- [2026] A.G. Katopodis, D. Ping, and S.W. May. A novel enzyme from bovine neurointermediate pituitary catalyzes dealkylation of α -hydroxyglycine derivatives, thereby functioning sequentially with peptidylglycine α -amidating monooxygenase in peptide amidation. *Biochemistry*, 29:6115–6120, 1990.
- [2027] T. Katsumata, A. Hasegawa, T. Fujiwara, T. Komatsu, M. Notomi, H. Abe, M. Natsume, and H. Kawaide. *Arabidopsis* CYP85A2 catalyzes lactonization reactions in the biosynthesis of 2-deoxy-7-oxalactone brassinosteroids. *Biosci. Biotechnol. Biochem.*, 72:2110–2117, 2008.

- [2028] Y. Katsuyama, K. Harmrolfs, D. Pistorius, Y. Li, and R. Muller. A semipinacol rearrangement directed by an enzymatic system featuring dual-function FAD-dependent monooxygenase. *Angew. Chem. Int. Ed. Engl.*, 51:9437–9440, 2012.
- [2029] F. Katzen and J. Beckwith. Transmembrane electron transfer by the membrane protein DsbD occurs via a disulfide bond cascade. *Cell*, 103:769–779, 2000.
- [2030] F. Katzen and J. Beckwith. Role and location of the unusual redox-active cysteines in the hydrophobic domain of the transmembrane electron transporter DsbD. *Proc. Natl. Acad. Sci. USA*, 100:10471–10476, 2003.
- [2031] H.M. Katzen, F. Tietze, and D. Stetten. Further studies on the properties of hepatic glutathione-insulin transhydrogenase. *J. Biol. Chem.*, 238:1006–1011, 1963.
- [2032] B. Kauffmann, A. Aubry, and F. Favier. The three-dimensional structures of peptide methionine sulfoxide reductases: current knowledge and open questions. *Biochim. Biophys. Acta*, 1703:249–260, 2005.
- [2033] B.T. Kaufman and R.C. Gardiner. Studies on dihydrofolic reductase. I. Purification and properties of dihydrofolic reductase from chicken liver. *J. Biol. Chem.*, 241:1319–1328, 1966.
- [2034] E.E. Kaufman, T. Nelson, H.M. Fales, and D.M. Levin. Isolation and characterization of a hydroxyacid-oxoacid transhydrogenase from rat kidney mitochondria. *J. Biol. Chem.*, 263:16872–16879, 1988.
- [2035] S. Kaufman. Studies on the mechanism of the enzymic conversion of phenylalanine to tyrosine. *J. Biol. Chem.*, 234:2677–2682, 1959.
- [2036] S. Kaufman. Phenylalanine hydroxylase. *Methods Enzymol.*, 5:809–816, 1962.
- [2037] S. Kaufman, S. Korkes, and A. del Campillo. Biosynthesis of dicarboxylic acids by carbon dioxide fixation. V. Further studies of the "malic" enzyme of *Lactobacillus arabinosus*. *J. Biol. Chem.*, 192:301–312, 1951.
- [2038] B. Kauppi, K. Lee, E. Carredano, R.E. Parales, D.T. Gibson, H. Eklund, and S. Ramaswamy. Structure of an aromatic-ring-hydroxylating dioxygenase - naphthalene 1,2-dioxygenase. *Structure*, 6:571–586, 1998.
- [2039] M.P. Kautsky and D.D. Hagerman. 17 β -Estradiol dehydrogenase of ovine ovaries. *J. Biol. Chem.*, 245:1978–1984, 1970.
- [2040] T. Kawachi and H. Rudney. Solubilization and purification of β -hydroxy- β -methylglutaryl coenzyme A reductase from rat liver. *Biochemistry*, 9:1700–1700, 1970.
- [2041] A. Kawaguchi, S. Tsubotani, Y. Seyama, T. Yamakawa, T. Osumi, T. Hashimoto, T. Kikuchi, M. Ando, and S. Okuda. Stereochemistry of dehydrogenation catalyzed by acyl-CoA oxidase. *J. Biochem. (Tokyo)*, 88:1481–1486, 1980.
- [2042] F. Kawai and X. Hu. Biochemistry of microbial polyvinyl alcohol degradation. *Appl. Microbiol. Biotechnol.*, 84:227–237, 2009.
- [2043] F. Kawai, T. Kimura, Y. Tani, H. Yamada, T. Ueno, and H. Fukami. Identification of reaction-products of polyethylene-glycol dehydrogenase. *Agric. Biol. Chem.*, 47:1669–1671, 1983.
- [2044] F. Kawai, H. Yamanaka, M. Ameyama, E. Shinagawa, K. Matsushita, and O. Adachi. Identification of the prosthetic group and further characterization of a novel enzyme, polyethylene-glycol dehydrogenase. *Agric. Biol. Chem.*, 49:1071–1076, 1985.
- [2045] S. Kawai, M. Goda-Tsutsumi, T. Yakushi, K. Kano, and K. Matsushita. Heterologous overexpression and characterization of a flavoprotein-cytochrome *c* complex fructose dehydrogenase of *Gluconobacter japonicus* NBRC3260. *Appl. Environ. Microbiol.*, 79:1654–1660, 2013.
- [2046] T. Kawano, S. Koyama, H. Takematsu, Y. Kozutsumi, H. Kawasaki, S. Kawashima, T. Kawasaki, and A. Suzuki. Molecular cloning of cytidine monophospho-*N*-acetylneuraminic acid hydroxylase. Regulation of species- and tissue-specific expression of *N*-glycolylneuraminic acid. *J. Biol. Chem.*, 270:16458–16463, 1995.
- [2047] S. Kawasaki, J. Ishikura, D. Chiba, T. Nishino, and Y. Niimura. Purification and characterization of an H₂O-forming NADH oxidase from *Clostridium aminovalericum*: existence of an oxygen-detoxifying enzyme in an obligate anaerobic bacteria. *Arch. Microbiol.*, 181:324–330, 2004.
- [2048] S. Kawata, J.M. Trzaskos, and J.L. Gaylor. Affinity chromatography of microsomal enzymes on immobilized detergent-solubilized cytochrome *b*₅. *J. Biol. Chem.*, 261:3790–3799, 1986.

- [2049] C.W. Kay, B. Mennenga, H. Gorisch, and R. Bittl. Characterisation of the PQQ cofactor radical in quinoprotein ethanol dehydrogenase of *Pseudomonas aeruginosa* by electron paramagnetic resonance spectroscopy. *FEBS Lett.*, 564:69–72, 2004.
- [2050] E.V. Kearns, S. Hugly, and C.R. Somerville. The role of cytochrome *b₅* in Δ^{12} desaturation of oleic acid by microsomes of safflower (*Carthamus tinctorius* L.). *Arch. Biochem. Biophys.*, 284:431–436, 1991.
- [2051] A. Keck, D. Conradt, A. Mahler, A. Stolz, R. Mattes, and J. Klein. Identification and functional analysis of the genes for naphthalenesulfonate catabolism by *Sphingomonas xenophaga* BN6. *Microbiology*, 152:1929–1940, 2006.
- [2052] N.Y. Kedishvili, O.V. Chumakova, S.V. Chetyrkin, O.V. Belyaeva, E.A. Lapshina, D.W. Lin, M. Matsumura, and P.S. Nelson. Evidence that the human gene for prostate short-chain dehydrogenase/reductase (PSDR1) encodes a novel retinal reductase (RalR1). *J. Biol. Chem.*, 277:28909–28915, 2002.
- [2053] B.B. Keele, J.M. McCord, and I. Fridovich. Further characterization of bovine superoxide dismutase and its isolation from bovine heart. *J. Biol. Chem.*, 246:2875–2880, 1971.
- [2054] D. Keilin and E.F. Hartree. Coupled oxidation of alcohol. *Proc. R. Soc. Lond. B Biol. Sci.*, 119:141–159, 1936.
- [2055] D. Keilin and E.F. Hartree. Properties of glucose oxidase (notatin). *Biochem. J.*, 42:221–229, 1948.
- [2056] D. Keilin and E.F. Hartree. Specificity of glucose oxidase (notatin). *Biochem. J.*, 50:331–341, 1952.
- [2057] D. Keilin and T. Mann. Laccase, a blue copper-protein oxidase from the latex of *Rhus succedanea*. *Nature (Lond.)*, 143:23–24, 1939.
- [2058] T. Keitel, A. Diehl, T. Knaute, J.J. Stezowski, W. Hohne, and H. Gorisch. X-ray structure of the quinoprotein ethanol dehydrogenase from *Pseudomonas aeruginosa*: basis of substrate specificity. *J. Mol. Biol.*, 297:961–974, 2000.
- [2059] Y. Keller, F. Bouvier, A. d’Harlingue, and B. Camara. Metabolic compartmentation of plastid prenyl lipid biosynthesis—evidence for the involvement of a multifunctional geranylgeranyl reductase. *Eur. J. Biochem.*, 251:413–417, 1998.
- [2060] J.T. Kellis, Vickery Jr., and L.E. Purification and characterization of human placental aromatase cytochrome *P-450*. *J. Biol. Chem.*, 262:4413–4420, 1987.
- [2061] S.L. Kelly, D.C. Lamb, A.J. Corran, B.C. Baldwin, L.W. Parks, and D.E. Kelly. Purification and reconstitution of activity of *Saccharomyces cerevisiae* P450 61, a sterol Δ^{22} -desaturase. *FEBS Lett.*, 377:217–220, 1995.
- [2062] J.D. Kemp, E. Hack, D.W. Sutton, and M. El-Wakil. Unusual amino acids and their relationship to tumorigenesis. *Proc. Int. Conf. Plant Pathol. Bact.*, 4th:183–188, 1979.
- [2063] J.D. Kemp, D.W. Sutton, and E. Hack. Purification and characterization of the crown gall specific enzyme nopaline synthase. *Biochemistry*, 18:3755–3760, 1979.
- [2064] S.W. Kengen, J. van der Oost, and W.M. de Vos. Molecular characterization of H₂O₂-forming NADH oxidases from *Archaeoglobus fulgidus*. *Eur. J. Biochem.*, 270:2885–2894, 2003.
- [2065] R.H. Kenten and P.J.G. Mann. Simple method for the preparation of horseradish peroxidase. *Biochem. J.*, 57:347–348, 1954.
- [2066] L. Kerscher and D. Oesterhelt. Purification and properties of two 2-oxoacid:ferredoxin oxidoreductases from *Halobacterium halobium*. *Eur. J. Biochem.*, 116:587–594, 1981.
- [2067] S.J. Kerscher, J.G. Okun, and U. Brandt. A single external enzyme confers alternative NADH:ubiquinone oxidoreductase activity in *Yarrowia lipolytica*. *J. Cell Sci.*, 112 (Pt 14):2347–2354, 1999.
- [2068] P.J. Kersten and D. Cullen. Cloning and characterization of cDNA encoding glyoxal oxidase, a H₂O₂-producing enzyme from the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Proc. Natl. Acad. Sci. USA*, 90:7411–7413, 1993.
- [2069] P.J. Kersten and T.K. Kirk. Involvement of a new enzyme, glyoxal oxidase, in extracellular H₂O₂ production by *Phanerochaete chrysosporium*. *J. Bacteriol.*, 169:2195–2201, 1987.

- [2070] P.J. Kersten, M. Tien, B. Kalyanaraman, and T.K. Kirk. The ligninase of *Phanerochaete chrysosporium* generates cation radicals from methoxybenzenes. *J. Biol. Chem.*, 260:2609–2612, 1985.
- [2071] P.J. Kersten, C. Witek, A. vanden Wymelenberg, and D. Cullen. *Phanerochaete chrysosporium* glyoxal oxidase is encoded by two allelic variants: structure, genomic organization, and heterologous expression of *glx1* and *glx2*. *J. Bacteriol.*, 177:6106–6110, 1995.
- [2072] M.A. Kertesz, K. Schmidt-Larbig, and T. Wuest. A novel reduced flavin mononucleotide-dependent methanesulfonate sulfonate encoded by the sulfur-regulated *msu* operon of *Pseudomonas aeruginosa*. *J. Bacteriol.*, 181:1464–1473, 1999.
- [2073] D.L. Kessel, J.L. Johnston, H.J. Cohen, and K.V. Rajagopalan. Visualization of hepatic sulfite oxidase in crude tissue preparation by electron paramagnetic resonance spectroscopy. *Biochim. Biophys. Acta*, 334:86–96, 1974.
- [2074] A.C. Kessler, P.K. Brown, L.K. Romana, and P.R. Reeves. Molecular cloning and genetic characterization of the *rfb* region from *Yersinia pseudotuberculosis* serogroup IIA, which determines the formation of the 3,6-dideoxyhexose abequose. *J. Gen. Microbiol.*, 137:2689–2695, 1991.
- [2075] H. Khairy, J.H. Wubbeler, and A. Steinbuechel. Biodegradation of the organic disulfide 4,4'-dithiodibutyric acid by *Rhodococcus* spp. *Appl. Environ. Microbiol.*, 81:8294–8306, 2015.
- [2076] H. Khairy, J.H. Wubbeler, and A. Steinbuechel. The NADH:flavin oxidoreductase Nox from *Rhodococcus erythropolis* MI2 is the key enzyme of 4,4'-dithiodibutyric acid degradation. *Lett. Appl. Microbiol.*, 63:434–441, 2016.
- [2077] B. Khaliullin, R. Ayikpoe, M. Tuttle, and J.A. Latham. Mechanistic elucidation of the mycofactocin-biosynthetic radical S-adenosylmethionine protein, MftC. *J. Biol. Chem.*, 292:13022–13033, 2017.
- [2078] S.S. Khandekar and L.D. Eirich. Purification and characterization of an anabolic fumarate reductase from *Methanobacterium thermoautotrophicum*. *Appl. Environ. Microbiol.*, 55:856–861, 1989.
- [2079] S.V. Khangulov, V.N. Gladyshev, G.C. Dismukes, and T.C. Stadtman. Selenium-containing formate dehydrogenase H from *Escherichia coli*: a molybdopterin enzyme that catalyzes formate oxidation without oxygen transfer. *Biochemistry*, 37:3518–3528, 1998.
- [2080] M. Khanna, K.N. Qin, R.W. Wang, and K.C. Cheng. Substrate specificity, gene structure, and tissue-specific distribution of multiple human 3 α -hydroxysteroid dehydrogenases. *J. Biol. Chem.*, 270:20162–20168, 1995.
- [2081] Y. Khatri, M. Ringle, M. Lisurek, J.P. von Kries, J. Zapp, and R. Bernhardt. Substrate hunting for the myxobacterial CYP260A1 revealed new 1 α -hydroxylated products from C-19 steroids. *ChemBioChem*, 17:90–101, 2016.
- [2082] A. Khindaria, G. Nie, and S.D. Aust. Detection and characterization of the lignin peroxidase compound II-veratryl alcohol cation radical complex. *Biochemistry*, 36:14181–14185, 1997.
- [2083] A. Khindaria, I. Yamazaki, and S.D. Aust. Veratryl alcohol oxidation by lignin peroxidase. *Biochemistry*, 34:16860–16869, 1995.
- [2084] A. Khindaria, I. Yamazaki, and S.D. Aust. Stabilization of the veratryl alcohol cation radical by lignin peroxidase. *Biochemistry*, 35:6418–6424, 1996.
- [2085] S. Khurana, D.B. Powers, S. Anderson, and M. Blaber. Crystal structure of 2,5-diketo-D-gluconic acid reductase A complexed with NADPH at 2.1-Å resolution. *Proc. Natl. Acad. Sci. USA*, 95:6768–6773, 1998.
- [2086] T. Kido, K. Hashizume, and K. Soda. Purification and properties of nitroalkane oxidase from *Fusarium oxysporum*. *J. Bacteriol.*, 133:53–58, 1978.
- [2087] C. Kiefer, S. Hessel, J.M. Lampert, K. Vogt, M.O. Lederer, D.E. Breithaupt, and J. von Lintig. Identification and characterization of a mammalian enzyme catalyzing the asymmetric oxidative cleavage of provitamin A. *J. Biol. Chem.*, 276:14110–14116, 2001.
- [2088] M.B. Kilgore, M.M. Augustin, G.D. May, J.A. Crow, and T.M. Kutchan. CYP96T1 of *Narcissus* sp. aff. *pseudonarcissus* catalyzes formation of the *para-para'* C-C phenol couple in the Amaryllidaceae alkaloids. *Front. Plant Sci.*, 7:225–225, 2016.

- [2089] W.W. Kilgore and M.P. Starr. Catabolism of galacturonic and glucuronic acids by *Erwinia carotovora*. *J. Biol. Chem.*, 234:2227–2235, 1959.
- [2090] W.W. Kilgore and M.P. Starr. Uronate oxidation by phytopathogenic pseudomonads. *Nature (Lond.)*, 183:1412–1413, 1959.
- [2091] C. Kim, W.W. Lorenz, J.T. Hoopes, and J.F. Dean. Oxidation of phenolate siderophores by the multicopper oxidase encoded by the *Escherichia coli* *yacK* gene. *J. Bacteriol.*, 183:4866–4875, 2001.
- [2092] G.T. Kim, S. Fujioka, T. Kozuka, F.E. Tax, S. Takatsuto, S. Yoshida, and H. Tsukaya. CYP90C1 and CYP90D1 are involved in different steps in the brassinosteroid biosynthesis pathway in *Arabidopsis thaliana*. *Plant J.*, 41:710–721, 2005.
- [2093] J. Kim and S.D. Copley. The orphan protein bis- γ -glutamylcystine reductase joins the pyridine nucleotide disulfide reductase family. *Biochemistry*, 52:2905–2913, 2013.
- [2094] J. Kim, D. Darley, T. Selmer, and W. Buckel. Characterization of (*R*)-2-hydroxyisocaproate dehydrogenase and a family III coenzyme A transferase involved in reduction of L-leucine to isocaproate by *Clostridium difficile*. *Appl. Environ. Microbiol.*, 72:6062–6069, 2006.
- [2095] J. Kim, C. Gherasim, and R. Banerjee. Decyanation of vitamin B₁₂ by a trafficking chaperone. *Proc. Natl. Acad. Sci. USA*, 105:14551–14554, 2008.
- [2096] J. Kim, H. Suh, S. Kim, K. Kim, C. Ahn, and J. Yim. Identification and characteristics of the structural gene for the *Drosophila* eye colour mutant *sepia*, encoding PDA synthase, a member of the ω class glutathione *S*-transferases. *Biochem. J.*, 398:451–460, 2006.
- [2097] J.J. Kim, M. Wang, and R. Paschke. Crystal structures of medium-chain acyl-CoA dehydrogenase from pig liver mitochondria with and without substrate. *Proc. Natl. Acad. Sci. USA*, 90:7523–7527, 1993.
- [2098] K.H. Kim, B.H. Ha, S.J. Kim, S.K. Hong, K.Y. Hwang, and E.E. Kim. Crystal structures of Enoyl-ACP reductases I (FabI) and III (FabL) from *B. subtilis*. *J. Mol. Biol.*, 406:403–415, 2011.
- [2099] K.H. Kim, V. Janiak, and M. Petersen. Purification, cloning and functional expression of hydroxyphenylpyruvate reductase involved in rosmarinic acid biosynthesis in cell cultures of *Coleus blumei*. *Plant Mol. Biol.*, 54:311–323, 2004.
- [2100] K.H. Kim, J.K. Park, B.H. Ha, J.H. Moon, and E.E. Kim. Crystallization and preliminary X-ray crystallographic analysis of enoyl-ACP reductase III (FabL) from *Bacillus subtilis*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 63:246–248, 2007.
- [2101] K.S. Kim, J.G. Pelton, W.B. Inwood, U. Andersen, S. Kustu, and D.E. Wemmer. The Rut pathway for pyrimidine degradation: novel chemistry and toxicity problems. *J. Bacteriol.*, 192:4089–4102, 2010.
- [2102] M.I. Kim, I. Shin, S. Cho, J. Lee, and S. Rhee. Structural and functional insights into (*S*)-ureidoglycolate dehydrogenase, a metabolic branch point enzyme in nitrogen utilization. *PLoS One*, 7:e52066–e52066, 2012.
- [2103] S. Kim, L. Benoiton, and W.K. Paik. α -Alkyllysine. Purification and properties of the enzyme. *J. Biol. Chem.*, 239:3790–3796, 1964.
- [2104] S. Kim and T.P. West. Pyrimidine catabolism in *Pseudomonas aeruginosa*. *FEMS Microbiol. Lett.*, 61:175–179, 1991.
- [2105] S.J. Kim and M. Shoda. Purification and characterization of a novel peroxidase from *Geotrichum candidum* dec 1 involved in decolorization of dyes. *Appl. Environ. Microbiol.*, 65:1029–1035, 1999.
- [2106] S.M. Kim, K.H. Paek, and S.B. Lee. Characterization of NADP⁺-specific L-rhamnose dehydrogenase from the thermoacidophilic Archaeon *Thermoplasma acidophilum*. *Extremophiles*, 16:447–454, 2012.
- [2107] S.Y. Kim, P. Zhao, M. Igarashi, R. Sawa, T. Tomita, M. Nishiyama, and T. Kuzuyama. Cloning and heterologous expression of the cyclooctatin biosynthetic gene cluster afford a diterpene cyclase and two P450 hydroxylases. *Chem. Biol.*, 16:736–743, 2009.
- [2108] T. Kim and S. Buratowski. Two *Saccharomyces cerevisiae* JmjC domain proteins demethylate histone H3 Lys³⁶ in transcribed regions to promote elongation. *J. Biol. Chem.*, 282:20827–20835, 2007.

- [2109] T.W. Kim, J.Y. Hwang, Y.S. Kim, S.H. Joo, S.C. Chang, J.S. Lee, S. Takatsuto, and S.K. Kim. *Arabidopsis* CYP85A2, a cytochrome P450, mediates the Baeyer-Villiger oxidation of castasterone to brassinolide in brassinosteroid biosynthesis. *Plant Cell*, 17:2397–2412, 2005.
- [2110] W. Kim, T.A. Major, and W.B. Whitman. Role of the precorrin 6-X reductase gene in cobamide biosynthesis in *Methanococcus maripaludis*. *Archaea*, 1:375–384, 2005.
- [2111] Y.A. Kim, H.J. Chung, Y.J. Kim, Y.K. Choi, Y.K. Hwang, S.W. Lee, and Y.S. Park. Characterization of recombinant *Dictyostelium discoideum* sepiapterin reductase expressed in *E. coli*. *Mol. Cells*, 10:405–410, 2000.
- [2112] Y.B. Kim, M.R. Uddina, Y. Kim, C.G. Park, and S.U. Park. Molecular cloning and characterization of tyrosine aminotransferase and hydroxyphenylpyruvate reductase, and rosmarinic acid accumulation in *Scutellaria baicalensis*. *Nat. Prod. Commun.*, 9:1311–1314, 2014.
- [2113] Y.M. Kim, E.C. Kim, and Y. Kim. The human lysyl oxidase-like 2 protein functions as an amine oxidase toward collagen and elastin. *Mol. Biol. Rep.*, 38:145–149, 2011.
- [2114] Y.O. Kim, H.J. Koh, S.H. Kim, S.H. Jo, J.W. Huh, K.S. Jeong, I.J. Lee, B.J. Song, and T.L. Huh. Identification and functional characterization of a novel, tissue-specific NAD⁺-dependent isocitrate dehydrogenase β subunit isoform. *J. Biol. Chem.*, 274:36866–36875, 1999.
- [2115] Y.S. Kim, N.H. Kim, S.J. Yeom, S.W. Kim, and D.K. Oh. *In vitro* characterization of a recombinant Blh protein from an uncultured marine bacterium as a β -carotene 15,15'-dioxygenase. *J. Biol. Chem.*, 284:15781–15793, 2009.
- [2116] Y.S. Kim and D.K. Oh. Substrate specificity of a recombinant chicken β -carotene 15,15'-monooxygenase that converts β -carotene into retinal. *Biotechnol. Lett.*, 31:403–408, 2009.
- [2117] Y.S. Kim, C.S. Park, and D.K. Oh. Retinal production from β -carotene by β -carotene 15,15'-dioxygenase from an unculturable marine bacterium. *Biotechnol. Lett.*, 32:957–961, 2010.
- [2118] Y.S. Kim, E.S. Seo, and D.K. Oh. Characterization of an apo-carotenoid 13,14-dioxygenase from *Novosphingobium aromaticivorans* that converts β -apo-8'-carotenal to β -apo-13-carotenone. *Biotechnol. Lett.*, 34:1851–1856, 2012.
- [2119] N. Kimmich, A. Das, I. Sevrioukova, Y. Meharena, S.G. Sligar, and T.L. Poulos. Electron transfer between cytochrome P450_{cin} and its FMN-containing redox partner, cindoxin. *J. Biol. Chem.*, 282:27006–27011, 2007.
- [2120] H.L. King, Dyar Jr., Wilken R.E., and D.R. Ketopantoyl lactone and ketopantoic acid reductases. Characterization of the reactions and purification of two forms of ketopantoyl lactone reductase. *J. Biol. Chem.*, 247:4689–4695, 1972.
- [2121] T.E. King and V.H. Cheldelin. Oxidation of acetaldehyde by *Aerobacter suboxydans*. *J. Biol. Chem.*, 220:177–191, 1956.
- [2122] M. Kinne, M. Poraj-Kobielska, E. Aranda, R. Ullrich, K.E. Hammel, K. Scheibner, and M. Hofrichter. Regioselective preparation of 5-hydroxypropranolol and 4'-hydroxydiclofenac with a fungal peroxygenase. *Bioorg. Med. Chem. Lett.*, 19:3085–3087, 2009.
- [2123] M. Kinne, M. Poraj-Kobielska, S.A. Ralph, R. Ullrich, M. Hofrichter, and K.E. Hammel. Oxidative cleavage of diverse ethers by an extracellular fungal peroxygenase. *J. Biol. Chem.*, 284:29343–29349, 2009.
- [2124] J.J. Kinzel and J.K. Bhattacharjee. Lysine biosynthesis in *Rhodotorula glutinis*: properties of pipercolic acid oxidase. *J. Bacteriol.*, 151:1073–1077, 1982.
- [2125] U. Kirchner, A.H. Westphal, R. Muller, and W.J. van Berkel. Phenol hydroxylase from *Bacillus thermoglucosidasius* A7, a two-protein component monooxygenase with a dual role for FAD. *J. Biol. Chem.*, 278:47545–47553, 2003.
- [2126] K. Kiritani, S. Narise, and R.P. Wagner. The reductoisomerase of *Neurospora crassa*. *J. Biol. Chem.*, 241:2047–2051, 1966.
- [2127] M. Y. Kiriuchin, L. V. Kletsova, A. Y. Chistoserdov, and Y. D. Tsygankov. Properties of glucose 6-phosphate and 6-phosphogluconate dehydrogenases of the obligate methylotroph *Methylobacillus flagellatum* KT. *FEMS Microbiol. Lett.*, 52:199–204, 1988.
- [2128] Y. Kishi, T. Goto, Y. Hirata, O. Shiromura, and F.H. Johnson. Cypridina bioluminescence. I. Structure of *Cypridina* luciferin. *Tetrahedron Lett.*, pages 3427–3436, 1966.

- [2129] S. Kishigami and K. Ito. Roles of cysteine residues of DsbB in its activity to reoxidize DsbA, the protein disulphide bond catalyst of *Escherichia coli*. *Genes Cells*, 1:201–208, 1996.
- [2130] S. Kishigami, E. Kanaya, M. Kikuchi, and K. Ito. DsbA-DsbB interaction through their active site cysteines. Evidence from an odd cysteine mutant of DsbA. *J. Biol. Chem.*, 270:17072–17074, 1995.
- [2131] H. Kita, M. Kamimoto, S. Senoh, T. Adachi, and Y. Takeda. Crystallization and some properties of 3,4-dihydroxyphenylacetate-2,3-oxygenase. *Biochem. Biophys. Res. Commun.*, 18:66–70, 1965.
- [2132] K. Kita, C.R. Vibat, S. Meinhardt, J.R. Guest, and R.B. Gennis. One-step purification from *Escherichia coli* of complex II (succinate: ubiquinone oxidoreductase) associated with succinate-reducible cytochrome *b*₅₅₆. *J. Biol. Chem.*, 264:2672–2677, 1989.
- [2133] W. Kitagawa, N. Kimura, and Y. Kamagata. A novel *p*-nitrophenol degradation gene cluster from a gram-positive bacterium, *Rhodococcus opacus* SAO101. *J. Bacteriol.*, 186:4894–4902, 2004.
- [2134] S. Kitamura and K. Tatsumi. Purification of *N*-hydroxy-2-acetylaminofluorene reductase from rabbit liver cytosol. *Biochem. Biophys. Res. Commun.*, 133:67–74, 1985.
- [2135] N. Kitaoka, H. Kawaide, N. Amano, T. Matsubara, K. Nabeta, K. Takahashi, and H. Matsuura. CYP94B3 activity against jasmonic acid amino acid conjugates and the elucidation of 12-*O*- β -glucopyranosyl-jasmonoyl-L-isoleucine as an additional metabolite. *Phytochemistry*, 99:6–13, 2014.
- [2136] N. Kitaoka, T. Matsubara, M. Sato, K. Takahashi, S. Wakuta, H. Kawaide, H. Matsui, K. Nabeta, and H. Matsuura. *Arabidopsis* CYP94B3 encodes jasmonoyl-L-isoleucine 12-hydroxylase, a key enzyme in the oxidative catabolism of jasmonate. *Plant Cell Physiol.*, 52:1757–1765, 2011.
- [2137] N. Kitaoka, Y. Wu, M. Xu, and R.J. Peters. Optimization of recombinant expression enables discovery of novel cytochrome P450 activity in rice diterpenoid biosynthesis. *Appl. Microbiol. Biotechnol.*, 99:7549–7558, 2015.
- [2138] J.P. Kitcher, P.W. Trudgill, and J.S. Rees. Purification and properties of 2-furoyl-coenzyme A hydroxylase from *Pseudomonas putida* F2. *Biochem. J.*, 130:121–132, 1972.
- [2139] A. Kitmitto, N. Myronova, P. Basu, and H. Dalton. Characterization and structural analysis of an active particulate methane monooxygenase trimer from *Methylococcus capsulatus* (Bath). *Biochemistry*, 44:10954–10965, 2005.
- [2140] M. Kito and L.I. Pizer. Purification and regulatory properties of the biosynthetic L-glycerol 3-phosphate dehydrogenase from *Escherichia coli*. *J. Biol. Chem.*, 244:3316–3323, 1969.
- [2141] K. Kiuchi, M. Noshikimi, and K. Yagi. Purification and characterization of L-gulonolactone oxidase from chicken kidney microsomes. *Biochemistry*, 21:5076–5082, 1982.
- [2142] K.I. Kivirikko, Y. Kishida, S. Sakakibara, and J. Prockop. Hydroxylation of (X-Pro-Gly)_n by procollagen proline hydroxylase. Effect of chain length, helical conformation and amino acid sequence in the substrate. *Biochim. Biophys. Acta*, 271:347–356, 1972.
- [2143] K.I. Kivirikko and J. Myllyharju. Prolyl 4-hydroxylases and their protein disulfide isomerase subunit. *Matrix Biol*, 16:357–368, 1998.
- [2144] K.I. Kivirikko and D.J. Prockop. Purification and partial characterization of the enzyme for the hydroxylation of proline in procollagen. *Arch. Biochem. Biophys.*, 118:611–618, 1967.
- [2145] H. Kiyohara, K. Nagao, and K. Yano. Isolation and some properties of NAD-linked 2-carboxybenzaldehyde dehydrogenase in *Alcaligenes faecalis* AFK 2 grown on phenanthrene. *J. Gen. Appl. Microbiol.*, 27:443–455, 1981.
- [2146] R. Kjonaas, C. Martinkus-Taylor, and R. Croteau. Metabolism of monoterpenes: conversion of *l*-menthone to *l*-menthol and *d*-neomenthol by stereospecific dehydrogenases from peppermint (*Mentha piperita*) leaves. *Plant Physiol.*, 69:1013–1017, 1982.
- [2147] R.B. Kjonaas, K.V. Venkatachalam, and R. Croteau. Metabolism of monoterpenes: oxidation of isopiperitenol to isopiperitenone, and subsequent isomerization to piperitenone by soluble enzyme preparations from peppermint (*Mentha piperita*) leaves. *Arch. Biochem. Biophys.*, 238:49–60, 1985.

- [2148] E. Klamann and F. Lingens. Degradation of (-)-ephedrine by *Pseudomonas putida*. Detection of (-)-ephedrine: NAD⁺-oxidoreductase from *Arthrobacter globiformis*. *Z. Naturforsch. C: Biosci.*, 35:80–87, 1980.
- [2149] S.J. Klebanoff. Myeloperoxidase: friend and foe. *J. Leukoc. Biol.*, 77:598–625, 2005.
- [2150] L.A. Kleczkowski and G.E. Edwards. Identification of hydroxypyruvate and glyoxylate reductases in maize leaves. *Plant Physiol.*, 91:278–286, 1989.
- [2151] L.A. Kleczkowski and D.D. Randall. Purification and characterization of a novel NADPH(NADH)-dependent hydroxypyruvate reductase from spinach leaves. Comparison of immunological properties of leaf hydroxypyruvate reductases. *Biochem. J.*, 250:145–152, 1988.
- [2152] L.A. Kleczkowski, D.D. Randall, and D.G. Blevins. Purification and characterization of a novel NADPH(NADH)-dependent glyoxylate reductase from spinach leaves. Comparison of immunological properties of leaf glyoxylate reductase and hydroxypyruvate reductase. *Biochem. J.*, 239:653–659, 1986.
- [2153] A. Klein, V.M. Fernandez, and R.K. Thauer. H₂-Forming N⁵,N¹⁰-methylenetetrahydromethanopterin dehydrogenase: mechanism of H₂-formation analyzed using hydrogen isotopes. *FEBS Lett.*, 368:203–206, 1995.
- [2154] D. Klein, B. Fink, B. Arold, W. Eisenreich, and W. Schwab. Functional characterization of enone oxidoreductases from strawberry and tomato fruit. *J. Agric. Food Chem.*, 55:6705–6711, 2007.
- [2155] A. Kletzin. Coupled enzymatic production of sulfite, thiosulfate, and hydrogen sulfide from sulfur: purification and properties of a sulfur oxygenase reductase from the facultatively anaerobic archaeobacterium *Desulfurolobus ambivalens*. *J. Bacteriol.*, 171:1638–1643, 1989.
- [2156] A. Kletzin. Molecular characterization of the sor gene, which encodes the sulfur oxygenase/reductase of the thermoacidophilic Archaeum *Desulfurolobus ambivalens*. *J. Bacteriol.*, 174:5854–5859, 1992.
- [2157] D.P. Kloer, S. Ruch, S. Al-Babili, P. Beyer, and G.E. Schulz. The structure of a retinal-forming carotenoid oxygenase. *Science*, 308:267–269, 2005.
- [2158] R.J. Klose, K. Yamane, Y. Bae, D. Zhang, H. Erdjument-Bromage, P. Tempst, J. Wong, and Y. Zhang. The transcriptional repressor JHDM3A demethylates trimethyl histone H3 lysine 9 and lysine 36. *Nature*, 442:312–316, 2006.
- [2159] R.J. Klose, Q. Yan, Z. Tothova, K. Yamane, H. Erdjument-Bromage, P. Tempst, D.G. Gilliland, Y., Kaelin Zhang, , and Jr. The retinoblastoma binding protein RBP2 is an H₃K4 demethylase. *Cell*, 128:889–900, 2007.
- [2160] M. Kluge, R. Ullrich, C. Dolge, K. Scheibner, and M. Hofrichter. Hydroxylation of naphthalene by aromatic peroxygenase from *Agroclybe aegerita* proceeds via oxygen transfer from H₂O₂ and intermediary epoxidation. *Appl. Microbiol. Biotechnol.*, 81:1071–1076, 2009.
- [2161] J. Knablein, H. Dobbek, S. Ehlert, and F. Schneider. Isolation, cloning, sequence analysis and X-ray structure of dimethyl sulfoxide trimethylamine *N*-oxide reductase from *Rhodobacter capsulatus*. *Biol. Chem.*, 378:293–302, 1997.
- [2162] D.B. Knaff and M. Hirasawa. Ferredoxin-dependent chloroplast enzymes. *Biochim. Biophys. Acta*, 1056:93–125, 1991.
- [2163] D.B. Knaff, R. Malkin, J.C. Myron, and M. Stoller. The role of plastoquinone and β-carotene in the primary reaction of plant photosystem II. *Biochim. Biophys. Acta*, 459:402–411, 1977.
- [2164] J.E. Knapp, D.T. Mitchell, M.A. Yazdi, S.R. Ernst, L.J. Reed, and M.L. Hackert. Crystal structure of the truncated cubic core component of the *Escherichia coli* 2-oxoglutarate dehydrogenase multienzyme complex. *J. Mol. Biol.*, 280:655–668, 1998.
- [2165] B. Kneidinger, M. Graninger, G. Adam, M. Puchberger, P. Kosma, S. Zayni, and P. Messner. Identification of two GDP-6-deoxy-D-lyxo-4-hexulose reductases synthesizing GDP-D-rhamnose in *Aneurinibacillus thermoaerophilus* L420-91^T. *J. Biol. Chem.*, 276:5577–5583, 2001.
- [2166] B. Kneidinger, K. O’Riordan, J. Li, J.R. Brisson, J.C. Lee, and J.S. Lam. Three highly conserved proteins catalyze the conversion of UDP-*N*-acetyl-D-glucosamine to precursors for the biosynthesis of O antigen in *Pseudomonas aeruginosa* O11 and capsule in *Staphylococcus aureus* type 5. Implications for the UDP-*N*-acetyl-L-fucosamine biosynthetic pathway. *J. Biol. Chem.*, 278:3615–3627, 2003.

- [2167] O. Kniemeyer and J. Heider. Ethylbenzene dehydrogenase, a novel hydrocarbon-oxidising molybdenum/iron-sulfur/heme enzyme. *J. Biol. Chem.*, 276:21381–21386, 2001.
- [2168] O. Kniemeyer and J. Heider. (S)-1-phenylethanol dehydrogenase of *Azoarcus* sp. strain EbN1, an enzyme of anaerobic ethylbenzene catabolism. *Arch. Microbiol.*, 176:129–135, 2001.
- [2169] H.R. Knobel, T. Egli, and J.R. van der Meer. Cloning and characterization of the genes encoding nitrilotriacetate monooxygenase of *Chelatobacter heintzii* ATCC 29600. *J. Bacteriol.*, 178:6123–6132, 1996.
- [2170] M. Knop, T.Q. Dang, G. Jeschke, and F.P. Seebeck. Copper is a cofactor of the formylglycine-generating enzyme. *ChemBioChem*, 18:161–165, 2017.
- [2171] M. Knop, P. Engi, R. Lemnaru, and F.P. Seebeck. *In vitro* reconstitution of formylglycine-generating enzymes requires copper(I). *ChemBioChem*, 16:2147–2150, 2015.
- [2172] S. Knorr, M. Sinn, D. Galetskiy, R.M. Williams, C. Wang, N. Muller, O. Mayans, D. Schleheck, and J.S. Hartig. Widespread bacterial lysine degradation proceeding via glutarate and L-2-hydroxyglutarate. *Nat. Commun.*, 9:5071–5071, 2018.
- [2173] W.E. Knox. The quinine-oxidizing enzyme and liver aldehyde oxidase. *J. Biol. Chem.*, 163:699–711, 1946.
- [2174] W.E. Knox and S.W. Edwards. Homogentisate oxidase of liver. *J. Biol. Chem.*, 216:479–487, 1955.
- [2175] K. Kobayashi and A. Yoshimoto. Studies on yeast sulfite reductase. IV. Structure and steady-state kinetics. *Biochim. Biophys. Acta*, 705:348–356, 1982.
- [2176] S. Kobayashi and O. Hayaishi. Anthranilic acid conversion to catechol (*Pseudomonas*). *Methods Enzymol.*, 17A:505–510, 1970.
- [2177] T. Kobayashi, H. Nakanishi, M. Takahashi, S. Kawasaki, N.K. Nishizawa, and S. Mori. In vivo evidence that *Ids3* from *Hordeum vulgare* encodes a dioxygenase that converts 2'-deoxymugineic acid to mugineic acid in transgenic rice. *Planta*, 212:864–871, 2001.
- [2178] M.J. Kobylarz, J.C. Grigg, S.J. Takayama, D.K. Rai, D.E. Heinrichs, and M.E. Murphy. Synthesis of L-2,3-diaminopropionic acid, a siderophore and antibiotic precursor. *Chem. Biol.*, 21:379–388, 2014.
- [2179] G.L.E. Koch, D.C. Shaw, and F. Gibson. Tyrosine biosynthesis in *Aerobacter aerogenes*. Purification and properties of chorismate mutase-prephenate dehydrogenase. *Biochim. Biophys. Acta*, 212:375–386, 1970.
- [2180] C.D. Kochakian, B.R. Carroll, and B. Uhri. Comparisons of the oxidation of C₁₉-hydroxysteroids by guinea pig liver homogenates. *J. Biol. Chem.*, 224:811–818, 1957.
- [2181] G. Kochs and H. Grisebach. Enzymic synthesis of isoflavones. *Eur. J. Biochem.*, 155:311–318, 1986.
- [2182] T. Kodera, S.V. Smirnov, N.N. Samsonova, Y.I. Kozlov, R. Koyama, M. Hibi, J. Ogawa, K. Yokozeki, and S. Shimizu. A novel L-isoleucine hydroxylating enzyme, L-isoleucine dioxygenase from *Bacillus thuringiensis*, produces (2S,3R,4S)-4-hydroxyisoleucine. *Biochem. Biophys. Res. Commun.*, 390:506–510, 2009.
- [2183] L.K. Koditschek and W.W. Umbreit. α -Glycerophosphate oxidase in *Streptococcus faecium* F 24. *J. Bacteriol.*, 93:1063–1068, 1969.
- [2184] T. Koeduka, T.J. Baiga, J.P. Noel, and E. Pichersky. Biosynthesis of *t*-anethole in anise: characterization of *t*-anol/iso Eugenol synthase and an *O*-methyltransferase specific for a C₇-C₈ propenyl side chain. *Plant Physiol.*, 149:384–394, 2009.
- [2185] T. Koeduka, E. Fridman, D.R. Gang, D.G. Vass ao, B.L. Jackson, C.M. Kish, I. Orlova, S.M. Spassova, N.G. Lewis, J.P. Noel, T.J. Baiga, N. Dudareva, and E. Pichersky. Eugenol and isoeugenol, characteristic aromatic constituents of spices, are biosynthesized via reduction of a coniferyl alcohol ester. *Proc. Natl. Acad. Sci. USA*, 103:10128–10133, 2006.
- [2186] T. Koeduka, G.V. Louie, I. Orlova, C.M. Kish, M. Ibdah, C.G. Wilkerson, M.E. Bowman, T.J. Baiga, J.P. Noel, N. Dudareva, and E. Pichersky. The multiple phenylpropene synthases in both *Clarkia breweri* and *Petunia hybrida* represent two distinct protein lineages. *Plant J.*, 54:362–374, 2008.

- [2187] T. Koeduka, K. Matsui, Y. Akakabe, and T. Kajiwara. Catalytic properties of rice α -oxygenase. A comparison with mammalian prostaglandin H synthases. *J. Biol. Chem.*, 277:22648–22655, 2002.
- [2188] T.C. Koekemoer, D. Litthauer, and W. Oelofsen. Isolation and characterization of adipose tissue glycerol-3-phosphate dehydrogenase. *Int. J. Biochem. Cell Biol.*, 27:625–632, 1995.
- [2189] K.J. Koelen and G.G. Gross. Partial purification and properties of tropine dehydrogenase from root cultures of *Datura stramonium*. *Planta Med.*, 44:227–230, 1982.
- [2190] A.L. Koen and C.R. Shaw. Retinol and alcohol dehydrogenases in retina and liver. *Biochim. Biophys. Acta*, 128:48–54, 1966.
- [2191] R.A. Koeth, B.S. Levison, M.K. Culley, J.A. Buffa, Z. Wang, J.C. Gregory, E. Org, Y. Wu, L. Li, J.D. Smith, W.H. Tang, J.A. DiDonato, A.J. Lusis, and S.L. Hazen. γ -Butyrobetaine is a proatherogenic intermediate in gut microbial metabolism of L-carnitine to TMAO. *Cell Metab.*, 20:799–812, 2014.
- [2192] J.W. Koetter and G.E. Schulz. Crystal structure of 6-hydroxy-D-nicotine oxidase from *Arthrobacter nicotinovorans*. *J. Mol. Biol.*, 352:418–428, 2005.
- [2193] H. Koga, E. Yamaguchi, K. Matsunaga, H. Aramaki, and T. Horiuchi. Cloning and nucleotide sequences of NADH-putidaredoxin reductase gene (*camA*) and putidaredoxin gene (*camB*) involved in cytochrome *P*-450_{cam} hydroxylase of *Pseudomonas putida*. *J. Biochem.*, 106:831–836, 1989.
- [2194] S. Koga, J. Ogawa, L.Y. Cheng, Y.M. Choi, H. Yamada, and S. Shimizu. Nucleoside oxidase, a hydrogen peroxide-forming oxidase, from *Flavobacterium meningosepticum*. *Appl. Environ. Microbiol.*, 63:4282–4286, 1997.
- [2195] Y. Koga, T. Kyuragi, M. Nishihara, and N. Sone. Did archaeal and bacterial cells arise independently from noncellular precursors? A hypothesis stating that the advent of membrane phospholipid with enantiomeric glycerophosphate backbones caused the separation of the two lines of descent. *J. Mol. Evol.*, 46:54–63, 1998.
- [2196] H.J. Koh, S.M. Lee, B.G. Son, S.H. Lee, Z.Y. Ryoo, K.T. Chang, J.W. Park, D.C. Park, B.J. Song, R.L. Veech, H. Song, and T.L. Huh. Cytosolic NADP⁺-dependent isocitrate dehydrogenase plays a key role in lipid metabolism. *J. Biol. Chem.*, 279:39968–39974, 2004.
- [2197] H.-P.E. Kohler, D. Kohler-Staub, and D.D. Focht. Degradation of 2-hydroxybiphenyl and 2,2'-dihydroxybiphenyl by *Pseudomonas* sp. strain HBP1. *Appl. Environ. Microbiol.*, 54:2683–2688, 1988.
- [2198] S.D. Kohlwein, S. Eder, C.S. Oh, C.E. Martin, K. Gable, D. Bacikova, and T. Dunn. Tsc13p is required for fatty acid elongation and localizes to a novel structure at the nuclear-vacuolar interface in *Saccharomyces cerevisiae*. *Mol. Cell Biol.*, 21:109–125, 2001.
- [2199] L.D. Kohn and W.B. Jakoby. L- and mesotartaric acid dehydrogenase (crystalline). *Methods Enzymol.*, 9:236–240, 1966.
- [2200] L.D. Kohn and W.B. Jakoby. Tartaric acid metabolism. VI. Crystalline oxalloglycolate reductive decarboxylase. *J. Biol. Chem.*, 243:2486–2493, 1968.
- [2201] L.D. Kohn and W.B. Jakoby. Tartaric acid metabolism. VII. Crystalline hydroxypyruvate reductase (D-glycerate dehydrogenase). *J. Biol. Chem.*, 243:2494–2499, 1968.
- [2202] L.D. Kohn and H.R. Kaback. Mechanisms of active transport in isolated bacterial membrane vesicles. XV. Purification and properties of the membrane-bound D-lactate dehydrogenase from *Escherichia coli*. *J. Biol. Chem.*, 248:7012–7017, 1973.
- [2203] L.D. Kohn, P.M. Packman, R.H. Allen, and W.B. Jakoby. Tartaric acid metabolism. V. Crystalline tartrate dehydrogenase. *J. Biol. Chem.*, 243:2479–2485, 1968.
- [2204] K.-D. Kohnert, H.-J. Hahn, H. Zühlke, S. Schmidt, and H. Fiedler. Breakdown of exogenous insulin by Langerhans islets of the pancreas in vitro. *Biochim. Biophys. Acta*, 338:68–77, 1974.
- [2205] H. Kohno, T. Furukawa, T. Yoshinaga, R. Tokunaga, and S. Taketani. Coproporphyrinogen oxidase. Purification, molecular cloning, and induction of mRNA during erythroid differentiation. *J. Biol. Chem.*, 268:21359–21363, 1993.

- [2206] P. Koivunen, M. Hirsila, V. Gunzler, K.I. Kivirikko, and J. Myllyharju. Catalytic properties of the asparaginyl hydroxylase (FIH) in the oxygen sensing pathway are distinct from those of its prolyl 4-hydroxylases. *J. Biol. Chem.*, 279:9899–9904, 2004.
- [2207] K. Kojima, W. Tsugawa, and K. Sode. Cloning and expression of glucose 3-dehydrogenase from *Halomonas* sp. α -15 in *Escherichia coli*. *Biochem. Biophys. Res. Commun.*, 282:21–27, 2001.
- [2208] Y. Kojima, N. Itada, and O. Hayaishi. Metapyrocatechase: a new catechol-cleaving enzyme. *J. Biol. Chem.*, 236:2223–2228, 1961.
- [2209] J.G. Koland, M.J. Miller, and R.B. Gennis. Reconstitution of the membrane-bound, ubiquinone-dependent pyruvate oxidase respiratory chain of *Escherichia coli* with the cytochrome *d* terminal oxidase. *Biochemistry*, 23:445–453, 1984.
- [2210] N.W. Kolar, A.C. Swart, J.I. Mason, and P. Swart. Functional expression and characterisation of human cytochrome P45017 α in *Pichia pastoris*. *J. Biotechnol.*, 129:635–644, 2007.
- [2211] P.E. Kolattukuday. Reduction of fatty acids to alcohols by cell-free preparations of *Euglena gracilis*. *Biochemistry*, 9:1095–1102, 1970.
- [2212] A.K. Koli, C. Yearby, W. Scott, and K.O. Donaldson. Purification and properties of three separate menadione reductases from hog liver. *J. Biol. Chem.*, 244:621–629, 1969.
- [2213] V.L. Kolossov and C.A. Rebeiz. Chloroplast biogenesis 84: solubilization and partial purification of membrane-bound [4-vinyl]chlorophyllide *a* reductase from etiolated barley leaves. *Anal. Biochem.*, 295:214–219, 2001.
- [2214] S. Kominami, H. Ochi, Y. Kobayashi, and S. Takemori. Studies on the steroid hydroxylation system in adrenal cortex microsomes. Purification and characterization of cytochrome *P*-450 specific for steroid C-21 hydroxylation. *J. Biol. Chem.*, 255:3386–3394, 1980.
- [2215] A.J. Komor, B.S. Rivard, R. Fan, Y. Guo, L. Que, Lipscomb Jr., and J.D. CmlI *N*-oxygenase catalyzes the final three steps in chloramphenicol biosynthesis without dissociation of intermediates. *Biochemistry*, 56:4940–4950, 2017.
- [2216] R. Komuniecki, S. Fekete, and J. Thissen-Parra. Purification and characterization of the 2-methyl branched-chain acyl-CoA dehydrogenase, an enzyme involved in NADH-dependent enoyl-CoA reduction in anaerobic mitochondria of the nematode, *Ascaris suum*. *J. Biol. Chem.*, 260:4770–4777, 1985.
- [2217] R. Komuniecki, J. McCrury, J. Thissen, and N. Rubin. Electron-transfer flavoprotein from anaerobic *Ascaris suum* mitochondria and its role in NADH-dependent 2-methyl branched-chain enoyl-CoA reduction. *Biochim. Biophys. Acta*, 975:127–131, 1989.
- [2218] H. Kondo, K. Kagotani, M. Oshima, and M. Ishimoto. Purification and some properties of taurine dehydrogenase from a bacterium. *J. Biochem. (Tokyo)*, 73:1269–1278, 1973.
- [2219] K.H. Kondo, M.H. Kai, Y. Setoguchi, G. Eggertsen, P. Sjöblom, T. Setoguchi, K.I. Okuda, and I. Björkhem. Cloning and expression of cDNA of human Δ^4 -3-oxosteroid 5 β -reductase and substrate specificity of the expressed enzyme. *Eur. J. Biochem.*, 219:357–363, 1994.
- [2220] N. Kondo, Y. Ohno, M. Yamagata, T. Obara, N. Seki, T. Kitamura, T. Naganuma, and A. Kihara. Identification of the phytosphingosine metabolic pathway leading to odd-numbered fatty acids. *Nat. Commun.*, 5:5338–5338, 2014.
- [2221] W. Kong, S. Shiota, Y. Shi, H. Nakayama, and K. Nakayama. A novel peroxiredoxin of the plant *Sedum lineare* is a homologue of *Escherichia coli* bacterioferritin co-migratory protein (Bcp). *Biochem. J.*, 351:107–114, 2000.
- [2222] J. Konishi, Y. Ishii, T. Onaka, Y. Ohta, M. Suzuki, and K. Maruhashi. Purification and characterization of dibenzothio-phene sulfone monooxygenase and FMN-dependent NADH oxidoreductase from the thermophilic bacterium *Paenibacillus* sp. strain A11-2. *J. Biosci. Bioeng.*, 90:607–613, 2000.
- [2223] J. Konishi, H. Ohta, and G. Tuchihasi. Asymmetric reduction of benzil to benzoin catalyzed by the enzyme system of a microorganism. *Chem. Lett.*, 14:1111–1112, 1985.
- [2224] Y. Kono and I. Fridovich. Isolation and characterization of the pseudocatalase of *Lactobacillus plantarum*. *J. Biol. Chem.*, 258:6015–6019, 1983.

- [2225] A.J. Koo, T.F. Cooke, and G.A. Howe. Cytochrome P450 CYP94B3 mediates catabolism and inactivation of the plant hormone jasmonoyl-L-isoleucine. *Proc. Natl. Acad. Sci. USA*, 108:9298–9303, 2011.
- [2226] A.J. Koo, C. Thireault, S. Zemelis, A.N. Poudel, T. Zhang, N. Kitaoka, F. Brandizzi, H. Matsuura, and G.A. Howe. Endoplasmic reticulum-associated inactivation of the hormone jasmonoyl-L-isoleucine by multiple members of the cytochrome P450 94 family in *Arabidopsis*. *J. Biol. Chem.*, 289:29728–29738, 2014.
- [2227] J.A. Koo, S.P. Schmidt, and G.B. Schuster. Bioluminescence of the firefly: key steps in the formation of the electronically excited state for model systems. *Proc. Natl. Acad. Sci. USA*, 75:30–33, 1978.
- [2228] J. Koolman and P. Karlson. Ecdysone oxidase, an enzyme from the blowfly *Calliphora erythrocephala* (Meigen). *Hoppe-Seyler's Z. Physiol. Chem.*, 35:1131–1131, 1975.
- [2229] F. Koopman, N. Wierckx, J.H. de Winde, and H.J. Ruijsenaars. Identification and characterization of the furfural and 5-(hydroxymethyl)furfural degradation pathways of *Cupriavidus basilensis* HMF14. *Proc. Natl. Acad. Sci. USA*, 107:4919–4924, 2010.
- [2230] V.K. Korboukh, N. Li, E.W. Barr, J.M. Bollinger, Krebs Jr., and C. A long-lived, substrate-hydroxylating peroxidase-iron(III/III) intermediate in the amine oxygenase, AurF, from *Streptomyces thioluteus*. *J. Am. Chem. Soc.*, 131:13608–13609, 2009.
- [2231] J.M. Korff and J. Jarabak. Isolation and properties of an NADP⁺-dependent PGI₂-specific 15-hydroxyprostaglandin dehydrogenase from rabbit kidney. *Methods Enzymol.*, 86:152–155, 1982.
- [2232] J. Körhle. Iodothyronine deiodinases. *Methods Enzymol.*, 347:125–167, 2002.
- [2233] S.B. Koritz. The conversion of prenenolone to progesterone by small particle from rat adrenal. *Biochemistry*, 3:1098–1102, 1964.
- [2234] A.W. Kormann, R.O. Hurst, and T.G. Flynn. Purification and properties of an NADP⁺-dependent glycerol dehydrogenase from rabbit skeletal muscle. *Biochim. Biophys. Acta*, 258:40–55, 1972.
- [2235] A. Kornberg and W.E. Pricer. Di- and triphosphopyridine nucleotide isocitric dehydrogenase in yeast. *J. Biol. Chem.*, 189:123–136, 1951.
- [2236] S. Korwar, B.L. Morris, H.I. Parikh, R.A. Coover, T.W. Doughty, I.M. Love, B.J. Hilbert, W.E. Royer, Kellogg Jr., Grossman G.E., Ellis S.R., and K.C. Design, synthesis, and biological evaluation of substrate-competitive inhibitors of C-terminal Binding Protein (CtBP). *Bioorg. Med. Chem.*, 24:2707–2715, 2016.
- [2237] T. Koshiba and H. Matsuyama. An in vitro system of indole-3-acetic acid formation from tryptophan in maize (*Zea mays*) coleoptile extracts. *Plant Physiol.*, 102:1319–1324, 1993.
- [2238] T. Koshiba, E. Saito, N. Ono, N. Yamamoto, and M. Sato. Purification and properties of flavin- and molybdenum-containing aldehyde oxidase from coleoptiles of maize. *Plant Physiol.*, 110:781–789, 1996.
- [2239] A. Koshkin, G.M. Knudsen, and P.R. Ortiz De Montellano. Intermolecular interactions in the AhpC/AhpD antioxidant defense system of *Mycobacterium tuberculosis*. *Arch. Biochem. Biophys.*, 427:41–47, 2004.
- [2240] A. Koshkin, C.M. Nunn, S. Djordjevic, and P.R. Ortiz de Montellano. The mechanism of *Mycobacterium tuberculosis* alkylhydroperoxidase AhpD as defined by mutagenesis, crystallography, and kinetics. *J. Biol. Chem.*, 278:29502–29508, 2003.
- [2241] H. Koskiniemi, T. Grocholski, G. Schneider, and J. Niemi. Expression, purification and crystallization of the cofactor-independent monooxygenase SnoaB from the nogalamycin biosynthetic pathway. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 65:256–259, 2009.
- [2242] V. Kostanjevecki, A. Brige, T.E. Meyer, M.A. Cusanovich, Y. Guisez, and J. van Beeumen. A membrane-bound flavocytochrome *c*-sulfide dehydrogenase from the purple phototrophic sulfur bacterium *Ectothiorhodospira vacuolata*. *J. Bacteriol.*, 182:3097–3103, 2000.
- [2243] T. Kosuge, M.G. Heskett, and E.E. Wilson. Microbial synthesis and degradation of indole-3-acetic acid. I. The conversion of L-tryptophan to indole-3-acetamide by an enzyme system from *Pseudomonas savastanoi*. *J. Biol. Chem.*, 241:3738–3744, 1966.

- [2244] T. Kotani, T. Yamamoto, H. Yurimoto, Y. Sakai, and N. Kato. Propane monooxygenase and NAD⁺-dependent secondary alcohol dehydrogenase in propane metabolism by *Gordonia* sp. strain TY-5. *J. Bacteriol.*, 185:7120–7128, 2003.
- [2245] T. Kotani, H. Yurimoto, N. Kato, and Y. Sakai. Novel acetone metabolism in a propane-utilizing bacterium, *Gordonia* sp. strain TY-5. *J. Bacteriol.*, 189:886–893, 2007.
- [2246] K. Koteva, G. Cox, J.K. Kelso, M.D. Surette, H.L. Zubyk, L. Ejim, P. Stogios, A. Savchenko, D. Sørensen, and G.D. Wright. Rox, a rifamycin resistance enzyme with an unprecedented mechanism of action. *Cell Chem Biol*, 25:403–412.e5, 2018.
- [2247] V.P. Kotsira and Y.D. Clonis. Oxalate oxidase from barley roots: purification to homogeneity and study of some molecular, catalytic, and binding properties. *Arch. Biochem. Biophys.*, 340:239–249, 1997.
- [2248] S. Kottogoda, E. Waligora, and M. Hyman. Metabolism of 2-methylpropene (isobutylene) by the aerobic bacterium *Mycobacterium* sp. strain ELW1. *Appl. Environ. Microbiol.*, 81:1966–1976, 2015.
- [2249] M. Koutmos, C. Gherasim, J.L. Smith, and R. Banerjee. Structural basis of multifunctionality in a vitamin B₁₂-processing enzyme. *J. Biol. Chem.*, 286:29780–29787, 2011.
- [2250] H. Koyama. Purification and characterization of a novel L-phenylalanine oxidase (deaminating and decarboxylating) from *Pseudomonas* sp. P-501. *J. Biochem. (Tokyo)*, 92:1235–1240, 1982.
- [2251] H. Koyama. A simple and rapid enzymatic determination of L-phenylalanine with a novel L-phenylalanine oxidase (deaminating and decarboxylating) from *Pseudomonas* sp. P-501. *Clin. Chim. Acta*, 1361:131–136, 1984.
- [2252] H. Koyama. Oxidation and oxygenation of L-amino acids catalyzed by a L-phenylalanine oxidase (deaminating and decarboxylating) from *Pseudomonas* sp. P-501. *J. Biochem. (Tokyo)*, 96:421–427, 1984.
- [2253] H. Koyama and H. Suzuki. Spectral and kinetic studies on *Pseudomonas* L-phenylalanine oxidase (deaminating and decarboxylating). *J. Biochem. (Tokyo)*, 100:859–866, 1986.
- [2254] Y. Kozutsumi, T. Kawano, T. Yamakawa, and A. Suzuki. Participation of cytochrome *b*₅ in CMP-*N*-acetylneuraminic acid hydroxylation in mouse liver cytosol. *J. Biochem. (Tokyo)*, 109:704–706, 1990.
- [2255] T. Krafft, A. Bowen, F. Theis, and J.M. Macy. Cloning and sequencing of the genes encoding the periplasmic-cytochrome B-containing selenate reductase of *Thauera selenatis*. *DNA*, 10:365–377, 2000.
- [2256] T. Krafft and J.M. Macy. Purification and characterization of the respiratory arsenate reductase of *Chrysiogenes arsenatis*. *Eur. J. Biochem.*, 255:647–653, 1998.
- [2257] W. Kramer, K. Sauber, K.H. Baringhaus, M. Kurz, S. Stengelin, G. Lange, D. Corsiero, F. Girbig, W. König, and C. Weyland. Identification of the bile acid-binding site of the ileal lipid-binding protein by photoaffinity labeling, matrix-assisted laser desorption ionization-mass spectrometry, and NMR structure. *J. Biol. Chem.*, 276:7291–7301, 2001.
- [2258] V.M. Kramlinger, L.D. Nagy, R. Fujiwara, K.M. Johnson, T.T. Phan, Y. Xiao, J.M. Enright, M.B. Toomey, J.C. Corbo, and F.P. Guengerich. Human cytochrome P450 27C1 catalyzes 3,4-desaturation of retinoids. *FEBS Lett.*, 590:1304–1312, 2016.
- [2259] Z. Krejčík, K. Hollemeyer, T.H. Smits, and A.M. Cook. Isethionate formation from taurine in *Chromohalobacter salexigenis*: purification of sulfoacetaldehyde reductase. *Microbiology*, 156:1547–1555, 2010.
- [2260] Z. Krejčík, K. Denger, S. Weinitschke, K. Hollemeyer, V. Pačes, A.M. Cook, and T.H.M. Smits. Sulfoacetate released during the assimilation of taurine-nitrogen by *Neptuniibacter caesariensis*: purification of sulfoacetaldehyde dehydrogenase. *Arch. Microbiol.*, 190:159–168, 2008.
- [2261] T.A. Krenitsky, S.M. Neil, G.B. Elion, and G.H. Hitchings. A comparison of the specificities of xanthine oxidase and aldehyde oxidase. *Arch. Biochem. Biophys.*, 150:585–599, 1972.
- [2262] V.L. Kretovich and K.M. Stepanovich. The enzyme catalyzing the reductive amination of oxypyruvate. *Izv. Akad. Nauk SSSR Biol.*, 2:295–300, 1966.
- [2263] W.L. Kretovich, T.I. Kariakina, M.K. Weinova, L.I. Sidelnikova, and O.W. Kazakova. The synthesis of aspartic acid in *Rhizobium lupini* bacteroids. *Plant Soil*, 61:145–156, 1981.

- [2264] H. Kries, F. Kellner, M.O. Kamileen, and S.E. O'Connor. Inverted stereocontrol of iridoid synthase in snapdragon. *J. Biol. Chem.*, 292:14659–14667, 2017.
- [2265] C. Kristensen, M. Morant, C.E. Olsen, C.T. Ekstrøm, D.W. Galbraith, B.L. Møller, and S. Bak. Metabolic engineering of dhurrin in transgenic *Arabidopsis* plants with marginal inadvertent effects on the metabolome and transcriptome. *Proc. Natl. Acad. Sci. USA*, 102:1779–1784, 2005.
- [2266] J.E. Krochko, G.D. Abrams, M.K. Loewen, S.R. Abrams, and A.J. Cutler. (+)-Abscisic acid 8'-hydroxylase is a cytochrome P₄₅₀ monooxygenase. *Plant Physiol.*, 118:849–860, 1998.
- [2267] H. Krugel, P. Krubasik, K. Weber, H.P. Saluz, and G. Sandmann. Functional analysis of genes from *Streptomyces griseus* involved in the synthesis of isorenieratene, a carotenoid with aromatic end groups, revealed a novel type of carotenoid desaturase. *Biochim. Biophys. Acta*, 1439:57–64, 1999.
- [2268] T. Kruse, K. Ho, H.D. Yoo, T. Johnson, M. Hippely, J.H. Park, R. Flavell, and S. Bobzin. In planta biocatalysis screen of P450s identifies 8-methoxypsoralen as a substrate for the CYP82C subfamily, yielding original chemical structures. *Chem. Biol.*, 15:149–156, 2008.
- [2269] I.C. Kuan and M. Tien. Stimulation of Mn peroxidase activity: a possible role for oxalate in lignin biodegradation. *Proc. Natl. Acad. Sci. USA*, 90:1242–1246, 1993.
- [2270] R.L. Kubiak and H.M. Holden. Combined structural and functional investigation of a C-3''-ketoreductase involved in the biosynthesis of dTDP-L-digitoxose. *Biochemistry*, 50:5905–5917, 2011.
- [2271] R.L. Kubiak, R.K. Phillips, M.W. Zmudka, M.R. Ahn, E.M. Maka, G.L. Pyeatt, S.J. Roggensack, and H.M. Holden. Structural and functional studies on a 3'-epimerase involved in the biosynthesis of dTDP-6-deoxy-D-allose. *Biochemistry*, 51:9375–9383, 2012.
- [2272] T. Kubota, Y. Tanaka, K. Hiraga, M. Inui, and H. Yukawa. Characterization of shikimate dehydrogenase homologues of *Corynebacterium glutamicum*. *Appl. Microbiol. Biotechnol.*, 97:8139–8149, 2013.
- [2273] Y. Kuchino, H. Kasai, K. Nihei, and S. Nishimura. Biosynthesis of the modified nucleoside Q in transfer RNA. *Nucleic Acids Res.*, 3:393–398, 1976.
- [2274] F. Kudo, Y. Yamamoto, K. Yokoyama, T. Eguchi, and K. Kakinuma. Biosynthesis of 2-deoxystreptomine by three crucial enzymes in *Streptomyces fradiae* NBRC 12773. *J. Antibiot. (Tokyo)*, 58:766–774, 2005.
- [2275] A.E. Kuhm, A. Stolz, K.L. Ngai, and H.J. Knackmuss. Purification and characterization of a 1,2-dihydroxynaphthalene dioxygenase from a bacterium that degrades naphthalenesulfonic acids. *J. Bacteriol.*, 173:3795–3802, 1991.
- [2276] A. Kühn, S. Yu, and F. Giffhorn. Catabolism of 1,5-anhydro-D-fructose in *Sinorhizobium morelense* S-30.7.5: discovery, characterization, and overexpression of a new 1,5-anhydro-D-fructose reductase and its application in sugar analysis and rare sugar synthesis. *Appl. Environ. Microbiol.*, 72:1248–1257, 2006.
- [2277] K. Kuhnel, W. Blankenfeldt, J. Terner, and I. Schlichting. Crystal structures of chloroperoxidase with its bound substrates and complexed with formate, acetate, and nitrate. *J. Biol. Chem.*, 281:23990–23998, 2006.
- [2278] M. Kuhner, K. Haufschildt, A. Neumann, S. Storbeck, J. Streif, and G. Layer. The alternative route to heme in the methanogenic archaeon *Methanosarcina barkeri*. *Archaea*, 2014:327637–327637, 2014.
- [2279] T. Kühnl, U. Koch, W. Heller, and E. Wellman. Chlorogenic acid biosynthesis: characterization of a light-induced microsomal 5-O-(4-coumaroyl)-D-quinic/shikimate 3'-hydroxylase from carrot (*Daucus carota* L.) cell suspension cultures. *Arch. Biochem. Biophys.*, 258:226–232, 1987.
- [2280] J. Kuivanen and P. Richard. The *yjjN* of *E. coli* codes for an L-galactonate dehydrogenase and can be used for quantification of L-galactonate and L-gulonate. *Appl. Biochem. Biotechnol.*, 173:1829–1835, 2014.
- [2281] T. Kumano, S. Hori, S. Watanabe, Y. Terashita, H.Y. Yu, Y. Hashimoto, T. Senda, M. Senda, and M. Kobayashi. FAD-dependent C-glycoside-metabolizing enzymes in microorganisms: Screening, characterization, and crystal structure analysis. *Proc. Natl. Acad. Sci. USA*, 118(40):e2106580118–, 2021.

- [2282] A. Kumar, A.M. Balakrishna, W. Narthey, M.S.S. Manimekalai, and G. Gruber. Redox chemistry of *Mycobacterium tuberculosis* alkylhydroperoxide reductase E (AhpE): Structural and mechanistic insight into a mycoredoxin-1 independent reductive pathway of AhpE via mycothiol. *Free Radic. Biol. Med.*, 97:588–601, 2016.
- [2283] R. Kumar, S. Zhao, M.W. Vetting, B.M. Wood, A. Sakai, K. Cho, J. Solbiati, S.C. Almo, J.V. Sweedler, M.P. Jacobson, J.A. Gerlt, and J.E. Cronan. Prediction and biochemical demonstration of a catabolic pathway for the osmoprotectant proline betaine. *MBio*, 5:e00933–e00913, 2014.
- [2284] V. Kumar, J.E. Carlson, K.A. Ohgi, T.A. Edwards, D.W. Rose, C.R. Escalante, M.G. Rosenfeld, and A.K. Aggarwal. Transcription corepressor CtBP is an NAD(+)-regulated dehydrogenase. *Mol. Cell*, 10:857–869, 2002.
- [2285] O.M. Kumiko, K. Budai, and N.B. Javitt. Cholesterol and 27-hydroxycholesterol 7 α -hydroxylation: evidence for two different enzymes. *J. Lipid Res.*, 34:581–588, 1993.
- [2286] H. Kumita, K. Matsuura, T. Hino, S. Takahashi, H. Hori, Y. Fukumori, I. Morishima, and Y. Shiro. NO reduction by nitric-oxide reductase from denitrifying bacterium *Pseudomonas aeruginosa*: characterization of reaction intermediates that appear in the single turnover cycle. *J. Biol. Chem.*, 279:55247–55254, 2004.
- [2287] E. Kun, J.M. Dechary, and H.C. Pitot. The oxidation of glycolic acid by a liver enzyme. *J. Biol. Chem.*, 210:269–280, 1954.
- [2288] S.P. Kunapuli and C.S. Vaidyanathan. Purification and characterization of a new indole oxygenase from the leaves of *Tecoma stans* L. *Plant Physiol.*, 71:19–23, 1983.
- [2289] W.-H. Kunau and P. Dommes. Degradation of unsaturated fatty acids. Identification of intermediates in the degradation of *cis*-4-decenyl-CoA by extracts of beef-liver mitochondria. *Eur. J. Biochem.*, 91:533–544, 1978.
- [2290] J.W. Kung, S. Baumann, M. von Bergen, M. Muller, P.L. Hagedoorn, W.R. Hagen, and M. Boll. Reversible biological Birch reduction at an extremely low redox potential. *J. Am. Chem. Soc.*, 132:9850–9856, 2010.
- [2291] J.W. Kung, J. Seifert, M. von Bergen, and M. Boll. Cyclohexanecarboxyl-coenzyme A (CoA) and cyclohex-1-ene-1-carboxyl-CoA dehydrogenases, two enzymes involved in the fermentation of benzoate and crotonate in *Syntrophus aciditrophicus*. *J. Bacteriol.*, 195:3193–3200, 2013.
- [2292] S. Kuno, M. Tashiro, H. Taniuchi, K. Horibata, O. Hayashi, S. Seno, T. Tokuyama, and T. Sakan. Enzymatic degradation of kynurenic acid. *Fed. Proc.*, 20:3–3, 1961.
- [2293] J. Kunow, D. Linder, K.O. Stetter, and R.K. Thauer. F₄₂₀H₂: quinone oxidoreductase from *Archaeoglobus fulgidus*. Characterization of a membrane-bound multisubunit complex containing FAD and iron-sulfur clusters. *Eur. J. Biochem.*, 223:503–511, 1994.
- [2294] L. Kunst, J. Browse, , and C.R. A mutant of *Arabidopsis* deficient in desaturation of palmitic acid in leaf lipids. *Plant Physiol.*, 90:943–947, 1989.
- [2295] T.T. Kuo and T. Kosuge. Factors influencing the production and further metabolism of indole-3-acetic acid by *Pseudomonas savastanoi*. *J. Gen. Appl. Microbiol.*, 15:51–63, 1969.
- [2296] S. Kuorelahti, N. Kalkkinen, M. Penttila, J. Londesborough, and P. Richard. Identification in the mold *Hypocrea jecorina* of the first fungal D-galacturonic acid reductase. *Biochemistry*, 44:11234–11240, 2005.
- [2297] D. Kupfer, G.K. Miranda, J. Navarro, D.E. Piccolo, and A.D. Theoharides. Effect of inducers and inhibitors of monooxygenase on the hydroxylation of prostaglandins in the guinea pig. Evidence for several monooxygenases catalyzing ω - and ω -1-hydroxylation. *J. Biol. Chem.*, 254:10405–10414, 1979.
- [2298] A. Kurata, T. Kurihara, H. Kamachi, and N. Esaki. 2-Haloacrylate reductase, a novel enzyme of the medium chain dehydrogenase/reductase superfamily that catalyzes the reduction of a carbon-carbon double bond of unsaturated organohalogen compounds. *J. Biol. Chem.*, 280:20286–20291, 2005.
- [2299] P. Kurdrid, S. Subudhi, A. Hongsthong, M. Ruengjitchawalya, and M. Tanticharoen. Functional expression of *Spirulina*- Δ^6 desaturase gene in yeast, *Saccharomyces cerevisiae*. *Mol. Biol. Rep.*, 32:215–226, 2005.
- [2300] M. Kurfurst, S. Ghisla, and J.W. Hastings. Characterization and postulated structure of the primary emitter in the bacterial luciferase reaction. *Proc. Natl. Acad. Sci. USA*, 81:2990–2994, 1984.

- [2301] S. Kurihara, S. Oda, K. Kato, H.G. Kim, T. Koyanagi, H. Kumagai, and H. Suzuki. A novel putrescine utilization pathway involves γ -glutamylated intermediates of *Escherichia coli* K-12. *J. Biol. Chem.*, 280:4602–4608, 2005.
- [2302] M Kurokawa, Y Fukumori, and T Yamanaka. A hydroxylamine - cytochrome *c* reductase occurs in the heterotrophic nitrifier *Arthrobacter globiformis*. *Plant Cell Physiol.*, 26:1439–1442, 1985.
- [2303] T. Kurokawa and J. Sakamoto. Purification and characterization of succinate:menaquinone oxidoreductase from *Corynebacterium glutamicum*. *Arch. Microbiol.*, 183:317–324, 2005.
- [2304] S. Kuroki, S. Matoba, M. Akiyoshi, Y. Matsumura, H. Miyachi, N. Mise, K. Abe, A. Ogura, D. Wilhelm, P. Koopman, M. Nozaki, Y. Kanai, Y. Shinkai, and M. Tachibana. Epigenetic regulation of mouse sex determination by the histone demethylase Jmjd1a. *Science*, 341:1106–1109, 2013.
- [2305] J.M. Kurth, J.A. Brito, J. Reuter, A. Flegler, T. Koch, T. Franke, E.M. Klein, S.F. Rowe, J.N. Butt, K. Denkmann, I.A. Pereira, M. Archer, and C. Dahl. Electron accepting units of the diheme cytochrome *c* TsdA, a bifunctional thiosulfate dehydrogenase/tetrathionate reductase. *J. Biol. Chem.*, 291:24804–24818, 2016.
- [2306] K. Kusai and T. Yamanaka. The oxidation mechanisms of thiosulphate and sulphide in *Chlorobium thiosulphatophilum*: roles of cytochrome *c*-551 and cytochrome *c*-553. *Biochim. Biophys. Acta*, 325:304–314, 1973.
- [2307] H. Kusakabe, K. Kodama, A. Kuninaka, H. Yoshino, H. Misono, and K. Soda. A new antitumor enzyme, L-lysine α -oxidase from *Trichoderma viride*. Purification and enzymological properties. *J. Biol. Chem.*, 255:976–981, 1980.
- [2308] H. Kusakabe, A. Kuninaka, and H. Yoshino. Purification and properties of a new enzyme, glutathione oxidase from *Penicillium* sp.K-6-5. *Agric. Biol. Chem.*, 46:2057–2067, 1982.
- [2309] H. Kusakabe, Y. Midorikawa, T. Fujishima, A. Kuninaka, and H. Yoshino. Purification and properties of a new enzyme, L-glutamate oxidase, from *Streptomyces* sp X-119-6 grown on wheat bran. *Agric. Biol. Chem.*, 47:1323–1328, 1983.
- [2310] T. Kusudo, T. Sakaki, D. Abe, T. Fujishima, A. Kittaka, H. Takayama, S. Hatakeyama, M. Ohta, and K. Inouye. Metabolism of A-ring diastereomers of $1\alpha,25$ -dihydroxyvitamin D₃ by CYP24A1. *Biochem. Biophys. Res. Commun.*, 321:774–782, 2004.
- [2311] T.M. Kutchan and H. Dittrich. Characterization and mechanism of the berberine bridge enzyme, a covalently flavinylated oxidase of benzophenanthridine alkaloid biosynthesis in higher plants. *J. Biol. Chem.*, 270:24475–24481, 1995.
- [2312] R. Krishnan Kutty, N.A. Devi, M. Veeraswamy, S. Ramesh, and P.V. Subba Rao. Degradation of (\pm)-synephrine by *Arthrobacter synephrinum*. Oxidation of 3,4-dihydroxyphenylacetate to 2-hydroxy-5-carboxymethyl-muconate semi-aldehyde. *Biochem. J.*, 167:163–170, 1977.
- [2313] C. Kutzbach and E.L.R. Stokstad. Partial purification of a 10-formyl-tetrahydrofolate: NADP oxidoreductase from mammalian liver. *Biochem. Biophys. Res. Commun.*, 30:111–117, 1968.
- [2314] C. Kutzbach and E.L.R. Stokstad. Mammalian methylenetetrahydrofolate reductase. Partial purification, properties, and inhibition by S-adenosylmethionine. *Biochim. Biophys. Acta*, 250:459–477, 1971.
- [2315] T. Kuwabara, M. Hasegawa, M. Kawano, and S. Takaichi. Characterization of violaxanthin de-epoxidase purified in the presence of Tween 20: effects of dithiothreitol and pepstatin A. *Plant Cell Physiol.*, 40:1119–1126, 1999.
- [2316] E. Kvam, K. Gable, T.M. Dunn, and D.S. Goldfarb. Targeting of Tsc13p to nucleus-vacuole junctions: a role for very-long-chain fatty acids in the biogenesis of microautophagic vesicles. *Mol. Biol. Cell*, 16:3987–3998, 2005.
- [2317] S.-S. Kwak, Y. Kamiya, A. Sakurai, N. Takahishi, and J.E. Graebe. Partial-purification and characterization of gibberellin 3 β -hydroxylase from immature seeds of *Phaseolus vulgaris* L. *Plant Cell Physiol.*, 29:935–943, 1988.
- [2318] H.S. Kwan and E.L. Barrett. Map locations and functions of *Salmonella typhimurium* men genes. *J. Bacteriol.*, 159:1090–1092, 1984.
- [2319] S. Kwiatkowski, M. Bozko, M. Zarod, A. Witecka, K. Kocdemir, A.K. Jagielski, and J. Drozak. Recharacterization of the mammalian cytosolic type 2 (*R*)- β -hydroxybutyrate dehydrogenase (BDH2) as 4-oxo-L-proline reductase (EC 1.1.1.104). *J. Biol. Chem.*, 298:101708–101708, 2022.

- [2320] O. Kwon, A. Kotsakis, and R. Meganathan. Ubiquinone (coenzyme Q) biosynthesis in *Escherichia coli*: identification of the *ubiF* gene. *FEMS Microbiol. Lett.*, 186:157–161, 2000.
- [2321] E.F. LaBelle, Hajira Jr., and A.K. Enzymatic reduction of alkyl and acyl derivatives of dihydroxyacetone phosphate by reduced pyridine nucleotides. *J. Biol. Chem.*, 247:5825–5834, 1972.
- [2322] B. Laber, W. Maurer, S. Scharf, K. Stepusin, and F.S. Schmidt. Vitamin B₆ biosynthesis: formation of pyridoxine 5'-phosphate from 4-(phosphohydroxy)-L-threonine and 1-deoxy-D-xylulose-5-phosphate by PdxA and PdxJ protein. *FEBS Lett.*, 449:45–48, 1999.
- [2323] A.L. Laborde and D.T. Gibson. Metabolism of dibenzothiophene by a *Beijerinckia* species. *Appl. Environ. Microbiol.*, 34:783–790, 1977.
- [2324] L. Lad, D.J. Schuller, H. Shimizu, J. Friedman, H. Li, P.R. Ortiz de Montellano, and T.L. Poulos. Comparison of the heme-free and -bound crystal structures of human heme oxygenase-1. *J. Biol. Chem.*, 278:7834–7843, 2003.
- [2325] D. Laempe, M. Jahn, and G. Fuchs. 6-Hydroxycyclohex-1-ene-1-carbonyl-CoA dehydrogenase and 6-oxocyclohex-1-ene-1-carbonyl-CoA hydrolase, enzymes of the benzoyl-CoA pathway of anaerobic aromatic metabolism in the denitrifying bacterium *Thauera aromatica*. *Eur. J. Biochem.*, 263:420–429, 1999.
- [2326] M.A. Lafferty and R.H. Garrett. Purification and properties of the *Neurospora crassa* assimilatory nitrite reductase. *J. Biol. Chem.*, 249:7555–7567, 1974.
- [2327] V. Lakshmi and C. Monder. Purification and characterization of the corticosteroid 11 β -dehydrogenase component of the rat liver 11 β -hydroxysteroid dehydrogenase complex. *Endocrinology*, 123:2390–2398, 1988.
- [2328] H.M. Lam and M.E. Winkler. Metabolic relationships between pyridoxine (vitamin B₆) and serine biosynthesis in *Escherichia coli* K-12. *J. Bacteriol.*, 172:6518–6528, 1990.
- [2329] L.K. Lam, Z. Zhang, P.G. Board, and L. Xun. Reduction of benzoquinones to hydroquinones via spontaneous reaction with glutathione and enzymatic reaction by *S*-glutathionyl-hydroquinone reductases. *Biochemistry*, 51:5014–5021, 2012.
- [2330] P.Y. Lam, F.Y. Zhu, W.L. Chan, H. Liu, and C. Lo. Cytochrome P450 93G1 is a flavone synthase II that channels flavanones to the biosynthesis of triclin *O*-linked conjugates in rice. *Plant Physiol.*, 165:1315–1327, 2014.
- [2331] W. W. Y Lam and T. D. H. Bugg. Chemistry of extradiol aromatic ring cleavage: isolation of a stable dienol ring fission intermediate and stereochemistry of its enzymatic hydrolytic cleavage. *J. Chem. Soc., Chem. Commun.*, 10:1163–1164, 1994.
- [2332] R.H. Lambalot, D.E. Cane, J.J. Aparicio, and L. Katz. Overproduction and characterization of the erythromycin C-12 hydroxylase, EryK. *Biochemistry*, 34:1858–1866, 1995.
- [2333] A. Lamberg, T. Pihlajaniemi, and K.I. Kivirikko. Site-directed mutagenesis of the α subunit of human prolyl 4-hydroxylase. Identification of three histidine residues critical for catalytic activity. *J. Biol. Chem.*, 270:9926–9931, 1995.
- [2334] H.J. Lambie, N.I. Heyer, S.D. Bull, D.W. Hough, and M.J. Danson. Metabolic pathway promiscuity in the archaeon *Sulfolobus solfataricus* revealed by studies on glucose dehydrogenase and 2-keto-3-deoxygluconate aldolase. *J. Biol. Chem.*, 278:34066–34072, 2003.
- [2335] M. Lammers and H. Follmann. The ribonucleotide reductases - a unique group of metalloenzymes essential for cell-proliferation. *Struct. Bonding*, 54:27–91, 1983.
- [2336] F. Lan, P.E. Bayliss, J.L. Rinn, J.R. Whetstine, J.K. Wang, S. Chen, S. Iwase, R. Alpatov, I. Issaeva, E. Canaani, T.M. Roberts, H.Y. Chang, and Y. Shi. A histone H3 lysine 27 demethylase regulates animal posterior development. *Nature*, 449:689–694, 2007.
- [2337] P. Lanciano, A. Magalon, P. Bertrand, B. Guigliarelli, and S. Grimaldi. High-stability semiquinone intermediate in nitrate reductase A (NarGHI) from *Escherichia coli* is located in a quinol oxidation site close to heme *b_L*. *Biochemistry*, 46:5323–5329, 2007.
- [2338] D. Lando, D.J. Peet, D.A. Whelan, J.J. Gorman, and M.L. Whitelaw. Asparagine hydroxylation of the HIF transactivation domain a hypoxic switch. *Science*, 295:858–861, 2002.

- [2339] J. Landry and R. Sternglanz. Yeast Fms1 is a FAD-utilizing polyamine oxidase. *Biochem. Biophys. Res. Commun.*, 303:771–776, 2003.
- [2340] S.G. Van Lanen, S. Lin, and B. Shen. Biosynthesis of the enediyne antitumor antibiotic C-1027 involves a new branching point in chorismate metabolism. *Proc. Natl. Acad. Sci. USA*, 105:494–499, 2008.
- [2341] S.G. Van Lanen, J.S. Reader, M.A. Swairjo, V. de Crécy-Lagard, B. Lee, and D. Iwata-Reuyl. From cyclohydrolase to oxidoreductase: discovery of nitrile reductase activity in a common fold. *Proc. Natl. Acad. Sci. USA*, 102:4264–4269, 2005.
- [2342] I. Lang, C. Gobel, A. Porzel, I. Heilmann, and I. Feussner. A lipoxygenase with linoleate diol synthase activity from *Nostoc* sp. PCC 7120. *Biochem. J.*, 410:347–357, 2008.
- [2343] M.A. Lang, J.E. Gielen, and D.W. Nebert. Genetic evidence for many unique liver microsomal *P*-450-mediated monooxygenase activities in heterogenic stock mice. *J. Biol. Chem.*, 256:12068–12075, 1981.
- [2344] M.A. Lang and D.W. Nebert. Structural gene products of the Ah locus. Evidence for many unique *P*-450-mediated monooxygenase activities reconstituted from 3-methylcholanthrene-treated C57BL/6N mouse liver microsomes. *J. Biol. Chem.*, 256:12058–12075, 1981.
- [2345] C. Lange, S. Kiesel, S. Peters, S. Virus, H. Scheer, D. Jahn, and J. Moser. Broadened substrate specificity of 3-hydroxyethyl bacteriochlorophyllide *a* dehydrogenase (BchC) indicates a new route for the biosynthesis of bacteriochlorophyll *a*. *J. Biol. Chem.*, 290:19697–19709, 2015.
- [2346] C.C. Lange, B.J. Schneider, and C.S. Orser. Verification of the role of PCP 4-monooxygenase in chlorine elimination from pentachlorophenol by *Flavobacterium* sp. strain ATCC 39723. *Biochem. Biophys. Res. Commun.*, 219:146–149, 1996.
- [2347] L.J. Langer, J.A. Alexander, and L.L. Engel. Human placental estradiol-17 β dehydrogenase. II. Kinetics and substrate specificities. *J. Biol. Chem.*, 234:2609–2614, 1959.
- [2348] B. Langkau, S. Ghisla, R. Buder, K. Ziegler, and G. Fuchs. 2-Aminobenzoyl-CoA monooxygenase/reductase, a novel type of flavoenzyme. Identification of the reaction products. *Eur. J. Biochem.*, 191:365–371, 1990.
- [2349] F.J.S. Lara. On the decomposition of pyrimidines by bacteria. II. Studies with cell-free enzyme preparations. *J. Bacteriol.*, 64:279–285, 1952.
- [2350] R. Larbat, A. Hehn, J. Hans, S. Schneider, H. Jugde, B. Schneider, U. Matern, and F. Bourgaud. Isolation and functional characterization of CYP71AJ4 encoding for the first *P*₄₅₀ monooxygenase of angular furanocoumarin biosynthesis. *J. Biol. Chem.*, 284:4776–4785, 2009.
- [2351] R. Larbat, S. Kellner, S. Specker, A. Hehn, E. Gontier, J. Hans, F. Bourgaud, and U. Matern. Molecular cloning and functional characterization of psoralen synthase, the first committed monooxygenase of furanocoumarin biosynthesis. *J. Biol. Chem.*, 282:542–554, 2007.
- [2352] A.T. Large, C.J.P. Jones, and M.J. Connock. The association of mannitol oxidase with a distinct organelle in the digestive gland of the terrestrial slug *Arion ater*. *Protoplasma*, 175:93–101, 1993.
- [2353] P.J. Large, C.A. Boulton, and M.J. Crabbe. The reduced nicotinamide-adenine dinucleotide phosphate- and oxygen-dependent *N*-oxygenation of trimethylamine by *Pseudomonas aminovorans*. *Biochem. J.*, 128:137P–138P, 1972.
- [2354] A. Larkin and B. Imperiali. Biosynthesis of UDP-GlcNAc(3NAc)A by WbpB, WbpE, and WbpD: enzymes in the Wbp pathway responsible for O-antigen assembly in *Pseudomonas aeruginosa* PAO1. *Biochemistry*, 48:5446–5455, 2009.
- [2355] J. Larner, W.T. Jackson, D.J. Graves, and J.R. Stammer. Inositol dehydrogenase from *Aerobacter aerogenes*. *Arch. Biochem. Biophys.*, 60:352–363, 1956.
- [2356] A. Larsson. Ribonucleotide reductase from regenerating rat liver. II. Substrate phosphorylation level and effect of deoxyadenosine triphosphate. *Biochim. Biophys. Acta*, 324:447–451, 1973.
- [2357] A. Larsson and P. Reichard. Enzymatic synthesis of deoxyribonucleotides. IX. Allosteric effects in the reduction of pyrimidine ribonucleotides by the ribonucleoside diphosphate reductase system of *Escherichia coli*. *J. Biol. Chem.*, 241:2533–2539, 1966.

- [2358] A. Larsson and P. Reichard. Enzymatic synthesis of deoxyribonucleotides. X. Reduction of purine ribonucleotides; allosteric behavior and substrate specificity of the enzyme system from *Escherichia coli* B. *J. Biol. Chem.*, 241:2540–2549, 1966.
- [2359] C. Larsson, I.L. Pählman, R. Ansell, M. Rigoulet, L. Adler, and L. Gustafsson. The importance of the glycerol 3-phosphate shuttle during aerobic growth of *Saccharomyces cerevisiae*. *Yeast*, 14:347–357, 1998.
- [2360] J.A. Latham, A.T. Iavarone, I. Barr, P.V. Juthani, and J.P. Klinman. PqqD is a novel peptide chaperone that forms a ternary complex with the radical *S*-adenosylmethionine protein PqqE in the pyrroloquinoline quinone biosynthetic pathway. *J. Biol. Chem.*, 290:12908–12918, 2015.
- [2361] D. Latowski, H.E. Akerlund, and K. Strzalka. Violaxanthin de-epoxidase, the xanthophyll cycle enzyme, requires lipid inverted hexagonal structures for its activity. *Biochemistry*, 43:4417–4420, 2004.
- [2362] D. Latowski, J. Kruk, K. Burda, M. Skrzynecka-Jaskiern, A. KostECKA-GUGALA, and K. Strzalka. Kinetics of violaxanthin de-epoxidation by violaxanthin de-epoxidase, a xanthophyll cycle enzyme, is regulated by membrane fluidity in model lipid bilayers. *Eur. J. Biochem.*, 269:4656–4665, 2002.
- [2363] P.C.K. Lau, D.S. Layne, and D.G. Williamson. A 3(17) α -hydroxysteroid dehydrogenase of female rabbit kidney cytosol. Purification and characterization of multiple forms of the enzyme. *J. Biol. Chem.*, 257:9444–9449, 1982.
- [2364] P.C.K. Lau, D.S. Layne, and D.G. Williamson. Comparison of the multiple forms of the soluble 3(17) α -hydroxysteroid dehydrogenases of female rabbit kidney and liver. *J. Biol. Chem.*, 257:9450–9456, 1982.
- [2365] W. Lau and E.S. Sattely. Six enzymes from mayapple that complete the biosynthetic pathway to the etoposide aglycone. *Science*, 349:1224–1228, 2015.
- [2366] S. Lautru, M. Gondry, R. Genet, and J.L. Pernodet. The albonoursin gene cluster of *S. noursei*. Biosynthesis of dike-topiperazine metabolites independent of nonribosomal peptide synthetases. *Chem. Biol.*, 9:1355–1364, 2002.
- [2367] J. Laville, C. Blumer, C. Von Schroetter, V. Gaia, G. Defago, C. Keel, and D. Haas. Characterization of the hcnABC gene cluster encoding hydrogen cyanide synthase and anaerobic regulation by ANR in the strictly aerobic biocontrol agent *Pseudomonas fluorescens* CHA0. *J. Bacteriol.*, 180:3187–3196, 1998.
- [2368] C.C. Lawrence, M. Bennati, H.V. Obias, G. Bar, R.G. Griffin, and J. Stubbe. High-field EPR detection of a disulfide radical anion in the reduction of cytidine 5'-diphosphate by the E441Q R1 mutant of *Escherichia coli* ribonucleotide reductase. *Proc. Natl. Acad. Sci. USA*, 96:8979–8984, 1999.
- [2369] C.C. Lawrence, W.J. Sobey, R.A. Field, J.E. Baldwin, and C.J. Schofield. Purification and initial characterization of proline 4-hydroxylase from *Streptomyces griseoviridis* P8648: a 2-oxoacid, ferrous-dependent dioxygenase involved in etamycin biosynthesis. *Biochem. J.*, 313:185–191, 1996.
- [2370] C.C. Lawrence and J. Stubbe. The function of adenosylcobalamin in the mechanism of ribonucleoside triphosphate reductase from *Lactobacillus leichmannii*. *Curr. Opin. Chem. Biol.*, 2:650–655, 1998.
- [2371] G. Layer, J. Moser, D.W. Heinz, D. Jahn, and W.D. Schubert. Crystal structure of coproporphyrinogen III oxidase reveals cofactor geometry of radical SAM enzymes. *EMBO J.*, 22:6214–6224, 2003.
- [2372] G. Layer, K. Verfürth, E. Mahlitz, and D. Jahn. Oxygen-independent coproporphyrinogen-III oxidase HemN from *Escherichia coli*. *J. Biol. Chem.*, 277:34136–34142, 2002.
- [2373] P.J. Lea and B.J. Mifflin. Alternative route for nitrogen assimilation in higher plants. *Nature (Lond.)*, 251:614–616, 1974.
- [2374] C. Leadbeater, L. McIver, D.J. Campopiano, S.P. Webster, R.L. Baxter, S.M. Kelly, N.C. Price, D.A. Lysek, M.A. Noble, S.K. Chapman, and A.W. Munro. Probing the NADPH-binding site of *Escherichia coli* flavodoxin oxidoreductase. *Biochem. J.*, 352:257–266, 2000.
- [2375] D. Leclerc, A. Wilson, R. Dumas, C. Gafuik, D. Song, D. Watkins, H.H.Q. Heng, J.M. Rommens, S.W. Scherer, D.S. Rosenblatt, , and R.A. Cloning and mapping of a cDNA for methionine synthase reductase, a flavoprotein defective in patients with homocystinuria. *Proc. Natl. Acad. Sci. USA*, 95:3059–3064, 1998.
- [2376] A.K. Lee, A.B. Banta, J.H. Wei, D.J. Kiemle, J. Feng, J.L. Giner, and P.V. Welander. C-4 sterol demethylation enzymes distinguish bacterial and eukaryotic sterol synthesis. *Proc. Natl. Acad. Sci. USA*, 115:5884–5889, 2018.

- [2377] B.H. Lee, W.K. Huh, S.T. Kim, J.S. Lee, and S.O. Kang. Bacterial production of D-erythroascorbic acid and L-ascorbic acid through functional expression of *Saccharomyces cerevisiae* D-arabinono-1,4-lactone oxidase in *Escherichia coli*. *Appl. Environ. Microbiol.*, 65:4685–4687, 1999.
- [2378] B.I. Lee, C. Chang, S.J. Cho, S.H. Eom, K.K. Kim, Y.G. Yu, and S.W. Suh. Crystal structure of the MJ0490 gene product of the hyperthermophilic archaeobacterium *Methanococcus jannaschii*, a novel member of the lactate/malate family of dehydrogenases. *J. Mol. Biol.*, 307:1351–1362, 2001.
- [2379] C.A. Lee, D. Neul, A. Clouser-Roche, D. Dalvie, M.R. Wester, Y. Jiang, J.P. Jones, Freiwald 3rd, Zientek S., Totah M., and R.A. Identification of novel substrates for human cytochrome P450 2J2. *Drug Metab. Dispos.*, 38:347–356, 2010.
- [2380] C.C. Lee, Y. Hu, and M.W. Ribbe. Vanadium nitrogenase reduces CO. *Science*, 329:642–642, 2010.
- [2381] C.C. Lee, Y. Hu, and M.W. Ribbe. Tracing the hydrogen source of hydrocarbons formed by vanadium nitrogenase. *Angew. Chem. Int. Ed. Engl.*, 50:5545–5547, 2011.
- [2382] D.S. Lee, A. Yamada, H. Sugimoto, I. Matsunaga, H. Ogura, K. Ichihara, S. Adachi, S.Y. Park, and Y. Shiro. Substrate recognition and molecular mechanism of fatty acid hydroxylation by cytochrome *P*₄₅₀ from *Bacillus subtilis*. Crystallographic, spectroscopic, and mutational studies. *J. Biol. Chem.*, 278:9761–9767, 2003.
- [2383] H.J. Lee, M.D. Lloyd, K. Harlos, I.J. Clifton, J.E. Baldwin, and C.J. Schofield. Kinetic and crystallographic studies on deacetoxycephalosporin C synthase (DAOCS). *J. Mol. Biol.*, 308:937–948, 2001.
- [2384] J. Lee, V. Sperandio, D.E. Frantz, J. Longgood, A. Camilli, M.A. Phillips, and A.J. Michael. An alternative polyamine biosynthetic pathway is widespread in bacteria and essential for biofilm formation in *Vibrio cholerae*. *J. Biol. Chem.*, 284:9899–9907, 2009.
- [2385] J. Lee and H. Zhao. Mechanistic studies on the conversion of arylamines into aryl nitro compounds by aminopyrrolnitrin oxygenase: identification of intermediates and kinetic studies. *Angew. Chem. Int. Ed. Engl.*, 45:622–625, 2006.
- [2386] J.K. Lee, B.S. Koo, and S.Y. Kim. Cloning and characterization of the *xy11* gene, encoding an NADH-preferring xylose reductase from *Candida parapsilosis*, and its functional expression in *Candida tropicalis*. *Appl. Environ. Microbiol.*, 69:6179–6188, 2003.
- [2387] J.L. Lee and M.J. Fasco. Metabolism of vitamin K and vitamin K 2,3-epoxide via interaction with a common disulfide. *Biochemistry*, 23:2246–2252, 1984.
- [2388] M. Lee, M. Lenman, A. Banas, M. Bafor, S. Singh, M. Schweizer, R. Nilsson, C. Liljenberg, A. Dahlqvist, P.O. Gummeson, S. Sjö Dahl, A. Green, and S. Szymne. Identification of non-heme di-iron proteins that catalyze triple bond and epoxy group formation. *Science*, 280:915–918, 1998.
- [2389] M.G. Lee, R. Villa, P. Trojer, J. Norman, K.P. Yan, D. Reinberg, L. Di Croce, and R. Shiekhattar. Demethylation of H₃K27 regulates polycomb recruitment and H2A ubiquitination. *Science*, 318:447–450, 2007.
- [2390] P.C. Lee, E. Holtzapple, and C. Schmidt-Dannert. Novel activity of *Rhodobacter sphaeroides* spheroidene monooxygenase CrtA expressed in *Escherichia coli*. *Appl. Environ. Microbiol.*, 76:7328–7331, 2010.
- [2391] S. Lee, S. Badiyan, D.R. Bevan, M. Herde, C. Gatz, and D. Tholl. Herbivore-induced and floral homoterpene volatiles are biosynthesized by a single P450 enzyme (CYP82G1) in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA*, 107:21205–21210, 2010.
- [2392] S. Lee, H. Jeon, P. Giri, U.J. Lee, H. Jung, S. Lim, S. Sarak, T.P. Khobragade, B.G. Kim, and H. Yun. The reductive amination of carbonyl compounds using native amine dehydrogenase from *Laribacter hongkongensis*. *Biotechnol. Bioprocess Eng.*, 26:384–391, 2021.
- [2393] S.-C. Lee and L. Levine. Prostaglandin metabolism. II. Identification of two 15-hydroxyprostaglandin dehydrogenase types. *J. Biol. Chem.*, 250:548–552, 1975.
- [2394] S.-C. Lee and L. Levine. Purification and regulatory properties of chicken heart prostaglandin E 9-ketoreductase. *J. Biol. Chem.*, 250:4549–4555, 1975.
- [2395] S.-C. Lee, S.-S. Pong, D. Katzen, K.-Y. Wu, and L. Levine. Distribution of prostaglandin E 9-ketoreductase and types I and II 15-hydroxyprostaglandin dehydrogenase in swine kidney medulla and cortex. *Biochemistry*, 14:142–145, 1975.

- [2396] S.A. Lee, O.V. Belyaeva, and N.Y. Kedishvili. Biochemical characterization of human epidermal retinol dehydrogenase 2. *Chem. Biol. Interact.*, 178:182–187, 2009.
- [2397] S.B. Lee, S.J. Cho, J.A. Kim, S.Y. Lee, S.M. Kim, and H.S. Lim. Metabolic pathway of 3,6-anhydro-L-galactose in agar-degrading microorganisms. *Biotechnol. Bioprocess Eng.*, 19:866–878, 2014.
- [2398] T.-C. Lee. Characterization of fatty alcohol:NAD⁺ oxidoreductase from rat liver. *J. Biol. Chem.*, 254:2892–2896, 1979.
- [2399] W.T. Lee and H.R. Levy. Lysine-21 of *Leuconostoc mesenteroides* glucose 6-phosphate dehydrogenase participates in substrate binding through charge-charge interaction. *Protein Sci.*, 1:329–334, 1992.
- [2400] Y. Lee and L.M. Sayre. Reaffirmation that metabolism of polyamines by bovine plasma amine oxidase occurs strictly at the primary amino termini. *J. Biol. Chem.*, 273:19490–19494, 1998.
- [2401] Y.C. Lee, M.J. Nelson, and E.E. Snell. Enzymes of vitamin B₆ degradation. Purification and properties of isopyridoxal dehydrogenase and 5-formyl-3-hydroxy-2-methylpyridine-4-carboxylic-acid dehydrogenase. *J. Biol. Chem.*, 261:15106–15111, 1986.
- [2402] J.M. Leeds, P.J. Brown, G.M. McGeehan, F.K. Brown, and J.S. Wiseman. Isotope effects and alternative substrate reactivities for tryptophan 2,3-dioxygenase. *J. Biol. Chem.*, 268:17781–17786, 1993.
- [2403] L. Lo Leggio, T.J. Simmons, J.C. Poulsen, K.E. Frandsen, G.R. Hemsworth, M.A. Stringer, P. von Freiesleben, M. Tovborg, K.S. Johansen, L. De Maria, P.V. Harris, C.L. Soong, P. Dupree, T. Tryfona, N. Lenfant, B. Henrissat, G.J. Davies, and P.H. Walton. Structure and boosting activity of a starch-degrading lytic polysaccharide monooxygenase. *Nat. Commun.*, 6:5961–5961, 2015.
- [2404] M. Lehmann, B. Tshisuaka, S. Fetzner, and F. Lingens. Molecular cloning of the isoquinoline 1-oxidoreductase genes from *Pseudomonas diminuta* 7, structural analysis of iorA and iorB, and sequence comparisons with other molybdenum-containing hydroxylases. *J. Biol. Chem.*, 270:14420–14429, 1995.
- [2405] M. Lehmann, B. Tshisuaka, S. Fetzner, P. Roger, and F. Lingens. Purification and characterization of isoquinoline 1-oxidoreductase from *Pseudomonas diminuta* 7, a novel molybdenum-containing hydroxylase. *J. Biol. Chem.*, 269:11254–11260, 1994.
- [2406] A.L. Lehninger and G.D. Greville. The enzymatic oxidation of *d*- and *l*-β-hydroxybutyrate. *Biochim. Biophys. Acta*, 12:188–202, 1953.
- [2407] A.L. Lehninger, H.C. Sudduth, and J.B. Wise. D-β-Hydroxybutyric dehydrogenase of mitochondria. *J. Biol. Chem.*, 235:2450–2455, 1960.
- [2408] I.E. Lehoux and B. Mitra. (*S*)-Mandelate dehydrogenase from *Pseudomonas putida*: mechanistic studies with alternate substrates and pH and kinetic isotope effects. *Biochemistry*, 38:5836–5848, 1999.
- [2409] B. Lei, M. Liu, S. Huang, and S.C. Tu. *Vibrio harveyi* NADPH-flavin oxidoreductase: cloning, sequencing and overexpression of the gene and purification and characterization of the cloned enzyme. *J. Bacteriol.*, 176:3552–3558, 1994.
- [2410] B. Lei and S.C. Tu. Mechanism of reduced flavin transfer from *Vibrio harveyi* NADPH-FMN oxidoreductase to luciferase. *Biochemistry*, 37:14623–14629, 1998.
- [2411] F.-J. Leinweber, R.C. Greenough, C.F. Schwender, H.R. Kaplan, and F.J. DiCarlo. Bunolol metabolism by cell-free preparations of human liver: biosynthesis of dihydrobunolol. *Xenobiotica*, 2:191–202, 1972.
- [2412] H. Leisch, R. Shi, S. Grosse, K. Morley, H. Bergeron, M. Cygler, H. Iwaki, Y. Hasegawa, and P.C. Lau. Cloning, Baeyer-Villiger biooxidations, and structures of the camphor pathway 2-oxo-Δ³-4,5,5-trimethylcyclopentenylacetyl-coenzyme A monooxygenase of *Pseudomonas putida* ATCC 17453. *Appl. Environ. Microbiol.*, 78:2200–2212, 2012.
- [2413] N. Leissing and E.T. McGuinness. Rapid affinity purification and properties of rat liver sorbitol dehydrogenase. *Biochim. Biophys. Acta*, 524:254–261, 1978.
- [2414] R. Lenz and B. Giese. Studies on the Mechanism of Ribonucleotide Reductases. *J. Am. Chem. Soc.*, 119:2784–2794, 1997.

- [2415] R. Lenz and M.H. Zenk. Purification and properties of codeinone reductase (NADPH) from *Papaver somniferum* cell cultures. *Eur. J. Biochem.*, 233:132–139, 1995.
- [2416] R. Lenz and M.H. Zenk. Stereoselective reduction of codeinone, the penultimate step during morphine biosynthesis in *Papaver somniferum*. *Tetrahedron Lett.*, 36:2449–2452, 1995.
- [2417] M.A. Leo, J.M. Lasker, J.L. Rauby, C.I. Kim, M. Black, and C.S. Lieber. Metabolism of retinol and retinoic acid by human liver cytochrome *P*₄₅₀IIC8. *Arch. Biochem. Biophys.*, 269:305–312, 1989.
- [2418] A.E. Leonard, B. Kelder, E.G. Bobik, L.T. Chuang, J.M. Parker-Barnes, J.M. Thurmond, P.E. Kroeger, J.J. Kopchick, Y.S. Huang, and P. Mukerji. cDNA cloning and characterization of human Δ^5 -desaturase involved in the biosynthesis of arachidonic acid. *Biochem. J.*, 347 Pt 3:719–724, 2000.
- [2419] J. Lequeu, M.L. Fauconnier, A. Chammai, R. Bronner, and E. Blee. Formation of plant cuticle: evidence for the occurrence of the peroxygenase pathway. *Plant J.*, 36:155–164, 2003.
- [2420] R.F. Lerud and H.R. Whiteley. Purification and properties of α -ketoglutarate reductase from *Micrococcus aerogenes*. *J. Bacteriol.*, 106:571–577, 1971.
- [2421] A.M. Leseney, D. Deme, O. Legue, R. Ohayon, P. Chanson, J.P. Sales, D. Pires de Carvalho, C. Dupuy, and A. Virion. Biochemical characterization of a Ca^{2+} /NAD(P)H-dependent H_2O_2 generator in human thyroid tissue. *Biochimie*, 81:373–380, 1999.
- [2422] R.K. Lesniak, S. Markolovic, K. Tars, and C.J. Schofield. Human carnitine biosynthesis proceeds via (2*S*,3*S*)-3-hydroxy-*N*^ε-trimethyllysine. *Chem. Commun. (Camb.)*, 53:440–442, 2016.
- [2423] D.J. Lessner, G.R. Johnson, R.E. Parales, J.C. Spain, and D.T. Gibson. Molecular characterization and substrate specificity of nitrobenzene dioxygenase from *Comamonas* sp. strain JS765. *Appl. Environ. Microbiol.*, 68:634–641, 2002.
- [2424] M.G. Leuenberger, C. Engeloch-Jarret, and W.D. Woggon. The reaction mechanism of the enzyme-catalysed central cleavage of β -carotene to retinal. *Angew. Chem.*, 40:2614–2616, 2001.
- [2425] T. Leungsakul, G.R. Johnson, and T.K. Wood. Protein engineering of the 4-methyl-5-nitrocatechol monooxygenase from *Burkholderia* sp. strain DNT for enhanced degradation of nitroaromatics. *Appl. Environ. Microbiol.*, 72:3933–3939, 2006.
- [2426] C. Leutwein and J. Heider. Anaerobic toluene-catabolic pathway in denitrifying *Thauera aromatica*: activation and β -oxidation of the first intermediate, (*R*)-(+)-benzylsuccinate. *Microbiology*, 145:3265–3271, 1999.
- [2427] C. Leutwein and J. Heider. (*R*)-Benzylsuccinyl-CoA dehydrogenase of *Thauera aromatica*, an enzyme of the anaerobic toluene catabolic pathway. *Arch. Microbiol.*, 178:517–524, 2002.
- [2428] J.H. Leveau and S.E. Lindow. Utilization of the plant hormone indole-3-acetic acid for growth by *Pseudomonas putida* strain 1290. *Appl. Environ. Microbiol.*, 71:2365–2371, 2005.
- [2429] E.Y. Levin, B. Levenberg, and S. Kaufman. The enzymatic conversion of 3,4-dihydroxyphenylethylamine to norepinephrine. *J. Biol. Chem.*, 235:2080–2086, 1960.
- [2430] G.M. Levis. 2-Hydroxy fatty acid oxidases of rat kidney. *Biochem. Biophys. Res. Commun.*, 38:470–477, 1970.
- [2431] C.C. Levy. Melilotate hydroxylase. Purification of the enzyme and the nature of the prosthetic group. *J. Biol. Chem.*, 242:747–753, 1967.
- [2432] C.C. Levy and P. Frost. The metabolism of coumarin by a microorganism. V. Melilotate hydroxylase. *J. Biol. Chem.*, 241:997–1003, 1966.
- [2433] C.C. Levy and G.D. Weinstein. The metabolism of coumarin by a microorganism. II. The reduction of *o*-coumaric acid to melilotic acid. *Biochemistry*, 3:1944–1947, 1964.
- [2434] H.R. Levy and P. Talalay. Enzymatic introduction of double bonds into steroid ring A. *J. Am. Chem. Soc.*, 79:2658–2659, 1957.
- [2435] H.R. Levy and P. Talalay. Bacterial oxidation of steroids. II. Studies on the enzymatic mechanisms of ring A dehydrogenation. *J. Biol. Chem.*, 234:2014–2021, 1959.

- [2436] H.R. Levy, V.E. Vought, X. Yin, and M.J. Adams. Identification of an arginine residue in the dual coenzyme-specific glucose-6-phosphate dehydrogenase from *Leuconostoc mesenteroides* that plays a key role in binding NADP⁺ but not NAD⁺. *Arch. Biochem. Biophys.*, 326:145–151, 1996.
- [2437] J. De Ley. 5-Ketogluconic acid reductase. *Methods Enzymol.*, 9:200–203, 1966.
- [2438] J. De Ley and M. Doudoroff. The metabolism of D-galactose in *Pseudomonas saccharophila*. *J. Biol. Chem.*, 227:745–757, 1957.
- [2439] D. Leys, J. Basran, and N.S. Scrutton. Channelling and formation of ‘active’ formaldehyde in dimethylglycine oxidase. *EMBO J.*, 22:4038–4048, 2003.
- [2440] C. Li and C.D. Lu. Arginine racemization by coupled catabolic and anabolic dehydrogenases. *Proc. Natl. Acad. Sci. USA*, 106:906–911, 2009.
- [2441] D.F. Li, J.Y. Zhang, Y.J. Hou, L. Liu, Y. Hu, S.J. Liu, C. Wang da, and W. Liu. Structures of aminophenol dioxygenase in complex with intermediate, product and inhibitor. *Acta Crystallogr. D Biol. Crystallogr.*, 69:32–43, 2013.
- [2442] F. Li, J. Hinderberger, H. Seedorf, J. Zhang, W. Buckel, and R.K. Thauer. Coupled ferredoxin and crotonyl coenzyme A (CoA) reduction with NADH catalyzed by the butyryl-CoA dehydrogenase/Etf complex from *Clostridium kluyveri*. *J. Bacteriol.*, 190:843–850, 2008.
- [2443] H. Li, J. Qiu, F. Chen, X. Lv, C. Fu, D. Zhao, X. Hua, and Q. Zhao. Molecular characterization and expression analysis of dihydroflavonol 4-reductase (DFR) gene in *Saussurea medusa*. *Mol. Biol. Rep.*, 39:2991–2999, 2012.
- [2444] J. Li, M.G. Biswas, A. Chao, D.W. Russell, and J. Chory. Conservation of function between mammalian and plant steroid 5 α -reductases. *Proc. Natl. Acad. Sci. USA*, 94:3554–3559, 1997.
- [2445] J. Li, B.G. Hansen, J.A. Ober, D.J. Kliebenstein, and B.A. Halkier. Subclade of flavin-monoxygenases involved in aliphatic glucosinolate biosynthesis. *Plant Physiol.*, 148:1721–1733, 2008.
- [2446] J. Li, Y. Huang, Y. Hou, X. Li, H. Cao, and Z. Cui. Novel gene clusters and metabolic pathway involved in 3,5,6-trichloro-2-pyridinol degradation by *Ralstonia* sp. strain T6. *Appl. Environ. Microbiol.*, 79:7445–7453, 2013.
- [2447] J. Li, H.J. Liao, Y. Tang, J.L. Huang, L. Cha, T.S. Lin, J.L. Lee, I.V. Kurnikov, M.G. Kurnikova, W.C. Chang, N.L. Chan, and Y. Guo. Epoxidation catalyzed by the nonheme iron(II)- and 2-oxoglutarate-dependent oxygenase, AsqJ: mechanistic elucidation of oxygen atom transfer by a ferryl intermediate. *J. Am. Chem. Soc.*, 142:6268–6284, 2020.
- [2448] L. Li. *Gonyaulax* luciferase: gene structure, protein expression, and purification from recombinant sources. *Methods Enzymol.*, 305:249–258, 2000.
- [2449] L. Li, X. Liu, W. Yang, F. Xu, W. Wang, L. Feng, M. Bartlam, L. Wang, and Z. Rao. Crystal structure of long-chain alkane monooxygenase (LadA) in complex with coenzyme FMN: unveiling the long-chain alkane hydroxylase. *J. Mol. Biol.*, 376:453–465, 2008.
- [2450] L. Li and C.C. Wang. A likely molecular basis of the susceptibility of *Giardia lamblia* towards oxygen. *Mol. Microbiol.*, 59:202–211, 2006.
- [2451] M. Li, T.A. Muller, B.A. Fraser, and R.P. Hausinger. Characterization of active site variants of xanthine hydroxylase from *Aspergillus nidulans*. *Arch. Biochem. Biophys.*, 470:44–53, 2008.
- [2452] N. Li, V.K. Korboukh, C., Bollinger Krebs, , and Jr. Four-electron oxidation of *p*-hydroxylaminobenzoate to *p*-nitrobenzoate by a peroxodiferric complex in AurF from *Streptomyces thioluteus*. *Proc. Natl. Acad. Sci. USA*, 107:15722–15727, 2010.
- [2453] Q. Li, R. Zallot, B.S. MacTavish, A. Montoya, D.J. Payan, Y. Hu, J.A. Gerlt, A. Angerhofer, V. de Crecy-Lagard, and S.D. Bruner. Epoxyqueuosine reductase QueH in the biosynthetic pathway to tRNA queuosine is a unique metalloenzyme. *Biochemistry*, 60:3152–3161, 2021.
- [2454] R. Li, M.A. Bianchet, P. Talalay, and L.M. Amzel. The three-dimensional structure of NAD(P)H:quinone reductase, a flavoprotein involved in cancer chemoprotection and chemotherapy: mechanism of the two-electron reduction. *Proc. Natl. Acad. Sci. USA*, 92:8846–8850, 1995.

- [2455] S. Li, H. Ouellet, D.H. Sherman, and L.M. Podust. Analysis of transient and catalytic desosamine-binding pockets in cytochrome *P*-450 PikC from *Streptomyces venezuelae*. *J. Biol. Chem.*, 284:5723–5730, 2009.
- [2456] S. Li, X. Yu, and G.A. Beattie. Glycine betaine catabolism contributes to *Pseudomonas syringae* tolerance to hyperosmotic stress by relieving betaine-mediated suppression of compatible solute synthesis. *J. Bacteriol.*, 195:2415–2423, 2013.
- [2457] T. Li, C.Y. Chang, D.Y. Jin, P.J. Lin, A. Khvorova, and D.W. Stafford. Identification of the gene for vitamin K epoxide reductase. *Nature*, 427:541–544, 2004.
- [2458] T. Li, L. Simonds, E.L. Kovrigin, and K.D. Noel. *In vitro* biosynthesis and chemical identification of UDP-*N*-acetyl-D-quinovosamine (UDP-D-QuiNAc). *J. Biol. Chem.*, 289:18110–18120, 2014.
- [2459] T.L. Li, O.W. Choroba, E.H. Charles, A.M. Sandercock, D.H. Williams, and J.B. Spencer. Characterisation of a hydroxymandelate oxidase involved in the biosynthesis of two unusual amino acids occurring in the vancomycin group of antibiotics. *Chem. Commun. (Camb.)*, pages 1752–1753, 2001.
- [2460] X. Li, W.T. Beeson, Phillips 4th, Marletta C.M., Cate M.A., and J.H. Structural basis for substrate targeting and catalysis by fungal polysaccharide monooxygenases. *Structure*, 20:1051–1061, 2012.
- [2461] X. Li, X. Xiong, K. Wang, L. Wang, X. Shu, S. Ma, and C. Yi. Transcriptome-wide mapping reveals reversible and dynamic *N*-methyladenosine methylome. *Nat. Chem. Biol.*, 2016.
- [2462] Y. Li, M. Ishida, H. Ashida, T. Ishikawa, H. Shibata, and Y. Sawa. A non-NadB type L-aspartate dehydrogenase from *Ralstonia eutropha* strain JMP134: molecular characterization and physiological functions. *Biosci. Biotechnol. Biochem.*, 75:1524–1532, 2011.
- [2463] Y. Li, N. Kawakami, H.J. Ogola, H. Ashida, T. Ishikawa, H. Shibata, and Y. Sawa. A novel L-aspartate dehydrogenase from the mesophilic bacterium *Pseudomonas aeruginosa* PAO1: molecular characterization and application for L-aspartate production. *Appl. Microbiol. Biotechnol.*, 90:1953–1962, 2011.
- [2464] Y. Li, S. Li, K. Thodey, I. Trenchard, A. Cravens, and C.D. Smolke. Complete biosynthesis of noscapine and halogenated alkaloids in yeast. *Proc. Natl. Acad. Sci. USA*, 115:E3922–E3931, 2018.
- [2465] Y. Li, N.M. Llewellyn, R. Giri, F. Huang, and J.B. Spencer. Biosynthesis of the unique amino acid side chain of butirosin: possible protective-group chemistry in an acyl carrier protein-mediated pathway. *Chem. Biol.*, 12:665–675, 2005.
- [2466] Y. Li, H.J. Ogola, and Y. Sawa. L-aspartate dehydrogenase: features and applications. *Appl. Microbiol. Biotechnol.*, 93:503–516, 2012.
- [2467] Y. Li and C.D. Smolke. Engineering biosynthesis of the anticancer alkaloid noscapine in yeast. *Nat. Commun.*, 7:12137–12137, 2016.
- [2468] Y.M. Li, J.C. Milne, L.L. Madison, R. Kolter, and C.T. Walsh. From peptide precursors to oxazole and thiazole-containing peptide antibiotics: microcin B17 synthase. *Science*, 274:1188–1193, 1996.
- [2469] J. Li-Hawkins, E.G. Lund, A.D. Bronson, and D.W. Russell. Expression cloning of an oxysterol 7 α -hydroxylase selective for 24-hydroxycholesterol. *J. Biol. Chem.*, 275:16543–16549, 2000.
- [2470] J. Liang and R.H. Burris. Hydrogen burst associated with nitrogenase-catalyzed reactions. *Proc. Natl. Acad. Sci. USA*, 85:9446–9450, 1988.
- [2471] X. Liang, C. Thorpe, and H. Schulz. 2,4-Dienoyl-CoA reductase from *Escherichia coli* is a novel iron-sulfur flavoprotein that functions in fatty acid β -oxidation. *Arch. Biochem. Biophys.*, 380:373–379, 2000.
- [2472] H.J. Liao, J. Li, J.L. Huang, M. Davidson, I. Kurnikov, T.S. Lin, J.L. Lee, M. Kurnikova, Y. Guo, N.L. Chan, and W.C. Chang. Insights into the desaturation of cyclopeptin and its C₃ epimer catalyzed by a non-heme iron enzyme: structural characterization and mechanism elucidation. *Angew. Chem. Int. Ed. Engl.*, 57:1831–1835, 2018.
- [2473] S. Liao, J.T. Dulaney, and H.G. Williams-Ashman. Purification and properties of a flavoprotein catalyzing the oxidation of reduced ribosyl nicotinamide. *J. Biol. Chem.*, 237:2981–2987, 1962.

- [2474] M. Libiad, P.K. Yadav, V. Vitvitsky, M. Martinov, and R. Banerjee. Organization of the human mitochondrial hydrogen sulfide oxidation pathway. *J. Biol. Chem.*, 289:30901–30910, 2014.
- [2475] C.A. Libreros-Minotta, J.P. Pardo, G. Mendoza-Hernandez, and J.L. Rendon. Purification and characterization of glutathione reductase from *Rhodospirillum rubrum*. *Arch. Biochem. Biophys.*, 298:247–253, 1992.
- [2476] B.R. Lichman, M.O. Kamileen, G.R. Titchiner, G. Saalbach, C.E.M. Stevenson, D.M. Lawson, and S.E. O'Connor. Uncoupled activation and cyclization in catmint reductive terpenoid biosynthesis. *Nat. Chem. Biol.*, 15:71–79, 2019.
- [2477] B.R. Lichman, S.E. O'Connor, and H. Kries. Biocatalytic strategies towards [4+2] cycloadditions. *Chemistry*, 25:6864–6877, 2019.
- [2478] S.S. Licht, S. Booker, and J. Stubbe. Studies on the catalysis of carbon-cobalt bond homolysis by ribonucleoside triphosphate reductase: evidence for concerted carbon-cobalt bond homolysis and thiyl radical formation. *Biochemistry*, 38:1221–1233, 1999.
- [2479] I. Lidbury, M.A. Mausz, D.J. Scanlan, and Y. Chen. Identification of dimethylamine monooxygenase in marine bacteria reveals a metabolic bottleneck in the methylated amine degradation pathway. *ISME J.*, 11:1592–1601, 2017.
- [2480] M. Liden, A. Romert, K. Tryggvason, B. Persson, and U. Eriksson. Biochemical defects in 11-*cis*-retinol dehydrogenase mutants associated with fundus albipunctatus. *J. Biol. Chem.*, 276:49251–49257, 2001.
- [2481] I. Lieberman and A. Kornberg. Enzymic synthesis and breakdown of a pyrimidine, orotic acid. I. Dihydro-orotic dehydrogenase. *Biochim. Biophys. Acta*, 12:223–234, 1953.
- [2482] J. Liepins, S. Kuorelahti, M. Penttila, and P. Richard. Enzymes for the NADPH-dependent reduction of dihydroxyacetone and D-glyceraldehyde and L-glyceraldehyde in the mould *Hypocrea jecorina*. *FEBS J.*, 273:4229–4235, 2006.
- [2483] C. Liers, C. Bobeth, M. Pecyna, R. Ullrich, and M. Hofrichter. DyP-like peroxidases of the jelly fungus *Auricularia auricula-judae* oxidize nonphenolic lignin model compounds and high-redox potential dyes. *Appl. Microbiol. Biotechnol.*, 85:1869–1879, 2010.
- [2484] A. Lieutaud, R. van Lis, S. Duval, L. Capowiez, D. Muller, R. Lebrun, S. Lignon, M.L. Fardeau, M.C. Lett, W. Nitschke, and B. Schoepp-Cothenet. Arsenite oxidase from *Ralstonia* sp. 22: characterization of the enzyme and its interaction with soluble cytochromes. *J. Biol. Chem.*, 285:20433–20441, 2010.
- [2485] J.G. Lim, Y.J. Bang, and S.H. Choi. Characterization of the *Vibrio vulnificus* 1-Cys peroxiredoxin Prx3 and regulation of its expression by the Fe-S cluster regulator IscR in response to oxidative stress and iron starvation. *J. Biol. Chem.*, 289:36263–36274, 2014.
- [2486] C.H. Lin, B. Li, S. Swanson, Y. Zhang, L. Florens, M.P. Washburn, S.M. Abmayr, and J.L. Workman. Heterochromatin protein 1a stimulates histone H3 lysine 36 demethylation by the *Drosophila* KDM4A demethylase. *Mol. Cell*, 32:696–706, 2008.
- [2487] E.C.C. Lin. An inducible D-arabitol dehydrogenase from *Aerobacter aerogenes*. *J. Biol. Chem.*, 236:31–36, 1961.
- [2488] E.C.C. Lin and B. Magasanik. The activation of glycerol dehydrogenase from *Aerobacter aerogenes* by monovalent cations. *J. Biol. Chem.*, 235:1820–1823, 1960.
- [2489] H.C. Lin, Y. Tsunematsu, S. Dhingra, W. Xu, M. Fukutomi, Y.H. Chooi, D.E. Cane, A.M. Calvo, K. Watanabe, and Y. Tang. Generation of complexity in fungal terpene biosynthesis: discovery of a multifunctional cytochrome P450 in the fumagillin pathway. *J. Am. Chem. Soc.*, 136:4426–4436, 2014.
- [2490] J.T. Lin, T.A. McKeon, M. Goodrich-Tanrikulu, and A.E. Stafford. Characterization of oleoyl-12-hydroxylase in castor microsomes using the putative substrate, 1-acyl-2-oleoyl-*sn*-glycero-3-phosphocholine. *Lipids*, 31:571–577, 1996.
- [2491] J.W. Lin, Y.F. Chao, and S.F. Weng. Nucleotide sequence of the *luxC* gene encoding fatty acid reductase of the *lux* operon from *Photobacterium leiognathi*. *Biochem. Biophys. Res. Commun.*, 191:314–318, 1993.
- [2492] M.C.M. Lin and C. Wagner. Purification and characterization of *N*-methylalanine dehydrogenase. *J. Biol. Chem.*, 250:3746–3751, 1975.

- [2493] S. Lin, S.G. Van Lanen, and B. Shen. Characterization of the two-component, FAD-dependent monooxygenase SgcC that requires carrier protein-tethered substrates for the biosynthesis of the enediyne antitumor antibiotic C-1027. *J. Am. Chem. Soc.*, 130:6616–6623, 2008.
- [2494] Y.M. Lin and J. Jarabak. Isolation of two proteins with 9-ketoprostaglandin reductase and NADP-linked 15-hydroxyprostaglandin dehydrogenase activities and studies on their inhibition. *Biochem. Biophys. Res. Commun.*, 81:1227–1234, 1978.
- [2495] K.E. Lind. Dihydropteridine reductase. Investigation of the specificity for quinoid dihydropteridine and the inhibition by 2,4-diaminopteridines. *Eur. J. Biochem.*, 25:560–562, 1972.
- [2496] P. Lindemann and M. Luckner. Biosynthesis of pregnane derivatives in somatic embryos of *Digitalis lanata*. *Phytochemistry*, 46:507–513, 1997.
- [2497] H. Linden. Carotenoid hydroxylase from *Haematococcus pluvialis*: cDNA sequence, regulation and functional complementation. *Biochim. Biophys. Acta*, 1446:203–212, 1999.
- [2498] R. Lindigkeit, A. Biller, M. Buch, H.M. Schiebel, M. Boppre, and T. Hartmann. The two facies of pyrrolizidine alkaloids: the role of the tertiary amine and its *N*-oxide in chemical defense of insects with acquired plant alkaloids. *Eur. J. Biochem.*, 245:626–636, 1997.
- [2499] P.F. Card Lindley, I. Zaitsev G. Zaitseva, B. Selin Lindgren V. Reinhammar, Yoshida E., and K. An X-ray structural study of human ceruloplasmin in relation to ferroxidase activity. *J. Biol. Inorg. Chem.*, 2:454–463, 1997.
- [2500] A. Lindqvist, Y.G. He, and S. Andersson. Cell type-specific expression of β -carotene 9',10'-monooxygenase in human tissues. *J. Histochem. Cytochem.*, 53:1403–1412, 2005.
- [2501] G. Lindstedt and S. Lindstedt. Cofactor requirements of γ -butyrobetaine hydroxylase from rat liver. *J. Biol. Chem.*, 245:4178–4186, 1970.
- [2502] S. Lindstedt and M. Rundgren. Blue color, metal content, and substrate binding in 4-hydroxyphenylpyruvate dioxygenase from *Pseudomonas* sp. strain P. J. 874. *J. Biol. Chem.*, 257:11922–11931, 1982.
- [2503] F. Lingens, E. Keller, and B. Keller. Arogenate dehydrogenase from *Phenylobacterium immobile*. *Methods Enzymol.*, 142:513–518, 1987.
- [2504] T. Link, G. Lohaus, I. Heiser, K. Mendgen, M. Hahn, and R.T. Voegele. Characterization of a novel NADP⁺-dependent D-arabitol dehydrogenase from the plant pathogen *Uromyces fabae*. *Biochem. J.*, 389:289–295, 2005.
- [2505] J.D. Lipscomb and A.B. Hooper. Resolution of multiple heme centers of hydroxylamine oxidoreductase from *Nitrosomonas*. 1. Electron paramagnetic resonance spectroscopy. *Biochemistry*, 21:3965–3972, 1982.
- [2506] M. Liss, S.B. Horwitz, and N.O. Kaplan. D-Mannitol 1-phosphate dehydrogenase and D-sorbitol 6-phosphate dehydrogenase in *Aerobacter aerogenes*. *J. Biol. Chem.*, 237:1342–1350, 1962.
- [2507] M. Lisurek, M.J. Kang, R.W. Hartmann, and R. Bernhardt. Identification of monohydroxy progesterones produced by CYP106A2 using comparative HPLC and electrospray ionisation collision-induced dissociation mass spectrometry. *Biochem. Biophys. Res. Commun.*, 319:677–682, 2004.
- [2508] M. Lisurek, B. Simgen, I. Antes, and R. Bernhardt. Theoretical and experimental evaluation of a CYP106A2 low homology model and production of mutants with changed activity and selectivity of hydroxylation. *ChemBioChem*, 9:1439–1449, 2008.
- [2509] H.N. Little. Oxidation of nitroethane by extracts from *Neurospora*. *J. Biol. Chem.*, 193:347–358, 1951.
- [2510] T.K. Littlejohn, O. Takikawa, R.J. Truscott, and M.J. Walker. Asp²⁷⁴ and His³⁴⁶ are essential for heme binding and catalytic function of human indoleamine 2,3-dioxygenase. *J. Biol. Chem.*, 278:29525–29531, 2003.
- [2511] C.-K. Liu, C.-A. Hsu, and M.T. Abbott. Catalysis of three sequential dioxygenase reactions by thymine 7-hydroxylase. *Arch. Biochem. Biophys.*, 159:180–187, 1973.
- [2512] C.J. Liu, D. Huhman, L.W. Sumner, and R.A. Dixon. Regiospecific hydroxylation of isoflavones by cytochrome *p*₄₅₀ 81E enzymes from *Medicago truncatula*. *Plant J.*, 36:471–484, 2003.

- [2513] H. Liu, S.J. Wang, J.J. Zhang, H. Dai, H. Tang, and N.Y. Zhou. Patchwork assembly of nag-like nitroarene dioxygenase genes and the 3-chlorocatechol degradation cluster for evolution of the 2-chloronitrobenzene catabolism pathway in *Pseudomonas stutzeri* ZWLR2-1. *Appl. Environ. Microbiol.*, 77:4547–4552, 2011.
- [2514] H. Liu, Y. Xin, and L. Xun. Distribution, diversity, and activities of sulfur dioxygenases in heterotrophic bacteria. *Appl. Environ. Microbiol.*, 80:1799–1806, 2014.
- [2515] H.-W. Liu and J.S. Thorson. Pathways and mechanisms in the biogenesis of novel deoxysugars by bacteria. *Annu. Rev. Microbiol.*, 48:223–256, 1994.
- [2516] J. Liu, Y. Wei, K. Ma, J. An, X. Liu, Y. Liu, E.L. Ang, H. Zhao, and Y. Zhang. Mechanistically diverse pathways for sulfoquinovose degradation in bacteria. *ACS Catal.*, 11:14740–14750, 2021.
- [2517] L. Liu, A. Hausladen, M. Zeng, L. Que, J. Heitman, and J.S. Stamler. A metabolic enzyme for S-nitrosothiol conserved from bacteria to humans. *Nature*, 410:490–494, 2001.
- [2518] L. Liu, H. Kim, A. Casta, Y. Kobayashi, L.S. Shapiro, and A.M. Christiano. Hairless is a histone H3K9 demethylase. *FASEB J.*, 28:1534–1542, 2014.
- [2519] L.K. Liu, Y. Dai, H. Abdelwahab, P. Sobrado, and J.J. Tanner. Structural evidence for rifampicin monooxygenase inactivating rifampicin by cleaving its ansa-bridge. *Biochemistry*, 57:2065–2068, 2018.
- [2520] M. Liu, B. Lei, Q. Ding, J.C. Lee, and S.C. Tu. *Vibrio harveyi* NADPH:FMN oxidoreductase: preparation and characterization of the apoenzyme and monomer-dimer equilibrium. *Arch. Biochem. Biophys.*, 337:89–95, 1997.
- [2521] P. Liu, M.P. Mehn, F. Yan, Z. Zhao, L. Que, Liu Jr., and H.W. Oxygenase activity in the self-hydroxylation of (S)-2-hydroxypropylphosphonic acid epoxidase involved in fosfomycin biosynthesis. *J. Am. Chem. Soc.*, 126:10306–10312, 2004.
- [2522] Q. Liu, D. Manzano, N. Tanic, M. Pesic, J. Bankovic, I. Pateraki, L. Ricard, A. Ferrer, R. de Vos, S. van de Krol, and H. Bouwmeester. Elucidation and in planta reconstitution of the parthenolide biosynthetic pathway. *Metab. Eng.*, 23:145–153, 2014.
- [2523] S. Liu, C. Zhang, T. Su, T. Wei, D. Zhu, K. Wang, Y. Huang, Y. Dong, K. Yin, S. Xu, P. Xu, and L. Gu. Crystal structure of DszC from *Rhodococcus* sp. XP at 1.79 Å. *Proteins*, 82:1708–1720, 2014.
- [2524] W. Liu, C.E. Rogge, B. Bambai, G. Palmer, A.L. Tsai, and R.J. Kulmacz. Characterization of the heme environment in *Arabidopsis thaliana* fatty acid α -dioxygenase-1. *J. Biol. Chem.*, 279:29805–29815, 2004.
- [2525] X. Liu, Y. Dong, X. Li, Y. Ren, Y. Li, W. Wang, L. Wang, and L. Feng. Characterization of the anthranilate degradation pathway in *Geobacillus thermodenitrificans* NG80-2. *Microbiology*, 156:589–595, 2010.
- [2526] Y.W. Liu, K. Denkmann, K. Kosciow, C. Dahl, and D.J. Kelly. Tetrathionate stimulated growth of *Campylobacter jejuni* identifies a new type of bi-functional tetrathionate reductase (TsdA) that is widely distributed in bacteria. *Mol. Microbiol.*, 88:173–188, 2013.
- [2527] J.R. Livingstone, T. Maruo, I. Yoshida, Y. Tarui, K. Hirooka, Y. Yamamoto, N. Tsutui, and E. Hirasawa. Purification and properties of betaine aldehyde dehydrogenase from *Avena sativa*. *J. Plant Res.*, 116:133–140, 2003.
- [2528] C.T. Lloyd, D.F. Iwig, B. Wang, M. Cossu, W.W. Metcalf, A.K. Boal, and S.J. Booker. Discovery, structure, and mechanism of a tetraether lipid synthase. *Nature*, 2022.
- [2529] M.D. Lloyd, S.J. Lipscomb, K.S. Hewitson, C.M. Hensgens, J.E. Baldwin, and C.J. Schofield. Controlling the substrate selectivity of deacetoxycephalosporin/deacetylcephalosporin C synthase. *J. Biol. Chem.*, 279:15420–15426, 2004.
- [2530] H.H. Locher, T. Leisinger, and A.M. Cook. 4-Sulphobenzoate 3,4-dioxygenase. Purification and properties of a desulphonative two-component enzyme system from *Comamonas testosteroni* T-2. *Biochem. J.*, 274:833–842, 1991.
- [2531] A.M. Loening, T.D. Fenn, and S.S. Gambhir. Crystal structures of the luciferase and green fluorescent protein from *Renilla reniformis*. *J. Mol. Biol.*, 374:1017–1028, 2007.
- [2532] P.C. Loewen, B.L. Triggs, C.S. George, and B.E. Hrabarchuk. Genetic mapping of *katG*, a locus that affects synthesis of the bifunctional catalase-peroxidase hydroperoxidase I in *Escherichia coli*. *J. Bacteriol.*, 162:661–667, 1985.

- [2533] Y.H. Loh, W. Zhang, X. Chen, J. George, and H.H. Ng. Jmjd1a and Jmjd2c histone H3 Lys 9 demethylases regulate self-renewal in embryonic stem cells. *Genes Dev.*, 21:2545–2557, 2007.
- [2534] A. Lomascolo, E. Dubreucq, and P. Galzy. Study of the Δ^{12} -desaturase system of *Lipomyces starkeyi*. *Lipids*, 31:253–259, 1996.
- [2535] M. Lombard, M. Fontecave, D. Touati, and V. Niviere. Reaction of the desulfoferrodoxin from *Desulfoarculus baarsii* with superoxide anion. Evidence for a superoxide reductase activity. *J. Biol. Chem.*, 275:115–121, 2000.
- [2536] J.B. Lombardini, T.P. Singer, and P.D. Boyer. Cystein oxygenase. II. Studies on the mechanism of the reaction with ^{18}O oxygen. *J. Biol. Chem.*, 244:1172–1175, 1969.
- [2537] M. London and P.B. Hudson. Purification and properties of solubilized uricase. *Biochim. Biophys. Acta*, 21:290–298, 1956.
- [2538] J.C. Loper. Histidinol dehydrogenase from *Salmonella typhimurium*. Crystallization and composition studies. *J. Biol. Chem.*, 243:3264–3272, 1968.
- [2539] J.M. Lord. Glycolate oxidoreductase in *Escherichia coli*. *Biochim. Biophys. Acta*, 267:227–237, 1972.
- [2540] E. Lorentzen, R. Hensel, T. Knura, H. Ahmed, and E. Pohl. Structural basis of allosteric regulation and substrate specificity of the non-phosphorylating glyceraldehyde 3-phosphate dehydrogenase from *Thermoproteus tenax*. *J. Mol. Biol.*, 341:815–828, 2004.
- [2541] W.W. Lorenz, R.O. McCann, M. Longiaru, and M.J. Cormier. Isolation and expression of a cDNA encoding *Renilla reniformis* luciferase. *Proc. Natl. Acad. Sci. USA*, 88:4438–4442, 1991.
- [2542] T. Lotan and J. Hirschberg. Cloning and expression in *Escherichia coli* of the gene encoding β -C-4-oxygenase, that converts β -carotene to the ketocarotenoid canthaxanthin in *Haematococcus pluvialis*. *FEBS Lett.*, 364:125–128, 1995.
- [2543] G.V. Louie, T.J. Baiga, M.E. Bowman, T. Koeduka, J.H. Taylor, S.M. Spassova, E. Pichersky, and J.P. Noel. Structure and reaction mechanism of basil eugenol synthase. *PLoS One*, 2:e993–, 2007.
- [2544] T.M. Louie, C.M. Webster, and L. Xun. Genetic and biochemical characterization of a 2,4,6-trichlorophenol degradation pathway in *Ralstonia eutropha* JMP134. *J. Bacteriol.*, 184:3492–3500, 2002.
- [2545] T.M. Louie, X.S. Xie, and L. Xun. Coordinated production and utilization of FADH₂ by NAD(P)H-flavin oxidoreductase and 4-hydroxyphenylacetate 3-monooxygenase. *Biochemistry*, 42:7509–7517, 2003.
- [2546] N. Lovallo and D.L. Cox-Foster. Alteration in FAD-glucose dehydrogenase activity and hemocyte behavior contribute to initial disruption of *Manduca sexta* immune response to *Cotesia congregata* parasitoids. *J. Insect Physiol.*, 45:1037–1048, 1999.
- [2547] H. Löw, I.L. Sun, P. Navas, C. Grebing, F.L. Crane, and D.J. Morré. Transplasmalemma electron transport from cells is part of a diferric transferrin reductase system. *Biochem. Biophys. Res. Commun.*, 139:1117–1123, 1986.
- [2548] A.Y.H. Lu, K.W. Junk, and M.J. Coon. Resolution of the cytochrome P-450-containing ω -hydroxylation system of liver microsomes into three components. *J. Biol. Chem.*, 244:3714–3721, 1969.
- [2549] A.Y.H. Lu, S.W. Kuntzman, M. Jacobson, and A.H. Conney. Reconstituted liver microsomal enzyme system that hydroxylates drugs, other foreign compounds, and endogenous substrates. II. Role of the cytochrome P-450 and P-448 fractions in drug and steroid hydroxylations. *J. Biol. Chem.*, 247:1727–1734, 1972.
- [2550] H. Lu, E. Chanco, and H. Zhao. CmlI is an N-oxygenase in the biosynthesis of chloramphenicol. *Tetrahedron*, 68:7651–7654, 2012.
- [2551] W.-P. Lu and D.P. Kelly. Properties and role of sulphite:cytochrome c oxidoreductase purified from *Thiobacillus versutus* (A2). *J. Gen. Microbiol.*, 130:1683–1692, 1984.
- [2552] W.-P. Lu and D.P. Kelly. Cellular location and partial purification of the ‘thiosulphate-oxidizing enzyme’ and ‘trithionate hydrolase’ from *Thiobacillus tepidarius*. *J. Gen. Microbiol.*, 134:877–885, 1988.
- [2553] J. Luba, V. Charrier, and A. Claiborne. Coenzyme A-disulfide reductase from *Staphylococcus aureus*: evidence for asymmetric behavior on interaction with pyridine nucleotides. *Biochemistry*, 38:2725–2737, 1999.

- [2554] C.E. Lubner, D.P. Jennings, D.W. Mulder, G.J. Schut, O.A. Zadovnyy, J.P. Hoben, M. Tokmina-Lukaszewska, L. Berry, D.M. Nguyen, G.L. Lipscomb, B. Bothner, A.K. Jones, A.F. Miller, P.W. King, M.W.W. Adams, and J.W. Peters. Mechanistic insights into energy conservation by flavin-based electron bifurcation. *Nat. Chem. Biol.*, 13:655–659, 2017.
- [2555] P. Lucas-Elío, D. Gómez, F. Solano, and A. Sanchez-Amat. The antimicrobial activity of marinocine, synthesized by *Marinomonas mediterranea*, is due to hydrogen peroxide generated by its lysine oxidase activity. *J. Bacteriol.*, 188:2493–2501, 2006.
- [2556] F. Lüddecke, A. Wülfig, M. Timke, F. Germer, J. Weber, A. Dikfidan, T. Rahnfeld, D. Linder, A. Meyerdierks, and J. Harder. Geraniol and geranial dehydrogenases induced in anaerobic monoterpene degradation by *Castellaniella defragrans*. *Appl. Environ. Microbiol.*, 78:2128–2136, 2012.
- [2557] Z. Luka, S. Pakhomova, L.V. Loukachevitch, M.E. Newcomer, and C. Wagner. Folate in demethylation: the crystal structure of the rat dimethylglycine dehydrogenase complexed with tetrahydrofolate. *Biochem. Biophys. Res. Commun.*, 449:392–398, 2014.
- [2558] E.V. Lukasheva and T.T. Berezov. L-Lysine α -oxidase: physicochemical and biological properties. *Biochemistry (Mosc.)*, 67:1152–1158, 2002.
- [2559] R. Lukačič, U. Matern, K.T. Junghanns, M.L. Heskamp, L. Britsch, G. Forkmann, and S. Martens. Purification and antigenicity of flavone synthase I from irradiated parsley cells. *Arch. Biochem. Biophys.*, 393:177–183, 2001.
- [2560] R. Lukačič, F. Wellmann, L. Britsch, S. Martens, and U. Matern. Flavonol synthase from *Citrus unshiu* is a bifunctional dioxygenase. *Phytochemistry*, 62:287–292, 2003.
- [2561] Y. Lukyanenko, J.J. Chen, and J.C. Hutson. Testosterone regulates 25-hydroxycholesterol production in testicular macrophages. *Biol. Reprod.*, 67:1435–1438, 2002.
- [2562] E.G. Lund, J.M. Guileyardo, and D.W. Russell. cDNA cloning of cholesterol 24-hydroxylase, a mediator of cholesterol homeostasis in the brain. *Proc. Natl. Acad. Sci. USA*, 96:7238–7243, 1999.
- [2563] E.G. Lund, T.A. Kerr, J. Sakai, W.P. Li, and D.W. Russell. cDNA cloning of mouse and human cholesterol 25-hydroxylases, polytopic membrane proteins that synthesize a potent oxysterol regulator of lipid metabolism. *J. Biol. Chem.*, 273:34316–34327, 1998.
- [2564] E.G. Lund, C. Xie, T. Kotti, S.D. Turley, J.M. Dietschy, and D.W. Russell. Knockout of the cholesterol 24-hydroxylase gene in mice reveals a brain-specific mechanism of cholesterol turnover. *J. Biol. Chem.*, 278:22980–22988, 2003.
- [2565] K. Lundell, R. Hansson, and K. Wikvall. Cloning and expression of a pig liver taurochenodeoxycholic acid 6 α -hydroxylase (CYP4A21): a novel member of the CYP4A subfamily. *J. Biol. Chem.*, 276:9606–9612, 2001.
- [2566] K. Lundell and K. Wikvall. Gene structure of pig sterol 12 α -hydroxylase (CYP8B1) and expression in fetal liver: comparison with expression of taurochenodeoxycholic acid 6 α -hydroxylase (CYP4A21). *Biochim. Biophys. Acta*, 1634:86–96, 2003.
- [2567] C.A.K. Lundgren, D. Sjostrand, O. Biner, M. Bennett, A. Rudling, A.L. Johansson, P. Brzezinski, J. Carlsson, C. von Ballmoos, and M. Hogbom. Scavenging of superoxide by a membrane-bound superoxide oxidase. *Nat. Chem. Biol.*, 14:788–793, 2018.
- [2568] B. Lupa, E.L. Hendrickson, J.A. Leigh, and W.B. Whitman. Formate-dependent H₂ production by the mesophilic methanogen *Methanococcus maripaludis*. *Appl. Environ. Microbiol.*, 74:6584–6590, 2008.
- [2569] S. Lupien, F. Karp, M. Wildung, and R. Croteau. Regiospecific cytochrome P450 limonene hydroxylases from mint (*Mentha*) species: cDNA isolation, characterization, and functional expression of (–)-4S-limonene-3-hydroxylase and (–)-4S-limonene-6-hydroxylase. *Arch. Biochem. Biophys.*, 368:181–192, 1999.
- [2570] G.A. Lyles. Mammalian plasma and tissue-bound semicarbazide-sensitive amine oxidases: biochemical, pharmacological and toxicological aspects. *Int. J. Biochem. Cell Biol.*, 28:259–274, 1996.
- [2571] W.S. Lynn and R.H. Brown. The conversion of progesterone to androgens by testes. *J. Biol. Chem.*, 232:1015–1030, 1958.

- [2572] R.M. Lyric and I. Suzuki. Enzymes involved in the metabolism of thiosulfate by *Thiobacillus thioparus*. I. Survey of enzymes and properties of sulfite: cytochrome *c* oxidoreductase. *Can. J. Biochem.*, 48:334–343, 1970.
- [2573] K. Ma and M.W. Adams. Sulfide dehydrogenase from the hyperthermophilic archaeon *Pyrococcus furiosus*: a new multifunctional enzyme involved in the reduction of elemental sulfur. *J. Bacteriol.*, 176:6509–6517, 1994.
- [2574] K. Ma and M.W.W. Adams. A hyperactive NAD(P)H:rubredoxin oxidoreductase from the hyperthermophilic archaeon *Pyrococcus furiosus*. *J. Bacteriol.*, 181:5530–5533, 1999.
- [2575] K. Ma, R.N. Schicho, R.M. Kelly, and M.W. Adams. Hydrogenase of the hyperthermophile *Pyrococcus furiosus* is an elemental sulfur reductase or sulfhydrogenase: evidence for a sulfur-reducing hydrogenase ancestor. *Proc. Natl. Acad. Sci. USA*, 90:5341–5344, 1993.
- [2576] K. Ma and R.K. Thauer. Purification and properties of N^5,N^{10} -methylenetetrahydromethanopterin reductase from *Methanobacterium thermoautotrophicum* (strain Marburg). *Eur. J. Biochem.*, 191:187–193, 1990.
- [2577] K. Ma and R.K. Thauer. Single step purification of methylenetetrahydromethanopterin reductase from *Methanobacterium thermoautotrophicum* by specific binding to blue sepharose CL-6B. *FEBS Lett.*, 268:59–62, 1990.
- [2578] K. Ma, R. Weiss, and M.W. Adams. Characterization of hydrogenase II from the hyperthermophilic archaeon *Pyrococcus furiosus* and assessment of its role in sulfur reduction. *J. Bacteriol.*, 182:1864–1871, 2000.
- [2579] K. Ma, Z.H. Zhou, and M.W. Adams. Hydrogen production from pyruvate by enzymes purified from the hyperthermophilic archaeon, *Pyrococcus furiosus*: A key role for NADPH. *FEMS Microbiol. Lett.*, 122:245–250, 1994.
- [2580] L. Ma, C. Huang, X.J. Wang, D.E. Xin, L.S. Wang, Q.C. Zou, Y.S. Zhang, M.D. Tan, Y.M. Wang, T.C. Zhao, D. Chatterjee, R.A. Altura, C. Wang, Y.S. Xu, J.H. Yang, Y.S. Fan, B.H. Han, J. Si, X. Zhang, J. Cheng, Z. Chang, and Y.E. Chin. Lysyl oxidase 3 is a dual-specificity enzyme involved in STAT3 deacetylation and deacetylimination modulation. *Mol. Cell*, 65:296–309, 2017.
- [2581] W.J. Maalcke, A. Dietl, S.J. Marritt, J.N. Butt, M.S. Jetten, J.T. Keltjens, T.R. Barends, and B. Kartal. Structural basis of biological NO generation by octaheme oxidoreductases. *J. Biol. Chem.*, 289:1228–1242, 2014.
- [2582] I.A. Macdonald, D.E. Mahony, J.F. Jellett, and C.E. Meier. NAD-dependent 3 α - and 12 α -hydroxysteroid dehydrogenase activities from *Eubacterium lentum* ATCC no. 25559. *Biochim. Biophys. Acta*, 489:466–476, 1977.
- [2583] I.A. Macdonald and P.D. Roach. Bile induction of 7 α - and 7 β -hydroxysteroid dehydrogenases in *Clostridium absonum*. *Biochim. Biophys. Acta*, 665:262–269, 1981.
- [2584] I.A. Macdonald, Y.P. Rochon, D.M. Hutchison, and L.V. Holdeman. Formation of ursodeoxycholic acid from chenodeoxycholic acid by a 7 β -hydroxysteroid dehydrogenase-elaborating *Eubacterium aerofaciens* strain cocultured with 7 α -hydroxysteroid dehydrogenase-elaborating organisms. *Appl. Environ. Microbiol.*, 44:1187–1195, 1982.
- [2585] I.A. Macdonald, C.N. Williams, and D.E. Mahony. 7 α -Hydroxysteroid dehydrogenase from *Escherichia coli* B: preliminary studies. *Biochim. Biophys. Acta*, 309:243–253, 1973.
- [2586] I.A. Macdonald, C.N. Williams, D.E. Mahony, and W.M. Christie. NAD- and NADP-dependent 7 α -hydroxysteroid dehydrogenases from *Bacteroides fragilis*. *Biochim. Biophys. Acta*, 384:12–24, 1975.
- [2587] I.K. Macdonald, S.K. Badyal, L. Ghamsari, P.C. Moody, and E.L. Raven. Interaction of ascorbate peroxidase with substrates: a mechanistic and structural analysis. *Biochemistry*, 45:7808–7817, 2006.
- [2588] J.C. MacDonald. Biosynthesis of pulcherriminic acid. *Biochem. J.*, 96:533–538, 1965.
- [2589] M.J. MacDonald and L.J. Brown. Calcium activation of mitochondrial glycerol phosphate dehydrogenase restudied. *Arch. Biochem. Biophys.*, 326:79–84, 1996.
- [2590] I.J. MacFarlane, E.M. Lees, and E.E. Conn. The in vitro biosynthesis of dhurrin, the cyanogenic glycoside of *Sorghum bicolor*. *J. Biol. Chem.*, 250:4708–4713, 1975.
- [2591] P. Macheroux, H.J. Plattner, A. Romaguera, and H. Diekmann. FAD and substrate analogs as probes for lysine N^6 -hydroxylase from *Escherichia coli* EN 222. *Eur. J. Biochem.*, 213:995–1002, 1993.

- [2592] Y. Machida and T. Nakanishi. Purification and properties of pyranose oxidase from *Coriolus versicolor*. *Agric. Biol. Chem.*, 48:2463–2470, 1984.
- [2593] J.J. MacKenzie and L.B. Sorensen. Guanosine 5'-phosphate reductase of human erythrocytes. *Biochim. Biophys. Acta*, 327:282–294, 1973.
- [2594] J. MacLachlan, A.T. Wotherspoon, R.O. Ansell, and C.J. Brooks. Cholesterol oxidase: sources, physical properties and analytical applications. *J. Steroid Biochem. Mol. Biol.*, 72:169–195, 2000.
- [2595] R.M. MacLeod, W. Farkas, I. Fridovitch, and P. Handler. Purification and properties of hepatic sulfite oxidase. *J. Biol. Chem.*, 236:1841–1846, 1961.
- [2596] A.R. Macrae. Isolation and properties of a 'malic' enzyme from cauliflower bud mitochondria. *Biochem. J.*, 122:495–501, 1971.
- [2597] J.M. Macy, S. Rech, G. Auling, M. Dorsch, E. Stackebrandt, and L.I. Sly. *Thauera selenatis* gen. nov., sp. nov., a member of the β subclass of Proteobacteria with a novel type of anaerobic respiration. *Int. J. Syst. Bacteriol.*, 43:135–142, 1993.
- [2598] V.K. Madan, P. Hillmer, and G. Gottschalk. Purification and properties of NADP-dependent L(+)-3-hydroxybutyryl-CoA dehydrogenase from *Clostridium kluyveri*. *Eur. J. Biochem.*, 32:51–56, 1973.
- [2599] S.L. Mader, A. Brauer, M. Groll, and V.R.I. Kaila. Catalytic mechanism and molecular engineering of quinolone biosynthesis in dioxygenase AsqJ. *Nat. Commun.*, 9:1168–1168, 2018.
- [2600] D. Madern. The putative L-lactate dehydrogenase from *Methanococcus jannaschii* is an NADPH-dependent L-malate dehydrogenase. *Mol. Microbiol.*, 37:1515–1520, 2000.
- [2601] K. Madhavan, M. Conscience-Egli, F. Sieber, and H. Ursprung. Farnesol metabolism in *Drosophila melanogaster*: ontogeny and tissue distribution of octanol dehydrogenase and aldehyde oxidase. *J. Insect Physiol.*, 19:235–241, 1973.
- [2602] K.M. Madyastha and V. Srivatsan. Studies on the metabolism of *l*-menthol in rats. *Drug Metab. Dispos.*, 16:765–772, 1988.
- [2603] K. Maeda-Yorita, K. Aki, H. Sagai, H. Misaki, and V. Massey. L-Lactate oxidase and L-lactate monooxygenase: mechanistic variations on a common structural theme. *Biochimie*, 77:631–642, 1995.
- [2604] S. Mafu, Y. Ding, K.M. Murphy, O. Yaacoobi, J.B. Addison, Q. Wang, Z. Shen, S.P. Briggs, J. Bohlmann, G. Castro-Falcon, C.C. Hughes, M. Betsiashvili, A. Huffaker, E.A. Schmelz, and P. Zerbe. Discovery, biosynthesis and stress-related accumulation of dolabradiene-derived defenses in maize. *Plant Physiol.*, 176:2677–2690, 2018.
- [2605] N.A. Magarvey, B. Haltli, M. He, M. Greenstein, and J.A. Hucul. Biosynthetic pathway for mannopeptimycins, lipoglycopeptide antibiotics active against drug-resistant gram-positive pathogens. *Antimicrob. Agents Chemother.*, 50:2167–2177, 2006.
- [2606] B. Magasanik, H.S. Moyed, and L.B. Gehring. Enzymes essential for the biosynthesis of nucleic acid guanine; inosine 5'-phosphate dehydrogenase of *Aerobacter aerogenes*. *J. Biol. Chem.*, 226:339–350, 1957.
- [2607] P.T. Magee and E.E. Snell. The bacterial degradation of pantothenic acid. IV. Enzymatic conversion of aldopantoate to α -ketoisovalerate. *Biochemistry*, 5:409–416, 1966.
- [2608] J. Mager and B. Magasanik. Guanosine 5'-phosphate reductase and its role in the interconversion of purine nucleotides. *J. Biol. Chem.*, 235:1474–1478, 1960.
- [2609] O.T. Magnusson, H. Toyama, M. Saeki, A. Rojas, J.C. Reed, R.C. Liddington, J.P. Klinman, and R. Schwarzenbacher. Quinone biogenesis: Structure and mechanism of PqqC, the final catalyst in the production of pyrroloquinoline quinone. *Proc. Natl. Acad. Sci. USA*, 101:7913–7918, 2004.
- [2610] O.T. Magnusson, H. Toyama, M. Saeki, R. Schwarzenbacher, and J.P. Klinman. The structure of a biosynthetic intermediate of pyrroloquinoline quinone (PQQ) and elucidation of the final step of PQQ biosynthesis. *J. Am. Chem. Soc.*, 126:5342–5343, 2004.
- [2611] R.P. Magnusson, A. Taurog, and M.L. Dorris. Mechanism of iodide-dependent catalytic activity of thyroid peroxidase and lactoperoxidase. *J. Biol. Chem.*, 259:197–205, 1984.

- [2612] W. Mah, J.C. Deme, D. Watkins, S. Fung, A. Janer, E.A. Shoubridge, D.S. Rosenblatt, and J.W. Coulton. Subcellular location of MMACHC and MMADHC, two human proteins central to intracellular vitamin B₁₂ metabolism. *Mol Genet Metab*, 108:112–118, 2013.
- [2613] H.R. Mahler. Studies on the fatty acid oxidizing system of animal tissue. IV. The prosthetic group of butyryl coenzyme A dehydrogenase. *J. Biol. Chem.*, 206:13–26, 1954.
- [2614] H.R. Mahler, G. Hübscher, and H. Baum. Studies on uricase. I. Preparation, purification, and properties of a cuproprotein. *J. Biol. Chem.*, 216:625–641, 1955.
- [2615] H.R. Mahler, B. Mackler, D.E. Green, and R.M. Bock. Studies on metalloflavoproteins. III. Aldehyde oxidase: a molybdo-flavoprotein. *J. Biol. Chem.*, 210:465–480, 1954.
- [2616] H.R. Mahler, I. Raw, R. Molinari, and D.F. do Amaral. Studies of electron transport enzymes. II. Isolation and some properties of a cytochrome-specific reduced diphosphopyridine nucleotide dehydrogenase from pig liver. *J. Biol. Chem.*, 233:230–239, 1958.
- [2617] S.S. Mahmoud and R.B. Croteau. Menthofuran regulates essential oil biosynthesis in peppermint by controlling a downstream monoterpene reductase. *Proc. Natl. Acad. Sci. USA*, 100:14481–14486, 2003.
- [2618] P.C. Mahon, K. Hirota, and G.L. Semenza. FIH-1: a novel protein that interacts with HIF-1 α and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev.*, 15:2675–2686, 2001.
- [2619] D.E. Mahony, C.E. Meier, I.A. Macdonald, and L.V. Holdeman. Bile salt degradation by nonfermentative clostridia. *Appl. Environ. Microbiol.*, 34:419–423, 1977.
- [2620] X. Mai and M.W. Adams. Characterization of a fourth type of 2-keto acid-oxidizing enzyme from a hyperthermophilic archaeon: 2-ketoglutarate ferredoxin oxidoreductase from *Thermococcus litoralis*. *J. Bacteriol.*, 178:5890–5896, 1996.
- [2621] X.H. Mai and M.W.W. Adams. Indolepyruvate ferredoxin oxidoreductase from the hyperthermophilic archaeon *Pyrococcus furiosus* - a new enzyme involved in peptide fermentation. *J. Biol. Chem.*, 269:16726–16732, 1994.
- [2622] M.D. Maines, N.G. Ibrahim, and K. Kappas. Solubilization and partial purification of heme oxygenase from rat liver. *J. Biol. Chem.*, 252:5900–5903, 1977.
- [2623] A. Maiti and A.C. Drohat. Thymine DNA glycosylase can rapidly excise 5-formylcytosine and 5-carboxylcytosine: potential implications for active demethylation of CpG sites. *J. Biol. Chem.*, 286:35334–35338, 2011.
- [2624] P.J. Mak, M.C. Gregory, I.G. Denisov, S.G. Sligar, and J.R. Kincaid. Unveiling the crucial intermediates in androgen production. *Proc. Natl. Acad. Sci. USA*, 112:15856–15861, 2015.
- [2625] M. Mäki, N. Järvinen, J. Rabinä, C. Roos, H. Maaheimo, P. Mattila, and R. Renkonen. Functional expression of *Pseudomonas aeruginosa* GDP-4-keto-6-deoxy-D-mannose reductase which synthesizes GDP-rhamnose. *Eur. J. Biochem.*, 269:593–601, 2002.
- [2626] Y. Maki, S. Yamamoto, M. Nozaki, and O. Hayaishi. Studies on monooxygenases. II. Crystallization and some properties of imidazole acetate monooxygenase. *J. Biol. Chem.*, 244:2942–2950, 1969.
- [2627] M. Makino, H. Sugimoto, Y. Shiro, S. Asamizu, H. Onaka, and S. Nagano. Crystal structures and catalytic mechanism of cytochrome P₄₅₀ StaP that produces the indolocarbazole skeleton. *Proc. Natl. Acad. Sci. USA*, 104:11591–11596, 2007.
- [2628] T.M. Makris, M. Chakrabarti, E. Munck, and J.D. Lipscomb. A family of diiron monooxygenases catalyzing amino acid β -hydroxylation in antibiotic biosynthesis. *Proc. Natl. Acad. Sci. USA*, 107:15391–15396, 2010.
- [2629] T.M. Makris, V.V. Vu, K.K. Meier, A.J. Komor, B.S. Rivard, E. Munck, L. Que, Lipscomb Jr., and J.D. An unusual peroxo intermediate of the arylamine oxygenase of the chloramphenicol biosynthetic pathway. *J. Am. Chem. Soc.*, 137:1608–1617, 2015.
- [2630] Z. Malik, C.J.P. Jones, and M.J. Connock. Assay and subcellular localization of H₂O₂ generating mannitol oxidase in the terrestrial slug *Arion ater*. *J. Exp. Zool.*, 242:9–15, 1987.
- [2631] E. Malito, A. Alfieri, M.W. Fraaije, and A. Mattevi. Crystal structure of a Baeyer-Villiger monooxygenase. *Proc. Natl. Acad. Sci. USA*, 101:13157–13162, 2004.

- [2632] D.H. Mallonee, M.A. Lijewski, and P.B. Hylemon. Expression in *Escherichia coli* and characterization of a bile acid-inducible 3 α -hydroxysteroid dehydrogenase from *Eubacterium* sp. strain VPI 12708. *Curr. Microbiol.*, 30:259–263, 1995.
- [2633] B.G. Malmström, L.-E. Andréasson, and B. Reinhammar. Copper-containing oxidases and superoxide dismutase. In P.D. Boyer, editor, *The Enzymes*, volume 12, pages 507–579. Academic Press, New York, 3rd edition, 1975.
- [2634] R. Mamoto, X. Hu, H. Chiue, Y. Fujioka, and F. Kawai. Cloning and expression of soluble cytochrome *c* and its role in polyvinyl alcohol degradation by polyvinyl alcohol-utilizing *Sphingopyxis* sp. strain 113P3. *J. Biosci. Bioeng.*, 105:147–151, 2008.
- [2635] Y. Manevich, S.I. Feinstein, and A.B. Fisher. Activation of the antioxidant enzyme 1-CYS peroxiredoxin requires glutathionylation mediated by heterodimerization with π GST. *Proc. Natl. Acad. Sci. USA*, 101:3780–3785, 2004.
- [2636] J. Mano, Y. Torii, S. Hayashi, K. Takimoto, K. Matsui, K. Nakamura, D. Inzé, E. Babiychuk, S. Kushnir, and K. Asada. The NADPH:quinone oxidoreductase P1- ζ -crystallin in *Arabidopsis* catalyzes the α,β -hydrogenation of 2-alkenals: detoxication of the lipid peroxide-derived reactive aldehydes. *Plant Cell Physiol.*, 43:1445–1455, 2002.
- [2637] K.M. Manoj. Chlorinations catalyzed by chloroperoxidase occur via diffusible intermediate(s) and the reaction components play multiple roles in the overall process. *Biochim. Biophys. Acta*, 1764:1325–1339, 2006.
- [2638] K.M. Manoj and L.P. Hager. Chloroperoxidase, a janus enzyme. *Biochemistry*, 47:2997–3003, 2008.
- [2639] D.J. Mansell, H.S. Toogood, J. Waller, J.M.X. Hughes, C.W. Levy, J.M. Gardiner, , and N.S. Biocatalytic asymmetric alkene reduction: crystal structure and characterization of a double bond reductase from *Nicotiana tabacum*. *ACS Catal.*, 3:370–379, 2013.
- [2640] R.L. Mansell, G.R. Babbel, and M.H. Zenk. Multiple forms and specificity of coniferyl alcohol dehydrogenase from cambial regions of higher plants. *Phytochemistry*, 15:1849–1853, 1976.
- [2641] H. Mao, J. Liu, F. Ren, R.J. Peters, and Q. Wang. Characterization of CYP71Z18 indicates a role in maize zealexin biosynthesis. *Phytochemistry*, 121:4–10, 2016.
- [2642] L.W. Mapson and E. Breslow. Properties of partially purified L-galactono- γ -lactone dehydrogenase. *Biochem. J.*, 65:29–29, 1957.
- [2643] L.W. Mapson, F.A. Isherwood, and Y.T. Chen. Biological synthesis of L-ascorbic acid: the conversion of L-galactono- γ -lactone into L-ascorbic acid by plant mitochondria. *Biochem. J.*, 56:21–28, 1954.
- [2644] B.N. Marbois and C.F. Clarke. The COQ7 gene encodes a protein in *Saccharomyces cerevisiae* necessary for ubiquinone biosynthesis. *J. Biol. Chem.*, 271:2995–3004, 1996.
- [2645] M. Marcia, J.D. Langer, D. Parcej, V. Vogel, G. Peng, and H. Michel. Characterizing a monotopic membrane enzyme. Biochemical, enzymatic and crystallization studies on *Aquifex aeolicus* sulfide:quinone oxidoreductase. *Biochim. Biophys. Acta*, 1798:2114–2123, 2010.
- [2646] J. Marcinkeviciene, L.M. Tinney, K.H. Wang, M.J. Rogers, and R.A. Copeland. Dihydroorotate dehydrogenase B of *Enterococcus faecalis*. Characterization and insights into chemical mechanism. *Biochemistry*, 38:13129–13137, 1999.
- [2647] P.I. Marcus and P. Talalay. Induction and purification of α - and β -hydroxysteroid dehydrogenases. *J. Biol. Chem.*, 218:661–674, 1956.
- [2648] M. Maremonti, G. Greco, and R. Wichmann. Characterisation of 2,5-diketo-D-gluconic acid reductase from *Corynebacterium* sp. *Biotechnology Letters*, 18:845–850, 1996.
- [2649] B. Maresca, E. Jacobson, G. Medoff, and G. Kobayashi. Cystine reductase in the dimorphic fungus *Histoplasma capsulatum*. *J. Bacteriol.*, 135:987–992, 1978.
- [2650] Z. Marijanovic, D. Laubner, G. Moller, C. Gege, B. Husen, J. Adamski, and R. Breitling. Closing the gap: identification of human 3-ketosteroid reductase, the last unknown enzyme of mammalian cholesterol biosynthesis. *Mol. Endocrinol.*, 17:1715–1725, 2003.
- [2651] F. Märki and C. Martius. Vitamin K-Reductase, Darstellung und Eigenschaften. *Biochem. Z.*, 333:111–135, 1960.

- [2652] S. Markolovic, Q. Zhuang, S.E. Wilkins, C.D. Eaton, M.I. Abboud, M.J. Katz, H.E. McNeil, R.K. Lesniak, C. Hall, W.B. Struwe, R. Konietzny, S. Davis, M. Yang, W. Ge, J.L.P. Benesch, B.M. Kessler, P.J. Ratcliffe, M.E. Cockman, R. Fischer, P. Wappner, R. Chowdhury, M.L. Coleman, and C.J. Schofield. The Jumonji-C oxygenase JMJD7 catalyzes (3S)-lysyl hydroxylation of TRAFAC GTPases. *Nat. Chem. Biol.*, 14:688–695, 2018.
- [2653] A. Markovitz. Biosynthesis of guanosine diphosphate D-rhamnose and guanosine diphosphate D-talomethylose from guanosine diphosphate α -D-mannose. *J. Biol. Chem.*, 239:2091–2098, 1964.
- [2654] P.J. Markovitz, D.T. Chuang, and R.P. Cox. Familial hyperlysinemias. Purification and characterization of the bifunctional aminoadipic semialdehyde synthase with lysine-ketoglutarate reductase and saccharopine dehydrogenase activities. *J. Biol. Chem.*, 259:11643–11646, 1984.
- [2655] A. Markus, D. Krekel, and F. Lingens. Purification and some properties of component A of the 4-chlorophenylacetate 3,4-dioxygenase from *Pseudomonas* species strain CBS. *J. Biol. Chem.*, 261:12883–12888, 1986.
- [2656] J. Marmur and R.D. Hotchkiss. Mannitol metabolism, a transferable property of pneumococcus. *J. Biol. Chem.*, 214:383–396, 1955.
- [2657] J.V. Marques, K.W. Kim, C. Lee, M.A. Costa, G.D. May, J.A. Crow, L.B. Davin, and N.G. Lewis. Next generation sequencing in predicting gene function in podophyllotoxin biosynthesis. *J. Biol. Chem.*, 288:466–479, 2013.
- [2658] H. Marrakchi, G. Laneelle, and A. Quemard. InhA, a target of the antituberculous drug isoniazid, is involved in a mycobacterial fatty acid elongation system, FAS-II. *Microbiology*, 146:289–296, 2000.
- [2659] L. Marrone, S. Siemann, M. Beecroft, and T. Viswanatha. Specificity of lysine:N-6-hydroxylase: A hypothesis for a reactive substrate intermediate in the catalytic mechanism. *Bioorg. Chem.*, 24:401–406, 1996.
- [2660] I.R. Marsh and M. Bradley. Substrate specificity of trypanothione reductase. *Eur. J. Biochem.*, 243:690–694, 1977.
- [2661] S. Martens and G. Forkmann. Cloning and expression of flavone synthase II from *Gerbera* hybrids. *Plant J.*, 20:611–618, 1999.
- [2662] S. Martens, G. Forkmann, L. Britsch, F. Wellmann, U. Matern, and R. Lukačín. Divergent evolution of flavonoid 2-oxoglutarate-dependent dioxygenases in parsley. *FEBS Lett.*, 544:93–98, 2003.
- [2663] S. Martens, G. Forkmann, U. Matern, and R. Lukačín. Cloning of parsley flavone synthase I. *Phytochemistry*, 58:43–46, 2001.
- [2664] E.S. Martens-Uzunova and P.J. Schaap. An evolutionary conserved D-galacturonic acid metabolic pathway operates across filamentous fungi capable of pectin degradation. *Fungal Genet. Biol.*, 45:1449–1457, 2008.
- [2665] J.F. Martín, S. Gutiérrez, F.J. Fernández, J. Velasco, F. Fierro, A.T. Marcos, and K. Kosalkova. Expression of genes and processing of enzymes for the biosynthesis of penicillins and cephalosporins. *Antonie Van Leeuwenhoek*, 65:227–243, 1994.
- [2666] J.L. Martin. Thioredoxin - a fold for all reasons. *Structure*, 3:245–250, 1995.
- [2667] R.C. Martin, M.C. Mok, G. Shaw, and D.W.S. Mok. An enzyme mediating the conversion of zeatin to dihydrozeatin in *Phaseolus* embryos. *Plant Physiol.*, 90:1630–1635, 1989.
- [2668] R.O. Martin and P.K. Stumpf. Fat metabolism in higher plants. XII. α -Oxidation of long chain fatty acids. *J. Biol. Chem.*, 234:2548–2554, 1959.
- [2669] I. Martineau, A. Belanger, A. Tchernof, and Y. Tremblay. Molecular cloning and expression of guinea pig cytochrome P450c21 cDNA (steroid 21-hydroxylase) isolated from the adrenals. *J. Steroid Biochem. Mol. Biol.*, 86:123–132, 2003.
- [2670] G. Martinez, H.A. Barker, and B.L. Horecker. A specific mannitol dehydrogenase from *Lactobacillus brevis*. *J. Biol. Chem.*, 238:1598–1603, 1963.
- [2671] M.J. Martínez, F.J. Ruiz-Dueñas, F. Guillén, and A.T. Martínez. Purification and catalytic properties of two manganese peroxidase isoenzymes from *Pleurotus eryngii*. *Eur. J. Biochem.*, 237:424–432, 1996.

- [2672] S. Martinez, M. Fellner, C.Q. Herr, A. Ritchie, J. Hu, and R.P. Hausinger. Structures and mechanisms of the non-heme Fe(II)- and 2-oxoglutarate-dependent ethylene-forming enzyme: substrate binding creates a twist. *J. Am. Chem. Soc.*, 139:11980–11988, 2017.
- [2673] T. Martinez, G. Fabrias, and F. Camps. Sex pheromone biosynthetic pathway in *Spodoptera littoralis* and its activation by a neurohormone. *J. Biol. Chem.*, 265:1381–1387, 1990.
- [2674] K. Maruyama. Isolation and identification of the reaction product of α -hydroxy- γ -carboxymuconic ϵ -semialdehyde dehydrogenase. *J. Biochem.*, 86:1671–1677, 1979.
- [2675] K. Maruyama. Purification and properties of 2-pyrone-4,6-dicarboxylate hydrolase. *J. Biochem. (Tokyo)*, 93:557–565, 1983.
- [2676] K. Maruyama, N. Ariga, M. Tsuda, and K. Deguchi. Purification and properties of α -hydroxy- γ -carboxymuconic ϵ -semialdehyde dehydrogenase. *J. Biochem. (Tokyo)*, 83:1125–1134, 1978.
- [2677] R. Maruyama, M. Nishizawa, Y. Itoi, S. Ito, and M. Inoue. Isolation and expression of a *Bacillus cereus* gene encoding benzil reductase. *Biotechnol. Bioeng.*, 75:630–633, 2001.
- [2678] R. Maruyama, M. Nishizawa, Y. Itoi, S. Ito, and M. Inoue. The enzymes with benzil reductase activity conserved from bacteria to mammals. *J. Biotechnol.*, 94:157–169, 2002.
- [2679] E. Masai, K. Momose, H. Hara, S. Nishikawa, Y. Katayama, and M. Fukuda. Genetic and biochemical characterization of 4-carboxy-2-hydroxymuconate-6-semialdehyde dehydrogenase and its role in the protocatechuate 4,5-cleavage pathway in *Sphingomonas paucimobilis* SYK-6. *J. Bacteriol.*, 182:6651–6658, 2000.
- [2680] G.T. Mashabela and F.P. Seebeck. Substrate specificity of an oxygen dependent sulfoxide synthase in ovolithol biosynthesis. *Chem. Commun. (Camb.)*, 49:7714–7716, 2013.
- [2681] K. Mashiguchi, K. Tanaka, T. Sakai, S. Sugawara, H. Kawaide, M. Natsume, A. Hanada, T. Yaeno, K. Shirasu, H. Yao, P. McSteen, Y. Zhao, K. Hayashi, Y. Kamiya, and H. Kasahara. The main auxin biosynthesis pathway in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA*, 108:18512–18517, 2011.
- [2682] H.S. Mason. Structures and functions of the phenolase complex. *Nature (Lond.)*, 177:79–81, 1956.
- [2683] M.G. Mason, P. Nicholls, C. Divne, B.M. Hallberg, G. Henriksson, and M.T. Wilson. The heme domain of cellobiose oxidoreductase: a one-electron reducing system. *Biochim. Biophys. Acta*, 1604:47–54, 2003.
- [2684] V. Massey. The composition of the ketoglutarate dehydrogenase complex. *Biochim. Biophys. Acta*, 38:447–460, 1960.
- [2685] V. Massey. Lipoyl dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 275–306. Academic Press, New York, 2nd edition, 1963.
- [2686] V. Massey, Q.H. Gibson, and C. Veeger. Intermediates in the catalytic action of lipoyl dehydrogenase (diaphorase). *Biochem. J.*, 77:341–351, 1960.
- [2687] V. Massey, G. Palmer, and R. Bennett. The purification and some properties of D-amino acid oxidase. *Biochim. Biophys. Acta*, 48:1–9, 1961.
- [2688] N. Mast, A.J. Annalora, D.T. Lodowski, K. Palczewski, C.D. Stout, and I.A. Pikuleva. Structural basis for three-step sequential catalysis by the cholesterol side chain cleavage enzyme CYP11A1. *J. Biol. Chem.*, 286:5607–5613, 2011.
- [2689] N. Mast, R. Norcross, U. Andersson, M. Shou, K. Nakayama, I. Bjorkhem, and I.A. Pikuleva. Broad substrate specificity of human cytochrome P_{450} 46A1 which initiates cholesterol degradation in the brain. *Biochemistry*, 42:14284–14292, 2003.
- [2690] B.S.S. Masters, M.H. Bilimoria, H. Kamen, and Q.H. Gibson. The mechanism of 1- and 2-electron transfers catalyzed by reduced triphosphopyridine nucleotide-cytochrome *c* reductase. *J. Biol. Chem.*, 240:4081–4088, 1965.
- [2691] B.S.S. Masters, H. Kamin, Q.H. Williams Gibson, , and Jr. Studies on the mechanism of microsomal triphosphopyridine nucleotide-cytochrome *c* reductase. *J. Biol. Chem.*, 240:921–931, 1965.
- [2692] D.S. Masters, Meister Jr., and A. Inhibition of homocysteine sulfonamide of glutamate synthase purified from *Saccharomyces cerevisiae*. *J. Biol. Chem.*, 257:8711–8715, 1982.

- [2693] S. Masuda, S.A. Strugnell, J.C. Knutson, R. St-Arnaud, and G. Jones. Evidence for the activation of 1 α -hydroxyvitamin D₂ by 25-hydroxyvitamin D-24-hydroxylase: delineation of pathways involving 1 α ,24-dihydroxyvitamin D₂ and 1 α ,25-dihydroxyvitamin D₂. *Biochim. Biophys. Acta*, 1761:221–234, 2006.
- [2694] S. Masuda, Y. Suzuki, Y. Fujitani, R. Mitsui, T. Nakagawa, M. Shintani, and A. Tani. Lanthanide-dependent regulation of methylothrophy in *Methylobacterium aquaticum* strain 22A. *mSphere*, 3:e00462–e00417, 2018.
- [2695] T. Masui, R. Herman, and E. Staple. The oxidation of 5 β -cholestane-3 α ,7 α ,12 α ,26-tetraol to 5 β -cholestane-3 α ,7 α ,12 α -triol-26-oic acid via 5 β -cholestane-3 α ,7 α ,12 α -triol-26-al by rat liver. *Biochim. Biophys. Acta*, 117:266–268, 1966.
- [2696] M. Matsubara, S. Katoh, M. Akino, and S. Kaufman. Sepiapterin reductase. *Biochim. Biophys. Acta*, 122:202–212, 1966.
- [2697] S. Matsubara, T. Morosinotto, R. Bassi, A.L. Christian, E. Fischer-Schliebs, U. Luttge, B. Orthen, A.C. Franco, F.R. Scarano, B. Forster, B.J. Pogson, and C.B. Osmond. Occurrence of the lutein-epoxide cycle in mistletoes of the Loranthaceae and Viscaceae. *Planta*, 217:868–879, 2003.
- [2698] H. Matsuda and Y. Suzuki. γ -Guanidinobutyraldehyde dehydrogenase of *Vicia faba* leaves. *Plant Physiol.*, 76:654–657, 1984.
- [2699] D. Matsui, A. Terai, and Y. Asano. L-Arginine oxidase from *Pseudomonas* sp. TPU 7192: Characterization, gene cloning, heterologous expression, and application to L-arginine determination. *Enzyme Microb. Technol.*, 82:151–157, 2016.
- [2700] T. Matsui, S. Nambu, Y. Ono, C.W. Goulding, K. Tsumoto, and M. Ikeda-Saito. Heme degradation by *Staphylococcus aureus* IsdG and IsdI liberates formaldehyde rather than carbon monoxide. *Biochemistry*, 52:3025–3027, 2013.
- [2701] J. Matsumoto, M. Higuchi, M. Shimada, Y. Yamamoto, and Y. Kamio. Molecular cloning and sequence analysis of the gene encoding the H₂O-forming NADH oxidase from *Streptococcus mutans*. *Biosci. Biotechnol. Biochem.*, 60:39–43, 1996.
- [2702] S. Matsumoto, M. Mizutani, K. Sakata, and B. Shimizu. Molecular cloning and functional analysis of the *ortho*-hydroxylases of *p*-coumaroyl coenzyme A/feruloyl coenzyme A involved in formation of umbelliferone and scopoletin in sweet potato, *Ipomoea batatas* (L.) Lam. *Phytochemistry*, 74:49–57, 2012.
- [2703] T. Matsumoto, C.D. Funk, O. Radmark, J.-O. Hoog, H. Jornvall, and B. Samuelsson. Molecular cloning and amino acid sequence of human 5-lipoxygenase. *Proc. Natl. Acad. Sci. USA*, 85:26–30, 1988.
- [2704] I. Matsunaga and Y. Shiro. Peroxide-utilizing biocatalysts: structural and functional diversity of heme-containing enzymes. *Curr. Opin. Chem. Biol.*, 8:127–132, 2004.
- [2705] I. Matsunaga, A. Ueda, N. Fujiwara, T. Sumimoto, and K. Ichihara. Characterization of the *ybdT* gene product of *Bacillus subtilis*: novel fatty acid β -hydroxylating cytochrome P450. *Lipids*, 34:841–846, 1999.
- [2706] I. Matsunaga, A. Yamada, D.S. Lee, E. Obayashi, N. Fujiwara, K. Kobayashi, H. Ogura, and Y. Shiro. Enzymatic reaction of hydrogen peroxide-dependent peroxygenase cytochrome P₄₅₀S: kinetic deuterium isotope effects and analyses by resonance Raman spectroscopy. *Biochemistry*, 41:1886–1892, 2002.
- [2707] I. Matsunaga, M. Yamada, E. Kusunose, T. Miki, and K. Ichihara. Further characterization of hydrogen peroxide-dependent fatty acid α -hydroxylase from *Sphingomonas paucimobilis*. *J. Biochem.*, 124:105–110, 1998.
- [2708] I. Matsunaga, M. Yamada, E. Kusunose, Y. Nishiuchi, I. Yano, and K. Ichihara. Direct involvement of hydrogen peroxide in bacterial α -hydroxylation of fatty acid. *FEBS Lett.*, 386:252–254, 1996.
- [2709] M. Matsuno, V. Compagnon, G.A. Schoch, M. Schmitt, D. Debayle, J.E. Bassard, B. Pollet, A. Hehn, D. Heintz, P. Ullmann, C. Lapiere, F. Bernier, J. Ehrling, and D. Werck-Reichhart. Evolution of a novel phenolic pathway for pollen development. *Science*, 325:1688–1692, 2009.
- [2710] A. Matsushima, Y. Sato, M. Otsuka, T. Watanabe, H. Yamamoto, and T. Hirata. An enone reductase from *Nicotiana tabacum*: cDNA cloning, expression in *Escherichia coli*, and reduction of enones with the recombinant proteins. *Bioorg. Chem.*, 36:23–28, 2008.
- [2711] K. Matsushima, Y. Ando, T. Hamasaki, and K. Yabe. Purification and characterization of two versiconal hemiacetal acetate reductases involved in aflatoxin biosynthesis. *Appl. Environ. Microbiol.*, 60:2561–2567, 1994.

- [2712] K. Matsushita and H.R. Kaback. D-Lactate oxidation and generation of the proton electrochemical gradient in membrane vesicles from *Escherichia coli* GR19N and in proteoliposomes reconstituted with purified D-lactate dehydrogenase and cytochrome *o* oxidase. *Biochemistry*, 25:2321–2327, 1986.
- [2713] K. Matsushita, Y. Kobayashi, M. Mizuguchi, H. Toyama, O. Adachi, K. Sakamoto, and H. Miyoshi. A tightly bound quinone functions in the ubiquinone reaction sites of quinoprotein alcohol dehydrogenase of an acetic acid bacterium, *Gluconobacter suboxydans*. *Biosci. Biotechnol. Biochem.*, 72:2723–2731, 2008.
- [2714] K. Matsushita, E. Shinagawa, O. Adachi, and M. Ameyama. Quinoprotein D-glucose dehydrogenase of the *Acinetobacter calcoaceticus* respiratory chain: membrane-bound and soluble forms are different molecular species. *Biochemistry*, 28:6276–6280, 1989.
- [2715] K. Matsushita, E. Shinagawa, and M. Ameyama. D-Gluconate dehydrogenases from bacteria, 2-keto-D-gluconate-yielding membrane-bound. *Methods Enzymol.*, 89:187–193, 1982.
- [2716] K. Matsushita, Y. Takaki, E. Shinagawa, M. Ameyama, and O. Adachi. Ethanol oxidase respiratory chain of acetic acid bacteria. Reactivity with ubiquinone of pyrroloquinoline quinone-dependent alcohol dehydrogenases purified from *Acetobacter aceti* and *Gluconobacter suboxydans*. *Biosci. Biotechnol. Biochem.*, 56:304–310, 1992.
- [2717] K. Matsushita, H. Toyama, and O. Adachi. Respiratory chains and bioenergetics of acetic acid bacteria. *Adv. Microb. Physiol.*, 36:247–301, 1994.
- [2718] K. Matsushita, H. Toyama, M. Ameyama, O. Adachi, A. Dewanti, and J.A. Duine. Soluble and membrane-bound quinoprotein D-glucose dehydrogenases of the *Acinetobacter calcoaceticus* : the binding process of PQQ to the apoenzymes. *Biosci. Biotechnol. Biochem.*, 59:1548–1555, 1995.
- [2719] K. Matsushita, T. Yakushi, H. Toyama, E. Shinagawa, and O. Adachi. Function of multiple heme *c* moieties in intramolecular electron transport and ubiquinone reduction in the quinohemoprotein alcohol dehydrogenase-cytochrome *c* complex of *Gluconobacter suboxydans*. *J. Biol. Chem.*, 271:4850–4857, 1996.
- [2720] K. Matsushita, T. Yamashita, N. Aoki, H. Toyama, and O. Adachi. Electron transfer from quinohemoprotein alcohol dehydrogenase to blue copper protein azurin in the alcohol oxidase respiratory chain of *Pseudomonas putida* HK5. *Biochemistry*, 38:6111–6118, 1999.
- [2721] Y. Matsuzaka, K. Okamoto, H. Tsuji, T. Mabuchi, A. Ozawa, G. Tamiya, and H. Inoko. Identification of the hRDH-E2 gene, a novel member of the SDR family, and its increased expression in psoriatic lesion. *Biochem. Biophys. Res. Commun.*, 297:1171–1180, 2002.
- [2722] A. Mattevi, G. Tedeschi, L. Bacchella, A. Coda, A. Negri, and S. Ronchi. Structure of L-aspartate oxidase: implications for the succinate dehydrogenase/fumarate reductase oxidoreductase family. *Structure*, 7:745–756, 1999.
- [2723] A. Matthews, R. Saleem-Batcha, J.N. Sanders, F. Stull, K.N. Houk, and R. Teufel. Aminoperoxide adducts expand the catalytic repertoire of flavin monooxygenases. *Nat. Chem. Biol.*, 16:556–563, 2020.
- [2724] P. Mattila, J. Rabinä, S. Hortling, J. Jelin, and R. Renkonen. Functional expression of *Escherichia coli* enzymes synthesizing GDP-L-fucose from inherent GDP-D-mannose in *Saccharomyces cerevisiae*. *Glycobiology*, 10:1041–1047, 2000.
- [2725] M. Matuschek, C. Wallwey, X. Xie, and S.M. Li. New insights into ergot alkaloid biosynthesis in *Claviceps purpurea*: an agroclavine synthase EasG catalyses, via a non-enzymatic adduct with reduced glutathione, the conversion of chanoclavine-I aldehyde to agroclavine. *Org. Biomol. Chem.*, 9:4328–4335, 2011.
- [2726] C.J. Mau, F. Karp, M. Ito, G. Honda, and R.B. Croteau. A candidate cDNA clone for (–)-limonene-7-hydroxylase from *Perilla frutescens*. *Phytochemistry*, 71:373–379, 2010.
- [2727] D.V. Mavrodi, R.F. Bonsall, S.M. Delaney, M.J. Soule, G. Phillips, and L.S. Thomashow. Functional analysis of genes for biosynthesis of pyocyanin and phenazine-1-carboxamide from *Pseudomonas aeruginosa* PAO1. *J. Bacteriol.*, 183:6454–6465, 2001.
- [2728] E.S. Maxwell, H.M. Kalckar, and J.L. Strominger. Some properties of uridine diphosphoglucose dehydrogenase. *Arch. Biochem. Biophys.*, 65:2–10, 1956.

- [2729] A.M. Mayer and E. Harel. Polyphenol oxidases in plants. *Phytochemistry*, 18:193–215, 1979.
- [2730] E. Mayer, S. Waldner-Sander, B. Keller, E. Keller, and F. Lingens. Purification of arogenate dehydrogenase from *Phenyllobacterium immobile*. *FEBS Lett.*, 179:208–212, 1985.
- [2731] J. Mayer, T. Huhn, M. Habeck, K. Denger, K. Hollemeyer, and A.M. Cook. 2,3-Dihydroxypropane-1-sulfonate degraded by *Cupriavidus pinatubonensis* JMP134: purification of dihydroxypropanesulfonate 3-dehydrogenase. *Microbiology*, 156:1556–1564, 2010.
- [2732] O. Mayol, K. Bastard, L. Beloti, A. Frese, J.P. Turkenburg, J.L. Petit, A. Mariage, A. Debard, V. Pellouin, A. Perret, V. de Berardinis, A. Zaparucha, G. Grogan, and C. Vergne-Vaxelaire. A family of native amine dehydrogenases for the asymmetric reductive amination of ketones. *Nature Catalysis*, 2:324–333, 2019.
- [2733] O. Mayol, S. David, E. Darii, A. Debard, A. Mariage, V. Pellouin, J.L. Petit, M. Salanoubat, V. de Berardinis, A. Zaparucha, and C. Vergne-Vaxelaire. Asymmetric reductive amination by a wild-type amine dehydrogenase from the thermophilic bacteria *Petrogala mobilis*. *Catalysis Science & Technology*, 6:7421–7428, 2016.
- [2734] P. Mayr, K. Bruggler, K.D. Kulbe, and B. Nidetzky. D-Xylose metabolism by *Candida intermedia*: isolation and characterisation of two forms of aldose reductase with different coenzyme specificities. *J. Chromatogr. B Biomed. Sci. Appl.*, 737:195–202, 2000.
- [2735] R. Mazmouz, F. Chapuis-Hugon, Mejean Pichon V., A., and O. The last step of the biosynthesis of the cyanotoxins cylindrospermopsin and 7-*epi*-cylindrospermopsin is catalysed by CyrI, a 2-oxoglutarate-dependent iron oxygenase. *ChemBioChem*, 12:858–862, 2011.
- [2736] R. Mazmouz, I. Essadik, D. Hamdane, A. Mejean, and O. Ploux. Characterization of CyrI, the hydroxylase involved in the last step of cylindrospermopsin biosynthesis: Binding studies, site-directed mutagenesis and stereoselectivity. *Arch. Biochem. Biophys.*, 647:1–9, 2018.
- [2737] J. Mazoch, R. Tesarik, V. Sedlacek, I. Kucera, and J. Turanek. Isolation and biochemical characterization of two soluble iron(III) reductases from *Paracoccus denitrificans*. *Eur. J. Biochem.*, 271:553–562, 2004.
- [2738] R.P. McAndrew, Y. Wang, A.W. Mohsen, M. He, J. Vockley, and J.J. Kim. Structural basis for substrate fatty acyl chain specificity: crystal structure of human very-long-chain acyl-CoA dehydrogenase. *J. Biol. Chem.*, 283:9435–9443, 2008.
- [2739] M.J. McBride, S.R. Pope, K. Hu, C.D. Okafor, E.P. Balskus, J.M. Bollinger, Boal Jr., and A.K. Structure and assembly of the diiron cofactor in the heme-oxygenase-like domain of the *N*-nitrosourea-producing enzyme SznF. *Proc. Natl. Acad. Sci. USA*, 118, 2021.
- [2740] M.J. McBride, D. Sil, T.L. Ng, A.M. Croke, G.E. Kenney, C.R. Tysoe, B. Zhang, E.P. Balskus, A.K. Boal, C., Bollinger Krebs, , and Jr. A peroxodiiron(III/III) intermediate mediating both *N*-hydroxylation steps in biosynthesis of the *N*-nitrosourea pharmacophore of streptozotocin by the multi-domain metalloenzyme SznF. *J. Am. Chem. Soc.*, 142:11818–11828, 2020.
- [2741] D.L. McCarthy, S. Navarrete, W.S. Willett, P.C. Babbitt, and S.D. Copley. Exploration of the relationship between tetrachlorohydroquinone dehalogenase and the glutathione *S*-transferase superfamily. *Biochemistry*, 35:14634–14642, 1996.
- [2742] M. McConn, S. Hugly, J. Browse, and C. Somerville. A mutation at the *fad8* locus of *Arabidopsis* identifies a second chloroplast ω -3 desaturase. *Plant Physiol.*, 106:1609–1614, 1994.
- [2743] J.R.D. McCormick, U. Hirsch, N.O. Sjolander, and A.P. Doerschuk. Cosynthesis of tetracyclines by pairs of *Streptomyces aureofaciens* mutants. *J. Am. Chem. Soc.*, 82:5006–5007, 1960.
- [2744] J.R.D. McCormick and G.O. Morton. Identity of cosynthetic factor I of *Streptomyces aureofaciens* and fragment FO from coenzyme F₄₂₀ of *Methanobacterium* species. *J. Am. Chem. Soc.*, 104:4014–4015, 1982.
- [2745] K.M. McCulloch, T. Mukherjee, T.P. Begley, and S.E. Ealick. Structure of the PLP degradative enzyme 2-methyl-3-hydroxypyridine-5-carboxylic acid oxygenase from *Mesorhizobium loti* MAFF303099 and its mechanistic implications. *Biochemistry*, 48:4139–4149, 2009.

- [2746] C.A. McDevitt, P. Hugenholtz, G.R. Hanson, and A.G. McEwan. Molecular analysis of dimethyl sulphide dehydrogenase from *Rhodovulum sulfidophilum*: its place in the dimethyl sulphoxide reductase family of microbial molybdopterin-containing enzymes. *Mol. Microbiol.*, 44:1575–1587, 2002.
- [2747] M.T. McDowell and J.C. Lagarias. Purification and biochemical properties of phytochromobilin synthase from etiolated oat seedlings. *Plant Physiol.*, 126:1546–1554, 2001.
- [2748] J.S. McFarlane, C.L. Davis, and A.L. Lamb. Staphylopine, pseudopaline, and yersinopine dehydrogenases: A structural and kinetic analysis of a new functional class of opine dehydrogenase. *J. Biol. Chem.*, 293:8009–8019, 2018.
- [2749] P. McGlynn and C.N. Hunter. Genetic analysis of the *bchC* and *bchA* genes of *Rhodobacter sphaeroides*. *Mol. Gen. Genet.*, 236:227–234, 1993.
- [2750] W. McIntire, D.E. Edmondson, and T.P. Singer. 8 α -O-Tyrosyl-FAD: a new form of covalently bound flavin from *p*-cresol methylhydroxylase. *J. Biol. Chem.*, 255:6553–6555, 1980.
- [2751] L. McIver, C. Leadbeater, D.J. Campopiano, R.L. Baxter, S.N. Daff, S.K. Chapman, and A.W. Munro. Characterisation of flavodoxin NADP⁺ oxidoreductase and flavodoxin; key components of electron transfer in *Escherichia coli*. *Eur. J. Biochem.*, 257:577–585, 1998.
- [2752] E.J. McKenna and M.J. Coon. Enzymatic ω -oxidation. IV. Purification and properties of the ω -hydroxylase of *Pseudomonas oleovorans*. *J. Biol. Chem.*, 245:3882–3889, 1970.
- [2753] K.J. McLean, P. Lafite, C. Levy, M.R. Cheesman, N. Mast, I.A. Pikuleva, D. Leys, and A.W. Munro. The Structure of *Mycobacterium tuberculosis* CYP125: molecular basis for cholesterol binding in a P450 needed for host infection. *J. Biol. Chem.*, 284:35524–35533, 2009.
- [2754] B. McMahon, M.E. Gallagher, and S.G. Mayhew. The protein coded by the PP2216 gene of *Pseudomonas putida* KT2440 is an acyl-CoA dehydrogenase that oxidises only short-chain aliphatic substrates. *FEMS Microbiol. Lett.*, 250:121–127, 2005.
- [2755] L.A. McNeill, K.S. Hewitson, J.M. Gleadle, L.E. Horsfall, N.J. Oldham, P.H. Maxwell, C.W. Pugh, P.J. Ratcliffe, and C.J. Schofield. The use of dioxygen by HIF prolyl hydroxylase (PHD1). *Bioorg. Med. Chem. Lett.*, 12:1547–1550, 2002.
- [2756] M.S. McReynolds and G.B. Kitto. Purification and properties of *Drosophila* malate dehydrogenases. *Biochim. Biophys. Acta*, 198:165–175, 1970.
- [2757] F.R. McSorley, P.B. Wyatt, A. Martinez, E.F. DeLong, B. Hove-Jensen, and D.L. Zechel. PhnY and PhnZ comprise a new oxidative pathway for enzymatic cleavage of a carbon-phosphorus bond. *J. Am. Chem. Soc.*, 134:8364–8367, 2012.
- [2758] A. Medina and D.J.D. Nicholas. Hyponitrite reductase in *Neurospora*. *Nature (Lond.)*, 179:533–534, 1957.
- [2759] A.E. Medlock and H.A. Dailey. Human coproporphyrinogen oxidase is not a metalloprotein. *J. Biol. Chem.*, 271:32507–32510, 1996.
- [2760] M. Meguro, H. Ito, A. Takabayashi, R. Tanaka, and A. Tanaka. Identification of the 7-hydroxymethyl chlorophyll *a* reductase of the chlorophyll cycle in *Arabidopsis*. *Plant Cell*, 23:3442–3453, 2011.
- [2761] Y.T. Meharena, H. Li, D.B. Hawkes, A.G. Pearson, J. De Voss, and T.L. Poulos. Crystal structure of P450_{cin} in a complex with its substrate, 1,8-cineole, a close structural homologue to D-camphor, the substrate for P450_{cam}. *Biochemistry*, 43:9487–9494, 2004.
- [2762] Y.T. Meharena, K.E. Slessor, S.M. Cavaignac, T.L. Poulos, and J.J. De Voss. The critical role of substrate-protein hydrogen bonding in the control of regioselective hydroxylation in p450cin. *J. Biol. Chem.*, 283:10804–10812, 2008.
- [2763] R.A. Meigs and K.J. Ryan. 16- α -Hydroxysteroid dehydrogenase of rat kidney. Purification, assay, and properties. *J. Biol. Chem.*, 241:4011–4015, 1966.
- [2764] A.K. Meisner, A. Saffert, P. Schreier, and A. Schon. Fatty acid α -dioxygenase from *Pisum sativum*: temporal and spatial regulation during germination and plant development. *J. Plant Physiol.*, 166:333–343, 2009.
- [2765] A. Meister, A.N. Radhakrishnan, and S.D. Buckley. Enzymatic synthesis of L-pipecolic acid and L-proline. *J. Biol. Chem.*, 229:789–800, 1957.

- [2766] A. Meister and D. Wellner. Flavoprotein amino acid oxidase. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 609–648. Academic Press, New York, 2nd edition, 1963.
- [2767] A. Melo and L. Glaser. The mechanism of 6-deoxyhexose synthesis. II. Conversion of deoxythymidine diphosphate 4-keto-6-deoxy-D-glucose to deoxythymidine diphosphate L-rhamnose. *J. Biol. Chem.*, 243:1475–1478, 1968.
- [2768] A.M. Melo, T.M. Bandejas, and M. Teixeira. New insights into type II NAD(P)H:quinone oxidoreductases. *Microbiol. Mol. Biol. Rev.*, 68:603–616, 2004.
- [2769] K.M. Meneely and A.L. Lamb. Biochemical characterization of a flavin adenine dinucleotide-dependent monooxygenase, ornithine hydroxylase from *Pseudomonas aeruginosa*, suggests a novel reaction mechanism. *Biochemistry*, 46:11930–11937, 2007.
- [2770] B. Mennenga, C.W. Kay, and H. Gorisch. Quinoprotein ethanol dehydrogenase from *Pseudomonas aeruginosa*: the unusual disulfide ring formed by adjacent cysteine residues is essential for efficient electron transfer to cytochrome *c*₅₅₀. *Arch. Microbiol.*, 191:361–367, 2009.
- [2771] S. Menon, M. Stahl, R. Kumar, G.-Y. Xu, and F. Sullivan. Stereochemical course and steady state mechanism of the reaction catalyzed by the GDP-fucose synthetase from *Escherichia coli*. *J. Biol. Chem.*, 274:26743–26750, 1999.
- [2772] J. Menting, R.K. Scopes, and T.W. Stevenson. Characterization of flavonoid 3',5'-hydroxylase in microsomal membrane fraction of *Petunia hybrida* flowers. *Plant Physiol.*, 106:633–642, 1994.
- [2773] C. Mercier, V. Chalansonnet, S. Orega, and C. Gilbert. Characteristics of major *Escherichia coli* reductases involved in aerobic nitro and azo reduction. *J. Appl. Microbiol.*, 115:1012–1022, 2013.
- [2774] F. Meschi, F. Wiertz, L. Klauss, C. Cavalieri, A. Blok, B. Ludwig, H.A. Heering, A. Merli, G.L. Rossi, and M. Ubbink. Amicyanin transfers electrons from methylamine dehydrogenase to cytochrome *c*-551i via a ping-pong mechanism, not a ternary complex. *J. Am. Chem. Soc.*, 132:14537–14545, 2010.
- [2775] R. Meskys, R.J. Harris, V. Casaite, J. Basran, and N.S. Scrutton. Organization of the genes involved in dimethylglycine and sarcosine degradation in *Arthrobacter* spp.: implications for glycine betaine catabolism. *Eur. J. Biochem.*, 268:3390–3398, 2001.
- [2776] J. Messens, G. Hayburn, A. Desmyter, G. Laus, and L. Wyns. The essential catalytic redox couple in arsenate reductase from *Staphylococcus aureus*. *Biochemistry*, 38:16857–16865, 1999.
- [2777] J. Messens, J.C. Martins, K. Van Belle, E. Brosens, A. Desmyter, M. De Gieter, J.M. Wieruszkeski, R. Willem, L. Wyns, and I. Zegers. All intermediates of the arsenate reductase mechanism, including an intramolecular dynamic disulfide cascade. *Proc. Natl. Acad. Sci. USA*, 99:8506–8511, 2002.
- [2778] J. Messens and S. Silver. Arsenate reduction: thiol cascade chemistry with convergent evolution. *J. Mol. Biol.*, 362:1–17, 2006.
- [2779] A. Messerschmidt, R. Ladenstein, R. Huber, M. Bolognesi, L. Avigliano, R. Petruzzelli, A. Rossi, and A. Finazzi-Agro. Refined crystal structure of ascorbate oxidase at 1.9 Å resolution. *J. Mol. Biol.*, 224:179–205, 1992.
- [2780] S. El Mestikawy, J. Glowinski, and M. Hamon. Tyrosine hydroxylase activation in depolarized dopaminergic terminals -involvement of Ca²⁺-dependent phosphorylation. *Nature (Lond.)*, 302:830–832, 1983.
- [2781] W.W. Metcalf, B.M. Griffin, R.M. Cicchillo, J. Gao, S.C. Janga, H.A. Cooke, B.T. Circello, B.S. Evans, W. Martens-Habbena, D.A. Stahl, and W.A. van der Donk. Synthesis of methylphosphonic acid by marine microbes: a source for methane in the aerobic ocean. *Science*, 337:1104–1107, 2012.
- [2782] J.G. Metz, M.R. Pollard, L. Anderson, T.R. Hayes, and M.W. Lassner. Purification of a jojoba embryo fatty acyl-coenzyme A reductase and expression of its cDNA in high erucic acid rapeseed. *Plant Physiol.*, 122:635–644, 2000.
- [2783] J. Meuer, H.C. Kuettner, J.K. Zhang, R. Hedderich, and W.W. Metcalf. Genetic analysis of the archaeon *Methanosarcina barkeri* Fusaro reveals a central role for Ech hydrogenase and ferredoxin in methanogenesis and carbon fixation. *Proc. Natl. Acad. Sci. USA*, 99:5632–5637, 2002.
- [2784] M. Meury, M. Knop, and F.P. Seebeck. Structural basis for copper-oxygen mediated C-H bond activation by the formylglycine-generating enzyme. *Angew. Chem. Int. Ed. Engl.*, 2017.

- [2785] A. Meyer, P. Cirpus, C. Ott, R. Schlecker, U. Zähringer, and E. Heinz. Biosynthesis of docosahexaenoic acid in *Euglena gracilis*: biochemical and molecular evidence for the involvement of a Δ^4 -fatty acyl group desaturase. *Biochemistry*, 42:9779–9788, 2003.
- [2786] G.B. Michaels, J.T. Peck Davidson, , and Jr. A flavin-sulfite adduct as an intermediate in the reaction catalyzed by adenyl sulfate reductase from *Desulfovibrio vulgaris*. *Biochem. Biophys. Res. Commun.*, 39:321–328, 1970.
- [2787] L.V. Michaelson, C.M. Lazarus, G. Griffiths, J.A. Napier, and A.K. Stobart. Isolation of a Δ^5 -fatty acid desaturase gene from *Mortierella alpina*. *J. Biol. Chem.*, 273:19055–19059, 1998.
- [2788] L.V. Michaelson, J.E. Markham, S. Zäuner, M. Matsumoto, M. Chen, E.B. Cahoon, and J.A. Napier. Identification of a cytochrome *b₅*-fusion desaturase responsible for the synthesis of triunsaturated sphingolipid long chain bases in the marine diatom *Thalassiosira pseudonana*. *Phytochemistry*, 90:50–55, 2013.
- [2789] L.V. Michaelson, S. Zäuner, J.E. Markham, R.P. Haslam, R. Desikan, S. Mugford, S. Albrecht, D. Warnecke, P. Sperling, E. Heinz, and J.A. Napier. Functional characterization of a higher plant sphingolipid Δ^4 -desaturase: defining the role of sphingosine and sphingosine-1-phosphate in *Arabidopsis*. *Plant Physiol.*, 149:487–498, 2009.
- [2790] J.L. Michalover and D.W. Ribbons. 3-Hydroxybenzoate 4-hydroxylase from *Pseudomonas testosteroni*. *Biochem. Biophys. Res. Commun.*, 55:888–896, 1973.
- [2791] W.P. Michalski and D.J.D. Nicholas. Molecular characterization of a copper-containing nitrite reductase from *Rhodospseudomonas sphaeroides* forma sp. Denitrificans. *Biochim. Biophys. Acta*, 828:130–137, 1985.
- [2792] C. Michel, G. van Echten-Deckert, J. Rother, K. Sandhoff, E., Merrill Wang, , and Jr. Characterization of ceramide synthesis. A dihydroceramide desaturase introduces the 4,5-*trans*-double bond of sphingosine at the level of dihydroceramide. *J. Biol. Chem.*, 272:22432–22437, 1997.
- [2793] G. Michel, A.W. Roszak, V. Sauvé, J. Maclean, A. Matte, J.R. Coggins, M. Cygler, and A.J. Laphorn. Structures of shikimate dehydrogenase AroE and its paralog YdiB. A common structural framework for different activities. *J. Biol. Chem.*, 278:19463–19472, 2003.
- [2794] E. Miclet, V. Stoven, P.A. Michels, F.R. Opperdoes, J.-Y. Lallemand, and F. Duffieux. NMR spectroscopic analysis of the first two steps of the pentose-phosphate pathway elucidates the role of 6-phosphogluconolactonase. *J. Biol. Chem.*, 276:34840–34846, 2001.
- [2795] T. Mieda, Y. Yabuta, M. Rapolu, T. Motoki, T. Takeda, K. Yoshimura, T. Ishikawa, and S. Shigeoka. Feedback inhibition of spinach L-galactose dehydrogenase by L-ascorbate. *Plant Cell Physiol.*, 45:1271–1279, 2004.
- [2796] M. Miethke, J. Hou, and M.A. Marahiel. The siderophore-interacting protein YqjH acts as a ferric reductase in different iron assimilation pathways of *Escherichia coli*. *Biochemistry*, 50:10951–10964, 2011.
- [2797] K. Miettinen, L. Dong, N. Navrot, T. Schneider, V. Burlat, J. Pollier, L. Woittiez, S. van der Krol, R. Lugan, T. Ilc, R. Verpoorte, K.M. Oksman-Caldentey, E. Martinoia, H. Bouwmeester, A. Goossens, J. Memelink, and D. Werck-Reichhart. The seco-iridoid pathway from *Catharanthus roseus*. *Nat. Commun.*, 5:3606–3606, 2014.
- [2798] L. Miguel and R. Meganathan. Electron donors and the quinone involved in dimethyl sulfoxide reduction in *Escherichia coli*. *Curr. Microbiol.*, 22:109–115, 1991.
- [2799] S.J. Mihalik, J.C. Morrell, D. Kim, K.A. Sacksteder, P.A. Watkins, and S.J. Gould. Identification of PAHX, a Refsum disease gene. *Nat. Genet.*, 17:185–189, 1997.
- [2800] S.J. Mihalik, A.M. Rainville, and P.A. Watkins. Phytanic acid α -oxidation in rat liver peroxisomes. Production of α -hydroxyphytanoyl-CoA and formate is enhanced by dioxxygenase cofactors. *Eur. J. Biochem.*, 232:545–551, 1995.
- [2801] M. Mihasan, C.B. Chiribau, T. Friedrich, V. Artenie, and R. Brandsch. An NAD(P)H-nicotine blue oxidoreductase is part of the nicotine regulon and may protect *Arthrobacter nicotinovorans* from oxidative stress during nicotine catabolism. *Appl. Environ. Microbiol.*, 73:2479–2485, 2007.
- [2802] B.N. Mijts, P.C. Lee, and C. Schmidt-Dannert. Identification of a carotenoid oxygenase synthesizing acyclic xanthophylls: combinatorial biosynthesis and directed evolution. *Chem. Biol.*, 12:453–460, 2005.

- [2803] B. Mikami and S. Ida. Purification and properties of ferredoxin-nitrate reductase from the cyanobacterium *Plectonema borganum*. *Biochim. Biophys. Acta*, 791:294–304, 1984.
- [2804] M.D. Mikkelsen, C.H. Hansen, U. Wittstock, and B.A. Halkier. Cytochrome P_{450} CYP79B2 from *Arabidopsis* catalyzes the conversion of tryptophan to indole-3-acetaldoxime, a precursor of indole glucosinolates and indole-3-acetic acid. *J. Biol. Chem.*, 275:33712–33717, 2000.
- [2805] D. Milbredt, E.P. Patallo, and K.H. van Pee. A tryptophan 6-halogenase and an amidotransferase are involved in thienodolin biosynthesis. *ChemBioChem*, 15:1011–1020, 2014.
- [2806] C.C. Milburn, H.J. Lambie, A. Theodossis, S.D. Bull, D.W. Hough, M.J. Danson, and G.L. Taylor. The structural basis of substrate promiscuity in glucose dehydrogenase from the hyperthermophilic archaeon *Sulfolobus solfataricus*. *J. Biol. Chem.*, 281:14796–14804, 2006.
- [2807] Z.D. Miles, R.M. McCarty, G. Molnar, and V. Bandarian. Discovery of epoxyqueuosine (oQ) reductase reveals parallels between halo-respiration and tRNA modification. *Proc. Natl. Acad. Sci. USA*, 108:7368–7372, 2011.
- [2808] D.L. Miller and V.W. Rodwell. Metabolism of basic amino acids in *Pseudomonas putida*. Intermediates in L-arginine catabolism. *J. Biol. Chem.*, 246:5053–5058, 1971.
- [2809] J.V. Miller, D.A. Estell, and R.A. Lazarus. Purification and characterization of 2,5-diketo-D-gluconate reductase from *Corynebacterium* sp. *J. Biol. Chem.*, 262:9016–9020, 1987.
- [2810] P.A. Miller, A. Saturnelli, J.H. Martin, L.A. Itscher, and N. Bohonos. A new family of tetracycline precursors. *N-demethylanhydrotetracyclines*. *Biochem. Biophys. Res. Commun.*, 16:285–291, 1964.
- [2811] P.A. Miller, N.O. Sjolander, S. Nalesnyk, N. Arnold, S. Johnson, A.P. Doerschuk, and J.R.D. McCormick. Cosynthetic factor I, a factor involved in hydrogen-transfer in *Streptomyces aureofaciens*. *J. Am. Chem. Soc.*, 82:5002–5003, 1960.
- [2812] R.E. Miller and E.R. Stadtman. Glutamate synthase from *Escherichia coli*. An iron-sulfide flavoprotein. *J. Biol. Chem.*, 247:7407–7419, 1972.
- [2813] R.W. Miller and R.R. Eady. Molybdenum and vanadium nitrogenases of *Azotobacter chroococcum*. Low temperature favours N_2 reduction by vanadium nitrogenase. *Biochem. J.*, 256:429–432, 1988.
- [2814] W.L. Miller, M.E. Kalafer, J.L. Gaylor, and C.V. Delwicke. Investigation of the component reactions of oxidative sterol demethylation. Study of the aerobic and anaerobic processes. *Biochemistry*, 6:2673–2678, 1967.
- [2815] W.L. Miller, C.Q. Wenzel, C. Daniels, S. Larocque, J.R. Brisson, and J.S. Lam. Biochemical characterization of WbpA, a UDP-*N*-acetyl-D-glucosamine 6-dehydrogenase involved in O-antigen biosynthesis in *Pseudomonas aeruginosa* PAO1. *J. Biol. Chem.*, 279:37551–37558, 2004.
- [2816] S. Milstien and S. Kaufman. Biosynthesis of tetrahydrobiopterin: conversion of dihydroneopterin triphosphate to tetrahydropterin intermediates. *Biochem. Biophys. Res. Commun.*, 128:1099–1107, 1985.
- [2817] T. Min, H. Kasahara, D.L. Bedgar, B. Youn, P.K. Lawrence, D.R. Gang, S.C. Halls, H. Park, J.L. Hilsenbeck, L.B. Davin, N.G. Lewis, and C. Kang. Crystal structures of pinoresinol-lariciresinol and phenylcoumaran benzylic ether reductases and their relationship to isoflavone reductases. *J. Biol. Chem.*, 278:50714–50723, 2003.
- [2818] J.M. Mingot, M.A. Penalva, and J.M. Fernandez-Canon. Disruption of *phacA*, an *Aspergillus nidulans* gene encoding a novel cytochrome P450 monooxygenase catalyzing phenylacetate 2-hydroxylation, results in penicillin overproduction. *J. Biol. Chem.*, 274:14545–14550, 1999.
- [2819] M. Miquel and J. Browse. *Arabidopsis* mutants deficient in polyunsaturated fatty acid synthesis. Biochemical and genetic characterization of a plant oleoyl-phosphatidylcholine desaturase. *J. Biol. Chem.*, 267:1502–1509, 1992.
- [2820] E. Misaka and K. Nakanishi. Studies on menadione reductase of bakers' yeast. I. Purification, crystallization and some properties. *J. Biochem. (Tokyo)*, 53:465–471, 1963.
- [2821] H. Misono, H. Hashimoto, H. Uehigashi, S. Nagata, and S. Nagasaki. Properties of L-lysine ϵ -dehydrogenase from *Agrobacterium tumefaciens*. *J. Biochem. (Tokyo)*, 105:1002–1008, 1989.

- [2822] H. Misono and S. Nagasaki. Occurrence of L-lysine ϵ -dehydrogenase in *Agrobacterium tumefaciens*. *J. Bacteriol.*, 150:398–401, 1982.
- [2823] H. Misono, H. Togawa, T. Yamamoto, and K. Soda. Occurrence of meso- α,ϵ -diaminopimelate dehydrogenase in *Bacillus sphaericus*. *Biochem. Biophys. Res. Commun.*, 72:89–93, 1976.
- [2824] H. Misono, H. Togawa, T. Yamamoto, and K. Soda. meso- α,ϵ -Diaminopimelate D-dehydrogenase: distribution and the reaction product. *J. Bacteriol.*, 137:22–27, 1979.
- [2825] H. Misono, H. Uehigashi, E. Morimoto, and S. Nagasaki. Purification and properties of L-lysine ϵ -dehydrogenase from *Agrobacterium tumefaciens*. *Agric. Biol. Chem.*, 49:2253–2255, 1985.
- [2826] M. Misset-Smits, P.W. Van Ophem, S. Sakuda, and J.A. Duine. Mycothiol, 1-*O*-(2'-[*N*-acetyl-L-cysteiny]amido-2'-deoxy- α -D-glucopyranosyl)-D-*myo*-inositol, is the factor of NAD/factor-dependent formaldehyde dehydrogenase. *FEBS Lett.*, 409:221–222, 1997.
- [2827] D. Missiakas, F. Schwager, and S. Raina. Identification and characterization of a new disulfide isomerase-like protein (DsbD) in *Escherichia coli*. *EMBO J.*, 14:3415–3424, 1995.
- [2828] A.G. Mitchell and C.E. Martin. Fah1p, a *Saccharomyces cerevisiae* cytochrome *b*₅ fusion protein, and its *Arabidopsis thaliana* homolog that lacks the cytochrome *b*₅ domain both function in the α -hydroxylation of sphingolipid-associated very long chain fatty acids. *J. Biol. Chem.*, 272:28281–28288, 1997.
- [2829] C. Mitoma. Studies on partially purified phenylalanine hydroxylase. *Arch. Biochem. Biophys.*, 60:476–484, 1956.
- [2830] C. Mitoma, H.S. Posner, H.C. Reitz, and S. Udenfriend. Enzymic hydroxylation of aromatic compounds. *Arch. Biochem. Biophys.*, 61:431–441, 1956.
- [2831] C. Mitoma and S. Udenfriend. Aryl-4-hydroxylase. *Methods Enzymol.*, 5:816–819, 1962.
- [2832] D. Mitra and C.S. Vaidyanathan. A new 4-nitrophenol 2-hydroxylase from a *Nocardia* sp. *Biochem. Int.*, 8:609–615, 1984.
- [2833] S. Mitsuhashi and B.D. Davis. Aromatic biosynthesis. XIII. Conversion of quinic acid to 5-dehydroquinic acid by quinic dehydrogenase. *Biochim. Biophys. Acta*, 15:268–280, 1954.
- [2834] K. Mitsukura, M. Suzuki, S. Shinoda, T. Kuramoto, T. Yoshida, and T. Nagasawa. Purification and characterization of a novel (*R*)-imine reductase from *Streptomyces* sp. GF3587. *Biosci. Biotechnol. Biochem.*, 75:1778–1782, 2011.
- [2835] J.R. Mitton, N.A. Scholan, and G.S. Boyd. The oxidation of cholesterol in rat liver sub-cellular particles. The cholesterol-7 α -hydroxylase enzyme system. *Eur. J. Biochem.*, 20:569–579, 1971.
- [2836] M.A. Mitz and R.L. Henrikson. Omega hydroxy fatty acid dehydrogenase. *Biochim. Biophys. Acta*, 46:45–50, 1961.
- [2837] T. Miyafusa, Y. Tanaka, M. Kuroda, T. Ohta, and K. Tsumoto. Expression, purification, crystallization and preliminary diffraction analysis of CapF, a capsular polysaccharide-synthesis enzyme from *Staphylococcus aureus*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 64:512–515, 2008.
- [2838] A. Miyanaga, S. Fujisawa, N. Furukawa, K. Arai, M. Nakajima, and H. Taguchi. The crystal structure of D-mandelate dehydrogenase reveals its distinct substrate and coenzyme recognition mechanisms from those of 2-ketopantoate reductase. *Biochem. Biophys. Res. Commun.*, 439:109–114, 2013.
- [2839] M. Miyata and T. Mori. Studies on denitrification. X. The "denitrifying enzyme" as a nitrite reductase and the electron donating system for denitrification. *J. Biochem. (Tokyo)*, 66:463–471, 1969.
- [2840] S. Miyata, Y. Suzuki, S. Kamisaka, and Y. Masuda. Indole-3-acetaldehyde oxidase of pea-seedlings. *Physiol. Plant.*, 51:402–406, 1981.
- [2841] K. Miyauchi, Y. Adachi, Y. Nagata, and M. Takagi. Cloning and sequencing of a novel meta-cleavage dioxygenase gene whose product is involved in degradation of γ -hexachlorocyclohexane in *Sphingomonas paucimobilis*. *J. Bacteriol.*, 181:6712–6719, 1999.
- [2842] K. Miyazaki. Bifunctional isocitrate-homoisocitrate dehydrogenase: a missing link in the evolution of β -decarboxylating dehydrogenase. *Biochem. Biophys. Res. Commun.*, 331:341–346, 2005.

- [2843] R. Miyazaki, T. Yamazaki, K. Yoshimatsu, K. Kojima, R. Asano, K. Sode, and W. Tsugawa. Elucidation of the intra- and inter-molecular electron transfer pathways of glucoside 3-dehydrogenase. *Bioelectrochemistry*, 122:115–122, 2018.
- [2844] M. Miyazawa and M. Shindo. Biotransformation of 1,8-cineole by human liver microsomes. *Nat. Prod. Lett.*, 15:49–53, 2001.
- [2845] M. Miyazawa, M. Shindo, and T. Shimada. Oxidation of 1,8-cineole, the monoterpene cyclic ether originated from *Eucalyptus polybractea*, by cytochrome P450 3A enzymes in rat and human liver microsomes. *Drug Metab. Dispos.*, 29:200–205, 2001.
- [2846] M. Miyazawa, M. Shindo, and T. Shimada. Roles of cytochrome P₄₅₀ 3A enzymes in the 2-hydroxylation of 1,4-cineole, a monoterpene cyclic ether, by rat and human liver microsomes. *Xenobiotica*, 31:713–723, 2001.
- [2847] T. Miyoshi, H. Sato, and T. Harada. Purification and characterization of 2-alkyne-1-ol dehydrogenase induced by 2-butene-1,4-diol in *Fusarium merismoides* B11. *Biochim. Biophys. Acta*, 358:231–239, 1974.
- [2848] M. Mizugaki, T. Nishimaki, T. Shiraishi, A. Kawaguchi, S. Okuda, and H. Yamanaka. Studies on the metabolism of unsaturated fatty acids. IX. Stereochemical studies of the reaction catalyzed by *trans*-2-enoyl-coenzyme A reductase of *Escherichia coli*. *J. Biochem. (Tokyo)*, 92:1649–1654, 1982.
- [2849] S. Mizushima. Purified D-glutamic-aspartic oxidase of *Aspergillus ustus*. *J. Gen. Appl. Microbiol.*, 3:233–239, 1957.
- [2850] S. Mizushima and K. Kitahara. Purification and properties of DPNH peroxidase in *Lactobacillus casei*. *J. Gen. Appl. Microbiol.*, 8:56–62, 1962.
- [2851] Y. Mizutani, A. Kihara, and Y. Igarashi. Identification of the human sphingolipid C4-hydroxylase, hDES2, and its up-regulation during keratinocyte differentiation. *FEBS Lett.*, 563:93–97, 2004.
- [2852] H. Mo, Y. Dai, S.S. Pochapsky, and T.C. Pochapsky. ¹H, ¹³C and ¹⁵N NMR assignments for a carbon monoxide generating metalloenzyme from *Klebsiella pneumoniae*. *J. Biomol. NMR*, 14:287–288, 1999.
- [2853] K. Möbius, R. Arias-Cartin, D. Breckau, A.L. Hännig, K. Riedmann, R. Biedendieck, S. Schroder, D. Becher, A. Magalon, J. Moser, M. Jahn, and D. Jahn. Heme biosynthesis is coupled to electron transport chains for energy generation. *Proc. Natl. Acad. Sci. USA*, 107:10436–10441, 2010.
- [2854] E. Mobus and E. Maser. Molecular cloning, overexpression, and characterization of steroid-inducible 3 α -hydroxysteroid dehydrogenase/carbonyl reductase from *Comamonas testosteroni*. A novel member of the short-chain dehydrogenase/reductase superfamily. *J. Biol. Chem.*, 273:30888–30896, 1998.
- [2855] F.F. Moebius, B.U. Fitzky, J.N. Lee, Y.K. Paik, and H. Glossmann. Molecular cloning and expression of the human Δ^7 -sterol reductase. *Proc. Natl. Acad. Sci. USA*, 95:1899–1902, 1998.
- [2856] D.J. Moffa, F.J. Lotspeich, and R.F. Krause. Preparation and properties of retinal-oxidizing enzyme from rat intestinal mucosa. *J. Biol. Chem.*, 245:439–447, 1970.
- [2857] M.E. Mohamed, A. Zaar, C. Ebenau-Jehle, and G. Fuchs. Reinvestigation of a new type of aerobic benzoate metabolism in the proteobacterium *Azoarcus evansii*. *J. Bacteriol.*, 183:1899–1908, 2001.
- [2858] S.K. Mohanty, C.L. Yu, S. Das, T.M. Louie, L. Gakhar, and M. Subramanian. Delineation of the caffeine C-8 oxidation pathway in *Pseudomonas* sp. strain CBB1 via characterization of a new trimethyluric acid monooxygenase and genes involved in trimethyluric acid metabolism. *J. Bacteriol.*, 194:3872–3882, 2012.
- [2859] S.G. Moinuddin, B. Youn, D.L. Bedgar, M.A. Costa, G.L. Helms, C. Kang, L.B. Davin, and N.G. Lewis. Secoisolariciresinol dehydrogenase: mode of catalysis and stereospecificity of hydride transfer in *Podophyllum peltatum*. *Org. Biomol. Chem.*, 4:808–816, 2006.
- [2860] J.W. Moir, L.C. Crossman, S. Spiro, and D.J. Richardson. The purification of ammonia monooxygenase from *Paracoccus denitrificans*. *FEBS Lett.*, 387:71–74, 1996.
- [2861] J.W. Moir, J.M. Wehrfritz, S. Spiro, and D.J. Richardson. The biochemical characterization of a novel non-haem-iron hydroxylamine oxidase from *Paracoccus denitrificans* GB17. *Biochem. J.*, 319:823–827, 1996.

- [2862] A.R. Moise, V. Kuksa, Y. Imanishi, and K. Palczewski. Identification of *all-trans*-retinol:*all-trans*-13,14-dihydroretinol saturase. *J. Biol. Chem.*, 279:50230–50242, 2004.
- [2863] D. Molenaar, M.E. van der Rest, and S. Petrovic. Biochemical and genetic characterization of the membrane-associated malate dehydrogenase (acceptor) from *Corynebacterium glutamicum*. *Eur. J. Biochem.*, 254:395–403, 1998.
- [2864] I.M Møller, , and J.M. Direct evidence for the presence of a rotenone-resistant NADH dehydrogenase on the inner surface of plant mitochondria. *Physiol. Plant.*, 54:267–274, 1982.
- [2865] D.T. Molowa, A.G. Shayne, and P.S. Guzelian. Purification and characterization of chlordecone reductase from human liver. *J. Biol. Chem.*, 261:12624–12627, 1986.
- [2866] C. Monder. α -Keto aldehyde dehydrogenase, an enzyme that catalyzes the enzymic oxidation of methylglyoxal to pyruvate. *J. Biol. Chem.*, 242:4603–4609, 1967.
- [2867] C. Monder and A. White. The 21-hydroxysteroid dehydrogenases of liver. A nicotinamide adenine dinucleotide phosphate dehydrogenase and two nicotinamide adenine dinucleotide dehydrogenases. *J. Biol. Chem.*, 240:71–77, 1965.
- [2868] L.C. Montemiglio, S. Gianni, B. Vallone, and C. Savino. Azole drugs trap cytochrome P450 EryK in alternative conformational states. *Biochemistry*, 49:9199–9206, 2010.
- [2869] G.M. Montero-Moran, M. Li, E. Rendon-Huerta, F. Jourdan, D.J. Lowe, A.W. Stumpff-Kane, M. Feig, C. Scazzocchio, and R.P. Hausinger. Purification and characterization of the Fe^{II}- and α -ketoglutarate-dependent xanthine hydroxylase from *Aspergillus nidulans*. *Biochemistry*, 46:5293–5304, 2007.
- [2870] B.L. Montgomery and J.C. Lagarias. Phytochrome ancestry: sensors of bilins and light. *Trends Plant Sci*, 7:357–366, 2002.
- [2871] M.J. Moonen, S.A. Synowsky, W.A. van den Berg, A.H. Westphal, A.J. Heck, R.H. van den Heuvel, M.W. Fraaije, and W.J. van Berkel. Hydroquinone dioxygenase from pseudomonas fluorescens ACB: a novel member of the family of nonheme-iron(II)-dependent dioxygenases. *J. Bacteriol.*, 190:5199–5209, 2008.
- [2872] D. Moonmangmee, Y. Fujii, H. Toyama, G. Theeragool, N. Lotong, K. Matsushita, and O. Adachi. Purification and characterization of membrane-bound quinoprotein cyclic alcohol dehydrogenase from *Gluconobacter frateurii* CHM 9. *Biosci. Biotechnol. Biochem.*, 65:2763–2772, 2001.
- [2873] E.C. Moore and R.B. Hurlbert. Regulation of mammalian deoxyribonucleotide biosynthesis by nucleotides as activators and inhibitors. *J. Biol. Chem.*, 241:4802–4809, 1966.
- [2874] E.C. Moore, P. Reichard, and L. Thelander. Enzymatic synthesis of deoxyribonucleotides. V. Purification and properties of thioredoxin reductase from *Escherichia coli* B. *J. Biol. Chem.*, 239:3445–3452, 1964.
- [2875] I.F. Moore, D.W. Hughes, and G.D. Wright. Tigecycline is modified by the flavin-dependent monooxygenase TetX. *Biochemistry*, 44:11829–11835, 2005.
- [2876] M.R. Moore, W.E. O'Brien, and L.G. Ljungdahl. Purification and characterization of nicotinamide adenine dinucleotide-dependent methylenetetrahydrofolate dehydrogenase from *Clostridium formicoaceticum*. *J. Biol. Chem.*, 249:5250–5253, 1974.
- [2877] S.J. Moore, A.D. Lawrence, R. Biedendieck, E. Deery, S. Frank, M.J. Howard, S.E. Rigby, and M.J. Warren. Elucidation of the anaerobic pathway for the corrin component of cobalamin (vitamin B₁₂). *Proc. Natl. Acad. Sci. USA*, 110:14906–14911, 2013.
- [2878] R. Morales, M.H. Charon, G. Kachalova, L. Serre, M. Medina, C. Gomez-Moreno, and M. Frey. A redox-dependent interaction between two electron-transfer partners involved in photosynthesis. *EMBO Rep.*, 1:271–276, 2000.
- [2879] M. Morant, K. Jørgensen, H. Schaller, F. Pinot, B.L. Møller, D. Werck-Reichhart, and S. Bak. CYP703 is an ancient cytochrome P450 in land plants catalyzing in-chain hydroxylation of lauric acid to provide building blocks for sporopollenin synthesis in pollen. *Plant Cell*, 19:1473–1487, 2007.
- [2880] R.A. Moreau and A.H.C. Huang. Oxidation of fatty alcohol in the cotyledons of jojoba seedlings. *Arch. Biochem. Biophys.*, 194:422–430, 1979.

- [2881] R.A. Moreau and A.H.C. Huang. Enzymes of wax ester catabolism in jojoba. *Methods Enzymol.*, 71:804–813, 1981.
- [2882] R.A. Moreau and P.K. Stumpf. Recent studies of the enzymic-synthesis of ricinoleic acid by developing castor beans. *Plant Physiol.*, 67:672–676, 1981.
- [2883] J.C. Moreno, H. Bikker, M.J. Kempers, A.S. van Trotsenburg, F. Baas, J.J. de Vijlder, T. Vulsma, and C. Ris-Stalpers. Inactivating mutations in the gene for thyroid oxidase 2 (THOX2) and congenital hypothyroidism. *N. Engl. J. Med.*, 347:95–102, 2002.
- [2884] L.R. Morgan, Weimorts Jr., Aubert D.M., and C.C. Oxidation of 3-hydroxyanthranilic acid by a soluble liver fraction from poikilothermic vertebrates. *Biochim. Biophys. Acta*, 100:393–402, 1965.
- [2885] H. Mori, T. Shibasaki, Y. Uozaki, K. Ochiai, and A. Ozaki. Detection of novel proline 3-hydroxylase activities in *Streptomyces* and *Bacillus* spp. by regio- and stereospecific hydroxylation of L-proline. *Appl. Environ. Microbiol.*, 62:1903–1907, 1996.
- [2886] H. Mori, T. Shibasaki, K. Yano, and A. Ozaki. Purification and cloning of a proline 3-hydroxylase, a novel enzyme which hydroxylates free L-proline to *cis*-3-hydroxy-L-proline. *J. Bacteriol.*, 179:5677–5683, 1997.
- [2887] N. Mori, B. Kawakami, Y. Tani, and H. Yamada. Purification and properties of dimethylglycine oxidase from *Cylindrocarpon didymum* M-1. *Agric. Biol. Chem.*, 44:1383–1389, 1980.
- [2888] N. Mori, M. Sano, Y. Tani, and H. Yamada. Purification and propertie of sarcosine oxidase from *Cylindrocarpon didymum* M-1. *Agric. Biol. Chem.*, 44:1391–1397, 1980.
- [2889] H. Morii, M. Nishihara, and Y. Koga. CTP:2,3-di-*O*-geranylgeranyl-*sn*-glycero-1-phosphate cytidyltransferase in the methanogenic archaeon *Methanothermobacter thermoautotrophicus*. *J. Biol. Chem.*, 275:36568–36574, 2000.
- [2890] T. Morikawa, M. Mizutani, N. Aoki, B. Watanabe, H. Saga, S. Saito, A. Oikawa, H. Suzuki, N. Sakurai, D. Shibata, A. Wadano, K. Sakata, and D. Ohta. Cytochrome P450 CYP710A encodes the sterol C-22 desaturase in *Arabidopsis* and tomato. *Plant Cell*, 18:1008–1022, 2006.
- [2891] T. Morinaga, H. Ashida, and K. Yoshida. Identification of two *scyllo*-inositol dehydrogenases in *Bacillus subtilis*. *Microbiology*, 156:1538–1546, 2010.
- [2892] H. Morise, O. Shimomura, F.H. Johnson, and J. Winant. Intermolecular energy transfer in the bioluminescent system of *Aequorea*. *Biochemistry*, 13:2656–2662, 1974.
- [2893] M. Morita, N. Hamada, K. Sakai, and Y. Watanabe. Purification and properties of secondary alcohol oxidase from a strain of *Pseudomonas*. *Agric. Biol. Chem.*, 43:1225–1235, 1979.
- [2894] M. Moritani. Demethylase. IV. Kinetics and reaction mechanism. *Hukuoka Acta Med.*, 43:651–658, 1952.
- [2895] M. Moritani. Demethylase. V. Specificity and its relation to amino acid oxidase. *Hukuoka Acta Med.*, 43:731–735, 1952.
- [2896] M. Moritani, T.-C. Tung, S. Fujii, H. Mito, N. Izumika, K. Kenmochi, and R. Hirohata. Specificity of rabbit kidney demethylase. *J. Biol. Chem.*, 209:485–492, 1954.
- [2897] H. Moriuchi, N. Koda, E. Okuda-Ashitaka, H. Daiyasu, K. Ogasawara, H. Toh, S. Ito, D.F. Woodward, and K. Watanabe. Molecular characterization of a novel type of prostamide/prostaglandin F synthase, belonging to the thioredoxin-like superfamily. *J. Biol. Chem.*, 283:792–801, 2008.
- [2898] P. Moroni, T. Buronfosse, C. Longin-Sauvageon, P. Delatour, and E. Benoit. Chiral sulfoxidation of albendazole by the flavin adenine dinucleotide-containing and cytochrome P450-dependent monooxygenases from rat liver microsomes. *Drug Metab. Dispos.*, 23:160–165, 1995.
- [2899] D.R. Morris and L.P. Hager. Chloroperoxidase. I. Isolation and properties of the crystalline glycoprotein. *J. Biol. Chem.*, 241:1763–1768, 1966.
- [2900] M. Morrison, H.B. Hamilton, and E. Stotz. The isolation and purification of lactoperoxidase by ion exchange chromatography. *J. Biol. Chem.*, 228:767–776, 1957.
- [2901] D. Morse and M. Mittag. Dinoflagellate luciferin-binding protein. *Methods Enzymol.*, 305:258–276, 2000.

- [2902] D. Morse, A.M. Pappenheimer, Hastings Jr., and J.W. Role of a luciferin-binding protein in the circadian bioluminescent reaction of *Gonyaulax polyedra*. *J. Biol. Chem.*, 264:11822–11826, 1989.
- [2903] M. Mortarino, A. Negri, G. Tedeschi, T. Simonic, S. Duga, H.G. Gassen, and S. Ronchi. L-Aspartate oxidase from *Escherichia coli*. I. Characterization of coenzyme binding and product inhibition. *Eur. J. Biochem.*, 239:418–426, 1996.
- [2904] P.N. Moschou, M. Sanmartin, A.H. Andriopoulou, E. Rojo, J.J. Sanchez-Serrano, and K.A. Roubelakis-Angelakis. Bridging the gap between plant and mammalian polyamine catabolism: a novel peroxisomal polyamine oxidase responsible for a full back-conversion pathway in *Arabidopsis*. *Plant Physiol.*, 147:1845–1857, 2008.
- [2905] T. Moses, J. Pollier, L. Almagro, D. Buyst, M. Van Montagu, M.A. Pedre no, J.C. Martins, J.M. Thevelein, and A. Goossens. Combinatorial biosynthesis of saponins and saponins in *Saccharomyces cerevisiae* using a C-16 α hydroxylase from *Bupleurum falcatum*. *Proc. Natl. Acad. Sci. USA*, 111:1634–1639, 2014.
- [2906] T. Moses, J. Pollier, A. Faizal, S. Apers, L. Pieters, J.M. Thevelein, D. Geelen, and A. Goossens. Unraveling the triterpenoid saponin biosynthesis of the African shrub *Maesa lanceolata*. *Mol. Plant*, 8:122–135, 2015.
- [2907] S.E. Moshier and D.J. Chapman. Biosynthetic studies on aromatic carotenoids. Biosynthesis of chlorobactene. *Biochem. J.*, 136:395–404, 1973.
- [2908] J. Moskovitz, V.K. Singh, J. Requena, B.J. Wilkinson, R.K. Jayaswal, and E.R. Stadtman. Purification and characterization of methionine sulfoxide reductases from mouse and *Staphylococcus aureus* and their substrate stereospecificity. *Biochem. Biophys. Res. Commun.*, 290:62–65, 2002.
- [2909] K. Moto, M.G. Suzuki, J.J. Hull, R. Kurata, S. Takahashi, M. Yamamoto, K. Okano, K. Imai, T. Ando, and S. Matsumoto. Involvement of a bifunctional fatty-acyl desaturase in the biosynthesis of the silkmoth, *Bombyx mori*, sex pheromone. *Proc. Natl. Acad. Sci. USA*, 101:8631–8636, 2004.
- [2910] Y. Motokawa and G. Kikuchi. Glycine metabolism by rat liver mitochondria. Reconstruction of the reversible glycine cleavage system with partially purified protein components. *Arch. Biochem. Biophys.*, 164:624–633, 1974.
- [2911] M.A. Moxley, J.J. Tanner, and D.F. Becker. Steady-state kinetic mechanism of the proline:ubiquinone oxidoreductase activity of proline utilization A (PutA) from *Escherichia coli*. *Arch. Biochem. Biophys.*, 516:113–120, 2011.
- [2912] H.S. Moyed and V. Williamson. Multiple 3-methyleneoxindole reductases of peas, differential inhibition by synthetic auxins. *J. Biol. Chem.*, 242:1075–1077, 1967.
- [2913] J. Moyle and M. Dixon. Purification of the isocitrate enzyme (triphosphopyridine nucleotide-linked isocitrate dehydrogenase-oxalosuccinate carboxylase). *Biochem. J.*, 63:548–552, 1956.
- [2914] G.C. Mueller and J.A. Miller. The reductive cleavage of 4-dimethylaminoazobenzene by rat liver: the intracellular distribution of the enzyme system and its requirements for triphosphopyridine nucleotide. *J. Biol. Chem.*, 180:1125–1136, 1949.
- [2915] G.C. Mueller and G. Rumney. Formation of 6 β -hydroxy and 6-keto derivatives of estradiol-16-C¹⁴ by mouse liver microsomes. *J. Am. Chem. Soc.*, 79:1004–1005, 1957.
- [2916] A.N. Mugo, J. Kobayashi, B. Mikami, Y. Yoshikane, T. Yagi, and K. Ohnishi. Crystal structure of 5-formyl-3-hydroxy-2-methylpyridine 4-carboxylic acid 5-dehydrogenase, an NAD(+)-dependent dismutase from *Mesorhizobium loti*. *Biochem. Biophys. Res. Commun.*, 456:35–40, 2015.
- [2917] I. Mukharji and R.B. Silverman. Purification of a vitamin K epoxide reductase that catalyzes conversion of vitamin K 2,3-epoxide to 3-hydroxy-2-methyl-3-phytyl-2,3-dihydronaphthoquinone. *Proc. Natl. Acad. Sci. USA*, 82:2713–2717, 1985.
- [2918] T. Mukherjee, Y. Zhang, S. Abdelwahed, S.E. Ealick, and T.P. Begley. Catalysis of a flavoenzyme-mediated amide hydrolysis. *J. Am. Chem. Soc.*, 132:5550–5551, 2010.
- [2919] R. Mukhopadhyay and B.P. Rosen. Arsenate reductases in prokaryotes and eukaryotes. *Environ Health Perspect*, 110 Suppl 5:745–748, 2002.

- [2920] S. Mukund and M.W.W. Adams. The novel tungsten-iron-sulfur protein of the hyperthermophilic archaeobacterium, *Pyrococcus furiosus*, is an aldehyde ferredoxin oxidoreductase - evidence for its participation in a unique glycolytic pathway. *J. Biol. Chem.*, 266:14208–14216, 1991.
- [2921] S. Mukund and M.W.W. Adams. Glyceraldehyde-3-phosphate ferredoxin oxidoreductase, a novel tungsten-containing enzyme with a potential glycolytic role in the hyperthermophilic archaeon *Pyrococcus furiosus*. *J. Biol. Chem.*, 270:8389–8392, 1995.
- [2922] F.H. Muller, T.M. Bandejas, T. Urich, M. Teixeira, C.M. Gomes, and A. Kletzin. Coupling of the pathway of sulphur oxidation to dioxygen reduction: characterization of a novel membrane-bound thiosulphate:quinone oxidoreductase. *Mol. Microbiol.*, 53:1147–1160, 2004.
- [2923] I. Muller, A. Kahnert, T. Pape, G.M. Sheldrick, W. Meyer-Klaucke, T. Dierks, M. Kertesz, and I. Uson. Crystal structure of the alkylsulfatase AtsK: insights into the catalytic mechanism of the Fe(II) α -ketoglutarate-dependent dioxygenase superfamily. *Biochemistry*, 43:3075–3088, 2004.
- [2924] M. Muller, M. Katzberg, M. Bertau, and W. Hummel. Highly efficient and stereoselective biosynthesis of (2*S*,5*S*)-hexanediol with a dehydrogenase from *Saccharomyces cerevisiae*. *Org. Biomol. Chem.*, 8:1540–1550, 2010.
- [2925] R. Müller, S. Haug, J. Eberspächer, and F. Lingens. Catechol-2,3-Dioxygenase aus Pyrazon-abbauenden Bakterien. *Hoppe-Seyler's Z. Physiol. Chem.*, 358:797–805, 1977.
- [2926] R. Müller, S. Schmitt, and F. Lingens. A novel non-heme iron-containing dioxygenase. Chloridazon-catechol dioxygenase from *Phenylobacterium immobilis* DSM 1986. *Eur. J. Biochem.*, 125:579–584, 1982.
- [2927] S. Muller and R.D. Walter. Purification and characterization of polyamine oxidase from *Ascaris suum*. *Biochem. J.*, 283:75–80, 1992.
- [2928] T.A. Muller, T. Fleischmann, J.R. van der Meer, and H.P. Kohler. Purification and characterization of two enantioselective α -ketoglutarate-dependent dioxygenases, RdpA and SdpA, from *Sphingomonas herbicidovorans* MH. *Appl. Environ. Microbiol.*, 72:4853–4861, 2006.
- [2929] T.A. Muller, M.I. Zavodszky, M. Feig, L.A. Kuhn, and R.P. Hausinger. Structural basis for the enantiospecificities of *R*- and *S*-specific phenoxypyruvate/ α -ketoglutarate dioxygenases. *Protein Sci.*, 15:1356–1368, 2006.
- [2930] E. Mulliez, M. Fontecave, J. Gaillard, and P. Reichard. An iron-sulfur center and a free radical in the active anaerobic ribonucleotide reductase of *Escherichia coli*. *J. Biol. Chem.*, 268:2296–2299, 1993.
- [2931] E. Mulliez, S. Ollagnier, M. Fontecave, R. Eliasson, and P. Reichard. Formate is the hydrogen donor for the anaerobic ribonucleotide reductase from *Escherichia coli*. *Proc. Natl. Acad. Sci. USA*, 92:8759–8762, 1995.
- [2932] E.F. Mulrooney, K.K. Poon, D.J. McNally, J.R. Brisson, and J.S. Lam. Biosynthesis of UDP-*N*-acetyl-L-fucosamine, a precursor to the biosynthesis of lipopolysaccharide in *Pseudomonas aeruginosa* serotype O11. *J. Biol. Chem.*, 280:19535–19542, 2005.
- [2933] J.W. Munos, S.J. Moon, S.O. Mansoorabadi, W. Chang, L. Hong, F. Yan, A. Liu, and H.W. Liu. Purification and characterization of the epoxidase catalyzing the formation of fosfomycin from *Pseudomonas syringae*. *Biochemistry*, 47:8726–8735, 2008.
- [2934] J.W. Munos, X. Pu, S.O. Mansoorabadi, H.J. Kim, and H.W. Liu. A secondary kinetic isotope effect study of the 1-deoxy-D-xylulose-5-phosphate reductoisomerase-catalyzed reaction: evidence for a retroaldol-aldol rearrangement. *J. Am. Chem. Soc.*, 131:2048–2049, 2009.
- [2935] A.W. Munro, M.A. Noble, L. Robledo, S.N. Daff, and S.K. Chapman. Determination of the redox properties of human NADPH-cytochrome P450 reductase. *Biochemistry*, 40:1956–1963, 2001.
- [2936] E. Murakami, U. Deppenmeier, and S.W. Ragsdale. Characterization of the intramolecular electron transfer pathway from 2-hydroxyphenazine to the heterodisulfide reductase from *Methanosarcina thermophila*. *J. Biol. Chem.*, 276:2432–2439, 2001.
- [2937] H. Murakami, K. Kita, and Y. Anraku. Cloning of *cybB*, the gene for cytochrome *b*₅₆₁ of *Escherichia coli* K12. *Mol. Gen. Genet.*, 198:1–6, 1984.

- [2938] H. Murakami, K. Kita, and Y. Anraku. Purification and properties of a diheme cytochrome b_{561} of the *Escherichia coli* respiratory chain. *J. Biol. Chem.*, 261:548–551, 1986.
- [2939] M. Murakami, K. Shibuya, T. Nakayama, T. Nishino, T. Yoshimura, and H. Hemmi. Geranylgeranyl reductase involved in the biosynthesis of archaeal membrane lipids in the hyperthermophilic archaeon *Archaeoglobus fulgidus*. *FEBS J.*, 274:805–814, 2007.
- [2940] N. Muraki, J. Nomata, K. Ebata, T. Mizoguchi, T. Shiba, H. Tamiaki, G. Kurisu, and Y. Fujita. X-ray crystal structure of the light-independent protochlorophyllide reductase. *Nature*, 465:110–114, 2010.
- [2941] H. Muramatsu, H. Mihara, R. Kakutani, M. Yasuda, M. Ueda, T. Kurihara, and N. Esaki. The putative malate/lactate dehydrogenase from *Pseudomonas putida* is an NADPH-dependent Δ^1 -piperidine-2-carboxylate/ Δ^1 -pyrroline-2-carboxylate reductase involved in the catabolism of D-lysine and D-proline. *J. Biol. Chem.*, 280:5329–5335, 2005.
- [2942] S. Murao and N. Tanaka. A new enzyme bilirubin oxidase produced by *Myrothecium verrucaria* MT-1. *Agric. Biol. Chem.*, 45:2383–2384, 1981.
- [2943] K. Murata, Y. Fukuda, M. Simosaka, K. Watanabe, T. Saikusa, and A. Kimura. Metabolism of 2-oxoaldehyde in yeasts. Purification and characterization of NADPH-dependent methylglyoxal-reducing enzyme from *Saccharomyces cerevisiae*. *Eur. J. Biochem.*, 151:631–636, 1985.
- [2944] C.D. Murphy, S.J. Moss, and D. O’Hagan. Isolation of an aldehyde dehydrogenase involved in the oxidation of fluoroacetaldehyde to fluoroacetate in *Streptomyces cattleya*. *Appl. Environ. Microbiol.*, 67:4919–4921, 2001.
- [2945] C.D. Murphy, C. Schaffrath, and D. O’Hagan. Fluorinated natural products: the biosynthesis of fluoroacetate and 4-fluorothreonine in *Streptomyces cattleya*. *Chemosphere*, 52:455–461, 2003.
- [2946] G.E. Murphy and G.J. Jensen. Electron cryotomography of the *E. coli* pyruvate and 2-oxoglutarate dehydrogenase complexes. *Structure*, 13:1765–1773, 2005.
- [2947] T. Murray-Stewart, Y. Wang, A. Goodwin, A. Hacker, A., Casero Meeker, , and Jr. Nuclear localization of human spermine oxidase isoforms - possible implications in drug response and disease etiology. *FEBS J.*, 275:2795–2806, 2008.
- [2948] A.S.N. Murthy, H.T. Keutmann, and B.A. Eipper. Further characterization of peptidylglycine α -amidating monooxygenase from bovine neurointermediate pituitary. *Mol. Endocrinol.*, 1:290–299, 1987.
- [2949] A.S.N. Murthy, R.E. Mains, and B.A. Eipper. Purification and characterization of peptidylglycine α -amidating monooxygenase from bovine neurointermediate pituitary. *J. Biol. Chem.*, 261:1815–1822, 1986.
- [2950] F.N. Musayev, M.L. Di Salvo, T.P. Ko, V. Schirch, and M.K. Safo. Structure and properties of recombinant human pyridoxine 5'-phosphate oxidase. *Protein Sci.*, 12:1455–1463, 2003.
- [2951] G. Mustafa, Y. Ishikawa, K. Kobayashi, C.T. Migita, M.D. Elias, S. Nakamura, S. Tagawa, and M. Yamada. Amino acid residues interacting with both the bound quinone and coenzyme, pyrroloquinoline quinone, in *Escherichia coli* membrane-bound glucose dehydrogenase. *J. Biol. Chem.*, 283:22215–22221, 2008.
- [2952] E. Muth, E. Morschel, and A. Klein. Purification and characterization of an 8-hydroxy-5-deazaflavin-reducing hydroge-nase from the archaeobacterium *Methanococcus voltae*. *Eur. J. Biochem.*, 169:571–577, 1987.
- [2953] J. Myllyharju and K.I. Kivirikko. Characterization of the iron- and 2-oxoglutarate-binding sites of human prolyl 4-hydroxylase. *EMBO J.*, 16:1173–1180, 1997.
- [2954] K. Nagahama, T. Ogawa, T. Fujii, M. Tazaki, S. Tanase, Y. Morino, and H. Fukuda. Purification and properties of an ethylene-forming enzyme from *Pseudomonas syringae* pv. *phaseolicola* PK2. *J. Gen. Microbiol.*, 137:2281–2286, 1991.
- [2955] J. Nagai and K. Bloch. Enzymatic desaturation of stearyl acyl carrier protein. *J. Biol. Chem.*, 243:4626–4633, 1968.
- [2956] S. Nagai and S. Black. A thiol-disulfide transhydrogenase from yeast. *J. Biol. Chem.*, 243:1942–1947, 1968.

- [2957] M. Nagano, Y. Ihara-Ohori, H. Imai, N. Inada, M. Fujimoto, N. Tsutsumi, H. Uchimiya, and M. Kawai-Yamada. Functional association of cell death suppressor, *Arabidopsis* Bax inhibitor-1, with fatty acid 2-hydroxylation through cytochrome *b5*. *Plant J.*, 58:122–134, 2009.
- [2958] M. Nagano, K. Takahara, M. Fujimoto, N. Tsutsumi, H. Uchimiya, and M. Kawai-Yamada. *Arabidopsis* sphingolipid fatty acid 2-hydroxylases (AtFAH1 and AtFAH2) are functionally differentiated in fatty acid 2-hydroxylation and stress responses. *Plant Physiol.*, 159:1138–1148, 2012.
- [2959] M. Nagano, H. Uchimiya, and M. Kawai-Yamada. Plant sphingolipid fatty acid 2-hydroxylases have unique characters unlike their animal and fungus counterparts. *Plant Signal. Behav.*, 7:1388–1392, 2012.
- [2960] S. Nagano, J.R. Cupp-Vickery, and T.L. Poulos. Crystal structures of the ferrous dioxygen complex of wild-type cytochrome P450*eryF* and its mutants, A245S and A245T: investigation of the proton transfer system in P450*eryF*. *J. Biol. Chem.*, 280:22102–22107, 2005.
- [2961] K. Nagao, N. Takizawa, and H. Kiyahara. Purification and properties of *cis*-phenanthrene dihydrodiol dehydrogenase in *Alcaligenes faecalis* AFK2. *Agric. Biol. Chem.*, 52:2621–2623, 1988.
- [2962] N. Nagata, R. Tanaka, S. Satoh, and A. Tanaka. Identification of a vinyl reductase gene for chlorophyll synthesis in *Arabidopsis thaliana* and implications for the evolution of *Prochlorococcus* species. *Plant Cell*, 17:233–240, 2005.
- [2963] T. Nagatsu, M. Levitt, and S. Udenfriend. Tyrosine hydroxylase. The initial step in norepinephrine biosynthesis. *J. Biol. Chem.*, 239:2910–2917, 1964.
- [2964] M. Nagel and J.R. Andreesen. Molybdenum-dependent degradation of nicotinic acid by *Bacillus* sp. DSM 2923. *FEMS Microbiol. Lett.*, 59:147–152, 1989.
- [2965] M. Nagel and J.R. Andreesen. Purification and characterization of the molybdoenzymes nicotinate dehydrogenase and 6-hydroxynicotinate dehydrogenase from *Bacillus niacini*. *Arch. Microbiol.*, 154:605–613, 1990.
- [2966] V. Nahoum, A. Gangloff, P. Legrand, D.W. Zhu, L. Cantin, B.S. Zhorov, V. Luu-The, F. Labrie, R. Breton, and S.X. Lin. Structure of the human 3 α -hydroxysteroid dehydrogenase type 3 in complex with testosterone and NADP at 1.25-Å resolution. *J. Biol. Chem.*, 276:42091–42098, 2001.
- [2967] U. Naik and S. Mavuinurve. α -Santonin 1,2-reductase and its role in the formation of dihydrosantonin and lumisantonin by *Pseudomonas cichorii* S. *Can. J. Microbiol.*, 33:658–662, 1987.
- [2968] P.M. Nair and C.S. Vaidyanathan. An indole oxidase isolated from the leaves of *Tecoma stans*. *Biochim. Biophys. Acta*, 81:496–506, 1964.
- [2969] P.M. Nair and C.S. Vaidyanathan. Isophenoxazine synthase. *Biochim. Biophys. Acta*, 81:507–516, 1964.
- [2970] P.M. Nair and L.C. Vining. Enzymic oxidation of catechol to diphenylenedioxide-2,3-quinone. *Arch. Biochem. Biophys.*, 106:422–427, 1964.
- [2971] P.M. Nair and L.C. Vining. Isophenoxazine synthase apoenzyme from *Pycnopus coccineus*. *Biochim. Biophys. Acta*, 96:318–327, 1965.
- [2972] S.R. Najle, A.D. Nusblat, C.B. Nudel, and A.D. Uttaro. The sterol-C7 desaturase from the ciliate *Tetrahymena thermophila* is a Rieske oxygenase, which is highly conserved in animals. *Mol. Biol. Evol.*, 30:1630–1643, 2013.
- [2973] H. Nakagawa and Y. Takeda. Phenol hydroxylase. *Biochim. Biophys. Acta*, 62:423–426, 1962.
- [2974] T. Nakagawa, R. Mitsui, A. Tani, K. Sasa, S. Tashiro, T. Iwama, T. Hayakawa, and K. Kawai. A catalytic role of XoxF1 as La³⁺-dependent methanol dehydrogenase in *Methylobacterium extorquens* strain AM1. *PLoS One*, 7:e50480–e50480, 2012.
- [2975] K. Nakahara, Y. Kitamura, Y. Yamagishi, H. Shoun, and T. Yasui. Levoglucosan dehydrogenase involved in the assimilation of levoglucosan in *Arthrobacter* sp. I-552. *Biosci. Biotechnol. Biochem.*, 58:2193–2196, 1994.
- [2976] C. Nakai, K. Hori, H. Kagamiyama, T. Nakazawa, and M. Nozaki. Purification, subunit structure, and partial amino acid sequence of metapyrocatechase. *J. Biol. Chem.*, 258:2916–2922, 1983.

- [2977] K. Nakajima, T. Hashimoto, and Y. Yamada. Two tropinone reductases with different stereospecificities are short-chain dehydrogenases evolved from a common ancestor. *Proc. Natl. Acad. Sci. USA*, 90:9591–9595, 1993.
- [2978] K. Nakamura and F. Bernheim. Studies on malonic semialdehyde dehydrogenase from *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta*, 50:147–152, 1961.
- [2979] L.K. Nakamura and D.D. Tyler. Induction of D-aldohexoside:cytochrome *c* oxidoreductase in *Agrobacterium tumefaciens*. *J. Bacteriol.*, 129:830–835, 1977.
- [2980] M. Nakamura, S. Maki, Y. Amano, Y. Ohkita, K. Niwa, T. Hirano, Y. Ohmiya, and H. Niwa. Firefly luciferase exhibits bimodal action depending on the luciferin chirality. *Biochem. Biophys. Res. Commun.*, 331:471–475, 2005.
- [2981] T. Nakamura. Purification and physico-chemical properties of laccase. *Biochim. Biophys. Acta*, 30:44–52, 1958.
- [2982] T. Nakamura. Stoichiometric studies on the action of laccase. *Biochim. Biophys. Acta*, 30:538–542, 1958.
- [2983] T. Nakamura, D. Komagata, S. Murakawa, K. Sakai, and A. Endo. Isolation and biosynthesis of 3 α -hydroxy-3,5-dihydromonacolin L. *J. Antibiot. (Tokyo)*, 43:1597–1600, 1990.
- [2984] T. Nakamura, T. Motoyama, S. Hirono, and I. Yamaguchi. Identification, characterization, and site-directed mutagenesis of recombinant pentachlorophenol 4-monooxygenase. *Biochim. Biophys. Acta*, 1700:151–159, 2004.
- [2985] W. Nakamura, S. Hosoda, and K. Hayashi. Purification and properties of rat liver glutathione peroxidase. *Biochim. Biophys. Acta*, 358:251–261, 1974.
- [2986] H. Nakanishi, H. Yamaguchi, T. Sasakuma, N.K. Nishizawa, and S. Mori. Two dioxygenase genes, *Ids3* and *Ids2*, from *Hordeum vulgare* are involved in the biosynthesis of mugineic acid family phytosiderophores. *Plant Mol. Biol.*, 44:199–207, 2000.
- [2987] M. Nakanishi, C. Yatome, N. Ishida, and Y. Kitade. Putative ACP phosphodiesterase gene (*acpD*) encodes an azoreductase. *J. Biol. Chem.*, 276:46394–46399, 2001.
- [2988] N. Nakanishi, H. Hasegawa, and S. Watabe. A new enzyme, NADPH-dihydropteridine reductase in bovine liver. *J. Biochem. (Tokyo)*, 81:681–685, 1977.
- [2989] H. Nakano, M. Wieser, B. Hurh, T. Kawai, T. Yoshida, T. Yamane, and T. Nagasawa. Purification, characterization and gene cloning of 6-hydroxynicotinate 3-monooxygenase from *Pseudomonas fluorescens* TN5. *Eur. J. Biochem.*, 260:120–126, 1999.
- [2990] M. Nakano and T.S. Danowski. Crystalline mammalian L-amino acid oxidase from rat kidney mitochondria. *J. Biol. Chem.*, 241:2075–2083, 1966.
- [2991] M. Nakano, E.J. Kelly, C. Wiek, H. Hanenberg, and A.E. Rettie. CYP4V2 in Bietti's crystalline dystrophy: ocular localization, metabolism of ω -3-polyunsaturated fatty acids, and functional deficit of the p.H331P variant. *Mol. Pharmacol.*, 82:679–686, 2012.
- [2992] M. Nakano, Y. Ushijima, M. Saga, Y. Tsutsumi, and H. Asami. Aliphatic L- α -hydroxyacid oxidase from rat livers: purification and properties. *Biochim. Biophys. Acta*, 167:9–22, 1968.
- [2993] Y. Nakano and K. Asada. Purification of ascorbate peroxidase in spinach chloroplasts; its inactivation in ascorbate-depleted medium and reactivation by monodehydroascorbate radical. *Plant Cell Physiol.*, 28:131–140, 1987.
- [2994] Y. Nakano, N. Suzuki, Y. Yoshida, T. Nezu, Y. Yamashita, and T. Koga. Thymidine diphosphate-6-deoxy-L-lyxo-4-hexulose reductase synthesizing dTDP-6-deoxy-L-talose from *Actinobacillus actinomycetemcomitans*. *J. Biol. Chem.*, 275:6806–6812, 2000.
- [2995] H. Nakao, S. Shinoda, and S. Yamamoto. Purification and some properties of carboxynorspermidine synthase participating in a novel biosynthetic pathway for norspermidine in *Vibrio alginolyticus*. *J. Gen. Microbiol.*, 137:1737–1742, 1991.
- [2996] K. Nakashima, H. Takei, O. Adachi, E. Shinagawa, and M. Ameyama. Determination of seminal fructose using D-fructose dehydrogenase. *Clin. Chim. Acta*, 151:307–310, 1985.

- [2997] T. Nakatsubo, M. Mizutani, S. Suzuki, T. Hattori, and T. Umezawa. Characterization of *Arabidopsis thaliana* pinoreosinol reductase, a new type of enzyme involved in lignan biosynthesis. *J. Biol. Chem.*, 283:15550–15557, 2008.
- [2998] W. Nakatsukasa and J.M. Akagi. Thiosulfate reductase isolated from *Desulfotomaculum nigrificans*. *J. Bacteriol.*, 98:429–433, 1969.
- [2999] T. Nakayama. Acetic acid bacteria. II. Intracellular distribution of enzymes related to acetic acid fermentation, and some properties of a highly purified triphosphopyridine nucleotide (TPN)-dependent aldehyde dehydrogenase. *J. Biochem. (Tokyo)*, 48:812–830, 1960.
- [3000] T. Nakayama, T. Sato, Y. Fukui, K. Yonekura-Sakakibara, H. Hayashi, Y. Tanaka, T. Kusumi, and T. Nishino. Specificity analysis and mechanism of aurone synthesis catalyzed by aureusidin synthase, a polyphenol oxidase homolog responsible for flower coloration. *FEBS Lett.*, 499:107–111, 2001.
- [3001] T. Nakayama, K. Yonekura-Sakakibara, T. Sato, S. Kikuchi, Y. Fukui, M. Fukuchi-Mizutani, T. Ueda, M. Nakao, Y. Tanaka, T. Kusumi, and T. Nishino. Aureusidin synthase: A polyphenol oxidase homolog responsible for flower coloration. *Science*, 290:1163–1166, 2000.
- [3002] T. Nakazawa, K. Hori, and O. Hayaishi. Studies on monooxygenases. V. Manifestation of amino acid oxidase activity by L-lysine monooxygenase. *J. Biol. Chem.*, 247:3439–3444, 1972.
- [3003] J.W. Nam and T.J. Kappock. Cloning and transcriptional analysis of *Crepis alpina* fatty acid desaturases affecting the biosynthesis of crepenynic acid. *J. Exp. Bot.*, 58:1421–1432, 2007.
- [3004] J.W. Nam, H. Nojiri, H. Noguchi, H. Uchimura, T. Yoshida, H. Habe, H. Yamane, and T. Omori. Purification and characterization of carbazole 1,9a-dioxygenase, a three-component dioxygenase system of *Pseudomonas resinovorans* strain CA10. *Appl. Environ. Microbiol.*, 68:5882–5890, 2002.
- [3005] Y. Namba, K. Yoshizawa, A. Ejima, T. Hayashi, and T. Kaneda. Coenzyme A- and nicotinamide adenine dinucleotide-dependent branched chain α -keto acid dehydrogenase. I. Purification and properties of the enzyme from *Bacillus subtilis*. *J. Biol. Chem.*, 244:4437–4447, 1969.
- [3006] S.C. Namboori and D.E. Graham. Acetamido sugar biosynthesis in the Euryarchaea. *J. Bacteriol.*, 190:2987–2996, 2008.
- [3007] S.C. Namboori and D.E. Graham. Enzymatic analysis of uridine diphosphate *N*-acetyl-D-glucosamine. *Anal. Biochem.*, 381:94–100, 2008.
- [3008] S. Nambu, T. Matsui, C.W. Goulding, S. Takahashi, and M. Ikeda-Saito. A new way to degrade heme: the *Mycobacterium tuberculosis* enzyme MhuD catalyzes heme degradation without generating CO. *J. Biol. Chem.*, 288:10101–10109, 2013.
- [3009] J.L. Napoli, R.T. Okita, B.S. Masters, and R.L. Horst. Identification of 25,26-dihydroxyvitamin D₃ as a rat renal 25-hydroxyvitamin D₃ metabolite. *Biochemistry*, 20:5865–5871, 1981.
- [3010] T. Nara, T. Hshimoto, and T. Aoki. Evolutionary implications of the mosaic pyrimidine-biosynthetic pathway in eukaryotes. *Gene*, 257:209–222, 2000.
- [3011] M. Nardini, G. Ricci, A.M. Caccuri, S.P. Solinas, L. Vesci, and D. Cavallini. Purification and characterization of a ketimine-reducing enzyme. *Eur. J. Biochem.*, 173:689–694, 1988.
- [3012] B. Nare, L. Hardy, and S.M. Beverley. The roles of pteridine reductase 1 and dihydrofolate reductase-thymidylate synthase in pteridine metabolism in the protozoan parasite *Leishmania major*. *J. Biol. Chem.*, 272:13883–13891, 1997.
- [3013] S.A. Narrod and W.B. Jakoby. Metabolism of ethanolamine. An ethanolamine oxidase. *J. Biol. Chem.*, 239:2189–2193, 1964.
- [3014] W. Nartey, S. Basak, N. Kamariah, M.S. Manimekalai, S. Robson, G. Wagner, B. Eisenhaber, F. Eisenhaber, and G. Gruber. NMR studies reveal a novel grab and release mechanism for efficient catalysis of the bacterial 2-Cys peroxiredoxin machinery. *FEBS J.*, 282:4620–4638, 2015.
- [3015] S. Narumiya and J.A. Salmon. Arachidonic acid-15-lipoxygenase from rabbit peritoneal polymorphonuclear leukocytes. *Methods Enzymol.*, 86:45–48, 1982.

- [3016] F.J. Ruiz-Due nas, S. Camarero, M. Pérez-Boada, M.J. Martínez, and A.T. Martínez. A new versatile peroxidase from *Pleurotus*. *Biochem. Soc. Trans.*, 29:116–122, 2001.
- [3017] F.J. Ruiz-Due nas, M.J. Martínez, and A.T. Martínez. Heterologous expression of *Pleurotus eryngii* peroxidase confirms its ability to oxidize Mn^{2+} and different aromatic substrates. *Appl. Environ. Microbiol.*, 65:4705–4707, 1999.
- [3018] F.J. Ruiz-Due nas, M.J. Martínez, and A.T. Martínez. Molecular characterization of a novel peroxidase isolated from the ligninolytic fungus *Pleurotus eryngii*. *Mol. Microbiol.*, 31:223–235, 1999.
- [3019] A. Nason. Nitrate reductases. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 587–607. Academic Press, New York, 2nd edition, 1963.
- [3020] A. Nason and H.J. Evans. Triphosphopyridine nucleotide-nitrate reductase in *Neurospora*. *J. Biol. Chem.*, 202:655–673, 1953.
- [3021] S. Nasu, F.D. Wicks, and R.K. Gholson. L-Aspartate oxidase, a newly discovered enzyme of *Escherichia coli*, is the B protein of quinolinate synthetase. *J. Biol. Chem.*, 257:626–632, 1982.
- [3022] C. Naumann, T. Hartmann, and D. Ober. Evolutionary recruitment of a flavin-dependent monooxygenase for the detoxification of host plant-acquired pyrrolizidine alkaloids in the alkaloid-defended arctiid moth *Tyria jacobaeae*. *Proc. Natl. Acad. Sci. USA*, 99:6085–6090, 2002.
- [3023] P. Naur, C.H. Hansen, S. Bak, B.G. Hansen, N.B. Jensen, H.L. Nielsen, and B.A. Halkier. CYP79B1 from *Sinapis alba* converts tryptophan to indole-3-acetaldoxime. *Arch. Biochem. Biophys.*, 409:235–241, 2003.
- [3024] P. Naur, B.L. Petersen, M.D. Mikkelsen, S. Bak, H. Rasmussen, C.E. Olsen, and B.A. Halkier. CYP83A1 and CYP83B1, two nonredundant cytochrome P450 enzymes metabolizing oximes in the biosynthesis of glucosinolates in *Arabidopsis*. *Plant Physiol.*, 133:63–72, 2003.
- [3025] R.M. Navarrete, J.A. Vara, and C.R. Hutchinson. Purification of an inducible L-valine dehydrogenase of *Streptomyces coelicolor* A3(2). *J. Gen. Microbiol.*, 136:273–281, 1990.
- [3026] F. Navarro, E. Martín-Figueroa, P. Candau, and F.J. Florencio. Ferredoxin-dependent iron-sulfur flavoprotein glutamate synthase (GlsF) from the cyanobacterium *Synechocystis* sp. PCC 6803: expression and assembly in *Escherichia coli*. *Arch. Biochem. Biophys.*, 379:267–276, 2000.
- [3027] I. Navarro, I. Font, G. Fabrias, and F. Camps. Stereospecificity of the (E)- and (Z)-11 myristoyl desaturases in the biosynthesis of *Spodoptera littoralis* sex pheromone. *J. Am. Chem. Soc.*, 119:11335–11336, 1997.
- [3028] A.S. Nayak, S. Sanjeev Kumar, M. Santosh Kumar, O. Anjaneya, and T.B. Karegoudar. A catabolic pathway for the degradation of chrysene by *Pseudoxanthomonas* sp. PNK-04. *FEMS Microbiol. Lett.*, 320:128–134, 2011.
- [3029] D.L. Neal and P.K. Kindel. D-Apiose reductase from *Aerobacter aerogenes*. *J. Bacteriol.*, 101:910–915, 1970.
- [3030] R.A. Neal. Bacterial metabolism of thiamine. 3. Metabolism of thiamine to 3-(2'-methyl-4'-amino-5'-pyrimidylmethyl)-4-methyl-thiazole-5-acetic acid (thiamine acetic acid) by a flavoprotein isolated from a soil microorganism. *J. Biol. Chem.*, 245:2599–2604, 1970.
- [3031] W.D. Neal and L.W. Parks. Sterol 24(28) methylene reductase in *Saccharomyces cerevisiae*. *J. Bacteriol.*, 129:1375–1378, 1977.
- [3032] K.H. Neilson and J.W. Hastings. Bacterial bioluminescence: its control and ecological significance. *Microbiol. Rev.*, 43:496–518, 1979.
- [3033] D.W. Nebert and H.V. Gelboin. Substrate-inducible microsomal aryl hydroxylase in mammalian cell culture. I. Assay and properties of induced enzyme. *J. Biol. Chem.*, 243:6242–6249, 1968.
- [3034] L. Ven Nederveelde, V. Verlinden, D. Philipp, and A. Debourg. Purification and characterization of yeast 3-methyl butanal reductases involved in the removal of wort carbonyls during fermentation. *Proc. 26th Congr.-Eur. Brew. Conv.*, pages 447–454, 1997.
- [3035] E. Negelein and H.-J. Wulff. Diphosphopyridinproteid ackkohol, acetaldehyd. *Biochem. Z.*, 293:351–389, 1937.

- [3036] F.B. Negm and W.H. Loescher. Detection and characterization of sorbitol dehydrogenase from apple callus tissue. *Plant Physiol.*, 64:69–73, 1979.
- [3037] F.B. Negm and W.H. Loescher. Characterization and partial-purification of aldose-6-phosphate reductase (alditol-6-phosphate-NADP 1-oxidoreductase) from apple leaves. *Plant Physiol.*, 67:139–142, 1981.
- [3038] E. Neidle, C. Hartnett, L.N. Ornston, A. Bairoch, M. Rekik, and S. Harayama. *cis*-Diol dehydrogenases encoded by the TOL pWW0 plasmid *xyiL* gene and the *Acinetobacter calcoaceticus* chromosomal *benD* gene are members of the short-chain alcohol dehydrogenase superfamily. *Eur. J. Biochem.*, 204:113–120, 1992.
- [3039] S.L. Neidleman, W.F. Amon, Geigert Jr., and J. Process for the production of fructose, 1981.
- [3040] M.J. Nelson and E.E. Snell. Enzymes of vitamin B₆ degradation. Purification and properties of 5-pyridoxic-acid oxygenase from *Arthrobacter* sp. *J. Biol. Chem.*, 261:15115–15120, 1986.
- [3041] A. Németh, Á. Svingor, M. Pócsik, J. Dobó, C Magyar, A. Szilaaagy, P. Gál, and P. Závodszy. Mirror image mutations reveal the significance of an intersubunit ion cluster in the stability of 3-isopropylmalate dehydrogenase. *FEBS Lett.*, 468:48–52, 2000.
- [3042] K.K. Nepal, T.J. Oh, and J.K. Sohng. Heterologous production of paromamine in *Streptomyces lividans* TK24 using kanamycin biosynthetic genes from *Streptomyces kanamyceticus* ATCC12853. *Mol. Cells*, 27:601–608, 2009.
- [3043] N.M. Nesbitt, C. Baleanu-Gogonea, R.M. Cicchillo, K. Goodson, D.F. Iwig, J.A. Broadwater, J.A. Haas, B.G. Fox, and S.J. Booker. Expression, purification, and physical characterization of *Escherichia coli* lipoyl(octanoyl)transferase. *Protein Expr. Purif.*, 39:269–282, 2005.
- [3044] H.A. Neufeld, L.F. Green, F.M. Latterell, and R.L. Weintraub. Thiooxidase, a new sulfhydryl-oxidizing enzyme from *Piricularia oryzae* and *Polyporus vesicolor*. *J. Biol. Chem.*, 232:1093–1099, 1958.
- [3045] W. Neuhauser, D. Haltrich, K.D. Kulbe, and B. Nidetzky. NAD(P)H-dependent aldose reductase from the xylose-assimilating yeast *Candida tenuis*. Isolation, characterization and biochemical properties of the enzyme. *Biochem. J.*, 326:683–692, 1997.
- [3046] H.Y. Neujahr and A. Gaal. Phenol hydroxylase from yeast. Purification and properties of the enzyme from *Trichosporon cutaneum*. *Eur. J. Biochem.*, 35:386–400, 1973.
- [3047] H.Y. Neujahr and A. Gaal. Phenol hydroxylase from yeast. Sulfhydryl groups in phenol hydroxylase from *Trichosporon cutaneum*. *Eur. J. Biochem.*, 58:351–357, 1975.
- [3048] A. Neumann, G. Wohlfarth, and G. Diekert. Purification and characterization of tetrachloroethene reductive dehalogenase from *Dehalospirillum multivorans*. *J. Biol. Chem.*, 271:16515–16519, 1996.
- [3049] C.S. Neumann, W. Jiang, J.R. Heemstra, Gontang Jr., Kolter E.A., Walsh R., and C.T. Biosynthesis of piperazic acid via N⁵-hydroxy-ornithine in *Kutzneria* spp. 744. *ChemBioChem*, 13:972–976, 2012.
- [3050] S. Neumann and H. Simon. On a non-pyridine nucleotide-dependent 2-oxoacid reductase of broad specificity from two *Proteus* species. *FEBS Lett.*, 167:29–32, 1985.
- [3051] S. Neumann, A. Wynen, H.G. Truper, and C. Dahl. Characterization of the *cys* gene locus from *Allochromatium vinosum* indicates an unusual sulfate assimilation pathway. *Mol. Biol. Rep.*, 27:27–33, 2000.
- [3052] D. Neusser, H. Schmidt, J. Spizek, J. Novotna, U. Peschke, S. Kaschabeck, P. Tichy, and W. Piepersberg. The genes *lmbB1* and *lmbB2* of *Streptomyces lincolnensis* encode enzymes involved in the conversion of L-tyrosine to propylproline during the biosynthesis of the antibiotic lincomycin A. *Arch. Microbiol.*, 169:322–332, 1998.
- [3053] A.M. Neville, J.C. Orr, and L.L. Engel. Δ^5 -3 β -Hydroxy steroid dehydrogenase activities of bovine adrenal cortex. *Biochem. J.*, 107:20–20, 1968.
- [3054] E.B. Newman, V. Kapoor, and R. Potter. Role of L-threonine dehydrogenase in the catabolism of threonine and synthesis of glycine by *Escherichia coli*. *J. Bacteriol.*, 126:1245–1249, 1976.
- [3055] L.M. Newman and L.P. Wackett. Purification and characterization of toluene 2-monoxygenase from *Burkholderia cepacia* G4. *Biochemistry*, 34:14066–14076, 1995.

- [3056] K. Ng, R. Ye, X.C. Wu, and S.L. Wong. Sorbitol dehydrogenase from *Bacillus subtilis*. Purification, characterization, and gene cloning. *J. Biol. Chem.*, 267:24989–24994, 1992.
- [3057] T.L. Ng, R. Rohac, A.J. Mitchell, A.K. Boal, and E.P. Balskus. An *N*-nitrosating metalloenzyme constructs the pharmacophore of streptozotocin. *Nature*, 566:94–99, 2019.
- [3058] D.T. Nguyen, J.C. Gopfert, N. Ikezawa, G. Macnevin, M. Kathiresan, J. Conrad, O. Spring, and D.K. Ro. Biochemical conservation and evolution of germacrene A oxidase in asteraceae. *J. Biol. Chem.*, 285:16588–16598, 2010.
- [3059] L.B. Nguyen, S. Shefer, G. Salen, J.Y. Chiang, and M. Patel. Cholesterol 7 α -hydroxylase activities from human and rat liver are modulated *in vitro* posttranslationally by phosphorylation/dephosphorylation. *Hepatology*, 24:1468–1474, 1996.
- [3060] L.B. Nguyen, S. Shefer, G. Salen, G. Ness, R.D. Tanaka, V. Packin, P. Thomas, V. Shore, and A. Batta. Purification of cholesterol 7 α -hydroxylase from human and rat liver and production of inhibiting polyclonal antibodies. *J. Biol. Chem.*, 265:4541–4546, 1990.
- [3061] D.J.D. Nicholas, A. Medina, and O.T.G. Jones. A nitrite reductase from *Neurospora crassa*. *Biochim. Biophys. Acta*, 37:468–476, 1960.
- [3062] D.J.D. Nicholas and A. Nason. Molybdenum and nitrate reductase. II. Molybdenum as a constituent of nitrate reductase. *J. Biol. Chem.*, 207:353–360, 1954.
- [3063] D.J.D. Nicholas and A. Nason. Diphosphopyridine nucleotide-nitrate reductase from *Escherichia coli*. *J. Bacteriol.*, 69:580–583, 1955.
- [3064] P. Nicholls and G.R. Schonbaum. Catalases. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 8, pages 147–225. Academic Press, New York, 2nd edition, 1963.
- [3065] B. Nidetzky, P. Mayr, P. Hadwiger, and A.E. Stutz. Binding energy and specificity in the catalytic mechanism of yeast aldose reductases. *Biochem. J.*, 344 Pt 1:101–107, 1999.
- [3066] T.D. Niehaus, S. Okada, T.P. Devarenne, D.S. Watt, V. Sviripa, and J. Chappell. Identification of unique mechanisms for triterpene biosynthesis in *Botryococcus braunii*. *Proc. Natl. Acad. Sci. USA*, 108:12260–12265, 2011.
- [3067] F.S. Nielsen, P.S. Andersen, and K.F. Jensen. The B form of dihydroorotate dehydrogenase from *Lactococcus lactis* consists of two different subunits, encoded by the *pyrDb* and *pyrK* genes, and contains FMN, FAD, and [FeS] redox centers. *J. Biol. Chem.*, 271:29359–29365, 1996.
- [3068] J.S. Nielsen and B.L. Møller. Cloning and expression of cytochrome P450 enzymes catalyzing the conversion of tyrosine to *p*-hydroxyphenylacetaldoxime in the biosynthesis of cyanogenic glucosides in *Triglochin maritima*. *Plant Physiol.*, 122:1311–1321, 2000.
- [3069] R. Niemetz, U. Altenschmidt, H. Herrmann, and G. Fuchs. Benzoyl-coenzyme-A 3-monooxygenase, a flavin-dependent hydroxylase. Purification, some properties and its role in aerobic benzoate oxidation via gentisate in a denitrifying bacterium. *Eur. J. Biochem.*, 227:161–168, 1995.
- [3070] J. Niemi, Y. Wang, K. Airas, K. Ylihonko, J. Hakala, and P. Mantsala. Characterization of aklavinone-11-hydroxylase from *Streptomyces purpurascens*. *Biochim. Biophys. Acta*, 1430:57–64, 1999.
- [3071] H.N. Nigg, J.A. Svoboda, M.J. Thompson, S.R. Dutky, J.N. Kaplanis, and W.E. Robbins. Ecdysone 20-hydroxylase from the midgut of the tobacco hornworm (*Manduca sexta* L.). *Experientia*, 32:438–439, 1976.
- [3072] M.W. Nirenberg and W.B. Jakoby. Enzymatic utilization of γ -hydroxybutyric acid. *J. Biol. Chem.*, 235:954–960, 1960.
- [3073] A. Nishida, T. Ishihara, and T. Hiroi. Studies on enzymes related to lignan biodegradation. *Baiomasu Henkan Keikaku Kenkyu Hokoku*, pages 38–59, 1987.
- [3074] H. Nishida, M. Nishiyama, N. Kobashi, T. Kosuge, T. Hoshino, and H. Yamane. A prokaryotic gene cluster involved in synthesis of lysine through the amino adipate pathway: a key to the evolution of amino acid biosynthesis. *Genome Res.*, 9:1175–1183, 1999.

- [3075] M. Nishihara and Y. Koga. *sn*-Glycerol-1-phosphate dehydrogenase in *Methanobacterium thermoautotrophicum*: key enzyme in biosynthesis of the enantiomeric glycerophosphate backbone of ether phospholipids of archaebacteria. *J. Biochem.*, 117:933–935, 1995.
- [3076] M. Nishihara and Y. Koga. Purification and properties of *sn*-glycerol-1-phosphate dehydrogenase from *Methanobacterium thermoautotrophicum*: characterization of the biosynthetic enzyme for the enantiomeric glycerophosphate backbone of ether polar lipids of Archaea. *J. Biochem.*, 122:572–576, 1997.
- [3077] M. Nishikimi, R. Fukuyama, S. Minoshima, N. Shimizu, and K. Yagi. Cloning and chromosomal mapping of the human nonfunctional gene for L-gulonolactone oxidase, the enzyme for L-ascorbic acid biosynthesis missing in man. *J. Biol. Chem.*, 269:13685–13688, 1994.
- [3078] Y. Nishimura and T. Eguchi. Biosynthesis of archaeal membrane lipids: digeranylgeranyl glycerophospholipid reductase of the thermoacidophilic archaeon *Thermoplasma acidophilum*. *J. Biochem.*, 139:1073–1081, 2006.
- [3079] Y. Nishimura and T. Eguchi. Stereochemistry of reduction in digeranylgeranyl glycerophospholipid reductase involved in the biosynthesis of archaeal membrane lipids from *Thermoplasma acidophilum*. *Bioorg. Chem.*, 35:276–283, 2007.
- [3080] H. Nishino, J. Nakaya, S. Nishi, T. Kurosawa, and T. Ishibashi. Temperature-induced differential kinetic properties between an initial burst and the following steady state in membrane-bound enzymes: studies on lathosterol 5-desaturase. *Arch. Biochem. Biophys.*, 339:298–304, 1997.
- [3081] T. Nishino, K. Okamoto, B.T. Eger, E.F. Pai, and T. Nishino. Mammalian xanthine oxidoreductase - mechanism of transition from xanthine dehydrogenase to xanthine oxidase. *FEBS J.*, 275:3278–3289, 2008.
- [3082] T. Nishio, A. Patel, Y. Wang, and P.C. Lau. Biotransformations catalyzed by cloned *p*-cymene monooxygenase from *Pseudomonas putida* F1. *Appl. Microbiol. Biotechnol.*, 55:321–325, 2001.
- [3083] Y. Nishiya and T. Imanaka. Cloning and sequencing of the sarcosine oxidase gene from *Arthrobacter* sp. TE1826. *J. Ferment. Bioeng.*, 75:239–244, 1993.
- [3084] Y. Nishiya and T. Imanaka. Alteration of substrate specificity and optimum pH of sarcosine oxidase by random and site-directed mutagenesis. *Appl. Environ. Microbiol.*, 60:4213–4215, 1994.
- [3085] Y. Nishiya and T. Imanaka. Purification and characterization of a novel glycine oxidase from *Bacillus subtilis*. *FEBS Lett.*, 438:263–266, 1998.
- [3086] T. Nishizawa, C.C. Aldrich, and D.H. Sherman. Molecular analysis of the rebeccamycin L-amino acid oxidase from *Lechevalieria aerocolonigenes* ATCC 39243. *J. Bacteriol.*, 187:2084–2092, 2005.
- [3087] T. Nishizawa, S. Gruschow, D.H. Jayamaha, C. Nishizawa-Harada, and D.H. Sherman. Enzymatic assembly of the bis-indole core of rebeccamycin. *J. Am. Chem. Soc.*, 128:724–725, 2006.
- [3088] Y. Nishizawa, T. Yabuki, E. Fukuda, and T. Wakagi. Gene expression and characterization of two 2-oxoacid:ferredoxin oxidoreductases from *Aeropyrum pernix* K1. *FEBS Lett.*, 579:2319–2322, 2005.
- [3089] B. Nisman. The Stickland reaction. *Bacteriol. Rev.*, 18:16–42, 1954.
- [3090] B. Nisman and J. Mager. Diphosphopyridine nucleotide and phosphate requirement for oxidation of amino-acids by cell-free extracts of obligate anaerobes. *Nature (Lond.)*, 169:243–244, 1952.
- [3091] S. Niwa, K. Takeda, M. Kosugi, E. Tsutsumi, T. Mogi, and K. Miki. Crystal structure of heme A synthase from *Bacillus subtilis*. *Proc. Natl. Acad. Sci. USA*, 115:11953–11957, 2018.
- [3092] B. Nobelmann and J.W. Lengeler. Sequence of the *gat* operon for galactitol utilization from a wild-type strain EC3132 of *Escherichia coli*. *Biochim. Biophys. Acta*, 1262:69–72, 1995.
- [3093] J. Nogales, A. Canales, J. Jiménez-Barbero, Pingarrón Serra B., García J. M., Díaz J. L., and E. Unravelling the gallic acid degradation pathway in bacteria: the *gal* cluster from *Pseudomonas putida*. *Mol. Microbiol.*, 79:359–374, 2011.
- [3094] J. Nogales, A. Canales, J. Jiménez-Barbero, J.L. García, and E. Díaz. Molecular characterization of the gallate dioxygenase from *Pseudomonas putida* KT2440. The prototype of a new subgroup of extradiol dioxygenases. *J. Biol. Chem.*, 280:35382–35390, 2005.

- [3095] K. Noge, M. Kato, N. Mori, M. Kataoka, C. Tanaka, Y. Yamasue, R. Nishida, and Y. Kuwahara. Geraniol dehydrogenase, the key enzyme in biosynthesis of the alarm pheromone, from the astigmatid mite *Carpoglyphus lactis* (Acari: Carpoglyphidae). *FEBS J.*, 275:2807–2817, 2008.
- [3096] M. Noguchi, T. Yoshida, and G. Kikuchi. Specific requirement of NADPH-cytochrome *c* reductase for the microsomal heme oxygenase reaction yielding biliverdin IX α . *FEBS Lett.*, 98:281–284, 1979.
- [3097] S. Noguchi, Z. Yamaizumi, T. Ohgi, T. Goto, Y. Nishimura, Y. Hirota, and S. Nishimura. Isolation of Q nucleoside precursor present in tRNA of an *E. coli* mutant and its characterization as 7-(cyano)-7-deazaguanosine. *Nucleic Acids Res.*, 5:4215–4223, 1978.
- [3098] E.A. Noltmann, C.J. Gubler, and S.A. Kuby. Glucose 6-phosphate dehydrogenase (Zwischenferment). I. Isolation of the crystalline enzyme from yeast. *J. Biol. Chem.*, 236:1225–1230, 1961.
- [3099] A. Noma, R. Ishitani, M. Kato, A. Nagao, O. Nureki, and T. Suzuki. Expanding role of the jumonji C domain as an RNA hydroxylase. *J. Biol. Chem.*, 285:34503–34507, 2010.
- [3100] J. Nomata, T. Mizoguchi, H. Tamiaki, and Y. Fujita. A second nitrogenase-like enzyme for bacteriochlorophyll biosynthesis: reconstitution of chlorophyllide *a* reductase with purified X-protein (BchX) and YZ-protein (BchY-BchZ) from *Rhodobacter capsulatus*. *J. Biol. Chem.*, 281:15021–15028, 2006.
- [3101] J. Nomata, T. Ogawa, M. Kitashima, K. Inoue, and Y. Fujita. NB-protein (BchN-BchB) of dark-operative protochlorophyllide reductase is the catalytic component containing oxygen-tolerant Fe-S clusters. *FEBS Lett.*, 582:1346–1350, 2008.
- [3102] T. Nomura, T. Kushiro, T. Yokota, Y. Kamiya, G.J. Bishop, and S. Yamaguchi. The last reaction producing brassinolide is catalyzed by cytochrome *P*-450s, CYP85A3 in tomato and CYP85A2 in *Arabidopsis*. *J. Biol. Chem.*, 280:17873–17879, 2005.
- [3103] S. Nørager, S. Arent, O. Björnberg, M. Ottosen, L. Lo Leggio, K.F. Jensen, and S. Larsen. *Lactococcus lactis* dihydroorotate dehydrogenase A mutants reveal important facets of the enzymatic function. *J. Biol. Chem.*, 278:28812–28822, 2003.
- [3104] R.C. Nordlie and H.J. Fromm. Ribitol dehydrogenase. II. Studies on the reaction mechanism. *J. Biol. Chem.*, 234:2523–2531, 1959.
- [3105] H. Nordlöv and S. Gatenbeck. Enzymatic synthesis of (+)- and (-)-bisdechlorogeodin with sulochrin oxidase from *Penicillium frequentans* and *Oospora sulphurea ochracea*. *Arch. Microbiol.*, 131:208–211, 1982.
- [3106] I. Nordlund, J. Powlowski, and V. Shingler. Complete nucleotide sequence and polypeptide analysis of multicomponent phenol hydroxylase from *Pseudomonas* sp. strain CF600. *J. Bacteriol.*, 172:6826–6833, 1990.
- [3107] A. Norin, P.W. Van Ophem, S.R. Piersma, B. Person, J.A. Duine, and H. Jornvall. Mycothiol-dependent formaldehyde dehydrogenase, a prokaryotic medium-chain dehydrogenase/reductase, phylogenetically links different eukaryotic alcohol dehydrogenase's - primary structure, conformational modelling and functional correlations. *Eur. J. Biochem.*, 248:282–289, 1997.
- [3108] A. Norin, S.R. Piersma, J.A. Duine, and H. Jornvall. Nicotinoprotein (NAD⁺-containing) alcohol dehydrogenase: structural relationships and functional interpretations. *Cell. Mol. Life Sci.*, 60:999–1006, 2003.
- [3109] C. Notheis, C. Drewke, and E. Leistner. Purification and characterization of the pyridoxol-5'-phosphate:oxygen oxidoreductase (deaminating) from *Escherichia coli*. *Biochim. Biophys. Acta*, 1247:265–271, 1995.
- [3110] L. Nover and M. Luckner. Mixed functional oxygenations during the biosynthesis of cyclopenin and cyclophenol, benzodiazepine alkaloids of *Penicillium cyclopium* westling. Incorporation of molecular oxygen and NIH-shift. *FEBS Lett.*, 3:292–296, 1969.
- [3111] N.J. Novick and M.E. Tyler. Partial purification and properties of an L-arabinose dehydrogenase from *Azospirillum brasilense*. *Can. J. Microbiol.*, 29:242–246, 1983.
- [3112] B. Nowak-Thompson, N. Chaney, J.S. Wing, S.J. Gould, and J.E. Loper. Characterization of the pyoluteorin biosynthetic gene cluster of *Pseudomonas fluorescens* Pf-5. *J. Bacteriol.*, 181:2166–2174, 1999.

- [3113] R.A. Mu noz Clares, L. González-Segura, C. Mújica-Jiménez, and L. Contreras-Diaz. Ligand-induced conformational changes of betaine aldehyde dehydrogenase from *Pseudomonas aeruginosa* and *Amaranthus hypochondriacus* L. leaves affecting the reactivity of the catalytic thiol. *Chem. Biol. Interact.*, pages 129–137, 2003.
- [3114] M. Nozaki, H. Kagamiyama, and O. Hayaishi. Metapyrocatechase. I. Purification, crystallization and some properties. *Biochem. Z.*, 338:582–590, 1963.
- [3115] T. Nubel, C. Klughammer, R. Huber, G. Hauska, and M. Schutz. Sulfide:quinone oxidoreductase in membranes of the hyperthermophilic bacterium *Aquifex aeolicus* (VF5). *Arch. Microbiol.*, 173:233–244, 2000.
- [3116] M.L. Nuccio, B.L. Russell, K.D. Nolte, B. Rathinasabapathi, D.A. Gage, and A.D. Hanson. Glycine betaine synthesis in transgenic tobacco expressing choline monoxygenase is limited by the endogenous choline supply. *Plant J.*, 16:101–110, 1998.
- [3117] M.L. Nuccio, B.L. Russell, K.D. Nolte, B. Rathinasabapathi, D.A. Gage, and A.D. Hanson. The endogenous choline supply limits glycine betaine synthesis in transgenic tobacco expressing choline. *Plant J.*, 16:487–496, 1998.
- [3118] D.H. Nugteren. Arachidonate lipoxygenase in blood platelets. *Biochim. Biophys. Acta*, 380:299–307, 1975.
- [3119] A.P. Nygaard. D-(-)-Lactate cytochrome *c* reductase, a flavoprotein from yeast. *J. Biol. Chem.*, 236:920–925, 1961.
- [3120] A.P. Nygaard. Lactate dehydrogenases of yeast. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 557–565. Academic Press, New York, 2nd edition, 1963.
- [3121] K. De Nys, E. Meyhi, G.P. Mannaerts, M. Franssen, and P.P. Van Veldhoven. Characterisation of human peroxisomal 2,4-dienoyl-CoA reductase. *Biochim. Biophys. Acta*, 1533:66–72, 2001.
- [3122] K. Ôba, S. Ishikawa, M. Nishikawa, H. Mizuno, and T. Yamamoto. Purification and properties of L-galactono- γ -lactone dehydrogenase, a key enzyme for ascorbic acid biosynthesis, from sweet potato roots. *J. Biochem. (Tokyo)*, 117:120–124, 1995.
- [3123] H. Obata, M. Uebayashi, and T. Kaneda. Purification and properties of 4-hydroxycyclohexanecarboxylate dehydrogenase from *Corynebacterium cyclohexanicum*. *Eur. J. Biochem.*, 174:451–458, 1988.
- [3124] V. Oberhauser, O. Voolstra, A. Bangert, J. von Lintig, and K. Vogt. NinaB combines carotenoid oxygenase and retinoid isomerase activity in a single polypeptide. *Proc. Natl. Acad. Sci. USA*, 105:19000–19005, 2008.
- [3125] S. Obermaier and M. Muller. Ibotenic acid biosynthesis in the fly agaric is initiated by glutamate hydroxylation. *Angew. Chem. Int. Ed. Engl.*, 59:12432–12435, 2020.
- [3126] M.M. O'Brien, P.J. Schofield, and M.R. Edwards. Polyol-pathway enzymes of human brain. Partial purification and properties of sorbitol dehydrogenase. *Biochem. J.*, 211:81–90, 1983.
- [3127] S.J. O'Brien and R.J. MacIntyre. The α -glycerophosphate cycle in *Drosophila melanogaster*. I. Biochemical and developmental aspects. *Biochem. Genet.*, 7:141–161, 1972.
- [3128] T.A. O'Brien, H.L. Schrock, P. Russell, R. Blake, Gennis 2nd, and R.B. Preparation of *Escherichia coli* pyruvate oxidase utilizing a thiamine pyrophosphate affinity column. *Biochim. Biophys. Acta*, 452:13–29, 1976.
- [3129] S. Ochoa. Enzymic mechanisms in the citric acid cycle. *Adv. Enzymol. Relat. Subj. Biochem.*, 15:183–270, 1954.
- [3130] S. Ochoa, A.H. Mehler, and A. Kornberg. Biosynthesis of dicarboxylic acids by carbon dioxide fixation. I. Isolation and properties of an enzyme from pigeon liver catalyzing the reversible oxidative decarboxylation of *l*-malic acid. *J. Biol. Chem.*, 174:979–1000, 1948.
- [3131] R.J. O'Connor and H. Halvorson. The substrate specificity of L-alanine dehydrogenase. *Biochim. Biophys. Acta*, 48:47–55, 1961.
- [3132] F. Oehme, P. Ellinghaus, P. Kolkhof, T.J. Smith, S. Ramakrishnan, J. Hutter, M. Schramm, and I. Flamme. Overexpression of PH-4, a novel putative proline 4-hydroxylase, modulates activity of hypoxia-inducible transcription factors. *Biochem. Biophys. Res. Commun.*, 296:343–349, 2002.
- [3133] M. Ogata, K. Arihara, and T. Yagi. D-Lactate dehydrogenase of *Desulfovibrio vulgaris*. *J. Biochem. (Tokyo)*, 89:1423–1431, 1981.

- [3134] T. Ogishima, S. Deguchi, and K. Okuda. Purification and characterization of cholesterol 7 α -hydroxylase from rat liver microsomes. *J. Biol. Chem.*, 262:7646–7650, 1987.
- [3135] H.J. Ogola, T. Kamiike, N. Hashimoto, H. Ashida, T. Ishikawa, H. Shibata, and Y. Sawa. Molecular characterization of a novel peroxidase from the cyanobacterium *Anabaena* sp. strain PCC 7120. *Appl. Environ. Microbiol.*, 75:7509–7518, 2009.
- [3136] M.M. Oh, E.E. Carey, and C.B. Rajashekar. Environmental stresses induce health-promoting phytochemicals in lettuce. *Plant Physiol. Biochem.*, 47:578–583, 2009.
- [3137] H. Ohara, R.A. Russell, K. Uchida, and H. Kondo. Purification and characterization of NAD-specific 6-phosphogluconate dehydrogenase from *Leuconostoc lactis* SHO-54. *J. Biosci. Bioeng.*, 98:126–128, 2004.
- [3138] N. Ohishi, T. Izumi, M. Minami, S. Kitamura, Y. Seyama, S. Ohkawa, S. Terao, H. Yotsumoto, F. Takaku, and T. Shimizu. Leukotriene A₄ hydrolase in the human lung. Inactivation of the enzyme with leukotriene A₄ isomers. *J. Biol. Chem.*, 262:10200–10205, 1987.
- [3139] S. Ohki, N. Ogino, S. Yamamoto, and O. Hayaishi. Prostaglandin hydroperoxidase, an integral part of prostaglandin endoperoxide synthetase from bovine vesicular gland microsomes. *J. Biol. Chem.*, 254:829–836, 1979.
- [3140] M. Ohmura, A. Hara, M. Nakagawa, and H. Sawada. Demonstration of 3 α (17 β)-hydroxysteroid dehydrogenase distinct from 3 α -hydroxysteroid dehydrogenase in hamster liver. *Biochem. J.*, 266:583–589, 1990.
- [3141] T. Ohnishi, B. Godza, B. Watanabe, S. Fujioka, L. Hategan, K. Ide, K. Shibata, T. Yokota, M. Szekeres, and M. Mizutani. CYP90A1/CPD, a brassinosteroid biosynthetic cytochrome P450 of *Arabidopsis*, catalyzes C-3 oxidation. *J. Biol. Chem.*, 287:31551–31560, 2012.
- [3142] T. Ohnishi, A.M. Szatmari, B. Watanabe, S. Fujita, S. Bancos, C. Koncz, M. Lafos, K. Shibata, T. Yokota, K. Sakata, M. Szekeres, and M. Mizutani. C-23 hydroxylation by *Arabidopsis* CYP90C1 and CYP90D1 reveals a novel shortcut in brassinosteroid biosynthesis. *Plant Cell*, 18:3275–3288, 2006.
- [3143] T. Ohnishi, B. Watanabe, K. Sakata, and M. Mizutani. CYP724B2 and CYP90B3 function in the early C-22 hydroxylation steps of brassinosteroid biosynthetic pathway in tomato. *Biosci. Biotechnol. Biochem.*, 70:2071–2080, 2006.
- [3144] Y. Ohno, S. Nakamichi, A. Ohkuni, N. Kamiyama, A. Naoe, H. Tsujimura, U. Yokose, K. Sugiura, J. Ishikawa, M. Akiyama, and A. Kihara. Essential role of the cytochrome P450 CYP4F22 in the production of acylceramide, the key lipid for skin permeability barrier formation. *Proc. Natl. Acad. Sci. USA*, 112:7707–7712, 2015.
- [3145] T. Ohshiro, Y. Ishii, T. Matsubara, K. Ueda, Y. Izumi, K. Kino, and K. Kirimura. Dibenzothiophene desulfurizing enzymes from moderately thermophilic bacterium *Bacillus subtilis* WU-S₂B: purification, characterization and overexpression. *J. Biosci. Bioeng.*, 100:266–273, 2005.
- [3146] T. Ohshiro, T. Kojima, K. Torii, H. Kawasoe, and Y. Izumi. Purification and characterization of dibenzothiophene (DBT) sulfone monooxygenase, an enzyme involved in DBT desulfurization, from *Rhodococcus erythropolis* D-1. *J. Biosci. Bioeng.*, 88:610–616, 1999.
- [3147] T. Ohshiro, J. Littlechild, E. Garcia-Rodriguez, M.N. Isupov, Y. Iida, T. Kobayashi, and Y. Izumi. Modification of halogen specificity of a vanadium-dependent bromoperoxidase. *Protein Sci.*, 13:1566–1571, 2004.
- [3148] Y. Ohta and D.W. Ribbons. Bacterial metabolism of resorcinolic compounds: purification and properties of orcinol hydroxylase and resorcinol hydroxylase from *Pseudomonas putida* ORC. *Eur. J. Biochem.*, 61:259–269, 1976.
- [3149] S. Ohtaki, H. Nakagawa, M. Nakamura, and I. Yamazaki. One- and two-electron oxidations of tyrosine, monoiodotyrosine, and diiodotyrosine catalyzed by hog thyroid peroxidase. *J. Biol. Chem.*, 257:13398–13403, 1982.
- [3150] K. Ojima, J. Breitenbach, H. Visser, Y. Setoguchi, K. Tabata, T. Hoshino, J. van den Berg, and G. Sandmann. Cloning of the astaxanthin synthase gene from *Xanthophyllomyces dendrorhous* (*Phaffia rhodozyma*) and its assignment as a β -carotene 3-hydroxylase/4-ketolase. *Mol. Genet. Genomics*, 275:148–158, 2006.
- [3151] T. Oka and F.J. Simpson. Quercetinase, a dioxygenase containing copper. *Biochem. Biophys. Res. Commun.*, 43:1–5, 1971.

- [3152] K. Okada and T. Hase. Cyanobacterial non-mevalonate pathway: (*E*)-4-hydroxy-3-methylbut-2-enyl diphosphate synthase interacts with ferredoxin in *Thermosynechococcus elongatus* BP-1. *J. Biol. Chem.*, 280:20672–20679, 2005.
- [3153] N. Okada, S. Noguchi, S. Nishimura, T. Ohgi, T. Goto, P.F. Crain, and J.A. McCloskey. Structure determination of a nucleoside Q precursor isolated from *E. coli* tRNA: 7-(aminomethyl)-7-deazaguanosine. *Nucleic Acids Res.*, 5:2289–2296, 1978.
- [3154] N. Okada, A. Shinmyo, H. Okada, and Y. Yamada. Purification and characterization of (*S*)-tetrahydroberberine oxidase from cultured *Coptis japonica* cells. *Phytochemistry*, 27:979–982, 1988.
- [3155] H. Okamoto and O. Hayaishi. Flavin adenine dinucleotide requirement for kynurenine hydroxylase of rat liver mitochondria. *Biochem. Biophys. Res. Commun.*, 29:394–399, 1967.
- [3156] K. Okamoto. Enzymic studies on the formation of 5-ketogluconic acid by *Acetobacter suboxydans*. II. 5-Ketogluconate reductase. *J. Biochem. (Tokyo)*, 53:448–448, 1963.
- [3157] S. Okamoto, F. Yu, H. Harada, T. Okajima, J. Hattan, N. Misawa, and R. Utsumi. A short-chain dehydrogenase involved in terpene metabolism from *Zingiber zerumbet*. *FEBS J.*, 278:2892–2900, 2011.
- [3158] T. Okamura, H. Noda, S. Fukuda, and M. Ohsugi. Aspartate dehydrogenase in vitamin B₁₂-producing *Klebsiella pneumoniae* IFO 13541. *J. Nutr. Sci. Vitaminol.*, 44:483–490, 1998.
- [3159] K. Okamura-Ikeda, K. Fujiwara, and Y. Motokawa. Purification and characterization of chicken liver T-protein, a component of the glycine cleavage system. *J. Biol. Chem.*, 257:135–139, 1982.
- [3160] K. Okamura-Ikeda, Y. Ohmura, K. Fujiwara, and Y. Motokawa. Cloning and nucleotide sequence of the *gcv* operon encoding the *Escherichia coli* glycine-cleavage system. *Eur. J. Biochem.*, 216:539–548, 1993.
- [3161] T. Okayasu, M. Nagao, T. Ishibashi, and Y. Imai. Purification and partial characterization of linoleoyl-CoA desaturase from rat liver microsomes. *Arch. Biochem. Biophys.*, 206:21–28, 1981.
- [3162] S. Okazaki, M. Sugawara, and K. Minamisawa. *Bradyrhizobium elkanii* *rtxC* gene is required for expression of symbiotic phenotypes in the final step of rhizobitoxine biosynthesis. *Appl. Environ. Microbiol.*, 70:535–541, 2004.
- [3163] A. Okuda and K. Okuda. Purification and characterization of Δ^4 -3-ketosteroid 5 β -reductase. *J. Biol. Chem.*, 259:7519–7524, 1984.
- [3164] K. Okuda and N. Hoshita. Oxidation of 5 β -cholestane-3 α ,7 α ,12 α -triol by rat-liver mitochondria. *Biochim. Biophys. Acta*, 164:381–388, 1968.
- [3165] N. Okumura, N.K. Nishizawa, Y. Umehara, T. Ohata, H. Nakanishi, T. Yamaguchi, M., and Mori. S. A dioxygenase gene (*Ids2*) expressed under iron deficiency conditions in the roots of *Hordeum vulgare*. *Plant Mol. Biol.*, 25:705–719, 1994.
- [3166] K. Olavarria, D. Valdes, and R. Cabrera. The cofactor preference of glucose-6-phosphate dehydrogenase from *Escherichia coli* – modeling the physiological production of reduced cofactors. *FEBS J.*, 279:2296–2309, 2012.
- [3167] S.E. O’Leary, K.A. Hicks, S.E. Ealick, and T.P. Begley. Biochemical characterization of the HpxO enzyme from *Klebsiella pneumoniae*, a novel FAD-dependent urate oxidase. *Biochemistry*, 48:3033–3035, 2009.
- [3168] C. Olive, M.E. Geroch, and H.R. Levy. Glucose 6-phosphate dehydrogenase from *Leuconostoc mesenteroides*. Kinetic studies. *J. Biol. Chem.*, 246:2047–2057, 1971.
- [3169] T.F. Oliveira, C. Vonrhein, P.M. Matias, S.S. Venceslau, I.A. Pereira, and M. Archer. The crystal structure of *Desulfovibrio vulgaris* dissimilatory sulfite reductase bound to DsrC provides novel insights into the mechanism of sulfate respiration. *J. Biol. Chem.*, 283:34141–34149, 2008.
- [3170] E.H. Oliw and H. Sprecher. Metabolism of polyunsaturated fatty acids by an (n-6)-lipoxygenase associated with human ejaculates. *Biochim. Biophys. Acta*, 1002:283–291, 1989.
- [3171] E.H. Oliw, C. Su, T. Skogstrom, and G. Benthin. Analysis of novel hydroperoxides and other metabolites of oleic, linoleic and linolenic acids by liquid chromatography-mass spectrometry with ion trap MSn. *Lipids*, 33:843–852, 1998.

- [3172] S. Ollagnier, E. Mulliez, P.P. Schmidt, R. Eliasson, J. Gaillard, C. Deronzier, T. Bergman, A. Graslund, P. Reichard, and M. Fontecave. Activation of the anaerobic ribonucleotide reductase from *Escherichia coli*. The essential role of the iron-sulfur center for S-adenosylmethionine reduction. *J. Biol. Chem.*, 272:24216–24223, 1997.
- [3173] A. Olomucki, D.B. Pho, R. Lebar, L. Delcambe, and N.V. Thoai. Arginine oxygénase décarboxylante. V. Purification et nature flavinique. *Biochim. Biophys. Acta*, 151:353–366, 1968.
- [3174] A. Olry, S. Boschi-Muller, M. Marraud, S. Sanglier-Cianferani, A. Van Dorsselaar, and G. Branlant. Characterization of the methionine sulfoxide reductase activities of PILB, a probable virulence factor from *Neisseria meningitidis*. *J. Biol. Chem.*, 277:12016–12022, 2002.
- [3175] P.J. Olsiewski, G.J. Kaczorowski, and C. Walsh. Purification and properties of D-amino acid dehydrogenase, an inducible membrane-bound iron-sulfur flavoenzyme from *Escherichia coli* B. *J. Biol. Chem.*, 255:4487–4494, 1980.
- [3176] J.A. Olson and C.B. Anfinsen. The crystallization and characterization of L-glutamic acid dehydrogenase. *J. Biol. Chem.*, 197:67–79, 1952.
- [3177] H. Olteanu and R. Banerjee. Human methionine synthase reductase, a soluble P-450 reductase-like dual flavoprotein, is sufficient for NADPH-dependent methionine synthase activation. *J. Biol. Chem.*, 276:35558–35563, 2001.
- [3178] H. Olteanu, T. Munson, and R. Banerjee. Differences in the efficiency of reductive activation of methionine synthase and exogenous electron acceptors between the common polymorphic variants of human methionine synthase reductase. *Biochemistry*, 41:13378–13385, 2002.
- [3179] T. Omori, H. Ishigooka, and Y. Minoda. Purification and some properties of 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoic acid(HOPDA) reducing enzyme from *Pseudomonas cruciviae* S93B1 involved in the degradation of biphenyl. *Agric. Biol. Chem.*, 50:1513–1518, 1986.
- [3180] T. Omura, E. Sanders, R.W. Estabrook, D.Y. Cooper, and O. Rosenthal. Isolation from adrenal cortex of a nonheme iron protein and a flavoprotein functional as a reduced triphosphopyridine nucleotide-cytochrome P-450 reductase. *Arch. Biochem. Biophys.*, 117:660–673, 1966.
- [3181] H.V. Onckelen, E. Prinsen, D. Inze, P. Ruedesheim, M.V. Lijsebettens, A. Follin, J. Schell, M.V. Montagu, and J.D. Greef. Agrobacterium T-DNA gene1 codes for tryptophan 2-monooxygenase activity in tobacco crown gall cells. *FEBS Lett.*, 198:357–360, 1986.
- [3182] R.N. Ondarza, R. Abney, and A.M. López-Colomé. Characterization of a NADPH-dependent coenzyme A-S-S-glutathione reductase from yeast. *Biochim. Biophys. Acta*, 191:239–248, 1969.
- [3183] R.N. Ondarza, E. Escamilla, J. Gutierrez, and G. De la Chica. CoAS-Sglutathione and GSSG reductases from rat liver. Two disulfide oxidoreductase activities in one protein entity. *Biochim. Biophys. Acta*, 341:162–171, 1974.
- [3184] E. Ono, M. Fukuchi-Mizutani, N. Nakamura, Y. Fukui, K. Yonekura-Sakakibara, M. Yamaguchi, T. Nakayama, T. Tanaka, T. Kusumi, and Y. Tanaka. Yellow flowers generated by expression of the aurone biosynthetic pathway. *Proc. Natl. Acad. Sci. USA*, 103:11075–11080, 2006.
- [3185] E. Ono, M. Nakai, Y. Fukui, N. Tomimori, M. Fukuchi-Mizutani, M. Saito, H. Satake, T. Tanaka, M. Katsuta, T. Umezawa, and Y. Tanaka. Formation of two methylenedioxy bridges by a *Sesamum* CYP81Q protein yielding a furofuran lignan, (+)-sesamin. *Proc. Natl. Acad. Sci. USA*, 103:10116–10121, 2006.
- [3186] T. Ono and K. Bloch. Solubilization and partial characterization of rat liver squalene epoxidase. *J. Biol. Chem.*, 250:1571–1579, 1975.
- [3187] W. Oonanant, J. Sucharitakul, J. Yuvaniyama, and P. Chaiyen. Crystallization and preliminary X-ray crystallographic analysis of 2-methyl-3-hydroxypyridine-5-carboxylic acid (MHPC) oxygenase from *Pseudomonas* sp. MA-1. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:312–314, 2005.
- [3188] P.W. Van Ophem, J. Van Beeumen, and J.A. Duine. Nicotinoprotein [NAD(P)-containing] alcohol/aldehyde oxidoreductases. Purification and characterization of a novel type from *Amycolatopsis methanolica*. *Eur. J. Biochem.*, 212:819–826, 1993.

- [3189] U.C. Oppermann and E. Maser. Characterization of a 3 α -hydroxysteroid dehydrogenase/carbonyl reductase from the gram-negative bacterium *Comamonas testosteroni*. *Eur. J. Biochem.*, 241:744–749, 1996.
- [3190] E. Ordonez, K. Van Belle, G. Roos, S. De Galan, M. Letek, J.A. Gil, L. Wyns, L.M. Mateos, and J. Messens. Arsenate reductase, mycothiol, and mycoredoxin concert thiol/disulfide exchange. *J. Biol. Chem.*, 284:15107–15116, 2009.
- [3191] C. Orii, S. Takenaka, S. Murakami, and K. Aoki. Metabolism of 4-amino-3-hydroxybenzoic acid by *Bordetella* sp. strain 10d: A different modified meta-cleavage pathway for 2-aminophenols. *Biosci. Biotechnol. Biochem.*, 70:2653–2661, 2006.
- [3192] L. Örnung. ω -Oxidation of cysteine-containing leukotrienes by rat-liver microsomes. Isolation and characterization of ω -hydroxy and ω -carboxy metabolites of leukotriene E₄ and *N*-acetylleukotriene E₄. *Eur. J. Biochem.*, 170:77–85, 1987.
- [3193] S. Osaki. Kinetic studies of ferrous ion oxidation with crystalline human ferroxidase (ceruloplasmin). *J. Biol. Chem.*, 241:5053–5059, 1966.
- [3194] S. Osaki and O. Walaas. Kinetic studies of ferrous ion oxidation with crystalline human ferroxidase. II. Rate constants at various steps and formation of a possible enzyme-substrate complex. *J. Biol. Chem.*, 242:2653–2657, 1967.
- [3195] M.J. Osborn and F.M. Huennekens. Participation of anhydroleucovorin in the hydroxymethyl tetrahydrofolic dehydrogenase system. *Biochim. Biophys. Acta*, 26:646–647, 1957.
- [3196] R. Oshima, S. Fushinobu, F. Su, L. Zhang, N. Takaya, and H. Shoun. Structural evidence for direct hydride transfer from NADH to cytochrome P450_{nor}. *J. Mol. Biol.*, 342:207–217, 2004.
- [3197] N. Oshino, Y. Imai, and R. Sato. Electron-transfer mechanism associated with fatty acid desaturation catalyzed by liver microsomes. *Biochim. Biophys. Acta*, 128:13–27, 1966.
- [3198] N. Oshino, Y. Imai, and R. Sato. A function of cytochrome b₅ in fatty acid desaturation by rat liver microsomes. *J. Biochem. (Tokyo)*, 69:155–167, 1971.
- [3199] U. Oster, R. Tanaka, A. Tanaka, and W. Rüdiger. Cloning and functional expression of the gene encoding the key enzyme for chlorophyll *b* biosynthesis (CAO) from *Arabidopsis thaliana*. *Plant J.*, 21:305–310, 2000.
- [3200] J. Østergaard, G. Persiau, M.W. Davey, G. Bauw, and M. Van Montagu. Isolation of a cDNA coding for L-galactono- γ -lactone dehydrogenase, an enzyme involved in the biosynthesis of ascorbic acid in plants. Purification, characterization, cDNA cloning, and expression in yeast. *J. Biol. Chem.*, 272:30009–30016, 1997.
- [3201] M.C. Ostrowski and W.S. Kistler. Properties of a flavoprotein sulfhydryl oxidase from rat seminal vesicle secretion. *Biochemistry*, 19:2639–2645, 1980.
- [3202] J. O’Sullivan, M. Unzeta, J. Healy, M.I. O’Sullivan, G. Davey, and K.F. Tipton. Semicarbazide-sensitive amine oxidases: enzymes with quite a lot to do. *Neurotoxicology*, 25:303–315, 2004.
- [3203] T. Osumi, T. Hashimoto, and N. Ui. Purification and properties of acyl-CoA oxidase from rat liver. *J. Biochem. (Tokyo)*, 87:1735–1746, 1980.
- [3204] K. Otani, T. Takahashi, T. Furuya, and S. Ayabe. Licodione synthase, a cytochrome P₄₅₀ monooxygenase catalyzing 2-hydroxylation of 5-deoxyflavanone, in cultured *Glycyrrhiza echinata* L. cells. *Plant Physiol.*, 105:1427–1432, 1994.
- [3205] Y. Otha and D.W. Ribbons. Crystallization of orchinol hydroxylase from *Pseudomonas putida*. *FEBS Lett.*, 11:189–192, 1970.
- [3206] K. Otsuka. Triphosphopyridine nucleotide-allyl and -ethyl alcohol dehydrogenases from *Escherichia coli*. *J. Gen. Appl. Microbiol.*, 4:211–215, 1958.
- [3207] L.A.B.M. Otten, D. Vreugdenhil, and R.A. Schilperoort. Properties of D(+)-lysopine dehydrogenase from crown gall tumour tissue. *Biochim. Biophys. Acta*, 485:268–277, 1977.
- [3208] K. Otto, K. Hofstetter, M. Rothlisberger, B. Witholt, and A. Schmid. Biochemical characterization of StyAB from *Pseudomonas* sp. strain VLB120 as a two-component flavin-diffusible monooxygenase. *J. Bacteriol.*, 186:5292–5302, 2004.

- [3209] G. Ottolina, S. Bianchi, B. Belloni, G. Carrea, and B. Danieli. First asymmetric oxidation of tertiary amines by cyclohexanone monooxygenase. *Tetrahedron Lett.*, 40:8483–8486, 1999.
- [3210] A. Oubrie and B.W. Dijkstra. Structural requirements of pyrroloquinoline quinone dependent enzymatic reactions. *Protein Sci.*, 9:1265–1273, 2000.
- [3211] A. Oubrie, H.J. Rozeboom, K.H. Kalk, E.G. Huizinga, and B.W. Dijkstra. Crystal structure of quinohemoprotein alcohol dehydrogenase from *Comamonas testosteroni*: structural basis for substrate oxidation and electron transfer. *J. Biol. Chem.*, 277:3727–3732, 2002.
- [3212] S. Ouchane, A.S. Steunou, M. Picaud, and C. Astier. Aerobic and anaerobic Mg-protoporphyrin monomethyl ester cyclases in purple bacteria: a strategy adopted to bypass the repressive oxygen control system. *J. Biol. Chem.*, 279:6385–6394, 2004.
- [3213] T. Ouchi, T. Tomita, A. Horie, A. Yoshida, K. Takahashi, H. Nishida, K. Lassak, H. Taka, R. Mineki, T. Fujimura, S. Kosono, C. Nishiyama, R. Masui, S. Kuramitsu, S.V. Albers, T. Kuzuyama, and M. Nishiyama. Lysine and arginine biosyntheses mediated by a common carrier protein in *Sulfolobus*. *Nat. Chem. Biol.*, 9:277–283, 2013.
- [3214] H. Ouellet, S. Guan, J.B. Johnston, E.D. Chow, P.M. Kells, A.L. Burlingame, J.S. Cox, L.M. Podust, and P.R. de Montellano. *Mycobacterium tuberculosis* CYP125A1, a steroid C27 monooxygenase that detoxifies intracellularly generated cholest-4-en-3-one. *Mol. Microbiol.*, 77:730–742, 2010.
- [3215] H.J. Ougham, D.G. Taylor, and P.W. Trudgill. Camphor revisited: involvement of a unique monooxygenase in metabolism of 2-oxo- Δ^3 -4,5,5-trimethylcyclopentenylacetic acid by *Pseudomonas putida*. *J. Bacteriol.*, 153:140–152, 1983.
- [3216] T. Oura and S. Kajiwara. Disruption of the sphingolipid Δ^8 -desaturase gene causes a delay in morphological changes in *Candida albicans*. *Microbiology*, 154:3795–3803, 2008.
- [3217] F.W. Outten, D.L. Huffman, J.A. Hale, and T.V. O’Halloran. The independent cue and cus systems confer copper tolerance during aerobic and anaerobic growth in *Escherichia coli*. *J. Biol. Chem.*, 276:30670–30677, 2001.
- [3218] J. Owaki, K. Uzura, Z. Minami, and K. Kusai. Partial-purification and characterization of dihydrouracil oxidase, a flavoprotein from *Rhodotorula glutinis*. *J. Ferment. Technol.*, 64:205–210, 1986.
- [3219] K.S. Oyedotun and B.D. Lemire. The quaternary structure of the *Saccharomyces cerevisiae* succinate dehydrogenase. Homology modeling, cofactor docking, and molecular dynamics simulation studies. *J. Biol. Chem.*, 279:9424–9431, 2004.
- [3220] E. Pahlich and K.W. Joy. Glutamate dehydrogenase from pea roots: purification and properties of the enzyme. *Can. J. Biochem.*, 49:127–138, 1971.
- [3221] E.F. Pai, R.H. Schirmer, and G.E. Schulz. Structural studies on crystalline glutathione reductase from human erythrocytes. In T.P. Singer and R.N. Ondarza, editors, *Mechanisms of Oxidizing Enzymes*, pages 17–22. Mechanisms of Oxidizing Enzymes, New York, 1978.
- [3222] Y.K. Paik, J.M. Trzaskos, A. Shafice, and J.L. Gaylor. Microsomal enzymes of cholesterol biosynthesis from lanosterol. Characterization, solubilization, and partial purification of NADPH-dependent $\Delta^{8,14}$ -steroid 14-reductase. *J. Biol. Chem.*, 259:13413–13423, 1984.
- [3223] A.H. Palamakumbura and P.C. Trackman. A fluorometric assay for detection of lysyl oxidase enzyme activity in biological samples. *Anal. Biochem.*, 300:245–251, 2002.
- [3224] N.J. Palleroni and M. Doudoroff. Metabolism of carbohydrates by *Pseudomonas saccharophilla*. III. Oxidation of D-arabinose. *J. Bacteriol.*, 74:180–185, 1957.
- [3225] N.R. Palosaari and P. Rogers. Purification and properties of the inducible coenzyme A-linked butyraldehyde dehydrogenase from *Clostridium acetobutylicum*. *J. Bacteriol.*, 170:2971–2976, 1988.
- [3226] F. Paltauf. Biosynthesis of plasmalogens from alkyl- and alkyl-acyl-glycerophosphoryl ethanolamine in the rat brain. *FEBS Lett.*, 17:118–120, 1971.

- [3227] F. Paltuaf, R.A. Prough, B.S. Masters, and J.M. Johnston. Evidence for the participation of cytochrome *b₅* in plasmalogen biosynthesis. *J. Biol. Chem.*, 249:2661–2662, 1974.
- [3228] J.J. Pan, J.O. Solbiati, G. Ramamoorthy, B.S. Hillerich, R.D. Seidel, J.E. Cronan, S.C. Almo, and C.D. Poulter. Biosynthesis of squalene from farnesyl diphosphate in bacteria: three steps catalyzed by three enzymes. *ACS Cent. Sci.*, 1:77–82, 2015.
- [3229] Z. Pan, A.M. Rimando, S.R. Baerson, M. Fishbein, and S.O. Duke. Functional characterization of desaturases involved in the formation of the terminal double bond of an unusual 16:3Δ(9,12,15) fatty acid isolated from *Sorghum bicolor* root hairs. *J. Biol. Chem.*, 282:4326–4335, 2007.
- [3230] A. Paneque, F.F. Del Campo, J.M. Ramirez, and M. Losada. Flavin nucleotide nitrate reductase from spinach. *Biochim. Biophys. Acta*, 109:79–85, 1965.
- [3231] A.H. Pang, S. Garneau-Tsodikova, and O.V. Tsodikov. Crystal structure of halogenase PltA from the pyoluteorin biosynthetic pathway. *J. Struct. Biol.*, 192:349–357, 2015.
- [3232] Y. Pang, I.S. Abeysinghe, J. He, X. He, D. Huhman, K.M. Mewan, L.W. Sumner, J. Yun, and R.A. Dixon. Functional characterization of proanthocyanidin pathway enzymes from tea and their application for metabolic engineering. *Plant Physiol.*, 161:1103–1116, 2013.
- [3233] K. Pant, A.M. Bilwes, S. Adak, D.J. Stuehr, and B.R. Crane. Structure of a nitric oxide synthase heme protein from *Bacillus subtilis*. *Biochemistry*, 41:11071–11079, 2002.
- [3234] H. Pape and J.L. Strominger. Enzymatic synthesis of cytidine diphosphate 3,6-dideoxyhexoses. V. Partial purification of the two protein components required for introduction of the 3-deoxy group. *J. Biol. Chem.*, 244:3598–3604, 1969.
- [3235] J.V. Parales, R.E. Parales, S.M. Resnick, and D.T. Gibson. Enzyme specificity of 2-nitrotoluene 2,3-dioxygenase from *Pseudomonas* sp. strain JS42 is determined by the C-terminal region of the α subunit of the oxygenase component. *J. Bacteriol.*, 180:1194–1199, 1998.
- [3236] R.E. Parales, K. Lee, S.M. Resnick, H. Jiang, D.J. Lessner, and D.T. Gibson. Substrate specificity of naphthalene dioxygenase: effect of specific amino acids at the active site of the enzyme. *J. Bacteriol.*, 182:1641–1649, 2000.
- [3237] R. Parham and C.A. Rebeiz. Chloroplast biogenesis: [4-vinyl] chlorophyllide *a* reductase is a divinyl chlorophyllide *a*-specific, NADPH-dependent enzyme. *Biochemistry*, 31:8460–8464, 1992.
- [3238] R. Parham and C.A. Rebeiz. Chloroplast biogenesis 72: a [4-vinyl]chlorophyllide *a* reductase assay using divinyl chlorophyllide *a* as an exogenous substrate. *Anal. Biochem.*, 231:164–169, 1995.
- [3239] H. Park, H. Lee, Y.T. Ro, and Y.M. Kim. Identification and functional characterization of a gene for the methanol : *N,N'*-dimethyl-4-nitrosoaniline oxidoreductase from *Mycobacterium* sp. strain JC1 (DSM 3803). *Microbiology*, 156:463–471, 2010.
- [3240] Y.J. Park, C.B. Yoo, S.Y. Choi, and H.B. Lee. Purifications and characterizations of a ferredoxin and its related 2-oxoacid:ferredoxin oxidoreductase from the hyperthermophilic archaeon, *Sulfolobus solfataricus* P1. *J. Biochem. Mol. Biol.*, 39:46–54, 2006.
- [3241] J.B. Parker and C.T. Walsh. Action and timing of BacC and BacD in the late stages of biosynthesis of the dipeptide antibiotic bacilysin. *Biochemistry*, 52:889–901, 2013.
- [3242] T. Parkkinen, H. Boer, J. Janis, M. Andberg, M. Penttila, A. Koivula, and J. Rouvinen. Crystal structure of uronate dehydrogenase from *Agrobacterium tumefaciens*. *J. Biol. Chem.*, 286:27294–27300, 2011.
- [3243] R.J. Parry and W. Li. An NADPH:FAD oxidoreductase from the valanimycin producer, *Streptomyces viridifaciens*. Cloning, analysis, and overexpression. *J. Biol. Chem.*, 272:23303–23311, 1997.
- [3244] R.J. Parry and W. Li. Purification and characterization of isobutylamine *N*-hydroxylase from the valanimycin producer *Streptomyces viridifaciens* MG456-hF10. *Arch. Biochem. Biophys.*, 339:47–54, 1997.
- [3245] R.J. Parry, W. Li, and H.N. Cooper. Cloning, analysis, and overexpression of the gene encoding isobutylamine *N*-hydroxylase from the valanimycin producer, *Streptomyces viridifaciens*. *J. Bacteriol.*, 179:409–416, 1997.

- [3246] K. Parschat, C. Canne, J. Hüttermann, R. Kappl, and S. Fetzner. Xanthine dehydrogenase from *Pseudomonas putida* 86: specificity, oxidation-reduction potentials of its redox-active centers, and first EPR characterization. *Biochim. Biophys. Acta*, 1544:151–165, 2001.
- [3247] J.F. Parsons, B.T. Greenhagen, K. Shi, K. Calabrese, H. Robinson, and J.E. Ladner. Structural and functional analysis of the pyocyanin biosynthetic protein PhzM from *Pseudomonas aeruginosa*. *Biochemistry*, 46:1821–1828, 2007.
- [3248] S.J. Parsons and R.O. Burns. Purification and properties of β -isopropylmalate dehydrogenase. *J. Biol. Chem.*, 244:996–1003, 1969.
- [3249] S.D. Parzen and A.S. Fox. Purification of xanthine dehydrogenase from *Drosophila melanogaster*. *Biochim. Biophys. Acta*, 92:465–471, 1964.
- [3250] S. Pascal, M. Taton, and A. Rahier. Plant sterol biosynthesis. Identification and characterization of two distinct microsomal oxidative enzymatic systems involved in sterol C4-demethylation. *J. Biol. Chem.*, 268:11639–11654, 1993.
- [3251] A. Paszcynski and J. Trojanowski. An affinity-column procedure for the purification of veratrate *O*-demethylase from fungi. *Microbios*, 18:111–121, 1977.
- [3252] A. Paszcynski, V.-B. Huynh, and R. Crawford. Comparison of ligninase-I and peroxidase-M2 from the white-rot fungus *Phanerochaete chrysosporium*. *Arch. Biochem. Biophys.*, 244:750–765, 1986.
- [3253] I. Patel, D. Kracher, S. Ma, S. Garajova, M. Haon, C.B. Faulds, J.G. Berrin, R. Ludwig, and E. Record. Salt-responsive lytic polysaccharide monooxygenases from the mangrove fungus *Pestalotiopsis* sp. NCi6. *Biotechnol Biofuels*, 9:108–108, 2016.
- [3254] M.P. Patel and J.S. Blanchard. Expression, purification, and characterization of *Mycobacterium tuberculosis* mycothione reductase. *Biochemistry*, 38:11827–11833, 1999.
- [3255] M.P. Patel and J.S. Blanchard. *Mycobacterium tuberculosis* mycothione reductase: pH dependence of the kinetic parameters and kinetic isotope effects. *Biochemistry*, 40:5119–5126, 2001.
- [3256] M.S. Patel and T.E. Roche. Molecular biology and biochemistry of pyruvate dehydrogenase complexes. *FASEB J.*, 4:3224–3233, 1990.
- [3257] R.N. Patel, C.T. Hou, P. Derelanko, and A. Felix. Purification and properties of a heme-containing aldehyde dehydrogenase from *Methylosinus trichosporium*. *Arch. Biochem. Biophys.*, 203:654–662, 1980.
- [3258] T.R. Patel and E.A. Barnsley. Naphthalene metabolism by pseudomonads: purification and properties of 1,2-dihydroxynaphthalene oxygenase. *J. Bacteriol.*, 143:668–673, 1980.
- [3259] T.R. Patel and D.T. Gibson. Purification and properties of (+)-*cis*-naphthalene dihydrodiol dehydrogenase of *Pseudomonas putida*. *J. Bacteriol.*, 119:879–888, 1974.
- [3260] J.A. Pateman, B.M. Rever, and D.J. Cove. Genetic and biochemical studies of nitrate reduction in *Aspergillus nidulans*. *Biochem. J.*, 104:103–111, 1967.
- [3261] S.S. Patil and M. Zucker. Potato phenolases. Purification and properties. *J. Biol. Chem.*, 240:3938–3943, 1965.
- [3262] R.N. Patkar, P.I. Benke, Z. Qu, Y.Y. Chen, F. Yang, S. Swarup, and N.I. Naqvi. A fungal monooxygenase-derived jasmonate attenuates host innate immunity. *Nat. Chem. Biol.*, 11:733–740, 2015.
- [3263] W.R. Patterson and T.L. Poulos. Crystal structure of recombinant pea cytosolic ascorbate peroxidase. *Biochemistry*, 34:4331–4341, 1995.
- [3264] K.G. Paul. Peroxidases. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 8, pages 227–274. Academic Press, New York, 2nd edition, 1963.
- [3265] H.H. Pauli and T.M. Kutchan. Molecular cloning and functional heterologous expression of two alleles encoding (*S*)-*N*-methylcoclaurine 3'-hydroxylase (CYP80B1), a new methyl jasmonate-inducible cytochrome *P*-450-dependent monooxygenase of benzyloisoquinoline alkaloid biosynthesis. *Plant J.*, 13:793–801, 1998.
- [3266] H.E. Pauly and G. Pfeleiderer. D-Glucose dehydrogenase from *Bacillus megaterium* M 1286: purification, properties and structure. *Hoppe-Seylers Z. Physiol. Chem.*, 356:1613–1623, 1975.

- [3267] F. Pauwels, B. Vergauwen, F. Vanrobaeys, B. Devreese, and J.J. Van Beeumen. Purification and characterization of a chimeric enzyme from *Haemophilus influenzae* Rd that exhibits glutathione-dependent peroxidase activity. *J. Biol. Chem.*, 278:16658–16666, 2003.
- [3268] J.W. Payne, H. Bolton, Campbell Jr., Xun J.A., and L. Purification and characterization of EDTA monooxygenase from the EDTA-degrading bacterium BNC1. *J. Bacteriol.*, 180:3823–3827, 1998.
- [3269] C.W. Payton and Y.-F. Chang. Δ^1 -Piperidine-2-carboxylate reductase of *Pseudomonas putida*. *J. Bacteriol.*, 149:864–871, 1982.
- [3270] S.L. Pealing, A.C. Black, F.D. Manson, F.B. Ward, S.K. Chapman, and G.A. Reid. Sequence of the gene encoding flavocytochrome *c* from *Shewanella putrefaciens*: a tetraheme flavoenzyme that is a soluble fumarate reductase related to the membrane-bound enzymes from other bacteria. *Biochemistry*, 31:12132–12140, 1992.
- [3271] S.L. Pealing, M.R. Cheesman, G.A. Reid, A.J. Thomson, F.B. Ward, and S.K. Chapman. Spectroscopic and kinetic studies of the tetraheme flavocytochrome *c* from *Shewanella putrefaciens* NCIMB400. *Biochemistry*, 34:6153–6158, 1995.
- [3272] A.J. Pease, B.R. Roa, W. Luo, and M.E. Winkler. Positive growth rate-dependent regulation of the *pdxA*, *ksgA*, and *pdxB* genes of *Escherichia coli* K-12. *J. Bacteriol.*, 184:1359–1369, 2002.
- [3273] T.A. Pechurskaya, O.P. Lukashevich, A.A. Gilep, and S.A. Usanov. Engineering, expression, and purification of "soluble" human cytochrome P45017 α and its functional characterization. *Biochemistry (Mosc.)*, 73:806–811, 2008.
- [3274] S.C. Peck, H.A. Cooke, R.M. Cicchillo, P. Malova, F. Hammerschmidt, S.K. Nair, and W.A. van der Donk. Mechanism and substrate recognition of 2-hydroxyethylphosphonate dioxygenase. *Biochemistry*, 50:6598–6605, 2011.
- [3275] S.C. Peck, K. Denger, A. Burrichter, S.M. Irwin, E.P. Balskus, and D. Schleheck. A glyceryl radical enzyme enables hydrogen sulfide production by the human intestinal bacterium *Bifidobacterium wadsworthia*. *Proc. Natl. Acad. Sci. USA*, 116:3171–3176, 2019.
- [3276] M.J. Pecyna, R. Ullrich, B. Bittner, A. Clemens, K. Scheibner, R. Schubert, and M. Hofrichter. Molecular characterization of aromatic peroxygenase from *Agroclybe aegerita*. *Appl. Microbiol. Biotechnol.*, 84:885–897, 2009.
- [3277] J.I. Pedersen, G. Eggertsen, U. Hellman, U. Andersson, and I. Björkhem. Molecular cloning and expression of cDNA encoding 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanoyl-CoA oxidase from rabbit liver. *J. Biol. Chem.*, 272:18481–18489, 1997.
- [3278] J.R. Pedrajas, C.A. Padilla, B. McDonagh, and J.A. Barcena. Glutaredoxin participates in the reduction of peroxides by the mitochondrial 1-CYS peroxiredoxin in *Saccharomyces cerevisiae*. *Antioxid Redox Signal*, 13:249–258, 2010.
- [3279] B. Pedre, L.A. van Bergen, A. Pallo, L.A. Rosado, V.T. Dufe, I.V. Molle, K. Wahni, H. Erdogan, M. Alonso, F.D. Proft, and J. Messens. The active site architecture in peroxiredoxins: a case study on *Mycobacterium tuberculosis* AhpE. *Chem. Commun. (Camb.)*, 52:10293–10296, 2016.
- [3280] J. Peek and D. Christendat. The shikimate dehydrogenase family: functional diversity within a conserved structural and mechanistic framework. *Arch. Biochem. Biophys.*, 566:85–99, 2015.
- [3281] O.B. Peersen, E.A. Pratt, H.T. Truong, C. Ho, and G.S. Rule. Site-specific incorporation of 5-fluorotryptophan as a probe of the structure and function of the membrane-bound D-lactate dehydrogenase of *Escherichia coli*: a ^{19}F nuclear magnetic resonance study. *Biochemistry*, 29:3256–3262, 1990.
- [3282] J. Peisach and W.G. Levine. A comparison of the enzymic activities of pig ceruloplasmin and *Rhus vernicifera* laccase. *J. Biol. Chem.*, 240:2284–2289, 1965.
- [3283] D.A. Pelletier and C.S. Harwood. 2-Hydroxycyclohexanecarboxyl coenzyme A dehydrogenase, an enzyme characteristic of the anaerobic benzoate degradation pathway used by *Rhodospseudomonas palustris*. *J. Bacteriol.*, 182:2753–2760, 2000.
- [3284] M.K. Pelletier and B.W. Shirley. Analysis of flavanone 3-hydroxylase in *Arabidopsis* seedlings. Coordinate regulation with chalcone synthase and chalcone isomerase. *Plant Physiol.*, 111:339–345, 1996.

- [3285] L. Pelosi, A.L. Ducluzeau, L. Loiseau, F. Barras, D. Schneider, I. Junier, and F. Pierrel. Evolution of Ubiquinone Biosynthesis: Multiple Proteobacterial Enzymes with Various Regioselectivities To Catalyze Three Contiguous Aromatic Hydroxylation Reactions. *mSystems*, 1, 2016.
- [3286] A.M. Pelzmann, F. Mickoleit, and O. Meyer. Insights into the posttranslational assembly of the Mo-, S- and Cu-containing cluster in the active site of CO dehydrogenase of *Oligotropha carboxidovorans*. *J. Biol. Inorg. Chem.*, 19:1399–1414, 2014.
- [3287] T.M. Penning and R.B. Sharp. Prostaglandin dehydrogenase activity of purified rat liver 3 α -hydroxysteroid dehydrogenase. *Biochem. Biophys. Res. Commun.*, 148:646–652, 1987.
- [3288] M.P. Pereira and E.D. Brown. Bifunctional catalysis by CDP-ribitol synthase: convergent recruitment of reductase and cytidyltransferase activities in *Haemophilus influenzae* and *Staphylococcus aureus*. *Biochemistry*, 43:11802–11812, 2004.
- [3289] M.P. Pereira, M.A. D’Elia, J. Troczynska, and E.D. Brown. Duplication of teichoic acid biosynthetic genes in *Staphylococcus aureus* leads to functionally redundant poly(ribitol phosphate) polymerases. *J. Bacteriol.*, 190:5642–5649, 2008.
- [3290] M. Pérez-Boada, F.J. Ruiz-Dueñas, R. Pogni, R. Basosi, T. Choinowski, M.J. Martínez, K. Piontek, and A.T. Martínez. Versatile peroxidase oxidation of high redox potential aromatic compounds: site-directed mutagenesis, spectroscopic and crystallographic investigation of three long-range electron transfer pathways. *J. Mol. Biol.*, 354:385–402, 2005.
- [3291] V.H. Perez-Espana, N. Sanchez-Leon, and J.P. Vielle-Calzada. CYP85A1 is required for the initiation of female gametogenesis in *Arabidopsis thaliana*. *Plant Signal Behav.*, 6:321–326, 2011.
- [3292] M.E. Perez-Perez, A. Mata-Cabana, A.M. Sanchez-Riego, M. Lindahl, and F.J. Florencio. A comprehensive analysis of the peroxiredoxin reduction system in the cyanobacterium *Synechocystis* sp. strain PCC 6803 reveals that all five peroxiredoxins are thioredoxin dependent. *J. Bacteriol.*, 191:7477–7489, 2009.
- [3293] R.N. Perham. Swinging arms and swinging domains in multifunctional enzymes: catalytic machines for multistep reactions. *Annu. Rev. Biochem.*, 69:961–1004, 2000.
- [3294] B.C. Persson and G.R. Bjork. Isolation of the gene (*miaE*) encoding the hydroxylase involved in the synthesis of 2-methylthio-*cis*-ribozeatin in tRNA of *Salmonella typhimurium* and characterization of mutants. *J. Bacteriol.*, 175:7776–7785, 1993.
- [3295] B.C. Persson, O. Olafsson, H.K. Lundgren, L. Hederstedt, and G.R. Bjork. The ms2io6A37 modification of tRNA in *Salmonella typhimurium* regulates growth on citric acid cycle intermediates. *J. Bacteriol.*, 180:3144–3151, 1998.
- [3296] B. Peschke and F. Lingens. Microbial metabolism of quinoline and related compounds. XII. Isolation and characterization of the quinoline oxidoreductase from *Rhodococcus* sp. B1 compared with the quinoline oxidoreductase from *Pseudomonas putida* 86. *Biol. Chem. Hoppe-Seyler*, 372:1081–1088, 1991.
- [3297] J. Petasch, E.M. Disch, S. Markert, D. Becher, T. Schweder, B. Huttel, R. Reinhardt, and J. Harder. The oxygen-independent metabolism of cyclic monoterpenes in *Castellaniella defragrans* 65Phen. *BMC Microbiol.*, 14:164–164, 2014.
- [3298] F. Peters, D. Heintz, J. Johannes, A. van Dorsselaer, and M. Boll. Genes, enzymes, and regulation of *para*-cresol metabolism in *Geobacter metallireducens*. *J. Bacteriol.*, 189:4729–4738, 2007.
- [3299] J.W. Peters, W.N. Lanzilotta, B.J. Lemon, and L.C. Seefeldt. X-ray crystal structure of the Fe-only hydrogenase (Cpl) from *Clostridium pasteurianum* to 1.8 Angstrom resolution. *Science*, 282:1853–1858, 1998.
- [3300] M. Petersen and A.W. Alfermann. Two new enzymes of rosmarinic acid biosynthesis from cell cultures of *Coleus blumei*: hydroxyphenylpyruvate reductase and rosmarinic acid synthase. *Z. Naturforsch. C: Biosci.*, 43:501–504, 1988.
- [3301] J.A. Peterson, M. Kusunose, E. Kusunose, and M.J. Coon. Enzymatic ω -oxidation. II. Function of rubredoxin as the electron carrier in ω -hydroxylation. *J. Biol. Chem.*, 242:4334–4340, 1967.
- [3302] J.A. Peterson, M.C. Lorence, and B. Amarnah. Putidaredoxin reductase and putidaredoxin. Cloning, sequence determination, and heterologous expression of the proteins. *J. Biol. Chem.*, 265:6066–6073, 1990.

- [3303] K.L. Peterson, E.E. Sergienko, Y. Wu, N.R. Kumar, A.W. Strauss, A.E. Oleson, W.W. Muhonen, J.B. Shabb, and D.K. Srivastava. Recombinant human liver medium-chain acyl-CoA dehydrogenase: purification, characterization, and the mechanism of interactions with functionally diverse C₈-CoA molecules. *Biochemistry*, 34:14942–14953, 1995.
- [3304] H. Petitdemange, H. Blusson, and R. Gay. Detection of NAD(P)H-rubredoxin oxidoreductases in *Clostridia*. *Anal. Biochem.*, 116:564–570, 1981.
- [3305] H. Petitdemange, R. Marczak, H. Blusson, and R. Gay. Isolation and properties of reduced nicotinamide adenine dinucleotide rubredoxin oxidoreductase of *Clostridium acetobutylicum*. *Biochem. Biophys. Res. Commun.*, 91:1258–1265, 1979.
- [3306] G.A. Petrini, S.G. Altabe, and A.D. Uttaro. *Trypanosoma brucei* oleate desaturase may use a cytochrome b₅-like domain in another desaturase as an electron donor. *Eur. J. Biochem.*, 271:1079–1086, 2004.
- [3307] M. Petrusma, L. Dijkhuizen, and R. van der Geize. *Rhodococcus rhodochrous* DSM 43269 3-ketosteroid 9 α -hydroxylase, a two-component iron-sulfur-containing monooxygenase with subtle steroid substrate specificity. *Appl. Environ. Microbiol.*, 75:5300–5307, 2009.
- [3308] I. Pettinati, J. Brem, M.A. McDonough, and C.J. Schofield. Crystal structure of human persulfide dioxygenase: structural basis of ethylmalonic encephalopathy. *Hum. Mol. Genet.*, 24:2458–2469, 2015.
- [3309] F.H. Pettit, S.J. Yeaman, and L.J. Reed. Purification and characterization of branched chain α -keto acid dehydrogenase complex of bovine kidney. *Proc. Natl. Acad. Sci. USA*, 75:4881–4885, 1978.
- [3310] A. Pfitzner, B. Krausch, and J. Stöckigt. Characteristics of vellosimine reductase, a specific enzyme involved in the biosynthesis of the *Rauwolfia* alkaloid sarpagine. *Tetrahedron*, 40:1691–1699, 1984.
- [3311] A. Pfitzner and J. Stöckigt. Partial-purification and characterization of geissoschizine dehydrogenase from suspension-cultures of *Catharanthus roseus*. *Phytochemistry*, 21:1585–1588, 1982.
- [3312] E.C. Pflieger, C. Piantadosi, and F. Snyder. The biocleavage of isomeric glyceryl ethers by soluble liver enzymes in a variety of species. *Biochim. Biophys. Acta*, 144:633–648, 1967.
- [3313] C.M. Phillips, W.T. Beeson, J.H. Cate, and M.A. Marletta. Cellobiose dehydrogenase and a copper-dependent polysaccharide monooxygenase potentiate cellulose degradation by *Neurospora crassa*. *ACS Chem. Biol.*, 6:1399–1406, 2011.
- [3314] D.M. Phillips, V. Lakshmi, and C. Monder. Corticosteroid 11 β -dehydrogenase in rat testis. *Endocrinology*, 125:209–216, 1989.
- [3315] D.R. Phillips, J.A. Duley, D.J. Fennell, and R.S. Holmes. The self-association of L- α hydroxyacid oxidase. *Biochim. Biophys. Acta*, 427:679–687, 1976.
- [3316] A. Pickl and P. Schönheit. The oxidative pentose phosphate pathway in the haloarchaeon *Haloferax volcanii* involves a novel type of glucose-6-phosphate dehydrogenase⁻-The archaeal Zwischenferment. *FEBS Lett.*, 589:1105–1111, 2015.
- [3317] A. Piérard and J.M. Wiame. Propriétés de la L(+)-alanine-déshydrogénase. *Biochim. Biophys. Acta*, 37:490–502, 1960.
- [3318] B.S. Pierce, B.P. Subedi, S. Sardar, and J.K. Crowell. The ‘Gln-type’ thiol dioxygenase from *Azotobacter vinelandii* is a 3-mercaptopropionic acid dioxygenase. *Biochemistry*, 54:7477–7490, 2015.
- [3319] E. Pierce, D.F. Becker, and S.W. Ragsdale. Identification and characterization of oxalate oxidoreductase, a novel thiamine pyrophosphate-dependent 2-oxoacid oxidoreductase that enables anaerobic growth on oxalate. *J. Biol. Chem.*, 285:40515–40524, 2010.
- [3320] M.A. Pierrel, Y. Batard, M. Kazmaier, C. Mignotte-Vieux, F. Durst, and D. Werck-Reichhart. Catalytic properties of the plant cytochrome P450 CYP73 expressed in yeast. Substrate specificity of a cinnamate hydroxylase. *Eur. J. Biochem.*, 224:835–844, 1994.
- [3321] S.R. Piersma, A. Norin, S. de Vries, H. Jornvall, and J.A. Duine. Inhibition of nicotinoprotein (NAD⁺-containing) alcohol dehydrogenase by *trans*-4-(*N,N*-dimethylamino)-cinnamaldehyde binding to the active site. *J. Protein Chem.*, 22:457–461, 2003.

- [3322] S.R. Piersma, A.J. Visser, S. de Vries, and J.A. Duine. Optical spectroscopy of nicotinoprotein alcohol dehydrogenase from *Amycolatopsis methanolica*: a comparison with horse liver alcohol dehydrogenase and UDP-galactose epimerase. *Biochemistry*, 37:3068–3077, 1998.
- [3323] D. Pigeon, R. Drissi-Daoudi, F. Gros, and J. Thibault. Copurification of tyrosine hydroxylase from rat pheochromocytoma by protein kinase. *C. R. Acad. Sci. III*, 302:435–438, 1986.
- [3324] V.P. Pigiet and R.R. Conley. Purification of thioredoxin, thioredoxin reductase, and glutathione reductase by affinity chromatography. *J. Biol. Chem.*, 252:6367–6372, 1977.
- [3325] I.A. Pikuleva, A. Babiker, M.R. Waterman, and I. Björkhem. Activities of recombinant human cytochrome P450c27 (CYP27) which produce intermediates of alternative bile acid biosynthetic pathways. *J. Biol. Chem.*, 273:18153–18160, 1998.
- [3326] I.A. Pikuleva, A. Puchkaev, and I. Björkhem. Putative helix F contributes to regioselectivity of hydroxylation in mitochondrial cytochrome *P*₄₅₀ 27A1. *Biochemistry*, 40:7621–7629, 2001.
- [3327] A. Pinilla, F. Camps, and G. Fabrias. Cryptoregiochemistry of the Δ^{11} -myristoyl-CoA desaturase involved in the biosynthesis of *Spodoptera littoralis* sex pheromone. *Biochemistry*, 38:15272–15277, 1999.
- [3328] H. Pinkenburg Abounaga el, Schiffels O., El-Refai J., Buckel A., Selmer W., and T. Effect of an oxygen-tolerant bifurcating butyryl coenzyme A dehydrogenase/electron-transferring flavoprotein complex from *Clostridium difficile* on butyrate production in *Escherichia coli*. *J. Bacteriol.*, 195:3704–3713, 2013.
- [3329] V. Pinta, M. Picaud, F. Reiss-Husson, and C. Astier. *Rubrivivax gelatinosus* *acsF* (previously orf358) codes for a conserved, putative binuclear-iron-cluster-containing protein involved in aerobic oxidative cyclization of Mg-protoporphyrin IX monomethylester. *J. Bacteriol.*, 184:746–753, 2002.
- [3330] M.C. Pirrung. Ethylene biosynthesis from 1-aminocyclopropanecarboxylic acid. *Acc. Chem. Res.*, 32:711–718, 1999.
- [3331] E.K. Pistorius and H. Voss. Some properties of a basic L-amino-acid oxidase from *Anacystis nidulans*. *Biochim. Biophys. Acta*, 611:227–240, 1980.
- [3332] L.I. Pizer. The pathway and control of serine biosynthesis in *Escherichia coli*. *J. Biol. Chem.*, 238:3934–3944, 1963.
- [3333] J.E. Plager and L.T. Samuels. Synthesis of C14-17-hydroxy-11-desoxycorticosterone and 17-hydroxycorticosterone by fractionated extracts of adrenal homogenates. *Arch. Biochem. Biophys.*, 42:477–478, 1953.
- [3334] H.J. Plattner, P. Pfefferle, A. Romaguera, S. Waschutzka, and H. Diekmann. Isolation and some properties of lysine N⁶-hydroxylase from *Escherichia coli* strain EN222. *Biol. Met.*, 2:1–5, 1989.
- [3335] G.W.E. Plaut. Isocitrate dehydrogenases. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 105–126. Academic Press, New York, 2nd edition, 1963.
- [3336] G.W.E. Plaut and S.-C. Sung. Diphosphopyridine nucleotide isocitric dehydrogenase from animal tissues. *J. Biol. Chem.*, 207:305–314, 1954.
- [3337] T. Pluskal, M. Ueno, and M. Yanagida. Genetic and metabolomic dissection of the ergothioneine and selenoneine biosynthetic pathway in the fission yeast, *S. pombe*, and construction of an overproduction system. *PLoS One*, 9:e97774–e97774, 2014.
- [3338] T.C. Pochapsky, S.S. Pochapsky, T. Ju, H. Mo, F. Al-Mjeni, and M.J. Maroney. Modeling and experiment yields the structure of acireductone dioxygenase from *Klebsiella pneumoniae*. *Nat. Struct. Biol.*, 9:966–972, 2002.
- [3339] B. Pogson, K.A. McDonald, M. Truong, G. Britton, and D. DellaPenna. *Arabidopsis* carotenoid mutants demonstrate that lutein is not essential for photosynthesis in higher plants. *Plant Cell*, 8:1627–1639, 1996.
- [3340] E. Pohl, N. Brunner, M. Wilmanns, and R. Hensel. The crystal structure of the allosteric non-phosphorylating glyceraldehyde-3-phosphate dehydrogenase from the hyperthermophilic archaeum *Thermoproteus tenax*. *J. Biol. Chem.*, 277:19938–19945, 2002.
- [3341] V. Pohlmann and M.A. Marahiel. δ -amino group hydroxylation of L-ornithine during coelichelin biosynthesis. *Org. Biomol. Chem.*, 6:1843–1848, 2008.

- [3342] F. Pojer, R. Kahlich, B. Kammerer, S.M. Li, and L. Heide. CloR, a bifunctional non-heme iron oxygenase involved in chlorobiocin biosynthesis. *J. Biol. Chem.*, 278:30661–30668, 2003.
- [3343] T.J. Poklepovich, M.A. Rinaldi, M.L. Tomazic, N.O. Favale, A.P. Turkewitz, C.B. Nudel, and A.D. Nusblat. The cytochrome *b*₅ dependent C-5(6) sterol desaturase DES5A from the endoplasmic reticulum of *Tetrahymena thermophila* complements ergosterol biosynthesis mutants in *Saccharomyces cerevisiae*. *Steroids*, 77:1313–1320, 2012.
- [3344] A. Pol, T.R. Barends, A. Dietl, A.F. Khadem, J. Eygensteyn, M.S. Jetten, and H.J. Op den Camp. Rare earth metals are essential for methanotrophic life in volcanic mudpots. *Environ. Microbiol.*, 16:255–264, 2014.
- [3345] A. Poletti, F. Celotti, C. Rumio, M. Rabuffetti, and L. Martini. Identification of type 1 5 α -reductase in myelin membranes of male and female rat brain. *Mol. Cell. Endocrinol.*, 129:181–190, 1997.
- [3346] L. Pollegioni, F. Tonin, and E. Rosini. Lignin-degrading enzymes. *FEBS J.*, 282:1190–1213, 2015.
- [3347] K. Pollmann, S. Beil, and D.H. Pieper. Transformation of chlorinated benzenes and toluenes by *Ralstonia* sp. strain PS12 *tecA* (tetrachlorobenzene dioxygenase) and *tecB* (chlorobenzene dihydrodiol dehydrogenase) gene products. *Appl. Environ. Microbiol.*, 67:4057–4063, 2001.
- [3348] K. Pollmann, V. Wray, and D.H. Pieper. Chloromethylmuconolactones as critical metabolites in the degradation of chloromethylcatechols: recalcitrance of 2-chlorotoluene. *J. Bacteriol.*, 187:2332–2340, 2005.
- [3349] S.H. Pomerantz. Separation, purification, and properties of two tyrosinases from hamster melanoma. *J. Biol. Chem.*, 238:2351–2357, 1963.
- [3350] S.H. Pomerantz. 3,4-Dihydroxy-L-phenylalanine as the tyrosinase cofactor. Occurrence in melanoma and binding constant. *J. Biol. Chem.*, 242:5308–5314, 1967.
- [3351] A.L. Pometto and D.L. Crawford. Whole-cell bioconversion of vanillin to vanillic acid by *Streptomyces viridosporus*. *Appl. Environ. Microbiol.*, 45:1582–1585, 1983.
- [3352] B. Pontoppidan and C.G. Kannangara. Purification and partial characterisation of barley glutamyl-tRNA^{Glu} reductase, the enzyme that directs glutamate to chlorophyll biosynthesis. *Eur. J. Biochem.*, 225:529–537, 1994.
- [3353] S. Pontremoli, A. de Flora, E. Grazi, G. Mangiarotti, A. Bonsignore, and B.L. Horecker. Purification and properties of β -L-hydroxy acid dehydrogenase. II. Isolation of β -keto-L-gluconic acid, an intermediate in L-xylulose biosynthesis. *J. Biol. Chem.*, 236:2975–2980, 1961.
- [3354] A.T. Poret-Peterson, J.E. Graham, J. Gullede, and M.G. Klotz. Transcription of nitrification genes by the methane-oxidizing bacterium, *Methylococcus capsulatus* strain Bath. *ISME J.*, 2:1213–1220, 2008.
- [3355] R.J. Porra, W. Schafer, E. Cmiel, I. Katheder, and H. Scheer. The derivation of the formyl-group oxygen of chlorophyll *b* in higher plants from molecular oxygen. Achievement of high enrichment of the 7-formyl-group oxygen from ¹⁸O₂ in greening maize leaves. *Eur. J. Biochem.*, 219:671–679, 1994.
- [3356] D.H. Porter, R.J. Cook, and C. Wagner. Enzymatic properties of dimethylglycine dehydrogenase and sarcosine dehydrogenase from rat liver. *Arch. Biochem. Biophys.*, 243:396–407, 1985.
- [3357] D.J.T. Porter and H.J. Bright. Propionate-3-nitronate oxidase from *Penicillium atrovenerum* is a flavoprotein which initiates the autoxidation of its substrate by O₂. *J. Biol. Chem.*, 262:14428–14434, 1987.
- [3358] A.S. Pott and C. Dahl. Sirohaem sulfite reductase and other proteins encoded by genes at the *dsr* locus of *Chromatium vinosum* are involved in the oxidation of intracellular sulfur. *Microbiology*, 144:1881–1894, 1998.
- [3359] J.R.M. Potts, R. Weklych, and E.E. Conn. The 4-hydroxylation of cinnamic acid by sorghum microsomes and the requirement for cytochrome P-450. *J. Biol. Chem.*, 249:5019–5026, 1974.
- [3360] V.H. Potty and J.H. Bruemmer. Oxidation of geraniol by an enzyme system from orange. *Phytochemistry*, 9:1001–1007, 1970.
- [3361] R. Poulson. The enzymic conversion of protoporphyrinogen IX to protoporphyrin IX in mammalian mitochondria. *J. Biol. Chem.*, 251:3730–3733, 1976.

- [3362] R. Poulson and W.J. Polglase. The enzymic conversion of protoporphyrinogen IX to protoporphyrin IX. Protoporphyrinogen oxidase activity in mitochondrial extracts of *Saccharomyces cerevisiae*. *J. Biol. Chem.*, 250:1269–1274, 1975.
- [3363] M. Povelainen, E.V. Eneyskaya, A.A. Kulminskaya, D.R. Ivanen, N. Kalkkinen, K.N. Neustroev, and A.N. Miasnikov. Biochemical and genetic characterization of a novel enzyme of pentitol metabolism: D-arabitol-phosphate dehydrogenase. *Biochem. J.*, 371:191–197, 2003.
- [3364] J. Powlowski, L. Sahlman, and V. Shingler. Purification and properties of the physically associated meta-cleavage pathway enzymes 4-hydroxy-2-ketovaleate aldolase and aldehyde dehydrogenase (acylating) from *Pseudomonas* sp. strain CF600. *J. Bacteriol.*, 175:377–385, 1993.
- [3365] J. Powlowski, J. Sealy, V. Shingler, and E. Cadieux. On the role of DmpK, an auxiliary protein associated with multicomponent phenol hydroxylase from *Pseudomonas* sp. strain CF600. *J. Biol. Chem.*, 272:945–951, 1997.
- [3366] J. Powlowski and V. Shingler. *In vitro* analysis of polypeptide requirements of multicomponent phenol hydroxylase from *Pseudomonas* sp. strain CF600. *J. Bacteriol.*, 172:6834–6840, 1990.
- [3367] J.B. Powlowski, S. Dagley, V. Massey, and D.P. Ballou. Properties of anthranilate hydroxylase (deaminating), a flavoprotein from *Trichosporon cutaneum*. *J. Biol. Chem.*, 262:69–74, 1987.
- [3368] A. Prado-Cabrero, A.F. Estrada, S. Al-Babili, and J. Avalos. Identification and biochemical characterization of a novel carotenoid oxygenase: elucidation of the cleavage step in the *Fusarium* carotenoid pathway. *Mol. Microbiol.*, 64:448–460, 2007.
- [3369] R.L. Prairie and P. Talalay. Enzymatic formation of testolactone. *Biochemistry*, 2:203–208, 1963.
- [3370] J. Preiss. Sugar nucleotide reaction in *Arthrobacter*. II. Biosynthesis of guanosine diphosphomannuronate. *J. Biol. Chem.*, 239:3127–3132, 1964.
- [3371] J. Preiss and G. Ashwell. Alginic acid metabolism in bacteria. II. The enzymatic reduction of 4-deoxy-L-erythro-5-hexoseulose uronic acid to 2-keto-3-deoxy-D-gluconic acid. *J. Biol. Chem.*, 237:317–321, 1962.
- [3372] J. Preiss and G. Ashwell. Polygalacturonic acid metabolism in bacteria. II. Formation and metabolism of 3-deoxy-D-glycero-2,5-hexodiulosonic acid. *J. Biol. Chem.*, 238:1577–1583, 1963.
- [3373] M. Prejano, T. Marino, and N. Russo. How can methanol dehydrogenase from *Methylacidiphilum fumariolicum* work with the alien Ce(III) ion in the active center? A theoretical study. *Chemistry*, 23:8652–8657, 2017.
- [3374] R. Premkumar, P.V. Subba Rao, N.S. Streeleela, and C.S. Vaidyanathan. m-Hydroxybenzoic acid 4-hydroxylase from *Aspergillus niger*. *Can. J. Biochem.*, 47:825–827, 1969.
- [3375] D.J. Prescott and P.R. Vagelos. Acyl carrier protein. *Adv. Enzymol. Relat. Areas Mol. Biol.*, 36:269–311, 1972.
- [3376] A. Preusser, U. Wagner, T. Ellsner, and H.P. Kleber. Crotonobetaine reductase from *Escherichia coli* consists of two proteins. *Biochim. Biophys. Acta*, 1431:166–178, 1999.
- [3377] A. Preusser-Kunze, M. Mariappan, B. Schmidt, S.L. Gande, K. Mutenda, D. Wenzel, K. von Figura, and T. Dierks. Molecular characterization of the human C α -formylglycine-generating enzyme. *J. Biol. Chem.*, 280:14900–14910, 2005.
- [3378] A. Pribat, I.K. Blaby, A. Lara-Nunez, J.F. Gregory, de Crecy-Lagard 3rd, Hanson V., and A.D. FolX and FolM are essential for tetrahydromapterin synthesis in *Escherichia coli* and *Pseudomonas aeruginosa*. *J. Bacteriol.*, 192:475–482, 2010.
- [3379] H. Priefert, J. Rabenhorst, and A. Steinbuchel. Molecular characterization of genes of *Pseudomonas* sp. strain HR199 involved in bioconversion of vanillin to protocatechuate. *J. Bacteriol.*, 179:2595–2607, 1997.
- [3380] M.A. Prieto and J.L. Garcia. Molecular characterization of 4-hydroxyphenylacetate 3-hydroxylase of *Escherichia coli*. A two-protein component enzyme. *J. Biol. Chem.*, 269:22823–22829, 1994.
- [3381] M.A. Prieto, A. Perez-Aranda, and J.L. Garcia. Characterization of an *Escherichia coli* aromatic hydroxylase with a broad substrate range. *J. Bacteriol.*, 175:2162–2167, 1993.
- [3382] M.I. Prieto, J. Martin, R. Bala na Fouce, and A. Garrido-Pertierra. Properties of γ -aminobutyraldehyde dehydrogenase from *Escherichia coli*. *Biochimie*, 69:1161–1168, 1987.

- [3383] M.I. Prieto-Santos, J. Martin-Checa, R. Bala na Fouce, and A. Garrido-Pertierra. A pathway for putrescine catabolism in *Escherichia coli*. *Biochim. Biophys. Acta*, 880:242–244, 1986.
- [3384] S.T. Prigge, B.A. Eipper, R.E. Mains, and L.M. Amzel. Dioxygen binds end-on to mononuclear copper in a precatalytic enzyme complex. *Science*, 304:864–867, 2004.
- [3385] S.T. Prigge, A.S. Kolhekar, B.A. Eipper, R.E. Mains, and L.M. Amzel. Amidation of bioactive peptides: the structure of peptidylglycine α -hydroxylating monooxygenase. *Science*, 278:1300–1305, 1997.
- [3386] K.N. Prodouz and R.H. Garrett. *Neurospora crassa* NAD(P)H-nitrite reductase. Studies on its composition and structure. *J. Biol. Chem.*, 256:9711–9717, 1981.
- [3387] D.E. Prosser and G. Jones. Enzymes involved in the activation and inactivation of vitamin D. *Trends Biochem. Sci.*, 29:664–673, 2004.
- [3388] D.E. Prosser, M. Kaufmann, B. O’Leary, V. Byford, and G. Jones. Single A326G mutation converts human CYP24A1 from 25-OH-D₃-24-hydroxylase into -23-hydroxylase, generating 1 α ,25-(OH)₂D₃-26,23-lactone. *Proc. Natl. Acad. Sci. USA*, 104:12673–12678, 2007.
- [3389] A. Pružinská, I. Anders, S. Aubry, N. Schenk, E. Tapernoux-Lüthi, T. Müller, B. Kräutler, and S. Hörtensteiner. In vivo participation of red chlorophyll catabolite reductase in chlorophyll breakdown. *Plant Cell*, 19:369–387, 2007.
- [3390] A. Pružinská, G. Tanner, I. Anders, M. Roca, and S. Hörtensteiner. Chlorophyll breakdown: pheophorbide *a* oxygenase is a Rieske-type iron-sulfur protein, encoded by the accelerated cell death 1 gene. *Proc. Natl. Acad. Sci. USA*, 100:15259–15264, 2003.
- [3391] K.J. Puan, H. Wang, T. Dairi, T. Kuzuyama, and C.T. Morita. *fdA* is an essential gene required in the 2-C-methyl-D-erythritol 4-phosphate pathway for isoprenoid biosynthesis. *FEBS Lett.*, 579:3802–3806, 2005.
- [3392] E. Puentes-Cala, M. Liebeke, S. Markert, and J. Harder. Limonene dehydrogenase hydroxylates the allylic methyl group of cyclic monoterpenes in the anaerobic terpene degradation by *Castellaniella defragrans*. *J. Biol. Chem.*, 293:9520–9529, 2018.
- [3393] E.L. Pugh and M. Kates. Characterization of a membrane-bound phospholipid desaturase system of *Candida lipolytica*. *Biochim. Biophys. Acta*, 380:442–453, 1975.
- [3394] U. Puistola, T.M. Turpeenniemi-Hujanen, R. Myllyla, and K.I. Kivirikko. Studies on the lysyl hydroxylase reaction. I. Initial velocity kinetics and related aspects. *Biochim. Biophys. Acta*, 611:40–50, 1980.
- [3395] U. Puistola, T.M. Turpeenniemi-Hujanen, R. Myllyla, and K.I. Kivirikko. Studies on the lysyl hydroxylase reaction. II. Inhibition kinetics and the reaction mechanism. *Biochim. Biophys. Acta*, 611:51–60, 1980.
- [3396] E. Purwantini and L. Daniels. Purification of a novel coenzyme F₄₂₀-dependent glucose-6-phosphate dehydrogenase from *Mycobacterium smegmatis*. *J. Bacteriol.*, 178:2861–2866, 1996.
- [3397] E. Purwantini, T.P. Gillis, and L. Daniels. Presence of F₄₂₀-dependent glucose-6-phosphate dehydrogenase in *Mycobacterium* and *Nocardia* species, but absence from *Streptomyces* and *Corynebacterium* species and methanogenic *Archaea*. *FEMS Microbiol. Lett.*, 146:129–134, 1997.
- [3398] H. Qian, U. Edlund, J. Powlowski, V. Shingler, and I. Sethson. Solution structure of phenol hydroxylase protein component P2 determined by NMR spectroscopy. *Biochemistry*, 36:495–504, 1997.
- [3399] L. Qin, Y. Zhu, Z. Ding, X. Zhang, S. Ye, and R. Zhang. Structure of iridoid synthase in complex with NADP⁺/8-oxogeraniol reveals the structural basis of its substrate specificity. *J. Struct. Biol.*, 194:224–230, 2016.
- [3400] X. Qin, L. Sun, X. Wen, X. Yang, Y. Tan, H. Jin, Q. Cao, W. Zhou, Z. Xi, and Y. Shen. Structural insight into unique properties of protoporphyrinogen oxidase from *Bacillus subtilis*. *J. Struct. Biol.*, 170:76–82, 2010.
- [3401] X. Qin and J.A. Zeevaart. The 9-*cis*-epoxycarotenoid cleavage reaction is the key regulatory step of abscisic acid biosynthesis in water-stressed bean. *Proc. Natl. Acad. Sci. USA*, 96:15354–15361, 1999.
- [3402] J. Qiu, Y. Ma, Y. Wen, L. Chen, L. Wu, and W. Liu. Functional identification of two novel genes from *Pseudomonas* sp. strain HZN6 involved in the catabolism of nicotine. *Appl. Environ. Microbiol.*, 78:2154–2160, 2012.

- [3403] J. Qiu, Y. Wei, Y. Ma, R. Wen, Y. Wen, and W. Liu. A novel (S)-6-hydroxynicotine oxidase gene from *Shinella* sp. strain HZN7. *Appl. Environ. Microbiol.*, 80:5552–5560, 2014.
- [3404] W. Qiu, B. Zhou, D. Darwish, J. Shao, and Y. Yen. Characterization of enzymatic properties of human ribonucleotide reductase holoenzyme reconstituted in vitro from hRRM1, hRRM2, and p53R2 subunits. *Biochem. Biophys. Res. Commun.*, 340:428–434, 2006.
- [3405] X. Qiu, H. Hong, and S.L. MacKenzie. Identification of a Δ^4 fatty acid desaturase from *Thraustochytrium* sp. involved in the biosynthesis of docosahexanoic acid by heterologous expression in *Saccharomyces cerevisiae* and *Brassica juncea*. *J. Biol. Chem.*, 276:31561–31566, 2001.
- [3406] X. Qiu, D.W. Reed, H. Hong, S.L. MacKenzie, and P.S. Covello. Identification and analysis of a gene from *Calendula officinalis* encoding a fatty acid conjugase. *Plant Physiol.*, 125:847–855, 2001.
- [3407] Y. Qu, M.L. Easson, J. Froese, R. Simionescu, T. Hudlicky, and V. De Luca. Completion of the seven-step pathway from tabersonine to the anticancer drug precursor vindoline and its assembly in yeast. *Proc. Natl. Acad. Sci. USA*, 112:6224–6229, 2015.
- [3408] Y. Qu and J.C. Spain. Biodegradation of 5-nitroanthranilic acid by *Bradyrhizobium* sp. strain JS329. *Appl. Environ. Microbiol.*, 76:1417–1422, 2010.
- [3409] Y. Qu and J.C. Spain. Molecular and biochemical characterization of the 5-nitroanthranilic acid degradation pathway in *Bradyrhizobium* sp. strain JS329. *J. Bacteriol.*, 193:3057–3063, 2011.
- [3410] R. Quaderer, S. Omura, H. Ikeda, and D.E. Cane. Pentalenolactone biosynthesis. Molecular cloning and assignment of biochemical function to PtlI, a cytochrome P_{450} of *Streptomyces avermitilis*. *J. Am. Chem. Soc.*, 128:13036–13037, 2006.
- [3411] R. Quatrini, C. Appia-Ayme, Y. Denis, E. Jedlicki, D.S. Holmes, and V. Bonnefoy. Extending the models for iron and sulfur oxidation in the extreme acidophile *Acidithiobacillus ferrooxidans*. *BMC Genomics*, 10:394–394, 2009.
- [3412] J.R. Quayle. Formate dehydrogenase. *Methods Enzymol.*, 9:360–364, 1966.
- [3413] J.R. Quayle and G.A. Taylor. Carbon assimilation by *Pseudomonas oxalaticus* (OX1). 5. Purification and properties of glyoxylic dehydrogenase. *Biochem. J.*, 78:611–615, 1961.
- [3414] J. Quehenberger, T. Reichenbach, N. Baumann, L. Rettenbacher, C. Divne, and O. Spadiut. Kinetics and predicted structure of a novel xylose reductase from *Chaetomium thermophilum*. *Int. J. Mol. Sci.*, 20, 2019.
- [3415] A. Quemard, J.C. Sacchettini, A. Dessen, C. Vilcheze, R. Bittman, W.R. Jacobs, Blanchard Jr., and J.S. Enzymatic characterization of the target for isoniazid in *Mycobacterium tuberculosis*. *Biochemistry*, 34:8235–8241, 1995.
- [3416] E. Quemener, Y. Amet, S. di Stefano, G. Fournier, H.H. Floch, and J.H. Abalain. Purification of testosterone 5 α -reductase from human prostate by a four-step chromatographic procedure. *Steroids*, 59:712–718, 1994.
- [3417] M.W. Quong, C.G. Miyada, A.C. Switchenko, and T.C. Goodman. Identification, purification, and characterization of a D-arabinitol-specific dehydrogenase from *Candida tropicalis*. *Biochem. Biophys. Res. Commun.*, 196:1323–1329, 1993.
- [3418] T. Raab, J.A. Lopez-Raez, D. Klein, J.L. Caballero, E. Moyano, W. Schwab, and J. Munoz-Blanco. FaQR, required for the biosynthesis of the strawberry flavor compound 4-hydroxy-2,5-dimethyl-3(2H)-furanone, encodes an enone oxidoreductase. *Plant Cell*, 18:1023–1037, 2006.
- [3419] E. Racker. Aldehyde dehydrogenase, a diphosphopyridine nucleotide-linked enzyme. *J. Biol. Chem.*, 177:883–892, 1949.
- [3420] E. Racker. Glutathione-homocystine transhydrogenase. *J. Biol. Chem.*, 217:867–874, 1955.
- [3421] E. Racker. Glutathione reductase from bakers' yeast and beef liver. *J. Biol. Chem.*, 217:855–865, 1955.
- [3422] T.R. Radabaugh and H.V. Aposhian. Enzymatic reduction of arsenic compounds in mammalian systems: reduction of arsenate to arsenite by human liver arsenate reductase. *Chem. Res. Toxicol.*, 13:26–30, 2000.
- [3423] E.R. Rafanan, Hutchinson Jr., Shen C.R., and B. Triple hydroxylation of tetracenomycin A2 to tetracenomycin C involving two molecules of O₂ and one molecule of H₂O. *Org. Lett.*, 2:3225–3227, 2000.

- [3424] S.W. Ragsdale, J.E. Clark, L.G. Ljungdahl, L.L. Lundie, and H.L. Drake. Properties of purified carbon monoxide dehydrogenase from *Clostridium thermoaceticum*, a nickel, iron-sulfur protein. *J. Biol. Chem.*, 258:2364–2369, 1983.
- [3425] S. Ragnathan and H.R. Levy. Purification and characterization of the NAD-preferring glucose 6-phosphate dehydrogenase from *Acetobacter hansenii* (*Acetobacter xylinum*). *Arch. Biochem. Biophys.*, 310:360–366, 1994.
- [3426] A. Rahier, M. Bergdoll, G. Genot, F. Bouvier, and B. Camara. Homology modeling and site-directed mutagenesis reveal catalytic key amino acids of 3 β -hydroxysteroid-dehydrogenase/C4-decarboxylase from *Arabidopsis*. *Plant Physiol.*, 149:1872–1886, 2009.
- [3427] A. Rahier, S. Darnet, F. Bouvier, B. Camara, and M. Bard. Molecular and enzymatic characterizations of novel bifunctional 3 β -hydroxysteroid dehydrogenases/C-4 decarboxylases from *Arabidopsis thaliana*. *J. Biol. Chem.*, 281:27264–27277, 2006.
- [3428] A. Rahier, M. Smith, and M. Taton. The role of cytochrome *b*₅ in 4 α -methyl-oxidation and C5(6) desaturation of plant sterol precursors. *Biochem. Biophys. Res. Commun.*, 236:434–437, 1997.
- [3429] M.A. Rahim and C.J. Sih. Mechanisms of steroid oxidation by microorganisms. XI. Enzymatic cleavage of the pregnane side chain. *J. Biol. Chem.*, 241:3615–3623, 1966.
- [3430] A.D. Rahimtula and J.L. Gaylor. Partial purification of a microsomal sterol 4 α -carboxylic acid decarboxylase. *J. Biol. Chem.*, 247:9–15, 1972.
- [3431] A. Raisig, G. Bartley, P. Scolnik, and G. Sandmann. Purification in an active state and properties of the 3-step phytoene desaturase from *Rhodobacter capsulatus* overexpressed in *Escherichia coli*. *J. Biochem.*, 119:559–564, 1996.
- [3432] A. Raisig and G. Sandmann. 4,4'-diapophytoene desaturase: catalytic properties of an enzyme from the C₃₀ carotenoid pathway of *Staphylococcus aureus*. *J. Bacteriol.*, 181:6184–6187, 1999.
- [3433] A. Raisig and G. Sandmann. Functional properties of diapophytoene and related desaturases of C₃₀ to C₄₀ carotenoid biosynthetic pathways. *Biochim. Biophys. Acta*, 1533:164–170, 2001.
- [3434] J.K. Raison, G. Henson, and K.G. Rienits. The oxidation of gentisaldehyde by nicotinamide-adenine dinucleotide-specific, aromatic aldehyde dehydrogenase from rabbit liver. *Biochim. Biophys. Acta*, 118:285–298, 1966.
- [3435] R. Rajagopal. Metabolism of indole-3-acetaldehyde. III. Some characteristics of the aldehyde oxidase of *Avena coleoptiles*. *Physiol. Plant.*, 24:272–281, 1971.
- [3436] K.V. Rajagopalan and P. Handler. Purification and properties of chicken liver xanthine dehydrogenase. *J. Biol. Chem.*, 242:4097–4107, 1967.
- [3437] L.J. Rajakovich, M.E. Pandelia, A.J. Mitchell, W.C. Chang, B. Zhang, A.K. Boal, C., Bollinger Krebs, , and Jr. A new microbial pathway for organophosphonate degradation catalyzed by two previously misannotated non-heme-iron oxygenases. *Biochemistry*, 58:1627–1647, 2019.
- [3438] J. Rajniak, B. Barco, N.K. Clay, and E.S. Sattely. A new cyanogenic metabolite in *Arabidopsis* required for inducible pathogen defence. *Nature*, 525:376–379, 2015.
- [3439] J. Rajniak, R.FH. Giehl, E. Chang, I. Murgia, N. von Wiren, and E.S. Sattely. Biosynthesis of redox-active metabolites in response to iron deficiency in plants. *Nat. Chem. Biol.*, 14:442–450, 2018.
- [3440] L. Ralston, S.T. Kwon, M. Schoenbeck, J. Ralston, D.J. Schenk, R.M. Coates, and J. Chappell. Cloning, heterologous expression, and functional characterization of 5-*epi*-aristolochene-1,3-dihydroxylase from tobacco (*Nicotiana tabacum*). *Arch. Biochem. Biophys.*, 393:222–235, 2001.
- [3441] M. Ramachandra, R. Seetharam, M.H. Emptage, and F.S. Sariaslani. Purification and characterization of a soybean flour-inducible ferredoxin reductase of *Streptomyces griseus*. *J. Bacteriol.*, 173:7106–7112, 1991.
- [3442] C.V. Ramakrishnan and S.M. Martin. Isocitric dehydrogenase in *Aspergillus niger*. *Arch. Biochem. Biophys.*, 55:403–407, 1955.
- [3443] T. Ramakrishnan and J.J.R. Campbell. Gluconic dehydrogenase of *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta*, 17:122–127, 1955.

- [3444] R. Ramaley, Y. Fujita, and E. Freese. Purification and properties of *Bacillus subtilis* inositol dehydrogenase. *J. Biol. Chem.*, 254:7684–7690, 1979.
- [3445] P.B. Raman, D.C. Sharma, and R.I. Dorfman. Studies on aldosterone biosynthesis in vitro. *Biochemistry*, 5:1795–1795, 1966.
- [3446] B.V. Ramasastri and R.L. Blakley. 5,10-Methylenetetrahydrofolic acid dehydrogenase from Bakers' yeast. I. Partial purification and some properties. *J. Biol. Chem.*, 237:1982–1988, 1962.
- [3447] J.M. Ramirez, F.F. Del Campo, A. Paneque, and M. Losada. Ferredoxin-nitrite reductase from spinach. *Biochim. Biophys. Acta*, 118:58–71, 1966.
- [3448] T. Rand, T. Halkier, and O.C. Hansen. Structural characterization and mapping of the covalently linked FAD cofactor in choline oxidase from *Arthrobacter globiformis*. *Biochemistry*, 42:7188–7194, 2003.
- [3449] N.K. Ranjith, Ch.V. Ramana, and Ch. Sasikala. Purification and characterization of 3,4-dihydroxyphenylalanine oxidative deaminase from *Rhodobacter sphaeroides* OU5. *Can. J. Microbiol.*, 54:829–834, 2008.
- [3450] K.S. Rao, M. Albro, T.M. Dwyer, and F.E. Frerman. Kinetic mechanism of glutaryl-CoA dehydrogenase. *Biochemistry*, 45:15853–15861, 2006.
- [3451] P.V. Rao, C.M., Zigler Krishna, , and Jr. Identification and characterization of the enzymatic activity of zeta-crystallin from guinea pig lens. A novel NADPH:quinone oxidoreductase. *J. Biol. Chem.*, 267:96–102, 1992.
- [3452] P.V.S. Rao and C.S. Vaidyanathan. Studies on the metabolism of *o*-aminophenol. Purification and properties of isophenoxazine synthase from *Bauhenia monandra*. *Arch. Biochem. Biophys.*, 118:388–394, 1967.
- [3453] J. Ras, P.W. van Ophem, W.N. Reijnders, R.J. Van Spanning, J.A. Duine, A.H. Stouthamer, and N. Harms. Isolation, sequencing, and mutagenesis of the gene encoding NAD- and glutathione-dependent formaldehyde dehydrogenase (GD-FALDH) from *Paracoccus denitrificans*, in which GD-FALDH is essential for methylotrophic growth. *J. Bacteriol.*, 177:247–251, 1995.
- [3454] A.G. Rasmusson, K.L. Soole, and T.E. Elthon. Alternative NAD(P)H dehydrogenases of plant mitochondria. *Annu. Rev. Plant Biol.*, 55:23–39, 2004.
- [3455] L.J. Rather, B. Knapp, W. Haehnel, and G. Fuchs. Coenzyme A-dependent aerobic metabolism of benzoate via epoxide formation. *J. Biol. Chem.*, 285:20615–20624, 2010.
- [3456] B. Rathinasabapathi, M. Burnet, B.L. Russell, D.A. Gage, P. Liao, G.J. Nye, P. Scott, J.H. Golbeck, and A.D. Hanson. Choline monooxygenase, an unusual iron-sulfur enzyme catalyzing the first step of glycine betaine synthesis in plants: Prosthetic group characterization and cDNA cloning. *Proc. Natl. Acad. Sci. USA*, 94:3454–3458, 1997.
- [3457] M. Ratliff, W. Zhu, R. Deshmukh, A. Wilks, and I. Stojiljkovic. Homologues of neisserial heme oxygenase in gram-negative bacteria: degradation of heme by the product of the *pigA* gene of *Pseudomonas aeruginosa*. *J. Bacteriol.*, 183:6394–6403, 2001.
- [3458] H. Rauchová, R. Fato, Z. Drahotá, and G. Lenaz. Steady-state kinetics of reduction of coenzyme Q analogs by glycerol-3-phosphate dehydrogenase in brown adipose tissue mitochondria. *Arch. Biochem. Biophys.*, 344:235–241, 1997.
- [3459] S. Ravasio, B. Curti, and M.A. Vanoni. Determination of the midpoint potential of the FAD and FMN flavin cofactors and of the 3Fe-4S cluster of glutamate synthase. *Biochemistry*, 40:5533–5541, 2001.
- [3460] S. Ravasio, L. Dossena, E. Martin-Figueroa, F.J. Florencio, A. Mattevi, P. Morandi, B. Curti, and M.A. Vanoni. Properties of the recombinant ferredoxin-dependent glutamate synthase of *Synechocystis* PCC6803. Comparison with the *Azospirillum brasilense* NADPH-dependent enzyme and its isolated α subunit. *Biochemistry*, 41:8120–8133, 2002.
- [3461] R.G. Ravdin and D.I. Crandall. The enzymatic conversion of homogentisic acid to 4-fumarylacetoacetic acid. *J. Biol. Chem.*, 189:137–149, 1951.
- [3462] M. Rawat and Y. Av-Gay. Mycothiol-dependent proteins in actinomycetes. *FEMS Microbiol. Rev.*, 31:278–292, 2007.

- [3463] H.C. Rawden, G.O. Kokwaro, S.A. Ward, and G. Edwards. Relative contribution of cytochromes *P*-450 and flavin-containing monooxygenases to the metabolism of albendazole by human liver microsomes. *Br. J. Clin. Pharmacol.*, 49:313–322, 2000.
- [3464] A.B. Rawitch, G. Pollock, S.X. Yang, and A. Taugo. Thyroid peroxidase glycosylation: the location and nature of the N-linked oligosaccharide units in porcine thyroid peroxidase. *Arch. Biochem. Biophys.*, 297:321–327, 1992.
- [3465] M. Ray and S. Ray. On the interaction of nucleotides and glycolytic intermediates with NAD-linked α -ketoaldehyde dehydrogenase. *J. Biol. Chem.*, 257:10571–10574, 1982.
- [3466] M. Ray and S. Ray. Purification and partial characterization of a methylglyoxal reductase from goat liver. *Biochim. Biophys. Acta*, 802:119–127, 1984.
- [3467] S. Ray and M. Ray. Purification and characterization of NAD and NADP-linked α -ketoaldehyde dehydrogenases involved in catalyzing the oxidation of methylglyoxal to pyruvate. *J. Biol. Chem.*, 257:10566–10570, 1982.
- [3468] J.K. Rayton and E.D. Harris. Induction of lysyl oxidase with copper. Properties of an in vitro system. *J. Biol. Chem.*, 254:621–626, 1979.
- [3469] C.J. Rebouche and A.G. Engel. Tissue distribution of carnitine biosynthetic enzymes in man. *Biochim. Biophys. Acta*, 630:22–29, 1980.
- [3470] M.A. Recny and L.P. Hager. Reconstitution of native *Escherichia coli* pyruvate oxidase from apoenzyme monomers and FAD. *J. Biol. Chem.*, 257:12878–12886, 1982.
- [3471] A.S. Reddy and T.L. Thomas. Expression of a cyanobacterial Δ^6 -desaturase gene results in γ -linolenic acid production in transgenic plants. *Nat. Biotechnol.*, 14:639–642, 1996.
- [3472] C.C. Reddy, J.S. Swan, and G.A. Hamilton. *myo*-Inositol oxygenase from hog kidney. I. Purification and characterization of the oxygenase and of an enzyme complex containing the oxygenase and D-glucuronate reductase. *J. Biol. Chem.*, 256:8510–8518, 1981.
- [3473] C.C. Reddy and C.S. Vaidyanathan. Purification, properties and induction of a specific benzoate-4-hydroxylase from *Aspergillus niger* (UBC 814). *Biochim. Biophys. Acta*, 384:46–57, 1975.
- [3474] C.S. Reddy, S.H. Lee, J.S. Yoon, J.K. Kim, S.W. Lee, M. Hur, S.C. Koo, J. Meilan, W.M. Lee, J.K. Jang, Y. Hur, S.U. Park, and A.YB. Kim. Molecular cloning and characterization of carotenoid pathway genes and carotenoid content in *Ixeris dentata* var. *albiflora*. *Molecules*, 22, 2017.
- [3475] T.L.P. Reddy, P.M. Suryanarayana, and T.A. Venkitasubramanian. Variations in the pathways of malate oxidation and phosphorylation in different species of *Mycobacteria*. *Biochim. Biophys. Acta*, 376:210–218, 1975.
- [3476] Y.V. Reddy, A.H. Al Temimi, P.B. White, and J. Mecinovic. Evidence that trimethyllysine hydroxylase catalyzes the formation of (2*S*,3*S*)-3-hydroxy-*N*^e-trimethyllysine. *Org. Lett.*, 19:400–403, 2017.
- [3477] D.W. Reed and P.L. Hartzell. The *Archaeoglobus fulgidus* D-lactate dehydrogenase is a Zn²⁺ flavoprotein. *J. Bacteriol.*, 181:7580–7587, 1999.
- [3478] L.J. Reed. A trail of research from lipoic acid to α -keto acid dehydrogenase complexes. *J. Biol. Chem.*, 276:38329–38336, 2001.
- [3479] L.J. Reed, F.H. Pettit, M.H. Eley, L. Hamilton, J.H. Collins, and R.M. Oliver. Reconstitution of the *Escherichia coli* pyruvate dehydrogenase complex. *Proc. Natl. Acad. Sci. USA*, 72:3068–3072, 1975.
- [3480] M. Rees. Studies of the hydroxylamine metabolism of *Nitrosomonas europaea*. I. Purification of hydroxylamine oxidase. *Biochemistry*, 7:353–366, 1968.
- [3481] C.D. Reeve, M.A. Carver, and D.J. Hopper. Stereochemical aspects of the oxidation of 4-ethylphenol by the bacterial enzyme 4-ethylphenol methylenehydroxylase. *Biochem. J.*, 269:815–819, 1990.
- [3482] J.N. Reeve, G.S. Beckler, D.S. Cram, P.T. Hamilton, J.W. Brown, J.A. Krzycki, A.F. Kolodziej, L. Alex, W.H. Orme-Johnson, and C.T. Walsh. A hydrogenase-linked gene in *Methanobacterium thermoautotrophicum* strain δ H encodes a polyferredoxin. *Proc. Natl. Acad. Sci. USA*, 86:3031–3035, 1989.

- [3483] C.D. Reeves, S.L. Ward, W.P. Reville, H. Suzuki, M. Marcus, O.V. Petrakovsky, S. Marquez, H. Fu, S.D. Dong, and L. Katz. Production of hybrid 16-membered macrolides by expressing combinations of polyketide synthase genes in engineered *Streptomyces fradiae* hosts. *Chem. Biol.*, 11:1465–1472, 2004.
- [3484] R.E. Reeves, F.E. Montalvo, and T.S. Lushbaugh. Nicotinamide-adenine dinucleotide phosphate-dependent alcohol dehydrogenase. Enzyme from *Entamoeba histolytica* and some enzyme inhibitors. *Int. J. Biochem.*, 2:55–64, 1971.
- [3485] R.E. Reeves, L.G. Warren, B. Susskind, and H.-S. Lo. An energy-conserving pyruvate-to-acetate pathway in *Entamoeba histolytica*. Pyruvate synthase and a new acetate thiokinase. *J. Biol. Chem.*, 252:726–731, 1977.
- [3486] M. Reher and P. Schonheit. Glyceraldehyde dehydrogenases from the thermoacidophilic euryarchaeota *Picrophilus torridus* and *Thermoplasma acidophilum*, key enzymes of the non-phosphorylative Entner-Doudoroff pathway, constitute a novel enzyme family within the aldehyde dehydrogenase superfamily. *FEBS Lett.*, 580:1198–1204, 2006.
- [3487] G.A. Reid, C.S. Miles, R.K. Moysey, K.L. Pankhurst, and S.K. Chapman. Catalysis in fumarate reductase. *Biochim. Biophys. Acta*, 1459:310–315, 2000.
- [3488] U. Rein, R. Gueta, K. Denger, J. Ruff, K. Hollemeyer, and A.M. Cook. Dissimilation of cysteate via 3-sulfolactate sulfo-lyase and a sulfate exporter in *Paracoccus pantotrophus* NKNCYSA. *Microbiology*, 151:737–747, 2005.
- [3489] M. Reinartz, J. Tschape, T. Bruser, H.G. Truper, and C. Dahl. Sulfide oxidation in the phototrophic sulfur bacterium *Chromatium vinosum*. *Arch. Microbiol.*, 170:59–68, 1998.
- [3490] A.M. Reiner. Metabolism of aromatic compounds in bacteria. Purification and properties of the catechol-forming enzyme, 3,5-cyclohexadiene-1,2-diol-1-carboxylic acid (NAD⁺) oxidoreductase (decarboxylating). *J. Biol. Chem.*, 247:4960–4965, 1972.
- [3491] D.F. Reingold, A. Kawasaki, and P. Needleman. A novel prostaglandin 11-keto reductase found in rabbit liver. *Biochim. Biophys. Acta*, 659:179–188, 1981.
- [3492] B. Reinhammar and B.G. Malmström. "Blue" copper-containing oxidases. In T.G. Spiro, editor, *Copper Proteins*, pages 109–149. Copper Proteins, New York, 1981.
- [3493] C.R. Reisch, M.J. Stoudemayer, V.A. Varaljay, I.J. Amster, M.A. Moran, and W.B. Whitman. Novel pathway for assimilation of dimethylsulphoniopropionate widespread in marine bacteria. *Nature*, 473:208–211, 2011.
- [3494] M.V. Relling, R. Evans, C. Dass, D.M. Desiderio, and J. Nemeč. Human cytochrome *P*₄₅₀ metabolism of teniposide and etoposide. *J. Pharmacol. Exp. Ther.*, 261:491–496, 1992.
- [3495] E. Rembeza, A. Boverio, M.W. Fraaije, and M.K.M. Engqvist. Discovery of two novel oxidases using a high-throughput activity screen. *ChemBioChem*, 2021.
- [3496] H. Rembold and F. Simmersbach. Catabolism of pteridine cofactors. II. A specific pterin deaminase in rat liver. *Biochim. Biophys. Acta*, 184:589–596, 1969.
- [3497] S. Ren, H. Liu, E. Licad, and M.A. Correia. Expression of rat liver tryptophan 2,3-dioxygenase in *Escherichia coli*: structural and functional characterization of the purified enzyme. *Arch. Biochem. Biophys.*, 333:96–102, 1996.
- [3498] S. Ren, D. Marques, K. Redford, P.B. Hylemon, G. Gil, Z.R. Vlahcevic, and W.M. Pandak. Regulation of oxysterol 7 α -hydroxylase (CYP7B1) in the rat. *Metabolism*, 52:636–642, 2003.
- [3499] V. Renganathan. Possible involvement of toluene-2,3-dioxygenase in defluorination of 3-fluoro-substituted benzenes by toluene-degrading *Pseudomonas* sp. strain T-12. *Appl. Exp. Microbiol.*, 55:330–334, 1989.
- [3500] M.L. Reniere, G.N. Ukpabi, S.R. Harry, D.F. Stec, R. Krull, D.W. Wright, B.O. Bachmann, M.E. Murphy, and E.P. Skaar. The IsdG-family of haem oxygenases degrades haem to a novel chromophore. *Mol. Microbiol.*, 75:1529–1538, 2010.
- [3501] A.G.C. Renwick and L.L. Engel. The partial purification of 17 α - and 17 β -estradiol dehydrogenase activities from chicken liver. *Biochim. Biophys. Acta*, 146:336–348, 1967.
- [3502] L. Requena and S. Bornemann. Barley (*Hordeum vulgare*) oxalate oxidase is a manganese-containing enzyme. *Biochem. J.*, 343:185–190, 1999.

- [3503] M. Resch, H. Dobbek, and O. Meyer. Structural and functional reconstruction in situ of the [CuSMoO₂] active site of carbon monoxide dehydrogenase from the carbon monoxide oxidizing eubacterium *Oligotropha carboxidovorans*. *J. Biol. Inorg. Chem.*, 10:518–528, 2005.
- [3504] H. Rettenmaier and F. Lingens. Purification and some properties of two isofunctional juglone hydroxylases from *Pseudomonas putida* J1. *Biol. Chem. Hoppe-Seyler*, 366:637–646, 1985.
- [3505] K. Reuter, M. Pittelkow, J. Bursy, A. Heine, T. Craan, and E. Bremer. Synthesis of 5-hydroxyectoine from ectoine: crystal structure of the non-heme iron(II) and 2-oxoglutarate-dependent dioxygenase EctD. *PLoS One*, 5:e10647–e10647, 2010.
- [3506] K.A. Reynolds, P. Wang, K.M. Fox, M.K. Speedie, Y. Lam, and H.G. Floss. Purification and characterization of a novel enoyl coenzyme A reductase from *Streptomyces collinus*. *J. Bacteriol.*, 174:3850–3854, 1992.
- [3507] T. Rezen, N. Debeljak, D. Kordis, and D. Rozman. New aspects on lanosterol 14 α -demethylase and cytochrome P450 evolution: lanosterol/cycloartenol diversification and lateral transfer. *J. Mol. Evol.*, 59:51–58, 2004.
- [3508] S.K. Rhee and G. Fuchs. Phenylacetyl-CoA:acceptor oxidoreductase, a membrane-bound molybdenum-iron-sulfur enzyme involved in anaerobic metabolism of phenylalanine in the denitrifying bacterium *Thauera aromatica*. *Eur. J. Biochem.*, 262:507–515, 1999.
- [3509] J.G. Rheinwald, A.M. Chakrabarty, and I.C. Gunsalus. A transmissible plasmid controlling camphor oxidation in *Pseudomonas putida*. *Proc. Natl. Acad. Sci. USA*, 70:885–889, 1973.
- [3510] R.E. Rhoads and S. Udenfriend. Decarboxylation of α -ketoglutarate coupled to collagen proline hydroxylase. *Proc. Natl. Acad. Sci. USA*, 60:1473–1478, 1968.
- [3511] D.W. Ribbons. Bacterial oxidation of 2,3-dihydroxybenzoic acid - a new oxygenase. *Biochem. J.*, 99:30–30, 1966.
- [3512] A.L. Ribeiro, G. Degiacomi, F. Ewann, S. Buroni, M.L. Incandela, L.R. Chiarelli, G. Mori, J. Kim, M. Contreras-Dominguez, Y.S. Park, S.J. Han, P. Brodin, G. Valentini, M. Rizzi, G. Riccardi, and M.R. Pasca. Analogous mechanisms of resistance to benzothiazinones and dinitrobenzamides in *Mycobacterium smegmatis*. *PLoS One*, 6:e26675–e26675, 2011.
- [3513] R.B. Richerson and D.M. Ziegler. Cysteamine dioxygenase. *Methods Enzymol.*, 143:410–415, 1987.
- [3514] W. Richmond. Preparation and properties of a cholesterol oxidase from *Nocardia* sp. and its application to the enzymatic assay of total cholesterol in serum. *Clin. Chem.*, 19:1350–1356, 1973.
- [3515] A. Richter, C. Schaff, Z. Zhang, A.E. Lipka, F. Tian, T.G. Kollner, C. Schnee, S. Preiss, S. Irmisch, G. Jander, W. Boland, J. Gershenzon, E.S. Buckler, and J. Degenhardt. Characterization of biosynthetic pathways for the production of the volatile homoterpenes DMNT and TMTT in *Zea mays*. *Plant Cell*, 28:2651–2665, 2016.
- [3516] C.D. Richter, J.W. Allen, C.W. Higham, A. Koppenhofer, R.S. Zajicek, N.J. Watmough, and S.J. Ferguson. Cytochrome cd1, reductive activation and kinetic analysis of a multifunctional respiratory enzyme. *J. Biol. Chem.*, 277:3093–3100, 2002.
- [3517] L.W. Rider, M.B. Ottosen, S.G. Gattis, and B.A. Palfey. Mechanism of dihydrouridine synthase 2 from yeast and the importance of modifications for efficient tRNA reduction. *J. Biol. Chem.*, 284:10324–10333, 2009.
- [3518] J.P. Ridge, M. Lin, E.I. Larsen, M. Fegan, A.G. McEwan, and L.I. Sly. A multicopper oxidase is essential for manganese oxidation and laccase-like activity in *Pedomicrobium* sp. ACM 3067. *Environ. Microbiol.*, 9:944–953, 2007.
- [3519] J. Riemer, N. Bulleid, and J.M. Herrmann. Disulfide formation in the ER and mitochondria: two solutions to a common process. *Science*, 324:1284–1287, 2009.
- [3520] D. Riendeau, A. Rodrigues, and E. Meighen. Resolution of the fatty acid reductase from *Photobacterium phosphoreum* into acyl protein synthetase and acyl-CoA reductase activities. Evidence for an enzyme complex. *J. Biol. Chem.*, 257:6908–6915, 1982.
- [3521] L.U. Rigo, L.R. Maréchal, M.M. Vieira, and L.A. Veiga. Oxidative pathway for L-rhamnose degradation in *Pallularia pullulans*. *Can. J. Microbiol.*, 31:817–822, 1985.

- [3522] L.U. Rigo, M. Nakano, L.A. Veiga, and D.S. Feingold. L-Rhamnose dehydrogenase of *Pullularia pullulans*. *Biochim. Biophys. Acta*, 445:286–293, 1976.
- [3523] E. Rilkis and D. Rittenberg. Some observations on the enzyme, hydrogenase. *J. Biol. Chem.*, 236:2526–2529, 1961.
- [3524] K.L. Ringer, M.E. McConkey, E.M. Davis, G.W. Rushing, and R. Croteau. Monoterpene double-bond reductases of the (-)-menthol biosynthetic pathway: isolation and characterization of cDNAs encoding (-)-isopiperitenone reductase and (+)-pulegone reductase of peppermint. *Arch. Biochem. Biophys.*, 418:80–92, 2003.
- [3525] R.L. Ringler. Studies on the mitochondrial α -glycerophosphate dehydrogenase. II. Extraction and partial purification of the dehydrogenase from pig brain. *J. Biol. Chem.*, 236:1192–1198, 1961.
- [3526] P. Rippert and M. Matringe. Purification and kinetic analysis of the two recombinant arogenate dehydrogenase isoforms of *Arabidopsis thaliana*. *Eur. J. Biochem.*, 269:4753–4761, 2002.
- [3527] J. Risteli, K. Tryggvason, and K.I. Kivirikko. Prolyl 3-hydroxylase: partial characterization of the enzyme from rat kidney cortex. *Eur. J. Biochem.*, 73:485–492, 1977.
- [3528] J. Risteli, K. Tryggvason, and K.I. Kivirikko. A rapid assay for prolyl 3-hydroxylase activity. *Anal. Biochem.*, 84:423–431, 1978.
- [3529] J. Rivas, M. G. Guerrero, A. Paneque, and M. Losada. Characterization of the nitrate-reducing system of the yeast *Torulopsis nitratophila*. *Plant Sci. Lett.*, 1:105–113, 1973.
- [3530] C. Rivera-Perez, M. Nouzova, M.E. Clifton, E.M. Garcia, E. LeBlanc, and F.G. Noriega. Aldehyde dehydrogenase 3 converts farnesal into farnesoic acid in the corpora allata of mosquitoes. *Insect Biochem. Mol. Biol.*, 43:675–682, 2013.
- [3531] D.K. Ro, G. Arimura, S.Y. Lau, E. Piers, and J. Bohlmann. Loblolly pine abietadienol/abietadienal oxidase PtAO (CYP720B1) is a multifunctional, multisubstrate cytochrome P450 monooxygenase. *Proc. Natl. Acad. Sci. USA*, 102:8060–8065, 2005.
- [3532] P.L. Roach, I.J. Clifton, V. Fulop, K. Harlos, G.J. Barton, J. Hajdu, I. Andersson, C.J. Schofield, and J.E. Baldwin. Crystal structure of isopenicillin N synthase is the first from a new structural family of enzymes. *Nature*, 375:700–704, 1995.
- [3533] D.A. Robb. Tyrosinase. In R. Lontie, editor, *Copper Proteins and Copper Enzymes*, volume 2, pages 207–240. CRC Press, Boca Raton, FL, 1984.
- [3534] L. Robbel, V. Helmetag, T.A. Knappe, and M.A. Marahiel. Consecutive enzymatic modification of ornithine generates the hydroxamate moieties of the siderophore erythrochelin. *Biochemistry*, 50:6073–6080, 2011.
- [3535] K.C. Robbins, E.L. Barnett, and N.H. Grant. Partial purification of porcine liver uricase. *J. Biol. Chem.*, 216:27–35, 1955.
- [3536] J. Roberts and H.J. Rosenfeld. Isolation, crystallization, and properties of indolyl-3-alkane α -hydroxylase. A novel tryptophan-metabolizing enzyme. *J. Biol. Chem.*, 252:2640–2647, 1977.
- [3537] S.A. Roberts, A. Weichsel, G. Grass, K. Thakali, J.T. Hazzard, G. Tollin, C. Rensing, and W.R. Montfort. Crystal structure and electron transfer kinetics of CueO, a multicopper oxidase required for copper homeostasis in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA*, 99:2766–2771, 2002.
- [3538] S.A. Roberts, G.F. Wildner, G. Grass, A. Weichsel, A. Ambrus, C. Rensing, and W.R. Montfort. A labile regulatory copper ion lies near the T1 copper site in the multicopper oxidase CueO. *J. Biol. Chem.*, 278:31958–31963, 2003.
- [3539] M.A. Robien, G.M. Clore, J.G. Omichinski, R.N. Perham, E. Appella, K. Sakaguchi, and A.M. Gronenborn. Three-dimensional solution structure of the E3-binding domain of the dihydrolipoamide succinyltransferase core from the 2-oxoglutarate dehydrogenase multienzyme complex of *Escherichia coli*. *Biochemistry*, 31:3463–3471, 1992.
- [3540] W.G. Robinson and M.J. Coon. Purification and properties of β -hydroxyisobutyric dehydrogenase. *J. Biol. Chem.*, 225:511–521, 1957.
- [3541] E. Rocca and F. Ghiretti. Purification and properties of D-glutamic acid oxidase from *Octopus vulgaris* Lam. *Arch. Biochem. Biophys.*, 77:336–349, 1958.

- [3542] B. Roche and E. Azoulay. Régulation des alcool-déshydrogénases chez *Saccharomyces cerevisiae*. *Eur. J. Biochem.*, 8:426–434, 1969.
- [3543] P.A. Roche, T.J. Moorehead, and G.A. Hamilton. Purification and properties of hog liver 4-hydroxyphenylpyruvate dioxygenase. *Arch. Biochem. Biophys.*, 216:62–73, 1982.
- [3544] D.C. Rockholm and H.Y. Yamamoto. Violaxanthin de-epoxidase. *Plant Physiol.*, 110:697–703, 1996.
- [3545] S. Rodoni, W. Mühlecker, M. Anderl, B. Krätler, D. Moser, H. Thomas, P. Matile, and S. Hörtensteiner. Chlorophyll breakdown in senescent chloroplasts. Cleavage of pheophorbide *a* in two enzymic steps. *Plant Physiol.*, 115:669–676, 1997.
- [3546] S. Rodoni, F. Vicentini, M. Schellenberg, P. Matile, and S. Hörtensteiner. Partial purification and characterization of red chlorophyll catabolite reductase, a stroma protein involved in chlorophyll breakdown. *Plant Physiol.*, 115:677–682, 1997.
- [3547] F. Rodriguez, D.L. Hallahan, J.A. Pickett, and F. Camps. Characterization of the Δ^{11} -palmitoyl-CoA-desaturase from *Spodoptera littoralis* (Lepidoptera:Noctuidae). *Insect Biochem. Mol. Biol.*, 22:143–148, 1992.
- [3548] H.M. Rodriguez, M. Vaysberg, A. Mikels, S. McCauley, A.C. Velayo, C. Garcia, and V. Smith. Modulation of lysyl oxidase-like 2 enzymatic activity by an allosteric antibody inhibitor. *J. Biol. Chem.*, 285:20964–20974, 2010.
- [3549] M. Rodriguez-Saiz, J.L. Barredo, M.A. Moreno, J.M. Fernandez-Canon, M.A. Penalva, and B. Diez. Reduced function of a phenylacetate-oxidizing cytochrome P450 caused strong genetic improvement in early phylogeny of penicillin-producing strains. *J. Bacteriol.*, 183:5465–5471, 2001.
- [3550] V.W. Rodwell. Δ^1 -piperidine-6-carboxylic acid and α -aminoadipic acid δ -semialdehyde. *Method Enzymol*, 17B:188–199, 1971.
- [3551] C.R. Roe and N.O. Kaplan. Purification and substrate specificities of bacterial hydroxysteroid dehydrogenases. *Biochemistry*, 8:5093–5103, 1969.
- [3552] W.L. Roelofs, W. Liu, G. Hao, H. Jiao, A.P. Linn Rooney, , and Jr. Evolution of moth sex pheromones via ancestral genes. *Proc. Natl. Acad. Sci. USA*, 99:13621–13626, 2002.
- [3553] M. Le Roes-Hill, C. Goodwin, and S. Burton. Phenoxazinone synthase: what's in a name. *Trends Biotechnol.*, 27:248–258, 2009.
- [3554] D. Roeser, A. Preusser-Kunze, B. Schmidt, K. Gasow, J.G. Wittmann, T. Dierks, K. von Figura, and M.G. Rudolph. A general binding mechanism for all human sulfatases by the formylglycine-generating enzyme. *Proc. Natl. Acad. Sci. USA*, 103:81–86, 2006.
- [3555] F. Rohdich, S. Hecht, K. Gärtner, P. Adam, C. Krieger, S. Amslinger, D. Arigoni, A. Bacher, and W. Eisenreich. Studies on the nonmevalonate terpene biosynthetic pathway: Metabolic role of IspH (LytB) protein. *Proc. Natl. Acad. Sci. USA*, 99:1158–1163, 2002.
- [3556] R.C. Röhrich, N. Englert, K. Troschke, A. Reichenberg, M. Hintz, F. Seeber, E. Balconi, A. Aliverti, G. Zanetti, U. Köhler, M. Pfeiffer, E. Beck, H. Jomaa, and J. Wiesner. Reconstitution of an apicoplast-localised electron transfer pathway involved in the isoprenoid biosynthesis of *Plasmodium falciparum*. *FEBS Lett.*, 579:6433–6438, 2005.
- [3557] T. Rohwerder and W. Sand. The sulfane sulfur of persulfides is the actual substrate of the sulfur-oxidizing enzymes from *Acidithiobacillus* and *Acidiphilium* spp. *Microbiology*, 149:1699–1710, 2003.
- [3558] K. Rojas-Jimenez, C. Sohlenkamp, O. Geiger, E. Martinez-Romero, D. Werner, and P. Vinuesa. A ClC chloride channel homolog and ornithine-containing membrane lipids of *Rhizobium tropici* CIAT899 are involved in symbiotic efficiency and acid tolerance. *Mol. Plant Microbe Interact.*, 18:1175–1185, 2005.
- [3559] S. Roje, S.Y. Chan, F. Kaplan, R.K. Raymond, D.W. Horne, D.R. Appling, and A.D. Hanson. Metabolic engineering in yeast demonstrates that *S*-adenosylmethionine controls flux through the methylenetetrahydrofolate reductase reaction *in vivo*. *J. Biol. Chem.*, 277:4056–4061, 2002.

- [3560] S. Roje, H. Wang, S.D. McNeil, R.K. Raymond, D.R. Appling, Y. Shachar-Hill, H.J. Bohnert, and A.D. Hanson. Isolation, characterization, and functional expression of cDNAs encoding NADH-dependent methylenetetrahydrofolate reductase from higher plants. *J. Biol. Chem.*, 274:36089–36096, 1999.
- [3561] M. Rolff, J. Schottenheim, H. Decker, and F. Tuczek. Copper-O₂ reactivity of tyrosinase models towards external monophenolic substrates: molecular mechanism and comparison with the enzyme. *Chem Soc Rev*, 40:4077–4098, 2011.
- [3562] A.H. Romano and W.J. Nickerson. Cystine reductase of pea seeds and yeast. *J. Biol. Chem.*, 208:409–416, 1954.
- [3563] M.C. Romano, R.D. Eckardt, P.E. Bender, T.B. Leonard, K.M. Straub, and J.F. Newton. Biochemical characterization of hepatic microsomal leukotriene B₄ hydroxylases. *J. Biol. Chem.*, 262:1590–1595, 1987.
- [3564] V. Romanov and R.P. Hausinger. NADPH-dependent reductive *ortho* dehalogenation of 2,4-dichlorobenzoic acid in *Corynebacterium sepedonicum* KZ-4 and Coryneform bacterium strain NTB-1 via 2,4-dichlorobenzoyl coenzyme A. *J. Bacteriol.*, 178:2656–2661, 1996.
- [3565] M.J. Romao, M. Archer, I. Moura, J.J. Moura, J. LeGall, R. Engh, M. Schneider, P. Hof, and R. Huber. Crystal structure of the xanthine oxidase-related aldehyde oxido-reductase from *D. gigas*. *Science*, 270:1170–1176, 1995.
- [3566] E. Romero, M. Fedkenheuer, S.W. Chocklett, J. Qi, M. Oppenheimer, and P. Sobrado. Dual role of NADP(H) in the reaction of a flavin dependent *N*-hydroxylating monooxygenase. *Biochim. Biophys. Acta*, 1824:850–857, 2012.
- [3567] S. Rondet, M. Taton, and A. Rahier. Identification, characterization, and partial purification of 4 α -carboxysterol-C3-dehydrogenase/ C⁴-decarboxylase from *Zea mays*. *Arch. Biochem. Biophys.*, 366:249–260, 1999.
- [3568] P.W. Roome, Philley Jr., Peterson J.C., and J.A. Purification and properties of putidaredoxin reductase. *J. Biol. Chem.*, 258:2593–2598, 1983.
- [3569] R.J. Roon, H.L. Even, and F. Larimore. Glutamate synthase: properties of the reduced nicotinamide adenine dinucleotide-dependent enzyme from *Saccharomyces cerevisiae*. *J. Bacteriol.*, 118:89–95, 1974.
- [3570] J.D. Ropp, I.C. Gunsalus, and S.G. Sligar. Cloning and expression of a member of a new cytochrome *P*-450 family: cytochrome *P*-450lin (CYP111) from *Pseudomonas incognita*. *J. Bacteriol.*, 175:6028–6037, 1993.
- [3571] F. Rosati, I. Bardazzi, P. De Blasi, L. Simi, D. Scarpi, A. Guarna, M. Serio, M.L. Racchi, and G. Danza. 5 α -Reductase activity in *Lycopersicon esculentum*: cloning and functional characterization of LeDET2 and evidence of the presence of two isoenzymes. *J. Steroid Biochem. Mol. Biol.*, 96:287–299, 2005.
- [3572] B. Rosche, B. Tshisuaka, S. Fetzner, and F. Lingens. 2-Oxo-1,2-dihydroquinoline 8-monooxygenase, a two-component enzyme system from *Pseudomonas putida* 86. *J. Biol. Chem.*, 270:17836–17842, 1995.
- [3573] Z.B. Rose and E. Racker. Formaldehyde dehydrogenase. *Methods Enzymol.*, 9:357–360, 1966.
- [3574] I.N. Rosenberg. Purification of iodotyrosine deiodinase from bovine thyroid. *Metabolism*, 19:785–798, 1970.
- [3575] L.L. Rosenberg and D.I. Arnon. The preparation and properties of a new glyceraldehyde-3-phosphate dehydrogenase from photosynthetic tissues. *J. Biol. Chem.*, 217:361–371, 1955.
- [3576] C. Rosenthal, U. Mueller, S. Panjikar, L. Sun, M. Ruppert, Y. Zhao, and J. Stockigt. Expression, purification, crystallization and preliminary X-ray analysis of perakine reductase, a new member of the aldo-keto reductase enzyme superfamily from higher plants. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 62:1286–1289, 2006.
- [3577] K.Z. Rosloniec, M.H. Wilbrink, J.K. Capyk, W.W. Mohn, M. Ostendorf, R. van der Geize, L. Dijkhuizen, and L.D. Eltis. Cytochrome P450 125 (CYP125) catalyses C26-hydroxylation to initiate sterol side-chain degradation in *Rhodococcus jostii* RHA1. *Mol. Microbiol.*, 74:1031–1043, 2009.
- [3578] S. Roth, K. Jung, H. Jung, R.K. Hommel, and H.P. Kleber. Crotonobetaine reductase from *Escherichia coli* - a new inducible enzyme of anaerobic metabolism of L(-)-carnitine. *Antonie Van Leeuwenhoek*, 65:63–69, 1994.
- [3579] R.A. Rothery, C.A. Trieber, and J.H. Weiner. Interactions between the molybdenum cofactor and iron-sulfur clusters of *Escherichia coli* dimethylsulfoxide reductase. *J. Biol. Chem.*, 274:13002–13009, 1999.
- [3580] H.A. Rothschild and E.S.G. Barron. The oxidation of betaine aldehyde by betaine aldehyde dehydrogenase. *J. Biol. Chem.*, 209:511–523, 1954.

- [3581] N. Rouhier, E. Gelhaye, and J.P. Jacquot. Glutaredoxin-dependent peroxiredoxin from poplar: protein-protein interaction and catalytic mechanism. *J. Biol. Chem.*, 277:13609–13614, 2002.
- [3582] P. Rowland, O. Björnberg, F.S. Nielsen, K.F. Jensen, and S. Larsen. The crystal structure of *Lactococcus lactis* dihydroorotate dehydrogenase A complexed with the enzyme reaction product throws light on its enzymatic function. *Protein Sci.*, 7:1269–1279, 1998.
- [3583] P. Rowland, S. Nørager, K.F. Jensen, and S. Larsen. Structure of dihydroorotate dehydrogenase B: electron transfer between two flavin groups bridged by an iron-sulphur cluster. *Structure*, 8:1227–1238, 2000.
- [3584] B. Rowley and A.F. Tucci. Homoisocitric dehydrogenase from yeast. *Arch. Biochem. Biophys.*, 141:499–510, 1970.
- [3585] R. Roy, A.L. Menon, and M.W.W. Adams. Aldehyde oxidoreductases from *Pyrococcus furiosus*. *Methods Enzymol.*, 331:132–144, 2001.
- [3586] J. Royo, G. Vancanneyt, A.G. Perez, C. Sanz, K. Stormann, S. Rosahl, and J.J. Sanchez-Serrano. Characterization of three potato lipoxygenases with distinct enzymatic activities and different organ-specific and wound-regulated expression patterns. *J. Biol. Chem.*, 271:21012–21019, 1996.
- [3587] A. Rozhkova and R. Glockshuber. Thermodynamic aspects of DsbD-mediated electron transport. *J. Mol. Biol.*, 380:783–788, 2008.
- [3588] K.L. Rozwadowski, G.G. Khachatourians, and G. Selvaraj. Choline oxidase, a catabolic enzyme in *Arthrobacter pascens*, facilitates adaptation to osmotic stress in *Escherichia coli*. *J. Bacteriol.*, 173:472–478, 1991.
- [3589] D.A. Rozwarski, C. Vilcheze, M. Sugantino, R. Bittman, and J.C. Sacchettini. Crystal structure of the *Mycobacterium tuberculosis* enoyl-ACP reductase, InhA, in complex with NAD⁺ and a C₁₆ fatty acyl substrate. *J. Biol. Chem.*, 274:15582–15589, 1999.
- [3590] P.A. Rubenstein and J.L. Strominger. Enzymatic synthesis of cytidine diphosphate 3,6-dideoxyhexoses. VII. Mechanistic roles of enzyme E1 and pyridoxamine 5'-phosphate in the formation of cytidine diphosphate-4-keto-3,6-dideoxy-D-glucose from cytidine diphosphate-4-keto-6-deoxy-D-glucose. *J. Biol. Chem.*, 249:3776–3781, 1974.
- [3591] S. Ruch, P. Beyer, H. Ernst, and S. Al-Babili. Retinal biosynthesis in Eubacteria: *in vitro* characterization of a novel carotenoid oxygenase from *Synechocystis* sp. PCC 6803. *Mol. Microbiol.*, 55:1015–1024, 2005.
- [3592] W. Rüdiger. Biosynthesis of chlorophyll *b* and the chlorophyll cycle. *Photosynth. Res.*, 74:187–193, 2002.
- [3593] J. Rudolph, A.H. Erbse, L.S. Behlen, and S.D. Copley. A radical intermediate in the conversion of pentachlorophenol to tetrachlorohydroquinone by *Sphingobium chlorophenolicum*. *Biochemistry*, 53:6539–6549, 2014.
- [3594] J. Rudolph, J. Kim, and S.D. Copley. Multiple turnovers of the nicotino-enzyme PdxB require α -keto acids as cosubstrates. *Biochemistry*, 49:9249–9255, 2010.
- [3595] M. Rueffer and M.H. Zenk. Berberine synthesis, the methylenedioxy group forming enzyme in berberine synthesis. *Tetrahedron Lett.*, 26:201–202, 1985.
- [3596] M. Rueffer and M.H. Zenk. Enzymatic formation of protopines by a microsomal cytochrome-*P*-450 system of *Corydalis vaginans*. *Tetrahedron Lett.*, 28:5307–5310, 1987.
- [3597] M. Rueffer and M.H. Zenk. Canadine synthase from *Thalictrum tuberosum* cell cultures catalyses the formation of the methylenedioxy bridge in berberine synthesis. *Phytochemistry*, 36:1219–1223, 1994.
- [3598] H.W. Ruelius, R.M. Kerwin, and F.W. Janssen. Carbohydrate oxidase, a novel enzyme from *Polyporus obtusus*. I. Isolation and purification. *Biochim. Biophys. Acta*, 167:493–500, 1968.
- [3599] J. Ruf and P. Carayon. Structural and functional aspects of thyroid peroxidase. *Arch. Biochem. Biophys.*, 445:269–277, 2006.
- [3600] A. Rugor, M. Tataruch, J. Staron, A. Dudzik, E. Niedzialkowska, P. Nowak, A. Hogendorf, A. Michalik-Zym, D.B. Napruszewska, A. Jarzebski, K. Szymanska, W. Bialas, and M. Szaleniec. Regioselective hydroxylation of cholecalciferol, cholesterol and other sterol derivatives by steroid C25 dehydrogenase. *Appl. Microbiol. Biotechnol.*, 101:1163–1174, 2017.

- [3601] A. Rugor, A. Wojcik-Augustyn, E. Niedzialkowska, S. Mordalski, J. Staron, A. Bojarski, and M. Szalaniec. Reaction mechanism of sterol hydroxylation by steroid C25 dehydrogenase - Homology model, reactivity and isoenzymatic diversity. *J. Inorg. Biochem.*, 173:28–43, 2017.
- [3602] Z. Rui, M. Sandy, B. Jung, and W. Zhang. Tandem enzymatic oxygenations in biosynthesis of epoxyquinone pharmacophore of manumycin-type metabolites. *Chem. Biol.*, 20:879–887, 2013.
- [3603] H. Ruis and O. Hoffmann-Ostenhof. Enzymic epimerization of sequoyitol to D-pinitol in *Trifolium incarnatum*. *Eur. J. Biochem.*, 7:442–448, 1969.
- [3604] M.E. Rumpho, G.E. Edwards, and W.H. Loescher. A pathway for photosynthetic carbon flow to mannitol in celery leaves. Activity and localization of key enzymes. *Plant Physiol.*, 73:869–873, 1983.
- [3605] M. Rupp and H. Gorisch. Purification, crystallisation and characterization of quinoprotein ethanol dehydrogenase from *Pseudomonas aeruginosa*. *Biol. Chem. Hoppe-Seyler*, 369:431–439, 1988.
- [3606] B.L. Russell, B. Rathinasabapathi, and A.D. Hanson. Osmotic stress induces expression of choline monooxygenase in sugar beet and amaranth. *Plant Physiol.*, 116:859–865, 1998.
- [3607] D.W. Russell. The enzymes, regulation, and genetics of bile acid synthesis. *Annu. Rev. Biochem.*, 72:137–174, 2003.
- [3608] D.W. Russell and E.E. Conn. The cinnamic acid 4-hydroxylase of pea seedlings. *Arch. Biochem. Biophys.*, 122:256–268, 1967.
- [3609] W.J. Rutter and H.A. Lardy. Purification and properties of pigeon liver malic enzyme. *J. Biol. Chem.*, 233:374–382, 1958.
- [3610] F.J. Ruzicka and H. Beinert. A new iron-sulfur flavoprotein of the respiratory chain. A component of the fatty acid β -oxidation pathway. *J. Biol. Chem.*, 252:8440–8445, 1977.
- [3611] K.J. Ryan and L.L. Engel. Hydroxylation of steroids at carbon 21. *J. Biol. Chem.*, 225:103–114, 1957.
- [3612] P.R. Ryan, Q. Liu, P. Sperling, B. Dong, S. Franke, and E. Delhaize. A higher plant Δ^8 sphingolipid desaturase with a preference for (Z)-isomer formation confers aluminum tolerance to yeast and plants. *Plant Physiol.*, 144:1968–1977, 2007.
- [3613] J.Y. Ryu, J. Seo, S. Park, J.H. Ahn, Y. Chong, M.J. Sadowsky, and H.G. Hur. Characterization of an isoeugenol monooxygenase (iem) from *Pseudomonas nitroreducens* Jin1 that transforms isoeugenol to vanillin. *Biosci. Biotechnol. Biochem.*, 77:289–294, 2013.
- [3614] J.Y. Ryu, J. Seo, T. Unno, J.H. Ahn, T. Yan, M.J. Sadowsky, and H.G. Hur. Isoeugenol monooxygenase and its putative regulatory gene are located in the eugenol metabolic gene cluster in *Pseudomonas nitroreducens* Jin1. *Arch. Microbiol.*, 192:201–209, 2010.
- [3615] M.T. Ryzlak and R. Pietruszko. Heterogeneity of glyceraldehyde-3-phosphate dehydrogenase from human brain. *Biochim. Biophys. Acta*, 954:309–324, 1988.
- [3616] M.T. Ryzlak and R. Pietruszko. Human brain "high K_m " aldehyde dehydrogenase: purification, characterization, and identification as NAD⁺-dependent succinic semialdehyde dehydrogenase. *Arch. Biochem. Biophys.*, 266:386–396, 1988.
- [3617] Barz W. Clemens S. Cytochrome P_{450} -dependent methylenedioxy bridge formation in *Cicer arietinum*. *Phytochemistry*, 41:457–460, 1996.
- [3618] L. Saa, A. Jaureguibeitia, E. Largo, M.J. Llama, and J.L. Serra. Cloning, purification and characterization of two components of phenol hydroxylase from *Rhodococcus erythropolis* UPV-1. *Appl. Microbiol. Biotechnol.*, 86:201–211, 2010.
- [3619] L.L. Saari and R.V. Klucas. Ferric leghemoglobin reductase from soybean root nodules. *Arch. Biochem. Biophys.*, 231:102–113, 1984.
- [3620] D.J. Sabo and J.A. Orlando. Isolation, purification, and some properties of reduced nicotinamide adenine dinucleotide phosphate-cytochrome c_2 reductase from *Rhodopseudomonas spheroides*. *J. Biol. Chem.*, 243:3742–3749, 1968.

- [3621] P. Sachelaru, E. Schiltz, and R. Brandsch. A functional *mobA* gene for molybdopterine cytosine dinucleotide cofactor biosynthesis is required for activity and holoenzyme assembly of the heterotrimeric nicotine dehydrogenases of *Arthrobacter nicotinovorans*. *Appl. Environ. Microbiol.*, 72:5126–5131, 2006.
- [3622] J.C. Sadana and W.D. McElroy. Nitrate reductase from *Achromobacter fischeri*. Purification and properties: function of flavins and cytochrome. *Arch. Biochem. Biophys.*, 67:16–34, 1957.
- [3623] J.C. Sadana and A.V. Morey. Purification and properties of the hydrogenase of *Desulfovibrio desulfuricans*. *Biochim. Biophys. Acta*, 50:153–163, 1961.
- [3624] L. Saelices, L. Youssar, I. Holdermann, S. Al-Babili, and J. Avalos. Identification of the gene responsible for torulene cleavage in the *Neurospora* carotenoid pathway. *Mol. Genet. Genomics*, 278:527–537, 2007.
- [3625] A. Saffert, J. Hartmann-Schreier, A. Schon, and P. Schreier. A dual function α -dioxygenase-peroxidase and NAD(+) oxidoreductase active enzyme from germinating pea rationalizing α -oxidation of fatty acids in plants. *Plant Physiol.*, 123:1545–1552, 2000.
- [3626] M.K. Safo, F.N. Musayev, and V. Schirch. Structure of *Escherichia coli* pyridoxine 5'-phosphate oxidase in a tetragonal crystal form: insights into the mechanistic pathway of the enzyme. *Acta Crystallogr. D Biol. Crystallogr.*, 61:599–604, 2005.
- [3627] H. Sagami, A. Kurisaki, and K. Ogura. Formation of dolichol from dehydrodolichol is catalyzed by NADPH-dependent reductase localized in microsomes of rat liver. *J. Biol. Chem.*, 268:10109–10113, 1993.
- [3628] D. Sagher, D. Brunell, J.F. Hejtmancik, M. Kantorow, N. Brot, and H. Weissbach. Thionin can serve as a reducing agent for the methionine sulfoxide reductases. *Proc. Natl. Acad. Sci. USA*, 103:8656–8661, 2006.
- [3629] M. Sagi, R. Fluhr, and S.H. Lips. Aldehyde oxidase and xanthin dehydrogenase in a *flacca* tomato mutant with deficient abscisic acid and wilted phenotype. *Plant Physiol.*, 120:571–577, 1999.
- [3630] K. Saito, M. Kobayashi, Z. Gong, Y. Tanaka, and M. Yamazaki. Direct evidence for anthocyanidin synthase as a 2-oxoglutarate-dependent oxygenase: molecular cloning and functional expression of cDNA from a red form of *Perilla frutescens*. *Plant J.*, 17:181–190, 1999.
- [3631] K. Saito and A. Komamine. Biosynthesis of stizolobinic acid and stizolobic acid in higher plants. An enzyme system(s) catalyzing the conversion of dihydroxyphenylalanine into stizolobinic acid and stizolobic acid from etiolated seedlings of *Stizolobium hassjoo*. *Eur. J. Biochem.*, 68:237–243, 1976.
- [3632] K. Saito and A. Komamine. Biosynthesis of stizolobinic acid and stizolobic acid in higher plants. *Eur. J. Biochem.*, 82:385–392, 1978.
- [3633] S. Saito, N. Hirai, C. Matsumoto, H. Ohgashi, D. Ohta, K. Sakata, and M. Mizutani. *Arabidopsis* CYP707As encode (+)-abscisic acid 8'-hydroxylase, a key enzyme in the oxidative catabolism of abscisic acid. *Plant Physiol.*, 134:1439–1449, 2004.
- [3634] T. Saito, T. Nishino, and K. Tsushima. Interconversion between NAD-dependent and O₂-dependent types of rat liver xanthine dehydrogenase and difference in kinetic and redox properties between them. *Adv. Exp. Med. Biol.*, 253B:179–183, 1989.
- [3635] Y. Saito, O. Hayaishi, and S. Rothberg. Studies on oxygenases: enzymatic formation of 3-hydroxy-L-kynurenine from L-kynurenine. *J. Biol. Chem.*, 229:921–934, 1957.
- [3636] Y. Saito, S. Ito, A.M. Koltunow, and H. Sakai. Crystallization and preliminary X-ray analysis of geraniol dehydrogenase from *Backhousia citriodora* (lemon myrtle). *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 67:665–667, 2011.
- [3637] K. Sakai, N. Hamada, and Y. Watanabe. Separation of secondary alcohol oxidase and oxidized poly(vinyl alcohol) hydrolase by hydrophobic and dye-ligand chromatographies. *Agric. Biol. Chem.*, 47:153–155, 1983.
- [3638] R. Sakai, M. Fukuzawa, R. Nakano, S. Tatsuki, and Y. Ishikawa. Alternative suppression of transcription from two desaturase genes is the key for species-specific sex pheromone biosynthesis in two *Ostrinia* moths. *Insect Biochem. Mol. Biol.*, 39:62–67, 2009.

- [3639] T. Sakaki, N. Kagawa, K. Yamamoto, and K. Inouye. Metabolism of vitamin D₃ by cytochromes P₄₅₀. *Front. Biosci.*, 10:119–134, 2005.
- [3640] T. Sakaki, N. Sawada, K. Takeyama, S. Kato, and K. Inouye. Enzymatic properties of mouse 25-hydroxyvitamin D₃ 1 α -hydroxylase expressed in *Escherichia coli*. *Eur. J. Biochem.*, 259:731–738, 1999.
- [3641] T. Sakamoto, D.A. Los, S. Higashi, H. Wada, I. Nishida, M. Ohmori, and N. Murata. Cloning of ω^3 desaturase from cyanobacteria and its use in altering the degree of membrane-lipid unsaturation. *Plant Mol. Biol.*, 26:249–263, 1994.
- [3642] T. Sakamoto, H. Wada, I. Nishida, M. Ohmori, and N. Murata. Δ^9 Acyl-lipid desaturases of cyanobacteria. Molecular cloning and substrate specificities in terms of fatty acids, *sn*-positions, and polar head groups. *J. Biol. Chem.*, 269:25576–25580, 1994.
- [3643] R. Sakaue, T. Nakatsu, Y. Yamaguchi, H. Kato, and N. Kajiyama. Crystallization and preliminary crystallographic analysis of bacterial fructosyl amino acid oxidase. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:196–198, 2005.
- [3644] M. Sakuma, S. Kametani, and H. Akanuma. Purification and some properties of a hepatic NADPH-dependent reductase that specifically acts on 1,5-anhydro-D-fructose. *J. Biochem. (Tokyo)*, 123:189–193, 1998.
- [3645] E. Sakuno, K. Yabe, and H. Nakajima. Involvement of two cytosolic enzymes and a novel intermediate, 5'-oxoaverantin, in the pathway from 5'-hydroxyaverantin to averufin in aflatoxin biosynthesis. *Appl. Environ. Microbiol.*, 69:6418–6426, 2003.
- [3646] E. Sakuradani, T. Abe, K. Iguchi, and S. Shimizu. A novel fungal ω^3 -desaturase with wide substrate specificity from arachidonic acid-producing *Mortierella alpina* 1S-4. *Appl. Microbiol. Biotechnol.*, 66:648–654, 2005.
- [3647] S.A. Salisbury, H.S. Forrest, W.B.T. Cruse, and O. Kennard. A novel coenzyme from bacterial primary alcohol dehydrogenases. *Nature (Lond.)*, 280:843–844, 1979.
- [3648] E. Saller, H.R. Laue, H.R. Schläfli Oppenberg, and A.M. Cook. Purification and some properties of (1R,2S)-dihydroxy-3,5-cyclohexadiene-1,4-dicarboxylate dehydrogenase from *Comamonas testosteroni* T-2. *FEMS Microbiol. Lett.*, 130:97–102, 1996.
- [3649] S.P. Salowe, W.J. Krol, D. Iwatareyul, and C.A. Townsend. Elucidation of the order of oxidations and identification of an intermediate in the multistep clavaminic acid synthase reaction. *Biochemistry*, 30:2281–2292, 1991.
- [3650] N.N. Samsonova, S.V. Smirnov, A.E. Novikova, and L.R. Ptitsyn. Identification of *Escherichia coli* K12 YdcW protein as a γ -aminobutyraldehyde dehydrogenase. *FEBS Lett.*, 579:4107–4112, 2005.
- [3651] D.R. Sanadi, J.W. Littlefield, and R.M. Bock. Studies on α -ketoglutaric oxidase. II. Purification and properties. *J. Biol. Chem.*, 197:851–862, 1952.
- [3652] C. Sanchez, L. Zhu, A.F. Brana, A.P. Salas, J. Rohr, C. Mendez, and J.A. Salas. Combinatorial biosynthesis of antitumor indolocarbazole compounds. *Proc. Natl. Acad. Sci. USA*, 102:461–466, 2005.
- [3653] A. Sanchez-Ferrer, J.N. Rodriguez-Lopez, F. Garcia-Canovas, and F. Garcia-Carmona. Tyrosinase: a comprehensive review of its mechanism. *Biochim. Biophys. Acta*, 1247:1–11, 1995.
- [3654] A.S. Sandelius, R. Barr, F.L. Crane, and D.J. Morr . Redox reactions of plasma membranes isolated from soybean hypocotyls by phase partition. *Plant Sci.*, 48:1–10, 1986.
- [3655] H.K. Sanders, G.E. Becker, and A. Nason. Glycine-cytochrome *c* reductase from *Nitrobacter agilis*. *J. Biol. Chem.*, 247:2015–2025, 1972.
- [3656] P. Sandstrom, W.H. Welch, G.J. Blomquist, and C. Tittiger. Functional expression of a bark beetle cytochrome P450 that hydroxylates myrcene to ipsdienol. *Insect Biochem. Mol. Biol.*, 36:835–845, 2006.
- [3657] F.J. Sangari, J. Aguero, and J.M. Garcia-Lobo. The genes for erythritol catabolism are organized as an inducible operon in *Brucella abortus*. *Microbiology*, 146:487–495, 2000.
- [3658] P.C. Sanghani, C.L. Stone, B.D. Ray, E.V. Pindel, T.D. Hurley, and W.F. Bosron. Kinetic mechanism of human glutathione-dependent formaldehyde dehydrogenase. *Biochemistry*, 39:10720–10729, 2000.

- [3659] F. De Santa, M.G. Totaro, E. Prosperini, S. Notarbartolo, G. Testa, and G. Natoli. The histone H3 lysine-27 demethylase Jmjd3 links inflammation to inhibition of polycomb-mediated gene silencing. *Cell*, 130:1083–1094, 2007.
- [3660] J.M. Santini, U. Kappler, S.A. Ward, M.J. Honeychurch, R.N. vanden Hoven, and P.V. Bernhardt. The NT-26 cytochrome *c*₅₅₂ and its role in arsenite oxidation. *Biochim. Biophys. Acta*, 1767:189–196, 2007.
- [3661] P. Santoro and G. Parisi. A new enzyme from *Drosophila melanogaster* - in vitro conversion of xanthommatin into its dihydroform by means of xanthommatin reductase. *J. Exp. Zool.*, 239:169–173, 1986.
- [3662] B.D. Sanwal and M.W. Zink. L-Leucine dehydrogenase of *Bacillus cereus*. *Arch. Biochem. Biophys.*, 94:430–435, 1961.
- [3663] S. Sardar, A. Weitz, M.P. Hendrich, and B.S. Pierce. Outer-sphere tyrosine 159 within the 3-mercaptopropionic acid dioxygenase S-H-Y motif gates substrate-coordination denticity at the non-heme iron active site. *Biochemistry*, 58:5135–5150, 2019.
- [3664] N.S. Sargent and F.K. Habib. Partial purification of human prostatic 5 α -reductase (3-oxo-5 α -steroid:NADP⁺ 4-ene-oxido-reductase; EC 1.3.1.22) in a stable and active form. *J. Steroid Biochem. Mol. Biol.*, 38:73–77, 1991.
- [3665] F. Sarni, C. Grand, and A.M. Baudet. Purification and properties of cinnamoyl-CoA reductase and cinnamyl alcohol dehydrogenase from poplar stems (*Populus X euramericana*). *Eur. J. Biochem.*, 139:259–265, 1984.
- [3666] K.-I. Sasajima and A.J. Sinskey. Oxidation of L-glucose by a *Pseudomonad*. *Biochim. Biophys. Acta*, 571:120–126, 1979.
- [3667] D. Sasaki, M. Fujihashi, Y. Iwata, M. Murakami, T. Yoshimura, H. Hemmi, and K. Miki. Structure and mutation analysis of archaeal geranylgeranyl reductase. *J. Mol. Biol.*, 409:543–557, 2011.
- [3668] K. Sato, Y. Yamada, K. Aida, and T. Uemara. Enzymatic studies on the oxidation of sugar and sugar alcohol. 8. Particle-bound L-sorbose dehydrogenase from *Gluconobacter suboxydans*. *J. Biochem. (Tokyo)*, 66:521–527, 1969.
- [3669] M. Sato, M. Takahara, N. Kanno, Y. Sato, and W.R. Ellington. Isolation of a new opine, β -alanopine, from the extracts of the muscle of the marine bivalve mollusc *Scapharca broughtonii*. *Comp. Biochem. Physiol.*, 88B:803–806, 1987.
- [3670] S. Sato, M. Murakami, T. Yoshimura, and H. Hemmi. Specific partial reduction of geranylgeranyl diphosphate by an enzyme from the thermoacidophilic archaeon *Sulfolobus acidocaldarius* yields a reactive prenyl donor, not a dead-end product. *J. Bacteriol.*, 190:3923–3929, 2008.
- [3671] T. Sato and Y. Kobayashi. The ars operon in the skin element of *Bacillus subtilis* confers resistance to arsenate and arsenite. *J. Bacteriol.*, 180:1655–1661, 1998.
- [3672] T. Sato, T. Nakayama, S. Kikuchi, Y. Fukui, K. Yonekura-Sakakibara, T. Ueda, T. Nishino, Y. Tanaka, and T. Kusumi. Enzymatic formation of aurones in the extracts of yellow snapdragon flowers. *Plant Sci.*, 160:229–236, 2001.
- [3673] T. Satoh, M. Horie, H. Watanabe, Y. Tsuchiya, and T. Kamei. Enzymatic properties of squalene epoxidase from *Saccharomyces cerevisiae*. *Biol. Pharm. Bull.*, 16:349–352, 1993.
- [3674] T. Satomura, R. Kawakami, H. Sakuraba, and T. Ohshima. Dye-linked D-proline dehydrogenase from hyperthermophilic archaeon *Pyrobaculum islandicum* is a novel FAD-dependent amino acid dehydrogenase. *J. Biol. Chem.*, 277:12861–12867, 2002.
- [3675] T. Satyanarayana and A.N. Radhakrishnan. Biosynthesis of valine and isoleucine in plants. 3. Reductoisomerase of *Phaseolus radiatus*. *Biochim. Biophys. Acta*, 110:380–388, 1965.
- [3676] A.H. Saunders, J.H. Golbeck, and D.A. Bryant. Characterization of BciB: a ferredoxin-dependent 8-vinylprotochlorophyllide reductase from the green sulfur bacterium *Chloroherpeton thalassium*. *Biochemistry*, 52:8442–8451, 2013.
- [3677] P.P. Saunders and H.P. Broquist. Saccharopine, an intermediate of the amino adipic acid pathway of lysine biosynthesis. IV. Saccharopine dehydrogenase. *J. Biol. Chem.*, 241:3435–3440, 1966.
- [3678] M. Sauter, B. Tshisuaka, S. Fetzner, and F. Lingens. Microbial metabolism of quinoline and related compounds. XX. Quinaldic acid 4-oxidoreductase from *Pseudomonas* sp. AK-2 compared to other procaryotic molybdenum-containing hydroxylases. *Biol. Chem. Hoppe Seyler*, 374:1037–1046, 1993.

- [3679] N. Savage. Preparation and properties of highly purified diaphorase. *Biochem. J.*, 67:146–155, 1957.
- [3680] C. Savino, L.C. Montemiglio, G. Sciara, A.E. Miele, S.G. Kendrew, P. Jemth, S. Gianni, and B. Vallone. Investigating the structural plasticity of a cytochrome P450: three-dimensional structures of P450 EryK and binding to its physiological substrate. *J. Biol. Chem.*, 284:29170–29179, 2009.
- [3681] N. Sawada, T. Kusudo, T. Sakaki, S. Hatakeyama, M. Hanada, D. Abe, M. Kamao, T. Okano, M. Ohta, and K. Inouye. Novel metabolism of $1\alpha,25$ -dihydroxyvitamin D₃ with C₂₄-C₂₅ bond cleavage catalyzed by human CYP24A1. *Biochemistry*, 43:4530–4537, 2004.
- [3682] N. Sawada, T. Sakaki, S. Kitanaka, K. Takeyama, S. Kato, and K. Inouye. Enzymatic properties of human 25-hydroxyvitamin D₃ 1α -hydroxylase coexpression with adrenodoxin and NADPH-adrenodoxin reductase in *Escherichia coli*. *Eur. J. Biochem.*, 265:950–956, 1999.
- [3683] N. Sawada, T. Sakaki, S. Yoneda, T. Kusudo, R. Shinkyō, M. Ohta, and K. Inouye. Conversion of vitamin D₃ to $1\alpha,25$ -dihydroxyvitamin D₃ by *Streptomyces griseolus* cytochrome P450SU-1. *Biochem. Biophys. Res. Commun.*, 320:156–164, 2004.
- [3684] Y. Sawada and S. Ayabe. Multiple mutagenesis of P₄₅₀ isoflavonoid synthase reveals a key active-site residue. *Biochem. Biophys. Res. Commun.*, 330:907–913, 2005.
- [3685] Y. Sawada, K. Kinoshita, T. Akashi, T. Aoki, and S. Ayabe. Key amino acid residues required for aryl migration catalysed by the cytochrome P450 2-hydroxyisoflavanone synthase. *Plant J.*, 31:555–564, 2002.
- [3686] Y. Sawada, T. Ohyama, and I. Yamazaki. Preparation and physicochemical properties of green pea superoxide dismutase. *Biochim. Biophys. Acta*, 268:305–312, 1972.
- [3687] A.K. Saxena, P. Saxena, and V.M. Monnier. Purification and characterization of a membrane-bound deglycating enzyme (1-deoxyfructosyl alkyl amino acid oxidase, EC 1.5.3) from a *Pseudomonas* sp. soil strain. *J. Biol. Chem.*, 271:32803–32809, 1996.
- [3688] O. Sayanova, R. Haslam, M. Venegas Caleron, and J.A. Napier. Cloning and characterization of unusual fatty acid desaturases from *Anemone leveillei*: identification of an acyl-coenzyme A C₂₀ Δ^5 -desaturase responsible for the synthesis of sciadonic acid. *Plant Physiol.*, 144:455–467, 2007.
- [3689] O. Sayanova, M.A. Smith, P. Lapinskas, A.K. Stobart, G. Dobson, W.W. Christie, P.R. Shewry, and J.A. Napier. Expression of a borage desaturase cDNA containing an N-terminal cytochrome b₅ domain results in the accumulation of high levels of Δ^6 -desaturated fatty acids in transgenic tobacco. *Proc. Natl. Acad. Sci. USA*, 94:4211–4216, 1997.
- [3690] C.G. Sellsell, W.S. Tambyrajah, J.M. Murray, C.M. Wilmot, S.E. Phillips, M.J. McPherson, and P.F. Knowles. Probing the catalytic mechanism of *Escherichia coli* amine oxidase using mutational variants and a reversible inhibitor as a substrate analogue. *Biochem. J.*, 365:809–816, 2002.
- [3691] R.C. Scarpulla and R.L. Sofer. Membrane-bound proline dehydrogenase from *Escherichia coli*. Solubilization, purification, and characterization. *J. Biol. Chem.*, 253:5997–6001, 1978.
- [3692] J.C. Schabort and D.J.J. Potgieter. Cucurbitacin B Δ^{23} -reductase from *Cucurbita maxima*. II. Cofactor requirements, enzyme kinetics, substrate specificity and other characteristics. *Biochim. Biophys. Acta*, 151:47–53, 1968.
- [3693] J.C. Schabort and D.J.J. Potgieter. β -Cyclopiazonate oxidocyclase from *Penicillium cyclopium*. II. Studies on electron acceptors, inhibitors, enzyme kinetics, amino acid composition, flavin prosthetic group and other properties. *Biochim. Biophys. Acta*, 250:329–345, 1971.
- [3694] J.C. Schabort, D.J.J. Potgieter, and V. de Villiers. Cucurbitacin B Δ^{23} -reductase from *Cucurbita maxima*. I. Assay methods, isolation and purification. *Biochim. Biophys. Acta*, 151:33–46, 1968.
- [3695] S. Schach, G. Schwarz, S. Fetzner, and F. Lingens. Microbial metabolism of quinoline and related compounds. XVII. Degradation of 3-methylquinoline by *Comamonas testosteroni* 63. *Biol. Chem. Hoppe Seyler*, 374:175–181, 1993.
- [3696] S. Schach, B. Tshisuaka, S. Fetzner, and F. Lingens. Quinoline 2-oxidoreductase and 2-oxo-1,2-dihydroquinoline 5,6-dioxygenase from *Comamonas testosteroni* 63. The first two enzymes in quinoline and 3-methylquinoline degradation. *Eur. J. Biochem.*, 232:536–544, 1995.

- [3697] S.B. Schach, B. Tshisuaka, S. Fetzner, and F. Lingens. Quinoline 2-oxidoreductase and 2-oxo-1,2-dihydroquinoline 5,6-dioxygenase from *Comamonas testosteroni* 63. The first two enzymes in quinoline and 3-methylquinoline degradation. *Eur. J. Biochem.*, 232:536–544, 1995.
- [3698] H. Schachter, J. Sarney, E.J. McGuire, and S. Roseman. Isolation of diphosphopyridine nucleotide-dependent L-fucose dehydrogenase from pork liver. *J. Biol. Chem.*, 244:4785–4792, 1969.
- [3699] F. Schafer, U. Breuer, D. Benndorf, M. von Bergen, H. Harms, and R.H. Muller. Growth of *Aquicola tertiaricarbonis* L108 on *tert*-butyl alcohol leads to the induction of a phthalate dioxygenase-related protein and its associated oxidoreductase subunit. *Eng. Life Sci.*, 7:512–519, 2007.
- [3700] F. Schafer, J. Schuster, B. Wurz, C. Hartig, H. Harms, R.H. Muller, and T. Rohwerder. Synthesis of short-chain diols and unsaturated alcohols from secondary alcohol substrates by the Rieske nonheme mononuclear iron oxygenase MdpJ. *Appl. Environ. Microbiol.*, 78:6280–6284, 2012.
- [3701] J. Schalk, S. de Vries, J.G. Kuenen, and M.S. Jetten. Involvement of a novel hydroxylamine oxidoreductase in anaerobic ammonium oxidation. *Biochemistry*, 39:5405–5412, 2000.
- [3702] H.-P. Schär and H. Zuber. Structure and function of L-lactate dehydrogenases from thermophilic and mesophilic bacteria. I) Isolation and characterization of lactate dehydrogenases from thermophilic and mesophilic bacilli. *Hoppe-Seyler's Z. Physiol. Chem.*, 360:795–807, 1979.
- [3703] N.L. Schauer and J.G. Ferry. FAD requirement for the reduction of coenzyme F₄₂₀ by formate dehydrogenase from *Methanobacterium formicicum*. *J. Bacteriol.*, 155:467–472, 1983.
- [3704] N.L. Schauer and J.G. Ferry. Composition of the coenzyme F₄₂₀-dependent formate dehydrogenase from *Methanobacterium formicicum*. *J. Bacteriol.*, 165:405–411, 1986.
- [3705] S. Schauer, S. Chaturvedi, L. Randau, J. Moser, M. Kitabatake, S. Lorenz, E. Verkamp, W.D. Schubert, T. Nakayashiki, M. Murai, K. Wall, H.-U. Thomann, D.W. Heinz, H. Inokuchi, D. Söll, and D. Jahn. *Escherichia coli* glutamyl-tRNA reductase. Trapping the thioester intermediate. *J. Biol. Chem.*, 277:48657–48663, 2002.
- [3706] M. Schedel, M. Vanselow, and H. G. Trueper. Siroheme sulfite reductase from *Chromatium vinosum*. Purification and investigation of some of its molecular and catalytic properties. *Arch. Microbiol.*, 121:29–36, 1979.
- [3707] U. Scheler, W. Brandt, A. Porzel, K. Rothe, D. Manzano, D. Bozic, D. Papaefthimiou, G.U. Balcke, A. Henning, S. Lohse, S. Marillonnet, A.K. Kanellis, A. Ferrer, and A. Tissier. Elucidation of the biosynthesis of carnosic acid and its reconstitution in yeast. *Nat. Commun.*, 7:12942–12942, 2016.
- [3708] S. Schenk, A. Hoelz, B. Krauss, and K. Decker. Gene structures and properties of enzymes of the plasmid-encoded nicotine catabolism of *Arthrobacter nicotinovorans*. *J. Mol. Biol.*, 284:1323–1339, 1998.
- [3709] T. Schenk, R. Müller, F. Mörsberger, M.K. Otto, and F. Lingens. Enzymatic dehalogenation of pentachlorophenol by extracts from *Arthrobacter sp.* strain ATCC 33790. *J. Bacteriol.*, 171:5487–5491, 1989.
- [3710] P. Schenkels and J.A. Duine. Nicotinoprotein (NADH-containing) alcohol dehydrogenase from *Rhodococcus erythropolis* DSM 1069: an efficient catalyst for coenzyme-independent oxidation of a broad spectrum of alcohols and the interconversion of alcohols and aldehydes. *Microbiology*, 146:775–785, 2000.
- [3711] L. Schepers, P.P. Van Veldhoven, M. Casteels, H.J. Eyssen, and G.P. Mannaerts. Presence of three acyl-CoA oxidases in rat liver peroxisomes. An inducible fatty acyl-CoA oxidase, a noninducible fatty acyl-CoA oxidase, and a noninducible trihydroxycoprostanoyl-CoA oxidase. *J. Biol. Chem.*, 265:5242–5246, 1990.
- [3712] B.M. Scher and B.L. Horecker. Pentose metabolism in *Candida*. 3. The triphosphopyridine nucleotide-specific polyol dehydrogenase of *Candida utilis*. *Arch. Biochem. Biophys.*, 116:117–128, 1966.
- [3713] J.W. Schertzer, S.A. Brown, and M. Whiteley. Oxygen levels rapidly modulate *Pseudomonas aeruginosa* social behaviours via substrate limitation of PqsH. *Mol. Microbiol.*, 77:1527–1538, 2010.
- [3714] V. Scheumann, H. Ito, A. Tanaka, S. Schoch, and W. Rüdiger. Substrate specificity of chlorophyll(ide) *b* reductase in etioplasts of barley (*Hordeum vulgare* L.). *Eur. J. Biochem.*, 242:163–170, 1996.

- [3715] V. Scheumann, S. Schoch, and W. Rüdiger. Chlorophyll *a* formation in the chlorophyll *b* reductase reaction requires reduced ferredoxin. *J. Biol. Chem.*, 273:35102–35108, 1998.
- [3716] A. Schiefner, Q. Sinz, I. Neumaier, W. Schwab, and A. Skerra. Structural basis for the enzymatic formation of the key strawberry flavor compound 4-hydroxy-2,5-dimethyl-3(2*H*)-furanone. *J. Biol. Chem.*, 288:16815–16826, 2013.
- [3717] A. Schirmer, M.A. Rude, X. Li, E. Popova, and S.B. del Cardayre. Microbial biosynthesis of alkanes. *Science*, 329:559–562, 2010.
- [3718] H.W. Schiwara and G.F. Domagk. Über den Abbau der Desoxyzucker durch Bakterienenzyme. V. Anreicherung und Charakterisierung einer NADP-abhängigen Abequosedehydrogenase aus *Pseudomonas putida*. *Hoppe-Seyler's Z. Physiol. Chem.*, 349:1321–1329, 1968.
- [3719] H.W. Schiwara, W. Domschke, and G.F. Domagk. Über die Zucker-Dehydrogenase in der Säugetierleber. I. Differenzierung verschiedener Zucker-Dehydrogenasen in der Schweineleber durch Disk-Elektrophorese und Ionenaustauschchromatographie. *Hoppe-Seyler's Z. Physiol. Chem.*, 349:1575–1581, 1968.
- [3720] H.R. Schläfli, M.A. Weiss, T. Leisinger, and A.M. Cook. Terephthalate 1,2-dioxygenase system from *Comamonas testosteroni* T-2; purification and some properties of the oxygenase component. *J. Bacteriol.*, 176:6644–6652, 1994.
- [3721] W. Schlegel, S. Krüger, and K. Korte. Purification of prostaglandin E₂ 9-oxoreductase from human decidua vera. *FEBS Lett.*, 171:141–144, 1984.
- [3722] W. Schlenzka, L. Shaw, S. Kelm, C.L. Schmidt, E. Bill, A.X. Trautwein, F. Lottspeich, and R. Schauer. CMP-N-acetylneuraminic acid hydroxylase: the first cytosolic Rieske iron-sulphur protein to be described in Eukarya. *FEBS Lett.*, 385:197–200, 1996.
- [3723] M.J. Schlesinger and M.J. Coon. Reduction of mevaldic acid to mevalonic acid by a partial purified enzyme from liver. *J. Biol. Chem.*, 236:2421–2424, 1961.
- [3724] A. Schmidt and A. Trebst. The mechanism of photosynthetic sulfate reduction by isolated chloroplasts. *Biochim. Biophys. Acta*, 180:529–535, 1969.
- [3725] E.W. Schmidt, J.T. Nelson, D.A. Rasko, S. Sudek, J.A. Eisen, M.G. Haygood, and J. Ravel. Patellamide A and C biosynthesis by a microcin-like pathway in *Prochloron didemni*, the cyanobacterial symbiont of *Lissoclinum patella*. *Proc. Natl. Acad. Sci. USA*, 102:7315–7320, 2005.
- [3726] H. Schmidt, T. Dresselhaus, F. Buck, and E. Heinz. Purification and PCR-based cDNA cloning of a plastidial *n*-6 desaturase. *Plant Mol. Biol.*, 26:631–642, 1994.
- [3727] H. Schmidt and E. Heinz. Desaturation of oleoyl groups in envelope membranes from spinach chloroplasts. *Proc. Natl. Acad. Sci. USA*, 87:9477–9480, 1990.
- [3728] H. Schmidt and E. Heinz. Involvement of ferredoxin in desaturation of lipid-bound oleate in chloroplasts. *Plant Physiol.*, 94:214–220, 1990.
- [3729] H.L. Schmidt, W. Stocklein, J. Danzer, P. Kirch, and B. Limbach. Isolation and properties of an H₂O-forming NADH oxidase from *Streptococcus faecalis*. *Eur. J. Biochem.*, 156:149–155, 1986.
- [3730] U. Schmitt, K. Jahnke, K. Rosenbaum, P.F. Cook, and K.D. Schnackerz. Purification and characterization of dihydropyrimidine dehydrogenase from *Alcaligenes eutrophus*. *Arch. Biochem. Biophys.*, 332:175–182, 1996.
- [3731] P. Schneckenburger, L. Shaw, and R. Schauer. Purification, characterization and reconstitution of CMP-N-acetylneuraminic acid hydroxylase from mouse liver. *Glycoconj. J.*, 11:194–203, 1994.
- [3732] B.L. Schneider, A.K. Kiupakis, and L.J. Reitzer. Arginine catabolism and the arginine succinyltransferase pathway in *Escherichia coli*. *J. Bacteriol.*, 180:4278–4286, 1998.
- [3733] B.L. Schneider and L. Reitzer. Pathway and enzyme redundancy in putrescine catabolism in *Escherichia coli*. *J. Bacteriol.*, 194:4080–4088, 2012.
- [3734] C. Schneider, P. Boger, and G. Sandmann. Phytoene desaturase: heterologous expression in an active state, purification, and biochemical properties. *Protein Expr. Purif.*, 10:175–179, 1997.

- [3735] K. Schneider and H.G. Schlegel. Purification and properties of soluble hydrogenase from *Alcaligenes eutrophus* H 16. *Biochim. Biophys. Acta*, 452:66–80, 1976.
- [3736] K.H. Schneider, G. Jakel, R. Hoffmann, and F. Giffhorn. Enzyme evolution in *Rhodobacter sphaeroides*: selection of a mutant expressing a new galactitol dehydrogenase and biochemical characterization of the enzyme. *Microbiology*, 141:1865–1873, 1995.
- [3737] S. Schneider and G. Fuchs. Phenylacetyl-CoA:acceptor oxidoreductase, a new α -oxidizing enzyme that produces phenylglyoxylate. Assay, membrane localization, and differential production in *Thauera aromatica*. *Arch. Microbiol.*, 169:509–516, 1998.
- [3738] K. Schnurr, J. Belkner, F. Ursini, T. Schewe, and H. Kuhn. The selenoenzyme phospholipid hydroperoxide glutathione peroxidase controls the activity of the 15-lipoxygenase with complex substrates and preserves the specificity of the oxygenation products. *J. Biol. Chem.*, 271:4653–4658, 1996.
- [3739] M. Schobert and H. Gorisch. Cytochrome c_{550} is an essential component of the quinoprotein ethanol oxidation system in *Pseudomonas aeruginosa*: cloning and sequencing of the genes encoding cytochrome c_{550} and an adjacent acetaldehyde dehydrogenase. *Microbiology*, 145:471–481, 1999.
- [3740] G. Schoch, S. Goepfert, M. Morant, A. Hehn, D. Meyer, P. Ullmann, and D. Werck-Reichhart. CYP98A3 from *Arabidopsis thaliana* is a 3'-hydroxylase of phenolic esters, a missing link in the phenylpropanoid pathway. *J. Biol. Chem.*, 276:36566–36574, 2001.
- [3741] C. Schoenbohm, S. Martens, C. Eder, G. Forkmann, and B. Weisshaar. Identification of the *Arabidopsis thaliana* flavonoid 3'-hydroxylase gene and functional expression of the encoded P450 enzyme. *Biol. Chem.*, 381:749–753, 2000.
- [3742] A. Schoendorf, C.D. Rithner, R.M. Williams, and R.B. Croteau. Molecular cloning of a cytochrome P_{450} taxane 10 β -hydroxylase cDNA from *Taxus* and functional expression in yeast. *Proc. Natl. Acad. Sci. USA*, 98:1501–1506, 2001.
- [3743] P.V. Schoenlein, B.B. Roa, and M.E. Winkler. Divergent transcription of *pdxB* and homology between the *pdxB* and *serA* gene products in *Escherichia coli* K-12. *J. Bacteriol.*, 171:6084–6092, 1989.
- [3744] C.R. Schopfer, G. Kochs, F. Lottspeich, and J. Ebel. Molecular characterization and functional expression of dihydroxypterocarpan 6 α -hydroxylase, an enzyme specific for pterocarpanoid phytoalexin biosynthesis in soybean (*Glycine max* L.). *FEBS Lett.*, 432:182–186, 1998.
- [3745] W. Schöpp, H. Sorger, H.-P. Kleber, and H. Aurich. Kinetische Untersuchungen zum Reaktionsmechanismus der Carnitin-dehydrogenase aus *Pseudomonas aeruginosa*. *Eur. J. Biochem.*, 10:56–60, 1969.
- [3746] I. Schröder, S. Rech, T. Krafft, and J.M. Macy. Purification and characterization of the selenate reductase from *Thauera selenatis*. *J. Biol. Chem.*, 272:23765–23768, 1997.
- [3747] A. Schryvers, E. Lohmeier, and J.H. Weiner. Chemical and functional properties of the native and reconstituted forms of the membrane-bound, aerobic glycerol-3-phosphate dehydrogenase of *Escherichia coli*. *J. Biol. Chem.*, 253:783–788, 1978.
- [3748] H.L. Schubert, E. Raux, A.A. Brindley, H.K. Leech, K.S. Wilson, C.P. Hill, and M.J. Warren. The structure of *Saccharomyces cerevisiae* Met8p, a bifunctional dehydrogenase and ferrochelataze. *EMBO J.*, 21:2068–2075, 2002.
- [3749] R. Schuegger, M. Nafisi, M. Mansourova, B.L. Petersen, C.E. Olsen, A. Svatos, B.A. Halkier, and E. Glawischnig. CYP71B15 (PAD3) catalyzes the final step in camalexin biosynthesis. *Plant Physiol.*, 141:1248–1254, 2006.
- [3750] K. Schuhle, M. Saft, B. Vogeli, T.J. Erb, and J. Heider. Benzylmalonyl-CoA dehydrogenase, an enzyme involved in bacterial auxin degradation. *Arch. Microbiol.*, 203:4149–4159, 2021.
- [3751] D.J. Schuller, G.A. Grant, and L.J. Banaszak. The allosteric ligand site in the V_{max} -type cooperative enzyme phosphoglycerate dehydrogenase. *Nat. Struct. Biol.*, 2:69–76, 1995.
- [3752] S. Schulman, B. Wang, W. Li, and T.A. Rapoport. Vitamin K epoxide reductase prefers ER membrane-anchored thioredoxin-like redox partners. *Proc. Natl. Acad. Sci. USA*, 107:15027–15032, 2010.
- [3753] L.W. Schultz, L. Liu, M. Cegielski, and J.W. Hastings. Crystal structure of a pH-regulated luciferase catalyzing the bioluminescent oxidation of an open tetrapyrrole. *Proc. Natl. Acad. Sci. USA*, 102:1378–1383, 2005.

- [3754] R.M. Schultz, F.V. Groman, and L.L. Engel. 3(17) β -Hydroxysteroid dehydrogenase of *Pseudomonas testosteroni*. A convenient purification and demonstration of multiple molecular forms. *J. Biol. Chem.*, 252:3775–3783, 1977.
- [3755] H.-U. Schulze, H.-H. Schott, and H. Staudinger. Isolierung und Charakterisierung einer NADH: Semidehydroascorbinsäure-Oxidoreduktase aus *Neurospora crassa*. *Hoppe-Seyler's Z. Physiol. Chem.*, 353:1931–1942, 1972.
- [3756] H.-M. Schumacher and M.H. Zenk. Partial purification and characterization of dihydrobenzophenanthridine oxidase from *Eschscholtzia tenuifolia* cell suspension cultures. *Plant Cell Reports*, 7:43–46, 1988.
- [3757] W. Schumacher and C. Holliger. The proton/electron ratio of the menaquinone-dependent electron transport from dihydrogen to tetrachloroethene in "*Dehalobacter restrictus*". *J. Bacteriol.*, 178:2328–2333, 1996.
- [3758] W. Schumacher, C. Holliger, A.J.B. Zehnder, and W.R. Hagen. Redox chemistry of cobalamin and iron-sulfur cofactors in the tetrachloroethene reductase of *Dehalobacter restrictus*. *FEBS Lett.*, 409:421–425, 1997.
- [3759] M. Schuman and V. Massey. Purification and characterization of glycolic acid oxidase from pig liver. *Biochim. Biophys. Acta*, 227:500–520, 1971.
- [3760] J. Schuster, F. Schafer, N. Hubler, A. Brandt, M. Rosell, C. Hartig, H. Harms, R.H. Muller, and T. Rohwerder. Bacterial degradation of *tert*-amyl alcohol proceeds via hemiterpene 2-methyl-3-buten-2-ol by employing the tertiary alcohol desaturase function of the Rieske nonheme mononuclear iron oxygenase MdpJ. *J. Bacteriol.*, 194:972–981, 2012.
- [3761] G.J. Schut and M.W. Adams. The iron-hydrogenase of *Thermotoga maritima* utilizes ferredoxin and NADH synergistically: a new perspective on anaerobic hydrogen production. *J. Bacteriol.*, 191:4451–4457, 2009.
- [3762] G.J. Schut, S.L. Bridger, and M.W. Adams. Insights into the metabolism of elemental sulfur by the hyperthermophilic archaeon *Pyrococcus furiosus*: characterization of a coenzyme A- dependent NAD(P)H sulfur oxidoreductase. *J. Bacteriol.*, 189:4431–4441, 2007.
- [3763] G.J. Schut, A.L. Menon, and M.W.W. Adams. 2-Keto acid oxidoreductases from *Pyrococcus furiosus* and *Thermococcus litoralis*. *Methods Enzymol.*, 331:144–158, 2001.
- [3764] C. Schwalb, S.K. Chapman, and G.A. Reid. The membrane-bound tetrahaem *c*-type cytochrome CymA interacts directly with the soluble fumarate reductase in *Shewanella*. *Biochem Soc Trans.*, 30:658–662, 2002.
- [3765] J.K. Schwartz, P.P. Wei, K.H. Mitchell, B.G. Fox, and E.I. Solomon. Geometric and electronic structure studies of the binuclear nonheme ferrous active site of toluene-4-monooxygenase: parallels with methane monooxygenase and insight into the role of the effector proteins in O₂ activation. *J. Am. Chem. Soc.*, 130:7098–7109, 2008.
- [3766] S.H. Schwartz, K.M. Leon-Kloosterziel, M. Koornneef, and J.A. Zeevaart. Biochemical characterization of the *aba2* and *aba3* mutants in *Arabidopsis thaliana*. *Plant Physiol.*, 114:161–166, 1997.
- [3767] S.H. Schwartz, X. Qin, and M.C. Loewen. The biochemical characterization of two carotenoid cleavage enzymes from *Arabidopsis* indicates that a carotenoid-derived compound inhibits lateral branching. *J. Biol. Chem.*, 279:46940–46945, 2004.
- [3768] S.H. Schwartz, B.C. Tan, D.A. Gage, J.A. Zeevaart, and D.R. McCarty. Specific oxidative cleavage of carotenoids by VP14 of maize. *Science*, 276:1872–1874, 1997.
- [3769] M. Schwarz, A.C. Wright, D.L. Davis, H. Nazer, I. Bjorkhem, and D.W. Russell. The bile acid synthetic gene 3 β -hydroxy- Δ^5 -C²⁷-steroid oxidoreductase is mutated in progressive intrahepatic cholestasis. *J. Clin. Invest.*, 106:1175–1184, 2000.
- [3770] R. Schwarzenbacher, F. Stenner-Liewen, H. Liewen, J.C. Reed, and R.C. Liddington. Crystal structure of PqqC from *Klebsiella pneumoniae* at 2.1 Å resolution. *Proteins*, 56:401–403, 2004.
- [3771] A.I. Scott, C.A. Roessner, N.J. Stolowich, J.B. Spencer, C. Min, and S.I. Ozaki. Biosynthesis of vitamin B₁₂. Discovery of the enzymes for oxidative ring contraction and insertion of the fourth methyl group. *FEBS Lett.*, 331:105–108, 1993.
- [3772] D.B.M. Scott and S.S. Cohen. The oxidative pathway of carbohydrate metabolism in *Escherichia coli*. 1. The isolation and properties of glucose 6-phosphate dehydrogenase and 6-phosphogluconate dehydrogenase. *Biochem. J.*, 55:23–33, 1953.

- [3773] D.B.M. Scott and S.S. Cohen. The oxidative pathway of carbohydrate metabolism in *Escherichia coli*. 5. Isolation and identification of ribulose phosphate produced from 6-phosphogluconate by the dehydrogenase of *E. coli*. *Biochem. J.*, 65:686–689, 1957.
- [3774] J.C. Scott, I.V. Greenhut, and J.H. Leveau. Functional characterization of the bacterial *iac* genes for degradation of the plant hormone indole-3-acetic acid. *J Chem Ecol*, 39:942–951, 2013.
- [3775] P. Scriba and H. Holzer. Gewinnung von α -Hydroxyäthyl-2-thiaminpyrophosphat mit Pyruvatoxydase aus Schweineherzmuskel. *Biochem. Z.*, 334:473–486, 1961.
- [3776] N.S. Scrutton and M.J. Sutcliffe. Trimethylamine dehydrogenase and electron transferring flavoprotein. *Subcell. Biochem.*, 35:145–181, 2000.
- [3777] F.P. Seebeck. *In vitro* reconstitution of mycobacterial ergothioneine biosynthesis. *J. Am. Chem. Soc.*, 132:6632–6633, 2010.
- [3778] H. Seedorf, A. Dreisbach, R. Hedderich, S. Shima, and R.K. Thauer. F₄₂₀H₂ oxidase (FprA) from *Methanobrevibacter arboriphilus*, a coenzyme F₄₂₀-dependent enzyme involved in O₂ detoxification. *Arch. Microbiol.*, 182:126–137, 2004.
- [3779] H. Seedorf, C.H. Hagemeyer, S. Shima, R.K. Thauer, E. Warkentin, and U. Ermler. Structure of coenzyme F₄₂₀H₂ oxidase (FprA), a di-iron flavoprotein from methanogenic Archaea catalyzing the reduction of O₂ to H₂O. *FEBS J.*, 274:1588–1599, 2007.
- [3780] H. Seedorf, J. Kahnt, A.J. Pierik, and R.K. Thauer. *Si*-face stereospecificity at C5 of coenzyme F₄₂₀ for F₄₂₀H₂ oxidase from methanogenic Archaea as determined by mass spectrometry. *FEBS J.*, 272:5337–5342, 2005.
- [3781] J.E. Seegmiller. Triphosphopyridine nucleotide-linked aldehyde dehydrogenase from yeast. *J. Biol. Chem.*, 201:629–637, 1953.
- [3782] M. Seemann, B. Tse Sum Bui, M. Wolff, M. Miginiac-Maslow, and M. Rohmer. Isoprenoid biosynthesis in plant chloroplasts via the MEP pathway: direct thylakoid/ferredoxin-dependent photoreduction of GcpE/IspG. *FEBS Lett.*, 580:1547–1552, 2006.
- [3783] M. Seemann, B. Tse Sum Bui, M. Wolff, D. Tritsch, N. Campos, A. Boronat, A. Marquet, and M. Rohmer. Isoprenoid biosynthesis through the methylerythritol phosphate pathway: the (*E*)-4-hydroxy-3-methylbut-2-enyl diphosphate synthase (GcpE) is a [4Fe-4S] protein. *Angew. Chem. Int. Ed. Engl.*, 41:4337–4339, 2002.
- [3784] M. Seemann, P. Wegner, V. Schünemann, B. Tse Sum Bui, M. Wolff, A. Marquet, A.X. Trautwein, and M. Rohmer. Isoprenoid biosynthesis in chloroplasts via the methylerythritol phosphate pathway: the (*E*)-4-hydroxy-3-methylbut-2-enyl diphosphate synthase (GcpE) from *Arabidopsis thaliana* is a [4Fe-4S] protein. *J. Biol. Inorg. Chem.*, 10:131–137, 2005.
- [3785] B. Seibold, M. Matthes, M.H. Eppink, F. Lingens, W.J. Van Berkel, and R. Müller. 4-Hydroxybenzoate hydroxylase from *Pseudomonas* sp. CBS3. Purification, characterization, gene cloning, sequence analysis and assignment of structural features determining the coenzyme specificity. *Eur. J. Biochem.*, 239:469–478, 1996.
- [3786] J. Seidel, G. Schmitt, M. Hoffmann, D. Jendrossek, and O. Einsle. Structure of the processive rubber oxygenase RoxA from *Xanthomonas* sp. *Proc. Natl. Acad. Sci. USA*, 110:13833–13838, 2013.
- [3787] M.M. Seidman, A. Toms, and J.M. Wood. Influence of side-chain substituents on the position of cleavage of the benzene ring by *Pseudomonas fluorescens*. *J. Bacteriol.*, 97:1192–1197, 1969.
- [3788] H.U. Seitz and D.E. Gaertner. Enzymes in cardenolide-accumulating shoot cultures of *Digitalis purpurea*. *Plant Cell*, 38:337–344, 1994.
- [3789] H. Seki, K. Ohyama, S. Sawai, M. Mizutani, T. Ohnishi, H. Sudo, T. Akashi, T. Aoki, K. Saito, and T. Muranaka. Licorice β -amyrin 11-oxidase, a cytochrome P₄₅₀ with a key role in the biosynthesis of the triterpene sweetener glycyrrhizin. *Proc. Natl. Acad. Sci. USA*, 105:14204–14209, 2008.
- [3790] H. Seki, S. Sawai, K. Ohyama, M. Mizutani, T. Ohnishi, H. Sudo, E.O. Fukushima, T. Akashi, T. Aoki, K. Saito, and T. Muranaka. Triterpene functional genomics in licorice for identification of CYP72A154 involved in the biosynthesis of glycyrrhizin. *Plant Cell*, 23:4112–4123, 2011.

- [3791] N. Seki, K. Mori, T. Kitamura, M. Miyamoto, and A. Kihara. Yeast Mpo1 is a novel dioxygenase that catalyzes the α -oxidation of a 2-hydroxy fatty acid in an Fe²⁺-dependent manner. *Mol. Cell Biol.*, 39:e00428–18–e00428–18, 2019.
- [3792] Y. Seki, N. Sogawa, and M. Ishimoto. Siroheme as an active catalyst in sulfite reduction. *J. Biochem.*, 90:1487–1492, 1981.
- [3793] H. Sekimoto, M. Seo, N. Kawakami, T. Komano, S. Desloire, S. Liotenberg, A. Marion-Poll, M. Caboche, Y. Kamiya, and T. Koshiba. Molecular cloning and characterization of aldehyde oxidases in *Arabidopsis thaliana*. *Plant Cell Physiol.*, 39:433–442, 1998.
- [3794] Y. Sekiwa-Iijima, Y. Aizawa, and K. Kubota. Geraniol dehydrogenase activity related to aroma formation in ginger (*Zingiber officinale* Roscoe). *J. Agric. Food Chem.*, 49:5902–5906, 2001.
- [3795] S.A. Selifonov. Microbial oxidation of adamantanone by *Pseudomonas putida* carrying the camphor catabolic plasmid. *Biochem. Biophys. Res. Commun.*, 186:1429–1436, 1992.
- [3796] O.Z. Sellinger and O.N. Miller. The metabolism of acetol phosphate. II. 1,2-Propanediol-1-phosphate dehydrogenase. *J. Biol. Chem.*, 234:1641–1646, 1959.
- [3797] L. Sene, M.G. Felipe, S.S. Silva, and M. Vitolo. Preliminary kinetic characterization of xylose reductase and xylitol dehydrogenase extracted from *Candida guilliermondii* FTI 20037 cultivated in sugarcane bagasse hydrolysate for xylitol production. *Appl. Biochem. Biotechnol.*, 91-93:671–680, 2001.
- [3798] M. Seo, S. Akaba, T. Oritani, M. Delarue, C. Bellini, M. Caboche, and T. Koshiba. Higher activity of an aldehyde oxidase in the auxin-overproducing superroot1 mutant of *Arabidopsis thaliana*. *Plant Physiol.*, 116:687–693, 1998.
- [3799] M. Seo, H. Koiwai, S. Akaba, T. Komano, T. Oritani, Y. Kamiya, and T. Koshiba. Abscisic aldehyde oxidase in leaves of *Arabidopsis thaliana*. *Plant J.*, 23:481–488, 2000.
- [3800] M. Seo, A.J. Peeters, H. Koiwai, T. Oritani, A. Marion-Poll, J.A. Zeevaart, M. Koornneef, Y. Kamiya, and T. Koshiba. The *Arabidopsis* aldehyde oxidase 3 (*AAO3*) gene product catalyzes the final step in abscisic acid biosynthesis in leaves. *Proc. Natl. Acad. Sci. USA*, 97:12908–12913, 2000.
- [3801] M.J. Seo, D. Zhu, S. Endo, H. Ikeda, and D.E. Cane. Genome mining in *Streptomyces*. Elucidation of the role of Baeyer-Villiger monooxygenases and non-heme iron-dependent dehydrogenase/oxygenases in the final steps of the biosynthesis of pentalenolactone and neopentalenolactone. *Biochemistry*, 50:1739–1754, 2011.
- [3802] B. Setlow and P. Setlow. Levels of acetyl coenzyme A, reduced and oxidized coenzyme A, and coenzyme A in disulfide linkage to protein in dormant and germinated spores and growing and sporulating cells of *Bacillus megaterium*. *J. Bacteriol.*, 132:444–452, 1977.
- [3803] A. Setya, M. Murillo, and T. Leustek. Sulfate reduction in higher plants: Molecular evidence for a novel 5-adenylylphosphosulfate (APS) reductase. *Proc. Natl. Acad. Sci. USA*, 93:13383–13388, 1996.
- [3804] S. Setyahadi, T. Ueyama, T. Arimoto, N. Mori, and Y. Kitamoto. Purification and properties of a new enzyme, D-carnitine dehydrogenase, from *Agrobacterium* sp. 525a. *Biosci. Biotechnol. Biochem.*, 61:1055–1058, 1997.
- [3805] E. Setzke, R. Hedderich, S. Heiden, and R.K. Thauer. H₂: heterodisulfide oxidoreductase complex from *Methanobacterium thermoautotrophicum*. Composition and properties. *Eur. J. Biochem.*, 220:139–148, 1994.
- [3806] W. Seubert, I. Lamberts, R. Kramer, and B. Ohly. On the mechanism of malonyl-CoA-independent fatty acid synthesis. I. The mechanism of elongation of long-chain fatty acids by acetyl-CoA. *Biochim. Biophys. Acta*, 164:498–517, 1968.
- [3807] C.S. Sevier, J.W. Cuozzo, A. Vala, F. Aslund, and C.A. Kaiser. A flavoprotein oxidase defines a new endoplasmic reticulum pathway for biosynthetic disulfide bond formation. *Nat. Cell Biol.*, 3:874–882, 2001.
- [3808] I.F. Sevrioukova, C. Garcia, H. Li, B. Bhaskar, and T.L. Poulos. Crystal structure of putidaredoxin, the [2Fe-2S] component of the P450_{cam} monooxygenase system from *Pseudomonas putida*. *J. Mol. Biol.*, 333:377–392, 2003.
- [3809] I.F. Sevrioukova, H. Li, and T.L. Poulos. Crystal structure of putidaredoxin reductase from *Pseudomonas putida*, the final structural component of the cytochrome P450_{cam} monooxygenase. *J. Mol. Biol.*, 336:889–902, 2004.

- [3810] I.F. Sevrioukova and J.A. Peterson. NADPH-P-450 reductase: Structural and functional comparisons of the eukaryotic and prokaryotic isoforms. *Biochimie*, 77:562–572, 1995.
- [3811] I.F. Sevrioukova and T.L. Poulos. Putidaredoxin reductase, a new function for an old protein. *J. Biol. Chem.*, 277:25831–25839, 2002.
- [3812] D.J. Seward, G. Cubberley, S. Kim, M. Schonewald, L. Zhang, B. Tripet, and D.L. Bentley. Demethylation of trimethylated histone H3 Lys⁴ *in vivo* by JARID1 JmjC proteins. *Nat. Struct. Mol. Biol.*, 14:240–242, 2007.
- [3813] T. Seyama, T. Kasama, T. Yamakawa, A. Kawaguchi, K. Saito, and S. Okuda. Origin of hydrogen atoms in the fatty acids synthesized with yeast fatty acid synthetase. *J. Biochem. (Tokyo)*, 82:1325–1329, 1977.
- [3814] A. Shafiee and C.R. Hutchinson. Macrolide antibiotic biosynthesis: isolation and properties of two forms of 6-deoxyerythronolide B hydroxylase from *Saccharopolyspora erythraea* (*Streptomyces erythreus*). *Biochemistry*, 26:6204–6210, 1987.
- [3815] S. Shak and I.M. Goldstein. Leukotriene B₄ ω-hydroxylase in human polymorphonuclear leukocytes. Partial purification and identification as a cytochrome P-450. *J. Clin. Invest.*, 76:1218–1228, 1985.
- [3816] S.L. Shames, A.H. Fairlamb, A. Cerami, and C.T. Walsh. Purification and characterization of trypanothione reductase from *Crithidia fasciculata*, a newly discovered member of the family of disulfide-containing flavoprotein reductases. *Biochemistry*, 25:3519–3526, 1986.
- [3817] T. Shanati, , and M. Enzymes and methods for the stereoselective reduction of carbonyl compounds, oxidation and stereoselective reductive amination - for the enantioselective preparation of alcohol amine compounds, 2019.
- [3818] T. Shanati and M.B. Ansorge-Schumacher. Biodegradation of ephedrine isomers by *Arthrobacter* sp. strain TS-15: discovery of novel ephedrine and pseudoephedrine dehydrogenases. *Appl. Environ. Microbiol.*, 86(6):e02487–19–, 2020.
- [3819] T. Shanati, C. Lockie, L. Beloti, G. Grogan, and M.B. Ansorge-Schumacher. Two enantiocomplementary ephedrine dehydrogenases from *Arthrobacter* sp. TS-15 with broad substrate specificity. *ACS Catal.*, 9:6202–6211, 2019.
- [3820] J. Shanklin and C. Somerville. Stearoyl-acyl-carrier-protein desaturase from higher plants is structurally unrelated to the animal and fungal homologs. *Proc. Natl. Acad. Sci. USA*, 88:2510–2514, 1991.
- [3821] H. Shao, R.A. Dixon, and X. Wang. Crystal structure of vestitone reductase from alfalfa (*Medicago sativa* L.). *J. Mol. Biol.*, 369:265–276, 2007.
- [3822] X. Shao, H.Y. Cao, F. Zhao, M. Peng, P. Wang, C.Y. Li, W.L. Shi, T.D. Wei, Z. Yuan, X.H. Zhang, X.L. Chen, J.D. Todd, and Y.Z. Zhang. Mechanistic insight into 3-methylmercaptopropionate metabolism and kinetical regulation of demethylation pathway in marine dimethylsulfoniopropionate-catabolizing bacteria. *Mol. Microbiol.*, 111:1057–1073, 2019.
- [3823] Y.H. Shao, L.Z. Guo, Y.Q. Zhang, H. Yu, B.S. Zhao, H.Q. Pang, and W.D. Lu. Glycine betaine monooxygenase, an unusual Rieske-type oxygenase system, catalyzes the oxidative N-demethylation of glycine betaine in *Chromohalobacter salexigens* DSM 3043. *Appl. Environ. Microbiol.*, 84, 2018.
- [3824] M.A. Sharaf and F. Sweet. Dual activity at an enzyme active site: 3β,20α-hydroxysteroid oxidoreductase from fetal blood. *Biochemistry*, 21:4615–4620, 1982.
- [3825] H.K. Sharma and C.S. Vaidyanathan. A new mode of ring cleavage of 2,3-dihydroxybenzoic acid in *Tecoma stans* (L.). Partial purification and properties of 2,3-dihydroxybenzoate 2,3-oxygenase. *Eur. J. Biochem.*, 56:163–171, 1975.
- [3826] M. Sharma, P. Abayakoon, J.P. Lingford, R. Epa, A. John, Y. Jin, E.D. Goddard-Borger, G.J. Davies, and S.J. Williams. Dynamic structural changes accompany the production of dihydroxypropanesulfonate by sulfolactaldehyde reductase. *ACS Catalysis*, 10:2826–2836, 2020.
- [3827] M. Sharma, J.P. Lingford, M. Petricevic, A.J.D. Snow, Y. Zhang, M.A. Jarva, J.W. Mui, N.E. Scott, E.C. Saunders, R. Mao, R. Epa, B.M. da Silva, D.E.V. Pires, D.B. Ascher, M.J. McConville, G.J. Davies, S.J. Williams, and E.D. Goddard-Borger. Oxidative desulfurization pathway for complete catabolism of sulfoquinovose by bacteria. *Proc. Natl. Acad. Sci. USA*, 119:e2116022119–, 2022.

- [3828] M.L. Sharma, S.M. Kaul, and O.P. Shukla. Metabolism of 2-hydroxypyridine by *Bacillus brevis* (INA). *Biol. Membr.*, 9:43–52, 1984.
- [3829] J.O. Sharp, C.M. Sales, J.C. LeBlanc, J. Liu, T.K. Wood, L.D. Eltis, W.W. Mohn, and L. Alvarez-Cohen. An inducible propane monooxygenase is responsible for *N*-nitrosodimethylamine degradation by *Rhodococcus* sp. strain RHA1. *Appl. Environ. Microbiol.*, 73:6930–6938, 2007.
- [3830] K.H. Sharp, P.C. Moody, K.A. Brown, and E.L. Raven. Crystal structure of the ascorbate peroxidase-salicylhydroxamic acid complex. *Biochemistry*, 43:8644–8651, 2004.
- [3831] K.B. Sharpless, T.E. Snyder, T.A. Spencer, K.K. Maheshwari, and J.A. Nelson. Biological demethylation of 4,4-dimethyl sterols, Evidence for enzymic epimerization of the 4 β -methyl group prior to its oxidative removal. *J. Am. Chem. Soc.*, 91:3394–3396, 1969.
- [3832] D.R.D. Shaw. Polyol dehydrogenases. 3. Galactitol dehydrogenase and D-iditol dehydrogenase. *Biochem. J.*, 64:394–405, 1956.
- [3833] J.P. Shaw and S. Harayama. Purification and characterisation of the NADH:acceptor reductase component of xylene monooxygenase encoded by the TOL plasmid pWW0 of *Pseudomonas putida* mt-2. *Eur. J. Biochem.*, 209:51–61, 1992.
- [3834] L. Shaw and P.C. Engel. The purification and properties of ox liver short-chain acyl-CoA dehydrogenase. *Biochem. J.*, 218:511–520, 1984.
- [3835] L. Shaw and R. Schauer. The biosynthesis of *N*-glycoloylneuraminic acid occurs by hydroxylation of the CMP-glycoside of *N*-acetylneuraminic acid. *Biol. Chem. Hoppe-Seyler*, 369:477–486, 1988.
- [3836] P.D. Shaw. Biosynthesis of nitro compounds. III. The enzymatic reduction of β -nitroacrylic acid to β -nitropropionic acid. *Biochemistry*, 6:2253–2260, 1967.
- [3837] S. Shefer, S. Hauser, and E.H. Mosbach. Studies on the biosynthesis of 5 α -cholestan-3 β -ol. I. Cholestenone 5 α -reductase of rat liver. *J. Biol. Chem.*, 241:946–952, 1966.
- [3838] B. Shen and C.R. Hutchinson. Tetracenomycin F1 monooxygenase: oxidation of a naphthacenone to a naphthacenequinone in the biosynthesis of tetracenomycin C in *Streptomyces glaucescens*. *Biochemistry*, 32:6656–6663, 1993.
- [3839] B. Shen and C.R. Hutchinson. Triple hydroxylation of tetracenomycin A2 to tetracenomycin C in *Streptomyces glaucescens*. Overexpression of the *tcmG* gene in *Streptomyces lividans* and characterization of the tetracenomycin A2 oxygenase. *J. Biol. Chem.*, 269:30726–30733, 1994.
- [3840] G. Shen, Y. Pang, W. Wu, Z. Deng, L. Zhao, Y. Cao, X. Sun, and K. Tang. Cloning and characterization of a flavanone 3-hydroxylase gene from *Ginkgo biloba*. *Biosci Rep*, 26:19–29, 2006.
- [3841] W. Shen, W. Liu, J. Zhang, J. Tao, H. Deng, H. Cao, and Z. Cui. Cloning and characterization of a gene cluster involved in the catabolism of *p*-nitrophenol from *Pseudomonas putida* DLL-E4. *Bioresour. Technol.*, 101:7516–7522, 2010.
- [3842] W. Shen, Y. Wei, M. Dauk, Z. Zheng, and J. Zou. Identification of a mitochondrial glycerol-3-phosphate dehydrogenase from *Arabidopsis thaliana*: evidence for a mitochondrial glycerol-3-phosphate shuttle in plants. *FEBS Lett.*, 536:92–96, 2003.
- [3843] D. Sheng, D.P. Ballou, and V. Massey. Mechanistic studies of cyclohexanone monooxygenase: chemical properties of intermediates involved in catalysis. *Biochemistry*, 40:11156–11167, 2001.
- [3844] S.A. Shepherd, B.R. Menon, H. Fisk, A.W. Struck, C. Levy, D. Leys, and J. Micklefield. A structure-guided switch in the regioselectivity of a tryptophan halogenase. *ChemBioChem*, 17:821–824, 2016.
- [3845] C.A. Sheppard, E.E. Trimmer, and R.G. Matthews. Purification and properties of NADH-dependent 5,10-methylenetetrahydrofolate reductase (MetF) from *Escherichia coli*. *J. Bacteriol.*, 181:718–725, 1999.
- [3846] N.H. Sherden, B. Lichman, L. Caputi, D. Zhao, M.O. Kamileen, C.R. Buell, and S.E. O'Connor. Identification of iridoid synthases from *Nepeta* species: Iridoid cyclization does not determine nepetalactone stereochemistry. *Phytochemistry*, 145:48–56, 2018.

- [3847] D.H. Sherman, S. Li, L.V. Yermalitskaya, Y. Kim, J.A. Smith, M.R. Waterman, and L.M. Podust. The structural basis for substrate anchoring, active site selectivity, and product formation by P450 PikC from *Streptomyces venezuelae*. *J. Biol. Chem.*, 281:26289–26297, 2006.
- [3848] M. Shettigar, S. Balotra, A. Kasprzak, S.L. Pearce, M.J. Lacey, M.C. Taylor, J.W. Liu, D. Cahill, J.G. Oakeshott, and G. Pandey. Oxidative catabolism of (+)-pinoresinol is initiated by an unusual flavocytochrome encoded by translationally coupled genes within a cluster of (+)-pinoresinol-coinduced genes in *Pseudomonas* sp. strain SG-MS2. *Appl. Environ. Microbiol.*, 86:e00375–20–, 2020.
- [3849] J. Shi, V. Vlamis-Gardikas, F. Aslund, A. Holmgren, and B.P. Rosen. Reactivity of glutaredoxins 1, 2, and 3 from *Escherichia coli* shows that glutaredoxin 2 is the primary hydrogen donor to ArsC-catalyzed arsenate reduction. *J. Biol. Chem.*, 274:36039–36042, 1999.
- [3850] S. Shi and S. Ehrh. Dihydrolipoamide acyltransferase is critical for *Mycobacterium tuberculosis* pathogenesis. *Infect. Immun.*, 74:56–63, 2006.
- [3851] M. Shibahara, J.A. Moody, and L.L. Smith. Microbial hydroxylations. V. 11 α -Hydroxylation of progesterone by cell-free preparations of *Aspergillus ochraceus*. *Biochim. Biophys. Acta*, 202:172–179, 1970.
- [3852] T. Shibasaki, H. Mori, S. Chiba, and A. Ozaki. Microbial proline 4-hydroxylase screening and gene cloning. *Appl. Environ. Microbiol.*, 65:4028–4031, 1999.
- [3853] D. Shibata, J. Steczko, F.E. Dixon, M. Hermodson, R. Yasdanparast, and B. Axelrod. Primary structure of soybean lipoxygenase-1. *J. Biol. Chem.*, 262:10080–10085, 1987.
- [3854] M. Shibuya, M. Hoshino, Y. Katsube, H. Hayashi, T. Kushiro, and Y. Ebizuka. Identification of β -amyrin and sophoradiol 24-hydroxylase by expressed sequence tag mining and functional expression assay. *FEBS J.*, 273:948–959, 2006.
- [3855] A.K. Shiemke, S.A. Cook, T. Miley, and P. Singleton. Detergent solubilization of membrane-bound methane monooxygenase requires plastoquinol analogs as electron donors. *Arch. Biochem. Biophys.*, 321:421–428, 1995.
- [3856] S. Shigeoka, Y. Nakano, and S. Kitaoka. Metabolism of hydrogen peroxide in *Euglena gracilis* Z by L-ascorbic acid peroxidase. *Biochem. J.*, 186:377–380, 1980.
- [3857] S. Shigeoka, Y. Nakano, and S. Kitaoka. Purification and some properties of L-ascorbic-acid-specific peroxidase in *Euglena gracilis*. *Z. Arch. Biochem. Biophys.*, 201:121–127, 1980.
- [3858] J.C. Shih and K. Chen. Regulation of MAO-A and MAO-B gene expression. *Curr. Med. Chem.*, 11:1995–2005, 2004.
- [3859] I. Shiio and H. Ozaki. Regulation of nicotinamide adenine dinucleotide phosphate-specific glutamate dehydrogenase from *Brevibacterium flavum*, a glutamate-producing bacterium. *J. Biochem. (Tokyo)*, 68:633–647, 1970.
- [3860] M. Shikita, H. Inano, and B. Tamaoki. Further studies on 20 α -hydroxysteroid dehydrogenase of rat testes. *Biochemistry*, 6:1760–1764, 1967.
- [3861] N. Shikura, J. Yamamura, and T. Nihira. *barS1*, a gene for biosynthesis of a γ -butyrolactone autoregulator, a microbial signaling molecule eliciting antibiotic production in *Streptomyces* species. *J. Bacteriol.*, 184:5151–5157, 2002.
- [3862] Y. Shima, M. Shiina, T. Shinozawa, Y. Ito, H. Nakajima, Y. Adachi, and K. Yabe. Participation in aflatoxin biosynthesis by a reductase enzyme encoded by *vrda* gene outside the aflatoxin gene cluster. *Fungal Genet. Biol.*, 46:221–231, 2009.
- [3863] M. Shimada and E.E. Conn. The enzymatic conversion of *p*-hydroxyphenylacetaldoxime to *p*-hydroxymandelonitrile. *Arch. Biochem. Biophys.*, 180:199–207, 1977.
- [3864] Y. Shimada, S. Fujioka, N. Miyauchi, M. Kushiro, S. Takatsuto, T. Nomura, T. Yokota, Y. Kamiya, G.J. Bishop, and S. Yoshida. Brassinosteroid-6-oxidases from *Arabidopsis* and tomato catalyze multiple C-6 oxidations in brassinosteroid biosynthesis. *Plant Physiol.*, 126:770–779, 2001.
- [3865] Y. Shimada, R. Nakano-Shimada, M. Ohbayashi, Y. Okinaka, S. Kiyokawa, and Y. Kikuchi. Expression of chimeric P450 genes encoding flavonoid-3', 5'-hydroxylase in transgenic tobacco and petunia plants¹. *FEBS Lett.*, 461:241–245, 1999.

- [3866] T. Shimakata and P.K. Stumpf. Purification and characterizations of β -ketoacyl-[acyl-carrier-protein] reductase, β -hydroxyacyl-[acyl-carrier-protein] dehydrase, and enoyl-[acyl-carrier-protein] reductase from *Spinacia oleracea* leaves. *Arch. Biochem. Biophys.*, 218:77–91, 1982.
- [3867] M. Shima, K. Ninomiya, O. Kuno, N. Kato, and C. Sakazawa. Existence of a novel enzyme, pyrroloquinoline quinone-dependent polyvinyl alcohol dehydrogenase, in a bacterial symbiont, *Pseudomonas* sp. strain VM15C. *Appl. Environ. Microbiol.*, 51:268–268, 1986.
- [3868] M. Shima, Y. Nishimura, N. Kato, and C. Sakazawa. Localization of polyvinyl alcohol oxidase produced by a bacterial symbiont *Pseudomonas* sp strain VM 15C. *Appl. Environ. Microbiol.*, 49:8–10, 1985.
- [3869] M. Shima, S. Onishi, N. Kato, and C. Sakazawa. Pyrroloquinoline quinone-dependent cytochrome reduction in polyvinyl alcohol-degrading *Pseudomonas* sp strain VM15C. *Appl. Environ. Microbiol.*, 55:275–278, 1989.
- [3870] M. Shimizu, T. Yamamoto, N. Okabe, K. Sakai, E. Koide, Y. Miyachi, M. Kurimoto, M. Mochizuki, S. Yoshino-Yasuda, S. Mitsui, A. Ito, H. Murano, N. Takaya, and M. Kato. Novel 4-methyl-2-oxopentanoate reductase involved in synthesis of the Japanese sake flavor, ethyl leucate. *Appl. Microbiol. Biotechnol.*, 2015.
- [3871] T. Shimizu, T. Izumi, Y. Seyama, K. Tadokoro, O. Rådmark, and B. Samuelsson. Characterization of leukotriene A₄ synthase from murine mast cells: evidence for its identity to arachidonate 5-lipoxygenase. *Proc. Natl. Acad. Sci. USA*, 83:4175–4179, 1986.
- [3872] T. Shimizu, O. Rådmark, and B. Samuelsson. Enzyme with dual lipoxygenase activities catalyzes leukotriene A₄ synthesis from arachidonic acid. *Proc. Natl. Acad. Sci. USA*, 81:689–693, 1984.
- [3873] T. Shimizu, T. Tomita, T. Kuzuyama, and M. Nishiyama. Crystal Structure of the LysY.LysW Complex from *Thermus thermophilus*. *J. Biol. Chem.*, 291:9948–9959, 2016.
- [3874] O. Shimomura, F. H. Johnson, , and Y. Purification and properties of aequorin, a bio-(chemi-) luminescent protein from the jellyfish, *Aequorea aequorea*. *Fed. Proc.*, 21:401–, 1962.
- [3875] O. Shimomura and F.H. Johnson. The structure of *Latia* luciferin. *Biochemistry*, 7:1734–1738, 1968.
- [3876] O. Shimomura and F.H. Johnson. Chemical nature of bioluminescence systems in coelenterates. *Proc. Natl. Acad. Sci. USA*, 72:1546–1549, 1975.
- [3877] O. Shimomura, F.H. Johnson, and Y. Kohama. Reactions involved in bioluminescence systems of limpet (*Latia neritoides*) and luminous bacteria. *Proc. Natl. Acad. Sci. USA*, 69:2086–2089, 1972.
- [3878] O. Shimomura, T. Masugi, F.H. Johnson, and Y. Haneda. Properties and reaction mechanism of the bioluminescence system of the deep-sea shrimp *Oplophorus gracilorostri*. *Biochemistry*, 17:994–998, 1978.
- [3879] E. Shimoni, U. Ravid, and Y. Shoham. Isolation of a *Bacillus* sp. capable of transforming isoeugenol to vanillin. *J. Biotechnol.*, 78:1–9, 2000.
- [3880] K. Shimura, A. Okada, K. Okada, Y. Jikumaru, K.W. Ko, T. Toyomasu, T. Sassa, M. Hasegawa, O. Kodama, N. Shibuya, J. Koga, H. Nojiri, and H. Yamane. Identification of a biosynthetic gene cluster in rice for momilactones. *J. Biol. Chem.*, 282:34013–34018, 2007.
- [3881] M. Shin, K. Tagawa, and D.I. Arnon. Crystallization of ferredoxin-TPN reductase and its role in the photosynthetic apparatus of chloroplasts. *Biochem. Z.*, 338:84–96, 1963.
- [3882] E. Shinagawa and M. Ameyama. 2-Keto-D-gluconate dehydrogenase from *Gluconobacter melanogenus*, membrane-bound. *Methods Enzymol.*, 89:194–198, 1982.
- [3883] E. Shinagawa and M. Ameyama. Purification and characterization of D-sorbitol dehydrogenase from membrane of *Gluconobacter suboxydans* var-alpha. *Agric. Biol. Chem.*, 46:135–141, 1982.
- [3884] E. Shinagawa, H. Toyama, K. Matsushita, P. Tuitemwong, G. Theeragool, and O. Adachi. A novel type of formaldehyde-oxidizing enzyme from the membrane of *Acetobacter* sp. SKU 14. *Biosci. Biotechnol. Biochem.*, 70:850–857, 2006.
- [3885] M. Shinjoh, M. Tazoe, and T. Hoshino. NADPH-dependent L-sorbose reductase is responsible for L-sorbose assimilation in *Gluconobacter suboxydans* IFO 3291. *J. Bacteriol.*, 184:861–863, 2002.

- [3886] R. Shinkyo, T. Sakaki, M. Kamakura, M. Ohta, and K. Inouye. Metabolism of vitamin D by human microsomal CYP2R1. *Biochem. Biophys. Res. Commun.*, 324:451–457, 2004.
- [3887] K. Shinoda, T. Hasegawa, H. Sato, M. Shinozaki, H. Kuramoto, Y. Takamiya, T. Sato, N. Nikaidou, T. Watanabe, and T. Hoshino. Biosynthesis of violacein: a genuine intermediate, protoviolaceinic acid, produced by VioABDE, and insight into VioC function. *Chem. Commun. (Camb.)*, pages 4140–4142, 2007.
- [3888] T. Shiotani and G. Weber. Purification and properties of dihydrothymine dehydrogenase from rat liver. *J. Biol. Chem.*, 256:219–224, 1981.
- [3889] Y. Shiro, M. Fujii, T. Iizuka, S. Adachi, K. Tsukamoto, K. Nakahara, and H. Shoun. Spectroscopic and kinetic studies on reaction of cytochrome P450_{nor} with nitric oxide. Implication for its nitric oxide reduction mechanism. *J. Biol. Chem.*, 270:1617–1623, 1995.
- [3890] O. Shoji, C. Wiese, T. Fujishiro, C. Shirataki, B. Wunsch, and Y. Watanabe. Aromatic C-H bond hydroxylation by P450 peroxxygenases: a facile colorimetric assay for monooxygenation activities of enzymes based on Russig's blue formation. *J. Biol. Inorg. Chem.*, 15:1109–1115, 2010.
- [3891] S. Shojima, N.-K. Nishizawa, S. Fushiya, S. Nozoe, T. Irifune, and S. Mori. In vitro biosynthesis of 2'-deoxymugineic acid from L-methionine and nicotianamine. *Plant Physiol.*, 93:1497–1503, 1990.
- [3892] H. Shoun and T. Tanimoto. Denitrification by the fungus *Fusarium oxysporum* and involvement of cytochrome P-450 in the respiratory nitrite reduction. *J. Biol. Chem.*, 266:11078–11082, 1991.
- [3893] Y. Shoyama, A. Takeuchi, F. Taura, T. Tamada, M. Adachi, R. Kuroki, Y. Shoyama, and S. Morimoto. Crystallization of Δ^1 -tetrahydrocannabinolic acid (THCA) synthase from *Cannabis sativa*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:799–801, 2005.
- [3894] Y. Shoyama, T. Tamada, K. Kurihara, A. Takeuchi, F. Taura, S. Arai, M. Blaber, Y. Shoyama, S. Morimoto, and R. Kuroki. Structure and function of 1-tetrahydrocannabinolic acid (THCA) synthase, the enzyme controlling the psychoactivity of *Cannabis sativa*. *J. Mol. Biol.*, 423:96–105, 2012.
- [3895] A.L. Shug, P.W. Wilson, D.E. Green, and H.R. Mahler. The role of molybdenum and flavin in hydrogenase. *J. Am. Chem. Soc.*, 76:3355–3356, 1954.
- [3896] O.P. Shukla, S. Muller, and R.D. Walter. Polyamine oxidase from *Acanthamoeba culbertsoni* specific for N⁸-acetylpermidine. *Mol. Biochem. Parasitol.*, 51:91–98, 1992.
- [3897] O. Sibbesen, B. Koch, B.A. Halkier, and B.L. Møller. Cytochrome P-450TYR is a multifunctional heme-thiolate enzyme catalyzing the conversion of L-tyrosine to *p*-hydroxyphenylacetaldehyde oxime in the biosynthesis of the cyanogenic glucoside dhurrin in *Sorghum bicolor* (L.) Moench. *J. Biol. Chem.*, 270:3506–3511, 1995.
- [3898] M.A. Siddiqui, S. Fujiwara, and T. Imanaka. Indolepyruvate ferredoxin oxidoreductase from *Pyrococcus* sp. K0d1 possesses a mosaic: Structure showing features of various oxidoreductases. *Mol. Gen. Genet.*, 254:433–439, 1997.
- [3899] G. Siebert, J. Dubuc, R.C. Warner, and G.W.E. Plaut. The preparation of isocitrate dehydrogenase from mammalian heart. *J. Biol. Chem.*, 226:965–975, 1957.
- [3900] J.N. Siedow, A.L. Umbach, and A.L. Moore. The active site of the cyanide-resistant oxidase from plant mitochondria contains a binuclear iron center. *FEBS Lett.*, 362:10–14, 1995.
- [3901] L.M. Siegel, M.J. Murphy, and H. Kamin. Reduced nicotinamide adenine dinucleotide phosphate-sulfite reductase of enterobacteria. I. The *Escherichia coli* hemoflavoprotein: molecular parameters and prosthetic groups. *J. Biol. Chem.*, 248:251–264, 1973.
- [3902] L.M. Siegel, D.C. Rueger, M.J. Barber, R.J. Krueger, N.R. Orme-Johnson, and W.H. Orme-Johnson. *Escherichia coli* sulfite reductase hemoprotein subunit. Prosthetic groups, catalytic parameters, and ligand complexes. *J. Biol. Chem.*, 257:6343–6350, 1982.
- [3903] J.L. Simala-Grant and J.H. Weiner. Kinetic analysis and substrate specificity of *Escherichia coli* dimethyl sulfoxide reductase. *Microbiology*, 142:3231–3239, 1996.

- [3904] M. Simianu, E. Murakami, J.M. Brewer, and S.W. Ragsdale. Purification and properties of the heme- and iron-sulfur-containing heterodisulfide reductase from *Methanosarcina thermophila*. *Biochemistry*, 37:10027–10039, 1998.
- [3905] A. Simon, U. Hellman, C. Wernstedt, and U. Eriksson. The retinal pigment epithelial-specific 11-*cis* retinol dehydrogenase belongs to the family of short chain alcohol dehydrogenases. *J. Biol. Chem.*, 270:1107–1112, 1995.
- [3906] A.N. Simonov, J.K. Holien, J.C. Yeung, A.D. Nguyen, C.J. Corbin, J. Zheng, V.L. Kuznetsov, R.J. Auchus, A.J. Conley, A.M. Bond, M.W. Parker, R.J. Rodgers, and L.L. Martin. Mechanistic scrutiny identifies a kinetic role for cytochrome *b*₅ regulation of human cytochrome P450c17 (CYP17A1, P450 17A1). *PLoS One*, 10:e0141252–e0141252, 2015.
- [3907] R.K. Sindhu and D.C. Walton. Xanthoxin metabolism in cell-free preparations from wild type and wilty mutants of tomato. *Plant Physiol.*, 88:178–182, 1988.
- [3908] D. Singh, A. Kumari, S. Ramaswamy, and G. Ramanathan. Expression, purification and substrate specificities of 3-nitrotoluene dioxygenase from *Diaphorobacter* sp. strain DS2. *Biochem. Biophys. Res. Commun.*, 445:36–42, 2014.
- [3909] J. Singh. Cytochrome oxidase from *Pseudomonas aeruginosa*. III. Reduction of hydroxylamine. *Biochim. Biophys. Acta*, 333:28–36, 1974.
- [3910] S. Singh, J. Stavrinos, D. Christendat, and D.S. Guttman. A phylogenomic analysis of the shikimate dehydrogenases reveals broadscale functional diversification and identifies one functionally distinct subclass. *Mol. Biol. Evol.*, 25:2221–2232, 2008.
- [3911] S.K. Singh, G. Grass, C. Rensing, and W.R. Montfort. Cuprous oxidase activity of CueO from *Escherichia coli*. *J. Bacteriol.*, 186:7815–7817, 2004.
- [3912] S.K. Singh, S.A. Roberts, S.F. McDevitt, A. Weichsel, G.F. Wildner, G.B. Grass, C. Rensing, and W.R. Montfort. Crystal structures of multicopper oxidase CueO bound to copper(I) and silver(I): functional role of a methionine-rich sequence. *J. Biol. Chem.*, 286:37849–37857, 2011.
- [3913] V.K. Singh and J. Moskovitz. Multiple methionine sulfoxide reductase genes in *Staphylococcus aureus*: expression of activity and roles in tolerance of oxidative stress. *Microbiology*, 149:2739–2747, 2003.
- [3914] J.W. Singleton and L. Laster. Biliverdin reductase of guinea pig liver. *J. Biol. Chem.*, 240:4780–4789, 1965.
- [3915] D. Sippel and O. Einsle. The structure of vanadium nitrogenase reveals an unusual bridging ligand. *Nat. Chem. Biol.*, 13:956–960, 2017.
- [3916] D. Sircar and A. Mitra. Evidence for *p*-hydroxybenzoate formation involving enzymatic phenylpropanoid side-chain cleavage in hairy roots of *Daucus carota*. *J. Plant Physiol.*, 165:407–414, 2008.
- [3917] S. Sirikantaramas, S. Morimoto, Y. Shoyama, Y. Ishikawa, Y. Wada, Y. Shoyama, and F. Taura. The gene controlling marijuana psychoactivity: molecular cloning and heterologous expression of Δ^1 -tetrahydrocannabinolic acid synthase from *Cannabis sativa* L. *J. Biol. Chem.*, 279:39767–39774, 2004.
- [3918] W.R. Sistrom and R.Y. Stanier. The mechanism of formation of β -keto adipic acid by bacteria. *J. Biol. Chem.*, 210:821–836, 1954.
- [3919] A. Sivak and O. Hoffmann-Ostenhof. Enzymes of *meso*-inositol catabolism in the yeast *Schwanniomyces occidentalis*. *Biochim. Biophys. Acta*, 53:426–428, 1961.
- [3920] J. Sivaraman, Y. Li, J. Banks, D.E. Cane, A. Matte, and M. Cygler. Crystal structure of *Escherichia coli* PdxA, an enzyme involved in the pyridoxal phosphate biosynthesis pathway. *J. Biol. Chem.*, 278:43682–43690, 2003.
- [3921] J. Siwinska, K. Siatkowska, A. Olry, J. Grosjean, A. Hehn, F. Bourgaud, A.A. Meharg, M. Carey, E. Lojkowska, and A. Ihnatowicz. Scopoletin 8-hydroxylase: a novel enzyme involved in coumarin biosynthesis and iron-deficiency responses in *Arabidopsis*. *J. Exp. Bot.*, 69:1735–1748, 2018.
- [3922] B.A. Skaggs, J.F. Alexander, C.A. Pierson, K.S. Schweitzer, K.T. Chun, C. Koegel, R. Barbuch, and M. Bard. Cloning and characterization of the *Saccharomyces cerevisiae* C-22 sterol desaturase gene, encoding a second cytochrome *P*-450 involved in ergosterol biosynthesis. *Gene*, 169:105–109, 1996.

- [3923] T. Skotland and T. Ljones. Direct spectrophotometric detection of ascorbate free radical formed by dopamine β -monooxygenase and by ascorbate oxidase. *Biochim. Biophys. Acta*, 630:30–35, 1980.
- [3924] S. Skramo, H.P. Hersleth, M. Hammerstad, K.K. Andersson, and A.K. Rohr. Cloning, expression, purification, crystallization and preliminary X-ray diffraction analysis of a ferredoxin/ flavodoxin-NADP(H) oxidoreductase (Bc0385) from *Bacillus cereus*. *Acta Crystallogr. F Struct. Biol. Commun.*, 70:777–780, 2014.
- [3925] C.R. Slack, P.G. Roughan, and J. Browse. Evidence for an oleoyl phosphatidylcholine desaturase in microsomal preparations from cotyledons of safflower (*Carthamus tinctorius*) seed. *Biochem. J.*, 179:649–656, 1979.
- [3926] J.C. Slaughter and D.D. Davies. The isolation and characterization of 3-phosphoglycerate dehydrogenase from peas. *Biochem. J.*, 109:743–748, 1968.
- [3927] M.C. Sleeman, P. Smith, B. Kellam, S.R. Chhabra, B.W. Bycroft, and C.J. Schofield. Biosynthesis of carbapenem antibiotics: new carbapenam substrates for carbapenam synthase (CarC). *ChemBioChem*, 5:879–882, 2004.
- [3928] M.K. Sluis, L.A. Sayavedra-Soto, and D.J. Arp. Molecular analysis of the soluble butane monooxygenase from ‘*Pseudomonas butanovora*’. *Microbiology*, 148:3617–3629, 2002.
- [3929] F.J. Small and S.A. Ensign. Alkene monooxygenase from *Xanthobacter* strain Py2: purification and characterization of a four-component system central to the bacterial metabolism of aliphatic alkenes. *J. Biol. Chem.*, 272:24913–24920, 1997.
- [3930] R.C. Smallridge, K.D. Burman, K.E. Ward, L. Wartofsky, R.C. Dimond, F.D. Wright, and K.R. Lathan. 3',5'-Diodothyronine to 3'-moniodothyronine conversion in the fed and fasted rat: enzyme characteristics and evidence for two distinct 5'-deiodinases. *Endocrinology*, 108:2336–2345, 1981.
- [3931] M.J. Smanski, Z. Yu, J. Casper, S. Lin, R.M. Peterson, Y. Chen, E. Wendt-Pienkowski, S.R. Rajski, and B. Shen. Dedicated *ent*-kaurene and *ent*-atiserene synthases for platensimycin and platencin biosynthesis. *Proc. Natl. Acad. Sci. USA*, 108:13498–13503, 2011.
- [3932] J.D. Smiley and G. Ashwell. Purification and properties of β -L-hydroxy acid dehydrogenase. II. Isolation of β -keto-L-gluconic acid, an intermediate in L-xylulose biosynthesis. *J. Biol. Chem.*, 236:357–364, 1961.
- [3933] S.V. Smirnov, P.M. Sokolov, V.A. Kotlyarova, N.N. Samsonova, T. Kodera, M. Sugiyama, T. Torii, M. Hibi, S. Shimizu, K. Yokozeki, and J. Ogawa. A novel L-isoleucine-4'-dioxygenase and L-isoleucine dihydroxylation cascade in *Pantoea ananatis*. *MicrobiologyOpen*, 2:471–481, 2013.
- [3934] E.L. Smith, B.M. Austen, K.M. Blumenthal, and J.F. Nyc. Glutamate dehydrogenases. In P.D. Boyer, editor, *The Enzymes*, volume 11, pages 293–367. Academic Press, New York, 3rd edition, 1975.
- [3935] L.D. Smith, N. Budgen, S.J. Bungard, M.J. Danson, and D.W. Hough. Purification and characterization of glucose dehydrogenase from the thermoacidophilic archaeobacterium *Thermoplasma acidophilum*. *Biochem. J.*, 261:973–977, 1989.
- [3936] L.T. Smith and N.O. Kaplan. Purification, properties, and kinetic mechanism of coenzyme A-linked aldehyde dehydrogenase from *Clostridium kluyveri*. *Arch. Biochem. Biophys.*, 203:663–675, 1980.
- [3937] M.A. Smith, A.R. Cross, O.T. Jones, W.T. Griffiths, S. Stymne, and K. Stobart. Electron-transport components of the 1-acyl-2-oleoyl-*sn*-glycero-3-phosphocholine Δ^{12} -desaturase (Δ^{12} -desaturase) in microsomal preparations from developing safflower (*Carthamus tinctorius* L.) cotyledons. *Biochem. J.*, 272:23–29, 1990.
- [3938] M.A. Smith, M. Dauk, H. Ramadan, H. Yang, L.E. Seamons, R.P. Haslam, F. Beaudoin, I. Ramirez-Erosa, and L. Forseille. Involvement of *Arabidopsis* acyl-coenzyme A desaturase-like2 (At2g31360) in the biosynthesis of the very-long-chain monounsaturated fatty acid components of membrane lipids. *Plant Physiol.*, 161:81–96, 2013.
- [3939] M.A. Smith, L. Jonsson, S. Stymne, and K. Stobart. Evidence for cytochrome *b*₅ as an electron donor in ricinoleic acid biosynthesis in microsomal preparations from developing castor bean (*Ricinus communis* L.). *Biochem. J.*, 287:141–144, 1992.
- [3940] M.E. Smith and D.M. Greenberg. Characterization of an enzyme reducing pyrroline-5-carboxylate to proline. *Nature (Lond.)*, 177:1130–1130, 1956.

- [3941] N. Smith, M. Mayhew, M.J. Holden, H. Kelly, H. Robinson, A. Heroux, V.L. Vilker, and D.T. Gallagher. Structure of C73G putidaredoxin from *Pseudomonas putida*. *Acta Crystallogr. D Biol. Crystallogr.*, 60:816–822, 2004.
- [3942] N. A. Smith and D. P. Kelly. Isolation and physiological characterization of autotrophic sulphur bacteria oxidizing dimethyldisulphide as sole source of energy. *J. Gen. Microbiol.*, 134:1407–1417, 1988.
- [3943] N. A. Smith and D. P. Kelly. Mechanism of oxidation of dimethyl disulphide by *Thiobacillus thioparus* E6. *J. Gen. Microbiol.*, 134:3031–3039, 1988.
- [3944] S.L. Smith, W.E. Bollenbacher, D.Y. Cooper, H. Schleyer, J.J. Wielgus, and L.I. Gilbert. Ecdysone 20-monooxygenase: characterization of an insect cytochrome *p*-450 dependent steroid hydroxylase. *Mol. Cell. Endocrinol.*, 15:111–133, 1979.
- [3945] S.T. Smith, K.V. Rajagopalan, and P. Handler. Purification and properties of xanthine dehydrogenase from *Micrococcus lactilyticus*. *J. Biol. Chem.*, 242:4108–4117, 1967.
- [3946] T.E. Smith and C. Mitoma. Partial purification and some properties of 4-ketoproline reductase. *J. Biol. Chem.*, 237:1177–1180, 1962.
- [3947] V.A. Smith and J. MacMillan. The partial-purification and characterization of gibberellin 2 β -hydroxylases from seeds of *Pisum sativum*. *Planta*, 167:9–18, 1986.
- [3948] F. Snyder, B. Malone, and C. Piantadosi. Tetrahydropteridine-dependent cleavage enzyme for *O*-alkyl lipids: substrate specificity. *Biochim. Biophys. Acta*, 316:259–265, 1973.
- [3949] R.J. Soberman, T.W. Harper, R.C. Murphy, and K.F. Austen. Identification and functional characterization of leukotriene B₄ 20-hydroxylase of human polymorphonuclear leukocytes. *Proc. Natl. Acad. Sci. USA*, 82:2292–2295, 1985.
- [3950] K.M. Sogi, Z.J. Gartner, M.A. Breidenbach, M.J. Appel, M.W. Schelle, and C.R. Bertozzi. *Mycobacterium tuberculosis* Rv3406 is a type II alkyl sulfatase capable of sulfate scavenging. *PLoS One*, 8:e65080–e65080, 2013.
- [3951] K.M. Sogi, C.M. Holsclaw, G.K. Fragiadakis, D.K. Nomura, J.A. Leary, and C.R. Bertozzi. Biosynthesis and regulation of sulfomenaquinone, a metabolite associated with virulence in *Mycobacterium tuberculosis*. *ACS Infect Dis*, 2:800–806, 2016.
- [3952] B. Sohling and G. Gottschalk. Purification and characterization of a coenzyme-A-dependent succinate-semialdehyde dehydrogenase from *Clostridium kluyveri*. *Eur. J. Biochem.*, 212:121–127, 1993.
- [3953] D.-E. Sok, J.B. Kang, and H.D. Shin. 15-Hydroxyeicosatetraenoic acid dehydrogenase activity in microsomal fraction of mouse liver homogenate. *Biochem. Biophys. Res. Commun.*, 156:524–529, 1988.
- [3954] J.R. Sokatch, L.E. Sanders, and V.P. Marshall. Oxidation of methylmalonate semialdehyde to propionyl coenzyme A in *Pseudomonas aeruginosa* grown on valine. *J. Biol. Chem.*, 243:2500–2506, 1968.
- [3955] J. Soll, G. Schultz, W. Rudiger, and J. Benz. Hydrogenation of geranylgeraniol : two pathways exist in spinach chloroplasts. *Plant Physiol.*, 71:849–854, 1983.
- [3956] R. Somack and R.N. Costilow. 2,4-Diaminopentanoic acid C4 dehydrogenase. Purification and properties of the protein. *J. Biol. Chem.*, 248:385–388, 1973.
- [3957] W.S. Somers, M.L. Stahl, and F.X. Sullivan. GDP-fucose synthetase from *Escherichia coli*: Structure of a unique member of the short-chain dehydrogenase/reductase family that catalyzes two distinct reactions at the same active site. *Structure*, 6:1601–1612, 1998.
- [3958] C.C. Somerville, S.F. Nishino, and J.C. Spain. Purification and characterization of nitrobenzene nitroreductase from *Pseudomonas pseudoalcaligenes* JS45. *J. Bacteriol.*, 177:3837–3842, 1995.
- [3959] C. Sommer and H. Gorisch. Enzymology of the degradation of (di)chlorobenzenes by *Xanthobacter flavus* 14p1. *Arch. Microbiol.*, 167:384–391, 1997.
- [3960] H. Song, A.S. Her, F. Raso, Z. Zhen, Y. Huo, and P. Liu. Cysteine oxidation reactions catalyzed by a mononuclear non-heme iron enzyme (OvoA) in ovothiol biosynthesis. *Org. Lett.*, 16:2122–2125, 2014.

- [3961] H. Song, M. Leninger, N. Lee, and P. Liu. Regioselectivity of the oxidative C-S bond formation in ergothioneine and ovolthiol biosyntheses. *Org. Lett.*, 15:4854–4857, 2013.
- [3962] M. Song, A.C. Kim, A.J. Gorzalski, M. MacLean, S. Young, M.D. Ginzel, G.J. Blomquist, and C. Tittiger. Functional characterization of myrcene hydroxylases from two geographically distinct *Ips pini* populations. *Insect Biochem. Mol. Biol.*, 43:336–343, 2013.
- [3963] X. Song, J. Lu, and W. Lai. Mechanistic insights into dioxygen activation, oxygen atom exchange and substrate epoxidation by AsqJ dioxygenase from quantum mechanical/molecular mechanical calculations. *Phys Chem Chem Phys*, 19:20188–20197, 2017.
- [3964] M. Sono. Spectroscopic and equilibrium studies of ligand and organic substrate binding to indolamine 2,3-dioxygenase. *Biochemistry*, 29:1451–1460, 1990.
- [3965] T. Sonoyama, B. Kageyama, S. Yagi, and K. Mitsushima. Biochemical aspects of 2-keto-L-gulonate accumulation from 2,5-diketo-D-gluconate by *Corynebacterium* sp. and its mutants. *Agric. Biol. Chem.*, 51:3039–3047, 1987.
- [3966] T. Sonoyama and K. Kobayashi. Purification and properties of two 2,5-diketo-D-gluconate reductases from a mutant strain derived from *Corynebacterium* sp. *J Ferment Technol.*, 65:311–317, 1987.
- [3967] J.F. Soodsma, C. Piantadosi, and F. Snyder. Partial characterization of the alkylglycerol cleavage enzyme system of rat liver. *J. Biol. Chem.*, 247:3923–3929, 1972.
- [3968] C.L. Soong, J. Ogawa, and S. Shimizu. Novel amidohydrolytic reactions in oxidative pyrimidine metabolism: analysis of the barbiturase reaction and discovery of a novel enzyme, ureidomalonase. *Biochem. Biophys. Res. Commun.*, 286:222–226, 2001.
- [3969] K. Sorefan, J. Booker, K. Haurogne, M. Goussot, K. Bainbridge, E. Foo, S. Chatfield, S. Ward, C. Beveridge, C. Rameau, and O. Leyser. MAX4 and RMS1 are orthologous dioxygenase-like genes that regulate shoot branching in *Arabidopsis* and pea. *Genes Dev.*, 17:1469–1474, 2003.
- [3970] D.Yu Sorokin, G.A. de Jong, L.A. Robertson, and G.J. Kuenen. Purification and characterization of sulfide dehydrogenase from alkaliphilic chemolithoautotrophic sulfur-oxidizing bacteria. *FEBS Lett.*, 427:11–14, 1998.
- [3971] J.C. Spain and D.T. Gibson. Pathway for bioremediation of *p*-nitrophenol in a *Moraxella* sp. *Appl. Environ. Microbiol.*, 57:812–819, 1991.
- [3972] F. Sparla, G. Tedeschi, and P. Trost. NAD(P)H:(quinone-acceptor) oxidoreductase of tobacco leaves is a flavin mononucleotide-containing flavoenzyme. *Plant Physiol.*, 112:249–258, 1996.
- [3973] L.G. Sparrow, P.P.K. Ho, T.K. Sundaram, D. Zach, E.J. Nyns, and E.E. Snell. The bacterial oxidation of vitamin B₆. VII. Purification, properties, and mechanism of action of an oxygenase which cleaves the 3-hydroxypyridine ring. *J. Biol. Chem.*, 244:2590–2600, 1969.
- [3974] T. Spector and V. Massey. *p*-Hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*. Evidence for an oxygenated flavin intermediate. *J. Biol. Chem.*, 247:5632–5636, 1972.
- [3975] T. Spector and V. Massey. *p*-Hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*. Reactivity with oxygen. *J. Biol. Chem.*, 247:7123–7127, 1972.
- [3976] T. Spector and V. Massey. Studies on the effector specificity of *p*-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*. *J. Biol. Chem.*, 247:4679–4687, 1972.
- [3977] D. Spencer. A reduced diphosphopyridine-specific nitrate reductase from germinating wheat. *Aust. J. Biol. Sci.*, 12:181–189, 1959.
- [3978] M.L. Speranza, S. Ronchi, and L. Minchiotti. Purification and characterization of yeast thioredoxin reductase. *Biochim. Biophys. Acta*, 327:274–281, 1973.
- [3979] P. Sperling, A. Blume, U. Zähringer, , and E. Further characterization of Δ^8 -sphingolipid desaturases from higher plants. *Biochem Soc Trans.*, 28:638–641, 2000.

- [3980] P. Sperling, M. Lee, T. Girke, U. Zähringer, S. Stymne, and E. Heinz. A bifunctional Δ^6 -fatty acyl acetylenase/desaturase from the moss *Ceratodon purpureus*. A new member of the cytochrome b_5 superfamily. *Eur. J. Biochem.*, 267:3801–3811, 2000.
- [3981] P. Sperling, B. Libisch, U. Zähringer, J.A. Napier, and E. Heinz. Functional identification of a Δ^8 -sphingolipid desaturase from *Borago officinalis*. *Arch. Biochem. Biophys.*, 388:293–298, 2001.
- [3982] P. Sperling, P. Ternes, H. Moll, S. Franke, U. Zähringer, and E. Heinz. Functional characterization of sphingolipid C4-hydroxylase genes from *Arabidopsis thaliana*. *FEBS Lett.*, 494:90–94, 2001.
- [3983] P. Sperling, U. Zähringer, and E. Heinz. A sphingolipid desaturase from higher plants. Identification of a new cytochrome b_5 fusion protein. *J. Biol. Chem.*, 273:28590–28596, 1998.
- [3984] J.F. Sperry and D.C. Robertson. Erythritol catabolism by *Brucella abortus*. *J. Bacteriol.*, 121:619–630, 1975.
- [3985] E. Spiess and H. Gorisch. Purification and characterization of chlorobenzene *cis*-dihydrodiol dehydrogenase from *Xanthobacter flavus* 14p1. *Arch. Microbiol.*, 165:201–205, 1996.
- [3986] E. Spiess, C. Sommer, and H. Gorisch. Degradation of 1,4-dichlorobenzene by *Xanthobacter flavus* 14p1. *Appl. Environ. Microbiol.*, 61:3884–3888, 1995.
- [3987] P. Spitteller, E. Glawischnig, A. Gierl, and W. Steglich. Studies on the biosynthesis of 2-hydroxy-1,4-benzoxazin-3-one (HBOA) from 3-hydroxyindolin-2-one in *Zea mays*. *Phytochemistry*, 57:373–376, 2001.
- [3988] G. Spohn, A. Kleinridders, F.T. Wunderlich, M. Watzka, F. Zaucke, K. Blumbach, C. Geisen, E. Seifried, C. Muller, M. Paulsson, J.C. Bruning, and J. Oldenburg. VKORC1 deficiency in mice causes early postnatal lethality due to severe bleeding. *Thromb Haemost.*, 101:1044–1050, 2009.
- [3989] T. Spolitak and D.P. Ballou. Evidence for catalytic intermediates involved in generating the chromopyrrolic acid scaffold of rebeccamycin by RebO and RebD. *Arch. Biochem. Biophys.*, 573:111–119, 2015.
- [3990] H. Sprecher. Metabolism of highly unsaturated *n*-3 and *n*-6 fatty acids. *Biochim. Biophys. Acta*, 1486:219–231, 2000.
- [3991] G. Spyrou, E. Haggård-Ljungquist, M. Krook, H. Jörnvall, E. Nilsson, and P. Reichard. Characterization of the flavin reductase gene (*fre*) of *Escherichia coli* and construction of a plasmid for overproduction of the enzyme. *J. Bacteriol.*, 173:3673–3679, 1991.
- [3992] S. Sridhara and T.T. Wu. Purification and properties of lactaldehyde dehydrogenase from *Escherichia coli*. *J. Biol. Chem.*, 244:5233–5238, 1969.
- [3993] B. St-Pierre and V. De Luca. A cytochrome *P*-450 monooxygenase catalyzes the first step in the conversion of tabersonine to vindoline in *Catharanthus roseus*. *Plant Physiol.*, 109:131–139, 1995.
- [3994] C.S. Stachow, I.L. Stevenson, and D. Day. Purification and properties of nicotinamide adenine dinucleotide phosphate-specific benzaldehyde dehydrogenase from *Pseudomonas*. *J. Biol. Chem.*, 242:5294–5300, 1967.
- [3995] R. Stadler and M.H. Zenk. The purification and characterization of a unique cytochrome *P*-450 enzyme from *Berberis stolonifera* plant cell cultures. *J. Biol. Chem.*, 268:823–831, 1993.
- [3996] T.C. Stadtman. Lysine metabolism by clostridia. XIIB 2,4-Diaminohexanoate dehydrogenase (2,4-diaminopentanoate dehydrogenase). *Adv. Enzymol. Relat. Areas Mol. Biol.*, 38:441–445, 1973.
- [3997] T.C. Stadtman, A. Cherkes, and C.B. Anfinsen. Studies on the microbiological degradation of cholesterol. *J. Biol. Chem.*, 206:511–523, 1954.
- [3998] T.C. Stadtman and P. Elliott. Studies on the enzymic reduction of amino acids. II. Purification and properties of a D-proline reductase and a proline racemase from *Clostridium sticklandii*. *J. Biol. Chem.*, 228:983–997, 1957.
- [3999] H.A. Stafford and H.H. Lester. Flavan-3-ol biosynthesis the conversion of (+)-dihydromyricetin to its flavan-3,4-diol (leucodelphinidin) and to (+)-gallocatechin by reductases extracted from tissue-cultures of *Ginkgo biloba* and *Pseudotsugamenziesii*. *Plant Physiol.*, 78:791–794, 1985.
- [4000] H.A. Stafford, A. Magaldi, and B. Vennesland. The enzymatic reduction of hydroxypyruvic acid to D-glyceric acid in higher plants. *J. Biol. Chem.*, 207:621–629, 1954.

- [4001] R.Y. Stanier and J.L. Ingraham. Protocatechuic acid oxidase. *J. Biol. Chem.*, 210:799–820, 1954.
- [4002] C.J. Stanley, L.C. Packman, M.J. Danson, C.E. Henderson, and R.N. Perham. Intramolecular coupling of active sites in the pyruvate dehydrogenase multienzyme complexes from bacterial and mammalian sources. *Biochem. J.*, 195:715–721, 1981.
- [4003] J.S. Stanley, J.L. York, and A.M. Benson. Nitroreductases and glutathione transferases that act on 4-nitroquinoline 1-oxide and their differential induction by butylated hydroxyanisole in mice. *Cancer Res.*, 52:58–63, 1992.
- [4004] C.R. Staples, E. Ameyibor, W. Fu, L. Gardet-Salvi, A.L. Stritt-Etter, P. Schurmann, D.B. Knaff, and M.K. Johnson. The function and properties of the iron-sulfur center in spinach ferredoxin: thioredoxin reductase: a new biological role for iron-sulfur clusters. *Biochemistry*, 35:11425–11434, 1996.
- [4005] A. Stapon, R. Li, and C.A. Townsend. Carbapenem biosynthesis: confirmation of stereochemical assignments and the role of CarC in the ring stereoinversion process from L-proline. *J. Am. Chem. Soc.*, 125:8486–8493, 2003.
- [4006] G.R. Stark and C.R. Dawson. Ascorbic acid oxidase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 8, pages 297–311. Academic Press, New York, 2nd edition, 1963.
- [4007] W.L. Starnes, P. Munk, S.B. Maul, G.N. Cunningham, D.J. Cox, and W. Shive. Threonine-sensitive aspartokinase-homoserine dehydrogenase complex, amino acid composition, molecular weight, and subunit composition of the complex. *Biochemistry*, 11:677–687, 1972.
- [4008] F.L.H. Stassen. Properties of highly purified lysyl oxidase from embryonic chick cartilage. *Biochim. Biophys. Acta*, 438:49–60, 1976.
- [4009] C. Steele, L. Gijzen, M. Qutob, D. Dixon, R.A. Molecular characterization of the enzyme catalyzing the aryl migration reaction of isoflavonoid biosynthesis in soybean. *Arch. Biochem. Biophys.*, 367:146–150, 1999.
- [4010] I.H. Steen, T. Lien, and N.-K. Birkeland. Biochemical and phylogenetic characterization of isocitrate dehydrogenase from a hyperthermophilic archaeon, *Archaeoglobus fulgidus*. *Arch. Microbiol.*, 168:412–420, 1997.
- [4011] D.J. Steenkamp and M. Husain. The effect of tetrahydrofolate on the reduction of electron transfer flavoprotein by sarcosine and dimethylglycine dehydrogenases. *Biochem. J.*, 203:707–715, 1982.
- [4012] D.J. Steenkamp and T.P. Singer. Participation of the iron-sulphur cluster and of the covalently bound coenzyme of trimethylamine dehydrogenase in catalysis. *Biochem. J.*, 169:361–369, 1978.
- [4013] N. Steffan, A. Grundmann, S. Afiyatullo, H. Ruan, and S.M. Li. FtmOx1, a non-heme Fe(II) and α -ketoglutarate-dependent dioxygenase, catalyses the endoperoxide formation of verruculogen in *Aspergillus fumigatus*. *Org. Biomol. Chem.*, 7:4082–4087, 2009.
- [4014] P. Steffens, N. Nagakura, and M.H. Zenk. The berberine bridge forming enzyme in tetrahydroprotoberberine biosynthesis. *Tetrahedron Lett.*, 25:951–952, 1984.
- [4015] S. Steiger, C. Astier, and G. Sandmann. Substrate specificity of the expressed carotenoid 3,4-desaturase from *Rubrivivax gelatinosus* reveals the detailed reaction sequence to spheroidene and spirilloxanthin. *Biochem. J.*, 349:635–640, 2000.
- [4016] S. Steiger, L. Perez-Fons, S.M. Cutting, P.D. Fraser, and G. Sandmann. Annotation and functional assignment of the genes for the C₃₀ carotenoid pathways from the genomes of two bacteria: *Bacillus indicus* and *Bacillus firmus*. *Microbiology*, 161:194–202, 2015.
- [4017] S. Steiger and G. Sandmann. Cloning of two carotenoid ketolase genes from *Nostoc punctiforme* for the heterologous production of canthaxanthin and astaxanthin. *Biotechnol. Lett.*, 26:813–817, 2004.
- [4018] R.A. Steiner, K.H. Kalk, and B.W. Dijkstra. Anaerobic enzyme-substrate structures provide insight into the reaction mechanism of the copper-dependent quercetin 2,3-dioxygenase. *Proc. Natl. Acad. Sci. USA*, 99:16625–16630, 2002.
- [4019] U. Steiner, W. Schliemann, and D. Strack. Assay for tyrosine hydroxylation activity of tyrosinase from betalain-forming plants and cell cultures. *Anal. Biochem.*, 238:72–75, 1996.
- [4020] C.R. Steinman and W.B. Jakoby. Yeast aldehyde dehydrogenase. I. Purification and crystallization. *J. Biol. Chem.*, 242:5019–5023, 1967.

- [4021] K. Stenklo, H.D. Thorell, H. Bergius, R. Aasa, and T. Nilsson. Chlorite dismutase from *Ideonella dechloratans*. *J. Biol. Inorg. Chem.*, 6:601–607, 2001.
- [4022] P. Stenmark, J. Grunler, J. Mattsson, P.J. Sindelar, P. Nordlund, and D.A. Berthold. A new member of the family of di-iron carboxylate proteins. Coq7 (clk-1), a membrane-bound hydroxylase involved in ubiquinone biosynthesis. *J. Biol. Chem.*, 276:33297–33300, 2001.
- [4023] S.L. Stenmark, D.L. Pierson, R.A. Jensen, and G.I. Glover. Blue-green bacteria synthesise L-tyrosine by the pretyrosine pathway. *Nature*, 247:290–292, 1974.
- [4024] J.R. Stern. Crystalline β -hydroxybutyrate dehydrogenase from pig heart. *Biochim. Biophys. Acta*, 26:448–449, 1957.
- [4025] J.R. Stern and R.W. O'Brien. Oxidation D-malic and β -alkylmalic acids wild-type and mutant strains of *Salmonella typhimurium* and by *Aerobacter aerogenes*. *J. Bacteriol.*, 98:147–151, 1969.
- [4026] J.D. Stewart. Cyclohexanone monooxygenase: a useful reagent for asymmetric Baeyer-Villiger reactions. *Curr. Org. Chem.*, 2:195–216, 1998.
- [4027] K. Stich and G. Forkmann. Biosynthesis of 3-deoxyanthocyanins with flower extracts from *Sinningia cardinalis*. *Phytochemistry*, 27:785–789, 1988.
- [4028] R.G. Stickland. Some properties of the malic enzyme of pigeon liver. 1. Conversion of malate into pyruvate. *Biochem. J.*, 73:646–654, 1959.
- [4029] R.G. Stickland. Some properties of the malic enzyme of pigeon liver. 2. Synthesis of malate from pyruvate. *Biochem. J.*, 73:654–659, 1959.
- [4030] A.L. Stigliani, G. Giorio, and C. D'Ambrosio. Characterization of P450 carotenoid β - and ϵ -hydroxylases of tomato and transcriptional regulation of xanthophyll biosynthesis in root, leaf, petal and fruit. *Plant Cell Physiol.*, 52:851–865, 2011.
- [4031] J.L. Still, M.V. Buell, W.E. Knox, and D.E. Green. Studies on the cyclophorase system. VII. D-Aspartic oxidase. *J. Biol. Chem.*, 179:831–837, 1949.
- [4032] J.L. Still and E. Sperling. On the prosthetic group of the D-aspartic oxidase. *J. Biol. Chem.*, 182:585–589, 1950.
- [4033] T.J. Stillman, P.D. Hempstead, P.J. Artymiuk, S.C. Andrews, A.J. Hudson, A. Treffry, J.R. Guest, and P.M. Harrison. The high-resolution X-ray crystallographic structure of the ferritin (EcFtnA) of *Escherichia coli*; comparison with human H ferritin (HuHF) and the structures of the Fe^{3+} and Zn^{2+} derivatives. *J. Mol. Biol.*, 307:587–603, 2001.
- [4034] C. Stines-Chaumeil, F. Talfournier, and G. Branlant. Mechanistic characterization of the MSDH (methylmalonate semi-aldehyde dehydrogenase) from *Bacillus subtilis*. *Biochem. J.*, 395:107–115, 2006.
- [4035] D.I. Stirling and H. Dalton. Properties of the methane mono-oxygenase from extracts of *Methylosinus trichosporium* OB3b and evidence for its similarity to the enzyme from *Methylococcus capsulatus* (Bath). *Eur. J. Biochem.*, 96:205–212, 1979.
- [4036] W. Stoffel, G. Assmann, and K. Bister. Metabolism of sphingosine bases. XVII. Stereospecificities in the introduction of the 4t-double bond into sphinganine yielding 4t-sphingenine (sphingosine). *Hoppe-Seyler's Z. Physiol. Chem.*, 352:1531–1544, 1971.
- [4037] W. Stoffel and W. Därr. 2-Alkenal reductase isolation, properties and specificities. *Hoppe-Seyler's Z. Physiol. Chem.*, 355:54–60, 1974.
- [4038] W. Stoffel, D. Le Kim, and G. Heyn. Metabolism of sphingosine bases. XIV. Sphinganine (dihydrosphingosine), an effective donor of the alk-1-enyl chain of plasmalogens. *Hoppe-Seyler's Z. Physiol. Chem.*, 351:875–883, 1970.
- [4039] W. Stoffel, D. Le Kim, and G. Sticht. Biosynthesis of dihydrosphingosine in vitro. *Hoppe-Seyler's Z. Physiol. Chem.*, 349:664–670, 1968.
- [4040] W. Stoffel, D. Le Kim, and G. Sticht. Metabolism of sphingosine bases. 8. Distribution, isolation and properties of D-3-oxosphinganine reductase. Stereospecificity of the NADPH-dependent reaction of 3-oxodihydrosphingosine (2-amino-1-hydroxyoctadecane-3-one). *Hoppe-Seyler's Z. Physiol. Chem.*, 349:1637–1644, 1968.

- [4041] W. Stoffel and D. LeKim. Studies on the biosynthesis of plasmalogens. Precursors in the biosynthesis of plasmalogens: on the stereospecificity of the biochemical dehydrogenation of the 1-*O*-alkyl glyceryl to the 1-*O*-alk-1'-enyl glyceryl ether bond. *Hoppe-Seylers Z. Physiol. Chem.*, 352:501–511, 1971.
- [4042] L. Stoffels, M. Krehenbrink, B.C. Berks, and G. Unden. Thiosulfate reduction in *Salmonella enterica* is driven by the proton motive force. *J. Bacteriol.*, 194:475–485, 2012.
- [4043] T. Stoisser, M. Brunsteiner, D.K. Wilson, and B. Nidetzky. Conformational flexibility related to enzyme activity: evidence for a dynamic active-site gatekeeper function of Tyr²¹⁵ in *Aerococcus viridans* lactate oxidase. *Sci. Rep.*, 6:27892–27892, 2016.
- [4044] A. Stojanowic, G.J. Mander, E.C. Duin, and R. Hedderich. Physiological role of the F₄₂₀-non-reducing hydrogenase (Mvh) from *Methanothermobacter marburgensis*. *Arch. Microbiol.*, 180:194–203, 2003.
- [4045] J.E. Stok and J. De Voss. Expression, purification, and characterization of BioI: a carbon-carbon bond cleaving cytochrome P450 involved in biotin biosynthesis in *Bacillus subtilis*. *Arch. Biochem. Biophys.*, 384:351–360, 2000.
- [4046] H. Stolterfoht, D. Schwendenwein, C.W. Sensen, F. Rudroff, and M. Winkler. Four distinct types of E.C. 1.2.1.30 enzymes can catalyze the reduction of carboxylic acids to aldehydes. *J. Biotechnol.*, 257:222–232, 2017.
- [4047] A. Stolz, B. Nortemann, and H.J. Knackmuss. Bacterial metabolism of 5-aminosalicylic acid. Initial ring cleavage. *Biochem. J.*, 282:675–680, 1992.
- [4048] J.F. Stolz and R.S. Oremland. Bacterial respiration of arsenic and selenium. *FEMS Microbiol. Rev.*, 23:615–627, 1999.
- [4049] J.M.H. Stoop, W.S. Chilton, and D.M. Pharr. Substrate specificity of the NAD-dependent mannitol dehydrogenase from celery. *Phytochemistry*, 43:1145–1150, 1996.
- [4050] J.M.H. Stoop and D.M. Pharr. Partial purification and characterization of mannitol: mannose 1-oxidoreductase from celeriac (*Apium graveolens* var. *rapaceum*) roots. *Arch. Biochem. Biophys.*, 298:612–619, 1992.
- [4051] J.M.H. Stoop, J.D. Williamson, M.A. Conkling, and D.M. Pharr. Purification of NAD-dependent mannitol dehydrogenase from celery suspension cultures. *Plant Physiol. (1995)*, 108:1219–1225, 1995.
- [4052] C. Stournaras, P. Maurer, and G. Kurz. 6-phospho-D-gluconate dehydrogenase from *Pseudomonas fluorescens*. Properties and subunit structure. *Eur. J. Biochem.*, 130:391–396, 1983.
- [4053] G.D. Straganz, A. Glieder, L. Brecker, D.W. Ribbons, and W. Steiner. Acetylaceton-cleaving enzyme Dke1: a novel C-C-bond-cleaving enzyme from *Acinetobacter johnsonii*. *Biochem. J.*, 369:573–581, 2003.
- [4054] M. Strassman and L.N. Ceci. Enzymatic formation of α -keto adipic acid from homoisocitric acid. *J. Biol. Chem.*, 240:4357–4361, 1965.
- [4055] F.B. Straub. Isolation and properties of a flavoprotein from heart muscle tissue. *Biochem. J.*, 33:787–792, 1939.
- [4056] G. Strauss and G. Fuchs. Enzymes of a novel autotrophic CO₂ fixation pathway in the phototrophic bacterium *Chloroflexus aurantiacus*, the 3-hydroxypropionate cycle. *Eur. J. Biochem.*, 215:633–643, 1993.
- [4057] H.J. Strecker. Glutamic dehydrogenase. *Arch. Biochem. Biophys.*, 46:128–140, 1953.
- [4058] H.J. Strecker. The interconversion of glutamic acid and proline. III. Δ^1 -Pyrroline-5-carboxylic acid dehydrogenase. *J. Biol. Chem.*, 235:3218–3223, 1960.
- [4059] H.J. Strecker and I. Harary. Bacterial butylene glycol dehydrogenase and diacetyl reductase. *J. Biol. Chem.*, 211:263–270, 1954.
- [4060] H.J. Strecker and S. Korke. Glucose dehydrogenase. *J. Biol. Chem.*, 196:769–784, 1952.
- [4061] B.R. Streit, R. Kant, M. Tokmina-Lukaszewska, A.I. Celis, M.M. Machovina, E.P. Skaar, B. Bothner, and J.L. DuBois. Time-resolved studies of IsdG protein identify molecular signposts along the non-canonical heme oxygenase pathway. *J. Biol. Chem.*, 291:862–871, 2016.
- [4062] S. Strickland and V. Massey. The mechanism of action of the flavoprotein melilotate hydroxylase. *J. Biol. Chem.*, 248:2953–2962, 1973.

- [4063] S. Strickland and V. Massey. The purification and properties of the flavoprotein melilotate hydroxylase. *J. Biol. Chem.*, 248:2944–2952, 1973.
- [4064] R.C. Strickler, D.F. Covey, and B. Tobias. Study of 3α , 20β -hydroxysteroid dehydrogenase with an enzyme-generated affinity alkylator: dual enzyme activity at a single active site. *Biochemistry*, 19:4950–4954, 1980.
- [4065] R.C. Strickler, B. Tobias, and D.F. Covey. Human placental 17β -estradiol dehydrogenase and 20α -hydroxysteroid dehydrogenase. Two activities at a single enzyme active site. *J. Biol. Chem.*, 256:316–321, 1981.
- [4066] M. Strieker, F. Kopp, C. Mahlert, L.O. Essen, and M.A. Marahiel. Mechanistic and structural basis of stereospecific C β -hydroxylation in calcium-dependent antibiotic, a daptomycin-type lipopeptide. *ACS Chem. Biol.*, 2:187–196, 2007.
- [4067] A. Strijewski. The steroid- 9α -hydroxylation system from *Nocardia* species. *Eur. J. Biochem.*, 128:125–135, 1982.
- [4068] P. Strittmatter. Microsomal cytochrome *b*₅ and cytochrome *b*₅ reductase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 8, pages 113–145. Academic Press, New York, 2nd edition, 1963.
- [4069] P. Strittmatter, L. Sputz, D. Corcoran, M.J. Rogers, B. Setlow, and R. Redline. Purification and properties of rat liver microsomal stearyl coenzyme A desaturase. *Proc. Natl. Acad. Sci. USA*, 71:4565–4569, 1974.
- [4070] P. Strittmatter and S.F. Velick. The purification and properties of microsomal cytochrome reductase. *J. Biol. Chem.*, 228:785–799, 1957.
- [4071] G.A. Strobel and T. Kosuge. Polyol metabolism in *Diplodia viticola* Desm. *Arch. Biochem. Biophys.*, 109:622–626, 1965.
- [4072] J.L. Strominger and L.W. Mapson. Uridine diphosphoglucose dehydrogenase of pea seedlings. *Biochem. J.*, 66:567–572, 1957.
- [4073] J.L. Strominger, E.S. Maxwell, J. Axelrod, and H.M. Kalckar. Enzymatic formation of uridine diphosphogluconic acid. *J. Biol. Chem.*, 224:79–90, 1957.
- [4074] N. Strushkevich, F. MacKenzie, T. Cherksova, I. Grabovec, S. Usanov, and H.W. Park. Structural basis for pregnenolone biosynthesis by the mitochondrial monooxygenase system. *Proc. Natl. Acad. Sci. USA*, 108:10139–10143, 2011.
- [4075] N. Strushkevich, S.A. Usanov, A.N. Plotnikov, G. Jones, and H.W. Park. Structural analysis of CYP2R1 in complex with vitamin D₃. *J. Mol. Biol.*, 380:95–106, 2008.
- [4076] J. Stubbe. Identification of two α -ketoglutarate-dependent dioxygenases in extracts of *Rhodotorula glutinis* catalyzing deoxyuridine hydroxylation. *J. Biol. Chem.*, 260:9972–9975, 1985.
- [4077] J. Stubbe, D. Ackles, R. Segal, and R.L. Blakley. On the mechanism of ribonucleoside triphosphate reductase from *Lactobacillus leichmannii*. Evidence for 3' C–H bond cleavage. *J. Biol. Chem.*, 256:4843–4846, 1981.
- [4078] J. Stubbe, M. Ator, and T. Krenitsky. Mechanism of ribonucleoside diphosphate reductase from *Escherichia coli*. Evidence for 3'-C–H bond cleavage. *J. Biol. Chem.*, 258:1625–1631, 1983.
- [4079] D. Stuehr, S. Pou, and G.M. Rosen. Oxygen reduction by nitric-oxide synthases. *J. Biol. Chem.*, 276:14533–14536, 2001.
- [4080] D.J. Stuehr, N.S. Kwon, C.F. Nathan, O.W. Griffith, P.L. Feldman, and J. Wiseman. N^ω-hydroxy-L-arginine is an intermediate in the biosynthesis of nitric oxide from L-arginine. *J. Biol. Chem.*, 266:6259–6263, 1991.
- [4081] U. Stuhlemmer and W. Kreis. Cardenolide formation and activity of pregnane-modifying enzymes in cell suspension cultures, shoot cultures and leaves of *Digitalis lanata*. *Plant Physiol.*, 34:85–91, 1996.
- [4082] S. Stymne and L.-A. Appelqvist. The biosynthesis of linoleate from oleoyl-CoA via oleoyl-phosphatidylcholine in microsomes of developing safflower seeds. *Eur. J. Biochem.*, 90:223–229, 1978.
- [4083] C. Su and E.H. Oliw. Purification and characterization of linoleate 8-dioxygenase from the fungus *Gaeumannomyces graminis* as a novel hemoprotein. *J. Biol. Chem.*, 271:14112–14118, 1996.
- [4084] C. Su and E.H. Oliw. Manganese lipoxygenase: Purification and characterization. *J. Biol. Chem.*, 273:13072–13079, 1998.

- [4085] J. Su, P. Bao, T. Bai, L. Deng, H. Wu, F. Liu, and J. He. CotA, a multicopper oxidase from *Bacillus pumilus* WH4, exhibits manganese-oxidase activity. *PLoS One*, 8:e60573–e60573, 2013.
- [4086] A. Subramanian, J. Wang, and G. Gil. STAT 5 and NF-Y are involved in expression and growth hormone-mediated sexually dimorphic regulation of cytochrome *P*₄₅₀ 3A10/lithocholic acid 6 β -hydroxylase. *Nucleic Acids Res.*, 26:2173–2178, 1998.
- [4087] V. Subramanian, T.-N. Liu, W.K. Yeh, and D.T. Gibson. Toluene dioxygenase: purification of an iron-sulfur protein by affinity chromatography. *Biochem. Biophys. Res. Commun.*, 91:1131–1139, 1979.
- [4088] V. Subramanian and C.S. Vaidyanathan. Anthranilate hydroxylase from *Aspergillus niger*: new type of NADPH-linked nonheme iron monooxygenase. *J. Bacteriol.*, 160:651–655, 1984.
- [4089] H. Sucipto, F. Kudo, and T. Eguchi. The last step of kanamycin biosynthesis: unique deamination reaction catalyzed by the α -ketoglutarate-dependent nonheme iron dioxygenase KanJ and the NADPH-dependent reductase KanK. *Angew. Chem. Int. Ed. Engl.*, 51:3428–3431, 2012.
- [4090] T. Suda, J.C. Robinson, and T.A. Fjellstedt. Purification and properties of α -ketoacid reductase, a newly discovered enzyme from human placenta. *Arch. Biochem. Biophys.*, 176:610–620, 1976.
- [4091] W.C. Suen, B.E. Haigler, and J.C. Spain. 2,4-Dinitrotoluene dioxygenase from *Burkholderia* sp. strain DNT: similarity to naphthalene dioxygenase. *J. Bacteriol.*, 178:4926–4934, 1996.
- [4092] Y. Sugai, Y. Katsuyama, and Y. Ohnishi. A nitrous acid biosynthetic pathway for diazo group formation in bacteria. *Nat. Chem. Biol.*, 12:73–75, 2016.
- [4093] Y. Sugano. DyP-type peroxidases comprise a novel heme peroxidase family. *Cell. Mol. Life Sci.*, 66:1387–1403, 2009.
- [4094] Y. Sugano, Y. Ishii, and M. Shoda. Role of H164 in a unique dye-decolorizing heme peroxidase DyP. *Biochem. Biophys. Res. Commun.*, 322:126–132, 2004.
- [4095] Y. Sugano, Y. Matsushima, K. Tsuchiya, H. Aoki, M. Hirai, and M. Shoda. Degradation pathway of an anthraquinone dye catalyzed by a unique peroxidase DyP from *Thanatephorus cucumeris* Dec 1. *Biodegradation*, 20:433–440, 2009.
- [4096] E. Sugimoto and L.I. Pizer. The mechanism of end product inhibition of serine biosynthesis. I. Purification and kinetics of phosphoglycerate dehydrogenase. *J. Biol. Chem.*, 243:2081–1089, 1968.
- [4097] H. Sugimoto, R. Shinkyo, K. Hayashi, S. Yoneda, M. Yamada, M. Kamakura, S. Ikushiro, Y. Shiro, and T. Sakaki. Crystal structure of CYP105A1 (P450SU-1) in complex with 1 α ,25-dihydroxyvitamin D₃. *Biochemistry*, 47:4017–4027, 2008.
- [4098] Y. Sugimoto, M. Yoshida, and B. Tamaoki. Purification of 5 β -reductase from hepatic cytosol fraction of chicken. *J. Steroid Biochem.*, 37:717–724, 1990.
- [4099] T. Sugisawa, T. Hoshino, and A. Fujiwara. Purification and properties of NADPH-linked L-sorbose reductase from *Gluconobacter melanogenus* N44-1. *Agric. Biol. Chem.*, 55:2043–2049, 1991.
- [4100] T. Sugisawa, T. Hoshino, S. Nomura, and A. Fujiwara. Isolation and characterization of membrane-bound L-sorbose dehydrogenase from *Gluconobacter melanogenus* UV10. *Agric. Biol. Chem.*, 55:363–370, 1991.
- [4101] M. Sugishima, C.T. Migita, X. Zhang, T. Yoshida, and K. Fukuyama. Crystal structure of heme oxygenase-1 from cyanobacterium *Synechocystis* sp. PCC 6803 in complex with heme. *Eur. J. Biochem.*, 271:4517–4525, 2004.
- [4102] M. Sugiura, M. Nakahara, C. Yamada, T. Arakawa, M. Kitaoka, and S. Fushinobu. Identification, functional characterization, and crystal structure determination of bacterial levoglucosan dehydrogenase. *J. Biol. Chem.*, 293:17375–17386, 2018.
- [4103] S. Sugiyama, K. Yano, and K. Arima. Metabolites of aromatic compounds by microbes. Part VII. Further studies of gentisic acid oxidase. *Bull. Agric. Chem. Soc. Jpn*, 24:249–254, 1960.
- [4104] S. Sugiyama, K. Yano, K. Komagata, and K. Arima. Metabolites of aromatic compounds by microbes. Part VII. Gentisic acid oxidase. *Bull. Agric. Chem. Soc. Jpn*, 24:243–248, 1960.
- [4105] T. Sugiyama and T. Yamano. Purification and crystallization of NADPH-adrenodoxin reductase from bovine adrenocortical mitochondria. *FEBS Lett.*, 52:145–148, 1975.

- [4106] K. Suhara, K. Ohashi, K. Takahashi, and M. Katagiri. Aromatase and nonaromatizing 10-demethylase activity of adrenal cortex mitochondrial P-450(11)beta. *Arch. Biochem. Biophys.*, 267:31–37, 1988.
- [4107] K. Suhara, S. Takemori, and M. Katagiri. The purification and properties of benzylalcohol dehydrogenase from *Pseudomonas* sp. *Arch. Biochem. Biophys.*, 130:422–429, 1969.
- [4108] Heering Suharti, de Vries H.A., and S. NO reductase from *Bacillus azotoformans* is a bifunctional enzyme accepting electrons from menaquinol and a specific endogenous membrane-bound cytochrome *c*₅₅₁. *Biochemistry*, 43:13487–13495, 2004.
- [4109] Strampraad Suharti, Schroder M.J., de Vries I., and S. A novel copper A containing menaquinol NO reductase from *Bacillus azotoformans*. *Biochemistry*, 40:2632–2639, 2001.
- [4110] N. Sukumar, Z.W. Chen, D. Ferrari, A. Merli, G.L. Rossi, H.D. Bellamy, A. Chistoserdov, V.L. Davidson, and F.S. Mathews. Crystal structure of an electron transfer complex between aromatic amine dehydrogenase and azurin from *Alcaligenes faecalis*. *Biochemistry*, 45:13500–13510, 2006.
- [4111] J.D. Sullivan and M. Ikawa. Purification and characterization of hexose oxidase from the red alga *Chondrus crispus*. *Biochim. Biophys. Acta*, 309:11–22, 1973.
- [4112] A. Sultana, I. Alexeev, I. Kursula, P. Mantsala, J. Niemi, and G. Schneider. Structure determination by multiwavelength anomalous diffraction of aclacinomycin oxidoreductase: indications of multidomain pseudomerohedral twinning. *Acta Crystallogr. D Biol. Crystallogr.*, 63:149–159, 2007.
- [4113] R.M. Summers, T.M. Louie, C.L. Yu, L. Gakhar, K.C. Louie, and M. Subramanian. Novel, highly specific *N*-demethylases enable bacteria to live on caffeine and related purine alkaloids. *J. Bacteriol.*, 194:2041–2049, 2012.
- [4114] R.M. Summers, T.M. Louie, C.L. Yu, and M. Subramanian. Characterization of a broad-specificity non-haem iron *N*-demethylase from *Pseudomonas putida* CBB5 capable of utilizing several purine alkaloids as sole carbon and nitrogen source. *Microbiology*, 157:583–592, 2011.
- [4115] R.M. Summers, S.K. Mohanty, S. Gopishetty, and M. Subramanian. Genetic characterization of caffeine degradation by bacteria and its potential applications. *Microb. Biotechnol.*, 8:369–378, 2015.
- [4116] C.B. Summitt, L.C. Johnson, T.J. Jonsson, D. Parsonage, R.P. Holmes, and W.T. Lowther. Proline dehydrogenase 2 (PRODH₂) is a hydroxyproline dehydrogenase (HYPDH) and molecular target for treating primary hyperoxaluria. *Biochem. J.*, 466:273–281, 2015.
- [4117] C.W. Sun, Z.W. Chen, Z.G. He, P.J. Zhou, and S.J. Liu. Purification and properties of the sulfur oxygenase/reductase from the acidothermophilic archaeon, *Acidianus* strain S5. *Extremophiles*, 7:131–134, 2003.
- [4118] L. Sun, M. Ruppert, Y. Sheludko, H. Warzecha, Y. Zhao, and J. Stockigt. Purification, cloning, functional expression and characterization of perakine reductase: the first example from the AKR enzyme family, extending the alkaloidal network of the plant *Rauvolfia*. *Plant Mol. Biol.*, 67:455–467, 2008.
- [4119] W. Sun and H.B. Dunford. Kinetics and mechanism of the peroxidase-catalyzed iodination of tyrosine. *Biochemistry*, 32:1324–1331, 1993.
- [4120] Z. Sun, E., Cunningham Gantt, , and Jr. Cloning and functional analysis of the β -carotene hydroxylase of *Arabidopsis thaliana*. *J. Biol. Chem.*, 271:24349–24352, 1996.
- [4121] H. Sund and H. Theorell. Alcohol dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrbäck, editors, *The Enzymes*, volume 7, pages 25–83. Academic Press, New York, 2nd edition, 1963.
- [4122] T.K. Sundaram and E.E. Snell. The bacterial oxidation of vitamin B₆. V. The enzymatic formation of pyridoxal and isopyridoxal from pyridoxine. *J. Biol. Chem.*, 244:2577–2584, 1969.
- [4123] M. Sundaramoorthy, J. Terner, and T.L. Poulos. The crystal structure of chloroperoxidase: a heme peroxidase⁻-cytochrome P450 functional hybrid. *Structure*, 3:1367–1377, 1995.
- [4124] F.W. Sunderman, Downs Jr., Reid J.R., Bibeau M.C., and L.M. Gas-chromatographic assay for heme oxygenase activity. *Clin. Chem.*, 28:2026–2032, 1982.

- [4125] O. Sundheim, C.B. Vågbø, M. Bjørås, M.M. Sousa, V. Talstad, P.A. Aas, F. Drabløs, H.E. Krokan, J.A. Tainer, and G. Slupphaug. Human ABH3 structure and key residues for oxidative demethylation to reverse DNA/RNA damage. *EMBO J.*, 25:3389–3397, 2006.
- [4126] J.A. Sundlov, D.M. Fontaine, T.L. Southworth, B.R. Branchini, and A.M. Gulick. Crystal structure of firefly luciferase in a second catalytic conformation supports a domain alternation mechanism. *Biochemistry*, 51:6493–6495, 2012.
- [4127] A.R. Sundquist and R.C. Fahey. The novel disulfide reductase bis- γ -glutamylcystine reductase and dihydrolipoamide dehydrogenase from *Halobacterium halobium*: purification by immobilized-metal-ion affinity chromatography and properties of the enzymes. *J. Bacteriol.*, 170:3459–3467, 1988.
- [4128] A.R. Sundquist and R.C. Fahey. The function of γ -glutamylcystine and bis- γ -glutamylcystine reductase in *Halobacterium halobium*. *J. Biol. Chem.*, 264:719–725, 1989.
- [4129] P.H. Sung, F.C. Huang, Y.Y. Do, and P.L. Huang. Functional expression of geraniol 10-hydroxylase reveals its dual function in the biosynthesis of terpenoid and phenylpropanoid. *J. Agric. Food Chem.*, 59:4637–4643, 2011.
- [4130] S. Supangat, Y.K. Choi, Y.S. Park, D. Son, C.D. Han, and K.H. Lee. Expression, purification, crystallization and preliminary X-ray analysis of sepiapterin reductase from *Chlorobium tepidum*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:202–204, 2005.
- [4131] J.M. Sutter, U. Johnsen, and P. Schönheit. Characterization of a pentonolactonase involved in D-xylose and L-arabinose catabolism in the haloarchaeon *Haloferax volcanii*. *FEMS Microbiol. Lett.*, 364, 2017.
- [4132] W.B. Sutton. Mechanism of action and crystalization of lactic oxidative decarboxylase from *Mycobacterium phlei*. *J. Biol. Chem.*, 226:395–405, 1957.
- [4133] S. Suye. Purification and properties of alcohol oxidase from *Candida methanosorbosa* M-2003. *Curr. Microbiol.*, 34:374–377, 1997.
- [4134] G.M.H. Suylen, P.J. Large, J.P. van Dijken, and J.G. Kuenen. Methylmercaptan oxidase, a key enzyme in the metabolism of methylated sulphur compounds by *Hyphomicrobium* EG. *J. Gen. Microbiol.*, 133:2989–2997, 1987.
- [4135] H. Suzuki, Y. Ohnishi, Y. Furusho, S. Sakuda, and S. Horinouchi. Novel benzene ring biosynthesis from C₃ and C₄ primary metabolites by two enzymes. *J. Biol. Chem.*, 281:36944–36951, 2006.
- [4136] I. Suzuki and M. Silver. The initial product and properties of the sulfur-oxidizing enzyme of thiobacilli. *Biochim. Biophys. Acta*, 122:22–33, 1966.
- [4137] K. Suzuki, T. Kaidoh, M. Katagiri, and T. Tsuchiya. O₂ incorporation into a long-chain fatty-acid during bacterial luminescence. *Biochim. Biophys. Acta*, 722:297–301, 1983.
- [4138] K. Suzuki, Y. Mano, and N. Shimazono. Conversion of L-gulonolactone to L-ascorbic acid; properties of the microsomal enzyme in rat liver. *J. Biochem. (Tokyo)*, 48:313–315, 1960.
- [4139] K. Suzuki, S. Takemori, and M. Katagiri. Mechanism of the salicylate hydroxylase reaction. IV. Fluorimetric analysis of the complex formation. *Biochim. Biophys. Acta*, 191:77–85, 1969.
- [4140] M. Suzuki. Purification and some properties of sarcosine oxidase from *Corynebacterium* sp. U-96. *J. Biochem. (Tokyo)*, 89:599–607, 1981.
- [4141] M. Suzuki, T. Hayakawa, J.P. Shaw, M. Rekik, and S. Harayama. Primary structure of xylene monooxygenase: similarities to and differences from the alkane hydroxylation system. *J. Bacteriol.*, 173:1690–1695, 1991.
- [4142] T. Suzuki. Purification and some properties of polyvinyl alcohol-degrading enzyme produced by *Pseudomonas O-3*. *Agric. Biol. Chem.*, 40:497–504, 1976.
- [4143] T. Suzuki. Oxidation of secondary alcohols by polyvinyl alcohol-degrading enzyme produced by *Pseudomonas O-3*. *Agric. Biol. Chem.*, 42:1187–1194, 1977.
- [4144] T. Suzuki, S. Yokoyama, Y. Kinoshita, H. Yamada, M. Hatsu, K. Takamizawa, and K. Kawai. Expression of *xyrA* gene encoding for D-xylose reductase of *Candida tropicalis* and production of xylitol in *Escherichia coli*. *J. Biosci. Bioeng.*, 87:280–284, 1999.

- [4145] Y. Suzuki, T. Yoda, A. Ruhul, and W. Sugiura. Molecular cloning and characterization of the gene coding for azoreductase from *Bacillus* sp. OY1-2 isolated from soil. *J. Biol. Chem.*, 276:9059–9065, 2001.
- [4146] A.V. Sviridov, T.V. Shushkova, N.F. Zelenkova, N.G. Vinokurova, I.G. Morgunov, I.T. Ermakova, and A.A. Leontievsky. Distribution of glyphosate and methylphosphonate catabolism systems in soil bacteria *Ochrobactrum anthropi* and *Achromobacter* sp. *Appl. Microbiol. Biotechnol.*, 93:787–796, 2012.
- [4147] M.A. Swairjo, R.R. Reddy, B. Lee, S.G. Van Lanen, S. Brown, V. de Crécy-Lagard, D. Iwata-Reuyl, and P. Schimmel. Crystallization and preliminary X-ray characterization of the nitrile reductase QueF: a queuosine-biosynthesis enzyme. *Acta Crystallogr. F Struct. Biol. Cryst. Commun.*, 61:945–948, 2005.
- [4148] S. Swaminathan, D. Morrone, Q. Wang, D.B. Fulton, and R.J. Peters. CYP76M7 is an *ent*-cassadiene C11 α -hydroxylase defining a second multifunctional diterpenoid biosynthetic gene cluster in rice. *Plant Cell*, 21:3315–3325, 2009.
- [4149] M.L. Sweat, L.T. Samuels, and R. Lumry. Preparation and characterisation of the enzyme which converts testosterone to androstendione. *J. Biol. Chem.*, 185:75–84, 1950.
- [4150] F. Sweet and B.S. Samant. Bifunctional enzyme activity at the same active site: study of 3 α and 20 β activity by affinity alkylation of 3 α , 20 β -hydroxysteroid dehydrogenase with 17-(bromoacetoxy)steroids. *Biochemistry*, 19:978–986, 1980.
- [4151] A.C. Swindell and J.L. Gaylor. Investigation of the component reactions of oxidative sterol demethylation. Formation and metabolism of 3-ketosteroid intermediates. *J. Biol. Chem.*, 243:5546–5555, 1968.
- [4152] C. Sygmund, M. Klausberger, A.K. Felice, and R. Ludwig. Reduction of quinones and phenoxy radicals by extracellular glucose dehydrogenase from *Glomerella cingulata* suggests a role in plant pathogenicity. *Microbiology*, 157:3203–3212, 2011.
- [4153] C. Sygmund, P. Staudigl, M. Klausberger, N. Pinotsis, K. Djinovic-Carugo, L. Gorton, D. Haltrich, and R. Ludwig. Heterologous overexpression of *Glomerella cingulata* FAD-dependent glucose dehydrogenase in *Escherichia coli* and *Pichia pastoris*. *Microb. Cell Fact.*, 10:106–106, 2011.
- [4154] M. Sylvestre, Y. Hurtubise, D. Barriault, J. Bergeron, and D. Ahmad. Characterization of active recombinant 2,3-dihydro-2,3-dihydroxybiphenyl dehydrogenase from *Comamonas testosteroni* B-356 and sequence of the encoding gene (bphB). *Appl. Environ. Microbiol.*, 62:2710–2715, 1996.
- [4155] C. Szameit, C. Miech, M. Balleininger, B. Schmidt, K. von Figura, and T. Dierks. The iron sulfur protein AtsB is required for posttranslational formation of formylglycine in the *Klebsiella* sulfatase. *J. Biol. Chem.*, 274:15375–15381, 1999.
- [4156] I.S.-Y. Sze and S. Dagley. Properties of salicylate hydroxylase and hydroxyquinol 1,2-dioxygenase purified from *Trichosporon cutaneum*. *J. Bacteriol.*, 159:353–359, 1984.
- [4157] B. Tabakoff and V.G. Erwin. Purification and characterization of a reduced nicotinamide adenine dinucleotide phosphate-linked aldehyde reductase from brain. *J. Biol. Chem.*, 245:3263–3268, 1970.
- [4158] C.W. Tabor and P.D. Kellogg. Identification of flavin adenine dinucleotide and heme in a homogeneous spermidine dehydrogenase from *Serratia marcescens*. *J. Biol. Chem.*, 245:5424–5433, 1970.
- [4159] H. Tabor and C.W. Tabor. Biosynthesis and metabolism of 1,4-diaminobutane, spermidine, spermine, and related amines. IIE2a Spermidine dehydrogenase. *Adv. Enzymol. Relat. Areas Mol. Biol.*, 36:225–226, 1972.
- [4160] M.E. Taga, N.A. Larsen, A.R. Howard-Jones, C.T. Walsh, and G.C. Walker. BluB cannibalizes flavin to form the lower ligand of vitamin B₁₂. *Nature*, 446:449–453, 2007.
- [4161] K. Tagawa and D.I. Arnon. Ferredoxin as electron carriers in photosynthesis and in the biological production and consumption of hydrogen gas. *Nature (Lond.)*, 195:537–543, 1962.
- [4162] K. Tagawa, M. Shin, and K. Okunuki. Peroxidases from wheat germ. *Nature (Lond.)*, 183:111–111, 1959.
- [4163] J.M. Tager and N. Rautanen. Sulfite oxidation by a plant mitochondrial system. I. Preliminary observations. *Biochim. Biophys. Acta*, 18:111–121, 1955.
- [4164] H. Taguchi and W.L. Armarego. Glyceryl-ether monooxygenase [EC 1.14.16.5]. A microsomal enzyme of ether lipid metabolism. *Med. Res. Rev.*, 18:43–89, 1998.

- [4165] T.M. Taha, T. Kanao, F. Takeuchi, and T. Sugio. Reconstitution of iron oxidase from sulfur-grown *Acidithiobacillus ferrooxidans*. *Appl. Environ. Microbiol.*, 74:6808–6810, 2008.
- [4166] H.-H. Tai and B. Yuan. Purification and assay of 9-hydroxyprostaglandin dehydrogenase from rat kidney. *Methods Enzymol.*, 86:113–117, 1982.
- [4167] G. Taibi, F. Di Gaudio, and C.M. Nicotra. Xanthine dehydrogenase processes retinol to retinoic acid in human mammary epithelial cells. *J. Enzyme Inhib. Med. Chem.*, 23:317–327, 2008.
- [4168] T. Takabe, Y. Iwasaki, T. Hibino, and T. Ando. Subunit composition of photosystem I complex that catalyzes light-dependent transfer of electrons from plastocyanin to ferredoxin. *J. Biochem.*, 110:622–627, 1991.
- [4169] M. Takahashi, M. Pischetsrieder, and V.M. Monnier. Molecular cloning and expression of amadoriase isoenzyme (fructosyl amine:oxygen oxidoreductase, EC 1.5.3) from *Aspergillus fumigatus*. *J. Biol. Chem.*, 272:12505–12507, 1997.
- [4170] S. Takahashi, T. Kuzuyama, H. Watanabe, and H. Seto. A 1-deoxy-D-xylulose 5-phosphate reductoisomerase catalyzing the formation of 2-C-methyl-D-erythritol 4-phosphate in an alternative nonmevalonate pathway for terpenoid biosynthesis. *Proc. Natl. Acad. Sci. USA*, 95:9879–9884, 1998.
- [4171] S. Takahashi, Y.S. Yeo, Y. Zhao, P.E. O'Maille, B.T. Greenhagen, J.P. Noel, R.M. Coates, and J. Chappell. Functional characterization of premnaspirodiene oxygenase, a cytochrome P_{450} catalyzing regio- and stereo-specific hydroxylations of diverse sesquiterpene substrates. *J. Biol. Chem.*, 282:31744–31754, 2007.
- [4172] S. Takahashi, Y. Zhao, P.E. O'Maille, B.T. Greenhagen, J.P. Noel, R.M. Coates, and J. Chappell. Kinetic and molecular analysis of 5-epiaristolochene 1,3-dihydroxylase, a cytochrome P_{450} enzyme catalyzing successive hydroxylations of sesquiterpenes. *J. Biol. Chem.*, 280:3686–3696, 2005.
- [4173] K. Takai, H. Ushiro, Y. Noda, S. Narumiya, T. Tokuyama, and O. Hayaishi. Crystalline hemoprotein from *Pseudomonas* that catalyzes oxidation of side chain of tryptophan and other indole derivatives. *J. Biol. Chem.*, 252:2648–2656, 1977.
- [4174] M. Takai, K. Kamimura, and T. Sugio. A new iron oxidase from a moderately thermophilic iron oxidizing bacterium strain TI-1. *Eur. J. Biochem.*, 268:1653–1658, 2001.
- [4175] N. Takakuwa, M. Kinoshita, Y. Oda, and M. Ohnishi. Isolation and characterization of the genes encoding Δ^8 -sphingolipid desaturase from *Saccharomyces kluyveri* and *Kluyveromyces lactis*. *Curr. Microbiol.*, 45:459–461, 2002.
- [4176] S. Takamatsu, L.H. Xu, S. Fushinobu, H. Shoun, M. Komatsu, D.E. Cane, and H. Ikeda. Pentalenic acid is a shunt metabolite in the biosynthesis of the pentalenolactone family of metabolites: hydroxylation of 1-deoxypentalenic acid mediated by CYP105D7 (SAV_7469) of *Streptomyces avermitilis*. *J. Antibiot. (Tokyo)*, 64:65–71, 2011.
- [4177] H. Takeda and O. Hayaishi. Crystalline L-lysine oxygenase. *J. Biol. Chem.*, 241:2733–2736, 1966.
- [4178] H. Takeda, S. Yamamoto, Y. Kojima, and O. Hayaishi. Studies on monooxygenases. I. General properties of crystalline L-lysine monooxygenase. *J. Biol. Chem.*, 244:2935–2941, 1969.
- [4179] S. Takemori, H. Yasuda, K. Mihara, K. Suzuki, and M. Katagiri. Mechanism of the salicylate hydroxylase reaction. 3. Characterization and reactivity of chemically or photochemically reduced enzyme-flavin. *Biochim. Biophys. Acta*, 191:69–76, 1969.
- [4180] S. Takemori, H. Yasuda, K. Mihara, K. Suzuki, and M. Katagiri. Mechanism of the salicylate hydroxylase reaction. II. The enzyme-substrate complex. *Biochim. Biophys. Acta*, 191:58–68, 1969.
- [4181] T. Takemura, N. Ikezawa, K. Iwasa, and F. Sato. Molecular cloning and characterization of a cytochrome P450 in sanguinarine biosynthesis from *Eschscholzia californica* cells. *Phytochemistry*, 91:100–108, 2013.
- [4182] S. Takenaka, S. Murakami, R. Shinke, K. Hatakeyama, H. Yukawa, and K. Aoki. Novel genes encoding 2-aminophenol 1,6-dioxygenase from *Pseudomonas* species AP-3 growing on 2-aminophenol and catalytic properties of the purified enzyme. *J. Biol. Chem.*, 272:14727–14732, 1997.
- [4183] M. Takeuchi, N. Asano, Y. Kameda, and K. Matsui. Physiological role of glucoside 3-dehydrogenase and cytochrome c_{551} in the sugar oxidizing system of *Flavobacterium saccharophilum*. *J. Biochem.*, 103:938–943, 1988.

- [4184] M. Takeuchi, K. Ninomiya, K. Kawabata, N. Asano, Y. Kameda, and K. Matsui. Purification and properties of glucoside 3-dehydrogenase from *Flavobacterium saccharophilum*. *J. Biochem.*, 100:1049–1055, 1986.
- [4185] T. Takeuchi, E.C. Weinbach, and L.S. Diamond. Pyruvate oxidase (CoA acetylating) in *Entamoeba histolytica*. *Biochem. Biophys. Res. Commun.*, 65:591–596, 1975.
- [4186] O. Takikawa, R. Yoshida, R. Kido, and O. Hayaishi. Tryptophan degradation in mice initiated by indoleamine 2,3-dioxygenase. *J. Biol. Chem.*, 261:3648–3653, 1986.
- [4187] A. Taku and R.A. Anwar. Biosynthesis of uridine diphospho-*N*-acetylmuramic acid. IV. Activation of uridine diphospho-*N*-acetylenolpyruvylglucosamine reductase by monovalent cations. *J. Biol. Chem.*, 248:4971–1976, 1973.
- [4188] A. Taku, K.G. Gunetileke, and R.A. Anwar. Biosynthesis of uridine diphospho-*N*-acetylmuramic acid. 3. Purification and properties of uridine diphospho-*N*-acetylenolpyruvyl-glucosamine reductase. *J. Biol. Chem.*, 245:5012–5016, 1970.
- [4189] Y. Takusagawa, M. Otagiri, S. Ui, T. Ohtsuki, A. Mimura, M. Ohkuma, and T. Kudo. Purification and characterization of L-2,3-butanediol dehydrogenase of *Brevibacterium saccharolyticum* C-1012 expressed in *Escherichia coli*. *Biosci. Biotechnol. Biochem.*, 65:1876–1878, 2001.
- [4190] P. Talalay and M.M. Dobson. Purification and properties of a α -hydroxysteroid dehydrogenase. *J. Biol. Chem.*, 205:823–837, 1953.
- [4191] K. Tamura, Y. Teranishi, S. Ueda, H. Suzuki, N. Kawano, K. Yoshimatsu, K. Saito, N. Kawahara, T. Muranaka, and H. Seki. Cytochrome P450 monooxygenase CYP716A141 is a unique β -amyrin C-16 β oxidase Involved in triterpenoid saponin biosynthesis in *Platycodon grandiflorus*. *Plant Cell Physiol.*, 58:874–884, 2017.
- [4192] B.C. Tan, S.H. Schwartz, J.A. Zeevaart, and D.R. McCarty. Genetic control of abscisic acid biosynthesis in maize. *Proc. Natl. Acad. Sci. USA*, 94:12235–12240, 1997.
- [4193] T. Tanahashi and M.H. Zenk. Elicitor induction and characterization of microsomal protopine-6-hydroxylase, the central enzyme in benzophenanthridine alkaloid biosynthesis. *Phytochemistry*, 29:1113–1122, 1990.
- [4194] K. Tanaka, M.A. Budd, M.L. Efron, and K.J. Isselbacher. Isovaleric acidemia: a new genetic defect of leucine metabolism. *Proc. Natl. Acad. Sci. USA*, 56:236–242, 1966.
- [4195] M. Tanaka and S. Tahara. FAD-dependent epoxidase as a key enzyme in fungal metabolism of prenylated flavonoids. *Phytochemistry*, 46:433–439, 1997.
- [4196] N. Tanaka and S. Murao. Reaction of bilirubin oxidase produced by *Myrothecium verrucaria* MT-1. *Agr. Biol. Chem.*, 49:843–844, 1985.
- [4197] R. Tanaka, U. Oster, E. Kruse, W. Rudiger, and B. Grimm. Reduced activity of geranylgeranyl reductase leads to loss of chlorophyll and tocopherol and to partially geranylgeranylated chlorophyll in transgenic tobacco plants expressing antisense RNA for geranylgeranyl reductase. *Plant Physiol.*, 120:695–704, 1999.
- [4198] S.W. Tanenbaum. The metabolism of *Acetobacter peroxidans*. I. Oxidative enzymes. *Biochim. Biophys. Acta*, 21:335–342, 1956.
- [4199] A. Tang and N.P. Curthoys. Identification of ζ -crystallin/NADPH:quinone reductase as a renal glutaminase mRNA pH response element-binding protein. *J. Biol. Chem.*, 276:21375–21380, 2001.
- [4200] H. Tang, Y. Tang, I.V. Kurnikov, H.J. Liao, N.L. Chan, M.G. Kurnikova, Y. Guo, and W.C. Chang. Harnessing the substrate promiscuity of dioxygenase AsqJ and developing efficient chemoenzymatic synthesis for quinolones. *ACS Catal.*, 11:7186–7192, 2021.
- [4201] H. Tang, L. Wang, W. Wang, H. Yu, K. Zhang, Y. Yao, and P. Xu. Systematic unraveling of the unsolved pathway of nicotine degradation in *Pseudomonas*. *PLoS Genet.*, 9:e1003923–e1003923, 2013.
- [4202] H. Tang, S. Wang, L. Ma, X. Meng, Z. Deng, D. Zhang, C. Ma, and P. Xu. A novel gene, encoding 6-hydroxy-3-succinoylpyridine hydroxylase, involved in nicotine degradation by *Pseudomonas putida* strain S16. *Appl. Environ. Microbiol.*, 74:1567–1574, 2008.

- [4203] H. Tang, Y. Yao, D. Zhang, X. Meng, L. Wang, H. Yu, L. Ma, and P. Xu. A novel NADH-dependent and FAD-containing hydroxylase is crucial for nicotine degradation by *Pseudomonas putida*. *J. Biol. Chem.*, 286:39179–39187, 2011.
- [4204] M.C. Tang, C.Y. Fu, and G.L. Tang. Characterization of SfmD as a heme peroxidase that catalyzes the regioselective hydroxylation of 3-methyltyrosine to 3-hydroxy-5-methyltyrosine in saframycin A biosynthesis. *J. Biol. Chem.*, 287:5112–5121, 2012.
- [4205] Z. Tang, S.G. Salamanca-Pinzon, Z.L. Wu, Y. Xiao, and F.P. Guengerich. Human cytochrome P450 4F11: heterologous expression in bacteria, purification, and characterization of catalytic function. *Arch. Biochem. Biophys.*, 494:86–93, 2010.
- [4206] Y. Tani, K. Tanaka, T. Yabutani, Y. Mishima, H. Sakuraba, T. Ohshima, and J. Motonaka. Development of a D-amino acids electrochemical sensor based on immobilization of thermostable D-proline dehydrogenase within agar gel membrane. *Anal. Chim. Acta*, 619:215–220, 2008.
- [4207] M. Tanigawa, T. Shinohara, M. Saito, K. Nishimura, Y. Hasegawa, S. Wakabayashi, M. Ishizuka, and Y. Nagata. D-Amino acid dehydrogenase from *Helicobacter pylori* NCTC 11637. *Amino Acids*, 38:247–255, 2010.
- [4208] H. Taniguchi, M. Hatanaka, S. Kuno, O. Hayaishi, M. Nakajima, and N. Kurihara. Enzymatic formation of catechol from anthranilic acid. *J. Biol. Chem.*, 239:2204–2211, 1964.
- [4209] H. Taniguchi, H. Mitsui, K. Nakamura, and F. Egami. *Ann. Acad. Sci. Fenn. Ser. A II*, 60:200–200, 1955.
- [4210] H. Taniuchi and O. Hayaishi. Studies on the metabolism of kynurenic acid. III. Enzymatic formation of 7,8-dihydroxykynurenic acid from kynurenic acid. *J. Biol. Chem.*, 238:283–293, 1963.
- [4211] G.J. Tanner, K.T. Francki, S. Abrahams, J.M. Watson, P.J. Larkin, and A.R. Ashton. Proanthocyanidin biosynthesis in plants: Purification of legume leucoanthocyanidin reductase and molecular cloning of its cDNA. *J. Biol. Chem.*, 278:31647–31656, 2003.
- [4212] G.J. Tanner and K.N. Kristiansen. Synthesis of 3,4-*cis*-[3H]leucocyanidin and enzymatic reduction to catechin. *Anal. Biochem.*, 209:274–277, 1993.
- [4213] J.J. Tanner, B. Lei, S.C. Tu, and K.L. Krause. Flavin reductase P: structure of a dimeric enzyme that reduces flavin. *Biochemistry*, 35:13531–13539, 1996.
- [4214] P. Tansakul and W. De-Eknamkul. Geranylgeraniol-18-hydroxylase: the last enzyme in the plau-notol biosynthetic pathway in *Croton sublyratus*. *Phytochemistry*, 47:1241–1246, 1998.
- [4215] L. Tao, A. Schenzle, J.M. Odom, and Q. Cheng. Novel carotenoid oxidase involved in biosynthesis of 4,4'-diapolycopene dialdehyde. *Appl. Environ. Microbiol.*, 71:3294–3301, 2005.
- [4216] L. Tao, H. Yao, H. Kasai, N. Misawa, and Q. Cheng. A carotenoid synthesis gene cluster from *Algoriphagus* sp. KK10202C with a novel fusion-type lycopene β -cyclase gene. *Mol. Genet. Genomics*, 276:79–86, 2006.
- [4217] M. Taton, T. Husselstein, P. Benveniste, and A. Rahier. Role of highly conserved residues in the reaction catalyzed by recombinant Δ^7 -sterol-C5(6)-desaturase studied by site-directed mutagenesis. *Biochemistry*, 39:701–711, 2000.
- [4218] M. Taton and A. Rahier. Plant sterol biosynthesis: identification and characterization of higher plant Δ^7 -sterol C5(6)-desaturase. *Arch. Biochem. Biophys.*, 325:279–288, 1996.
- [4219] F. Taura, S. Morimoto, and Y. Shoyama. Purification and characterization of cannabidiolic-acid synthase from *Cannabis sativa* L.. Biochemical analysis of a novel enzyme that catalyzes the oxidocyclization of cannabigerolic acid to cannabidiolic acid. *J. Biol. Chem.*, 271:17411–17416, 1996.
- [4220] F. Taura, S. Shoyama Morimoto, Mechoulam Y., and R. First direct evidence for the mechanism of Δ^1 -tetrahydrocannabinolic acid biosynthesis. *J. Am. Chem. Soc.*, 117:9766–9767, 1995.
- [4221] F. Taura, S. Sirikantaramas, Y. Shoyama, K. Yoshikai, Y. Shoyama, and S. Morimoto. Cannabidiolic-acid synthase, the chemotype-determining enzyme in the fiber-type *Cannabis sativa*. *FEBS Lett.*, 581:2929–2934, 2007.
- [4222] A. Taurog, M.L. Dorris, and D.R. Doerge. Mechanism of simultaneous iodination and coupling catalyzed by thyroid peroxidase. *Arch. Biochem. Biophys.*, 330:24–32, 1996.

- [4223] S. Tavares, T. Grotkjær, T. Obsen, R.P. Haslam, J.A. Napier, and N. Gunnarsson. Metabolic engineering of *Saccharomyces cerevisiae* for production of eicosapentaenoic acid, using a novel Δ^5 -desaturase from *Paramecium tetraurelia*. *Appl. Environ. Microbiol.*, 77:1854–1861, 2011.
- [4224] P. Tavladoraki, M.N. Rossi, G. Saccuti, M.A. Perez-Amador, F. Polticelli, R. Angelini, and R. Federico. Heterologous expression and biochemical characterization of a polyamine oxidase from *Arabidopsis* involved in polyamine back conversion. *Plant Physiol.*, 141:1519–1532, 2006.
- [4225] P. Tavladoraki, M.E. Schinina, F. Cecconi, S. Di Agostino, F. Manera, G. Rea, P. Mariottini, R. Federico, and R. Angelini. Maize polyamine oxidase: primary structure from protein and cDNA sequencing. *FEBS Lett.*, 426:62–66, 1998.
- [4226] A.B. Taylor, D.M. Benglis, Dhandayuthapani Jr., Hart S., and P.J. Structure of *Mycobacterium tuberculosis* methionine sulfoxide reductase A in complex with protein-bound methionine. *J. Bacteriol.*, 185:4119–4126, 2003.
- [4227] D.G. Taylor and P.W. Trudgill. Camphor revisited: studies of 2,5-diketocamphane 1,2-monooxygenase from *Pseudomonas putida* ATCC 17453. *J. Bacteriol.*, 165:489–497, 1986.
- [4228] M.B. Taylor and E. Juni. Stereoisomeric specificities of 2,3-butanediol dehydrogenase. *Biochim. Biophys. Acta*, 39:448–457, 1960.
- [4229] W.H. Taylor, M.L. Taylor, and D.F. Eames. Two functionally different dihydroorotic dehydrogenases in bacteria. *J. Bacteriol.*, 91:2251–2256, 1966.
- [4230] T.T. Tchen and K. Bloch. On the conversion of squalene to lanosterol in vitro. *J. Biol. Chem.*, 226:921–930, 1957.
- [4231] E.P. Tchesnokov, M. Fellner, E. Siakkou, T. Kleffmann, L.W. Martin, S. Aloï, I.L. Lamont, S.M. Wilbanks, and G.N. Jameson. The cysteine dioxygenase homologue from *Pseudomonas aeruginosa* is a 3-mercaptopropionate dioxygenase. *J. Biol. Chem.*, 290:24424–24437, 2015.
- [4232] A. Tchigvintsev, A. Singer, G. Brown, R. Flick, E. Evdokimova, K. Tan, C.F. Gonzalez, A. Savchenko, and A.F. Yakunin. Biochemical and structural studies of uncharacterized protein PA0743 from *Pseudomonas aeruginosa* revealed NAD⁺-dependent L-serine dehydrogenase. *J. Biol. Chem.*, 287:1874–1883, 2012.
- [4233] B.W. te Brömmelstroet, W.J. Geerts, J.T. Keltjens, C. van der Drift, and G.D. Vogels. Purification and properties of 5,10-methylenetetrahydromethanopterin dehydrogenase and 5,10-methylenetetrahydromethanopterin reductase, two coenzyme F₄₂₀-dependent enzymes, from *Methanosarcina barkeri*. *Biochim. Biophys. Acta*, 1079:293–302, 1991.
- [4234] B.W. te Brömmelstroet, C.M. Hensgens, W.J. Geerts, J.T. Keltjens, C. van der Drift, and G.D. Vogels. Purification and properties of 5,10-methylenetetrahydromethanopterin cyclohydrolase from *Methanosarcina barkeri*. *J. Bacteriol.*, 172:564–571, 1990.
- [4235] B.W. te Brömmelstroet, C.M. Hensgens, J.T. Keltjens, C. van der Drift, and G.D. Vogels. Purification and properties of 5,10-methylenetetrahydromethanopterin reductase, a coenzyme F₄₂₀-dependent enzyme, from *Methanobacterium thermoautotrophicum* strain ΔH^* . *J. Biol. Chem.*, 265:1852–1857, 1990.
- [4236] G. Tedeschi, A. Negri, M. Mortarino, F. Ceciliani, T. Simonic, L. Faotto, and S. Ronchi. L-Aspartate oxidase from *Escherichia coli*. II. Interaction with C4 dicarboxylic acids and identification of a novel L-aspartate: fumarate oxidoreductase activity. *Eur. J. Biochem.*, 239:427–433, 1996.
- [4237] J. Teixeira and G. Gil. Cloning, expression, and regulation of lithocholic acid 6 β -hydroxylase. *J. Biol. Chem.*, 266:21030–21036, 1991.
- [4238] A.H. Al Temimi, B.J. Pieters, Y.V. Reddy, P.B. White, and J. Mecinovic. Substrate scope for trimethyllysine hydroxylase catalysis. *Chem. Commun. (Camb.)*, 52:12849–12852, 2016.
- [4239] D.W. Tempest, J.L. Meers, and C.M. Brown. Synthesis of glutamate in *Aerobacter aerogenes* by a hitherto unknown route. *Biochem. J.*, 117:405–407, 1970.
- [4240] H.B. ten Brink, H.L. Dekker, H.E. Schoemaker, and R. Wever. Oxidation reactions catalyzed by vanadium chloroperoxidase from *Curvularia inaequalis*. *J. Inorg. Biochem.*, 80:91–98, 2000.
- [4241] H.B. ten Brink, A. Tuynman, H.L. Dekker, W. Hemrika, Y. Izumi, T. Oshiro, H.E. Schoemaker, and R. Wever. Enantioselective sulfoxidation catalyzed by vanadium haloperoxidases. *Inorg. Chem.*, 37:6780–6784, 1998.

- [4242] K.H. Teoh, D.R. Polichuk, D.W. Reed, G. Nowak, and P.S. Covello. *Artemisia annua* L. (Asteraceae) trichome-specific cDNAs reveal CYP71AV1, a cytochrome P450 with a key role in the biosynthesis of the antimalarial sesquiterpene lactone artemisinin. *FEBS Lett.*, 580:1411–1416, 2006.
- [4243] H. Teramoto, M. Inui, and H. Yukawa. Regulation of expression of genes involved in quinate and shikimate utilization in *Corynebacterium glutamicum*. *Appl. Environ. Microbiol.*, 75:3461–3468, 2009.
- [4244] M. Teramoto, N. Rahlert, N. Misawa, and G. Sandmann. 1-Hydroxy monocyclic carotenoid 3,4-dehydrogenase from a marine bacterium that produces myxol. *FEBS Lett.*, 570:184–188, 2004.
- [4245] A. Terebieniec, T. Chroumpi, A. Dilokpimol, M.V. Aguilar-Pontes, M.R. Makela, and R.P. de Vries. Characterization of D-xylose reductase, XyrB, from *Aspergillus niger*. *Biotechnol Rep (Amst)*, 30:e00610–e00610, 2021.
- [4246] P. Ternes, S. Franke, U. Zähringer, P. Sperling, and E. Heinz. Identification and characterization of a sphingolipid Δ^4 -desaturase family. *J. Biol. Chem.*, 277:25512–25518, 2002.
- [4247] M.J. Terry, J.A. Wahleithner, and J.C. Lagarias. Biosynthesis of the plant photoreceptor phytochrome. *Arch. Biochem. Biophys.*, 306:1–15, 1993.
- [4248] A. Tersteegen, D. Linder, R.K. Thauer, and R. Hedderich. Structures and functions of four anabolic 2-oxoacid oxidoreductases in *Methanobacterium thermoautotrophicum*. *Eur. J. Biochem.*, 244:862–868, 1997.
- [4249] R. Teufel, J.W. Kung, D. Kockelkorn, B.E. Alber, and G. Fuchs. 3-hydroxypropionyl-coenzyme A dehydratase and acryloyl-coenzyme A reductase, enzymes of the autotrophic 3-hydroxypropionate/4-hydroxybutyrate cycle in the *Sulfolobales*. *J. Bacteriol.*, 191:4572–4581, 2009.
- [4250] R. Teufel, V. Mascaraque, W. Ismail, M. Voss, J. Perera, W. Eisenreich, W. Haehnel, and G. Fuchs. Bacterial phenylalanine and phenylacetate catabolic pathway revealed. *Proc. Natl. Acad. Sci. USA*, 107:14390–14395, 2010.
- [4251] A.M. Thariath, K.L. Fatum, M.A. Valvano, and T. Viswanatha. Physico-chemical characterization of a recombinant cytoplasmic form of lysine: N^6 -hydroxylase. *Biochim. Biophys. Acta*, 1203:27–35, 1993.
- [4252] R. Theiler, J.C. Cook, L.P. Hager, and J.F. Siuda. Halohydrocarbon synthesis by bromoperoxidase. *Science*, 202:1094–1096, 1978.
- [4253] A. Theodossis, C.C. Milburn, N.I. Heyer, H.J. Lambie, D.W. Hough, M.J. Danson, and G.L. Taylor. Preliminary crystallographic studies of glucose dehydrogenase from the promiscuous Entner-Doudoroff pathway in the hyperthermophilic archaeon *Sulfolobus solfataricus*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, 61:112–115, 2005.
- [4254] A.D. Theoharides and D. Kupfer. Evidence for different hepatic microsomal monooxygenases catalyzing ω - and (ω -1)-hydroxylations of prostaglandins E1 and E2. Effects of inducers of monooxygenase on the kinetic constants of prostaglandin hydroxylation. *J. Biol. Chem.*, 256:2168–2175, 1981.
- [4255] H. Theorell. Das gelbe Oxydationsferment. *Biochem. Z.*, 278:263–290, 1935.
- [4256] H. Theorell. The preparation and some properties of crystalline horse-radish peroxidase. *Ark. Kemi Mineral. Geol.*, 16A No. 2:1–11, 1943.
- [4257] H. Theorell. Kinetics and equilibria in the liver alcohol dehydrogenase system. *Adv. Enzymol. Relat. Subj. Biochem.*, 20:31–49, 1958.
- [4258] H. Theorell and Å. Åkesson. Molecular weight and FMN content of crystalline "old yellow enzyme". *Arch. Biochem. Biophys.*, 65:439–448, 1956.
- [4259] H. Theorell, R.T. Holman, and Å. Åkesson. Crystalline lipoxidase. *Acta Chem. Scand.*, 1:571–576, 1947.
- [4260] N.V. Thoai and A. Olomucki. Arginine décarboxy-oxydase. I. Caractères et nature de l'enzyme. *Biochim. Biophys. Acta*, 59:533–544, 1962.
- [4261] N.V. Thoai and A. Olomucki. Arginine décarboxy-oxydase. II. Oxydation de la canavanine et de l'homoarginine en β -guanidopropionamide et en δ -guanidovaleramide. *Biochim. Biophys. Acta*, 59:545–552, 1962.
- [4262] J.B. Thoden and H.M. Holden. Structural and functional studies of WlbA: A dehydrogenase involved in the biosynthesis of 2,3-diacetamido-2,3-dideoxy-D-mannuronic acid. *Biochemistry*, 49:7939–7948, 2010.

- [4263] J.B. Thoden and H.M. Holden. Biochemical and structural characterization of WlbA from *Bordetella pertussis* and *Chromobacterium violaceum*: enzymes required for the biosynthesis of 2,3-diacetamido-2,3-dideoxy-D-mannuronic acid. *Biochemistry*, 50:1483–1491, 2011.
- [4264] M.G. Thomas, M.D. Burkart, and C.T. Walsh. Conversion of L-proline to pyrrolyl-2-carboxyl-S-PCP during undecylprodigiosin and pyoluteorin biosynthesis. *Chem. Biol.*, 9:171–184, 2002.
- [4265] P.E. Thomas, A.Y.H. Lu, D. Ryan, S.B. West, J. Kawalek, and W. Levin. Immunochemical evidence for six forms of rat liver cytochrome P_{450} obtained using antibodies against purified rat liver cytochromes P_{450} and P_{448} . *Mol. Pharmacol.*, 12:746–758, 1976.
- [4266] S.R. Thomas, P.M. McTamney, J.M. Adler, N. Laronde-Leblanc, and S.E. Rokita. Crystal structure of iodotyrosine deiodinase, a novel flavoprotein responsible for iodide salvage in thyroid glands. *J. Biol. Chem.*, 284:19659–19667, 2009.
- [4267] S.R. Thomas and R. Stocker. Redox reactions related to indoleamine 2,3-dioxygenase and tryptophan metabolism along the kynurenine pathway. *Redox Rep.*, 4:199–220, 1999.
- [4268] A.J. Thompson, A.C. Jackson, R.A. Parker, D.R. Morpeth, A. Burbidge, and I.B. Taylor. Abscisic acid biosynthesis in tomato: regulation of zeaxanthin epoxidase and 9-*cis*-epoxycarotenoid dioxygenase mRNAs by light/dark cycles, water stress and abscisic acid. *Plant Mol. Biol.*, 42:833–845, 2000.
- [4269] A.J. Thompson, A.C. Jackson, R.C. Symonds, B.J. Mulholland, A.R. Dadswell, P.S. Blake, A. Burbidge, and I.B. Taylor. Ectopic expression of a tomato 9-*cis*-epoxycarotenoid dioxygenase gene causes over-production of abscisic acid. *Plant J.*, 23:363–374, 2000.
- [4270] E.A. Thompson, Siiteri Jr., and P.K. The involvement of human placental microsomal cytochrome $P-450$ in aromatization. *J. Biol. Chem.*, 249:5373–5378, 1974.
- [4271] H. Thompson, A. Tersteegen, R.K. Thauer, and R. Hedderich. Two malate dehydrogenases in *Methanobacterium thermoautotrophicum*. *Arch. Microbiol.*, 170:38–42, 1998.
- [4272] J. Thompson. N^5 -(L-1-Carboxyethyl)-L-ornithine:NADP⁺ oxidoreductase from *Streptococcus lactis*. Purification and partial characterization. *J. Biol. Chem.*, 264:9592–9601, 1989.
- [4273] J.E. Thompson, G.S. Basarab, A. Andersson, Y. Lindqvist, and D.B. Jordan. Trihydroxynaphthalene reductase from *Magnaporthe grisea*: realization of an active center inhibitor and elucidation of the kinetic mechanism. *Biochemistry*, 36:1852–1860, 1997.
- [4274] M.G. Thompson, J.M. Blake-Hedges, P. Cruz-Morales, J.F. Barajas, S.C. Curran, C.B. Eiben, N.C. Harris, V.T. Benites, J.W. Gin, W.A. Sharpless, F.F. Twigg, W. Skyrud, R.N. Krishna, J.H. Pereira, E.EK. Baidoo, C.J. Petzold, P.D. Adams, A.P. Arkin, A.M. Deutschbauer, and J.D. Keasling. Massively parallel fitness profiling reveals multiple novel enzymes in *Pseudomonas putida* lysine metabolism. *mBio*, 10, 2019.
- [4275] R.E. Thompson and W.R. Carper. Glucose dehydrogenase from pig liver. I. Isolation and purification. *Biochim. Biophys. Acta*, 198:397–406, 1970.
- [4276] R.N. Thorneley, N.H. Bergstrom, R.R. Eady, and D.J. Lowe. Vanadium nitrogenase of *Azotobacter chroococcum*. MgATP-dependent electron transfer within the protein complex. *Biochem. J.*, 257:789–794, 1989.
- [4277] C. Thorpe and J.J. Kim. Structure and mechanism of action of the acyl-CoA dehydrogenases. *FASEB J.*, 9:718–725, 1995.
- [4278] J.S. Thorson, S.F. Lo, O. Ploux, X. He, and H.W. Liu. Studies of the biosynthesis of 3,6-dideoxyhexoses: molecular cloning and characterization of the asc (ascarylose) region from *Yersinia pseudotuberculosis* serogroup VA. *J. Bacteriol.*, 176:5483–5493, 1994.
- [4279] J.S. Thrower, R. Blalock, and J.P. Klinman. Steady-state kinetics of substrate binding and iron release in tomato ACC oxidase. *Biochemistry*, 40:9717–9724, 2001.
- [4280] T.T. Thuy, K. Liou, T.J. Oh, D.H. Kim, D.H. Nam, J.C. Yoo, and J.K. Sohng. Biosynthesis of dTDP-6-deoxy- β -D-allose, biochemical characterization of dTDP-4-keto-6-deoxyglucose reductase (GerKI) from *Streptomyces* sp. KCTC 0041BP. *Glycobiology*, 17:119–126, 2007.

- [4281] J.L. Thweatt, B.H. Ferlez, J.H. Golbeck, and D.A. Bryant. BciD is a radical *S*-adenosyl-L-methionine (SAM) enzyme that completes bacteriochlorophyllide *e* biosynthesis by oxidizing a methyl group into a formyl group at C-7. *J. Biol. Chem.*, 292:1361–1373, 2017.
- [4282] P. Tiainen, A. Pasanen, R. Sormunen, and J. Myllyharju. Characterization of recombinant human prolyl 3-hydroxylase isoenzyme 2, an enzyme modifying the basement membrane collagen IV. *J. Biol. Chem.*, 283:19432–19439, 2008.
- [4283] B. Tian, A. Strid, and L.A. Eriksson. Catalytic roles of active-site residues in 2-methyl-3-hydroxypyridine-5-carboxylic acid oxygenase: an ONIOM/DFT study. *J. Phys. Chem. B*, 115:1918–1926, 2011.
- [4284] B. Tian, Y. Tu, A. Strid, and L.A. Eriksson. Hydroxylation and ring-opening mechanism of an unusual flavoprotein monooxygenase, 2-methyl-3-hydroxypyridine-5-carboxylic acid oxygenase: a theoretical study. *Chemistry*, 16:2557–2566, 2010.
- [4285] L. Tian, V. Musetti, J. Kim, M. Magallanes-Lundback, and D. DellaPenna. The *Arabidopsis* LUT1 locus encodes a member of the cytochrome *P*₄₅₀ family that is required for carotenoid ϵ -ring hydroxylation activity. *Proc. Natl. Acad. Sci. USA*, 101:402–407, 2004.
- [4286] E.C. Tidswell, G.J. Salter, D.B. Kell, and J.G. Morris. Enantioselectivity of sulcatone reduction by some anaerobic bacteria. *Enzyme Microb. Technol.*, 21:143–147, 1997.
- [4287] E.C. Tidswell, A.N. Thompson, and J.G. Morris. Selection in chemostat culture of a mutant strain of *Clostridium trybutyricum* improved in its reduction of ketones. *J. Appl. Microbiol. Biotechnol.*, 35:317–322, 1991.
- [4288] K. Tiemann, W. Hinderer, and W. Barz. Isolation of NADPH:isoflavone oxidoreductase, a new enzyme of pterocarpan biosynthesis in cell suspensions of *Cicer arietinum*. *FEBS Lett.*, 213:324–328, 1987.
- [4289] A. Tietz, M. Lindberg, and E.P. Kennedy. A new pteridine-requiring enzyme system for the oxidation of glyceryl ethers. *J. Biol. Chem.*, 239:4081–4090, 1964.
- [4290] T.V. Tikhonova, D.Y. Sorokin, W.R. Hagen, M.G. Khrenova, G. Muyzer, T.V. Rakitina, I.G. Shabalin, A.A. Trofimov, S.I. Tsallagov, and V.O. Popov. Trinuclear copper biocatalytic center forms an active site of thiocyanate dehydrogenase. *Proc. Natl. Acad. Sci. USA*, 2020.
- [4291] S.-M. Ting, O.N. Miller, and O.Z. Sellinger. The metabolism of lactaldehyde. VII. The oxidation of D-lactaldehyde in rat liver. *Biochim. Biophys. Acta*, 97:407–415, 1965.
- [4292] S.-M. Ting, O.Z. Sellinger, and O.N. Miller. The metabolism of lactaldehyde. VI. The reduction of D- and L-lactaldehyde in rat liver. *Biochim. Biophys. Acta*, 89:217–225, 1964.
- [4293] A.J. Tipping and M.J. McPherson. Cloning and molecular analysis of the pea seedling copper amine oxidase. *J. Biol. Chem.*, 270:16939–16946, 1995.
- [4294] K.F. Tipton, S. Boyce, J. O’Sullivan, G.P. Davey, and J. Healy. Monoamine oxidases: certainties and uncertainties. *Curr. Med. Chem.*, 11:1965–1982, 2004.
- [4295] W. Tischer, J. Bader, and H. Simon. Purification and some properties of a hitherto-unknown enzyme reducing the carbon-carbon double bond of α,β -unsaturated carboxylate anions. *Eur. J. Biochem.*, 97:103–112, 1979.
- [4296] D. Tischler, R. Kermer, J.A. Groning, S.R. Kaschabek, W.J. van Berkel, and M. Schlomann. StyA1 and StyA2B from *Rhodococcus opacus* ICP: a multifunctional styrene monooxygenase system. *J. Bacteriol.*, 192:5220–5227, 2010.
- [4297] K. Tittmann, G. Wille, R. Golbik, A. Weidner, S. Ghisla, and G. Hübner. Radical phosphate transfer mechanism for the thiamin diphosphate- and FAD-dependent pyruvate oxidase from *Lactobacillus plantarum*. Kinetic coupling of intercofactor electron transfer with phosphate transfer to acetyl-thiamin diphosphate via a transient FAD semiquinone/hydroxyethyl-ThDP radical pair. *Biochemistry*, 44:13291–13303, 2005.
- [4298] D.R. Tocher, M.J. Leaver, and P.A. Hodgson. Recent advances in the biochemistry and molecular biology of fatty acyl desaturases. *Prog. Lipid Res.*, 37:73–117, 1998.
- [4299] C.J. Toews. The kinetics and reaction mechanism of the nicotinamide-adenine dinucleotide phosphate-specific glycerol dehydrogenase of rat skeletal muscle. *Biochem. J.*, 105:1067–1073, 1967.

- [4300] P.J. Du Toit and J.P. Kotzé. The isolation and characterization of sorbitol-6-phosphate dehydrogenase from *Clostridium pasteurianum*. *Biochim. Biophys. Acta*, 206:333–342, 1970.
- [4301] T. Tokieda, T. Niimura, F. Takamura, and T. Yamaha. Purification and some properties of cyclohexylamine oxidase from a *Pseudomonas* sp. *J. Biochem. (Tokyo)*, 81:851–858, 1977.
- [4302] A. Toll, K. Wikvall, E. Sudjana-Sugiaman, K.H. Kondo, and I. Björkhem. 7α hydroxylation of 25-hydroxycholesterol in liver microsomes. Evidence that the enzyme involved is different from cholesterol 7α -hydroxylase. *Eur. J. Biochem.*, 224:309–316, 1994.
- [4303] S. Tomita, M. Tsujita, and Y. Ichikawa. Retinal oxidase is identical to aldehyde oxidase. *FEBS Lett.*, 336:272–274, 1993.
- [4304] G.M. Tomkins. A mammalian 3α -hydroxysteroid dehydrogenase. *J. Biol. Chem.*, 218:437–447, 1956.
- [4305] G.M. Tomkins. The enzymatic reduction of Δ^4 -3-ketosteroids. *J. Biol. Chem.*, 225:13–24, 1957.
- [4306] G.M. Tomkins, P.J. Michael, and J.F. Curran. Studies on the nature of steroid 11- β hydroxylation. *Biochim. Biophys. Acta*, 23:655–656, 1957.
- [4307] G.M. Tonge, D.E.F. Harrison, and I.J. Higgins. Purification and properties of the methane mono-oxygenase enzyme system from *Methylosinus trichosporium* OB3b. *Biochem. J.*, 161:333–344, 1977.
- [4308] T. Tonon, D. Harvey, T.R. Larson, and I.A. Graham. Identification of a very long chain polyunsaturated fatty acid Δ^4 -desaturase from the microalga *Pavlova lutheri*. *FEBS Lett.*, 553:440–444, 2003.
- [4309] T. Tonon, D. Harvey, R. Qing, Y. Li, T.R. Larson, and I.A. Graham. Identification of a fatty acid Δ^{11} -desaturase from the microalga *Thalassiosira pseudonana*. *FEBS Lett.*, 563:28–34, 2004.
- [4310] T. Tonon, O. Sayanova, L.V. Michaelson, R. Qing, D. Harvey, T.R. Larson, Y. Li, J.A. Napier, and I.A. Graham. Fatty acid desaturases from the microalga *Thalassiosira pseudonana*. *FEBS J.*, 272:3401–3412, 2005.
- [4311] H.S. Toogood, A. van Thiel, J. Basran, M.J. Sutcliffe, N.S. Scrutton, and D. Leys. Extensive domain motion and electron transfer in the human electron transferring flavoprotein.medium chain Acyl-CoA dehydrogenase complex. *J. Biol. Chem.*, 279:32904–32912, 2004.
- [4312] R.E. Toomey and S.J. Wakil. Studies on the mechanism of fatty acid synthesis. XV. Preparation and general properties of β -ketoacyl acyl carrier protein reductase from *Escherichia coli*. *Biochim. Biophys. Acta*, 116:189–197, 1966.
- [4313] N. Toriyama. [Metabolism of quinoline derivatives. On the reducing enzyme of 4-nitroquinoline-*N*-oxide]. *Nichidai Igaku Zasshi*, 24:423–432, 1965.
- [4314] E Torres and M. Ayala. In *Biocatalysis based on heme peroxidases*, pages 7–110. Springer, Berlin, 2010.
- [4315] S. Tottey, M.A. Block, M. Allen, T. Westergren, C. Albrieux, H.V. Scheller, S. Merchant, and P.E. Jensen. *Arabidopsis* CHL27, located in both envelope and thylakoid membranes, is required for the synthesis of protochlorophyllide. *Proc. Natl. Acad. Sci. USA*, 100:16119–16124, 2003.
- [4316] O. Touster, V.H. Reynolds, and R.M. Hutcheson. The reduction of L-xylulose to xylitol by guinea pig liver mitochondria. *J. Biol. Chem.*, 221:697–709, 1956.
- [4317] C.A. Townsend. New reactions in clavulanic acid biosynthesis. *Curr. Opin. Chem. Biol.*, 6:583–589, 2002.
- [4318] H. Toyama, L. Chistoserdova, and M.E. Lidstrom. Sequence analysis of pqq genes required for biosynthesis of pyrroloquinoline quinone in *Methylobacterium extorquens* AM1 and the purification of a biosynthetic intermediate. *Microbiology*, 143:595–602, 1997.
- [4319] H. Toyama, A. Fujii, K. Matsushita, E. Shinagawa, M. Ameyama, and O. Adachi. Three distinct quinoprotein alcohol dehydrogenases are expressed when *Pseudomonas putida* is grown on different alcohols. *J. Bacteriol.*, 177:2442–2450, 1995.
- [4320] H. Toyama, H. Fukumoto, M. Saeki, K. Matsushita, O. Adachi, and M.E. Lidstrom. PqqC/D, which converts a biosynthetic intermediate to pyrroloquinoline quinone. *Biochem. Biophys. Res. Commun.*, 299:268–272, 2002.

- [4321] N. Traitcheva, H. Jenke-Kodama, J. He, E. Dittmann, and C. Hertweck. Non-colinear polyketide biosynthesis in the aureothin and neo-aureothin pathways: an evolutionary perspective. *ChemBioChem*, 8:1841–1849, 2007.
- [4322] T. Tralau, P. Lafite, C. Levy, J.P. Combe, N.S. Scrutton, and D. Leys. An internal reaction chamber in dimethylglycine oxidase provides efficient protection from exposure to toxic formaldehyde. *J. Biol. Chem.*, 284:17826–17834, 2009.
- [4323] U.C. Tran, B. Marbois, P. Gin, M. Gulmezian, T. Jonassen, and C.F. Clarke. Complementation of *Saccharomyces cerevisiae* coq7 mutants by mitochondrial targeting of the *Escherichia coli* UbiF polypeptide: two functions of yeast Coq7 polypeptide in coenzyme Q biosynthesis. *J. Biol. Chem.*, 281:16401–16409, 2006.
- [4324] T. Trautwein, F. Krauss, F. Lottspeich, and H. Simon. The (2R)-hydroxycarboxylate-viologen-oxidoreductase from *Proteus vulgaris* is a molybdenum-containing iron-sulphur protein. *Eur. J. Biochem.*, 222:1025–1032, 1994.
- [4325] E.P. Treacy, B.R. Akerman, L.M. Chow, R. Youil, C. Bibeau, J. Lin, A.G. Bruce, M. Knight, D.M. Danks, J.R. Cashman, and S.M. Forrest. Mutations of the flavin-containing monooxygenase gene (FMO3) cause trimethylaminuria, a defect in detoxication. *Hum. Mol. Genet.*, 7:839–845, 1998.
- [4326] C. Trefzer, H. Škovierová, S. Buroni, A. Bobovská, S. Nenci, E. Molteni, F. Pojer, M.R. Pasca, V. Makarov, S.T. Cole, G. Riccardi, K. Mikušová, and K. Johnsson. Benzothiazinones are suicide inhibitors of mycobacterial decaprenylphosphoryl- β -D-ribofuranose 2'-oxidase DprE1. *J. Am. Chem. Soc.*, 134:912–915, 2012.
- [4327] L.R. Treiber, R.A. Reamer, C.S. Rooney, and H.G. Ramjit. Origin of monacolin L from *Aspergillus terreus* cultures. *J. Antibiot. (Tokyo)*, 42:30–36, 1989.
- [4328] P. Trickey, M.A. Wagner, M.S. Jorns, and F.S. Mathews. Monomeric sarcosine oxidase: structure of a covalently flavinylated amine oxidizing enzyme. *Structure*, 7:331–345, 1999.
- [4329] C. Tricot, C. Vander Wauven, R. Wattiez, P. Falmagne, and V. Stalon. Purification and properties of a succinyltransferase from *Pseudomonas aeruginosa* specific for both arginine and ornithine. *Eur. J. Biochem.*, 224:853–861, 1994.
- [4330] B.C. Tripathy and C.A. Rebeiz. Chloroplast biogenesis 60. Conversion of divinyl protochlorophyllide to monovinyl protochlorophyllide in green(ing) barley, a dark monovinyl/light divinyl plant species. *Plant Physiol.*, 87:89–94, 1988.
- [4331] K.E. Tripodi, L.V. Buttiglieri, S.G. Altabe, and A.D. Uttaro. Functional characterization of front-end desaturases from trypanosomatids depicts the first polyunsaturated fatty acid biosynthetic pathway from a parasitic protozoan. *FEBS J.*, 273:271–280, 2006.
- [4332] S. Trippett, S. Dagley, and D.A. Stopher. The bacterial oxidation of nicotinic acid. *Biochem. J.*, 76:9–9, 1960.
- [4333] M.G. Tromp, G. Olafsson, B.E. Krenn, and R. Wever. Some structural aspects of vanadium bromoperoxidase from *Ascophyllum nodosum*. *Biochim. Biophys. Acta*, 1040:192–198, 1990.
- [4334] M.K. Trower, R.M. Buckland, R. Higgins, and M. Griffin. Isolation and characterization of a cyclohexane-metabolizing *Xanthobacter* sp. *Appl. Environ. Microbiol.*, 49:1282–1289, 1985.
- [4335] J.J. Truglio, K. Theis, S. Leimkuhler, R. Rappa, K.V. Rajagopalan, and C. Kisker. Crystal structures of the active and alloxanthine-inhibited forms of xanthine dehydrogenase from *Rhodobacter capsulatus*. *Structure*, 10:115–125, 2002.
- [4336] W.R. Tschantz, J.A. Digits, H.J. Pyun, R.M. Coates, and P.J. Casey. Lysosomal prenylcysteine lyase is a FAD-dependent thioether oxidase. *J. Biol. Chem.*, 276:2321–2324, 2001.
- [4337] B. Tshisuaka, R. Kappl, J. Huttermann, and F. Lingens. Quinoline oxidoreductase from *Pseudomonas putida* 86: an improved purification procedure and electron paramagnetic resonance spectroscopy. *Biochemistry*, 32:12928–12934, 1993.
- [4338] A. Tsuchii and K. Takeda. Rubber-degrading enzyme from a bacterial culture. *Appl. Environ. Microbiol.*, 56:269–274, 1990.
- [4339] Y. Tsuda and H.C. Friedmann. Ornithine metabolism by *Clostridium sticklandii*. Oxidation of ornithine to 2-amino-4-ketopentanoic acid via 2,4-diaminopentanoic acid; participation of B₁₂ coenzyme, pyridoxal phosphate, and pyridine nucleotide. *J. Biol. Chem.*, 245:5914–5926, 1970.

- [4340] W. Tsugawa, S. Horiuchi, M. Tanaka, H. Wake, and K. Sode. Purification of a marine bacterial glucose dehydrogenase from *Cytophaga marinoflava* and its application for measurement of 1,5-anhydro-D-glucitol. *Appl. Biochem. Biotechnol.*, 56:301–310, 1996.
- [4341] F.I. Tsuji, R.V. Lynch, and C.L. Stevens. Some properties of luciferase from the bioluminescent crustacean, *Cypridina hilgendorfi*. *Biochemistry*, 13:5204–5209, 1974.
- [4342] H. Tsuji, T. Ogawa, N. Bando, and K. Sasaoka. Purification and properties of 4-aminobenzoate hydroxylase, a new monooxygenase from *Agaricus bisporus*. *J. Biol. Chem.*, 261:13203–13209, 1986.
- [4343] K. Tsukada. D-Amino acid dehydrogenases of *Pseudomonas fluorescens*. *J. Biol. Chem.*, 241:4522–4528, 1966.
- [4344] Y. Tsukada, J. Fang, H. Erdjument-Bromage, M.E. Warren, C.H. Borchers, P. Tempst, and Y. Zhang. Histone demethylation by a family of JmjC domain-containing proteins. *Nature*, 439:811–816, 2006.
- [4345] Y. Tsukatani, J. Harada, J. Nomata, H. Yamamoto, Y. Fujita, T. Mizoguchi, and H. Tamiaki. *Rhodobacter sphaeroides* mutants overexpressing chlorophyllide *a* oxidoreductase of *Blastochloris viridis* elucidate functions of enzymes in late bacteriochlorophyll biosynthetic pathways. *Sci. Rep.*, 5:9741–9741, 2015.
- [4346] Y. Tsukatani, H. Yamamoto, J. Harada, T. Yoshitomi, J. Nomata, M. Kasahara, T. Mizoguchi, Y. Fujita, and H. Tamiaki. An unexpectedly branched biosynthetic pathway for bacteriochlorophyll *b* capable of absorbing near-infrared light. *Sci. Rep.*, 3:1217–1217, 2013.
- [4347] X. Tu, P.A. Hubbard, J.J. Kim, and H. Schulz. Two distinct proton donors at the active site of *Escherichia coli* 2,4-dienoyl-CoA reductase are responsible for the formation of different products. *Biochemistry*, 47:1167–1175, 2008.
- [4348] P.K. Tubbs and G.D. Greville. Dehydrogenation of D-lactate by a soluble enzyme from kidney mitochondria. *Biochim. Biophys. Acta*, 34:290–291, 1959.
- [4349] P.K. Tubbs and G.D. Greville. The oxidation of D- α -hydroxy acids in animal tissues. *Biochem. J.*, 81:104–114, 1961.
- [4350] J.J. Turnbull, M.J. Nagle, J.F. Seibel, R.W. Welford, G.H. Grant, and C.J. Schofield. The C-4 stereochemistry of leucocyanidin substrates for anthocyanidin synthase affects product selectivity. *Bioorg. Med. Chem. Lett.*, 13:3853–3857, 2003.
- [4351] J.J. Turnbull, J. Nakajima, R.W. Welford, M. Yamazaki, K. Saito, and C.J. Schofield. Mechanistic studies on three 2-oxoglutarate-dependent oxygenases of flavonoid biosynthesis: anthocyanidin synthase, flavonol synthase, and flavanone 3 β -hydroxylase. *J. Biol. Chem.*, 279:1206–1216, 2004.
- [4352] J.J. Turnbull, W.J. Sobey, R.T. Aplin, A. Hassan, J.L. Firmin, C.J. Schofield, and A.G. Prescott. Are anthocyanidins the immediate products of anthocyanidin synthase? *Chem. Commun.*, pages 2473–2474, 2000.
- [4353] J.F. Turner and J.E. King. Inosine 5-phosphate dehydrogenase of pea seeds. *Biochem. J.*, 79:147–147, 1961.
- [4354] J.M. Turner. Microbial metabolism of amino ketones. Aminoacetone formation from 1-aminopropan-2-ol by a dehydrogenase in *Escherichia coli*. *Biochem. J.*, 99:427–433, 1966.
- [4355] J.M. Turner. Microbial metabolism of amino ketones. L-1-Aminopropan-2-ol dehydrogenase and L-threonine dehydrogenase in *Escherichia coli*. *Biochem. J.*, 104:112–121, 1967.
- [4356] A. Tuynman, J.L. Spelberg, I.M. Kooter, H.E. Schoemaker, and R. Wever. Enantioselective epoxidation and carbon-carbon bond cleavage catalyzed by *Coprinus cinereus* peroxidase and myeloperoxidase. *J. Biol. Chem.*, 275:3025–3030, 2000.
- [4357] H. Twilfer, F.-H. Bernhardt, and K. Gersonde. An electron-spin-resonance study on the redox-active centers of the 4-methoxybenzoate monooxygenase from *Pseudomonas putida*. *Eur. J. Biochem.*, 119:595–602, 1981.
- [4358] C.A. Tyson, J.D. Lipscomb, and I.C. Gunsalus. The role of putidaredoxin and P450_{cam} in methylene hydroxylation. *J. Biol. Chem.*, 247:5777–5784, 1972.
- [4359] H. Uchida, D. Kondo, A. Yamashita, Y. Nagaosa, T. Sakurai, Y. Fujii, K. Fujishiro, K. Aisaka, and T. Uwajima. Purification and characterization of an aldehyde oxidase from *Pseudomonas* sp. KY 4690. *FEMS Microbiol. Lett.*, 229:31–36, 2003.

- [4360] K. Uchida, T. Shimizu, R. Makino, K. Sakaguchi, T. Iizuka, Y. Ishimura, T. Nozawa, and M. Hatano. Magnetic and natural circular dichroism of L-tryptophan 2,3-dioxygenases and indoleamine 2,3-dioxygenase. I. Spectra of ferric and ferrous high spin forms. *J. Biol. Chem.*, 258:2519–2525, 1983.
- [4361] S. Uchida and B. Vennesland. Properties of triphosphopyridine nucleotide-linked dihydroorotic dehydrogenase. *J. Biol. Chem.*, 237:2018–2024, 1962.
- [4362] S. Udenfriend and J.R. Cooper. The enzymic conversion of phenylalanine to tyrosine. *J. Biol. Chem.*, 194:503–511, 1952.
- [4363] D.W. Udway, L. K. Casillas, and C.A. Townsend. Synthesis of 11-hydroxyl *O*-methylsterigmatocystin and the role of a cytochrome *P*-450 in the final step of aflatoxin biosynthesis. *J. Am. Chem. Soc.*, 124:5294–5303, 2002.
- [4364] T. Ueda, E.T. Lode, and M.J. Coon. Enzymatic ω -oxidation. VI. Isolation of homogeneous reduced diphosphopyridine nucleotide-rubredoxin reductase. *J. Biol. Chem.*, 247:2109–2116, 1972.
- [4365] T. Ueda, E.T. Lode, and M.J. Coon. Enzymatic oxidation. VII. Reduced diphosphopyridine nucleotide-rubredoxin reductase: properties and function as an electron carrier in hydroxylation. *J. Biol. Chem.*, 247:5010–5016, 1972.
- [4366] K. Uehara and S. Hosomi. D-Erythulose reductase from beef liver. *Methods Enzymol.*, 89:232–237, 1982.
- [4367] K. Uehara and M. Takeda. L-Xylose dehydrogenase in bakers' yeast. *J. Biochem. (Tokyo)*, 52:461–463, 1962.
- [4368] K. Uehara, T. Tanimoto, and H. Sato. Studies on D-tetrose metabolism. IV. Purification and some properties of D-erythulose reductase from beef liver. *J. Biochem. (Tokyo)*, 75:333–345, 1974.
- [4369] K. Uehara, J. Watanabe, Y. Mogi, and Y. Tsukioka. Identification and characterization of an enzyme involved in the biosynthesis of the 4-hydroxy-2(or 5)-ethyl-5(or 2)-methyl-3(2*H*)-furanone in yeast. *J. Biosci. Bioeng.*, 123:333–341, 2017.
- [4370] T. Uetz, R. Schneider, M. Snozzi, and T. Egli. Purification and characterization of a two-component monooxygenase that hydroxylates nitrilotriacetate from "Chelatobacter" strain ATCC 29600. *J. Bacteriol.*, 174:1179–1188, 1992.
- [4371] S. Ui, Y. Okajima, A. Mimura, H. Kanai, T. Kobayashi, , and T. Sequence analysis of the gene for and characterization of D-acetoin forming *meso*-2,3-butanediol dehydrogenase of *Klebsiella pneumoniae* expressed in *Escherichia coli*. *J. Ferment. Bioeng.*, 83:32–37, 1997.
- [4372] A.J. Ullah, R.I. Murray, P.K. Bhattacharyya, G.C. Wagner, and I.C. Gunsalus. Protein components of a cytochrome *P*-450 linalool 8-methyl hydroxylase. *J. Biol. Chem.*, 265:1345–1351, 1990.
- [4373] R. Ullrich, C. Dolge, M. Kluge, and M. Hofrichter. Pyridine as novel substrate for regioselective oxygenation with aromatic peroxigenase from *Agrocybe aegerita*. *FEBS Lett.*, 582:4100–4106, 2008.
- [4374] R. Ullrich and M. Hofrichter. The haloperoxidase of the agaric fungus *Agrocybe aegerita* hydroxylates toluene and naphthalene. *FEBS Lett.*, 579:6247–6250, 2005.
- [4375] R. Ullrich, J. Nuske, K. Scheibner, J. Spantzel, and M. Hofrichter. Novel haloperoxidase from the agaric basidiomycete *Agrocybe aegerita* oxidizes aryl alcohols and aldehydes. *Appl. Environ. Microbiol.*, 70:4575–4581, 2004.
- [4376] Y. Umena, K. Yorita, T. Matsuoaka, A. Kita, K. Fukui, and Y. Morimoto. The crystal structure of L-lactate oxidase from *Aerococcus viridans* at 2.1 Å resolution reveals the mechanism of strict substrate recognition. *Biochem. Biophys. Res. Commun.*, 350:249–256, 2006.
- [4377] A. Upadhyay, F.L. Fontes, M. Gonzalez-Juarrero, M.R. McNeil, D.C. Crans, M. Jackson, and D.C. Crick. Partial saturation of menaquinone in *Mycobacterium tuberculosis*: function and essentiality of a novel reductase, MenJ. *ACS Cent. Sci.*, 1:292–302, 2015.
- [4378] Y. Urugami, T. Senda, K. Sugimoto, N. Sato, V. Nagarajan, E. Masai, M. Fukuda, and Y. Mitsu. Crystal structures of substrate free and complex forms of reactivated BphC, an extradiol type ring-cleavage dioxygenase. *J. Inorg. Biochem.*, 83:269–279, 2001.
- [4379] K. Urich. [D-Glutamate oxidase from the antennal gland of the crayfish *Oronectes limosus*: purification and characterization]. *Z. Naturforsch. B*, 23:1508–1511, 1968.

- [4380] T. Urich, T.M. Bandejas, S.S. Leal, R. Rachel, T. Albrecht, P. Zimmermann, C. Scholz, M. Teixeira, C.M. Gomes, and A. Kletzin. The sulphur oxygenase reductase from *Acidianus ambivalens* is a multimeric protein containing a low-potential mononuclear non-haem iron centre. *Biochem. J.*, 381:137–146, 2004.
- [4381] F. Ursini, M. Maiorino, and C. Gregolin. The selenoenzyme phospholipid hydroperoxide glutathione peroxidase. *Biochim. Biophys. Acta*, 839:62–70, 1985.
- [4382] K. Uyeda and J.C. Rabinowitz. Pyruvate-ferredoxin oxidoreductase. 3. Purification and properties of the enzyme. *J. Biol. Chem.*, 246:3111–3119, 1971.
- [4383] K. Uyeda and J.C. Rabinowitz. Pyruvate-ferredoxin oxidoreductase. IV. Studies on the reaction mechanism. *J. Biol. Chem.*, 246:3120–3125, 1971.
- [4384] G. Vaaje-Kolstad, L.A. Bohle, S. Gaseidnes, B. Dalhus, M. Bjoras, G. Mathiesen, and V.G. Eijsink. Characterization of the chitinolytic machinery of *Enterococcus faecalis* V583 and high-resolution structure of its oxidative CBM33 enzyme. *J. Mol. Biol.*, 416:239–254, 2012.
- [4385] G. Vaaje-Kolstad, B. Westereng, S.J. Horn, Z. Liu, H. Zhai, M. Sorlie, and V.G. Eijsink. An oxidative enzyme boosting the enzymatic conversion of recalcitrant polysaccharides. *Science*, 330:219–222, 2010.
- [4386] K. Vackova, A. Mehta, and M. Kutacek. Tryptophan aminotransferase and tryptophan dehydrogenase - activities in some cell compartments of spinach leaves - the effect of calcium-ions on tryptophan dehydrogenase. *Biol. Plant.*, 27:154–158, 1985.
- [4387] A. Vadas, H.G. Monbouquette, E. Johnson, and I. Schroder. Identification and characterization of a novel ferric reductase from the hyperthermophilic Archaeon *Archaeoglobus fulgidus*. *J. Biol. Chem.*, 274:36715–36721, 1999.
- [4388] F.H. Vaillancourt, J. Yin, and C.T. Walsh. SyrB2 in syringomycin E biosynthesis is a nonheme Fe^{II} α -ketoglutarate- and O₂-dependent halogenase. *Proc. Natl. Acad. Sci. USA*, 102:10111–10116, 2005.
- [4389] Z. Vajo, L.M. King, T. Jonassen, D.J. Wilkin, N. Ho, A. Munnich, C.F. Clarke, and C.A. Francomano. Conservation of the *Caenorhabditis elegans* timing gene *clk-1* from yeast to human: a gene required for ubiquinone biosynthesis with potential implications for aging. *Mamm Genome*, 10:1000–1004, 1999.
- [4390] K. Valegaard, A.C.T. van Scheltinga, M.D. Lloyd, T. Hara, S. Ramaswamy, A. Perrakis, A. Thompson, H.-J. Lee, J.E. Baldwin, C.J. Schofield, J. Hajdu, and I. Andersson. Structure of a cephalosporin synthase. *Nature*, 394:805–809, 1998.
- [4391] R.C. Valentine, L.E. Mortenson, and J.E. Carnahan. The hydrogenase system of *Clostridium pasteurianum*. *J. Biol. Chem.*, 238:1141–1144, 1963.
- [4392] M.P. Valley, S.E. Tichy, and P.F. Fitzpatrick. Establishing the kinetic competency of the cationic imine intermediate in nitroalkane oxidase. *J. Am. Chem. Soc.*, 127:2062–2066, 2005.
- [4393] F. Valverde, M. Losada, and A. Serrano. Cloning by functional complementation in *E. coli* of the *gap2* gene of *Synechocystis* PCC 6803 supports an amphibolic role for cyanobacterial NAD(P)-dependent glyceraldehyde-3-phosphate dehydrogenase. In P. Mathis, editor, *Photosynthesis: From Light to Biosphere*, volume 1, pages 959–962. Kluwer Academic Publishers, 1995.
- [4394] F. Valverde, M. Losada, and A. Serrano. Functional complementation of an *Escherichia coli* gap mutant supports an amphibolic role for NAD(P)-dependent glyceraldehyde-3-phosphate dehydrogenase of *Synechocystis* sp. strain PCC 6803. *J. Bacteriol.*, 179:4513–4522, 1997.
- [4395] J. Vamecq and J.P. Draye. The enzymatic and mass spectrometric identification of 2-oxophytanic acid, a product of the peroxisomal oxidation of *l*-2-hydroxyphytanic acid. *Biomed. Environ. Mass Spectrom.*, 15:345–351, 1988.
- [4396] W.H.J. van Berkel and W.J.J. van den Tweel. Purification and characterisation of 3-hydroxyphenylacetate 6-hydroxylase: a novel FAD-dependent monooxygenase from a *Flavobacterium* sp. *Eur. J. Biochem.*, 201:585–592, 1991.
- [4397] W.J.H. van Berkel, M.H.M. Eppink, W.J. Middelhoven, J. Vervoort, and I.M.C.M. Rietjens. Catabolism of 4-hydroxybenzoate in *Candida parapsilosis* proceeds through initial oxidative decarboxylation by a FAD-dependent 4-hydroxybenzoate 1-hydroxylase. *FEMS Microbiol. Lett.*, 121:207–216, 1994.

- [4398] E. van Bloois, D.E. Torres Pazmino, R.T. Winter, and M.W. Fraaije. A robust and extracellular heme-containing peroxidase from *Thermobifida fusca* as prototype of a bacterial peroxidase superfamily. *Appl. Microbiol. Biotechnol.*, 86:1419–1430, 2010.
- [4399] R.H. van den Heuvel, D. Ferrari, R.T. Bossi, S. Ravasio, B. Curti, M.A. Vanoni, F.J. Florencio, and A. Mattevi. Structural studies on the synchronization of catalytic centers in glutamate synthase. *J. Biol. Chem.*, 277:24579–24583, 2002.
- [4400] R.H. van den Heuvel, D.I. Svergun, M.V. Petoukhov, A. Coda, B. Curti, S. Ravasio, M.A. Vanoni, and A. Mattevi. The active conformation of glutamate synthase and its binding to ferredoxin. *J. Mol. Biol.*, 330:113–128, 2003.
- [4401] R.H. van den Heuvel, A.H. Westphal, A.J. Heck, M.A. Walsh, S. Rovida, W.J. van Berkel, and A. Mattevi. Structural studies on flavin reductase PheA2 reveal binding of NAD in an unusual folded conformation and support novel mechanism of action. *J. Biol. Chem.*, 279:12860–12867, 2004.
- [4402] C. van der Drift, P.E. van Helvoort, and G.D. Vogels. S-Ureidoglycolate dehydrogenase: purification and properties. *Arch. Biochem. Biophys.*, 145:465–469, 1971.
- [4403] M.J. van der Werf and A.M. Boot. Metabolism of carveol and dihydrocarveol in *Rhodococcus erythropolis* DCL14. *Microbiology*, 146:1129–1141, 2000.
- [4404] M.J. van der Werf, H.J. Swarts, and J.A. de Bont. *Rhodococcus erythropolis* DCL14 contains a novel degradation pathway for limonene. *Appl. Environ. Microbiol.*, 65:2092–2102, 1999.
- [4405] C.G. van Ginkel, G.B. Rikken, A.G.M. Kron, and S.W.M. Kengen. Purification and characterization of chlorite dismutase: a novel oxygen-generating enzyme. *Arch. Microbiol.*, 166:321–326, 1996.
- [4406] D.J. van Haaster, P.J. Silva, P.L. Hagedoorn, J.A. Jongejan, and W.R. Hagen. Reinvestigation of the steady-state kinetics and physiological function of the soluble NiFe-hydrogenase I of *Pyrococcus furiosus*. *J. Bacteriol.*, 190:1584–1587, 2008.
- [4407] J. van Heijenoort. Recent advances in the formation of the bacterial peptidoglycan monomer unit. *Nat. Prod. Rep.*, 18:503–519, 2001.
- [4408] R. van Heyningen and A. Pirie. Reduction of glutathione coupled with oxidative decarboxylation of malate in cattle lens. *Biochem. J.*, 53:436–444, 1953.
- [4409] J.E. van Hylckama Vlieg, J. Kingma, W. Kruizinga, and D.B. Janssen. Purification of a glutathione S-transferase and a glutathione conjugate-specific dehydrogenase involved in isoprene metabolism in *Rhodococcus* sp. strain AD45. *J. Bacteriol.*, 181:2094–2101, 1999.
- [4410] M.F.M. van Iersel, M.H.M. Eppink, W.J.H. van Berkel, F.M. Rombouts, and T. Abee. Purification and characterization of a novel NADP-dependent branched-chain alcohol dehydrogenase from *Saccharomyces cerevisiae*. *Appl. Environ. Microbiol.*, 63:4079–4082, 1997.
- [4411] M.A.G. van Kleef and J.A. Duine. Bacterial NAD(P)-independent quinate dehydrogenase is a quinoprotein. *Arch. Microbiol.*, 150:32–36, 1988.
- [4412] P.W. van Ophem and J.A. Duine. NAD- and co-substrate (GSH or factor)-dependent formaldehyde dehydrogenases from methylotrophic microorganisms act as a class III alcohol dehydrogenase. *FEMS Microbiol. Lett.*, 116:87–94, 1994.
- [4413] L.M. van Staalduinen, S.K. Novakowski, and Z. Jia. Structure and functional analysis of YcfD, a novel 2-oxoglutarate/Fe²⁺(+)-dependent oxygenase involved in translational regulation in *Escherichia coli*. *J. Mol. Biol.*, 426:1898–1910, 2014.
- [4414] E.E. van Tamelen, J.D. Willett, R.B. Clayton, and K.E. Lord. Enzymic conversion of squalene 2,3-oxide to lanosterol and cholesterol. *J. Am. Chem. Soc.*, 88:4752–4754, 1966.
- [4415] N. van Thoai, C. Huc, D.B. Pho, and A. Olomucki. Octopine déhydrogénase. Purification et propriétés catalytiques. *Biochim. Biophys. Acta*, 191:46–57, 1969.
- [4416] J.J. van Thor, T.H. Geerlings, H.C. Matthijs, and K.J. Hellingwerf. Kinetic evidence for the PsaE-dependent transient ternary complex photosystem I/Ferredoxin/Ferredoxin:NADP⁺ reductase in a cyanobacterium. *Biochemistry*, 38:12735–12746, 1999.

- [4417] P.G. Vance, B.B. Keele, and K.V. Rajagopalan. Superoxide dismutase from *Streptococcus mutans*. Isolation and characterization of two forms of the enzyme. *J. Biol. Chem.*, 247:4782–4786, 1972.
- [4418] I. Vancurová, A. Vancura, J. Volc, J. Neuzil, M. Flieger, G. Basarová, and V. Behal. Isolation and characterization of valine dehydrogenase from *Streptomyces aureofaciens*. *J. Bacteriol.*, 170:5192–5196, 1988.
- [4419] I. Vancurova, J. Volc, M. Flieger, J. Neuzil, J. Novotna, J. Vlach, and V. Behal. Isolation of pure anhydrotetracycline oxygenase from *Streptomyces aureofaciens*. *Biochem. J.*, 253:263–267, 1988.
- [4420] R.N. vanden Hoven and J.M. Santini. Arsenite oxidation by the heterotroph *Hydrogenophaga* sp. str. NT-14: the arsenite oxidase and its physiological electron acceptor. *Biochim. Biophys. Acta*, 1656:148–155, 2004.
- [4421] A.S. Vangnai and D.J. Arp. An inducible 1-butanol dehydrogenase, a quinohaemoprotein, is involved in the oxidation of butane by ‘*Pseudomonas butanovora*’. *Microbiology*, 147:745–756, 2001.
- [4422] A.S. Vangnai, D.J. Arp, and L.A. Sayavedra-Soto. Two distinct alcohol dehydrogenases participate in butane metabolism by *Pseudomonas butanovora*. *J. Bacteriol.*, 184:1916–1924, 2002.
- [4423] A.S. Vangnai, L.A. Sayavedra-Soto, and D.J. Arp. Roles for the two 1-butanol dehydrogenases of *Pseudomonas butanovora* in butane and 1-butanol metabolism. *J. Bacteriol.*, 184:4343–4350, 2002.
- [4424] A.S. Vangnai, H. Toyama, W. De-Eknamkul, N. Yoshihara, O. Adachi, and K. Matsushita. Quinate oxidation in *Gluconobacter oxydans* IFO3244: purification and characterization of quinoprotein quinate dehydrogenase. *FEMS Microbiol. Lett.*, 241:157–162, 2004.
- [4425] M.A. Vanoni and B. Curti. Glutamate synthase: a complex iron-sulfur flavoprotein. *Cell. Mol. Life Sci.*, 55:617–638, 1999.
- [4426] F.A. Vazquez-Flota and V. De Luca. Developmental and light regulation of desacetoxyvindoline 4-hydroxylase in *Catharanthus roseus* (L.) G. Don. Evidence of a multilevel regulatory mechanism. *Plant Physiol.*, 117:1351–1361, 1998.
- [4427] J.M. Vega and R.H. Garrett. Siroheme: a prosthetic group of the *Neurospora crassa* assimilatory nitrite reductase. *J. Biol. Chem.*, 250:7980–7989, 1975.
- [4428] J.M. Vega, M.G. Guerrero, E. Leadbetter, and M. Losada. Reduced nicotinamide-adenine dinucleotide-nitrite reductase from *Azotobacter chroococcum*. *Biochem. J.*, 133:701–708, 1973.
- [4429] S.F. Velick and C. Furfine. Glyceraldehyde 3-phosphate dehydrogenase. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 7, pages 243–273. Academic Press, New York, 2nd edition, 1963.
- [4430] T. Vellosillo, M. Martinez, M.A. Lopez, J. Vicente, T. Cascon, L. Dolan, M. Hamberg, and C. Castresana. Oxylipins produced by the 9-lipoxygenase pathway in *Arabidopsis* regulate lateral root development and defense responses through a specific signaling cascade. *Plant Cell*, 19:831–846, 2007.
- [4431] M.A. Vences-Guzman, Z. Guan, E. Ormeno-Orrillo, N. Gonzalez-Silva, I.M. Lopez-Lara, E. Martinez-Romero, O. Geiger, and C. Sohlenkamp. Hydroxylated ornithine lipids increase stress tolerance in *Rhizobium tropici* CIAT899. *Mol. Microbiol.*, 79:1496–1514, 2011.
- [4432] S.S. Venceslau, Y. Stockdreher, C. Dahl, and I.A. Pereira. The “bacterial heterodisulfide” DsrC is a key protein in dissimilatory sulfur metabolism. *Biochim. Biophys. Acta*, 1837:1148–1164, 2014.
- [4433] M. Venegas-Caleron, A.M. Muro-Pastor, R. Garces, and E. Martinez-Force. Functional characterization of a plastidial ω -3 desaturase from sunflower (*Helianthus annuus*) in cyanobacteria. *Plant Physiol. Biochem.*, 44:517–525, 2006.
- [4434] P. Venkatasubramanian, L. Daniels, and J.P. Rosazza. Reduction of carboxylic acids by *Nocardia* aldehyde oxidoreductase requires a phosphopantetheinylated enzyme. *J. Biol. Chem.*, 282:478–485, 2007.
- [4435] C. Verduyn, R. Van Kleef, J. Frank, H. Schreuder, J.P. Van Dijken, and W.A. Scheffers. Properties of the NAD(P)H-dependent xylose reductase from the xylose-fermenting yeast *Pichia stipitis*. *Biochem. J.*, 226:669–677, 1985.
- [4436] B. Vergauwen, F. Pauwels, F. Jacquemotte, T.E. Meyer, M.A. Cusanovich, R.G. Bartsch, and J.J. Van Beeumen. Characterization of glutathione amide reductase from *Chromatium gracile*. Identification of a novel thiol peroxidase (Prx/Grx) fueled by glutathione amide redox cycling. *J. Biol. Chem.*, 276:20890–20897, 2001.

- [4437] B. Vergauwen, F. Van Petegem, H. Remaut, F. Pauwels, and J.J. Van Beeumen. Crystallization and preliminary X-ray crystallographic analysis of glutathione amide reductase from *Chromatium gracile*. *Acta Crystallogr. D Biol. Crystallogr.*, 58:339–340, 2002.
- [4438] M.F. Verhagen, T. O'Rourke, and M.W. Adams. The hyperthermophilic bacterium, *Thermotoga maritima*, contains an unusually complex iron-hydrogenase: amino acid sequence analyses versus biochemical characterization. *Biochim. Biophys. Acta*, 1412:212–229, 1999.
- [4439] N. Verma and P. Reeves. Identification and sequence of *rfbS* and *rfbE*, which determine antigenic specificity of group A and group D salmonellae. *J. Bacteriol.*, 171:5694–5701, 1989.
- [4440] M. Véron, F. Falcoz-Kelly, and G.N. Cohen. The threonine-sensitive homoserine dehydrogenase and aspartokinase activities of *Escherichia coli* K12. The two catalytic activities are carried by two independent regions of the polypeptide chain. *Eur. J. Biochem.*, 28:520–527, 1972.
- [4441] N.D. Vetter, D.M. Langill, S. Anjum, J. Boisvert-Martel, R.C. Jagdhane, E. Omene, H. Zheng, K.E. van Straaten, I. Asiamah, E.S. Krol, D.A. Sanders, and D.R. Palmer. A previously unrecognized kanosamine biosynthesis pathway in *Bacillus subtilis*. *J. Am. Chem. Soc.*, 135:5970–5973, 2013.
- [4442] J.L. Vey, A. Al-Mestarihi, Y. Hu, M.A. Funk, B.O. Bachmann, and T.M. Iverson. Structure and mechanism of ORF36, an amino sugar oxidizing enzyme in everninomicin biosynthesis. *Biochemistry*, 49:9306–9317, 2010.
- [4443] G. Vialart, A. Hehn, A. Olry, K. Ito, C. Krieger, R. Larbat, C. Paris, B. Shimizu, Y. Sugimoto, M. Mizutani, and F. Bourgaud. A 2-oxoglutarate-dependent dioxygenase from *Ruta graveolens* L. exhibits *p*-coumaroyl CoA 2'-hydroxylase activity (C2'H): a missing step in the synthesis of umbelliferone in plants. *Plant J.*, 70:460–470, 2012.
- [4444] B.A. Vick and D.C. Zimmerman. Characterization of 12-oxo-phytodienoic acid reductase in corn - the jasmonic acid pathway. *Plant Physiol.*, 80:202–205, 1986.
- [4445] T.J. Vickers, G. Orsomando, R.D. de la Garza, D.A. Scott, S.O. Kang, A.D. Hanson, and S.M. Beverley. Biochemical and genetic analysis of methylenetetrahydrofolate reductase in *Leishmania* metabolism and virulence. *J. Biol. Chem.*, 281:38150–38158, 2006.
- [4446] H.B. Vickery. A suggested new nomenclature for the isomers of isocitric acid. *J. Biol. Chem.*, 237:1739–1741, 1962.
- [4447] A. Vidal-Cros, F. Viviani, G. Labesse, M. Boccara, and M. Gaudry. Polyhydroxynaphthalene reductase involved in melanin biosynthesis in *Magnaporthe grisea*. Purification, cDNA cloning and sequencing. *Eur. J. Biochem.*, 219:985–992, 1994.
- [4448] M. Vidal-Lieria and N. van Uden. Inositol dehydrogenase from the yeast *Cryptococcus melibiosum*. *Biochim. Biophys. Acta*, 293:295–303, 1973.
- [4449] P.M. Vignais, B. Billoud, and J. Meyer. Classification and phylogeny of hydrogenases. *FEMS Microbiol. Rev.*, 25:455–501, 2001.
- [4450] E. Vijgenboom, J.E. Busch, and G.W. Canters. In vitro studies disprove the obligatory role of azurin in denitrification in *Pseudomonas aeruginosa* and show that azu expression is under the control of RpoS and ANR. *Microbiology*, 143:2853–2863, 1997.
- [4451] C. Vilcheze, H.R. Morbidoni, T.R. Weisbrod, H. Iwamoto, M. Kuo, J.C., Jacobs Sacchettini, , and Jr. Inactivation of the *inhA*-encoded fatty acid synthase II (FASII) enoyl-acyl carrier protein reductase induces accumulation of the FASI end products and cell lysis of *Mycobacterium smegmatis*. *J. Bacteriol.*, 182:4059–4067, 2000.
- [4452] C.A. Vilee and J.M. Spencer. Some properties of the pyridine nucleotide-specific 17 β -hydroxy steroid dehydrogenase of guinea pig liver. *J. Biol. Chem.*, 235:3615–3619, 1960.
- [4453] A. Virion, F. Courtin, D. Deme, J.L. Michot, J. Kaniewski, and J. Pommier. Spectral characteristics and catalytic properties of thyroid peroxidase-H₂O₂ compounds in the iodination and coupling reactions. *Arch. Biochem. Biophys.*, 242:41–47, 1985.
- [4454] T. Vitali, E. Maffioli, G. Tedeschi, and M.A. Vanoni. Properties and catalytic activities of MICAL1, the flavoenzyme involved in cytoskeleton dynamics, and modulation by its CH, LIM and C-terminal domains. *Arch. Biochem. Biophys.*, 593:24–37, 2016.

- [4455] J. Vlasits, C. Jakopitsch, M. Bernroitner, M. Zamocky, P.G. Furtmuller, and C. Obinger. Mechanisms of catalase activity of heme peroxidases. *Arch. Biochem. Biophys.*, 500:74–81, 2010.
- [4456] J. Vockley, W. A. Mohsen al, B. Binzak, J. Willard, and A. Fauq. Mammalian branched-chain acyl-CoA dehydrogenases: molecular cloning and characterization of recombinant enzymes. *Methods Enzymol.*, 324:241–258, 2000.
- [4457] M. Vogel, M. Lawson, W. Sippl, U. Conrad, and W. Roos. Structure and mechanism of sanguinarine reductase, an enzyme of alkaloid detoxification. *J. Biol. Chem.*, 285:18397–18406, 2010.
- [4458] K. Vogl and D.A. Bryant. Biosynthesis of the biomarker okenone: χ -ring formation. *Geobiology*, 10:205–215, 2012.
- [4459] R.N. Vogt, D.J. Steenkamp, R. Zheng, and J.S. Blanchard. The metabolism of nitrosothiols in the *Mycobacteria*: identification and characterization of S-nitrosomycothioliol reductase. *Biochem. J.*, 374:657–666, 2003.
- [4460] A. Volbeda, M.H. Charon, C. Piras, E.C. Hatchikian, M. Frey, and J.C. Fontecillacamps. Crystal-structure of the nickel-iron hydrogenase from *Desulfovibrio gigas*. *Nature*, 373:580–587, 1995.
- [4461] J. Volc, E. Kubátová, G. Daniel, P. Sedmera, and D. Haltrich. Screening of basidiomycete fungi for the quinone-dependent sugar C-2/C-3 oxidoreductase, pyranose dehydrogenase, and properties of the enzyme from *Macrolepiota rhacodes*. *Arch. Microbiol.*, 176:178–186, 2001.
- [4462] J. Volc, E. Kubátová, D. Wood, and G. Daniel. Pyranose 2-dehydrogenase, a novel sugar oxidoreductase from the basidiomycete fungus *Agaricus bisporus*. *Arch. Microbiol.*, 167:119–125, 1997.
- [4463] J. Volc, P. Sedmera, P. Halada, G. Daniel, V. P#345;ikyrllová, and D. Haltrich. C-3 oxidation of non-reducing sugars by a fungal pyranose dehydrogenase: spectral characterization. *J. Mol. Catal., B Enzym.*, 17:91–100, 2002.
- [4464] J. Volc, P. Sedmera, P. Halada, V. P#345;ikyrllová, and G. Daniel. C-2 and C-3 oxidation of D-Glc, and C-2 oxidation of D-Gal by pyranose dehydrogenase from *Agaricus bisporus*. *Carbohydr. Res.*, 310:151–156, 1998.
- [4465] J. Volc, P. Sedmera, P. Halada, V. P#345;ikyrllová, and D. Haltrich. Double oxidation of D-xylose to D-glycero-pentos-2,3-diulose (2,3-diketo-D-xylose) by pyranose dehydrogenase from the mushroom *Agaricus bisporus*. *Carbohydr. Res.*, 329:219–225, 2000.
- [4466] W.A. Volk and J.L. Larsen. β -Keto-L-gulonic acid as an intermediate in the bacterial metabolism of ascorbic acid. *J. Biol. Chem.*, 237:2454–2457, 1962.
- [4467] G. Volkers, G.J. Palm, M.S. Weiss, G.D. Wright, and W. Hinrichs. Structural basis for a new tetracycline resistance mechanism relying on the TetX monooxygenase. *FEBS Lett.*, 585:1061–1066, 2011.
- [4468] S. von Horsten, M.L. Lippert, Y. Geisselbrecht, K. Schuhle, I. Schall, L.O. Essen, and J. Heider. Inactive pseudoenzyme subunits in heterotetrameric BbsCD, a novel short-chain alcohol dehydrogenase involved in anaerobic toluene degradation. *FEBS J.*, 2021.
- [4469] G. von Schumann, S. Gao, and J. Stöckigt. Vomilenine reductase - a novel enzyme catalyzing a crucial step in the biosynthesis of the therapeutically applied antiarrhythmic alkaloid ajmaline. *J. Bioorg. Med. Chem.*, 10:1913–1918, 2002.
- [4470] J.P. von Wartburg and B. Wermoth. Aldehyde reductase. In W.B. Jakoby, editor, *Enzymatic Basis of Detoxication*, volume 1, pages 249–260. Academic Press, New York, 1980.
- [4471] D. von Wettstein, S. Gough, and C.G. Kannangara. Chlorophyll biosynthesis. *Plant Cell*, 7:1039–1057, 1995.
- [4472] J. Vonck, N. Arfman, G.E. De Vries, J. Van Beeumen, E.F. Van Bruggen, and L. Dijkhuizen. Electron microscopic analysis and biochemical characterization of a novel methanol dehydrogenase from the thermotolerant *Bacillus* sp. C1. *J. Biol. Chem.*, 266:3949–3954, 1991.
- [4473] J.E. Vorhaben, J.F. Scott, D.D. Smith, and J.W. Campbell. Mannitol oxidase: partial purification and characterisation of the membrane-bound enzyme from the snail *Helix aspersa*. *Int. J. Biochem.*, 18:337–344, 1986.
- [4474] J.A. Vorholt and R.K. Thauer. The active species of 'CO₂' utilized by formylmethanofuran dehydrogenase from methanogenic Archaea. *Eur. J. Biochem.*, 248:919–924, 1997.

- [4475] S. Vouquier, J. Mary, and B. Friguet. Subcellular localization of methionine sulphoxide reductase A (MsrA): evidence for mitochondrial and cytosolic isoforms in rat liver cells. *Biochem. J.*, 373:531–537, 2003.
- [4476] J.A. Vranka, L.Y. Sakai, and H.P. Bachinger. Prolyl 3-hydroxylase 1, enzyme characterization and identification of a novel family of enzymes. *J. Biol. Chem.*, 279:23615–23621, 2004.
- [4477] A. Vrielink. Cholesterol oxidase: structure and function. *Subcell. Biochem.*, 51:137–158, 2010.
- [4478] J.M. Vrtis, A.K. White, W.W. Metcalf, and W.A. van der Donk. Phosphite dehydrogenase: An unusual phosphoryl transfer reaction. *J. Am. Chem. Soc.*, 123:2672–2673, 2001.
- [4479] V.V. Vu, W.T. Beeson, E.A. Span, E.R. Farquhar, and M.A. Marletta. A family of starch-active polysaccharide monooxygenases. *Proc. Natl. Acad. Sci. USA*, 111:13822–13827, 2014.
- [4480] S. Vujcic, P. Liang, P. Diegelman, D.L. Kramer, and C.W. Porter. Genomic identification and biochemical characterization of the mammalian polyamine oxidase involved in polyamine back-conversion. *Biochem. J.*, 370:19–28, 2003.
- [4481] H. Wada, Z. Gombos, and N. Murata. Enhancement of chilling tolerance of a cyanobacterium by genetic manipulation of fatty acid desaturation. *Nature*, 347:200–203, 1990.
- [4482] H. Wada, H. Schmidt, E. Heinz, and N. Murata. *In vitro* ferredoxin-dependent desaturation of fatty acids in cyanobacterial thylakoid membranes. *J. Bacteriol.*, 175:544–547, 1993.
- [4483] H. Wada and E.E. Snell. The enzymatic oxidation of pyridoxine and pyridoxamine phosphates. *J. Biol. Chem.*, 236:2089–2095, 1961.
- [4484] Y. Wada, S. Iwai, Y. Tamura, T. Ando, T. Shinoda, K. Arai, and H. Taguchi. A new family of D-2-hydroxyacid dehydrogenases that comprises D-mandelate dehydrogenases and 2-ketopantoate reductases. *Biosci. Biotechnol. Biochem.*, 72:1087–1094, 2008.
- [4485] R. Waditee, Y. Tanaka, K. Aoki, T. Hibino, H. Jikuya, J. Takano, T. Takabe, and T. Takabe. Isolation and functional characterization of *N*-methyltransferases that catalyze betaine synthesis from glycine in a halotolerant photosynthetic organism *Aphanothece halophytica*. *J. Biol. Chem.*, 278:4932–4942, 2003.
- [4486] A.F.V. Wagner, M. Frey, F.A. Neugebauer, W. Schäfer, and J. Knappe. The free radical in pyruvate formate-lyase is located on glycine-734. *Proc. Natl. Acad. Sci. USA*, 89:996–1000, 1992.
- [4487] M. Wagner, D. Sonntag, R. Grimm, A. Eckerskorn Pich, Söhling C., Andreesen B., and J.R. Substrate-specific selenoprotein B of glycine reductase from *Eubacterium acidaminophilum*. *Eur. J. Biochem.*, 260:38–49, 1999.
- [4488] M.A. Wagner, P. Trickey, Z.W. Chen, F.S. Mathews, and M.S. Jorns. Monomeric sarcosine oxidase: 1. Flavin reactivity and active site binding determinants. *Biochemistry*, 39:8813–8824, 2000.
- [4489] T. Wagner, U. Ermler, and S. Shima. The methanogenic CO₂ reducing-and-fixing enzyme is bifunctional and contains 46 [4Fe-4S] clusters. *Science*, 354:114–117, 2016.
- [4490] N. Wajih, S.M. Hutson, and R. Wallin. Disulfide-dependent protein folding is linked to operation of the vitamin K cycle in the endoplasmic reticulum. A protein disulfide isomerase-VKORC1 redox enzyme complex appears to be responsible for vitamin K₁ 2,3-epoxide reduction. *J. Biol. Chem.*, 282:2626–2635, 2007.
- [4491] S.J. Wakil and R. Bressler. Studies on the mechanism of fatty acid synthesis. X. Reduced triphosphopyridine nucleotide-acetoacetyl coenzyme A reductase. *J. Biol. Chem.*, 237:687–693, 1962.
- [4492] S.J. Wakil, D.E. Green, S. Mii, and H.R. Mahler. Studies on the fatty acid oxidizing system of animal tissues. VI. β -Hydroxyacyl coenzyme A dehydrogenase. *J. Biol. Chem.*, 207:631–638, 1954.
- [4493] R.J. Walczak, M.L. Dickens, N.D. Priestley, and W.R. Strohl. Purification, properties, and characterization of recombinant *Streptomyces* sp. strain C5 DoxA, a cytochrome *P*-450 catalyzing multiple steps in doxorubicin biosynthesis. *J. Bacteriol.*, 181:298–304, 1999.
- [4494] A.J. Waldman, Y. Pechersky, P. Wang, J.X. Wang, and E.P. Balskus. The cremeomycin biosynthetic gene cluster encodes a pathway for diazo formation. *ChemBiochem*, 16:2172–2175, 2015.

- [4495] C.J. Walker, K.E. Mansfield, I.N. Rezzano, C.M. Hanamoto, K.M. Smith, and P.A. Castelfranco. The magnesium-protoporphyrin IX (oxidative) cyclase system. Studies on the mechanism and specificity of the reaction sequence. *Biochem. J.*, 255:685–692, 1988.
- [4496] D.A. Walker. Physiological studies on acid metabolism. 7. Malic enzyme from *Kalanchoë crenata*: effects of carbon dioxide concentration. *Biochem. J.*, 74:216–223, 1960.
- [4497] G.A. Walker and G.L. Kilgour. Pyridine nucleotide oxidizing enzymes of *Lactobacillus casei*. II. Oxidase and peroxidase. *Arch. Biochem. Biophys.*, 131:534–539, 1965.
- [4498] G.C. Walker and D.J.D. Nicholas. Hydroxylamine reductase from *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta*, 49:361–368, 1961.
- [4499] G.C. Walker and D.J.D. Nicholas. Nitrite reductase from *Pseudomonas aeruginosa*. *Biochim. Biophys. Acta*, 49:350–360, 1961.
- [4500] L. Wall and E.A. Meighen. Subunit structure of the fatty-acid reductase complex from *Photobacterium phosphoreum*. *Biochemistry*, 25:4315–4321, 1986.
- [4501] K.K. Wallace, Z.Y. Bao, H. Dai, R. Digate, G. Schuler, M.K. Speedie, and K.A. Reynolds. Purification of crotonyl-CoA reductase from *Streptomyces collinus* and cloning, sequencing and expression of the corresponding gene in *Escherichia coli*. *Eur. J. Biochem.*, 233:954–962, 1995.
- [4502] D.P. Wallach and V.R. Brown. A novel preparation of human platelet lipoxygenase. Characteristics and inhibition by a variety of phenyl hydrazones and comparisons with other lipoxygenases. *Biochim. Biophys. Acta*, 663:361–372, 1981.
- [4503] J.G. Wallis and J. Browse. The Δ^8 -desaturase of *Euglena gracilis*: an alternate pathway for synthesis of 20-carbon polyunsaturated fatty acids. *Arch. Biochem. Biophys.*, 365:307–316, 1999.
- [4504] C. Wallwey, C. Heddergott, X. Xie, A.A. Brakhage, and S.M. Li. Genome mining reveals the presence of a conserved gene cluster for the biosynthesis of ergot alkaloid precursors in the fungal family Arthrodermataceae. *Microbiology*, 158:1634–1644, 2012.
- [4505] C. Wallwey, M. Matuschek, and S.M. Li. Ergot alkaloid biosynthesis in *Aspergillus fumigatus*: conversion of chanoclavine-I to chanoclavine-I aldehyde catalyzed by a short-chain alcohol dehydrogenase FgaDH. *Arch. Microbiol.*, 192:127–134, 2010.
- [4506] C. Wallwey, M. Matuschek, X.L. Xie, and S.M. Li. Ergot alkaloid biosynthesis in *Aspergillus fumigatus*: Conversion of chanoclavine-I aldehyde to festuclavine by the festuclavine synthase FgaFS in the presence of the old yellow enzyme FgaOx3. *Org. Biomol. Chem.*, 8:3500–3508, 2010.
- [4507] D.A. Walsh and H.J. Sallach. Purification and properties of chicken liver D-3-phosphoglycerate dehydrogenase. *Biochemistry*, 4:1076–1085, 1965.
- [4508] A. Walt and M.L. Kahn. The *fixA* and *fixB* genes are necessary for anaerobic carnitine reduction in *Escherichia coli*. *J. Bacteriol.*, 184:4044–4047, 2002.
- [4509] A.C. Walz, R.A. Demel, B. de Kruijff, and R. Mutzel. Aerobic *sn*-glycerol-3-phosphate dehydrogenase from *Escherichia coli* binds to the cytoplasmic membrane through an amphipathic α -helix. *Biochem. J.*, 365:471–479, 2002.
- [4510] J.T. Wan and J.T. Jarrett. Electron acceptor specificity of ferredoxin (flavodoxin):NADP⁺ oxidoreductase from *Escherichia coli*. *Arch. Biochem. Biophys.*, 406:116–126, 2002.
- [4511] A.Y. Wang, Y.Y. Cronan Chang, and Jr. Role of the tetrameric structure of *Escherichia coli* pyruvate oxidase in enzyme activation and lipid binding. *J. Biol. Chem.*, 266:10959–10966, 1991.
- [4512] C. Wang, W.C. Chang, Y. Guo, H. Huang, S.C. Peck, M.E. Pandelia, G.M. Lin, H.W. Liu, C., Bollinger Krebs, and Jr. Evidence that the fosfomycin-producing epoxidase, HppE, is a non-heme-iron peroxidase. *Science*, 342:991–995, 2013.
- [4513] C.W. Wang and J.C. Liao. Alteration of product specificity of *Rhodobacter sphaeroides* phytoene desaturase by directed evolution. *J. Biol. Chem.*, 276:41161–41164, 2001.

- [4514] G.Q. Wang, J.F. Chen, B. Yi, H.X. Tan, L. Zhang, and W.S. Chen. HPPR encodes the hydroxyphenylpyruvate reductase required for the biosynthesis of hydrophilic phenolic acids in *Salvia miltiorrhiza*. *Chin J Nat Med*, 15:917–927, 2017.
- [4515] J. Wang, X. Chai, U. Eriksson, and J.L. Napoli. Activity of human 11-*cis*-retinol dehydrogenase (Rdh5) with steroids and retinoids and expression of its mRNA in extra-ocular human tissue. *Biochem. J.*, 338:23–27, 1999.
- [4516] J. Wang, Y. Liu, Y. Cai, F. Zhang, G. Xia, and F. Xiang. Cloning and functional analysis of geraniol 10-hydroxylase, a cytochrome P450 from *Swertia mussotii* Franch. *Biosci. Biotechnol. Biochem.*, 74:1583–1590, 2010.
- [4517] J. Wang, X. Wang, Q. Ouyang, W. Liu, J. Shan, H. Tan, X. Li, and G. Chen. *N*-nitrosation mechanism catalyzed by non-heme iron-containing enzyme SznF involving intramolecular oxidative rearrangement. *Inorg. Chem.*, 60:7719–7731, 2021.
- [4518] K.F. Wang, T.A. Dailey, and H.A. Dailey. Expression and characterization of the terminal heme synthetic enzymes from the hyperthermophile *Aquifex aeolicus*. *FEMS Microbiol. Lett.*, 202:115–119, 2001.
- [4519] L. Wang, H. Erlandsen, J. Haavik, P.M. Knappskog, and R.C. Stevens. Three-dimensional structure of human tryptophan hydroxylase and its implications for the biosynthesis of the neurotransmitters serotonin and melatonin. *Biochemistry*, 41:12569–12574, 2002.
- [4520] L. Wang, R.L. White, and L.C. Vining. Biosynthesis of the dideoxysugar component of jadomycin B: genes in the jad cluster of *Streptomyces venezuelae* ISP5230 for L-digitoxose assembly and transfer to the angucycline aglycone. *Microbiology*, 148:1091–1103, 2002.
- [4521] M. Wang, D.L. Roberts, R. Paschke, T.M. Shea, B.S. Masters, and J.J. Kim. Three-dimensional structure of NADPH-cytochrome P450 reductase: prototype for FMN- and FAD-containing enzymes. *Proc. Natl. Acad. Sci. USA*, 94:8411–8416, 1997.
- [4522] P. Wang, G. Bashiri, X. Gao, M.R. Sawaya, and Y. Tang. Uncovering the enzymes that catalyze the final steps in oxytetracycline biosynthesis. *J. Am. Chem. Soc.*, 135:7138–7141, 2013.
- [4523] P. Wang, C.D. Denoya, M.R. Morgenstern, D.D. Skinner, K.K. Wallace, R. Digate, S. Patton, N. Banavali, G. Schuler, M.K. Speedie, and K.A. Reynolds. Cloning and characterization of the gene encoding 1-cyclohexenylcarbonyl coenzyme A reductase from *Streptomyces collinus*. *J. Bacteriol.*, 178:6873–6881, 1996.
- [4524] P. Wang, W. Zhang, J. Zhan, and Y. Tang. Identification of OxyE as an ancillary oxygenase during tetracycline biosynthesis. *ChemBioChem*, 10:1544–1550, 2009.
- [4525] Q. Wang, P. Ding, A.V. Perepelov, Y. Xu, Y. Wang, Y.A. Knirel, L. Wang, and L. Feng. Characterization of the dTDP-D-fucofuranose biosynthetic pathway in *Escherichia coli* O52. *Mol. Microbiol.*, 70:1358–1367, 2008.
- [4526] Q. Wang, M.L. Hillwig, and R.J. Peters. CYP99A3: functional identification of a diterpene oxidase from the momilactone biosynthetic gene cluster in rice. *Plant J.*, 65:87–95, 2011.
- [4527] Q. Wang, M.L. Hillwig, Y. Wu, and R.J. Peters. CYP701A8: a rice *ent*-kaurene oxidase paralog diverted to more specialized diterpenoid metabolism. *Plant Physiol.*, 158:1418–1425, 2012.
- [4528] R. Wang and D.J.D. Nicholas. Some properties of nitrite and hydroxylamine reductases from *Derxia gummosa*. *Phytochemistry*, 25:2463–2469, 1986.
- [4529] S. Wang, H. Huang, J. Kahnt, and R.K. Thauer. *Clostridium acidurici* electron-bifurcating formate dehydrogenase. *Appl. Environ. Microbiol.*, 79:6176–6179, 2013.
- [4530] S. Wang, H. Huang, J. Moll, and R.K. Thauer. NADP⁺ reduction with reduced ferredoxin and NADP⁺ reduction with NADH are coupled via an electron-bifurcating enzyme complex in *Clostridium kluyveri*. *J. Bacteriol.*, 192:5115–5123, 2010.
- [4531] S. Wang, J. Tionson, and M.E. Rasche. Discovery and characterization of the first archaeal dihydromethanopterin reductase, an iron-sulfur flavoprotein from *Methanosarcina mazei*. *J. Bacteriol.*, 196:203–209, 2014.
- [4532] S. Wang, R. Wang, T. Liu, C. Lv, J. Liang, C. Kang, L. Zhou, J. Guo, G. Cui, Y. Zhang, D. Werck-Reichhart, L. Guo, and L. Huang. CYP76B74 catalyzes the 3''-hydroxylation of geranylhydroquinone in shikonin biosynthesis. *Plant Physiol.*, 179:402–414, 2019.

- [4533] S. Wang, Y. Wu, and F.W. Outten. Fur and the novel regulator YqjI control transcription of the ferric reductase gene *yqjH* in *Escherichia coli*. *J. Bacteriol.*, 193:563–574, 2011.
- [4534] T.P. Wang and J.O. Lampen. Metabolism of pyrimidines by a soil bacterium. *J. Biol. Chem.*, 194:775–783, 1952.
- [4535] T.P. Wang and J.O. Lampen. Uracil oxidase and the isolation of barbituric acid from uracil oxidation. *J. Biol. Chem.*, 194:785–791, 1952.
- [4536] X.L. Wang and J.M. Quan. Intermediate-assisted multifunctional catalysis in the conversion of flavin to 5,6-dimethylbenzimidazole by BluB: a density functional theory study. *J. Am. Chem. Soc.*, 133:4079–4091, 2011.
- [4537] Y. Wang, A. Hacker, T. Murray-Stewart, B. Frydman, A. Valasinas, A.V. Fraser, P.M., Casero Woster, , and Jr. Properties of recombinant human *N*¹-acetylpolyamine oxidase (hPAO): potential role in determining drug sensitivity. *Cancer Chemother. Pharmacol.*, 56:83–90, 2005.
- [4538] Y. Wang, T. Murray-Stewart, W. Devereux, A. Hacker, B. Frydman, P.M., Casero Woster, , and Jr. Properties of purified recombinant human polyamine oxidase, PAOh1/SMO. *Biochem. Biophys. Res. Commun.*, 304:605–611, 2003.
- [4539] Y.Z. Wang, Y. Zhou, and G.J. Zylstra. Molecular analysis of isophthalate and terephthalate degradation by *Comamonas testosteroni* YZW-D. *Environ. Health Perspect.* 103, Suppl., 5:9–12, 1995.
- [4540] Z.Q. Wang, R.J. Lawson, M.R. Buddha, C.C. Wei, B.R. Crane, A.W. Munro, and D.J. Stuehr. Bacterial flavodoxins support nitric oxide production by *Bacillus subtilis* nitric-oxide synthase. *J. Biol. Chem.*, 282:2196–2202, 2007.
- [4541] O. Warburg and W. Christian. Isolierung und Krystallisation des Proteins des oxydierenden Gärungsferments. *Biochem. Z.*, 303:40–68, 1939.
- [4542] D.E. Ward, C.J. Donnelly, M.E. Mullendore, J. van der Oost, W.M., Crane de Vos, , and 3rd. The NADH oxidase from *Pyrococcus furiosus*. Implications for the protection of anaerobic hyperthermophiles against oxidative stress. *Eur. J. Biochem.*, 268:5816–5823, 2001.
- [4543] M.J. Wargo, B.S. Szwegold, and D.A. Hogan. Identification of two gene clusters and a transcriptional regulator required for *Pseudomonas aeruginosa* glycine betaine catabolism. *J. Bacteriol.*, 190:2690–2699, 2008.
- [4544] H. Wariishi, L. Akileswaran, and M.H. Gold. Manganese peroxidase from the basidiomycete *Phanerochaete chrysosporium*: spectral characterization of the oxidized states and the catalytic cycle. *Biochemistry*, 27:5365–5370, 1988.
- [4545] H. Wariishi, L. Marquez, H.B. Dunford, and M.H. Gold. Lignin peroxidase compounds II and III. Spectral and kinetic characterization of reactions with peroxides. *J. Biol. Chem.*, 265:11137–11142, 1990.
- [4546] K.L. Warkentin and T.P. Fondy. Isolation and characterization of cytoplasmic L-glycerol-3-phosphate dehydrogenase from rabbit-renal-adipose tissue and its comparison with the skeletal-muscle enzyme. *Eur. J. Biochem.*, 36:97–109, 1973.
- [4547] B.J. Warn-Cramer, L.A. Macrander, and M.T. Abbott. Markedly different ascorbate dependencies of the sequential α -ketoglutarate dioxygenase reactions catalyzed by an essentially homogeneous thymine 7-hydroxylase from *Rhodotorula glutinis*. *J. Biol. Chem.*, 258:10551–10557, 1983.
- [4548] H.M. Warneck and H.U. Seitz. 3β -Hydroxysteroid oxidoreductase in suspension cultures of *Digitalis lanata* EHRH. *Z. Naturforsch. C: Biosci.*, 45:963–972, 1990.
- [4549] C.K. Warner, D.T. Watts, and V. Finnerty. Molybdenum hydroxylases in *Drosophila*. I. Preliminary studies of pyridoxal oxidase. *Mol. Gen. Genet.*, 180:449–453, 1980.
- [4550] M.J. Warren, E. Raux, H.L. Schubert, and J.C. Escalante-Semerena. The biosynthesis of adenosylcobalamin (vitamin B₁₂). *Nat. Prod. Rep.*, 19:390–412, 2002.
- [4551] F. Watanabe, Y. Oki, Y. Nakano, and S. Kitaoka. Occurrence and characterization of cyanocobalamin reductase (NADPH; CN-eliminating) involved in decyanation of cyanocobalamin in *Euglena gracilis*. *J. Nutr. Sci. Vitaminol.*, 34:1–10, 1988.
- [4552] H. Watanabe and J.W. Hastings. Specificities and properties of three reduced pyridine nucleotide-flavin mononucleotide reductases coupling to bacterial luciferase. *Mol. Cell. Biochem.*, 44:181–187, 1982.

- [4553] K. Watanabe, T. Shimizu, and O. Hayaishi. Enzymatic conversion of prostaglandin-D₂ to prostaglandin-F_{2α} in the rat lung. *Biochem. Int.*, 2:603–610, 1981.
- [4554] K. Watanabe, T. Shimizu, S. Iguchi, H. Wakatsuka, M. Hayashi, and O. Hayaishi. An NADP-linked prostaglandin D dehydrogenase in swine brain. *J. Biol. Chem.*, 255:1779–1882, 1980.
- [4555] K. Watanabe, R. Yoshida, T. Shimizu, and O. Hayaishi. Enzymatic formation of prostaglandin F_{2α} from prostaglandin H₂ and D₂. Purification and properties of prostaglandin F synthetase from bovine lung. *J. Biol. Chem.*, 260:7035–7041, 1985.
- [4556] S. Watanabe, F. Fukumori, H. Nishiwaki, Y. Sakurai, K. Tajima, and Y. Watanabe. Novel non-phosphorylative pathway of pentose metabolism from bacteria. *Sci. Rep.*, 9:155–155, 2019.
- [4557] S. Watanabe, T. Kodaki, and K. Makino. Cloning, expression, and characterization of bacterial L-arabinose 1-dehydrogenase involved in an alternative pathway of L-arabinose metabolism. *J. Biol. Chem.*, 281:2612–2623, 2006.
- [4558] S. Watanabe and K. Makino. Novel modified version of nonphosphorylated sugar metabolism - an alternative L-rhamnose pathway of *Sphingomonas* sp. *FEBS J.*, 276:1554–1567, 2009.
- [4559] S. Watanabe, M. Saimura, and K. Makino. Eukaryotic and bacterial gene clusters related to an alternative pathway of nonphosphorylated L-rhamnose metabolism. *J. Biol. Chem.*, 283:20372–20382, 2008.
- [4560] S. Watanabe, Y. Tanimoto, S. Yamauchi, Y. Tozawa, S. Sawayama, and Y. Watanabe. Identification and characterization of *trans*-3-hydroxy-L-proline dehydratase and Δ^1 -pyrroline-2-carboxylate reductase involved in *trans*-3-hydroxy-L-proline metabolism of bacteria. *FEBS Open Bio*, 4:240–250, 2014.
- [4561] J.D. Watkins and J. Jarabak. The effect of NaCl intake on 9-ketoprostaglandin reductase activity in the rabbit kidney. *Prostaglandins*, 30:335–349, 1985.
- [4562] K. Watschinger, M.A. Keller, A. Hermetter, G. Golderer, G. Werner-Felmayer, and E.R. Werner. Glyceryl ether monooxygenase resembles aromatic amino acid hydroxylases in metal ion and tetrahydrobiopterin dependence. *Biol. Chem.*, 390:3–10, 2009.
- [4563] C. Vander Wauven, A. Jann, D. Haas, T. Leisinger, and V. Stalon. *N*²-succinylornithine in ornithine catabolism of *Pseudomonas aeruginosa*. *Arch. Microbiol.*, 150:400–404, 1988.
- [4564] C. Vander Wauven and V. Stalon. Occurrence of succinyl derivatives in the catabolism of arginine in *Pseudomonas cepacia*. *J. Bacteriol.*, 164:882–886, 1985.
- [4565] J.M. Weber, J.O. Leung, S.J. Swanson, K.B. Idler, and J.B. McAlpine. An erythromycin derivative produced by targeted gene disruption in *Saccharopolyspora erythraea*. *Science*, 252:114–117, 1991.
- [4566] S.R. Wecksler, S. Stoll, A.T. Iavarone, E.M. Imsand, H. Tran, R.D. Britt, and J.P. Klinman. Interaction of PqqE and PqqD in the pyrroloquinoline quinone (PQQ) biosynthetic pathway links PqqD to the radical SAM superfamily. *Chem. Commun. (Camb.)*, 46:7031–7033, 2010.
- [4567] R.T. Wedding. Malic enzymes of higher plants: characteristics, regulation, and physiological function. *Plant Physiol.*, 90:367–371, 1989.
- [4568] R.T. Wedding and M.K. Black. Physical and kinetic properties and regulation of the NAD malic enzyme purified from leaves of *Crassula argentea*. *Plant Physiol.*, 72:1021–1028, 1983.
- [4569] G. Weeks and S.J. Wakil. Studies on the mechanism of fatty acid synthesis. 18. Preparation and general properties of the enoyl acyl carrier protein reductases from *Escherichia coli*. *J. Biol. Chem.*, 243:1180–1189, 1968.
- [4570] J.G. Vande Weghe and D.W. Ow. A fission yeast gene for mitochondrial sulfide oxidation. *J. Biol. Chem.*, 274:13250–13257, 1999.
- [4571] M.C. Weghoff, J. Bertsch, and V. Muller. A novel mode of lactate metabolism in strictly anaerobic bacteria. *Environ. Microbiol.*, 17:670–677, 2015.

- [4572] J. Wehrfritz, J.P. Carter, S. Spiro, and D.J. Richardson. Hydroxylamine oxidation in heterotrophic nitrate-reducing soil bacteria and purification of a hydroxylamine-cytochrome *c* oxidoreductase from a *Pseudomonas* species. *Arch. Microbiol.*, 166:421–424, 1996.
- [4573] J.M. Wehrfritz, A. Reilly, S. Spiro, and D.J. Richardson. Purification of hydroxylamine oxidase from *Thiosphaera pantotropha*. Identification of electron acceptors that couple heterotrophic nitrification to aerobic denitrification. *FEBS Lett.*, 335:246–250, 1993.
- [4574] Y. Wei, G. Mathies, K. Yokoyama, J. Chen, R.G. Griffin, and J. Stubbe. A chemically competent thiosulfuranyl radical on the *Escherichia coli* class III ribonucleotide reductase. *J. Am. Chem. Soc.*, 136:9001–9013, 2014.
- [4575] H. Weil-Malherbe. The oxidation of *l*(-)- α -hydroxyglutaric acid in animal tissues. *Biochem. J.*, 31:2080–2094, 1937.
- [4576] R. Weimberg and M. Doudoroff. The oxidation of L-arabinose by *Pseudomonas saccharophila*. *J. Biol. Chem.*, 217:607–624, 1955.
- [4577] S. Weinitschke, K. Hollemeyer, B. Kusian, B. Bowien, T.H. Smits, and A.M. Cook. Sulfoacetate is degraded via a novel pathway involving sulfoacetyl-CoA and sulfoacetaldehyde in *Cupriavidus necator* H16. *J. Biol. Chem.*, 285:35249–35254, 2010.
- [4578] D. Weiss, A. Baumert, M. Vogel, and W. Roos. Sanguinarine reductase, a key enzyme of benzophenanthridine detoxification. *Plant Cell Environ.*, 29:291–302, 2006.
- [4579] H. Weissbach, L. Resnick, and N. Brot. Methionine sulfoxide reductases: history and cellular role in protecting against oxidative damage. *Biochim. Biophys. Acta*, 1703:203–212, 2005.
- [4580] R. Welle and H. Grisebach. Induction of phytoalexin synthesis in soybean: enzymatic cyclization of prenylated pterocarpans to glyceollin isomers. *Arch. Biochem. Biophys.*, 263:191–198, 1988.
- [4581] C.L. Wellington and J.T. Beatty. Promoter mapping and nucleotide sequence of the *bchC* bacteriochlorophyll biosynthesis gene from *Rhodobacter capsulatus*. *Gene*, 83:251–261, 1989.
- [4582] F. Wellmann, M. Griesser, W. Schwab, S. Martens, W. Eisenreich, U. Matern, and R. Lukačín. Anthocyanidin synthase from *Gerbera hybrida* catalyzes the conversion of (+)-catechin to cyanidin and a novel procyanidin. *FEBS Lett.*, 580:1642–1648, 2006.
- [4583] F. Wellmann, R. Lukačín, T. Moriguchi, L. Britsch, E. Schiltz, and U. Matern. Functional expression and mutational analysis of flavonol synthase from *Citrus unshiu*. *Eur. J. Biochem.*, 269:4134–4142, 2002.
- [4584] F. Wellmann, U. Matern, and R. Lukačín. Significance of C-terminal sequence elements for Petunia flavanone 3 β -hydroxylase activity. *FEBS Lett.*, 561:149–154, 2004.
- [4585] D. Wellner and A. Meister. Crystalline L-amino acid oxidase of *Crotalus adamanteus*. *J. Biol. Chem.*, 235:2013–2018, 1960.
- [4586] C. Welte and U. Deppenmeier. Re-evaluation of the function of the F₄₂₀ dehydrogenase in electron transport of *Methanosarcina mazei*. *FEBS J.*, 278:1277–1287, 2011.
- [4587] H. Wengenmayer, J. Ebel, and H. Grisebach. Enzymic synthesis of lignin precursors. Purification and properties of a cinnamoyl-CoA: NADPH reductase from cell suspension cultures of soybean (*Glycinemax*). *Eur. J. Biochem.*, 65:529–536, 1976.
- [4588] M.J. Van Der Werf. Purification and characterization of a Baeyer-Villiger mono-oxygenase from *Rhodococcus erythropolis* DCL14 involved in three different monocyclic monoterpene degradation pathways. *Biochem. J.*, 347:693–701, 2000.
- [4589] B. Wermuth. Purification and properties of an NADPH-dependent carbonyl reductase from human brain. Relationship to prostaglandin 9-ketoreductase and xenobiotic ketone reductase. *J. Biol. Chem.*, 256:1206–1213, 1981.
- [4590] E.R. Werner, A. Hermetter, H. Prast, G. Golderer, and G. Werner-Felmayer. Widespread occurrence of glyceryl ether monooxygenase activity in rat tissues detected by a novel assay. *J. Lipid Res.*, 48:1422–1427, 2007.

- [4591] E.R. Werner, M. Schmid, G. Werner-Felmayer, B. Mayer, and H. Wachter. Synthesis and characterization of ³H-labelled tetrahydrobiopterin. *Biochem. J.*, 304:189–193, 1994.
- [4592] J. Wesche, E. Hammer, D. Becher, G. Burchhardt, and F. Schauer. The *bphC* gene-encoded 2,3-dihydroxybiphenyl-1,2-dioxygenase is involved in complete degradation of dibenzofuran by the biphenyl-degrading bacterium *Ralstonia* sp. SBUG 290. *J. Appl. Microbiol.*, 98:635–645, 2005.
- [4593] T.P. West. Isolation and characterization of an *Escherichia coli* B mutant strain defective in uracil catabolism. *Can. J. Microbiol.*, 44:1106–1109, 1998.
- [4594] T.P. West. Pyrimidine base catabolism in *Pseudomonas putida* biotype B. *Antonie Van Leeuwenhoek*, 80:163–167, 2001.
- [4595] A. Westendorf, D. Benndorf, R.H. Muller, and W. Babel. The two enantiospecific dichlorprop/ α -ketoglutarate-dioxygenases from *Delftia acidovorans* MC1 – protein and sequence data of RdpA and SdpA. *Microbiol. Res.*, 157:317–322, 2002.
- [4596] E.L. Westman, D.J. McNally, A. Charchoglyan, D. Brewer, R.A. Field, and J.S. Lam. Characterization of WbpB, WbpE, and WbpD and reconstitution of a pathway for the biosynthesis of UDP-2,3-diacetamido-2,3-dideoxy-D-mannuronic acid in *Pseudomonas aeruginosa*. *J. Biol. Chem.*, 284:11854–11862, 2009.
- [4597] A.L. Wheeler, R.M. Long, R.E. Ketchum, C.D. Rithner, R.M. Williams, and R. Croteau. Taxol biosynthesis: differential transformations of taxadien-5 α -ol and its acetate ester by cytochrome *P*₄₅₀ hydroxylases from *Taxus* suspension cells. *Arch. Biochem. Biophys.*, 390:265–78, 2001.
- [4598] G.L. Wheeler, M.A. Jones, and N. Smirnoff. The biosynthetic pathway of vitamin C in higher plants. *Nature*, 393:365–369, 1998.
- [4599] M.H. Wheeler and G.A. Greenblatt. The inhibition of melanin biosynthetic reactions in *Pyricularia oryzae* by compounds that prevent rice blast disease. *Exp. Mycol.*, 12:151–160, 1988.
- [4600] J.R. Whetstine, A. Nottke, F. Lan, M. Huarte, S. Smolikov, Z. Chen, E. Spooner, E. Li, G. Zhang, M. Colaiacovo, and Y. Shi. Reversal of histone lysine trimethylation by the JMJD2 family of histone demethylases. *Cell*, 125:467–481, 2006.
- [4601] E.H. White, F. McCapra, G.F. Field, and W.D. McElroy. The structure and synthesis of firefly luciferin. *J. Am. Chem. Soc.*, 83:2402–2403, 1961.
- [4602] E.H. White, E. Rapaport, T.A. Hopkins, and H.H. Seliger. Chemi- and bioluminescence of firefly luciferin. *J. Am. Chem. Soc.*, 91:2178–2180, 1969.
- [4603] H. White, G. Strobl, R. Feicht, and H. Simon. Carboxylic acid reductase: a new tungsten enzyme catalyses the reduction of non-activated carboxylic acids to aldehydes. *Eur. J. Biochem.*, 184:89–96, 1989.
- [4604] R.H. White. L-Aspartate semialdehyde and a 6-deoxy-5-ketohexose 1-phosphate are the precursors to the aromatic amino acids in *Methanocaldococcus jannaschii*. *Biochemistry*, 43:7618–7627, 2004.
- [4605] G.M. Whited and D.T. Gibson. Separation and partial characterization of the enzymes of the toluene-4-monooxygenase catabolic pathway in *Pseudomonas mendocina* KR1. *J. Bacteriol.*, 173:3017–3020, 1991.
- [4606] G.M. Whited and D.T. Gibson. Toluene-4-monooxygenase, a three-component enzyme system that catalyzes the oxidation of toluene to *p*-cresol in *Pseudomonas mendocina* KR1. *J. Bacteriol.*, 173:3010–3016, 1991.
- [4607] G.M. Whited, W.R. McCombie, L.D. Kwart, and D.T. Gibson. Identification of *cis*-diols as intermediates in the oxidation of aromatic acids by a strain of *Pseudomonas putida* that contains a TOL plasmid. *J. Bacteriol.*, 166:1028–1039, 1986.
- [4608] G.C. Whiting and R.A. Coggins. A new nicotinamide-adenine dinucleotide-dependent hydroaromatic dehydrogenase of *Lactobacillus plantarum* and its role in formation of (-)-*t*-3,*t*-4-dihydroxycyclohexane-*c*-1-carboxylate. *Biochem. J.*, 141:35–42, 1974.
- [4609] D.S. Whitlon, J.A. Sadowski, and J.W. Suttie. Mechanism of coumarin action: significance of vitamin K epoxide reductase inhibition. *Biochemistry*, 17:1371–1377, 1978.
- [4610] M. Whittaker, D. Bergmann, D. Arciero, and A.B. Hooper. Electron transfer during the oxidation of ammonia by the chemolithotrophic bacterium *Nitrosomonas europaea*. *Biochim. Biophys. Acta*, 1459:346–355, 2000.

- [4611] M.M. Whittaker, P.J. Kersten, N. Nakamura, J. Sanders-Loehr, E.S. Schweizer, and J.W. Whittaker. Glyoxal oxidase from *Phanerochaete chrysosporium* is a new radical-copper oxidase. *J. Biol. Chem.*, 271:681–687, 1996.
- [4612] J.T. Whitteck, P. Malova, S.C. Peck, R.M. Cicchillo, F. Hammerschmidt, and W.A. van der Donk. On the stereochemistry of 2-hydroxyethylphosphonate dioxygenase. *J. Am. Chem. Soc.*, 133:4236–4239, 2011.
- [4613] E. Whittle, E.B. Cahoon, S. Subrahmanyam, and J. Shanklin. A multifunctional acyl-acyl carrier protein desaturase from *Hedera helix* L. (English ivy) can synthesize 16- and 18-carbon monoene and diene products. *J. Biol. Chem.*, 280:28169–28176, 2005.
- [4614] D.J. Wichelecki, J.A. Vendiola, A.M. Jones, N. Al-Obaidi, S.C. Almo, and J.A. Gerlt. Investigating the physiological roles of low-efficiency D-mannonate and D-gluconate dehydratases in the enolase superfamily: pathways for the catabolism of L-gulonate and L-idonate. *Biochemistry*, 53:5692–5699, 2014.
- [4615] D.J. Wichelecki, M.W. Vetting, L. Chou, N. Al-Obaidi, J.T. Bouvier, S.C. Almo, and J.A. Gerlt. ATP-binding cassette (ABC) transport system solute-binding protein-guided identification of novel D-altritol and galactitol catabolic pathways in *Agrobacterium tumefaciens* C58. *J. Biol. Chem.*, 290:28963–28976, 2015.
- [4616] D.K. Wicht. The reduced flavin-dependent monooxygenase SfnG converts dimethylsulfone to methanesulfinate. *Arch. Biochem. Biophys.*, 604:159–166, 2016.
- [4617] B.M. Wickwire, C. Wagner, and H.P. Broquist. Pipecolic acid biosynthesis in *Rhizoctonia leguminicola*. II. Saccharopine oxidase: a unique flavin enzyme involved in pipecolic acid biosynthesis. *J. Biol. Chem.*, 265:14748–14753, 1990.
- [4618] P.F. Widboom, E.N. Fielding, Y. Liu, and S.D. Bruner. Structural basis for cofactor-independent dioxygenation in vancomycin biosynthesis. *Nature*, 447:342–345, 2007.
- [4619] E. Widemann, B. Grausem, H. Renault, E. Pineau, C. Heinrich, R. Lugan, P. Ullmann, L. Miesch, Y. Aubert, M. Miesch, T. Heitz, and F. Pinot. Sequential oxidation of jasmonoyl-phenylalanine and jasmonoyl-isoleucine by multiple cytochrome P450 of the CYP94 family through newly identified aldehyde intermediates. *Phytochemistry*, 117:388–399, 2015.
- [4620] G.J. Wiederrecht and G.M. Brown. Purification and properties of the enzymes from *Drosophila melanogaster* that catalyze the conversion of dihydroneopterin triphosphate to the pyrimidodiazepine precursor of the drosopterins. *J. Biol. Chem.*, 259:14121–14127, 1984.
- [4621] B. Wieland, C. Feil, E. Gloria-Maercker, G. Thumm, M. Lechner, J.M. Bravo, K. Poralla, and F. Gotz. Genetic and biochemical analyses of the biosynthesis of the yellow carotenoid 4,4'-diaponeurosporene of *Staphylococcus aureus*. *J. Bacteriol.*, 176:7719–7726, 1994.
- [4622] G.J. Wigmore and D.W. Ribbons. *p*-Cymene pathway in *Pseudomonas putida*: selective enrichment of defective mutants by using halogenated substrate analogs. *J. Bacteriol.*, 143:816–824, 1980.
- [4623] K. Wikvall. Purification and properties of a 3β -hydroxy- Δ^5 -C₂₇-steroid oxidoreductase from rabbit liver microsomes. *J. Biol. Chem.*, 256:3376–3380, 1981.
- [4624] K. Wikvall. Hydroxylations in biosynthesis of bile acids. Isolation of a cytochrome *P*-450 from rabbit liver mitochondria catalyzing 26-hydroxylation of C₂₇-steroids. *J. Biol. Chem.*, 259:3800–3804, 1984.
- [4625] M.C. Wilce, D.M. Dooley, H.C. Freeman, J.M. Guss, H. Matsunami, W.S. McIntire, C.E. Ruggiero, K. Tanizawa, and H. Yamaguchi. Crystal structures of the copper-containing amine oxidase from *Arthrobacter globiformis* in the holo and apo forms: implications for the biogenesis of topaquinone. *Biochemistry*, 36:16116–16133, 1997.
- [4626] J. Wilcoxon, B. Zhang, and R. Hille. Reaction of the molybdenum- and copper-containing carbon monoxide dehydrogenase from *Oligotropha carboxidovorans* with quinones. *Biochemistry*, 50:1910–1916, 2011.
- [4627] D.R. Wilken, H.L. King, Dyar Jr., and R.E. Ketopantoic acid and ketopantoyl lactone reductases. Stereospecificity of transfer of hydrogen from reduced nicotinamide adenine dinucleotide phosphate. *J. Biol. Chem.*, 250:2311–2314, 1975.
- [4628] S.E. Wilkins, M.S. Islam, J.M. Gannon, S. Markolovic, R.J. Hopkinson, W. Ge, C.J. Schofield, and R. Chowdhury. JMJD5 is a human arginyl C-3 hydroxylase. *Nat. Commun.*, 9:1180–1180, 2018.

- [4629] B.A. Williams, C. Elliot, L. Burri, Y. Kido, K. Kita, A.L. Moore, and P.J. Keeling. A broad distribution of the alternative oxidase in microsporidian parasites. *PLoS Pathog.*, 6:e1000761–e1000761, 2010.
- [4630] C.H. Williams, Kamin Jr., and H. Microsomal triphosphopyridine nucleotide-cytochrome *c* reductase in liver. *J. Biol. Chem.*, 237:587–595, 1962.
- [4631] D.R. Williams, P.W. Trudgill, and D.G. Taylor. Metabolism of 1,8-cineole by *Rhodococcus* species: ring cleavage reactions. *J. Gen. Microbiol.*, 135:1957–1967, 1989.
- [4632] F.R. Williams and L.P. Hager. Crystalline flavin pyruvate oxidase from *Escherichia coli*. I. Isolation and properties of the flavoprotein. *Arch. Biochem. Biophys.*, 116:168–176, 1966.
- [4633] G.J. Williams, S.D. Breazeale, C.R.H. Raetz, and J.H. Naismith. Structure and function of both domains of ArnA, a dual function decarboxylase and a formyltransferase, involved in 4-amino-4-deoxy-L-arabinose biosynthesis. *J. Biol. Chem.*, 280:23000–23008, 2005.
- [4634] P.A. Williams, L. Coates, F. Mohammed, R. Gill, P.T. Erskine, A. Coker, S.P. Wood, C. Anthony, and J.B. Cooper. The atomic resolution structure of methanol dehydrogenase from *Methylobacterium extorquens*. *Acta Crystallogr. D Biol. Crystallogr.*, 61:75–79, 2005.
- [4635] P.A. Williams, V. Fulop, Y.C. Leung, C. Chan, J.W.B. Moir, G. Howlett, S.J. Ferguson, S.E. Radford, and J. Hajdu. Pseudospecific docking surfaces on electron transfer proteins as illustrated by pseudoazurin, cytochrome *c*-550 and cytochrome *cd1* nitrite reductase. *Nat. Struct. Biol.*, 2:975–982, 1995.
- [4636] S.J. Williams, R.H. Senaratne, J.D. Mougous, L.W. Riley, and C.R. Bertozzi. 5'-Adenosinephosphosulfate lies at a metabolic branchpoint in mycobacteria. *J. Biol. Chem.*, 277:32606–32615, 2002.
- [4637] J.D. Williamson, J.M.H. Stoop, M.O. Massel, M.A. Conkling, and D.M. Pharr. Sequence analysis of a mannitol dehydrogenase cDNA from plants reveals a function for the pathogenesis-related protein ELI3. *Proc. Natl. Acad. Sci. USA*, 92:7148–7152, 1995.
- [4638] A.K. Willingham and J.T. Matschiner. Changes in phylloquinone epoxidase activity related to prothrombin synthesis and microsomal clotting activity in the rat. *Biochem. J.*, 140:435–441, 1974.
- [4639] R.C. Wilmouth, J.J. Turnbull, R.W. Welford, I.J. Clifton, A.G. Prescott, and C.J. Schofield. Structure and mechanism of anthocyanidin synthase from *Arabidopsis thaliana*. *Structure*, 10:93–103, 2002.
- [4640] N.W. Winkler and A. Markovitz. Guanosine diphosphate-4-keto-D-rhamnose reductase. A non-stereoselective enzyme. *J. Biol. Chem.*, 246:5868–5876, 1971.
- [4641] T. Winzer, M. Kern, A.J. King, T.R. Larson, R.I. Teodor, S.L. Donninger, Y. Li, A.A. Dowle, J. Cartwright, R. Bates, D. Ashford, J. Thomas, C. Walker, T.A. Bowser, and I.A. Graham. Morphinan biosynthesis in opium poppy requires a P450-oxidoreductase fusion protein. *Science*, 349:309–312, 2015.
- [4642] S. Wischgoll, U. Demmer, E. Warkentin, R. Gunther, M. Boll, and U. Ermler. Structural basis for promoting and preventing decarboxylation in glutaryl-coenzyme A dehydrogenases. *Biochemistry*, 49:5350–5357, 2010.
- [4643] S. Wischgoll, M. Taubert, F. Peters, N. Jehmlich, M. von Bergen, and M. Boll. Decarboxylating and nondecarboxylating glutaryl-coenzyme A dehydrogenases in the aromatic metabolism of obligately anaerobic bacteria. *J. Bacteriol.*, 191:4401–4409, 2009.
- [4644] F. Wissing. Cyanide production from glycine by a homogenate from a *Pseudomonas* species. *J. Bacteriol.*, 121:695–699, 1975.
- [4645] J.H. Wissler. D-Xylose:NADP oxidoreductase of arterial vessels and eye lens: a new enzyme and a final link in ATP-independent cycling of reducing equivalents in aldose-polyol-ketose interconversion. *Hoppe-Seyler's Z. Physiol. Chem.*, 358:1300–1301, 1977.
- [4646] J.H. Wissler. Direct spectrophotometric and specific quantitative determination of free and bound D-xylose by analytical application of a new enzyme, D-xylose:NADP-oxidoreductase. *Fresenius' Z. Anal. Chem.*, 290:179–180, 1978.
- [4647] M. Witschel, S. Nagel, and T. Egli. Identification and characterization of the two-enzyme system catalyzing oxidation of EDTA in the EDTA-degrading bacterial strain DSM 9103. *J. Bacteriol.*, 179:6937–6943, 1997.

- [4648] U. Wittstock and B.A. Halkier. Cytochrome P_{450} CYP79A2 from *Arabidopsis thaliana* L. Catalyzes the conversion of L-phenylalanine to phenylacetaldoxime in the biosynthesis of benzylglucosinolate. *J. Biol. Chem.*, 275:14659–14666, 2000.
- [4649] A.J. Wittwer and C. Wagner. Identification of the folate-binding proteins of rat liver mitochondria as dimethylglycine dehydrogenase and sarcosine dehydrogenase. Flavoprotein nature and enzymatic properties of the purified proteins. *J. Biol. Chem.*, 256:4109–4115, 1981.
- [4650] A.J. Wittwer and C. Wagner. Identification of the folate-binding proteins of rat liver mitochondria as dimethylglycine dehydrogenase and sarcosine dehydrogenase. Purification and folate-binding characteristics. *J. Biol. Chem.*, 256:4102–4108, 1981.
- [4651] G. Wohlfarth, G. Geerligs, and G. Diekert. Purification and properties of a NADH-dependent 5,10-methylenetetrahydrofolate reductase from *Peptostreptococcus productus*. *Eur. J. Biochem.*, 192:411–417, 1990.
- [4652] Z. Wojdyla and T. Borowski. On how the binding cavity of AsqJ dioxygenase controls the desaturation reaction regioselectivity: a QM/MM study. *J. Biol. Inorg. Chem.*, 23:795–808, 2018.
- [4653] J.B. Wolfe and N.O. Kaplan. Hexose phosphate and hexose reductase. A. D-Mannitol-1-phosphate dehydrogenase from *E. coli*. *Methods Enzymol.*, 1:346–348, 1955.
- [4654] J.B. Wolfe and N.O. Kaplan. D-Mannitol 1-phosphate dehydrogenase from *Escherichia coli*. *J. Biol. Chem.*, 218:849–869, 1956.
- [4655] R.G. Wolfe and J.B. Nielsens. Some molecular and kinetic properties of heart malic dehydrogenase. *J. Biol. Chem.*, 221:61–69, 1956.
- [4656] J.B. Wolff, , and N.O. Hexitol metabolism in *Escherichia coli*. *J. Bacteriol.*, 71:557–564, 1956.
- [4657] M. Wolff, M. Seemann, T.S.B. Bui, Y. Frapart, D. Tritsch, A. Garcia Estrabot, M. Rodríguez-Concepción, A. Boronat, A. Marquet, and M. Rohmer. Isoprenoid biosynthesis via the methylerythritol phosphate pathway: the (*E*)-4-hydroxy-3-methylbut-2-enyl diphosphate reductase (LytB/IspH) from *Escherichia coli* is a [4Fe-4S] protein. *FEBS Lett.*, 541:115–120, 2003.
- [4658] W.A. Wolken and M.J. van der Werf. Geraniol biotransformation-pathway in spores of *Penicillium digitatum*. *Appl. Microbiol. Biotechnol.*, 57:731–737, 2001.
- [4659] J. Wollam, L. Magomedova, D.B. Magner, Y. Shen, V. Rottiers, D.L. Motola, D.J. Mangelsdorf, C.L. Cummins, and A. Antebi. The Rieske oxygenase DAF-36 functions as a cholesterol 7-desaturase in steroidogenic pathways governing longevity. *Aging Cell*, 10:879–884, 2011.
- [4660] B. Wong, J.S. Murray, M. Castellanos, and K.D. Croen. D-Arabitol metabolism in *Candida albicans*: studies of the biosynthetic pathway and the gene that encodes NAD-dependent D-arabitol dehydrogenase. *J. Bacteriol.*, 175:6314–6320, 1993.
- [4661] P.Y.-K. Wong. Purification and partial characterization of prostaglandin D₂ 11-keto reductase in rabbit liver. *Biochim. Biophys. Acta*, 659:169–178, 1981.
- [4662] P.Y.-K. Wong. Purification of PGD₂ 11-ketoreductase from rabbit liver. *Methods Enzymol.*, 86:117–125, 1982.
- [4663] H.A. Woo, W. Jeong, T.S. Chang, K.J. Park, S.J. Park, J.S. Yang, and S.G. Rhee. Reduction of cysteine sulfinic acid by sulfiredoxin is specific to 2-Cys peroxiredoxins. *J. Biol. Chem.*, 280:3125–3128, 2005.
- [4664] J.L. Wood and D. Cavallini. Enzymic oxidation of cysteamine to hypotaurine in the absence of a cofactor. *Arch. Biochem. Biophys.*, 119:368–372, 1967.
- [4665] T. Wood. Catalysis of pentose phosphate pathway reactions by cytoplasmic fractions from muscle, uterus and liver of the rat, and the presence of a reduced nicotinamide-adenine dinucleotide phosphate-triose phosphate oxidoreductase in rat muscle. *Biochem. J.*, 138:71–76, 1974.
- [4666] W.A. Wood, M.J. McDonough, and L.B. Jacobs. Ribitol and D-arabitol utilization by *Aerobacter aerogenes*. *J. Biol. Chem.*, 236:2190–2195, 1961.

- [4667] Z.A. Wood, E. Schröder, J.R. Harris, and L.B. Poole. Structure, mechanism and regulation of peroxiredoxins. *Trends Biochem. Sci.*, 28:32–40, 2003.
- [4668] R. Woodyer, M. Simurdiak, W.A. van der Donk, and H. Zhao. Heterologous expression, purification, and characterization of a highly active xylose reductase from *Neurospora crassa*. *Appl. Environ. Microbiol.*, 71:1642–1647, 2005.
- [4669] B. Worsdorfer, M. Lingaraju, N.H. Yennawar, A.K. Boal, C. Krebs, J.M. Bollinger, Pandelia Jr., and M.E. Organophosphonate-degrading PhnZ reveals an emerging family of HD domain mixed-valent diiron oxygenases. *Proc. Natl. Acad. Sci. USA*, 110:18874–18879, 2013.
- [4670] D.J. Worthington and M.A. Rosemeyer. Glutathione reductase from human erythrocytes. Catalytic properties and aggregation. *Eur. J. Biochem.*, 67:231–238, 1976.
- [4671] W.D. Wosilait. The reduction of vitamin K₁ by an enzyme from dog liver. *J. Biol. Chem.*, 235:1196–1201, 1960.
- [4672] J.W. Wray and R.H. Abeles. A bacterial enzyme that catalyzes formation of carbon monoxide. *J. Biol. Chem.*, 268:21466–21469, 1993.
- [4673] J.W. Wray and R.H. Abeles. The methionine salvage pathway in *Klebsiella pneumoniae* and rat liver. Identification and characterization of two novel dioxygenases. *J. Biol. Chem.*, 270:3147–3153, 1995.
- [4674] J.F. Wu, C.Y. Jiang, B.J. Wang, Y.F. Ma, Z.P. Liu, and S.J. Liu. Novel partial reductive pathway for 4-chloronitrobenzene and nitrobenzene degradation in *Comamonas* sp. strain CNB-1. *Appl. Environ. Microbiol.*, 72:1759–1765, 2006.
- [4675] J.F. Wu, C.W. Sun, C.Y. Jiang, Z.P. Liu, and S.J. Liu. A novel 2-aminophenol 1,6-dioxygenase involved in the degradation of *p*-chloronitrobenzene by *Comamonas* strain CNB-1: purification, properties, genetic cloning and expression in *Escherichia coli*. *Arch. Microbiol.*, 183:1–8, 2005.
- [4676] K. Wu, R. Knox, X.Z. Sun, P. Joseph, A.K. Jaiswal, D. Zhang, P.S. Deng, and S. Chen. Catalytic properties of NAD(P)H:quinone oxidoreductase-2 (NQO₂), a dihydronicotinamide riboside dependent oxidoreductase. *Arch. Biochem. Biophys.*, 347:221–228, 1997.
- [4677] S. Wu, L. Wang, R. Gan, T. Tong, H. Bian, Z. Li, S. Du, Z. Deng, and S. Chen. Signature arsenic detoxification pathways in *Halomonas* sp. strain GFAJ-1. *mBio*, 9, 2018.
- [4678] S.-H. Wu, M.T. McDowell, and J.C. Lagarias. Phycocyanobilin is the natural chromophore precursor of phytochrome from the green alga *Mesotaenium caldariorum*. *J. Biol. Chem.*, 272:25700–25705, 1997.
- [4679] T. Wu, V. Yankovskaya, and W.S. McIntire. Cloning, sequencing, and heterologous expression of the murine peroxisomal flavoprotein, N¹-acetylated polyamine oxidase. *J. Biol. Chem.*, 278:20514–20525, 2003.
- [4680] T.P. Wu, T. Wang, M.G. Seetin, Y. Lai, S. Zhu, K. Lin, Y. Liu, S.D. Byrum, S.G. Mackintosh, M. Zhong, A. Tackett, G. Wang, L.S. Hon, G. Fang, J.A. Swenberg, and A.Z. Xiao. DNA methylation on *N*-adenine in mammalian embryonic stem cells. *Nature*, 532:329–333, 2016.
- [4681] X. Wu and V.M. Monnier. Enzymatic deglycation of proteins. *Arch. Biochem. Biophys.*, 419:16–24, 2003.
- [4682] X.B. Wu, K.Q. Fan, Q.H. Wang, and K.Q. Yang. C-terminus mutations of *Acremonium chrysogenum* deacetoxy/deacetylcephalosporin C synthase with improved activity toward penicillin analogs. *FEMS Microbiol. Lett.*, 246:103–110, 2005.
- [4683] Y. Wu, M.L. Hillwig, Q. Wang, and R.J. Peters. Parsing a multifunctional biosynthetic gene cluster from rice: biochemical characterization of CYP71Z6 & 7. *FEBS Lett.*, 585:3446–3451, 2011.
- [4684] Y. Wu, Q. Wang, M.L. Hillwig, and R.J. Peters. Picking sides: distinct roles for CYP76M6 and CYP76M8 in rice oryzalexin biosynthesis. *Biochem. J.*, 454:209–216, 2013.
- [4685] Z. Wu, D. Lee, J. Joo, J.H. Shin, W. Kang, S. Oh, D.Y. Lee, S.J. Lee, S.S. Yea, H.S. Lee, T. Lee, and K.H. Liu. CYP2J2 and CYP2C19 are the major enzymes responsible for metabolism of albendazole and fenbendazole in human liver microsomes and recombinant P450 assay systems. *Antimicrob. Agents Chemother.*, 57:5448–5456, 2013.
- [4686] Z. Wu, Y. Yang, N. Shaw, S. Bhattacharya, L. Yan, K. West, K. Roth, N. Noy, J. Qin, and J.W. Crabb. Mapping the ligand binding pocket in the cellular retinaldehyde binding protein. *J. Biol. Chem.*, 278:12390–12396, 2003.

- [4687] M. Wust, D.B. Little, M. Schalk, and R. Croteau. Hydroxylation of limonene enantiomers and analogs by recombinant (–)-limonene 3- and 6-hydroxylases from mint (*Mentha*) species: evidence for catalysis within sterically constrained active sites. *Arch. Biochem. Biophys.*, 387:125–136, 2001.
- [4688] K.L. Wüthrich, L. Bovet, P.E. Hunziker, I.S. Donnison, and S. Hörtensteiner. Molecular cloning, functional expression and characterisation of RCC reductase involved in chlorophyll catabolism. *Plant J.*, 21:189–198, 2000.
- [4689] P. Wyk and P. Reeves. Identification and sequence of the gene for abequose synthase, which confers antigenic specificity on group B salmonellae: homology with galactose epimerase. *J. Bacteriol.*, 171:5687–5693, 1989.
- [4690] R.L. Wykle and J.M. Schremmer Lockmiller. The biosynthesis of plasmalogens by rat brain: involvement of the microsomal electron transport system. *Biochim. Biophys. Acta*, 380:291–298, 1975.
- [4691] D. Wyrambik and H. Grisebach. Purification and properties of isoenzymes of cinnamyl-alcohol dehydrogenase from soybean-cell-suspension cultures. *Eur. J. Biochem.*, 59:9–15, 1975.
- [4692] D. Wyrambik and H. Grisebach. Enzymic synthesis of lignin precursors. Further studies on cinnamyl-alcohol dehydrogenase from soybean-cell-suspension cultures. *Eur. J. Biochem.*, 97:503–509, 1979.
- [4693] Z.Q. Xia, M.A. Costa, H.C. Pelissier, L.B. Davin, and N.G. Lewis. Secoisolariciresinol dehydrogenase purification, cloning, and functional expression. Implications for human health protection. *J. Biol. Chem.*, 276:12614–12623, 2001.
- [4694] Z.X. Xia, Y.N. He, W.W. Dai, S.A. White, G.D. Boyd, and F.S. Mathews. Detailed active site configuration of a new crystal form of methanol dehydrogenase from *Methylophilus* W3A1 at 1.9 Å resolution. *Biochemistry*, 38:1214–1220, 1999.
- [4695] Y. Xiang, Z. Zhu, G. Han, H. Lin, L. Xu, and C.D. Chen. JMJD3 is a histone H3K27 demethylase. *Cell Res.*, 17:850–857, 2007.
- [4696] D.Y. Xie, S.B. Sharma, and R.A. Dixon. Anthocyanidin reductases from *Medicago truncatula* and *Arabidopsis thaliana*. *Arch. Biochem. Biophys.*, 422:91–102, 2004.
- [4697] D.Y. Xie, S.B. Sharma, N.L. Paiva, D. Ferreira, and R.A. Dixon. Role of anthocyanidin reductase, encoded by BANYULS in plant flavonoid biosynthesis. *Science*, 299:396–399, 2003.
- [4698] X. Xie, C. Wallwey, M. Matuschek, K. Steinbach, and S.M. Li. Formyl migration product of chanoclavine-I aldehyde in the presence of the old yellow enzyme FgaOx3 from *Aspergillus fumigatus*: a NMR structure elucidation. *Magn. Reson. Chem.*, 49:678–681, 2011.
- [4699] Y. Xin, H. Liu, F. Cui, H. Liu, and L. Xun. Recombinant *Escherichia coli* with sulfide:quinone oxidoreductase and persulfide dioxygenase rapidly oxidises sulfide to sulfite and thiosulfate via a new pathway. *Environ. Microbiol.*, 18:5123–5136, 2016.
- [4700] F. Xing, S.L. Hiley, T.R. Hughes, and E.M. Phizicky. The specificities of four yeast dihydrouridine synthases for cytoplasmic tRNAs. *J. Biol. Chem.*, 279:17850–17860, 2004.
- [4701] F. Xing, M.R. Martzen, and E.M. Phizicky. A conserved family of *Saccharomyces cerevisiae* synthases effects dihydrouridine modification of tRNA. *RNA*, 8:370–381, 2002.
- [4702] M. Xing, Y. Wei, Y. Zhou, J. Zhang, L. Lin, Y. Hu, Hua G., A. N. Nanjaraj Urs, D. Liu, F. Wang, C. Guo, Y. Tong, M. Li, Y. Liu, E.L. Ang, H. Zhao, Z. Yuchi, and Y. Zhang. Radical-mediated C-S bond cleavage in C₂ sulfonate degradation by anaerobic bacteria. *Nat. Commun.*, 10:1609–1609, 2019.
- [4703] C. Xu, K. Liu, W. Tempel, M. Demetriades, W. Aik, C.J. Schofield, and J. Min. Structures of human ALKBH5 demethylase reveal a unique binding mode for specific single-stranded N⁶-methyladenosine RNA demethylation. *J. Biol. Chem.*, 289:17299–17311, 2014.
- [4704] J. Xu, G. Li, P. Wang, H. Velazquez, X. Yao, Y. Li, Y. Wu, A. Peixoto, S. Crowley, and G.V. Desir. Renalase is a novel, soluble monoamine oxidase that regulates cardiac function and blood pressure. *J. Clin. Invest.*, 115:1275–1280, 2005.
- [4705] L. Xu, E.P. Go, J. Finney, H. Moon, M. Lantz, K. Rebecchi, H. Desaire, and M. Mure. Post-translational modifications of recombinant human lysyl oxidase-like 2 (rhLOXL2) secreted from *Drosophila* S2 cells. *J. Biol. Chem.*, 288:5357–5363, 2013.

- [4706] N. Xu, E.G. Ahuja, P. Janning, D.V. Mavrodi, L.S. Thomashow, and W. Blankenfeldt. Trapped intermediates in crystals of the FMN-dependent oxidase PhzG provide insight into the final steps of phenazine biosynthesis. *Acta Crystallogr. D Biol. Crystallogr.*, 69:1403–1413, 2013.
- [4707] Q. Xu, T. Eguchi, I.I. Mathews, C.L. Rife, H.J. Chiu, C.L. Farr, J. Feuerhelm, L. Jaroszewski, H.E. Klock, M.W. Knuth, M.D. Miller, D. Weekes, M.A. Elsliger, A.M. Deacon, A. Godzik, S.A. Lesley, and I.A. Wilson. Insights into substrate specificity of geranylgeranyl reductases revealed by the structure of digeranylgeranyl glycerophospholipid reductase, an essential enzyme in the biosynthesis of archaeal membrane lipids. *J. Mol. Biol.*, 404:403–417, 2010.
- [4708] Y. Xu, M.W. Mortimer, T.S. Fisher, M.L. Kahn, F.J. Brockman, and L. Xun. Cloning, sequencing, and analysis of a gene cluster from *Chelatobacter heintzii* ATCC 29600 encoding nitrilotriacetate monooxygenase and NADH:flavin mononucleotide oxidoreductase. *J. Bacteriol.*, 179:1112–1116, 1997.
- [4709] B. Xue, A.P. Rooney, M. Kajikawa, N. Okada, and W.L. Roelofs. Novel sex pheromone desaturases in the genomes of corn borers generated through gene duplication and retroposon fusion. *Proc. Natl. Acad. Sci. USA*, 104:4467–4472, 2007.
- [4710] Y. Xue, D. Wilson, L. Zhao, Sherman Liu Hw, and D.H. Hydroxylation of macrolactones YC-17 and narbomycin is mediated by the *pikC*-encoded cytochrome P450 in *Streptomyces venezuelae*. *Chem. Biol.*, 5:661–667, 1998.
- [4711] L. Xun, S.M. Belchik, R. Xun, Y. Huang, H. Zhou, E. Sanchez, C. Kang, and P.G. Board. *S*-Glutathionyl-(chloro)hydroquinone reductases: a novel class of glutathione transferases. *Biochem. J.*, 428:419–427, 2010.
- [4712] L. Xun and E.R. Sandvik. Characterization of 4-hydroxyphenylacetate 3-hydroxylase (HpaB) of *Escherichia coli* as a reduced flavin adenine dinucleotide-utilizing monooxygenase. *Appl. Environ. Microbiol.*, 66:481–486, 2000.
- [4713] L. Xun, E. Topp, and C.S. Orser. Confirmation of oxidative dehalogenation of pentachlorophenol by a *Flavobacterium* pentachlorophenol hydroxylase. *J. Bacteriol.*, 174:5745–5747, 1992.
- [4714] L. Xun, E. Topp, and C.S. Orser. Diverse substrate range of a *Flavobacterium* pentachlorophenol hydroxylase and reaction stoichiometries. *J. Bacteriol.*, 174:2898–2902, 1992.
- [4715] L. Xun, E. Topp, and C.S. Orser. Purification and characterization of a tetrachloro-*p*-hydroquinone reductive dehalogenase from a *Flavobacterium* sp. *J. Bacteriol.*, 174:8003–8007, 1992.
- [4716] L. Xun and C.M. Webster. A monooxygenase catalyzes sequential dechlorinations of 2,4,6-trichlorophenol by oxidative and hydrolytic reactions. *J. Biol. Chem.*, 279:6696–6700, 2004.
- [4717] K. Yabe, Y. Matsuyama, Y. Ando, H. Nakajima, and T. Hamasaki. Stereochemistry during aflatoxin biosynthesis: conversion of norsolorinic acid to averufin. *Appl. Environ. Microbiol.*, 59:2486–2492, 1993.
- [4718] I. Yadid, J. Rudolph, K. Hlouchova, and S.D. Copley. Sequestration of a highly reactive intermediate in an evolving pathway for degradation of pentachlorophenol. *Proc. Natl. Acad. Sci. USA*, 110:E2182–E2190, 2013.
- [4719] T. Yagi. Formate: cytochrome oxidoreductase of *Desulfovibrio vulgaris*. *J. Biochem. (Tokyo)*, 66:473–478, 1969.
- [4720] T. Yagi. Purification and properties of cytochrome *c*-553, an electron acceptor for formate dehydrogenase of *Desulfovibrio vulgaris*, Miyazaki. *Biochim. Biophys. Acta*, 548:96–105, 1979.
- [4721] T. Yagi, G.M. Kishore, and E.E. Snell. The bacterial oxidation of vitamin B₆. 4-Pyridoxic acid dehydrogenase: a membrane-bound enzyme from *Pseudomonas* MA-1. *J. Biol. Chem.*, 258:9419–9425, 1983.
- [4722] E.W. Yamada and W.B. Jakoby. Aldehyde oxidation. V. Direct conversion of malonic semialdehyde to acetyl-coenzyme A. *J. Biol. Chem.*, 235:589–594, 1960.
- [4723] H. Yamada. Putrescine oxidase (*Micrococcus rubens*). *Methods Enzymol.*, 17B:726–730, 1971.
- [4724] M. Yamada, Y. Okada, T. Yoshida, and T. Nagasawa. Biotransformation of isoeugenol to vanillin by *Pseudomonas putida* IE27 cells. *Appl. Microbiol. Biotechnol.*, 73:1025–1030, 2007.
- [4725] M. Yamada, Y. Okada, T. Yoshida, and T. Nagasawa. Purification, characterization and gene cloning of isoeugenol-degrading enzyme from *Pseudomonas putida* IE27. *Arch. Microbiol.*, 187:511–517, 2007.

- [4726] M. Yamada, K. Sumi, K. Matsushita, O. Adachi, and Y. Yamada. Topological analysis of quinoprotein glucose-dehydrogenase in *Escherichia coli* and its ubiquinone-binding site. *J. Biol. Chem.*, 268:12812–12817, 1993.
- [4727] Y. Yamada, K. Aida, and T. Uemura. Enzymatic studies on the oxidation of sugar and sugar alcohol. I. Purification and properties of particle-bound fructose dehydrogenase. *J. Biochem. (Tokyo)*, 61:636–646, 1967.
- [4728] Y. Yamada, K. Iizuka, K. Aida, and T. Uemura. Enzymatic studies on the oxidation of sugar and sugar alcohol. 3. Purification and properties of L-sorbose oxidase from *Trametes sanguinea*. *J. Biochem. (Tokyo)*, 62:223–229, 1967.
- [4729] M. Yamaguchi. Studies on regulatory functions of malic enzymes. IV. Effects of sulfhydryl group modification on the catalytic function of NAD-linked malic enzyme from *Escherichia coli*. *J. Biochem.*, 86:325–333, 1979.
- [4730] M. Yamaguchi and H. Fujisawa. Characterization of NADH-cytochrome *c* reductase, a component of benzoate 1,2-dioxygenase system from *Pseudomonas arvilla* C-1. *J. Biol. Chem.*, 253:8848–8853, 1978.
- [4731] M. Yamaguchi and H. Fujisawa. Purification and characterization of an oxygenase component in benzoate 1,2-dioxygenase system from *Pseudomonas arvilla* C-1. *J. Biol. Chem.*, 255:5058–5063, 1980.
- [4732] M. Yamaguchi and H. Fujisawa. Subunit structure of oxygenase component in benzoate-1,2-dioxygenase system from *Pseudomonas arvilla* C-1. *J. Biol. Chem.*, 257:12497–12502, 1982.
- [4733] T. Yamaguchi, Y. Kuwahara, and Y. Asano. A novel cytochrome P450, CYP3201B1, is involved in (*R*)-mandelonitrile biosynthesis in a cyanogenic millipede. *FEBS Open Bio*, 7:335–347, 2017.
- [4734] T. Yamaguchi, K. Yamamoto, and Y. Asano. Identification and characterization of CYP79D16 and CYP71AN24 catalyzing the first and second steps in L-phenylalanine-derived cyanogenic glycoside biosynthesis in the Japanese apricot, *Prunus mume* Sieb. et Zucc. *Plant Mol. Biol.*, 86:215–223, 2014.
- [4735] H. Yamamoto, K. Inoue, S.M. Li, and L. Heide. Geranylhydroquinone 3''-hydroxylase, a cytochrome *P*-450 monooxygenase from *Lithospermum erythrorhizon* cell suspension cultures. *Planta*, 210:312–317, 2000.
- [4736] H. Yamamoto, N. Katano, A. Ooi, and K. Inoue. Secologanin synthase which catalyzes the oxidative cleavage of loganin into secologanin is a cytochrome *P*-450. *Phytochemistry*, 53:7–12, 2000.
- [4737] H. Yamamoto, N. Katano, Y. Ooi, and K. Inoue. Transformation of loganin and 7-deoxyloganin into secologanin by *Lonicera japonica* cell suspension cultures. *Phytochemistry*, 50:417–422, 1999.
- [4738] H. Yamamoto, A. Yatou, and K. Inoue. 8-Dimethylallylnaringenin 2'-hydroxylase, the crucial cytochrome *P*₄₅₀ monooxygenase for lavandulylated flavanone formation in *Sophora flavescens* cultured cells. *Phytochemistry*, 58:671–676, 2001.
- [4739] H.Y. Yamamoto and R.M. Higashi. Violaxanthin de-epoxidase. Lipid composition and substrate specificity. *Arch. Biochem. Biophys.*, 190:514–522, 1978.
- [4740] I. Yamamoto, T. Saiki, S.-M. Liu, and L.G. Ljungdahl. Purification and properties of NADP-dependent formate dehydrogenase from *Clostridium thermoaceticum*, a tungsten-selenium-iron protein. *J. Biol. Chem.*, 258:1826–1832, 1983.
- [4741] S. Yamamoto and K. Bloch. Studies on squalene epoxidase of rat liver. *J. Biol. Chem.*, 245:1670–1674, 1970.
- [4742] S. Yamamoto and O. Hayaishi. Tryptophan pyrrolase of rabbit intestine. D- and L-tryptophan-cleaving enzyme or enzymes. *J. Biol. Chem.*, 242:5260–5266, 1967.
- [4743] S. Yamamoto, M. Katagiri, H. Maeno, and O. Hayaishi. Salicylate hydroxylase, a monooxygenase requiring flavin adenine dinucleotide. *J. Biol. Chem.*, 240:3408–3413, 1965.
- [4744] K. Yamanaka, M. Gino, and R. Kaneda. A specific NAD-D-xylose dehydrogenase from *Arthrobacter* sp. *Agric. Biol. Chem.*, 41:1493–1499, 1977.
- [4745] K. Yamanaka and R. Minoshima. Identification and characterization of a nicotinamide adenine dinucleotide-dependent para-hydroxybenzyl alcohol-dehydrogenase from *Rhodospseudomonas acidophila* M402. *Agric. Biol. Chem.*, 48:1161–1171, 1984.
- [4746] T. Yamanaka and K. Okunuki. Isolation of a cytochrome peroxidase from *Thiobacillus novellus*. *Biochim. Biophys. Acta*, 220:354–356, 1970.

- [4747] T. Yamanaka, T. Yoshioka, and K. Kimura. Purification of sulphite cytochrome c reductase of *Thiobacillus novellus* and reconstitution of its sulphite oxidase system with the purified constituents. *Plant and Cell Physiol.*, 22:631–622, 1981.
- [4748] K. Yamane, C. Toumazou, Y. Tsukada, H. Erdjument-Bromage, P. Tempst, J. Wong, and Y. Zhang. JHDM2A, a JmjC-containing H₃K9 demethylase, facilitates transcription activation by androgen receptor. *Cell*, 125:483–495, 2006.
- [4749] Y. Yamanishi, H. Mihara, M. Osaki, H. Muramatsu, N. Esaki, T. Sato, Y. Hizukuri, S. Goto, and M. Kanehisa. Prediction of missing enzyme genes in a bacterial metabolic network. Reconstruction of the lysine-degradation pathway of *Pseudomonas aeruginosa*. *FEBS J.*, 274:2262–2273, 2007.
- [4750] S. Yamano, E. Kaguera, T. Ishida, and S. Toki. Purification and characterization of guinea pig liver morphine 6-dehydrogenase. *J. Biol. Chem.*, 260:5259–5264, 1985.
- [4751] S. Yamano, F. Nishida, and S. Toki. Guinea-pig liver morphine 6-dehydrogenase as a naloxone reductase. *Biochem. Pharmacol.*, 35:4321–4326, 1986.
- [4752] M. Yamashita, H. Omura, E. Okamoto, Y. Furuya, M. Yabuuchi, K. Fukahi, and Y. Murooka. Isolation, characterization, and molecular cloning of a thermostable xylitol oxidase from *Streptomyces* sp. IKD472. *J. Biosci. Bioeng.*, 89:350–360, 2000.
- [4753] I. Yamazaki and L.H. Piette. Mechanism of free radical formation and disappearance during the ascorbic acid oxidase and peroxidase reactions. *Biochim. Biophys. Acta*, 50:62–69, 1961.
- [4754] S. Yamazaki. A selenium-containing hydrogenase from *Methanococcus vannielii*. Identification of the selenium moiety as a selenocysteine residue. *J. Biol. Chem.*, 257:7926–7929, 1982.
- [4755] A. Yan, Z. Guan, and C.R.H. Raetz. An undecaprenyl phosphate-aminoarabinose flippase required for polymyxin resistance in *Escherichia coli*. *J. Biol. Chem.*, 282:36077–36089, 2007.
- [4756] F. Yan, S.J. Moon, P. Liu, Z. Zhao, J.D. Lipscomb, A. Liu, and H.W. Liu. Determination of the substrate binding mode to the active site iron of (*S*)-2-hydroxypropylphosphonic acid epoxidase using ¹⁷O-enriched substrates and substrate analogues. *Biochemistry*, 46:12628–12638, 2007.
- [4757] W. Yan, G.F. Jang, F. Haeseleer, N. Esumi, J. Chang, M. Kerrigan, M. Campochiaro, P. Campochiaro, K. Palczewski, and D.J. Zack. Cloning and characterization of a human β , β -carotene-15,15'-dioxygenase that is highly expressed in the retinal pigment epithelium. *Genomics*, 72:193–202, 2001.
- [4758] Z. Yan, M. Wang, and J.G. Ferry. A ferredoxin- and F₄₂₀H₂-dependent, electron-bifurcating, heterodisulfide reductase with homologs in the domains Bacteria and Archaea. *mBio*, 8:e02285–16–, 2017.
- [4759] H. Yanagawa and F. Egami. Asparagusate dehydrogenases and lipoyl dehydrogenase from asparagus mitochondria. *Biochim. Biophys. Acta*, 384:342–352, 1975.
- [4760] H. Yanagawa and F. Egami. Asparagusate dehydrogenases and lipoyl dehydrogenase from asparagus mitochondria. Physical, chemical, and enzymatic properties. *J. Biol. Chem.*, 251:3637–3644, 1976.
- [4761] K. Yanagibashi, M. Haniu, J.E. Shively, W.H. Shen, and P. Hall. The synthesis of aldosterone by the adrenal cortex. Two zones (fasciculata and glomerulosa) possess one enzyme for 11 β -, 18-hydroxylation, and aldehyde synthesis. *J. Biol. Chem.*, 261:3556–3562, 1986.
- [4762] C.C. Yang, L.C. Packman, and N.S. Scrutton. The primary structure of *Hyphomicrobium X* dimethylamine dehydrogenase. Relationship to trimethylamine dehydrogenase and implications for substrate recognition. *Eur. J. Biochem.*, 232:264–271, 1995.
- [4763] F.C. Yang, Y.L. Chen, S.L. Tang, C.P. Yu, P.H. Wang, W. Ismail, C.H. Wang, J.Y. Ding, C.Y. Yang, C.Y. Yang, and Y.R. Chiang. Integrated multi-omics analyses reveal the biochemical mechanisms and phylogenetic relevance of anaerobic androgen biodegradation in the environment. *ISME J.*, 10:1967–1983, 2016.
- [4764] H.C. Yang, J.F. Hainfeld, J.S. Wall, and P.A. Frey. Quaternary structure of pyruvate dehydrogenase complex from *Escherichia coli*. *J. Biol. Chem.*, 260:16049–16051, 1985.
- [4765] T. Yang, Y.H. Shao, L.Z. Guo, X.L. Meng, H. Yu, and W.D. Lu. Role of *N,N*-dimethylglycine and its catabolism to sarcosine in *Chromohalobacter salexigens* DSM 3043. *Appl. Environ. Microbiol.*, 86, 2020.

- [4766] W. Yang, I.F. Moore, K.P. Koteva, D.C. Bareich, D.W. Hughes, and G.D. Wright. TetX is a flavin-dependent monooxygenase conferring resistance to tetracycline antibiotics. *J. Biol. Chem.*, 279:52346–52352, 2004.
- [4767] X. Yang and K. Ma. Characterization of an exceedingly active NADH oxidase from the anaerobic hyperthermophilic bacterium *Thermotoga maritima*. *J. Bacteriol.*, 189:3312–3317, 2007.
- [4768] Y. Yang, R. Yatsunami, A. Ando, N. Miyoko, T. Fukui, S. Takaichi, and S. Nakamura. Complete biosynthetic pathway of the C₅₀ carotenoid bacterioruberin from lycopene in the extremely halophilic archaeon *Haloarcula japonica*. *J. Bacteriol.*, 197:1614–1623, 2015.
- [4769] Y. Yang, S. Yuan, T. Chen, P. Ma, G. Shang, and Y. Dai. Cloning, heterologous expression, and functional characterization of the nicotinate dehydrogenase gene from *Pseudomonas putida* KT2440. *Biodegradation*, 20:541–549, 2009.
- [4770] Y. Yang, M. Zhang, G. Eggertsen, and J.Y. Chiang. On the mechanism of bile acid inhibition of rat sterol 12 α -hydroxylase gene (CYP8B1) transcription: roles of α -fetoprotein transcription factor and hepatocyte nuclear factor 4 α . *Biochim. Biophys. Acta*, 1583:63–73, 2002.
- [4771] Y. Yang, G. Zhao, T.K. Man, and M.E. Winkler. Involvement of the *gapA*- and *epd* (*gapB*)-encoded dehydrogenases in pyridoxal 5'-phosphate coenzyme biosynthesis in *Escherichia coli* K-12. *J. Bacteriol.*, 180:4294–4299, 1998.
- [4772] Z. Yang, X. Chi, M. Funabashi, S. Baba, K. Nonaka, P. Pahari, J. Unrine, J.M. Jacobsen, G.I. Elliott, J. Rohr, and S.G. Van Lanen. Characterization of LipL as a non-heme, Fe(II)-dependent α -ketoglutarate:UMP dioxygenase that generates uridine-5'-aldehyde during A-90289 biosynthesis. *J. Biol. Chem.*, 286:7885–7892, 2011.
- [4773] Z. Yang, A. Savchenko, A. Yakunin, R. Zhang, A. Edwards, C. Arrowsmith, and L. Tong. Aspartate dehydrogenase, a novel enzyme identified from structural and functional studies of TM1643. *J. Biol. Chem.*, 278:8804–8808, 2003.
- [4774] Z. Yang, J. Unrine, K. Nonaka, and S.G. Van Lanen. Fe(II)-dependent, uridine-5'-monophosphate α -ketoglutarate dioxygenases in the synthesis of 5'-modified nucleosides. *Methods Enzymol.*, 516:153–168, 2012.
- [4775] Z.M. Yang and C.E. Bauer. *Rhodobacter capsulatus* genes involved in early steps of the bacteriochlorophyll biosynthetic pathway. *J. Bacteriol.*, 172:5001–5010, 1990.
- [4776] H. Yaniv and C. Gilvarg. Aromatic biosynthesis. XIV. 5-Dehydroshikimic reductase. *J. Biol. Chem.*, 213:787–795, 1955.
- [4777] A. Yarzabal, C. Appia-Ayme, J. Ratouchniak, and V. Bonnefoy. Regulation of the expression of the *Acidithiobacillus ferrooxidans* *rus* operon encoding two cytochromes *c*, a cytochrome oxidase and rusticyanin. *Microbiology*, 150:2113–2123, 2004.
- [4778] A. Yarzabal, G. Bresseur, J. Ratouchniak, K. Lund, D. Lemesle-Meunier, J.A. DeMoss, and V. Bonnefoy. The high-molecular-weight cytochrome *c* *Cyc2* of *Acidithiobacillus ferrooxidans* is an outer membrane protein. *J. Bacteriol.*, 184:313–317, 2002.
- [4779] H. Yasui, K. Takai, R. Yoshida, and O. Hayaishi. Interferon enhances tryptophan metabolism by inducing pulmonary indoleamine 2,3-dioxygenase: its possible occurrence in cancer patients. *Proc. Natl. Acad. Sci. USA*, 83:6622–6626, 1986.
- [4780] S. Yasumoto, H. Seki, Y. Shimizu, E.O. Fukushima, and T. Muranaka. Functional characterization of CYP716 family P450 enzymes in triterpenoid biosynthesis in tomato. *Front. Plant Sci.*, 8:21–21, 2017.
- [4781] T. Yasuta, S. Okazaki, H. Mitsui, K. Yuhashi, H. Ezura, and K. Minamisawa. DNA sequence and mutational analysis of rhizobitoxine biosynthesis genes in *Bradyrhizobium elkanii*. *Appl. Environ. Microbiol.*, 67:4999–5009, 2001.
- [4782] M.G. Yates and A. Nason. Electron transport systems of the chemoautotroph *Ferrobacillus ferrooxidans*. II. Purification and properties of a heat-labile iron-cytochrome *c* reductase. *J. Biol. Chem.*, 241:4872–4880, 1966.
- [4783] S. Ye, F. Von Delft, A. Brooun, M.W. Knuth, R.V. Swanson, and D.E. McRee. The crystal structure of shikimate dehydrogenase (AroE) reveals a unique NADPH binding mode. *J. Bacteriol.*, 185:4144–4151, 2003.
- [4784] C.M. Yeager, P.J. Bottomley, D.J. Arp, and M.R. Hyman. Inactivation of toluene 2-monooxygenase in *Burkholderia cepacia* G4 by alkynes. *Appl. Environ. Microbiol.*, 65:632–639, 1999.

- [4785] A.P. Yeh, Y. Hu, F.E. Jenney, Adams Jr., Rees M.W.W., and D.C. Structures of the superoxide reductase from *Pyrococcus furiosus* in the oxidized and reduced states. *Biochemistry*, 39:2499–2508, 2000.
- [4786] E. Yeh, S. Garneau, and C.T. Walsh. Robust *in vitro* activity of RebF and RebH, a two-component reductase/halogenase, generating 7-chlorotryptophan during rebeccamycin biosynthesis. *Proc. Natl. Acad. Sci. USA*, 102:3960–3965, 2005.
- [4787] W.K. Yeh, S.K. Ghag, and S.W. Queener. Enzymes for epimerization of isopenicillin N, ring expansion of penicillin N, and 3'-hydroxylation of deacetoxycephalosporin C. Function, evolution, refolding, and enzyme engineering. *Ann. N.Y. Acad. Sci.*, 672:396–408, 1992.
- [4788] Y.-C. Yeh and D.M. Greenberg. Purification and properties of N^5, N^{10} -methylenetetrahydrofolate dehydrogenase of calf thymus. *Biochim. Biophys. Acta*, 105:279–291, 1965.
- [4789] W.S. Yew, A.A. Fedorov, E.V. Fedorov, J.F. Rakus, R.W. Pierce, S.C. Almo, and J.A. Gerlt. Evolution of enzymatic activities in the enolase superfamily: L-fuconate dehydratase from *Xanthomonas campestris*. *Biochemistry*, 45:14582–14597, 2006.
- [4790] C. Yi, G. Jia, G. Hou, Q. Dai, W. Zhang, G. Zheng, X. Jian, C.G. Yang, Q. Cui, and C. He. Iron-catalysed oxidation intermediates captured in a DNA repair dioxygenase. *Nature*, 468:330–333, 2010.
- [4791] C. Yi, C.G. Yang, and C. He. A non-heme iron-mediated chemical demethylation in DNA and RNA. *Acc. Chem. Res.*, 42:519–529, 2009.
- [4792] D.T. Yin, S. Urresti, M. Lafond, E.M. Johnston, F. Derikvand, L. Ciano, J.G. Berrin, B. Henrissat, P.H. Walton, G.J. Davies, and H. Brumer. Structure-function characterization reveals new catalytic diversity in the galactose oxidase and glyoxal oxidase family. *Nat. Commun.*, 6:10197–10197, 2015.
- [4793] Q.J. Yin, W.J. Zhang, X.Q. Qi, S.D. Zhang, T. Jiang, X.G. Li, Y. Chen, C.L. Santini, H. Zhou, I.M. Chou, and L.F. Wu. High hydrostatic pressure inducible trimethylamine *N*-oxide reductase improves the pressure tolerance of piezosensitive bacteria *Vibrio fluvialis*. *Front. Microbiol.*, 8:2646–2646, 2017.
- [4794] N. Yokochi, Y. Yoshikane, S. Matsumoto, M. Fujisawa, K. Ohnishi, and T. Yagi. Gene identification and characterization of 5-formyl-3-hydroxy-2-methylpyridine 4-carboxylic acid 5-dehydrogenase, an NAD^+ -dependent dismutase. *J. Biochem.*, 145:493–503, 2009.
- [4795] K. Yokoyama, M. Numakura, F. Kudo, D. Ohmori, and T. Eguchi. Characterization and mechanistic study of a radical SAM dehydrogenase in the biosynthesis of butirosin. *J. Am. Chem. Soc.*, 129:15147–15155, 2007.
- [4796] K. Yokoyama, D. Ohmori, F. Kudo, and T. Eguchi. Mechanistic study on the reaction of a radical SAM dehydrogenase BtrN by electron paramagnetic resonance spectroscopy. *Biochemistry*, 47:8950–8960, 2008.
- [4797] S. Yokoyama, T. Miyazawa, Y. Iitaka, Z. Yamaizumi, H. Kasai, and S. Nishimura. Three-dimensional structure of hypermodified nucleoside Q located in the wobbling position of tRNA. *Nature*, 282:107–109, 1979.
- [4798] K. Yoneda, R. Kawakami, Y. Tagashira, H. Sakuraba, S. Goda, and T. Ohshima. The first archaeal L-aspartate dehydrogenase from the hyperthermophile *Archaeoglobus fulgidus*: gene cloning and enzymological characterization. *Biochim. Biophys. Acta*, 1764:1087–1093, 2006.
- [4799] K. Yoneda, H. Sakuraba, H. Tsuge, N. Katunuma, and T. Ohshima. Crystal structure of archaeal highly thermostable L-aspartate dehydrogenase/NAD/citrate ternary complex. *FEBS J.*, 274:4315–4325, 2007.
- [4800] T. Yonetani. Cytochrome c peroxidase. *Adv. Enzymol. Relat. Areas Mol. Biol.*, 33:309–335, 1970.
- [4801] Y.-N. Yong. Detection of a pteridine oxidase in plants. *Plant Sci. Lett.*, 18:169–175, 1980.
- [4802] H. Yoon, C.D. Anderson, and B.M. Anderson. Kinetic studies of *Haemophilus influenzae* 6-phosphogluconate dehydrogenase. *Biochim. Biophys. Acta*, 994:75–80, 1989.
- [4803] T. Yorifuji, K. Koike, T. Sakurai, and K. Yokoyama. 4-Aminobutyraldehyde and 4-guanidinobutyraldehyde dehydrogenases for arginine degradation in *Pseudomonas putida*. *Agric. Biol. Chem.*, 50:2009–2016, 1986.
- [4804] J.L. York, A.P. Grollman, and C. Bublitz. TPN-L-gulonate dehydrogenase. *Biochim. Biophys. Acta*, 47:298–306, 1961.

- [4805] A. Yoshida and E. Freese. Enzymic properties of alanine dehydrogenase of *Bacillus subtilis*. *Biochim. Biophys. Acta*, 96:248–262, 1965.
- [4806] A. Yoshida, T. Tomita, H. Atomi, T. Kuzuyama, and M. Nishiyama. Lysine biosynthesis of *Thermococcus kodakarensis* with the capacity to function as an ornithine biosynthetic system. *J. Biol. Chem.*, 291:21630–21643, 2016.
- [4807] K. Yoshida, S. Ueda, and I. Maeda. Carotenoid production in *Bacillus subtilis* achieved by metabolic engineering. *Biotechnol. Lett.*, 31:1789–1793, 2009.
- [4808] K. Yoshida, M. Yamaguchi, T. Morinaga, M. Ikeuchi, M. Kinehara, and H. Ashida. Genetic modification of *Bacillus subtilis* for production of D-*chiro*-inositol, an investigational drug candidate for treatment of type 2 diabetes and polycystic ovary syndrome. *Appl. Environ. Microbiol.*, 72:1310–1315, 2006.
- [4809] K.-I. Yoshida, H. Oshima, and P. Troen. Studies of the human testis. XIII. Properties of nicotinamide adenine dinucleotide (reduced form)-linked 17 α -hydroxylation. *J. Clin. Endocrinol. Metab.*, 50:895–899, 1980.
- [4810] M. Yoshida, T. Oikawa, H. Obata, K. Abe, H. Mihara, and N. Esaki. Biochemical and genetic analysis of the γ -resorcylate (2,6-dihydroxybenzoate) catabolic pathway in *Rhizobium* sp. strain MTP-10005: identification and functional analysis of its gene cluster. *J. Bacteriol.*, 189:1573–1581, 2007.
- [4811] N. Yoshida, S. Akazawa, T. Katsuragi, and Y. Tani. Characterization of two fructosyl-amino acid oxidase homologs of *Schizosaccharomyces pombe*. *J. Biosci. Bioeng.*, 97:278–280, 2004.
- [4812] T. Yoshida, S. Takahashi, and J. Kikuchi. Partial purification and reconstitution of the heme oxygenase system from pig spleen microsomes. *J. Biochem. (Tokyo)*, 75:1187–1191, 1974.
- [4813] Y. Yoshida and Y. Aoyama. Yeast cytochrome P-450 catalyzing lanosterol 14 α -demethylation. I. Purification and spectral properties. *J. Biol. Chem.*, 259:1655–1660, 1984.
- [4814] Y. Yoshida, Y. Nakano, T. Nezu, Y. Yamashita, and T. Koga. A novel NDP-6-deoxyhexosyl-4-ulose reductase in the pathway for the synthesis of thymidine diphosphate-D-fucose. *J. Biol. Chem.*, 274:16933–16939, 1999.
- [4815] S. Yoshihara and K. Tatsumi. Purification and characterization of hepatic aldehyde oxidase in male and female mice. *Arch. Biochem. Biophys.*, 338:29–34, 1997.
- [4816] K. Yoshikawa, S. Takei, S. Hasegawa-Ishii, Y. Chiba, A. Furukawa, N. Kawamura, M. Hosokawa, D.F. Woodward, K. Watanabe, and A. Shimada. Preferential localization of prostamide/prostaglandin F synthase in myelin sheaths of the central nervous system. *Brain Res.*, 1367:22–32, 2011.
- [4817] A. Yoshimoto, T. Ogasawara, I. Kitamura, T. Oki, T. Inui, T. Takeuchi, and H. Umezawa. Enzymatic conversion of aclacinomycin A to Y by a specific oxidoreductase in *Streptomyces*. *J. Antibiot. (Tokyo)*, 32:472–481, 1979.
- [4818] A. Yoshimoto and R. Sato. Studies on yeast sulfite reductase. I. Purification and characterization. *Biochim. Biophys. Acta*, 153:555–575, 1968.
- [4819] T. Yoshiyama-Yanagawa, S. Enya, Y. Shimada-Niwa, S. Yaguchi, Y. Haramoto, T. Matsuya, K. Shiomi, Y. Sasakura, S. Takahashi, M. Asashima, H. Kataoka, and R. Niwa. The conserved Rieske oxygenase DAF-36/Neverland is a novel cholesterol-metabolizing enzyme. *J. Biol. Chem.*, 286:25756–25762, 2011.
- [4820] K.-S. You. Stereospecificity for nicotinamide nucleotides in enzymatic and chemical hydride transfer reactions. *CRC Crit. Rev. Biochem.*, 17:313–451, 1985.
- [4821] S.Y. You, S. Cosloy, and H. Schulz. Evidence for the essential function of 2,4-dienoyl-coenzyme A reductase in the β -oxidation of unsaturated fatty acids *in vivo*. Isolation and characterization of an *Escherichia coli* mutant with a defective 2,4-dienoyl-coenzyme A reductase. *J. Biol. Chem.*, 264:16489–16495, 1989.
- [4822] Z. You, S. Omura, H. Ikeda, and D.E. Cane. Pentalenolactone biosynthesis. Molecular cloning and assignment of biochemical function to PtlH, a non-heme iron dioxygenase of *Streptomyces avermitilis*. *J. Am. Chem. Soc.*, 128:6566–6567, 2006.
- [4823] Z. You, S. Omura, H. Ikeda, and D.E. Cane. Pentalenolactone biosynthesis: Molecular cloning and assignment of biochemical function to PtlF, a short-chain dehydrogenase from *Streptomyces avermitilis*, and identification of a new biosynthetic intermediate. *Arch. Biochem. Biophys.*, 459:233–240, 2007.

- [4824] Z. You, S. Omura, H. Ikeda, D.E. Cane, and G. Jogl. Crystal structure of the non-heme iron dioxygenase PtlH in pentalenolactone biosynthesis. *J. Biol. Chem.*, 282:36552–36560, 2007.
- [4825] M.B. Youdim and Y.S. Bakhle. Monoamine oxidase: isoforms and inhibitors in Parkinson's disease and depressive illness. *Br. J. Pharmacol.*, 147 Suppl. 1:S287–S296, 2006.
- [4826] M.B. Youdim, D. Edmondson, and K.F. Tipton. The therapeutic potential of monoamine oxidase inhibitors. *Nat. Rev. Neurosci.*, 7:295–309, 2006.
- [4827] B. Youn, S.G. Moinuddin, L.B. Davin, N.G. Lewis, and C. Kang. Crystal structures of apo-form and binary/ternary complexes of *Podophyllum* secoisolariciresinol dehydrogenase, an enzyme involved in formation of health-protecting and plant defense lignans. *J. Biol. Chem.*, 280:12917–12926, 2005.
- [4828] I.G. Young and F. Gibson. Regulation of the enzymes involved in the biosynthesis of 2,3-dihydroxybenzoic acid in *Aerobacter aerogenes* and *Escherichia coli*. *Biochim. Biophys. Acta*, 177:401–411, 1969.
- [4829] J. Yourno and I. Ino. Purification and crystallization of histidinol dehydrogenase from *Salmonella typhimurium* LT-2. *J. Biol. Chem.*, 243:3273–3276, 1968.
- [4830] B. Yu, M. Ruppert, and J. Stöckigt. Deoxysarpagine hydroxylase — a novel enzyme closing a short side pathway of alkaloid biosynthesis in *Rauvolfia*. *Bioorg. Med. Chem.*, 10:2479–2483, 2002.
- [4831] C.A. Yu and I.C. Gunsalus. Monooxygenases. VII. Camphor ketolactonase I and the role of three protein components. *J. Biol. Chem.*, 244:6149–6152, 1969.
- [4832] C.L. Yu, Y. Kale, S. Gopishetty, T.M. Louie, and M. Subramanian. A novel caffeine dehydrogenase in *Pseudomonas* sp. strain CBB1 oxidizes caffeine to trimethyluric acid. *J. Bacteriol.*, 190:772–776, 2008.
- [4833] F. Yu, S. Okamoto, H. Harada, K. Yamasaki, N. Misawa, and R. Utsumi. *Zingiber zerumbet* CYP71BA1 catalyzes the conversion of α -humulene to 8-hydroxy- α -humulene in zerumbone biosynthesis. *Cell. Mol. Life Sci.*, 68:1033–1040, 2011.
- [4834] H. Yu, S. Zhao, and L. Guo. Novel gene encoding 5-aminosalicylate 1,2-dioxygenase from *Comamonas* sp. strain QT12 and catalytic properties of the purified enzyme. *J. Bacteriol.*, 200, 2018.
- [4835] J. Yu, P.K. Chang, J.W. Cary, D. Bhatnagar, and T.E. Cleveland. *avnA*, a gene encoding a cytochrome *P*-450 monooxygenase, is involved in the conversion of averantin to averufin in aflatoxin biosynthesis in *Aspergillus parasiticus*. *Appl. Environ. Microbiol.*, 63:1349–1356, 1997.
- [4836] J. Yu, P.K. Chang, K.C. Ehrlich, J.W. Cary, B. Montalbano, J.M. Dyer, D. Bhatnagar, and T.E. Cleveland. Characterization of the critical amino acids of an *Aspergillus parasiticus* cytochrome *P*-450 monooxygenase encoded by *ordA* that is involved in the biosynthesis of aflatoxins B1, G1, B2, and G2. *Appl. Environ. Microbiol.*, 64:4834–4841, 1998.
- [4837] L. Yu, S. Mah, T. Otani, and P. Dedon. The benzoxazolinone of C-1027 confers intercalative DNA binding. *J. Am. Chem. Soc.*, 117:8877–8878, 1995.
- [4838] X. Yu, T. Liu, F. Zhu, and C. Khosla. *In vitro* reconstitution and steady-state analysis of the fatty acid synthase from *Escherichia coli*. *Proc. Natl. Acad. Sci. USA*, 108:18643–18648, 2011.
- [4839] Y. Yu, X. Hou, X. Ni, and H. Xia. Biosynthesis of 3'-deoxy-carbamoylkanamycin C in a *Streptomyces tenebrarius* mutant strain by *tacB* gene disruption. *J. Antibiot. (Tokyo)*, 61:63–69, 2008.
- [4840] B. Yuan, N. Yokochi, Y. Yoshikane, K. Ohnishi, and T. Yagi. Molecular cloning, identification and characterization of 2-methyl-3-hydroxypyridine-5-carboxylic-acid-dioxygenase-coding gene from the nitrogen-fixing symbiotic bacterium *Mesorhizobium loti*. *J. Biosci. Bioeng.*, 102:504–510, 2006.
- [4841] H. Yuan, G. Fu, P.T. Brooks, I. Weber, and G. Gadda. Steady-state kinetic mechanism and reductive half-reaction of D-arginine dehydrogenase from *Pseudomonas aeruginosa*. *Biochemistry*, 49:9542–9550, 2010.
- [4842] H. Yuan, Y. Xin, D. Hamelberg, and G. Gadda. Insights on the mechanism of amine oxidation catalyzed by D-arginine dehydrogenase through pH and kinetic isotope effects. *J. Am. Chem. Soc.*, 133:18957–18965, 2011.

- [4843] T. Yubisui, M. Tamura, and M. Takeshita. Characterization of a second form of NADPH-flavin reductase purified from human erythrocytes. *Biochem. Int.*, 15:1–8, 1987.
- [4844] C.J. Yue, X. Zhou, and J.J. Zhong. Protopanaxadiol 6-hydroxylase and its role in regulating the ginsenoside heterogeneity in *Panax notoginseng* cells. *Biotechnol. Bioeng.*, 100:933–940, 2008.
- [4845] D.Y. Yum, B.Y. Lee, and J.G. Pan. Identification of the yqhE and yafB genes encoding two 2,5-diketo-D-gluconate reductases in *Escherichia coli*. *Appl. Environ. Microbiol.*, 65:3341–3346, 1999.
- [4846] E.J. Yun, S. Lee, H.T. Kim, J.G. Pelton, S. Kim, J. Choi I, G. Ko H, and K.H. Kim. The novel catabolic pathway of 3,6-anhydro-L-galactose, the main component of red macroalgae, in a marine bacterium. *Environ. Microbiol.*, 17:1677–1688, 2014.
- [4847] T. Yura and H.J. Vogel. Pyrroline-5-carboxylate reductase of *Neurospora crassa*: partial purification and some properties. *J. Biol. Chem.*, 234:335–338, 1959.
- [4848] Y.Y., Cronan Chang, , and Jr. Sulfhydryl chemistry detects three conformations of the lipid binding region of *Escherichia coli* pyruvate oxidase. *Biochemistry*, 36:11564–11573, 1997.
- [4849] A. Zaar, J. Gescher, W. Eisenreich, A. Bacher, and G. Fuchs. New enzymes involved in aerobic benzoate metabolism in *Azoarcus evansii*. *Mol. Microbiol.*, 54:223–238, 2004.
- [4850] O. Zaborina, D.L. Daubaras, A. Zago, L. Xun, , and K. , Klem,T., Nikolic, D. and Chakrabarty, A.M. Novel pathway for conversion of chlorohydroxyquinol to maleylacetate in *Burkholderia cepacia* AC1100. *J. Bacteriol.*, 180:4667–4675, 1998.
- [4851] T.M. Zabriskie and M.D. Jackson. Lysine biosynthesis and metabolism in fungi. *Nat. Prod. Rep.*, 17:85–97, 2000.
- [4852] M. Zachariou and R.K. Scopes. Glucose-fructose oxidoreductase: a new enzyme isolated from *Zymomonas mobilis* that is responsible for sorbitol production. *J. Bacteriol.*, 167:863–869, 1986.
- [4853] J.A. Zahn, D.M. Arciero, A.B. Hooper, and A.A. DiSpirito. Evidence for an iron center in the ammonia monooxygenase from *Nitrosomonas europaea*. *FEBS Lett.*, 397:35–38, 1996.
- [4854] M.M. Zakaria, T. Stegemann, C. Sievert, L.H. Kruse, E. Kaltenecker, U. Girreser, S.S. Cicek, M. Nimtz, and D. Ober. Insights into polyamine metabolism: homospermidine is double-oxidized in two discrete steps by a single copper-containing amine oxidase in pyrrolizidine alkaloid biosynthesis. *Plant Cell*, 34:2364–2382, 2022.
- [4855] R.A. Zakharyan and H.V. Aposhian. Enzymatic reduction of arsenic compounds in mammalian systems: the rate-limiting enzyme of rabbit liver arsenic biotransformation is MMA(V) reductase. *Chem. Res. Toxicol.*, 12:1278–1283, 1999.
- [4856] R. Zallot, R. Ross, W.H. Chen, S.D. Bruner, P.A. Limbach, and V. De Crecy-Lagard. Identification of a novel epoxyqueuosine reductase family by comparative genomics. *ACS Chem. Biol.*, 12:844–851, 2017.
- [4857] N. Zamboni, E. Fischer, D. Laudert, S. Aymerich, H.P. Hohmann, and U. Sauer. The *Bacillus subtilis* yqjI gene encodes the NADP⁺-dependent 6-P-gluconate dehydrogenase in the pentose phosphate pathway. *J. Bacteriol.*, 186:4528–4534, 2004.
- [4858] E. Zameitat, A.J. Pierik, K. Zocher, and M. Löffler. Dihydroorotate dehydrogenase from *Saccharomyces cerevisiae*: spectroscopic investigations with the recombinant enzyme throw light on catalytic properties and metabolism of fumarate analogues. *FEMS Yeast Res.*, 7:897–904, 2007.
- [4859] L.O. Zamir, R. Tiberio, K.A. Devor, F. Sauriol, S. Ahmad, and R.A. Jensen. Structure of D-prephenyllactate. A carboxy-cyclohexadienyl metabolite from *Neurospora crassa*. *J. Biol. Chem.*, 263:17284–17290, 1988.
- [4860] V.G. Zannoni and W.W. Weber. Isolation and properties of aromatic α -keto acid reductase. *J. Biol. Chem.*, 241:1340–1344, 1966.
- [4861] A. Zapun, J.C. Bardwell, and T.E. Creighton. The reactive and destabilizing disulfide bond of DsbA, a protein required for protein disulfide bond formation *in vivo*. *Biochemistry*, 32:5083–5092, 1993.
- [4862] S. Zäuner, W. Jochum, T. Bigorowski, and C. Benning. A cytochrome *b*₅-containing plastid-located fatty acid desaturase from *Chlamydomonas reinhardtii*. *Eukaryot Cell*, 11:856–863, 2012.

- [4863] S. Zayni, K. Steiner, A. Pfostl, A. Hofinger, P. Kosma, C. Schaffer, and P. Messner. The dTDP-4-dehydro-6-deoxyglucose reductase encoding *fcd* gene is part of the surface layer glycoprotein glycosylation gene cluster of *Geobacillus tepidamans* GS5-97T. *Glycobiology*, 17:433–443, 2007.
- [4864] I. Zegers, J.C. Martins, R. Willem, L. Wyns, and J. Messens. Arsenate reductase from *S. aureus* plasmid pI258 is a phosphatase drafted for redox duty. *Nat. Struct. Biol.*, 8:843–847, 2001.
- [4865] S. Zehner, A. Kotzsch, B. Bister, R.D. Sussmuth, C. Mendez, J.A. Salas, and K.H. van Pee. A regioselective tryptophan 5-halogenase is involved in pyrroindomycin biosynthesis in *Streptomyces rugosporus* LL-42D005. *Chem. Biol.*, 12:445–452, 2005.
- [4866] I. Zelitch. Oxidation and reduction of glycolic and glyoxylic acids in plants. II. Glyoxylic acid reductase. *J. Biol. Chem.*, 201:719–726, 1953.
- [4867] I. Zelitch. The isolation and action of crystalline glyoxylic acid reductase from tobacco leaves. *J. Biol. Chem.*, 216:553–575, 1955.
- [4868] E.A. Zeller. Diamine oxidases. In P.D. Boyer, H. Lardy, and K. Myrback, editors, *The Enzymes*, volume 8, pages 313–335. Academic Press, New York, 2nd edition, 1963.
- [4869] J. Zeng and J. Zhan. Characterization of a tryptophan 6-halogenase from *Streptomyces toxytricini*. *Biotechnol. Lett.*, 33:1607–1613, 2011.
- [4870] Z. Zeng, H. Chen, H. Yang, Y. Chen, W. Yang, X. Feng, H. Pei, and P.V. Welander. Identification of a protein responsible for the synthesis of archaeal membrane-spanning GDGT lipids. *Nat. Commun.*, 13:1545–1545, 2022.
- [4871] F. Zepeck, T. Grawert, J. Kaiser, N. Schramek, W. Eisenreich, A. Bacher, and F. Rohdich. Biosynthesis of isoprenoids. purification and properties of IspG protein from *Escherichia coli*. *J. Org. Chem.*, 70:9168–9174, 2005.
- [4872] J. Zeyer, H.P. Kocher, and N. Timmis. Influence of para-substituents on the oxidative metabolism of *o*-nitrophenols by *Pseudomonas putida* B2. *Appl. Environ. Microbiol.*, 52:334–339, 1986.
- [4873] H. Zhang, P.F. Coville, R.J. Walker, J.O. Miners, D.J. Birkett, and S. Wanwimolruk. Evidence for involvement of human CYP3A in the 3-hydroxylation of quinine. *Br. J. Clin. Pharmacol.*, 43:245–252, 1997.
- [4874] H. Zhang, Y. Zhao, H. Cao, G. Mou, and H. Yin. Expression and characterization of a lytic polysaccharide monooxygenase from *Bacillus thuringiensis*. *Int. J. Biol. Macromol.*, 79:72–75, 2015.
- [4875] J. Zhang, L. Dai, J. Yang, C. Liu, Y. Men, Y. Zeng, Y. Cai, Y. Zhu, and Y. Sun. Oxidation of cucurbitadienol catalyzed by CYP87D18 in the biosynthesis of mogrosides from *Siraitia grosvenorii*. *Plant Cell Physiol.*, 57:1000–1007, 2016.
- [4876] J.F. Zhang, W.Q. Chen, and H. Chen. Gene cloning and expression of a glucoside 3-dehydrogenase from *Sphingobacterium faecium* ZJF-D6, and used it to produce *N-p*-nitrophenyl-3-ketovalidamine. *World J. Microbiol. Biotechnol.*, 33:21–21, 2017.
- [4877] J.F. Zhang, Y.G. Zheng, Y.P. Xue, and Y.C. Shen. Purification and characterization of the glucoside 3-dehydrogenase produced by a newly isolated *Stenotrophomonas maltophilia* CCTCC M 204024. *Appl. Microbiol. Biotechnol.*, 71:638–645, 2006.
- [4878] J.J. Zhang, H. Liu, Y. Xiao, X.E. Zhang, and N.Y. Zhou. Identification and characterization of catabolic para-nitrophenol 4-monooxygenase and para-benzoquinone reductase from *Pseudomonas* sp. strain WBC-3. *J. Bacteriol.*, 191:2703–2710, 2009.
- [4879] L. Zhang, T. Kudo, N. Takaya, and H. Shoun. The B' helix determines cytochrome P450_{nor} specificity for the electron donors NADH and NADPH. *J. Biol. Chem.*, 277:33842–33847, 2002.
- [4880] L. Zhang, X. Lu, J. Lu, H. Liang, Q. Dai, G.L. Xu, C. Luo, H. Jiang, and C. He. Thymine DNA glycosylase specifically recognizes 5-carboxylcytosine-modified DNA. *Nat. Chem. Biol.*, 8:328–330, 2012.
- [4881] L. Zhang, K.J. Nelson, K.V. Rajagopalan, and G.N. George. Structure of the molybdenum site of *Escherichia coli* trimethylamine *N*-oxide reductase. *Inorg. Chem.*, 47:1074–1078, 2008.

- [4882] L. Zhang, W.R. Tschantz, and P.J. Casey. Isolation and characterization of a prenylcysteine lyase from bovine brain. *J. Biol. Chem.*, 272:23354–23359, 1997.
- [4883] M. Zhang, C. Gao, X. Guo, S. Guo, Z. Kang, D. Xiao, J. Yan, F. Tao, W. Zhang, W. Dong, P. Liu, C. Yang, C. Ma, and P. Xu. Increased glutarate production by blocking the glutaryl-CoA dehydrogenation pathway and a catabolic pathway involving L-2-hydroxyglutarate. *Nat. Commun.*, 9:2114–2114, 2018.
- [4884] Q. Zhang, T. Iwasaki, T. Wakagi, and T. Oshima. 2-oxoacid:ferredoxin oxidoreductase from the thermoacidophilic archaeon, *Sulfolobus* sp. strain 7. *J. Biochem.*, 120:587–599, 1996.
- [4885] W. Zhang, B.D. Ames, and C.T. Walsh. Identification of phenylalanine 3-hydroxylase for *meta*-tyrosine biosynthesis. *Biochemistry*, 50:5401–5403, 2011.
- [4886] W. Zhang, K. Watanabe, X. Cai, M.E. Jung, Y. Tang, and J. Zhan. Identifying the minimal enzymes required for anhydrotetracycline biosynthesis. *J. Am. Chem. Soc.*, 130:6068–6069, 2008.
- [4887] X. Zhang, M.S. Carter, M.W. Vetting, B. San Francisco, S. Zhao, N.F. Al-Obaidi, J.O. Solbiati, J.J. Thiaville, V. de Crecy-Lagard, M.P. Jacobson, S.C. Almo, and J.A. Gerlt. Assignment of function to a domain of unknown function: DUF1537 is a new kinase family in catabolic pathways for acid sugars. *Proc. Natl. Acad. Sci. USA*, 113:E4161–E4169, 2016.
- [4888] Y. Zhang, K.H. Teoh, D.W. Reed, L. Maes, A. Goossens, D.J. Olson, A.R. Ross, and P.S. Covelto. The molecular cloning of artemisinic aldehyde $\Delta^{11(13)}$ reductase and its role in glandular trichome-dependent biosynthesis of artemisinin in *Artemisia annua*. *J. Biol. Chem.*, 283:21501–21508, 2008.
- [4889] Y.W. Zhang, M.K. Tiwari, H. Gao, S.S. Dhiman, M. Jeya, and J.K. Lee. Cloning and characterization of a thermostable H₂O-forming NADH oxidase from *Lactobacillus rhamnosus*. *Enzyme Microb. Technol.*, 50:255–262, 2012.
- [4890] Z. Zhang and D.B. McCormick. Uptake and metabolism of *N*-(4'-pyridoxyl)amines by isolated rat liver cells. *Arch. Biochem. Biophys.*, 294:394–397, 1992.
- [4891] Z.H. Zhang, J.N. Barlow, J.E. Baldwin, and C.J. Schofield. Metal-catalyzed oxidation and mutagenesis studies on the iron(II) binding site of 1-aminocyclopropane-1-carboxylate oxidase. *Biochemistry*, 36:15999–16007, 1997.
- [4892] Z.H. Zhang, J.S. Ren, D.K. Stammers, J.E. Baldwin, K. Harlos, and C.J. Schofield. Structural origins of the selectivity of the trifunctional oxygenase clavaminic acid synthase. *Nat. Struct. Biol.*, 7:127–133, 2000.
- [4893] Z.H. Zhang, C.J. Schofield, J.E. Baldwin, P. Thomas, and P. John. Expression, purification and characterization of 1-aminocyclopropane-1-carboxylate oxidase from tomato in *Escherichia coli*. *Biochem. J.*, 307:77–85, 1995.
- [4894] B. Zhao, F.P. Guengerich, A. Bellamine, D.C. Lamb, M. Izumikawa, L. Lei, L.M. Podust, M. Sundaramoorthy, J.A. Kalaitzis, L.M. Reddy, S.L. Kelly, B.S. Moore, D. Stec, M. Voehler, J.R. Falck, T. Shimada, and M.R. Waterman. Binding of two flavin substrate molecules, oxidative coupling, and crystal structure of *Streptomyces coelicolor* A3(2) cytochrome P450 158A2. *J. Biol. Chem.*, 280:11599–11607, 2005.
- [4895] B. Zhao, F.P. Guengerich, M. Voehler, and M.R. Waterman. Role of active site water molecules and substrate hydroxyl groups in oxygen activation by cytochrome P450 158A2: a new mechanism of proton transfer. *J. Biol. Chem.*, 280:42188–42197, 2005.
- [4896] B. Zhao, D.C. Lamb, L. Lei, S.L. Kelly, H. Yuan, D.L. Hachey, and M.R. Waterman. Different binding modes of two flavin substrate molecules in cytochrome P450 158A1 (CYP158A1) compared to CYP158A2. *Biochemistry*, 46:8725–8733, 2007.
- [4897] B. Zhao, X. Lin, L. Lei, D.C. Lamb, S.L. Kelly, M.R. Waterman, and D.E. Cane. Biosynthesis of the sesquiterpene antibiotic albaflavenone in *Streptomyces coelicolor* A3(2). *J. Biol. Chem.*, 283:8183–8189, 2008.
- [4898] G. Zhao, R.C. Bruckner, and M.S. Jorns. Identification of the oxygen activation site in monomeric sarcosine oxidase: role of Lys²⁶⁵ in catalysis. *Biochemistry*, 47:9124–9135, 2008.
- [4899] G. Zhao, A.J. Pease, N. Bharani, and M.E. Winkler. Biochemical characterization of gapB-encoded erythrose 4-phosphate dehydrogenase of *Escherichia coli* K-12 and its possible role in pyridoxal 5'-phosphate biosynthesis. *J. Bacteriol.*, 177:2804–2812, 1995.

- [4900] G. Zhao and M.E. Winkler. A novel α -ketoglutarate reductase activity of the *serA*-encoded 3-phosphoglycerate dehydrogenase of *Escherichia coli* K-12 and its possible implications for human 2-hydroxyglutaric aciduria. *J. Bacteriol.*, 178:232–239, 1996.
- [4901] P. Zhao, K. Inoue, I. Kouno, and H. Yamamoto. Characterization of leachianone G 2''-dimethylallyltransferase, a novel prenyl side-chain elongation enzyme for the formation of the lavandulyl group of sophoraflavanone G in *Sophora flavescens* Ait. cell suspension cultures. *Plant Physiol.*, 133:1306–1313, 2003.
- [4902] Q. Zhao, X.L. Yang, W.D. Holtzclaw, and P. Talalay. Unexpected genetic and structural relationships of a long-forgotten flavoenzyme to NAD(P)H:quinone reductase (DT-diaphorase). *Proc. Natl. Acad. Sci. USA*, 94:1669–1674, 1997.
- [4903] S. Zhao, Z. Lin, W. Ma, D. Luo, and Q. Cheng. Cloning and characterization of long-chain fatty alcohol oxidase LjFAO1 in *Lotus japonicus*. *Biotechnol. Prog.*, 24:773–779, 2008.
- [4904] X.-J. Zhao, T. Kawashiro, and T. Ishizaki. Mutual inhibition between quinine and etoposide by human liver microsomes. Evidence for cytochrome P4503A4 involvement in their major metabolic pathways. *Drug Metab. Dispos.*, 26:188–191, 1998.
- [4905] X.-J. Zhao, H. Yokoyama, K. Chiba, S. Wanwimolruk, and T. Ishizaki. Identification of human cytochrome P_{450} isoforms involved in the 3-hydroxylation of quinine by human liver microsomes and nine recombinant human cytochromes P_{450} . *J. Pharmacol. Exp. Ther.*, 279:1327–1334, 1996.
- [4906] Y. Zhao. Auxin biosynthesis: a simple two-step pathway converts tryptophan to indole-3-acetic acid in plants. *Mol. Plant*, 5:334–338, 2012.
- [4907] Y. Zhao, A.K. Hull, N.R. Gupta, K.A. Goss, J. Alonso, J.R. Ecker, J. Normanly, J. Chory, and J.L. Celenza. Trp-dependent auxin biosynthesis in *Arabidopsis*: involvement of cytochrome P_{450} s CYP79B2 and CYP79B3. *Genes Dev.*, 16:3100–3112, 2002.
- [4908] G. Zheng, J.A. Dahl, Y. Niu, P. Fedorcsak, C.M. Huang, C.J. Li, C.B. Vagbo, Y. Shi, W.L. Wang, S.H. Song, Z. Lu, R.P. Bosmans, Q. Dai, Y.J. Hao, X. Yang, W.M. Zhao, W.M. Tong, X.J. Wang, F. Bogdan, K. Furu, Y. Fu, G. Jia, X. Zhao, J. Liu, H.E. Krokan, A. Klungland, Y.G. Yang, and C. He. ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. *Mol. Cell*, 49:18–29, 2013.
- [4909] H. Zheng, O. Rowland, and L. Kunst. Disruptions of the *Arabidopsis* enoyl-CoA reductase gene reveal an essential role for very-long-chain fatty acid synthesis in cell expansion during plant morphogenesis. *Plant Cell*, 17:1467–1481, 2005.
- [4910] Y.J. Zhong, J.C. Huang, J. Liu, Y. Li, Y. Jiang, Z.F. Xu, G. Sandmann, and F. Chen. Functional characterization of various algal carotenoid ketolases reveals that ketolating zeaxanthin efficiently is essential for high production of astaxanthin in transgenic *Arabidopsis*. *J. Exp. Bot.*, 62:3659–3669, 2011.
- [4911] J. Zhou, M. Gunsior, B.O. Bachmann, C.A. Townsend, and E.I. Solomon. Substrate binding to the α -ketoglutarate-dependent non-heme iron enzyme clavaminic synthase 2: Coupling mechanism of oxidative decarboxylation and hydroxylation. *J. Am. Chem. Soc.*, 120:13539–13540, 1998.
- [4912] J. Zhou, S.S. Kang, P.W. Wong, B. Fournier, and R. Rozen. Purification and characterization of methylenetetrahydrofolate reductase from human cadaver liver. *Biochem Med Metab Biol*, 43:234–242, 1990.
- [4913] J. Zhou, W.L. Kelly, B.O. Bachmann, M. Gunsior, C.A. Townsend, and E.I. Solomon. Spectroscopic studies of substrate interactions with clavaminic synthase 2, a multifunctional α -KG-dependent non-heme iron enzyme: Correlation with mechanisms and reactivities. *J. Am. Chem. Soc.*, 123:7388–7398, 2001.
- [4914] N.Y. Zhou, J. Al-Dulayymi, M.S. Baird, and P.A. Williams. Salicylate 5-hydroxylase from *Ralstonia* sp. strain U2: a monooxygenase with close relationships to and shared electron transport proteins with naphthalene dioxygenase. *J. Bacteriol.*, 184:1547–1555, 2002.
- [4915] N.Y. Zhou, A. Jenkins, C.K.N.C.K. Chion, and D.J. Leak. The alkene monooxygenase from *Xanthobacter* strain Py2 is closely related to aromatic monooxygenases and catalyzes aromatic monohydroxylation of benzene, toluene, and phenol. *Appl. Environ. Microbiol.*, 65:1589–1595, 1999.
- [4916] R. Zhou and J.E. Linz. Enzymatic function of the nor-1 protein in aflatoxin biosynthesis in *Aspergillus parasiticus*. *Appl. Environ. Microbiol.*, 65:5639–5641, 1999.

- [4917] X.R. Zhou, S.S. Robert, J.R. Petrie, D.M. Frampton, M.P. Mansour, S.I. Blackburn, P.D. Nichols, A.G. Green, and S.P. Singh. Isolation and characterization of genes from the marine microalga *Pavlova salina* encoding three front-end desaturases involved in docosahexaenoic acid biosynthesis. *Phytochemistry*, 68:785–796, 2007.
- [4918] Y. Zhou, Y. Wei, A.N. Nanjaraj Urs, L. Lin, T. Xu, Y. Hu, E.L. Ang, H. Zhao, Z. Yuchi, and Y. Zhang. Identification and characterization of a new sulfoacetaldehyde reductase from the human gut bacterium *Bifidobacterium kashiwanohense*. *Biosci. Rep.*, 39, 2019.
- [4919] C. Zhu, S. Yamamura, M. Nishihara, H. Koiwa, and G. Sandmann. cDNAs for the synthesis of cyclic carotenoids in petals of *Gentiana lutea* and their regulation during flower development. *Biochim. Biophys. Acta*, 1625:305–308, 2003.
- [4920] D. Zhu, M.J. Seo, H. Ikeda, and D.E. Cane. Genome mining in streptomyces. Discovery of an unprecedented P450-catalyzed oxidative rearrangement that is the final step in the biosynthesis of pentalenolactone. *J. Am. Chem. Soc.*, 133:2128–2131, 2011.
- [4921] J. Zhu and H.F. DeLuca. Vitamin D 25-hydroxylase - Four decades of searching, are we there yet? *Arch. Biochem. Biophys.*, 523:30–36, 2012.
- [4922] J. Zhu, G.M. Lippa, A.M. Gulick, and P.A. Tipton. Examining reaction specificity in PvcB, a source of diversity in isonitrile-containing natural products. *Biochemistry*, 54:2659–2669, 2015.
- [4923] Q. Zhu, M.L. Hillwig, Y. Doi, and X. Liu. Aliphatic halogenase enables late-stage C-H functionalization: selective synthesis of a brominated fischerindole alkaloid with enhanced antibacterial activity. *ChemBioChem*, 17:466–470, 2016.
- [4924] T. Zhu, X. Cheng, Y. Liu, Z. Deng, and D. You. Deciphering and engineering of the final step halogenase for improved chlortetracycline biosynthesis in industrial *Streptomyces aureofaciens*. *Metab. Eng.*, 19:69–78, 2013.
- [4925] X. Zhu, W. De Laurentis, K. Leang, J. Herrmann, K. Ihlefeld, K.H. van Pee, and J.H. Naismith. Structural insights into regioselectivity in the enzymatic chlorination of tryptophan. *J. Mol. Biol.*, 391:74–85, 2009.
- [4926] X. Zhu, J. Liu, and W. Zhang. *De novo* biosynthesis of terminal alkyne-labeled natural products. *Nat. Chem. Biol.*, 11:115–120, 2015.
- [4927] X. Zhu, M. Su, K. Manickam, and W. Zhang. Bacterial genome mining of enzymatic tools for alkyne biosynthesis. *ACS Chem. Biol.*, 10:2785–2793, 2015.
- [4928] Y. Zhu, E. Jameson, M. Crosatti, H. Schafer, K. Rajakumar, T.D. Bugg, and Y. Chen. Carnitine metabolism to trimethylamine by an unusual Rieske-type oxygenase from human microbiota. *Proc. Natl. Acad. Sci. USA*, 111:4268–4273, 2014.
- [4929] J. Zi and R.J. Peters. Characterization of CYP76AH4 clarifies phenolic diterpenoid biosynthesis in the Lamiaceae. *Org. Biomol. Chem.*, 11:7650–7652, 2013.
- [4930] D.M. Ziegler and F.H. Pettit. Microsomal oxidases. I. The isolation and dialkylarylamine oxygenase activity of pork liver microsomes. *Biochemistry*, 5:2932–2938, 1966.
- [4931] G.A. Ziegler, C. Vornhein, I. Hanukoglu, and G.E. Schulz. The structure of adrenodoxin reductase of mitochondrial P450 systems: electron transfer for steroid biosynthesis. *J. Mol. Biol.*, 289:981–990, 1999.
- [4932] D.C. Zimmerman. Specificity of flaxseed lipoxidase. *Lipids*, 5:392–397, 1970.
- [4933] M.W. Zink and B.D. Sanwal. The distribution and substrate specificity of L-leucine dehydrogenase. *Arch. Biochem. Biophys.*, 99:72–77, 1962.
- [4934] C. Zirngibl, R. Hedderich, and R.K. Thauer. N^5, N^{10} -Methylenetetrahydromethanopterin dehydrogenase from *Methanobacterium thermoautotrophicum* has hydrogenase activity. *FEBS Lett.*, 261:112–116, 1990.
- [4935] G. Zocher, R. Winkler, C. Hertweck, and G.E. Schulz. Structure and action of the *N*-oxygenase AurF from *Streptomyces thioluteus*. *J. Mol. Biol.*, 373:65–74, 2007.
- [4936] M. Zolli, D.J. Kobric, and E.D. Brown. Reduction precedes cytidylyl transfer without substrate channeling in distinct active sites of the bifunctional CDP-ribitol synthase from *Haemophilus influenzae*. *Biochemistry*, 40:5041–5048, 2001.

- [4937] A. Zöphel, M.C. Kennedy, H. Beinert, and P.M.H. Kroneck. Investigations on microbial sulfur respiration. 1. Activation and reduction of elemental sulfur in several strains of Eubacteria. *Arch. Microbiol.*, 150:72–77, 1988.
- [4938] C. Zubietta, R. Joseph, S.S. Krishna, D. McMullan, M. Kapoor, H.L. Axelrod, M.D. Miller, P. Abdubek, C. Acosta, T. Astakhova, D. Carlton, H.J. Chiu, T. Clayton, M.C. Deller, L. Duan, Y. Elias, M.A. Elsliger, J. Feuerhelm, S.K. Grzechnik, J. Hale, G.W. Han, L. Jaroszewski, K.K. Jin, H.E. Klock, M.W. Knuth, P. Kozbial, A. Kumar, D. Marciano, A.T. Morse, K.D. Murphy, E. Nigoghossian, L. Okach, S. Oommachen, R. Reyes, C.L. Rife, P. Schimmel, C.V. Trout, H. van den Bedem, D. Weekes, A. White, Q. Xu, K.O. Hodgson, J. Wooley, A.M. Deacon, A. Godzik, S.A. Lesley, and I.A. Wilson. Identification and structural characterization of heme binding in a novel dye-decolorizing peroxidase, TyrA. *Proteins*, 69:234–243, 2007.
- [4939] D. Zucchini, G. Caprini, R.J. Pasterkamp, G. Tedeschi, and M.A. Vanoni. Kinetic and spectroscopic characterization of the putative monooxygenase domain of human MICAL-1. *Arch. Biochem. Biophys.*, 515:1–13, 2011.
- [4940] M.H.J. Zuidweg. Hydroxylation of Reichstein's compound S with cell-free preparations from *Curvularia lunata*. *Biochim. Biophys. Acta*, 152:144–158, 1968.
- [4941] W.G. Zumft. Cell biology and molecular basis of denitrification. *Microbiol. Mol. Biol. Rev.*, 61:533–616, 1997.
- [4942] W.G. Zumft and P.M. Kroneck. Respiratory transformation of nitrous oxide (N₂O) to dinitrogen by bacteria and archaea. *Adv. Microb. Physiol.*, 52:107–227, 2007.
- [4943] W.G. Zumft and L.E. Mortenson. The nitrogen-fixing complex of bacteria. *Biochim. Biophys. Acta*, 416:1–52, 1975.
- [4944] W.G. Zumft, A. Paneque, P.J. Aparicio, and M. Losada. Mechanism of nitrate reduction in *Chlorella*. *Biochem. Biophys. Res. Commun.*, 36:980–986, 1969.
- [4945] D. Zweytick, , and C. , Kohlwein. S.D. and Daum, G. Biochemical characterization and subcellular localization of the sterol C-24(28) reductase, erg4p, from the yeast *Saccharomyces cerevisiae*. *FEBS Lett.*, 470:83–87, 2000.

Index

- Δ^{12} acyl-lipid conjugase (11*E*,13*E*-forming), 487
 Δ^4 -3-oxosteroid 5 β -reductase, 159
 α -*N*-dichloroacetyl-*p*-aminophenylserinol *N*-oxygenase, 517
(-)-4'-demethyl-deoxypodophyllotoxin 4-hydroxylase, 443
(-)-*endo*-fenchol dehydrogenase, 70
(13*S*,14*R*)-13-*O*-acetyl-1-hydroxy-*N*-methylcanadine 8-hydroxylase, 453
(2*R*)-3-sulfolactate dehydrogenase (NADP⁺), 74
(2*S*)-[(*R*)-hydroxy(phenyl)methyl]succinyl-CoA dehydrogenase, 96
(3*R*)-2'-hydroxyisoflavanone reductase, 76
(3*S*)-3-amino-3-(3-chloro-4-hydroxyphenyl)propanoyl-[peptidyl-carrier protein SgcC2] monooxygenase, 409
(3*S*,4*R*)-3,4-dihydroxycyclohexa-1,5-diene-1,4-dicarboxylate dehydrogenase, 170
(*E*)-2-methylbutanal oxime monooxygenase, 417
(*E*)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase (ferredoxin), 530
(*E*)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase (flavodoxin), 531
(*R*)-2-hydroxy-fatty-acid dehydrogenase, 22
(*R*)-2-hydroxyglutarate—pyruvate transhydrogenase, 124
(*R*)-3-[(carboxymethyl)amino]fatty acid dioxygenase/decarboxylase, 359
(*R*)-3-hydroxyacid-ester dehydrogenase, 59
(*R*)-6-hydroxynicotine oxidase, 241
(*S*)-1-hydroxy-*N*-methylcanadine 13-hydroxylase, 452
(*S*)-1-phenylethanol dehydrogenase, 68
(*S*)-2-hydroxy-acid oxidase, 103
(*S*)-2-hydroxy-fatty-acid dehydrogenase, 22
(*S*)-2-hydroxyglutarate dehydrogenase, 114
(*S*)-2-hydroxypropylphosphonic acid epoxidase, 302
(*S*)-3-hydroxyacid-ester dehydrogenase, 59
(*S*)-6-hydroxynicotine oxidase, 241
(*S*)-8-oxocitronellyl enol synthase, 185
(*S*)-*N*-methylcanadine 1-hydroxylase, 453
(+)-*trans*-carveol dehydrogenase, 58
(+)-abscisic acid 8'-hydroxylase, 444
(+)-borneol dehydrogenase, 42
(+)-camphor 6-*endo*-hydroxylase, 461
(+)-camphor 6-*exo*-hydroxylase, 388
(+)-lariciresinol reductase, 551
(+)-larreatricin hydroxylase, 512
(+)-menthofuran synthase, 446
(+)-neomenthol dehydrogenase, 44
(+)-pinoselinol hydroxylase, 532
(+)-pinoselinol reductase, 551
(+)-piperitol/(+)-sesamin synthase, 499
(+)-pulegone reductase, 175
(+)-sabinene 3-hydroxylase, 387
(+)-sabinol dehydrogenase, 49
(+)-thujan-3-ol dehydrogenase, 70
(-)-borneol dehydrogenase, 49
(-)-isopiperitenone reductase, 175
(-)-menthol dehydrogenase, 44
(-)-menthol monooxygenase, 374
[50S ribosomal protein L16]-arginine 3-hydroxylase, 352
Botryococcus squalene synthase, 179
N-[(2*S*)-2-amino-2-carboxyethyl]-L-glutamate dehydrogenase, 238
all-trans- ζ -carotene desaturase, 207
all-trans-10'-apo- β -carotenal 13,14-cleaving dioxygenase, 328
all-trans-8'-apo- β -carotenal 15,15'-oxygenase, 329
all-trans-retinol 13,14-reductase, 206
all-trans-retinol 3,4-desaturase, 493
all-trans-retinol dehydrogenase (NAD⁺), 24
cis-1,2-dihydro-1,2-dihydroxynaphthalene dehydrogenase, 165
cis-1,2-dihydrobenzene-1,2-diol dehydrogenase, 163
cis-1,2-dihydroxy-4-methylcyclohexa-3,5-diene-1-carboxylate dehydrogenase, 172
cis-2,3-dihydrobiphenyl-2,3-diol dehydrogenase, 170
cis-2-enoyl-CoA reductase (NADPH), 166
cis-3,4-dihydrophenanthrene-3,4-diol dehydrogenase, 169
sn-1 acyl-lipid ω -3 desaturase (ferredoxin), 488
sn-1 linoleoyl-lipid 6-desaturase, 491
sn-1 oleoyl-lipid 12-desaturase, 491
sn-1 stearoyl-lipid 9-desaturase, 486
sn-2 acyl-lipid ω -3 desaturase (ferredoxin), 488
sn-2 palmitoyl-lipid 9-desaturase, 485
trans-1,2-dihydrobenzene-1,2-diol dehydrogenase, 163
trans-2-enoyl-CoA reductase (NAD⁺), 168
trans-2-enoyl-CoA reductase (NADPH), 167
trans-4-coumaroyl-CoA 2-hydroxylase, 355
D-2-hydroxyacid dehydrogenase (NAD⁺), 76
D-2-hydroxyacid dehydrogenase (NADP⁺), 58
D-2-hydroxyacid dehydrogenase (quinone), 113
D-2-hydroxyglutarate dehydrogenase, 124
D-*chiro*-inositol 1-dehydrogenase, 81
D-*threo*-aldose 1-dehydrogenase, 27
L-2-aminoadipate reductase, 145
L-2-hydroxycarboxylate dehydrogenase (NAD⁺), 74
L-2-hydroxycarboxylate dehydrogenase [NAD(P)⁺], 82
L-2-hydroxyglutarate dehydrogenase, 117
L-*erythro*-3,5-diaminohexanoate dehydrogenase, 213
11-*cis*-retinol dehydrogenase, 68
15-*cis*-phytoene desaturase, 192
2-(*R*)-hydroxypropyl-CoM dehydrogenase, 57
2-(*S*)-hydroxypropyl-CoM dehydrogenase, 57
3 α (or 20 β)-hydroxysteroid dehydrogenase, 13
3 β (or 20 α)-hydroxysteroid dehydrogenase, 45
3(or 17) α -hydroxysteroid dehydrogenase, 45
3(or 17) β -hydroxysteroid dehydrogenase, 13
3-(3-hydroxyphenyl)propanoate hydroxylase, 384
3-(*cis*-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate dehydrogenase, 177
3-[(*E*)-2-isocyanoethenyl]-1*H*-indole synthase, 504

3-[(Z)-2-isocyanoethenyl]-1H-indole synthase, 504
 3-*aci*-nitropropanoate oxidase, 269
 4-(γ -L-glutamylamino)butanoyl-[BtrI acyl-carrier protein] monooxygenase, 408
 4-(γ -glutamylamino)butanal dehydrogenase, 146
 5-*O*-(4-coumaroyl)-D-quininate 3'-monooxygenase, 433
 5-*exo*-hydroxycamphor dehydrogenase, 71
 5a,11a-dehydrotetracycline 5-monooxygenase, 400
 5a,11a-dehydrotetracycline reductase, 202
 6-*endo*-hydroxycineole dehydrogenase, 52
 8-*epi*-inunolide synthase, 454
 9,9'-*dicis*- ζ -carotene desaturase, 192
 9-*cis*- β -carotene 9',10'-cleaving dioxygenase, 327
 9-*cis*-epoxycarotenoid dioxygenase, 323

 A-factor type γ -butyrolactone 1'-reductase (1*S*-forming), 91
 abieta-7,13-dien-18-al dehydrogenase, 140
 abieta-7,13-dien-18-ol hydroxylase, 446
 abieta-7,13-diene hydroxylase, 446
 abscisic-aldehyde oxidase, 151
trans-acenaphthene-1,2-diol dehydrogenase, 292
 acetaldehyde dehydrogenase (acetylating), 127
 acetoacetyl-CoA reductase, 9
 acetone monooxygenase (methyl acetate-forming), 398
N-acetyl- γ -glutamyl-phosphate reductase, 133
 acetylacetone-cleaving enzyme, 322
N-acetylhexosamine 1-dehydrogenase, 51
 acetyloxidase, 269
*N*¹-acetylpolyamine oxidase, 242
*N*⁸-acetylspermidine oxidase (propane-1,3-diamine-forming), 243
 acireductone dioxygenase [iron(II)-requiring], 324
 acireductone dioxygenase (Ni²⁺-requiring), 323
 aclacinomycin-A oxidase, 190
 aclacinomycin-N oxidase, 109
 acrylate reductase, 186
 acrylyl-CoA reductase (NADH), 179
 acrylyl-CoA reductase (NADPH), 176
 acyl-[acyl-carrier-protein] 4-desaturase, 481
 acyl-[acyl-carrier-protein] 6-desaturase, 485
 acyl-CoA 11-(*E*)-desaturase, 484
 acyl-CoA 11-(*Z*)-desaturase, 479
 acyl-CoA 15-desaturase, 481
 acyl-CoA 5-desaturase, 489
 acyl-CoA 6-desaturase, 478
 acyl-CoA (8-3)-desaturase, 490
 acyl-CoA (9+3)-desaturase, 479
 acyl-CoA dehydrogenase (NADP⁺), 160
 acyl-CoA oxidase, 188
 acyl-lipid Δ^{12} -acetylenase, 489
 acyl-lipid Δ^6 -acetylenase, 489
 acyl-lipid ω -3 desaturase (cytochrome *b*₅), 485
 acyl-lipid ω -6 desaturase (cytochrome *b*₅), 484
 acyl-lipid ω -(9-4) desaturase, 481
 acyl-lipid (11-3)-desaturase, 479
 acyl-lipid (7-3)-desaturase, 487
 acyl-lipid (8-3)-desaturase, 486
 acyl-lipid (9+3)-(*E*)-desaturase, 487
 acyl-lipid (9-3)-desaturase, 491
 acyl-lipid (n+3)-(Z)-desaturase (ferredoxin), 484
 acylglycerone-phosphate reductase, 23
N-acylhexosamine oxidase, 106
N-acylmannosamine 1-dehydrogenase, 50
 adenylyl-sulfate reductase, 290
 adenylyl-sulfate reductase (glutathione), 282
 adenylyl-sulfate reductase (thioredoxin), 282
 AdoMet-dependent heme synthase, 203
 adrenodoxin-NADP⁺ reductase, 538
 aerobic 5,6-dimethylbenzimidazole synthase, 330
 aerobic carbon monoxide dehydrogenase, 154
 aflatoxin B synthase, 439
 agroclavine dehydrogenase, 237
 aklaviketone reductase, 80
 aklavinone 12-hydroxylase, 391
 alanine dehydrogenase, 211
 β -alanopine dehydrogenase, 232
 alanopine dehydrogenase, 230
 albendazole monooxygenase, 371
 albendazole monooxygenase (hydroxylating), 427
 albendazole monooxygenase (sulfoxide-forming), 427
 albonoursin synthase, 189
 alcohol dehydrogenase, 1
 alcohol dehydrogenase (azurin), 114
 alcohol dehydrogenase (cytochrome *c*), 99
 alcohol dehydrogenase (NADP⁺), 1
 alcohol dehydrogenase [NAD(P)⁺], 17
 alcohol dehydrogenase (nicotinoprotein), 123
 alcohol dehydrogenase (quinone), 112
 alcohol oxidase, 103
 alcohol-forming fatty acyl-CoA reductase, 142
 aldehyde dehydrogenase (FAD-independent), 158
 aldehyde dehydrogenase (NAD⁺), 125
 aldehyde dehydrogenase (NADP⁺), 126
 aldehyde dehydrogenase [NAD(P)⁺], 126
 aldehyde dehydrogenase (quinone), 153
 aldehyde ferredoxin oxidoreductase, 155
 aldehyde oxidase, 149
 alditol oxidase, 108
 aldose 1-dehydrogenase (NAD⁺), 27
 aldose 1-dehydrogenase [NAD(P)⁺], 79
 aldose reductase, 5
 aldose-6-phosphate reductase (NADPH), 43
 aliphatic glucosinolate *S*-oxygenase, 401
 alkan-1-ol dehydrogenase (acceptor), 120
 alkane 1-monooxygenase, 459
 alkanesulfonate monooxygenase, 406
 (4-alkanoyl-5-oxo-2,5-dihydrofuran-3-yl)methyl phosphate reductase, 183
 2-alkenal reductase (NADP⁺), 180
 2-alkenal reductase [NAD(P)⁺], 173
 alkene monooxygenase, 377
 alkyl sulfatase, 359
 2-alkyl-3-oxoalkanoate reductase, 91

alkylglycerol monooxygenase, 471
N-alkylglycine oxidase, 244
 2-alkyn-1-ol dehydrogenase, 35
 allyl-alcohol dehydrogenase, 14
 D-altritol 5-dehydrogenase, 90
 D-amino acid dehydrogenase (quinone), 224
 2-amino-1-hydroxyethylphosphonate dioxygenase (glycine-forming), 330
 2-amino-4-deoxychorismate dehydrogenase, 201
 4-amino-4-deoxyprephenate dehydrogenase, 185
 3-amino-4-hydroxybenzoate 2-monooxygenase, 404
 2-amino-5-chlorophenol 1,6-dioxygenase, 329
 5-amino-6-(5-phosphoribosylamino)uracil reductase, 41
 4-amino-L-phenylalanyl-[CmlP-peptidyl-carrier-protein] 3-hydroxylase, 516
 L-amino-acid dehydrogenase, 212
 D-amino-acid oxidase, 219
 L-amino-acid oxidase, 218
 [amino-group carrier protein]-5-phospho-L-glutamate reductase, 148
 [amino-group carrier protein]-6-phospho-L-2-aminoadipate reductase, 147
 L-aminoadipate-semialdehyde dehydrogenase, 132
 2-aminobenzenesulfonate 2,3-dioxygenase, 363
 4-aminobenzoate 1-monooxygenase, 371
 4-aminobenzoate *N*-oxygenase, 517
 aminobutyraldehyde dehydrogenase, 129
 aminocyclopropanecarboxylate oxidase, 473
 2-aminoethylphosphonate dioxygenase, 351
 aminomuconate-semialdehyde dehydrogenase, 132
 2-aminophenol 1,6-dioxygenase, 329
o-aminophenol oxidase, 293
 (*R*)-aminopropanol dehydrogenase, 17
 5-aminosalicylate 1,2-dioxygenase, 332
 ammonia monooxygenase, 511
 amorpho-4,11-diene 12-monooxygenase, 438
tert-amyl alcohol desaturase, 492
 β -amyirin 11-oxidase, 449
 β -amyirin 16 α -hydroxylase, 454
 β -amyirin 16 β -monooxygenase, 424
 β -amyirin 24-hydroxylase, 444
 β -amyirin 28-monooxygenase, 441
 β -amyirin 6 β -monooxygenase, 424
 anaerobic carbon monoxide dehydrogenase, 155
 anaerobic magnesium-protoporphyrin IX monomethyl ester cyclase, 548
 androst-4-ene-3,17-dione monooxygenase, 507
 angelicin synthase, 447
 3,6-anhydro- α -L-galactose dehydrogenase, 144
 1,5-anhydro-D-fructose reductase, 55
 1,5-anhydro-D-fructose reductase (1,5-anhydro-D-mannitol-forming), 63
 anhydrotetracycline 6-monooxygenase, 372
 anthocyanidin reductase [(2*R*,3*R*)-flavan-3-ol-forming], 174
 anthocyanidin reductase [(2*S*)-flavan-3-ol-forming], 183
 anthocyanidin synthase, 502
 anthranilate 1,2-dioxygenase (deaminating, decarboxylating), 361
 anthranilate 3-monooxygenase (deaminating), 372
 anthranilate 3-monooxygenase (FAD), 407
 anthraniloyl-CoA monooxygenase, 373
 D-apionate oxidoisomerase, 93
 apiose 1-reductase, 25
 D-apiose dehydrogenase, 93
 8'-apo- β -carotenoid 14',13'-cleaving dioxygenase, 327
 β -apo-4'-carotenal oxygenase, 142
 8'-apo-carotenoid 13,14-cleaving dioxygenase, 331
 D-arabinitol 2-dehydrogenase, 53
 L-arabinitol 2-dehydrogenase, 4
 D-arabinitol 4-dehydrogenase, 3
 L-arabinitol 4-dehydrogenase, 4
 D-arabinitol dehydrogenase (NADP⁺), 61
 D-arabinono-1,4-lactone oxidase, 107
 L-arabinose 1-dehydrogenase, 12
 D-arabinose 1-dehydrogenase (NAD⁺), 26
 D-arabinose 1-dehydrogenase (NADP⁺), 95
 D-arabinose 1-dehydrogenase [NAD(P)⁺], 26
 L-arabinose 1-dehydrogenase [NAD(P)⁺], 83
 D-arabitol-phosphate dehydrogenase, 65
 arachidonate 12-lipoxygenase, 319
 arachidonate 15-lipoxygenase, 319
 arachidonate 5-lipoxygenase, 319
 arachidonate 8-lipoxygenase, 321
 aralkylamine dehydrogenase (azurin), 225
 arginine 2-monooxygenase, 334
 D-arginine dehydrogenase, 226
 L-arginine dehydrogenase, 216
 L-arginine hydroxylase, 350
 L-arginine oxidase, 223
 argenate dehydrogenase, 168
 argenate dehydrogenase (NADP⁺), 174
 argenate dehydrogenase [NAD(P)⁺], 175
 aromatase, 408
 aromatic 2-oxoacid reductase, 25
 aromatic aldoxime *N*-monooxygenase, 419
 arsenate reductase (azurin), 542
 arsenate reductase (cytochrome *c*), 541
 arsenate reductase (donor), 543
 arsenate reductase (glutathione/glutaredoxin), 541
 arsenate reductase (thioredoxin), 542
 artemisinic aldehyde $\Delta^{11(13)}$ -reductase, 178
 aryl-alcohol dehydrogenase, 20
 aryl-alcohol dehydrogenase (NADP⁺), 21
 aryl-alcohol oxidase, 101
 aryl-aldehyde dehydrogenase, 131
 aryl-aldehyde oxidase, 150
 (aryl)acrylate reductase, 201
 L-ascorbate oxidase, 293
 L-ascorbate peroxidase, 299
 L-asparagine hydroxylase, 349
 asparagusate reductase, 274
 L-aspartate *N*-monooxygenase (nitrosuccinate-forming), 404

aspartate dehydrogenase, 215
 D-aspartate oxidase, 218
 L-aspartate oxidase, 221
 aspartate-semialdehyde dehydrogenase, 127
 asperlicin C monooxygenase, 396
 assimilatory dimethylsulfide *S*-monooxygenase, 403
 assimilatory sulfite reductase (ferredoxin), 287
 assimilatory sulfite reductase (NADPH), 272
 aurachin B dehydrogenase, 87
 aurachin C monooxygenase/isomerase, 397
 aureusidin synthase, 545
 averantin hydroxylase, 439
 azobenzene reductase, 263

 bacterial luciferase, 406
 bacterial non-heme ferritin, 521
 bacterial sulfide:quinone reductase, 285
 bacteriochlorophyllide *a* dehydrogenase, 88
 bacteriochlorophyllide *c* C-7¹-hydroxylase, 533
 benzaldehyde dehydrogenase (NAD⁺), 131
 benzaldehyde dehydrogenase (NADP⁺), 126
 benzene 1,2-dioxygenase, 361
 benzil reductase [(*R*)-benzoin forming], 70
 benzil reductase [(*S*)-benzoin forming], 70
 benzoate 1,2-dioxygenase, 362
 benzoate 4-monooxygenase, 432
p-benzoquinone reductase (NADPH), 258
 benzoyl-CoA 2,3-epoxidase, 394
 benzoyl-CoA 3-monooxygenase, 376
 benzoyl-CoA reductase, 194
 benzyl-2-methyl-hydroxybutyrate dehydrogenase, 47
 benzylmalonyl-CoA dehydrogenase, 190
 (*R*)-benzylsuccinyl-CoA dehydrogenase, 197
 berbaminine synthase, 497
 berberine reductase, 233
 betaine reductase, 547
 betaine-aldehyde dehydrogenase, 126
 biflavin synthase, 498
 bilirubin oxidase, 188
 biliverdin reductase, 164
 biochanin-A reductase, 168
 biphenyl 2,3-dioxygenase, 364
 biphenyl-2,3-diol 1,2-dioxygenase, 320
 bis- γ -glutamylcystine reductase, 275
 2,3-bis-*O*-geranylgeranyl-*sn*-glycero-phospholipid reductase, 195
 2,3-bis-*O*-geranylgeranyl-*sn*-glycerol 1-phosphate reductase [NAD(P)H], 180
 botryococcene synthase, 179
 branched-chain α -keto acid dehydrogenase system, 130
 brassinolide synthase, 457
 brassinosteroid 6-oxygenase, 457
 bromide peroxidase, 301
 bursehernin 5'-monooxygenase, 443
 butanal dehydrogenase, 136
 butane monooxygenase (soluble), 399
 (*R,R*)-butanediol dehydrogenase, 2
 (*S,S*)-butanediol dehydrogenase, 17
 1-butanol dehydrogenase (cytochrome *c*), 100
 1-butanol dehydrogenase (quinone), 113
 butanoyl-CoA dehydrogenase complex (NAD⁺, ferredoxin), 182
tert-butyl alcohol monooxygenase, 399
 γ -butyrobetaine dioxygenase, 341

 C-19 steroid 1 α -hydroxylase, 463
 caffeate 3,4-dioxygenase, 317
 caffeine dehydrogenase, 530
 caffeoyl-CoA reductase, 182
 calcidiol 1-monooxygenase, 463
 calcium-regulated photoprotein, 339
 camalexin synthase, 493
 camphor 5-monooxygenase, 459
 (*S*)-canadine synthase, 498
 cannabidiolic acid synthase, 545
 (5*R*)-carbapenam-3-carboxylate synthase, 501
 carbazole 1,9a-dioxygenase, 365
 carbonyl reductase (NADPH), 39
 3-carboxyethylcatechol 2,3-dioxygenase, 315
*N*⁵-(carboxyethyl)ornithine synthase, 232
 carboxylate reductase, 158
 carboxylate reductase (NADP⁺), 131
 5-carboxymethyl-2-hydroxymuconic-semialdehyde dehydrogenase, 137
 carboxynorspermidine synthase, 236
 carlactone synthase, 328
 (*S*)-carnitine 3-dehydrogenase, 54
 carnitine 3-dehydrogenase, 24
 carnitine monooxygenase, 401
 carnosic acid synthase, 424
 β -carotene 15,15'-dioxygenase, 326
 β -carotene 3-hydroxylase, 465
 β -carotene 4-ketolase, 516
 carotenoid χ -ring synthase, 210
 carotenoid ϵ hydroxylase, 450
 carotenoid ϕ -ring synthase, 210
 carotenoid isomeroxygenase, 327
 carotenoid-9',10'-cleaving dioxygenase, 328
 carveol dehydrogenase, 52
 carvone reductase, 207
ent-cassa-12,15-diene 11-hydroxylase, 437
ent-cassadiene hydroxylase, 425
 catalase, 297
 DCPH peroxidase, 302
 catechol 1,2-dioxygenase, 312
 catechol 2,3-dioxygenase, 312
 catechol oxidase, 292
 catechol oxidase (dimerizing), 103
 CDP-4-dehydro-6-deoxyglucose reductase, 523
 CDP-abequose synthase, 75
 CDP-paratose synthase, 75
 cellobiose dehydrogenase (acceptor), 119
 chanoclavine-I aldehyde reductase, 180
 chanoclavine-I dehydrogenase, 72

(S)-cheilanthifoline synthase, 497
 chlorate reductase, 552
 chlordecone reductase, 48
 chloridazon-catechol dioxygenase, 320
 chloride peroxidase, 298
 chlorite O₂-lyase, 322
 7-chloro-L-tryptophan 6-halogenase, 496
 7-chloro-L-tryptophan oxidase, 223
 chloroacetanilide *N*-alkylformylase, 465
 chlorobenzene dihydrodiol dehydrogenase, 185
 chlorobenzene dioxygenase, 366
 4-chlorophenylacetate 3,4-dioxygenase, 362
 chlorophyllide *a* reductase, 196
 chlorophyll(ide) *b* reductase, 63
 chlorophyllide-*a* oxygenase, 383
 cholest-4-en-3-one 26-monooxygenase [(25*R*)-3-oxocholest-4-en-26-oate forming], 466
 cholest-4-en-3-one 26-monooxygenase [(25*S*)-3-oxocholest-4-en-26-oate forming], 467
 cholest-5-ene-3β,7α-diol 3β-dehydrogenase, 39
 5β-cholestane-3α,7α-diol 12α-hydroxylase, 445
 cholestanetriol 26-monooxygenase, 462
 cholesterol 24-hydroxylase, 412
 cholesterol 25-monooxygenase, 511
 cholesterol 7α-monooxygenase, 411
 cholesterol 7-desaturase, 483
 cholesterol monooxygenase (side-chain-cleaving), 460
 cholesterol oxidase, 101
 choline dehydrogenase, 117
 choline monooxygenase, 460
 choline oxidase, 104
 1,8-cineole 2-*endo*-monooxygenase, 443
 1,8-cineole 2-*exo*-monooxygenase, 422
trans-cinnamate 2-monooxygenase, 368
trans-cinnamate 4-monooxygenase, 432
 cinnamoyl-CoA reductase, 134
 cinnamyl-alcohol dehydrogenase, 42
 clavamate synthase, 345
 CMP-*N*-acetylneuraminate monooxygenase, 474
 CoA-disulfide reductase, 275
 CoA-glutathione reductase, 274
 cobalt-precorrin-6A reductase, 181
 codeine 3-*O*-demethylase, 348
 codeinone reductase (NADPH), 52
 coenzyme F₄₂₀ hydrogenase, 309
 coenzyme F₄₂₀ oxidoreductase (ferredoxin), 248
 coenzyme F₄₂₀:CoB-CoM heterodisulfide,ferredoxin reductase, 289
 coenzyme F₄₂₀:methanophenazine dehydrogenase, 250
 coenzyme F₄₂₀H₂ oxidase, 245
 [Co(II) methylated amine-specific corrinoid protein] reductase, 523
 columbamine oxidase, 544
 coniferyl-alcohol dehydrogenase, 41
 coniferyl-aldehyde dehydrogenase, 138
 coproporphyrinogen dehydrogenase, 202
 coproporphyrinogen III oxidase (coproporphyrin-forming), 190
 coproporphyrinogen oxidase, 187
 corticosterone 18-monooxygenase, 459
 (S)-corytuberine synthase, 492
 costunolide synthase, 448
 2-coumarate reductase, 161
 crocetin dialdehyde synthase, 331
 crotonobetainyl-CoA reductase, 200
 crotonyl-CoA carboxylase/reductase, 176
 crotonyl-CoA reductase, 176
 cucurbitacin Δ²³-reductase, 160
 cucurbitadienol 11-hydroxylase, 426
p-cumate 2,3-dioxygenase, 365
 cuproxidase, 522
 cyanocobalamin reductase, 519
 cyclic alcohol dehydrogenase (quinone), 112
 cyclic dehydropanthinyl futasoline synthase, 547
 cyclohex-1-ene-1-carbonyl-CoA dehydrogenase, 200
 cyclohexane-1,2-diol dehydrogenase, 37
 cyclohexane-1-carbonyl-CoA dehydrogenase (electron-transfer flavoprotein), 200
 cyclohexane-1-carbonyl-CoA reductase (NADP⁺), 185
 cyclohexanol dehydrogenase, 52
 cyclohexanone dehydrogenase, 205
 cyclohexanone monooxygenase, 369
 cyclohexylamine oxidase, 220
 cyclooctat-9-en-7-ol 5-monooxygenase, 515
 cyclooctatin synthase, 516
 (-)-cyclopinine synthase, 360
 cyclopentanol dehydrogenase, 35
 cyclopentanone monooxygenase, 369
 β-cyclopiasonate dehydrogenase, 549
p-cymene methyl-monooxygenase, 465
 cypemycin cysteine dehydrogenase (decarboxylating), 209
 cysteamine dioxygenase, 316
 cysteine dioxygenase, 317
 cysteine-type anaerobic sulfatase-maturing enzyme, 290
 L-cysteinyl-L-histidinylsulfoxide synthase, 513
 cystine reductase, 273
 cytochrome-*b*₅ reductase, 254
 cytochrome-*c* peroxidase, 297
 cytochrome-*c*₃ hydrogenase, 309
 cytokinin dehydrogenase, 251
 dammarenediol 12-hydroxylase, 440
 deacetoxycephalosporin-C hydroxylase, 346
 deacetoxycephalosporin-C synthase, 501
 deacetoxyvindoline 4-hydroxylase, 345
 3''-deamino-3''-oxonicotianamine reductase, 61
 decanoyl-[acyl-carrier protein] acetylenase, 500
 decaprenylphospho-β-D-*erythro*-pentofuranosid-2-ulose 2-reductase, 73
 decaprenylphospho-β-D-ribofuranose 2-dehydrogenase, 115
 dehydro coenzyme F₄₂₀ reductase, 201
 13,14-dehydro-15-oxoprostaglandin 13-reductase, 169
 2-dehydro-3-deoxy-D-gluconate 5-dehydrogenase, 28

2-dehydro-3-deoxy-D-gluconate 6-dehydrogenase, 28
 2-dehydro-3-deoxy-L-fuconate 4-dehydrogenase, 97
 2-dehydro-3-deoxy-L-galactonate 5-dehydrogenase, 86
 2-dehydro-3-deoxy-L-rhamnonate dehydrogenase (NAD⁺), 89
 5-dehydro-6-demethoxyfumagillol dioxygenase, 361
 5-dehydro-6-demethoxyfumagillol synthase, 458
 3-dehydro-L-gulonate 2-dehydrogenase, 28
 3-dehydro-bile acid $\Delta^{4,6}$ -reductase, 183
 3,4-dehydroadipyl-CoA semialdehyde dehydrogenase (NADP⁺), 141
 7-dehydrocholesterol reductase, 163
 5-dehydrofumagillol 5-reductase, 97
 dehydrogluconate dehydrogenase, 117
 6-dehydroglucose reductase, 96
 2'-dehydrokanamycin reductase, 78
 2-dehydropantoate 2-reductase, 36
 (R)-dehydropantoate dehydrogenase, 132
 2-dehydropantolactone reductase, 79
 2-dehydropantolactone reductase (Re-specific), 36
 2-dehydropantolactone reductase (Si-specific), 46
 3-dehydroquininate synthase II, 216
 1,2-dehydroreticuline synthase, 493
 1,2-dehydroreticulium reductase (NADPH), 232
 3-dehydrosphinganine reductase, 23
 3-demethoxyubiquinol 3-hydroxylase, 515
 demethylphyloquinone reductase, 259
 3-deoxy- α -D-manno-octulosonate 8-oxidase, 109
 1-deoxy-11 β -hydroxypentalenate dehydrogenase, 74
 2-deoxy-*scyllo*-inosamine dehydrogenase, 72
 2-deoxy-*scyllo*-inosamine dehydrogenase (AdoMet-dependent), 124
 2-deoxy-D-gluconate 3-dehydrogenase, 28
 1-deoxy-D-xylulose-5-phosphate reductoisomerase, 56
 7-deoxycylindrospermopsin hydroxylase, 357
 13-deoxydaunorubicin hydroxylase, 391
 6-deoxyerythronolide B hydroxylase, 469
 deoxyhypusine monooxygenase, 510
 7-deoxyloganate 7-hydroxylase, 430
 2'-deoxymugineic-acid 2'-dioxygenase, 346
 deoxynogalonate monooxygenase, 338
 1-deoxypentalenic acid 11 β -hydroxylase, 349
 (-)-deoxypodophyllotoxin synthase, 503
 deoxysarpagine hydroxylase, 444
 diacetyl reductase [(R)-acetoin forming], 66
 diacetyl reductase [(S)-acetoin forming], 66
 diamine oxidase, 223
 2,5-diamino-6-(ribosylamino)-4(3H)-pyrimidinone 5'-phosphate reductase, 65
 2,4-diaminopentanoate dehydrogenase, 213
 2,4-diaminopentanoate dehydrogenase (NAD⁺), 216
 diaminopimelate dehydrogenase, 214
 diapolycopene oxygenase, 511
 4,4'-diapolycopenoate synthase, 159
 4,4'-diapophytoene desaturase (4,4'-diapolycopene-forming), 197
 dibenzothiophene dihydrodiol dehydrogenase, 171
 dibenzothiophene monooxygenase, 411
 dibenzothiophene sulfone monooxygenase, 411
 dichloroarcyriaflavin A synthase, 337
 2,4-dichlorobenzoyl-CoA reductase, 543
 dichlorochromopyrrolate synthase, 548
 2,4-dichlorophenol 6-monooxygenase, 369
 (R)-dichlorprop dioxygenase (2-oxoglutarate), 351
 (S)-dichlorprop dioxygenase (2-oxoglutarate), 350
 2,5-didehydrogluconate reductase (2-dehydro-D-gluconate-forming), 58
 2,5-didehydrogluconate reductase (2-dehydro-L-gulonate-forming), 76
 2,4-dienoyl-CoA reductase [(2E)-enoyl-CoA-producing], 166
 2,4-dienoyl-CoA reductase [(3E)-enoyl-CoA-producing], 186
 diethyl 2-methyl-3-oxosuccinate reductase, 49
 diferric-transferrin reductase, 519
 2,3-dihydro-2,3-dihydroxybenzoate dehydrogenase, 165
 dihydroanticapsin dehydrogenase, 85
 dihydrobenzophenanthridine oxidase, 242
 15,16-dihydrobiliverdin:ferredoxin oxidoreductase, 193
 dihydrobunolol dehydrogenase, 35
 dihydrocarveol dehydrogenase, 64
 dihydroceramide fatty acyl 2-hydroxylase, 475
 dihydrochelirubine 12-monooxygenase, 435
cis-dihydroethylcatechol dehydrogenase, 172
 dihydroflavonol 4-reductase, 47
 dihydrofolate reductase, 227
 dihydrolipoyl dehydrogenase, 272
 β -dihydromenaquinone-9 ω -hydroxylase, 466
 dihydromethanophenazine:CoB-CoM heterodisulfide reductase, 288
 dihydromethanopterin reductase (acceptor), 252
 dihydromethanopterin reductase [NAD(P)⁺], 237
 dihydromonacolin L hydroxylase, 441
 dihydromonapterin reductase, 238
 7,8-dihydroneopterin oxygenase, 330
 dihydroorotate dehydrogenase (fumarate), 202
 dihydroorotate dehydrogenase (NAD⁺), 162
 dihydroorotate dehydrogenase (NADP⁺), 162
 dihydroorotate dehydrogenase (quinone), 191
 dihydrophenazinedicarboxylate synthase, 295
 6,7-dihydropteridine reductase, 234
 dihydropyrimidine dehydrogenase (NAD⁺), 159
 dihydropyrimidine dehydrogenase (NADP⁺), 159
 dihydrorhizobitoxine desaturase, 496
 dihydrosanguinarine 10-monooxygenase, 434
 dihydrouracil oxidase, 188
 1,2-dihydrovomilenine reductase, 173
 2,4-dihydroxy-1,4-benzoxazin-3-one-glucoside dioxygenase, 354
 2,3-dihydroxy-2,3-dihydro-*p*-cumate dehydrogenase, 171
 5,6-dihydroxy-3-methyl-2-oxo-1,2,5,6-tetrahydroquinoline dehydrogenase, 172
 1,2-dihydroxy-6-methylcyclohexa-3,5-dienecarboxylate dehydrogenase, 172
 3,4-dihydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione 4,5-dioxygenase, 317
 2,4'-dihydroxyacetophenone dioxygenase, 321

2,3-dihydroxybenzoate 2,3-dioxygenase, 318
 2,3-dihydroxybenzoate 3,4-dioxygenase, 315
 1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate dehydrogenase, 164
 2,3-dihydroxyindole 2,3-dioxygenase, 317
 7,8-dihydroxykynurenate 8,8a-dioxygenase, 314
 1,2-dihydroxynaphthalene dioxygenase, 324
 3,4-dihydroxyphenylacetate 2,3-dioxygenase, 315
 (3,5-dihydroxyphenyl)acetyl-CoA 1,2-dioxygenase, 330
 3,4-dihydroxyphenylalanine oxidative deaminase, 336
 3,9-dihydroxypterocarpan 6a-monooxygenase, 432
 2,6-dihydroxypyridine 3-monooxygenase, 368
 2,5-dihydroxypyridine 5,6-dioxygenase, 314
 3 β ,22 α -dihydroxysteroid 3-dehydrogenase, 501
 diiodophenylpyruvate reductase, 22
 2,5-diketocamphane 1,2-monooxygenase, 436
 3,6-diketocamphane 1,2-monooxygenase, 450
N,N-dimethyl phenylurea *N*-demethylase, 470
 dimethyl sulfide:cytochrome *c*₂ reductase, 278
 8-dimethylallylnaringenin 2'-hydroxylase, 446
 dimethylamine dehydrogenase, 248
 dimethylamine monooxygenase, 401
 4-(dimethylamino)phenylazoxybenzene reductase, 264
 dimethylglycine dehydrogenase, 249
 dimethylglycine oxidase, 241
N,N-dimethylglycine/sarcosine dehydrogenase (ferredoxin), 248
 dimethylmalate dehydrogenase, 19
 dimethylnonatriene synthase, 423
 dimethylsulfone monooxygenase, 415
 dimethylsulfone reductase, 276
 2,4-dinitrotoluene dioxygenase, 365
 dinoflagellate luciferase, 337
 2,5-dioxoalate dehydrogenase, 131
 dissimilatory dimethyl sulfide monooxygenase, 384
 dissimilatory dimethyldisulfide reductase, 277
 dissimilatory sulfite reductase, 290
S-disulfanyl-L-cysteine oxidoreductase, 278
 4,4'-dithiodibutanoate disulfide reductase, 276
 3,8-divinyl chlorophyllide *a* reductase, 196
 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (ferredoxin), 195
 3,8-divinyl protochlorophyllide *a* 8-vinyl-reductase (NADPH), 174
 DNA *N*⁶-methyladenine demethylase, 352
 DNA oxidative demethylase, 348
 docosahexaenoic acid ω -hydroxylase, 428
 dolabradiene monooxygenase, 451
 dopamine β -monooxygenase, 472
 drimenol monooxygenase, 426
 dTDP-3,4-didehydro-2,6-dideoxy- α -D-glucose 3-reductase, 85
 dTDP-4-dehydro-6-deoxy- α -D-gulose 4-ketoreductase, 80
 dTDP-4-dehydro-6-deoxyglucose reductase, 56
 dTDP-4-dehydrorhamnose reductase, 29
 dTDP-6-deoxy-L-talose 4-dehydrogenase (NAD⁺), 74
 dTDP-6-deoxy-L-talose 4-dehydrogenase (NADP⁺), 29
 dTDP-6-deoxy-L-talose 4-dehydrogenase [NAD(P)⁺], 75
 dTDP-galactose 6-dehydrogenase, 40
 dye decolorizing peroxidase, 301
 ecdysone 20-monooxygenase, 509
 ecdysone oxidase, 103
 ectoine hydroxylase, 353
 electron-transferring-flavoprotein dehydrogenase, 247
 endo-cleaving rubber dioxygenase, 332
 enduracididine β -hydroxylase, 350
 2-enoate reductase, 165
 enoyl-[acyl-carrier-protein] reductase (NADH), 161
 enoyl-[acyl-carrier-protein] reductase (NADPH), 181
 enoyl-[acyl-carrier-protein] reductase (NADPH, *Re*-specific), 167
 enoyl-[acyl-carrier-protein] reductase (NADPH, *Si*-specific), 161
 enzyme-thiol transhydrogenase (glutathione-disulfide), 281
 (1*R*,2*S*)-ephedrine 1-dehydrogenase, 94
 ephedrine dehydrogenase, 230
 5-epiaristolochene 1,3-dihydroxylase, 448
 epoxyqueuosine reductase, 535
 D-erythritol 1-phosphate dehydrogenase, 89
 erythromycin 12-hydroxylase, 387
 D-erythronate 2-dehydrogenase, 91
 erythrose-4-phosphate dehydrogenase, 139
 erythrose reductase, 35
 estradiol 17 α -dehydrogenase, 32
 17 β -estradiol 17-dehydrogenase, 15
 estradiol 6 β -monooxygenase, 507
 ethanolamine oxidase, 220
 ethylbenzene hydroxylase, 534
 ethylenediaminetetraacetate monooxygenase, 414
 eugenol synthase, 69
 eukaryotic sulfide quinone oxidoreductase, 286
 eupatolide synthase, 454
 L-evernosamine nitrososynthase, 392
 exo-cleaving rubber dioxygenase, 331
 F₄₂₀H₂:quinone oxidoreductase, 115
 F-actin monooxygenase, 398
 factor-independent urate hydroxylase, 269
 FAD reductase (NADH), 235
 FAD reductase [NAD(P)H], 237
 FAD-dependent urate hydroxylase, 382
 farnesal dehydrogenase, 145
 farnesoate epoxidase, 442
 farnesol dehydrogenase (NAD⁺), 78
 farnesol dehydrogenase (NADP⁺), 46
 farnesylcysteine lyase, 280
 fatty acid α -dioxygenase, 333
 fatty-acid peroxidase, 297
 fatty-acid peroxygenase, 306
 fenbendazole monooxygenase (4'-hydroxylating), 427
 ferredoxin hydrogenase, 309
 ferredoxin—NAD⁺ reductase, 537
 ferredoxin—NADP⁺ reductase, 537
 ferredoxin—NAD(P)⁺ reductase (naphthalene dioxygenase ferredoxin-specific), 538

ferredoxin—nitrate reductase, 271
 ferredoxin—nitrite reductase, 271
 ferredoxin:CoB-CoM heterodisulfide reductase, 287
 ferredoxin:protochlorophyllide reductase (ATP-dependent), 194
 ferredoxin:thioredoxin reductase, 287
 ferric-chelate reductase (NADH), 520
 ferric-chelate reductase (NADPH), 520
 ferric-chelate reductase [NAD(P)H], 521
 ferroxidase, 521
 ferruginol monooxygenase, 423
 ferruginol synthase, 456
 feruloyl-CoA 6-hydroxylase, 354
 festuclavine dehydrogenase, 237
 firefly luciferase, 335
 flavanoid 3',5'-hydroxylase, 429
 flavanone 2-hydroxylase, 452
 flavanone 3-dioxygenase, 343
 flavanone 4-reductase, 50
 flavin reductase (NADH), 234
 flavin reductase (NADPH), 233
 flavin-containing monooxygenase, 367
 flavodoxin—NADP⁺ reductase, 540
 flavone synthase I, 502
 flavone synthase II, 500
 flavonoid 3'-monooxygenase, 429
 flavonol synthase, 502
 fluoren-9-ol dehydrogenase, 54
 fluoroacetaldehyde dehydrogenase, 138
 FMN reductase (NADH), 236
 FMN reductase (NADPH), 235
 FMN reductase [NAD(P)H], 235
 FMN-dependent NADH-azoreductase, 266
 formaldehyde dehydrogenase, 134
 formaldehyde dismutase, 157
 formate dehydrogenase, 525
 formate dehydrogenase (coenzyme F₄₂₀), 533
 formate dehydrogenase (cytochrome), 148
 formate dehydrogenase (cytochrome-*c*-553), 527
 formate dehydrogenase (hydrogenase), 533
 formate dehydrogenase (NAD⁺, ferredoxin), 526
 formate dehydrogenase (NADP⁺), 526
 formate dehydrogenase-N, 530
 [formate-*C*-acetyltransferase]-activating enzyme, 553
 formate:CoB-CoM heterodisulfide,ferredoxin reductase, 289
 5-formyl-3-hydroxy-2-methylpyridine 4-carboxylic acid 5-dehydrogenase, 146
 4-formylbenzenesulfonate dehydrogenase, 137
 2-formylbenzoate dehydrogenase, 141
 formylglycine-generating enzyme, 280
 formylmethanofuran dehydrogenase, 157
 formyltetrahydrofolate dehydrogenase, 228
 fraxetin 5-hydroxylase, 452
 fructose 5-dehydrogenase, 114
 fructose 5-dehydrogenase (NADP⁺), 27
 fructosyl amine oxidase (fructosamine-forming), 246
 fructosyl amine oxidase (glucosone-forming), 245
 fructuronate reductase, 14
 L-fucose dehydrogenase, 97
 fumarate reductase (CoM/CoB), 191
 fumarate reductase (cytochrome), 187
 fumarate reductase (NADH), 160
 fumitremorgin C monooxygenase, 439
 fumitremorgin C synthase, 498
 2-furoyl-CoA dehydrogenase, 204
 galactitol 2-dehydrogenase, 4
 galactitol 2-dehydrogenase (L-tagatose-forming), 90
 galactitol-1-phosphate 5-dehydrogenase, 53
 L-galactonate 5-dehydrogenase, 92
 L-galactonolactone dehydrogenase, 187
 L-galactonolactone oxidase, 189
 D-galactose 1-dehydrogenase, 12
 L-galactose 1-dehydrogenase, 69
 galactose 1-dehydrogenase (NADP⁺), 27
 galactose oxidase, 102
 D-galacturonate reductase, 80
 gallate dioxygenase, 324
 GDP-4-dehydro-6-deoxy-D-mannose reductase, 60
 GDP-4-dehydro-D-rhamnose reductase, 40
 GDP-6-deoxy-D-talose 4-dehydrogenase, 30
 GDP-L-colitose synthase, 78
 GDP-L-fucose synthase, 58
 GDP-mannose 6-dehydrogenase, 29
 geissoschizine dehydrogenase, 166
 gentisate 1,2-dioxygenase, 313
 geranial dehydrogenase, 143
 geraniol 8-hydroxylase, 430
 geraniol dehydrogenase (NAD⁺), 76
 geraniol dehydrogenase (NADP⁺), 39
 geranylgeraniol 18-hydroxylase, 447
 geranylgeranyl diphosphate reductase, 176
 geranylgeranyl-bacteriochlorophyllide *a* reductase, 182
 geranylhydroquinone 3''-hydroxylase, 455
 germacrene A acid 8β-hydroxylase, 453
 germacrene A hydroxylase, 433
 gibberellin 2β-dioxygenase, 344
 gibberellin 3β-dioxygenase, 344
 gibberellin-44 dioxygenase, 344
 gluconate 2-dehydrogenase, 46
 gluconate 2-dehydrogenase (acceptor), 117
 gluconate 5-dehydrogenase, 16
 glucose 1-dehydrogenase (FAD, quinone), 113
 glucose 1-dehydrogenase (NAD⁺), 26
 glucose 1-dehydrogenase (NADP⁺), 26
 glucose 1-dehydrogenase [NAD(P)⁺], 12
 glucose 1-dehydrogenase (PQQ, quinone), 111
 glucose oxidase, 101
 glucose-6-phosphate 3-dehydrogenase, 79
 glucose-6-phosphate dehydrogenase (coenzyme-F₄₂₀), 115
 glucose-6-phosphate dehydrogenase (NAD⁺), 86
 glucose-6-phosphate dehydrogenase (NADP⁺), 12
 glucose-6-phosphate dehydrogenase [NAD(P)⁺], 80

glucose-fructose oxidoreductase, 121
 glucose/galactose 1-dehydrogenase, 79
 glucoside 3-dehydrogenase (acceptor), 119
 glucoside 3-dehydrogenase (cytochrome *c*), 100
 glucuronate reductase, 5
 glucuronolactone reductase, 5
 L-glutamate γ -semialdehyde dehydrogenase, 143
 L-glutamate 3(*R*)-hydroxylase, 359
 glutamate dehydrogenase, 211
 glutamate dehydrogenase (NADP⁺), 212
 glutamate dehydrogenase [NAD(P)⁺], 212
 D-glutamate oxidase, 220
 L-glutamate oxidase, 220
 glutamate synthase (ferredoxin), 225
 glutamate synthase (NADH), 214
 glutamate synthase (NADPH), 213
 glutamate-5-semialdehyde dehydrogenase, 133
 D-glutamate(D-aspartate) oxidase, 221
 γ -glutamyl mercynylcysteine *S*-oxide synthase, 513
 glutamyl-tRNA reductase, 138
 glutarate dioxygenase, 355
 glutarate-semialdehyde dehydrogenase, 129
 glutaredoxin-dependent peroxiredoxin, 303
 glutaryl-CoA dehydrogenase (acceptor), 209
 glutaryl-CoA dehydrogenase (ETF), 198
 glutathione amide reductase, 275
 glutathione amide-dependent peroxidase, 301
 glutathione dehydrogenase (ascorbate), 284
 glutathione oxidase, 279
 glutathione peroxidase, 298
 glutathione—CoA-glutathione transhydrogenase, 281
 glutathione—cystine transhydrogenase, 281
 glutathione—homocystine transhydrogenase, 280
 glutathione-dependent peroxiredoxin, 304
 glutathione-disulfide reductase, 273
 2-glutathionyl-2-methylbut-3-en-1-ol dehydrogenase, 88
 glutathionyl-hydroquinone reductase, 286
 glyceollin synthase, 444
 glyceraldehyde dehydrogenase (FAD-containing), 158
 D-glyceraldehyde dehydrogenase (NADP⁺), 144
 D/L-glyceraldehyde reductase, 82
 glyceraldehyde-3-phosphate dehydrogenase (arsenate-transferring), 148
 glyceraldehyde-3-phosphate dehydrogenase (ferredoxin), 155
 glyceraldehyde-3-phosphate dehydrogenase (NADP⁺), 127
 glyceraldehyde-3-phosphate dehydrogenase [NAD(P)⁺], 144
 glyceraldehyde-3-phosphate dehydrogenase (NAD(P)⁺) (phosphorylating), 136
 glyceraldehyde-3-phosphate dehydrogenase (NADP⁺) (phosphorylating), 128
 glyceraldehyde-3-phosphate dehydrogenase (phosphorylating), 127
 glycerate dehydrogenase, 7
 glycerol 2-dehydrogenase (NADP⁺), 34
 glycerol dehydrogenase, 2
 glycerol dehydrogenase (acceptor), 120
 glycerol dehydrogenase (NADP⁺), 17
sn-glycerol-1-phosphate dehydrogenase, 55
 glycerol-3-phosphate 1-dehydrogenase (NADP⁺), 38
 glycerol-3-phosphate dehydrogenase, 111
 glycerol-3-phosphate dehydrogenase (NAD⁺), 3
 glycerol-3-phosphate dehydrogenase [NAD(P)⁺], 21
 glycerol-3-phosphate oxidase, 105
 glycine betaine monooxygenase, 405
 glycine cleavage system, 217
 glycine dehydrogenase, 213
 glycine dehydrogenase (aminomethyl-transferring), 224
 glycine dehydrogenase (cyanide-forming), 226
 glycine dehydrogenase (cytochrome), 217
 glycine oxidase, 222
 glycine reductase, 546
 L-glycol dehydrogenase, 39
 glycolaldehyde dehydrogenase, 129
 glycolate dehydrogenase, 119
 C-glycoside oxidase, 110
 glyoxylate dehydrogenase (acylating), 128
 glyoxylate oxidase, 150
 glyoxylate reductase, 7
 glyoxylate reductase (NADP⁺), 18
 glyphosate oxidoreductase, 245
 GMP reductase, 264
 grixazone synthase, 295
 γ -guanidinobutyraldehyde dehydrogenase, 136
 L-gulonate 3-dehydrogenase, 12
 L-gulonate 5-dehydrogenase, 84
 L-gulonolactone oxidase, 102
 H₂:CoB-CoM heterodisulfide,ferredoxin reductase, 289
 2-haloacrylate reductase, 180
 2-halobenzoate 1,2-dioxygenase, 363
 hapalindole-type alkaloid chlorinase, 505
 heme *a* synthase, 535
 heme oxygenase (biliverdin-IX- β and δ -forming), 515
 heme oxygenase (biliverdin-producing), 410
 heme oxygenase (biliverdin-producing, ferredoxin), 464
 heme oxygenase (mycobilin-producing), 514
 heme oxygenase (staphylobilin-producing), 512
 3-heptyl-3-hydroxy-4(1*H*)-quinolone synthase, 391
 mercynylcysteine *S*-oxide synthase, 545
 hex-5-enoyl-[acyl-carrier protein] acetylenase, 489
 (1*Z*)-hexadec-11-enoyl-CoA conjugase, 482
 hexadecanal dehydrogenase (acylating), 133
 hexadecanol dehydrogenase, 35
 2-hexadecenal reductase, 164
 hexose oxidase, 101
 histidinol dehydrogenase, 6
 [histone H3]-*N*⁶,*N*⁶-dimethyl-L-lysine⁴ FAD-dependent demethylase, 517
 [histone H3]-dimethyl-L-lysine³⁶ demethylase, 347
 [histone H3]-dimethyl-L-lysine⁹ demethylase, 355
 [histone H3]-trimethyl-L-lysine²⁷ demethylase, 357
 [histone H3]-trimethyl-L-lysine³⁶ demethylase, 357

[histone H3]-trimethyl-L-lysine⁴ demethylase, 356
 [histone H3]-trimethyl-L-lysine⁹ demethylase, 356
 homogentisate 1,2-dioxygenase, 313
 homoisocitrate dehydrogenase, 20
 homomethionine *N*-monooxygenase, 417
 homoserine dehydrogenase, 2
 homospermidine oxidase, 224
 α -humulene 10-hydroxylase, 438
 hydrazine dehydrogenase, 268
 hydrazine synthase, 268
 hydrogen dehydrogenase, 308
 hydrogen dehydrogenase (NADP⁺), 308
 hydrogen dehydrogenase [NAD(P)⁺], 308
 hydrogen peroxide-dependent heme synthase, 202
 hydrogen:quinone oxidoreductase, 309
 hydrogenase (acceptor), 311
 hydrogenase (NAD⁺, ferredoxin), 308
 hydroperoxy fatty acid reductase, 302
 hydroquinone 1,2-dioxygenase, 327
 3-hydroxy acid dehydrogenase, 84
 3 α -hydroxy bile acid-CoA-ester 3-dehydrogenase, 87
 2-hydroxy fatty acid dioxygenase, 477
 3 β -hydroxy- Δ^5 -steroid dehydrogenase, 32
 3-hydroxy-1,2-didehydro-2,3-dihydrotabersonine reductase, 125
 2-hydroxy-1,4-benzoquinone reductase, 258
 2-hydroxy-1,4-benzoxazin-3-one monooxygenase, 437
N-hydroxy-2-acetamidofluorene reductase, 264
 1-hydroxy-2-isopentenylcarotenoid 3,4-desaturase, 210
 3-hydroxy-2-methylbutyryl-CoA dehydrogenase, 38
 2-hydroxy-2-methylpropanal dehydrogenase, 146
 3-hydroxy-2-methylpyridine-5-carboxylate monooxygenase, 402
 3-hydroxy-2-methylquinolin-4-one 2,4-dioxygenase, 322
 1-hydroxy-2-naphthoate 1,2-dioxygenase, 320
 1-hydroxy-2-naphthoate hydroxylase, 385
 [1-hydroxy-2-(trimethylamino)ethyl]phosphonate dioxygenase (glycine-betaine-forming), 333
 4-hydroxy-3-methylbut-2-en-1-yl diphosphate reductase, 531
 7 β -hydroxy-3-oxochol-24-oyl-CoA 4-desaturase, 184
 2-hydroxy-3-oxopropionate reductase, 15
 4-hydroxy-3-prenylbenzoate synthase, 338
 4-hydroxy-3-prenylphenylpyruvate oxygenase, 331
 6-hydroxy-3-succinoylpyridine 3-monooxygenase, 388
 2-hydroxy-4-carboxymuconate semialdehyde hemiacetal dehydrogenase, 68
 3-hydroxy-4-methylanthranilyl-[aryl-carrier protein] 5-monooxygenase, 397
 3-hydroxy-4-oxoquinoline 2,4-dioxygenase, 322
 3 β -hydroxy-5 α -steroid dehydrogenase, 59
 3 α -hydroxy-5 β -androstane-17-one 3 α -dehydrogenase, 33
 3 β -hydroxy-5 β -steroid dehydrogenase, 59
 8-hydroxy-5-deazaflavin:NADPH oxidoreductase, 235
 2-hydroxy-5-methyl-1-naphthoate 7-hydroxylase, 467
 4-hydroxy-6-methylpretetramide 12a-monooxygenase, 400
 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate reductase, 167
 3-hydroxy-9,10-secoandrosta-1,3,5(10)-triene-9,17-dione monooxygenase, 408
 3¹-hydroxy-L-isoleucine 4-dioxygenase, 359
 4-hydroxy-tetrahydrodipicolinate reductase, 525
 4-hydroxyacetophenone monooxygenase, 379
 hydroxyacid-oxoacid transhydrogenase, 121
 3-hydroxyacyl-CoA dehydrogenase, 9
 3-hydroxyanthranilate 3,4-dioxygenase, 313
 3-hydroxyanthranilate oxidase, 293
 5'-hydroxyaverantin dehydrogenase, 77
 4-hydroxybenzaldehyde dehydrogenase (NAD⁺), 137
 4-hydroxybenzaldehyde dehydrogenase (NADP⁺), 145
 4-hydroxybenzoate 1-hydroxylase, 377
 3-hydroxybenzoate 2-monooxygenase, 509
 4-hydroxybenzoate 3-monooxygenase, 366
 4-hydroxybenzoate 3-monooxygenase [NAD(P)H], 372
 3-hydroxybenzoate 4-monooxygenase, 370
 3-hydroxybenzoate 6-monooxygenase, 370
 4-hydroxybenzoate brominase (decarboxylating), 494
 4-hydroxybenzoyl-CoA reductase, 114
 3-hydroxybenzyl-alcohol dehydrogenase, 22
 2-hydroxybiphenyl 3-monooxygenase, 374
 3-hydroxybutyrate dehydrogenase, 8
 4-hydroxybutyrate dehydrogenase, 15
 3-hydroxybutyryl-CoA dehydrogenase, 34
 1-hydroxycarotenoid 3,4-desaturase, 207
 3 β -hydroxycholanate 3-dehydrogenase (NAD⁺), 86
 3 β -hydroxycholanate 3-dehydrogenase (NADP⁺), 87
 3 α -hydroxycholanate dehydrogenase (NAD⁺), 13
 3 α -hydroxycholanate dehydrogenase (NADP⁺), 86
 24-hydroxycholesterol 7 α -hydroxylase, 412
 25/26-hydroxycholesterol 7 α -hydroxylase, 413
 hydroxycinnamoyl-CoA reductase, 184
 6-hydroxycyclohex-1-ene-1-carbonyl-CoA dehydrogenase, 81
 4-hydroxycyclohexanecarboxylate dehydrogenase, 48
 hydroxycyclohexanecarboxylate dehydrogenase, 36
 2-hydroxycyclohexanone 2-monooxygenase, 377
 3-hydroxycyclohexanone dehydrogenase, 121
 2'-hydroxydaidzein reductase, 169
 ω -hydroxydecanoate dehydrogenase, 16
 2-hydroxyethylphosphonate dioxygenase, 328
 8-hydroxygeraniol dehydrogenase, 70
 3 α -hydroxyglycylrrhettinate dehydrogenase, 49
 6-hydroxyhexanoate dehydrogenase, 54
 6 β -hydroxyhyoscyamine epoxidase, 504
 15-hydroxyicosatetraenoate dehydrogenase, 50
 2-hydroxyindolin-2-one monooxygenase, 437
 3-hydroxyisobutyrate dehydrogenase, 8
 2-hydroxyisoflavanone synthase, 431
 2'-hydroxyisoflavone reductase, 168
 12-hydroxyjasmonoyl-L-amino acid 12-hydroxylase, 420
 hydroxylamine dehydrogenase, 267
 hydroxylamine oxidase, 268
 hydroxylamine oxidase (cytochrome), 269
 hydroxylamine reductase, 271
 hydroxylamine reductase (NADH), 264
 hydroxymalonate dehydrogenase, 36
 4-hydroxymandelate oxidase, 109

4-hydroxymandelate oxidase (decarboxylating), 104
 4-hydroxymandelate synthase, 322
 7-hydroxymethyl chlorophyll *a* reductase, 531
 4-(hydroxymethyl)benzenesulfonate dehydrogenase, 54
 5-(hydroxymethyl)furfural oxidase, 109
 2-hydroxymethylglutarate dehydrogenase, 62
 hydroxymethylglutaryl-CoA reductase, 20
 hydroxymethylglutaryl-CoA reductase (NADPH), 8
 S-(hydroxymethyl)glutathione dehydrogenase, 61
 S-(hydroxymethyl)mycothiol dehydrogenase, 66
 (hydroxymethyl)phosphonate dioxygenase, 332
 2-hydroxymuconate-6-semialdehyde dehydrogenase, 142
 4-hydroxymuconic-semialdehyde dehydrogenase, 137
 6'''-hydroxyneomycin C oxidase, 108
 6-hydroxynicotinate 3-monooxygenase, 383
 6-hydroxynicotinate dehydrogenase, 527
 6-hydroxynicotinate reductase, 193
 4-hydroxyphenylacetaldehyde dehydrogenase, 135
 4-hydroxyphenylacetaldehyde oxime monooxygenase, 415
 4-hydroxyphenylacetate 1-monooxygenase, 369
 4-hydroxyphenylacetate 3-monooxygenase, 407
 3-hydroxyphenylacetate 6-hydroxylase, 376
 4-hydroxyphenylpyruvate dioxygenase, 318
 4-hydroxyphenylpyruvate oxidase, 151
 hydroxyphenylpyruvate reductase, 51
 hydroxyphytanate oxidase, 105
 3-hydroxypimeloyl-CoA dehydrogenase, 54
 17 α -hydroxyprogesterone deacetylase, 414
 hydroxyproline dehydrogenase, 247
 3-hydroxypropionate dehydrogenase, 15
 3-hydroxypropionate dehydrogenase (NADP⁺), 64
 15-hydroxyprostaglandin dehydrogenase (NAD⁺), 31
 15-hydroxyprostaglandin dehydrogenase (NADP⁺), 42
 15-hydroxyprostaglandin-D dehydrogenase (NADP⁺), 42
 15-hydroxyprostaglandin-I dehydrogenase (NADP⁺), 49
 6-hydroxypseudooxynicotine dehydrogenase, 252
 2-hydroxypyridine 5-monooxygenase, 509
 hydroxypyruvate reductase, 18
 hydroxyquinol 1,2-dioxygenase, 320
 4-hydroxyquinoline 3-monooxygenase, 376
 2-hydroxyquinoline 5,6-dioxygenase, 364
 2-hydroxyquinoline 8-monooxygenase, 376
 4-hydroxysphinganine ceramide fatty acyl 2-hydroxylase, 475
 hydroxysqualene dehydroxylase, 532
 22 α -hydroxysteroid 23-monooxygenase, 447
 3 α -hydroxysteroid 3-dehydrogenase, 78
 3 β -hydroxysteroid 3-dehydrogenase, 57
 3 α -hydroxysteroid 3-dehydrogenase (*Re*-specific), 46
 3 α -hydroxysteroid 3-dehydrogenase (*Si*-specific), 13
 11 β -hydroxysteroid dehydrogenase, 32
 12 α -hydroxysteroid dehydrogenase, 38
 12 β -hydroxysteroid dehydrogenase, 51
 16 α -hydroxysteroid dehydrogenase, 32
 20 α -hydroxysteroid dehydrogenase, 33
 7 α -hydroxysteroid dehydrogenase, 34
 21-hydroxysteroid dehydrogenase (NAD⁺), 33
 3 α (17 β)-hydroxysteroid dehydrogenase (NAD⁺), 51
 21-hydroxysteroid dehydrogenase (NADP⁺), 33
 7 β -hydroxysteroid dehydrogenase (NADP⁺), 43
 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating), 36
 3 β -hydroxysteroid-4 β -carboxylate 3-dehydrogenase (decarboxylating), 92
 11-hydroxysugiol 20-monooxygenase, 425
 4-hydroxythreonine-4-phosphate dehydrogenase, 55
 hyoscyamine (6*S*)-dioxygenase, 344
 hyponitrite reductase, 263
 hypoxia-inducible factor-asparagine dioxygenase, 348
 hypoxia-inducible factor-proline dioxygenase, 347
 icosanoyl-CoA 5-desaturase, 480
 D-idoitol 2-dehydrogenase, 4
 L-idoitol 2-dehydrogenase, 4
 L-idonate 5-dehydrogenase, 56
 L-idonate 5-dehydrogenase (NAD⁺), 80
 3-(imidazol-5-yl)lactate dehydrogenase, 25
 imidazoleacetate 4-monooxygenase, 367
 IMP dehydrogenase, 44
 indanol dehydrogenase, 25
 indole 2,3-dioxygenase, 316
 indole-2-monooxygenase, 449
 indole-3-acetaldehyde oxidase, 150
 indole-3-acetaldehyde reductase (NADH), 41
 indole-3-acetaldehyde reductase (NADPH), 41
 indole-3-acetate monooxygenase, 400
 indole-3-carbonyl nitrile 4-hydroxylase, 453
 indole-3-pyruvate monooxygenase, 389
 indoleamine 2,3-dioxygenase, 323
 indolepyruvate ferredoxin oxidoreductase, 156
 indolin-2-one monooxygenase, 450
 inositol 2-dehydrogenase, 5
 scyllo-inositol 2-dehydrogenase (NAD⁺), 81
 scyllo-inositol 2-dehydrogenase (NADP⁺), 82
 inositol oxygenase, 339
 iodide peroxidase, 298
 iodotyrosine deiodinase, 543
 ipsdienol dehydrogenase, 85
 ipsdienol synthase, 414
 iron—cytochrome-*c* reductase, 291
 iron:rusticyanin reductase, 522
 isobutylamine *N*-monooxygenase, 413
 isocitrate dehydrogenase (NAD⁺), 10
 isocitrate dehydrogenase (NADP⁺), 11
 isocitrate—homoisocitrate dehydrogenase, 61
 isoeugenol monooxygenase, 332
 isoeugenol synthase, 69
 isoflavone 2'-hydroxylase, 432
 isoflavone 3'-hydroxylase, 431
 ent-isokaurene C2/C3-hydroxylase, 427
 L-isoleucine 3¹-dioxygenase, 358
 L-isoleucine 4-hydroxylase, 351
 isoleucine *N*-monooxygenase, 416

isopenicillin-N synthase, 543
 isopiperitenol dehydrogenase, 48
 isopropanol dehydrogenase (NADP⁺), 18
 3-isopropylmalate dehydrogenase, 19
 isopyridoxal dehydrogenase (5-pyridoxate-forming), 147
 isopyridoxal dehydrogenase (5-pyridoxolactone-forming), 92
 isoquinoline 1-oxidoreductase, 205
 isovaleryl-CoA dehydrogenase, 198
epi-isozizaene 5-monooxygenase, 470

 jasmonic acid 12-hydroxylase, 399
 jasmonoyl-L-amino acid 12-hydroxylase, 420
 juglone 3-hydroxylase, 528

 kanamycin B dioxygenase, 349
ent-kaurene monooxygenase, 430
ent-kaurenoic acid monooxygenase, 436
 ketol-acid reductoisomerase (NAD⁺), 84
 ketol-acid reductoisomerase (NADP⁺), 20
 ketol-acid reductoisomerase [NAD(P)⁺], 84
 3-ketosteroid 9 α -monooxygenase, 467
 ketosteroid monooxygenase, 375
 kynurenate-7,8-dihydrodiol dehydrogenase, 163
 kynurenine 3-monooxygenase, 368
 kynurenine 7,8-hydroxylase, 506

 laccase, 293
 lactaldehyde dehydrogenase, 130
 lactaldehyde reductase, 18
 lactaldehyde reductase (NADPH), 14
 lactate 2-monooxygenase, 334
 D-lactate dehydrogenase, 7
 L-lactate dehydrogenase, 7
 D-lactate dehydrogenase (acceptor), 118
 D-lactate dehydrogenase (cytochrome), 98
 L-lactate dehydrogenase (cytochrome), 98
 D-lactate dehydrogenase (cytochrome *c*-553), 99
 lactate dehydrogenase (NAD⁺, ferredoxin), 97
 D-lactate dehydrogenase (quinone), 113
 L-lactate oxidase, 101
 lactate—malate transhydrogenase, 118
 lanthanide-dependent methanol dehydrogenase, 100
 (–)-lariciresinol reductase, 551
 laurate 7-monooxygenase, 442
 leghemoglobin reductase, 255
 leucine dehydrogenase, 213
 leucoanthocyanidin reductase, 524
 leukotriene-B₄ 20-monooxygenase, 433
 leukotriene-E₄ 20-monooxygenase, 372
 levoglucosan dehydrogenase, 95
 lignin peroxidase, 300
 lignostilbene $\alpha\beta$ -dioxygenase, 321
 limonene 1,2-monooxygenase, 381
 (*S*)-limonene 3-monooxygenase, 434
 (*R*)-limonene 6-monooxygenase, 421
 (*S*)-limonene 6-monooxygenase, 421
 (*S*)-limonene 7-monooxygenase, 421
 limonene dehydrogenase, 535
 limonene-1,2-diol dehydrogenase, 64
 linalool 8-monooxygenase, 430
 linoleate 10*R*-lipoxygenase, 326
 linoleate 11-lipoxygenase, 321
 linoleate 13*S*-lipoxygenase, 315
 linoleate 8*R*-lipoxygenase, 325
 linoleate 9*S*-lipoxygenase, 325
 linolenate 9*R*-lipoxygenase, 326
 linoleoyl-lipid Δ^{12} conjugase (11*E*,13*Z*-forming), 482
 linoleoyl-lipid Δ^9 conjugase, 481
 lipoyl-dependent peroxiredoxin, 304
 lithocholate 6 β -hydroxylase, 445
 long-chain acyl-[acyl-carrier-protein] reductase, 141
 long-chain acyl-CoA ω -monooxygenase, 442
 long-chain acyl-CoA dehydrogenase, 199
 long-chain acyl-protein thioester reductase, 135
 long-chain alkane monooxygenase, 413
 long-chain fatty acid ω -monooxygenase, 429
 long-chain-3-hydroxyacyl-CoA dehydrogenase, 45
 long-chain-alcohol dehydrogenase, 41
 long-chain-alcohol oxidase, 104
 long-chain-aldehyde dehydrogenase, 134
Cypridina-luciferin 2-monooxygenase, 335
Oplophorus-luciferin 2-monooxygenase, 336
Watasenia-luciferin 2-monooxygenase, 335
Latia-luciferin monooxygenase (demethylating), 508
 lupanine 17-hydroxylase (cytochrome *c*), 526
 luteothin monooxygenase, 469
 lysine 2-monooxygenase, 334
 lysine 6-dehydrogenase, 215
 L-lysine 6-oxidase, 222
 L-lysine *N*⁶-monooxygenase (NADPH), 376
 lysine dehydrogenase, 214
 L-lysine oxidase, 221
 D-lysopine dehydrogenase, 230
 lytic cellulose monooxygenase (C1-hydroxylating), 513
 lytic cellulose monooxygenase (C4-dehydrogenating), 514
 lytic chitin monooxygenase, 513
 lytic starch monooxygenase, 514

 magnesium-protoporphyrin IX monomethyl ester (oxidative) cyclase, 378
 malate dehydrogenase, 9
 D-malate dehydrogenase (decarboxylating), 19
 malate dehydrogenase (decarboxylating), 10
 malate dehydrogenase (NADP⁺), 19
 malate dehydrogenase [NAD(P)⁺], 64
 malate dehydrogenase (oxaloacetate-decarboxylating), 10
 malate dehydrogenase (oxaloacetate-decarboxylating) (NADP⁺), 10
 malate dehydrogenase (quinone), 111
 maleylacetate reductase, 165
 malonate-semialdehyde dehydrogenase, 128
 malonate-semialdehyde dehydrogenase (acetyating), 129
 malonyl-CoA reductase (malonate semialdehyde-forming), 140

mandelate 4-monooxygenase, 472
 (*R*)-mandelate dehydrogenase, 83
 (*S*)-mandelate dehydrogenase, 122
 (*R*)-mandelonitrile oxidase, 110
 manganese oxidase, 521
 manganese peroxidase, 299
 mannitol 2-dehydrogenase, 16
 mannitol 2-dehydrogenase (NADP⁺), 30
 mannitol dehydrogenase, 54
 mannitol dehydrogenase (cytochrome), 98
 D-mannitol oxidase, 107
 mannitol-1-phosphate 5-dehydrogenase, 5
 mannose-6-phosphate 6-reductase, 48
 mannuronate reductase, 29
 marmesin synthase, 425
 medium-chain acyl-CoA dehydrogenase, 199
 melilotate 3-monooxygenase, 367
 menaquinone-9 β -reductase, 210
 3-mercaptopropionate dioxygenase, 333
 mercury(II) reductase, 519
 meromycolic acid enoyl-[acyl-carrier-protein] reductase, 184
 methane monooxygenase (particulate), 474
 methane monooxygenase (soluble), 370
 methanesulfonate monooxygenase (FMNH₂), 415
 methanesulfonate monooxygenase (NADH), 382
 methanethiol oxidase, 279
 methanol dehydrogenase, 52
 methanol dehydrogenase (cytochrome *c*), 99
 methanol dehydrogenase (nicotinoprotein), 124
 5,10-methenyltetrahydromethanopterin hydrogenase, 310
 L-methionine (*R*)-*S*-oxide reductase, 283
 L-methionine (*S*)-*S*-oxide reductase, 283
 [methionine synthase] reductase, 520
 4-methoxybenzoate monooxygenase (*O*-demethylating), 508
 4'-methoxyisoflavone 2'-hydroxylase, 431
 methyl farnesoate epoxidase, 442
 2-methyl-1,2-propanediol dehydrogenase, 89
 5-methyl-1-naphthoate 3-hydroxylase, 392
 2-methyl-1-pyrroline reductase, 238
 3-methyl-2-oxobutanoate dehydrogenase (2-methylpropanoyl-transferring), 152
 3-methyl-2-oxobutanoate dehydrogenase (ferredoxin), 155
 4-methyl-5-nitrocatechol 5-monooxygenase, 395
N-methyl-L-amino-acid oxidase, 240
 3-methyl-L-tyrosine peroxygenase, 307
 methyl-branched lipid ω -hydroxylase, 462
*N*⁶-methyl-lysine oxidase, 240
N-methylalanine dehydrogenase, 214
 methylamine dehydrogenase (amicyanin), 225
 4-methylaminobutanoate oxidase (formaldehyde-forming), 244
 4-methylaminobutanoate oxidase (methylamine-forming), 244
 methylarsonate reductase, 541
 3-methylbutanal reductase, 56
N-methylcoclaurine 3'-monooxygenase, 435
 methylcytosine dioxygenase, 360
 methylecgonone reductase, 73
 2-methylene-furan-3-one reductase, 181
 3-methyleneoxindole reductase, 163
 methylenetetrahydrofolate dehydrogenase (NAD⁺), 230
 methylenetetrahydrofolate dehydrogenase (NADP⁺), 228
 methylenetetrahydrofolate reductase (ferredoxin), 247
 methylenetetrahydrofolate reductase (NADH), 239
 methylenetetrahydrofolate reductase (NADPH), 239
 methylenetetrahydrofolate reductase [NAD(P)H], 231
 methylenetetrahydromethanopterin dehydrogenase, 249
 5,10-methylenetetrahydromethanopterin reductase, 249
 methylglutamate dehydrogenase, 251
 (methyl)glyoxal oxidase, 151
 methylglyoxal reductase (NADH), 18
 methylglyoxal reductase (NADPH), 60
 β -methylindole-3-pyruvate reductase, 88
 methylmalonate-semialdehyde dehydrogenase (CoA-acylating), 131
 5-methylphenazine-1-carboxylate 1-monooxygenase, 396
 4-methylphenol dehydrogenase (hydroxylating), 532
 methylphosphonate hydroxylase, 358
 methylphosphonate synthase, 329
 6-methylpretetramide 4-monooxygenase, 400
 4 α -methylsterol monooxygenase, 476
 4 β -methylsterol monooxygenase, 403
 (2*S*)-methylsuccinyl-CoA dehydrogenase, 200
 (methylsulfanyl)alkanaldoxime *N*-monooxygenase, 418
 3-(methylsulfanyl)propanoyl-CoA 2-dehydrogenase, 211
 methyltetrahydroprotoberberine 14-monooxygenase, 434
 4-methylthio 2-oxobutanoate reductase (NADH), 95
 methylxanthine *N*¹-demethylase, 390
 methylxanthine *N*³-demethylase, 391
 7-methylxanthine demethylase, 384
 mevaldate reductase, 8
 mevaldate reductase (NADPH), 8
 momilactone-A synthase, 63
 monacolin L hydroxylase, 441
 monoamine oxidase, 219
 monocyclic monoterpene ketone monooxygenase, 381
 monodehydroascorbate reductase (NADH), 258
 monoprenyl isoflavone epoxidase, 510
 morphine 6-dehydrogenase, 47
 mRNA *N*¹-methyladenine demethylase, 353
 mRNA *N*⁶-methyladenine demethylase, 353
 mugineic-acid 3-dioxygenase, 346
 mycocyclosin synthase, 498
 [mycofactocin precursor peptide]-tyrosine decarboxylase, 203
 mycoredoxin, 542
 mycoredoxin-dependent peroxiredoxin, 305
 mycothione reductase, 275
 myeloperoxidase, 306
 NADH oxidase (H₂O₂-forming), 256
 NADH oxidase (H₂O-forming), 256
 NADH peroxidase, 296
 NADH-dependent peroxiredoxin, 303
 NADH:quinone reductase (non-electrogenic), 259

NAD(P)⁺ transhydrogenase, 253
 NAD(P)⁺ transhydrogenase (*Rel/Si*-specific), 253
 NAD(P)⁺ transhydrogenase (*Si*-specific), 252
 NAD(P)⁺ transhydrogenase (ferredoxin), 253
 NADP-retinol dehydrogenase, 65
 NADPH dehydrogenase, 261
 NAD(P)H dehydrogenase (quinone), 257
 NADPH dehydrogenase (quinone), 259
 NAD(P)H oxidase (H₂O₂-forming), 255
 NAD(P)H oxidase (H₂O-forming), 255
 NADPH peroxidase, 297
 NAD(P)H sulfur oxidoreductase (CoA-dependent), 276
 NADPH—cytochrome-*c*₂ reductase, 255
 NADPH—hemoprotein reductase, 254
 NADPH:quinone reductase, 258
 (*S*)-nandinine synthase, 499
 naphthalene 1,2-dioxygenase, 363
 neopentalenolactone D synthase, 390
 nepetalactol dehydrogenase, 93
 nepetalactol monooxygenase, 451
 nicotinate dehydrogenase, 524
 nicotinate dehydrogenase (cytochrome), 526
 nicotine blue oxidoreductase, 71
 nicotine dehydrogenase, 218, 250
 nitrate reductase (cytochrome), 291
 nitrate reductase (NADH), 262
 nitrate reductase (NADPH), 263
 nitrate reductase [NAD(P)H], 262
 nitrate reductase (quinone), 270
 nitric oxide dioxygenase, 364
 nitric oxide reductase (cytochrome *c*), 267
 nitric oxide reductase (menaquinol), 270
 nitric oxide reductase [NAD(P)⁺, nitrous oxide-forming], 265
 nitric-oxide synthase (flavodoxin), 420
 nitric-oxide synthase (NADPH), 373
 nitrilotriacetate monooxygenase, 407
 nitrite dismutase, 270
 nitrite reductase (cytochrome; ammonia-forming), 266
 nitrite reductase (NADH), 265
 nitrite reductase [NAD(P)H], 263
 nitrite reductase (NO-forming), 266
 β-nitroacrylate reductase, 162
 nitroalkane oxidase, 268
 nitroarene dioxygenase, 365
 nitrobenzene nitroreductase, 265
 4-nitrocatechol 4-monooxygenase, 388
 nitrogenase, 539
 nitrogenase (flavodoxin), 540
 nitronate monooxygenase, 337
 2-nitrophenol 2-monooxygenase, 371
 4-nitrophenol 2-monooxygenase, 371
 4-nitrophenol 4-monooxygenase, 389
 nitroquinoline-*N*-oxide reductase, 264
 5-nitrosalicylate dioxygenase, 326
 nitrosourea synthase, 405
 nitrous-oxide reductase, 267
 non-specific polyamine oxidase, 243
 D-nopaline dehydrogenase, 231
 noranthrone monooxygenase, 338
 noroxomaritidine synthase, 492
 norsolorinic acid ketoreductase, 77
 noscapine synthase, 92
 nucleoside oxidase, 105
 nucleoside oxidase (H₂O₂-forming), 107
 octanol dehydrogenase, 17
 D-octopine dehydrogenase, 229
 oleate 10*S*-lipoxygenase, 329
 opine dehydrogenase, 233
 orcinol 2-monooxygenase, 367
 L-ornithine *N*⁵-monooxygenase (NADPH), 393
 L-ornithine *N*⁵-monooxygenase [NAD(P)H], 393
 ornithine lipid ester-linked acyl 2-hydroxylase, 354
 oryzalexin D synthase, 440
 oryzalexin E synthase, 440
 oxalate oxidase, 149
 oxalate oxidoreductase, 156
 oxalglycolate reductase (decarboxylating), 21
 oxazoline dehydrogenase, 190
 11-oxo-β-amyrin 30-oxidase, 438
 3-oxo-Δ¹-steroid hydratase/dehydrogenase, 536
 3-oxo-5α-steroid 4-dehydrogenase (acceptor), 204
 3-oxo-5α-steroid 4-dehydrogenase (NADP⁺), 164
 3-oxo-5β-steroid 4-dehydrogenase, 204
 3-oxo-5,6-dehydrosuberil-CoA semialdehyde dehydrogenase, 144
 20-oxo-5-*O*-mycaminosyltylactone 23-monooxygenase, 468
 2-oxo-acid reductase, 122
 2-oxoacid oxidoreductase (ferredoxin), 156
 3-oxoacyl-[acyl-carrier-protein] reductase, 23
 3-oxoacyl-[acyl-carrier-protein] reductase (NADH), 45
 2-oxoadipate dioxygenase/decarboxylase, 333
 2-oxoadipate reductase, 37
 2-oxoaldehyde dehydrogenase (NAD⁺), 130
 2-oxoaldehyde dehydrogenase (NADP⁺), 135
 3-oxocholoyl-CoA 4-desaturase, 184
 6-oxocineole dehydrogenase, 375
 8-oxocoformycin reductase, 50
 8-oxogeranial reductase, 186
 oxoglutarate dehydrogenase (NADP⁺), 135
 oxoglutarate dehydrogenase (succinyl-transferring), 152
 2-oxoglutarate dehydrogenase system, 147
 2-oxoglutarate dioxygenase (ethene-forming), 338
 2-oxoglutarate reductase, 88
 2-oxoglutarate synthase, 154
 2-oxoglutarate/L-arginine monooxygenase/decarboxylase (succinate-forming), 503
 6-oxohexanoate dehydrogenase, 137
 12-oxophytodienoate reductase, 168
 4-oxoproline reductase, 23
 2-oxopropyl-CoM reductase (carboxylating), 273
 3-oxosteroid 1-dehydrogenase, 204

palmitoyl-CoA 14-(*E/Z*)-desaturase, 487
 palmitoyl-[glycerolipid] 3-(*E*)-desaturase, 490
 palmitoyl-[glycerolipid] 7-desaturase, 490
 pantoate 4-dehydrogenase, 24
 (*R*)-pantolactone dehydrogenase (flavin), 121
 paromamine 6'-oxidase, 108
 pentachlorophenol monooxygenase, 374
 pentalenene oxygenase, 468
 pentalenic acid synthase, 461
 pentalenolactone D synthase, 389
 pentalenolactone F synthase, 349
 pentalenolactone synthase, 480
 peptide-aspartate β -dioxygenase, 345
 peptide-methionine (*R*)-*S*-oxide reductase, 283
 peptide-methionine (*S*)-*S*-oxide reductase, 282
 peptide-tryptophan 2,3-dioxygenase, 318
 peptidyl-lysine (3*S*)-dioxygenase, 355
 peptidylglycine monooxygenase, 473
 perakine reductase, 69
 perillyl-alcohol dehydrogenase, 31
 peroxidase, 297
 persulfide dioxygenase, 316
Methanosarcina-phenazine hydrogenase, 310
 phenol 2-monooxygenase (FADH₂), 410
 phenol 2-monooxygenase (NADH), 403
 phenol 2-monooxygenase (NADPH), 367
 phenylacetaldehyde dehydrogenase, 133
 phenylacetaldehyde oxime monooxygenase, 418
 phenylacetate 2-hydroxylase, 422
 phenylacetone monooxygenase, 380
 phenylacetonitrile α -monooxygenase, 428
 phenylacetyl-CoA 1,2-epoxidase, 386
 phenylacetyl-CoA dehydrogenase, 530
 phenylalanine 2-monooxygenase, 336
 phenylalanine 3-monooxygenase, 472
 phenylalanine 4-monooxygenase, 470
 phenylalanine *N*-monooxygenase, 417
 phenylalanine dehydrogenase, 215
 phenylglyoxylate dehydrogenase (acylating), 136
 3-phenylpropanoate dioxygenase, 364
 pheophorbide *a* oxygenase, 463
 phloroglucinol reductase, 170
 phosphatidylcholine 12-monooxygenase, 474
 4-phospho-D-erythronate 3-dehydrogenase, 91
 4-phospho-D-threonate 3-dehydrogenase, 90
 phosphoadenylyl-sulfate reductase (thioredoxin), 281
 4-phosphoerythronate dehydrogenase, 62
 phosphogluconate 2-dehydrogenase, 11
 phosphogluconate dehydrogenase (NAD⁺-dependent, decarboxylating), 75
 phosphogluconate dehydrogenase (NADP⁺-dependent, decarboxylating), 11
 phosphogluconate dehydrogenase [NAD(P)⁺-dependent, decarboxylating], 77
 phosphoglycerate dehydrogenase, 21
 phospholipid-hydroperoxide glutathione peroxidase, 299
 phosphonate dehydrogenase, 541
 phosphonoacetaldehyde reductase (NADH), 67
 photosystem I, 554
 photosystem II, 294
 phthalate 4,5-*cis*-dihydrodiol dehydrogenase, 171
 phthalate 4,5-dioxygenase, 362
 phycoerythrin:ferredoxin oxidoreductase, 194
 phycoerythrobilin synthase, 194
 phycoerythrobilin:ferredoxin oxidoreductase, 193
 phylloquinone ω -hydroxylase, 428
 phylloquinone monooxygenase (2,3-epoxidizing), 508
 phytanoyl-CoA dioxygenase, 345
 phytochromobilin:ferredoxin oxidoreductase, 193
 phytoene desaturase (ζ -carotene-forming), 208
 phytoene desaturase (3,4-didehydrolycopene-forming), 208
 phytoene desaturase (lycopene-forming), 208
 phytoene desaturase (neurosporene-forming), 207
 pikromycin synthase, 468
 9 β -pimara-7,15-diene oxidase, 437
syn-pimaradiene 3-monooxygenase, 425
 pimeloyl-[acyl-carrier protein] synthase, 419
 pimeloyl-CoA dehydrogenase, 171
 α -pinene monooxygenase, 387
 D-pinitol dehydrogenase, 31
 (-)-pinoresinol reductase, 551
 L-pipecolate dehydrogenase, 250
 L-pipecolate oxidase, 241
 1-piperidine-2-carboxylate/1-pyrroline-2-carboxylate reductase (NADPH), 231
 1-piperidine-2-carboxylate/1-pyrroline-2-carboxylate reductase [NAD(P)H], 227
 plant 3 β -hydroxysteroid-4 α -carboxylate 3-dehydrogenase (decarboxylating), 93
 plant 4 α -monomethylsterol monooxygenase, 477
 plant 4,4-dimethylsterol C-4 α -methyl-monooxygenase, 476
 plant seed peroxygenase, 306
 plasmalyethanolamine desaturase, 500
 (-)-pluviatolide synthase, 499
 polyamine oxidase (propane-1,3-diamine-forming), 243
 polyprenol reductase, 178
 2-polyprenylphenol 6-hydroxylase, 402
 polyvinyl alcohol dehydrogenase (cytochrome), 99
 polyvinyl-alcohol oxidase, 106
 PqqA peptide cyclase, 548
 pre-mycofactocin synthase, 223
 precorrin-2 dehydrogenase, 174
 precorrin-3B synthase, 379
 precorrin-6A reductase, 170
 prenaspirodiene oxygenase, 448
 prenylcysteine oxidase, 279
 prephenate dehydrogenase, 162
 prephenate dehydrogenase (NADP⁺), 162
 preQ₁ synthase, 265
 primary-amine oxidase, 222
 procollagen-lysine 5-dioxygenase, 342
 procollagen-proline 3-dioxygenase, 343

procollagen-proline 4-dioxygenase, 341
 progesterone 11 α -monooxygenase, 507
 progesterone monooxygenase, 506
 proline 3-hydroxylase, 347
 L-proline *cis*-4-hydroxylase, 353
 L-proline *trans*-4-hydroxylase, 354
 D-proline dehydrogenase, 252
 proline dehydrogenase, 247
 D-proline reductase, 546
 L-prolyl-[peptidyl-carrier protein] dehydrogenase, 200
 propanal dehydrogenase (CoA-propanoylating), 143
 propane 2-monooxygenase, 398
 1,3-propanediol dehydrogenase, 43
 propanediol-phosphate dehydrogenase, 3
 prosolanapyrone-II oxidase, 108
 prostaglandin-E₂ 9-reductase, 40
 prostaglandin-endoperoxide synthase, 506
 prostaglandin-F synthase, 40
 prostamide/prostaglandin F_{2 α} synthase, 302
 protein dithiol oxidoreductase (disulfide-forming), 283
 protein dithiol:quinone oxidoreductase DsbB, 286
 protein-L-histidine (3*S*)-3-hydroxylase, 360
 [protein]-arginine 3-hydroxylase, 358
 protein-disulfide reductase, 274
 protein-disulfide reductase (glutathione), 280
 protein-lysine 6-oxidase, 221
 protoasukamycin 4-monooxygenase, 396
 protocatechuate 3,4-dioxygenase, 313
 protocatechuate 4,5-dioxygenase, 314
 protochlorophyllide reductase, 166
 protodeoxyviolaceinate monooxygenase, 396
 protopanaxadiol 6-hydroxylase, 440
 protopine 6-monooxygenase, 434
 protoporphyrinogen IX dehydrogenase (quinone), 192
 protoporphyrinogen oxidase, 188
 pseudobaptigenin synthase, 496
 pseudoephedrine dehydrogenase, 94
 pseudooxynicotine dehydrogenase, 218
 psoralen synthase, 445
 pteridine oxidase, 527
 pteridine reductase, 234
 pulcherriminic acid synthase, 462
 putidaredoxin—NAD⁺ reductase, 537
 putrescine oxidase, 220
 pyranose dehydrogenase (acceptor), 122
 pyranose oxidase, 102
 pyridoxal 4-dehydrogenase, 24
 pyridoxal 5'-phosphate synthase, 219
 pyridoxal oxidase, 150
 5-pyridoxate monooxygenase, 402
 4-pyridoxic acid dehydrogenase, 125
 pyridoxine 4-dehydrogenase, 16
 pyridoxine 4-oxidase, 103
 pyridoxine 5-dehydrogenase, 118
 pyrimidine oxygenase, 512
 pyrimidine-deoxynucleoside 1'-dioxygenase, 343
 pyrimidine-deoxynucleoside 2'-dioxygenase, 342
 pyrimidodiazepine synthase, 246
 pyrogallol 1,2-oxygenase, 319
 pyrogallol hydroxytransferase, 552
 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein] brominase, 495
 1*H*-pyrrole-2-carbonyl-[peptidyl-carrier protein] chlorinase, 494
 pyrrole-2-carboxylate monooxygenase, 384
 1-pyrroline-2-carboxylate reductase [NAD(P)H], 238
 pyrroline-5-carboxylate reductase, 227
 pyrroloquinoline-quinone synthase, 189
 pyruvate dehydrogenase (acetyl-transferring), 152
 pyruvate dehydrogenase (NADP⁺), 135
 pyruvate dehydrogenase (quinone), 153
 pyruvate dehydrogenase system, 147
 pyruvate oxidase, 149
 pyruvate oxidase (CoA-acetylating), 150
 pyruvate synthase, 154
 6-pyruvoyltetrahydropterin 2'-reductase, 47
 quercetin 2,3-dioxygenase, 317
 questin monooxygenase, 373
 quinaldate 4-oxidoreductase, 206
 quinate/shikimate dehydrogenase (NAD⁺), 6
 quinate/shikimate dehydrogenase [NAD(P)⁺], 60
 quinate/shikimate dehydrogenase (quinone), 112
 quinine 3-monooxygenase, 422
 quinoline 2-oxidoreductase, 205
 quinoline-4-carboxylate 2-oxidoreductase, 206
 red chlorophyll catabolite reductase, 195
 renalase, 256
 resorcinol 4-hydroxylase (FADH₂), 412
 resorcinol 4-hydroxylase (NADH), 397
 resorcinol 4-hydroxylase (NADPH), 396
 respiratory dimethylsulfoxide reductase, 285
 reticuline oxidase, 544
 retinal dehydrogenase, 133
 L-rhamnose 1-dehydrogenase, 37
 L-rhamnose 1-dehydrogenase (NADP⁺), 83
 L-rhamnose 1-dehydrogenase [NAD(P)⁺], 83
 ribitol 2-dehydrogenase, 14
 ribitol-5-phosphate 2-dehydrogenase, 30
 ribitol-5-phosphate 2-dehydrogenase (NADP⁺), 90
 riboflavin reductase [NAD(P)H], 236
 ribonucleoside-diphosphate reductase, 528
 ribonucleoside-triphosphate reductase (formate), 116
 ribonucleoside-triphosphate reductase (thioredoxin), 528
 ribose 1-dehydrogenase (NADP⁺), 26
 ribosyldihydronicotinamide dehydrogenase (quinone), 295
 rifampicin monooxygenase, 395
 rifamycin-B oxidase, 293
 rubredoxin—NAD⁺ reductase, 536
 rubredoxin—NAD(P)⁺ reductase, 537
 saccharopine dehydrogenase (NAD⁺, L-glutamate-forming), 229
 saccharopine dehydrogenase (NAD⁺, L-lysine-forming), 228

saccharopine dehydrogenase (NADP⁺, L-glutamate-forming), 229
saccharopine dehydrogenase (NADP⁺, L-lysine-forming), 229
L-saccharopine oxidase, 244
salicylaldehyde dehydrogenase, 138
salicylate 1-monooxygenase, 366
salicylate 5-hydroxylase, 390
salicyloyl-CoA 5-hydroxylase, 394
salutaridine reductase (NADPH), 53
salutaridine synthase, 497
salviol synthase, 424
ent-sandaracopimaradiene 3-hydroxylase, 426
sanguinarine reductase, 181
 α -santonin 1,2-reductase, 169
sarcosine dehydrogenase, 249
sarcosine oxidase (5,10-methylenetetrahydrofolate-forming), 245
sarcosine oxidase (formaldehyde-forming), 240
sarcosine reductase, 546
scopoletin 8-hydroxylase, 354
secoisolariciresinol dehydrogenase, 72
secologanin synthase, 496
secondary-alcohol dehydrogenase (coenzyme-F₄₂₀), 116
secondary-alcohol oxidase, 104
secondary-alkyl amine dehydrogenase [NAD(P)⁺], 217
selenate reductase, 553
senecionine *N*-oxygenase, 381
sepiapterin reductase (*L*-erythro-7,8-dihydrobiopterin forming), 33
sepiapterin reductase (*L*-threo-7,8-dihydrobiopterin forming), 71
sequoyitol dehydrogenase, 31
serine 2-dehydrogenase, 212
L-serine 3-dehydrogenase (NAD⁺), 85
serine 3-dehydrogenase (NADP⁺), 59
serine-type anaerobic sulfatase-maturing enzyme, 116
shikimate dehydrogenase (NADP⁺), 6
short-chain 2-methylacyl-CoA dehydrogenase, 198
short-chain acyl-CoA dehydrogenase, 197
soluble quinoprotein glucose dehydrogenase, 123
D-sorbitol dehydrogenase (acceptor), 120
sorbitol-6-phosphate 2-dehydrogenase, 30
L-sorbose 1-dehydrogenase, 123
sorbose 5-dehydrogenase (NADP⁺), 27
sorbose dehydrogenase, 119
L-sorbose oxidase, 102
sorbose reductase, 62
spermidine dehydrogenase, 251
spermine oxidase, 243
spheroidene monooxygenase, 461
sphingolipid 10-desaturase, 483
sphingolipid 4-desaturase, 482
sphingolipid 8-(*E*)-desaturase, 483
sphingolipid 8-(*E/Z*)-desaturase, 486
sphingolipid C4-monooxygenase, 475
squalene monooxygenase, 409
stachydrine *N*-demethylase, 404
staphylopine dehydrogenase, 238
stearoyl-[acyl-carrier-protein] 9-desaturase, 478
stearoyl-CoA 9-desaturase, 478
steroid 11 β -monooxygenase, 459
steroid 15 β -monooxygenase, 460
steroid 17 α -monooxygenase, 410
steroid 21-monooxygenase, 409
steroid 22S-hydroxylase, 456
steroid 9 α -monooxygenase, 509
steroid C-25 hydroxylase, 536
sterol 14 α -demethylase, 449
sterol 14 α -demethylase (ferredoxin), 469
sterol 22-desaturase, 490
 Δ^7 -sterol 5(6)-desaturase, 483
 Δ^{14} -sterol reductase, 172
 $\Delta^{24(24^1)}$ -sterol reductase, 173
 Δ^{24} -sterol reductase, 173
stizolobate synthase, 318
stizolobinate synthase, 318
strombine dehydrogenase, 231
(*S*)-stylopine synthase, 496
styrene monooxygenase, 407
succinate dehydrogenase, 191
succinate-semialdehyde dehydrogenase (acylating), 140
succinate-semialdehyde dehydrogenase (NAD⁺), 130
succinate-semialdehyde dehydrogenase (NADP⁺), 141
succinate-semialdehyde dehydrogenase [NAD(P)⁺], 128
3-succinoylsemialdehyde-pyridine dehydrogenase, 142
succinylglutamate-semialdehyde dehydrogenase, 139
sugiol synthase, 424
sulcatone reductase, 55
sulfhydrogenase, 310
sulfide dehydrogenase, 276
sulfide-cytochrome-*c* reductase (flavocytochrome *c*), 277
sulfiredoxin, 288
sulfite dehydrogenase (cytochrome), 277
sulfite dehydrogenase (quinone), 285
sulfite oxidase, 279
sulfite reductase (coenzyme F₄₂₀), 288
sulfoacetaldehyde dehydrogenase, 139
sulfoacetaldehyde dehydrogenase (acylating), 142
sulfoacetaldehyde reductase (NADH), 97
sulfoacetaldehyde reductase (NADPH), 68
4-sulfobenzoate 3,4-dioxygenase, 362
sulfolactaldehyde 3-reductase, 82
3-sulfolactaldehyde dehydrogenase, 145
(*S*)-sulfolactate dehydrogenase, 67
sulfopropanediol 3-dehydrogenase, 67
sulfoquinovose 1-dehydrogenase, 86
sulfoquinovose monooxygenase, 457
sulfur oxygenase/reductase, 324
sulochrin oxidase [(+)-bisdechlorogeodin-forming], 544
sulochrin oxidase [(-)-bisdechlorogeodin-forming], 544
superoxide dismutase, 518
superoxide oxidase, 295
superoxide reductase, 518

tabersonine 16-hydroxylase, 435
 tabersonine 3-oxygenase, 421
 tagaturonate reductase, 14
meso-tartrate dehydrogenase, 160
 tartrate dehydrogenase, 21
 taurine dehydrogenase, 226
 taurine dioxygenase, 345
 taurochenodeoxycholate 6 α -hydroxylase, 422
 tauropine dehydrogenase, 232
 taxadiene 5 α -hydroxylase, 456
 taxane 10 β -hydroxylase, 435
 taxane 13 α -hydroxylase, 436
 taxifolin 8-monoxygenase, 369
 taxoid 14 β -hydroxylase, 386
 taxoid 2 α -hydroxylase, 458
 taxoid 7 β -hydroxylase, 458
 terephthalate 1,2-dioxygenase, 363
 testosterone 17 β -dehydrogenase (NADP⁺), 15
 tetracenomycin A2 monoxygenase-dioxygenase, 393
 tetracenomycin-F1 monoxygenase, 338
 tetrachlorobenzoquinone reductase, 90
 tetrachloroethene reductive dehalogenase, 550
 tetrachlorohydroquinone reductive dehalogenase, 547
 tetracycline 11a-monoxygenase, 399
 tetracycline 7-halogenase, 492
 tetraether lipid synthase, 549
 tetrahydroberberine oxidase, 188
 tetrahydrocannabinolic acid synthase, 545
 tetrahydroxynaphthalene reductase, 53
 thebaine 6-*O*-demethylase, 348
 thiamine oxidase, 105
 thiocyanate desulfurase, 278
 thiol oxidase, 279
 thiomorpholine-carboxylate dehydrogenase, 232
 thiophene-2-carbonyl-CoA monoxygenase, 510
 thioredoxin-dependent peroxiredoxin, 303
 thioredoxin-disulfide reductase, 274
 thioredoxin:protein disulfide reductase, 284
 thiosulfate dehydrogenase, 277
 thiosulfate dehydrogenase (quinone), 284
 thiosulfate reductase (cytochrome), 278
 thiosulfate reductase (quinone), 285
 D-threitol dehydrogenase (NAD⁺), 89
 L-threonate 2-dehydrogenase, 91
 L-threonate 3-dehydrogenase, 28
 L-threonine 3-dehydrogenase, 23
 L-threonyl-[L-threonyl-carrier protein] 4-chlorinase, 505
 thymine dioxygenase, 342
 thyroxine 5'-deiodinase, 550
 thyroxine 5-deiodinase, 550
 toluene 2-monoxygenase, 402
 toluene 4-monoxygenase, 401
 toluene dioxygenase, 362
 toluene methyl-monoxygenase, 465
 torulene dioxygenase, 325
 2,4,6-trichlorophenol monoxygenase, 455
 3,5,6-trichloropyridin-2-ol monoxygenase, 455
 3 α ,7 α ,12 α -trihydroxy-5 β -cholestanoyl-CoA 24-hydroxylase, 534
 (2,2,3-trimethyl-5-oxocyclopent-3-enyl)acetyl-CoA 1,5-monoxygenase, 388
 trimethylamine dehydrogenase, 248
 trimethylamine monoxygenase, 386
 trimethylamine-*N*-oxide reductase, 267
 [2-(trimethylamino)ethyl]phosphonate dioxygenase, 358
 4-trimethylammoniumbutyraldehyde dehydrogenase, 134
 trimethyllysine dioxygenase, 343
 trimethyltridecatetraene synthase, 423
 1,3,7-trimethyluric acid 5-monoxygenase, 395
 tRNA 2-(methylsulfanyl)-*N*⁶-isopentenyladenosine³⁷ hydroxylase, 518
 tRNA^{Phe} (7-(3-amino-3-carboxypropyl)wyosine³⁷-*C*²)-hydroxylase, 350
 tRNA-dihydrouridine^{16/17} synthase [NAD(P)⁺], 177
 tRNA-dihydrouridine^{20a/20b} synthase [NAD(P)⁺], 177
 tRNA-dihydrouridine²⁰ synthase [NAD(P)⁺], 178
 tRNA-dihydrouridine⁴⁷ synthase [NAD(P)⁺], 177
 tropinone reductase I, 44
 tropinone reductase II, 50
 trypanothione-disulfide reductase, 274
 tryprostatin B 6-hydroxylase, 439
 tryptamine 4-monoxygenase, 515
 tryptophan α , β -oxidase, 189
 tryptophan 2'-dioxygenase, 339
 tryptophan 2,3-dioxygenase, 314
 tryptophan 2-monoxygenase, 334
 tryptophan 5-halogenase, 495
 tryptophan 5-monoxygenase, 471
 tryptophan 6-halogenase, 495
 tryptophan 7-halogenase, 480
 tryptophan *N*-monoxygenase, 450
 tryptophan dehydrogenase, 215
Renilla-type luciferase, 334
 tyrosinase, 473
 tyrosine 3-monoxygenase, 471
 tyrosine *N*-monoxygenase, 415
 L-tyrosine isonitrile desaturase, 503
 L-tyrosine isonitrile desaturase/decarboxylase, 503
 L-tyrosine peroxygenase, 307
 L-tyrosine reductase, 146
 ubiquinol oxidase (non-electrogenic), 294
 UDP-2-acetamido-2,6- β -*L*-arabino-hexul-4-ose reductase, 81
 UDP-*N*-acetyl- α -D-quinovosamine dehydrogenase, 95
 UDP-*N*-acetyl-2-amino-2-deoxyglucuronate dehydrogenase, 73
 UDP-*N*-acetyl-D-mannosamine dehydrogenase, 74
 UDP-*N*-acetylglucosamine 3-dehydrogenase, 82
 UDP-*N*-acetylglucosamine 6-dehydrogenase, 30
 UDP-*N*-acetylmuramate dehydrogenase, 179
 UDP-glucose 6-dehydrogenase, 6
 UDP-glucuronic acid dehydrogenase (UDP-4-keto-hexauronic acid decarboxylating), 66
 ultra-long-chain fatty acid ω -hydroxylase, 456

unspecific monooxygenase, 405
 unspecific peroxygenase, 306
 uracil/thymine dehydrogenase, 534
 ureidoglycolate dehydrogenase, 34
 ureidoglycolate dehydrogenase (NAD⁺), 77
 uridine-5'-phosphate dioxygenase, 352
 urocanate reductase, 209
 uronate dehydrogenase, 43
 (*S*)-usnate reductase, 43

 validamycin A dioxygenase, 353
 valine *N*-monooxygenase, 416
 valine dehydrogenase (NAD⁺), 216
 valine dehydrogenase (NADP⁺), 212
 vanadium-dependent nitrogenase, 539
 vanillate monooxygenase, 379
 vanillin dehydrogenase, 138
 vanillyl-alcohol oxidase, 107
 vellosimine dehydrogenase, 58
 verruculogen synthase, 349
 versatile peroxidase, 300
 versiconal hemiacetal acetate reductase, 77
 very-long-chain 3-oxoacyl-CoA reductase, 72
 very-long-chain acyl-CoA dehydrogenase, 199
 very-long-chain acyl-lipid ω-9 desaturase, 500
 very-long-chain enoyl-CoA reductase, 178
 vinorine hydroxylase, 435
 violacein synthase, 398
 violaxanthin de-epoxidase, 552
 vitamin D 1,25-hydroxylase, 464
 vitamin D 25-hydroxylase, 412
 vitamin D₃ 24-hydroxylase, 463
 vitamin-K-epoxide reductase (warfarin-insensitive), 529
 vitamin-K-epoxide reductase (warfarin-sensitive), 529
 vomifoliol dehydrogenase, 47
 vomilenine reductase, 234

 xanthine dehydrogenase, 524
 xanthine dioxygenase, 352
 xanthine oxidase, 527
 xanthommatin reductase, 167
 xanthoxin dehydrogenase, 62
 D-xylose 1-dehydrogenase, 37
 L-xylose 1-dehydrogenase, 25
 D-xylose 1-dehydrogenase (NADP⁺, D-xylono-1,4-lactone-forming),
 94
 D-xylose 1-dehydrogenase (NADP⁺, D-xylono-1,5-lactone-forming),
 38
 D-xylose reductase (NADH), 96
 D-xylose reductase (NADPH), 96
 D-xylose reductase [NAD(P)H], 66
 D-xylulose reductase, 3
 L-xylulose reductase, 3

 zealexin A1 synthase, 451
 zeatin reductase, 172
 zeaxanthin 4-ketolase, 516
 zeaxanthin epoxidase, 464
 zerumbone synthase, 71